

Glycosylated Porphyrin Derivatives and Their Photodynamic Activity in Cancer Cells

Seenuvasan Vedachalam,^a Bo-Hwa Choi,^b Kalyan Kumar Pasunooti,^a Kun Mei Ching,^b Kijoon Lee,^c Ho Sup Yoon,^{*b} Xue-Wei Liu,^{*a}

^a*Division of Chemistry and Biological Chemistry, School of Physical and Mathematical Sciences, Nanyang Technological University, 21 Nanyang Link, Singapore 637371*

^b*Division of Structural and Computational Biology, School of Biological Sciences School of Biological Sciences, Nanyang Technological University, 60 Nanyang Drive, Singapore 637551*

^c*Division of Bioengineering, School of Chemical and Biomedical Engineering, 62 Nanyang Drive, Nanyang Technological University, Singapore 637457*

Supporting Information

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Experimental Section

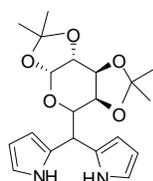
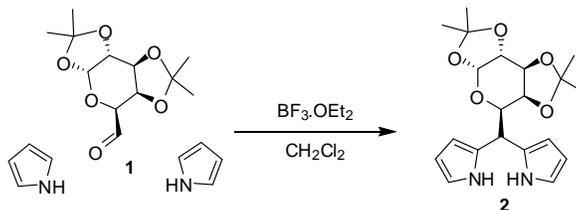
General method and materials

General: All the reactions were carried out in a flame or oven dried glassware under an argon or nitrogen atmosphere with freshly distilled dry solvents under anhydrous conditions unless otherwise indicated. Evaporation of organic solutions was achieved by rotary evaporation with a water bath temperature below 40 °C. Product purification by flash column chromatography was accomplished using silica gel 60 (0.010–0.063 nm). Analytical thin-layer chromatography was performed on E. Merck silica gel 60 F₂₅₄ plates (0.25 mm). Chromatograms were visualized by fluorescence quenching with UV light at 254 nm or by staining using base solution of potassium permanganate. Porphyrinic compounds were visualized as green emerald spots by dipping in a solution of Ce(III)sulfate (1.0 g), ammonium molybdate (21.0 g), 96% sulfuric acid (31.0 mL), and distilled water (500 mL). IR spectra were recorded using FTIR Restige-21 (Shimadzu). NMR spectra were recorded at room temperature on 300 MHz Bruker ACF 300, 400 MHz Bruker DPX 400, 500 MHz Bruker AMX 500, and 400 MHz JEOL ECA 400 NMR spectrometers. The residual solvent signals were taken as the reference (7.26 ppm for ¹H NMR spectra and 77.0 ppm for ¹³C NMR spectra in CDCl₃). Sometimes the TMS signal at 0.0 ppm was used as an internal standard for ¹H NMR spectra. Chemical shift (δ) is reported in ppm, coupling constants (J) are given in Hz. The following abbreviations classify the multiplicity: s = singlet, d = doublet, t = triplet, m = multiplet or unresolved, br = broad signal. HRMS (ESI) spectra were recorded on a Finnigan/MAT LCQ quadrupole ion trap mass spectrometer, coupled with the TSP4000 HPLC system and the Crystal 310 CE system.

Materials: All solvents were distilled under argon from the following drying agents immediately before use: Dichloromethane was distilled from calcium hydride. Technical grade solvents were used for chromatography and were distilled prior to use. All benzaldehyde were purchased from commercial suppliers and used without further purification. Sugar aldehyde was prepared galactose isopropylidene protection¹ followed by IBX oxidation². BF₃·Et₂O solution and DDQ were purchased from commercial suppliers and used without further purification. Starting material dipyrrolyl methane unit (**2**) was prepared from condensation freshly distilled pyrrole with 1,2:3,4-di-*O*-isopropylidene- α -D-galacto-hexadialdo-1,5-pyranose (**1**) purified through silica gel column and perfectly dried prior to use.

Synthesis and spectral details of dipyrryl methane:

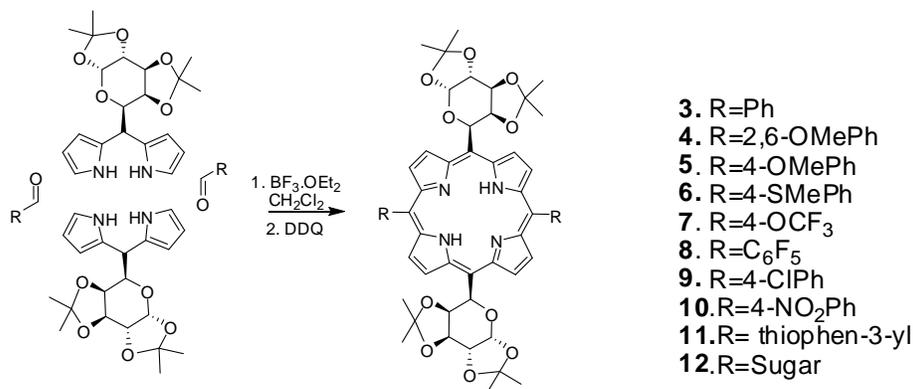
Synthesis of 1,2:3,4-di-*O*-isopropylidene-5,5-dipyrryl-6-deoxy- α -D-galactopyranose (**2**):



To a solution of freshly distilled pyrrole (1.35 mL, 19.38 mmol) and 1,2:3,4-di-*O*-isopropylidene- α -D-galacto-hexadialdo-1,5-pyranose (**1**) (1 g, 3.88 mmol) in CH_2Cl_2 (100 mL) at ambient temperature with stirring under N_2 was added $\text{BF}_3 \cdot \text{etheral}$ solution (48 μL , 0.39 mmol). After 3 h stirring, the bright orange reaction mixture was quenched by addition of a saturated aqueous NaHCO_3 solution (10 mL) and then diluted with CH_2Cl_2 (100 mL). The organic layer was separated, washed with water (2x50 mL). Then the combined organic layer were dried (MgSO_4), filtered, evaporated, and purified by flash chromatography (7:3 hexane/EtOAc) to give 913 mg (63%) of **2** as a white solid; $^1\text{H NMR}$ (300 MHz, CDCl_3): δ 8.83 (s, 1H, NH), 8.49 (s, 1H, NH), 6.71-6.69 (m, 2H, Py-CH), 6.15-6.13 (m, 2H, Py-CH), 6.09 (d, $J = 1.1$ Hz, 1H, Py-CH), 6.02 (s, 1H, Py-CH), 5.65 (d, $J = 5.0$ Hz, 1H, Sug-CH), 4.55 (dd, $J_1 = 8.0$ Hz, $J_2 = 2.3$ Hz), 4.48 (d, $J = 10.0$ Hz, 1H, Sug-CH), 4.32 (dd, $J_1 = 5.0$ Hz, $J_2 = 2.3$ Hz, 1H, Sug-CH), 4.13 (dd, $J_1 = 10.0$ Hz, $J_2 = 1.3$ Hz, 1H), 3.91 (dd, $J_1 = 8.0$ Hz, $J_2 = 1.6$ Hz, 1H, Sug-CH), 1.55 (s, 3H, $-\text{CH}_3$), 1.51 (s, 3H, $-\text{CH}_3$), 1.35 (s, 6H, $-\text{CH}_3$); $^{13}\text{C NMR}$ (75MHz, CDCl_3): δ 131.0, 129.6, 116.6, 116.5, 109.1, 108.8, 108.1, 107.7, 107.6, 107.0, 96.9, 71.6, 70.8, 70.7, 70.3, 38.1, 25.9, 25.8, 24.8, 24.5; **IR** (neat): ν_{max} 3417, 1643, 1384, 1213, 717 cm^{-1} ; **HRMS** (ESI): m/z ($\text{M}+\text{H}$) $^+$ Calcd for $\text{C}_{20}\text{H}_{27}\text{N}_2\text{O}_5$: 375.1920, found: 375.1917.

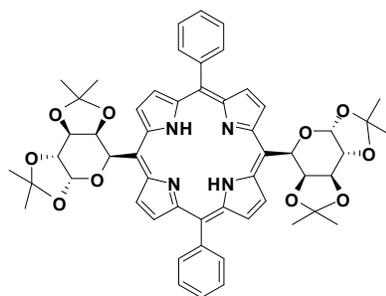
Synthesis of sugar porphyrin conjugates & spectral details:

General procedure:



To a solution of 1,2:3,4-di-*O*-isopropylidene-5,5-dipyrrolyl-6-deoxy- α -D-galactopyranose (**2**) (200 mg, 0.53 mmol) in 250 mL of CH_2Cl_2 were added sequentially aromatic aldehyde (0.53 mmol) and $\text{BF}_3 \cdot \text{etheral}$ solution (6.7 μL , 0.05 mmol) while a stream of pure argon was passing. The reaction vessel was carefully shielded from light, and stirring was continued for 3 h. Then, triethylamine (7.4 μL , 0.05 mmol) and 2,3-dichloro-5,6-dicyano-*p*-benzoquinone (DDQ) (132.90 mg, 0.59 mmol) were added, and the reaction mixture was stirred at room temperature for an additional 3 h. The solvent was evaporated under vacuum, and the resulting dark-violet solid was purified by column chromatography on silica gel to give porphyrin compound as a purple solid (5-16% yields).

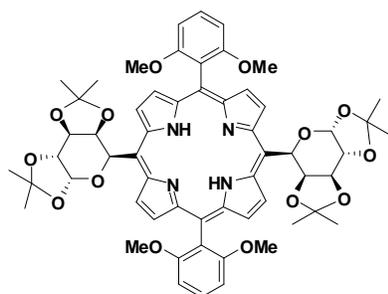
5,15-[Bis(phenyl)]-10 α ,20 β -[bis(1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose-6-yl)] porphyrin (**3**):



Prepared according to general procedure using benzaldehyde; Purple solid; (58 mg, 12% yield); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ 9.69 (d, $J = 4.2$ Hz, 4H, H- β), 8.82 (d, $J = 4.8$ Hz, 4H, H- β), 8.19 (d, $J = 6.3$ Hz, 4H, Ph-CH), 7.79-7.72 (m, 4H, Ph-CH), 7.68 (s, 2H, H-5'), 6.26 (d, $J = 5.1$ Hz, 2H, H-1'), 5.21 (d, $J = 1.2$ Hz, 2H, H-4'), 5.14 (d, $J = 1.8$ Hz, 2H, H-3'), 4.79 (dd, $J_1 = 5.0$ Hz, $J_2 = 2.0$ Hz, 2H, H-2'), 1.86 (s, 6H, -CH₃), 1.7 (s, 6H, -CH₃), 1.55 (s, 6H, -CH₃), 1.19 (s, 6H, -CH₃), -2.67 (s, 2H, NH); $^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ 162.3, 143.3, 134.5, 131.7, 129.6,

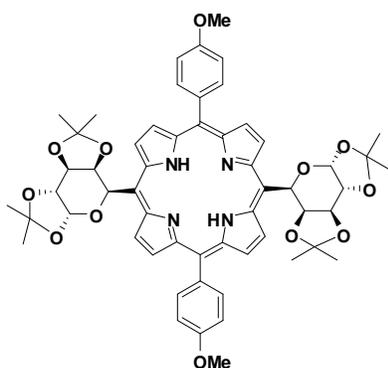
127.5, 126.3, 119.4, 113.7, 109.6, 109.0, 97.8, 71.9, 71.4, 26.8, 25.9, 25.1, 23.4; **HRMS** (ESI): m/z (M+H)⁺ Calcd for C₅₄H₅₅N₄O₁₀: 919.3918, found: 919.3948; **UV-VIS** (CHCl₃) λ_{max} (log ϵ): 406 (4.478), 516 (4.136), 549 (3.749), 589 (3.672), 644 (3.549); **IR** (neat): ν_{max} 3437, 2989, 1732, 1597, 1483, 1382, 1064, 802 cm⁻¹.

5,15-[Bis(2,6-dimethoxyphenyl)]-10 α ,20 β -[bis(1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose-6 yl)]porphyrin (4):



Prepared according to general procedure using 2,6-dimethoxybenzaldehyde; Purple solid; (72 mg, 13% yield); **¹H NMR** (300 MHz, CDCl₃): δ 9.59 (s, 4H, H- β), 8.77 (d, J = 4.8 Hz, 4H, H- β), 7.72 (t, J = 8.5 Hz, 2H, Ph-CH), 7.64 (s, 2H, H-5'), 7.01-6.99 (m, 4H, Ph-CH), 6.23 (d, J = 4.9 Hz, 2H, H-1'), 5.26 (d, J = 7.8 Hz, 2H, H-4'), 5.8 (dd, J_1 = 7.8 Hz, J_2 = 1.9 Hz, 2H, H-3'), 4.76 (dd, J_1 = 4.9 Hz, J_2 = 2.0 Hz, 2H, H-2'), 3.49 (s, 12H, -OCH₃), 1.84 (s, 6H, -CH₃), 1.70 (s, 6H, -CH₃), 1.52 (s, 6H, -CH₃), 1.21 (s, 6H, -CH₃), -2.48 (s, 2H, NH); **¹³C NMR** (125 MHz, CDCl₃): δ 160.6, 146.5, 145.2, 130.7, 129.8, 129.5, 121.3, 112.3, 111.3, 109.4, 108.9, 104.2, 97.7, 76.6, 71.9, 71.8, 71.4, 56.0, 26.8, 25.9, 25.1, 23.4; **HRMS** (ESI): m/z (M+H)⁺ Calcd for C₅₈H₆₃N₄O₁₄: 1039.4341, found: 1039.4347; **UV-VIS** (CHCl₃) λ_{max} (log ϵ): 412 (4.533), 516 (4.205), 546 (3.607), 590 (3.768), 644 (3.526); **IR** (neat): ν_{max} 3435, 2927, 1633, 1469, 1382, 1249, 1109, 1064 cm⁻¹.

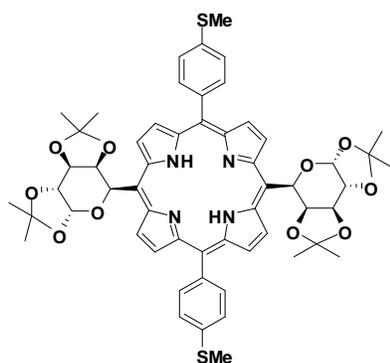
5,15-[Bis(4-methoxyphenyl)]-10 α ,20 β -[bis(1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose-6-yl)]porphyrin (5):



Prepared according to general procedure using 4-methoxybenzaldehyde; Purple solid; (41 mg, 8% yield); **¹H NMR** (500 MHz, CDCl₃): δ 9.67 (s, 4H, H- β), 8.84 (d, J = 4.6 Hz, 4H, H- β), 8.08 (d, J = 8.0 Hz, 4H, Ph-CH), 7.66 (s, 2H, H-5'), 7.24 (d, J = 7.6 Hz, 4H, Ph-CH), 6.25 (d, J = 4.8 Hz, 2H, H-1'), 5.21 (d, J = 7.8 Hz, 2H, H-4'), 5.12 (d, J = 6.7 Hz, 2H, H-3'), 4.78 (d, J = 3.1 Hz, 2H, H-2'), 4.08 (s, 6H, -CH₃), 1.85 (s, 6H, -CH₃), 1.68 (s, 6H, -CH₃), 1.53 (s, 6H, -CH₃), 1.17 (s, 6H, -CH₃), -2.69 (s, 2H, NH); **¹³C NMR**

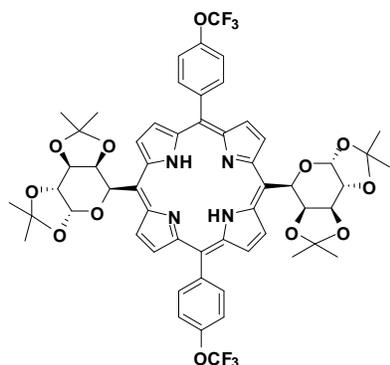
(125 MHz, CDCl₃): δ 159.3, 135.7, 135.5, 119.2, 113.5, 112.0, 111.8, 109.6, 109.0, 97.8, 71.9, 71.4, 55.6, 26.8, 25.9, 25.1, 23.4. **HRMS** (ESI): m/z (M+H)⁺ Calcd for C₅₆H₅₉N₄O₁₂: 979.4129, found: 979.4128; **UV-VIS** (CHCl₃) λ_{max} (log ϵ): 414 (4.494), 518 (3.514), 550 (3.412), 590 (3.160), 646 (2.890). **IR** (neat): ν_{max} 3435, 2922, 1643, 1462, 1379, 1247, 1174, 1066 cm⁻¹.

