Electronic Supplementary Material (ESI) for Nanoscale. This journal is © The Royal Society of Chemistry 2015

#### **Electronic Supplementary Information**

# Bipyridine hydrogel for selective and visible detection and absorption of Cd<sup>2+</sup>

Qingqing Miao,<sup>a</sup> Ziye Wu,<sup>c</sup> Zijuan Hai,<sup>a</sup> Changlu Tao,<sup>d</sup> Qingpan Yuan,<sup>a</sup> Yadi Gong,<sup>a</sup> Yafeng Guan,<sup>b</sup> Jun Jiang, \*<sup>c</sup> and Gaolin Liang \*<sup>a</sup>

<sup>a</sup>CAS Key Laboratory of Soft Matter Chemistry, Department of Chemistry, University of Science and Technology of China, Hefei, Anhui 230026, China

<sup>b</sup>Key Laboratory of Separation Sciences for Analytical Chemistry, Dalian Institute of Chemical Physics, Chinese Academy of Sciences, Dalian 116023, China

<sup>c</sup>Department of Chemical Physics, University of Science and Technology of China, 96 Jinzhai Road, Hefei, Anhui 230026, China.

<sup>d</sup>Center for Integrative Imaging, Hefei National Laboratory for Physical Sciences at the Microscale & School of Life Sciences, University of Science and Technology of China, Hefei, Anhui 230026, China

E-mail: jiangj1@ustc.edu.cn (J.J.), gliang@ustc.edu.cn (G.-L. L.).

## **Contents:**

- 1. Synthetic route for 1.
- 2. Supporting Figures.
- 3. Supporting Tables.
- 4. References.

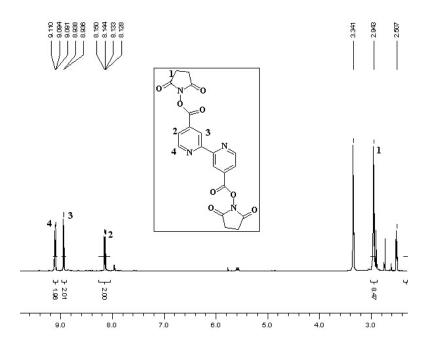
## 1. Synthetic route for 1.

Preparation of 1.

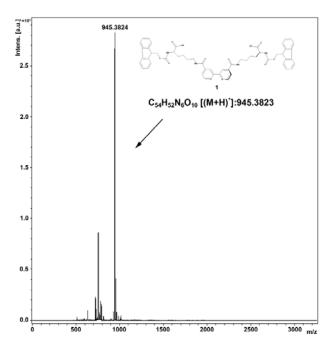
Scheme S1. Synthetic route for compound 1.

Synthesis of 4,4'-Dicarboxysuccinimidyl-2,2'-bipyridine:

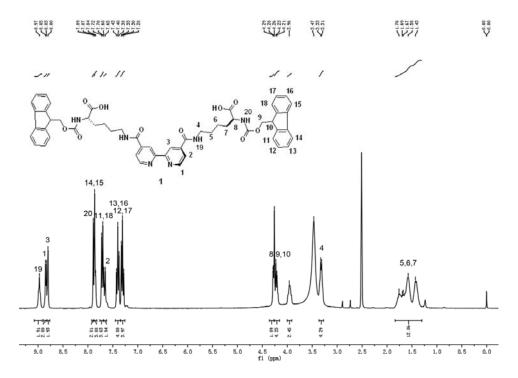
# 2. Supporting Figures.



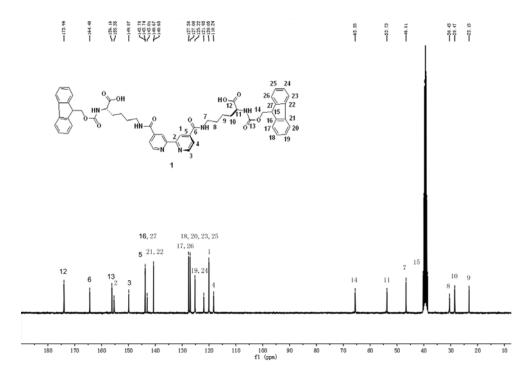
**Figure S1**. <sup>1</sup>H NMR spectrum of compound 4,4'-dicarboxysuccinimidyl-2,2'-bipyridine.



