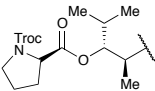
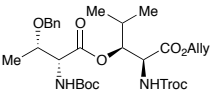
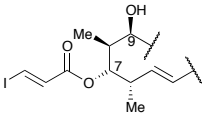
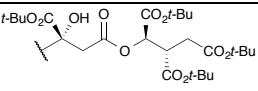
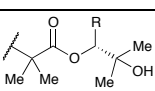
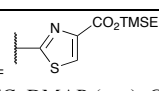
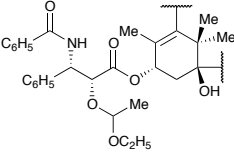
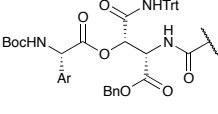
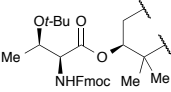
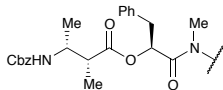
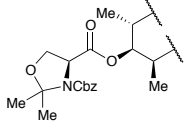
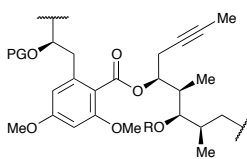
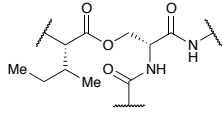
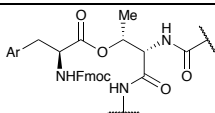
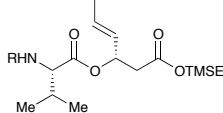
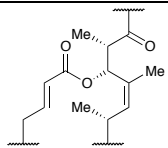
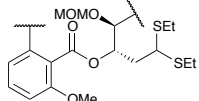
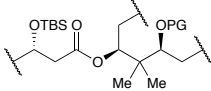
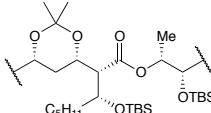
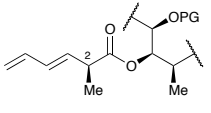
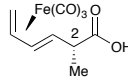
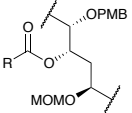
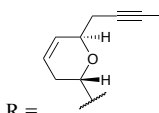
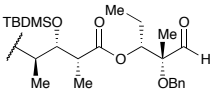
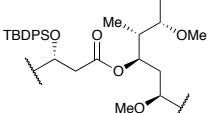
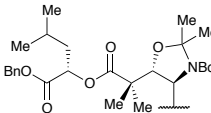
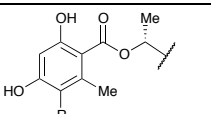


