# **Supplementary Material**

Asymmetric synthesis of orthogonally protected *trans*-cyclopropane  $\gamma$ -amino acids *via* intramolecular ring closure

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#### **Experimental**

For reactions conducted under anhydrous conditions glassware was dried overnight in an oven at 130 °C and was allowed to cool in a dessicator over anhydrous KOH. Anhydrous reactions were carried out under argon. Solvents were BOC standard reagent grade and distilled before use. Reagents/solvents for anhydrous reactions were dried as follows: THF was distilled from sodium wire with benzophenone as indicator. Ether was distilled from a mixture of CaH<sub>2</sub> and LiAlH<sub>4</sub>. Dichloromethane, hexane, acetonitrile, toluene, pyridine, N,N-dimethylformamide, triethylamine, dimethylsulfoxide and diisopropylamine were dried and stored over 4 Å molecular sieves. Methanol was dried and stored over 3 Å molecular sieves. *n*-Butyllithium was titrated against diphenylacetic acid before use.<sup>1</sup> Sulfate buffer was prepared by dissolving 1.5 mol of Na<sub>2</sub>SO<sub>4</sub> in 0.5 mol H<sub>2</sub>SO<sub>4</sub> and adding water to give a total volume of 2000 cm<sup>3</sup>. Thin layer chromatography (TLC) was carried out on commercially available pre-coated glass plates (Merck  $60F_{254}$ ). The quoted  $R_{\rm f}$  values are rounded to the nearest 0.05. Dry Column Vacuum Chromatography (DCVC) was performed according to the published procedure.<sup>2</sup> A larger diameter column than that recommended was generally necessary with phosphine oxides due to their tendency to streak on the columns. <sup>1</sup>H, <sup>13</sup>C, APT, DEPT, HMOC, COSY, and NOE NMR spectra were recorded on Bruker Avance 400 (5 mm QNP probe), Bruker Avance 500 (5 mm dual <sup>13</sup>C-<sup>1</sup>H cryo probe) and Bruker Avance 700 (5 mm conventional geometry dual <sup>13</sup>C-<sup>1</sup>H probe) Fourier transform spectrometers using an internal deuterium lock. <sup>31</sup>P NMR spectra were recorded on a Bruker Avance 400 (5 mm QNP probe) Fourier transform spectrometer using 85% H<sub>3</sub>PO<sub>4</sub> as external standard. Solvents were used as internal standard when assigning NMR spectra ( $\delta_{\rm H}$ : CDCl<sub>3</sub> 7.26 ppm, DMSO- $d_6$  2.50;  $\delta_{\rm C}$ : CDCl<sub>3</sub> 77.0 ppm, DMSO- $d_6$  39.4 ppm). Mestre-C 4.5.6 software,<sup>3</sup> was used for assigning spectra. J values are given in Hz and rounded to the nearest 0.5 Hz. LC-MS was run on a Waters Alliance LC/MS system consisting of a Waters 2795 Separations Module, a Waters 2996 Photodiode Array Detector and a Waters Micromass ZQ on a C18 analytical Reverse Phase Supercosil<sup>TM</sup> ABZ+PLUS column  $(3.3 \text{ cm} \times 4.6 \text{ mm}, 3 \mu \text{m})$  using the following gradient: 0.00-0.70 min 100% solvent A, 0.70-4.20 min 100% solvent A to 100% solvent B, 4.20-7.70 min 100% solvent B, 7.70-8.00 min 100% solvent B to 100% solvent A (solvent A: 10 mM ammonium acetate in water containing 0.1% formic acid; solvent B: 95% acetonitrile in water) with a flow rate of 1 cm<sup>3</sup>/min. EI and LSIMS mass spectra were recorded on a Kratos concept 1H double focusing magnetic sector instrument using a MACH 3 data system. +ESI mass spectra were recorded using a Bruker Bio-Apex II FT-ICR instrument or a Micromass Q-Tof 1 machine. Microanalyses were carried out on a CE440 Elemental Analyser from Exeter Analytical, INC. The calculated values were adjusted for residual solvents. Melting points were measured on a microscope hot stage melting point apparatus (C. Reichert Optische Werke AG) and are uncorrected. Infra-red spectra were recorded using a Perkin Elmer Spectrum One (FT-IR) spectrometer with a universal ATR sampling accessory. Optical rotations were recorded on a Perkin Elmer 241 polarimeter using to the sodium D line (589 nm) at 23 °C and are given in units of 10<sup>-1</sup> deg dm<sup>2</sup> g<sup>-1</sup>. X-ray Crystallographic Data was measured on a Nonius Kappa CCD diffractometer at 180(2) K. Analytical chiral HPLC was carried out on a Daicel Chiralpak AD column (0.46 cm × 25 cm) and guard column with a Spectra-Physics SP8800 pump, a Spectra-Physics SP8450 UV detection system and a ChromJet single channel integrator with a flow rate of 1  $cm^3/min$ .

