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Active site analysis

Glucose-6-phosphate isomerase (G6I) is plays a crucial role in glycolysis, gluconeogenesis and pentose phosphate pathway regulation. These pathways are important for ATP production in this parasite. Thus targeting G6I seems to promising for drug discovery and design as anti*leishmanial* therapy. Active site analysis of human G6PI (PDB ID 1IAT) shows that it is more deep and its electrostatic potential also differs with *Leishmania* G6PI (modbase ID Q4QGN9) (Figure 1 and 2).

Phosphomannomutase (PMM) is responsible for mannose-6-phosphate to mannose-1-phosphate which helps in biosynthesis of glycoconjugates. These glycoconjugates form cell surface of prokaryote. Thus PMM consider as potential novel target in anti-*Leishmania* drug development. Kedzierski *et al* suggest that PMM inhibitors design challenging mission because of similarity between human and *Leishmania* PMM [1]. Human PMM (PDB ID 2FUC) probable active site has Asp12, Ser47, Lys189, Asp218 residues (Figure 3A) and *L. mexicana* PMM (PDB ID 2AMY) shows the presence of Asp10, Lys188, Ser46 residues (Figure 3B) respectively both the results are according to uniprot database and experimental result of Handman *et al.* [1, 2]. *L. mexicana* PMM active site volume is larger and deeper than human Phosphomannomutase (Figure 4A and 4B). There electrostatic potential difference is clearly visible in Figure 4C and 4D.

Binding site

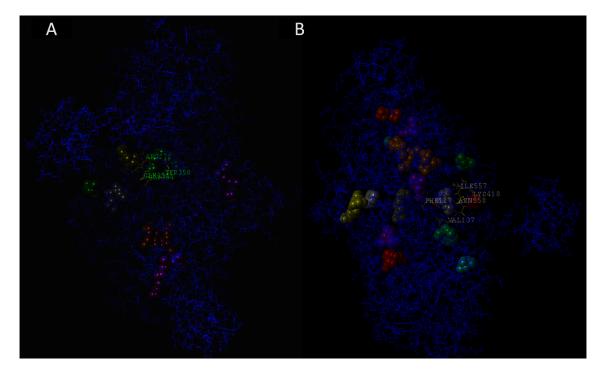


Figure 1. Glucose 6 phosphate isomerise binding site analysis (A) Human Glucose 6 phosphate isomerise (B) *Leishmania* Glucose 6 phosphate isomerise

Cavity Depth and MEP analysis

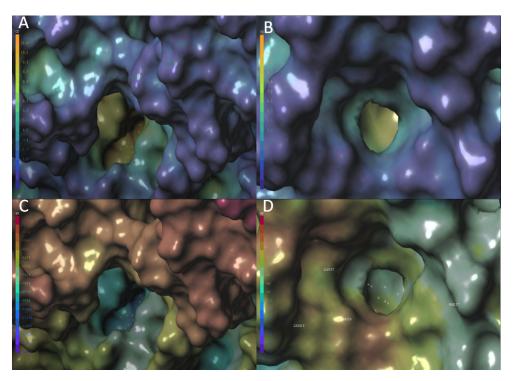


Figure 2 Glucose 6 phosphate isomerise cavity depth and MEP analysis (A) Human Glucose 6 phosphate isomerise cavity depth is high compare to *Leishmania* G6PI (B) *Leishmania* Glucose 6 phosphate isomerise cavity is relatively narrow (C) Human Glucose 6 phosphate isomerise MEP surface (D) *Leishmania* Glucose 6 phosphate isomerise MEP surface

Binding site

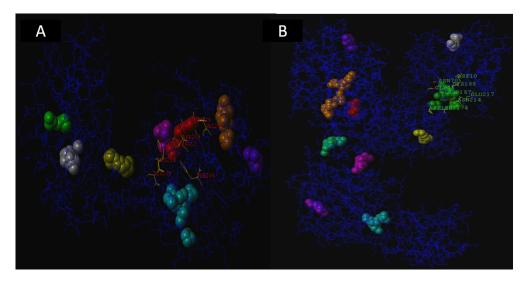


Figure 3 Phosphomannomutase binding site analysis (A) Analysis shows eight binding sites of Human Phosphomannomutase (B) ten binding sites are identified in *Leishmania* Phosphomannomutase

