

# A red light-controlled synthetic gene expression switch for plant systems

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<b>Supplementary Table 1</b>	Expression vectors and oligonucleotides designed and used in this study.
<b>Supplementary Figure 1</b>	Effect of clarithromycin on constitutive gene expression in <i>N. tabacum</i>
<b>Supplementary Figure 2</b>	Spectrum of the white light source

## Supplementary Table 1

### Expression vectors and oligonucleotides designed and used in this study.

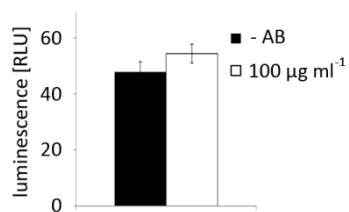
Plasmid	Description	Ref. or source
auxin sensor	Plasmid encoding P <sub>CaMV35S</sub> -controlled optimized auxin sensor (P <sub>CaMV35S</sub> -RLuc-p2A-SM[L2min17]-FLuc-pA)	1
pKM002	Vector encoding SEAP under control of a modified P <sub>Tet</sub> (tetO <sub>13</sub> -394bp-P <sub>hCMVmin</sub> -SEAP-pA)	2
pKM006	Vector encoding SEAP under the control of a modified P <sub>Tet</sub> (tetO <sub>13</sub> -422bp-P <sub>hCMVmin</sub> -SEAP-pA)	2
pKM022	Bicistronic vector encoding PhyB(1-650)-VP16-NLS and TetR-PIF6(1-100)-HA under control of P <sub>SV40</sub> (P <sub>SV40</sub> -PhyB(1-650)-VP16-NLS-IRES <sub>pV</sub> -TetR-PIF6(1-100)-HA-pA)	2
pKM033	Vector encoding VEGF <sub>121</sub> under control of a modified P <sub>Tet</sub> (tetO <sub>13</sub> -422bp-P <sub>hCMVmin</sub> -VEGF <sub>121</sub> -pA)	2
pKM081	Vector encoding SEAP under control of a modified P <sub>ETR</sub> (etr <sub>8</sub> -P <sub>hCMVmin</sub> -SEAP-pA)	3
pKM082	Vector encoding SEAP under control of a modified P <sub>ETR</sub> (etr <sub>8</sub> -386bp-P <sub>hCMVmin</sub> -SEAP-pA) A 372 bp fragment was amplified from <i>CFP</i> using oligos oKM090 (5'-caagtagctagcCCCTGAAGTTCATCTGCACC-3') and oKM003 (5'-caagtcgctagcTCTTGAAGTTGGCCTTGATGC-3'), digested ( <i>NheI</i> ) and ligated ( <i>NheI</i> ) into pKM081.	This work
pKM271	Vector for P <sub>CaMV35S</sub> -controlled expression of PiP-VP16-NLS (P <sub>CaMV35S</sub> -PiP-VP16-NLS-pA) PiP-VP16-NLS was amplified from pMF156 using oligos oKM373 (5'-caagtcaccggatcatatggccaccATGAGTCGAGGAGAGGTGCG-3') and oKM374 (5'-caagtcgattctacaccttctcttcttcttggcCCACCGTACTCGTCAATTCC-3'), digested ( <i>NdeI/EcoRI</i> ) and ligated ( <i>NdeI/EcoRI</i> ) into pMZ824.	This work
pKM272	Vector encoding FLuc under control of a modified P <sub>PTR</sub> (PIR <sub>3</sub> -P <sub>HSP70min</sub> -FLuc-pA) FLuc was excised ( <i>EcoRI/HindIII</i> ) from pMZ836 and ligated ( <i>EcoRI/HindIII</i> ) into pMF199.	This work
pKM295	Vector encoding VEGF <sub>121</sub> under the control of a modified P <sub>ETR</sub> (etr <sub>8</sub> -P <sub>hCMVmin</sub> -VEGF <sub>121</sub> -pA) VEGF <sub>121</sub> was excised ( <i>EcoRI/NotI</i> ) from pKM033 and ligated ( <i>EcoRI/NotI</i> ) into pMZ836.	This work
