Probing Aromatic, Hydrophobic, and Steric Effects on the Self-Assembly of an Amyloid- β Fragment Peptide

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Figure S1. Reverse sedimentation data for solubilization of peptide monomers from resuspended fibrils derived from $A\beta(16-22)$ variants indicating increase in monomer concentration over time.



Figure S2. HPLC trace (215 nm) of A β (16–22) Wild-type.



Figure S3. HPLC trace (215 nm) of Aβ(16–22) F₅-Phe 19.



Figure S4. HPLC trace (215 nm) of $A\beta(16-22)$ Cha 19.









Figure S7. HPLC trace (215 nm) of Aβ(16–22) F₅-Phe 20.



Figure S8. HPLC trace (215 nm) of $A\beta(16-22)$ Cha 20.



Figure S9. HPLC trace (215 nm) of Aβ(16–22) F₅-Phe 19,20.



Figure S10. HPLC trace (215 nm) of $A\beta(16-22)$ Cha 19,20.



Peptide	ide Sequence Retention Gradient (solution A: water/0.5% TFA		Gradient (solution A: water/0.5% TFA; solution
		time (min)	B: acetonitrile/0.5% TFA)
1	Ac-KLVFFAE-NH ₂	11.9	Isocratic 5% B over 5 minutes, increase 5-95% B over 10 minutes, maintain at 95% B over 5 minutes
2	Ac-KLVAFAE-NH ₂	10.9	Isocratic 5% B over 5 minutes, increase 5-95% B over 10 minutes, maintain at 95% B over 5 minutes
3	Ac-KLVAFAE-NH ₂	11.0	Isocratic 5% B over 5 minutes, increase 5-95% B over 10 minutes, maintain at 95% B over 5 minutes
4	Ac-KLV(F ₅ -Phe)FAE-NH ₂	12.2	Isocratic 5% B over 5 minutes, increase 5-95% B over 10 minutes, maintain at 95% B over 5 minutes
5	Ac-KLV(Cha)FAE-NH ₂	12.4	Isocratic 5% B over 5 minutes, increase 5-95% B over 10 minutes, maintain at 95% B over 5 minutes
6	Ac-KLVF(F ₅ -Phe)AE-NH ₂	12.4	Isocratic 5% B over 5 minutes, increase 5-95% B over 10 minutes, maintain at 95% B over 5 minutes
7	Ac-KLV(Cha)FAE-NH ₂	12.3	Isocratic 5% B over 5 minutes, increase 5-95% B over 10 minutes, maintain at 95% B over 5 minutes
8	Ac-KLV(F ₅ -Phe) (F ₅ - Phe)AE-NH ₂	12.7	Isocratic 5% B over 5 minutes, increase 5-95% B over 10 minutes, maintain at 95% B over 5 minutes
9	Ac-KLV(Cha)(Cha)AE- NH ₂	12.8	Isocratic 5% B over 5 minutes, increase 5-95% B over 10 minutes, maintain at 95% B over 5 minutes

 Table S1. Analytical HPLC conditions for peptides 1-9.

Figure S11. HPLC trace for co-injection of peptides 1-5.



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Peak #	Retention time	Mass (observed)	Peptide	
1	10.9-11	817.32, 910.01	Ala 19, Tyr 19	
2	11.9	893.06	Wild-type	
3	12.2	983.00	F ₅ -Phe 19	
4	12.4	899.14	Cha 19	

Figure S12. HPLC trace for co-injection of peptides 1 & 6-9.



Table S3. MALDI mass spectra characterization of each peak from Figure S11.

Peak #	Retention time	Mass (observed)	Peptide
1	11.9	894.52	Wild-type
2	12.4	900.58, 984.50	Cha 20, F ₅ -Phe 20
3	12.8	906.58, 1096.40	Cha 19,20, F ₅ -Phe 19,20

Figure S13. ESI mass spectrum of $A\beta(16-22)$ Wild-type.



Figure S14. ESI mass spectrum of $A\beta(16-22)$ F₅-Phe 19.



Figure S15. ESI mass spectrum of $A\beta(16-22)$ Cha 19.



Figure S16. ESI mass spectrum of $A\beta(16-22)$ Ala 19.



Figure S17. ESI mass spectrum of $A\beta(16-22)$ Tyr 19.



Figure S18. ESI mass spectrum of A β (16–22) Wild-type ([1-¹³C] Leu17, Ala21).



Figure S19. ESI mass spectrum of $A\beta(16-22)$ F₅-Phe 19 ([1-¹³C] Leu17, Ala21).



Figure S20. ESI mass spectrum of A β (16–22) Cha 19 ([1-¹³C] Leu17, Ala21).



Figure S21. ESI mass spectrum of A β (16–22) Wild-type ([1-¹³C] Leu17, Phe20).



Figure S22. ESI mass spectrum of A β (16–22) F₅-Phe 19 ([1-¹³C] Leu17, Phe20).



Figure S23. ESI mass spectrum of A β (16–22) Cha 19 ([1-¹³C] Leu17, Phe20).