5,15-[Bis(4-(methylthio)phenyl)]-10 α ,20 β -[bis(1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose-6-yl)]porphyrin (6):



Prepared according to general procedure using 4-(methylthio)benzaldehyde; Purple solid; (37 mg, 7% yield); **¹H NMR** (300 MHz, CDCl₃): δ 9.70 (d, J = 3.6 Hz, 4H, H- β), 8.86 (d, J = 4.9 Hz, 4H, H- β), 8.11 (d, J = 8.1 Hz, 4H, Ph-CH), 7.69 (s, 2H, H-5'), 7.64 (d, J = 8.3 Hz, 4H, Ph-CH), 6.27 (d, J = 5.0 Hz, 2H, H-1'), 5.22 (d, J = 8.2 Hz, 2H, H-4'), 5.15 (d, J = 7.8 Hz, 2H, H-3'), 4.80 (dd, J_1 = 5.0 Hz, J_2 = 1.8 Hz, 2H, H-2'), 2.78 (s, 6H, -CH₃), 1.87 (s, 6H, -CH₃), 1.71 (s, 6H, -CH₃), 1.56 (s, 6H, -CH₃), 1.20 (s, 6H, -CH₃), -2.69 (s, 2H, NH); **¹³C NMR** (125 MHz, CDCl₃): δ 140.0, 138.0, 135.1, 134.8, 131.6, 129.6, 125.2, 124.2, 118.8, 113.7, 109.6, 109.0, 107.0, 105.1, 97.8, 71.9, 71.3, 26.7, 25.8, 25.1, 23.3, 15.92; **HRMS** (ESI): m/z (M+H)⁺ Calcd for C₅₆H₅₉N₄O₁₀S₂: 1011.3673, found: 1011.3665; **UV-VIS** (CHCl₃) λ_{max} (log ϵ): 414 (4.540), 518 (4.282), 550 (3.817), 591 (3.796), 645 (3.653); **IR** (neat): ν_{max} 3439, 2924, 1643, 1456, 1382, 1257, 1163, 1066 cm⁻¹.

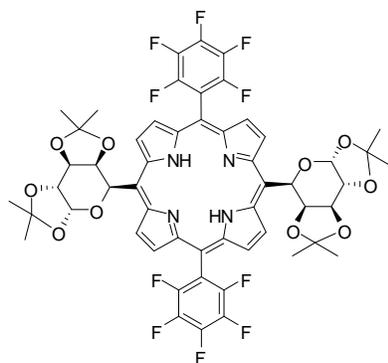
5,15-[Bis(4-trifluoromethoxyphenyl)]-10 α ,20 β [bis(1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose-6-yl)]porphyrin (7):



Prepared according to general procedure using 4-trifluoromethoxybenzaldehyde; Purple solid; (29 mg, 5% yield); **¹H NMR** (500 MHz, CDCl₃): δ 9.71 (s, 4H, H- β), 8.77 (d, J = 4.8 Hz, 4H, H- β), 8.20 (d, J = 8.2 Hz, 4H, Ph-CH), 7.67 (s, 2H, H-5'), 7.60 (d, J = 7.9 Hz, 4H, Ph-CH), 6.25 (d, J = 4.9 Hz, 2H, H-1'), 5.20 (dd, J_1 = 7.9 Hz, J_2 = 1.5 Hz, 2H, H-4'), 5.13 (dd, J_1 = 7.8 Hz, J_2 = 1.9 Hz, 2H, H-3'), 4.79 (dd, J_1 = 5.0 Hz, J_2 = 2.0 Hz,

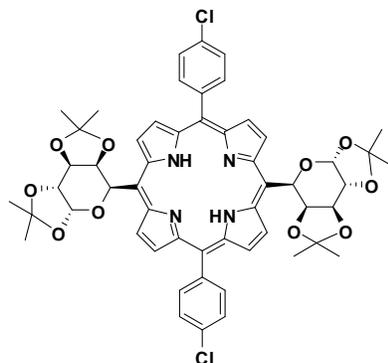
2H, H-2'), 1.85 (s, 6H, -CH₃), 1.69 (s, 6H, -CH₃), 1.53 (s, 6H, -CH₃), 1.18 (s, 6H, -CH₃), -2.73 (s, 2H, NH); ¹³C NMR (100 MHz, CDCl₃): δ 149.8, 141.9, 135.5, 130.1, 129.2, 117.9, 114.2, 109.7, 109.1, 97.8, 71.9, 71.3, 26.8, 25.9, 25.1, 23.4; HRMS (ESI): m/z (M+H)⁺ Calcd for C₅₆H₅₃N₄O₁₂F₆: 1087.3564, found: 1087.3545; UV-VIS (CHCl₃) λ_{max} (log ε): 403 (4.538), 516 (4.211), 550 (3.827), 591 (3.757), 645 (3.487); IR (neat): ν_{max} = 3435, 2922, 1643, 1382, 1257, 1066, 804 cm⁻¹.

5,15-[Bis(pentafluorophenyl)]-10α,20β-[bis(1,2:3,4-di-O-isopropylidene-α-D-galactopyranose-6-yl)]porphyrin (8):



Prepared according to general procedure using pentafluorobenzaldehyde; Purple solid; (64 mg, 11% yield); ¹H NMR (500 MHz, CDCl₃): δ 9.86 (s, 4H, H-β), 8.87 (d, *J* = 4.6 Hz, 4H, H-β), 7.72 (s, 2H, H-5'), 6.31 (d, *J* = 4.8 Hz, 2H, H-1'), 5.26 (d, *J* = 7.7 Hz, 2H, H-4'), 5.19 (d, *J* = 7.6 Hz, 2H, H-3'), 4.85 (d, *J* = 4.8 Hz, 2H, H-2'), 1.91 (s, 6H, -CH₃), 1.76 (s, 6H, -CH₃), 1.58 (s, 6H, -CH₃), 1.24 (s, 6H, -CH₃), -2.63 (s, 2H, -NH); ¹³C NMR (75 MHz, CDCl₃): δ 147.9 (d, *J* = 58.6 Hz), 144.3 (t, *J* = 54.1 Hz), 139.6 (d, *J* = 86.3), 135.7, 131.6, 130.0, 117.6 (dt, *J*₁ = 39.0 Hz, *J*₂ = 3.6 Hz), 115.3, 109.8, 109.2, 101.5, 97.8, 77.3, 76.6, 71.9, 71.8, 71.3, 26.8, 25.9, 25.0, 23.4; HRMS (ESI): m/z (M+H)⁺ Calcd for C₅₄H₄₅N₄O₁₀F₁₀: 1099.2976, found: 1099.2979; UV-VIS (CHCl₃) λ_{max} (log ε): 405 (4.521), 513 (4.163), 543 (3.422), 586 (3.701), 640 (3.163); IR (neat): ν_{max} 3439, 2077, 1645, 1494, 1257, 1163, 1064 cm⁻¹.

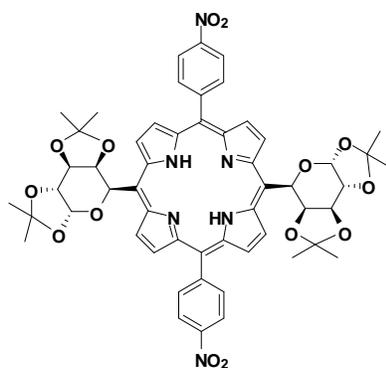
5,15-[Bis(4-chlorophenyl)]-10α,20β-[bis(1,2:3,4-di-O-isopropylidene-α-D-galactopyranose-6-yl)]Porphyrin (9):



Prepared according to general procedure using 4-chlorobenzaldehyde; Purple solid; (36 mg, 7% yield); ¹H NMR (500 MHz, CDCl₃): δ 9.70 (s, 4H, H-β), 8.79 (d, *J* = 4.6 Hz, 4H, H-β), 8.10 (d, *J* = 7.0 Hz, 4H, Ph-CH), 7.72 (d, *J* = 7.0 Hz, 4H, Ph-CH), 7.66 (s, 2H, H-5'), 6.25 (d, *J* = 4.9 Hz, 2H, H-1'), 5.20 (d, *J* = 7.7 Hz, 2H, H-4'), 5.13 (d, *J* = 7.8 Hz, 2H, H-3'), 4.79 (d,

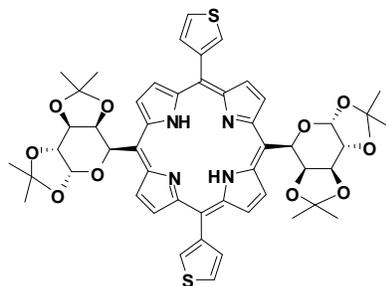
$J = 4.9$ Hz, 2H, H-2'), 1.85 (s, 6H, -CH₃), 1.69 (s, 6H, -CH₃), 1.52 (s, 6H, -CH₃), 1.18 (s, 6H, -CH₃), -2.74 (s, 2H, NH); ¹³C NMR (75 MHz, CDCl₃): δ 141.7, 135.4, 134.0, 131.9, 130.2, 126.6, 117.9, 114.0, 109.6, 109.0, 97.8, 71.9, 71.3, 26.7, 25.8, 25.1, 23.3; HRMS (ESI): m/z (M+H)⁺ Calcd for C₅₄H₅₃N₄O₁₀Cl₂: 987.3139, found: 919.3138; UV-VIS (CHCl₃) λ_{max} (log ϵ): 416 (4.480), 516 (3.453), 548 (3.051), 590 (2.979), 644 (2.723); IR (neat): ν_{max} 3439, 2922, 1714, 1643, 1462, 1379, 1068, 702 cm⁻¹.

5,15-[Bis(4-nitrophenyl)]-10 α ,20 β -[bis(1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose-6-yl)]porphyrin (10):



Prepared according to general procedure using 4-nitrobenzaldehyde; Purple solid; (32 mg, 6% yield); ¹H NMR (300 MHz, CDCl₃): δ 9.74 (d, $J = 3.9$ Hz, 4H, H- β), 8.72 (d, $J = 4.9$ Hz, 4H, H- β), 8.63 (d, $J = 8.6$ Hz, 4H, Ph-CH), 8.36 (d, $J = 8.5$ Hz, 4H, Ph-CH), 7.66 (s, 2H, H-5'), 6.25 (d, $J = 5.0$ Hz, 2H, H-1'), 5.18 (d, $J = 9.5$ Hz, 2H, H-4'), 5.14 (d, $J = 1.8$ Hz, 2H, H-3'), 4.80 (dd, $J_1 = 5.0$ Hz, $J_2 = 1.8$ Hz, 2H, H-2') 1.85 (s, 6H, -CH₃), 1.69 (s, 6H, -CH₃), 1.53 (s, 6H, -CH₃), 1.19 (s, 6H, -CH₃), -2.72 (s, 2H, NH); ¹³C NMR (125 MHz, CDCl₃): δ 150.0, 147.7, 135.0, 121.6, 116.8, 114.9, 109.7, 109.1, 97.8, 71.8, 71.8, 71.3, 26.7, 25.8, 25.0, 23.3. HRMS (ESI): m/z (M+H)⁺ Calcd for C₅₄H₅₃N₆O₁₄: 1009.3620, found: 1009.3619; UV-VIS (CHCl₃) λ_{max} (log ϵ): 419 (4.506), 518 (3.577), 550 (3.103), 590 (3.106), 645 (2.811); IR (neat): ν_{max} 3439, 2958, 1714, 1643, 1519, 1462, 1066, cm⁻¹.

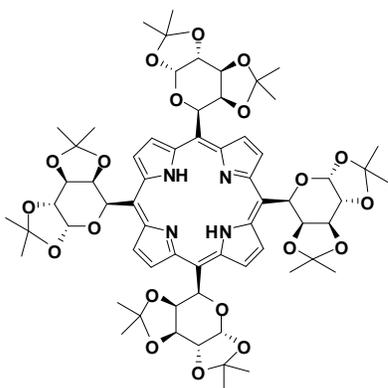
5,15-[Bis(3-thiophene)]-10 α ,20 β -[bis(1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose-6-yl)]porphyrin (11):



Prepared according to general procedure using 3-formyl thiophene; Purple solid; (24 mg, 5% yield); ¹H NMR (300 MHz, CDCl₃): δ 9.68 (d, $J = 3.9$ Hz, 4H, H- β), 8.94 (d, $J = 4.9$ Hz, 4H, H- β), 7.98-7.96 (m, 4H, Ar-CH), 7.70 (dd, $J_1 = 4.8$ Hz, $J_2 = 3.0$ Hz, 4H, H- β), 7.66 (d, $J = 1.1$ Hz, 2H, H-5'), 6.25 (d, $J = 4.9$ Hz, 2H, H-1'), 5.20 (dd, $J_1 = 8.0$ Hz, $J_2 = 1.5$ Hz, 2H, H-

4'), 5.12 (dd, $J_1 = 7.9$ Hz, $J_2 = 2.0$ Hz, 2H, H-3'), 4.78 (dd, $J_1 = 5.0$ Hz, $J_2 = 2.0$ Hz, 2H, H-2'), 1.86 (s, 6H, -CH₃), 1.69 (s, 6H, -CH₃), 1.53 (s, 6H, -CH₃), 1.18 (s, 6H, -CH₃), -2.7 (s, 2H, NH); ¹³C NMR (75 MHz, CDCl₃): δ 162.3, 143.3, 134.8, 131.4, 130.1, 128.1, 122.8, 113.8, 113.7, 109.6, 109.6, 109.0, 97.8, 76.5, 71.9, 71.3, 26.7, 25.9, 25.1, 23.3; HRMS (ESI): m/z (M+H)⁺ Calcd for C₅₀H₅₁N₄O₁₀S₂: 931.3047, found: 931.3044; UV-VIS (CHCl₃) λ_{max} (log ϵ): 412 (4.501), 518 (3.767), 550 (3.480), 591 (3.329), 646 (3.089); IR (neat): ν_{max} 3437, 2918, 1643, 1454, 1382, 1255, 1064 cm⁻¹.

5 α ,10 β ,15 α ,20 β -Tetrakis(1,2:3,4-di-*O*-isopropylidene- α -D-galactopyranose-6-yl)porphyrin (12):



Prepared according to general procedure with slight modification. Here, we used instead of aromatic aldehyde another

1,2:3,4-di-*O*-isopropylidene- α -D-galacto-

hexadialdo-1,5-pyranose (**1**); Purple solid; (104 mg, 16% yield); ¹H NMR (300 MHz, CDCl₃): δ 9.81 (s, 8H), 7.77 (s, 4H, H-5'), 6.32 (d, $J = 4.9$ Hz, 4H, H-1'), 5.28 (d, $J = 8.7$ Hz, 4H, H-4'), 5.18 (dd, $J_1 = 7.9$ Hz, $J_2 = 1.8$ Hz, 4H, H-3'), 4.84 (dd, $J_1 = 5.0$ Hz, $J_2 = 1.9$ Hz, 4H, H-2'), 1.94 (s, 12H, -

CH₃), 1.85 (s, 12H, -CH₃), 1.59 (s, 12H, -CH₃), 1.26 (s, 12H, -CH₃), -2.88 (s, 2H, NH); ¹³C NMR (75 MHz, CDCl₃): δ 130.0, 112.5, 109.6, 109.0, 97.7, 76.6, 72.2, 71.9, 71.5, 26.9, 26.1, 25.1, 23.5; HRMS (ESI): m/z (M+H)⁺ Calcd for C₆₄H₇₉N₄O₂₀: 1223.5288, found: 1223.5266; UV-VIS (CHCl₃) λ_{max} (log ϵ): 406 (4.519), 519 (4.017), 552 (3.381), 591 (3.591), 646 (3.437); IR (neat): ν_{max} 3441, 2989, 2073, 1643, 1382, 1255, 1064, cm⁻¹.

Biology Methods and Materials:

Cell Cultures:

Human cancer cell lines including HCT116 and HeLa were maintained in Dulbecco's Modified Eagle Medium (DMEM) containing 10% fetal bovine serum and 1% penicillin/streptomycin in a humidified 5% CO₂ incubator at 37 °C.

Photocytotoxicity Assay.

Cells were seeded onto 96-well plates at a density of about 2×10^4 cells per well and incubated in the dark in medium containing 5% serum together with compounds for 24 h at 37°C. Cells were rinsed with phosphate buffered saline (PBS) and then exposed to broad-spectrum green light (480-550 nm) generated by two layers of green cellophane-filtered 50 W halogen lamp using a dose rate of 13 mW/cm². Cell viability was determined using the CellTiter 96 Aqueous One Solution Reagent kit (Promega, Madison, WI) according to the manufacturer's instructions, 24 h after light exposure by measuring absorbance at 490 nm.

Intracellular Localization and Image Analysis.

Cells plated on coverslips in a 6-well plate were incubated with 1 μM of compound for 24 h. For intracellular localization in HeLa cells, cells incubated with compound for 24 h were loaded with 100 nM MitoTracker Deep Red (Molecular Probes) for 15 min or with 100 nM LysoTracker Red (Molecular Probes) for 1 h at 37 °C. The slides were washed three times with PBS and were visualized by at 60 x magnification on a Zeiss LSM META confocal laser scanning microscopy (Zeiss, Oberkochen, Germany).

Measurement of Apoptosis.

Apoptosis was performed as previously described from our group.³ In brief, cells treated with 1 μM compound were incubated for 24 h and then illuminated. After 24 h, cells were collected and apoptosis was examined by using Annexin V-FLUOS staining kit (Roche, Penzberg, Germany). Cells were counter-stained with propidium iodide followed by fluorescence activated cell sorter (FACS) analysis on a flow cytometer (BD LDR II, BD Biosciences, San

Jose, CA). For visualization of apoptotic cells, cells were seeded on coverslips within a 6-well plate. After fixation in 3.7% paraformaldehyde, cells were washed with PBS and permeabilized with 0.2% Triton X-100, washed again with PBS, and mounted by ProLong Gold antifade reagent with DAPI (Molecular probes, Eugene, Oregon). The stained nuclei were observed and photographed under a fluorescence microscope (Nikon Inc., Melville, NY). Apoptosis was measured as the percentage of annexin V-positive and PI-negative cell population. For all experiments, at least 10,000 events were collected per sample.

