Electronic Supplementary Information

The Discovery of Novel 10,11-Dihydro-5H-dibenz[b,f]azepine SIRT2 Inhibitors

Paolo Di Fruscia,^a Ka-Kei Ho,^b Sasiwan Laohasinnarong,^b Mattaka Khongkow,^b Sebastian H.B. Kroll,^a Suhail A. Islam,^c Michael J. E. Sternberg,^c Karin Schmidtkunz,^d Manfred Jung,^d Eric W.-F. Lam,^b and Matthew J. Fuchter^{a*}

^aDepartment of Chemistry, Imperial College London, London, United Kingdom

^bDepartment of Surgery and Cancer, Imperial College London, London, United Kingdom

^cDepartment of Bioinformatics, Imperial College London, London, United Kingdom

^dInstitute of Pharmaceutical Sciences, Albert-Ludwigs-Universität, Freiburg, Germany

*E-mail: m.fuchter@imperial.ac.uk

Table of Contents

A. Homology Modeling and Docking Studies	Page	S 1
B. Enzymatic Screening Assays	Page	S 6
C. Cellular Screens	Page	S8
D. Synthetic Schemes	Page	S9
E. Synthetic Procedures and Compound Characterization	Page	S11
F. ¹ H NMR Spectra of Tested Compounds Without Elemental Analysis	Page	S31
G. References	Page	S35

A. Homology Modeling and Docking Studies

The human SIRT1 amino acid sequence was retrieved from the UniProt database (UniProt ID: Q96EB6)¹ and a 3D comparative model of the human SIRT1 enzyme was predicted by the PHYRE server using a yeast Sir2 X-ray crystal structure (PDB entry code: 2HJH, chain A)² as template. 2HJH showed high sequence identity with the target protein (Fig. S1) and high Blast³ and SAS⁴ scores (Table S1).

SIRT1 234	RKKRKDINTIEDAVKLLQECKKIIVLTGAGVSVSCGIPDFRSRDGIYARLAVDFPDLPDP R + + TI+ ++ L +KI+VLTGAGVS S GIPDFRS +G Y++ + L DP	293
Sir2 235		292
SIRT1 294	QAMFDIEYFRKDPRPFFKFAKEIYPGQFQPSLCHKFIALSDKEGKLLRNYTQNIDTLEQV Q +F+ F DP F+ A + P + S H FI + +GKLLRNYTQNID LE	353
Sir2 293	•	352
SIRT1 354	AGIQRIIQCHGSFATASCLICKYKVDCEAVRGDIFNQVVPRCPRCPADAGI +++QCHGSFATA+C+ C + + E + I N +P CP C	401
Sir2 353		412
SIRT1 402	EPLAIMKPEIVFFGENLPEQFHRAMKYDKDEVDLLIVIGS	441
Sir2 413	++KP+I FFGE LP +FH++++ D E DLLI IG+ NKVGVAASQGSMSERPPYILNSYGVLKPDITFFGEALPNKFHKSIREDILECDLLICIGT	472
SIRT1 442		
Sir2 473	SLKV PV+ I + +P VPQ+LINR+P+ H FD+ LLG CD I + + G SLKVAPVSEIVNMVPSHVPQVLINRDPVKHAEFDLSLLGYCDDIAAMVAQKCG 525	

Fig. S1 Sequence alignment between target SIRT1 and template Sir2 catalytic domain.

PDB ID **BLAST SAS** Smith-%aa-Score Z-Score E-Value Waterman Length E-Value Identity Overlap Score 2HJH A 219 3e-57 764 43.8 276 405.4 8.4e-16 325

Table S1 BLAST and SAS scoring of the selected template structure

Dockings of inhibitor **8** were carried out using GOLD 4.0⁵ The proteins used in the present study, SIRT2 X-ray crystal structure (PDB ID: 1J8F, chain B) and SIRT1 homology model, were initialised and optimised with GOLD. No degree of flexibility was adopted (fully rigid proteins). The molecular structure of **8** was generated with ChemBioDraw Ultra 12.0⁶ and energy-minimized with ChemBio3D Ultra 12.0⁶ using the MMFF94 force field. *GoldScore* was chosen as primary scoring function and *ChemScore* as rescoring function. The search efficiency of the genetic algorithm was set at 200%, which represents the highest efficiency the software can employ.

The binding site was defined on the C_{α} of His363 for SIRT1 and C_{α} of His187 for SIRT2, with a radius of 20 Å. Water molecules within the binding site definition of the SIRT2 protein were retained and considered in the docking studies, using the settings "toggle" and "spin". For the ligand, the maximum degree of flexibility (fully flexible ligand) was adopted, and 10 docking runs were performed. The top-ranked poses were visually analysed with PyMOL.⁷ The stereochemical quality of the protein structures was evaluated with the PROCHECK⁸ tool: the Ramachandran plot for the SIRT1 homology model yielded 88.6% of residues in the "most favoured regions", 10.3% in "additional allowed regions", 1.1% in "generously allowed regions" and 0.0% in "disallowed regions", indicating a general high quality of the model (Fig. S2).

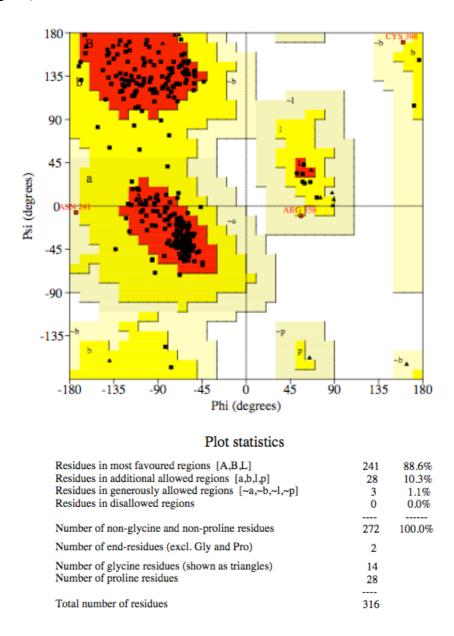


Fig. S2 Ramachandran plot of SIRT1 homology model.

The Ramachandran plot for the SIRT2 X-ray structure yielded 91.9% of residues in the "most favoured regions", 7.4% in "additional allowed regions", 0.4% in "generously allowed regions" and 0.4% in "disallowed regions". The 0.4% in "disallowed regions" represents Ser311, which is not located in the catalytic site (Fig. S3). Overall, it is expected that a homology model would have a lower quality Ramachandran plot than the template (X-ray crystal structure).

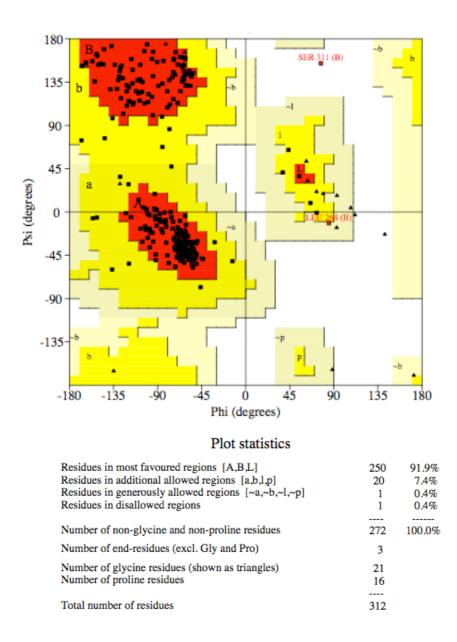


Fig. S3 Ramachandran plot of SIRT2 crystal structure (PDB ID: 1J8F, chain B).

PDBsum server⁹ was used to predict and analyse the secondary structure elements of SIRT1 (Fig. S4) and SIRT2 (Fig. S5) structures.

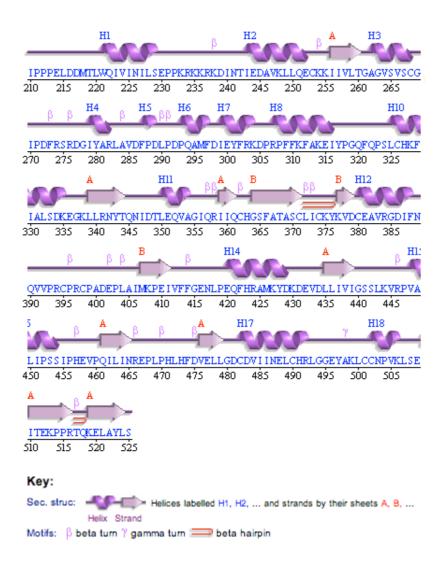


Fig. S4 Secondary structure elements of human SIRT1 homology model.

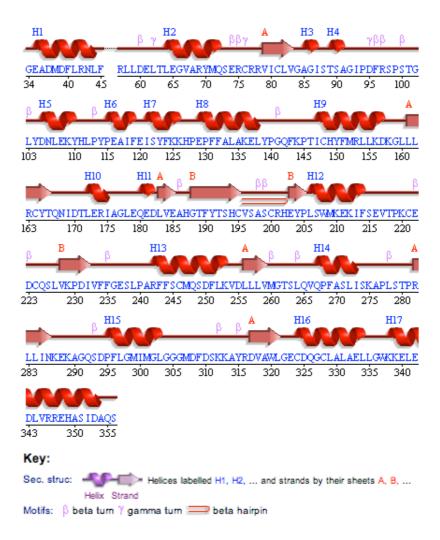


Fig. S5 Secondary structure elements of human SIRT2 crystal structure (PDB ID: 1J8F, chain B).

