## Electronic Supplementary Information Exploring a Direct Injection Method for Microfluidic Generation of Polymer Microgels

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#### 1. Materials

SU-8 50 photoresist was supplied from MicroChem Corp. (Newton, MA, USA). Poly(dimethyl siloxane) (PDMS; Sylgard 184) was obtained from Dow Corning (Midland, MI, USA). The ultra-low gelling temperature agarose (SeaPrep) was obtained from Lonza Rockland Inc. (Rockland, ME, USA). Phosphate buffered saline (PBS), mineral oil, sorbitan monooleate (Span 80) and methyl- $\beta$ -cyclodextrin (MCD) were supplied by from Sigma (St. Louis, MO, USA). Sodium dodecyl sulfate (SDS) and glycerol were obtained from Sigma-Aldrich Co. (St. Louis, MO, USA). ABIL EM90 was donated by Evonic Goldschmidt Corp. (Hopewell, VA, USA). Poly(ethylene glycol) end-terminated with octadecyl unimer groups (PEG-ODU; molecular weight 20 kg/mol) was synthesized and purified as described elsewhere.<sup>1,2</sup>

#### 2. Dimensions of channels in the microfluidic devices

We used MF devices with a similar design for the preparation of agarose and PEG-ODU microgels. The height of the microchannels was 60  $\mu$ m. The dimensions of the microchannels were W<sub>o</sub>=67  $\mu$ m, W<sub>a1</sub>=60  $\mu$ m, W<sub>s</sub>=100  $\mu$ m and W<sub>o</sub>'=100  $\mu$ m

(Fig. S1). The widths of the side channels used for direct injections were  $W_{a2}$ =100 µm (Fig. S1a) and  $W_{a2}$ '=30 µm (Fig. S1b) for the preparation of agarose microgels and PEG-ODU microgels, respectively.



**Fig. S1**. Design of MF device used for the preparation of microgels. (a) and (b) shows the widths of the injection channels used for the preparation of agarose microgels and PEG-ODU microgels, respectively.

#### 3. Viscosity of agarose solutions



**Fig. S2.** Variation in viscosity of agarose solutions, plotted as a function of weight concentration of agarose. The shear rate was  $125 \text{ s}^{-1}$ , the temperature is  $37 \text{ }^{\circ}\text{C}$ .

#### 4. Size distribution of agarose microgels

Experiment was conducted as  $C_1=2$  wt.%;  $C_2=5$  wt.%; and  $Q_i$ ,  $Q_{ii}$  and  $Q_{iii}$  of 0.1, 0.05 and 0.05 mL/hr, respectively. Injection was conducted at  $Q_{iii}=0.05$  mL/hr, at which the maximum volume of  $1.85 \times 10^5 \,\mu\text{m}^3$  of 5 wt.% agarose solution was injected to the primary plugs (at  $V_2/V_3=0.28$ ).  $D_d$  is the diameter of the microgels. The data points of  $D_d$  were fitted with Gaussian distribution. The polydispersity of the microgels was 3.4%.



Fig. S3. Distribution of diameters of agarose microgels

# 5. Viscosity measurements of MCD+PEG-ODU and SDS+MCD+PEG-ODU solutions

Viscosities of mixed solutions PEG-ODU+MCD at different  $\alpha$  (defined as molar ratio between MCD and PEG-ODU) were measured by adding a particular amount of MCD (solid) to a PEG-ODU solution, while maintaining the concentration of PEG-ODU at 8 wt.%. The solution was mixed (for 10 min at 37 °C) to achieve complete MCD dissolution.

Similarly, viscosities of PEG-ODU+MCD+SDS mixtures in water at different  $\beta$  (defined as the molar ratio of SDS-to-MCD) were measured by adding a particular amount of SDS (solid) to the solution of PEG-ODU and MCD, while maintaining the constant concentrations of PEG-ODU and MCD at 4 wt.% and 0.5 wt.%, respectively. After addition of SDS, the system was mixed for 10 min to achieve full SDS dissolution.

#### 6. Videos of injections of agarose and SDS solutions in primary plugs

The scale bars in the videos of injections of agarose and SDS solutions are 200 and  $100 \ \mu m$ , respectively.

#### **Notes and References**

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