Protein-Induced Structural Evolution of Silver Sulfide at

Nanoscale: From Hollow Particles to Solid Spheres

Jun Chen^a, Yifei Kong^b, Jiajia ji^a, Jing Ruan^a, Kan Wang^a, Feng Gao^a and Daxiang Cui^{*a}

^a Department of Bio-Nano Science and Engineering, National Key Laboratory of Nano/Micro Fabrication Technology, Key Laboratory for Thin Film and Microfabrication of Ministry of Education,Institute of Micro/Nano Science and Technology, Shanghai Jiao Tong University, Shanghai, 200240, P. R. China.

^b School of Chemistry and Astbury Centre for Structural Molecular Biology, University of Leeds, LS2 9JT, Leeds, United Kingdom.

Supporting Information

Experiment Section

Chemicals: Bovine pancreatic RNase A was purchased from Sigma-Aldrich. Silver nitrate (AgNO₃) (\geq 99.8%, M_w = 169.87 A.R.) and thioacetamide (TAA) (\geq 99.0%, M_w = 75.13) were purchased from Shanghai Chemical Reagents Company. Solutions were prepared by using purified water as the solvent with a resistivity of 18 m Ω . All chemicals were used as received without further purification.

Synthesis Ag_2S nanospheres: AgNO₃ (5 mL, 50 mM) was introduced into 5 mL of 12.5 mgmL⁻¹ RNase A solution with vigorous stirring. The precursor solution was then kept at room temperature for pumping vacuum for 30 min. After that, freshly prepared TAA solution (5 mL, 50 mM) was injected. The as-prepared samples were stored in dark at 60°C.

Characterization: X-ray powder diffraction (XRD) measurements were performed on a Bruker D&Advance X-ray powder diffractometer with graphite monochromatized Cu K α (λ =0.15406 nm). Scanning electron microscopy (SEM) images were taken on a JSM-5600LV scanning electron microscope equipped with an X-ray energy dispersive spectroscopy (EDS) at an accelerating voltage of 20 kV. Transmission electron microscopy (TEM), accompanied by selected-area electron diffraction (SAED), was carried out on a JEOL JEM 100CX-II transmission electron microscope; high resolution TEM (HRTEM) images were taken on a JEOL JEM-2010 transmission electron microscope. The UV–vis absorbance of the samples was measured on a Shimadzu UV-2450 UV–visible spectrophotometer. The FT-IR spectra were recorded on a Bio-Rad FTS-40 Fourier transform infrared spectrograph in the wavelength range of 4000~950 cm⁻¹. Circular dichroism (CD) spectra (190-260 nm) were recorded on a Jasco J-810 spectropolarimeter. The native polyacrylamide gel electrophoresis (PAGE) for RNase A and different stage Ag₂S samples were both run in a 15% acrylamide gel at 130 V for 80 min. Atom force microscopy (AFM) imaging was performed using a Digital Instruments Multimode Scanning Probe Microscope equipped with a Nanoscope IIIA controller and a vertical engage J-scanner.



Fig.S1 HR-TEM images of a) hollow and b) solid Ag₂S NPs.



Fig.S2 SEM images of Ag₂S NPs stored for a)20 days and b) 30 days. Scale bar is 100 nm. Size distribution of Ag₂S NPs stored for c) 1 day, d) 20 days and e) 30 days.







Fig.S4 EDS spectra of Ag₂S NPs stored for a) 1 day, b) 20 days and c) 30 days.

Assignments	-OH (cm ⁻¹)	amide A (cm ⁻¹)	amide I (cm ⁻¹)	amide II (cm ⁻¹)	-NO ₃ (cm ⁻¹)
Pure RNase A	3411	3150	1647	1539	
RNase $A-Ag^+$	3273	3273	1636	1516	1369
RNase A-Ag ₂ S	3423		1624	1542	1317

Table S1. The main FT-IR peaks of pure RNase A, RNase A-Ag+ and RNase A-Ag₂S (30 days).

Table S2. The percentages of the secondary structures of pure RNase A, RNase A-Ag⁺ and RNase A-Ag₂S (30 days) corresponding to the CD spectra

and Krase <i>R-Rg</i> ₂ 5 (50 days) corresponding to the CD spectra.							
Assignments	α-helix (%)	β-sheet (%)	turn (%)	random (%)			
Pure RNase A	9.7	31.4	12.5	38.9			
RNase $A-Ag^+$	13.1	30.8	12.5	37.9			
RNase A-Ag ₂ S	20.8	25.8	12.5	39.5			



Fig.S5 AFM image of as-prepared Ag_2S NPs stored for 30 days



Fig.S6 a) TEM and b) SEM images of Ag₂S precipitation without RNase A.



Fig.S7 a) TEM image of Ag₂S NPs stored for 20 days at a) 40°C and b) 80°C.



Fig. S8 TEM images of Ag₂S NPs stored for a) 20 days and b) 25 days.