pKM300	Bicistronic vector encoding PhyB(1-650)-VP16-NLS and E-PIF6(1-100)-HA under control of P <sub>SV40</sub> (P <sub>SV40</sub> -PhyB(1-650)-VP16-NLS-IRES <sub>pV</sub> -E-PIF6(1-100)-HA-pA) IRES <sub>pV</sub> was amplified from pKM022 using oligos oKM400 (5'-ACCCACCCAGAGGCC-3') and oKM401 (5'-gatatcGCCGCAATCCAATTCGCTTTATG-3'), while PIF6(1-100)-HA was amplified from pKM022 using oligos oKM402 (5'-ATGATGTTCTTACCAACCGATTATTGTTG-3') and oKM403 (5'-AAAAACCTCCCACACCTCCCC-3'). E was amplified from pWW043 using oligos oKM404 (5'-atcacagattgtatcataaagcgaattggattgctggcgatatcgCCACCATGCCCGCCCC-3') and oKM405 (5'-tgatcgcttaacctgcaacaataatcggttgtaagaacatcataccagcactaccagcactaccagcactgtaattaaGCTGTACGCGGACGCATGTG-3') and the three fragments were cloned into <i>KpnI/MfeI</i> -digested pKM022 by Gibson cloning.	This work
pKM301	Bicistronic vector encoding PhyB(1-650)-VP16-NLS and PiP-PIF6(1-100)-HA under control of P <sub>SV40</sub> (P <sub>SV40</sub> -PhyB(1-650)-VP16-NLS-IRES <sub>pV</sub> -PiP-PIF6(1-100)-HA-pA) IRES <sub>pV</sub> and PIF6(1-100) were amplified as described for pKM300. PiP was amplified from pMF150 using oligos oKM406 (5'-atcacagattgtatcataaagcgaattggattgctggcgatatcgCCACCATGAGTCGAGGAGAG-3') and oKM407 (5'-tgatcgcttaacctgcaacaataatcggttgtaagaacatcataccagcactaccagcactaccagcactgtaattaaGGCCTGTTCGACCATCGC-3') and all fragments were cloned into <i>KpnI/MfeI</i> -digested pKM022 by Gibson cloning.	This work
pMF150	Vector encoding PiP under control of P <sub>hCMV</sub> (P <sub>hCMV</sub> -PiP-pA)	4
pMF156	Vector encoding PiP-VP16 under control of P <sub>SV40</sub> (P <sub>SV40</sub> -PiP-VP16-pA)	4
pMF199	Vector encoding SEAP under control of a modified P <sub>PTR</sub> (PIR <sub>3</sub> -P <sub>HSP70min</sub> -SEAP-pA)	4
pMK052	Vector encoding P <sub>EF1α</sub> -controlled TIR1 (P <sub>EF1α</sub> -TIR1-pA)	5
pMZ802	Vector encoding FLuc under control of a modified P <sub>Tet</sub> (tetO <sub>13</sub> -P <sub>hCMVmin</sub> -FLuc-pA) FLuc was amplified from pSW209 using oligos oMZ807 (5'-tcagcttccgctcaattggctGCGCTGGCGCTAGCATGGAAG-3') and oMZ808 (5'-gggaccagcagccatggcagGGTTTACACGGCGATCTTTCCGCC-3'), while the backbone of pKM002 was amplified using oMZ809 (5'-gtgtaaacccgtccatggcgtCGTGGTCCCCGCGTTGCTTC-3') and oMZ810 (5'-cgccagcagccaattgagcGGAAGCTGACTCTAGAGGATCCCC-3'). Finally, both fragments were fused by Gibson cloning.	This work
pMZ824	Vector for P <sub>CaMV35S</sub> -controlled expression of E-VP16-NLS (P <sub>CaMV35S</sub> -E-VP16-NLS-pA) E-VP16-NLS was amplified from pWW035 using oligos oMZ809 (5'-	This work

