# Boronic acid based photoinduced electron transfer (PET) fluorescence sensors for saccharides

Joseph D. Larkin, Karine A. Frimat, Thomas M. Fyles, Stephen E. Flower and Tony D. James

#### 2-o-Tolyl-[1,3,2] dioxaborinane

To a stirred solution of *o*-tolylboronic acid (2.50 g, 18.40 mmol) in toluene (70 cm<sup>3</sup>) was added 1,3propanediol (2.80 g, 36.80 mmol). The mixture was heated and stirred under Dean and Stark conditions for 1 hour. After cooling, the reaction mixture was washed with water (50 cm<sup>3</sup>) and dried (MgSO<sub>4</sub>). The solvent was removed under reduced pressure affording 2-*o*-tolyl-[1,3,2] dioxaborinane as a dark yellow oil (2.97 g, 91.7 %).  $\delta_{\rm H}$ (400 MHz; CDCl<sub>3</sub>) 2.04-2.05 (2H, m), 2.55 (3H, s), 4.19 (4H, t, *J* 5.4 Hz), 7.16-7.20 (2H, m), 7.29-7.31 (1H, m), 7.32-7.77 (1H, m);  $\delta_{\rm C}$ (100MHz; CDCl<sub>3</sub>) 22.5, 27.5, 61.9, 124.5, 129.8, 134.6, 143.7; *m/z* (EI<sup>+</sup>) 176 (100 %, M<sup>+</sup>).

### 2-(2-Bromomethyl-phenyl)-[1,3,2] dioxaborinane (13)

To a stirred solution of 2-*o*-tolyl-[1,3,2] dioxaborinane (2.94 g, 16.70 mmol) in benzene (60 cm<sup>3</sup>) was added *N*-bromosuccinimide (3.10 g, 17.60 mmol) and AIBN (0.30 g). The mixture was heated and stirred under reflux for 2 hours. The resulting mixture was then cooled on an ice-water-bath and the precipitate was removed by filtration. The filtrate solvent was removed under reduced pressure and the crude product was purified by silica gel chromatography eluting with n-hexane:chloroform (1:5) to afford the dioxaborinane **13** as a yellow oil (2.55 g, 60.1 %), (Found: C, 47.1; H, 4.6. Calc. For C<sub>10</sub>H<sub>12</sub>BBrO<sub>2</sub>: C, 47.1; H, 4.7 %).  $\delta_{\rm H}$ (400 MHz; CDCl<sub>3</sub>) 2.06-2.52 (2H, m), 4.19 (4H, t, *J* 5.4 Hz), 4.92 (2H, s), 7.25-7.92 (4H, m);  $\delta_{\rm C}$ (100 MHz; CDCl<sub>3</sub>) 27.3, 27.5, 62.1, 127.4, 130.0, 130.2, 135.2, 143.2; *m/z* (EI<sup>+</sup>) 253 (1.8 %, [M – H]<sup>+</sup> and 175 (96, [M – Br]<sup>+</sup>).

### 9,10-Bis[(methylamino)methyl]anthracene (27)

Methylamine (7.5 cm<sup>3</sup> of a 2.0 moldm<sup>-3</sup> solution in methanol, 15.00 mmol) was added under argon atmosphere to a solution of 9,10-dialdehyde anthracene **25** (1.17 g, 5.00 mmol) in methanol (20 cm<sup>3</sup>) a stirred round-bottomed flask at room temperature. After stirring overnight, a solution of sodium borohydride (0.93 g, 25.00 mmol) in dry methanol (10 cm<sup>3</sup>) was added in one portion and the reaction mixture was stirred for 4 hours before being poured onto 50 cm<sup>3</sup> of water. The aqueous layer was extracted with dichloromethane (3 x 50 cm<sup>3</sup>). The combined organic extracts were dried (MgSO<sub>4</sub>), filtered, and concentrated under reduced pressure to give the diamine **27** as a yelloworange powder (1.10 g, 83.2 %), mp 139-140°C.  $\delta_{\rm H}(300 \text{ MHz}, \text{CDCl}_3)$  2.58 (6H, s), 4.70 (4H, s), 7.59 (4H, dd, J 6.9 Hz, J 3.3 Hz), 8.41 (4H, dd, J 6.9 Hz, J 3.3 Hz, );  $\delta_{C}$ (75 MHz, CDCl<sub>3</sub>) 37.2, 48.0, 124.9, 125.72, 130.2, 132.0; *m*/*z* (FAB<sup>+</sup>) 265 (32 %, [M + H]<sup>+</sup>) and 234 (100, [M - CH<sub>3</sub>NH]<sup>+</sup>).