B. Enzymatic Screening Assays

All compounds were evaluated for their ability to inhibit recombinant sirtuins using a

homogeneous fluorescent deacetylase assay. Stock solutions of inhibitors were prepared in

DMSO. The assay was carried out in 96-well plates: $60~\mu L$ reaction volume contained the

fluorescent histone deacetylase substrate ZMAL (10.5 μM), NAD⁺ (500 μM), and SIRT2 or

SIRT1. Total substrate conversion was driven to about 15% - 30% to assure initial state

conditions. After 4 h incubation at 37 °C, the deacetylation reaction was stopped, and the

metabolite formed (ZML, the deacetylated form of ZMAL) was developed using a tryptic

digest for 20 min to form a different fluorophore. Finally, fluorescence was measured in a

plate reader (BMG Polarstar) with excitation at λ =390 nm and emission at λ =460 nm. The

amount of remaining substrate in the positive control with inhibitor versus negative control

without inhibitor (only DMSO) was employed to calculate inhibition. All IC₅₀ determinations

were carried out in triplicate (pre-test determinations in duplicate). IC₅₀ data were analyzed

using GraphPad Prism software.

Materials, buffers and enzymes:

Fluorescent histone deacetylase substrate ZMAL, 12.6 mM in DMSO; stock solution of AMC

(7-Amino-4-methylcoumarin; Fluka), 12.6 mM in DMSO; stock solution of nicotinamide,

120 mM in DMSO; all stored at -80 °C.

Sirtuin buffer pH 8.0:

25 mM Tris-HCl

137 mM NaCl

2.7 mM KCl

1 mM MgCl₂

Trypsin buffer pH 8.0:

50 mM Tris-HCl

100 mM NaCl

Recombinant hSIRT1 or recombinant hSIRT2 (in-house purification). Stock solution of

trypsin from bovine pancreas (10000 BAEE units/mg) 6 mg/mL in trypsin buffer. Inhibitor

solution in DMSO as 10 mM stock. Microplate reader Polarstar galaxy (BMG

Labtechnologies, Germany) with an excitation filter of 390 nm an emission filter of 460 nm.

S6

Black 96-well micro plates (Greiner or PerkinElmer).

SIRT2

Plasmid pEV1440 (5.5 kb) was provided from the Lab of Prof. Dr. E. Verdin, Gladstone Institute, San Francisco, USA. Here, full length human SIRT2 cDNA was cloned into pHEX-2T with BamHI/EcoRI (original vector pGEX-2T from Phamacia in which the GST-encoding sequence was replaced by 6 x His). The resulting protein was an N-tagged 6 x His-Sirt2. We transformed pEV1440 into E. coli DH5α (Invitrogen) for plasmid purification and the purified plasmid was transformed into E. coli BL21 (DE3) for protein purification. First the protein was purified with affinity chromatography (Ni-NTA Superflow, Qiagen). In a second step the eluted protein was loaded on a PD10 desalting column (GE Healthcare) for buffer exchange. The activity was determined and the active fractions were pooled. The enzyme was analyzed with SDS page. The IC50 with NA was determined and the activity with or without NAD+ was tested.

SIRT1

Plasmid pTe34 (6961 bp) was provided from Prof. Dr. A. Salminen (University Kuopio, Finland). Here, a human SIRT1 ORF fragment was inserted into pGEX2T (Amersham). The resulting protein was an N-tagged GST-SIRT1 fusion protein. Transformations for plasmid or protein purification were performed like described for SIRT2. The protein was purified by using Glutathione Sepharose 4B Beads (Amersham Biosciences) for affinity chromatography. Further purification and and analyses were performed like described before.

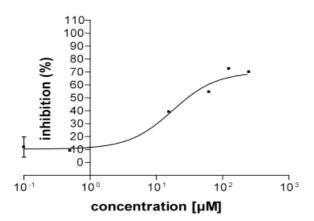


Fig. S6 Compound 8 concentration-SIRT2 inhibition curve.

Compound 8 (SIRT2 Fluorescence data)											
		fluorescence	fluorescenze	fluorescence	fluorescence	conversion [%] 1	conversion [%] 1	inhibition 1	inhibition 2		
		data1	data	data1-	data2-	related to	related to	related to	related to	BMG Polarstar	
				blank	blank	100% conversion	100% conversion	conversion	conversion	Ex: 390	Em: 460
blank		1253	1253	0	0					Tryp. Inkub [min]:	20
100% conversion		57982	57982	56728	56728					Inkub.time [min]:	240
conversion	DMSO	19114	19114	17861	17861	31.5	31.5			Inkub. Temp [°C]:	37
8	250 µM	6701	6478	5448	5225	9.6	9.2	69.5	70.7	Gain:	2151
8	125µM	6321	5976	5068	4723	8.9	8.3	71.6	73.6		
8	62,5 µM	9558	9199	8305	7946	14.6	14.0	53.5	55.5		
8	31,25 µM	14111	15632	12858	14379	22.7	25.3	28.0			
8	15,6 µM	12089	12143	10836	10890	19.1	19.2	39.3	39.0		
8	0,5 µM	17492	17492	16239	16239	28.6	28.6	9.1	9.1		
8	0,1µM	15582	18378	14329	17125	25.3	30.2	19.8	4.1		

Fig. S7 Fluorescence and percentage data, of SIRT2 inhibition, related to different concentrations of 8.

C. Cellular Screens

SIRT inhibitors, EX-527, salermide and sirtinol were synthesized by us, while AK-7 was acquired from AKos GmbH. The inhibitors were dissolved in DMSO at a concentration of 20 mM for stock solutions and then appropriately diluted.

- Cell viability analysis

MCF-7 cells were incubated with growth media supplemented with DMSO or the indicated inhibitor at 30 μ M for 0, 24, 48 and 72 hours. At each time point, cells were washed three times with PBS and once with EDTA, followed by trypsinisation. Cells were then suspended with PBS and cell numbers were determined using a haemocytometer. Representative data from three independent experiments are shown.

- Cell cycle analysis

Cell cycle analysis was performed by propidium iodide (PI) staining followed by FACS analysis. Both floating and adherent cells were harvested by trypsinisation, washed with PBS, fixed and permeablised with 90% cold ethanol. Cells were then stained with propidium iodide (0.2 mg/mL in PBS) containing 0.01 mg/mL RNase A. The stained cells were acquired using a Bectron Dickinson FACS flow cytometor and analysed for the cell cycle distribution by FacsDiva software (Becton Dickinson).

D. Synthetic Schemes

 $\textbf{Reagents and conditions:} \ (a) \ ClSO_2NCO, \ DCM, \ 0 \ ^{\circ}C, \ 1 \ h. \ (b) \ AgNO_3, \ BzCl, \ MeCN, \ rt, \ 16 \ h.$

 $\textbf{Reagents and conditions:} (a) \ H_2O_2, \ KOH, \ EtOH, \ 60\ ^{\circ}C, \ 2\ h. \ (b) \ EtCOO'Bu, \ H_2SO_4, \ 42\ ^{\circ}C, \ 1\ h. \ (c) \ NCS, \ SiO_2, \ DCM, \ rt, \ 16\ h. \ (d) \ NCS, \ SiO_2, \ DCM, \ rt, \ 16\ h. \ (e) \ NCS, \ SiO_3, \ DCM, \ rt, \ 16\ h. \ (e) \ NCS, \ SiO_4, \ NCS, \ SiO_5, \ DCM, \ rt, \ 16\ h. \ (e) \ NCS, \ SiO_5, \ DCM, \ rt, \ 16\ h. \ (e) \ NCS, \ SiO_5, \ NCS, \ SiO_5, \ DCM, \ rt, \ 16\ h. \ (e) \ NCS, \ SiO_5, \ NCS, \ SiO_5, \ NCS, \ SiO_5, \ NCS, \ NCS$

Scheme S3

 $\textbf{Reagents and conditions:} \ (a) \ AgNO_3, \ BzCl, \ MeCN, \ rt, \ 16 \ h. \ (b) \ H_2O_2, \ KOH, \ EtOH, \ 60 \ ^{\circ}C, \ 2 \ h.$

Scheme S4

 $\textbf{Reagents and conditions:} \ (a) \ i) \ NaHMDS, \ THF, \ 0 \ ^{\circ}C \ 30 \ min; \ ii) \ MeI, \ rt, \ 16 \ h. \ (b) \ H_2O_2, \ KOH, \ EtOH, \ 60 \ ^{\circ}C, \ 2 \ h. \ (b) \ H_2O_3, \ KOH, \ EtOH, \ 60 \ ^{\circ}C, \ 2 \ h. \ (c) \ H_2O_4, \ KOH, \ EtOH, \ 60 \ ^{\circ}C, \ 2 \ h. \ (c) \ H_2O_5, \ KOH, \ EtOH, \ 60 \ ^{\circ}C, \ 2 \ h. \ (c) \ H_2O_5, \ KOH, \ EtOH, \ 60 \ ^{\circ}C, \ 2 \ h. \ (c) \ H_2O_5, \ KOH, \ EtOH, \ 60 \ ^{\circ}C, \ 2 \ h. \ (c) \ H_2O_5, \ KOH, \ EtOH, \ 60 \ ^{\circ}C, \ 2 \ h. \ (c) \ H_2O_5, \ KOH, \ EtOH, \ 60 \ ^{\circ}C, \ 2 \ h. \ (c) \ H_2O_5, \ KOH, \ EtOH, \ 60 \ ^{\circ}C, \ 2 \ h. \ (c) \ H_2O_5, \ KOH, \ EtOH, \ 60 \ ^{\circ}C, \ 2 \ h. \ (c) \ H_2O_5, \ KOH, \ EtOH, \ 60 \ ^{\circ}C, \ 2 \ h. \ (c) \ H_2O_5, \ KOH, \ EtOH, \ 60 \ ^{\circ}C, \ 2 \ h. \ (c) \ H_2O_5, \ KOH, \ EtOH, \ 60 \ ^{\circ}C, \ 2 \ h. \ (c) \ H_2O_5, \ KOH, \ EtOH, \ 60 \ ^{\circ}C, \ 2 \ h. \ (c) \ H_2O_5, \ H$

Scheme S5

Reagents and conditions: (a) H₂O₂, KOH, EtOH, 60 °C, 1 h. (b) DIBAL-H, DCM, 0 °C, 30 min. (c) NaBH₄, MeOH, rt, 1 h.