	gtgtaaaccgctccatggcgtCGTGGTCCCCGCGTTGCTTC-3') and oMZ8123 (5'-tcacgtcgtgctagccgttgcttacaccttctcttcttcttggCCCACCGTACTCGTCAATTCCAAG-3'), while the backbone of pSW209 was amplified using oligos oMZ873 (5'-gcaacggctagcagcagctgaTGC GGCAGCGGCCGAATTCC-3') and oMZ874 (5'-tggtgcatatgcgtcgttcaGGTGGTACCAAGCTTACCTAGCC-3'). Finally, both fragments were fused by Gibson cloning.	
pMZ827	Vector encoding P <sub>CaMV35S</sub> -controlled nuclear-targeted E-PIF6(1-100) (P <sub>CaMV35S</sub> -E-PIF6(1-100)-NLS-pA) E-Pif6 was amplified from pKM300 using oligos oMZ895 (5'-tgaacgacgcatatgacaacaATGCCCGCCCAAGCTCAAG-3') and oMZ8127 (5'-tcacgtcgtgctagccgttgcttacaccttctcttcttcttggGTCAACATGTTTATTGCTTCCAACATGTTTG-3'), while the backbone of pSW209 was amplified using oligos oMZ873 and oMZ874. Finally, both fragments were fused by Gibson cloning.	This work
pMZ828	Vector encoding P <sub>CaMV35S</sub> -controlled nuclear-targeted PhyB(1-650)-VP16 (P <sub>CaMV35S</sub> -PhyB(1-650)-VP16-NLS-pA) PhyB-VP16-NLS was amplified from pKM300 using oligos oMZ856 (5'-gccatggtgagcagcGTGACTTAGATCACACCTCCG-3') and oMZ8123 (5'-tcacgtcgtgctagccgttgcttacaccttctcttcttcttggCCCACCGTACTCGTCAATTCCAAG-3'), while the backbone of pSW209 was amplified using oligos oMZ873 and oMZ874. Finally, both fragments were fused by Gibson cloning.	This work
pMZ833	Vector for P <sub>CaMV35S</sub> -controlled expression of TetR-VP16-NLS (P <sub>CaMV35S</sub> -TetR-VP16-NLS-pA) Tet-VP16 was amplified from pSAM200 using oligos oMZ891 (5'-tgaacgacgcatatgacaacaCGGCCGCCACCATGTCTAGATTAG-3') and oMZ8123 (5'-tcacgtcgtgctagccgttgcttacaccttctcttcttcttggCCCACCGTACTCGTCAATTCCAAG-3'), while the backbone of pSW209 was amplified using oligos oMZ873 and oMZ874. Finally, both fragments were fused by Gibson cloning.	This work
pMZ836	Vector encoding FLuc under control of a modified P <sub>ETR</sub> (etr <sub>8</sub> -P <sub>hCMVmin</sub> -FLuc-pA) FLuc was amplified from pSW209 using oligos oMZ807 and oMZ808, while the backbone of pKM081 was amplified using oligos oMZ809 (5'-gtgtaaaccgctccatggcgtCGTGGTCCCCGCGTTGCTTC-3') and oMZ810 (5'-cgccagcgcagccaattgagcGGAAGCTGACTCTAGAGGATCCCC-3'). Finally, both fragments were fused by Gibson cloning.	This work
pMZ837	Vector encoding P <sub>CaMV35S</sub> -driven expression of miRNA <sub>TIR1</sub> (P <sub>CaMV35S</sub> -miRNA <sub>TIR1</sub> -pA) For the design of an miRNA targeting <i>N. tabacum</i> TIR1 the online tool CentroidFold <sup>6</sup> was used and miRNA cloning was performed by a modification of a previously described protocol. <sup>7</sup> First, the 5'-stem sequence was amplified from pRS300 <sup>7</sup> using oligos oMZ880 (5'-tgaacgacgcatatgacaacaGAGGTCGACGGTATCGATAAGCTTG-3') and oMZ8163 (5'-cggtagacaaattggatcattgattctcttgggtgcaactgactgattgtCTCTCTTTTTGTATTCCAATTTCTT-3'). In doing so, the miRNA that was contained in the 5'-overhang of oMZ1861 was introduced. In the same way, the 3'-stem sequence was amplified from pRS300 using oligos oMZ883 (5'-tcacgtcgtgctagccgttgCGGATAACAATTTACACAGGAAACAG-3') and oMZ8164 (5'-gaagctaattgaatcatatcagcactgtgagtgctcaactgacaggattgaTCTACATATATTCCTAAAACATCAAA-3'). Next, both PCR-products were extended by overlapping loop sequences by amplification with oMZ880 and oMZ882 (5'-cgagtctagtttgaatttggcgactcggtatttggatgaatgagtcgGAAGCTAATTGAATCATATCACGACCTGTGAG-3') or oMZ883 and oMZ885 (5'-cgagtcgcaaaattcaactagactcgtaaatgaatgaatgatgCGGTAGACAAATTGGATCATTGATTCTCTTTG-3'), respectively. Finally, both fragments were fused and introduced into the oMZ873/oMZ874-amplified backbone of pSW209 by Gibson cloning.	This work
pMZ839	Vector encoding miRNA <sub>TIR1</sub> under control of a modified P <sub>ETR</sub> (etr <sub>8</sub> -P <sub>hCMVmin</sub> -miRNA <sub>TIR1</sub> -pA) miRNA <sub>TIR1</sub> was amplified from pMZ837 using oligos oMZ8118 (5'-gctcaattggctgctgctggcCCACCTGAACGACGCATATGACAAC-3') and oMZ8119 (5'-acgcatggagcgggtttacacCATCACGTCTGCTAGCCGTTGC-3') and fused with the oMZ809/oMZ810-amplified backbone of pKM081 by Gibson cloning.	This work
pMZ841	Vector encoding TIR1 under control of a modified P <sub>ETR</sub> (etr <sub>8</sub> -P <sub>hCMVmin</sub> -TIR1-pA) TIR1 was excised ( <i>EcoRI/XbaI</i> ) from pMK052 and ligated ( <i>EcoRI/SpeI</i> ) into pKM081.	This work
pSW209	Vector encoding firefly luciferase and renilla luciferase separated by a 2A-peptide under control of P <sub>CaMV35S</sub> (P <sub>CaMV35S</sub> -FLuc-p2A-RLuc-pA).	1
pWW035	Vector encoding P <sub>SV40</sub> -driven expression of E-VP16 (P <sub>SV40</sub> -E-VP16-pA)	8
pWW043	Vector encoding P <sub>SV40</sub> -driven expression of E-KRAB (P <sub>SV40</sub> -E-KRAB-pA)	8