Cavity Depth Analysis and MEP analysis

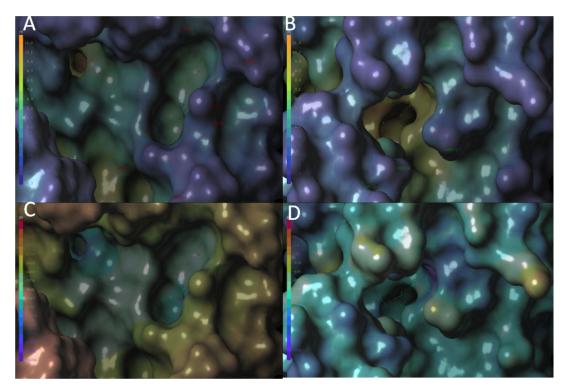


Figure 4 Phosphomannomutase active site analysis (A) Human Phosphomannomutase cavity depth (B) *Leishmania* Phosphomannomutase cavity depth was higher (C) Human Phosphomannomutase MEP surface (D) *Leishmania* Phosphomannomutase MEP surface

Druggability Analysis

DogSiteScorer is a new algorithm for predicting the pockets and its druggability which is highly important in pharmaceutical research. Druggability score is ranges from zero to one. The higher score indicates more druggablity of the pocket. Apart from that it also calculates the volume, surface lipo surface and depth of pocket [3].

Results

Druggablity results of Homoserine kinase, L-ribulokinase and Phospholipid:diacylglycerol acyltransferase are promising and gives better scope for rational drug designing (Corresponding figures 5,6 and 7). Druggability of *leishmania* targets is given in table 1 and 2.

S.	Non	Total	Pockets	Volume	Surface	Lipo	Depth	Drug
No.	Homologous	No. of		[ų]	[Ų]	surface	[Å]	Score
	Targets	Pockets				[Ų]		

1	Phospholipid:	17	P0	3787.70	4461.43	3121.1	38.40	0.81
	diacylglycerol					2		
	acyltransferase,		P1	332.99	674.11	302.96	16.15	0.67
	putative							
2	Homoserine	10	P0	950.21	1324.74	824.22	25.77	0.82
	kinase, putative		P1	770.69	984.92	819.60	20.81	0.84
3	L-ribulokinase,	18	P0	1805.85	2043.88	1310.5	26.71	0.81
	putative					5		
			P1	897.56	1157.53	822.44	24.99	0.84

Table 2 Other Leishmanial targets druggablity analysis

S.	Non	Total	Pockets	Volume	Surface	Lipo	Depth	Drug
No.	Homologous	No. of		[ų]	[Ų]	surface	[Å]	Score
	Targets	Pockets				[Ų]		
1	Nucleoside 16 diphosphate	16	PO	871.37	1168.96	712.42	18.25	0.57
	kinase B (NDKb)		P1	849.58	953.35	588.92	21.19	0.54
2	Glucose-6- phosphate isomerase (G6I)	20	PO	691.20	765.84	495.81	24.53	0.38
			P1	447.83	545.82	379.04	20.36	0.28
3	Phosphomanno mutase	11	PO	365.31	417.08	231.14	12.33	0.21
			P1	318.14	720.48	440.07	18.44	0.18

References

- 1. L. Kedzierski, R. L. Malby, B. J. Smith, M. A. Perugini, A. N. Hodder, T. Ilg, P. M. Colman, E. Handman, *J. Mol. Biol.*, 2006, **363**, 215-27.
- 2. The UniProt Consortium, Nucl. Acids Res., 2012, 40, D71-5
- 3. A. Volkamer, D. Kuhn, T. Grombacher, F. Rippmann, M. Rarey, J. Chem. Inf. Model., 2012, **52**, 360-372.