Unprecedented inhibition of resistant Penicillin Binding Proteins by *bis*-2oxoazetidinyl macrocycles

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Experimental Section

General

Experiments were performed under argon atmosphere in flame-dried glassware. All solvents, including anhydrous solvents, and reagents were purchased from Acros Organics, Alfa Aesar, Fluka, Sigma-Aldrich or VWR, and used without any further purification. TLC analyses were performed on aluminum plates coated with silica gel $60F_{254}$ (Merck) and visualized with a KMnO₄ solution and UV (254 nm) detection, and column chromatography was performed on silica gel (40-63 or 63-200 µm) purchased from Rocc. Melting points (mp) were determined on a Büchi B-540 apparatus calibrated with caffeine, vanillin, and phenacetin. [α]_D was measured on Perkin-Elmer 241 MC or 343 polarimeter, at 20 °C, in CHCl₃ or CH₃OH. Concentrations are given in g/100 mL. Nuclear magnetic resonance (¹H and ¹³C) spectra were recorded at 300 MHz for proton and 75 MHz for carbon (Bruker Avance 300) or 500 MHz for proton and 125 MHz for carbon (Bruker Avance 500) using deuterated chloroform (CDCl₃). Chemical shifts are reported in parts per million relative to residual CHCl₃ in CDCl₃ (7.26 and 77.16 ppm). NMR coupling constants (*J*) are reported in hertz. Infrared (IR) spectra were recorded using FTIR-8400S Shimadzu apparatus. Products were analyzed as thin films deposited on a Se-Zn crystal by evaporation from CH₂Cl₂ solutions. High Resolution Mass Spectrometry (HRMS) analyses were performed at the University of Mons Hainaut (Belgium) or at the University College London (UK).

General procedure for the preparation of bis-acylated compounds 3

Trifluoroacetic acid (13.57 mmol) was added to **2** (0.68 mmol) dissolved in CH_2Cl_2 (7 mL) at 0 °C. The mixture was warmed to rt and stirred for 2 h. Concentration of the reaction solution afforded the crude trifluoroacetate salt as a viscous oil. Then alkenoyl chloride (1.02 mmol) was added to a stirred solution of the crude trifluoroacetate salt and triethylamine (2.04 mmol) in CH_2Cl_2 (5 mL) cooled at 0 °C. The mixture was then warmed to r.t. and stirred overnight. The mixture was then diluted with CH_2Cl_2 (25 mL), and sequentially washed with HCl 2 M solution (30 mL), saturated aqueous NaHCO₃ (30 mL), and brine (40 mL). After drying over MgSO₄ and removing the solvent under reduced pressure, the residue was purified by flash column chromatography (hexane/EtOAc 3/2), to provide **3** as a white solid.

3a, **3e** and **3i** have been described previously.¹

The isolated yields and the spectral data for **3b-d**, **3f-h** are as follows.

N-[(3S)-2-oxo-1-(pent-4-enoyl)azetidin-3-yl]hex-5-enamide (3b). Yield: 83%; mp 101.5-102.2 °C; [α]_D +16.4 (*c* 2.9 in CHCl₃); IR: v 3298, 3078-2844, 1798, 1780, 1693, 1652, 1541 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 1.68-1.78 (m, 2H), 2.04-2.12 (m, 2H), 2.21-2.26 (m, 2H), 2.37-2.44 (m, 2H), 2.79-2.86 (m, 2H), 3.67 (dd, J = 3.9, 7.4 Hz, 1H), 3.85 (m, 1H), 4.70 (td, J = 3.8, 7.0 Hz, 1H), 4.96-5.10 (m, 4H), 5.68-5.89 (m, 2H), 6.38 (d, J = 7.3 Hz, 1H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 24.2, 27.9, 33.6, 35.0, 35.9, 44.9, 55.9, 115.7, 115.9, 136.5, 137.6, 164.8, 170.6, 173.6; MS (ESI) *m/z* (%) 551 [2M + Na]⁺ (36), 287 [M + Na]⁺ (100), 265 [M + H]⁺ (12); HRMS (ESI) calcd. C₁₄H₂₀N₂O₃Na [M + Na]⁺ 287.1372, found 287.1380.

N-[(3S)-2-oxo-1-(pent-4-enoyl)azetidin-3-yl]hept-6-enamide (3c). Yield: 85%; mp 98.5-99.7 °C; $[\alpha]_D$ +15.3 (*c* 3.4 in CHCl₃); IR: v 3313-3300, 2980-2856, 1798, 1697, 1663, 1641, 1526 cm⁻¹; ¹H NMR (300

¹ A. Sliwa, G. Dive, J.-L. Habib Jiwan, J. Marchand-Brynaert, *Tetrahedron*, 2010, **66**, 9519.

MHz, CDCl₃, 20 °C): δ 1.37-1.47 (m, 2H), 1.60-1.70 (m, 2H), 2.03-2.10 (m, 2H), 2.22-2.27 (m, 2H), 2.39-2.46 (m, 2H), 2.75-2.93 (m, 2H), 3.68 (dd, *J* = 3.9, 7.4 Hz, 1H), 3.87 (m, 1H), 4.71 (td, *J* = 3.8, 7.0 Hz, 1H), 4.93-5.11 (m, 4H), 5.71-5.90 (m, 2H), 6.09 (d, *J* = 7.0 Hz, 1H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 24.8, 28.0, 28.5, 33.5, 35.8, 36.0, 45.1, 55.9, 115.0, 115.9, 136.5, 138.3, 164.7, 170.6, 173.6; MS (ESI) *m*/*z* (%) 579 [2M + Na]⁺ (27), 301 [M + Na]⁺ (100), 279 [M + H]⁺ (31); HRMS (ESI) calcd. C₁₅H₂₂N₂O₃Na [M + Na]⁺ 301.1528, found 301.1534.

