

Electronic Supplementary Material (ESI) for New Journal of Chemistry

**Assembly of indole fluorophore in situ for hydrogen sulfide signaling through substrate triggered intramolecular reduction/cyclization cascade: A sensitive and selective probe in aqueous solution**

**Ji Zhou, Yuanyuan Luo, Qiang Li, Jiaoning Shen, Rui Wang, Yufang Xu\* and Xuhong Qian\***

Shanghai Key Laboratory of Chemical Biology, State Key Laboratory of Bioreactor Engineering, School of Pharmacy, East China University of Science and Technology, Shanghai, 200237, China.

Fax: +86 21 6425 2603; Tel: +86 21 6425 3589; E-mail: [xhqian@ecust.edu.cn](mailto:xhqian@ecust.edu.cn); [yfxu@ecust.edu.cn](mailto:yfxu@ecust.edu.cn)

**Table of Contents:**

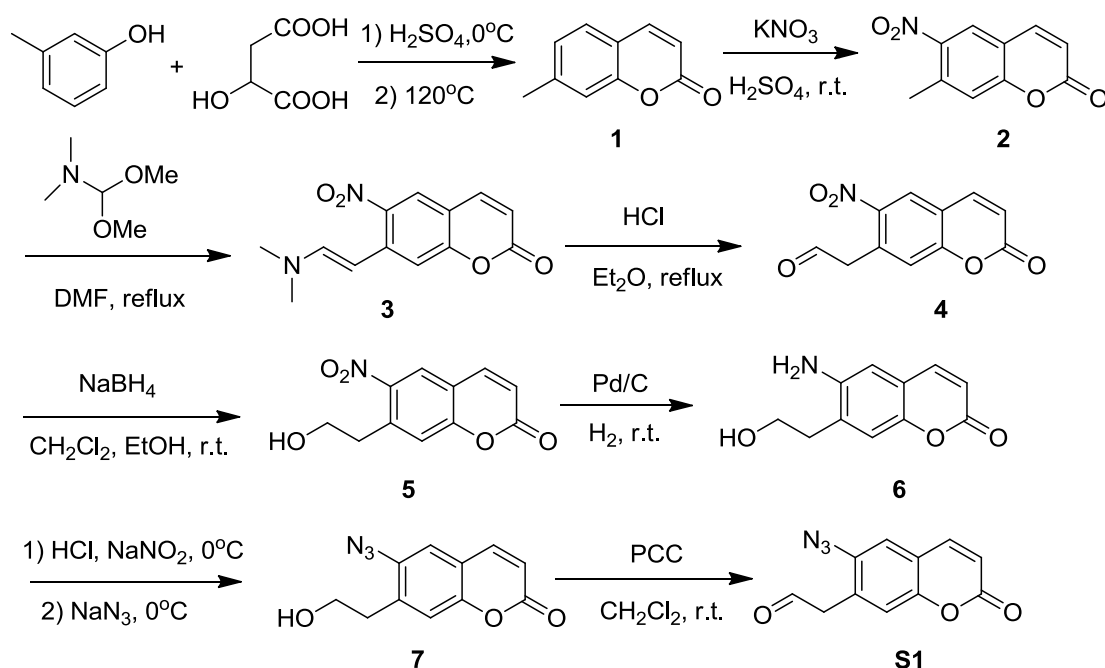
<b>1. Materials and instruments</b> .....	2
<b>2. Synthesis</b> .....	2
<b>3. Methods and data</b> .....	5
<b>4. NMR and MS spectra</b> .....	8
<b>5. References:</b> .....	17

## 1. Materials and instruments

**Materials:** All chemical reagents were purchased from Aladdin Corporation and J&K Corporation without further purification. All the organic solvents were of analytical grade. DMF was distilled using CaH<sub>2</sub> to remove the water before used. Thin-layer chromatography was performed on silica gel plates. Silica gel P60 (Qingdao, 300-400 mesh) was used for column chromatography. Water was purified by a Milli-Q system.

**Instruments:** Fluorescence spectra were collected by a Varian Cary Eclipse Fluorescence Spectrometer. Absorption spectra were recorded by a Varian Cary 100 UV-Vis spectrophotometer. <sup>1</sup>H-NMR and <sup>13</sup>C-NMR spectra were taken in CDCl<sub>3</sub> and DMSO-*d*<sub>6</sub> at 25 °C on a Bruker AV-400 spectrometer in NMR Facility of East China University of Science and Technology (ECUST). The chemical shifts were reported in ppm (TMS as internal standard). Mass spectra were performed in the Analysis Center of East China University of Science and Technology (ECUST).

## 2. Synthesis



Scheme S1 The synthesis of probe S1

### Synthesis of 7-methylcoumarin (compound 1)<sup>1</sup>

To a solution of 2-hydroxysuccinic acid (25.40 g, 0.19 mol) in m-cresol (20 mL, 0.19 mol) at 0 °C was added concentrated sulfuric acid (50 mL) dropwise. The reaction mixture was stirred at room temperature for 0.5 h, then heated up to 120 °C for 3 h until there was no bubble out. After cooling down to room temperature, the mixture was poured onto ice. The product was

extracted with EtOAc, washed with brine and dried over Na<sub>2</sub>SO<sub>4</sub>. After the solvent was removed in vacuo, the residue was recrystallized with 95% ethanol to give the title compound (12 g, 40%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.69 (d, *J* = 9.6 Hz, 1H), 7.38 (d, *J* = 7.6 Hz, 1H), 7.16 (s, 1H), 7.11 (d, *J* = 7.6 Hz, 1H), 6.37 (d, *J* = 9.6 Hz, 1H), 2.47 (s, 3H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 161.08, 154.12, 143.46, 143.09, 127.57, 125.62, 116.96, 116.45, 115.35, 21.75.

#### **Synthesis of 7-methyl-6-nitrocoumarin (compound 2)<sup>2</sup>**

To a solution of compound 1 (9.31 g, 58.2 mmol) in concentrated sulfuric acid (60 mL) at 0 °C was added KNO<sub>3</sub> (7.08 g, 69.8 mmol) portionwise. The reaction mixture was stirred at 0 °C for 2 h, and poured onto ice. The mixture was filtered and the filter cake was washed with water (3×900 mL). The filter cake was dried off to give the title compound (8.4 g, 70%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>): δ 8.54 (s, 1H), 8.16 (d, *J* = 9.6 Hz, 1H), 7.56 (s, 1H), 6.63 (d, *J* = 9.6 Hz, 1H), 2.63 (s, 3H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>): δ 159.58, 156.02, 145.24, 143.53, 138.09, 125.84, 120.52, 117.88, 117.72, 20.66.

#### **Synthesis of 7-formylmethyl-6-nitrocoumarin (compound 4)<sup>2</sup>**

Compound 2 (6 g, 28.8 mmol) and N, N-Dimethylformamidedimethyl acetal (7.2 mL, 51.84 mmol) were refluxed under Ar in 60 mL of anhydrous DMF for 2 h. The reaction mixture was concentrated in vacuo to get the crude product 3, 60 mL of Et<sub>2</sub>O and 60 mL of 3 N aq. HCl were added and the mixture was refluxed for 2 h. 400 mL of CH<sub>2</sub>Cl<sub>2</sub> and 400 mL of water were then added. The organic layer was washed with brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and concentrated in vacuo. The residue was purified by silica gel flash chromatography with 1:3 PE / CH<sub>2</sub>Cl<sub>2</sub> to give the title compound (2.1 g, 30%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>): δ 9.75 (s, 1H), 8.64 (s, 1H), 8.20 (d, *J* = 9.6 Hz, 1H), 7.57 (s, 1H), 6.67 (d, *J* = 9.6 Hz, 1H), 4.35 (s, 2H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>): δ 198.7, 159.4, 156.2, 145.1, 143.4, 133.6, 126.1, 121.6, 118.7, 118.5, 48.2.

#### **Synthesis of 7-hydroxyethyl-6-nitrocoumarin (compound 5)<sup>2</sup>**

To a solution of compound 4 (1.5 g, 6.4 mmol) in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (50 mL) at 0 °C was added NaBH<sub>4</sub> (242 mg, 6.4 mmol) suspension in EtOH (45 mL) dropwise over 30 min. The reaction was completed in 5 min, and quenched with 1N HCl. The solution was concentrated in vacuo. Then CH<sub>2</sub>Cl<sub>2</sub> and water were added. The organic layer was washed with brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and concentrated in vacuo. The residue was purified by silica gel flash chromatography with 1:20 EtOAc/ CH<sub>2</sub>Cl<sub>2</sub> to give the title compound (902 mg, 60%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>): δ 8.43 (s, 1H), 8.13 (d, *J* = 9.6 Hz, 1H), 7.52 (s, 1H), 6.62 (d, *J* = 9.6 Hz, 1H), 4.82 (s, 1H), 3.68 (t, *J* = 6.4 Hz, 2H), 3.10 (t, *J* = 6.4 Hz, 2H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>): δ 159.55, 155.62, 146.05, 143.39, 138.99, 125.50, 120.38, 118.01, 117.81,

61.07, 35.95.

### **Synthesis of 7-hydroxyethyl-6-aminocoumarin (compound 6)<sup>2</sup>**

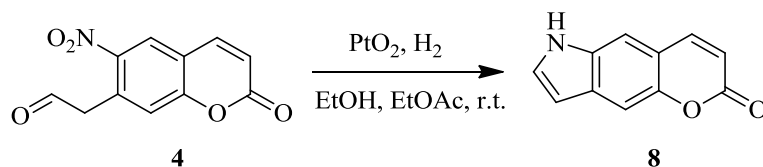
Compound 5 (900 mg, 3.8 mmol) was dissolved in 50 mL of MeOH / EtOAc (1:1). The mixture was hydrogenated at 1 atm of H<sub>2</sub> using Pd/C (45 mg) as the catalyst. After completion of the reaction as evidence by TLC, the solution was filtered on diatomite and the diatomite cake was washed with methanol. Then the filtrate was concentrated in vacuo to give the title compound (624 mg, 80%) without any further purification. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>): δ 7.87 (d, *J* = 9.2 Hz, 1H), 7.04 (s, 1H), 6.81 (s, 1H), 6.31 (d, *J* = 9.2 Hz, 1H), 5.06 (s, 2H), 3.66 (br, 2H), 2.70 (t, *J* = 6.8 Hz, 2H).