**Table S1.** Collection of complex ester couplings in total synthesis.

Entry	Substructure	Natural Product	Procedure <sup>a</sup>	Yield	Comment
1		<b>Virginia mycin M2</b> 1996 <sup>182</sup>	DCC, DMAP (cat.) 0 °C to rt, 10 h	92%	
2		<b>A83586C</b> 1997 <sup>183</sup> <b>Azinothricin and Kettapeptin</b> 2009 <sup>184</sup>	DCC, DMAP <sup>b</sup> 0 °C to rt	83–88%	5–10% epimerization at the $\alpha$ -carbon of the threonine-unit
3		<b>Elaiophylin</b> (macrocyclic core) 1997 <sup>185</sup>	DCC, DMAP, -20 °C, 16 h then Ti(O <i>i</i> -Pr) <sub>4</sub> , 20 °C, 16 h <sup>b</sup>	44% (2 steps)	esterification followed by equilibration (C9-OH less hindered than C7-OH) esterification under kinetic conditions: C9:C7 ester = 94:6 after equilibration: C9:C7 ester = 31:69  <i>Unsuccessful:</i> <ul style="list-style-type: none"> <li>C9-OH Bz-protected: esterification failed under a variety of conditions, presumably due to steric congestion</li> </ul>
4		<b>Citrafungin A</b> 2007 and 2008 <sup>129</sup>	DCC (6.5 equiv.), DMAP•HCl (9.8 equiv.), ROH (2 equiv.) rt, 4 h	50%	
5		<b>Lyngbyabellin A</b> 2001 <sup>127,186</sup>	R =  DCC, DMAP (cat.), CSA (cat.) rt, 19 h	64%	Keck procedure
		<b>Lyngbyabellin B</b> 2002 <sup>186</sup>	R = CO <sub>2</sub> Allyl TCBC, TEA, DMAP rt to 60 °C, 13 h	66%	<i>Unsuccessful:</i> <ul style="list-style-type: none"> <li>Keck conditions, which have successfully been employed for a related coupling in the synthesis of Lyngbyabellin A (<i>vide supra</i>)</li> </ul>
6		<b>Taxol</b> 1988 <sup>21</sup>	ROH (6 equiv.), DPC (6 equiv.), DMAP (2 equiv.) 73 °C, 100 h	80% <sup>c</sup>	Modification of Kim's esterification <sup>22</sup>  <i>Unsuccessful:</i> <ul style="list-style-type: none"> <li>various ester coupling conditions that are usually successful for hindered substrates (not specified in the paper)</li> <li>Kim's esterification (unmodified)</li> </ul>
7		<b>Ramoplanin A2</b> 2003 <sup>104</sup>	acid (2 equiv.), EDC (3 equiv.), DMAP (0.3 equiv.) 0 °C, 1 h	87%	diastereomeric ratio 9:1  <i>Unsuccessful:</i> <ul style="list-style-type: none"> <li>mixed anhydrides</li> <li>Mitsunobu</li> <li>acyl fluoride</li> <li>Yamaguchi</li> <li>Corey-Nicolaou</li> <li>EDC or DCC without DMAP</li> <li>higher temperature (25 °C)</li> <li>longer reaction time</li> <li>more DMAP (0.5–2 equiv.)</li> </ul> lower conversion, lower <i>dr</i> or $\beta$ -elimination