 $\label{eq:conditions: Academic Conditions: Academic Conditions: Conditions:$

E. Synthetic Procedures and Compound Characterization

Melting points were obtained on a Reichert-Thermovar melting point apparatus and are uncorrected. Infrared spectra were recorded on a Perkin Elmer (Spectrum Express Version 1.03.00) spectrometer with automated background substraction. Reported absorptions are strong or medium strength unless stated otherwise and given in wavenumbers (cm⁻¹). ¹H NMR spectra were recorded on a Bruker DRX-400 spectrometer operating at 400 MHz. ¹³C NMR spectra were recorded on a Bruker DRX-400 spectrometer operating at 100 MHz. Chemical shifts (δ) are quoted in parts per million (ppm) and are referenced to a residual solvent peak. CDCl₃ (δ_H : 7.26, δ_C : 77.0), DMSO- d_6 (δ_H : 2.50, δ_C : 39.5). Low and high resolution mass spectrometry (EI, ESI) were recorded using a Micromass Platform II and Micromass AutoSpec-Q spectrometer. Elemental analyses were determined by the University of North London Analytical Service. All manipulations of air or moisture sensitive materials were carried out in oven or flame dried glassware under an inert atmosphere of nitrogen or argon. Syringes, which were used to transfer reagents and solvents, were purged with nitrogen prior to use. Reaction solvents were distilled from CaH₂ (dichloromethane, triethylamine), Na/Ph₂CO (tetrahydrofuran, diethyl ether) or obtained as dry or anhydrous from Sigma-Aldrich Chemical Company (N,N-dimethylformamide, acetonitrile) or BDH (ethanol). All reagents were obtained from commercial suppliers and used as obtained if purity was $\geq 98\%$. All flash-column chromatography was carried out on BDH silica gel 60, particle size 0.040 -0.063 mm unless otherwise stated. Thin layer chromatography (TLC) was performed on precoated aluminium backed or glass backed plates (Merck Kieselgel 60 F₂₅₄), and visualised with ultraviolet light (254 nm) or potassium permanganate (KMnO₄), vanillin or phosphomolybdic acid (PMA) stains as deemed appropriate.

2-Bromo-10,11-dihydro-5H-dibenz[b,f]azepine (5)

To a solution of **4** (5.00 g, 25.64 mmol) in DCM (2 L) were added NBS (4.60 g, 25.64 mmol) and SiO₂ (50.00 g). The resulting mixture was stirred at room temperature for 1 h. The suspension was filtered through a pad of celite and the filtrate washed with brine, dried over MgSO₄, filtered and evaporated under reduced pressure to afford the title compound (6.30 g) as a blue solid, which was used in the next step without purification: MS (ESI) m/z 274 (M+H)⁺; HRMS (ESI) m/z calc for C₁₄H₁₃BrN 274.0231, found: 274.0224.

10,11-Dihydro-5H-dibenz[b,f]azepine-2-carbonitrile (6)

Degassed DMF (30 mL) was added to a mixture of **5** (6.20 g, 22.71 mmol), Zn(CN)₂ (2.70 g, 22.71 mmol) and Pd(PPh₃)₄ (5.20 g, 4.54 mmol). The resulting suspension was stirred at 90 °C for 2 h. Sat. Na₂CO₃ (aq.) (200 mL) was added and the crude product extracted with Et₂O (x 2). The combined organics were washed with brine, dried over MgSO₄, filtered and the solvent evaporated under reduced pressure. The resultant residue was purified by silica gel flash-column chromatography with hexanes:DCM (4:6) as eluent to afford **6** (2.00 g, 36% over two steps) as a white solid: Elem. anal. calc for C₁₅H₁₂N₂: C, 81.79; H, 5.49; N, 12.72; found: C, 81.80; H, 5.41; N, 12.82; IR (neat) 3340, 2216 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 2.96 (*app*-s, 4H), 6.78 (*app*-t, J = 7.3 Hz, 1H), 7.01 - 7.13 (m, 4H), 7.41 - 7.46 (m, 2H), 8.99 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 34.8, 35.3, 99.0, 118.6, 119.3, 120.5, 120.6, 127.3, 127.8, 129.8, 130.7, 131.0, 135.1, 141.6, 147.3; MS (ESI) m/z 221 (M+H)⁺; HRMS (ESI) m/z calc for C₁₅H₁₃N₂ 221.1079, found: 221.1067.

8-Nitro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carbonitrile (7)

To a solution of **6** (480 mg, 2.18 mmol) in MeCN (10 mL) were added AgNO₃ (405 mg, 2.40 mmol) and BzCl (277 μ L, 2.40 mmol) at 0 °C. The resulting mixture was stirred at room temperature for 16 h. MeCN was evaporated and the resultant residue dissolved in EtOAc and filtered under reduced pressure. The filtrate was washed with sat. Na₂CO₃ (aq.), brine, dried over MgSO₄, filtered and the solvent evaporated under reduced pressure. The crude material was purified by silica gel flash-column chromatography with hexanes:DCM (1:9) as eluent to afford **7** (200 mg, 35%) as an orange solid: Elem. anal. calc for C₁₅H₁₁N₃O₂: C, 67.92; H, 4.18; N, 15.84; found: C, 67.81; H, 4.09; N, 15.73; IR (neat) 3339, 2220, 1495, 1289 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 2.96 - 3.14 (m, 4H), 7.20 (*app*-t, J = 8.5 Hz, 2H), 7.52 - 7.62 (m, 2H), 7.97 - 8.06 (m, 2H), 9.80 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 34.4, 34.7,

101.9, 119.2, 119.9, 120.1, 123.5, 126.9, 128.7, 130.3, 131.3, 134.9, 139.3, 145.5, 148.2; MS (EI) m/z 265 M^{*+} ; HRMS (EI) m/z calc for $C_{15}H_{11}N_3O_2$ 265.0851, found: 265.0848.

8-Nitro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (8)

$$H_2N$$
 N
 N
 N
 N
 N
 N

To a solution of 7 (100 mg, 0.38 mmol) in EtOH (5 mL) were added KOH (192 mg, 3.42 mmol) and 35% wt. H_2O_2 (aq.) (0.8 mL, 9.50 mmol). The resultant mixture was stirred at 60 °C for 1 h. EtOH was evaporated and the residue partitioned between EtOAc and water. The organic layer was washed with brine, dried over MgSO₄, filtered and evaporated under reduced pressure to provide a residue which was purified by silica gel flash-column chromatography with DCM:EtOAc (3:7) as eluent to give the title compound (100 mg, 95%) as an orange crystalline solid: mp 248 °C (from EtOAc:MeOH 2:1); Elem. anal. calc for $C_{15}H_{13}N_3O_3$: C, 63.60; H, 4.63; N, 14.83; found: C, 63.66; H, 4.60; N, 14.83; IR (neat) 3447, 3339, 3296, 1643, 1496, 1310 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 3.00 - 3.14 (m, 4H), 7.09 - 7.19 (m, 3H), 7.64 - 7.69 (m, 2H), 7.71 - 7.80 (br s, 1H), 7.96 - 8.04 (m, 2H), 9.62 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 34.5, 34.7, 118.1, 118.7, 123.1, 126.0, 126.5, 126.6, 127.4, 128.7, 130.1, 138.0, 143.2, 148.5, 167.3; MS (ESI) m/z 284 (M+H)⁺; HRMS (ESI) m/z calc for $C_{15}H_{14}N_3O_3$ 284.1035, found: 284.1023.

8-Amino-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (9)

$$H_2N$$
 N
 N
 N
 N

To a solution of **8** (60 mg, 0.21 mmol) in MeOH (5 mL) were added NH₄HCO₂ (134 mg, 2.10 mmol) and Pd/C (12 mg). The resulting mixture was stirred at 50 °C for 1 h. The mixture was filtered through a pad of celite and the filtrate evaporated. The residue was partitioned between EtOAc and water. The organic phase was washed with brine, dried over MgSO₄, filtered and the solvent evaporated under reduced pressure. The crude material was purified by silica gel flash-column chromatography with EtOAc as eluent to afford **9** (20 mg, 37%) as

a brown solid: IR (neat) 3321, 3197, 1641 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 2.79 - 2.99 (m, 4H), 4.56 (s, 2H), 6.30 - 6.38 (m, 2H), 6.72 (d, J = 8.4 Hz, 1H), 6.82 (d, J = 8.4 Hz, 1H), 6.87 (s, 1H), 7.45 - 7.53 (m, 2H), 7.55 (s, 1H), 8.16 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 34.9, 35.9, 113.3, 115.8, 116.7, 120.0, 122.2, 124.6, 126.7, 131.2, 131.3, 132.6, 142.4, 146.8, 168.2; MS (ESI) m/z 254 (M+H)⁺; HRMS (ESI) m/z calc for C₁₅H₁₆N₃O 254.1293, found: 254.1299.

2,8-Dibromo-10,11-dihydro-5H-dibenz[b,f]azepine (10)

Following the procedure described for the preparation of 2-bromo-10,11-dihydro-5H-dibenz[b,f]azepine (**5**), compound **4** (1.00 g, 5.13 mmol) was treated with NBS (1.87 g, 10.52 mmol) and SiO₂ (20.00 g) in DCM (1 L) to give, after purification by silica gel flash-column chromatography with hexanes:DCM (7:3) as eluent and crystallisation from CHCl₃, the title compound (1.60 g, 88%) as white crystals: ¹H NMR (400 MHz, DMSO-d₆) δ 2.92 (s, 4H), 6.91 (d, J = 8.0 Hz, 2H), 7.17 - 7.21 (m, 4H), 8.58 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 34.6, 110.2, 120.5, 129.7, 130.7, 132.9, 142.3; MS (ESI) m/z 352 (M+H)⁺; HRMS (ESI) m/z calc for C₁₄H₁₂Br₂N 351.9336, found: 351.9330.

