

## SUPPLEMENTARY INFORMATION

### Synthesis and biological activities of ferrocenyl derivatives of paclitaxel

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## Experimental

Commercially available solvents (HPLC grade – ALDRICH) and reagents (ALDRICH or AKSCI (USA) or HBCChem (USA)) were used as received. Microanalyses (C, H, and N) and MS spectra were performed at The Centre of Molecular and Macromolecular Studies in Łódź (Polish Academy of Sciences), Poland. MS spectra (MALDI-TOF) were recorded on Voyager-Elite, PerSeptive Biosystems Inc. (Framingham, MA, USA), pulse laser at 337 nm was used.  $^1\text{H}$  and  $^{13}\text{C}$  nuclear magnetic resonance spectra were recorded on Bruker Avance III 600 MHz spectrometer operating at 600.26 and 150.95 MHz respectively. Chemical shifts were determined relative to the solvent residual peaks.

## Synthesis

### General procedure of acylation of taxoids with ferrocene acids.

#### 2'-ferrocenoylpaclitaxel **3**

To a solution of 85.4 mg (0.10 mmol) of paclitaxel **1** and 25.4 mg (0.11 mmol) of ferrocenecarboxylic acid and 13 mg (0.11 mmol) of DMAP in 5 ml of anhydrous dichloromethane, diisopropylcarbodiimide (DIC) (25.2 mg, 0.20 mmol, 31  $\mu\text{l}$ ) was added and the resulted solution was stirred at 0°C. After 4 h the organic solvent was evaporated and the product was purified by chromatography on silica gel (50 g) using n-hexane – ethyl acetate 1-1 as eluent. Yield: 71 mg, 67%, of an orange crystals.

$^1\text{H}$  NMR (600.26 MHz,  $\text{CDCl}_3$ ):  $\delta$  1.15 (3 H, s), 1.25 (3 H, s), 1.70 (3 H, s), 1.90 (1 H, t,  $J=12.23$  Hz), 2.03 (3 H, s), 2.14 (1 H, dd,  $J=15.25$ , 8.85 Hz), 2.23 (3 H, s), 2.38 (1 H, dd,

$J=15.25, 9.22$  Hz), 2.50 (3 H, s), 2.58 (1 H, ddd,  $J=15.06, 9.03, 6.78$  Hz), 3.85 (1 H, d,  $J=6.78$  Hz), 4.15 (5 H, s), 4.22 (1 H, d,  $J=8.28$  Hz), 4.33 (1 H, d,  $J=8.28$  Hz), 4.47 (3 H, br. s.), 4.79 (1 H, br. s.), 4.82 (1 H, br. s.), 4.99 (1 H, d,  $J=9.03$  Hz), 5.55 (1 H, d,  $J=3.01$  Hz), 5.70 (1 H, d,  $J=7.15$  Hz), 6.00 (1 H, d,  $J=6.40$  Hz), 6.28 - 6.31 (1 H, m), 6.33 (1 H, s), 7.00 (1 H, d,  $J=8.28$  Hz), 7.33 - 7.38 (1 H, m), 7.42 - 7.49 (6 H, m), 7.51 - 7.56 (3 H, m), 7.59 - 7.66 (1 H, m), 7.81 (2 H, d,  $J=7.53$  Hz), 8.14 (2 H, d,  $J=7.53$  Hz)

$^{13}\text{C}$  NMR (150.95 MHz,  $\text{CDCl}_3$ ):  $\delta$  9.61, 14.18, 14.91, 20.80, 22.17, 22.76, 26.82, 35.51, 35.63, 43.18, 45.60, 53.13, 58.54, 60.36, 68.84, 69.99, 70.14, 70.35, 71.69, 71.90, 72.01, 72.11, 74.13, 75.16, 75.62, 76.43, 79.22, 81.08, 84.46, 126.42, 127.02, 128.49, 128.73, 128.80, 129.10, 129.22, 130.21, 132.02, 132.70, 133.64, 133.83, 137.29, 143.00, 167.04, 168.22, 169.75, 170.86, 171.10, 171.22, 203.83

IR (KBr)  $\nu_{\text{max}}/\text{cm}^{-1}$ : 3447 (NH), 1720 (C=O)

Elemental analysis calculated for  $\text{C}_{58}\text{H}_{59}\text{FeNO}_{15}$  C-65.35, H – 5.58, N – 1.31 found C – 65.40, H – 5.62, N – 1.28

MS (MALDI, HCHA): calculated for  $\text{C}_{58}\text{H}_{59}\text{FeNO}_{15}$ : 1065.32, found: 1065.28

## 2'-(3-ferrocenoylpropiolyl)paclitaxel 4

It was prepared in the same method as **3** starting from 3-ferrocenoylpropionic acid (32mg, 0.2 mmol) instead of ferrocenecarboxylic acid. Yield: 77.4 mg, 69% (orange crystals).

$^1\text{H}$  NMR (600.26 MHz,  $\text{CDCl}_3$ ):  $\delta$  1.14 (3 H, s), 1.23 (3 H, s), 1.68 (3 H, s), 1.82 (1 H, br. s.), 1.86 (1 H, s), 1.87 - 1.90 (1 H, m), 1.93 (3 H, s), 2.14 (1 H, dd,  $J=15.43, 9.03$  Hz), 2.22 (3 H, s), 2.34 (1 H, dd,  $J=15.43, 9.41$  Hz), 2.44 (3 H, s), 2.49 (1 H, br. s.), 2.51 - 2.58 (1 H, m), 2.79 - 2.86 (2 H, m), 2.99 - 3.06 (1 H, m), 3.06 - 3.14 (1 H, m), 3.81 (1 H, d,  $J=6.78$  Hz), 4.22 (5 H, s), 4.31 (1 H, d,  $J=8.28$  Hz), 4.39 - 4.46 (1 H, m), 4.52 (2 H, d,  $J=1.13$  Hz), 4.77 (2 H, dd,  $J=8.66, 1.13$  Hz), 4.96 (1 H, d,  $J=8.66$  Hz), 5.52 (1 H, d,  $J=3.76$  Hz), 5.68 (1 H, d,  $J=7.15$  Hz), 5.95 (1 H, dd,  $J=8.66, 3.39$  Hz), 6.25 (1 H, t,  $J=8.85$  Hz), 6.28 (1 H, s), 7.08 (1 H, d,  $J=8.66$  Hz), 7.33 (1 H, td,  $J=5.46, 2.63$  Hz), 7.40 - 7.45 (6 H, m), 7.49 - 7.55 (3 H, m), 7.59 - 7.64 (1 H, m), 7.80 (2 H, d,  $J=7.15$  Hz), 8.14 (2 H, d,  $J=7.53$  Hz)

$^{13}\text{C}$  NMR (150.95 MHz,  $\text{CDCl}_3$ ):  $\delta$  9.94, 14.18, 21.01, 22.60, 23.47, 26.37, 27.47, 28.16, 33.87, 35.58, 36.87, 42.16, 43.10, 46.41, 57.60, 60.33, 69.11, 69.29, 69.94, 71.79, 71.86, 72.29, 72.31, 74.49, 74.58, 75.06, 76.56, 78.24, 78.91, 80.33, 80.99, 84.22, 126.50, 128.17,

128.68, 128.86, 129.27, 130.18, 133.60, 135.52, 139.18, 156.90, 167.06, 168.14, 169.65,  
171.13, 172.20, 201.47, 211.58

IR (KBr)  $\nu_{\text{max}}/\text{cm}^{-1}$ : 3432 (NH), 1741 (C=O), 1725 (C=O)

Elemental analysis calculated for  $C_{61}H_{63}\text{FeNO}_{16}$  C-65.30, H – 5.66, N – 1.25 found C – 65.18,  
H – 5.72, N – 1.38

MS (MALDI, HCHA): calculated for  $C_{61}H_{63}\text{FeNO}_{16}$ : 1121.35, found: 1121.45

### **2'-ferrocenoyldocetaxel 5**

It was prepared in the same method as **3** starting from docetaxel (80.5 mg, 0.10 mmol) instead  
of paclitaxel. Yield: 67mg, 66% (orange crystals).

$^1\text{H}$  NMR (600.26 MHz.  $CDCl_3$ ):  $\delta$  1.14 (3 H, s), 1.25 (3 H, s), 1.37 (9 H, s), 1.67 (3H, s), 1.77  
(3 H, s), 1.87 (1 H, t,  $J=12.42$  Hz), 2.03 (3 H, s), 2.11 - 2.19 (1 H, m), 2.34 (1 H, br. s.), 2.48  
(3 H, br. s.), 2.56 - 2.65 (1 H, m), 3.97 (1 H, d,  $J=7.15$  Hz), 4.11 (5 H, s), 4.19 - 4.25 (2 H, m),  
4.29 (1 H, d,  $J=4.52$  Hz), 4.34 (1 H, d,  $J=8.66$  Hz), 4.44 (2 H, br. s.), 4.76 (1 H, br. s.), 4.81 (1  
H, br. s.), 4.99 (1 H, d,  $J=9.04$  Hz), 5.25 (1 H, s), 5.43 (1 H, br. s.), 5.55 (1 H, br. s.), 5.71 (1  
H, d,  $J=7.15$  Hz), 6.25 - 6.35 (1 H, m), 7.30 - 7.35 (1 H, m), 7.38 - 7.42 (2 H, m), 7.43 - 7.47  
(2 H, m), 7.52 (2 H, t,  $J=7.72$  Hz), 7.60 - 7.64 (1 H, m), 8.13 (2 H, d,  $J=7.53$  Hz)

$^{13}\text{C}$  NMR (150.95 MHz.  $CDCl_3$ ):  $\delta$  9.95, 14.15, 20.93, 20.99, 22.68, 26.34, 28.17, 35.58,  
36.85, 43.09, 46.42, 57.57, 60.36, 68.92, 69.98, 70.10, 70.44, 71.71, 71.80, 71.85, 74.37,  
74.48, 75.09, 76.54, 78.95, 81.00, 84.23, 126.24, 128.18, 128.66, 128.91, 129.30, 130.16,  
133.57, 135.43, 167.02, 168.14, 169.69, 170.95, 171.13, 211.53,

IR (KBr)  $\nu_{\text{max}}/\text{cm}^{-1}$ : 3447 (NH), 1720 (C=O)

Elemental analysis calculated for  $C_{54}H_{61}\text{FeNO}_{15}$  C-63.59, H – 6.03, N – 1.37 found C – 63.52,  
H – 6.11, N – 1.42

MS (MALDI, HCHA): calculated for  $C_{54}H_{61}\text{FeNO}_{15}$ : 1119.34, found: 1119.34

### **2'-(3-ferrocenylpropioyl)docetaxel 6**

It was prepared in the same method as **4** starting from docetaxel (80.5 mg, 0.10 mmol) instead  
of paclitaxel. Yield: 74 mg, 69% (orange crystals).

