Septanose Carbohydrates

Recognition of Septanose Carbohydrates by Concanavalin A

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| | |

Details of STD Experiments 500MHz Brüker

| Current Data | Parameters |
|---------------|--------------------|
| NAME | MWP-ii-150-STD |
| PROCNO | 1 |
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| USER | IIIIII Su |
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| FZ-ACQUISICIO | |
| Date | 20050208 |
| INSTRUM | spect |
| | |
| PULPROG | stddiffesgp.3~ |
| TD | 2048 |
| NS | 128 |
| SWH | 3501.401 Hz |
| FW | 125000.00 Hz |
| RG | 200 |
| DMOV | 2.975 usec |
| DIGTYP | HADC+ |
| DR | 18 |
| DE | 6.00 usec |
| D1 | 7.00000000 sec |
| d12 | 0.00002000 sec |
| D20 | 7.0000000 sec |
| d31 | 7 00000000 500 |
| | 7:00000000 sec |
| | 2 |
| NBL | 12 50 50 |
| PI P10 | 13.50 usec |
| P12 | 2000.00 usec |
| p29 | 50000.00 usec |
| PL1 | -1.00 dB |
| PL29 | 4.35 dB |
| SP1 | 37.00 dB |
| SPOFF1 | 0.00 Hz |
| NUC2 | 1H |
| PL2 | 120.00 dB |
| SP13 | 37.00 dB |
| SPOFF13 | 0.00 Hz |
| GPNAM2 | SINE.100 |
| GPX1 | 0.00 % |
| GPX3 | |
| CDV2 | |
| GF12 CDF1 | |
| GP21 | 40.00 8 |
| GPZ3 | 11.00 % |
| P30 | 3000.00 usec |
| F1_Acquistion | Parameters |
| | 1 |
| SEO1 | L 500 1224 MTT- |
| STUL | GH 1 000 |
| DOM DOM | SW I.999 ppm |
| FOV | 20.00 cm |
| F2-Processing | Parameters |
| ST | 4096 |
| SF | 500.1300000 MHz |
| SD SD | |
| 5K | 0.00 HZ |

| Time | 11.42 | | | |
|-------------------------|--------------------------------|--|--|--|
| PROBHD | 5mm BBO BB-IH Z-GRD Z8007/0104 | | | |
| AQ_mod SOLVENT DS | DQD D2O 8 | | | |
| FIDRES | 1.709668 Hz | | | |
| AQ | 0.2926472 sec | | | |
| DW | 142.800 usec | | | |
| DECIM | 48 | | | |
| DIGMOD | digital | | | |
| DDR | 2 208 2 K | | | |
| 1E d11 | 0 0300000 sec | | | |
| D16 | 0.00020000 sec | | | |
| D29 | 0.05000000 sec | | | |
| DELTA1 | 0.00000000 sec | | | |
| 15 | 140 | | | |
| NUC1 | 1H | | | |
| p2 | 27.00 usec | | | |
| P17 | 2000.00 usec | | | |
| PL0 | 120.00 dB | | | |
| PL10 | 4.35 dB | | | |
| SF01 | 500.1323541 MHz | | | |
| SPNAM1 | Squa100.1000 | | | |
| FQ2LIST | freqlisti | | | |
| PI3 GEO2 | 50000.00 USEC | | | |
| SFUZ SDNAM13 | Gaug1 100 | | | |
| GPNAM1 | Sine.100 | | | |
| GPNAM3 | Sine.100 | | | |
| GPX2 | 0.00 % | | | |
| GPY1 | 0.00 % | | | |
| GPY3 | 0.00 % | | | |
| GPZ2 | 31.00 % | | | |
| P16 | 1000.00 usec | | | |
| ጥD | 2 | | | |
| FIDRES | 500.000000 Hz | | | |
| SWH | 1000.00 Hz | | | |
| FnMODE | QF | | | |