Figure S24. MALDI mass spectrum of $A\beta(16-22)$ F₅-Phe 20.



Figure S25. MALDI mass spectrum of $A\beta(16-22)$ Cha 20.



Figure S26. MALDI mass spectrum of $A\beta(16-22)$ F₅-Phe 20 ([1-¹³C] Lys16, Ala21).







Figure S28. MALDI mass spectrum of $A\beta(16-22)$ F₅-Phe 19,20.



Figure S29. MALDI mass spectrum of $A\beta(16-22)$ Cha 19,20.



Figure S30. MALDI mass spectrum of Aβ(16–22) F₅-Phe 19,20 ([1-¹³C] Lys16, Ala21).







Table S4. Calculated and observed m/z for peptides 1–11.

Peptide	Sequence	Calculated	Observed
1	Ac-KLVFFAE-NH ₂	894.50 (M ⁺ H)	894.49
2	Ac-KLV(F ₅ -Phe)FAE-NH ₂	984.45 (M ⁺ H)	984.50
3	Ac-KLV(Cha)FAE-NH ₂	900.55 (M ⁺ H)	900.70
4	Ac-KLVAFAE-NH ₂	818.46 (M ⁺ H)	818.45
5	Ac-KLVYFAE-NH ₂	910.49 (M ⁺ H)	910.45
6	Ac-K*LVFF*AE-NH ₂	895.60 (M ⁺)	895.70
7	Ac-K*LV(F ₅ -Phe)F*AE-NH ₂	986.45 (M ⁺ H)	986.50
8	Ac-K*LV(Cha)F*AE-NH ₂	902.55 (M ⁺ H)	902.70
9	Ac-K*LVF*FAE-NH ₂	895.60 (M ⁺)	895.50
10	Ac-K*LV(F ₅ -Phe)*FAE-NH ₂	985.45 (M ⁺)	985.55
11	Ac-K*LV(Cha)*FAE-NH ₂	902.55 (M ⁺ H)	902.45
12	Ac-KLVF(F ₅ -Phe)AE-NH ₂	1006.44 (M ⁺ H)	1006.21
13	Ac-KLVF(Cha)AE-NH ₂	900.55 (M ⁺ H)	900.38
14	Ac-*KLVF(F ₅ -Phe)*AE-NH ₂	986.45 (M ⁺ H)	986.49
15	Ac-*KLVF(Cha)*AE-NH ₂	902.55 (M ⁺ H)	902.58
16	Ac-KLV(F ₅ -Phe) (F ₅ -Phe)AE-NH ₂	906.60 (M ⁺ H)	906.40
17	Ac-KLV(Cha) (Cha)AE-NH ₂	1096.39 (M ⁺ Na)	1096.09
18	Ac-*KLV(F ₅ -Phe) (F ₅ -Phe)*AE-NH ₂	908.60 (M ⁺ H)	908.70
19	Ac-*KLV(Cha)(Cha)*AE-NH ₂	1076.50 (M ⁺ H)	1076.47

* Indicates [1-¹³C] label.





Figure S33. HPLC concentration curve $A\beta(16-22)$ F₅-Phe 19 and F₅-Phe 20.







Figure S35. HPLC concentration curve $A\beta(16-22)$ Ala 19.







Figure S37. HPLC concentration curve $A\beta(16-22)$ F₅-Phe 19,20.



Figure S38. HPLC concentration curve $A\beta(16-22)$ Cha 19,20.



Preparation of HPLC concentration curves

Concentration curves were produced using a protocol adopted from Wetzel *et al.* [O'Nuallain, B.; Thakur, A.M.; Williams, A. D.; Bhattacharyya, A. M.; Chen, S.; Thiagarajan, G.; Wetzel, R. (2006) *Methods Enzymol.* 413, 34–74]. The curves are instrument specific and are only valid for the exact instrument and column combination used here (Shimadzu HPLC system: SPD-20A UV-vis detector, SIL-10ADVP auto-injector, LC-10ATVP solvent delivery modules; Column: Waters X-Bridge BEH300 Prep C18 10mm).

Freshly dissociated peptide was dissolved in DMSO. Serial dilutions of this DMSO stock were subsequently prepared injected in triplicate on an HPLC column monitored by UV at 215 nm. Each injection was followed by a blank (neat DMSO) to insure there was no carry-over from previous injections. The peak areas (based on UV absorbance) were plotted against the amount of peptide injected, providing linear concentration curves. At higher concentrations these curves diverge from linearity, and in these cases, lower concentrations are necessary to insure accurate determination of concentration. At least solutions corresponding to distinct points on the concentration curve were then submitted for amino acid analysis (American International Biotechnology Services, Richmod, VA) to determine the precise amount of peptide in each solution. These values and the known injection volumes were correlated to the concentration curves, allowing precise determination of peptide quantity as a function of UV absorbance in HPLC eluent. Peptide concentrations from HPLC sedimentation assays were determined using these concentration curves.