### **Synthesis of 7-hydroxyethyl-6-azidecoumarin (compound 7)<sup>3</sup>**

To a 0 °C solution of compound 6 (400 mg, 1.95 mmol) in EtOAc (1 mL) and water (1 mL) was added concentrated hydrochloric acid (2 mL). The mixture was stirred for 15 min. To this solution was added a solution of sodium nitrite (245 mg, 3.51 mmol) in water (0.74 mL) over 5 min. After completion of the addition, the reaction was stirred for an additional 1.5 h. A solution of sodium azide (230 mg, 3.51 mmol) in water (0.7 mL) was added over 5 min. After 1 h, the reaction mixture was diluted with water (100 mL), extracted with ethyl acetate (2×50 mL). The combined organic layer was washed with dilute sodium hydroxide solution, then with brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated in vacuo. The residue was purified by silica gel flash chromatography with 1:30 EtOAc/ CH<sub>2</sub>Cl<sub>2</sub> to give the title compound (225 mg, 50%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>): δ 8.07 (d, *J* = 9.6 Hz, 1H), 7.67 (s, 1H), 7.32 (s, 1H), 6.50 (d, *J* = 9.6 Hz, 1H), 4.73 (t, *J* = 5.2 Hz, 1H), 3.61 (q, *J* = 6.4 Hz, 2H), 2.77 (t, *J* = 6.8 Hz, 2H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>): δ 160.32, 150.89, 143.75, 135.91, 134.90, 118.99, 118.40, 117.77, 116.78, 60.70, 34.81. HRMS (EI): Calcd for C<sub>11</sub>H<sub>9</sub>N<sub>3</sub>O<sub>3</sub> 231.0644; Found, 231.0646.

### **Synthesis of 7-formylmethyl-6-azidecoumarin (compound S1)<sup>4</sup>**

To a solution of compound 7 (231 mg, 1 mmol) in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (30 mL) was added PCC (323 mg, 1.5 mmol). Then the mixture was stirred at room temperature for 4 h and filtered on diatomite. Then the diatomite cake was washed with CH<sub>2</sub>Cl<sub>2</sub>. To the filtrate was added water (100 mL). The organic layer was washed with brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, and concentrated in vacuo. The residue was purified by silica gel flash chromatography with 1:2 PE/ CH<sub>2</sub>Cl<sub>2</sub> to give the title compound (45 mg, 20%). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 9.78 (s, 1H), 7.70 (d, *J* = 9.6 Hz, 1H), 7.27 (s, 1H), 7.20 (s, 1H), 6.50 (d, *J* = 9.6 Hz, 1H), 3.81 (br, 2H). <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 196.90, 160.04, 150.96, 141.93, 135.55, 128.71, 120.09, 119.06, 117.86, 116.19, 46.11. HRMS (EI): Calcd for C<sub>11</sub>H<sub>7</sub>N<sub>3</sub>O<sub>3</sub> 229.0487; Found, 229.0486.



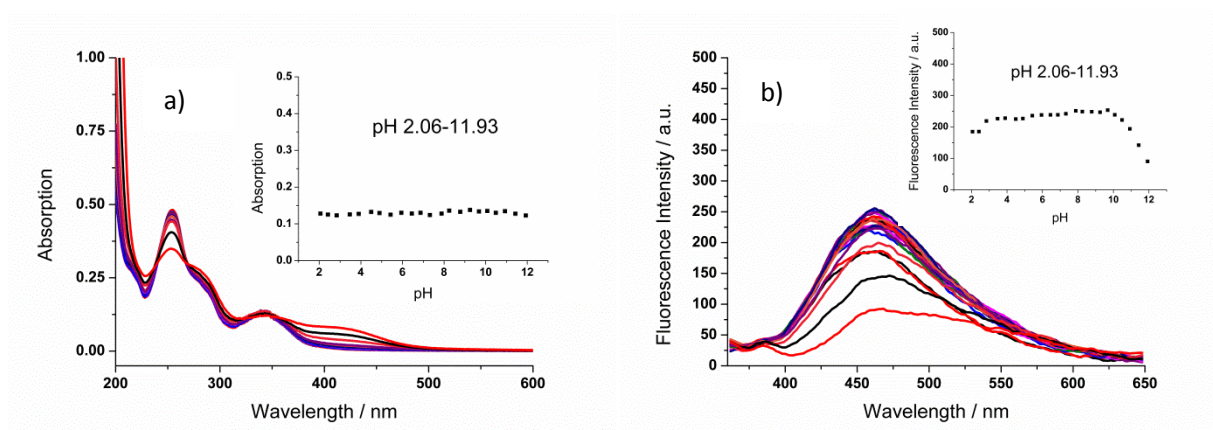
Scheme S2 The synthesis of compound 8

### Synthesis of compound 8<sup>2</sup>

To a solution of compound 4 (300 mg, 1.3 mmol) in 20 mL of EtOH / EtOAc (1:1) was added PtO<sub>2</sub> (20 mg, 0.09 mmol) as the catalyst. The mixture was hydrogenated at 1 atm of H<sub>2</sub> for 4h. The reaction mixture was concentrated in vacuo. The residue was purified by silica gel flash chromatography with 1:5 PE/ CH<sub>2</sub>Cl<sub>2</sub> to give the title compound (168 mg, 69%). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>): δ 11.53 (s, 1H), 8.15 (d, *J* = 9.6 Hz, 1H), 7.75 (s, 1H), 7.65 (s, 1H), 7.53 (s, 1H), 6.56 (s, 1H), 6.32 (d, *J* = 9.6 Hz, 1H). <sup>13</sup>C NMR (100 MHz, DMSO-*d*<sub>6</sub>): δ 161.45, 147.91, 146.05, 133.55, 131.13, 130.98, 114.23, 113.30, 110.72, 105.79, 101.91.

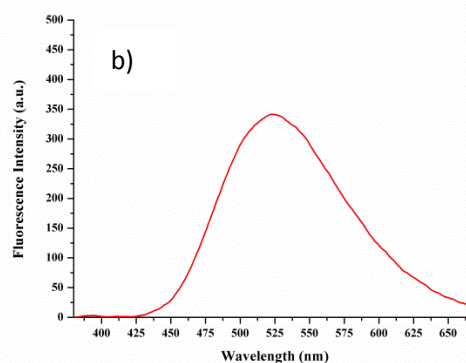
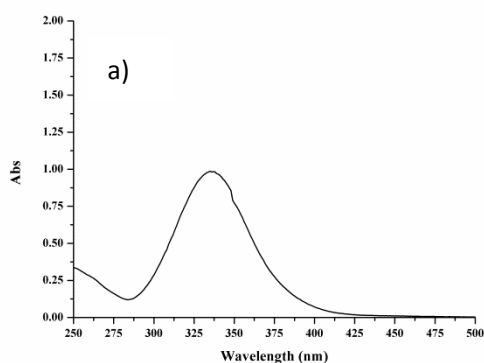
### 3. Methods and data

#### (1) The pH titration



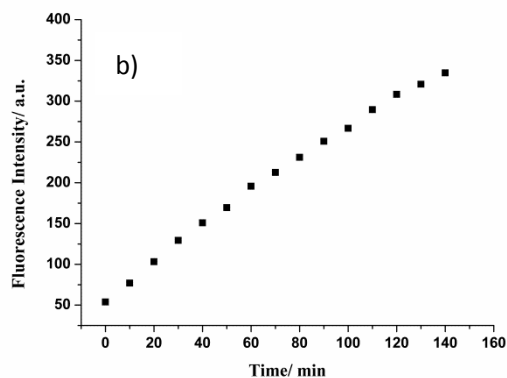
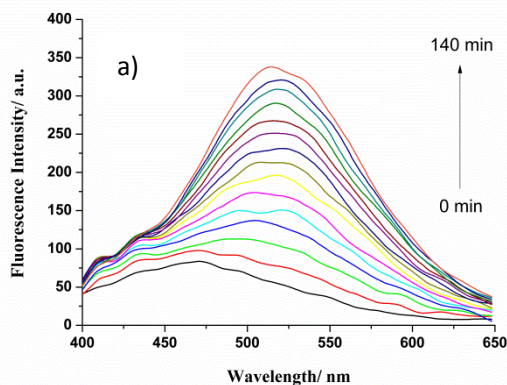
**Fig.S1** (a) Absorption response of **S1** (30 μM) to various pH in water with 5% CH<sub>3</sub>CN as the co-solvent. The insert figure was about the absorption responses of the probe **S1** to different pH. (b) Fluorescence response of **S1** (30 μM) to various pH in water with 5% CH<sub>3</sub>CN as the co-solvent. The insert figure was about the fluorescence responses of the probe **S1** to different pH. The excitation wavelength was 340 nm. pH 2.06-11.93, slit:10 nm, 10 nm.

#### (2) The absorption and emission spectrum of the reaction product compound 8



**Fig.S2** (a) The absorption spectrum of the reaction product compound **8**. (b) The emission spectrum of compound **8** (60  $\mu\text{M}$ ) in the water. The excitation wavelength was 340 nm. Slit: 5 nm, 10 nm.

(3) Fluorescence changes of **S1** to different concentrations of  $\text{NaSH}$  in the water (with 5%  $\text{CH}_3\text{CN}$ ).