| PPARMOD | 2D | 2D NMR | Plot Parameters |
|---------------|-----------------|--|-----------------|
| OFFSET | 8.207 ppm | CX2 | 15.00 cm |
| HzpPT | 0.854834 Hz | CX1 | 15.00 cm |
| - | | F2PLO | 9.800 ppm |
| | | F210 | 4901.27 Hz |
| | | F2PHT | mqq 0.2.0- |
| พกพ | гм | тонт 1 21 111 1 12 111 | -100 03 Hz |
| | | | 9 800 000 |
| | 0.50 HZ | I IIIO | 3.800 ppm |
| PH_MOO | | 55B | 0 |
| PHCU | 0.000 degrees | GB | 0 |
| BC_mod | no | PKNL | TRUE |
| FT_mod | fqc | PHC1 | 0.000 degrees |
| ME_mod | no | BCFW | 0.000 ppm |
| NCOEF | 0 | FCOR | 0.5 |
| ABSF1 | 0.000 ppm | COROFF | S 0.00 Hz |
| ABSG | 0 | LPBIN | 0 |
| AZFE | 0.100 ppm | ABSF2 | 0.000 ppm |
| Tdeff | 2048 | ABSL | 0 |
| STSR | 0 | AZFW | 0.100 mag |
| DSCAL | srea | That we have a second s | 0.100 ppm |
| DC | 1 00 | | 4006 |
| PC | 1.00 | 5151 | 4090 |
| ML | 0.00 Cm | SREGLS | T IH.CDCL3 |
| INTBC | yes | PSIGN | pos. |
| ISEN | 128 | MAXI | 10000.00 cm |
| AUNMP | proc_1d | INTSCL | 1 |
| NOISF1 | 0.000 ppm | REVERS | E FALSE |
| SIGF1 | 0.000 ppm | SINO | 400 |
| ASSFAC | 0 | NOISF2 | 0.000 ppm |
| ASSFACX | 0 | SIGF2 | mqq 000.0 |
| ратмор | proc | ASSFAC | т 0 |
| NSD | p100 | ASSWID | - - 0 |
| | 0 | DC DC | 0 |
| | 0 | | 0 |
| TMI | 0 | NZP | 0 |
| ALPHA | 0 | TI | |
| YMAX_p | 5868959 | TM2 | 0 |
| C_proc | -1 | GAMMA | 0 |
| S_DEV | 85793.000 | YMIN_p | 0 |
| NLEV | 6 | MEAN | 0.000 |
| TOPLEV | 0.00 % | AQORDE | R 3-2-1 |
| | | LEV0 | 0.00 |
| F1 Processing | Parameters | | |
| ST | 1024 | | |
| SF | 500.1300000 MHz | | |
| SP | 0 00 Hz | MC 2 | OF |
| SK SW p | | | 5 707 ppm |
| aw_b | 1000.00 HZ | UFFSEI | 0.076562 |
| WDW | SINE | HZPPT | 0.976562 |
| LB | 0.00 HZ | XDIM | 1024 |
| Tdeff | 2 | SSB | 0 |
| STSR | 0 | GB | 0 |
| BC_mod | no | TDoff | 0 |
| PHC0 | 0.000 degrees | STSI | 1024 |
| ABSF1 | mqq 000.0 | PH mod | mc |
| ABSG | 0 | PHC1 | 0.000 degrees |
| FT mod | fac | ARSF2 | 0.000 mman |
| SYMM | -90 -90 | ARST | 0.000 ppm |
| тм1 | 110 | | ਹ ਸ ਦੁਆਰਦ |
| ME mod | 0 | | E FALSE |
| | no | TILT | FALSE |
| TLRIN | 0 | 'I'M2 | 0 |
| | | NCOEF | 0 |

| F1LO | 4901.27 Hz |
|---------------|-----------------|
| F1PHI | -0.200 ppm |
| F1HI | -100.03 Hz |
| F2PPMCM | 0.66667 ppm/cm |
| F1PPMCM | 0.66667 ppm/cm |
| F1HZCM | 333.41998 Hz/cm |
| scaling in F1 | [ppm/cm] |



References and details for the preparation of 3-7

The preparation of **3-6** have been reported in the literature.

3, **6**: DeMatteo, M.; Snyder, N. L.; Morton, M.; Baldisseri, D. M.; Hadad, C. M.; Peczuh, M. W.; Septanose Carbohydrates: Synthesis and Conformational Studies of Methyl- α -D-*Glycero*-D-idoseptanoside and Methyl β -D-*Glycero*-D-guloseptanoside. *J. Org. Chem.* **2005**, *70*, 24-38.

4: Castro. S.; Peczuh, M. W.; Sequential Cyclization-Elimination Route to Carbohydrate Based Oxepines *J. Org. Chem.* **2005**, *70*, 3312-3315.

5: Peczuh, M. W.; Snyder, N. L.; Fyvie, W. S. Synthesis, crystal structure and reactivity of a D-xylose based oxepine. *Carbohydr. Res.* **2004**, *339*, 1163-1171.



