

Supporting Information

Modular Synthesis of Biologically Active Phosphatidic Acid Probes Using Click Chemistry

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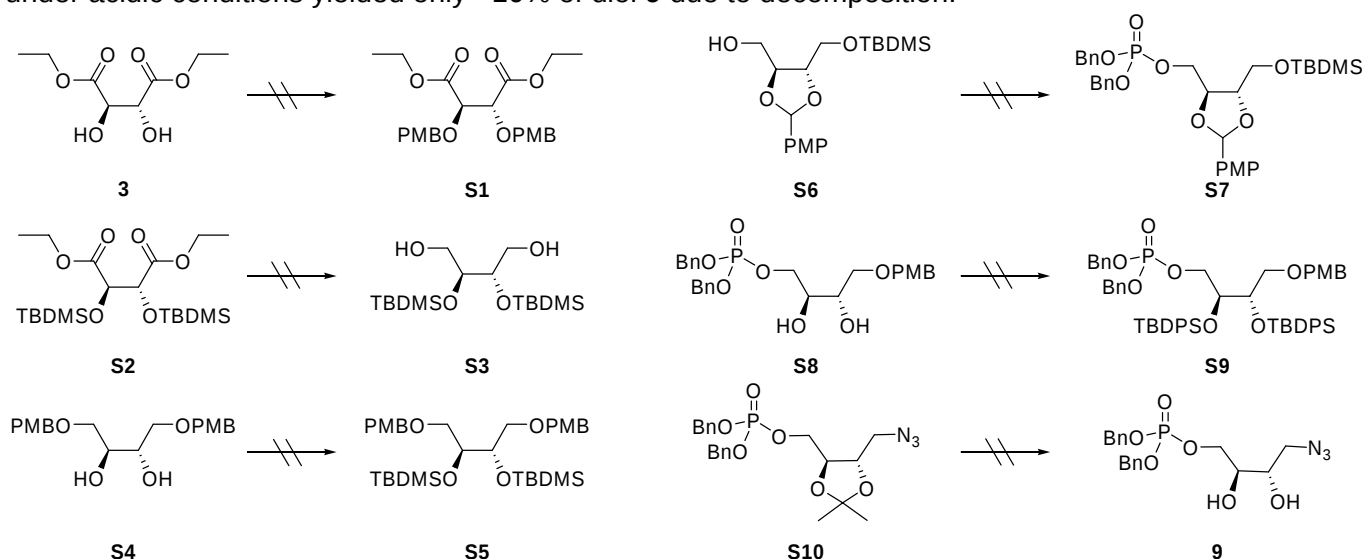
^cDepartment of Chemistry and Biochemistry and the Walther Center for Cancer Research, University of Notre Dame, Notre Dame, IN 46656

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I. Protecting Group Strategies Explored for the Interior Diol of Scaffold 2

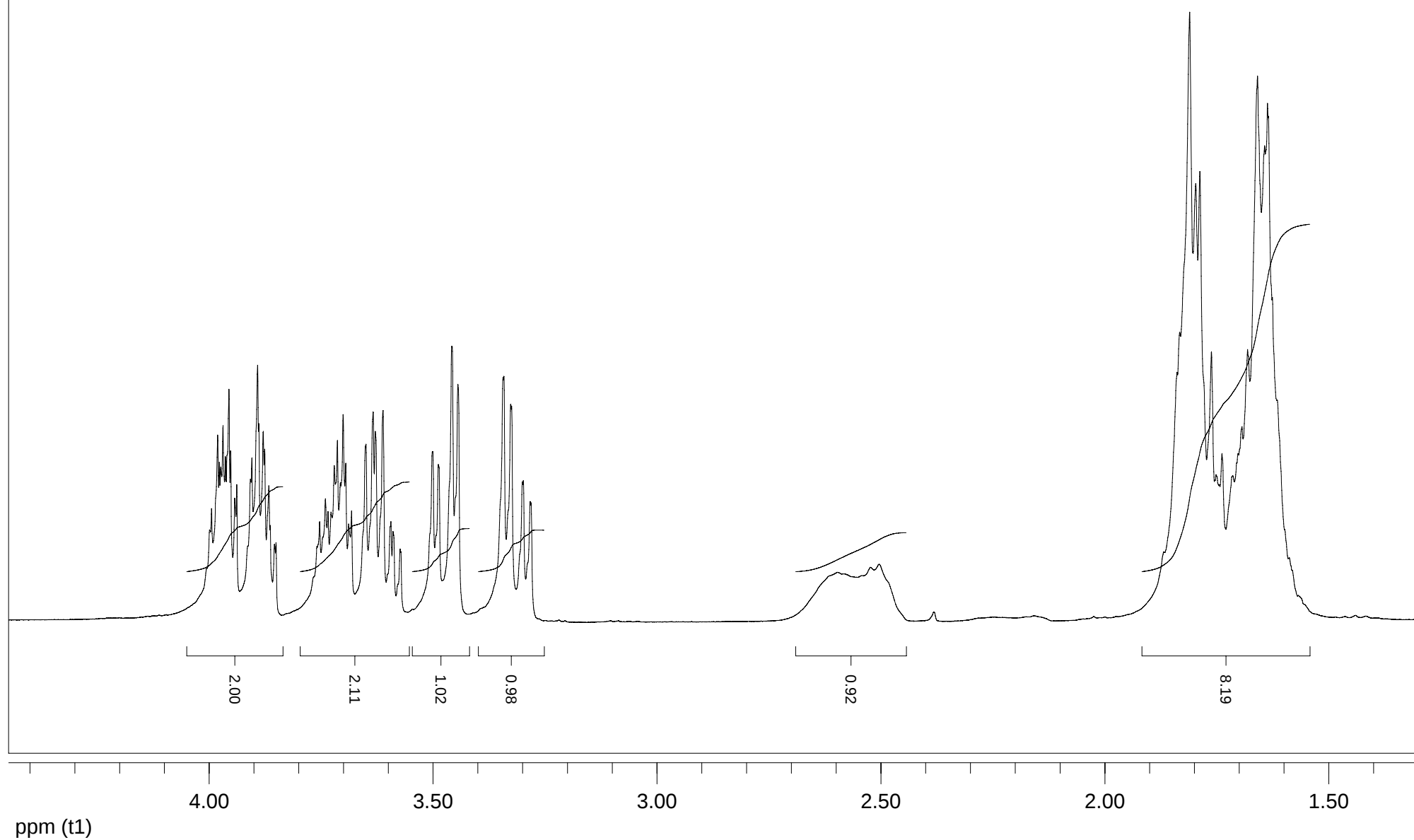
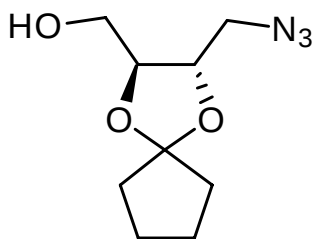
A key aspect of this synthesis was the determination of the optimal protecting group for the interior diol of **8** that could be removed without affecting the phosphotriester and azide groups. Several approaches were explored to determine that which is most effective. These routes are outlined in Scheme S1, in which the steps that were unsuccessful are indicated. First, the installation of two *p*-methoxybenzyl protecting groups onto the diol of **3** to **S1** was ineffective. While two silyl protecting groups could be introduced following literature procedures^{1,2} to **S2**, these were removed during ester reduction in attempting to access **S3**. However, the introduction of two silyl groups onto the diol of **S4** to access **S5** was unsuccessful. While *p*-methoxybenzylidene-protected derivative **S6** was synthesized, all oxidation conditions attempted for the production of phosphotriester **S7** led to removal of this group. Bis-silylation of **S8** to **S9** was once again unsuccessful. Finally, deprotection of the acetonide group of **S10** under acidic conditions yielded only ~10% of diol **9** due to decomposition.

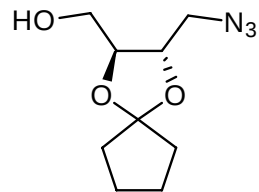


Scheme S1. Protecting group strategies explored in the synthesis of modular PA core 2.

References Cited

- 1 J. E. Baldwin; R. M. Adlington; A. T. Russell; M. L. Smith. *J. Chem. Soc., Chem. Commun.* 1994, 85-86.
- 2 T. Hiayama; T. Minami; K. Takahashi. *Bull. Chem. Soc. Japan* 1995, **68**, 364-372.





119.828

78.543

76.066

62.035

52.011

37.291

37.211

23.496

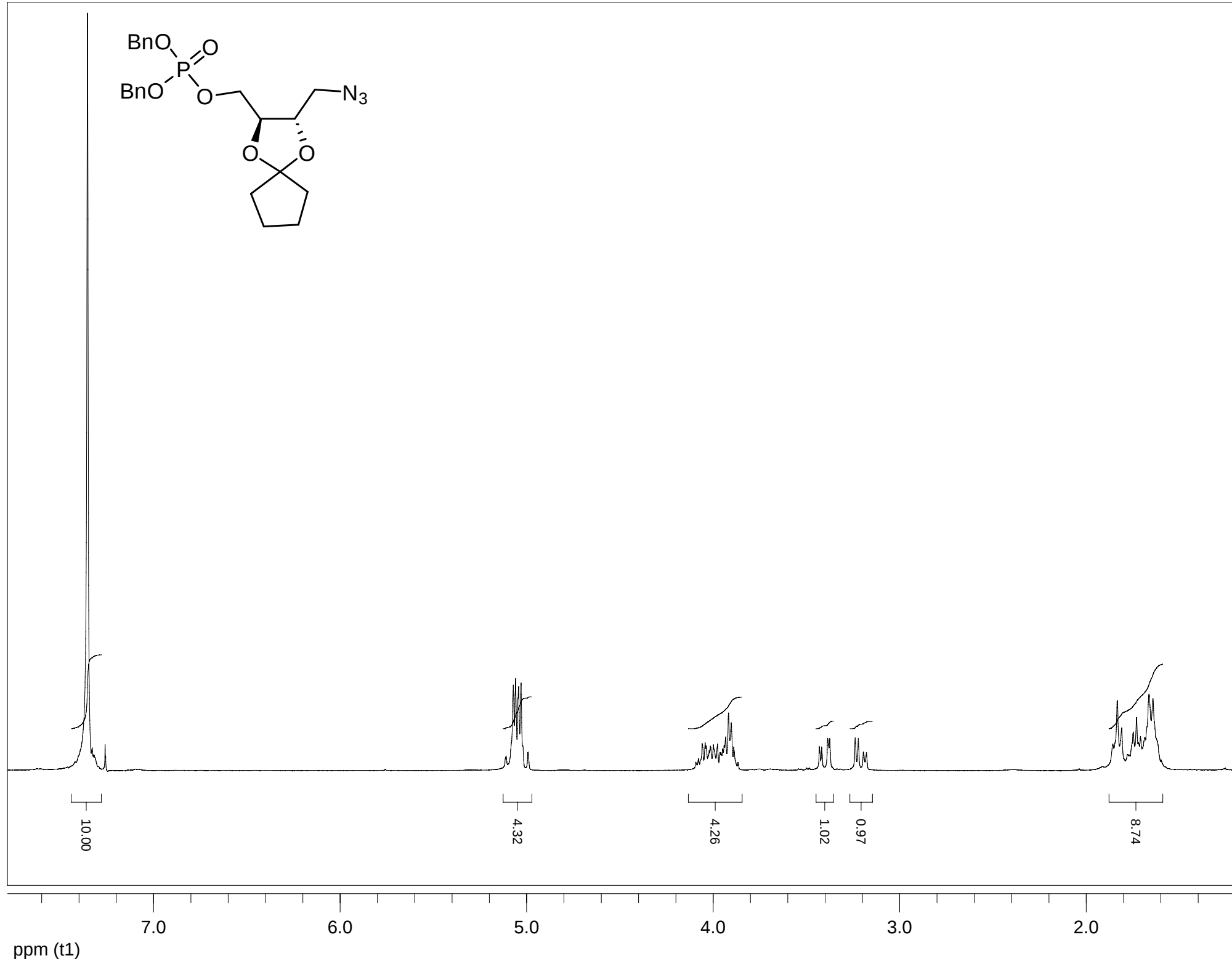
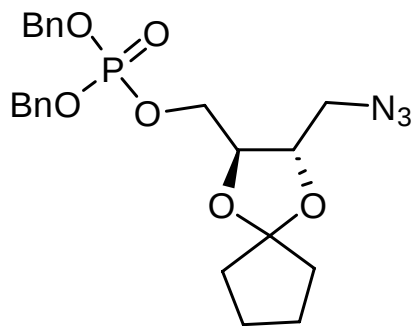
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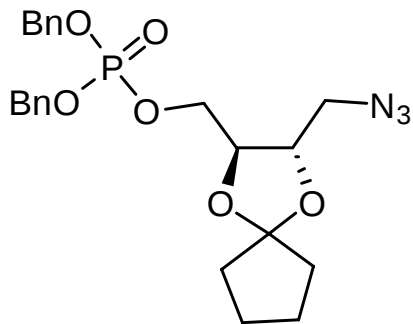
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50

0



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135.557
128.677
128.626
128.038
120.234



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75.855

69.547
69.474
66.673
66.598

51.788

37.278
37.086

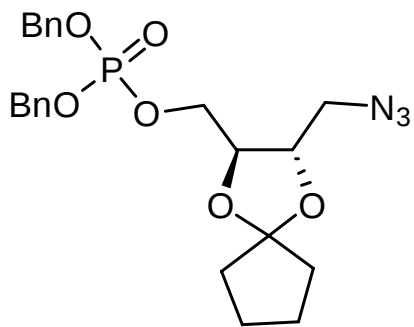
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23.360

0 S5

ppm (t1)

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50



0.143

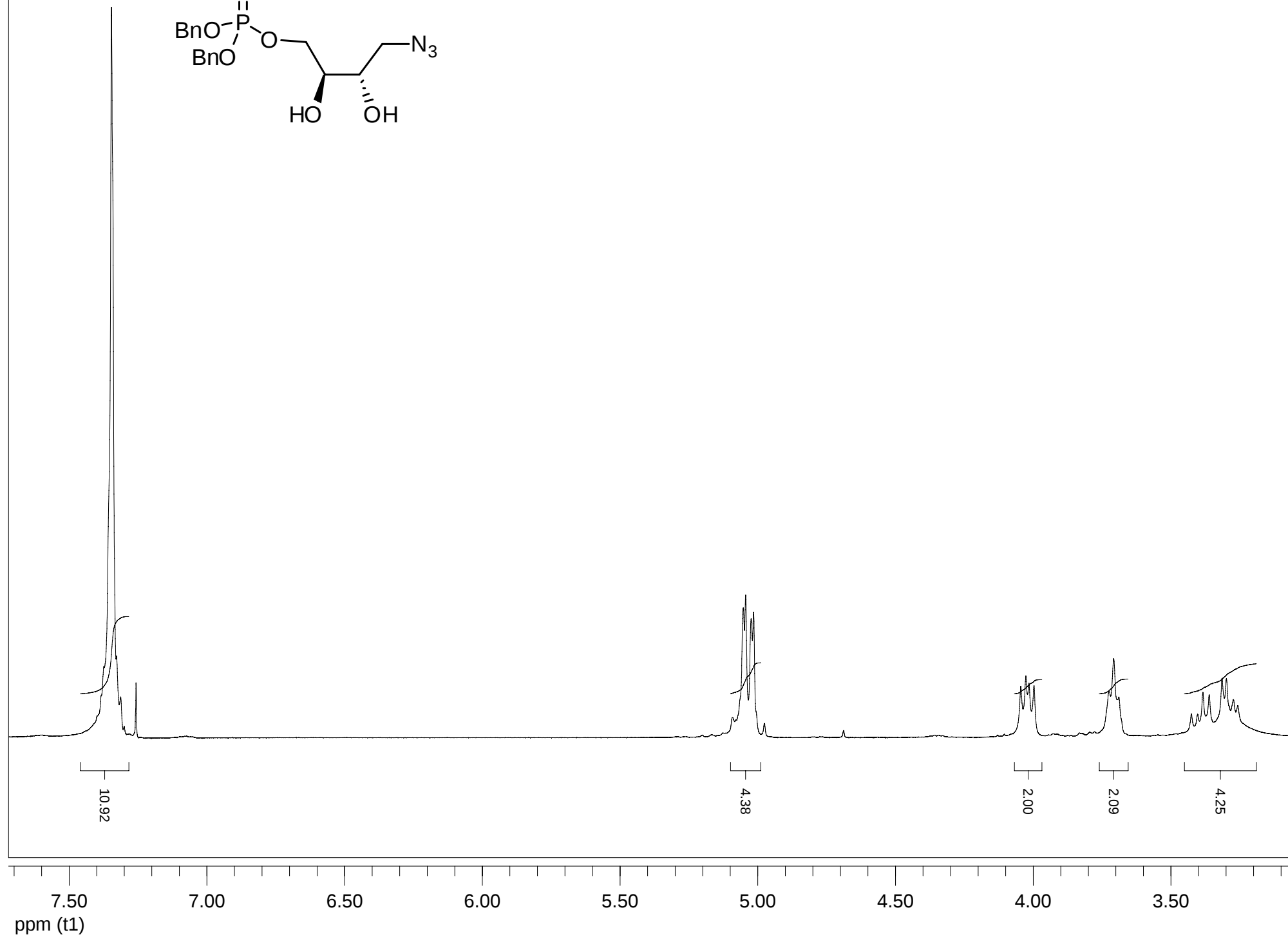
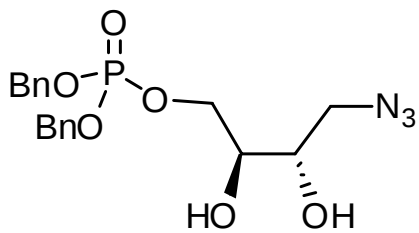
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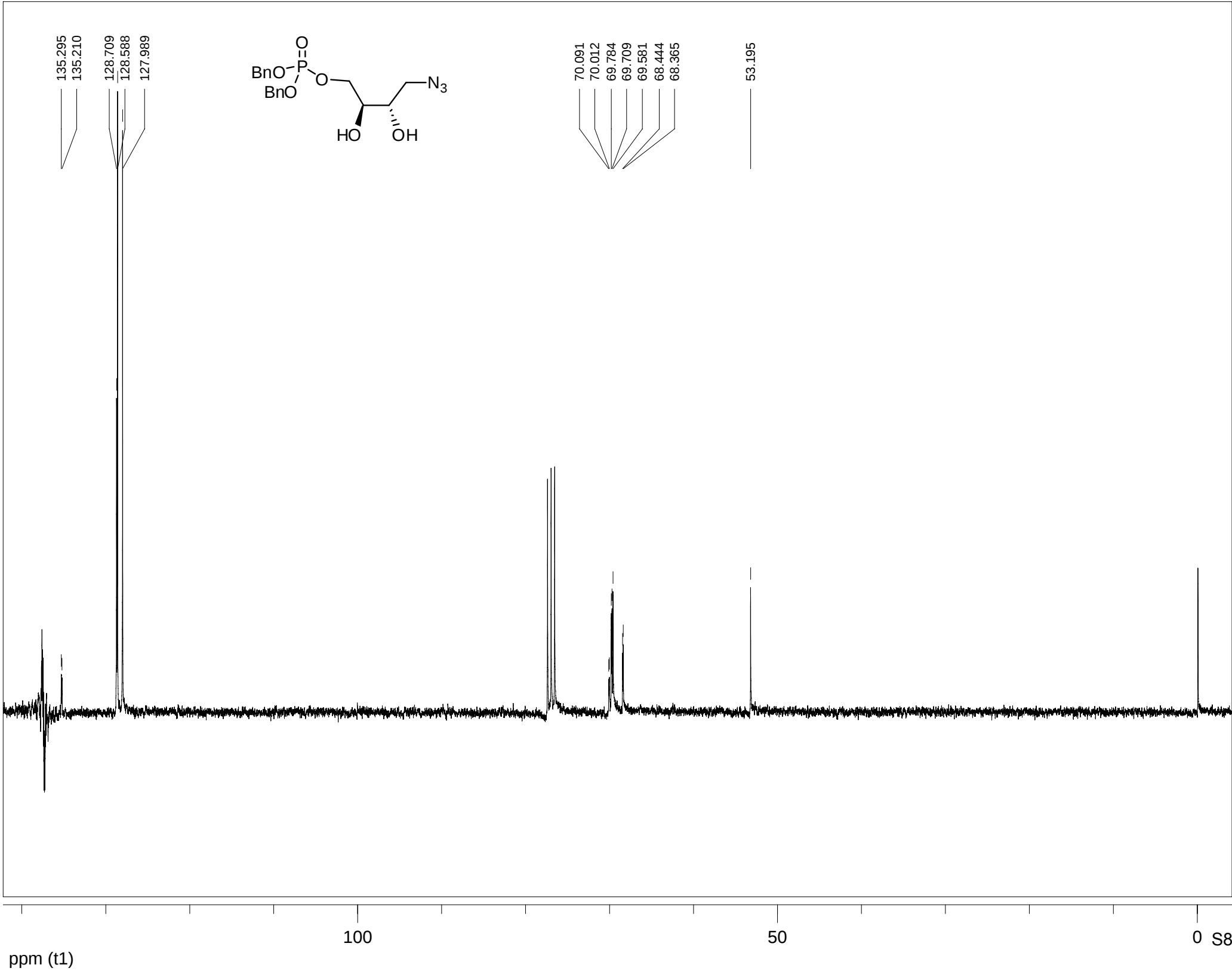
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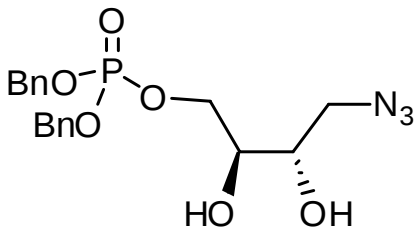
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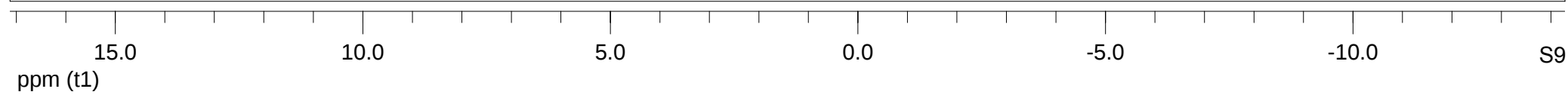
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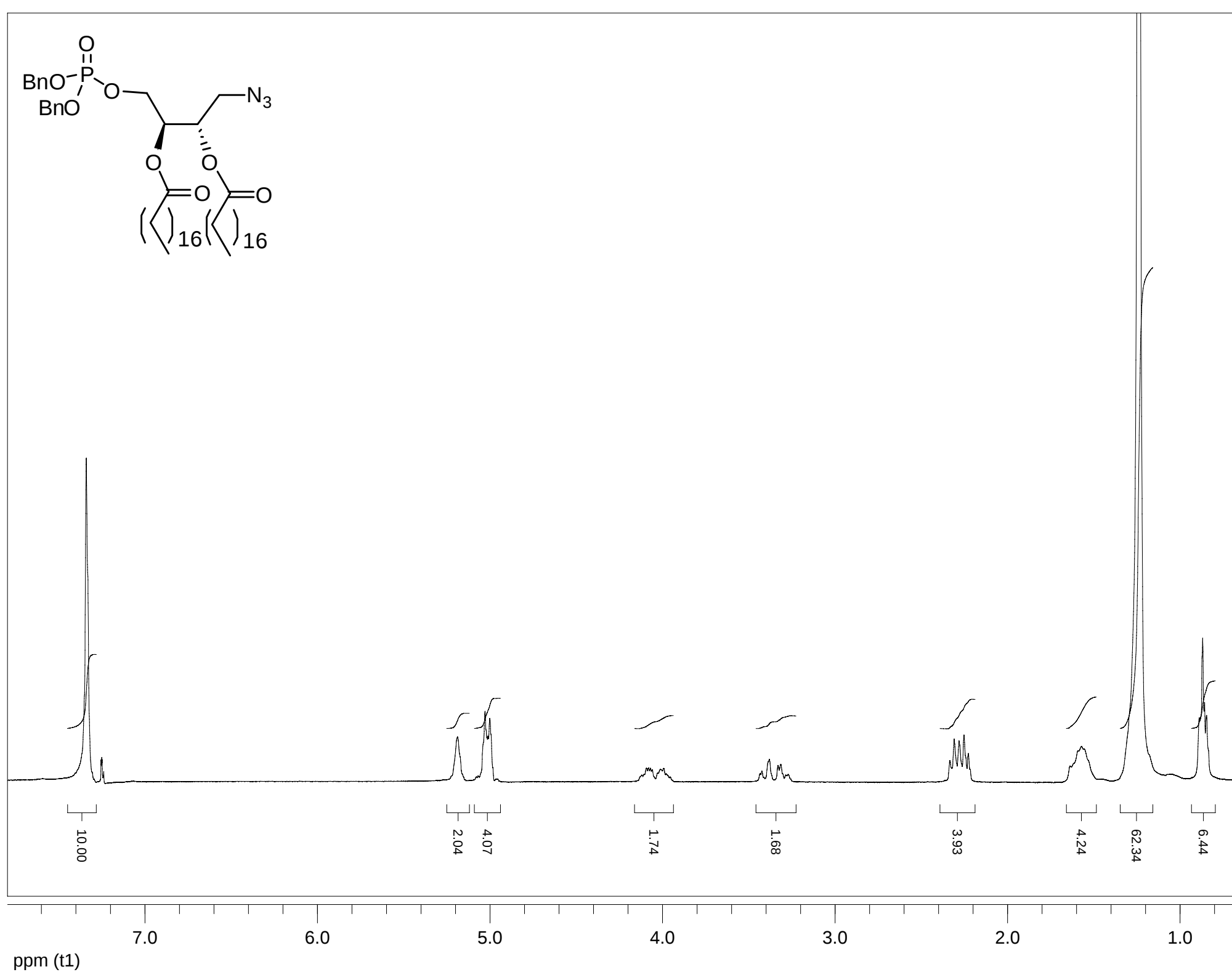
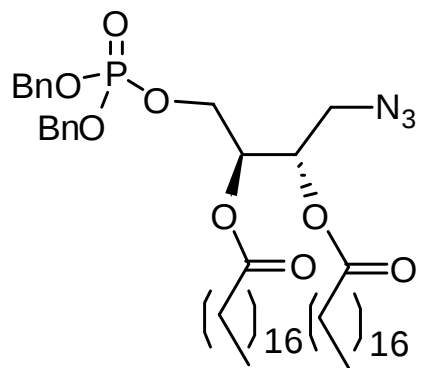