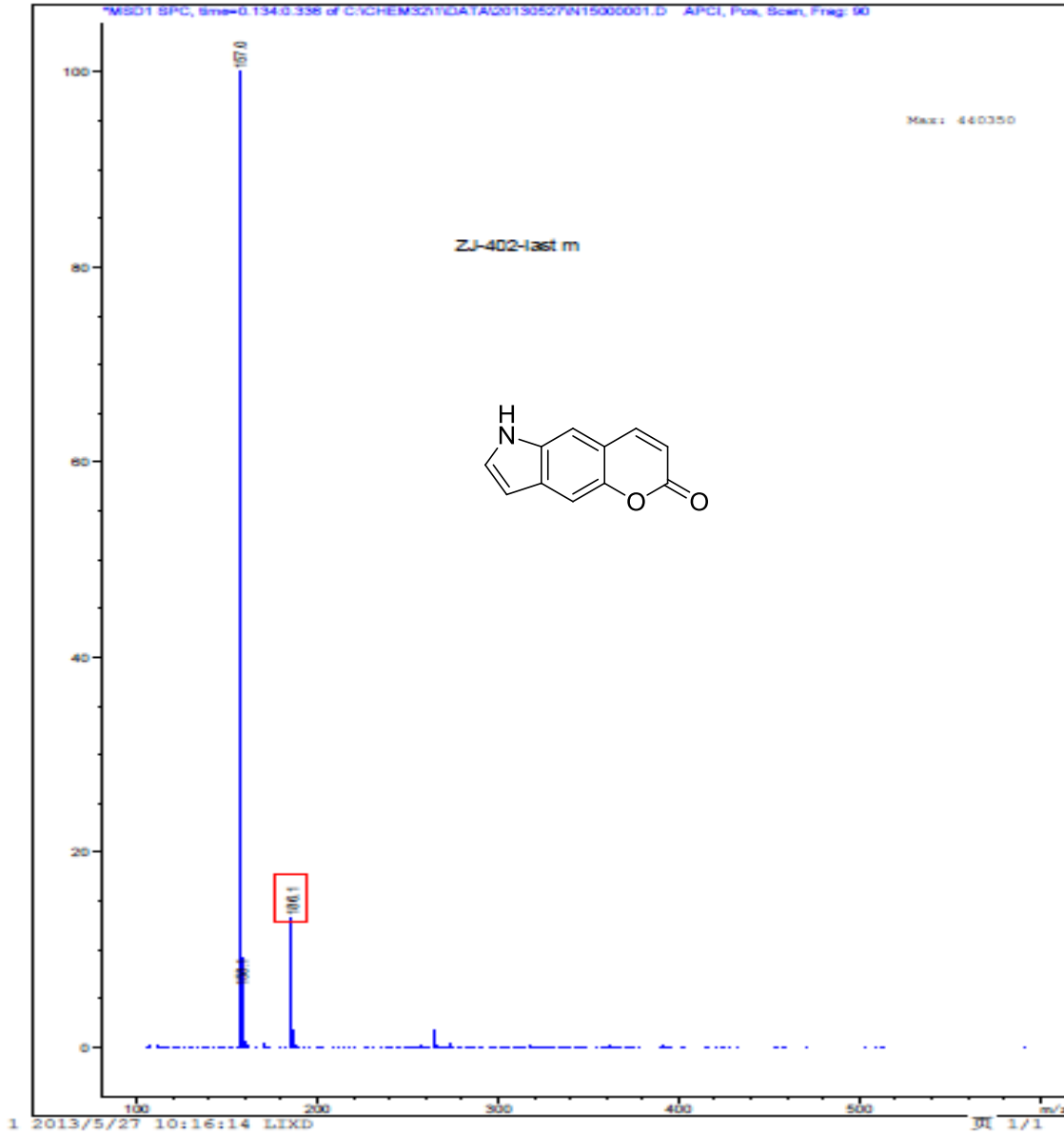
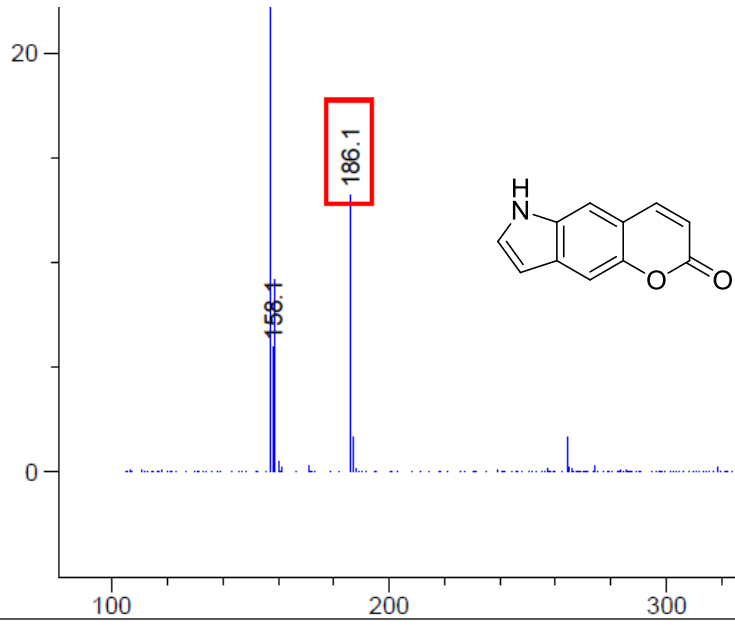


**Fig.S3** (a) The fluorescence responses of **S1** (15  $\mu\text{M}$ ) with  $\text{H}_2\text{S}$  (50  $\mu\text{M}$ ) in the water with 5%  $\text{CH}_3\text{CN}$  as a co-solvent at 25 $^\circ\text{C}$ . The data were recorded every 10 min. (b) Fluorescence responses ( $I_{520\text{ nm}}$ ) of **S1** (15  $\mu\text{M}$ ) to  $\text{H}_2\text{S}$  (50  $\mu\text{M}$ ). Excitation wavelength was 340 nm. Slit: 10 nm, 10 nm.

(4) The detection limit

The concentration of  $\text{H}_2\text{S}$  ranges from nano- to millimolar levels in the blood. The detection limit was calculated according to the previous literature<sup>5</sup>. The fluorescence intensity of **S1** was measured by ten times and the standard deviation was calculated. The fluorescence intensity at 520 nm was plotted as a concentration of  $\text{H}_2\text{S}$ . By using detection limit  $3\sigma/k$ , the detection limit was calculated as 0.22  $\mu\text{M}$ .  $\sigma$  is the standard deviation of the fluorescence intensity of **S1**,  $k$  is the slope between the fluorescence intensity at 520 nm versus the  $\text{H}_2\text{S}$  concentration.

(5) MS (ESI) of the product resulting from the reaction between the probe **S1** and  $\text{H}_2\text{S}$ .



The reaction was performed in CH<sub>3</sub>OH and water.

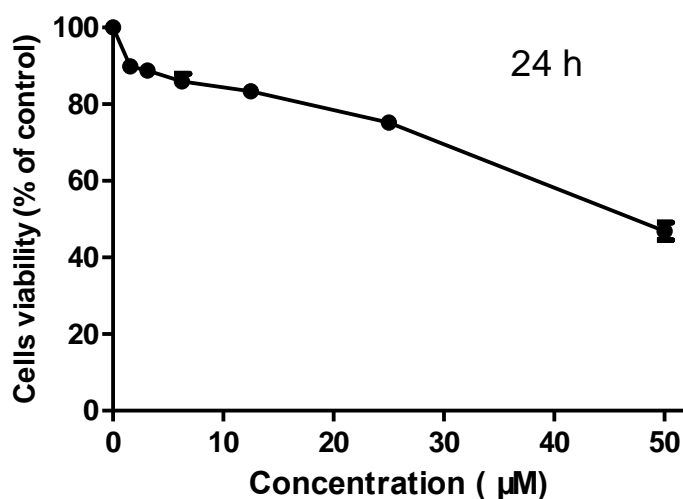
#### (6) Cell Culture and Imaging

HeLa cells were obtained from American Type Culture collection, and grown in Dulbecco's modification of Eagle's medium Dulbecco (DMEM/high: with 4500 mg/L Glucose, 4.0 mM L-Glutamine, and 110 mg/L Sodium Pyruvate) supplemented with 10% foetal bovine serum (FBS). Cells were incubated in a 5% CO<sub>2</sub> humidified incubator at 37 °C and typically passaged with sub-cultivation ratio of 1:4 every two days.

For fluorescence microscopy, HeLa cells were seeded in 24-well culture plate for one night. The cells were first incubated with **S1** (15 µM) for 20 min at 37 °C and washed with phosphate buffer (pH 7.4). Then the cells were treated with or without NaHS (250 µM) for another 40 min at 37 °C and washed with phosphate buffer (pH 7.4) for three times. Fluorescence imaging was performed with Nikon Ti-S with Xenon lamp. Exposure time is 20 s for green emission.

#### (7) Cell viability assay

MCF-7 cells (1×10<sup>4</sup>) were seeded into a 96-well plate and treated with various concentrations of **S1** for 24 h. Cells were irradiated under light emitted from a 100-watt quartz-halogen lamp. The fluence rate was 240 mW/cm<sup>2</sup> measured by a photo radiometer (Delta Ohm, Padua, Italy). Cell viability was measured by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. The IC<sub>50</sub> value was 47.2 µM.