N-[(3S)-1-(hex-5-enoyl)-2-oxoazetidin-3-yl]pent-4-enamide (3d). Yield: 85%; mp 95.1-96.3 °C; [α]_D +17.8 (*c* 3.1 in CHCl₃); IR: v 3300, 3080-2872, 1798, 1778, 1693, 1654, 1537 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 1.70-1.80 (m, 2H), 2.07-2.14 (m, 2H), 2.29-2.40 (m, 4H), 2.69-2.74 (td, J = 2.8, 7.4 Hz, 2H), 3.65 (dd, J = 3.9, 7.4 Hz, 1H), 3.84 (m, 1H), 4.70 (td, J = 3.9, 7.0 Hz, 1H), 4.95-5.09 (m, 4H), 5.70-5.86 (m, 2H), 6.41 (br s, 1H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 23.2, 29.2, 33.1, 35.0, 36.0, 44.9, 55.8, 115.5, 116.2, 136.5, 137.7, 164.7, 171.3, 173.0; MS (ESI) m/z (%) 551 [2M + Na]⁺ (85), 287 [M + Na]⁺ (100), 265 [M + H]⁺ (21); HRMS (ESI) calcd. C₁₄H₂₀N₂O₃Na [M + Na]⁺ 287.1372, found 287.1367.

N-[(3S)-1-(hex-5-enoyl)-2-oxoazetidin-3-yl]hept-6-enamide (3f). Yield: 82%; mp 85.8-86.5 °C; $[\alpha]_{\rm D}$ +15.3 (*c* 3.3 in CHCl₃); IR: v 3296, 3080-2854, 1797, 1780, 1693, 1651, 1537 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 1.36-1.46 (m, 2H), 1.59-1.69 (m, 2H), 1.71-1.80 (m, 2H), 2.02-2.15 (m, 4H), 2.21-2.26 (m, 2H), 2.69-2.75 (m, 2H), 3.66 (dd, *J* = 3.9, 7.4 Hz, 1H), 3.85 (m, 1H), 4.70 (td, *J* = 3.9, 7.0 Hz, 1H), 4.92-5.06 (m, 4H), 5.70-5.84 (m, 2H), 6.24 (d, *J* = 7.1 Hz, 1H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 23.2, 24.8, 28.4, 33.1, 33.5, 35.8, 36.0, 45.0, 55.8, 115.0, 115.5, 137.7, 138.3, 164.7, 171.2, 173.6; MS (ESI) *m/z* (%) 607 [2M + Na]⁺ (21), 315 [M + Na]⁺ (100), 293 [M + H]⁺ (24); HRMS (ESI) calcd. C₁₆H₂₄N₂O₃Na [M + Na]⁺ 315.1685, found 315.1684.

N-[(3S)-1-(hept-6-enoyl)-2-oxoazetidin-3-yl]pent-4-enamide (3g). Yield: 70%; mp 105.2-106.6 °C; $[α]_D$ +17.3 (*c* 3.3 in CHCl₃); IR: v 3296, 2978-2852, 1799, 1780, 1691, 1653, 1541 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 1.39-1.49 (m, 2H), 1.62-1.72 (m, 2H), 2.03-2.10 (m, 2H), 2.30-2.41 (m, 4H), 2.69-2.75 (m, 2H), 3.65 (dd, *J* = 3.9, 7.4 Hz, 1H), 3.85 (m, 1H), 4.71 (td, *J* = 3.9, 7.0 Hz, 1H), 4.91-5.10 (m, 4H), 5.71-5.87 (m, 2H), 6.30 (d, *J* = 7.0 Hz, 1H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 23.5, 28.4, 29.2, 33.5, 35.1, 36.5, 45.0, 55.8, 114.8, 116.2, 136.5, 138.5, 164.7, 171.3, 173.0; MS (ESI) *m/z* (%) 579 [2M + Na]⁺ (53), 301 [M + Na]⁺ (100), 279 [M + H]⁺ (22); HRMS (ESI) calcd. C₁₅H₂₂N₂O₃Na [M + Na]⁺ 301.1528, found 301.1530.

N-[(3S)-1-(hept-6-enoyl)-2-oxoazetidin-3-yl]hex-5-enamide (3h). Yield: 79%; mp 87.1-87.9 °C; [α]_D +14.1 (*c* 3.0 in CHCl₃); IR: v 3296, 3065-2852, 1798, 1780, 1693, 1655, 1535 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 1.39-1.49 (m, 2H), 1.62-1.78 (m, 4H), 2.03-2.12 (m, 4H), 2.21-2.26 (m, 2H), 2.69-2.75 (m, 2H), 3.66 (dd, J = 3.9, 7.4 Hz, 1H), 3.85 (m, 1H), 4.70 (td, J = 3.9, 7.0 Hz, 1H), 4.91-5.04 (m, 4H), 5.68-5.85 (m, 2H), 6.26 (d, J = 7.0 Hz, 1H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 23.5, 24.2, 28.4, 33.0, 33.5, 35.0, 36.5, 45.0, 55.8, 114.8, 115.7, 137.6, 138.4, 164.8, 171.3, 173.5; MS (ESI) *m/z* (%) 607 [2M + Na]⁺ (100), 315 [M + Na]⁺ (78), 293 [M + H]⁺ (11); HRMS (ESI) calcd. C₁₆H₂₄N₂O₃Na [M + Na]⁺ 315.1685, found 315.1693.

General procedure of RCM for the preparation of bis-acylated compounds 4

Grubbs catalyst (second generation) (0.05 equiv) was added to a stirred solution of **3** (1 equiv) in dry CH_2Cl_2 (5 mM) and the solution was stirred at reflux under argon for 4 h. Then a second addition of Grubbs catalyst (0.05 equiv) was made and the mixture was additionally stirred at reflux for 20 h. Then the solvent was removed under reduced pressure and the crude product was purified thrice by column chromatography (EtOAc/MeOH 98/2), to provide **4** as pale-brown oil.

4e and **4i** have been described previously.¹

The isolated yields and the spectral data for **4b-d**, **4f-h** are as follows.