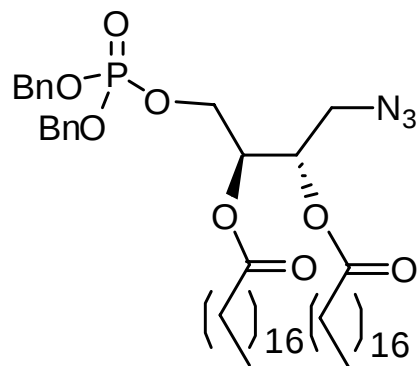




0.923







172.667
172.616

135.594
135.505

128.664
128.631
128.022
127.975

69.984
69.879
69.733
69.609
69.534
65.084
65.014

50.556

34.041
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29.123
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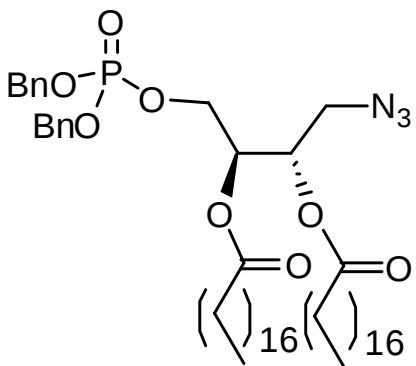
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150

100

50

0 S11



0.041

ppm (t1)

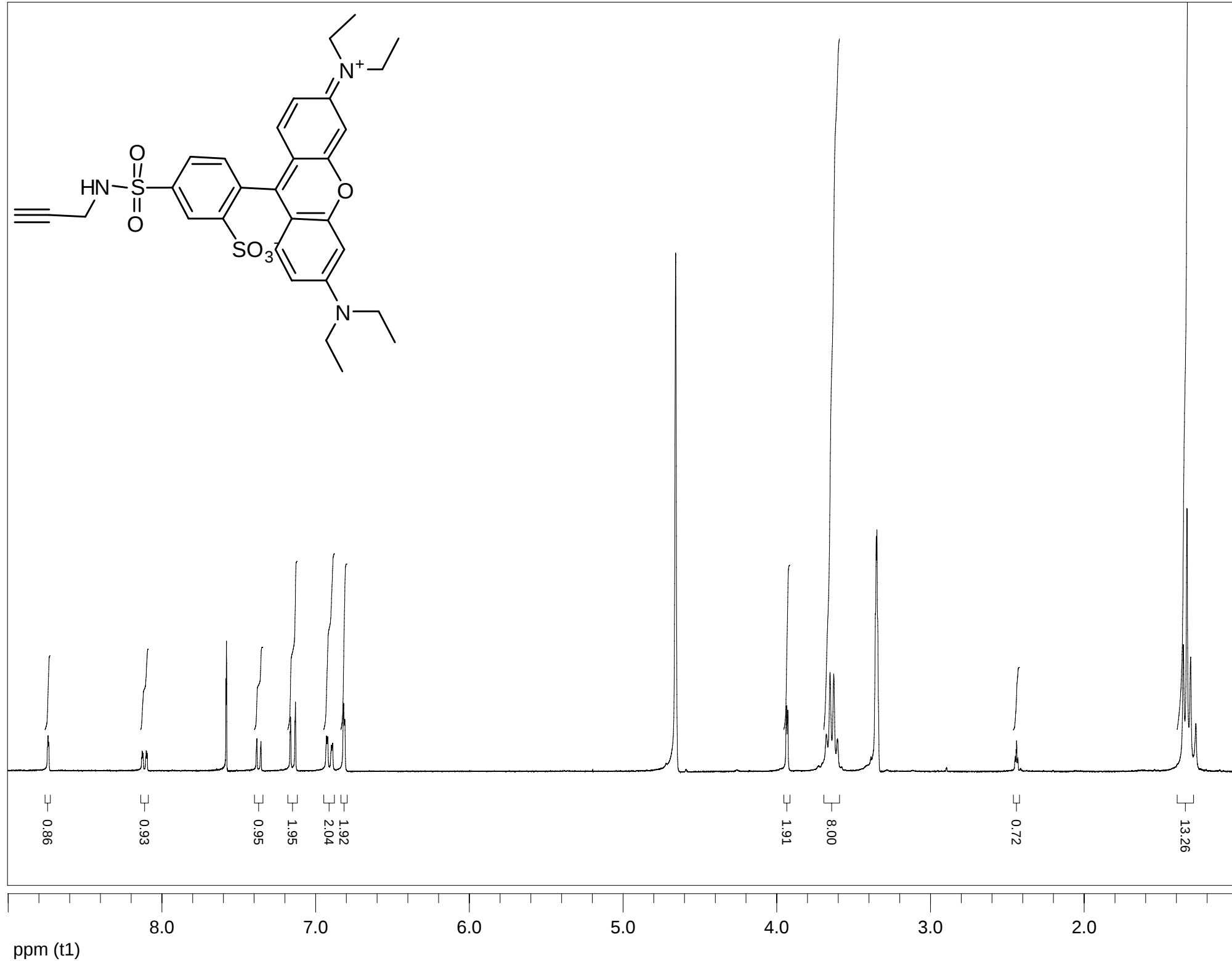
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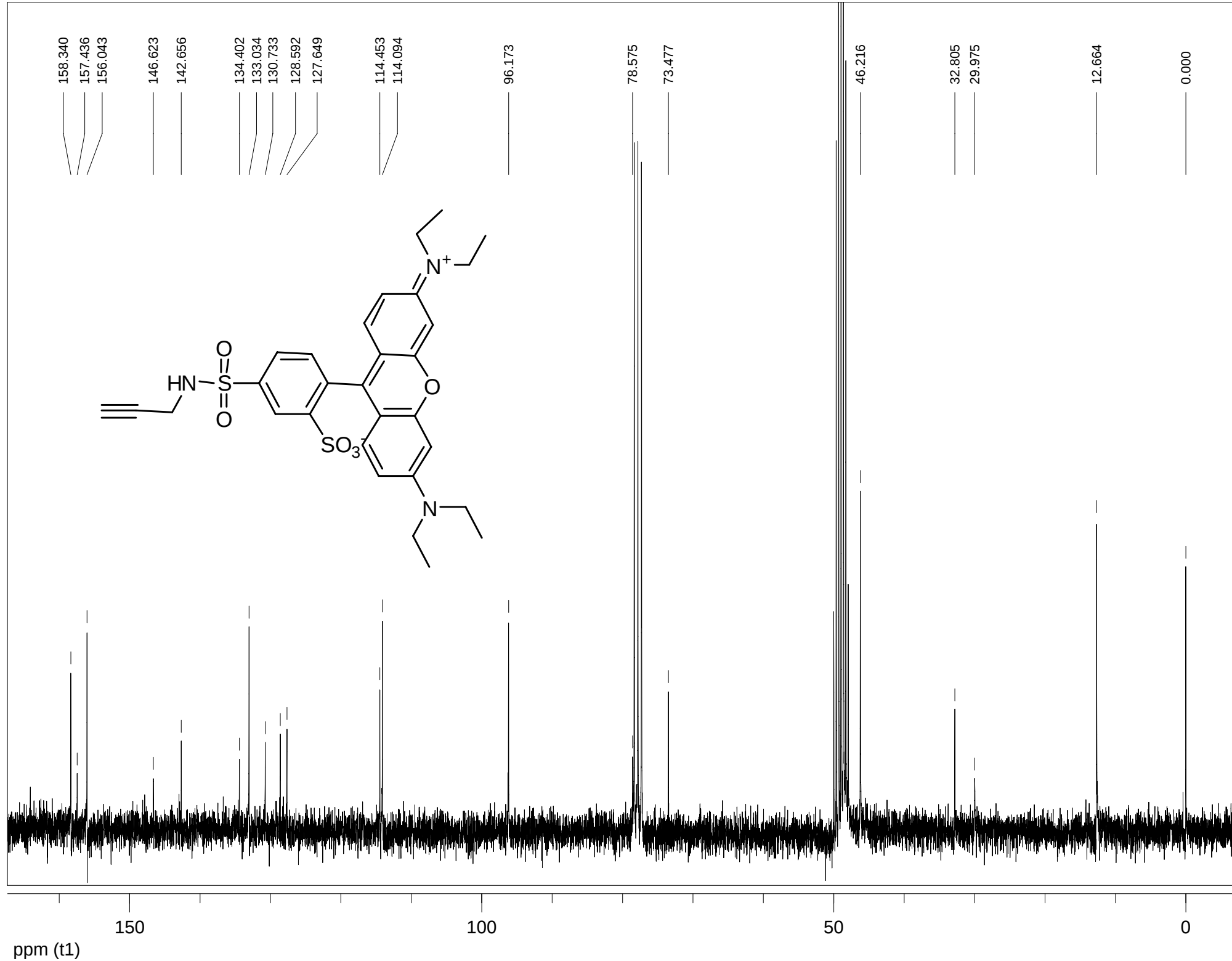
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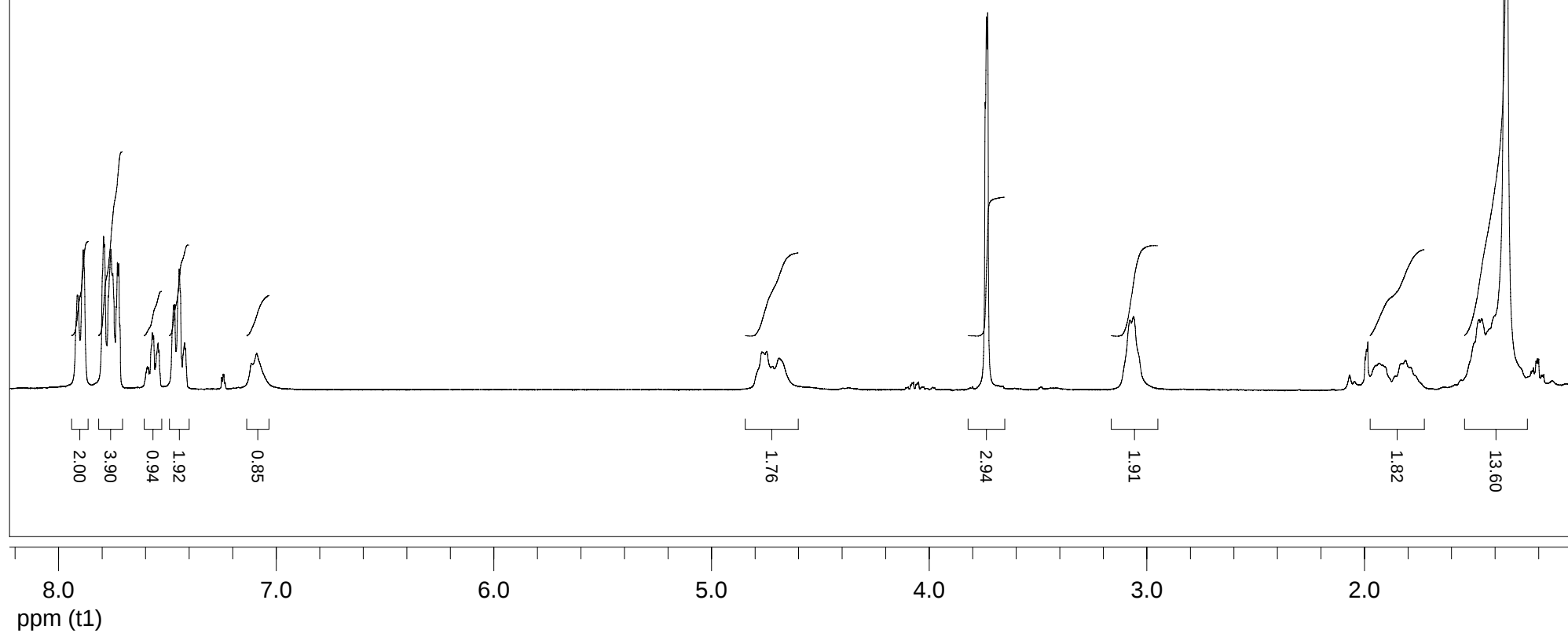
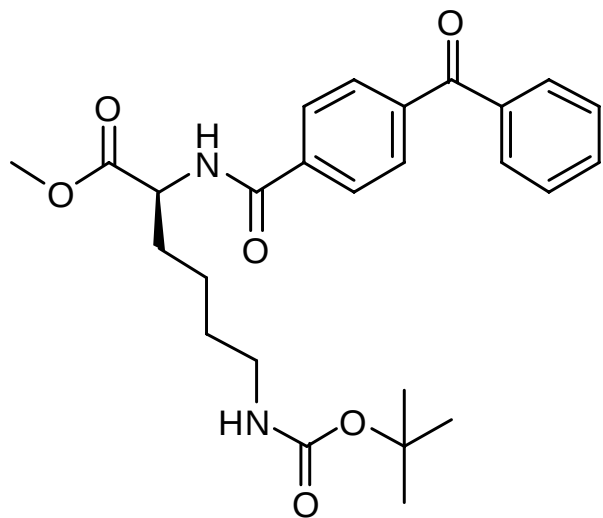
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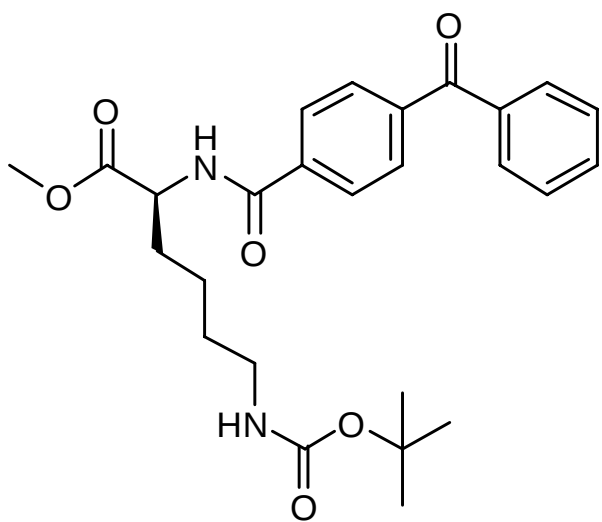
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S12









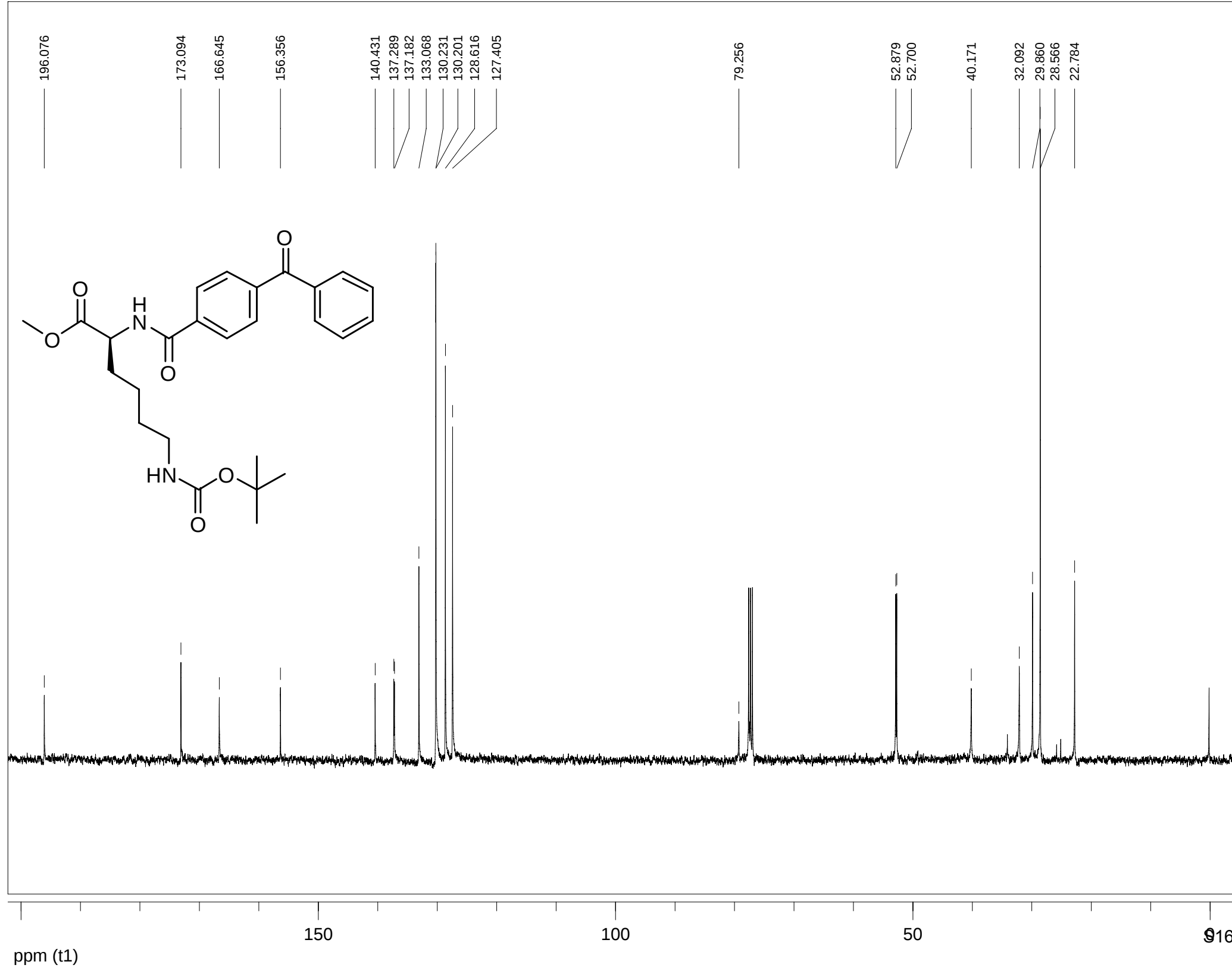
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127.405