<sup>1</sup>H NMR (600.26 MHz, *CDCl*<sub>3</sub>) δ 1.13 (3 H, s), 1.24 (3 H, s), 1.35 (9 H, s), 1.68 (2 H, s), 1.75 (3 H, s), 1.85 (1 H, t, *J*=12.42 Hz), 1.95 (3 H, s), 2.05 (2 H, s), 2.14 - 2.22 (1 H, m), 2.32 (1 H, br. s.), 2.43 (3 H, br. s.), 2.53 - 2.61 (1 H, m), 2.75 - 2.83 (2 H, m), 2.94 - 3.01 (1 H, m), 3.03 - 3.11 (1 H, m), 3.94 (1 H, d, *J*=6.78 Hz), 4.17 - 4.21 (2 H, m), 4.23 (5 H, s), 4.33 (1 H, d, *J*=8.66 Hz), 4.52 (2 H, s), 4.78 (1 H, br. s.), 4.80 (1 H, br. s.), 4.96 (1 H, d, *J*=9.03 Hz), 5.20 (1 H, s), 5.37 (1 H, br. s.), 5.45 (2 H, br. s.), 5.70 (1 H, d, *J*=6.78 Hz), 6.26 (1 H, br. s.), 7.33 (3 H, d, *J*=7.53 Hz), 7.38 - 7.43 (2 H, m), 7.48 - 7.54 (2 H, m), 7.59 - 7.64 (1 H, m), 8.13 (2 H, d, *J*=7.53 Hz)

<sup>13</sup>C NMR (150.95 MHz, *CDCl*<sub>3</sub>) δ 211.58, 201.47, 172.20, 171.13, 169.65, 168.14, 167.06, 156.90, 139.18, 135.52, 133.60, 130.18, 129.27, 128.86, 128.68, 128.17, 126.50, 84.22, 80.99, 80.33, 78.91, 78.24, 76.56, 75.06, 74.58, 74.49, 72.31, 72.29, 71.86, 71.79, 69.94, 69.29, 69.11, 60.37, 57.60, 46.41, 43.10, 42.20, 36.87, 35.58, 33.84, 28.16, 27.47, 26.37, 22.60, 20.89, 14.18, 9.94,

IR (KBr) ν<sub>max</sub>/cm<sup>-1</sup>: 3442 (NH), 1741 (C=O), 1717 (C=O)

Elemental analysis calculated for C<sub>57</sub>H<sub>65</sub>FeNO<sub>16</sub> C-63.63, H – 6.09, N – 1.30 found C – 63.57, H – 6.28, N – 1.24

MS (MALDI, HCHA): calculated for C<sub>57</sub>H<sub>65</sub>FeNO<sub>16</sub>: 1075.67, found: 1075.37

### Cytotoxic activity

The sensitivity of the parental and multidrug-resistant cancer cell lines was determined by a modified MTT-reduction assay. Shortly, the cells suspended in 100 µl of medium were seeded on the 96-well plates at the density of 5000 cells/well. The cells were allowed to attach for 24 hours and then the investigated compounds at the desired concentration was administered. The stock solutions of the taxanes were prepared in dimethylsulphoxide (DMSO) and care was taken to keep solvent concentration equal in all wells including controls. The final DMSO concentration did not exceed 0.1% v/v and was proven to be non-toxic to the cells. After 70 hours of incubation, MTT was added to the medium to a final concentration of 1.1 mmole/l. After further 2 hours the medium was removed and the formazane crystals were dissolved in 100 µl of DMSO. The absorbance was measured at 580 nm analytical wavelength and 720 nm reference wavelength. The results were turned into percentage of controls and the IC<sub>50</sub> values for each cell line and drug were calculated by GraphPad Prism 4.03 software (GraphPad Inc. La Jolla, California, USA).

### Tubulin affinity

The tubulin affinity was measured with Tubulin Polymerization Assay Biochem Fluorescence Based Kit (Cytoskeleton Inc., Denver, Colorado, USA) according to the manufacturer's instructions. The investigated compounds were added to a reaction medium in a final concentration range from 0.6 up to 3.0  $\mu$ M. The tubulin polymerization rate was determined from the linear portion of a polymerization curve and plotted against the drug concentration. The affinity constant was calculated by GraphPad Prism 4.03 software (GraphPad Inc. La Jolla, California, USA).