Immunoblot Analysis.

Cells were resuspended in a lysis buffer (20 mM Tris-HCl, pH 7.5, 150 mM NaCl, 0.5% Triton X-100, 1 mM EDTA, 1 mM PMSF) containing protease inhibitors on ice for 40 min. The clear cell lysates were obtained after centrifuging for 15 min at 15,000 rpm. The lysates (30 μ g of protein) were resolved by SDS-polyacrylamide gel electrophoresis (SDS-PAGE) and were transferred to nitrocellulose membranes. The membranes were blocked with 5% dry milk in TBS-T (20 mM Tris-HCl, pH 7.5, 140 mM NaCl, and 0.05% tween-20) and subsequently incubated with primary antibody followed by a goat anti-rabbit or goat anti-mouse IgG conjugated to horseradish peroxidase, and the immunoreactive bands were visualized by the SUPEX Western blotting detection kit (Neuronex, Korea)

- 1) J. B. Malcolm R. Banks, J. I. G. Cadogan, Ian M. Dawson, Suneel Gaur, Ian Gosney, Robert and K. J. G. a. P. K. G. H. Gould, *J. Chem. Soc., Chem. Commun.*, 1993, 1146.
- 2) J. T. Suri, S. Mitsumori, K. Albertshofer, F. Tanaka and C. F. Barbas, *J. Org. Chem.*, 2006, **71**, 3822-3828.
- 3) a) B.-H. Choi, L. Feng and H. S. Yoon, *J. Biol. Chem.*, 2010, **285**, 9770; b) B.-H. Choi, W. Kim, Q. C. Wang, D.-C. Kim, S. N. Tan, J. W. H. Yong, K.-T. Kim and Yoon, H. S. *Cancer Lett.* 2008, **261**, 37.

NMR, UV, HRMS spectrum of sugar porphyrin conjugates

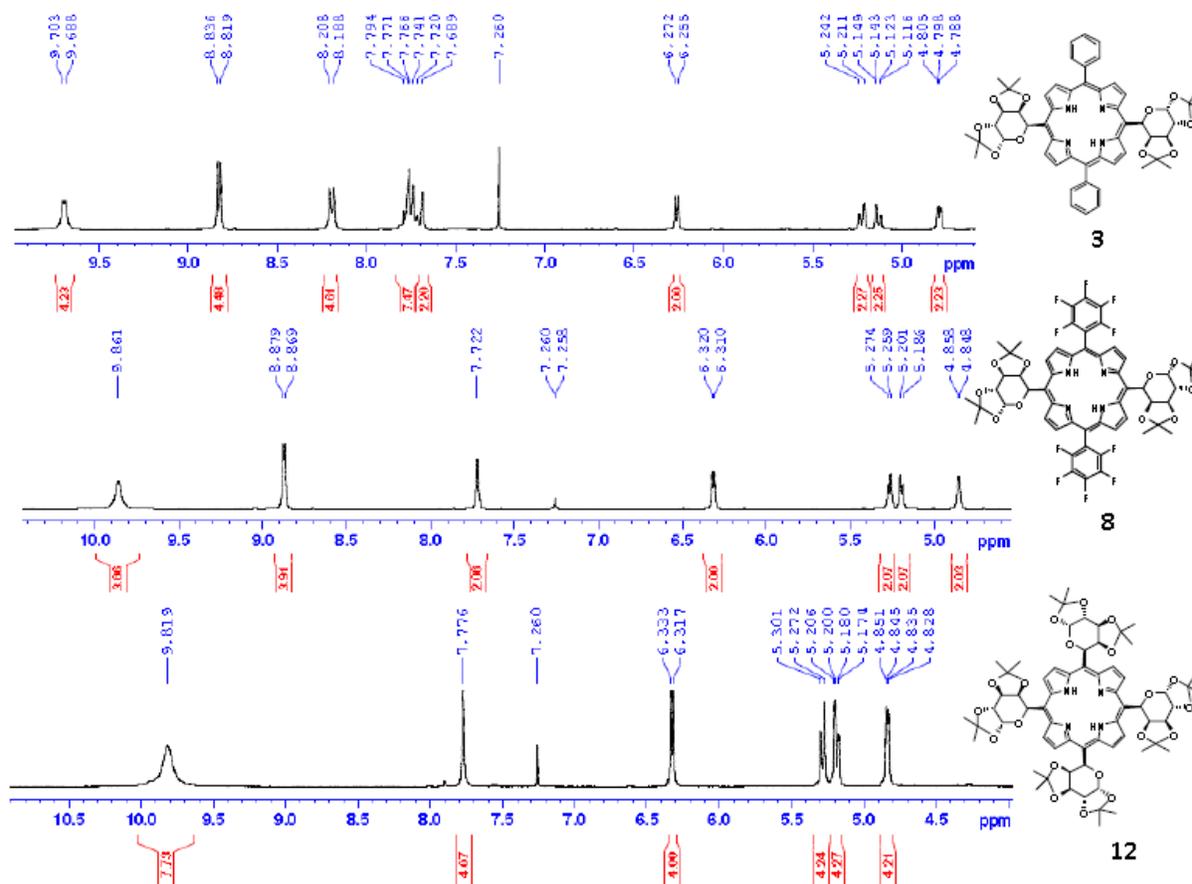
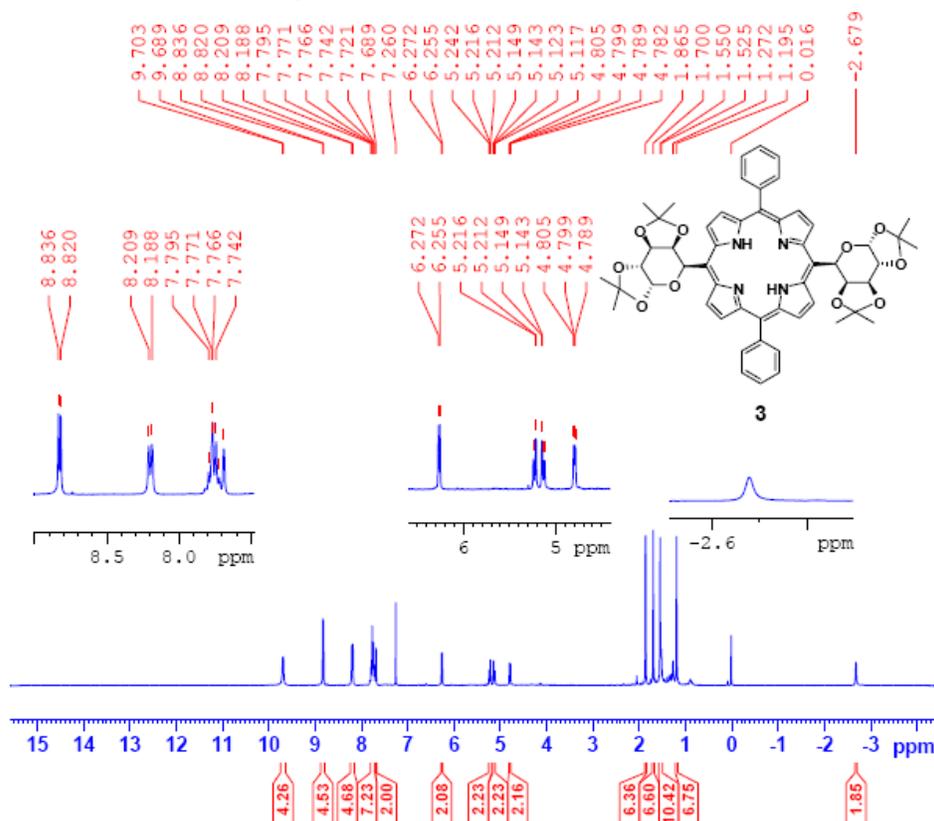


Fig. 1S. Partial ^1H NMR spectra of compound **3**, **8** and **12**

Furthermore, comparative NMR diagram illustrates that the partial ^1H NMR spectra of compounds **3**, **8** and **12**, thereby evidencing the diagnostic signals. For compounds **3** and **8**, two types of β , β' pyrrole protons appeared in the most deshielded aromatic region, but compound **12** displayed single peak. Compound **3** displayed Ar-H signal next to the β, β' pyrrole proton, but it disappeared in compound **8** due to the replacement of Ar-H by Ar-F. For all the compounds, sugar H-5' appeared in the deshielded region due to ring-current obtained by highly conjugated aromatic system. Successively, anomeric proton followed by the sugar methylene protons appears towards the shielded region. This picture further evidencing compounds **3** & **8** possess C_2 -symmetry where as compound **12** shows highly D_2 symmetry.

¹H Nmr, CDC13, 300MHz, Compound 3



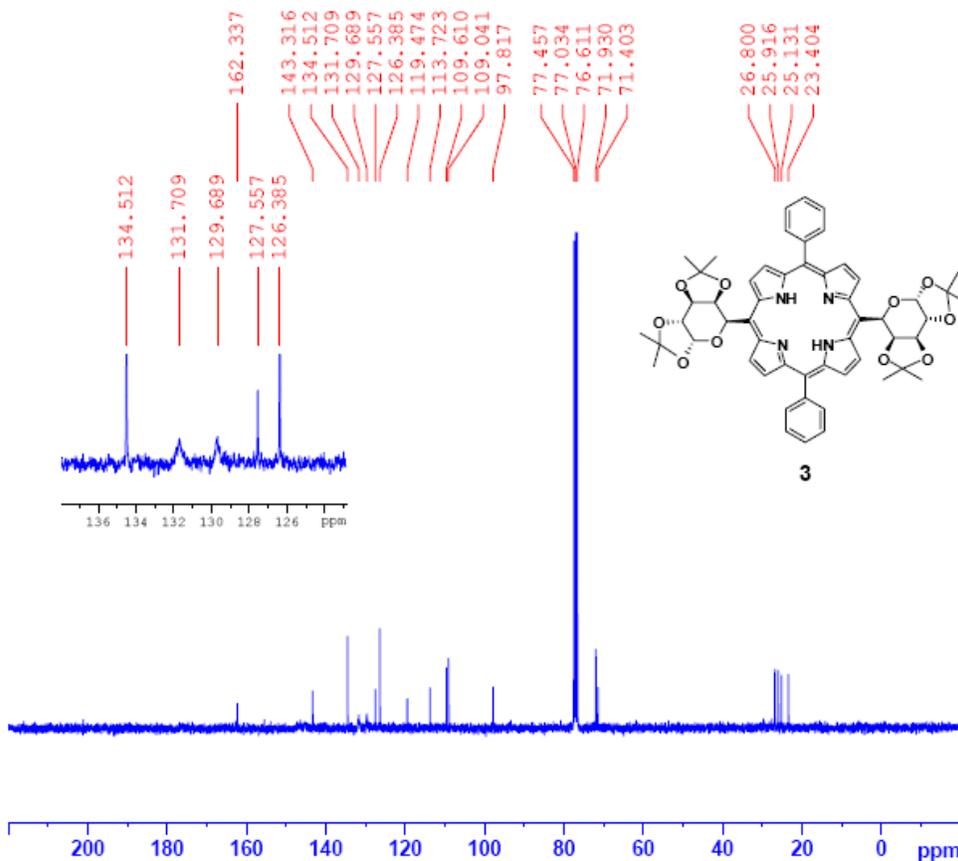
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DS       0
SWH      5995.204 Hz
FIDRES   0.299760 Hz
AQ       1.6680501 sec
RG       322.5
DW       83.400 usec
DE       6.00 usec
TE       297.8 K
D1       1.00000000 sec
TDO      1

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P1       11.00 usec
PL1      -1.00 dB
SFO1     300.1316870 MHz

F2 - Processing parameters
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SF       300.1300061 MHz
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SSB      0
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FC       1.00
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¹³C Nmr, CDC13, 300MHz, Compound 3



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EXPNO    1
PROCNO   1

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PULPROG zgpg30
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SOLVENT CDCl3
NS       8
DS       0
SWH      18115.941 Hz
FIDRES   0.276427 Hz
AQ       1.8098436 sec
RG       32768
DW       27.600 usec
DE       6.00 usec
TE       297.7 K
D1       2.00000000 sec
d11      0.03000000 sec
DELTA    1.89999998 sec
TDO      120

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NUC1     13C
P1       10.00 usec
PL1      -1.00 dB
SFO1     75.4752965 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2     1H
PCPD2    80.00 usec
FL2      -1.00 dB
FL12     16.23 dB
FL13     19.23 dB
SFO2     300.1312005 MHz

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SF       75.4677490 MHz
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SSB      0
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FC       1.40
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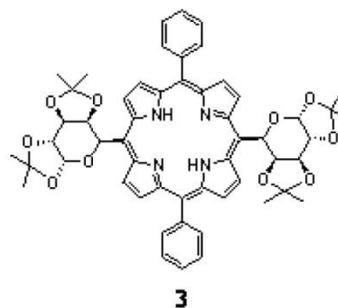
Elemental Composition Report

Page 1

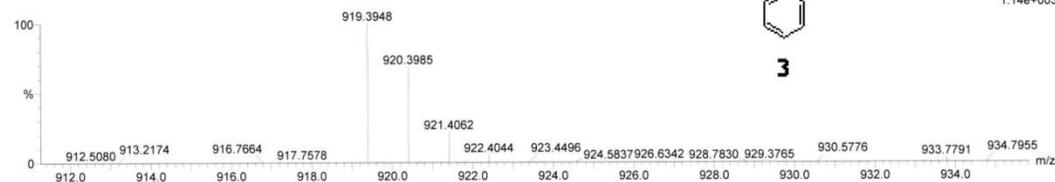
Single Mass Analysis

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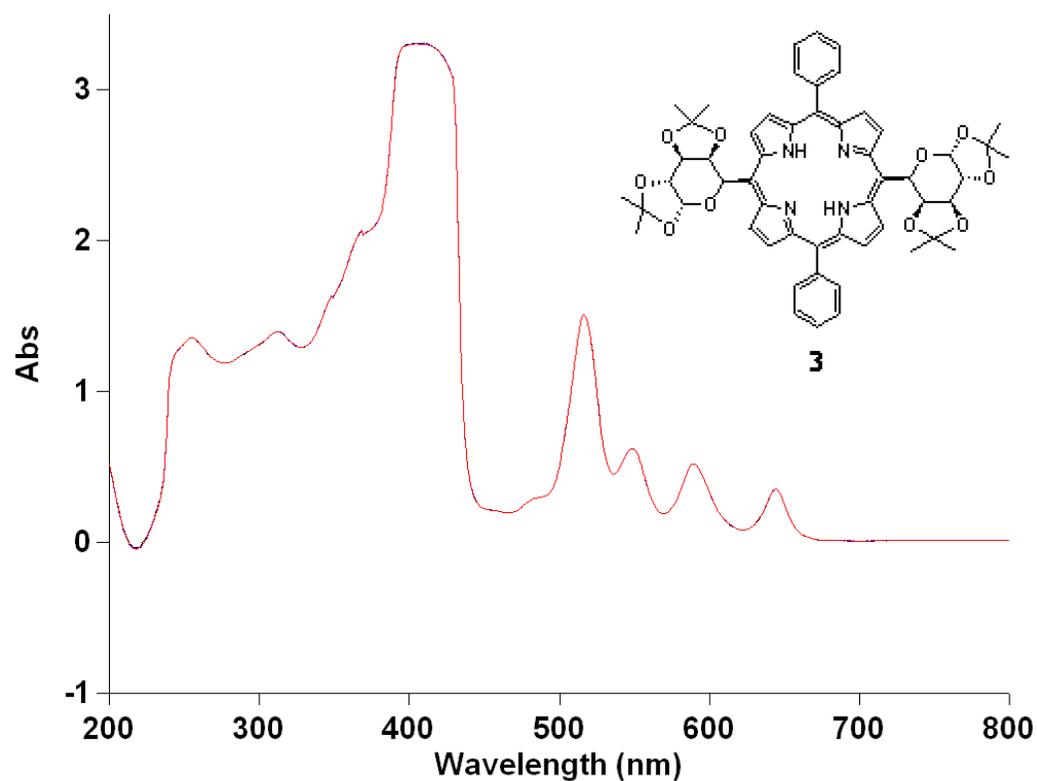
Monoisotopic Mass, Even Electron Ions
64 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)
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C: 0-54 H: 0-55 N: 0-5 O: 0-12
MW919
S3 3 (0.082) Cm (2.10)