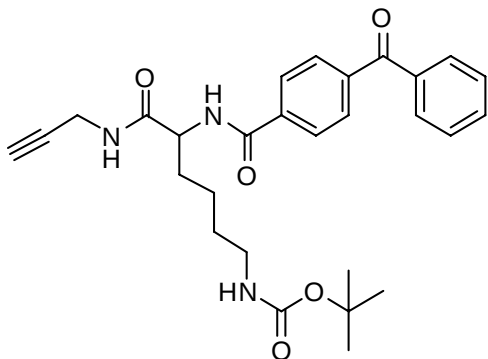
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52.700

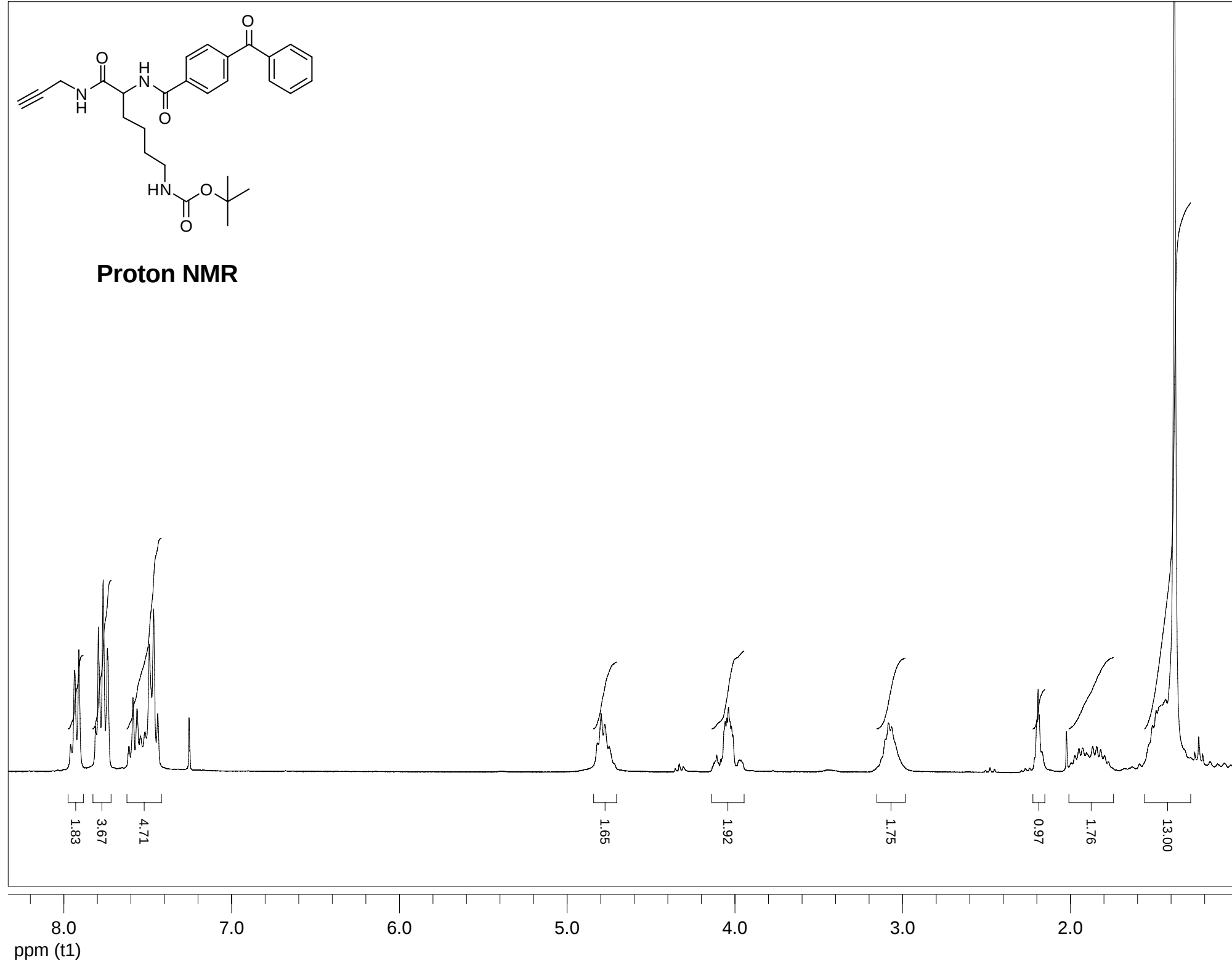
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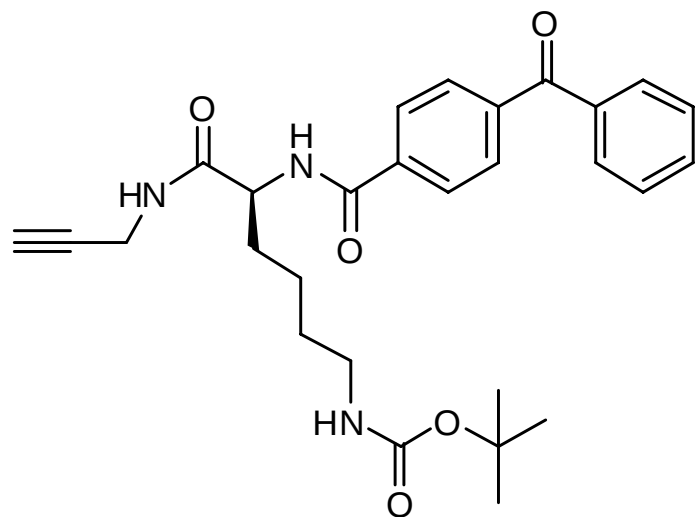
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29.860
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22.784





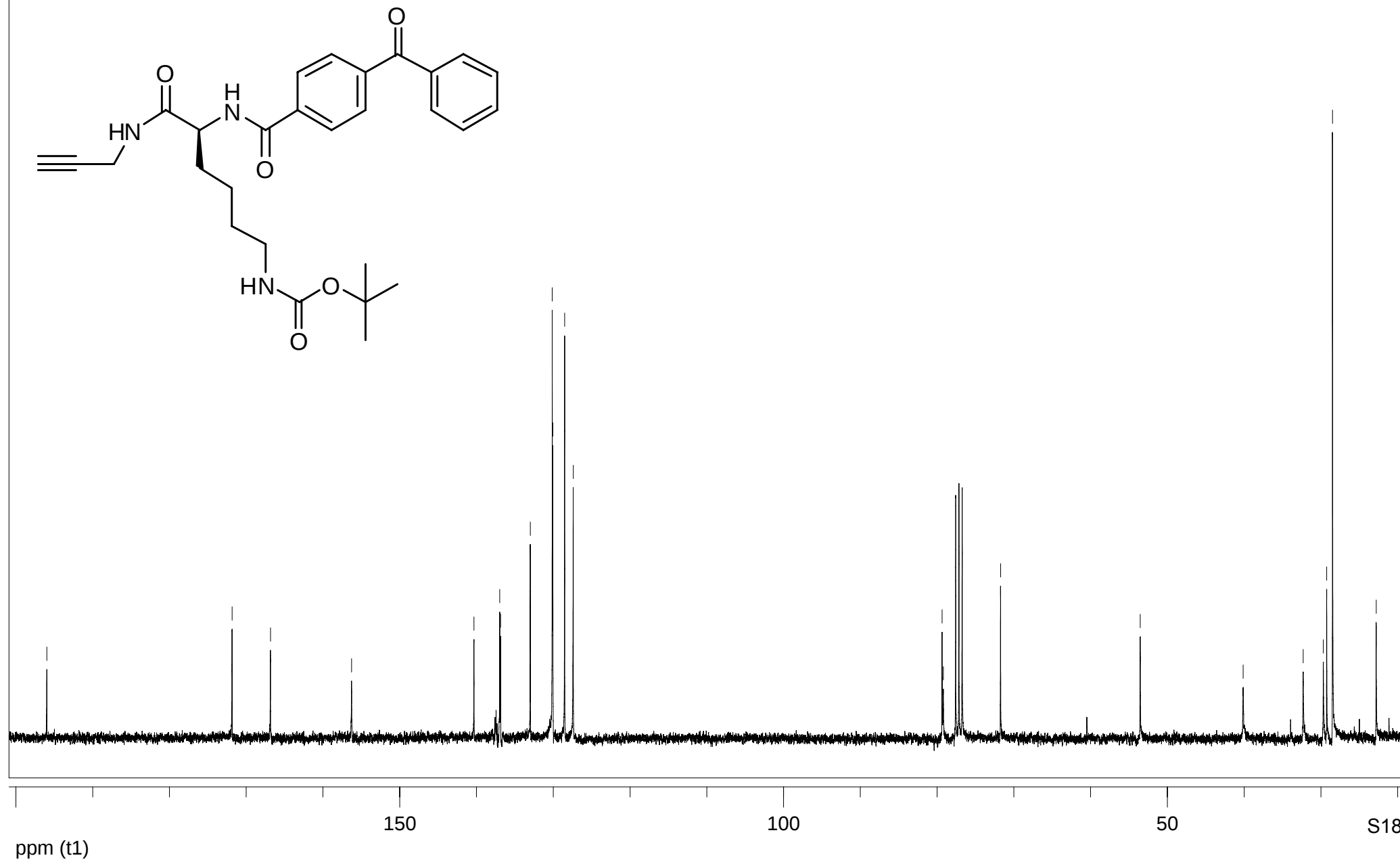
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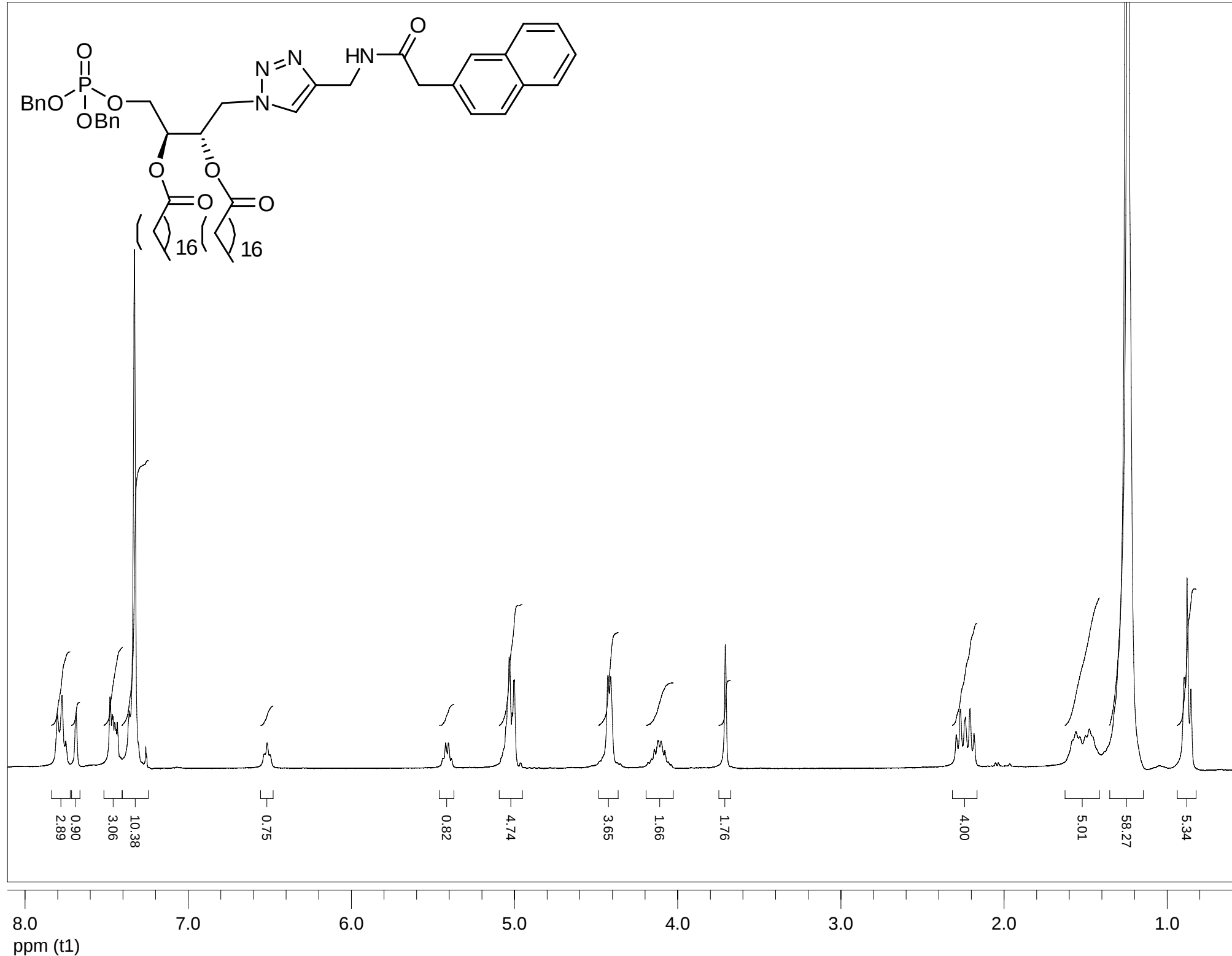
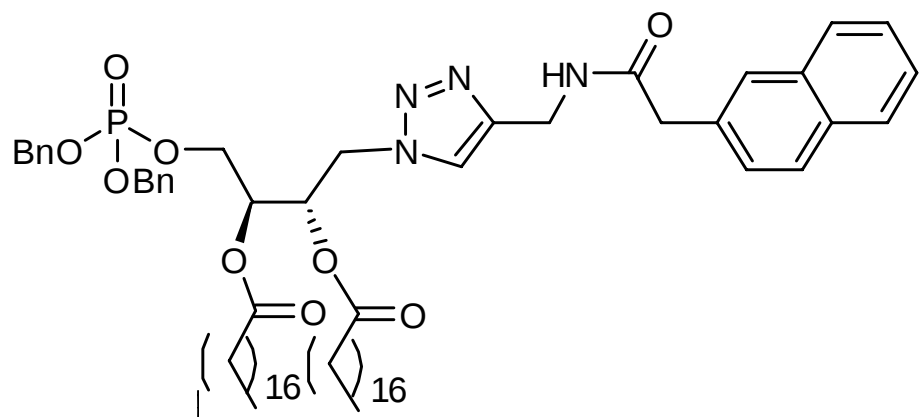


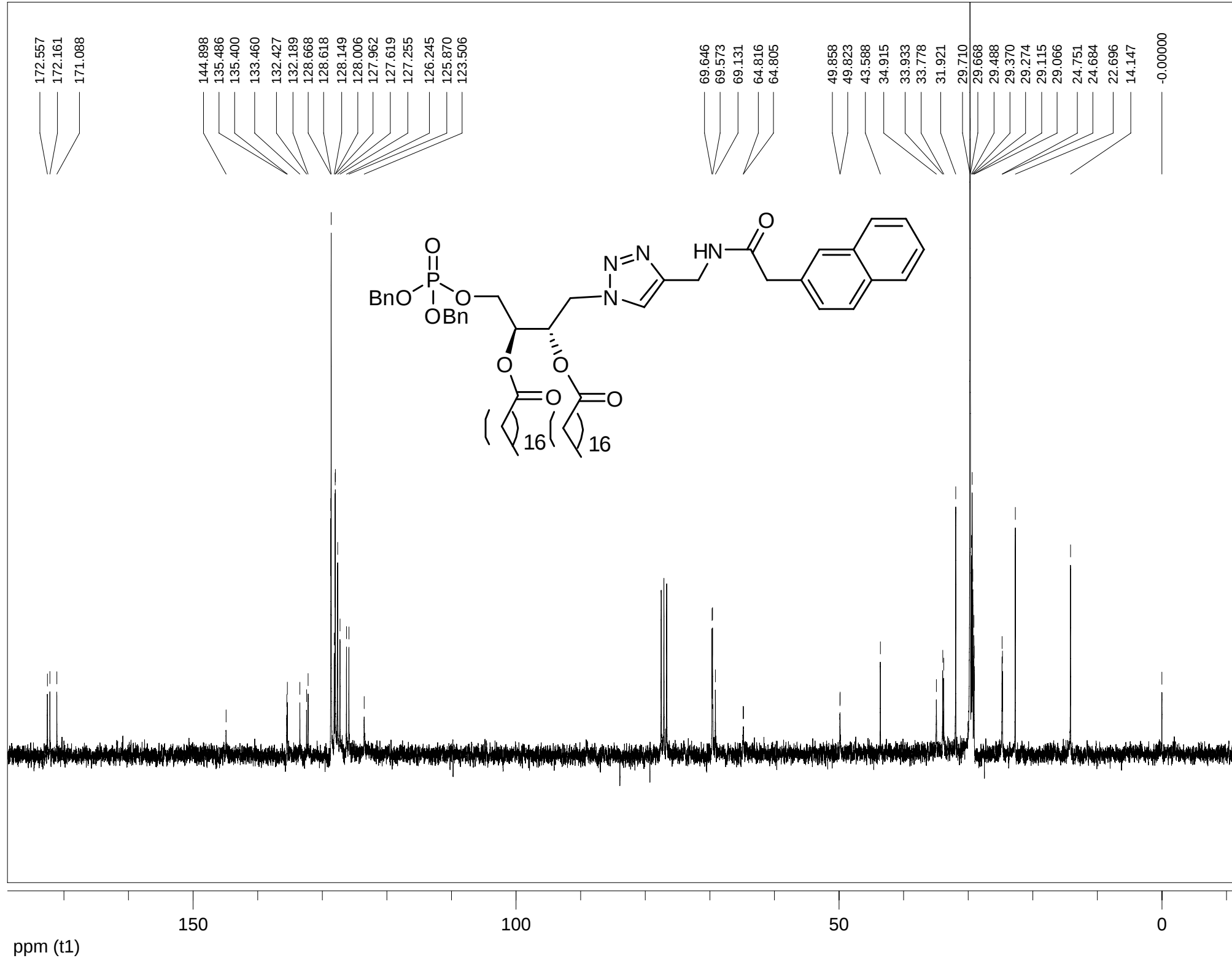


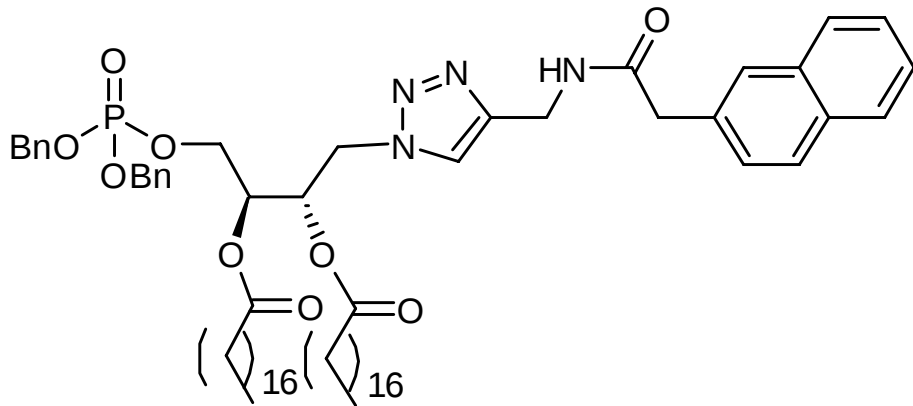
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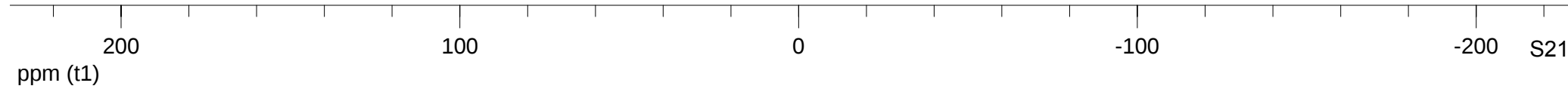


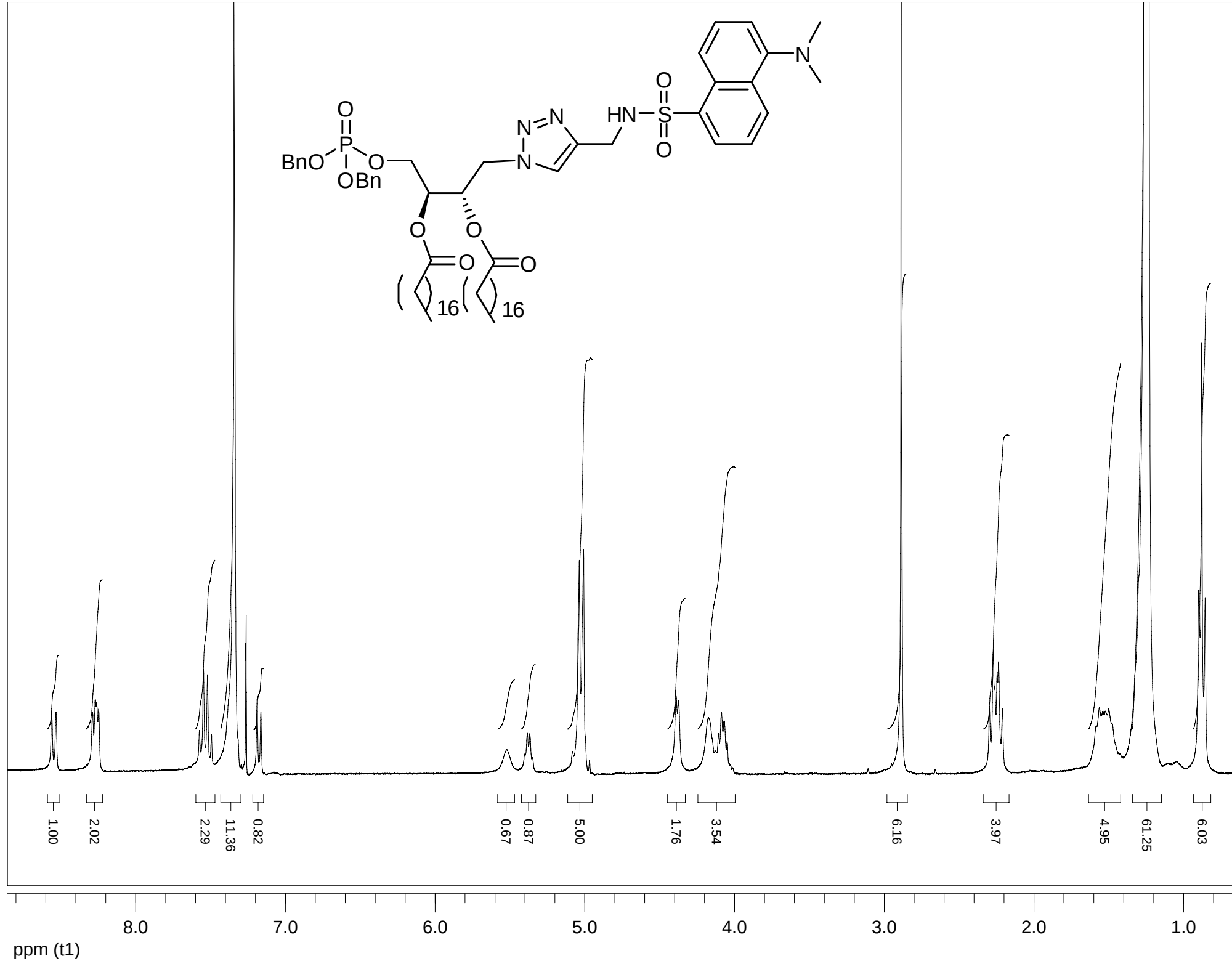
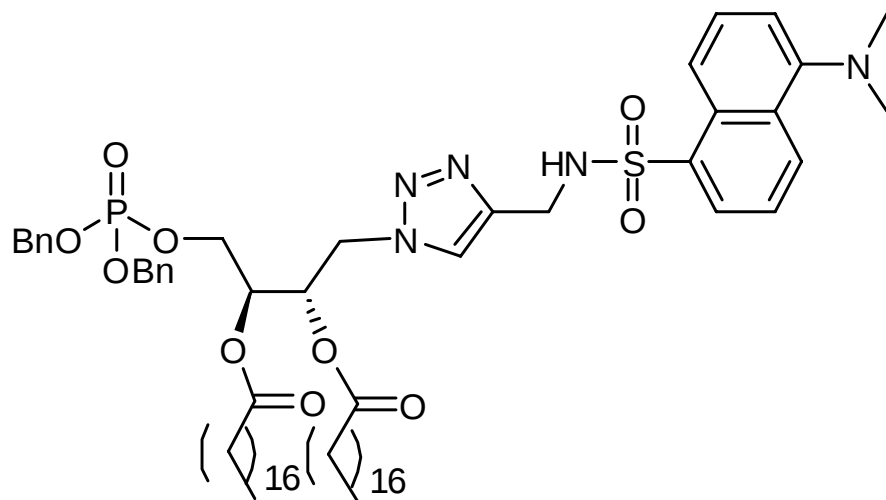