Non-symmetrical cyclic dimer (4b). Yield: 35%; $[\alpha]_D$ -44.7 (*c* 1.0 in CHCl₃); IR: v 3350-3255, 2983-2925, 1796, 1744, 1724, 1720, 1709, 1668, 1537, 1444 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, 20 °C): δ 1.61-1.81 (m, 4H), 1.98-2.50 (m, 12H), 2.64-3.16 (m, 4H), 3.48-3.69 (m, 2H), 3.77-3.97 (m, 2H), 4.71-5.11 (m, 2H), 5.29-5.52 (m, 4H), 6.80-7.11 and 7.35-7.48 (m, 2H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 23.9, 24.3, 27.2, 27.4, 31.0, 31.7, 34.1, 35.0, 35.8, 36.0, 44.9, 45.5, 55.3, 129.2, 129.7, 130.2, 130.5, 130.7, 130.8, 165.4, 165.6, 171.2, 171.5, 173.7, 174.0; MS (ESI) *m*/*z* (%) 495 [M + Na]⁺ (100), 481 [M + Na - CH₂]⁺ (16), 473 [M + H]⁺ (50), 459 [M + H - CH₂]⁺ (8), 445 [M + H - CO]⁺ (11); HRMS (ESI) calcd. C₂₄H₃₂N₄O₆Na [M + Na]⁺ 495.2220, found 495.2221.

Non-symmetrical cyclic dimer (4c). Yield: 33%; $[\alpha]_D$ -10.5 (*c* 2.0 in CHCl₃); IR: v 3284, 2981, 1780, 1693, 1512 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, 20 °C): δ 1.24-1.39 (m, 4H), 1.54-1.62 (m, 4H), 1.99-2.08 (m, 4H), 2.21-2.36 (m, 8H), 2.67-2.93 (m, 4H), 3.53-3.64 (m, 2H), 3.82-3.95 (m, 2H), 4.83-5.01 (m, 2H), 5.34-5.41 (m, 4H), 6.92-7.13 (m, 2H); ¹³C (125 MHz, CDCl₃, 20 °C): δ 22.4, 24.3, 24.6, 25.1, 26.2, 26.8, 27.4, 27.9, 28.4, 28.9, 29.5, 29.8, 31.7, 31.8, 35.4, 35.9, 36.1, 36.4, 45.2, 45.6, 55.5, 55.6, 127.9, 128.7, 129.8, 128.9, 130.7, 131.3, 131.8, 165.3, 165.5, 171.1, 171.4, 173.9, 174.0; MS (ESI) *m/z* (%) 523 [M + Na]⁺ (100); HRMS (ESI) calcd. C₂₆H₃₆N₄O₆Na [M + Na]⁺ 523.2533, found 523.2513.

Non-symmetrical cyclic dimer (4d). Yield: 21%; $[\alpha]_D$ -43.0 (*c* 1.1 in CHCl₃); IR: v 3390-3258, 2962-2858, 1796, 1745, 1734, 1712-1647, 1539, 1437 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, 20 °C): δ 1.56-2.44 (m, 16H), 2.55-3.05 (m, 4H), 3.39-3.67 (m, 2H), 3.77-3.92 (m, 2H), 4.83-5.15 (m, 2H), 5.29-5.60 (m, 4H), 6.82-7.21 and 7.30-7.40 (m, 2H); ¹³C (125 MHz, CDCl₃, 20 °C): δ 22.5, 23.0, 23.1, 24.0, 26.0, 26.1, 28.1, 28.3, 31.0, 31.2, 34.7, 35.0, 35.1, 35.9, 36.4, 36.8, 44.8, 45.3, 45.7, 55.2, 55.4, 55.5, 55.7, 129.0, 130.0, 130.2, 130.5, 130.8, 130.9, 131.0, 165.2, 165.3, 171.3, 171.5, 173.5, 173.6; MS (ESI) *m/z* (%) 495 [M + Na]⁺ (100), 481 [M + Na - CH₂]⁺ (17), 473 [M + H]⁺ (9), 445 [M + H - CO]⁺ (9); HRMS (ESI) calcd. C₂₄H₃₂N₄O₆Na [M + Na]⁺ 495.2220, found 495.2221.

Non-symmetrical cyclic dimer (4f). Yield: 57%; $[\alpha]_D$ -11.6 (*c* 1.0 in CHCl₃); IR: v 3408-3231, 2960-2833, 1789, 1720, 1693, 1674, 1531, 1456 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 1.24-1.81 (m, 12H), 1.95-2.34 (m, 12H), 2.56-2.77 (m, 4H), 3.48-3.69 (m, 2H), 3.79-3.93 (m, 2H), 4.65-4.99 (m, 2H), 5.23-5.47 (m, 4H), 6.82-7.24 (m, 2H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 22.4, 22.9, 24.3, 24.7, 26.4, 28.3, 28.8, 31.3, 31.4, 31.6, 31.9, 35.1, 35.2, 35.7, 35.9, 36.1, 36.4, 44.8, 45.3, 55.2, 55.3, 129.3, 129.9, 130.8, 131.9, 165.2, 165.5, 171.4, 171.7, 174.2, 174.4; MS (ESI) *m*/*z* (%) 551 [M + Na]⁺ (100), 537 [M + Na - CH₂]⁺ (25), 529 [M + H]⁺ (9); HRMS (ESI) calcd. C₂₈H₄₀N₄O₆Na [M + Na]⁺ 551.2846, found 551.2851.

Non-symmetrical cyclic dimer (4g). Yield: 41%; $[\alpha]_D$ -3.8 (*c* 3.6 in CHCl₃); IR: v 3310, 2924, 1794, 1689, 1529 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 1.24-1.43 (m, 4H), 1.54-1.66 (m, 4H), 1.96-2.14 (m, 4H), 2.20-3.07 (m, 12H), 3.41-3.89 (m, 4H), 4.67-5.14 (m, 2H), 5.35-5.61 (m, 4H), 6.64-7.20 (m, 2H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 23.0, 23.4, 27.6, 27.7, 28.1, 28.2, 28.4, 28.5, 29.8, 31.4, 31.8, 35.3, 36.1, 36.8, 45.0, 45.3, 55.2, 55.6, 129.3, 129.9, 130.7, 131.2, 132.0, 165.2, 171.7, 171.8, 171.9, 173.3, 173.4; MS (ESI) *m*/*z* (%) 523 [M + Na]⁺ (100), 509 [M + Na - CH₂]⁺ (42), 501 [M + H]⁺ (39), 487 [M + H - CH₂]⁺ (21), 473 [M + H - CO]⁺ (21), 459 [M + H - CH₂ - CO]⁺ (8); HRMS (ESI) calcd. C₂₆H₃₆N₄O₆Na [M + Na]⁺ 523.2533, found 523.2533.