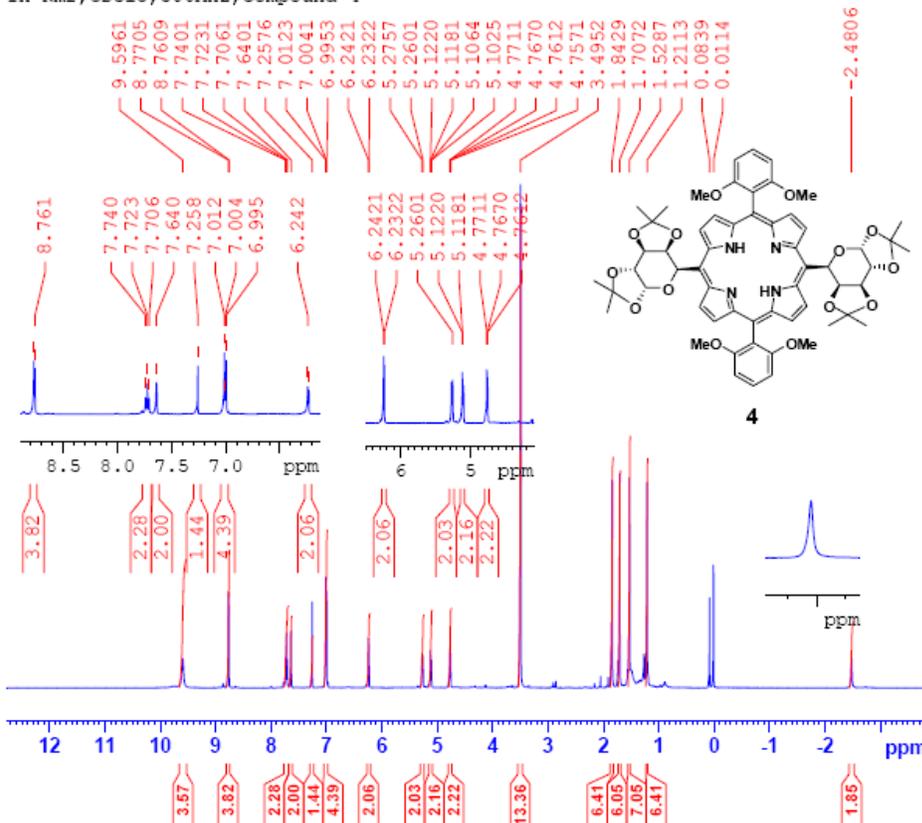
1: TOF MS ES+
1.14e+003



Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
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¹H Nmr, CDCl₃, 500MHz, compound 4

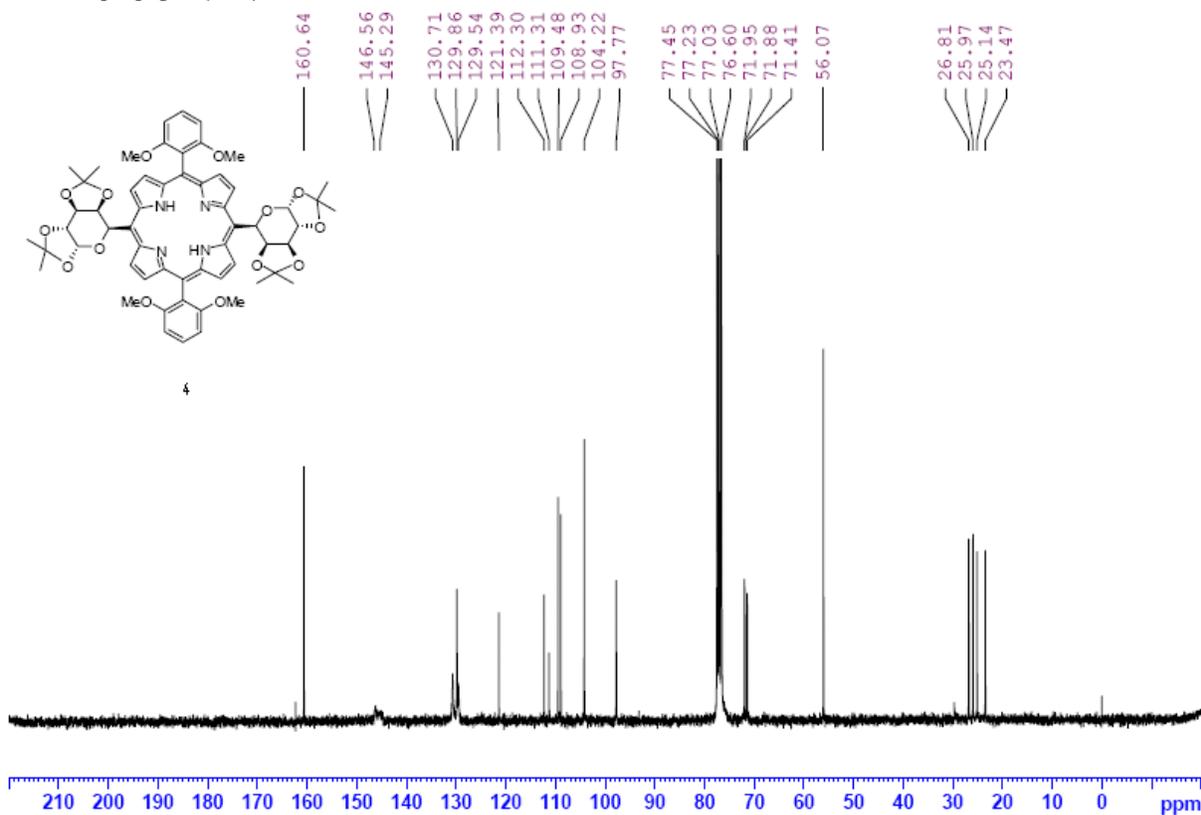


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PROCNO 1

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NS 16
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SWH 10000.000 Hz
FIDRES 0.533390 Hz
AQ 0.9374500 sec
RG 203
DW 50.000 usec
DE 6.00 usec
TE 296.2 K
D1 1.00000000 sec
TDO 1

===== CHANNEL f1 =====
NUC1 1H
P1 6.80 usec
PL1 0.00 dB
SFO1 500.1328782 MHz

F2 - Processing parameters
SI 32768
SF 500.1300142 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Elemental Composition Report

Page 1

Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0
Element prediction: Off
Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

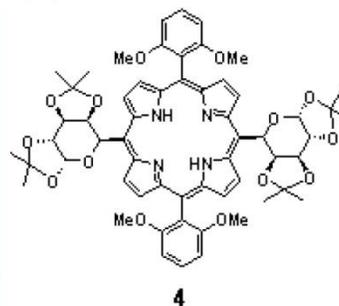
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Elements Used:

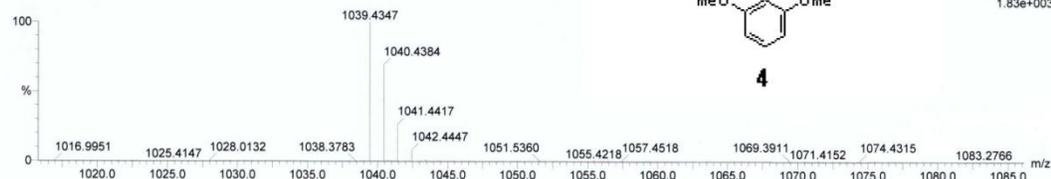
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MW1038

SNV-7.5 (0.120) Cm (5.14)



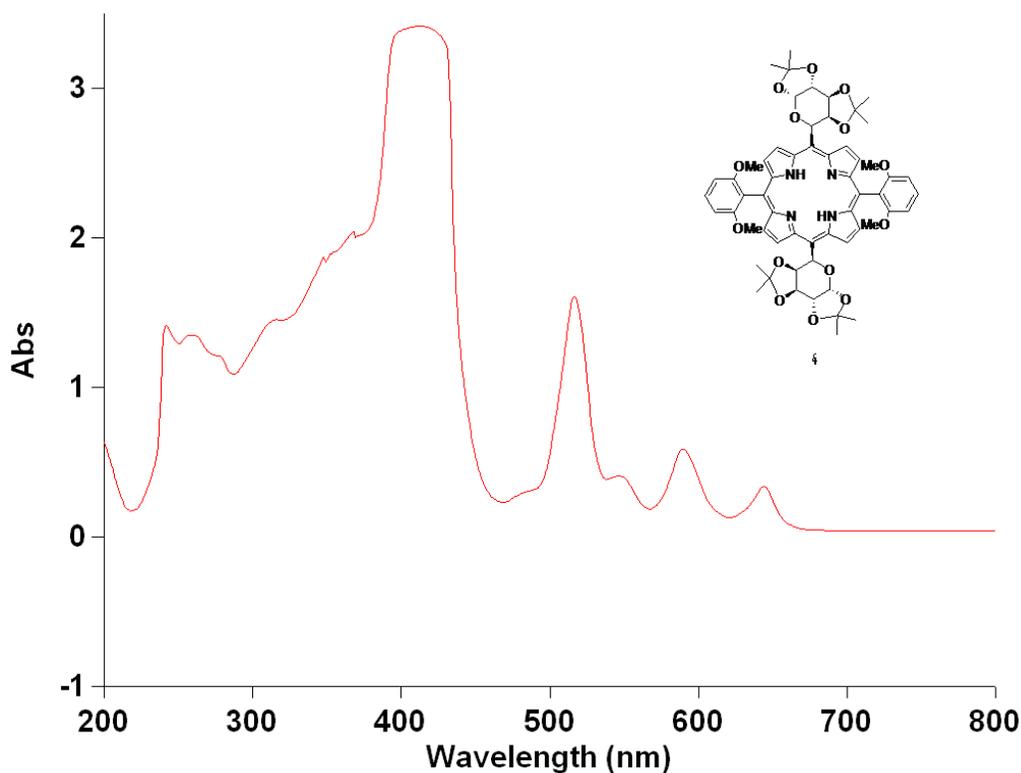
1: TOF MS ES+
1.83e+003



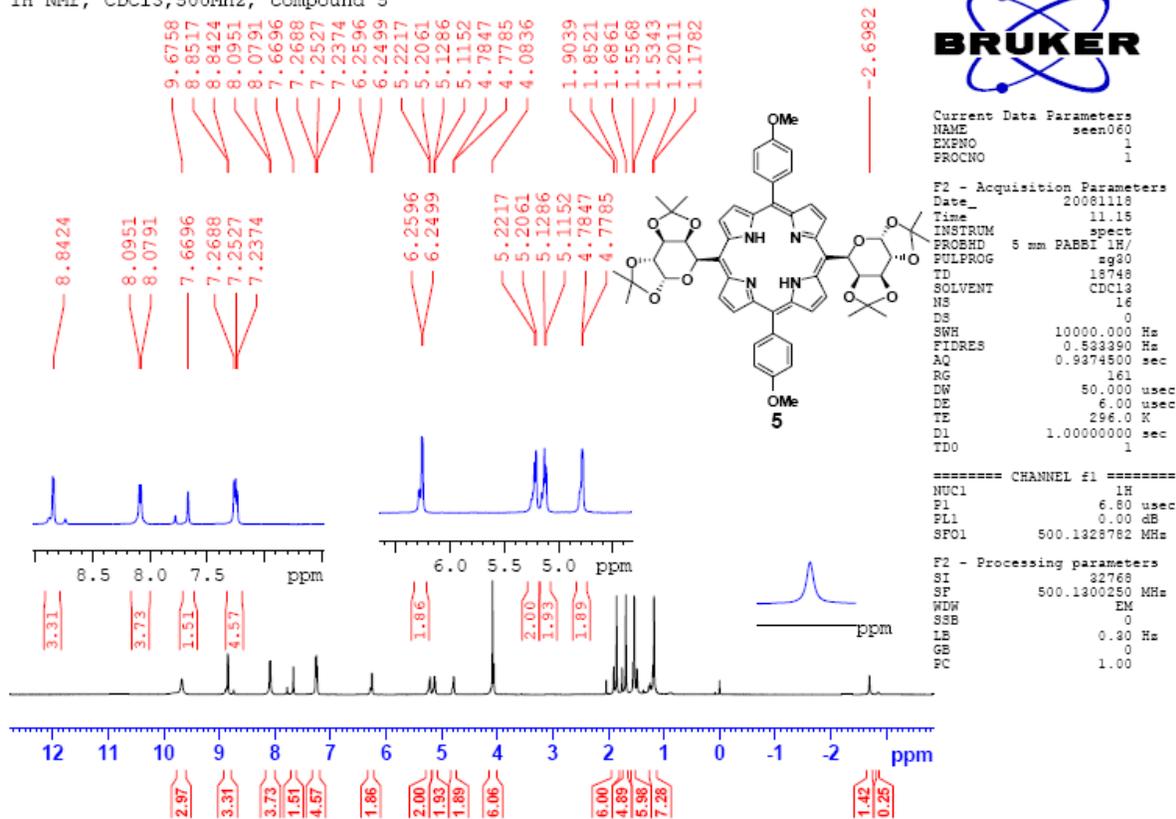
Minimum:

Maximum:

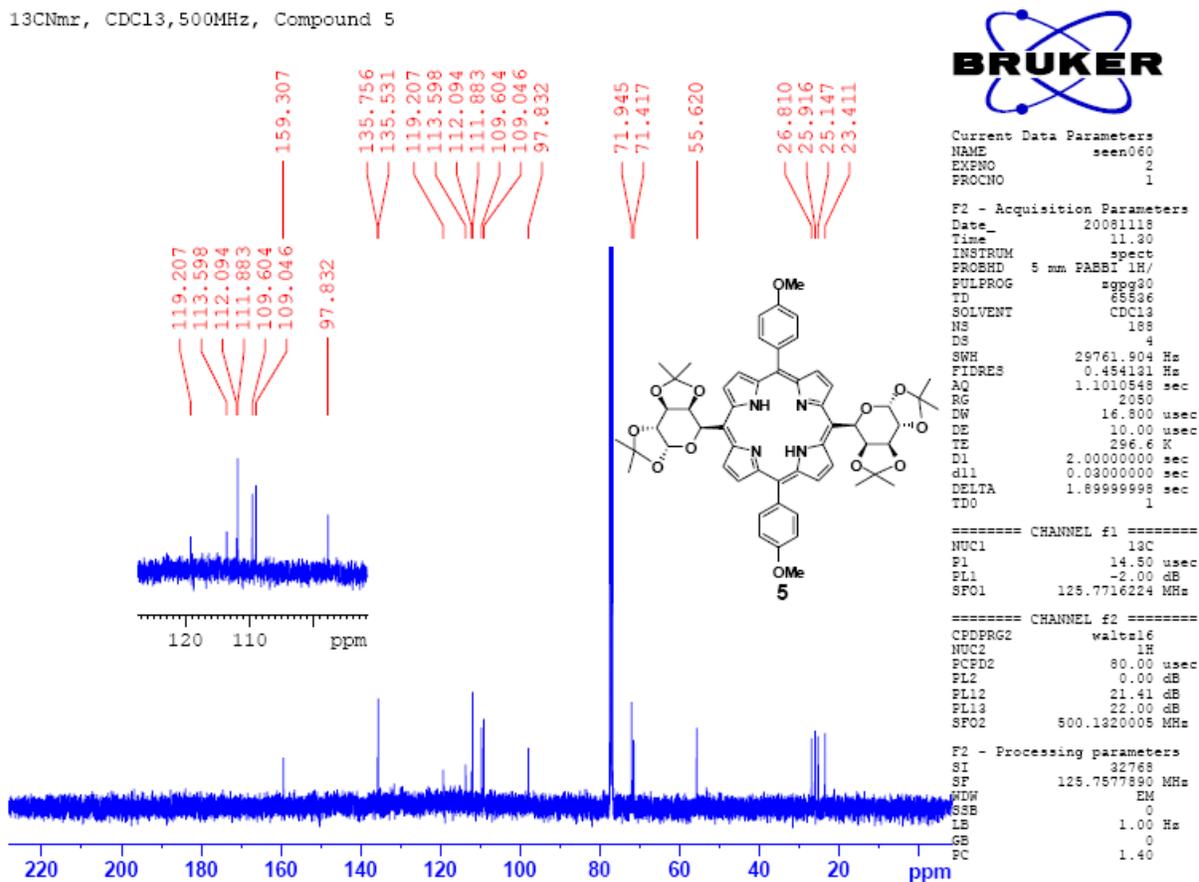
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
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¹H NMR, CDCl₃, 500MHz, compound 5



¹³C NMR, CDCl₃, 500MHz, Compound 5



Elemental Composition Report

Page 1

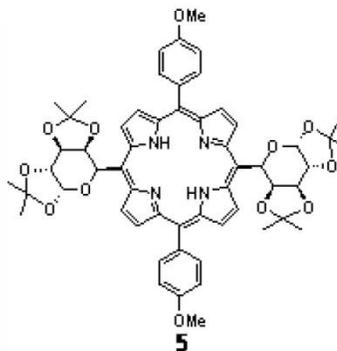
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Element prediction: Off
Number of isotope peaks used for i-FIT = 3

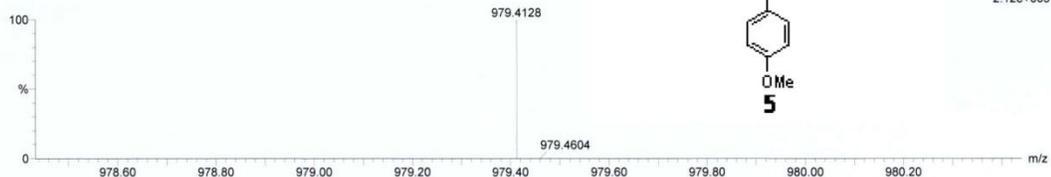
Monoisotopic Mass, Even Electron Ions
101 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)