# Method 1: Asymmetric dihydroxylation (AD)

By a method analogous to that reported by Sharpless,<sup>4</sup> the substrate (1 mmol) is dissolved in *t*-BuOH (10 cm<sup>3</sup>). Water (10 cm<sup>3</sup>) is added and the mixture cooled to 0 °C. A mixture of K<sub>2</sub>OsO<sub>4</sub>·2 H<sub>2</sub>O (1 mol%), K<sub>3</sub>Fe(CN)<sub>6</sub> (3 eq.), K<sub>2</sub>CO<sub>3</sub> (3 eq.), MeSO<sub>2</sub>NH<sub>2</sub> (1 eq.) and (DHQD)<sub>2</sub>PHAL (2 mol%) is added to the cooled solution and it is stirred vigorously until completion. Sodium sulfite (~10 eq.) is added and the reaction allowed to warm to room temperature with vigorous stirring. The slurry is transferred to a separatory funnel and the phases are separated. The organic phase is concentrated *in vacuo* and the residue dissolved in dichloromethane (20 cm<sup>3</sup>) and transferred to a separatory funnel with the aqueous phase and water (10 cm<sup>3</sup>). Extracted with dichloromethane (2 × 20 cm<sup>3</sup>). The combined organic extracts are washed with aqueous sulfate buffer (20 cm<sup>3</sup>), saturated aqueous NaHCO<sub>3</sub> (20 cm<sup>3</sup>), dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and evaporated under reduced pressure. The residue is purified by column chromatography.

## Method 2: Racemic dihydroxylation

According to the procedure by Warren<sup>5,6</sup> racemic dihydroxylations were performed at room temperature and  $(DHQD)_2PHAL$  was replaced with quinuclidine (5 mol%). Sodium sulfite (~10 eq.) is added and the reaction allowed to warm to room temperature with vigorous stirring. The slurry is transferred to a separatory funnel with water (20 cm<sup>3</sup>) and extracted with ethyl acetate (3 × 20 cm<sup>3</sup>). The combined organic extracts are washed with aqueous sulfate buffer (20 cm<sup>3</sup>), saturated aqueous NaHCO<sub>3</sub> (20 cm<sup>3</sup>), dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and evaporated under reduced pressure and the residue purified by column chromatography.

## Method 3: Diphenylphosphinoylation of alcohols

To a stirred solution of the alcohol (1 mmol) in anhydrous dichloromethane (10 cm<sup>3</sup>) under argon is added Et<sub>3</sub>N (2 eq.), DMAP (0.2 eq.) and diphenylphosphinoyl chloride (1.1 eq.). When the reaction has gone to completion water (10 cm<sup>3</sup>) is added and the mixture transferred to a separatory funnel and extracted with dichloromethane ( $3 \times 20$  cm<sup>3</sup>). The combined organic phases are washed with saturated aqueous sulfate buffer (25 cm<sup>3</sup>), saturated aqueous NaHCO<sub>3</sub> (25 cm<sup>3</sup>), dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and the solvent removed *in vacuo* to give the crude product that is purified by column chromatography.

# Method 4: Synthesis of cyclic sulfites

Thionyl chloride (1.5 eq.) is added to a stirred solution of the diol (1 mmol) and pyridine (4 eq.) in dichloromethane (5 cm<sup>3</sup>) at room temperature under argon. When the reaction has gone to completion saturated aqueous NH<sub>4</sub>Cl (5 cm<sup>3</sup>) is added and the mixture transferred to a separatory funnel with water (5 cm<sup>3</sup>) and extracted with dichloromethane (3 × 10 cm<sup>3</sup>). The combined organic phases are washed with aqueous sulfate buffer (10 cm<sup>3</sup>), dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and the solvent removed *in vacuo* to give the crude product that is purified by column chromatography.

