

Supplemental Information

Supplemental Table S1. Putative pentose transporters from BLAST search.

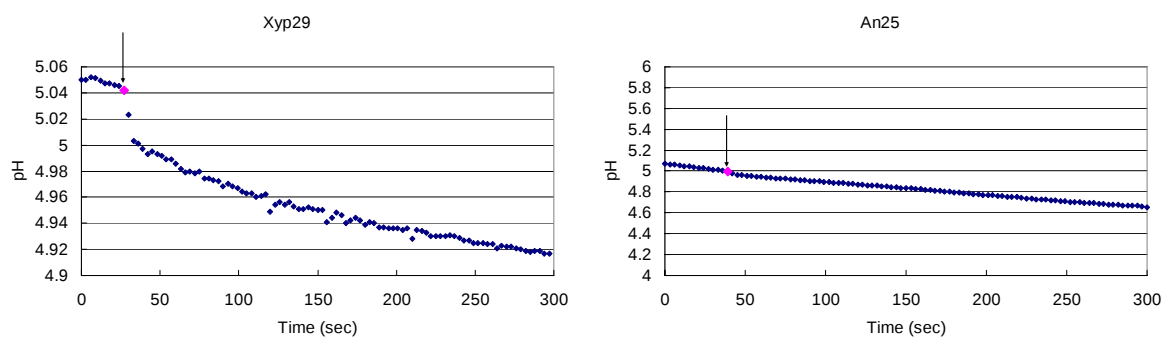
a. BLAST search results using *AUT1* (Locus tag PICST_87108) from *Pichia stipitis* as a probe

Gene Name	Origin	% identity with <i>AUT1</i>	Annotation from NCBI	Length (cDNA, nt)	Locus Tag
<i>Ap31 (SUT2)</i>	<i>P. stipitis</i>	31	sugar uptake (tentative)	1653	ABN66266
<i>Ap26</i>	<i>P. stipitis</i>	26	sugar transporter	1404	XP_001387242
<i>An49</i>	<i>N. crassa</i>	49	hypothetical protein NCU01494, similar to MFS sugar transporter	2025	EAA26691
<i>An41</i>	<i>N. crassa</i>	41	hypothetical protein NCU09287, similar to galactose-proton symporter	1968	EAA28903
<i>An29-2</i>	<i>N. crassa</i>	29	hypothetical protein NCU04963, similar to MFS monosaccharide transporter	1584	EAA30175
<i>An28-3</i>	<i>N. crassa</i>	28	hypothetical protein NCU02188, conserved hypothetical protein	1458	EAA30346
<i>An25</i>	<i>N. crassa</i>	25	sugar transporter	1653	EAA35128

b. BLAST search results using *GXS1* from *Candida intermedia* as a probe

Gene Name	Origin	% identity with <i>GXS1</i>	Annotation from NCBI	Length (cDNA, nt)	Locus Tag
<i>Xy50</i>	<i>N. crassa</i>	50	hypothetical protein NCU04537 similar to monosaccharide transporter	1620	EAA26741
<i>Xy31</i>	<i>N. crassa</i>	31	hypothetical protein NCU06138, similar to MFS monosaccharide transporter	1752	EAA30764
<i>Xy33</i>	<i>N. crassa</i>	33	hypothetical protein NCU00988, similar to MFS quinate transporter	1614	EAA34662
<i>Xyp37 (SUT3)</i>	<i>P. stipitis</i>	37	sugar uptake (tentative)	1653	ABN67990
<i>Xyp33 (XUT3)</i>	<i>P. stipitis</i>	33	sugar transporter, putative xylose uptake (tentative); predicted transporter (major facilitator superfamily)	1656	EAZ63115
<i>Xyp32 (XUT1)</i>	<i>P. stipitis</i>	32	sugar transporter, high affinity, putative; xylose uptake (tentative)	1701	ABN67554
<i>Xyp30 (STL1)</i>	<i>P. stipitis</i>	30	sugar transporter, strongly conserved	1587	ABN65745
<i>Xyp31 (XUT2)</i>	<i>P. stipitis</i>	31	sugar transporter, xylose transporter (tentative) similarly to <i>GXS1</i> (<i>STL1</i>)	1404	AAVQ01000002
<i>Xyp29 (STL12) (XUT6)</i>	<i>P. stipitis</i>	29	sugar transporter, putative (<i>STL12</i>); xylose uptake (tentative)	1635	ABN68560
<i>Xyp30-1 (HGT3)</i>	<i>P. stipitis</i>	30	high affinity xylose transporter (putative), xylose uptake (tentative)	1587	ABN68686
<i>Xyp28 (XUT7)</i>	<i>P. stipitis</i>	28	xylose transporter, high affinity, putative similarity to <i>STL13</i> , high affinity sugar transporters	1251	EAZ63044