8		<b>Viequeamide A</b> 2013 <sup>187</sup> <u>coupling junction:</u> [L-Thr – (2 <i>S</i> )-Dhoya] (See also entry 33)	acid (5 equiv.), EDC (10 equiv.), DMAP (10 equiv.) rt, o/n	81%	<i>Unsuccessful:</i> <ul style="list-style-type: none"> <li>acid-Cl/DMAP/DIPEA</li> </ul> <i>Low yields:</i> <ul style="list-style-type: none"> <li>DCC/PPY</li> <li>TCBC/DMAP/DIPEA</li> <li>EDC/DMAP (3 equiv. each)</li> </ul>

9		<b>Plusbacin A<sub>3</sub></b> <sup>188</sup> (2007)	EDC, DMAP (cat.) -15 °C to rt, 12 h	84%	EDC added in 5 portions over 1 h
10		<b>Pipecolidepsin A</b> 2013 <sup>109</sup>	solid phase acid (15 equiv.), DIC (15 equiv.), DMAP (0.5 equiv.), CH <sub>2</sub> Cl <sub>2</sub> /DMF 9:1 45 °C, 2.3 h	98%	11% epimerization  <i>Unsuccessful:</i> • MSNT/NMI • acyl fluoride/DIPEA  <i>Lower yield:</i> • less equivalents of DIC • rt
11		<b>Symplocamide A</b> 2010 <sup>189</sup>	solid phase acid (10 equiv.), DIC (10 equiv.), DMAP (1 equiv.), CH <sub>2</sub> Cl <sub>2</sub> /DMF 9:1 rt, 4 x 2 h	92%	
12		<b>Iejimalides</b> 2011 <sup>190,124</sup>	A: EDC (2 equiv.), DMAP (2 equiv.), DMAP•HCl (2.1 equiv.), high vacuum, Ar (g), then acid (1.3 equiv.), ROH, rt, 48 h (sealed flask under Ar (g) in the dark) <sup>190</sup>  B: acid, EDC, PPY (cat.), 0 °C to rt 10 min then conc. under a stream of argon, addition of ROH rt, 18 h <sup>124</sup>	78% <sup>d</sup>	<i>Unsuccessful:</i> <sup>190</sup> • Yamaguchi/Yonemitsu ( <i>note:</i> Yonemitsu successful for R = Boc <sup>191,192</sup> ) • Mukaiyama's salt • DCC/DMAP • EDC/DMAP • EDC/DMAP/Sc(OTf) <sub>3</sub>  <i>Low to moderate yield:</i> <sup>190</sup> • Keck (DCC (1 equiv.), DMAP (cat.), DMAP•HCl (cat.)): DCC-urea byproduct difficult to separate • EDC (1 equiv.), DMAP (cat.), DMAP•HCl (cat.): low yield (20%) • As procedure A, but alcohol : acid = 1:1 instead of 1.3:1 (60%)  reaction has to be monitored closely, conditions have to be scrupulously met
13		<b>Iriomoteolide-3a</b> 2009 <sup>112,193</sup>	EDC, PPY, rt (PG = TBS) <sup>b,112</sup>  <i>Alternatively:</i> TCBC, TEA, DMAP, rt, 36 h (PG = MOM) <sup>193</sup>	72% <sup>112</sup>  93% <sup>182</sup>	
14		<b>506BD</b> 1991 <sup>114</sup>	DIC (5 equiv.), PPY (3 equiv.), acid (4 equiv.) -20 °C, 20 h	92%	
15		<b>Veraguamide A</b> 2012 <sup>194</sup>	EDC 0 °C to rt <sup>b</sup>	55%	<i>Unsuccessful:</i> • DCC/PPY • Yamaguchi • oxalyl chloride/DMAP
16		<b>Halipeptin A</b> 2005 <sup>132</sup>	acid-Cl, DMAP (cat.), DIPEA -15 °C <sup>b</sup>	95% <sup>d</sup>	<i>Unsuccessful:</i> • Yamaguchi • other activated esters • higher temp (0 °C): racemization • more DMAP (1 equiv.): racemization • higher temp (0 °C) and less DMAP (0.2 equiv.): low conversion