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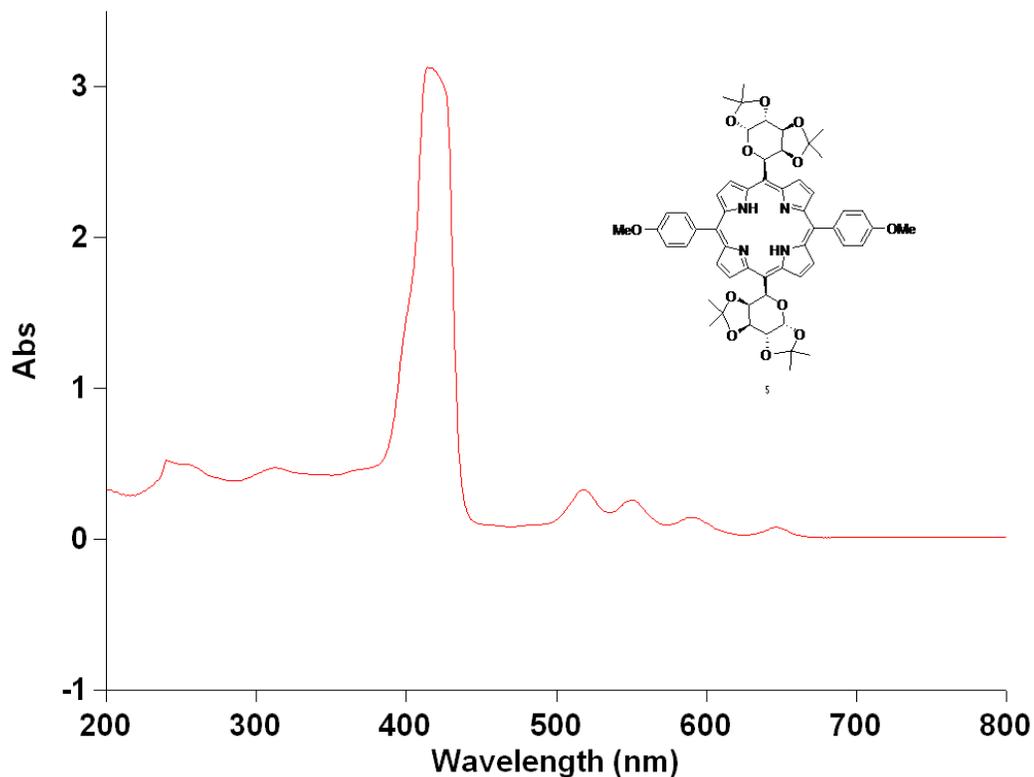
MW978
2-srv 3 (0.083) Cm (2:9)



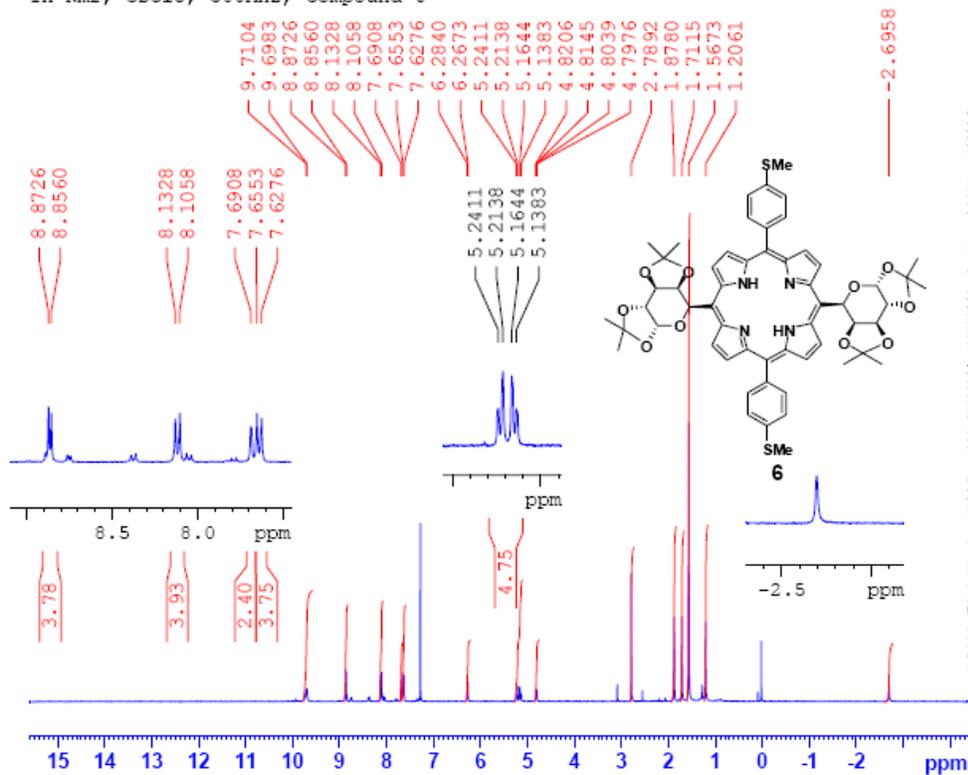
1: TOF MS ES+
2.12e+003



Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
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¹H Nmr, CDCl₃, 300MHz, Compound 6



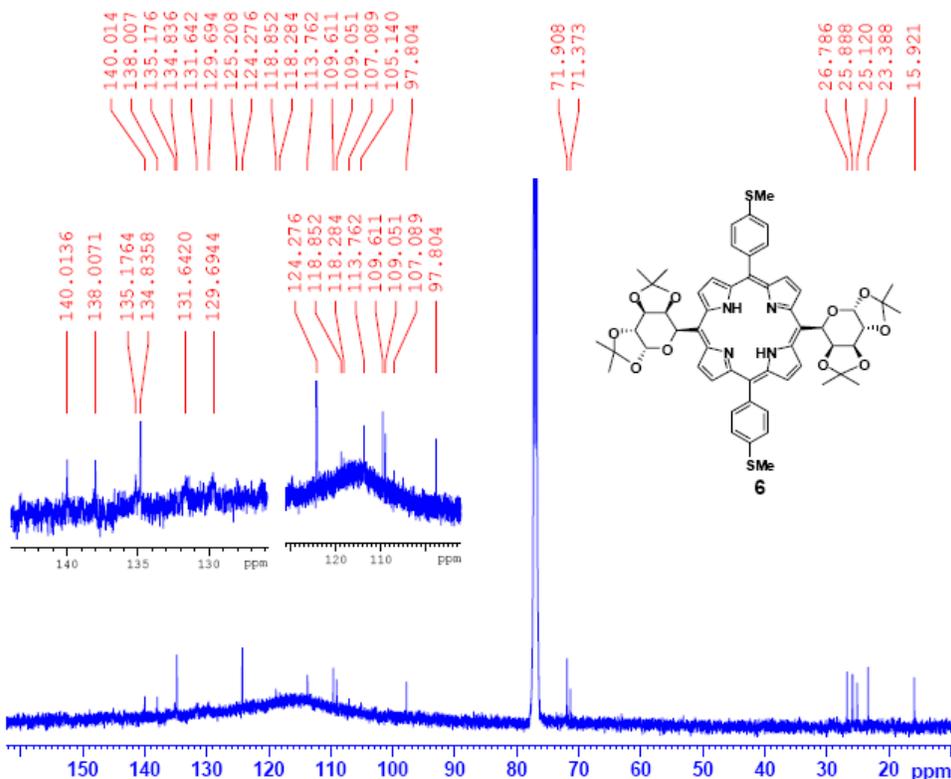
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PROCNO 1

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FIDRES 0.299760 Hz
AQ 1.6680501 sec
RG 456.1
DW 82.400 usec
DE 6.00 usec
TE 297.0 K
D1 1.00000000 sec
TD0 1

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P1 11.00 usec
PL1 -1.00 dB
SFO1 300.1316870 MHz

F2 - Processing parameters
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SF 300.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹³C Nmr, CDCl₃, 500Mhz, compound 6



Current Data Parameters
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EXPNO 1
PROCNO 1

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AQ 1.1010548 sec
RG 2050
DW 16.800 usec
DE 10.00 usec
TE 296.0 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
TD0 1

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NUC1 ¹³C
P1 14.50 usec
PL1 -2.00 dB
SFO1 125.7716224 MHz

===== CHANNEL f2 =====
CFDPRG2 waltz16
NUC2 ¹H
PCPD2 80.00 usec
PL2 0.00 dB
PL12 21.41 dB
PL13 22.00 dB
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.7577890 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Elemental Composition Report

Page 1

Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

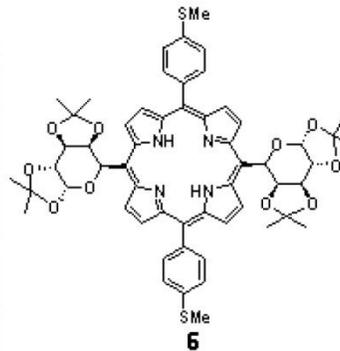
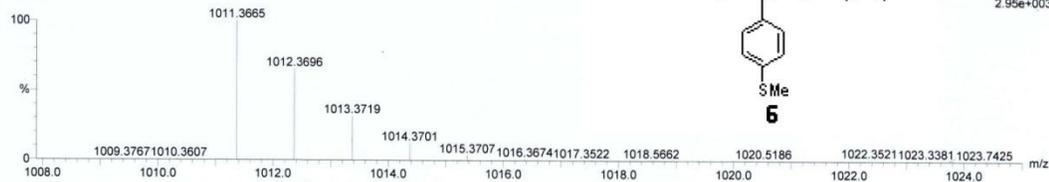
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MW1011

SNV-9.5 (0.120) Cm (5:10)

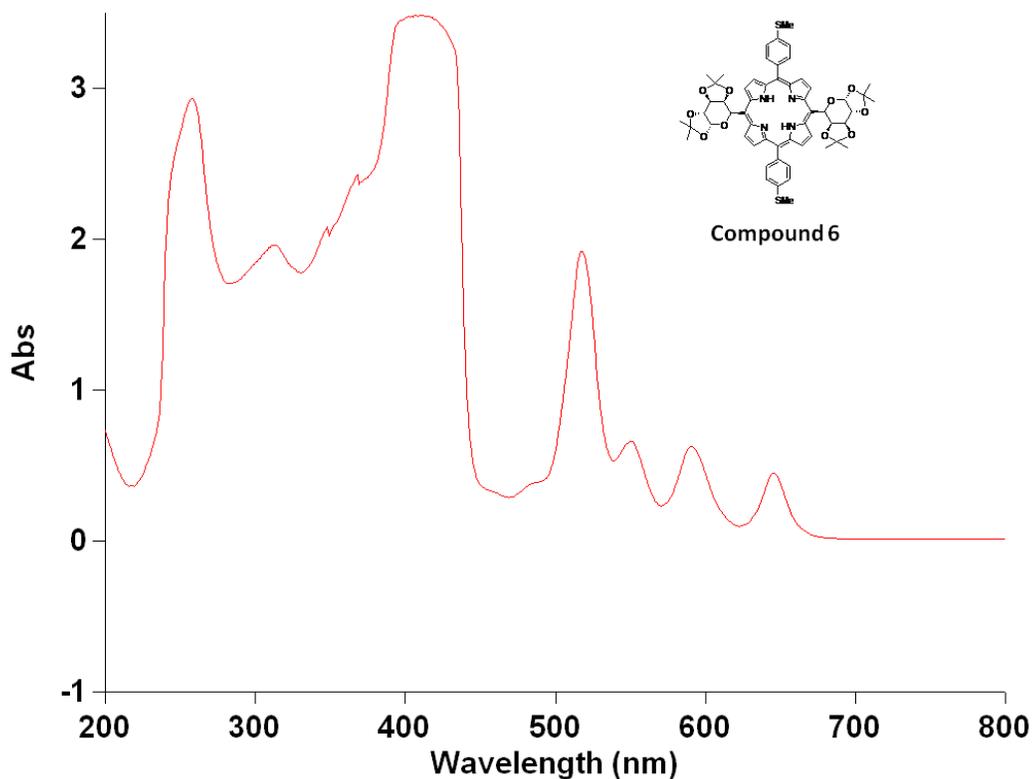


1: TOF MS ES+
2.95e+003

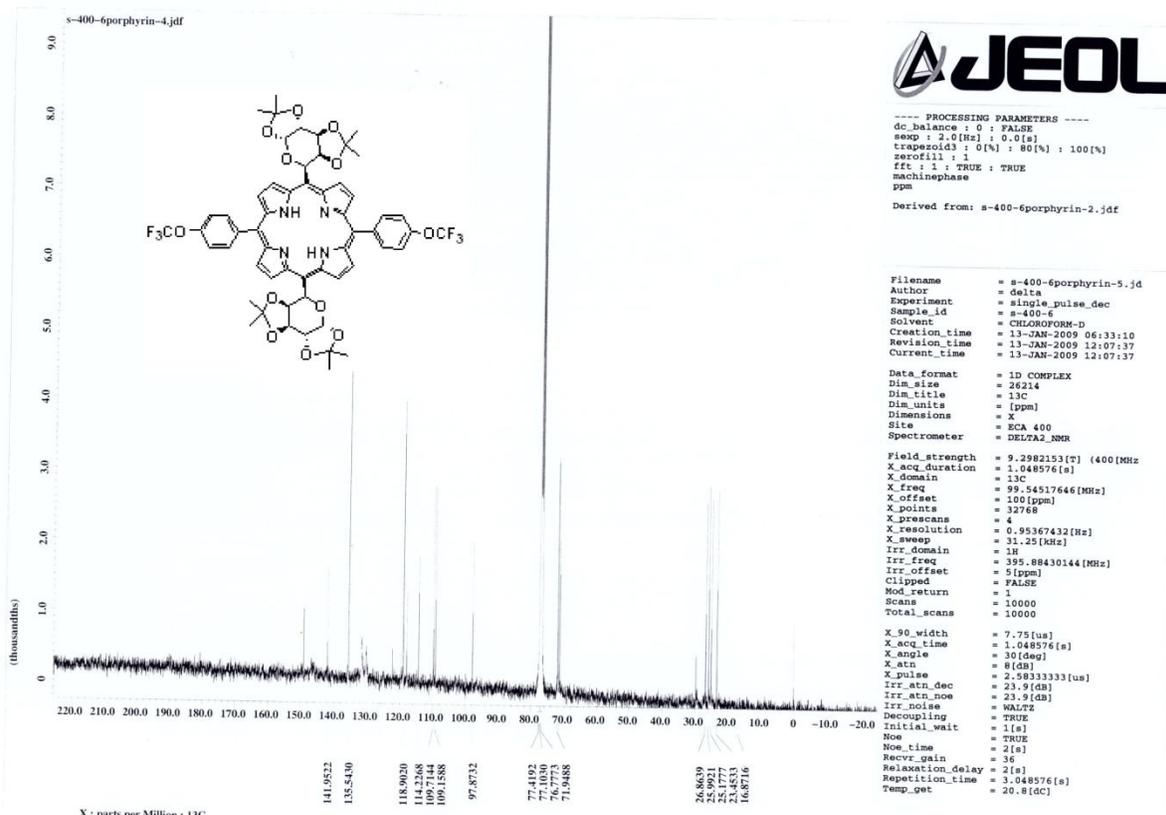
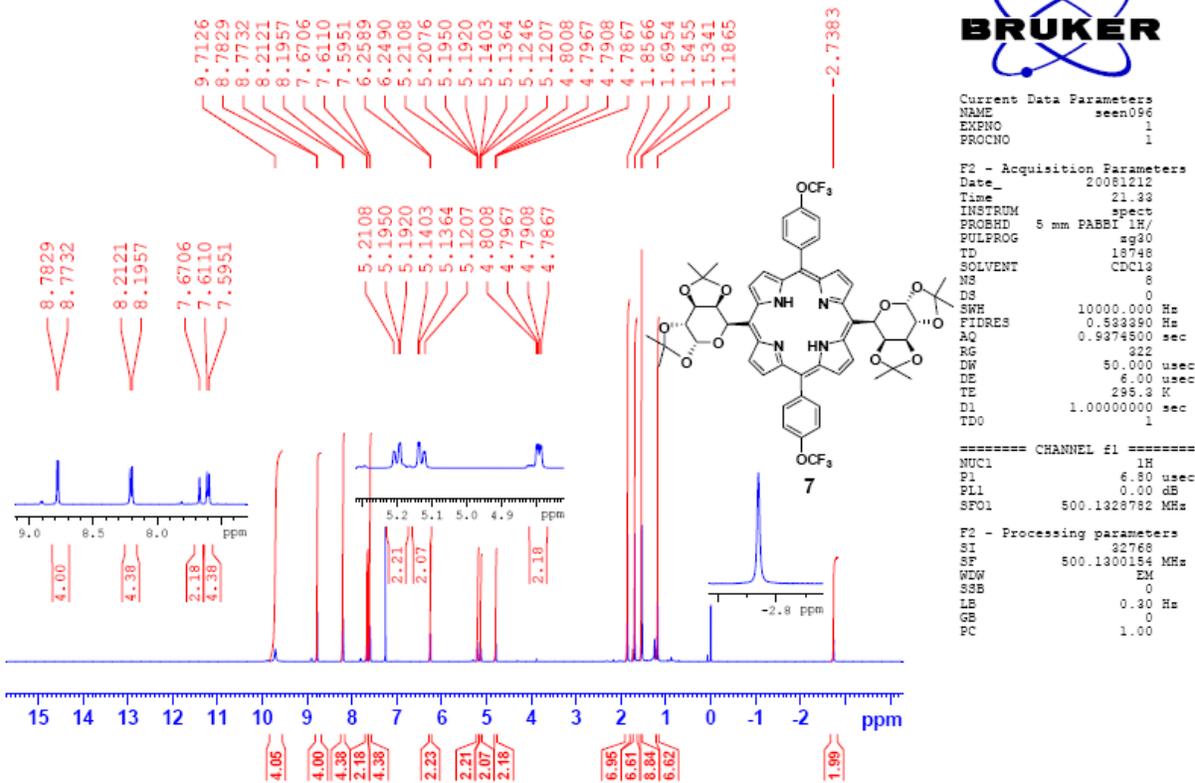
Minimum:

Maximum:

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
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X ¹H Nmr, CDC13, 300MHz, compound 7



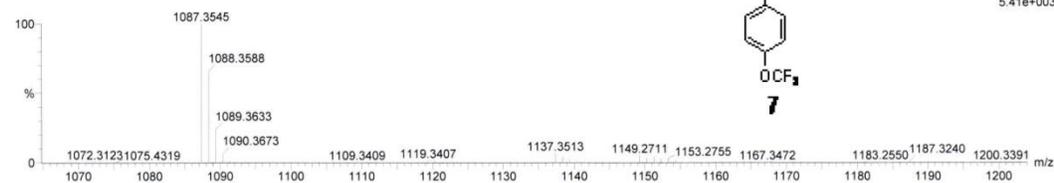
Elemental Composition Report

Single Mass Analysis

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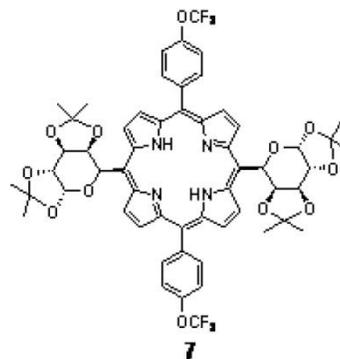
Monoisotopic Mass, Even Electron Ions
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MW1086
SNV-11.5 (0.119) Cm (5:12)

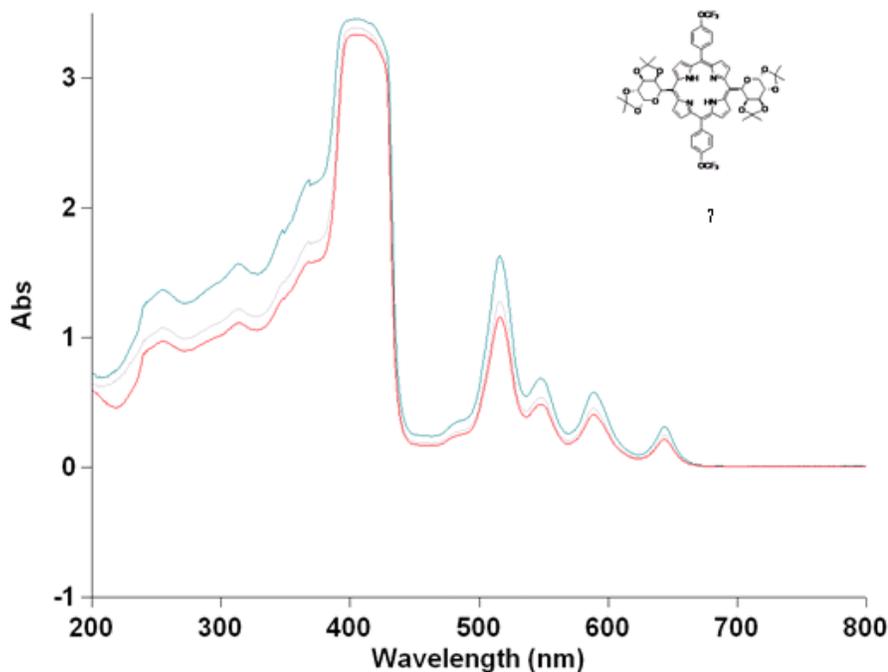


Minimum: -1.5
Maximum: 5.0 5.0 50.0

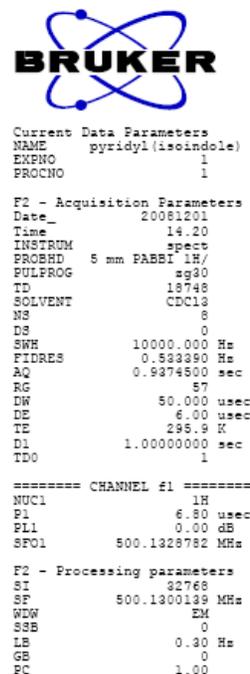
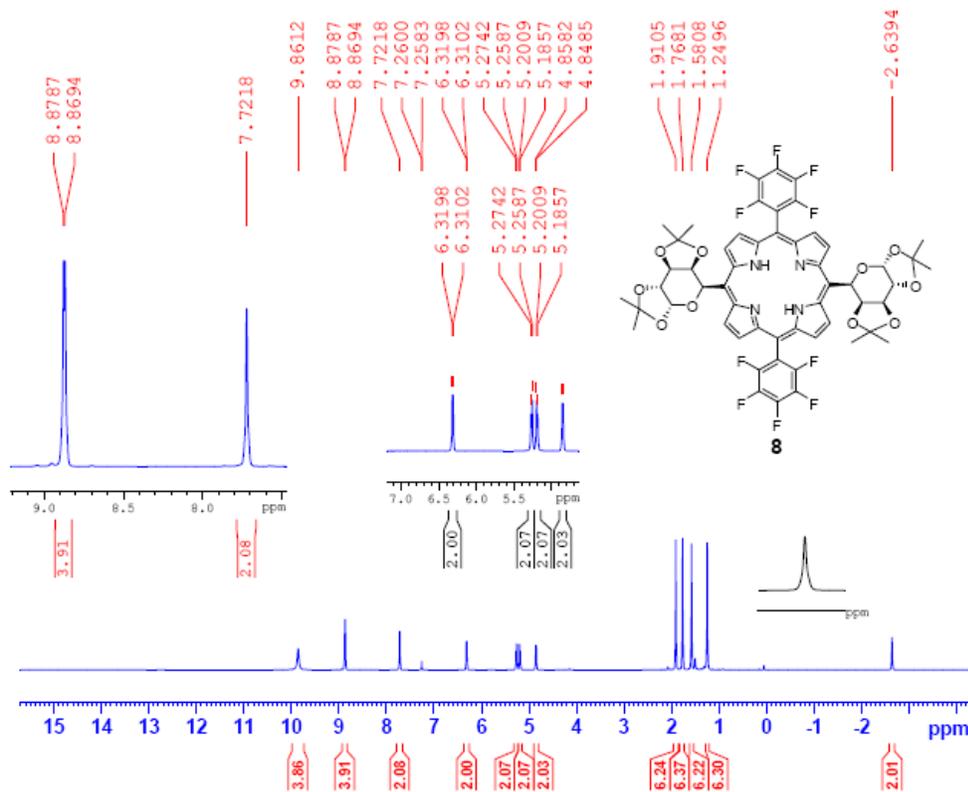
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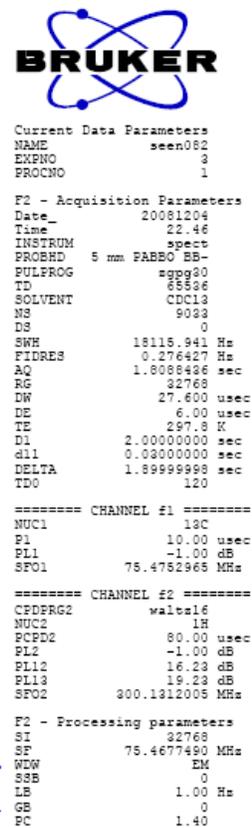
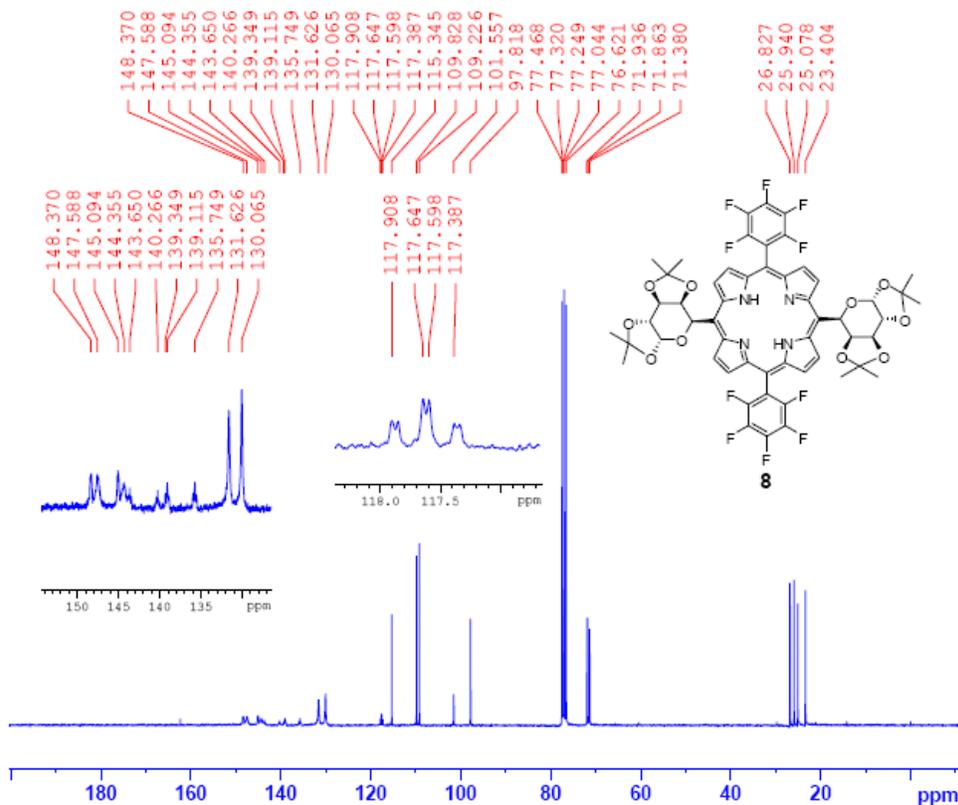
1: TOF MS ES+
5.41e+003



¹H Nmr, 500MHz, CDCl₃, compound 8



¹³C Nmr, CDCl₃, 300MHz, compound 8



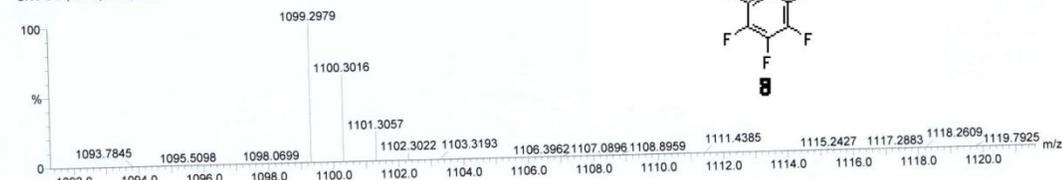
Elemental Composition Report

Single Mass Analysis

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Number of isotope peaks used for i-FIT = 3

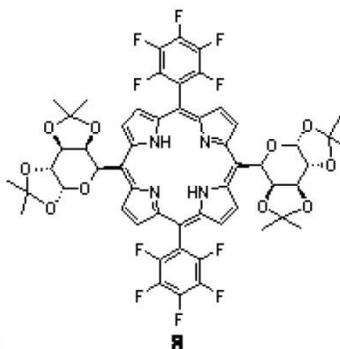
Monoisotopic Mass, Even Electron Ions
573 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)
Elements Used:
C: 0-54 H: 0-45 N: 0-4 O: 0-10 F: 0-10

MW1098
SNV-6.6 (0.139) Cm (6:19)

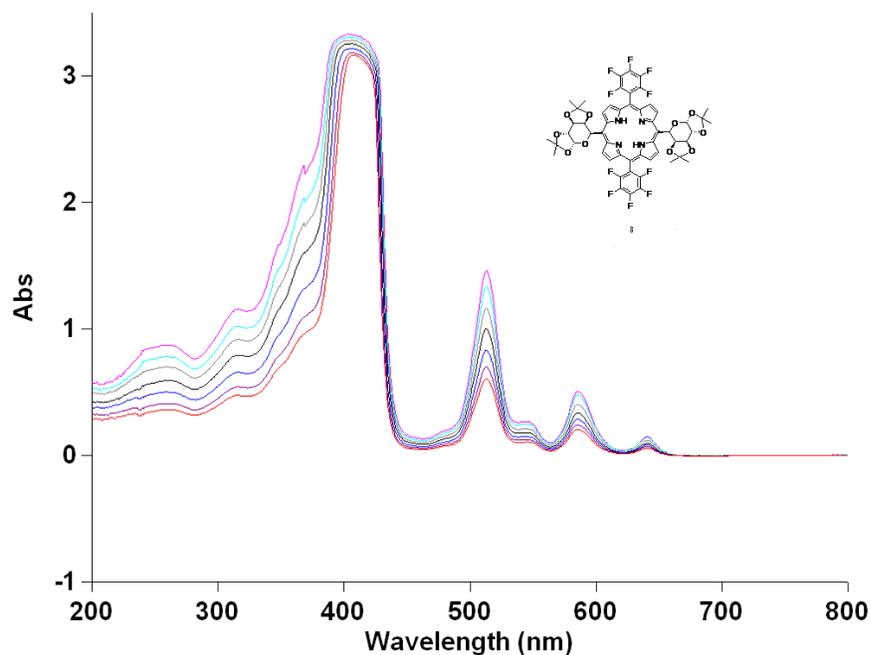


Minimum: -1.5
Maximum: 50.0

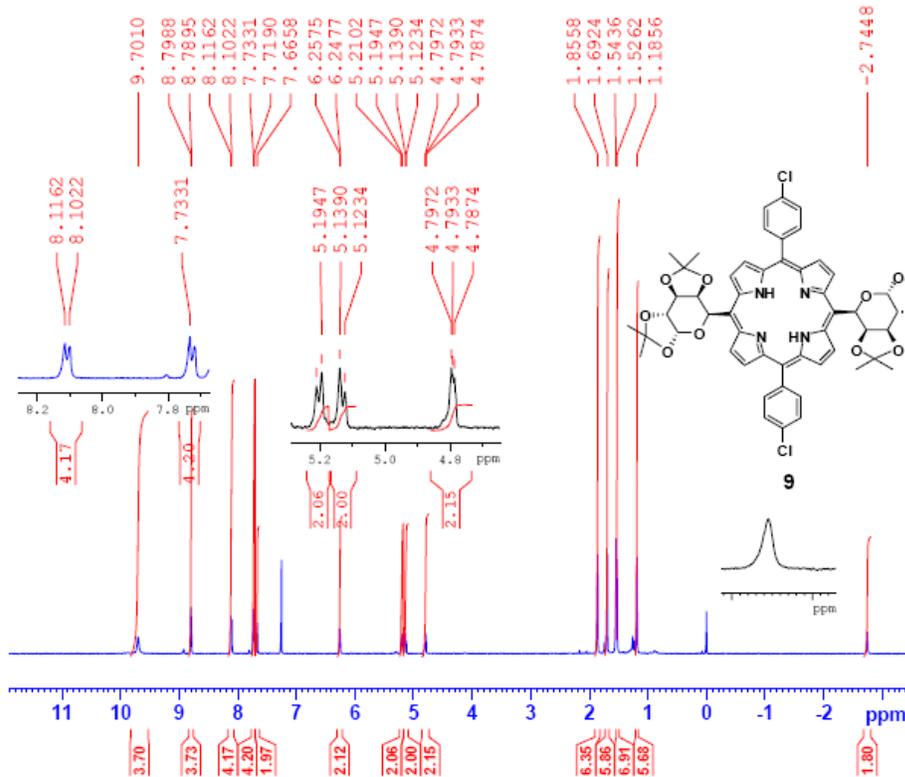
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
1099.2979	1099.2976	0.3	0.3	29.5	93.9	0.0	C54 H45 N4 O10 F10