0.005





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172.352

151.974

144.489

135.623

135.535

134.707

130.580

129.936

129.673

129.568

128.751

128.715

128.578

128.108

128.068

123.526

123.216

118.920

115.341

69.758

69.684

69.156

65.010

64.942

49.909

45.486

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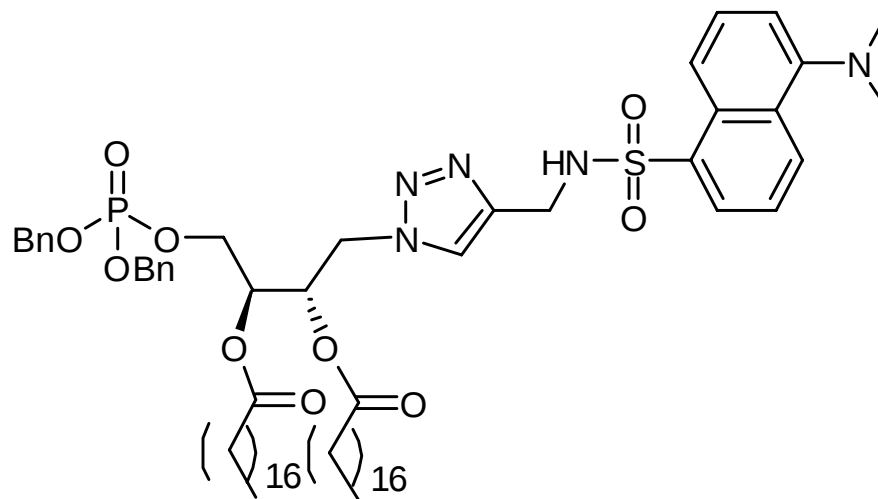
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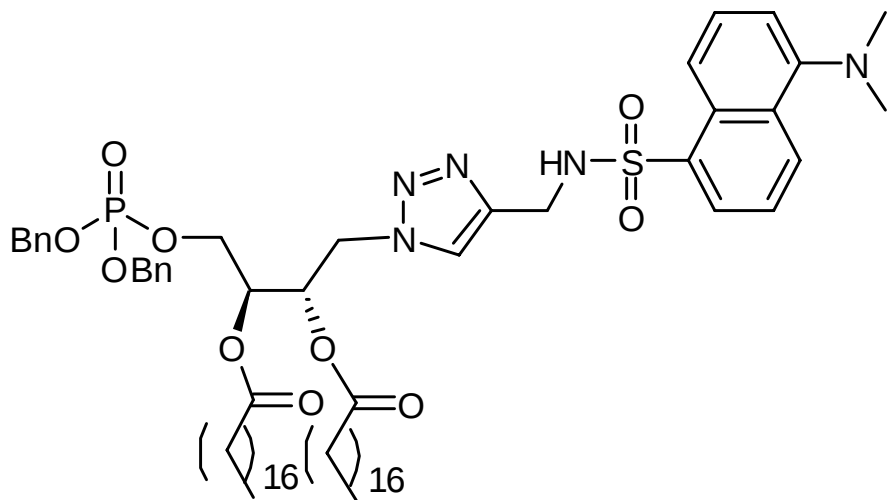
150

100

50

S23

ppm (t1)



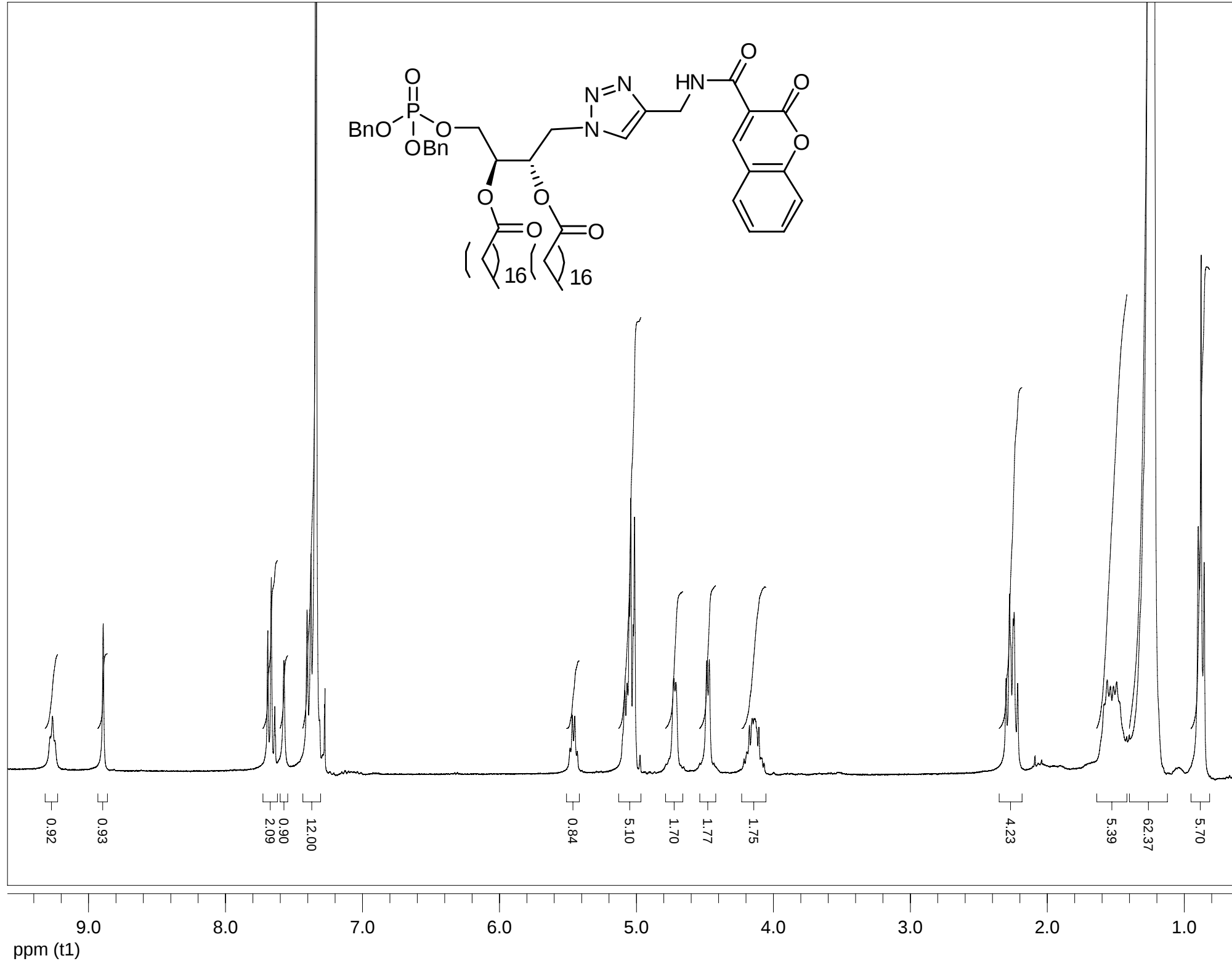
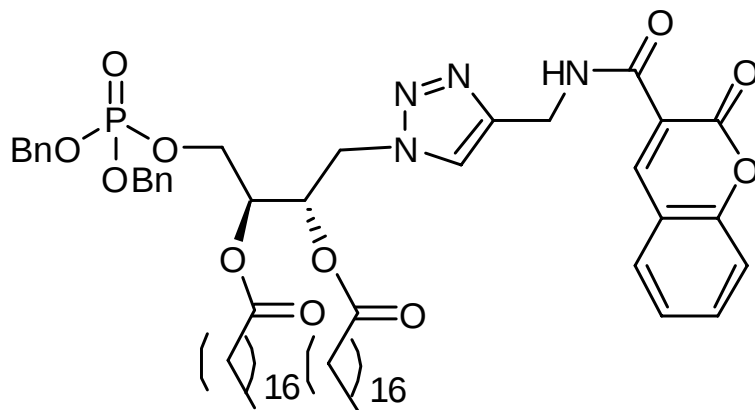
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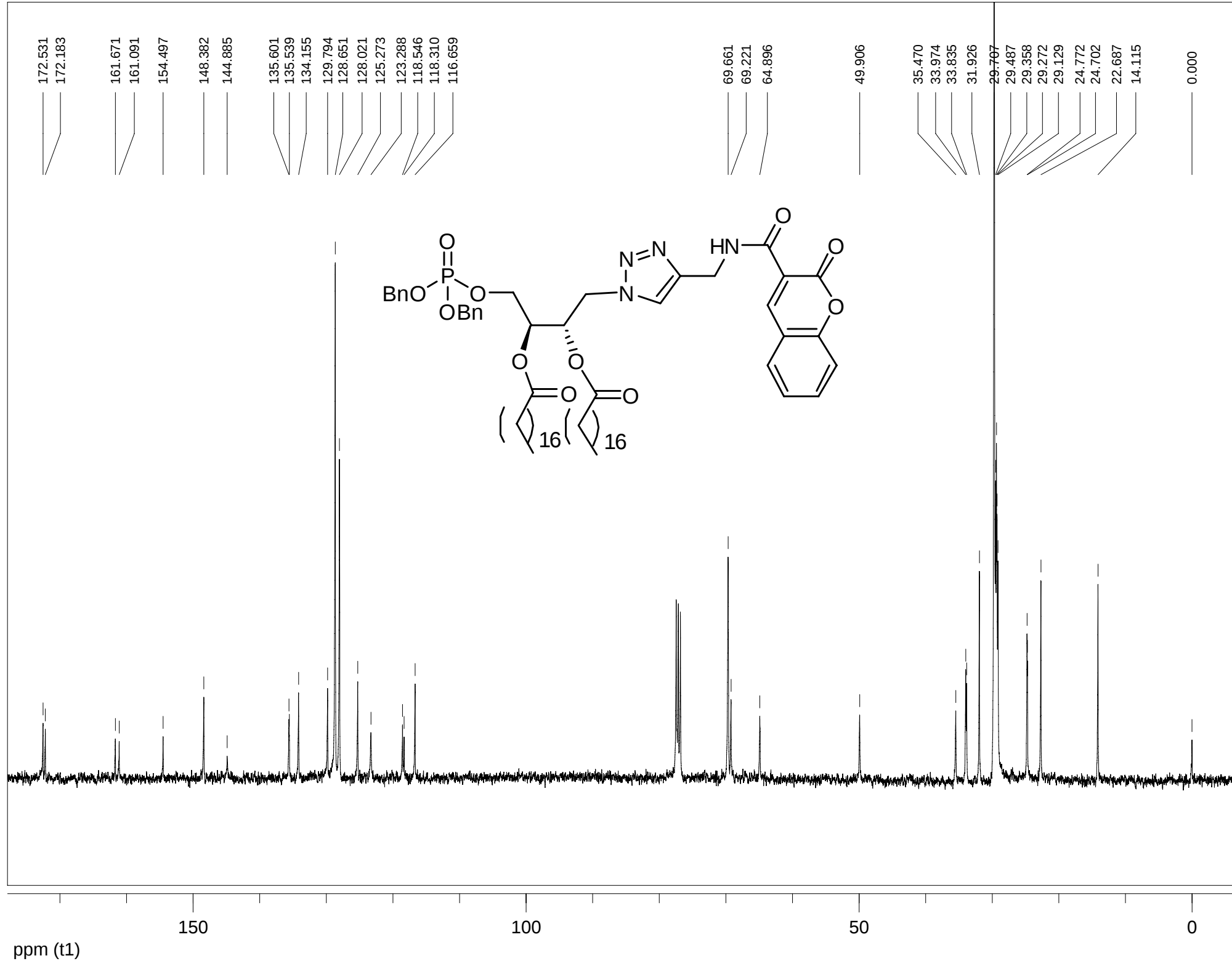
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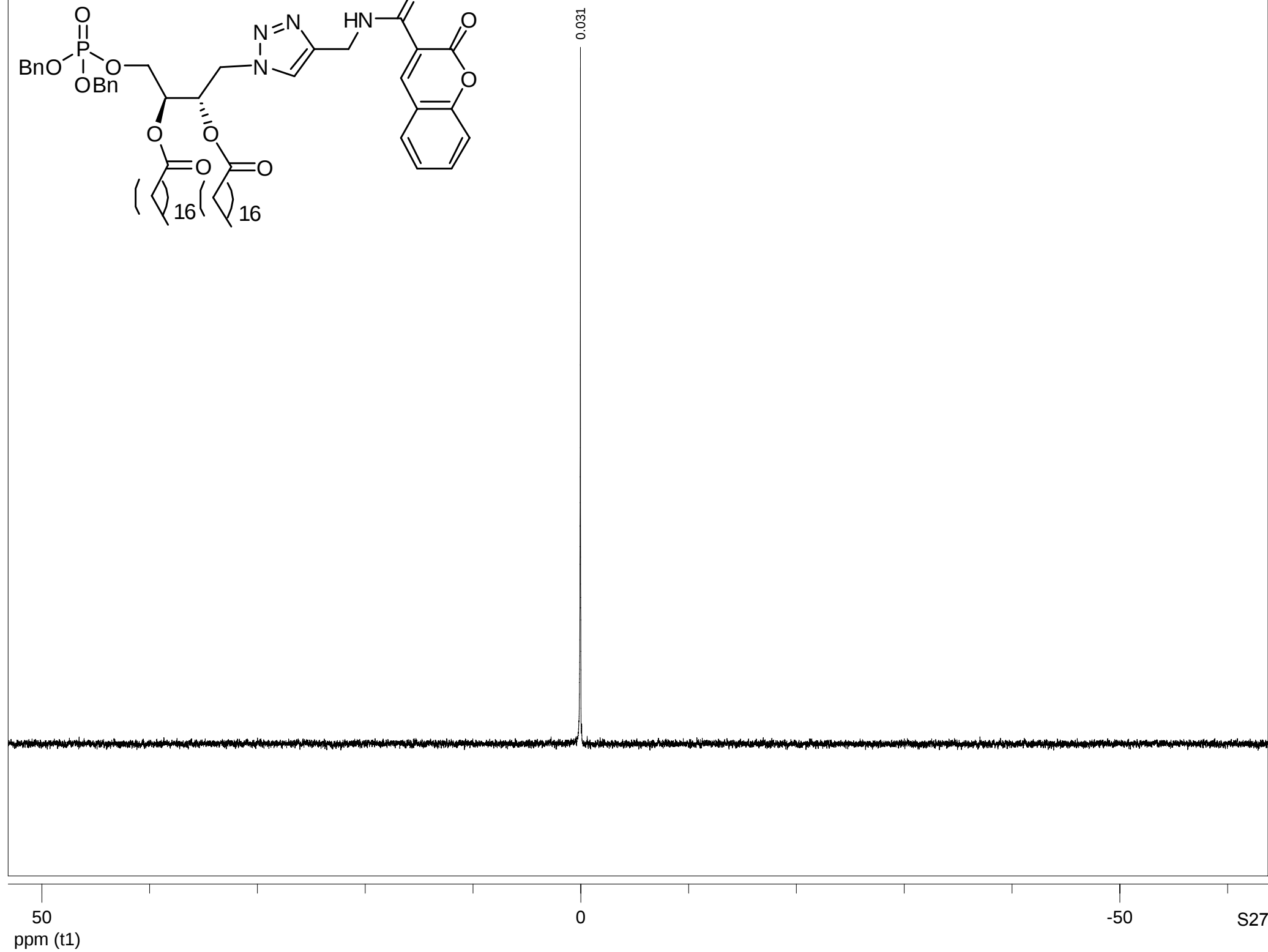
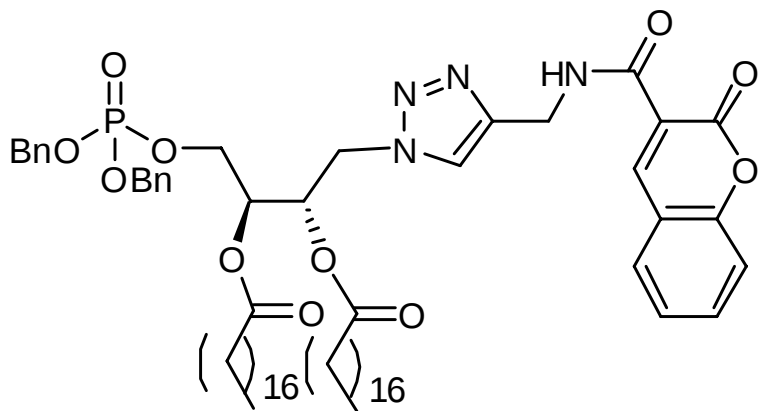
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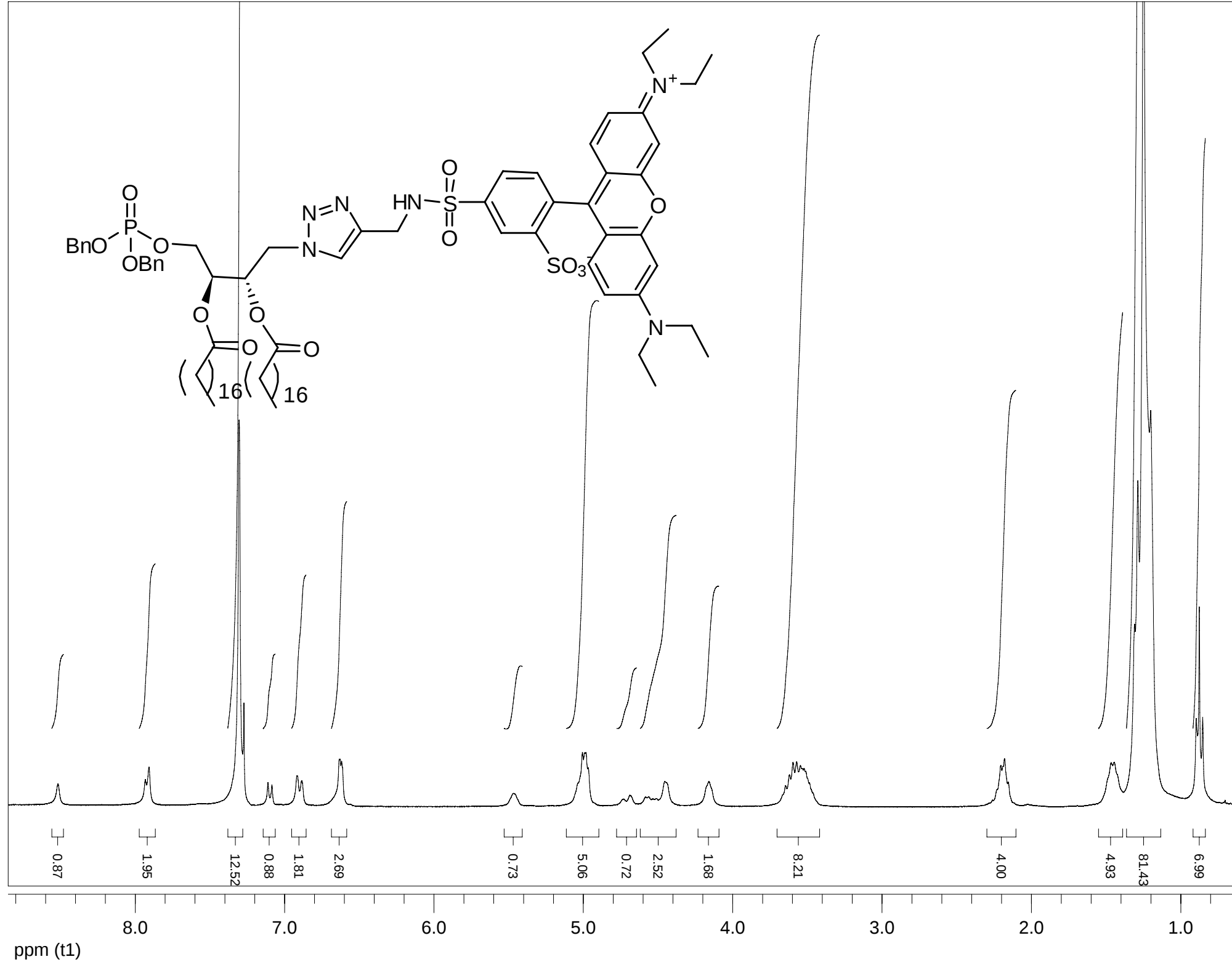
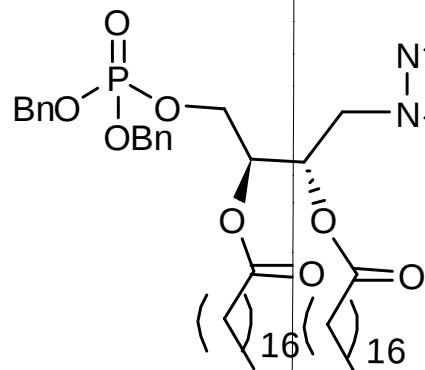
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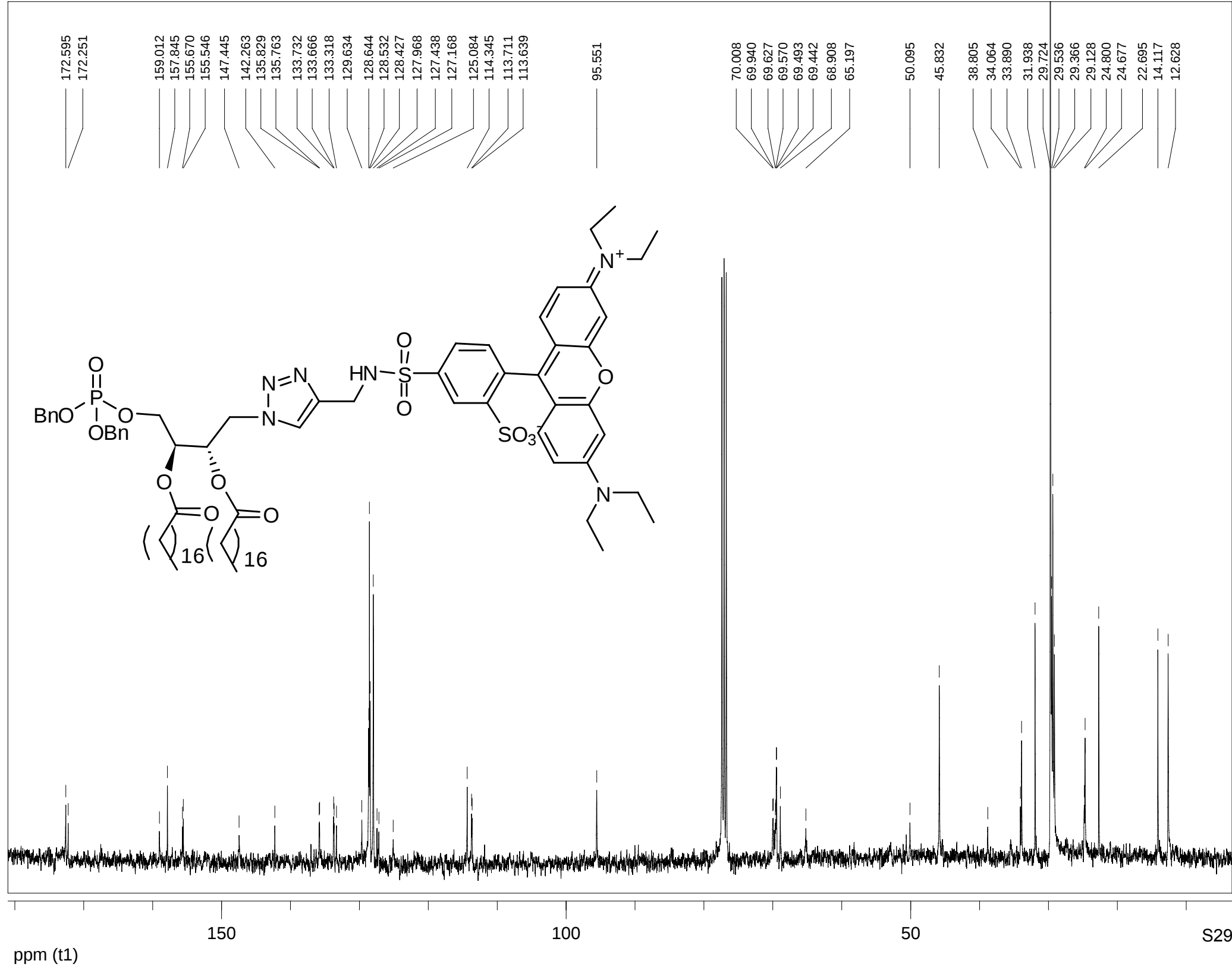
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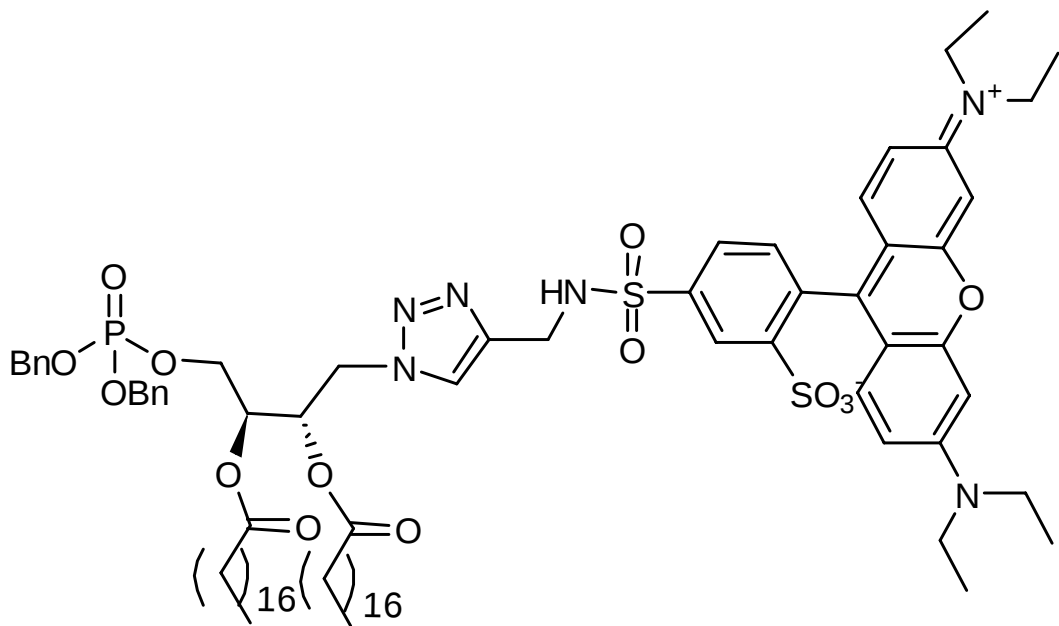