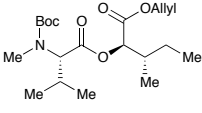
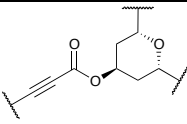
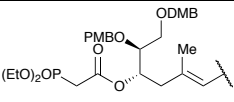
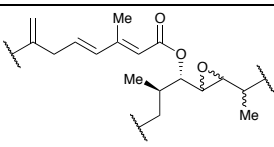
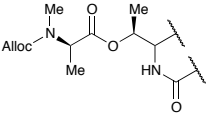
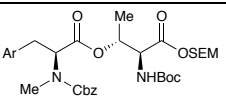
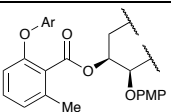
17		<b>Grassypeptolide</b> 2010 <sup>195</sup>	acid-Cl, DMAP (cat.), NMM 0 °C to rt <sup>b</sup>	90%	<i>Unsuccessful:</i> <ul style="list-style-type: none"> <li>EDC/DMAP</li> <li>DCC/DMAP</li> <li>Mukaiyama's reagent</li> </ul>
18		<b>LL15G256γ</b> 2008 <sup>196</sup>	acid, (COCl) <sub>2</sub> , DMF/CH <sub>2</sub> Cl <sub>2</sub> , then ROH, TEA, DMAP, CH <sub>2</sub> Cl <sub>2</sub>	85%	Garner's protective protocol for serine was crucial for the coupling  <i>Unsuccessful:</i> <ul style="list-style-type: none"> <li>Coupling of diprotected serine (TBS and Cbz or Fmoc) under Keck and Yamaguchi conditions led to racemization and low yields</li> </ul>
19	  R = TBS, PG = TBDPS <sup>138</sup> R = H, PG = TIPS <sup>137</sup>	<b>Cruentaren A</b> 2007 <sup>137,138</sup>	2,4,6-trifluoro-1,3,5-triazene (5 equiv.), pyridine (1.1 equiv.), CH <sub>2</sub> Cl <sub>2</sub> -25 °C, 10 min then sodium salt of the alcohol (1.1 equiv.), THF rt, 1 h	91%	<i>Unsuccessful:</i> <ul style="list-style-type: none"> <li>Mitsunobu</li> <li>Yamaguchi</li> <li>Trost esterification</li> <li>carbodiimide based reagents</li> <li>acid chloride (lactone formation)</li> <li>activated esters and thioesters</li> <li>PySSPy/PPh<sub>3</sub>, AgBF<sub>4</sub></li> <li>di-2-thienyl carbonate (2-DTC), Hf(OTf)<sub>2</sub>, DMAP</li> <li>PG = MOM or TCA (lactone formation)</li> </ul> <i>Modest yields:</i> <ul style="list-style-type: none"> <li>Mukaiyama's reagent</li> <li>sodium salt of the alcohol/imidazolide derivative of the acid, 0 °C to rt, 3 d (65%) (for R = H and PG = TIPS)</li> </ul>
20		<b>MA026</b> 2013 <sup>143</sup>	acid (1.15 equiv.), HBTU (4 equiv.), HOBt (4 equiv.), NMM (5 equiv.), CH <sub>2</sub> Cl <sub>2</sub> /DMF 4:1 rt, 48 h	40% (71% brsm)	<i>Unsuccessful:</i> <ul style="list-style-type: none"> <li>EDC</li> <li>mixed anhydrides</li> <li>Yamaguchi</li> <li>Mukaiyama</li> <li>Shiina's protocol</li> </ul> <i>Low conversion:</i> <ul style="list-style-type: none"> <li>EDC/HOBt/NMM</li> </ul>
21		<b>Daptomycin</b> 2013 <sup>144</sup>	acid (2.2 equiv.), PyBOP (4 equiv.), DIPEA (4 equiv.), rt, 12 h	87%	<i>Unsuccessful:</i> <ul style="list-style-type: none"> <li>coupling of the unnatural amino acid kynurenine</li> </ul>
22		<b>FK228</b> 2008 <sup>152</sup>	R = Fmoc-D-Val-D-Cys(Tr)-Dhb-: MSNT, N-Me-imidazole 0 °C, 5 h to rt, 100 min	34%	For R = Fmoc-D-Val-D-Cys(Tr)-Dhb-: <ul style="list-style-type: none"> <li>prolonged reaction time resulted in epimerization</li> <li>42% unreacted alcohol</li> </ul> R = Fmoc: DCC, DMAP (cat.) 0 °C, 3 h 85%
23		<b>(+)-Migrastatin</b> 2003, <sup>161</sup> 2004 <sup>162</sup> and 2007 <sup>197</sup>	acid (3 equiv.), TCBC (3 equiv.), DIPEA (2.8 equiv.), rt, 3 h then ROH, pyridine (4 equiv.) rt, 24 h	66–74%	<i>Unsuccessful:</i> <ul style="list-style-type: none"> <li>acid chloride/DMAP, pyridine or AgCN</li> <li>EDC or DCC</li> <li>Mukaiyama's reagent</li> <li>Keck coupling</li> <li>decomposition of SM or partial isomerisation to the β,γ-unsaturated ester</li> </ul>
24		<b>Oximidines I and II</b> 2003 <sup>198</sup>	TCBC, TEA, CH <sub>2</sub> Cl <sub>2</sub> , rt, 3 h then ROH, DMAP, PhMe, 90 °C, 16 h	38%	