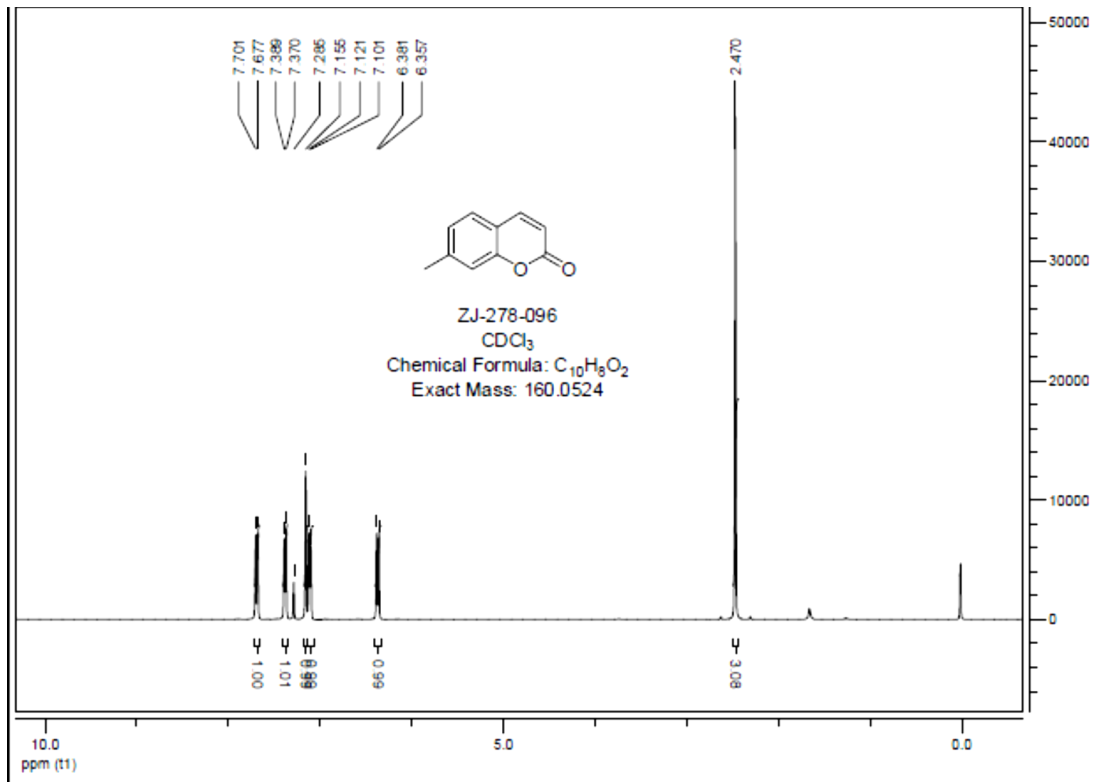


IC<sub>50</sub>=47.2 µM

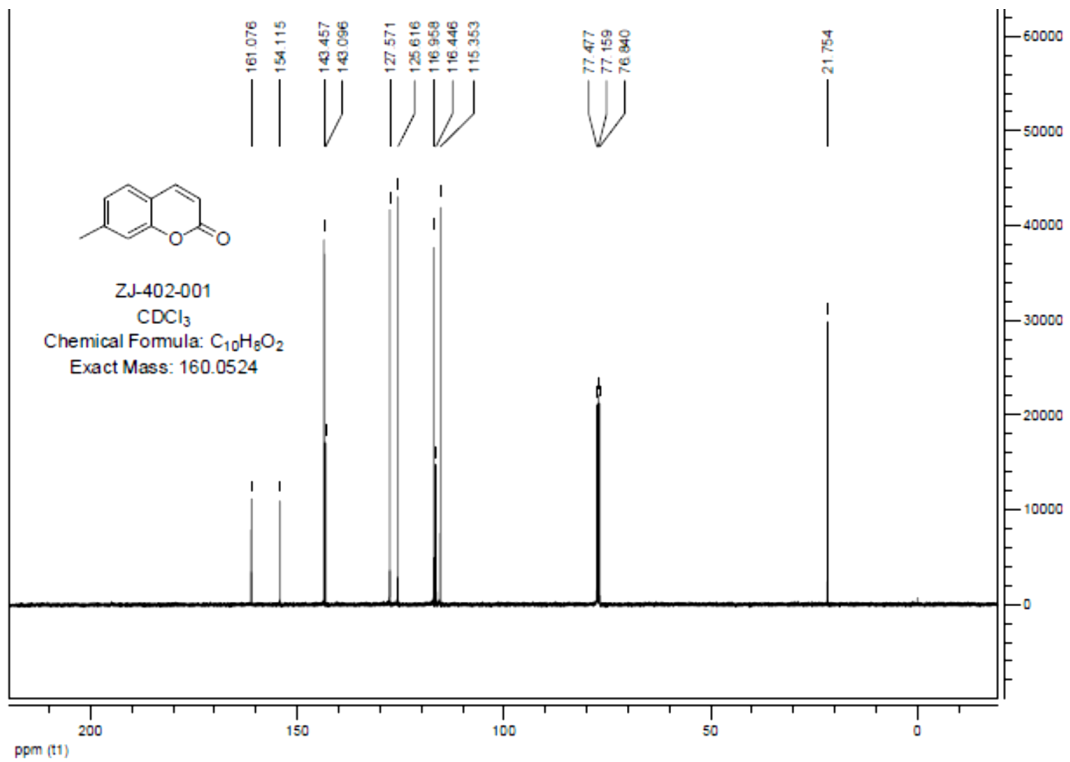
#### 4. NMR and MS spectra



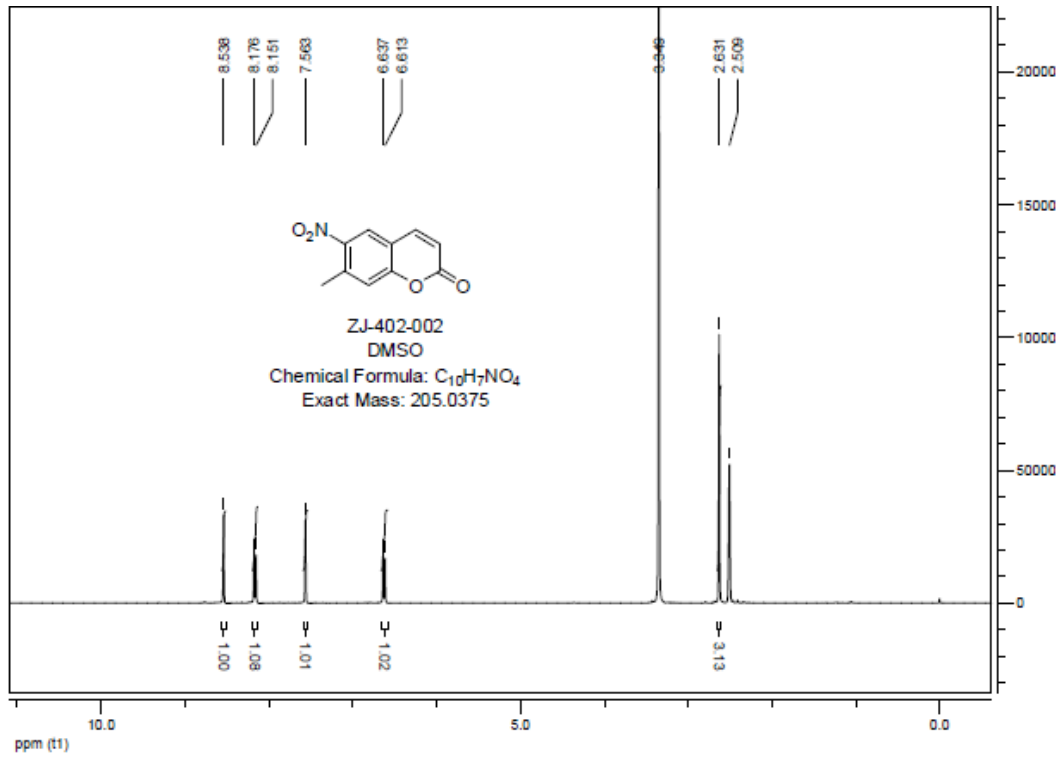
### Compound 1-<sup>1</sup>H NMR



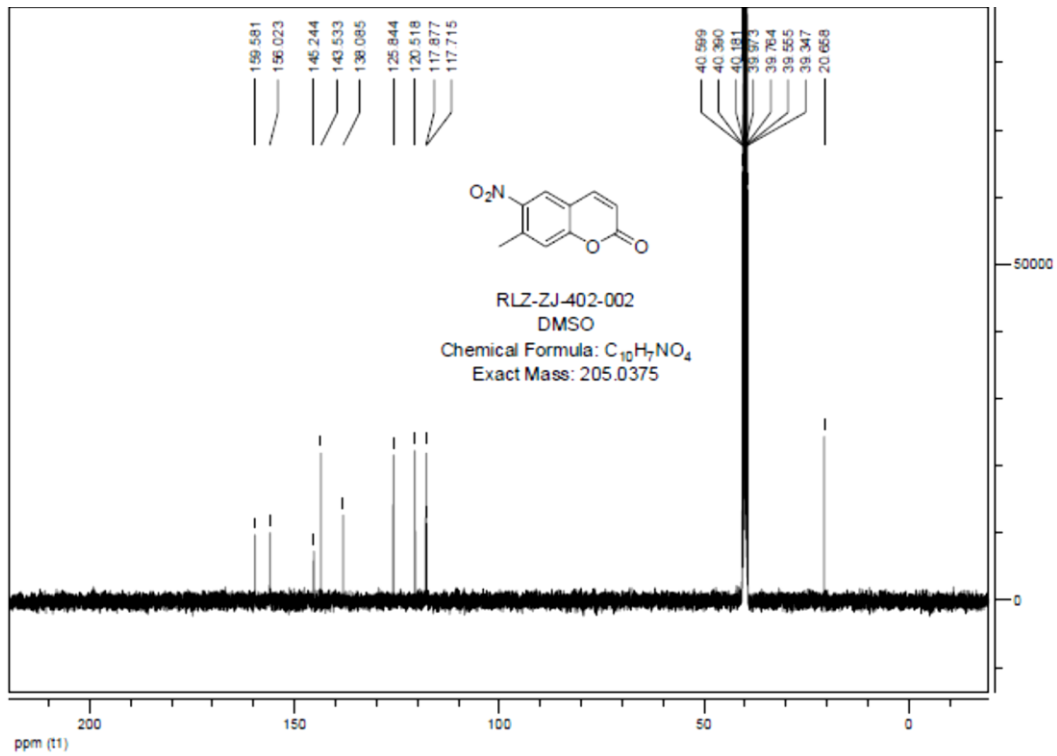
### Compound 1-<sup>13</sup>C NMR



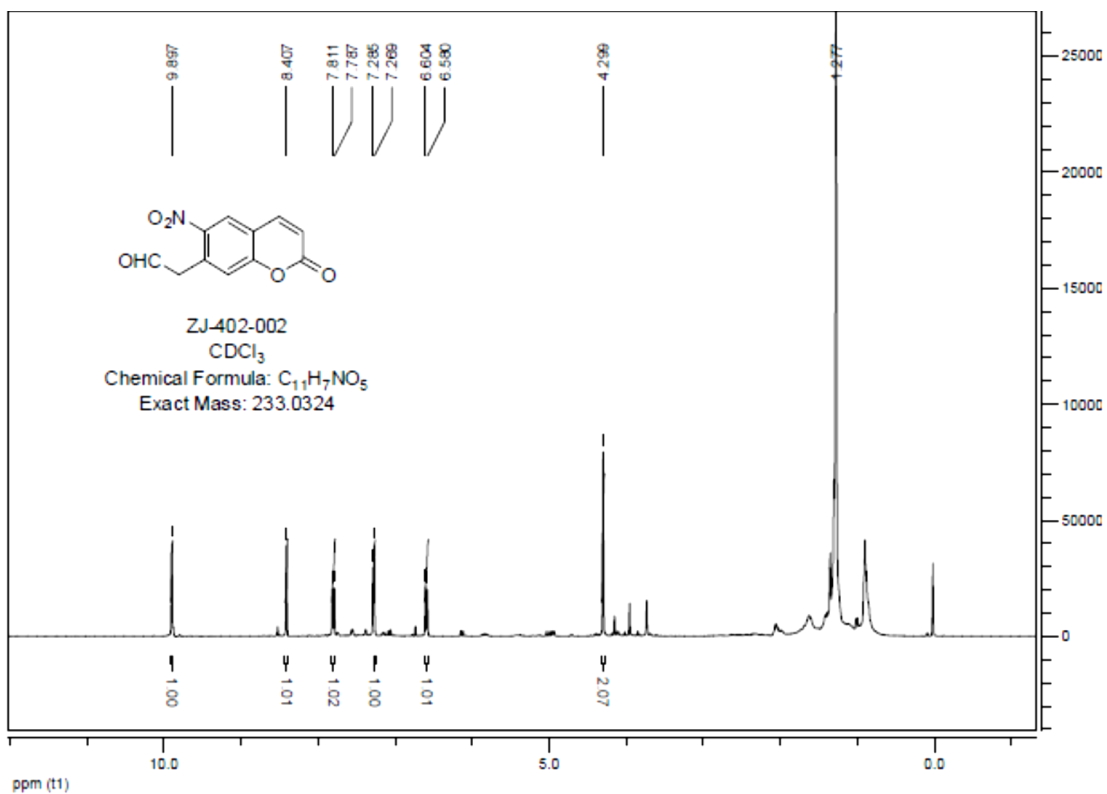
### Compound 2-<sup>1</sup>H NMR



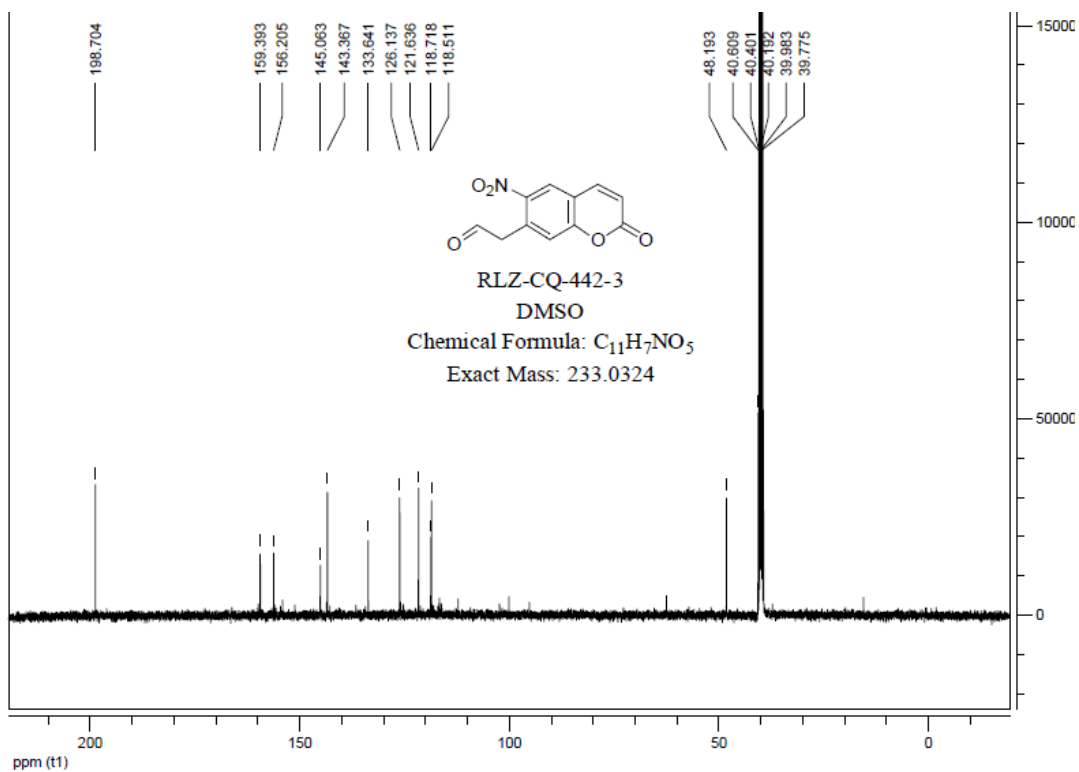