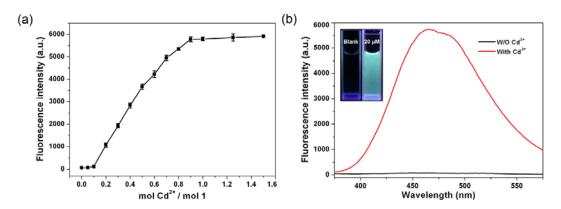
**Figure S2**. HR-MALDI-TOF/MS spectrum of **1**.



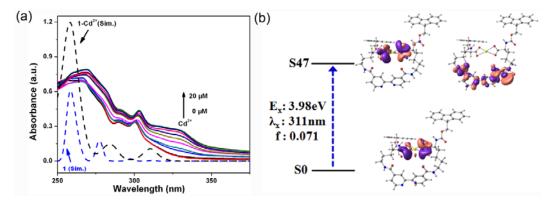
**Figure S3**. <sup>1</sup>H NMR spectrum of compound **1**.



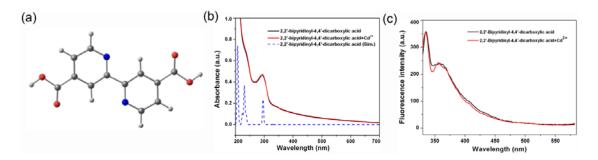
**Figure S4**. <sup>13</sup>C NMR spectrum of compound **1**.



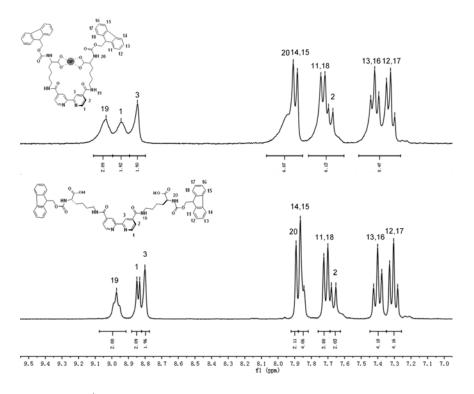
**Figure S5.** (a) Correlation of fluorescence intensities at 470 nm of **1** (20 μM,  $\lambda_{ex}$  = 300 nm) in the presence of various concentrations of Cd<sup>2+</sup> (0, 1, 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 25 or 30 μM) in phosphate buffer (10 mM, pH 7.5) containing 10% ethanol at RT. (b) Fluorescence spectra of **1** (20 μM,  $\lambda_{ex}$  = 300 nm) in the presence of 0, or 20 μM Cd<sup>2+</sup> in phosphate buffer (10 mM, pH 7.5) containing 10% ethanol at RT. The inset fluorescent photographs show the fluorescence changes of **1** at 20 μM before and after addition of 20 μM Cd<sup>2+</sup> under a UV lamp.



**Figure S6.** (a) UV-vis absorption spectra of **1** (20  $\mu$ M) in the presence of various concentrations of Cd<sup>2+</sup> in phosphate buffer (10 mM, pH 7.5) containing 10% ethanol at RT. The simulated (Sim.) spectra of **1** and **1**-Cd<sup>2+</sup> are given with dashed curves. (b) The transition energy and orbitals responsible for the absorption peak at around 311 nm of **1**-Cd<sup>2+</sup>.



**Figure S7.** (a) Theoretically optimized molecular structure of 2,2'-bipyridinyl-4,4'-dicarboxylic. (b) UV-vis absorption spectra of 2,2'-bipyridinyl-4,4'-dicarboxylic acid (20 μM) in the presence of  $Cd^{2+}$  at 0, or 20 μM in phosphate buffer (10 mM, pH 7.5) containing 10% ethanol at RT. The simulated spectrum is given as blue dashed line. (c) The corresponding fluorescence spectra of 2,2'-bipyridinyl-4,4'-dicarboxylic acid (20 μM) in the presence of  $Cd^{2+}$  at 0, or 20 μM, excited at 300 nm.