Non-symmetrical cyclic dimer (4h). Yield: 31%; $[\alpha]_D$ -5.2 (*c* 1.0 in CHCl₃); IR: v 3389-3221, 2986-2885, 1796, 1747, 1720, 1697, 1683, 1539, 1521 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, 20 °C): δ 1.35-1.87 (m, 12H), 1.96-2.31 (m, 12H), 2.59-2.79 (m, 4H), 3.43-3.68 (m, 2H), 3.75-3.89 (m, 2H), 4.63-4.98 (m, 1H), 5.18-5.49 (m, 5H), 6.57-7.18 and 7.30-7.53 (m, 2H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 22.6, 23.6, 26.1, 26.7, 27.9, 28.1, 31.1, 31.4, 32.0, 34.1, 34.5, 36.2, 36.4, 36.6, 45.4, 55.0, 130.1, 130.5, 130.7, 131.9, 165.7, 172.2, 173.3; MS (ESI) *m/z* (%) 551 [M + Na]⁺ (100), 537 [M + Na - CH₂]⁺ (45), 529 [M + H]⁺ (32), 515 [M + H - CH₂]⁺ (13), 501 [M + H - CO]⁺ (23); HRMS (ESI) calcd. C₂₈H₄₀N₄O₆Na [M + Na]⁺ 551.2846, found 551.2826.

General Procedure for Hydrogenation

To a stirred solution of compound obtained by RCM (1 eq.) in methanol (0.03 M) was added 10% Pd/C (0.05 eq.). After being stirred under hydrogen atmosphere (P = 1 atm) for 3 h at room temperature, the mixture was filtered through a short pad of Celite and concentrated under reduced pressure. The residue was purified by column chromatography (EtOAc/MeOH 98/2), to provide products as colourless oils.

5e and **5i** have been described previously.¹

The isolated yields and the spectral data for **5b-d**, **5f-h** are as follows.

Non-symmetrical cyclic dimer (5b). Yield: 48%; $[\alpha]_D$ -26.1 (*c* 1.0 in CHCl₃); IR: v 3294, 2924, 1791, 1742, 1720, 1705, 1647, 1533, 1435 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, 20 °C): δ 1.23-1.40 (m, 12H), 1.58-1.80 (m, 8H), 2.18-2.29 (m, 4H), 2.52-2.66 (m, 2H), 2.76-2.92 (m, 2H), 3.56-3.67 (m, 2H), 3.64-3.95 (m, 2H), 4.89-5.03 (m, 2H), 6.92-7.14 (m, 2H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 23.4, 24.3, 24.5, 24.9, 26.8, 27.1, 27.2, 27.3, 27.4, 27.5, 27.7, 35.4, 35.6, 35.7, 36.2, 45.2, 45.6, 55.5, 165.2, 165.3, 171.8, 172.1, 174.1, 174.2; MS (ESI) *m*/*z* (%) 499 [M + Na]⁺ (100), 485 [M + Na - CH₂]⁺ (15), 477 [M + H]⁺ (7); HRMS (ESI) calcd. C₂₄H₃₆N₄O₆Na [M + Na]⁺ 499.2533, found 499.2516.

Non-symmetrical cyclic dimer (5c). Yield: 52%; $[\alpha]_D$ +7.9 (*c* 1.6 in CHCl₃); IR: v 3287, 2923, 1796, 1726, 1693, 1672, 1535 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 1.27-1.32 (m, 16H), 1.62-1.74 (m, 8H), 2.22-2.27 (m, 4H), 2.54-2.85 (m, 4H), 3.57-3.69 (m, 2H), 3.85-3.98 (m, 2H), 4.76-4.81 (m, 1H), 4.92-4.98 (m, 1H), 6.64-6.85 (m, 2H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 23.9, 24.2, 25.0, 27.4, 27.7, 28.1, 28.2, 28.6, 35.5, 35.9, 36.0, 36.3, 45.1, 45.7, 55.6, 55.8, 165.1, 165.2, 171.8, 172.1, 174.0, 174.2; MS (ESI) *m*/*z* (%) 527 [M + Na]⁺ (100), 505 [M + H]⁺ (11); HRMS (ESI) calcd. C₂₆H₄₀N₄O₆Na [M + Na]⁺ 527.2846, found 527.2864.

Non-symmetrical cyclic dimer (5d). Yield: 22%; $[\alpha]_D$ -17.7 (*c* 0.6 in CHCl₃); IR: v 3263, 2924, 1796, 1747, 1731, 1718, 1705, 1647, 1541, 1437 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 1.26-1.47 (m, 10H), 1.56-1.87 (m, 10H), 2.21-2.32 (m, 4H), 2.49-2.67 (m, 2H), 2.80-3.01 (m, 2H), 3.57-3.68 (m, 2H), 3.84-3.94 (m, 2H), 4.82-5.20 (m, 2H), 6.50-7.09 (m, 2H); ¹³C (125 MHz, CDCl₃, 20 °C): δ 23.3, 23.5, 23.6, 23.8, 23.9, 24.1, 24.3, 24.4, 24.5, 24.6, 26.5, 26.7, 26.8, 26.9, 27.0, 27.0, 27.1, 27.2, 27.3, 27.4, 27.5, 27.7, 35.2, 35.4, 35.6, 35.7, 35.8, 36.1, 45.3, 45.5, 55.4, 55.5, 55.6, 55.7, 165.1, 165.2, 165.3, 171.8, 173.8, 173.9, 174.0, 174.1; MS (ESI) *m/z* (%) 499 [M + Na]⁺ (100), 485 [M + Na - CH₂]⁺ (17), 477 [M + H]⁺ (10), 449 [M + H - CO]⁺ (7); HRMS (ESI) calcd. C₂₄H₃₆N₄O₆Na [M + Na]⁺ 499.2533, found 499.2518.

