Supplementary Information

Cytotoxicity of nucleus-targeting fluorescent gold nanoclusters

Jing-Ya Zhao,^a Ran Cui, ^a Zhi-Ling Zhang, ^a Mingxi Zhang, ^b Zhi-Xiong Xie^c and

Dai-Wen Pang *a

aKey Laboratory of Analytical Chemistry for Biology and Medicine (Ministry of Education),

College of Chemistry and Molecular Sciences, State Key Laboratory of Virology, and Wuhan

Institute of Biotechnology, Wuhan University, Wuhan, 430072, P. R. China

^bState Key Laboratory of Advanced Technology for Materials Synthesis and Processing, Wuhan

University of Technology, Wuhan 430070, P. R. China

^cCollege of Life Sciences, Wuhan University, Wuhan, 430072, P. R. China

*Corresponding author. Tel.: +86 27 68756759; fax: +86 27 68754067.

E-mail address: dwpang@whu.edu.cn (D. W. Pang)

Figures



Fig. S1. Fluorescence photograph (A) and intensity (B) of $Au_{25}NCs$ in BR buffer at pH value from 4.5 to 11.5. (C) Fluorescence intensity of $Au_{25}NCs$ in aqueous solution with different concentration of NaCl. (D) Fluorescence spectra of $Au_{25}NCs$ stored at 4 °C for different time.



Fig. S2. Time evolution of the photoluminescence spectra for the preparation of fluorescent AuNCs by adopting Au(I) complex (A) and HAuCl₄ (B) as gold precursor.



Fig. S3. MALDI-TOF mass spectra of WGA and $Au_{25}NCs$. The MALDI-TOF mass spectra were performed using matrixes of α -cyano-4-hydroxycinnamic acid in positive ion mode.



Fig. S4. Hydrodynamic sizes of Au₂₅NCs before (A) and after (B) conjugation with streptavidin.



Fig. S5. Fluorescence microscopic images of A549 cells (a-c), Vero cells (d-f) and MDCK cells (g-i) treated with TAT-Au₂₅NCs (A), SA-Au₂₅NCs (B), a mixture of biotin-TAT and Au₂₅NCs (C) and Au₂₅NCs (D).



Fig. S6. (A) Photostability comparison between TAT-Au₂₅NCs and Alexa 488-IgG, Syto 13 and Syto82. TAT-Au₂₅NCs: Cal 27 cells were incubated with 2.4 μmol/L TAT-Au₂₅NCs for 1 h at 37 °C and then the cells were fixed by paraformaldehyde; Alexa 488-IgG: Paraformaldehyde-fixed and BSA-blocked Cal 27 cells were incubated sequentially with a mouse anti-epithelial-cell-adhesion-molecular antibody and Alexa 488 conjugated to goat anti-mouse immunoglobulin G (Alexa 488-IgG) for 1 h each; Syto 13: Cal 27 cells were incubated with 500 nM Syto 13 nucleic acid dye for 30 min at 37 °C and then the cells were fixed by paraformaldehyde; Syto 82: Cal 27 cells were incubated with 500 nM Syto 13 nucleic acid dye for 30 min at 37 °C and then the cells were fixed by paraformaldehyde. Then the specimens were continuously illuminated for 3 min with a 488 nm laser (laser energy 13 mW). Fluorescence images were acquired using an Andor Revolution XD laser confocal microscope at 5 s intervals. (B) Photostability curves of TAT-Au₂₅NCs, Alexa 488-IgG, Syto 13 nucleic acid dye and Syto 82 nucleic acid dye. Quantitative analysis of the changes in fluorescence intensities were performed by the ImageJ software.



Fig. S7. MTT assay of A549 cells, MDCK cells and Vero cells treated with different concentrations of biotin-TAT (A), TAT-SA (B) and Au₂₅NCs(C) for 24 h.



Fig. S8. Microscopic images of untreated (A), biotin-TAT treated (B) and TAT-SA treated (C) Vero cells at 0 min (a) and 420 min (b) post-treating. Vero cells were incubated with 4.8 μ mol/L biotin-TAT (B) and 2.4 μ mol/L TAT-SA (C) for 1 h and then were supplied with fresh medium and observed on the microscope.



Fig. S9. Appoptosis assay on cells exposed to different concentrations of TAT-Au₂₅NCs for 2 h. Cells treated with 1 mM H_2O_2 and 0.1 μ M actinomycin D were used as the necrosis positive control and apoptosis positive control, respectively.