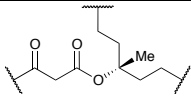
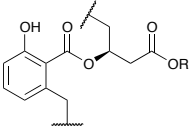
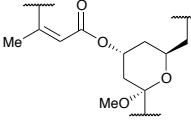
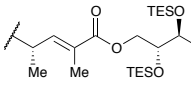
25		<b>Rhizopodin</b> 2012 <sup>199,200</sup> and 2013 <sup>201,202,203</sup>	TCBC, TEA, DMAP <sup>b,201</sup>	63–96%	PG = TBS or TES
26		<b>Filipin III</b> 1999 <sup>204</sup>	TCBC, TEA, THF, rt, 16 h then ROH, DMAP, PhMe, rt, 2 h	70%	<i>Unsuccessful:</i> • DCC/DMAP • BOP/DMAP  <i>Low yield:</i> • PyBroP (30% yield)
27		<b>Amphidinolide E</b> 2007 <sup>205</sup>	(CO) <sub>3</sub> Fe-complexed dienic acid (see below), TCBC, TEA, DMAP 0 °C, 1 h to rt, 1 h   followed by oxidative decomplexation of the (CO) <sub>3</sub> Fe unit (96%)	94%	<i>Note:</i> inversion at C2 of the acid component!  <i>Unsuccessful:</i> • excess amounts of acid and various coupling reagents • Yonemitsu modification of the Yamaguchi reaction • mild peptide coupling conditions (EDC•methiodide (EDC•MeI)/DMAP, PyBrOP/DIPEA, DCC/HOBt), • Otera's transesterification catalyst • tributyltin ether of the alcohol/acyl fluoride • lithium alkoxide/acyl fluoride • Kita's conditions isomerization to the fully conjugated species
28		<b>(-)-Laulimalide</b> 2012 <sup>170e,171</sup>	 R =	18%	<i>Unsuccessful:</i> • Steglich and related protocols • acyl chloride or acyl fluoride • Ghosez reagent <sup>48f,g</sup> (allene formation) • Kita's conditions  <i>Low yields/only traces of product:</i> • transesterification from methyl ester in presence of Otera's catalyst (13%) • Shiina mixed anhydride
			R = CH <sub>2</sub> PO(OCH <sub>2</sub> CF <sub>3</sub> ) <sub>2</sub> acid (2.3 equiv.), TCBC (2.9 equiv.), DIPEA (4.6 equiv.), THF, rt, 10 min then DMAP (6.9 equiv.), C <sub>6</sub> H <sub>6</sub> , rt, 2 h	99%	presumably via a more reactive ketene intermediate from phosphonoacetic acid
29		<b>Methynolide</b> 1986 <sup>206</sup>	TCBC, TEA, DMAP rt, 20 h <sup>b</sup>	58%	<i>Unsuccessful:</i> • DCC/DMAP: 70% yield but impurities difficult to remove • other esterification reagents (not specified)
30		<b>Mycalolide B</b> 2010 <sup>207</sup>	MNBA, TEA, DMAP rt <sup>b</sup>	55%	
31		<b>Kitastatin and Respirantin</b> 2014 <sup>208</sup>	MNBA, DMAP, TEA rt, 40 h	93%	
32		<b>Pochonin C</b> <sup>179,180</sup> and <b>Radicol</b> <sup>180</sup> 2004 (R = H)	P( <i>m</i> ClPh) <sub>3</sub> , DIAD 23 °C, 3 h	84%	<i>Lower yield:</i> • 2-OH group protected (Mitsunobu or carbodiimide conditions) • different strategy for R = Cl (Pochonin A <sup>49(b)</sup> ): persilylation of the benzoic acid