## 9,10-Bis[[N-methyl-N-(o-boronobenzyl)amino]methyl]anthracene (1)

To a stirred solution of diamine **27** (0.53 g, 2.00 mmol) in 50 cm<sup>3</sup> of dry acetonitrile was added the benzyl bromide **13** (1.52 g, 6.00 mmol), followed by K<sub>2</sub>CO<sub>3</sub> (1.10 g, 8.00 mmol). The reaction mixture was then allowed to stir under reflux overnight. The acetonitrile was removed under reduced pressure and water (50 cm<sup>3</sup>) was added. The aqueous layer was then extracted with dichloromethane (3 x 50 cm<sup>3</sup>) and the organic layers were combined, dried (MgSO<sub>4</sub>) and then concentrated under reduced pressure to afford crude **1**. The crude material was purified by trituration with ethyl acetate to give **1** as a yellow orange powder (0.30 g, 28.2 %), mp 188-190°C (decomp.).  $\delta_{\rm H}(300 \text{ MHz}; \text{CDCl}_3/\text{ CD}_3\text{OD} 1:1) 2.19$  (6H, s), 3.90 (4H, s), 4.46 (4H, s,), 7.28-8.38 (16H, m); *m*/*z* (FAB<sup>+</sup>) 645 (24 %, [M – 4 × H<sub>2</sub>O + 2 × glycerol + H]<sup>+</sup>) and 205 (100, [CH<sub>2</sub>AnthCH<sub>2</sub> + H]<sup>+</sup>).



**Figure 1S.** Relative fluorescence intensity vs. Saccharide concentration profile of sensor **1** ( $1 \times 10^{-7}$  moldm<sup>-3</sup>) (**n**) D- glucose, (**o**) D-fructose, (**d**) D-galactose; (**v**) D-mannose in 52.1 wt% MeOH pH 8.21 phosphate buffer at 25 °C;  $\lambda_{ex} = 370$  nm,  $\lambda_{em} = 422$  nm.



**Figure 2S.** Relative fluorescence intensity vs. Saccharide concentration profile of sensor **3** ( $1 \times 10^{-7}$  moldm<sup>-3</sup>) (**n**) D- glucose, (**o**) D-fructose, (**d**) D-galactose; (**V**) D-mannose in 52.1 wt% MeOH pH 8.21 phosphate buffer at 25 °C;  $\lambda_{ex} = 271$  nm,  $\lambda_{em} = 339$  nm.



**Figure 3S.** Relative fluorescence intensity vs. Saccharide concentration profile of sensor 4 ( $1 \times 10^{-7}$  moldm<sup>-3</sup>) (**n**) D- glucose, (**o**) D-fructose, (**A**) D-galactose; (**V**) D-mannose in 52.1 wt% MeOH pH 8.21 phosphate buffer at 25 °C;  $\lambda_{ex} = 370$  nm,  $\lambda_{em} = 416$  nm.



**Figure 4S.** Relative fluorescence intensity vs. Saccharide concentration profile of sensor  $5(1 \times 10^{-7} \text{ moldm}^{-3})$  (**n**) D- glucose, (**o**) D-fructose, (**d**) D-galactose; (**v**) D-mannose in 52.1 wt% MeOH pH 8.21 phosphate buffer at 25 °C;  $\lambda_{ex} = 343 \text{ nm}$ ,  $\lambda_{em} = 397 \text{ nm}$ .



**Figure 5S.** Relative fluorescence intensity vs. Saccharide concentration profile of sensor **6** ( $1 \times 10^{-7}$  moldm<sup>-3</sup>) (**n**) D- glucose, (**o**) D-fructose, (**A**) D-galactose; (**V**) D-mannose in 52.1 wt% MeOH pH 8.21 phosphate buffer at 25 °C;  $\lambda_{ex} = 271$  nm,  $\lambda_{em} = 343$  nm.



**Figure 6S.** Relative fluorescence intensity vs. Saccharide concentration profile of sensor 7 ( $1 \times 10^{-7}$  moldm<sup>-3</sup>) (**n**) D- glucose, (**o**) D-fructose, (**d**) D-galactose; (**V**) D-mannose in 52.1 wt% MeOH pH 8.21 phosphate buffer at 25 °C;  $\lambda_{ex} = 370$  nm,  $\lambda_{em} = 417$  nm.