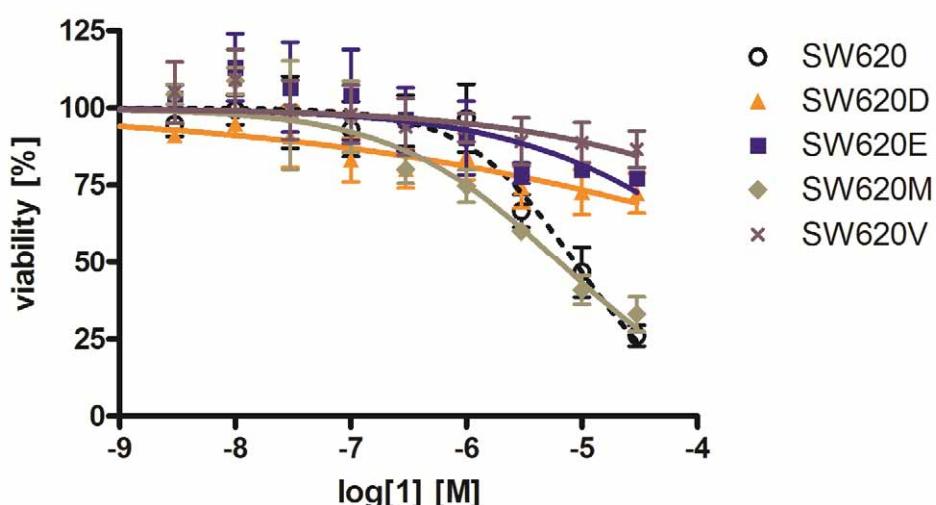


Figure S1. Viability of cancer cell lines in the presence of **1**

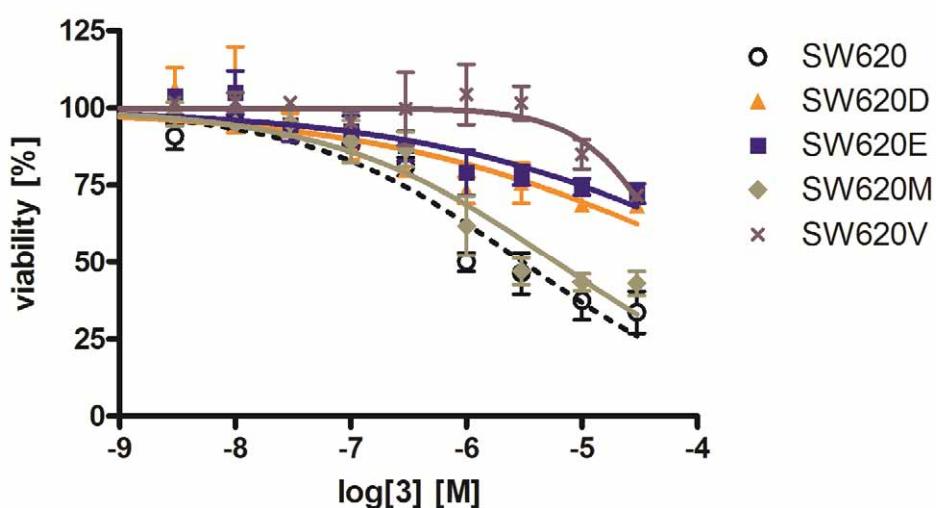


Figure S2. Viability of cancer cell lines in the presence of **3**

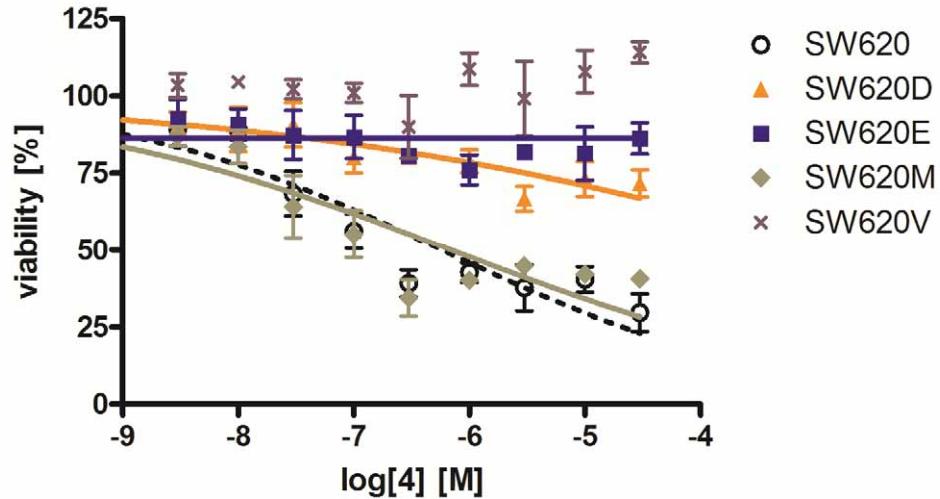


Figure S3. Viability of cancer cell lines in the presence of **4**

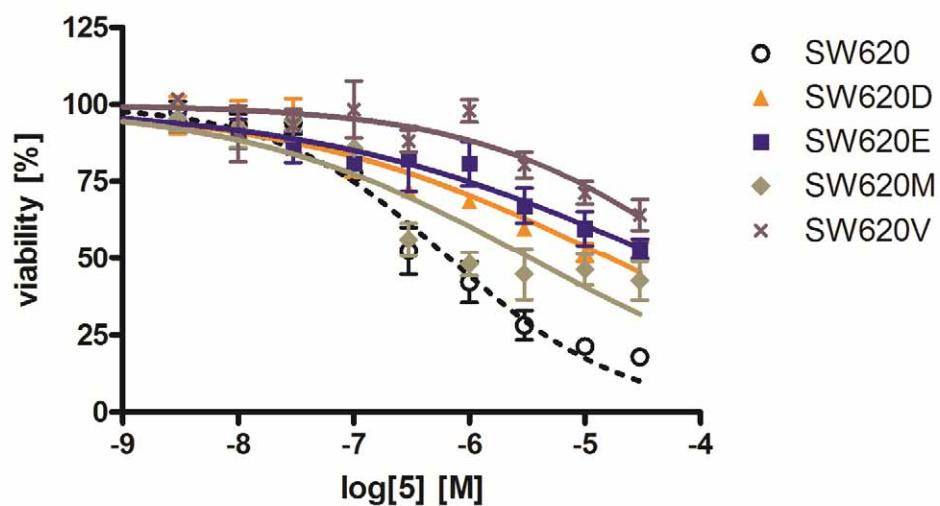


Figure S4. Viability of cancer cell lines in the presence of **5**

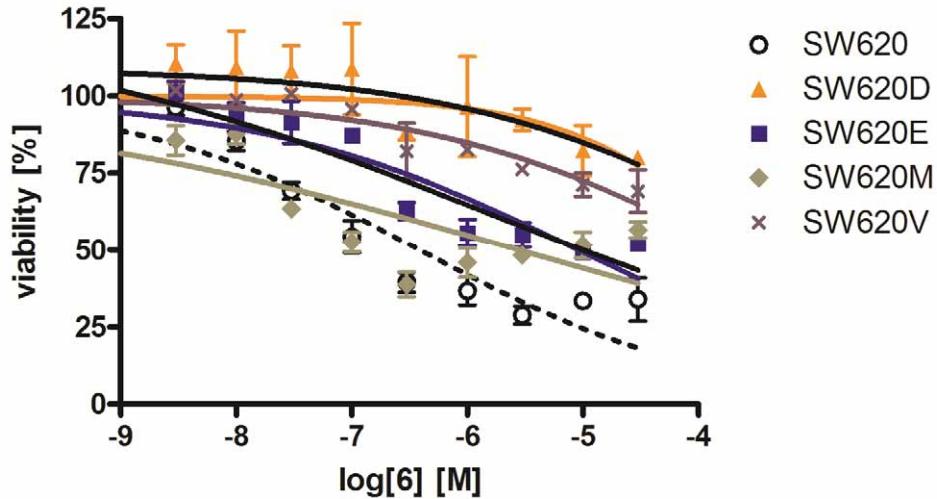


Figure S5. Viability of cancer cell lines in the presence of **6**

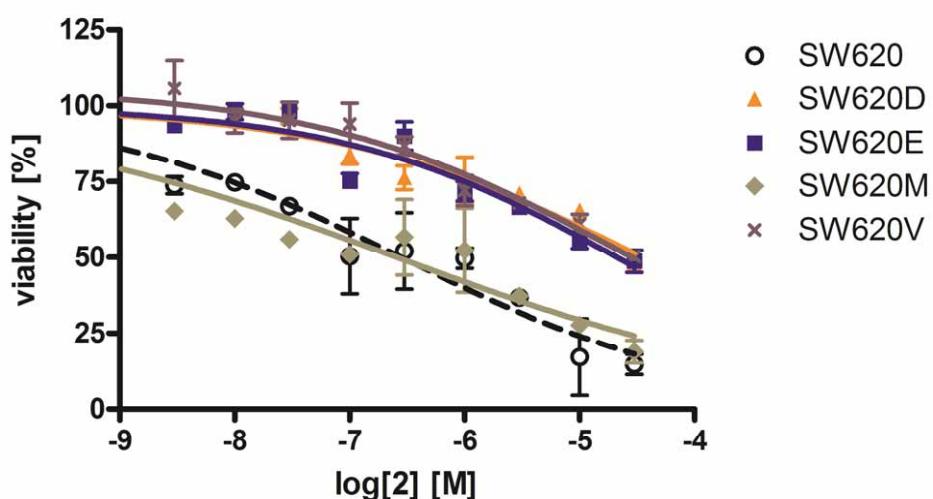


Figure S6. Figure S1. Viability of cancer cell lines in the presence of **2**

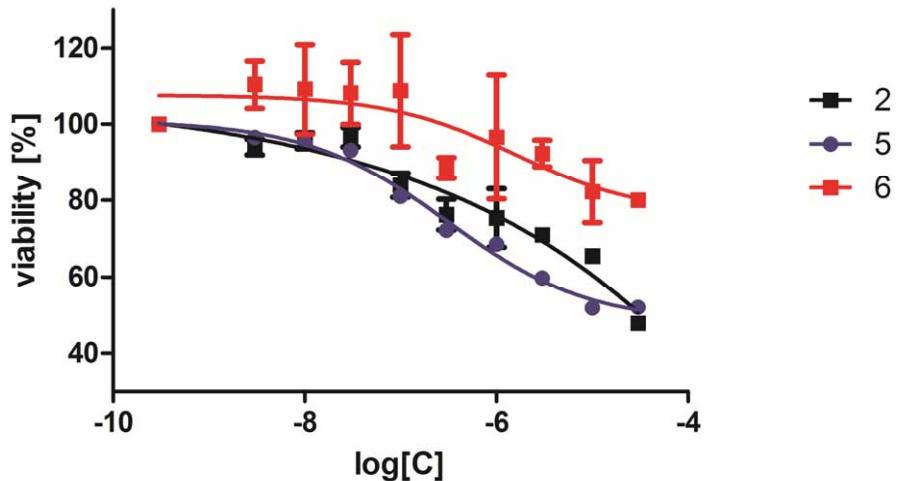


Figure S7. Viability of SW620D cell line in the presence of **2**, **5** and **6**

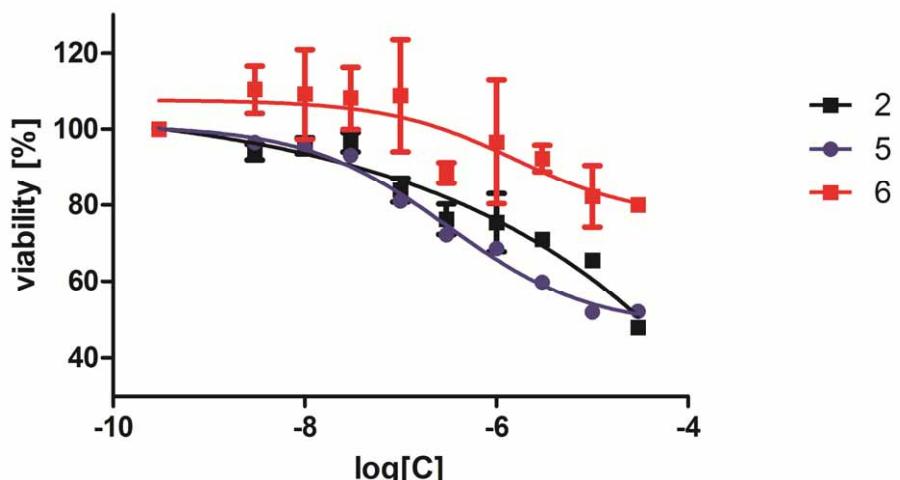
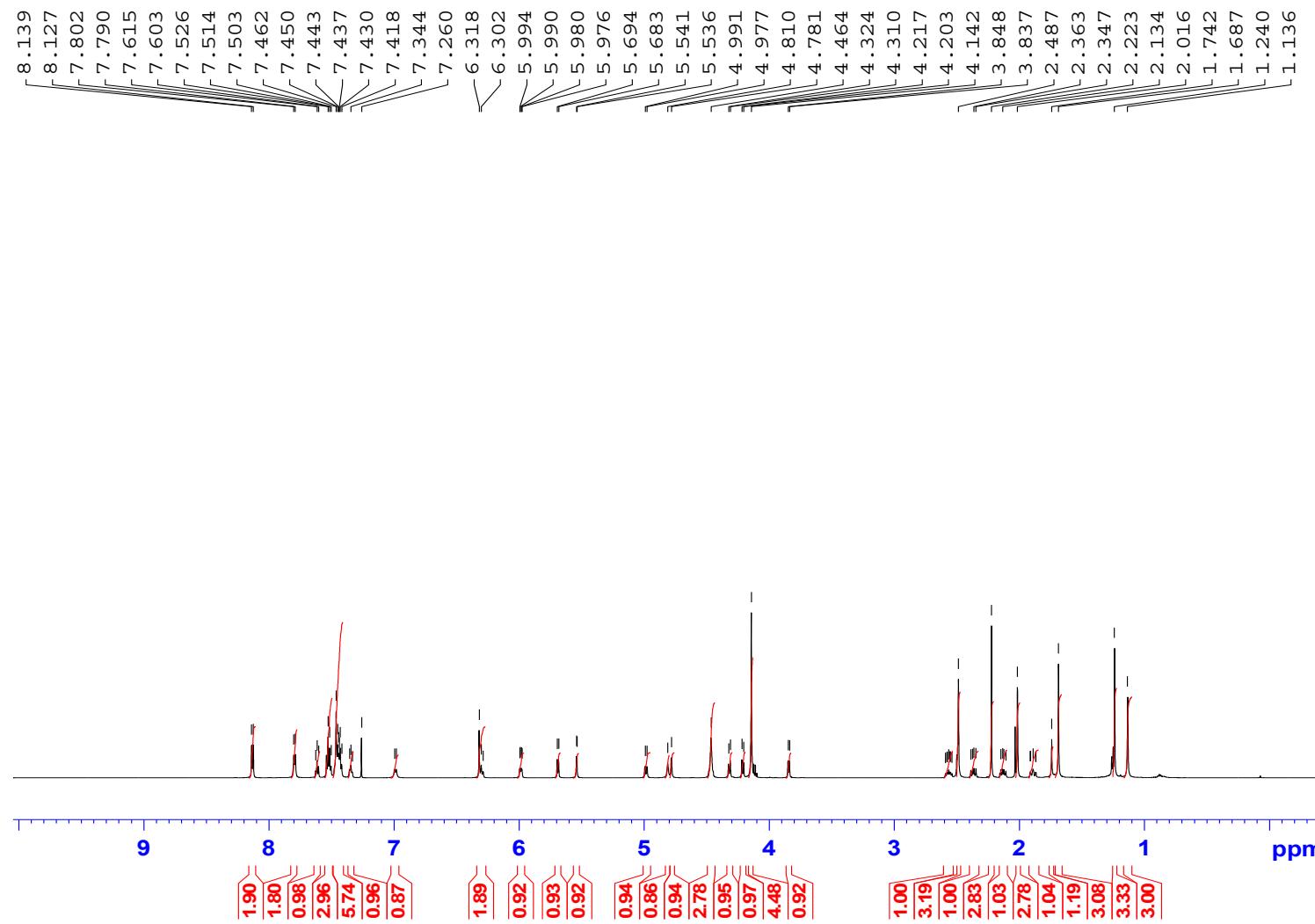