Non-symmetrical cyclic dimer (5f). Yield: 63%; $[\alpha]_D$ -17.3 (*c* 1.0 in CHCl₃); IR: v 3369-3258, 2935-2850, 1790, 1745, 1731, 1724, 1710, 1649, 1533, 1456 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 1.20-1.39 (m, 20H), 1.52-1.76 (m, 8H), 2.18-2.27 (m, 4H), 2.52-2.85 (m, 4H), 3.58-3.68 (m, 2H), 3.80-3.92 (m, 2H), 4.66-4.88 (m, 2H), 6.81-6.99 (m, 2H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 23.8, 24.9, 25.0, 27.9, 28.0, 28.1, 28.2, 28.3, 28.4, 35.7, 35.8, 36.2, 36.4, 45.2, 45.3, 55.7, 165.0, 165.1, 171.8, 174.2; MS (ESI) *m*/*z* (%) 555 [M + Na]⁺ (100), 541 [M + Na - CH₂]⁺ (27), 533 [M + H]⁺ (20); HRMS (ESI) calcd. C₂₈H₄₄N₄O₆Na [M + Na]⁺ 555.3159, found 555.3134.

Non-symmetrical cyclic dimer (5g). Yield: 60%; $[\alpha]_D$ -23.9 (*c* 1 in CHCl₃); IR: v 3355, 2938, 1794, 1776, 1716, 1682, 1535 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 1.21-1.41 (m, 16H), 1.52-1.77 (m, 8H), 2.21-2.26 (m, 4H), 2.51-2.84 (m, 4H), 3.61-3.73 (m, 2H), 3.83-3.89 (m, 4H), 4.71-4.92 (m, 2H), 6.86-7.08 (m, 2H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 23.9, 24.5, 24.7, 24.9, 25.0, 27.7, 27.8, 28.0, 28.1, 28.2, 35.7, 35.9, 36.3, 45.1, 45.3, 55.6, 55.8, 165.2, 171.8, 174.1, 174.2; MS (ESI) *m/z* (%) 527 [M + Na]⁺ (100), 513 [M + Na - CH₂]⁺ (72), 505 [M + H]⁺ (14); HRMS (ESI) calcd. C₂₆H₄₀N₄O₆Na [M + Na]⁺ 527.2846, found 527.2874.

Non-symmetrical cyclic dimer (5h). Yield: 36%; $[\alpha]_D$ -18.5 (*c* 0.5 in CHCl₃); IR: v 3362-3223, 2924-2860, 1794, 1695, 1683, 1535, 1461 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 1.20-1.43 (m, 20H), 1.57-1.77 (m, 8H), 2.21-2.30 (m, 4H), 2.53-2.68 (m, 2H), 2.75-2.83 (m, 2H), 3.62-3.69 (m, 2H), 3.84-3.92 (m, 2H), 4.73-4.90 (m, 2H), 6.40-6.61 (m, 2H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 23.8, 23.9, 24.8, 24.9, 27.7, 27.9, 28.0, 28.1, 28.2, 28.3, 35.8, 36.4, 45.3, 55.7, 55.8, 165.0, 171.8, 171.9, 174.0, 174.1; MS (ESI) *m*/*z* (%) 555 [M + Na]⁺ (100), 541 [M + Na - CH₂]⁺ (47), 533 [M + H]⁺ (7); HRMS (ESI) calcd. C₂₈H₄₄N₄O₆Na [M + Na]⁺ 555.3159, found 555.3162.

N-[(3S)-2-oxo-1-(pent-4-enoyl)azetidin-3-yl]heptanamide (6). Following the general procedure for the preparation of bis-acylated compounds, **6** was obtained as a white solid. Yield: 80%; mp 115.7-116.5 °C; $[\alpha]_D$ +18.2 (*c* 1.5 in CHCl₃); IR: v 3294-3288, 2951-2867, 1797, 1780, 1693, 1651, 1539 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 0.84-0.88 (m, 3H), 1.23-1.43 (m, 6H), 1.55-1.65 (m, 2H), 2.19-2.25 (m, 2H), 2.37-2.44 (m, 2H), 2.73-2.88 (m, 2H), 3.67 (dd, *J* = 3.8, 7.4 Hz, 1H), 3.85 (m, 1H), 4.69 (td, *J* = 3.8, 6.9 Hz, 1H), 4.97-5.10 (m, 2H), 5.75-5.89 (m, 1H), 6.37 (br d, *J* = 7.3 Hz, 1H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 14.1, 22.6, 25.3, 28.0, 28.9, 31.6, 36.0, 45.0, 55.9, 115.9, 136.5, 164.8, 170.6, 173.8; MS (ESI) *m/z* (%) 303 [M + Na]⁺ (31), 281 [M + H]⁺ (100), 253 [M + H - CO]⁺ (17); HRMS (ESI) calcd. C₁₅H₂₄N₂O₃Na [M + Na]⁺ 303.1685, found 303.1681.

General procedure of CM for the preparation of non-cyclic dimers

Grubbs catalyst (second generation) (0.05 equiv) was added to a stirred solution of **6** or **9** (1 equiv) in dry CH_2Cl_2 (10 mM) and the solution was stirred at reflux under argon for 4 h. Then a second addition of Grubbs catalyst (0.05 equiv) was made and the mixture was additionally stirred at reflux for 8 h. Then the solvent was removed under reduced pressure and the crude product was purified thrice by column chromatography (hexane/EtOAc 2/8), to provide **7** or **10** as a pale-brown solid.

$N-[(3S)-1-\{8-[(3S)-3-heptanamido-2-oxoazetidin-1-yl]-8-oxooct-4-enoyl\}-2-oxoazetidin-3-indicating a statistical statistical$

yl]heptanamide (**7**). Yield : 77%; mp 135.4-136.2 °C; $[\alpha]_D$ +3.3 (*c* 1.0 in CHCl₃); IR: v 3310, 2920, 1798, 1724, 1688, 1666, 1529 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 0.85-0.89 (m, 6H), 1.28-1.35 (m, 12H), 1.57-1.67 (m, 4H), 2.20-2.25 (m, 4H), 2.33-2.39 (m, 4H), 2.72-2.86 (m, 4H), 3.60 (m, 2H), 3.90 (m, 2H), 4.88-4.94 (m, 2H), 5.46 (m, 2H), 6.53 and 6.63 (2 br d, *J* = 7.5 Hz, 2H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 14.1, 22.6, 25.4, 27.1, 29.0, 31.6, 36.0, 36.1, 45.5, 55.6, 129.2, 165.2, 171.0, 173.8; MS (ESI) *m/z* (%) 555 [M + Na]⁺ (100), 533 [M + H]⁺ (7); HRMS (ESI) calcd. C₂₈H₄₄N₄O₆Na [M + Na]⁺ 555.3159, found 555.3157.