**Figure 7S.** Relative fluorescence intensity vs. Saccharide concentration profile of sensor **8** ( $1 \times 10^{-7}$  moldm<sup>-3</sup>) (**n**) D- glucose, (**o**) D-fructose, (**d**) D-galactose; (**V**) D-mannose in 52.1 wt% MeOH pH 8.21 phosphate buffer at 25 °C;  $\lambda_{ex} = 343$  nm,  $\lambda_{em} = 397$  nm.



**Figure 8S.** Fluorescence Intensity  $(I_F)$  of Sensor **3** with increasing concentration of D-glucose at pH 8.21.



Figure 9S. Fluorescence Intensity  $(I_F)$  of Sensor 3 with increasing concentration of D-galactose at pH 8.21.



**Figure 10S.** Fluorescence Intensity  $(I_F)$  of Sensor **3** with increasing concentration of D-mannose at pH 8.21.



**Figure 11S.** Fluorescence Intensity  $(I_F)$  of Sensor **3** with increasing concentration of D-fructose at pH 8.21.



Figure 12S. CD Spectra of Sensor 3 in the presence of D-or L-glucose.



**Figure 13S.** Fluorescence Intensity  $(I_F)$  of Sensor 4 with increasing concentration of D-glucose at pH 8.21.



Figure 14S. Fluorescence Intensity  $(I_F)$  of Sensor 4 with increasing concentration of D-galactose at pH 8.21.



**Figure 15S.** Fluorescence Intensity  $(I_F)$  of Sensor 4 with increasing concentration of D-mannose at pH 8.21.



**Figure 16S.** Fluorescence Intensity  $(I_F)$  of Sensor 4 with increasing concentration of D-fructose at pH 8.21.



Figure 17S. CD Spectra of Sensor 4 in the presence of D-or L-glucose.



**Figure 18S.** Fluorescence Intensity  $(I_F)$  of Sensor 5 with increasing concentration of D-glucose at pH 8.21.



Figure 19S. Fluorescence Intensity  $(I_F)$  of Sensor 5 with increasing concentration of D-galactose at pH 8.21.



**Figure 20S.** Fluorescence Intensity  $(I_F)$  of Sensor **5** with increasing concentration of D-mannose at pH 8.21.



Figure 21S. Fluorescence Intensity  $(I_F)$  of Sensor 5 with increasing concentration of D-fructose at pH 8.21.



Figure 22S. CD Spectra of Sensor 5 in the presence of D-or L-glucose.



**Figure 23S.** Fluorescence Intensity  $(I_F)$  of Sensor 6 with increasing concentration of D-glucose at pH 8.21.



**Figure 24S.** Fluorescence Intensity  $(I_F)$  of Sensor 6 with increasing concentration of D-galactose at pH 8.21.



**Figure 25S.** Fluorescence Intensity  $(I_F)$  of Sensor 6 with increasing concentration of D-mannose at pH 8.21.



**Figure 26S.** Fluorescence Intensity  $(I_F)$  of Sensor 6 with increasing concentration of D-fructose at pH 8.21.



Figure 27S. CD Spectra of Sensor 6 in the presence of D-or L-glucose.



Figure 28S. CD Spectra of Sensor 6 in the presence of D-galactose.



Figure 29S. Fluorescence Intensity  $(I_F)$  of Sensor 7 with increasing concentration of D-glucose at pH 8.21.



**Figure 30S.** Fluorescence Intensity  $(I_F)$  of Sensor 7 with increasing concentration of D-galactose at pH 8.21.



Figure 31S. Fluorescence Intensity  $(I_F)$  of Sensor 7 with increasing concentration of D-fructose at pH 8.21.



Figure 32S. CD Spectra of Sensor 7 in the presence of D-and L-glucose.



Figure 33S. CD Spectra of Sensor 7 in the presence of D-galactose.



Figure 34S. Fluorescence Intensity  $(I_F)$  of Sensor 8 with increasing concentration of D-glucose at pH 8.21.



**Figure 35S.** Fluorescence Intensity  $(I_F)$  of Sensor 8 with increasing concentration of D-galactose at pH 8.21.



**Figure 36S.** Fluorescence Intensity  $(I_F)$  of Sensor 8 with increasing concentration of D-mannose at pH 8.21.



**Figure 37S.** Fluorescence Intensity  $(I_F)$  of Sensor **8** with increasing concentration of D-fructose at pH 8.21.



Figure 38S. CD Spectra of Sensor 8 in the presence of D-and L-glucose.