10,11-Dihydro-5H-dibenz[b,f]azepine-2,8-dicarbonitrile (11)

Following the procedure described for the preparation of 10,11-dihydro-5H-dibenz[b,f]azepine-2-carbonitrile (6), compound 10 (1.50 g, 4.25 mmol) was treated with Zn(CN)₂ (994 mg, 8.50 mmol) and Pd(PPh₃)₄ (982 mg, 0.85 mmol) in degassed DMF (5 mL) to give, after purification by silica gel flash-column chromatography with hexanes:DCM (1:9) as eluent, compound 11 (0.80 g, 77%) as a white solid: Elem. anal. calc for C₁₆H₁₁N₃: C, 78.35; H, 4.52; N, 17.13; found: C, 78.27; H, 4.47; N, 17.10; IR (neat) 3329, 2217 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 3.00 (s, 4H), 7.16 (d, J = 8.0 Hz, 2H), 7.51 - 7.57 (m, 4H), 9.53 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 34.5, 101.1, 119.7, 120.1, 129.6, 131.3,

134.9, 146.0; MS (ESI) m/z 245 M^+ ; HRMS (ESI) m/z calc for $C_{16}H_{11}N_3$ 245.0953, found: 245.0945.

10,11-Dihydro-5H-dibenz[b,f]azepine-2,8-dicarboxamide (12)

Following the procedure described for the preparation of 8-nitro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (**8**), compound **11** (150 mg, 0.61 mmol) was treated with KOH (307 mg, 5.49 mmol) and 35% wt. H_2O_2 (aq.) (1.3 mL, 15.25 mol) to give, after purification by silica gel flash-column chromatography with EtOAc:MeOH (9:1) as eluent, the desired compound **12** (60 mg, 35%) as a white solid: IR (neat) 3439, 3458, 3316, 1643, 1598 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 2.98 - 3.02 (s, 4H), 6.96 - 7.10 (m, 4H), 7.56 - 7.63 (m, 4H), 7.69 (s, 2H), 8.99 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 35.5, 118.1, 124.7, 126.9, 127.6, 130.8, 145.1, 168.0; MS (ESI) m/z 282 (M+H)⁺; HRMS (ESI) m/z calc for $C_{16}H_{16}N_3O_2$ 282.1243, found: 282.1237.

10,11-Dihydro-5H-dibenz[b,f]azepine-4-carbaldehyde (13)

To a solution of **4** (6.0 g, 30.77 mmol) in Et₂O (200 mL) was added *n*-BuLi (38 mL, 92.31 mmol) at -78 °C and the resulting mixture was stirred at room temperature for four days. DMF (3.6 mL, 46.16 mmol) was added at -78 °C and the reaction was allowed to stir at room temperature for additional 24 h. 0.5N HCl (aq.) (150 mL) was added. The organic phase was separated from the aqueous solution, washed with brine, dried over MgSO₄, filtered and the solvent evaporated under reduced pressure. The resultant residue was purified by silica gel flash-column chromatography with hexanes:DCM (7:3) as eluent to yield **13** (5.9 g, 85%) as an orange oil: Elem. anal. calc for $C_{15}H_{13}NO$: C, 80.69; H, 5.87; N, 6.27; found: C, 80.76; H, 5.84; N, 6.14; IR (neat) 1655 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 3.06 - 3.12 (m, 4H), 6.78 (*app*-t, J = 7.5 Hz, 1H), 6.87 (*app*-t, J = 7.2 Hz, 1H), 7.02 (d, J = 8.0 Hz, 1H), 7.07 (d, J = 7.6

Hz, 1H), 7.16 (*app*-t, J = 8.2 Hz, 1H), 7.22 (d, J = 7.3 Hz, 1H), 7.42 (d, J = 7.8 Hz, 1H), 9.87 (s, 1H), 11.27 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 34.9, 36.0, 117.0, 120.0, 120.4, 121.0, 127.1, 129.2, 130.1, 130.8, 135.8, 136.9, 140.7, 145.8, 194.6; MS (ESI) m/z 224 (M+H)⁺; HRMS (ESI) m/z calc for C₁₅H₁₄NO 224.1075, found: 224.1073.

(10,11-Dihydro-5H-dibenz[b,f]azepin-4-yl)methanol (14)

To a solution of **13** (50 mg, 0.22 mmol) in MeOH (2 mL) was added NaBH₄ (8 mg, 0.22 mmol) and the resulting mixture was stirred at room temperature for 2 h. H₂O was added and the solution concentrated. The crude product was extracted with EtOAc. The organic phase was washed with brine, dried over MgSO₄, filtered and the solvent evaporated under reduced pressure. The residue was purified by silica gel flash-column chromatography with hexanes:EtOAc (9:1) as eluent, to afford the title compound (45 mg, 90%) as a white solid: Elem. anal. calc for C₁₅H₁₅NO: C, 79.97; H, 6.71; N, 6.22; found: C, 80.07; H, 6.67; N, 6.15; IR (neat) 3381, 3216 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 1.76 (s, 1H), 3.00 - 3.15 (m, 4H), 4.78 (s, 2H), 6.74 (*app*-t, J = 7.1 Hz, 2H), 6.83 (d, J = 7.8 Hz 1H), 6.96 - 7.22 (m, 4H), 7.63 - 7.77 (br s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 35.0, 35.4, 65.5, 118.7, 118.9, 119.1, 126.5, 126.8, 127.5, 127.6, 130.8, 130.9, 131.5, 142.4, 142.8; MS (ESI) m/z 226 (M+H)⁺; HRMS (ESI) m/z calc for C₁₅H₁₆NO 226.1232, found: 226.1230.

10,11-Dihydro-5H-dibenz[b,f]azepine-4-carboxylic acid (15)

To a solution of **13** (2.0 g, 8.97 mmol) in *t*-butanol (48 mL) and H_2O (12 mL) were added NaH_2PO_4 (9.6 g, 71.76 mmol), 2-methyl-2-butene (9.4 mL, 89.70 mmol) and $NaClO_2$ (3.2 g, 35.88 mmol). The mixture was stirred at room temperature for 16 h. *t*-Butanol was evaporated and the resulting residue partitioned between Et_2O and 2.0M NaOH (aq.). The aqueous phase

was washed with Et₂O (x 2), acidified with conc. HCl to acidic pH and filtered. The filter cake was washed with cold water and n-hexane, and purified by silica gel flash-column chromatography with DCM:MeOH (97:3) as eluent to give **15** (1.5 g, 71 %) as a yellow solid: Elem. anal. calc for C₁₅H₁₃NO₂: C, 75.30; H, 5.48; N, 5.85; found: C, 75.25; H, 5.40; N, 5.86; IR (neat) 2947, 1661 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 3.02 (app-s, 4H), 6.72 - 6.82 (m, 2H), 6.85 (d, J = 7.7 Hz, 1H), 7.05 - 7.15 (m, 2H), 7.29 (d, J = 7.4 Hz, 1H), 7.83 (d, J = 8.0 Hz, 1H), 10.98 (s, 1H), 13.15 - 13.32 (br s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 34.9, 35.5, 113.5, 117.9, 119.7, 120.3, 127.4, 129.3, 130.5, 130.6, 130.9, 136.0, 141.5, 146.3, 171.3; MS (ESI) m/z 238 (M-H)⁻; HRMS (ESI) m/z calc for C₁₅H₁₂NO₂ 238.0868, found: 238.0872.

10,11-Dihydro-5H-dibenz[b,f]azepine-4-carboxamide (16)

A solution of **15** (550 mg, 2.30 mmol), 0.5M NH₃ sol. in 1,4-dioxane (20 mL), HOBt (373 mg, 2.76 mmol) and EDC•HCl (527 mg, 2.76 mmol) was stirred at room temperature for 16 h. The solvent was evaporated and the residue partitioned between EtOAc and sat. Na₂CO₃ (aq.). The organic layer was washed with brine, dried over MgSO₄, filtered and the solvent evaporated under reduced pressure. The resultant residue was purified by silica gel flash-column chromatography with DCM:EtOAc (95:5) as eluent to yield **16** (400 mg, 82 %) as a yellow solid: Elem. anal. calc for C₁₅H₁₄N₂O: C, 75.61; H, 5.92; N, 11.76; found: C, 75.55; H, 5.86; N, 11.70; IR (neat) 3389, 3361, 3208, 1637 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 2.99 (*app*-s, 4H), 6.69 - 6.78 (m, 3H), 7.02 - 7.11 (m, 2H), 7.19 (d, J = 7.1 Hz, 1H), 7.50 - 7.62 (m, 2H), 8.06 - 8.16 (br s, 1H), 11.21 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 35.1, 35.2, 117.8, 118.0, 119.3, 119.4, 127.4, 127.8, 128.4, 131.0, 131.4, 133.8, 142.1, 144.7, 172.6; MS (ESI) m/z 239 (M+H)⁺; HRMS (ESI) m/z calc for C₁₅H₁₅N₂O 239.1184, found: 239.1183.

1-(10,11-Dihydro-5H-dibenz[b,f]azepin-5-yl)-2,2,2-trifluoroethanone (17)

To a solution of **4** (3.00 g, 15.38 mmol) in DCM (30 mL) were added at 0 °C TFAA (4.3 mL, 30.77 mmol), DIPEA (6.7 mL, 38.46 mmol) and DMAP (cat.). The mixture was stirred at room temperature for 16 h. 1N HCl (aq.) (50 mL) was added. The organic phase was separated from the acidic aqueous solution, washed with brine, dried over MgSO₄, filtered and the solvent evaporated under reduced pressure. The resultant residue was purified by silica gel flash-column chromatography with hexanes:EtOAc (9:1) as eluent to yield **17** (3.28 g, 73%) as a yellow solid: Elem. anal. calc for $C_{16}H_{12}F_3NO$: C, 65.98; H, 4.15; N, 4.81; found: C, 65.94; H, 4.10; N, 4.76; IR (neat) 1695 cm⁻¹; 1H NMR (400 MHz, CDCl₃) δ 2.82 - 2.99 (m, 2H), 3.33 - 3.58 (m, 2H), 7.13 - 7.46 (m, 8H); ^{13}C NMR (100 MHz, CDCl₃) δ 29.7, 30.8, 116.5 (q), 127.0, 127.1, 127.3, 127.6, 128.5, 129.5, 129.7, 131.4, 134.2, 137.9, 138.8, 139.4, 156.7 (q); MS (ESI) m/z 292 (M+H)⁺; HRMS (ESI) m/z calc for $C_{16}H_{13}F_3NO$ 292.0949, found: 292.0942.