### Compound 2-<sup>13</sup>C NMR



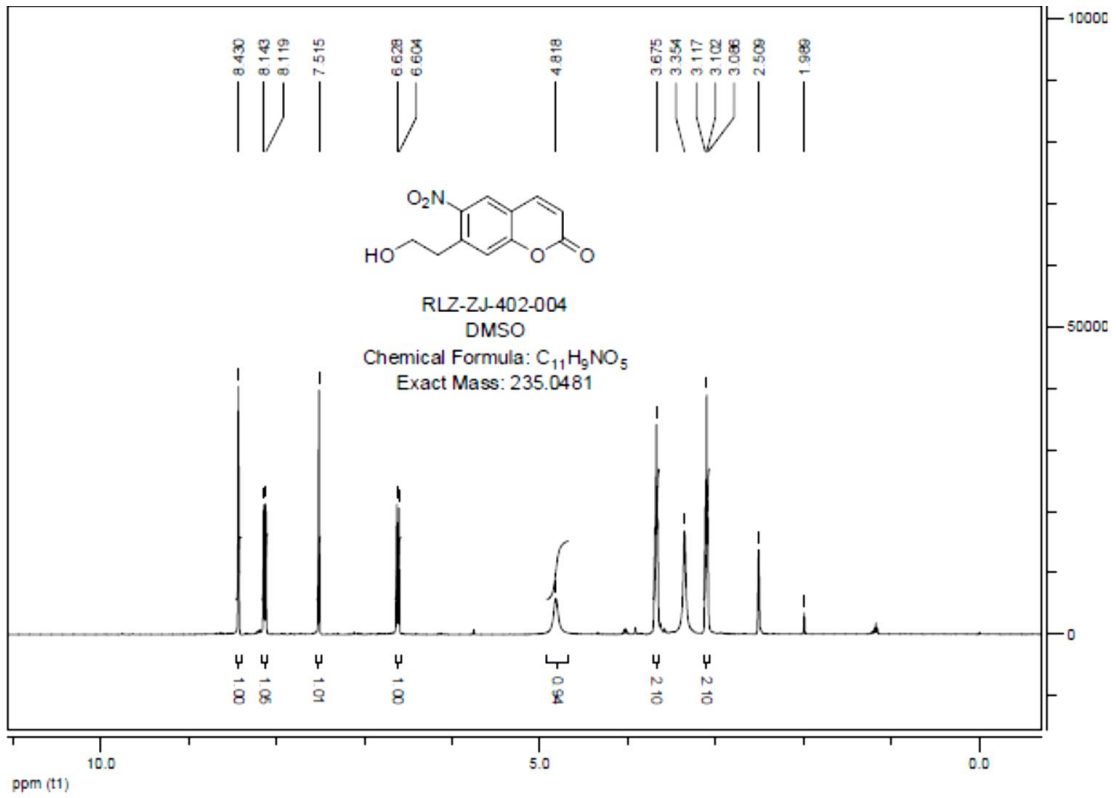
### Compound 4-<sup>1</sup>H NMR



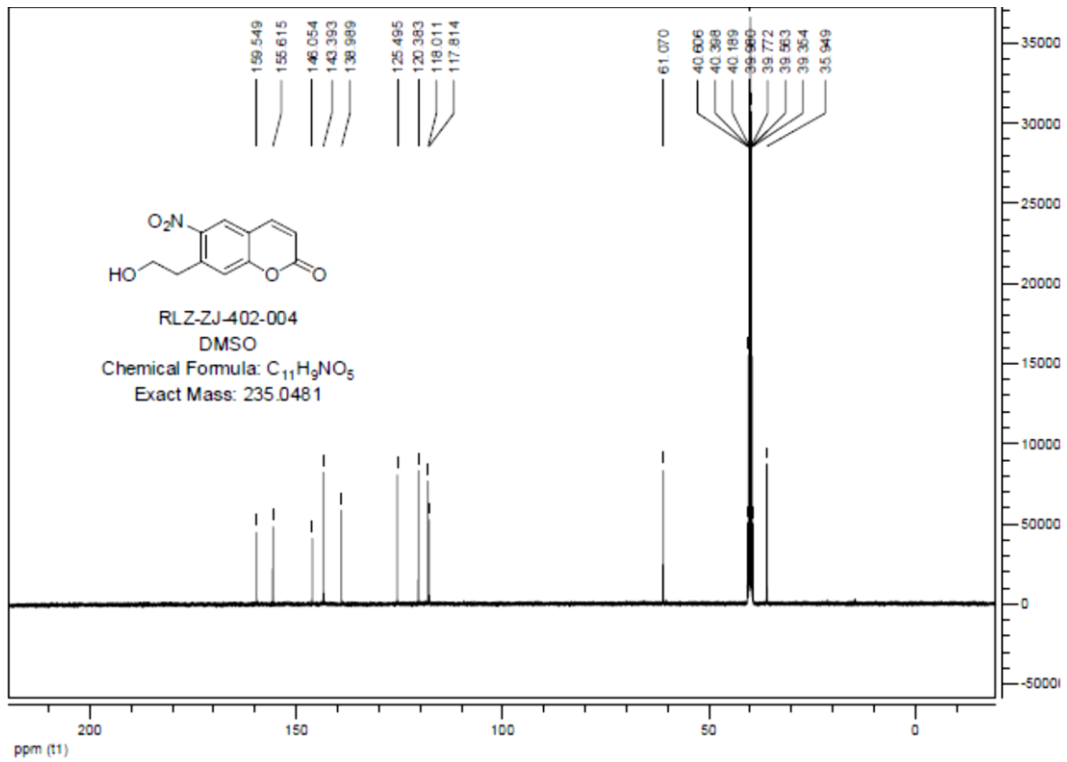
### Compound 4-<sup>13</sup>C NMR



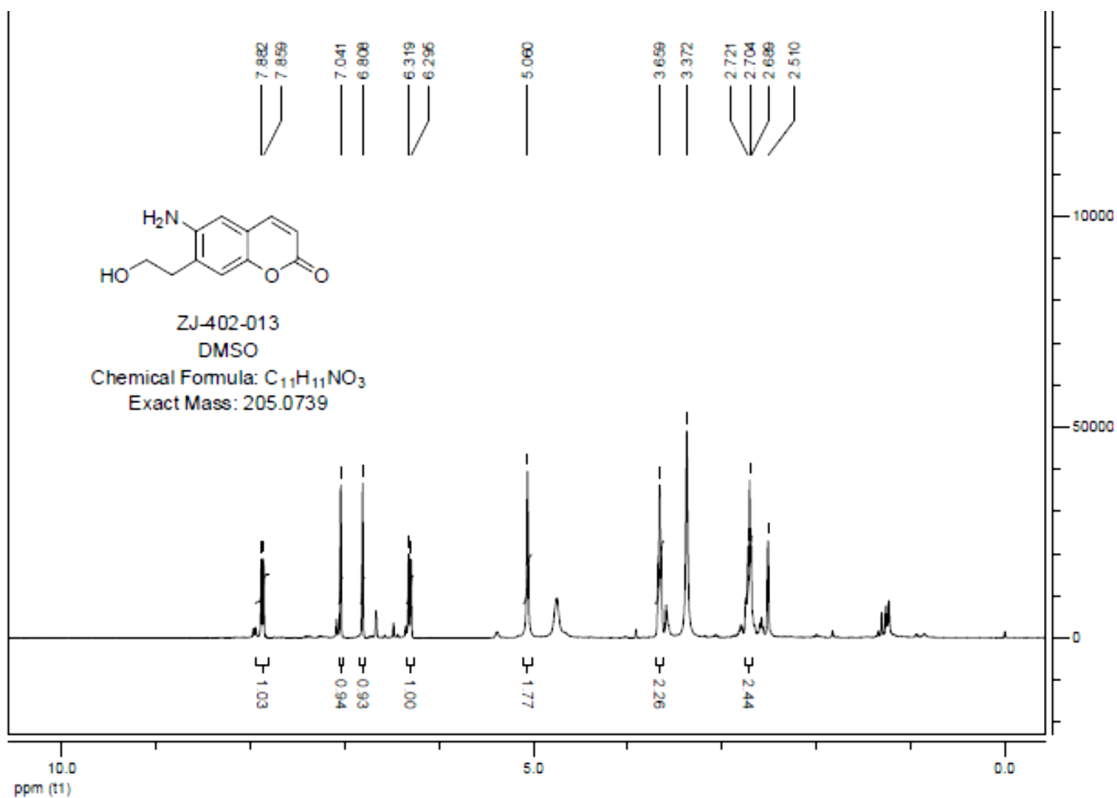
### Compound 5-<sup>1</sup>H NMR



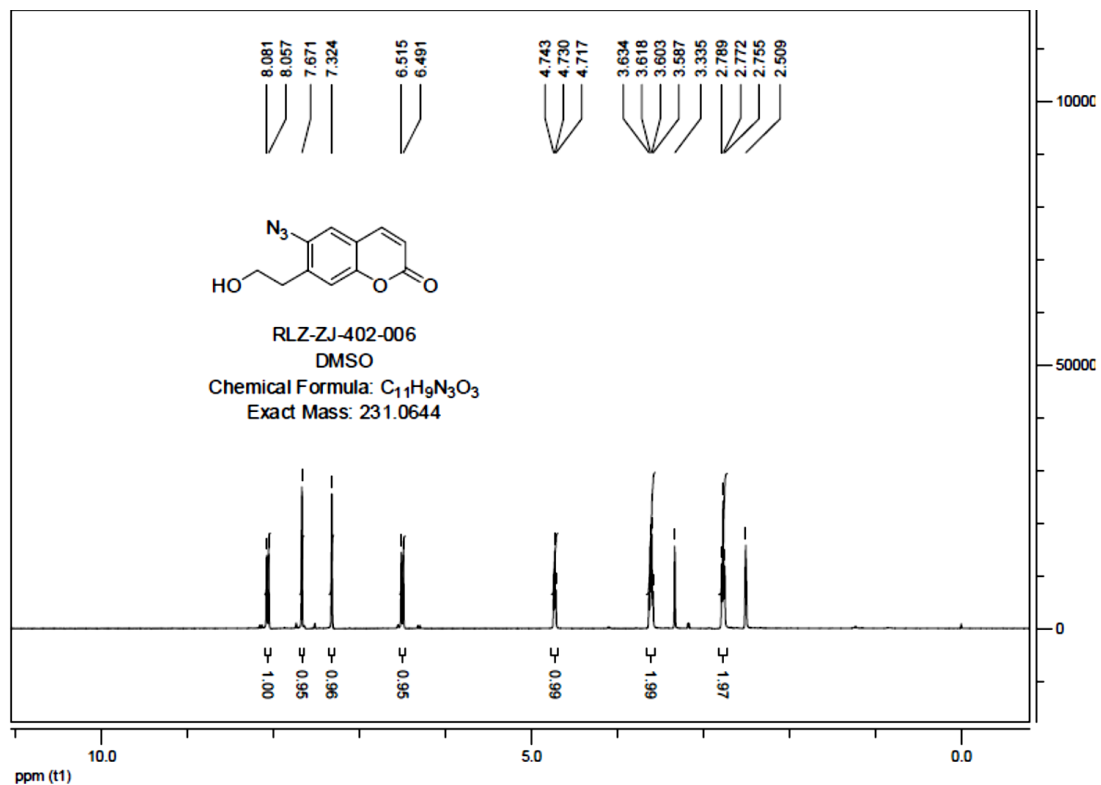