**Figure S8.** <sup>1</sup>H NMR spectra (300 MHz) of **1** (bottom) and **1** upon addition of  $Cd^{2+}$  (top) in  $d_6$ -DMSO.

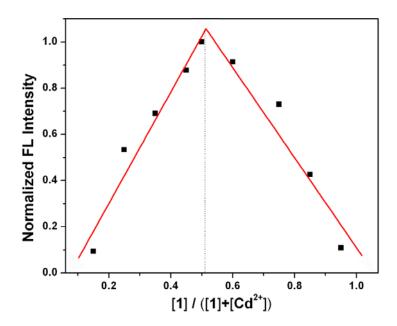
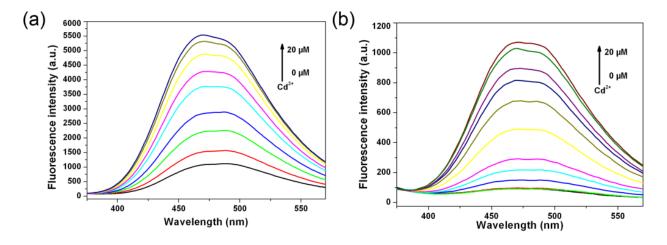
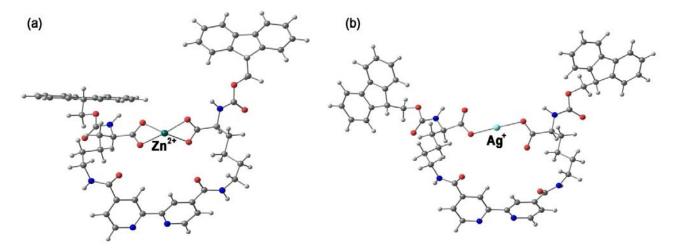


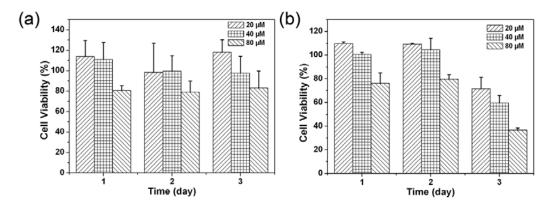
Figure S9. Job' plots of fluorescence intensity changes at varying mole ratios of 1 with  $Cd^{2+}([1] + [Cd^{2+}] = 40 \mu M)$ .



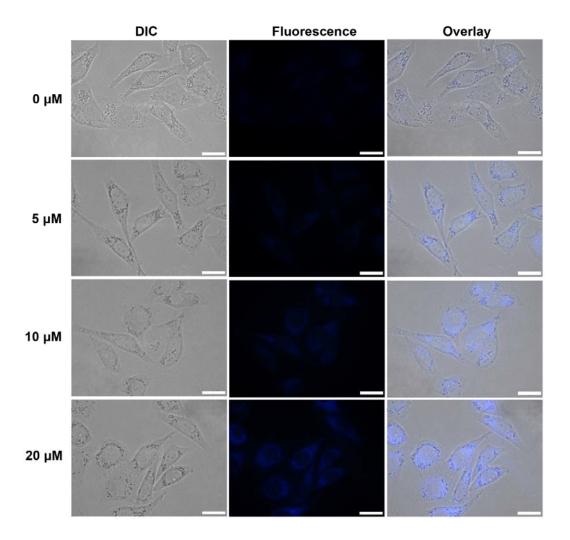
**Figure S10.** (a) Fluorescence spectra of **1** (20  $\mu$ M,  $\lambda_{ex}$  = 300 nm) in the presence of various concentrations of Cd<sup>2+</sup> in phosphate buffer (10 mM, pH 6) containing 10% ethanol at room temperature. (b) Fluorescence spectra of **1** (20  $\mu$ M,  $\lambda_{ex}$  = 300 nm) in the presence of various concentrations of Cd<sup>2+</sup> in phosphate buffer (10 mM, pH 9) containing 10% ethanol at room temperature.