1-(3-(Chlorosulphonyl)-10,11-dihydro-5H-dibenz[b,f] azepin-5-yl)-2,2,2-trifluoroethanone~(18)

17 (150 mg, 0.52 mmol) was added to CISO₃H (2 mL) at 0 °C and the resultant mixture was allowed to stir at 0 °C for 3 h. The solution was poured dropwise onto crushed ice and the product extracted with EtOAc. The organic phase was washed with brine, dried over MgSO₄, filtered and the solvent evaporated under reduced pressure. The resultant residue was purified by silica gel flash-column chromatography with hexanes:EtOAc (7:3) as eluent to yield the title compound (130 mg, 65%) as a white solid: ¹H NMR (400 MHz, CDCl₃) δ 2.85 - 3.09 (m, 2H), 3.38 - 3.62 (m, 2H), 7.17 - 7.61 (m, 5H), 7.82 - 8.12 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 29.7, 31.5, 116.5 (q), 127.0, 127.1, 127.3, 127.6, 128.5, 129.5, 129.7, 131.4, 134.2, 137.9, 139.2, 143.0, 156.7 (q).

N-tert-Butyl-5-trifluoroacetyl-10,11-dihydro-5H-dibenz[b,f]azepine-3-sulphonamide (19)

To a solution of **18** (80 mg, 0.21 mmol) in DCM (2 mL) was added t-BuNH₂ (88 μ L, 0.84 mmol). The mixture was stirred at room temperature for 16 h. The solution was washed with brine, dried over MgSO₄, filtered and the solvent evaporated under reduced pressure to afford **19** (82 mg, 93%) as a white solid, which was used in the next step without purification.

N-tert-Butyl-10,11-dihydro-5H-dibenz[b,f]azepine-3-sulphonamide (20)

To a solution of **19** (60 mg, 0.14 mmol) in MeOH:THF:H₂O (2:1:1 mL) was added K₂CO₃ (580 mg, 4.20 mmol). The resultant mixture was stirred at room temperature for 16 h. The solvent was concentrated under reduced pressure and the resulting residue partitioned between EtOAc and H₂O. The organic phase was washed with brine, dried over MgSO₄, filtered and the solvent evaporated under reduced pressure. The crude material was purified by silica gel flash-column chromatography with DCM:EtOAc (97:3) as eluent to afford **20** (43 mg, 92%) as a white solid: Elem. anal. calc for C₁₈H₂₂N₂O₂S: C, 65.42; H, 6.71; N, 8.48; found: C, 65.58; H, 6.60; N, 8.44; IR (neat) 3366, 3261, 1299, 1151 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 1.13 (s, 9H), 2.91 - 3.04 (m, 4H), 6.70 (*app*-t, J = 7.3 Hz, 1H), 6.97 - 7.09 (m, 4H), 7.17 (d, J = 8.0 Hz, 1H), 7.36 (s, 1H), 7.49 (d, J = 1.8 Hz, 1H), 8.68 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 29.8, 34.4, 34.8, 53.1, 115.6, 115.7, 118.2, 119.0, 126.7, 128.1, 130.3, 130.8, 131.1, 142.2, 142.5, 142.9; MS (ESI) m/z 329 (M+H)⁻; HRMS (ESI) m/z calc for C₁₈H₂₁N₂O₂S 329.1329, found: 329.1333.

1-(3-Nitro-10,11-dihydro-5H-dibenz[b,f]azepin-5-yl)-2,2,2-trifluoroethanone (21)

To a solution of 17 (200 mg, 0.69 mmol) in Ac₂O (3 mL) was added at 0 °C HNO₃ (1 mL). The resulting mixture was stirred at room temperature for 16 h, after which time the reaction was found to be complete (by TLC). The solution was poured dropwise onto crushed ice and the product extracted with EtOAc. The organic layer was washed with sat. NaHCO₃ (aq.), brine, dried over MgSO₄, filtered and the solvent evaporated under reduced pressure to give a crude product (250 mg) which was used in the next step without purification.

3-Nitro-10,11-dihydro-5H-dibenz[b,f]azepine (22)

Following the procedure described for the preparation of *N-tert*-butyl-10,11-dihydro-5H-dibenz[b,f]azepine-3-sulphonamide (**20**), compound **21** (200 mg, crude material) was treated with K_2CO_3 (1.64 g) in MeOH:THF: H_2O (2:1:1 mL) to give, after purification by silica gel flash-column chromatography with hexanes:DCM (6:4) as eluent, the title compound (60 mg, 45% over two steps) as an orange solid: Elem. anal. calc for $C_{14}H_{12}N_2O_2$: C, 69.99; H, 5.03; N, 11.66; found: C, 69.89; H, 4.86; N, 11.54; IR (neat) 3366, 1489, 1318 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 2.93 - 3.11 (m, 4H), 6.75 (*app*-t, J = 7.2 Hz, 1H), 6.95 - 7.14 (m, 3H), 7.26 (d, J = 8.5 Hz, 1H), 7.46 (d, J = 7.2 Hz, 1H), 7.90 (s, 1H), 8.88 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 34.4, 35.5, 112.5, 112.5, 118.8, 120.1, 127.4, 129.0, 130.7, 132.2, 135.1, 142.2, 144.3, 147.0; MS (ESI) m/z 241 (M+H)⁺; HRMS (ESI) m/z calc for $C_{14}H_{13}N_2O_2$ 241.0977, found: 241.0979.

10,11-Dihydro-5H-dibenz[b,f]azepine-2-carboxamide (23)

Following the procedure described for the preparation of 8-nitro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (**8**), compound **6** (100 mg, 0.45 mmol) was treated with KOH (0.22 g, 3.63 mmol) and 35% wt. H_2O_2 (aq.) (1.1 mL, 11.35 mol) in EtOH (2.5 mL) to give, after purification by silica gel flash-column chromatography with DCM:EtOAc (4:6) as eluent, the desired compound (104 mg, 96%) as a white solid: Elem. anal. calc for $C_{15}H_{14}N_2O$: C, 75.61; H, 5.92; N, 11.76; found: C, 75.51; H, 5.88; N, 11.64; IR (neat) 3359, 3153, 1638 cm⁻¹; ¹H NMR (400MHz, DMSO-d₆) δ 2.98 (*app*-s, 4H), 6.72 (*app*-t, J = 7.6 Hz, 1H), 6.93 - 7.13 (m, 5H), 7.54 - 7.74 (m, 3H), 8.65 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 35.2, 35.8, 117.6, 118.8, 119.7, 123.8, 126.6, 126.8, 127.2, 129.1, 130.7, 130.9, 142.5, 145.9, 168.1; MS (ESI) m/z 239 (M+H)⁺; HRMS (ESI) m/z calc for $C_{15}H_{15}N_2O$ 239.1184, found: 239.1181.

N-tert-Butyl-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (24)

To a solution of **6** (40 mg, 0.14 mmol) in EtCO₂t-Bu (1 mL) was added conc. H₂SO₄ (cat.) and the resulting mixture was stirred at 42 °C for 1 h. Water (2 mL) was added and the crude product extracted with EtOAc (x2). The combined organics were washed with brine, dried over MgSO₄, filtered and evaporated under reduced pressure. The resultant residue was purified by silica gel flash-column chromatography with hexanes:EtOAc (7:3) as eluent to afford **24** (10 mg, 19%) as a pale yellow solid: IR (neat) 3326, 1594 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 1.46 (s, 9H), 2.88 - 3.28 (*app*-br s, 4H), 5.84 (s, 1H), 6.08 - 6.43 (br s, 1H), 6.52 - 6.95 (m, 3H), 6.99 - 7.16 (m, 2H), 7.35 - 7.53 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 29.0, 34.8, 35.3, 51.4, 117.5, 118.3, 120.3, 125.5, 126.0, 127.0, 127.4, 129.2, 130.0, 130.6, 141.5, 144.9, 166.5; MS (ESI) m/z 295 (M+H)⁺; HRMS (ESI) m/z calc for C₁₉H₂₃N₂O 295.1810, found: 295.1795.

2-Nitro-10,11-dihydro-5H-dibenz[b,f]azepine (25)

Following the procedure described for the preparation of 8-nitro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carbonitrile (7), compound **4** (300 mg, 1.54 mmol) was treated with AgNO₃ (286 mg, 1.69 mmol) and BzCl (195 μ L, 1.69 mmol) in anhydrous MeCN (10 mL) to give, after purification by silica gel flash-column chromatography with hexanes:DCM (6:4) as eluent, the title compound (100 mg, 27%) as an orange solid: Elem. anal. calc for C₁₄H₁₂N₂O₂: C, 69.99; H, 5.03; N, 11.66; found: C, 70.03; H, 4.96; N, 11.75; IR (neat) 3355, 1494, 1290 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 2.94 - 3.09 (m, 4H), 6.84 (*app*-t, J = 7.2 Hz, 1H), 7.05 - 7.18 (m, 4H), 7.91 - 7.98 (m, 2H), 9.39 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 34.7, 35.5, 118.0, 119.8, 121.4, 123.6, 126.9, 127.3, 127.4, 130.5, 130.6, 137.8, 141.0, 149.6; MS (ESI) m/z 241 (M+H)⁺; HRMS (ESI) m/z calc for C₁₄H₁₃N₂O₂ 241.0977, found: 241.0968.

10,11-Dihydro-5H-dibenz[b,f]azepine-5-carboxamide (26)

To a solution of dibenz[b,f]azepine **4** (100 mg, 0.50 mmol) in DCM (3.5 mL) was added CISO₂NCO (53 μ L, 0.60 mmol) at 0 °C and the resulting mixture was stirred for 1 h. Water (3 mL) was added and the mixture allowed to stir for an additional 1 h. The organic phase was separated, washed with brine, dried over MgSO₄ and the solvent evaporated to give a residue which was purified by silica gel flash-column chromatography with DCM:MeOH (95:5) as eluent to afford **26** (95 mg, 80%) as a white solid: Elem. anal. calc for C₁₅H₁₄N₂O: C, 75.61; H, 5.92; N, 11.76; found: C, 75.56; H, 5.95; N, 11.84; IR (neat) 3471, 3352, 1648 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 2.84 (*app*-s, 2H), 3.42 (*app*-s, 2H), 4.82 (s, 2H), 7.18 - 7.28 (m, 6H), 7.36 - 7.44 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 30.7, 127.1, 128.1, 128.5, 130.4, 140.6, 157.6; MS (EI) m/z 238 M⁺; HRMS (EI) m/z calc for C₁₅H₁₄N₂O 238.1106, found: 238.1100.