### Compound 5-<sup>13</sup>C NMR



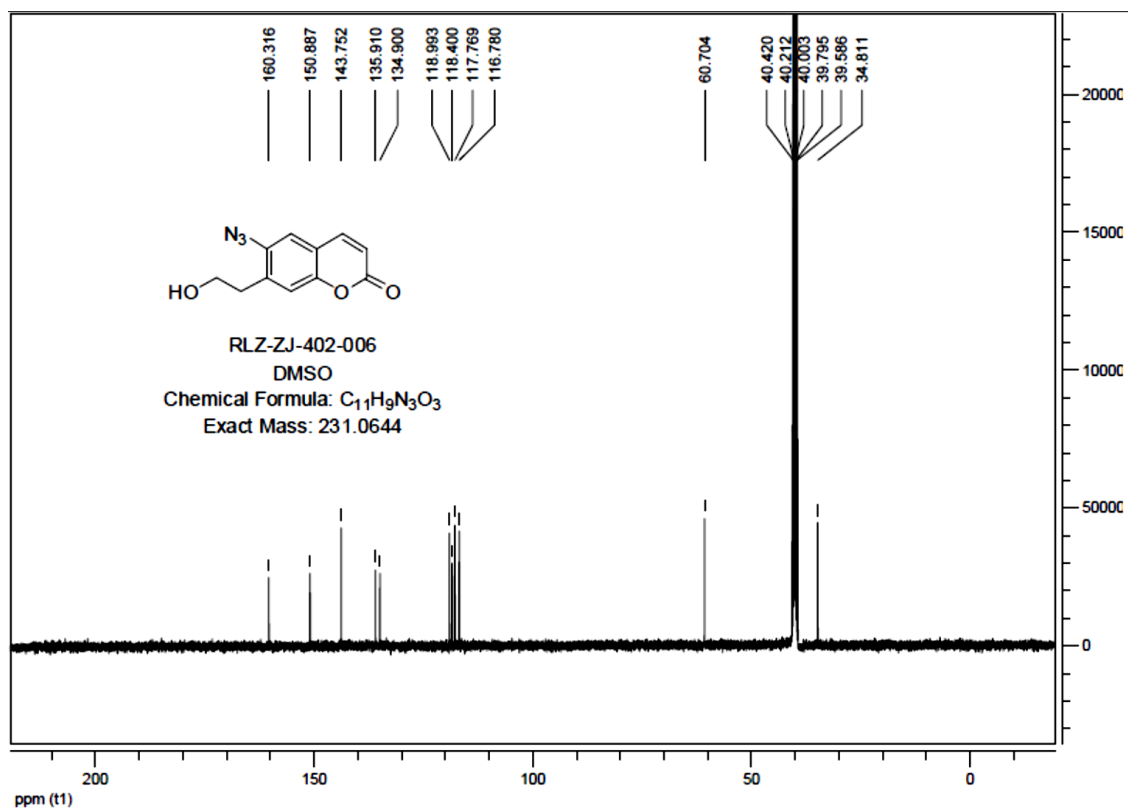
### Compound 6-<sup>1</sup>H NMR



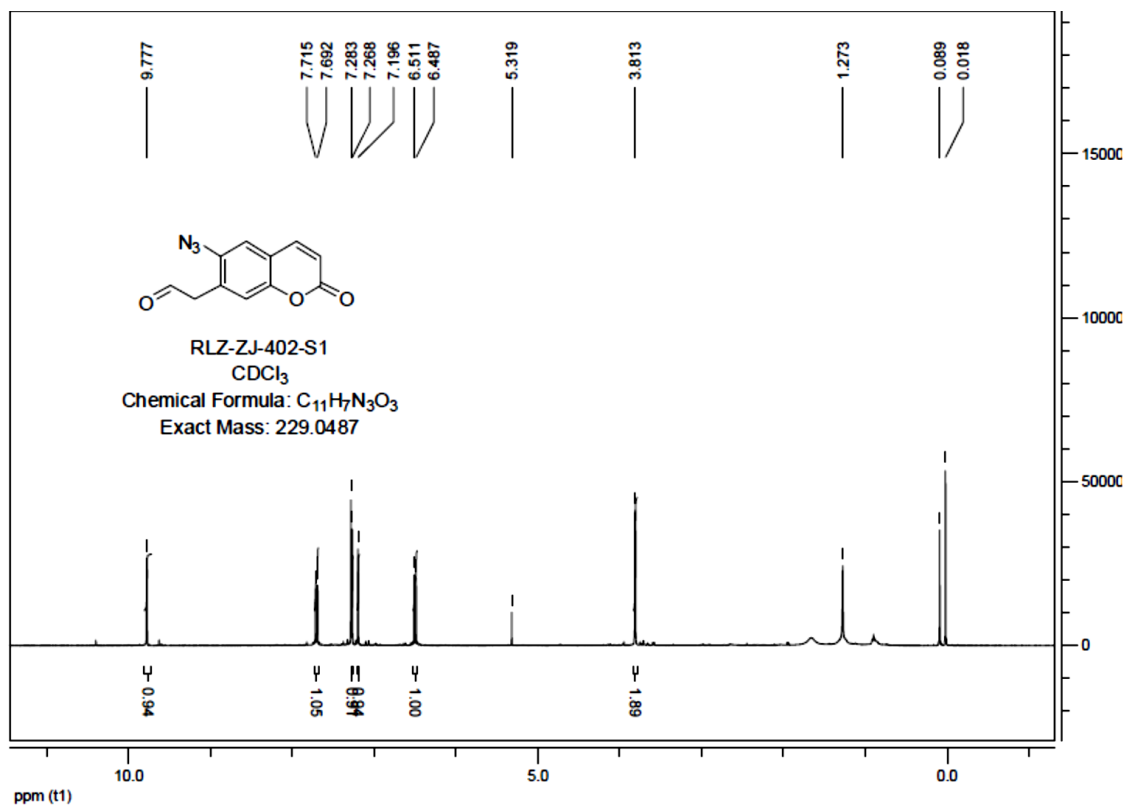
### Compound 7-<sup>1</sup>H NMR



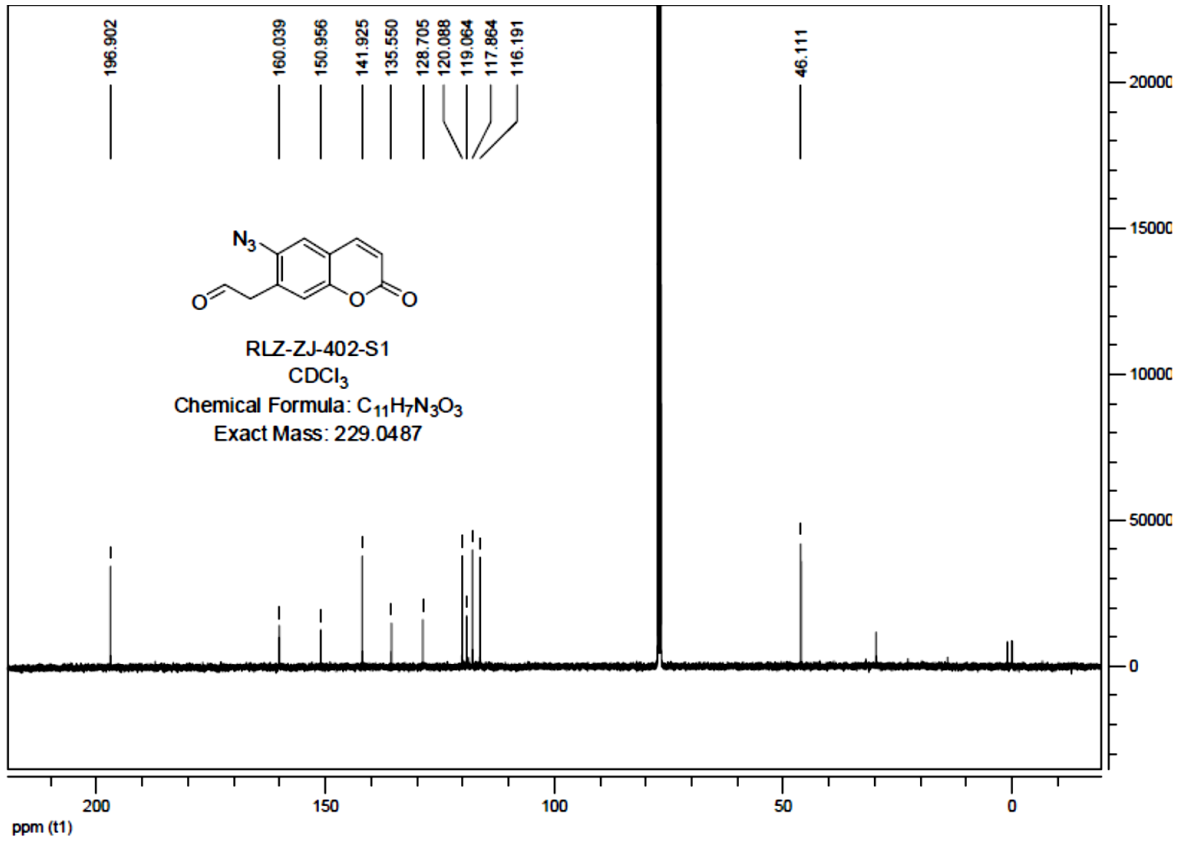
### Compound 7-<sup>13</sup>C NMR



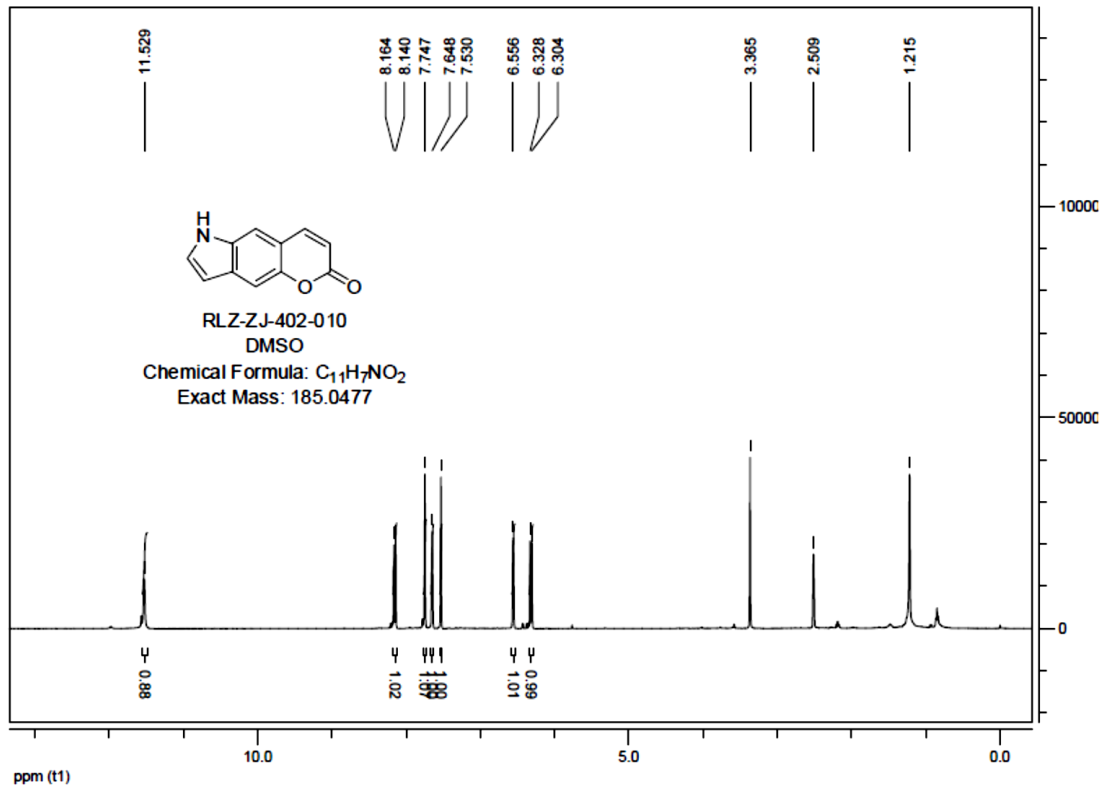
### Compound S1-<sup>1</sup>H NMR



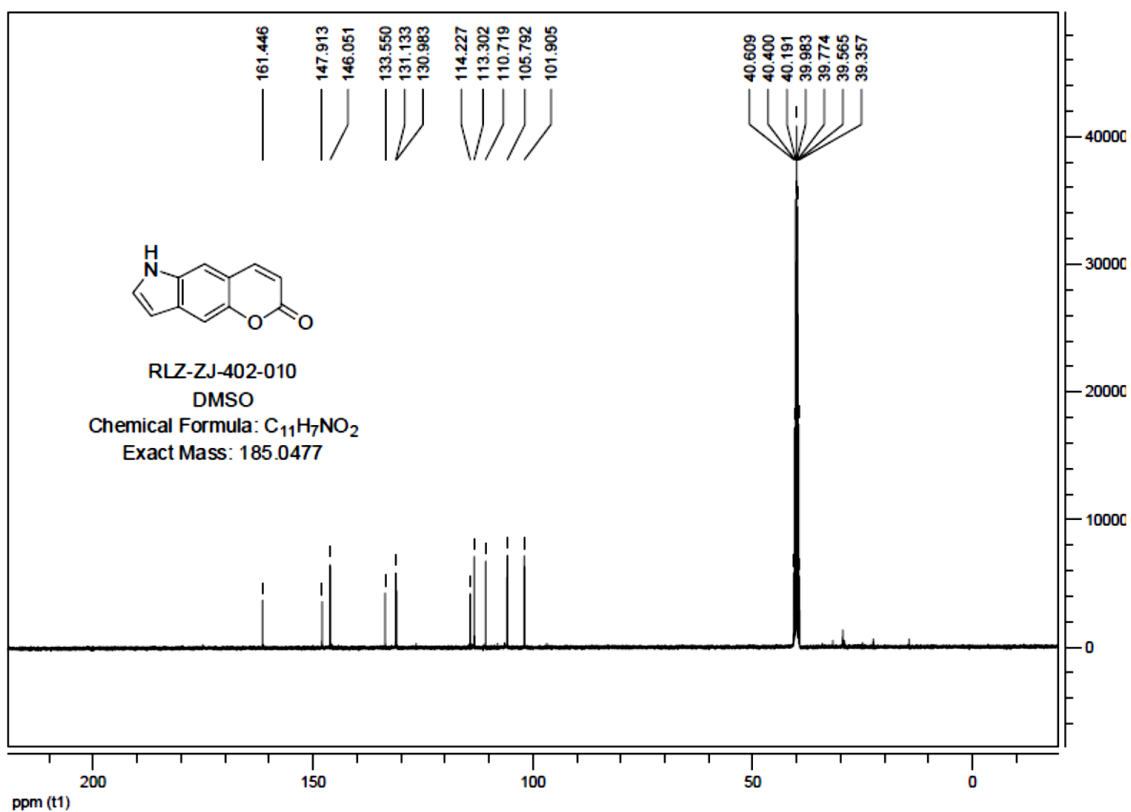
