Supplementary Information

Encoding Molecular Information in Plasmonic Nanostructures for Anti-counterfeiting Application

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Fig. S1 Tapping mode AFM topographic image of (A) silver film, (C) silver nanowires and the corresponding cross-section profles of (B) silver film and (D) nanowires. (E) Tilted SEM image of the as-prepared Ag nanowires – 100-nm Ag deposited on polymer nanowires. (F) Schematic of the nanowire used in simulation.



Fig. S2 2D Raman imaging "Taiji" patterns functionalized with (A - D) 2-NT molecules and (E - H) 4-MBT molecules. (A, B, E, F) SERS mapping images collected at 1078 cm⁻¹, a Raman window where both molecules have overlapping molecular vibrations. (C, D, G, H) SERS mapping images collected at 1381 cm⁻¹, a unique fingerprint spectral window that is specific to 2-NT, and not observed in 4-MBT. (A, C, E, G) Raman mappings performed using *x*-polarization. (B, D, F, H) Raman mappings performed using *y*-polarization. Each Raman map area is $60 \times 60 \ \mu\text{m}^2$. All scale bars are 10 μm .



Figure S3. The darkfield optical images of silver "Taiji" structures functionalized with 2-NT and 4-MBT. All scale bars are $10 \,\mu$ m.

Calculation of SERS enhancement factor

The experimental Raman enhancement factor (EF) for the Ag coated polymer substrate is estimated using the equation: $\text{EF} = \left(\frac{I_{\text{SERS}}}{N_{\text{SERS}}}\right) / \left(\frac{I_{\text{ref}}}{N_{\text{ref}}}\right)$, where I_{SERS} is the intensity of the specific

Raman band from the analyte adsorbed on a SERS active substrate, and N_{SERS} is the number of molecules contributing to I_{SERS}. Similarly, I_{ref} is the intensity of the same Raman band from the bulk analyte, normalized with the laser power and acquisition time, and N_{ref} is the number of molecules that yield I_{ref}. Here, 1079 cm⁻¹ Raman band is selected for EF calculation. For a more comprehensive estimate of EF, we utilize the average intensity from 10 sample points of the substrate. The average I_{SERS} is around 337.8 counts per second (cps). The radius of the diffraction-limited laser beam is obtained by measuring the intensity profile of 100-nm fluorescence beads. The diameter of the laser beam is measured at ~520 nm, and the area is A = $\pi \times r^2 = 2.12 \times 10^5$ nm². Taking 4.5×10^{14} molecules/cm² for a monolayer of 4-MBT on silver (assume the binding of 4-MBT on silver is similar to that of gold),¹ the number of molecules excited under the laser spot is approximated at 9.56 ×10⁵ molecules.

The average I_{ref} is measured at 0.57 cps using an 1 M 4-MBT ethanolic solution as a reference solution. For the determination of N_{ref} , the confocal volume is obtained by measuring the intensity profile of fluorescence beads immersed in ethanol solution, which simulates the condition of normal Raman signal (I_{ref}) measurement. The measured laser diameter and the focal depth (*h*) are 910 nm and 4320 nm, respectively. The effective excitation volume (*V*) is $V = A \times h$ 2.81 × 10⁹ nm³. The larger confocal volume in ethanol solution than that in ideal dry condition is caused by the distortion of laser spot in solution.²⁻⁶ The number of 4-MBT molecules in ethanol (1 mol·L⁻¹) excited by the laser beam is therefore $N_{ref} = 1.69 \times 10^9$ molecules. The average enhancement factor, based on mean intensity from 10 sample points, is estimated to be ~ 1.05 × 10⁶.

REFERENCES

- 1. K. Seo and E. Borguet, J. Phys. Chem. C, 2007, 111, 6335-6342.
- 2. J. S. Teguh, F. Liu, B. G. Xing, E. K. L. Yeow, *Chem.-Asian J.* 2011, 7, 975-981.
- 3. R. A. Alvarez-Puebla, J. Phys. Chem. Lett. 2012, 3, 857-866.

- 4. N. J. Everall, *Analyst* 2010, **135**, 2512-2522.
- 5. H. X. Lin, J. M. Li, B. J. Liu, D. Y. Liu, J. X. Liu, A. Terfort, Z. X. Xie, Z. Q. Tian and B. Ren, *Phys. Chem. Chem. Phys.*, 2013, **15**, 4130-4135.
- 6. X. M. Lin, Y. Cui, Y. H. Xu, B. Ren and Z. Q. Tian, *Anal. Bioanal. Chem.*, 2009, **394**, 1729-1745.