8-Chloro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (27)

$$H_2N$$
 N
 CI

Following the procedure described for the preparation of 8-nitro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (**8**), compound **34** (20 mg, 0.08 mmol) was treated with KOH (36 mg, 0.64 mmol) and 35% wt. H_2O_2 (aq.) (0.2 mL, 2.00 mmol) in EtOH (1 mL) to give, after purification by silica gel flash-column chromatography with DCM:EtOAc (4:6) as eluent, the desired compound **27** (18 mg, 86%) as a white solid: IR (neat) 3469, 3324, 3182, 1635 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 2.97 (*app*-s, 4H), 6.88 - 7.18 (m, 5H), 7.49 - 7.74 (m, 3H), 8.78 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 34.8, 35.3, 117.7, 120.3, 122.9, 124.3, 126.7, 126.9, 127.0, 130.0, 130.9, 131.0, 141.6, 145.3, 168.1; MS (ESI) m/z 273 (M+H)⁺; HRMS (ESI) m/z calc for $C_{15}H_{14}CIN_2O$ 273.0795, found: 273.0786.

8-Cyano-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (28)

$$H_2N$$
 N
 CN

Following the procedure described for the preparation of 8-nitro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (**8**), compound **11** (150 mg, 0.61 mmol) was treated with KOH (307 mg, 5.49 mmol) and 35% wt. H_2O_2 (aq.) (1.3 mL, 15.25 mol) to give, after purification by silica gel flash-column chromatography with EtOAc:MeOH (9:1) as eluent, the desired compound **28** (40 mg, 25%) as a brown solid: IR (neat) 3421, 3345, 2216, 1649 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 2.99 (*app*-s, 4H), 6.99 - 7.16 (m, 3H), 7.43 - 7.53 (m, 2H), 7.59 - 7.67 (m, 2H), 7.73 (s, 1H), 9.28 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 35.0, 35.1, 100.0, 118.6, 119.1, 120.3, 125.7, 126.9, 128.4, 128.8, 130.7, 131.1, 135.0, 144.2, 146.7, 167.9; MS (ESI) m/z 264 (M+H)⁺; HRMS (ESI) m/z calc for $C_{16}H_{14}N_3O$ 264.1137, found: 264.1128.

5-Methyl-8-nitro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (29)

$$H_2N$$
 N
 N
 N
 N
 N
 N

Following the procedure described for the preparation of 8-nitro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (**8**), compound **36** (25 mg, 0.09 mmol) was treated with KOH (45 mg, 0.81 mmol) and 35% wt. H_2O_2 (aq.) (0.2 mL, 2.25 mmol) in EtOH (1 mL) to give, after purification by silica gel flash-column chromatography with DCM:EtOAc (4:6) as eluent, the desired compound **29** (23 mg, 88%) as a yellow solid: Elem. anal. calc for $C_{16}H_{15}N_3O_3$: C, 64.64; H, 5.09; N, 14.13; found: C, 64.74; H, 5.00; N, 14.07; IR (neat) 3423, 3191, 1648, 1498, 1315 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 3.09 - 3.21 (m, 4H), 3.45 (s, 3H), 7.23 - 7.31 (m, 3H), 7.70 - 7.77 (m, 2H), 7.85 - 7.91 (br s, 1H), 7.98 - 8.06 (m, 2H); ¹³C NMR (100 MHz, DMSO-d₆) δ 31.6, 34.6, 42.6, 118.9, 121.3, 123.0, 126.4, 126.7, 128.7, 129.6, 130.8, 134.8, 140.2, 150.2, 152.9, 167.8; MS (ESI) m/z 298 (M+H)⁺; HRMS (ESI) m/z calc for $C_{16}H_{16}N_3O_3$ 298.1192, found: 298.1186.

6-Nitro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (30)

Following the procedure described for the preparation of 8-nitro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (**8**), compound **35** (30 mg, 0.11 mmol) was treated with KOH (55 mg, 0.99 mmol) and 35% wt. H_2O_2 (aq.) (0.25 mL, 2.75 mmol) in EtOH (1 mL) to give, after purification by silica gel flash-column chromatography with EtOAc as eluent, the title compound (23 mg, 73%) as a red solid: ¹H NMR (400 MHz, DMSO-d₆) δ 3.03 - 3.17 (m, 4H), 6.94 (*app*-t, J = 7.9 Hz, 1H), 7.09 (d, J = 8.2 Hz, 1H), 7.13 - 7.21 (br s, 1H), 7.50 (d, J = 7.0 Hz, 1H), 7.64 - 7.72 (m, 2H), 7.76 - 7.84 (br s, 1H), 8.01 (d, J = 8.0 Hz, 1H), 10.14 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 34.2, 34.6, 119.2, 119.8, 124.7, 126.3, 126.5, 128.2, 130.4, 133.8, 136.1, 136.5, 138.7, 142.3, 167.2; MS (ESI) m/z 284 (M+H)⁺; HRMS (ESI) m/z calc for $C_{15}H_{14}N_3O_3$ 284.1035, found: 284.1037.

6-(Hydroxymethyl)-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (31)

Following the procedure described for the preparation of (10,11-Dihydro-5H-dibenz[b,f]azepin-4-yl)methanol (14), compound 40 (40 mg, 0.15 mmol) was treated with NaBH₄ (6 mg, 0.15 mmol) in MeOH (1 mL) to give, after purification by silica gel flash-column chromatography with DCM:MeOH (9:1) as eluent, the desired compound 31 (35 mg, 87%) as colourless crystals: m.p. 201-203 °C (from EtOH); Elem. anal. calc for $C_{16}H_{16}N_2O_2$: C, 71.62; H, 6.01; N, 10.44; found: C, 71.68; H, 6.04; N, 10.40; IR (neat) 3360, 3193, 1665 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 3.01 (s, 4H), 4.64 (d, J = 5.1 Hz, 2H), 5.74 (t, J = 5.0 Hz, 1H), 6.76 (t, J = 7.4 Hz, 1H), 6.84 (d, J = 8.7 Hz, 1H), 6.96 - 7.11 (m, 3H), 7.55 - 7.63 (m, 2H), 7.67 (br s, 1H), 8.32 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 34.2, 35.3, 62.6, 117.6, 119.7, 123.5, 125.4, 126.5, 126.8, 128.9, 129.3, 130.7, 131.2, 140.8, 144.8, 167.5; MS (ESI) m/z 269 (M+H)⁺; HRMS (ESI) m/z calc for $C_{16}H_{17}N_2O_2$ 269.1290, found: 269.1292.

(10,11-Dihydro-5H-dibenz[b,f]azepine-2,6-diyl)dimethanol (32)

Following the procedure described for the preparation of (10,11-Dihydro-5H-dibenz[b,f]azepin-4-yl)methanol (**14**), compound **41** (90 mg, 0.36 mmol) was reduced with NaBH₄ (27 mg, 0.72 mmol) in MeOH (5 mL) to give, after purification by silica gel flash-column chromatography with DCM:MeOH (95:5) as eluent, compound **32** (80 mg, 88%) as a white solid: IR (neat) 3192 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 2.93 - 3.02 (m, 4H), 4.35 (d, J = 6.3 Hz, 2H), 4.61 (d, J = 5.2 Hz, 2H), 4.93 (t, J = 6.3 Hz, 1H), 5.66 (t, J = 5.2 Hz, 1H), 6.69 (app-t, J = 8.3 Hz, 1H), 6.78 (d, J = 8.2 Hz, 1H), 6.94 - 7.04 (m, 4H), 7.93 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 34.8, 35.4, 63.1, 63.3, 118.6, 119.2, 126.0, 126.9, 127.2, 128.6, 129.8, 129.9, 130.8, 132.8, 141.6, 142.4; MS (ESI) m/z 256 (M+H)⁺; HRMS (ESI) m/z calc for C₁₆H₁₈NO₂ 256.1338, found: 256.1327.

(10,11-Dihydro-5H-dibenz[b,f]azepine-2,8-diyl)dimethanol (33)

Following the procedure described for the preparation of (10,11-Dihydro-5H-dibenz[b,f]azepin-4-yl)methanol (**14**), compound **37** (50 mg, 0.20 mmol) was reduced with NaBH₄ (15 mg, 0.40 mmol) in MeOH (3 mL) to give, after purification by silica gel flash-column chromatography with DCM:Et₂O (1:1) as eluent, compound **33** (47 mg, 93%) as a yellow solid: IR (neat) 3367 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 2.94 (s, 4H), 4.34 (d, J = 5.8 Hz, 4H), 4.90 (t, J = 5.8 Hz, 2H), 6.86 - 6.99 (m, 6H), 8.18 (s, 1H); ¹³C NMR (100 MHz, DMSO-d₆) δ 35.4, 63.2, 118.0, 125.9, 127.6, 129.6, 132.6, 142.2; MS (ESI) m/z 256 (M+H)⁺; HRMS (ESI) m/z calc for C₁₆H₁₈NO₂ 256.1338, found: 256.1330.