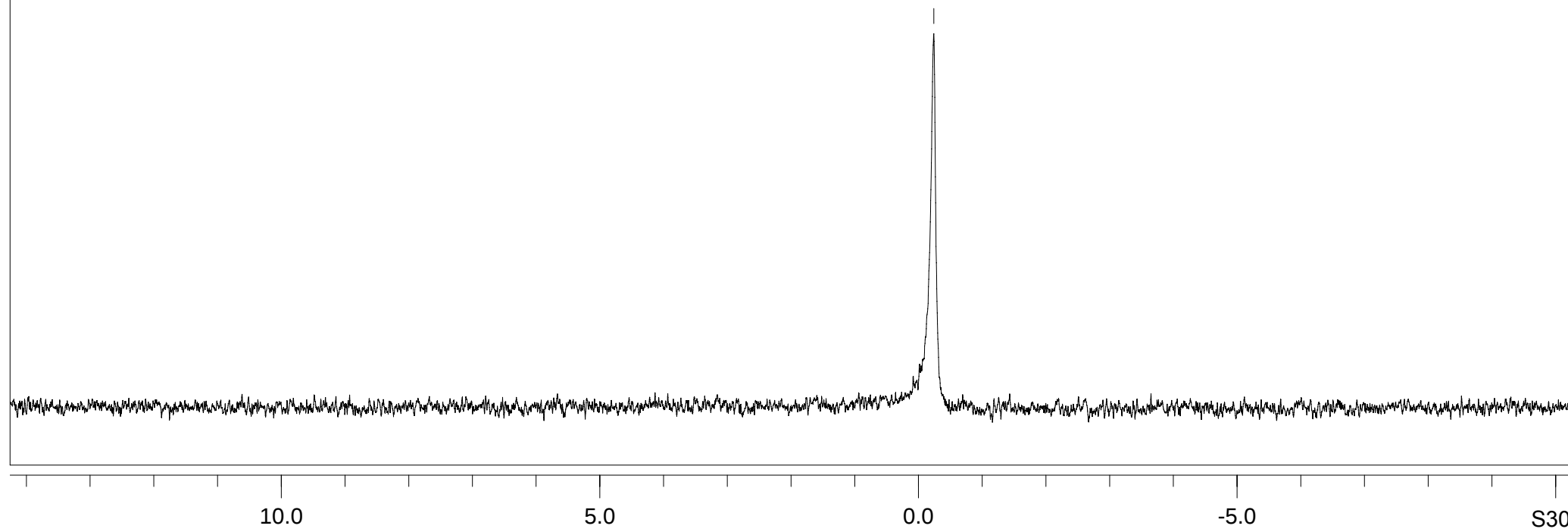




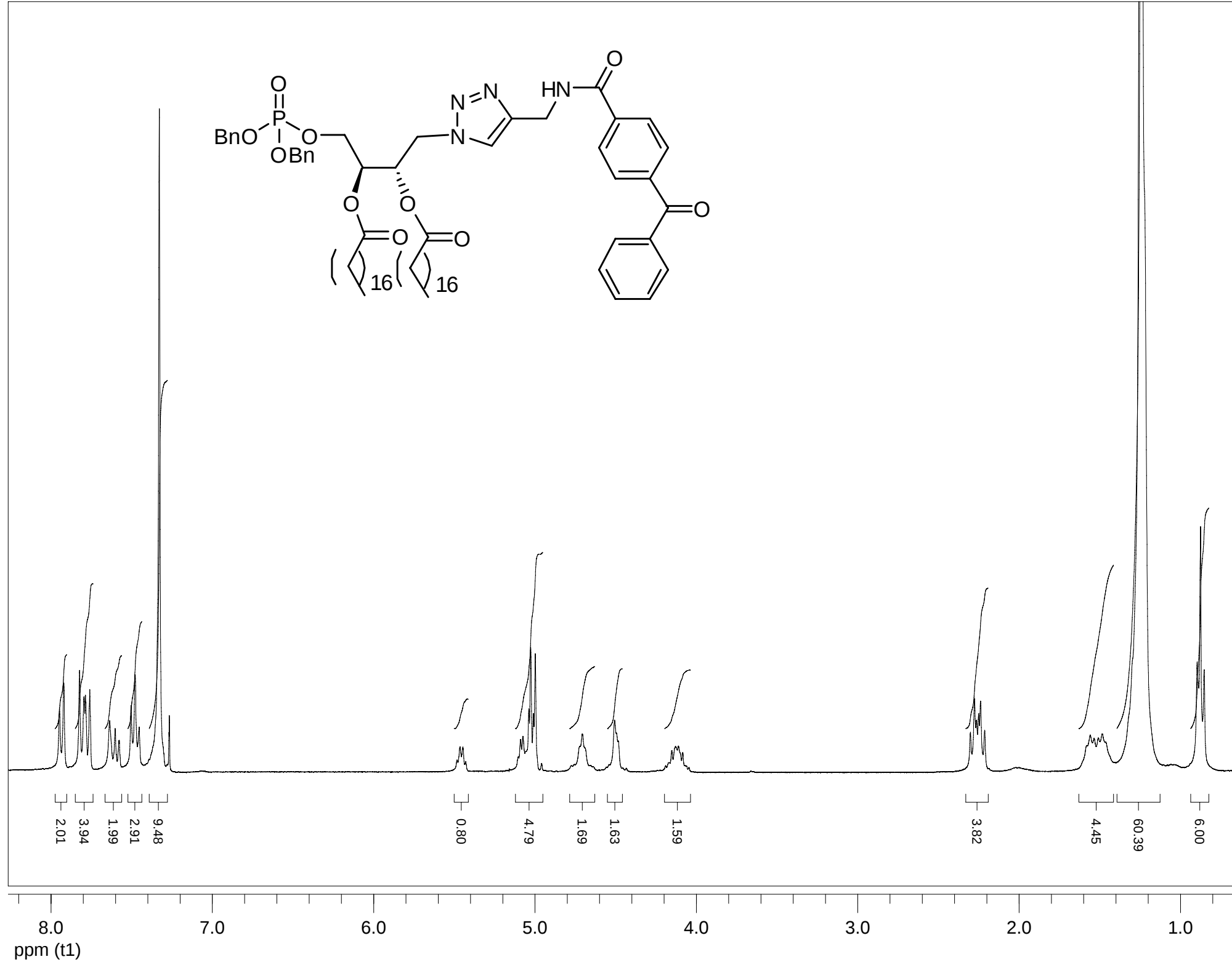
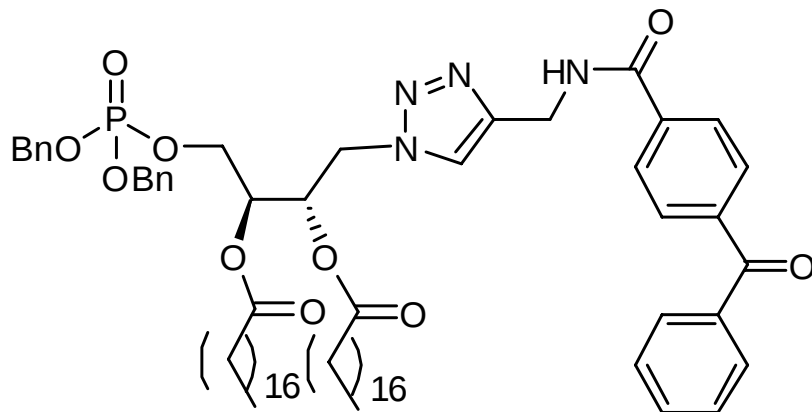


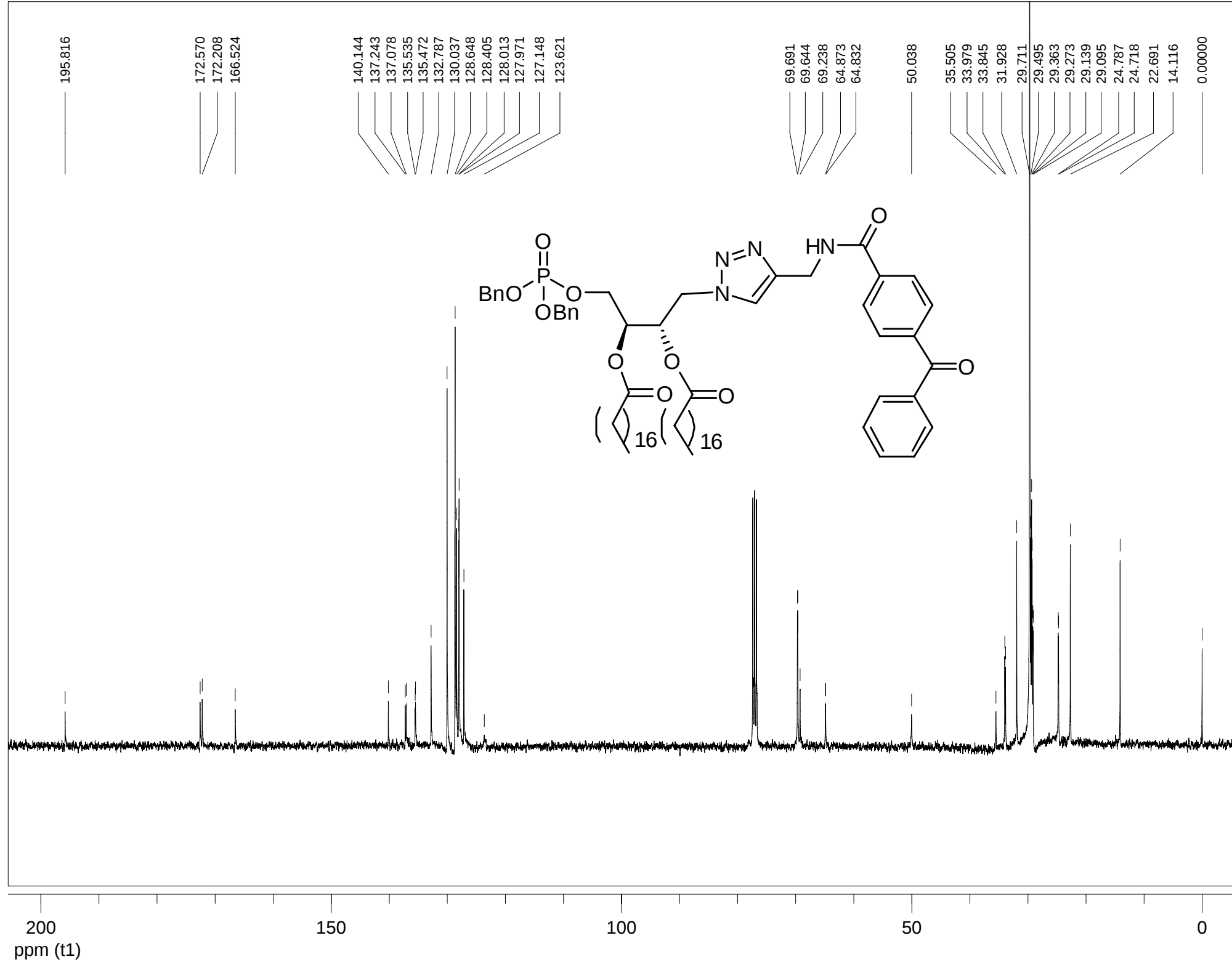


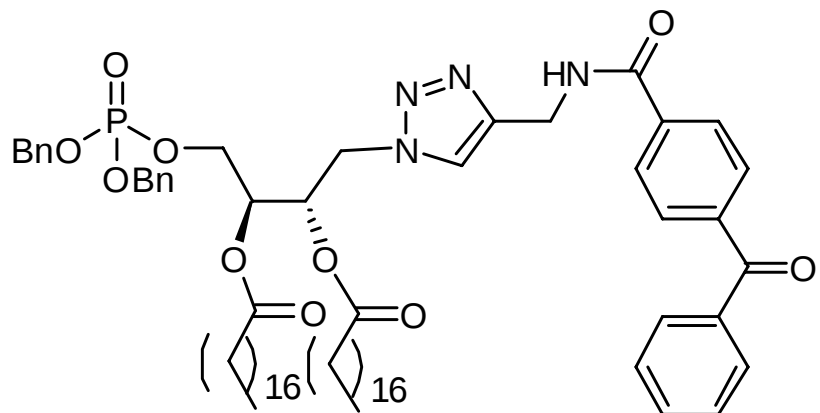
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S30