$N-[(3S)-1-\{8-[(3S)-3-heptanamido-2-oxoazetidin-1-yl]-8-oxooctanoyl\}-2-oxoazetidin-3-indicating a statement of the statement$

yl]heptanamide (8). Following the general procedure for hydrogenation, compound **8** was obtained as a pale-brown solid. Yield: 60%; mp 145.8-146.4 °C; $[\alpha]_D$ +6.4 (*c* 2.0 in CHCl₃); IR: v 3283, 2930, 1789, 1697, 1650, 1547 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 0.84-0.88 (m, 6H), 1.27-1.36 (m, 16H), 1.56-1.65 (m, 8H), 2.19-2.26 (m, 4H), 2.63-2.76 (m, 4H), 3.60-3.65 (m, 2H), 3.84-3.90 (m, 2H), 4.78-4.85 (m, 2H), 6.56-6.70 (m, 2H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 14.1, 22.6, 23.9, 25.4, 28.5, 29.0, 31.6, 36.0, 36.3, 45.2, 55.7, 165.0, 171.5, 174.0; MS (ESI) *m*/*z* (%) 557 [M + Na]⁺ (100); HRMS (ESI) calcd. C₂₈H₄₆N₄O₆Na 557.3315, found 557.3323.

tert-butyl N-[(3S)-2-oxo-1-pentanoylazetidin-3-yl]carbamate. To a stirred solution of **1** (553 mg, 2.97 mmol) in dry CH₂Cl₂ (25 mL) were added pyridine (0.48 mL, 5.94 mmol) and valeroyl chloride (0.71 mL, 5.94 mmol). The mixture was stirred for 12 h at rt and then diluted with CH₂Cl₂ (30 mL) and the organic layer was washed with HCl 2 M solution (50 mL), saturated aqueous NaHCO₃ (50 mL), and brine (50 mL). After drying over MgSO₄ and removing the solvent under reduced pressure, the residue was purified by flash column chromatography (hexane/EtOAc 4/1), to provide the product as a white solid (321 mg, 40%). mp 114.8-115.3 °C; $[\alpha]_D$ +10.1 (*c* 1.0 in CHCl₃); IR: v 3362, 2976-2933, 1797, 1780, 1688, 1537 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 0.91 (t, *J* = 7.4Hz, 3H), 1.33-1.43 (m, 11H), 1.64 (m, 2H), 2.72 (m, 2H), 3.64 (dd, *J* = 3.8, 7.4 Hz, 1H), 3.86 (m, 1H), 4.45 and 4.68 (2 br s, 1H, rotamers), 5.17 (br d, *J* = 7.5Hz, 1H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 13.9, 22.4, 26.1, 28.3, 36.5, 45.6, 56.6, 81.2, 154.7, 165.1, 171.5; MS (ESI) *m/z* (%) 293 [M + Na]⁺ (100), 279 [M + H]⁺ (8); HRMS (ESI) calcd. C₁₃H₂₁N₂O₄[M - H]⁺ 269.1501, found 269.1494.

N-[(3S)-2-oxo-1-pentanoylazetidin-3-yl]hept-6-enamide (9). Following the general procedure for the preparation of bis-acylated compounds, starting from tert-butyl N-[(3S)-2-oxo-1-pentanoylazetidin-3-yl]carbamate, compound **9** was obtained as a white solid. Yield: 89%; mp 101.0-101.9 °C; $[\alpha]_D$ +15.8 (*c* 1.0 in CHCl₃); IR: v 3282, 2959-2923, 1780, 1728, 1718, 1693, 1655, 1537 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 0.91 (t, *J* = 7.3 Hz, 3H), 1.31-1.46 (m, 4H), 1.59-1.69 (m, 4H), 2.02-2.09 (m, 2H), 2.24 (t, *J* = 7.6 Hz, 2H), 2.68-2.74 (m, 2H), 3.66 (dd, *J* = 3.9, 7.4 Hz, 1H), 3.85 (m, 1H), 4.71 (td, *J* = 3.8, 6.8

Hz, 1H), 4.93-5.02 (m, 2H), 5.70-5.84 (m, 1H), 6.21 (br d, J = 7.0 Hz, 1H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 13.9, 22.4, 24.8, 26.2, 28.5, 33.5, 35.8, 36.4, 45.0, 55.8, 115.0, 138.3, 164.7, 171.5, 173.6; MS (ESI) m/z (%) 303 [M + Na]⁺ (100), 281 [M + H]⁺ (65), 253 [M + H - CO]⁺ (22); HRMS (ESI) calcd. C₁₅H₂₄N₂O₃Na [M + Na]⁺ 303.1685, found 303.1670.

N,N'-bis[(3S)-2-oxo-1-pentanoylazetidin-3-yl]dodec-6-enediamide (10). Following the general procedure for CM, compound 10 was obtained as a pale-brown solid. Yield: 74%; mp 146.1-146.8 °C; $[\alpha]_D$ +5.7 (*c* 1.0 in CHCl₃); IR: v 3292, 2962-2868, 1796, 1693, 1676, 1656, 1537 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 0.91 (t, *J* = 7.4 Hz, 6H), 1.24-1.43 (m, 8H), 1.58-1.72 (m, 8H), 2.00 (m, 4H), 2.20-2.25 (m, 4H), 2.68-2.73 (m, 4H), 3.63-3.67 (m, 2H), 3.82-3.87 (m, 2H), 4.70 (m, 2H), 5.34 (m, 2H), 6.54-7.03 (m, 2H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 13.9, 22.4, 24.7, 26.2, 28.7, 32.0, 34.3, 35.9, 36.4, 44.9, 55.8, 130.1, 130.5, 131.3, 164.8, 165.0, 171.6, 174.0, 174.1; MS (ESI) *m/z* (%) 555 [M + Na]⁺ (100), 541 [M + Na - CH₂]⁺ (57); HRMS (ESI) calcd. C₂₈H₄₄N₄O₆Na [M + Na]⁺ 555.3159, found 555.3160.