Figure S8. Viability of SW620E cell line in the presence of **2**, **5** and **6**

<sup>1</sup>H NMR of 3



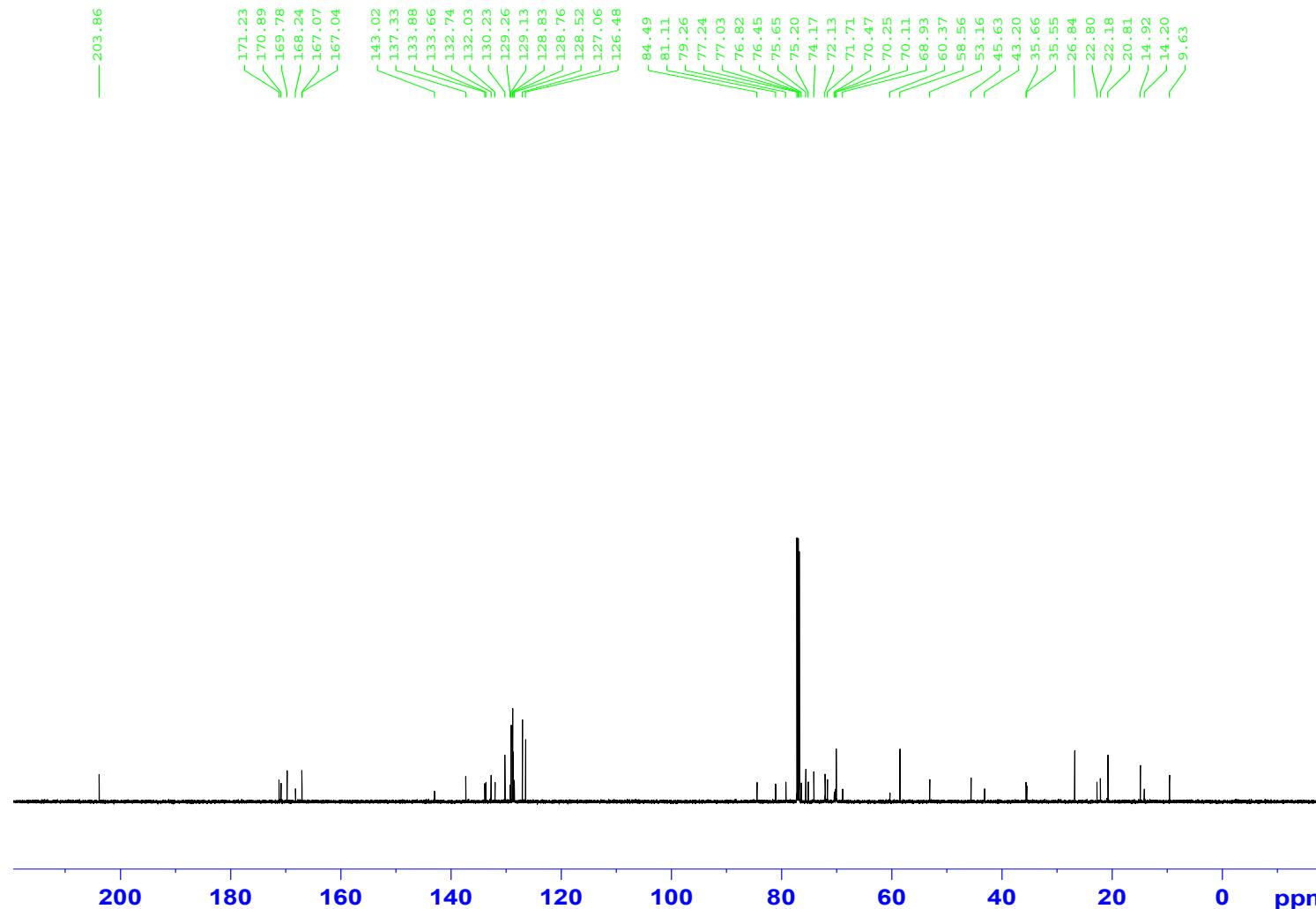
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FIDRES 0.188225 Hz  
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DW 40.533 usec  
DE 8.50 usec  
TE 302.0 K  
D1 1.0000000 sec  
TDO 1

===== CHANNEL f1 =====  
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P1 9.40 usec  
PL1 -3.00 dB  
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SFO1 600.2637069 MHz

F2 - Processing parameters  
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SSB 0  
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PC 1.00

<sup>13</sup>C NMR of 3



Current Data Parameters  
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PROCNO 1

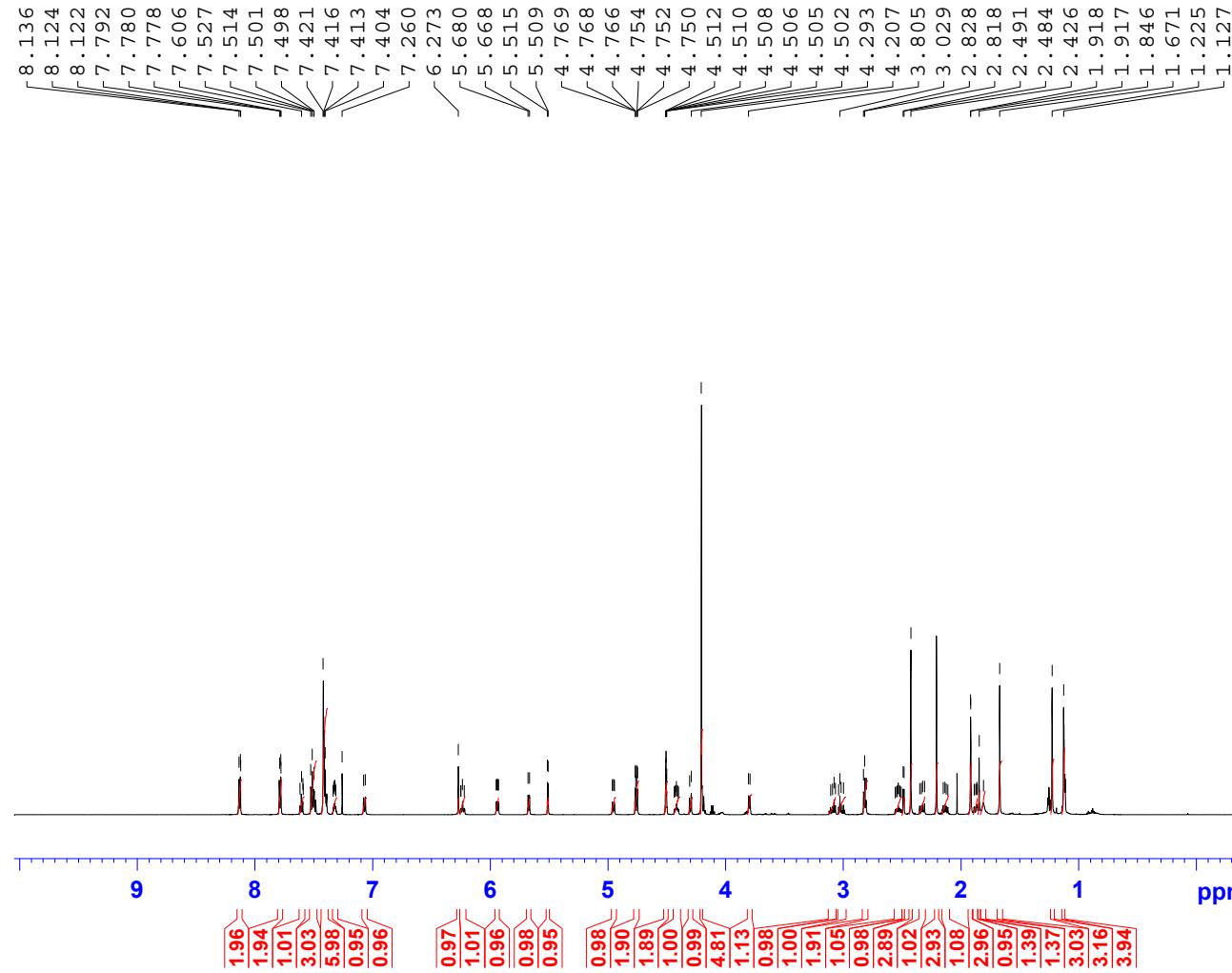
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D1 2.0000000 sec  
D11 0.03000000 sec  
TDO 1

===== CHANNEL f1 ======  
NUC1 <sup>13</sup>C  
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SFO1 150.9505906 MHz

===== CHANNEL f2 ======  
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NUC2 <sup>1H</sup>  
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PL2 -3.00 dB  
PL12 16.26 dB  
PL13 18.00 dB  
PL2W 30.57242203 W  
PL12W 0.36251819 W  
PL13W 0.24284537 W  
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F2 - Processing parameters  
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GB 0  
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<sup>1</sup>H NMR of 4