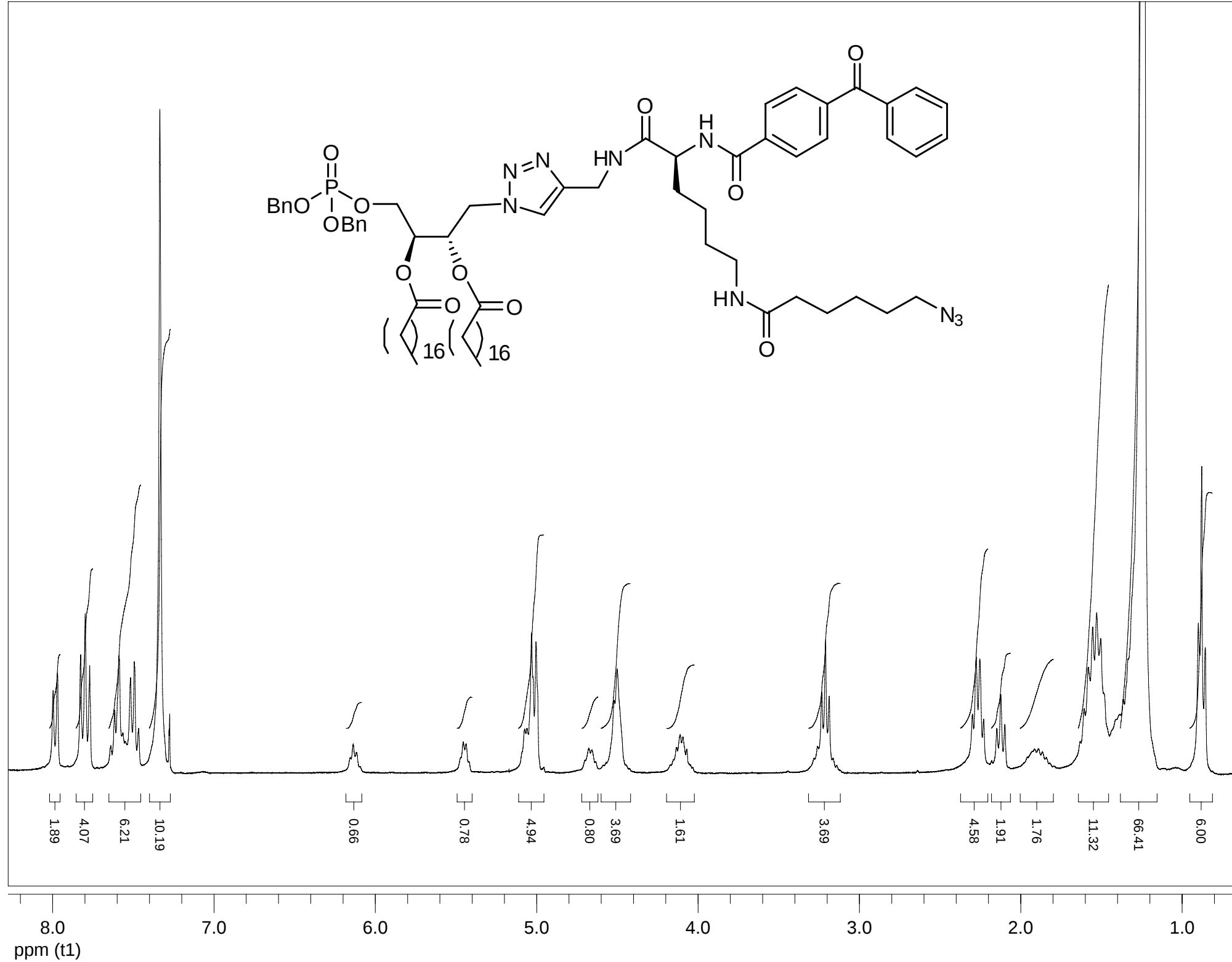
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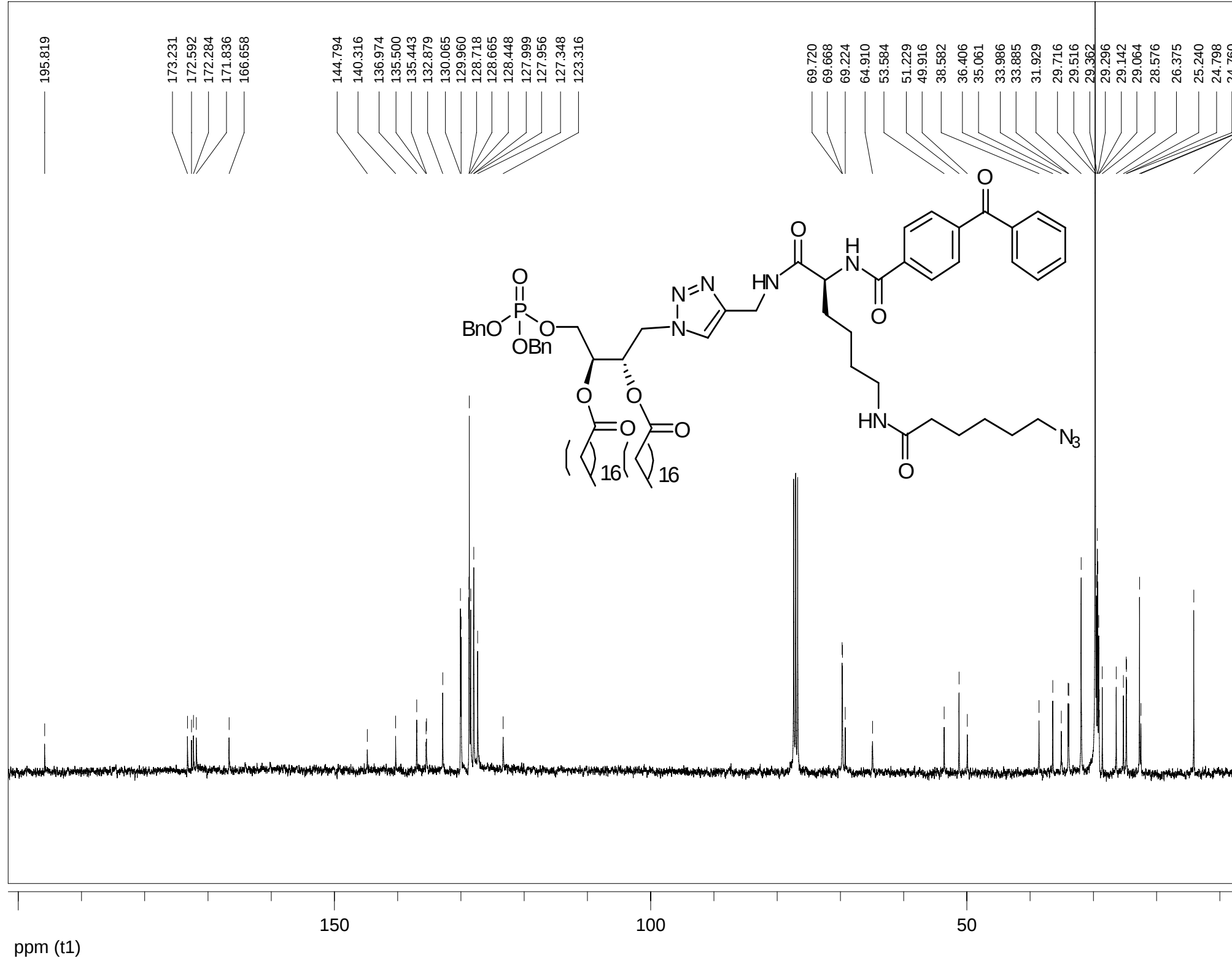
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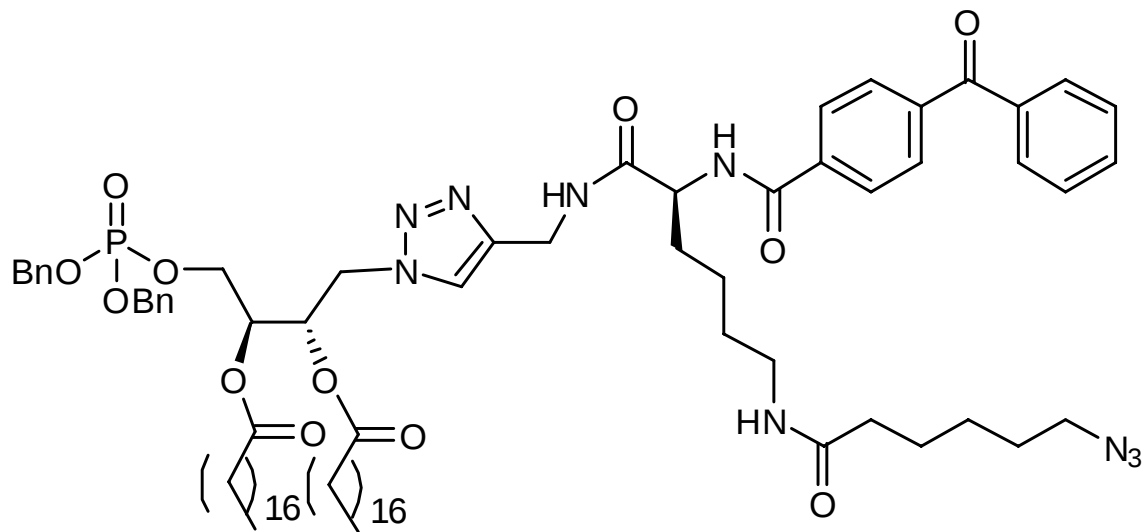
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S33

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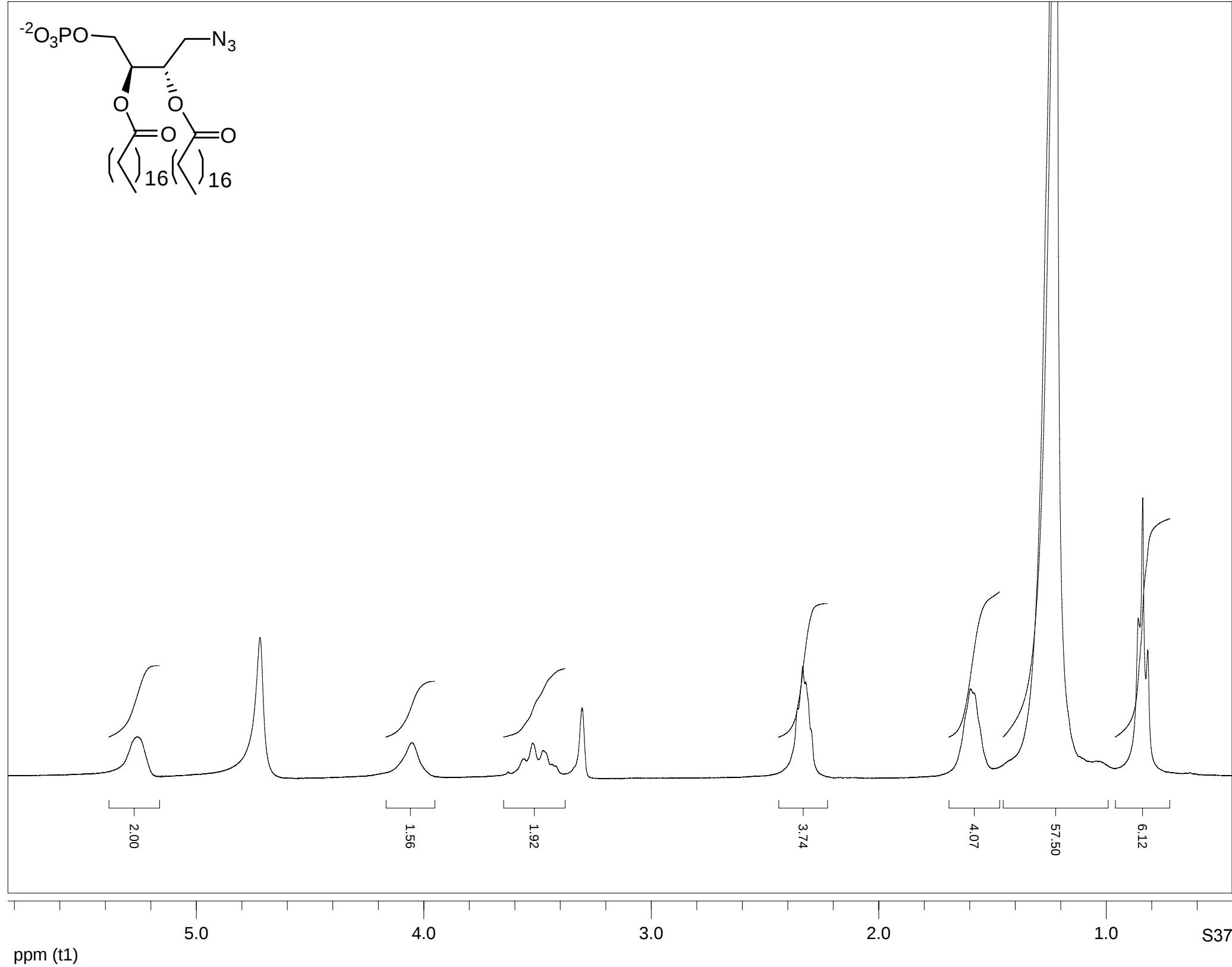
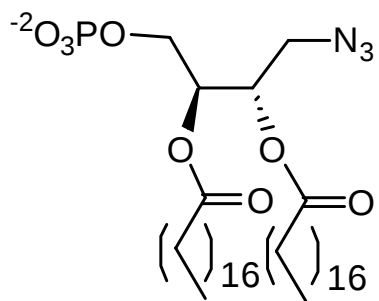
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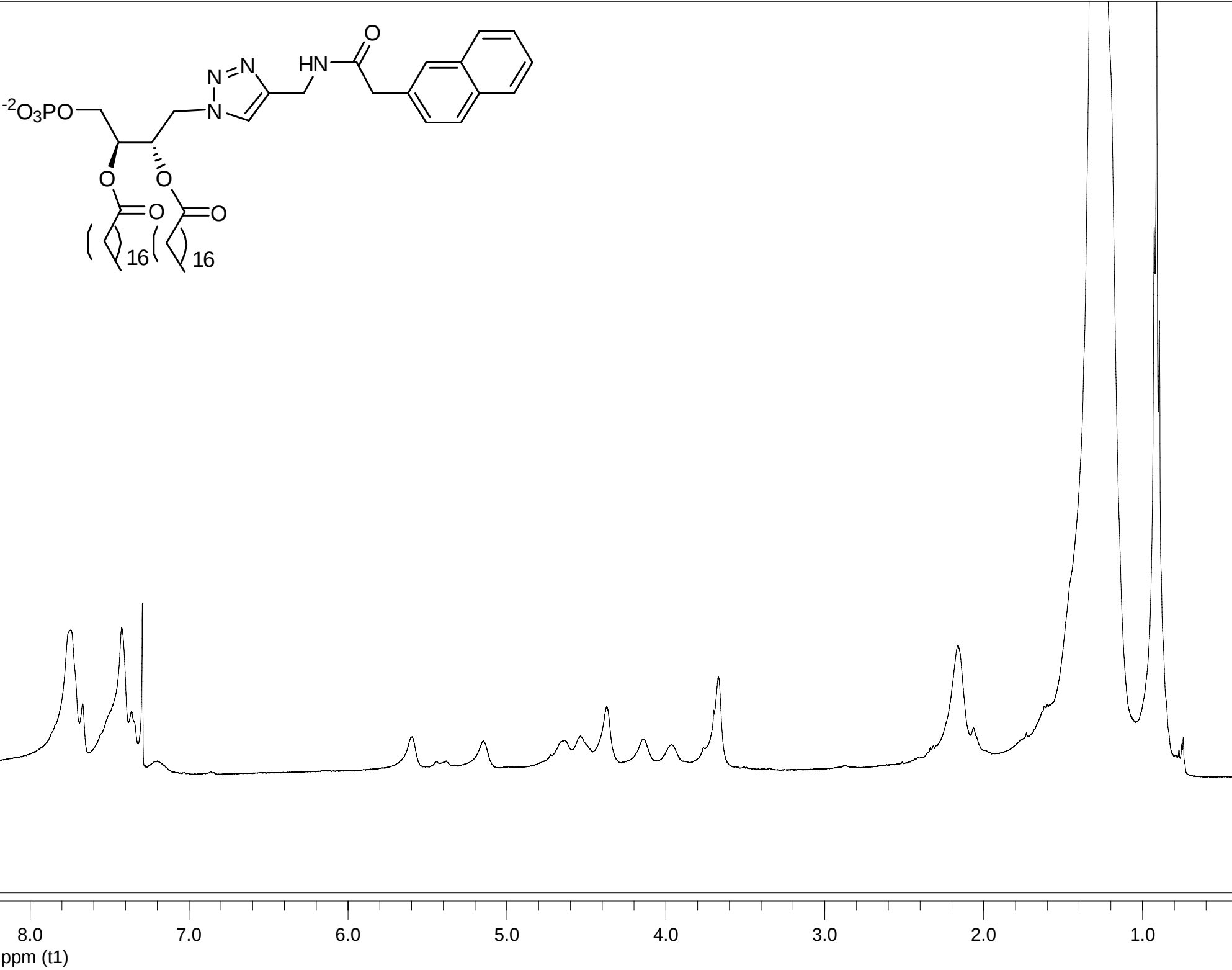
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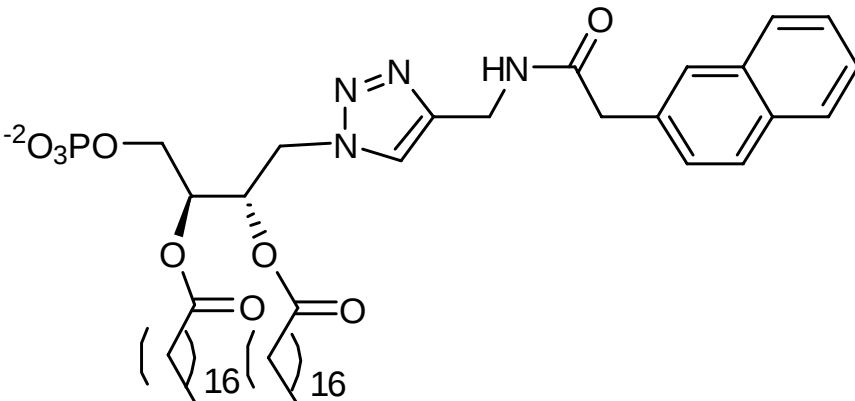
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S36

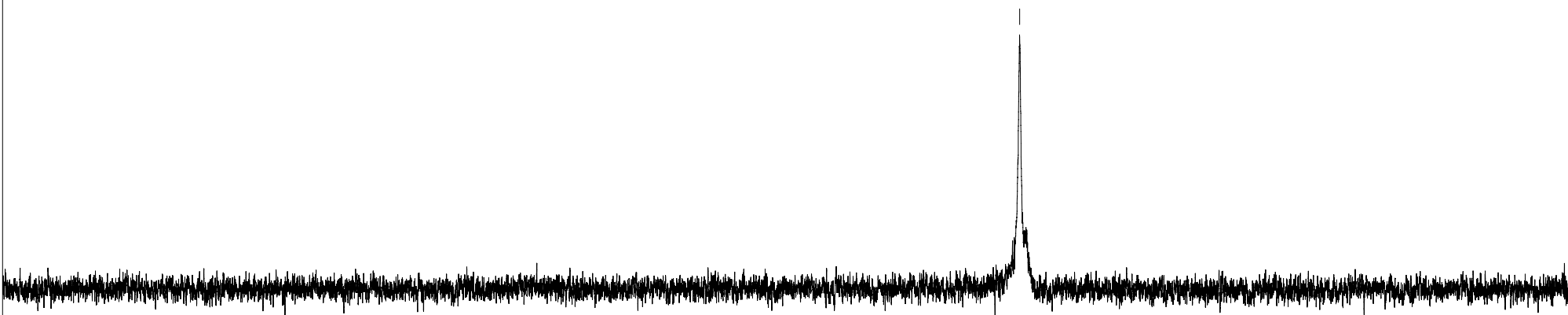
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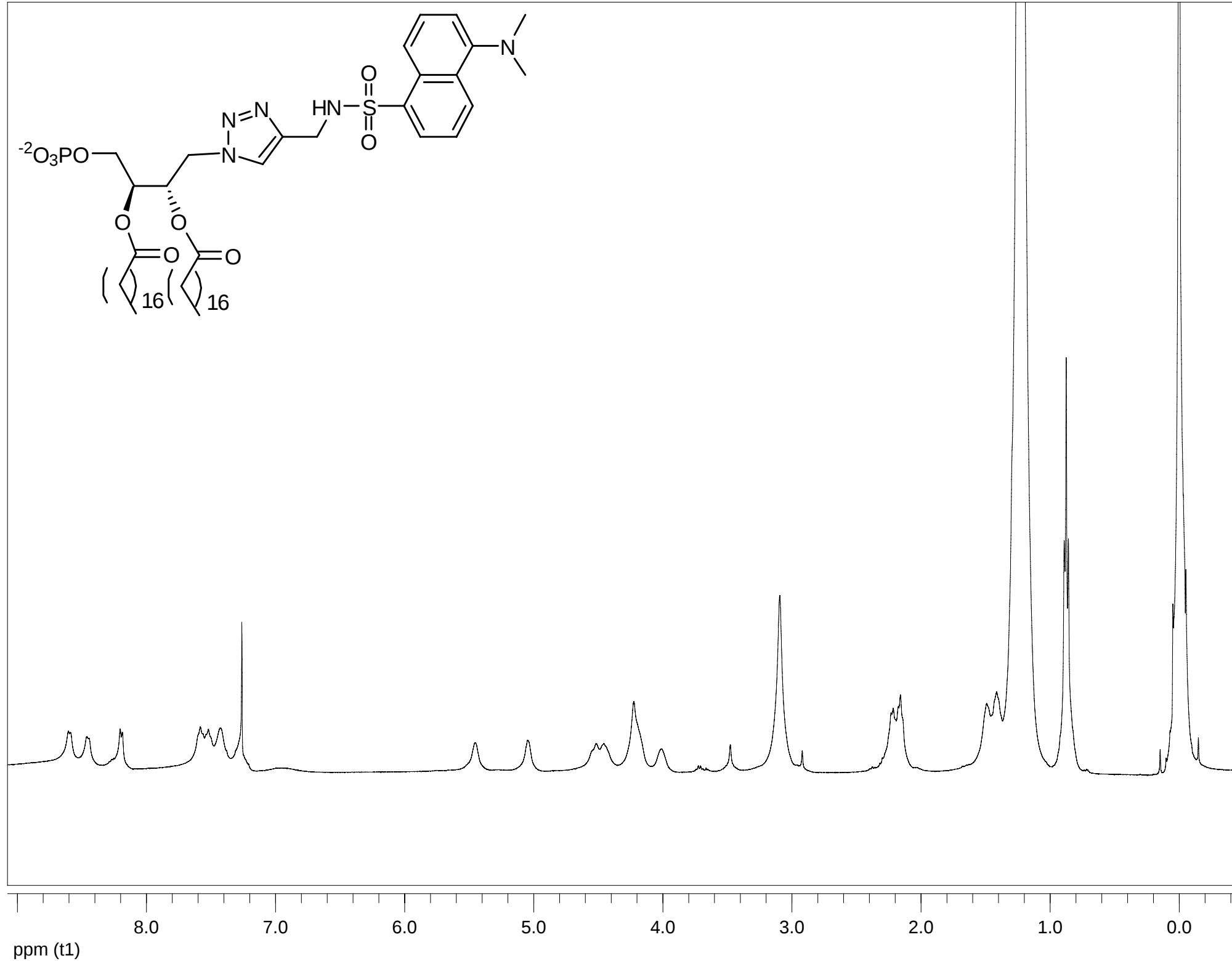
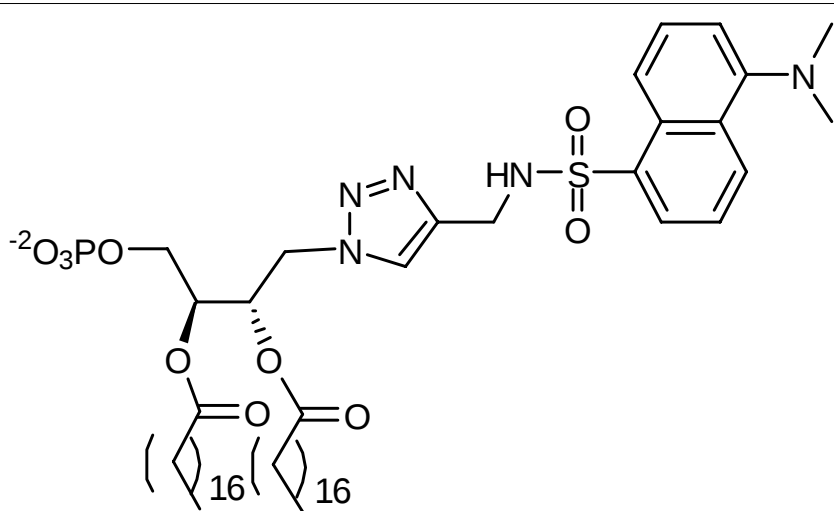