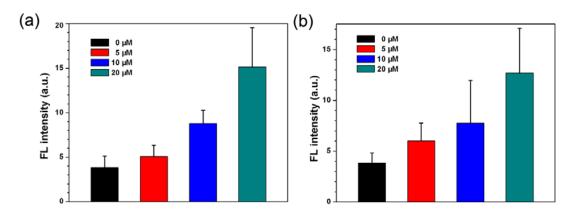
**Figure S11.** (a)  $Zn^{2+}$  forms effective bondings with four oxygen atoms in the **1**- $Zn^{2+}$  complex. (b)  $Ag^{+}$  forms weakly bondings with two oxygen atoms in the **1**- $Ag^{+}$  complex.



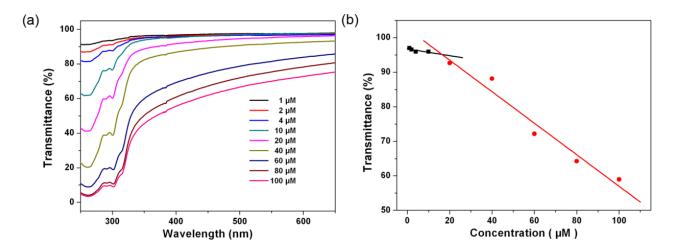
**Figure S12.** MTT assay of **1** on (a) HepG2 cells, and (b) LoVo cells. Cell viability values (%) estimated by MTT proliferation test at concentrations of 20, 40 and 80 μM of **1**. HepG2 cells, or LoVo cells were cultured in the presence of **1** for 1, 2 and 3 day at 37 °C under 5% CO<sub>2</sub>. These experiments were performed in triplicate. Results are representative of three independent experiments. Error bars represent standard deviations.



**Figure S13.** Differential interference contrast (DIC) images (left), fluorescence images (middle, DAPI channel), and merged images (right) of LoVo cells incubated with 0, 5, 10, or 20  $\mu$ M of Cd<sup>2+</sup> in serum-free medium for 30 min at 37 °C, washed with PBS for three times, then incubated with 20  $\mu$ M 1 in serum-free medium for 0.5 h at 37 °C prior to imaging, respectively. Scale bar: 20  $\mu$ m.



**Figure S14.** (a) The average fluorescence intensity of HepG2 cells in Figure 4. (b) The average fluorescence intensity of LoVo cells in Figure S13.



**Figure S15**. (a) UV-vis transmittance spectra of **1** at various concentrations in water (pH 5.5). (b) Concentration-dependent optical transmittance of **1** at 425 nm in water (pH 5.5).

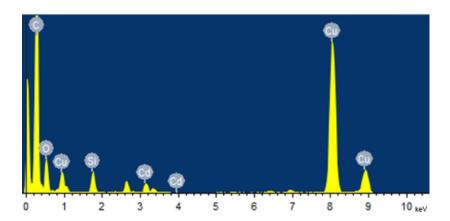
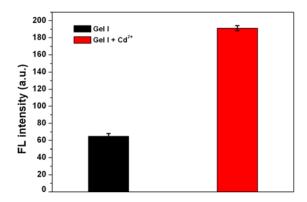


Figure S16. Energy-dispersive X-ray spectroscopic (EDS) elemental analysis of nanofibers in Figure 6c.



**Figure S17.** The average fluorescence intensity in Figures 6b & d.

# 3. Supporting Tables.

**Table S1**. HPLC condition for the purification of **1**.

| Time (minute) | Flow (mL/min.) | H <sub>2</sub> O % | CH <sub>3</sub> CN% |
|---------------|----------------|--------------------|---------------------|
| 0             | 3.0            | 30                 | 70                  |
| 3             | 3.0            | 30                 | 70                  |
| 35            | 3.0            | 0                  | 100                 |
| 37            | 3.0            | 0                  | 100                 |
| 38            | 3.0            | 30                 | 70                  |
| 40            | 3.0            | 30                 | 70                  |

