## Supporting Information for

## Encapsulation and Controlled Release of L-Leuprolide from Poly(β-

## hydroxyalkanoate)s: Impact of Microstructure and Chemical Functionalities

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**Figure S1-S5.** <sup>13</sup>C $\{^{1}H\}$  NMR and <sup>1</sup>H NMR spectra of *i*-PHB and *i*-P(-HB-*co*-HB<sup>allyl</sup>, 11 mol%)

**Figure S6.** Example of RI detector responses during Gel Permation Chromotography (GPC) of *i*-PHB and *i*-P(-HB-*co*-HB<sup>allyl</sup>, 11 mol%)

**Figure S7.** SEM images of microparticles based on syndiotactic PHAs, prepared by "solvent evaporation" mode

**Figure S8.** Gaussian distributions of microparticles size. A: *s*-P(HB-*co*-HB<sup>allyl</sup>), **B**: *s*-P(HB-*co*-HB<sup>diOH</sup>), **C**: *s*-P(HB-*co*-HB<sup>allyl</sup>)/LeA, **D**: *s*-P(HB-*co*-HB<sup>diOH</sup>)/LeA.

**Figure S9.** SEM images of polymer microparticles based on isotactic PHAs, prepared by the "solvent evaporation" method.

Figures S10-S11. DSC traces of *i*-PHB and of *i*-PHB/LeA microparticles

**Figures S12-S15.** Viscoelastic properties of (co)polymers and (co)polymers encapsulated with LeA: storage modulus G' and loss modulus G" as a function of frequency

**Figure S16.** Effect of the molecular weight on the release profiles of LeA from LeA-loaded *s*-P(-HB<sup>diOH</sup>, 11 mol% diOH) prepared by "co-precipitation" method

Entry	[BL <sup>R</sup> ]	(co)polymer [BL <sup>R</sup> ] <sup>b</sup> in copolymer (mol%)	$M_{n exp}^{c}$ (× 10 <sup>3</sup> g/mol)	$M_{ m w}/M_{ m n}^{ m c}$	$P_r/P_m^d$
1	rac	s-P(HB-co-HB <sup>allyl</sup> )	9.2	1.24	84/16
2	rac	$s-P(HB-co-HB^{diOH})$	11.2	1.28	82/18
3	rac	$s-P(HB-co-HB^{allyl})$	33.5	1.50	84/16
4	rac	$s-P(HB-co-HB^{diOH})$	38.4	1.68	nd
5	rac	$s-P(HB-co-HB^{allyl})$	61.2	1.62	82/18
6	rac	$s-P(HB-co-HB^{diOH})$	69.6	1.76	nd
7	rac	$a-P(HB-co-HB^{allyl})$	25.4	1.45	55/45
8	rac	a-P(HB- $co$ -HB <sup>diOH</sup> )	21.0	1.52	53/47
9	R	i-P(HB- $co$ -HB <sup>allyl</sup> )	30.1	1.30	nd
10	R	i-P(HB- $co$ -HB <sup>diOH</sup> )	35.1	1.41	nd
11	R	<i>i</i> -P(HB)	4.2	1.06	03/97
12	R	0 <i>i</i> -P(HB)	8.5	1.13	04/96
13	R	0 <i>i</i> -P(HB)	15.9	1.11	04/96
14	R	0 <i>i</i> -P(HB)	28.0	1.13	05/95

**Table S1.** Homo- and copolymerization of β-butyrolactone and *rac*-allyl-butyrolactone.<sup>a</sup>

<sup>[a]</sup>  $[BL^R] = 2.44 \text{ mol/L}$ ; polymerization carried out at 20 °C; the results shown for each entry are representative of at least two reproducible (± 5%) runs. <sup>[b]</sup> Comonomer content determined by <sup>1</sup>H NMR in CDCl<sub>3</sub>. <sup>[c]</sup> Average-number molecular weight in g.mol<sup>-1</sup> (uncorrected) determined by GPC vs. PS standards. <sup>[d]</sup>  $P_r$  is the probability of racemic linkages between monomer units and is determined by <sup>13</sup>C{<sup>1</sup>H} NMR spectroscopy.



**Figure S1.** <sup>13</sup>C{<sup>1</sup>H} NMR spectrum (125 MHz, CDCl<sub>3</sub>, 40 °C) of a highly isotactic PHB ( $P_m = 0.95$ ) sample prepared by ROP of *R*-BL<sup>Me</sup> with complex **1**.



**Figure S2.** Methyl, Methylene, and carbonyl regions of the <sup>13</sup>C{<sup>1</sup>H} NMR spectrum (125 MHz, CDCl<sub>3</sub>, 40 °C) of a highly isotactic PHB ( $P_m = 0.95$ ) sample prepared by ROP of *R*-BL<sup>Me</sup> with complex **1**.



**Figure S3.** <sup>1</sup>H NMR spectrum (500 MHz, CDCl<sub>3</sub>, 20 °C) of an isotactic-rich ( $P_m = 0.85$ ) *i*-P(HB*co*-HB<sup>allyl</sup>, 11 mol%) sample prepared by ROP of *R*-BL<sup>Me</sup> and *rac*-BL<sup>allyl</sup> with complex **1** (Table S1, entry 9).