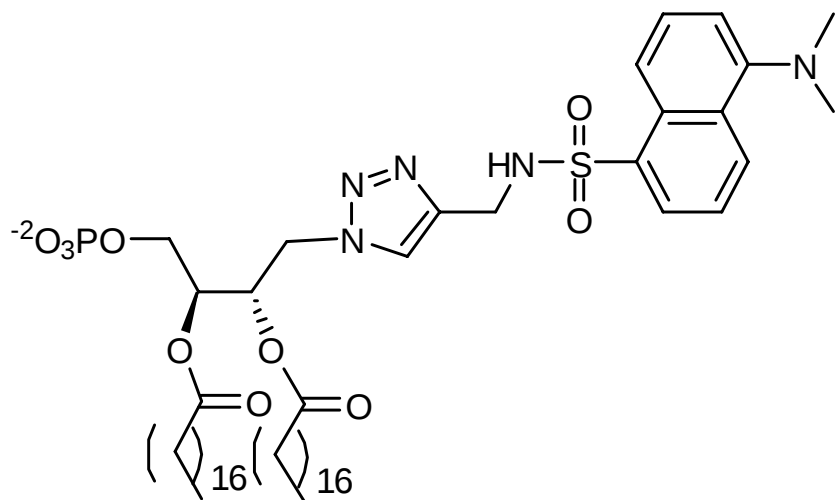




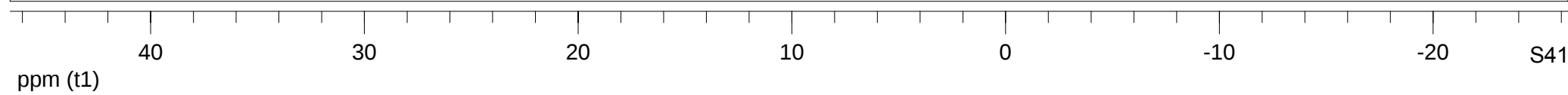
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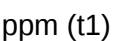


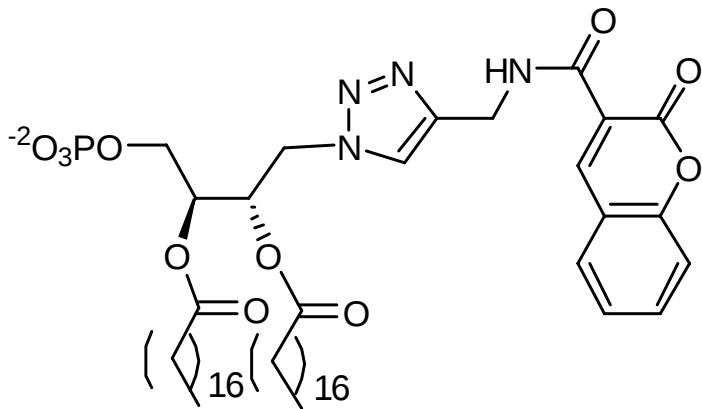




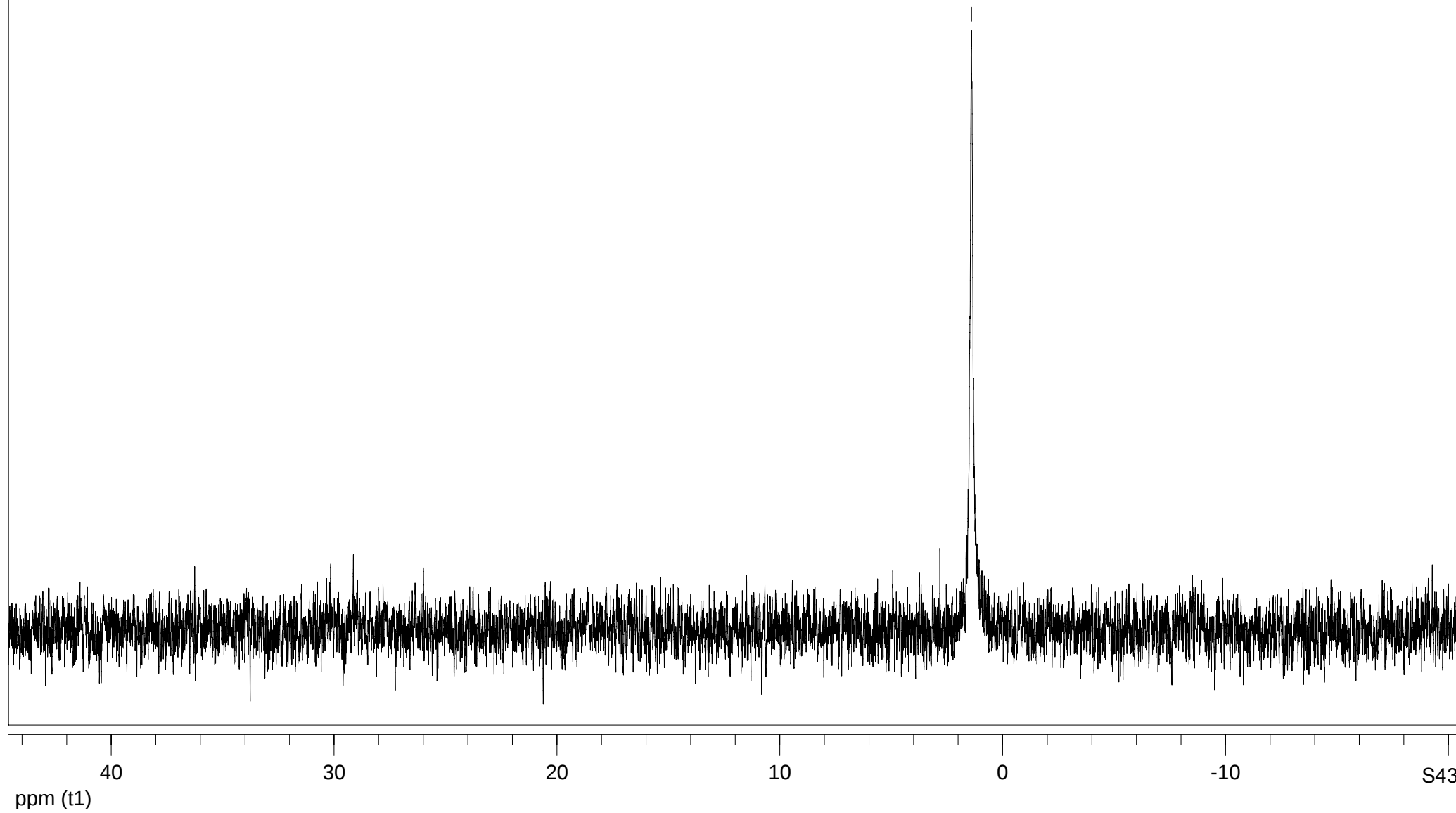
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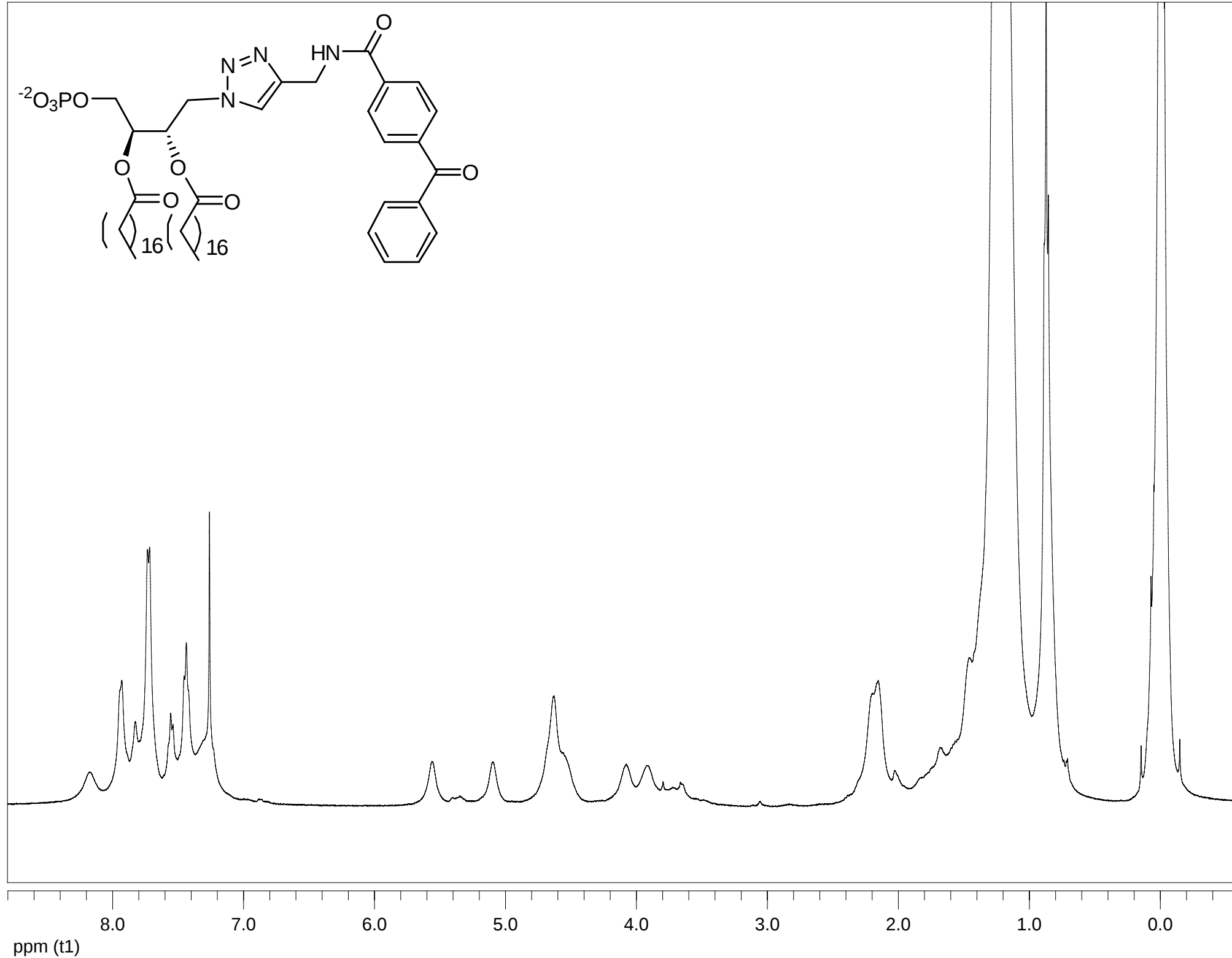
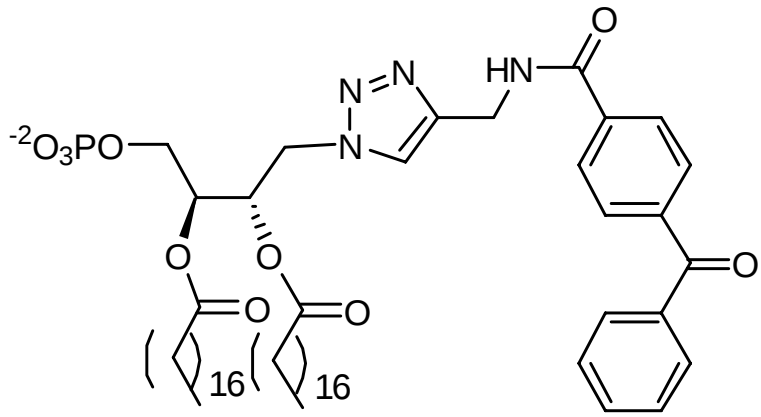


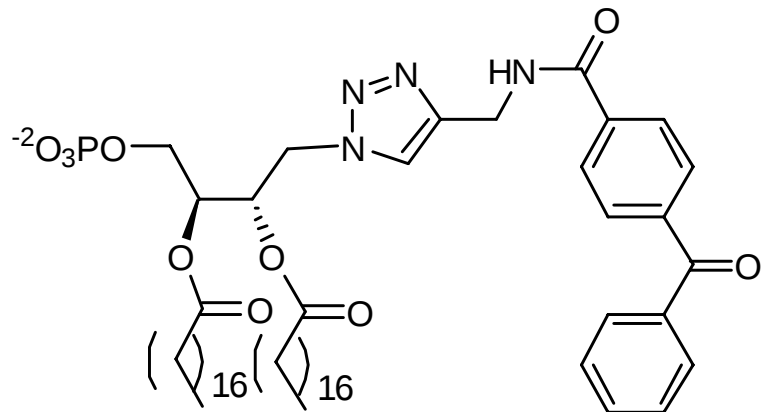




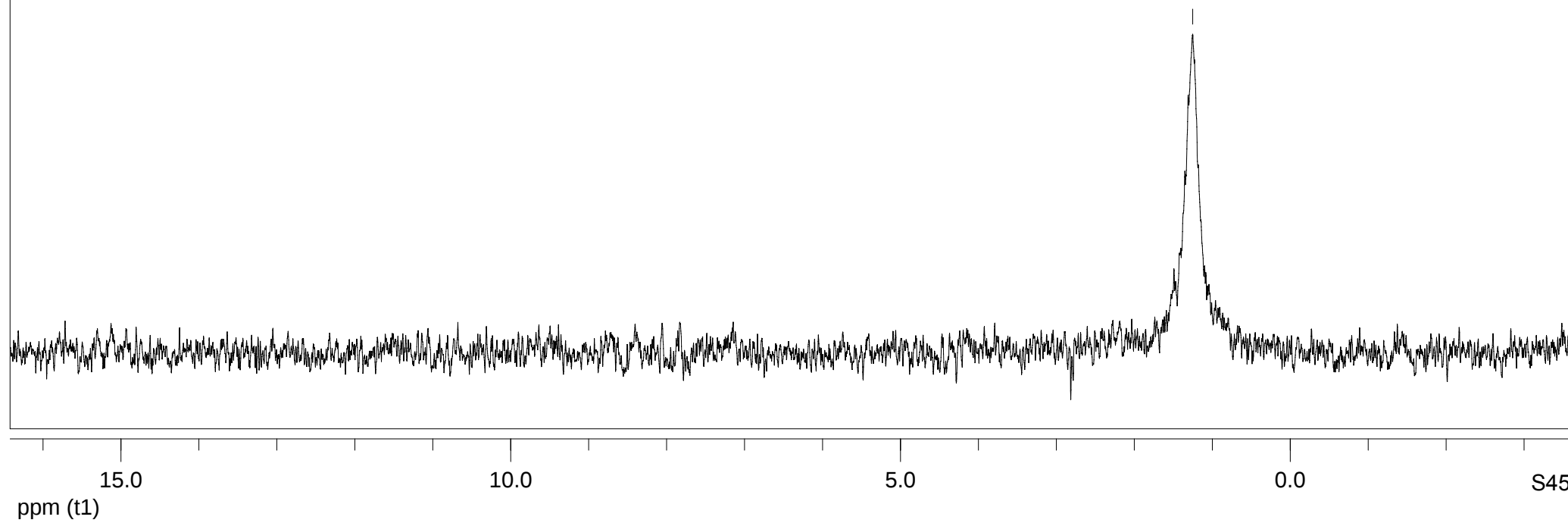
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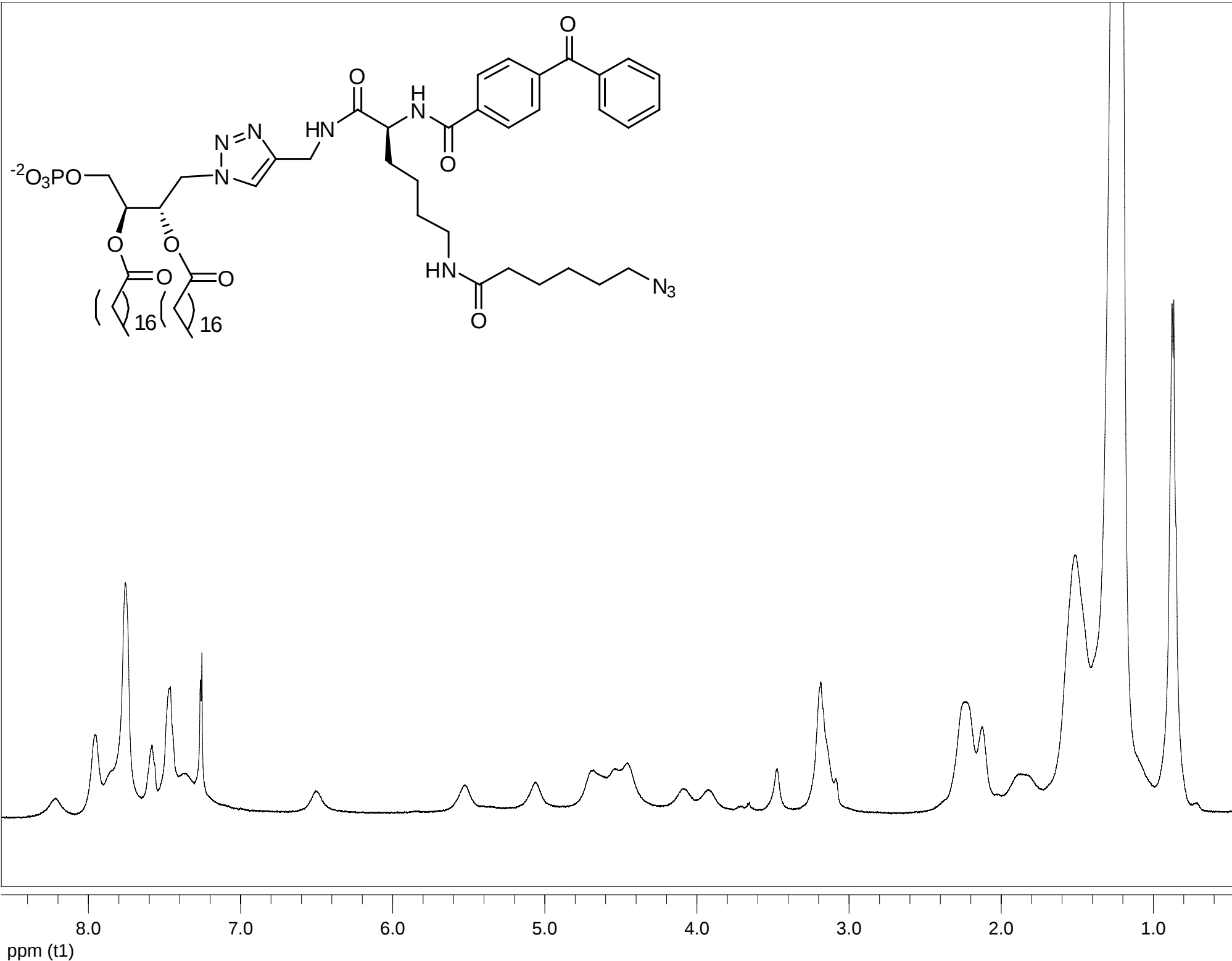
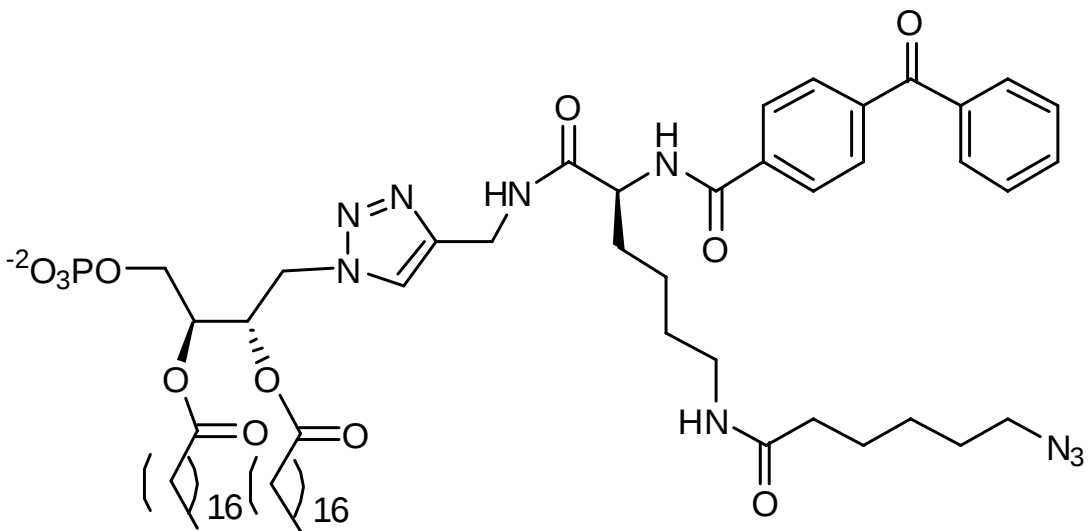