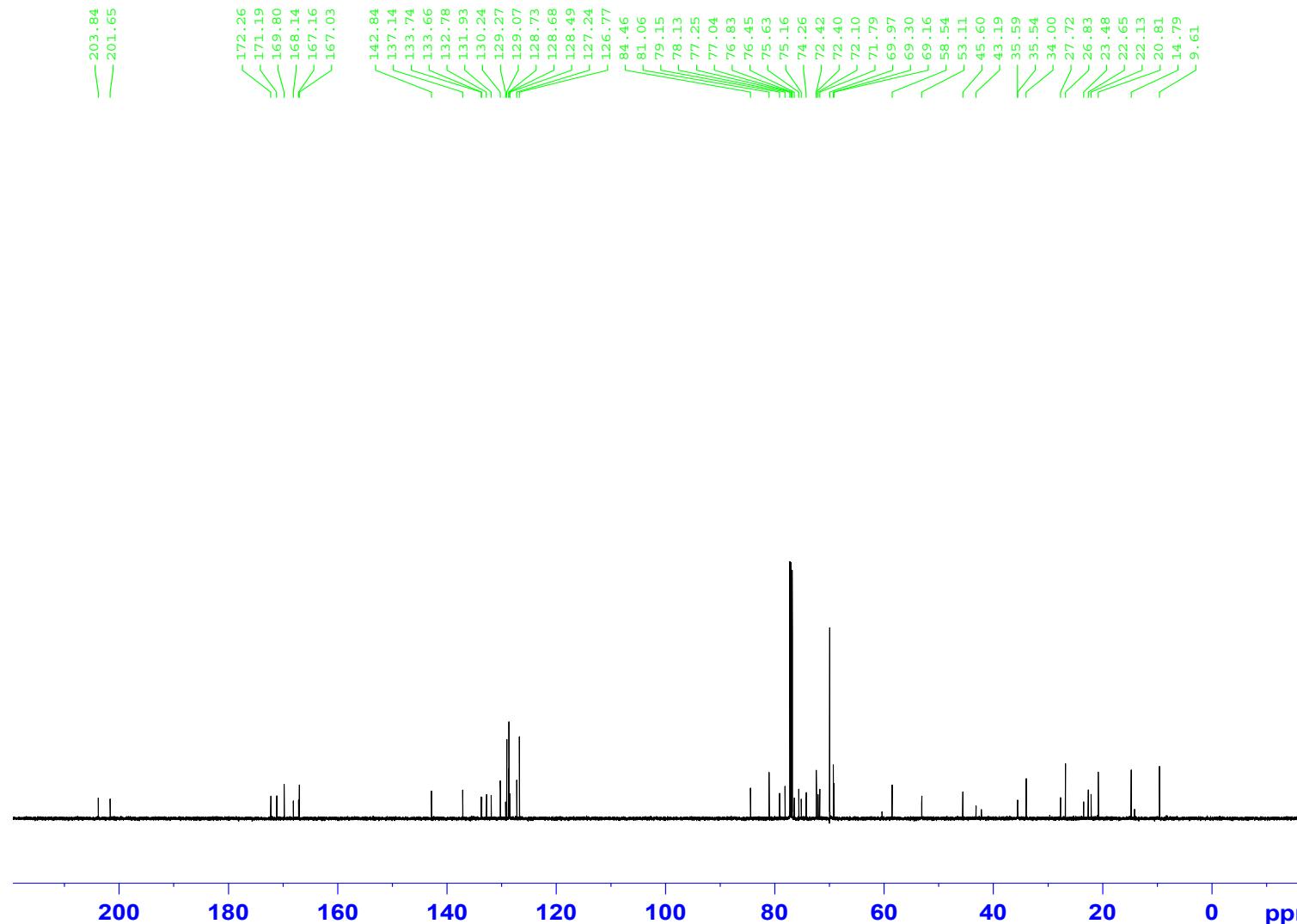
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DS 2  
SWH 12335.526 Hz  
FIDRES 0.188225 Hz  
AQ 2.6564426 sec  
RG 80.6  
DW 40.533 usec  
DE 8.50 usec  
TE 300.0 K  
D1 1.0000000 sec  
TD0 1

===== CHANNEL f1 =====  
NUC1 1H  
P1 9.40 usec  
PL1 -3.00 dB  
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SFO1 600.2637069 MHz

F2 - Processing parameters  
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<sup>13</sup>C NMR of 4



Current Data Parameters  
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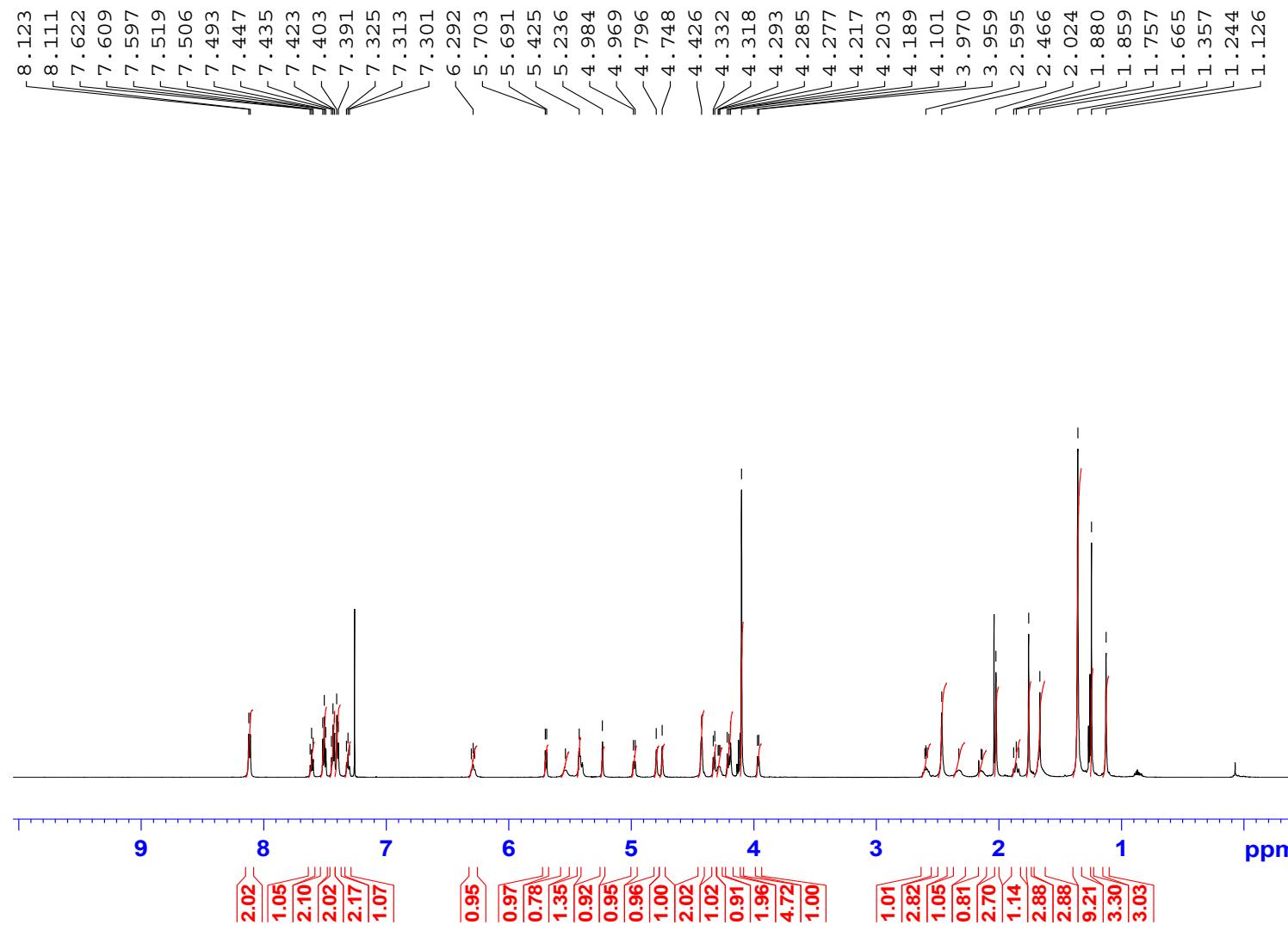
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SWH 36057.691 Hz  
FIDRES 0.550197 Hz  
AQ 0.9088159 sec  
RG 2050  
DW 13.867 usec  
DE 8.50 usec  
TE 301.2 K  
D1 2.0000000 sec  
D11 0.0300000 sec  
TD0 1

===== CHANNEL f1 ======  
NUC1 <sup>13</sup>C  
P1 10.50 usec  
PL1 0 dB  
PL1W 91.93504333 W  
SFO1 150.9505906 MHz

===== CHANNEL f2 ======  
CPDPRG2 waltz16  
NUC2 <sup>1</sup>H  
PCPD2 90.00 usec  
PL2 -3.00 dB  
PL12 16.26 dB  
PL13 18.00 dB  
PL2W 30.57242203 W  
PL12W 0.36251819 W  
PL13W 0.24284537 W  
SFO2 600.2624010 MHz

F2 - Processing parameters  
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WDW no  
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GB 0  
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<sup>1</sup>H NMR of 5