1: TOF MS ES+
2.33e+003



¹H NMR, CDCl₃, 500 MHz, compound 9



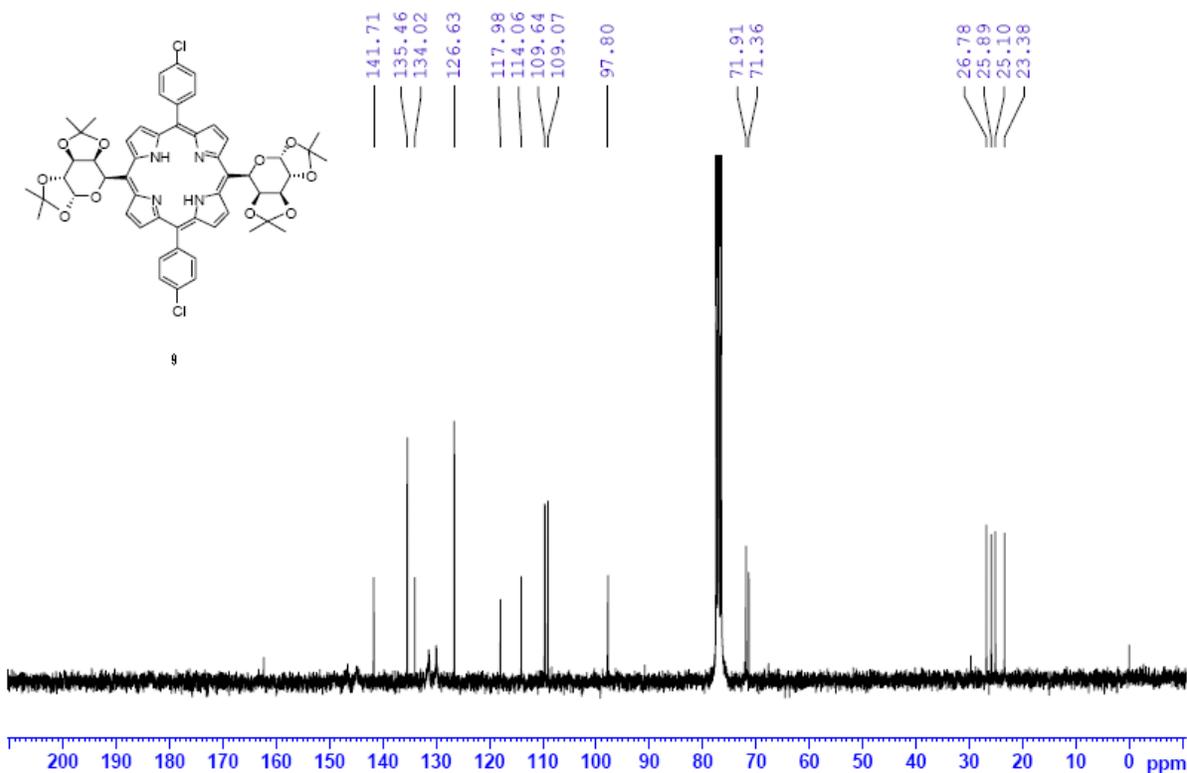
Current Data Parameters
NAME seen094
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20081211
Time 18.27
INSTRUM spect
PROBHD 5 mm PABBI 1H/
PULPROG zg30
TD 18748
SOLVENT CDCl3
NS 8
DS 0
SWH 10000.000 Hz
FIDRES 0.593390 Hz
AQ 0.9374500 sec
RG 2050
DW 50.000 usec
DE 6.000 usec
TE 295.8 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 6.80 usec
PL1 0.00 dB
SFO1 500.1328782 MHz

F2 - Processing parameters
SI 32768
SF 500.1320159 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

seen093, ¹³C, p chloro



Elemental Composition Report

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Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

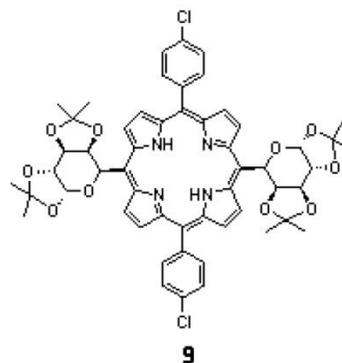
273 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)

Elements Used:

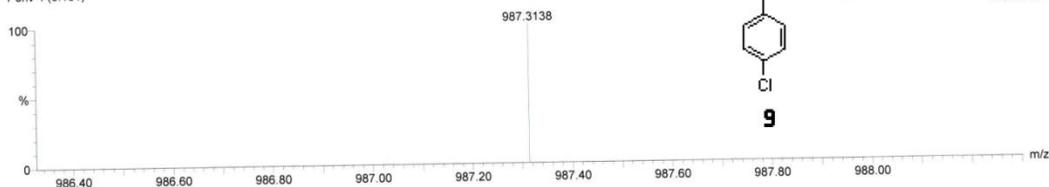
C: 0-54 H: 0-53 N: 0-4 O: 0-10 S: 0-1 Cl: 0-2

MW986

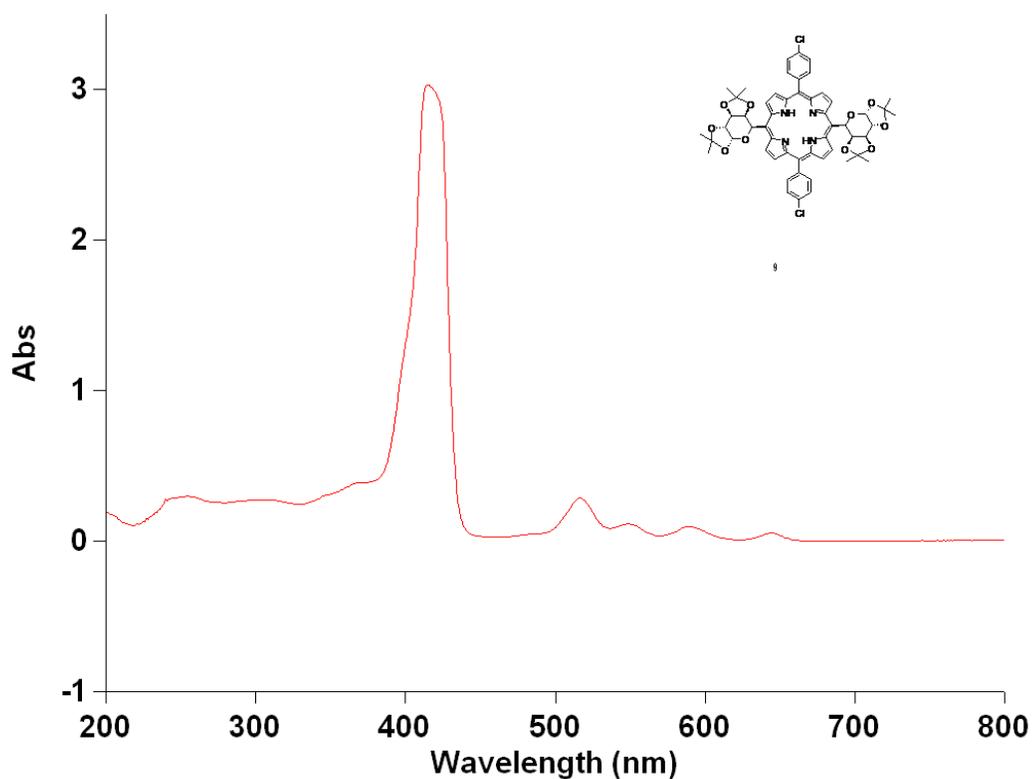
1-srv 4 (0.101)



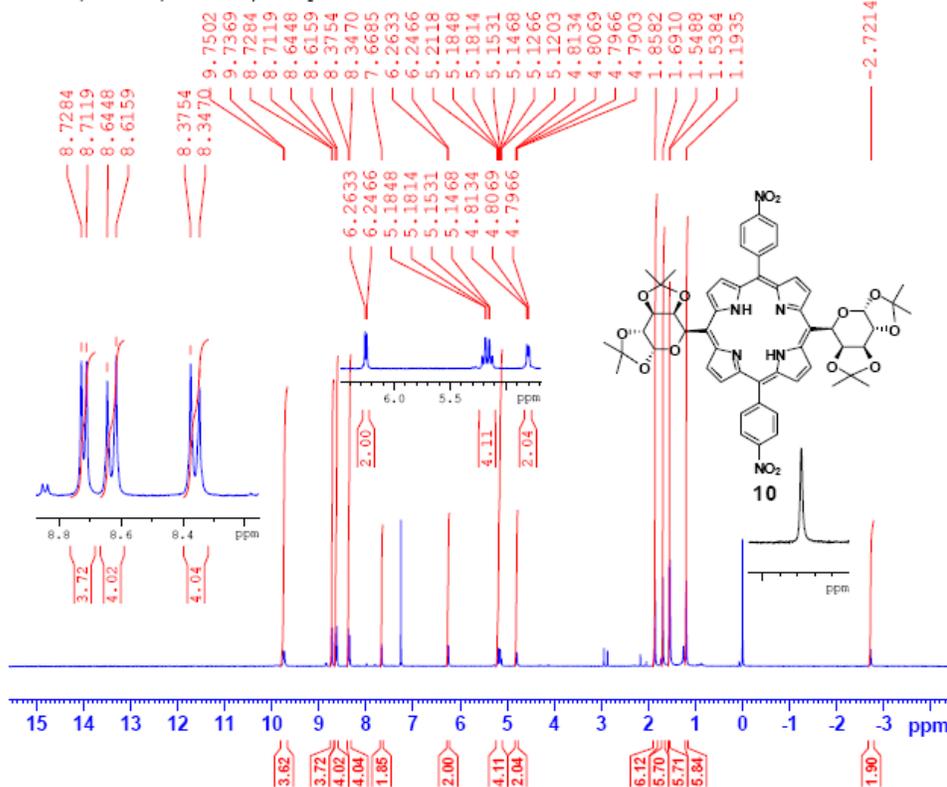
1: TOF MS ES+
8.38e+002



Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
987.3138	987.3139	-0.1	-0.1	29.5	25.4	0.0	C54 H53 N4 O10 Cl2



¹H NMR, CDCl₃, 300MHz, compound 10



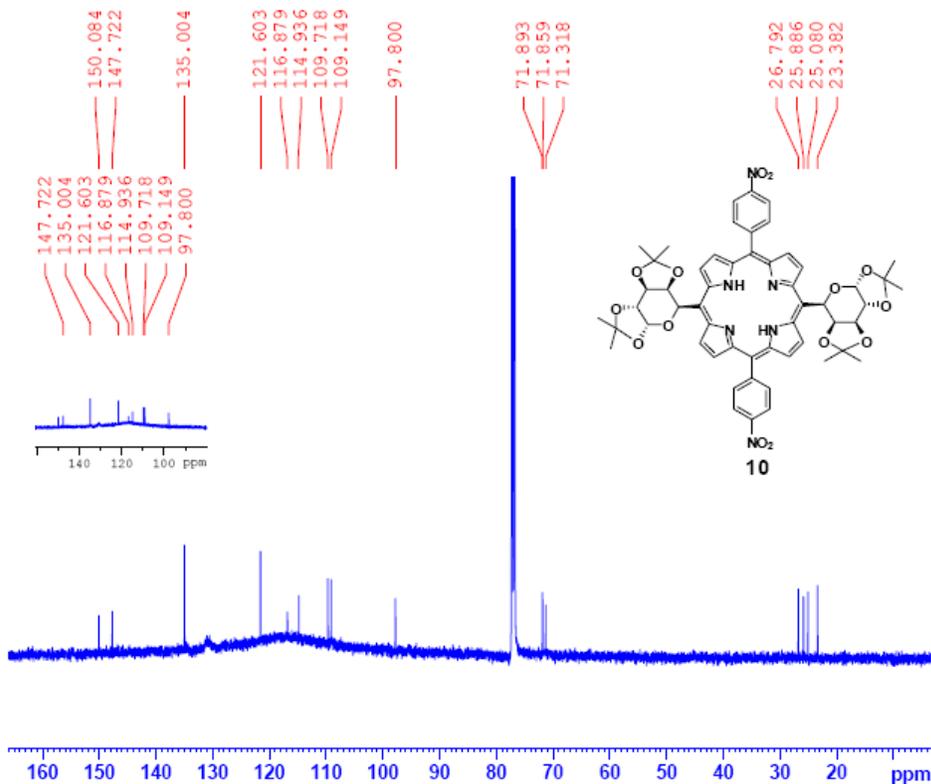
Current Data Parameters
NAME seen110
EXNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20081224
Time 13.59
INSTRUM spect
PROBHD 5 mm FABB0 BB-
PULPROG zg30
TD 20000
SOLVENT CDCl3
NS 16
DS 0
SWH 5995.204 Hz
FIDRES 0.299760 Hz
AQ 1.6680501 sec
RG 456.1
DW 456.100 usec
DE 63.400 usec
TE 296.9 K
D1 1.0000000 sec
TDO 1

===== CHANNEL f1 =====
NUC1 1H
P1 11.00 usec
PL1 -1.00 dB
SFO1 300.1316870 MHz

F2 - Processing parameters
SI 32768
SF 300.1300068 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹³C NMR, CDCl₃, 500MHz, compound 10



Current Data Parameters
NAME seen110
EXNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20081226
Time 22.33
INSTRUM spect
PROBHD 5 mm FABB1 1H/
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 10000
DS 4
SWH 29761.904 Hz
FIDRES 0.454131 Hz
AQ 1.1010548 sec
RG 2050
DW 16.800 usec
DE 10.00 usec
TE 296.1 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.8999999 sec
TDO 1

===== CHANNEL f1 =====
NUC1 13C
P1 14.50 usec
PL1 -2.00 dB
SFO1 125.7716224 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 0.00 dB
PL12 21.41 dB
PL13 22.00 dB
SFO2 500.1320005 MHz

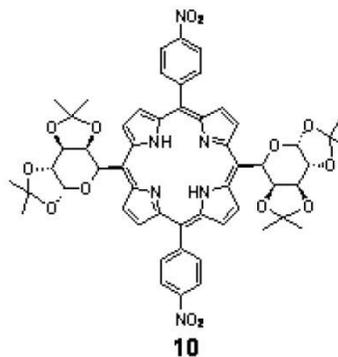
F2 - Processing parameters
SI 32768
SF 125.7577890 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

Elemental Composition Report

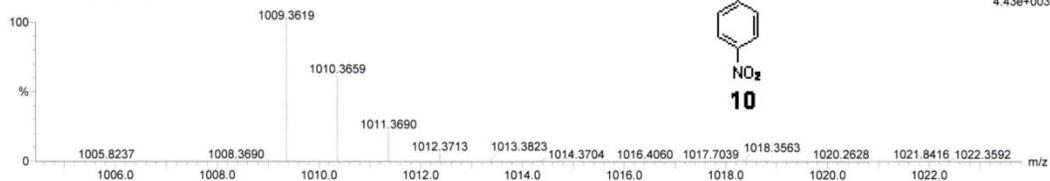
Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0
Element prediction: Off
Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions
91 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)
Elements Used:
C: 0-54 H: 0-53 N: 0-6 O: 0-14
MW1008
SNV-8.3 (0.082) Cm (3.13)

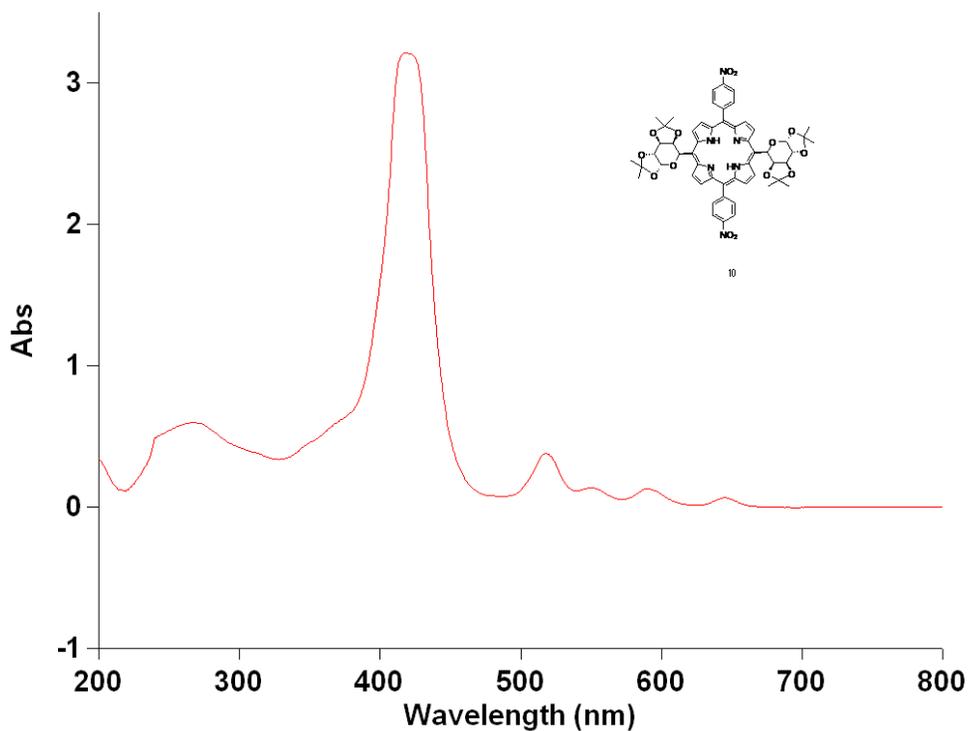


1: TOF MS ES+
4.43e+003

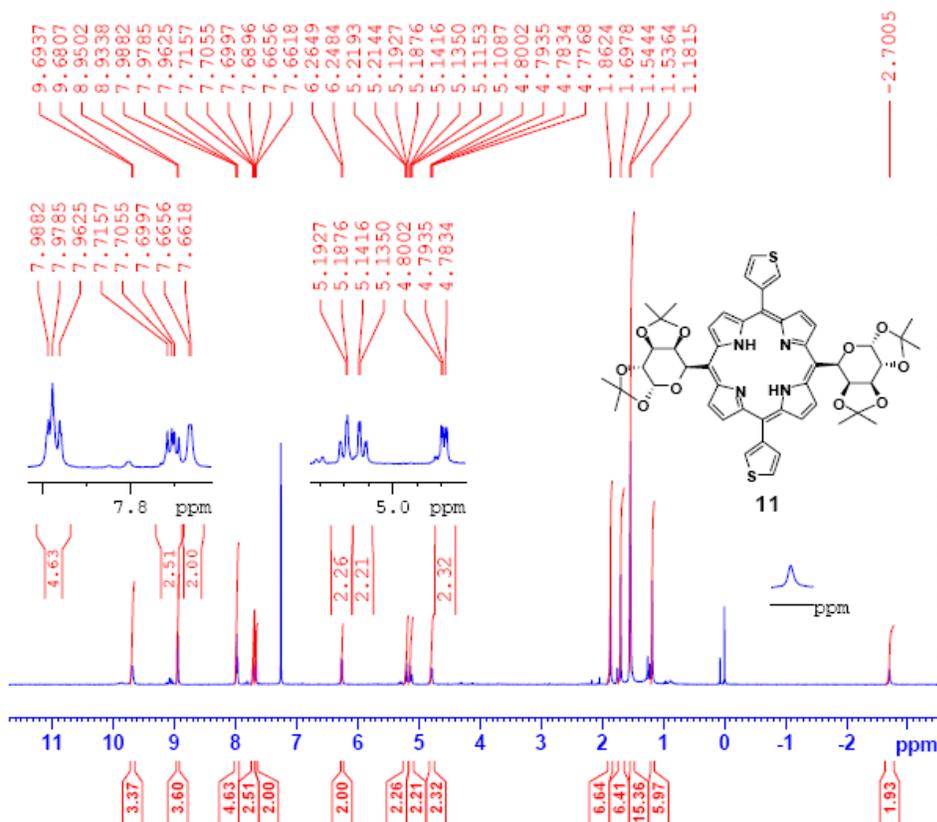


Minimum: -1.5
Maximum: 5.0 5.0 50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
1009.3619	1009.3620	-0.1	-0.1	31.5	107.6	0.0	C ₅₄ H ₅₃ N ₆ O ₁₄