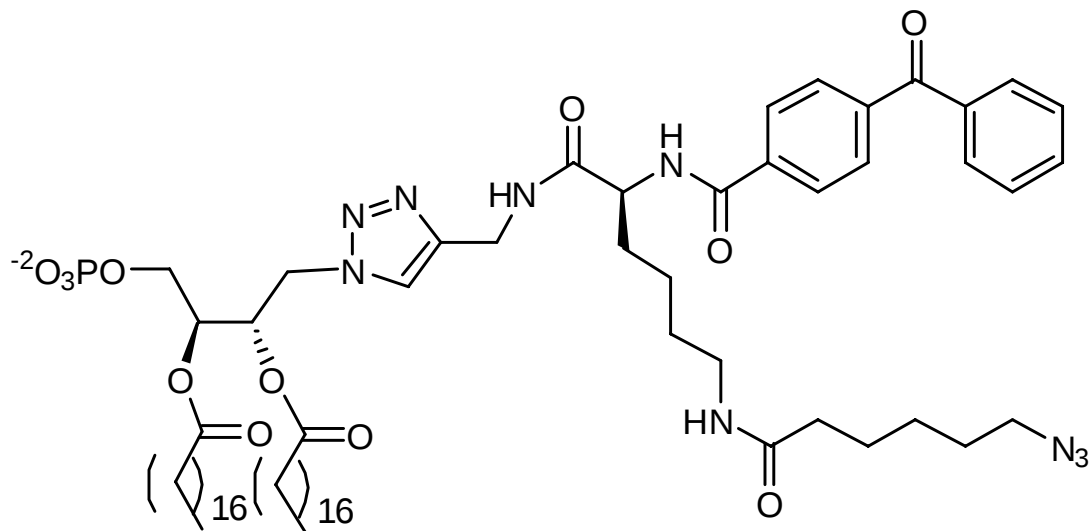




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1.124



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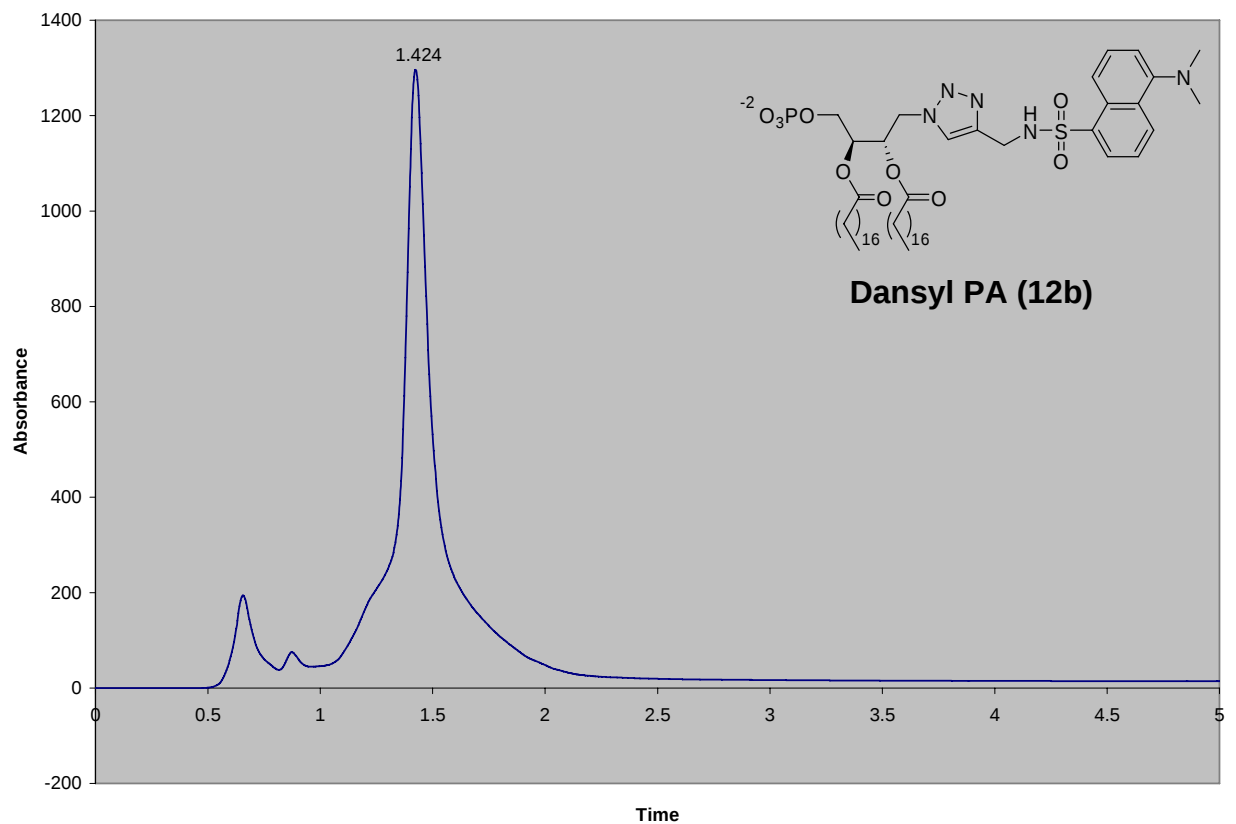
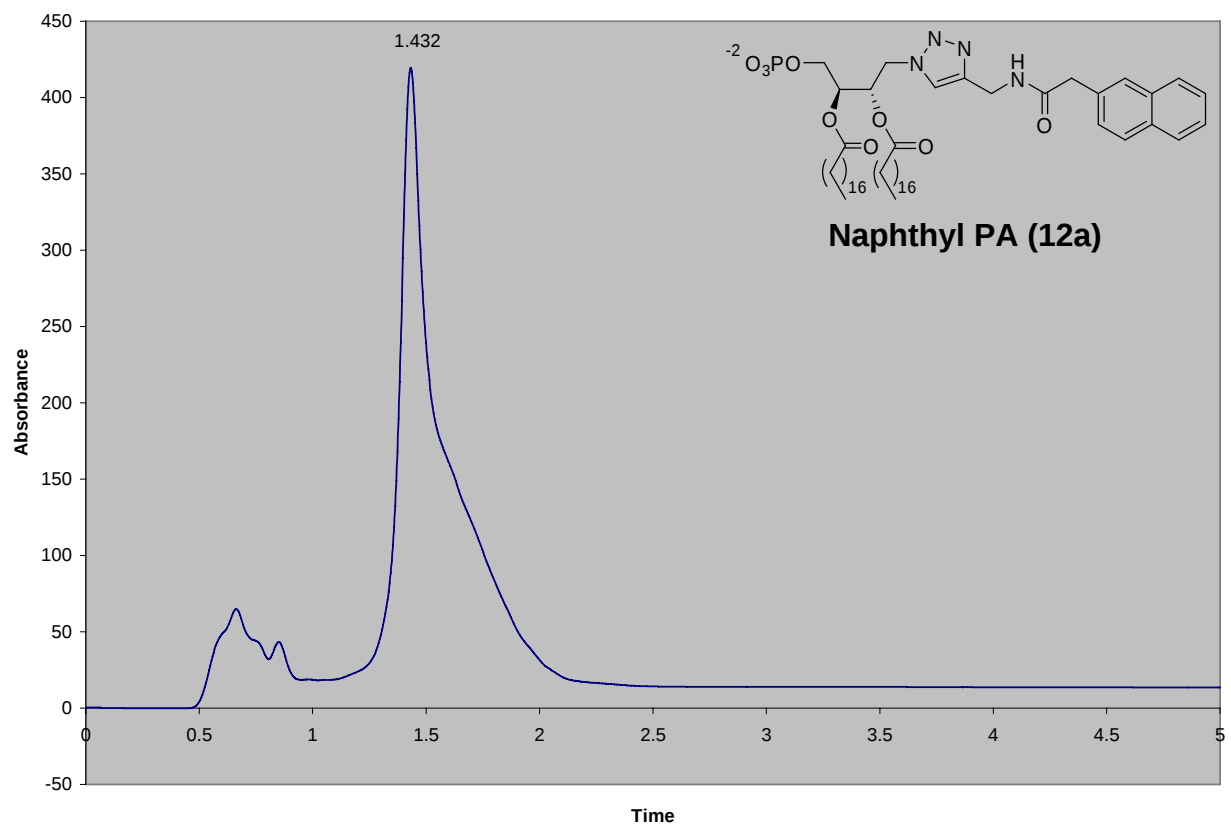
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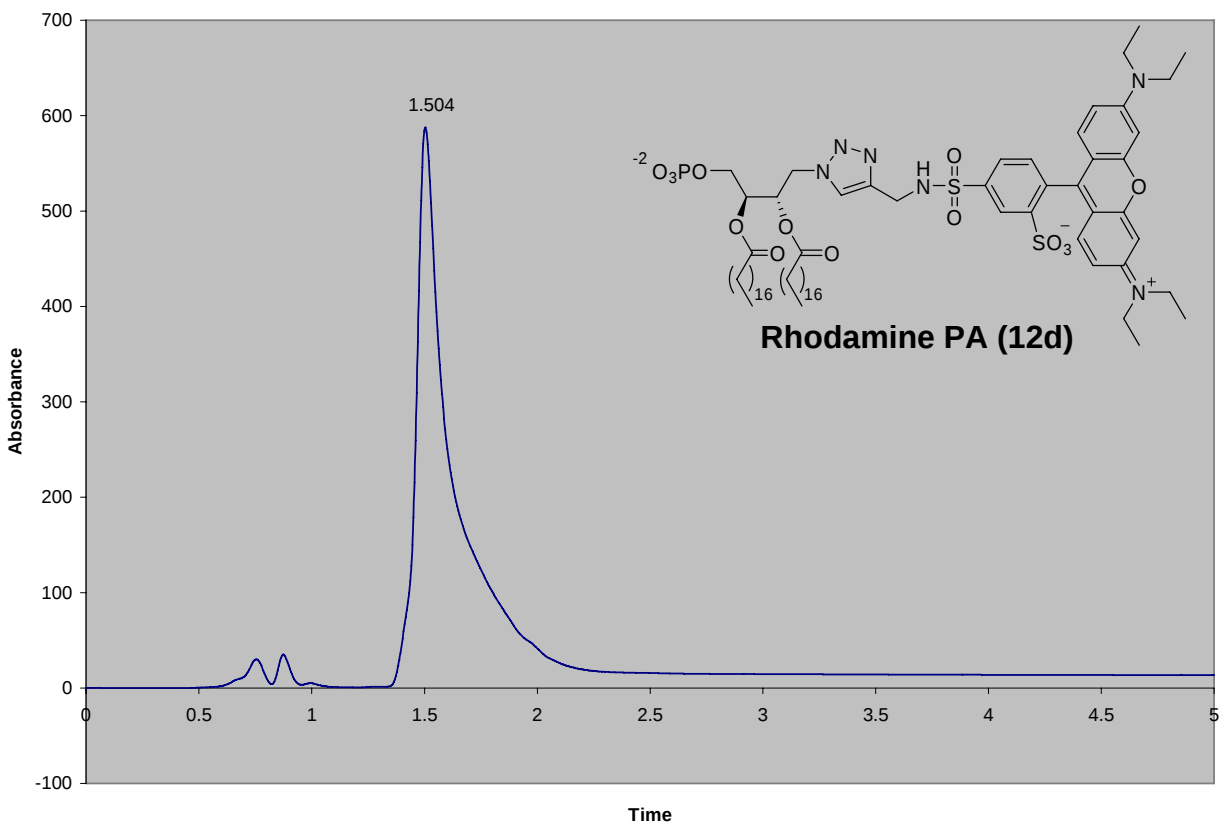
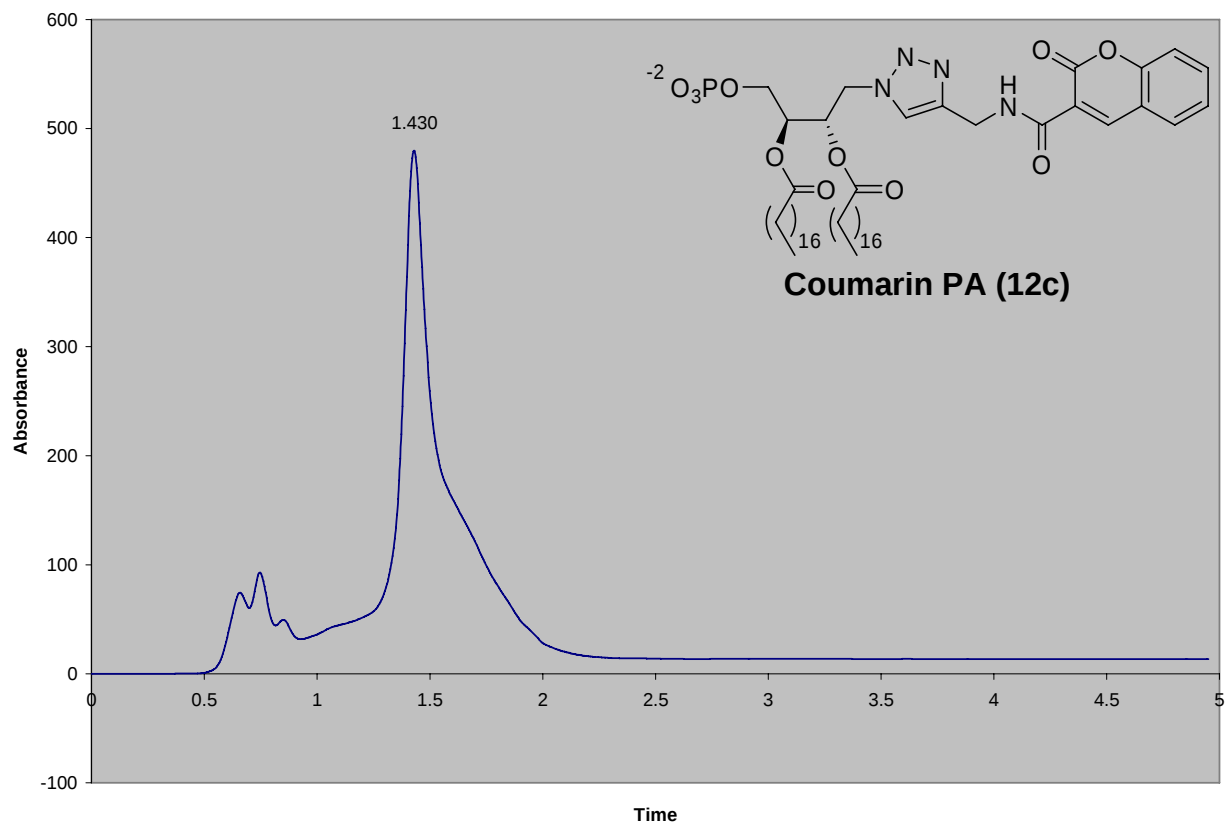
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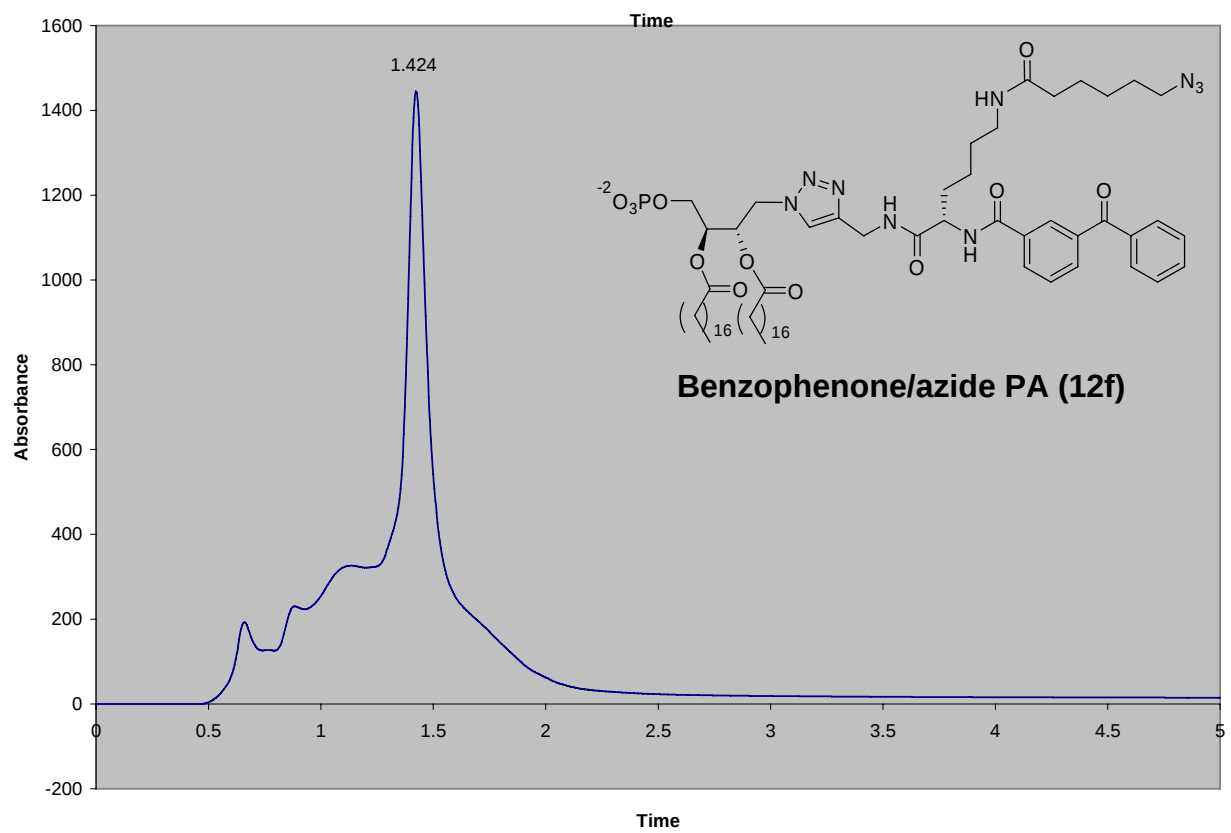
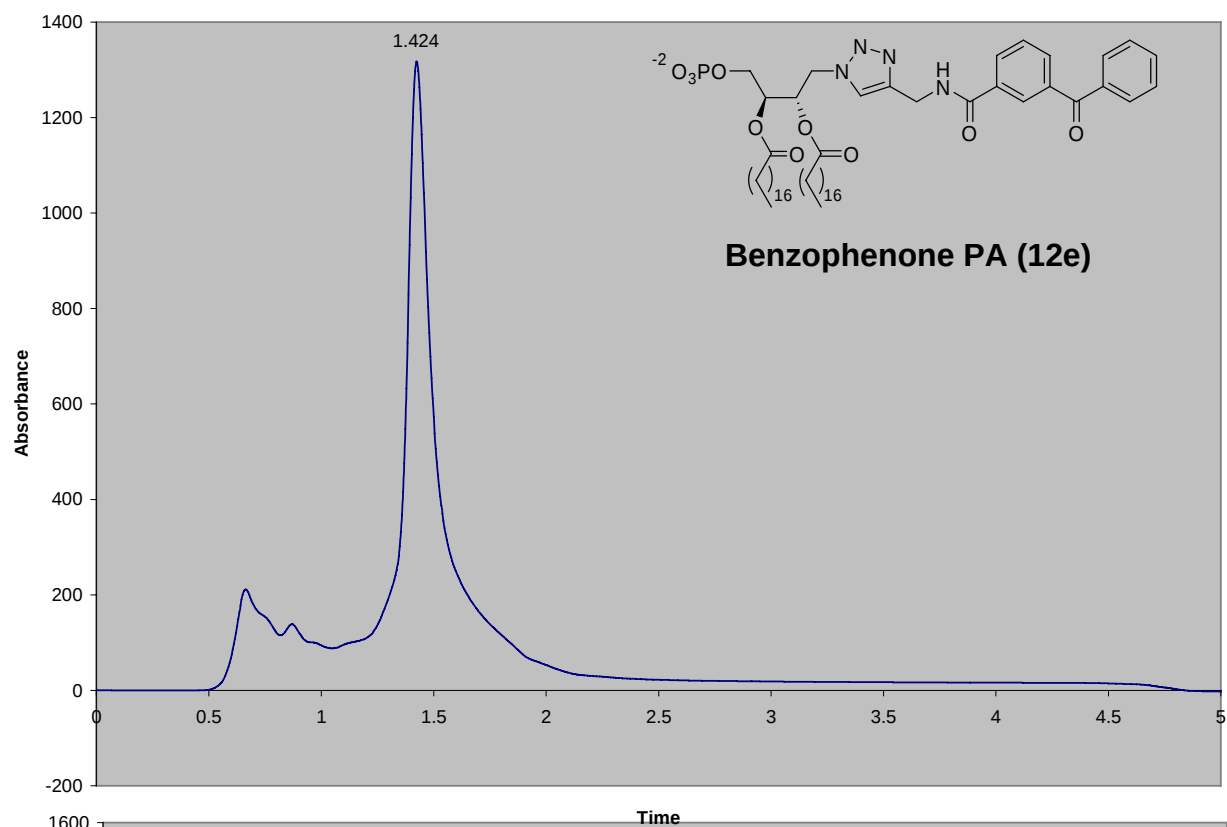
S47

ppm (t1)

III. HPLC Traces of Derivatized PA Probes







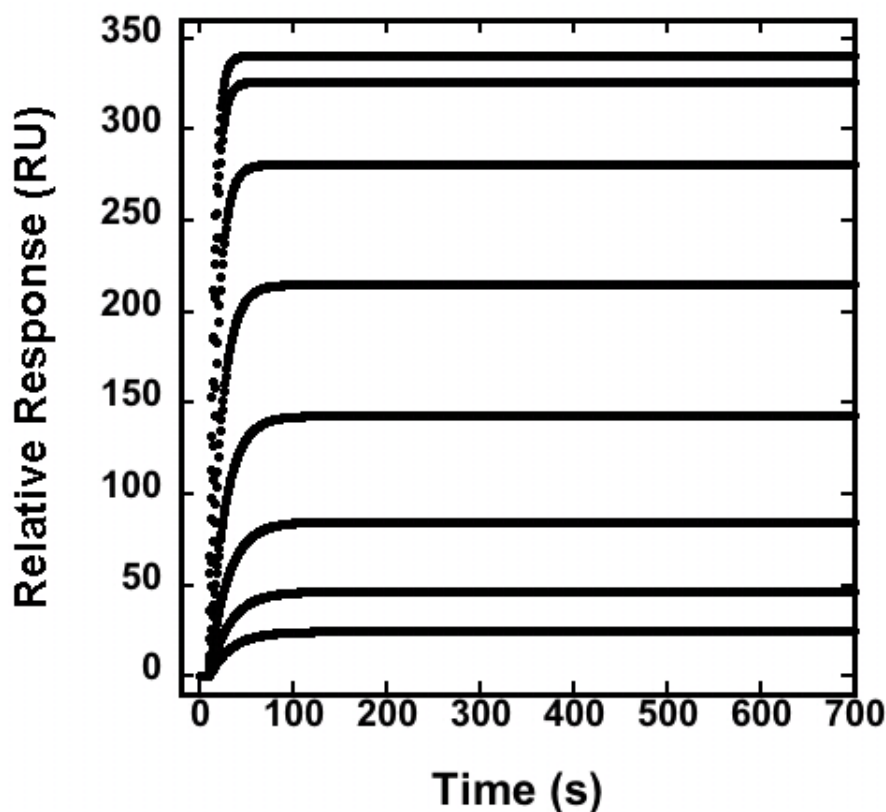


Figure S51. Equilibrium SPR analyses of PKC α -C2. The PKC α -C2 was injected at 10 μ l/min at varying concentrations (50, 100, 200, 400, 800, 1500, 3000, and 4000 nM from bottom to top) over the POPC:POPE:Dansyl-PA (40:40:20) surface using a POPC:POPE (50:50) surface as a control. Subtraction of the POPC:POPE (50:50) response for each protein concentration was performed to yield the binding sensorgrams shown. R_{eq} values were then measured to determine K_d by a nonlinear least-squares analysis of the binding isotherm (See Figure 3).