N,N'-bis[(3S)-2-oxo-1-pentanoylazetidin-3-yl]dodecanediamide (11). Following the general procedure for hydrogenation, compound **11** was obtained as a pale-brown solid. Yield: 82%; mp 165.4-166.3 °C; $[\alpha]_D$ +12.9 (*c* 1.0 in CHCl₃); IR: v 3300, 2930-2849, 1780, 1730, 1693, 1654, 1539 cm⁻¹; ¹H NMR (300 MHz, CDCl₃, 20 °C): δ 0.91 (t, *J* = 7.3 Hz, 6H), 1.26-1.42 (m, 16H), 1.58-1.68 (m, 8H), 2.19-2.24 (m, 4H), 2.67-2.72 (m, 4H), 3.63-3.67 (m, 2H), 3.82-3.86 (m, 2H), 4.67-4.73 (m, 2H), 6.53-6.70 (m, 2H); ¹³C (75 MHz, CDCl₃, 20 °C): δ 13.9, 22.3, 25.2, 25.3, 26.2, 27.8, 28.8, 29.0, 29.1, 35.8, 35.9, 36.4, 44.9, 55.8, 165.0, 171.6, 174.1; MS (ESI) *m/z* (%) 557 [M + Na]⁺ (100), 541 [M + Na – CH₂]⁺ (70); HRMS (ESI) calcd. C₂₈H₄₆N₄O₆Na [M + Na]⁺ 557.3315, found 557.3314.

Computational chemistry

All the calculations have been performed with the Gaussian03 suite of programs.² The B3LYP/6-31G(d) optimization of the dimers has been made with redundant internal coordinates in order to ensure a rapid convergence of each component of the gradient lower than 2.10-6 a.u. For the localization of the TS at the RHF/MINI-1' level, 3N-6 variables of the Z-matrix have been used. The eigenvector components associated to the negative eigenvalue of the Hessian matrix well combine the variables describing the pseudo 8-membered ring of the H transfer from Ser49 to the nitrogen of the β -lactam.

Acyl-enzymes modeling

The crystal structure of R39 complexed with a monobactam (unpublished results) was used as a template to model the acyl-enzyme between R39 and compound **5e**. The interactions made by the compound amide group with Asn300 side chain on one side, and with the backbone carbonyl of Thr413 on the other side were preserved. The remaining dihedral angles were adapted to avoid steric clashes between the compound and the protein. A similar procedure was used to model the acyl-enzyme between PBP2a and **5e**.

² Gaussian 03, Revision D.02, M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, J. A. Montgomery, Jr., T. Vreven, K. N. Kudin, J. C. Burant, J. M. Millam, S. S. Iyengar, J. Tomasi, V. Barone, B. Mennucci, M. Cossi, G. Scalmani, N. Rega, G. A. Petersson, H. Nakatsuji, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, M. Klene, X. Li, J. E. Knox, H. P. Hratchian, J. B. Cross, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, P. Y. Ayala, K. Morokuma, G. A. Voth, P. Salvador, J. J. Dannenberg, V. G. Zakrzewski, S. Dapprich, A. D. Daniels, M. C. Strain, O. Farkas, D. K. Malick, A. D. Rabuck, K. Raghavachari, J. B. Foresman, J. V. Ortiz, Q. Cui, A. G. Baboul, S. Clifford, J. Cioslowski, B. B. Stefanov, G. Liu, A. Liashenko, P. Piskorz, I. Komaromi, R. L. Martin, D. J. Fox, T. Keith, M. A. Al-Laham, C. Y. Peng, A. Nanayakkara, M. Challacombe, P. M. W. Gill, B. Johnson, W. Chen, M. W. Wong, C. Gonzalez, and J. A. Pople, Gaussian, Inc., Wallingford CT, 2004.

Biochemical evaluation

Assay with resistant PBPs

Purified PBP5 from *Enterococcus faecium* D63r and PBP2a from methicillin-resistant *Staphylococcus aureus* ATCC 43300 were used as target proteins to test inhibitory activity of synthesized β -lactams. Each of the purified PBPs (2.5 μ M) were first incubated with 1 mM potential inhibitor in 100 mM phosphate buffer, 0.01% Triton X-100,³ pH 7, for 4 h at 30 °C. Then, 25 μ M fluorescein-labeled ampicillin was added to detect the residual penicillin binding activity (RA). The samples were further incubated for 30 min at 37 °C in a total volume of 20 μ L. Denaturation buffer was added (0.1 M Tris/HCl, pH 6.8, containing 25% glycerol, 2% SDS, 20% β -mercaptoethanol and 0.02% bromophenol blue) and the samples were heated to 100 °C for 1 min. The samples were then loaded onto a 10% SDS-acrylamide gel (10 x 7 cm) and electrophoresis was performed for 45 min at 180 V (12 mA). Detection and quantification of the RAs were done with Molecular Image FX equipment and Quantity One software (BioRad, Hercules, CA, USA). Three independent experiments were carried out for each inhibitor.

³ B. Y. Feng, B. K. Shoichet, *Nat. Protoc.* 2006, **1**, 550.

Assay with R39

All assays with R39 have been done in microtiter plates 96-wells (Brand, Wertheim, Germany). 20 mM of the tested compounds have been solved in DMF. Finally 7.5 µL of the solution have been used in the assay. The final concentration of the compounds in the assays was 100 µM. The final concentration of DMF in the assays was 0.25%. R39 (3.5 nM) was incubated in the presence of the potential inhibitors in 10 mM sodium phosphate buffer (pH 7.2) with 100 mM NaCl, 100 mM D-alanine, 0.01 mg/ml BSA and 0.01% Triton for 60 min at 25 °C. This preincubation was realized, in order to detect also slow binding inhibitors. After the preincubation the residual activity RA of R39 was determined by observing the hydrolysis of the thiolester S2d substrate (i.e. N-benzoyl-D-Alanyl-thioglycolate), in the presence of DTNB, catalyzed by the non-inhibited enzyme. The initial rate of hydrolysis of 1 mM S2d in the presence of 1 mM DTNB was determined by monitoring the increase of absorbance at 412 nm (DTNB: $\varepsilon[\Delta \varepsilon] =$ 13600 M⁻¹ s⁻¹) using a microplate absorbance reader (Power Wave X, Biotek Instruments, Winooski, U.S.A.). The rate of spontaneous hydrolysis of S2d in the presence of the inhibitors was also determined in absence of R39. All assays have been done three times. The determination of RA of R39 in absence of inhibitors has been done six times on each plate. In order to detect false positives which could be slow binding non-competitive promiscuous inhibitors, the assays have been done in the presence of 0.01% Triton-X-100. 3,4



N-benzoyl-D-alanyl-thioglycolate (S2d):

5-(3-Carboxy-4-nitrophenyl)disulfanyl-2-nitrobenzoic acid (DTNB):

Hydrolysis of the thiolester S2d substrate by R39:



Formed thiol reacts with DTNB, affording 2-nitro-5-thiobenzoate, which ionizes in water at neutral and alkaline pH. This ion has a yellow color and is quantified by measuring the absorbance:



CO₂H

HO₂C

⁴ B. K. Shoichet, *Drug Discovery Today* 2006, **11**, 607.