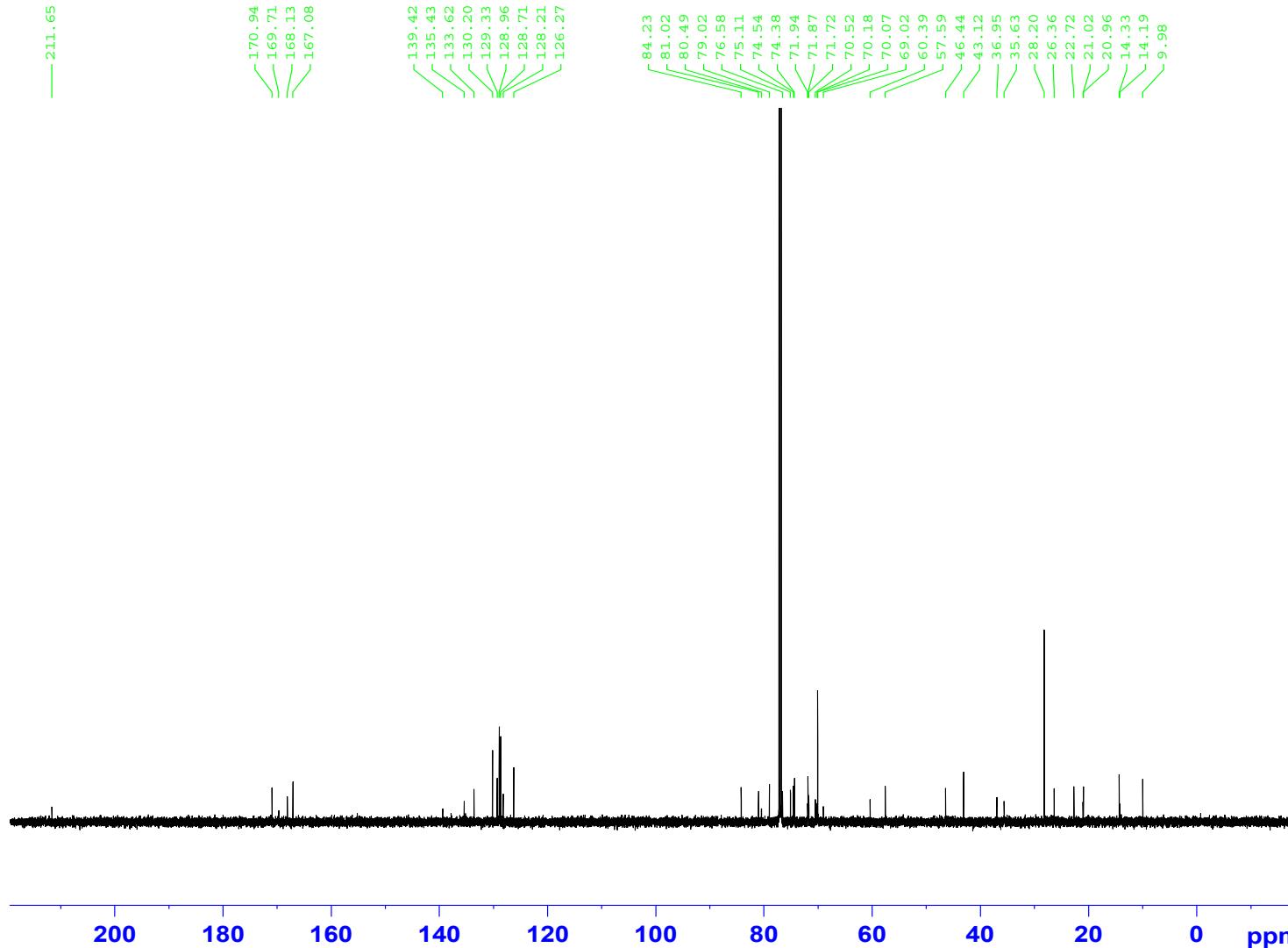
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Date\_ 20111212  
Time 20.52  
INSTRUM spect  
PROBHD 5 mm PABBO BB-  
PULPROG zg30  
TD 65536  
SOLVENT CDCl<sub>3</sub>  
NS 16  
DS 2  
SWH 12335.526 Hz  
FIDRES 0.188225 Hz  
AQ 2.6564426 sec  
RG 203  
DW 40.533 usec  
DE 8.50 usec  
TE 302.0 K  
D1 1.0000000 sec  
TD0 1

===== CHANNEL f1 =====  
NUC1 1H  
P1 9.40 usec  
PL1 -3.00 dB  
PL1W 30.57242203 W  
SFO1 600.2637069 MHz

F2 - Processing parameters  
SI 32768  
SF 600.2600174 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

<sup>13</sup>C NMR of 5



Current Data Parameters  
NAME dp-11-051  
EXPNO 10  
PROCNO 1

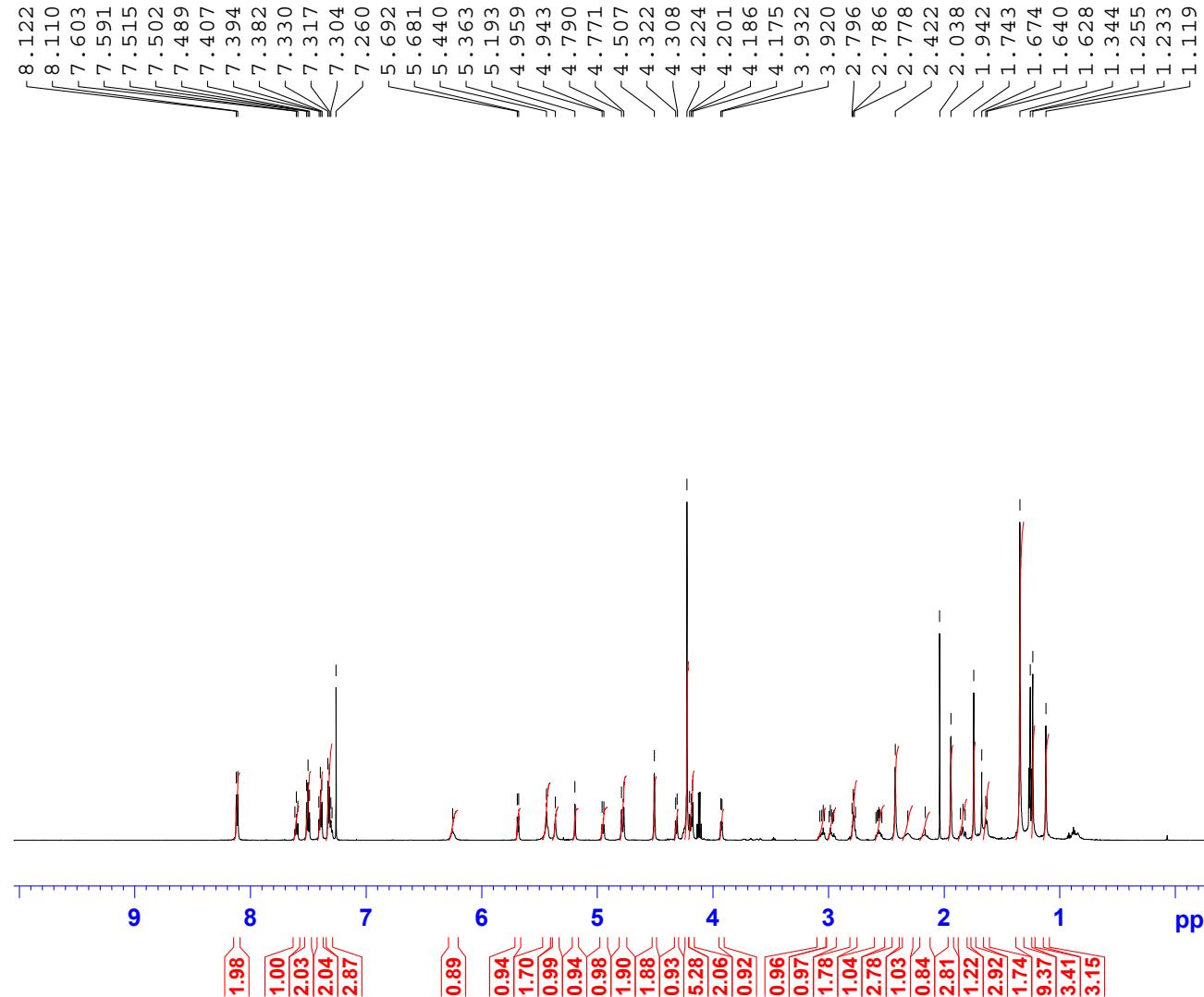
F2 - Acquisition Parameters  
Date\_ 20111212  
Time\_ 20.38  
INSTRUM spect  
PROBHD 5 mm PABBO BB-  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 1024  
DS 4  
SWH 36057.691 Hz  
FIDRES 0.550197 Hz  
AQ 0.9088159 sec  
RG 2050  
DW 13.867 usec  
DE 8.50 usec  
TE 302.0 K  
D1 2.0000000 sec  
D11 0.0300000 sec  
TDO 1

===== CHANNEL f1 =====  
NUC1 <sup>13</sup>C  
P1 10.50 usec  
PL1 0 dB  
PL1W 91.93504333 W  
SFO1 150.9505906 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 90.00 usec  
PL2 -3.00 dB  
PL12 16.26 dB  
PL13 18.00 dB  
PL2W 30.57242203 W  
PL12W 0.36251819 W  
PL13W 0.24284537 W  
SFO2 600.2624010 MHz

F2 - Processing parameters  
SI 32768  
SF 150.9354970 MHz  
WDW no  
SSB 0  
LB 0 Hz  
GB 0  
PC 1.40