derivative, followed by conversion to the acid chloride and esterification with the alcohol  
 a) TBSCl, DIPEA, b) (COCl)<sub>2</sub>, DMF/CH<sub>2</sub>Cl<sub>2</sub> c) ROH, TEA (29% over 3 steps)

*Poor selectivity:*

- classical Mitsunobu conditions: alkylation of the *para*-phenol

33		<b>Viequeamide A</b> 2013 <sup>187</sup> <u>coupling junction:</u> [ <i>N</i> -Me-L-Val – (2 <i>R</i> ,3 <i>S</i> )-HMPA] (See also entry 8)	acid (3 equiv.), DEAD (3 equiv.), PPh <sub>3</sub> (4.5 equiv.) 0 °C to rt, 14 h	91%	
34		<b>Leucascandrolide A</b> 2003 <sup>176c-d</sup>	acid (4.5 equiv.), DEAD (3.3 equiv.), PPh <sub>3</sub> (20 equiv.), THF/PhMe = 1.5:1, 0 °C to 20 °C, 14 h	90%	From C5 equatorial alcohol  <i>Unsuccessful:</i> from C5 axial alcohol: <ul style="list-style-type: none"> <li>• various activated acid derivatives,</li> <li>• acid chloride</li> </ul>
35		<b>(+)-Zampanolide and (+)-Dactylolide</b> 2002 <sup>91</sup>	acid (15 equiv.), PPh <sub>3</sub> (6.5 equiv.), DEAD (7 equiv.) rt, o/n	99%	<i>Note:</i> complete retention of configuration!  Saturation of the reaction medium with acid, PPh <sub>3</sub> and DEAD necessary to achieve full conversion of the starting material  <i>Unsuccessful:</i> <ul style="list-style-type: none"> <li>• standard Mitsunobu protocol (premixing alcohol and acid followed by addition of PPh<sub>3</sub> and DEAD)</li> <li>• more DEAD and PPh<sub>3</sub></li> <li>• prolonged reaction times</li> <li>• reversed order of reagent addition</li> </ul> <i>Incomplete consumption of the alcohol:</i> <ul style="list-style-type: none"> <li>• heating to 40–50 °C: elimination side product</li> </ul>
36		<b>Amphidinolide A</b> (different stereoisomers) 2002 <sup>209</sup> , 2004 <sup>210</sup> , 2005 <sup>211</sup> and 2006 <sup>212</sup>	[RuCl <sub>2</sub> ( <i>p</i> -cymene)] <sub>2</sub> (cat.), HCCOEt, PhMe, rt, 2 h ROH, CSA (cat.), ClCH <sub>2</sub> CH <sub>2</sub> Cl rt, 2 h <sup>209</sup>	42–66%	<i>Unsuccessful:</i> <ul style="list-style-type: none"> <li>• Keck or Mitsunobu: complex mixtures<sup>212</sup></li> <li>• any other esterification which employed basic conditions: extensive isomerisation of the triene<sup>209</sup></li> </ul>
37		<b>Callipeltin B</b> 2006 <sup>213</sup>	solid phase acid (4 equiv.), MSNT (4 equiv.), <i>N</i> -Me-imidazole (8 equiv.), rt, 5 min then ROH (resin bound), N <sub>2</sub> , rt, 2.5 h	quant.	
38		<b>Didemmins A, B, C and Tamandarin A</b> 1990 <sup>214</sup> and 1999 <sup>215</sup>	isoprenyl chloroformate, TEA, DMAP (cat.), 0 °C, 1 h  <i>Alternatively:</i> TCBC, TEA, THF, rt, 1 h then ROH, DMAP, C <sub>6</sub> H <sub>5</sub> , rt, 4 h	87%  81%	Castro's esterification procedure <sup>216</sup>  <i>Unsuccessful:</i> <ul style="list-style-type: none"> <li>• carbodiimides: poor yields after long reaction times (48 h)</li> </ul>
39		<b>Aspercyclide C</b> 2005 <sup>217</sup>	<i>N</i> -methyl-2-chloropyridinium iodide, TEA, MeCN, reflux <sup>b</sup>	82%	

40		<b>Lyngbyaloside B macrocycle</b> 2013 <sup>96</sup>	ROH (1.2 equiv.), PhMe, reflux, 12 h azeotropic removal of EtOH	77%	transesterification from ethylester and tertiary alcohol via an acyl ketene intermediate (thermolysis of the ethylester)  <i>Lower yield:</i> <ul style="list-style-type: none"> <li>• equimolar amounts of alcohol and acid component (68%)</li> </ul>
41		<b>Lobatamide C<sup>1</sup></b> 2002 <sup>218</sup>	R = H: ROH (1 equiv.), NBu <sub>4</sub> OH, MeOH, rt, 20 min, azeotropic removal of water, then cyanomethylester of the acid component (1 equiv.), Na <sub>2</sub> CO <sub>3</sub> , DMF/2-butanone 80 °C, 2 h	43% <sup>f</sup>	<i>Unsuccessful:</i> <ul style="list-style-type: none"> <li>• for R = Et: base catalyzed fragment couplings: elimination side reactions</li> </ul>
42		<b>Latrunculin B, C, and M</b> 2007 <sup>219</sup>	ROH, Tf <sub>2</sub> O (1.4 equiv.), pyridine (2.3 equiv.), CH <sub>2</sub> Cl <sub>2</sub> -20 °C, 1 h then Na salt of the acid (3.5 equiv.), [15]crown-5, THF rt, o/n	58% (2 steps)	small amounts of an elimination product as byproduct  <i>Unsuccessful:</i> <ul style="list-style-type: none"> <li>• Mitsunobu esterification</li> </ul>
43		<b>(+)-TMC-151C</b> 2011 <sup>220</sup>	Piv <sub>2</sub> O, TEA, CH <sub>2</sub> Cl <sub>2</sub> , 0 °C, 14 h then ROH, DMAP, PhMe, rt, 2 days	60% (2 steps)	

a: information concerning the stoichiometry is only given, when one of the coupling partners was used in a > 3-fold excess or if strict demands on the stoichiometry were reported; b: no detailed information provided; c: based on 50% conversion. 60% yield at 85% conversion; d: 2 steps, starting from the methylester of the acid; e: based on 90% conversion of the alcohol; f: after deprotection of a DEIPS group

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