Compound	Absolute Energy of Open Precursors (a.u.)			
	i	ii	iii	iv
3a	-841.345666490	-841.344443521	-841.353210506	-841.352690676
3b	-880.656567829	-880.653009341	-880.663916481	-880.661489845
3c	-919.971821898	-919.970354436	-919.979335270	-919.978698126
3d	-880.656474399	-880.655707513	-880.664624120	-880.664457265
3e	-919.971291898	-919.968972012	-919.978967831	-919.977524714
3f	-959.284718263	-959.283092592	-959.292076753	-959.291873437
3g	-919.971522432	-919.970387861	-919.979169246	-919.979055387
3h	-959.285225983	-959.283693496	-959.292508879	-959.292301476
3i	-998.596454880	-998.595138399	-998.604239590	-998.603651658

Absolute energies of the precursors in the selected conformations

Absolute energies of HH and HT unsaturated and saturated dimers

Unsaturated	Saturated	Absolute Energy	Absolute Energy	Absolute Energy	Absolute Energy
Dimer	Dimer	of HH Unsaturated	of HT Unsaturated	of HH Saturated	of HT Saturated
		Dimer (a.u.)	Dimer (a.u.)	Dimer (a.u.)	Dimer (a.u.)
4b	5b	-1604.15438264	-1604.14594029	-1606.61653396	-1606.60549081
4 c	5c	-1682.76433472	-1682.78664441	-1685.22705340	-1685.24077746
4d	5d	-1604.12946450	-1604.15983383	-1606.59401209	-1606.61375676
4e	5e	-1682.78001976	-1682.79530381	-1685.23800902	-1685.25186658
4f	5f	-1761.40880740	-1761.39708436	-1763.86072030	-1763.84755629
4 g	5g	-1682.75924146	-1682.78188747	-1685.21531345	-1685.23860424
4h	5h	-1761.39571506	-1761.39276448	-1763.85536192	-1763.85467780
4i	5i	-1840.03971085	-1840.03584304	-1842.49592968	-1842.49350722

Absolute and relative energies of the precursors of linear dimers in the selected conformations

Geometry	Absolute Energy	Relative Energy	Absolute Energy	Relative Energy
	of 6 (a.u.)	of 6 (kcal/mol)	of 9 (a.u.)	of 9 (kcal/mol)
i	-921.208638229	4.93	-921.209144878	4.56
ii	-921.207482127	5.66	-921.207458348	5.62
iii	-921.216498978	0.00	-921.216415416	0.00
iv	-921.215812176	0.43	-921.216085773	0.21

Absolute, relative energies and heat of formation of linear dimers 7 and 10

Compound	Absolute Energy	Relative Energy	Heat of Formation
	(a.u.)	(kcal/mol)	(kcal/mol)
7	-1763.84693322	0.00	0.15
10	-1763.84628580	0.41	0.45

The heats of formation of the dimers have been calculated with respect to the most stable conformation of their respective open precursor which is the *iii* conformation.

Stereo view of compound 5e in the model of R39 active site



Legend: C of **5e** are purple, others in yellow, H are in grey, O in red, N in blue



Molecule 5f docked as an acyl-enzyme complex in PBP2a cavity

Acyl-enzyme complexes between **5f** and PBP2a. a) **5f**, HH conformer b) **5f**, HT conformer.

Legend: Residues surrounding the alkyl chain of the macrocycle at the bottom of the active site are highlighted with carbon atoms coloured green, nitrogen blue and oxygen red. The carbon atoms of **5f** are coloured yellow and hydrogen white. The active serine 403 is behind the ligand and the covalent bond between the serine and the ligand is coloured black.

The flexibility of the macrocycle **5f** of both HH and HT isomers allows its docking in the active site as acyl-enzymes. The docking highlights the possibility for the whole macrocycle to completely fill the active cavity with the closed lactam cycle sandwiched between Tyr446 and Met641. The figures show the macrocycle lying between a series of residues, especially Glu602 and Gln613 at the entrance of the cavity, without making hydrogen bonds with these residues. The important catalytic residues of the conserved motifs (Ser403, Lys406, Ser462 and Lys597) lie behind the macrocycle and are not shown on the figures.

A modification of the geometry has been applied to dock this molecule into PBP2a. Starting from these two different geometries, a reoptimization at the B3LYP/6-31G(d) level leads to two other local minima with relative energies of 4.56 kcal and 5.16 kcal (Fig. below).



Dimers 4 and 5. Relative energies are given in kcal/mol







Experiments of temperature coefficients for amide protons

Temperature coefficients of the NH chemical shifts were recorded at 500 MHz in 1,1,2,2-tetrachloroethane- d_2 by heating at 10°C intervals from 298 to 358 K.



The temperature coefficients of the two amide protons of compound **11** are -3.0 and -2.6 ppb/ $^{\circ}$ C. The temperature coefficients of the two amide protons of compound **5c** are -2.6 and -3.9 ppb/ $^{\circ}$ C.

In non-competitive solvents, intramolecular hydrogen-bonded amide protons typically exhibit a relatively large temperature dependence of the chemical shift relative to free amide protons which exhibit a small temperature dependence of the chemical shift.⁵

We can observe two different evolutions for the NH of the compound 5c, it could be explained by the presence of one intramolecular hydrogen-bonded amide proton since the second amide proton is not involved in an intramolecular H bond.

⁵ T. K. Chakraborty, A. Ghosh, R. Nagaraj, A. R. Sankar, A. C. Kunwar, Tetrahedron 2001, **57**, 9169 and references therein.





















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ppm 180 170 70 30 20