8-Chloro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carbonitrile (34)

Following the procedure described for the preparation of 2-Bromo-10,11-dihydro-5H-dibenz[b,f]azepine (**5**), compound **6** (80 mg, 0.36 mmol) was treated with NCS (105 mg, 0.80 mmol) and SiO₂ (1.60 g) in DCM (20 mL) to give, after purification by silica gel flash-column chromatography with hexanes:Et₂O (6:4) as eluent, the desired compound **34** (30 mg, 33%) as a brown solid: IR (neat) 3347, 2212 cm⁻¹; ¹H NMR (400 MHz, MeOH-d₄) δ 3.00 (*app*-s, 4H), 6.88 (*app*-t, J = 8.0 Hz, 2H), 6.99 - 7.05 (m, 2H), 6.24 - 7.32 (m, 2H); ¹³C NMR (100 MHz, MeOH-d₄) δ 34.5, 35.1, 99.5, 118.1, 120.0, 120.1, 125.0, 126.7, 127.4, 129.6, 130.7, 131.0, 134.8, 140.0, 146.7; MS (ESI) m/z 254 M⁺; HRMS (ESI) m/z calc for C₁₅H₁₁ClN₂ 254.0611, found: 254.0597.

6-Nitro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carbonitrile (35)

Following the procedure described for the preparation of 8-Nitro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carbonitrile (7), compound **6** (480 mg, 2.18 mmol) was treated with AgNO₃ (405 mg, 2.40 mmol) and BzCl (277 μ L, 2.40 mmol) in anhydrous MeCN (10 mL) to give, after purification by silica gel flash-column chromatography with hexanes:DCM (1:9) as eluent, the desired compound **35** (51 mg, 10%) as an orange solid: ¹H NMR (400 MHz, CDCl₃) δ 3.05 - 3.20 (m, 4H), 6.89 (*app*-t, *J* = 8.6 Hz, 1H), 7.02 (d, *J* = 8.4 Hz, 1H), 7.34 (d, *J* = 7.3 Hz, 1H), 7.38 (s, 1H), 7.43 (*app*-t, *J* = 8.3 Hz, 1H), 8.13 (d, *J* = 8.5 Hz, 1H), 10.79 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 34.6, 35.1, 103.8, 119.1, 119.7, 121.0, 125.4, 129.6, 131.1, 133.2, 134.7, 136.2, 136.8, 139.0, 143.9; MS (EI) m/z 265 M*+; HRMS (EI) m/z calc for C₁₅H₁₂N₃O₂ 265.0851, found: 265.0851.

5-Methyl-8-nitro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carbonitrile (36)

To a solution of 7 (100 mg, 0.38 mmol) in THF (3 mL) was added 1.0M (sol. in THF) NaHMDS (0.49 mL, 0.49 mmol) at 0 °C and the resulting mixture was stirred at 0 °C for 10 min. MeI (36 μ L, 0.57 mmol) was added and the reaction mixture allowed to stir at room temperature for 2 h. Water (5 mL) was added and the crude product extracted with Et₂O. The organic phase was washed with brine, dried over MgSO₄, filtered and the solvent evaporated under reduced pressure. The resultant residue was purified by silica gel flash-column chromatography with hexanes:DCM (3:7) as eluent to yield the title compound (100 mg, 95%) as a yellow solid: IR (neat) 2225, 1480, 1316 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 3.16 - 3.25 (m, 4H), 3.49 (s, 3H), 7.13 (d, J = 9.0 Hz, 1H), 7.16 (d, J = 8.5 Hz, 1H), 7.42 (d, J = 1.9 Hz, 1H), 7.49 (dd, J = 8.7, 2.2 Hz, 1H), 8.00 (d, J = 2.7 Hz, 1H), 8.06 (dd, J = 9.0, 2.5 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 32.2, 33.5, 42.1, 106.1, 118.9, 119.1, 120.7, 122.7, 125.7, 131.0, 132.1, 133.3, 134.5, 141.7, 150.9, 152.3; MS (EI) m/z 279 M**; HRMS (EI) m/z calc for C₁₆H₁₄N₃O₂ 279.1008, found: 279.1003.

10,11-Dihydro-5H-dibenz[b,f]azepine-2,8-dicarbaldehyde (37)

To a solution of **11** (200 mg, 0.82 mmol) in DCM (2 mL) was added 1.0M (sol. in DCM) DIBAL-H (4.1 mL, 4.10 mmol) at 0 °C. 2.0M NaOH (aq.) (10 mL) was added after 30 min and the mixture allowed to stir at room temperature for 1 h. The organic layer was washed with brine, dried over MgSO₄, filtered and evaporated under reduced pressure. The resultant residue was purified by silica gel flash-column chromatography with DCM:Et₂O (9:1) as eluent to afford **37** (70 mg, 34%) as a yellow solid: IR (neat) 3313, 1663 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 3.08 (*app*-s, 4H), 7.22 (d, *J* = 8.5 Hz, 2H), 7.61 - 7.68 (m, 4H), 9.64 (s, 1H), 9.78 (s, 2H); ¹³C NMR (100 MHz, DMSO-d₆) δ 35.1, 119.6, 128.7, 128.9, 129.0, 133.5, 147.4, 191.2; MS (ESI) m/z 252 (M+H)⁺; HRMS (ESI) m/z calc for C₁₆H₁₄NO₂ 252.1025, found: 252.1016.

8-Bromo-10,11-dihydro-5H-dibenz[b,f]azepine-4-carbaldehyde (38)

Following the procedure described for the preparation of 2-Bromo-10,11-dihydro-5H-dibenz[b,f]azepine (**5**), compound **13** (2.50 g, 11.21 mmol) was treated with NBS (2.10 g, 11.77 mmol) and SiO₂ (25.00 g) in DCM (300 mL) to give, after purification by silica gel flash-column chromatography with hexanes:DCM (7:3) as eluent, the desired compound **38** (2.10 g, 60%) as a yellow solid: Elem. anal. calc for $C_{15}H_{12}BrNO$: C, 59.62; H, 4.00; N, 4.64; found: C, 59.71; H, 4.05; N, 4.70; IR (neat) 1624 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 2.99 - 3.10 (m, 4H), 6.80 (*app*-t, J = 8.0 Hz, 1H), 6.87 (d, J = 8.1 Hz, 1H), 7.17 - 7.25 (m, 3H), 7.42 (d, J = 7.9 Hz, 1H), 9.86 (s, 1H), 11.24 (br s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 34.6, 35.7, 113.2, 117.5, 120.2, 121.5, 129.0, 129.8, 132.5, 132.6, 135.9, 137.0, 140.0, 145.2, 194.7; MS (ESI) m/z 302 (M+H)⁺; HRMS (ESI) m/z calc for $C_{15}H_{13}BrNO$ 302.0181, found: 302.0175.

6-Formyl-10,11-dihydro-5H-dibenz[b,f]azepine-2-carbonitrile (39)

Following the procedure described for the preparation of 10,11-dihydro-5H-dibenz[b,f]azepine-2-carbonitrile (6), compound **38** (1.70 g, 5.63 mmol) was treated with Zn(CN)₂ (660 mg, 5.63 mmol) and Pd(PPh₃)₄ (1.30 g, 1.13 mmol) in degassed DMF (15 mL) to give, after purification by silica gel flash-column chromatography with hexanes:DCM (3:7) as eluent, compound **39** (1.10 g, 75%) as a yellow solid: Elem. anal. calc for C₁₆H₁₂N₂O: C, 77.40; H, 4.87; N, 11.28; found: C, 77.53; H, 4.76; N, 11.20; IR (neat) 2213, 1656 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 2.89-3.23 (m, 4H), 6.93 (*app*-t, J = 8.3 Hz, 1H), 7.02 (d, J = 8.1 Hz, 1H), 7.29 (d, J = 8.0 Hz, 1H), 7.35 (s, 1H), 7.41 (d, J = 7.8 Hz, 1H), 7.49 (d, J = 8.2 Hz, 1H), 9.90 (s, 1H), 11.44 (br s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 34.8, 35.2, 103.0, 119.0, 119.5, 120.5, 121.0, 129.8, 130.3, 131.0, 134.3, 135.9, 137.0, 144.1, 144.8, 195.0; MS (EI) m/z 248 M*+; HRMS (EI) m/z calc for C₁₆H₁₂N₂O 248.0950, found: 248.0958.

6-Formyl-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (40)

Following the procedure described for the preparation of 8-nitro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (**8**), compound **39** (150 mg, 0.60 mmol) was treated with KOH (302 mg, 5.40 mmol) and 35% wt. H_2O_2 (aq.) (1.3 mL, 15.00 mmol) in EtOH (5 mL) to give, after purification by silica gel flash-column chromatography with DCM:MeOH (95:5) as eluent, the title compound (118 mg, 73%) as a yellow solid: Elem. anal. calc for $C_{16}H_{14}N_2O_2$: C, 72.16; H, 5.30; N, 10.52; found: C, 72.18; H, 5.30; N, 10.43; IR (neat) 3346, 3169, 1645, 1607 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 3.14 (*app*-s, 4H), 5.67 (br s, 1H), 5.85 (br s, 1H), 6.90 (*app*-t, J = 8.1 Hz, 1H), 7.05 (d, J = 8.0 Hz, 1H), 7.28 (s, 1H), 7.49 (d, J = 8.2 Hz, 1H), 7.57 (d, J = 8.3 Hz, 1H), 7.64 (s, 1H), 9.91 (s, 1H), 11.44 (br s, 1H); ¹³C NMR (100 MHz, CD₃OD/CDCl₃) δ 35.2, 35.7, 118.7, 120.0, 120.9, 125.0, 126.7, 126.8, 130.0, 130.6,

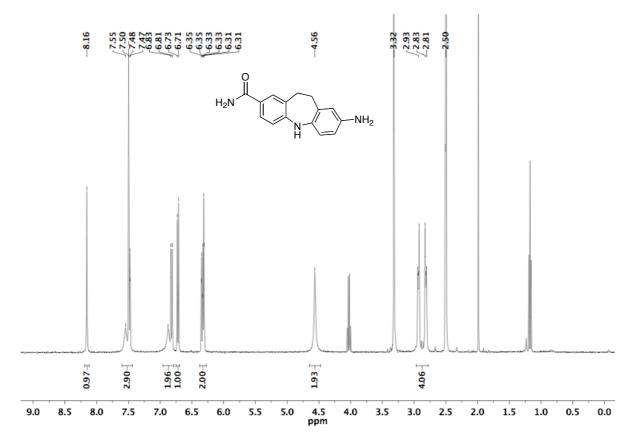
136.2, 137.4, 144.4, 144.8, 170.6, 195.5; MS (ESI) m/z 267 (M+H) $^+$; HRMS (ESI) m/z calc for $C_{16}H_{15}N_2O_2$ 267.1134, found: 267.1125.