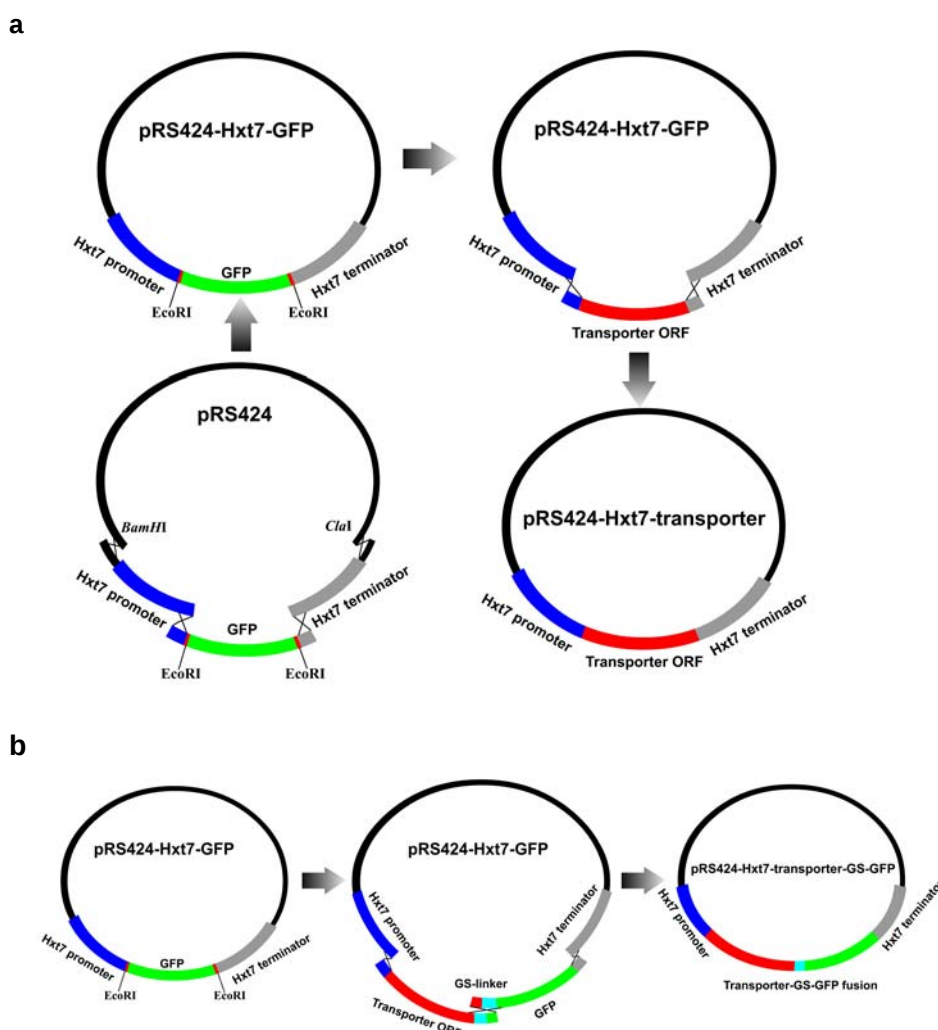
Supplemental Figure S2. Symporter assay of An25 and Xyp29. pH change in un-buffered cell suspension of *S. cerevisiae* EBY.VW4000 strains harboring D-xylose-specific transporter overexpressing plasmids. The pink point indicates when the sugar solution is added into the system.



Supplemental Figure S3. Plasmid construction using the DNA assembler method.

a. Cloning of putative pentose transporters. The pRS424-HXT7-GFP plasmid was used for cloning of putative pentose transporters. In this plasmid, a HXT7 promoter, a GFP gene flanked with the *EcoRI* site at both ends, and a HXT7 terminator, were assembled into the pRS424 shuttle vector linearized by *ClaI* and *BamHI*. PCR products of the putative pentose transporters flanked with DNA fragments sharing sequence identity to the HXT7 promoter and terminator were co-transferred into CEN.PK2-1C with *EcoRI* digested pRS424-HXT7-GFP using the standard lithium acetate method.

b. Construction of plasmids containing transporter-GFP fusion proteins. The GS-linker was added to the N-terminus of the GFP open reading frame by a PCR primer, resulting in a PCR product of GS-linker-GFP flanked with nucleotide sequences identical to the transporters at the 5'-end and the HXT7 terminator at the 3'-end. Transporter genes were amplified from the original pRS424-HXT7-transporter constructs to generate DNA fragments of the transporters flanked with nucleotide sequences identical to the HXT7 promoter at the 5'-end and the GS-linker-GFP at the 3'-end. These two fragments were then co-transferred into CEN.PK2-1C with pRS424-HXT7-GFP digested with *EcoRI*. The resulting transformation mixture was plated on SC-Trp supplemented with 2% D-glucose.