¹H NMR, CDCl₃, 300MHz, Compound 11



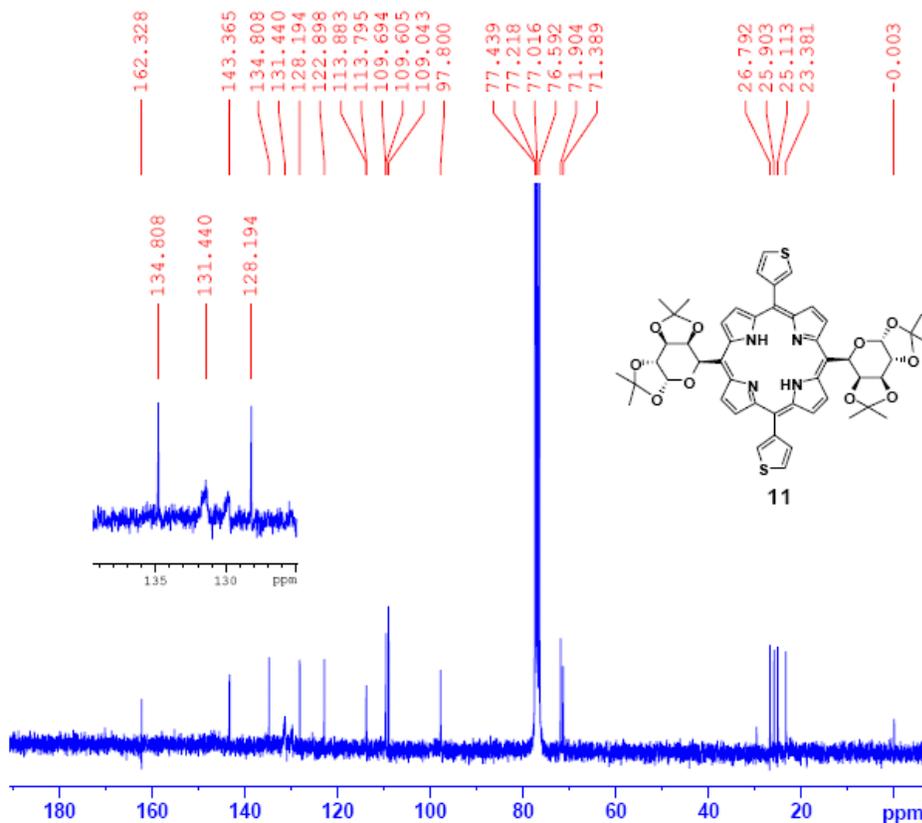
Current Data Parameters
NAME seen108
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20081223
Time 18.42
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 20000
SOLVENT CDCl₃
NS 16
DS 0
SWH 5995.204 Hz
FIDRES 0.299760 Hz
AQ 1.6680501 sec
RG 456.1
DW 83.403 usec
DE 6.00 usec
TE 297.1 K
D1 1.00000000 sec
TDO 1

===== CHANNEL #1 =====
NUC1 1H
P1 11.00 usec
PL1 -1.00 dB
SFO1 300.1316870 MHz

F2 - Processing parameters
SI 32768
SF 300.1300072 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
FC 1.00

¹³C NMR, CDCl₃, 300 MHz, compound11



Current Data Parameters
NAME seen090
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters
Date_ 20081209
Time 23.09
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl₃
NS 666
DS 0
SWH 18115.941 Hz
FIDRES 0.276427 Hz
AQ 1.8088436 sec
RG 32768
DW 27.600 usec
DE 6.00 usec
TE 297.5 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999999 sec
TDO 120

===== CHANNEL #1 =====
NUC1 13C
P1 10.00 usec
PL1 -1.00 dB
SFO1 75.4752965 MHz

===== CHANNEL #2 =====
CFDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -1.00 dB
PL12 16.23 dB
PL13 19.23 dB
SFO2 300.1312005 MHz

F2 - Processing parameters
SI 32768
SF 75.4677490 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
FC 1.40

Elemental Composition Report

Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0
Element prediction: Off
Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

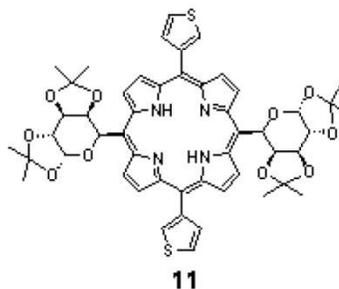
144 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)

Elements Used:

C: 0-50 H: 0-51 N: 0-4 O: 0-10 S: 0-2

MW930

3-snv 6 (0.139) Cm (6.36)

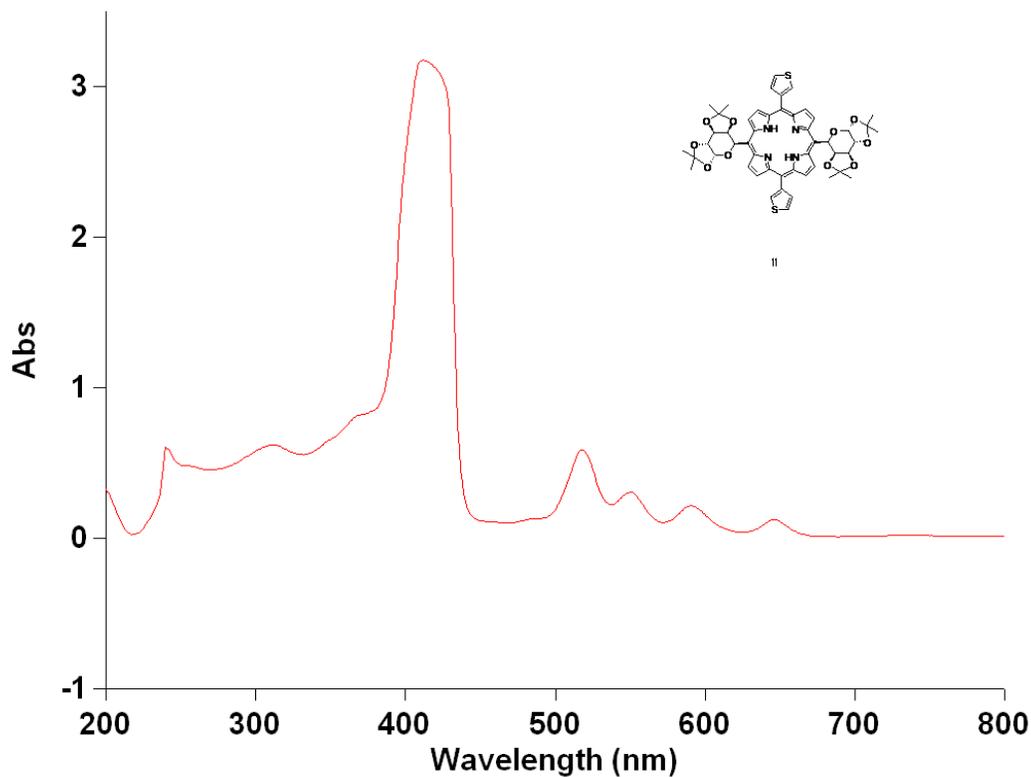


1: TOF MS ES+
3.67e+002

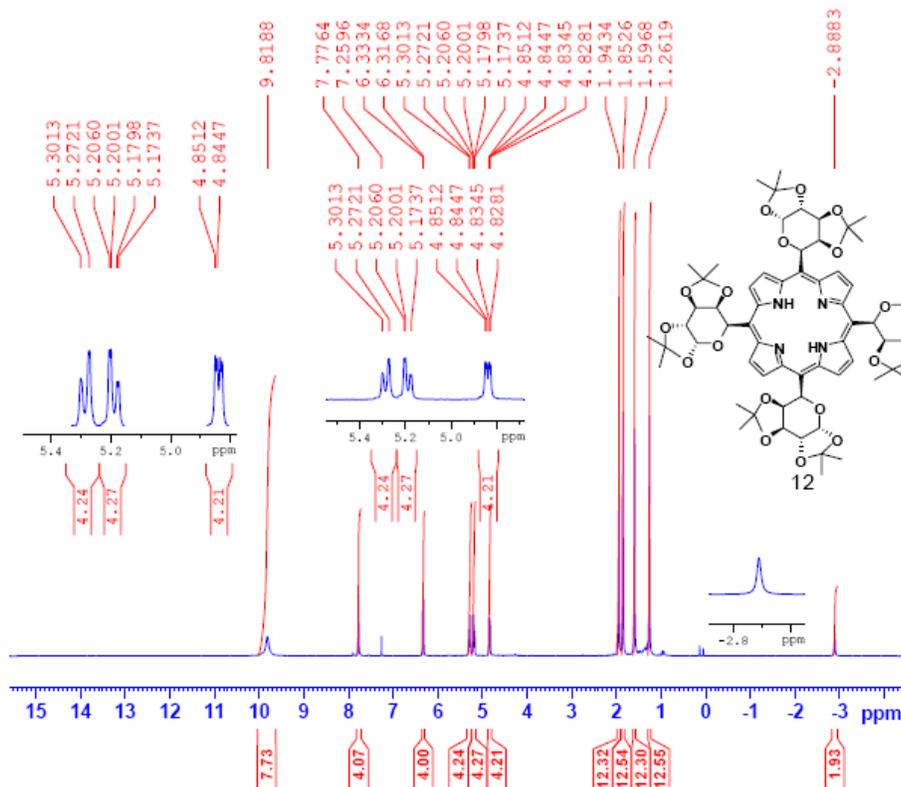


Minimum: -1.5
Maximum: 5.0 5.0 50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
931.3044	931.3047	-0.3	-0.3	27.5	24.6	0.0	C50 H51 N4 O10 S2



¹H NMR, CDCl₃, 300MHz, compound 12



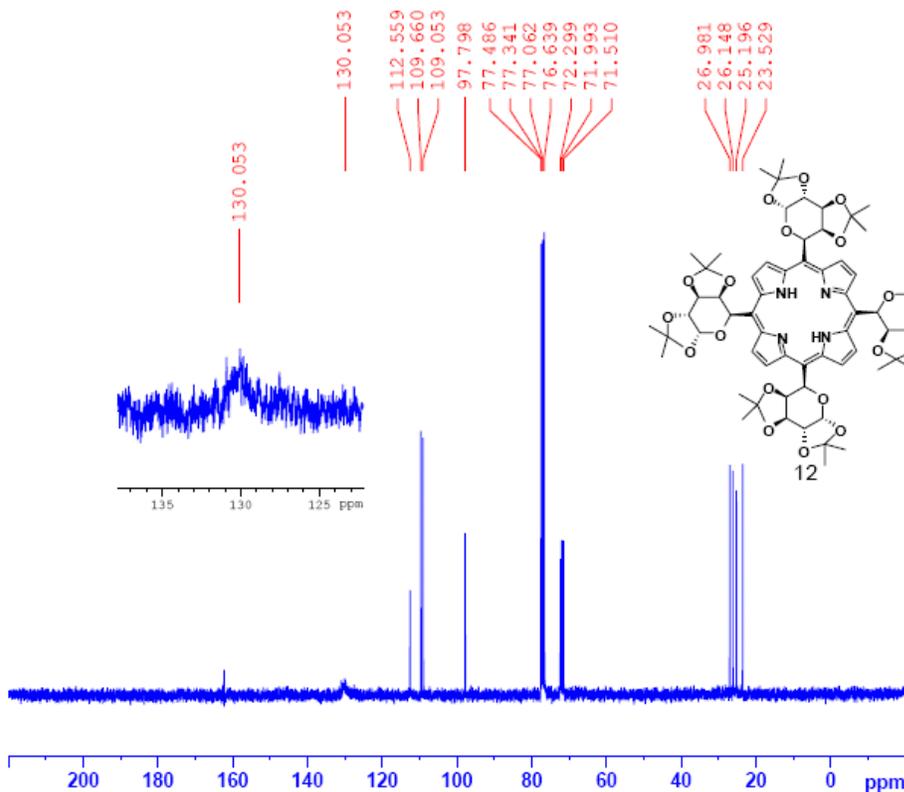
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Current Data Parameters
NAME      seen071
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20081129
Time     13.58
INSTRUM spect
PROBHD   5 mm PABBO BB-
PULPROG zg30
TD       20000
SOLVENT  CDCl3
NS       8
DS       0
SWH      5995.204 Hz
FIDRES   0.299760 Hz
AQ       1.6680501 sec
RG       64
DW       89.400 usec
DE       6.00 usec
TE       297.2 K
D1       1.00000000 sec
TDO      1

===== CHANNEL f1 =====
NUC1     1H
P1       11.00 usec
PL1      -1.00 dB
SFO1     300.1316870 MHz

F2 - Processing parameters
SI       32768
SF       300.1300062 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00
```

¹³C NMR, CDCl₃, 300MHz, compound 12



```
Current Data Parameters
NAME      seen071
EXPNO    2
PROCNO   1

F2 - Acquisition Parameters
Date_    20081129
Time     14.09
INSTRUM spect
PROBHD   5 mm PABBO BB-
PULPROG zgpg30
TD       65536
SOLVENT  CDCl3
NS       278
DS       0
SWH      18115.941 Hz
FIDRES   0.276427 Hz
AQ       1.8089496 sec
RG       18390.4
DW       27.600 usec
DE       6.00 usec
TE       297.9 K
D1       2.00000000 sec
d11      0.03000000 sec
DELTA    1.89999998 sec
TDO      120

===== CHANNEL f1 =====
NUC1     13C
P1       10.00 usec
PL1      -1.00 dB
SFO1     75.4752965 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2     1H
PCPD2    80.00 usec
PL2      -1.00 dB
PL12     16.23 dB
PL13     19.23 dB
SFO2     300.1312005 MHz

F2 - Processing parameters
SI       32768
SF       75.4677490 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40
```

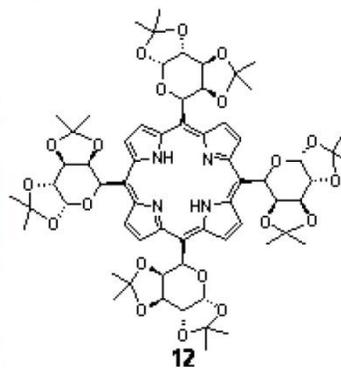
Elemental Composition Report

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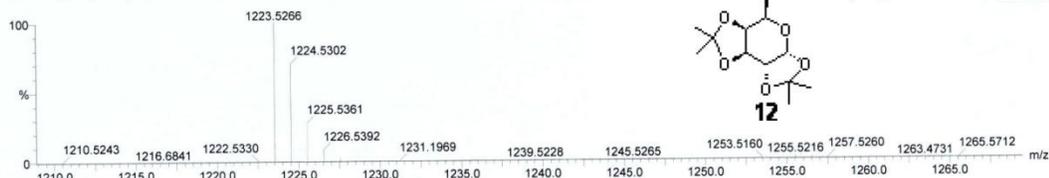
Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 50.0
Element prediction: Off
Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions
91 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)
Elements Used:
C: 0-64 H: 0-79 N: 0-4 O: 0-20
MW1222
SNV-10.3 (0.083) Cm (3:11)



1: TOF MS ES+
1.38e+004



Minimum: -1.5
Maximum: 50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
1223.5266	1223.5288	-2.2	-1.8	27.5	158.2	0.0	C ₆₄ H ₇₉ N ₄ O ₂₀