Methyl 4,5,7-tri-O-benzyl-3-deoxy-β-D-glycero-D-guloseptanoside (S2) Oxepine S1¹ (0.050 g, 0.116 mmol) was azeotroped from toluente (3 x 5 mL), dissolved in dry CH₂Cl₂ (5 mL), and cooled to 0 °C. DMDO (0.278 mL of a 0.2 M solution in CH₂Cl₂) was added and the mixture was stirred at 0 °C for 30 min. The solvent was removed under reduced pressure and a solution of NaOCH₃ (0.010 g) in CH₃OH (5 mL) was added to the residue. The mixture was stirred overnight (12 h) at rt. The reaction was quenched with water (2 mL) and the solvent was removed under reduced pressure. The residue was dissolved in CH₂Cl₂ (15 mL) and washed with water (2 x 15 mL), dried (Na₂SO₄) and the solvent was removed under reduced pressure. The residue was removed under reduced pressure. The residue was dissolved in CH₂Cl₂ (15 mL) and washed with water (2 x 15 mL), dried (Na₂SO₄) and the solvent was removed under reduced pressure. The residue was generated pressure. The residue was purified by column chromatography (1:3 EtOAc-Hexanes) to give a clear and colorless oil (0.034 g, 61%).!R=0.65 (1:1 EtOAc-Hexanes); [α]_D -6.79° (*c* 2.40, CHCl₃); ¹H NMR 400 MHz (CDCl) δ 7.34-7.17 (m, 15H), 4.65 (d, 1H, *J* = 12.1 Hz), 4.55 (m, 3H), 4.51 (d, 1H, *J* = 2.9 Hz), 4.31 (d, 1H, *J* = 11.4 Hz), 4.17 (d, 1H, *J* = 7.5 Hz), 3.89 (ddd, 1H, *J* = 13.0, 8.9, 4.2 Hz), 3.83 (dd, 1H, *J* = 7.2, 4.4 Hz), 3.70-3.68 (m, 1H), 3.63-3.61 (m, 2H), 3.53-3.50 (m, 4H) 2.20 (ddd, 1H, *J* = 10.5, 6.9, 2.7 Hz), 1.89 (dd, 1H *J* = 14.7, 9.1 Hz); ¹³C NMR 100 MHz (CDCl) δ 138.5, 138.0, 128.6 (2), 128.5, 128.1, 128.0 (2), 127.9, 127.8 (2), 110.3, 80.7, 79.9, 76.2, 73.5, 73.0, 71.7, 71.3, 70.0, 56.2, 31.6; FAB-MS *m/z* [M+H]⁺ calcd 479.2434, found 479.2431.

Methyl 3-deoxy-β-D-glycero-D-guloseptanoside (7) 10% Pd/C (0.006 g) was added to a solution of S2 (0.034 g, 0.071 mmol) in CH₃OH (5 mL).!The reaction was placed under an H₂ atmosphere via a balloon and the mixture was stirred for 4 h at rt. The balloon was removed from the flask and the mixture was filtered through a short pad of celite.!The celite was washed with additional CH₃OH (4 x 5 mL). The solvent was removed from the combined filtrates by rotary evaporation under reduced pressure to give a clear, colorless oil (0.015 g, 98%). [α]_D -6.2° (*c* 1.54, CH₃OH); ¹H NMR 400 MHz (CD₃OD) δ 4.2 (d, 1H, *J* = 6.7 Hz), 3.84 (dd, 1H, *J* = 11.8, 2.6 Hz), 3.80-3.74 (m, 2H), 3.65 (dd, 1H, *J* = 11.8, 6.7 Hz), 3.5 (s, 3H), 3.41-3.35 (m, 1H), 3.20 (dd, 1H, *J* = 8.2, 8.2 Hz), 2.02 (ddd, 1H, *J* = 14.7, 10.5, 4.8 Hz), 1.81 (ddd, 1H, *J* = 14.5, 3.6, 1.4 Hz); ¹³C NMR 100 MHz (CD₃OD) δ 112.3, 84.2, 76.6, 71.7, 70.7, 64.3, 56.4, 35.7; FAB-MS *m*/*z* [M-H]⁺ calcd 207.0869, found 207.0886.

⁽¹⁾ Peczuh, M. W.; Snyder, N. L. Carbohydrate Based Oxepines: Ring Expanded Glycals for the Synthesis of Septanose Saccharides. *Tetrahedron Lett.* **2003**, *44*, 4057-4061.













Isothermal Titration Calorimetry

Measurements were made with MicroCal VP-ITC. The buffer used in all titrations was 50 mM 3,3dimethylglutarate pH 5.2, 250 mM NaCl, 1 mM CaCl₂, 1 mM MnCl₂. For a given experiment, a solution of ConA ranging in concentration from 200 mM to 300 mM was placed in the cell and titrated with a solution of the saccharide ligand (1-7) over a course of 35 7 μ L injections. A delay time of 5 min between injections was utilized. The cell volume was 1.4167 mL. Heat was generated upon each injection which resulted from protein-ligand association. Heats of dilution, collected from a blank titration, were then subtracted from the titration data before analysis. Titration data was then integrated to give a binding curve. Nonlinear least squares fits of the data, using Origin software from the calorimeter manufacturer, provided K_a (binding constant), Δ H (enthalpy of binding), and N (stoichiometry) values.

Note on the value of *c*:

The binding isotherm of an ITC experiment is characterized by the value of c, which is defined as:

$c = K_{a}[M]n$

where K_a is the association constant of the interaction, [M] is the concentration of the macromolecule, and *n* is the stoichiometry of the interaction. The concentration of the macromolecule (ConA) in the present system is expressed in the terms of the monomer binding sites making the n value in the above expression 1. The *c* value has no units and is a way to describe the shape of the binding isotherm in ITC. This shape is used in determining the inflection point of a titration. Optimal *c* values range from between 1 and 1000; values below this range generate isotherms that are too linear, making the equivalence point hard to determine. Conversely, values above this range of *c* have too few points near inflection and make determination of thermodynamic parameters similarly difficult.



S-14





ITC 12.3 mM *4* / 146 mM ConA at 298 K This plot is representative of ITC data acquired for 4, 5, and 6

ITC 25 mM 6 / 200 μ M ConA at 298 K (same data as Figure 2.)