Supplemental Table S4. Alignment of amino acid sequence of cloned An25 and Xyp29 with their corresponding NCBI entries.

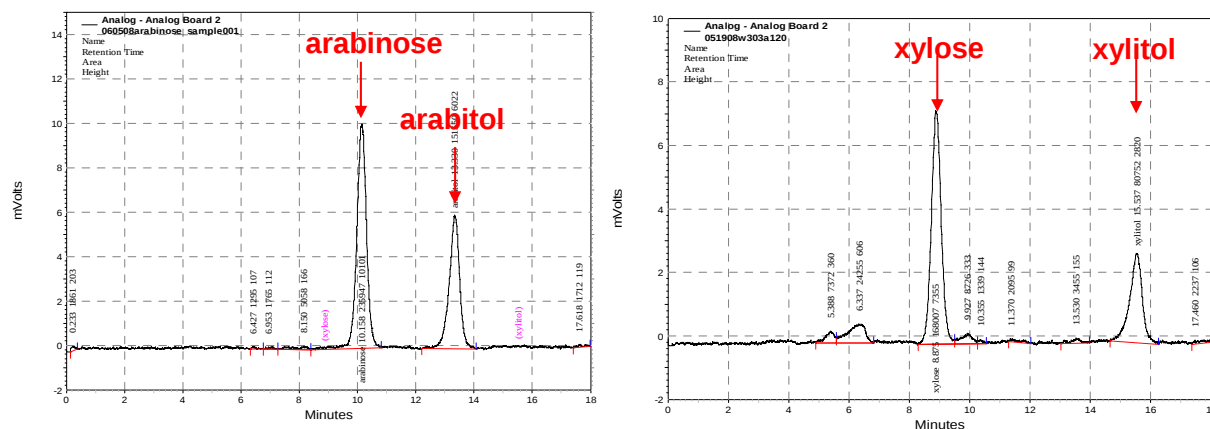
a. Align An25 amino acid sequences (97.3% identity)

An25_NCBI	10	20	30	40	50	60
	M	A	P	P	K	F
An25_cloned	10	20	30	40	50	60
	M	A	P	P	K	F
An25_NCBI	70	80	90	100	110	120
	L	I	T	A	I	E
An25_cloned	70	80	90	100	110	120
	L	I	T	A	I	E
An25_NCBI	130	140	150	160		
	R	V	V	M	---	D
An25_cloned	130	140	150	160		
	R	V	V	M	G	I
An25_NCBI	170	180	190	200	210	220
	V	W	R	F	P	I
An25_cloned	170	180	190	200	210	220
	V	W	R	F	P	I
An25_NCBI	230	240	250	260	270	280
	N	L	I	V	D	S
An25_cloned	230	240	250	260	270	280
	N	L	I	V	D	S
An25_NCBI	290	300	310	320	330	340
	F	Q	D	S	I	G
An25_cloned	290	300	310	320	330	340
	F	Q	D	S	I	G
An25_NCBI	350	360	370	380	390	400
	L	I	P	D	D	P
An25_cloned	350	360	370	380	390	400
	L	I	P	D	D	P
An25_NCBI	410	420	430	440	450	460
	L	I	V	M	V	T
An25_cloned	410	420	430	440	450	460
	L	I	V	M	V	T
An25_NCBI	470	480	490	500	510	520
	Y	V	T	A	A	K
An25_cloned	470	480	490	500	510	520
	Y	V	T	A	A	K
An25_NCBI	530	540	550			
	D	V	V	D	E	H
An25_cloned	530	540	550			
	D	V	V	D	E	H

a. Align Xyp29 amino acid sequences (99.6% identity)