E, macrolide-responsive repressor protein; *etr*, operator sequence binding E; FLuc, firefly luciferase; HA, human influenza hemagglutinin-derived epitope tag; IRES<sub>PV</sub>, polioviral internal ribosome entry site; KRAB, transcriptional repressor domain from human Kox1; NLS, nuclear localization signal from simian virus 40 large T antigen; pA, polyadenylation signal; p2A, foot-and-mouth disease virus-derived self-processing 2A peptide; P<sub>CaMV35S</sub>, cauliflower mosaic virus 35S promoter; PiP, pristinamycin-induced protein, P<sub>hCMV</sub>, human cytomegalovirus immediate early promoter; P<sub>hCMVmin</sub>, minimal human cytomegalovirus immediate early promoter; P<sub>HSP70min</sub>, minimal heat-shock protein 70 promoter from *Drosophila*; PhyB, Phytochrome B; PhyB(1-650), N-terminus of Phytochrome B with amino acids 1-650; PIF6, Phytochrome-interacting-factor 6; PIF6(1-100), N-terminus of Phytochrome-interacting-factor 6 with amino acids 1-100; P<sub>SV40</sub>, simian virus 40 early promoter; PIR, operator sequence binding PiP; P<sub>Tet</sub>, tetracycline-responsive promoter; RLuc, renilla luciferase; SEAP, human placental secreted alkaline phosphatase; SM, auxin sensor module; tetO, operator sequence binding TetR; TetR, tetracycline repressor protein; TIR1, auxin receptor transport inhibitor response 1; VEGF<sub>121</sub>, 121 amino acids splice variant of human vascular endothelial growth factor; VP16, *Herpes simplex* virus-derived transactivation domain.

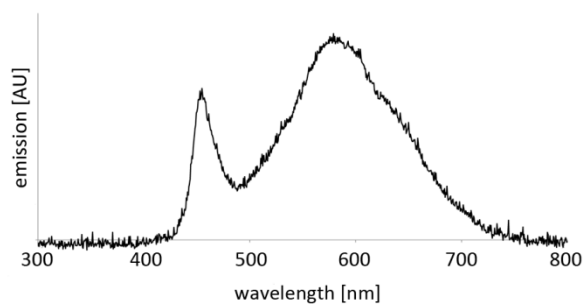
Uppercase in oligos, annealing sequence; underlined sequence, restriction site.

## Supplementary Figure 1



**Effect of clarithromycin on constitutive gene expression in *N. tabacum*.** 125,000 protoplasts were transformed for constitutive firefly luciferase expression. After incubation for 24 h in the absence (-AB) or presence of 100 µg ml<sup>-1</sup> clarithromycin, the firefly luciferase luminescence was quantified. Data are means ± SEM (n=12).

## Supplementary Figure 2



**Spectrum of the white light source.** The light spectrum between 300 nm and 800 nm was recorded using an Avaspec-ULS2048 spectroradiometer (Avatec).

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