**Table S2**. Using **1** to detect of Cd<sup>2+</sup> in pond water (from east campus of USTC).

| Sample        | Level added (µM) | Level found (µM) | Recovery (%) | SD (µM) | RSD (%) |
|---------------|------------------|------------------|--------------|---------|---------|
| pond water 10 |                  | 9.71             | 97           | 0.039   | 0.406   |
|               | 10               | 9.66             | 96           |         |         |
|               |                  | 9.64             | 96           |         |         |

**Table S3**. Statistics of sensitivity of **1** and other reported methods for the detection of Cd<sup>2+</sup>.

|  | LOD   | The limit of detection (LOD) of Cd <sup>2+</sup>       |  |  |
|--|---|--|--|--|
| Method   |   |  |  |  |
|  | In this paper   | $2.10 \times 10^{-8} \mathrm{M}$                       |  |  |
| Fluorometry  | Cheng <i>et al.</i> reported a small molecule-based chemosensors <sup>1</sup> | $1.00 \times 10^{-7} \text{ M}$                        |  |  |
|  | Varriale <i>et al.</i> reported protein-based sensing systems <sup>2</sup>    | $5.00 \times 10^{-7} \text{ M}$                        |  |  |
| Atomic absorp  | otion spectrometry (AAS) <sup>3</sup>   | $4.20 \times 10^{-11} \text{ M}$                       |  |  |
| Inductively co   | oupled plasmas-Atomic fluorescence spectrometry                               |  |  |  |
| (ICP-AFS) <sup>4</sup>   |   | $3.56 \times 10^{-9} \mathrm{M}$                       |  |  |
| Inductively  | coupled plasmas-atomic emission   |  |  |  |
| spectrometry(  | ICP-AES) <sup>5</sup>   | $1.78 \times 10^{-6} \mathrm{M}$                       |  |  |
| Inductively coupled plasma mass spectrometry (ICP-MS) <sup>6</sup> |   |  |  |  |
|  |   | $8.90 \times 10^{-11} \text{ M}$                       |  |  |
| X-ray fluoresc   | rence <sup>7</sup>  | $1.78 \times 10^{-4} - 1.27 \times 10^{-3} \mathrm{M}$ |  |  |
| Liquid chi   | romatography with electrochemical or  | $8.90 \times 10^{-7}$ M for electrochemical            |  |  |
| spectrophotometric detection <sup>8</sup>                          |   | detection;   |  |  |
|  |   | $1.78 \times 10^{-8}$ M for spectrophotometric         |  |  |
|  |   | detection  |  |  |
| Surface enhanced Raman scattering (SERS) <sup>9</sup>              |   | $1.00 \times 10^{-6} \mathrm{M}$                       |  |  |

#### 4. References.

- 1 T. Y. Cheng, Y. F. Xu, S. Y. Zhang, W. P. Zhu, X. H. Qian and L. P. Duan, *J. Am. Chem. Soc.*, 2008, **130**, 16160-16161.
- 2 A. Varriale, M. Staiano, M. Rossi and S. D'Auria, Anal. Chem., 2007, 79, 5760-5762.

- 3 D. Colbert, K. S. Johnson and K. H. Coale, Anal. Chim. Acta 1998, 377, 255-262.
- 4 A. Montaser and V. A. Fassel, Anal. Chem., 1976, 48, 1490-1499.
- 5 A. C. Davis, C. P. Calloway and B. T. Jones, *Talanta*, 2007, **71**, 1144-1149.
- 6 S. N. Willie, Y. Iida and J. W. McLaren, Atom. Spectrosc., 1998, 19, 67-72.
- 7 L. Ahlgren and S. Mattsson, *Phys. Med. Biol.*, 1981, **26**, 19-26.
- 8 A. M. Bond and G. G. Wallace, Anal. Chem., 1984, 56, 2085-2090.
- 9 J. Yin, T. Wu, J. B. Song, Q. Zhang, S. Y. Liu, R. Xu and H. W. Duan, *Chem. Mater.*, 2011, **23**, 4756-4764.