<sup>1</sup>H NMR of 6



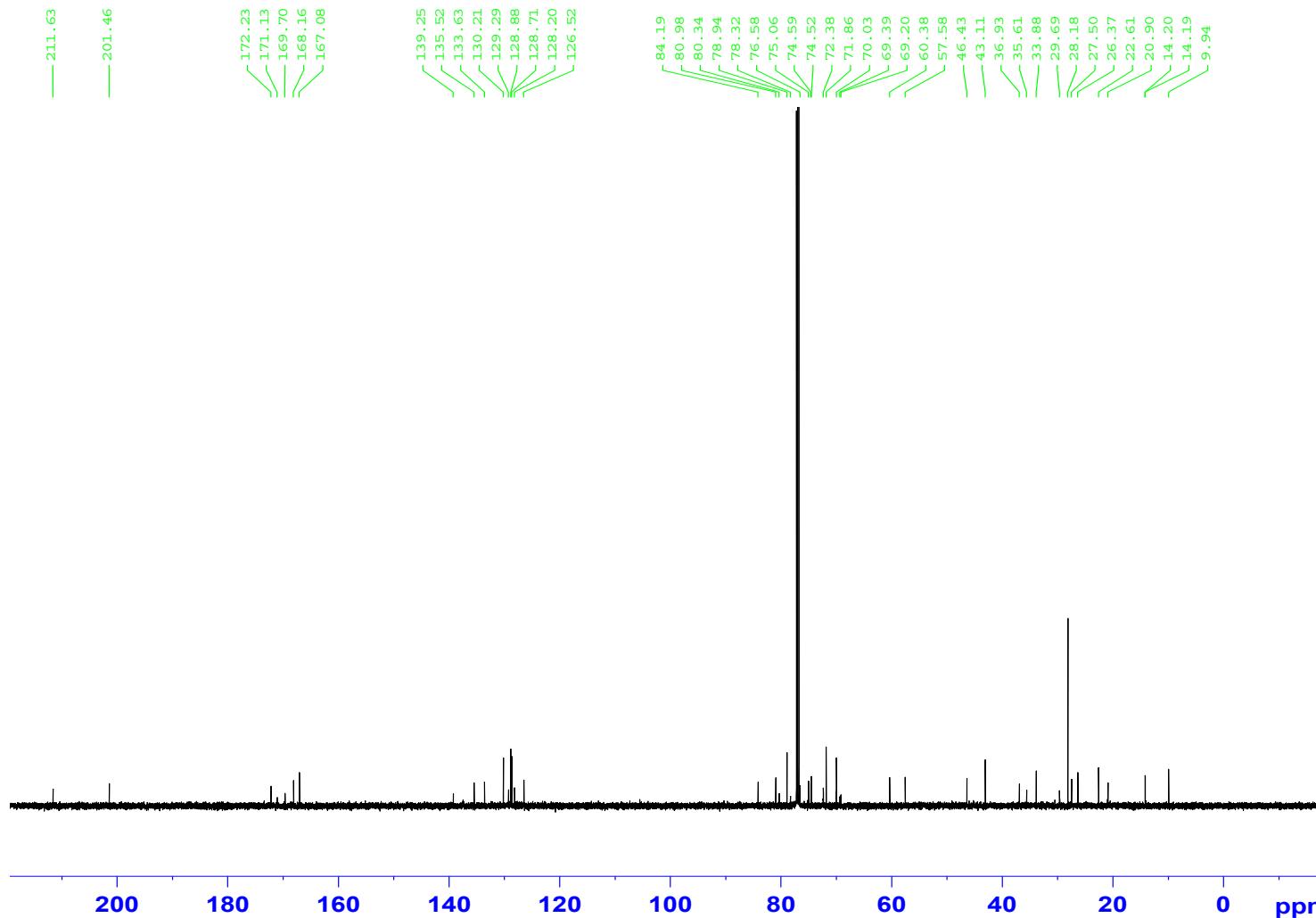
Current Data Parameters  
NAME dp-11-053  
EXPNO 12  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 2011213  
Time 16.00  
INSTRUM spect  
PROBHD 5 mm PABBO BB-  
PULPROG zg30  
TD 65536  
SOLVENT CDCl3  
NS 16  
DS 2  
SWH 12335.526 Hz  
FIDRES 0.188225 Hz  
AQ 2.6564426 sec  
RG 181  
DW 40.533 usec  
DE 8.50 usec  
TE 301.9 K  
D1 1.0000000 sec  
TDO 1

===== CHANNEL f1 ======  
NUC1 1H  
P1 9.40 usec  
PL1 -3.00 dB  
PL1W 30.57242203 W  
SFO1 600.2637069 MHz

F2 - Processing parameters  
SI 32768  
SF 600.2600174 MHz  
WDW EM  
SSB 0 0.30 Hz  
LB 0  
GB PC 1.00

<sup>13</sup>C NMR of **6**



Current Data Parameters  
NAME dp-11-053  
EXPNO 10  
PROCNO 1

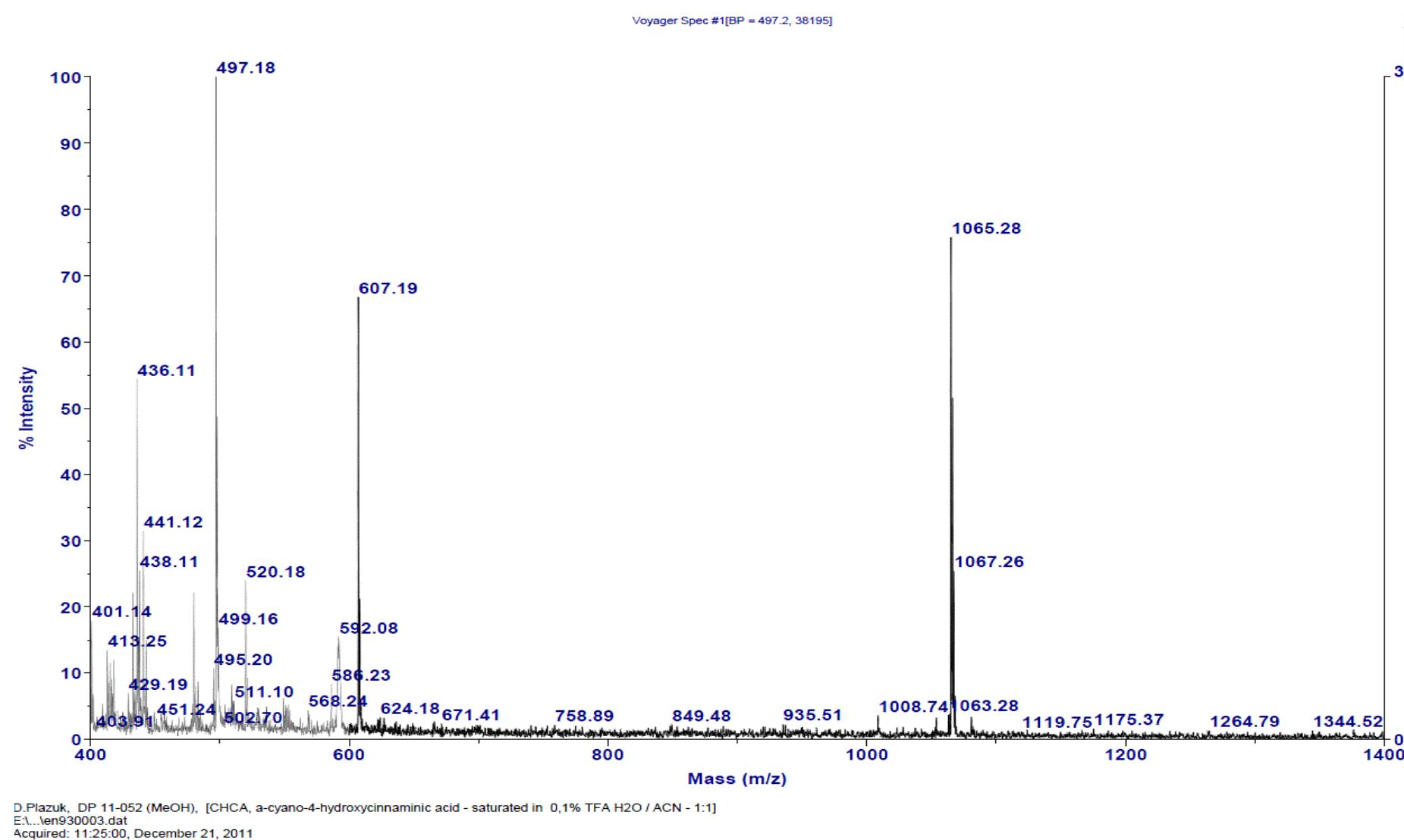
F2 - Acquisition Parameters  
Date\_ 20111213  
Time 15.45  
INSTRUM spect  
PROBHD 5 mm PABBO BB-  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 1024  
DS 4  
SWH 36057.691 Hz  
FIDRES 0.550197 Hz  
AQ 0.9088159 sec  
RG 2050  
DW 13.867 usec  
DE 8.50 usec  
TE 302.0 K  
D1 2.00000000 sec  
D11 0.03000000 sec  
TDO 1

===== CHANNEL f1 =====  
NUC1 13C  
P1 10.50 usec  
PL1 0 dB  
PL1W 91.93504333 W  
SFO1 150.9505906 MHz

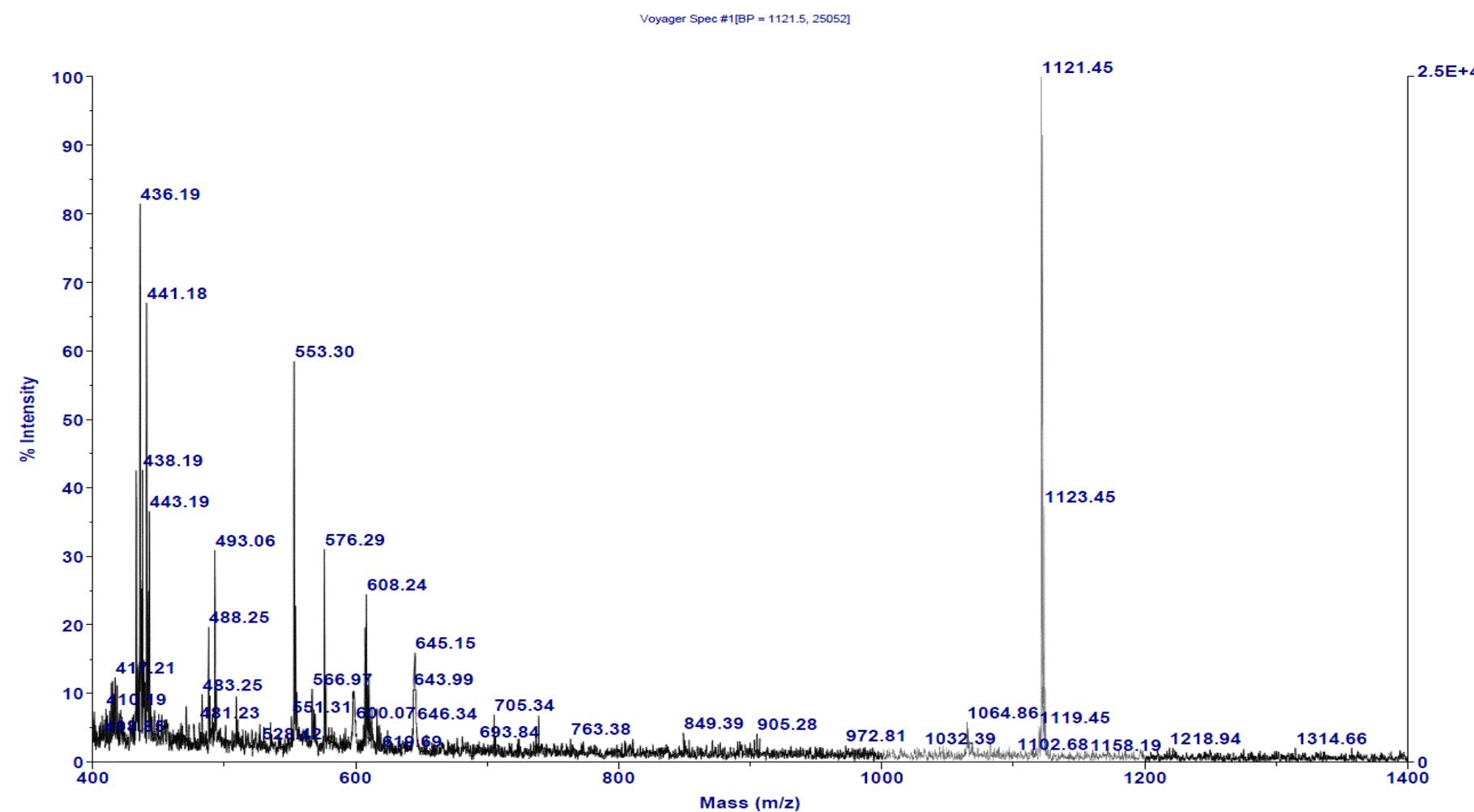
===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 90.00 usec  
PL2 -3.00 dB  
PL12 16.26 dB  
PL13 18.00 dB  
PL2W 30.57242203 W  
PL12W 0.36251819 W  
PL13W 0.24284537 W  
SFO2 600.2624010 MHz