**Figure S4.** <sup>13</sup>C{<sup>1</sup>H} NMR spectrum (125 MHz, CDCl<sub>3</sub>, 40 °C) of an isotactic-rich ( $P_m = 0.85$ ) *i*-P(HB-*co*-HB<sup>allyl</sup>, 11 mol%) sample prepared by ROP of *R*-BL<sup>Me</sup> and *rac*-BL<sup>allyl</sup> with complex **1** (Table S1, entry 9).



**Figure S5.** Methyl, Methylene, and carbonyl regions of the <sup>13</sup>C{<sup>1</sup>H} NMR spectrum (125 MHz, CDCl<sub>3</sub>, 40 °C) of an isotactic-rich ( $P_{\rm m} = 0.85$ ) *i*-P(HB-*co*-HB<sup>allyl</sup>, 11 mol%) sample prepared by ROP of *R*-BL<sup>Me</sup> and *rac*-BL<sup>allyl</sup> with complex **1**(Table S1, entry 9).



**Figure S6.** RI detector responses during Gel Permeation Chromotography (GPC) (THF, 25 °C) of an isotactic-rich ( $P_{\rm m} = 0.85$ ) *i*-P(HB-*co*-HB<sup>allyl</sup>, 11 mol%) sample prepared by ROP of *R*-BL<sup>Me</sup> and *rac*-BL<sup>allyl</sup> with complex **1**(Table S1, entry 9).



**Figure S7.** SEM images of microparticles prepared by the "solvent evaporation" method, the size of the bar is 10-100  $\mu$ m. A: *s*-P(HB-*co*-HB<sup>allyl</sup>, 11 mol% allyl) (Table S1, entry 3). B: *s*-P(HB-*co*-HB<sup>allyl</sup>, 11 mol% allyl) (Table S1, entry 3) containing 7 wt% of LeA. C: *s*-P(HB-*co*-HB<sup>diOH</sup>, 11 mol% diOH) (Table S1, entry 4). D: *s*-P(HB-*co*-HB<sup>diOH</sup>, 11 mol% diOH) (Table S1, entry 4). D: *s*-P(HB-*co*-HB<sup>diOH</sup>, 11 mol% diOH) (Table S1, entry 4).

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**Figure S8.** Gaussian distributions of microparticles size. A: *s*-P(HB-*co*-HB<sup>allyl</sup>), **B**: *s*-P(HB-*co*-HB<sup>diOH</sup>), **C**: *s*-P(HB-*co*-HB<sup>allyl</sup>)/LeA, **D**: *s*-P(HB-*co*-HB<sup>diOH</sup>)/LeA.



**Figure S9.** SEM images of polymer microparticles prepared by the "solvent evaporation" method, the size of the bar is 50-100  $\mu$ m. A: *i*-PHB (Table S1, entry 14). B: *i*-PHB (Table S1, entry 14) containing 7 wt% of LeA. C: *i*-P(HB-*co*-HB<sup>allyl</sup>, 11 mol% allyl) (Table S1, entry 9). D: *i*-P(HB-*co*-HB<sup>diOH</sup>, 11 mol% diOH) (Table S1, entry 10).



**Figure S10.** DSC trace (run from 23 °C to 200 °C at 10 °C/min) of *i*-PHB prepared by ROP of R-BL<sup>Me</sup> with complex **1** (Table S1, entry 14).



**Figure S11.** DSC trace (run from 23 °C to 200 °C at 10 °C/min) of *i*-PHB (Table S1, entry 14)/LeA microparticles.



**Figure S12.** Storage modulus G' as a function of frequency of s-P(HB-co-HB<sup>allyl</sup> (11 mol% allyl) and s-P(HB-co-HB<sup>diOH</sup> (11 mol% diOH) in 10 wt% chloroform solutions



**Figure S13.** Loss modulus G" as a function of frequency of *s*-P(HB-*co*-HB<sup>allyl</sup> (11 mol% allyl) and *s*-P(HB-*co*-HB<sup>diOH</sup> (11 mol% diOH) in 10 wt% chloroform solutions.

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**Figure S14.** Storage modulus G' as a function of frequency of s-P(HB-co-HB<sup>diOH</sup> (11 mol% diOH) and s-P(HB-co-HB<sup>diOH</sup> (11 mol% diOH) encapsulated with LeA (7 wt%), in 10wt% chloroform solutions.



**Figure S15.** Loss modulus G'' as a function of frequency of s-P(HB-co-HB<sup>diOH</sup> (11 mol% diOH) and s-P(HB-co-HB<sup>diOH</sup> (11 mol% diOH) encapsulated with LeA (7 wt%) in 10 wt% chloroform solutions.



**Figure S16.** Effect of the molecular weight on the release profiles of LeA from LeA-loaded *s*-P(HB-*co*-HB<sup>diOH</sup>, 11 mol% diOH) prepared by "co-precipitation" method,.  $\Delta M_n = 11\ 200\ \text{g.mol}^{-1}$ , (Table S1, entry 2);  $\blacksquare M_n = 38\ 400\ \text{g.mol}^{-1}$ , (Table S1, entry 4);  $\Box M_n = 69\ 600\ \text{g.mol}^{-1}$ , (Table S1, entry 6).