## **Electronic Supplementary Information**

# Anti-colorectal Cancer Activity of an Organometallic Osmium Arene Azopyridine Complex

Steve D. Shnyder,\*<sup>a</sup> Ying Fu,<sup>b</sup> Abraha Habtemariam,<sup>b</sup> Sabine H. van Rijt,<sup>b</sup> Patricia A. Cooper,<sup>a</sup> Paul M. Loadman,<sup>a</sup> and Peter J. Sadler<sup>\*b</sup>

<sup>a</sup> Institute of Cancer Therapeutics, University of Bradford, Richmond Road, Bradford, BD7 1DP, U.K. E-mail: <u>S.D.Shnyder@Bradford.ac.uk</u>; Fax: +44 1274233234 ; Tel: +44 1274 235898

<sup>b</sup> Department of Chemistry, University of Warwick, Gibbet Hill Road, Coventry, CV4 7AL, U.K. E-mail: <u>P.J.Sadler@warwick.ac.uk</u>; Fax: +44 24 76523819; Tel: +44 76523818

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#### Materials

The A2780 human ovarian carcinoma and A549 human lung cancer cell lines were purchased from European Collection of Animal Cell Cultures (Salisbury, UK), RPMI-1640 media and trypsin from Invitrogen, bovine serum from Biosera, penicillin, streptomycin, L-buthionine sulfoximine (L-BSO), trichloroacetic acid (TCA) and sulforhodamine B (SRB) from Sigma-Aldrich, and tris[hydroxymethyl]aminomethane from Formedium. Complex **1** (FY026) was synthesised and characterised as previously described (ref 8).

#### **Experimental Procedures**

**ICP-MS**. ICP-MS analyses were carried out on an Agilent Technologies 7500 Series ICP-MS instrument. The water used for ICP-MS analysis was double deionised (DDW) using a USF Elga UHQ water deionizer. The osmium Specpure plasma standard (Alfa Aesar, 1000 ppm in 5% HCl) was diluted with DDW to 20 ppm. The standards for calibration were freshly prepared by diluting this stock solution with 3% HNO<sub>3</sub> in DDW. The concentrations used were 100, 60, 20, 10, 5, 4, 2, 1, 0.4 and 0.1 ppb.

**ROS Detection.** DCFH-DA was handled under an N<sub>2</sub> atmosphere, dissolved in DMSO to give a 10 mM stock solution. A2780 cells were seeded at 5000 cells/well into black 96-well plate and incubated for 24 h at 310 K, 5% CO<sub>2</sub>, high humidity. Cells were incubated for 20 min with DCFH-DA (10  $\mu$ M). To remove extracellular probe, the cells were washed twice with PBS. The cells were then kept in PBS solution and either the osmium complex **1**, or **1** plus L-BSO (50  $\mu$ M), or hydrogen peroxide (50  $\mu$ M, as the positive control) were added. The fluorescence was recorded over a period of 4 h at 310 K by excitation at 480 nm and emission at 530 nm using a TECAN plate reader.

**Combination treatment with 1 and L-BSO.** To determine the effect of decreased GSH levels on the cytotoxicity of **1**, A2780 human ovarian and A549 human lung cancer cells were co-incubated with 50  $\mu$ M L-BSO and various concentrations of **1** for 24 h. Cell viability was then determined using the sulforhodamine (SRB) assay.

**Redox potentials.** The electrochemical study was carried out using the same conditions as for the ruthenium analog reported previously.<sup>1</sup>

**Animals.** Six to ten week old female BALB/cOlaHsd-Foxn1<sup>nu</sup> mice (Harlan, Blackthorn, UK), maintained with food (Teklad 2018 diet, Harlan) and water *ad libitum*, were used according to UKCCCR guidelines for the welfare of animals <sup>2</sup>. Regulated procedures were carried out under a UK Home Office Project License.

**Tumour System.** HCT-116 (human colon adenocarcinoma line) tumours were excised from a donor animal, placed in sterile physiological saline containing antibiotics and cut into small fragments of approximately 2 mm<sup>3</sup>. Under brief general inhalation anaesthesia, fragments were implanted in the left flank of each mouse using a trocar. Once the tumours could accurately be measured by calipers (mean tumour volume of 32 mm<sup>3</sup>), the mice were allocated into groups by restricted randomisation to keep group mean tumour size variation to a minimum.

**Plasma and Tissue Distribution Studies.** Tumour-bearing mice (n = 3 per time point) were sacrificed at 5 min, 60 min and 240 min following intravenous administration of a single 10 mg kg<sup>-1</sup> dose of FY026. Tumour, liver, lung, kidneys and plasma were collected, weighed and stored at 193 K until analyzed. Osmium concentrations in different samples were measured by ICP-MS using the methodology described above. Tumour and tissue samples were homogenized in phosphate buffered saline, and

digested in HNO<sub>3</sub> at 353 K overnight to give clear solutions. These were then diluted as appropriate for evaluation by ICP-MS.

**Chemotherapy Studies.** The compounds were administered by single injections, with the day of therapy designated day 0. The maximum soluble dose of FY026 when administered as a single intravenous injection was established as 40 mgkg<sup>-1</sup>. The efficacy of FY026 was compared with the standard anticancer agent cisplatin which was administered as a single dose intraperitoneally at its maximum tolerated dose of 8 mg kg<sup>-1</sup>. The effects of therapy were assessed as previously described.<sup>3</sup> Briefly, daily 2-dimensional caliper measurements of the tumours were taken, with volumes calculated using the formula  $(a^2 \ge b) / 2$ , where *a* is the smaller and *b* the larger diameter of the tumour. Tumour volume was then normalised to the respective volume on day 0, and semi-log plots of relative tumour volume (RTV) versus time were made. Mann-Whitney U tests were performed to determine the statistical significance of any differences in growth rate (based on tumour volume doubling time) between control and treated groups, and between the 2 compounds.



**Fig. S1** Concentration-versus-time point profiles of complex 1 in kidney, lung, liver, plasma and tumour. Points represent the mean concentration for three mice. Data are in Table 1.



Fig. S2 Relative changes in DCF fluorescence detected over time after exposure to 1  $\mu$ M complex 1 (FY026), 1  $\mu$ M 1 with 50  $\mu$ M L-BSO, and 50  $\mu$ M H<sub>2</sub>O<sub>2</sub>. For each experiment, the fluorescence was averaged over 8 wells.

### References

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