Xyp29_NCBI	10	20	30	40	50	60
	MSSVEKSAETASYTSQVSASGSAKTNSYLGLRGHKLNFVSCFAGVGFLFLFGYDQGV MGS					
Xyp29_cloned	10	20	30	40	50	60
	MSSVEKSAETASYTSQVSASGSAKTNSYLGLRGHKLNFVSCFAGVGFLFLFGYDQGV MGS					
Xyp29_NCBI	70	80	90	100	110	120
	LLTLPSFENTFPAMKASNNATLQGAVIALYEIGCMSSSLATIYLGDR LGRLKIMFIGCVI					
Xyp29_cloned	70	80	90	100	110	120
	LLTLPSFENTFPAMKASNNATLQGAVIALYEIGCMSSSLATIYLGDR LGRLKIMFIGCVI					
Xyp29_NCBI	130	140	150	160	170	180
	VCIGAALQASAFTIAHLTVARIITGLGTGFITSTVPVYQSECSPA KKRGLIMMEGSLIA					
Xyp29_cloned	130	140	150	160	170	180
	VCIGAALQASAFTIAHLTVARIITGLGTGFITSTVPVYQSECSPA KKRGLIMMEGSLIA					
Xyp29_NCBI	190	200	210	220	230	240
	LGIAISYWIDFGFYFLRNDGLHSSASWRAPIALQCVFAVLLISTV FFFPESPRWLLNKGR					
Xyp29_cloned	190	200	210	220	230	240
	LGIAISYWIDFGFYFLRNDGLHSSASWRAPIALQCVFAVLLISTV FFFPESPRWLLNKGR					
Xyp29_NCBI	250	260	270	280	290	300
	TEEAREVFSALYDLPADSEKISIQIEEIQA AIDLERQAGEGFVLKELFTQGPARNLQRVA					
Xyp29_cloned	250	260	270	280	290	300
	TEEAREVFSALYDLPADSEKISIQIEEIQA AIDLERQAGEGFVLKELFTQGPARNLQRVA					
Xyp29_NCBI	310	320	330	340	350	360
	LSCWSQIMQQITGINIITYYAGTIFESYIGMSPFMSRILAALNG TEYFLVSLIAFYTVR					
Xyp29_cloned	310	320	330	340	350	360
	LSCWSQIMQQITGINIITYYAGTIFESYIGMSPFMSRILAALNG TEYFLVSLIAFYTVR					
Xyp29_NCBI	370	380	390	400	410	420
	LGRRFLLFWGAIAMALVMAGLTVTVKLAGEGNTHAGVGA AVLLFAFNSFFGVSWLGGSWL					
Xyp29_cloned	370	380	390	400	410	420
	LGRRFLLFWGAIAMALVMAGLTVTVKLAGEGNTHAGVGA AVLLFAFNSFFGVSWLGGSWL					
Xyp29_NCBI	430	440	450	460	470	480
	LPPELLSLKLRAPGAALSTASNWAFNFMVVMITPVGFQSIGSY TYLIFAAINLLMAPVIY					
Xyp29_cloned	430	440	450	460	470	480
	LPPELLSLKLRAPGAALSTASNWAFNFMVVMITPVGFQSIGSY TYLIFAAINLLMAPVIY					
Xyp29_NCBI	490	500	510	520	530	540
	FLYPETKGRSLEEMDIIFNQCPVWEPWKVVQIARDLPIMHSE VLDHEKDVIIKSRIEHV					
Xyp29_cloned	490	500	510	520	530	540
	FLYPETKGRSLEEMDIIFNQCPVWEPWKVVQIARDLPIMHSE VLDHEKNVIIKKSRIEHV					
Xyp29_NCBI	ENIS					
Xyp29_cloned	ENIS					

Supplemental Figure S5. Representative chromatogram of HPLC analysis of intracellular sugar concentrations.



Sugar and corresponding sugar alcohol concentrations were determined using Shimadzu HPLC equipped with a Bio-Rad HPX-87C column (Bio-Rad Laboratories, Hercules, CA) and Shimadzu ELSD-LTII low temperature-evaporative light scattering detector following the manufacturer's protocol. The HPX-87C column was kept at 85 °C using a Shimadzu CTO-20AC column oven. Deionized water was used as mobile phase at a constant flow rate of 0.2 mL/min. During the HPLC process, nitrogen was provided to the ELSD detector at a constant pressure of 350 kPa. 20 uL of filtered sample was injected into the HPLC system with a Shimadzu SIL-20AC HT auto sampler, and each run was stopped at 20 minutes after the injection. The concentration of the sugars and corresponding sugar alcohols were determined using a standard curve generated using a series of external standards. The sugar uptake activity was calculated as mg sugar extracted through osmosis per mL of cell culture at OD~10.