F2 - Processing parameters  
SI 32768  
SF 150.9354970 MHz  
WDW no  
SSB 0 Hz  
LB 0 Hz  
GB 0  
PC 1.40

## MALDI-MS for 3

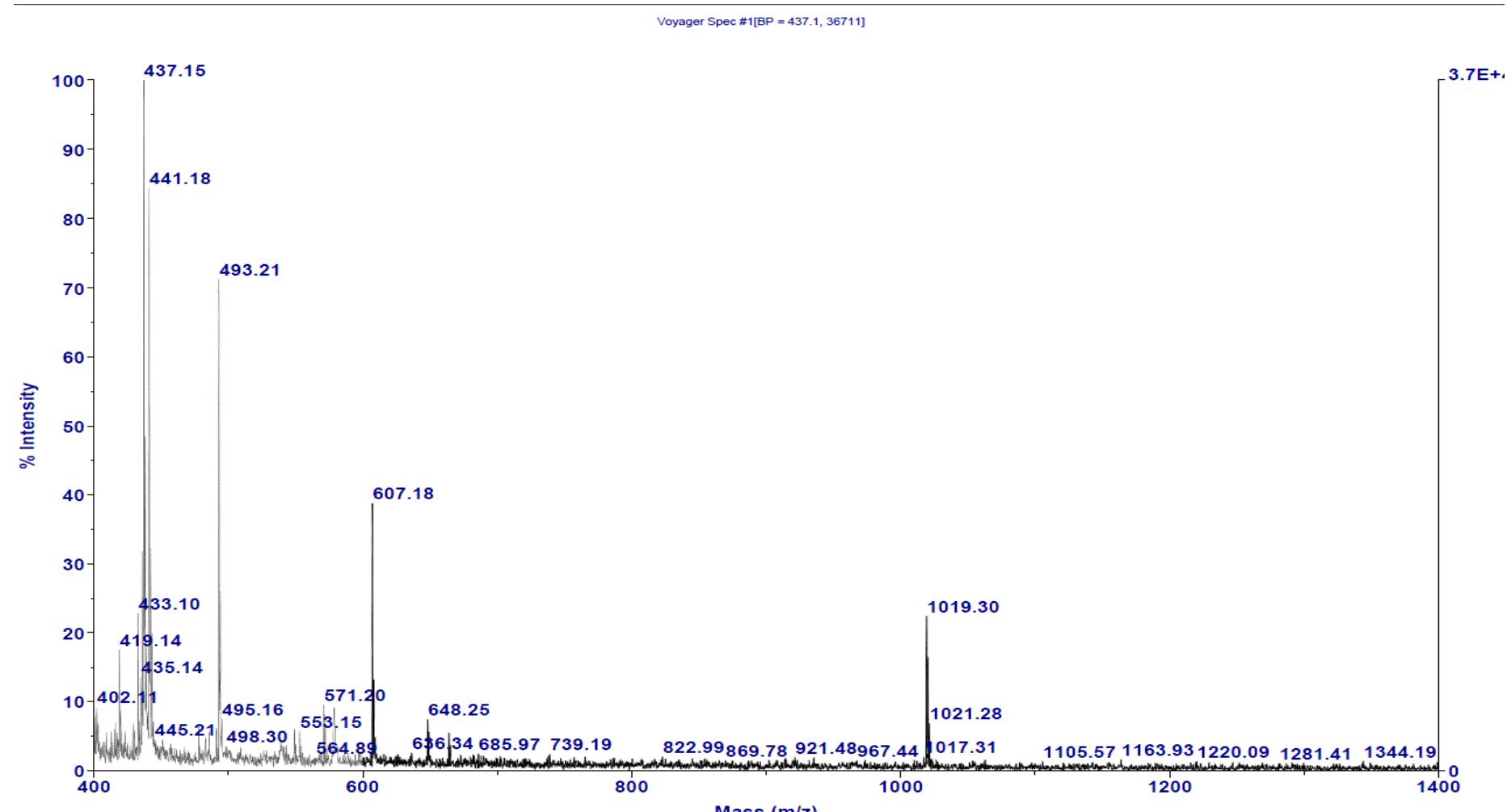


## MALDI-MS for 4



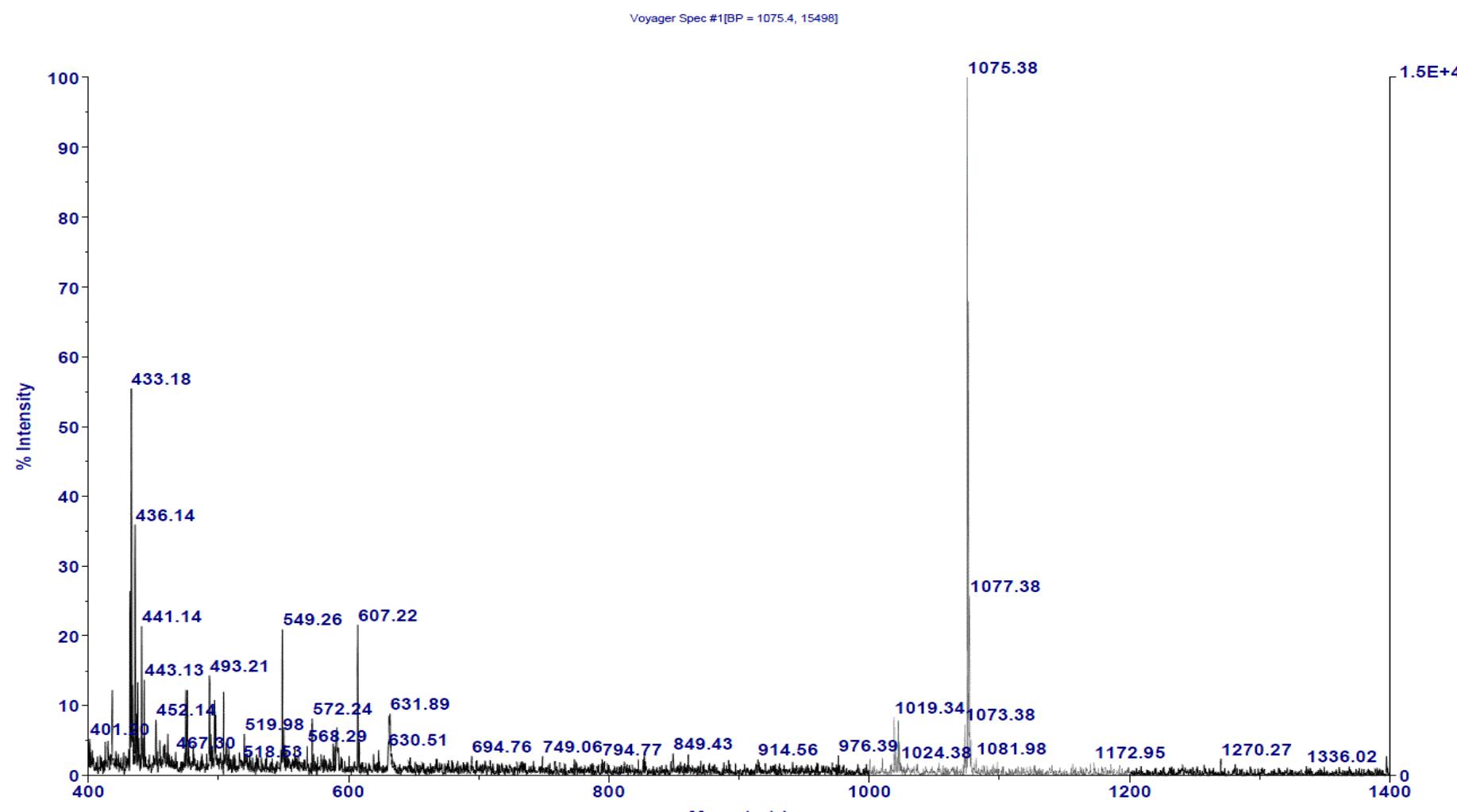
D.Plazuk, DP 1161 (MeOH), [CHCA, a-cyano-4-hydroxycinnamic acid - saturated in 0,1% TFA H<sub>2</sub>O / ACN - 1:1]  
E:\...\Ven950003.dat  
Acquired: 11:20:00, December 21, 2011

## MALDI-MS for 5



D.Plazuk, DP 11-051 (MeOH), [CHCA,  $\alpha$ -cyano-4-hydroxycinnamic acid - saturated in 0.1% TFA H<sub>2</sub>O / ACN - 1:1]  
E:\...len920002.dat  
Acquired: 11:28:00, December 21, 2011

## MALDI-MS for 6



D.Plazuk, DP 11-053 (MeOH), [CHCA, α-cyano-4-hydroxycinnamic acid - saturated in 0.1% TFA H<sub>2</sub>O / ACN - 1:1]  
E:\...len940001.dat  
Acquired: 11:21:00, December 21, 2011