### Compound S1-<sup>13</sup>C NMR



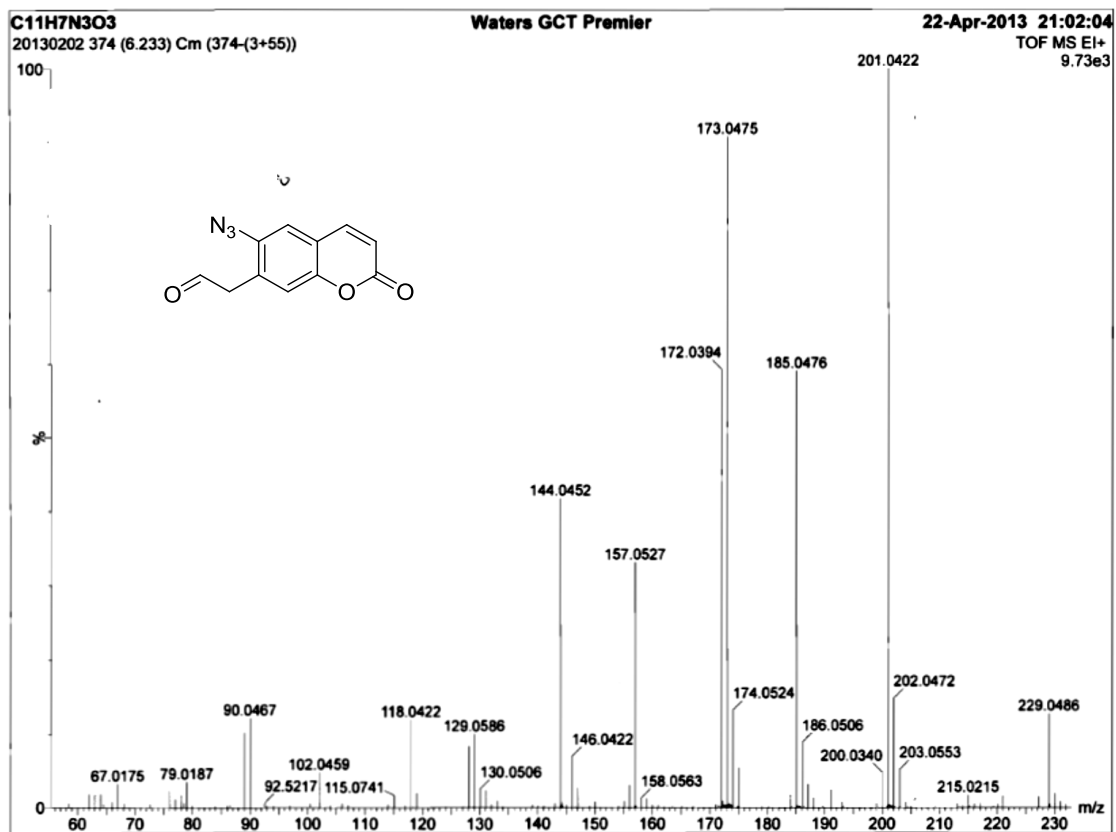
### Compound 8-<sup>1</sup>H NMR



### Compound 8-<sup>13</sup>C NMR

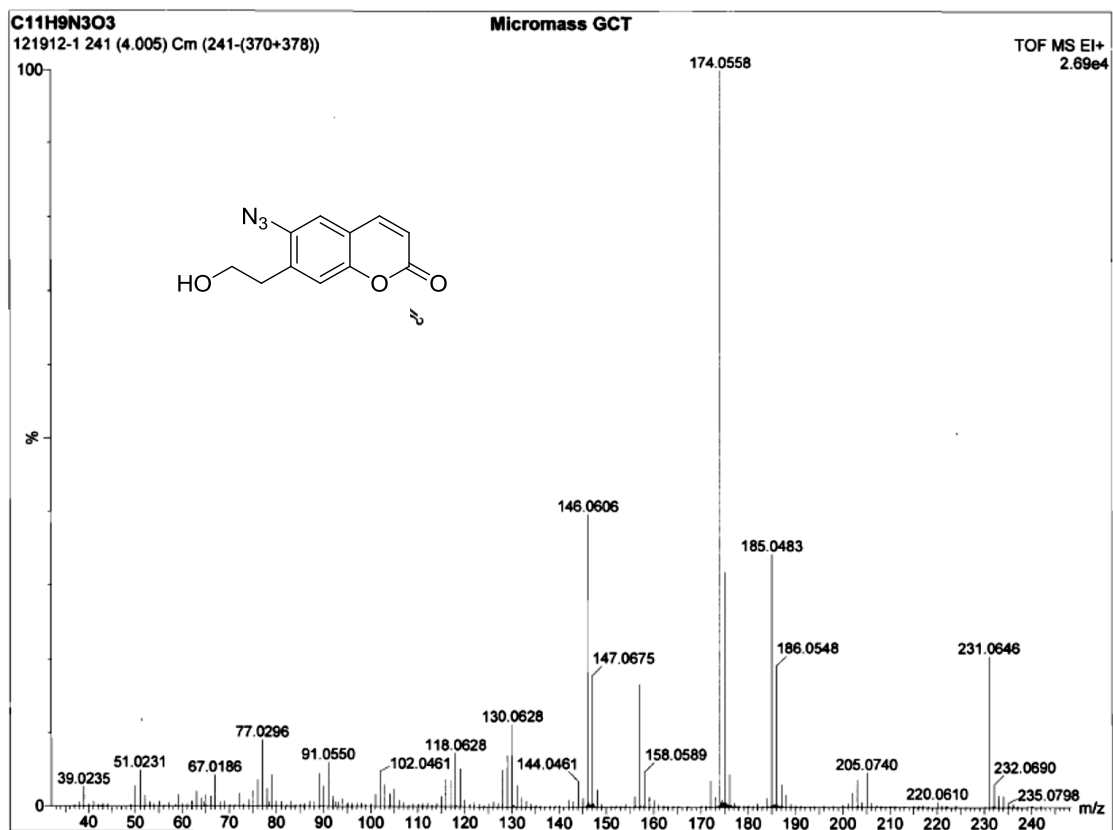


### Compound S1-HRMS (EI)





## Compound 7-HRMS (EI)



### 5. References:

1. P. Yates and T. S. Macas, *Canadian Journal of Chemistry*, 1988, **66**, 1.
2. G. Chen, D. J. Yee, N. G. Gubernator and D. Sames, *J. Am. Chem. Soc.*, 2005, **127**, 4544.
3. H. Cheng, J. Wan, M.-I. Lin, Y. Liu, X. Lu, J. Liu, Y. Xu, J. Chen, Z. Tu, Y.-S. E. Cheng and K. Ding, *J. Med. Chem.*, 2012, **55**, 2144.
4. M. Alajar n, J. Cabrera, A. Pastor and J. M. Villalgordo, *Tetrahedron Lett.*, 2007, **48**, 3495.
5. T. Liu, Z. Xu, D. R. Spring and J. Cui, *Org. Lett.*, 2013, **15**, 2310.