6-(Hydroxymethyl)-10,11-dihydro-5H-dibenz[b,f]azepine-2-carbaldehyde (41)

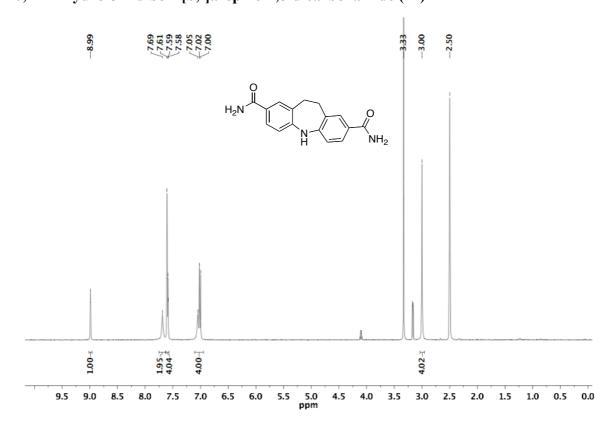
Following the procedure described for the preparation of 10,11-Dihydro-5H-dibenz[b,f]azepine-2,8-dicarbaldehyde (37), compound 39 (100 mg, 0.40 mmol) was treated with DIBAL-H (2.0 mL, 2.00 mmol) in DCM (2 mL) to give, after purification by silica gel flash-column chromatography with DCM:MeOH (95:5) as eluent, the title compound (90 mg, 88%) as a yellow oil: IR (neat) 3296, 1657 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 3.14 (*app*-s, 4H), 4.85 (s, 2H), 6.83 - 6.93 (m, 2H), 7.05 (d, J = 7.2 Hz, 1H), 7.11 (d, J = 7.1 Hz, 1H), 7.58 (s, 1H), 7.62 (d, J = 9.3 Hz, 1H), 8.48 (br s, 1H), 9.78 (s, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 34.8, 35.8, 65.4, 118.8, 120.8, 126.6, 127.4, 127.5, 127.6, 129.0, 130.6, 132.5, 133.8, 141.1, 148.0, 190.6; MS (ESI) m/z 254 (M+H)⁺; HRMS (ESI) m/z calc for C₁₆H₁₆NO₂ 254.1181, found: 254.1179.

F. ¹H NMR Spectra of Tested Compounds Without Elemental Analysis

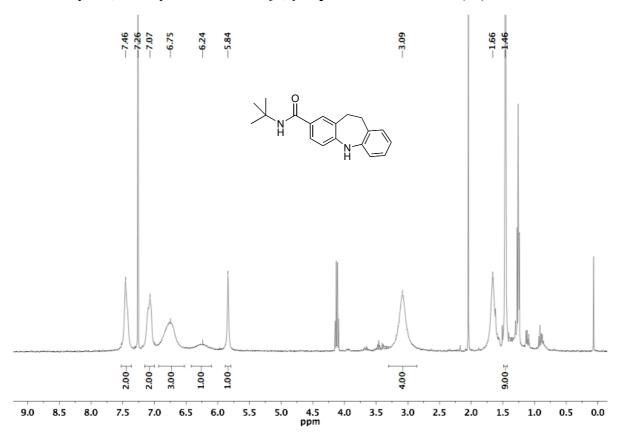
8-Amino-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (9)



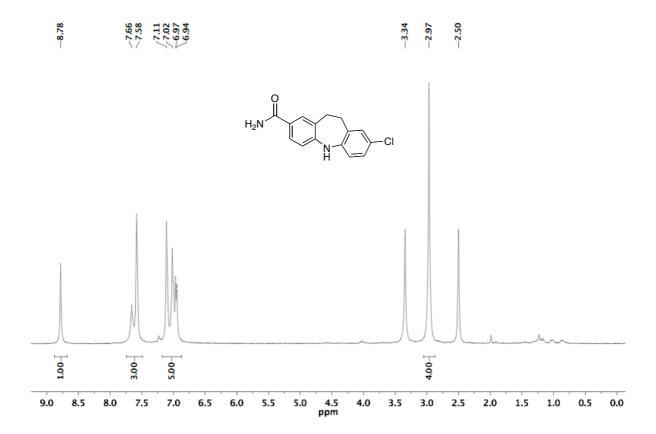
10,11-Dihydro-5H-dibenz[b,f]azepine-2,8-dicarboxamide (12)



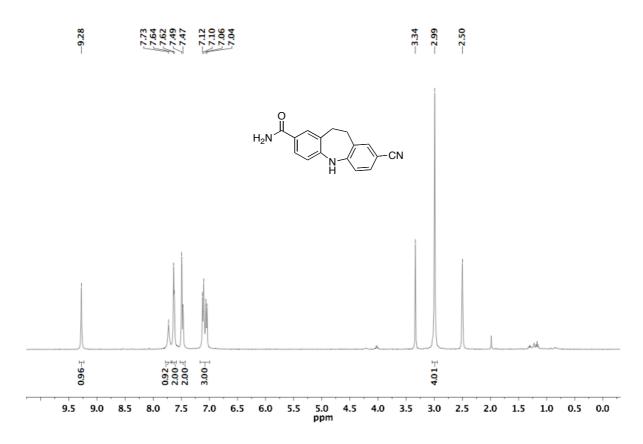
N-tert-Butyl-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (24)



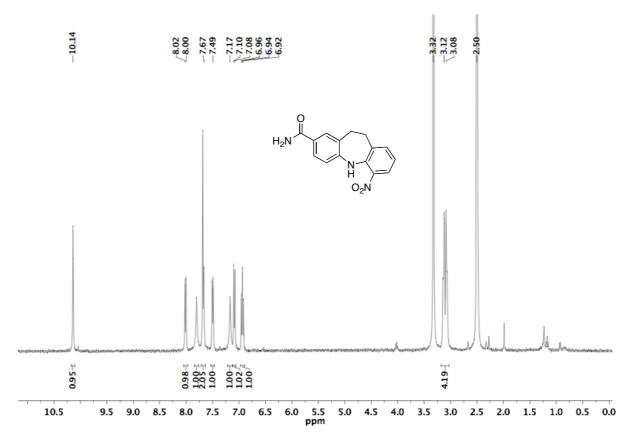
8-Chloro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (27)



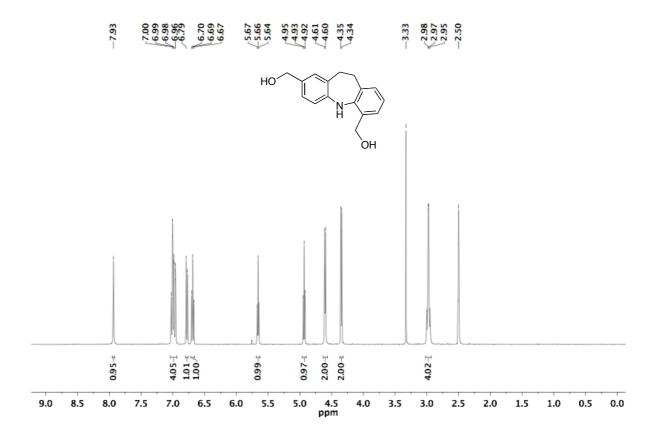
8-Cyano-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (28)



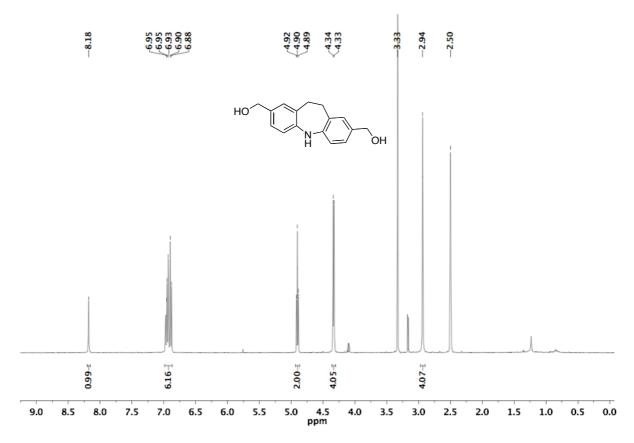
6-Nitro-10,11-dihydro-5H-dibenz[b,f]azepine-2-carboxamide (30)



(10,11-Dihydro-5H-dibenz[b,f]azepine-2,6-diyl)dimethanol (32)



(10,11-Dihydro-5H-dibenz[b,f]azepine-2,8-diyl)dimethanol (33)



G. References

- 1 Universal Protein Resource (http://www.uniprot.org/).
- 2 B. E. Hall, J. R. Buchberger, S. A. Gerber, A. L. B. Ambrosio, S. P. Gygi, D. Filman, D. Moazed, and T. Ellenberger, unpublished work.
- 3 Protein-protein BLAST (blastp), National Center for Biotechnology information, Bethesda, US (http://www.ncbi.nlm.nih.gov/BLAST/Blast.cgi/).
- 4 SAS, European Bioinformatics Institute, Cambridge, UK (http://www.ebi.ac.uk/thornton-srv/databases/sas/).
- 5 G. Jones, P. Willett, R. C. Glen, and R. Taylor, *J. Mol. Biol.*, 1997, **267**, 727-748.
- 6 CambridgeSoft, ChemBioOffice Ultra 2010 Suite (http://www.cambridgesoft.com/).
- 7 DeLano WL (2002) The PyMOL Molecular Graphics System. DeLano Scientific, Palo Alto, CA (http://www.pymol.org/).
- 8 R. A. Laskowski, M. W. MacArthur, D. S. Moss, and J. M. Thornton, *J. App. Cryst.*, 1993, **26**, 283-291.
- 9 PDBsum, European Bioinformatics Institute, Cambridge, UK (http://www.ebi.ac.uk/pdbsum/).