#### Method 5: Mesylation of alcohols

To a stirred solution of the alcohol (1 mmol) in anhydrous dichloromethane (5 cm<sup>3</sup>) under argon is added anhydrous pyridine (10 eq.) and methanesulfonyl chloride (1.1 eq.). When the reaction has gone to completion sulfate buffer (20 cm<sup>3</sup>) is added and the mixture transferred to a separatory funnel with water (10 cm<sup>3</sup>) and extracted with dichloromethane ( $3 \times 20$  cm<sup>3</sup>). The combined organic phases are washed with saturated aqueous NaHCO<sub>3</sub> (20 cm<sup>3</sup>), dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and the

solvent removed *in vacuo* to give the crude product that is re-dissolved in toluene and concentrated *in vacuo* to remove pyridine traces. The product is purified by column chromatography.

# Method 6: Tosylation of alcohols

To a stirred solution of the alcohol (1 mmol) in anhydrous dichloromethane (10 cm<sup>3</sup>) under argon is added triethylamine (2 eq.), DMAP (0.2 eq.) and 4-methylphenylsulfonyl chloride (1.1 eq.). When the reaction has gone to completion sulfate buffer (20 cm<sup>3</sup>) is added and the mixture transferred to a separatory funnel with water (10 cm<sup>3</sup>) and extracted with dichloromethane (3 × 20 cm<sup>3</sup>). The combined organic phases are washed with saturated aqueous NaHCO<sub>3</sub> (20 cm<sup>3</sup>), dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and the solvent removed *in vacuo* to give the crude product that is purified by column chromatography.

#### Method 7 a-c: Cyclopropanation by intramolecular ring closure

**7a (LDA):** The substrate (1 mmol) is dissolved in anhydrous THF (10 cm<sup>3</sup>) and cooled to -78 °C with stirring under argon. Freshly prepared LDA cooled to -78 °C is added by cannula. After stirring at -78 °C for 1-2 hours the reaction mixture is allowed to slowly warm to room temperature overnight. The reaction is quenched with saturated aqueous NH<sub>4</sub>Cl (20 cm<sup>3</sup>), extracted with ethyl acetate (3 × 20 cm<sup>3</sup>), dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated *in vacuo* to give the crude product, which is purified by column chromatography.

**7b (NaHMDS):** Same procedure as method **7a** except that sodium hexamethyldisilazide (2.0 M in THF, 1.05 eq.) is employed instead of LDA, and the temperature is maintained at -78 °C for 3-4 hours after the addition of base.

7c (KHMDS): Same procedure as method 7a except that potassium hexamethyldisilazide (0.5 M in toluene, 1.05 eq.) is employed instead of LDA, and the temperature is maintained at -78 °C for 3-4 hours after the addition of base.

# Method 8: Opening of cyclic sulfites with sodium azide

The cyclic sulfite (1 mmol) is dissolved in anhydrous DMF (5 cm<sup>3</sup>). NaN<sub>3</sub> (2 eq.) is added and the reaction mixture heated to 60 °C with stirring under argon for 48 hours. When the reaction mixture has cooled to room temperature it is transferred to a separatory funnel with water (20 cm<sup>3</sup>) and extracted with ethyl acetate (3 × 20 cm<sup>3</sup>). The combined organic phases are washed with aqueous

sulfate buffer (20 cm<sup>3</sup>), dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated *in vacuo*. Residual DMF is removed on a high vacuum pump and the residue purified by column chromatography.

#### Method 9: Synthesis of Mosher's amides

Mosher's amide derivatives using racemic Mosher's acid ( $\alpha$ -methoxy- $\alpha$ -trifluoromethylphenylacetic acid) and (R)-(+)-Mosher's acid were prepared and analysed using <sup>1</sup>H and <sup>19</sup>F NMR. Moshers acid (2.14 mmol) is dissolved in anhydrous dichloromethane (10 cm<sup>3</sup>) and cooled to 0 °C. Oxalyl chloride (21.4 mmol) is added followed by 1 drop of DMF. After stirring for 1 hour the reaction mixture is concentrated *in vacuo* and the residue suspended in hexane (2 × 25 cm<sup>3</sup>) and concentrated *in vacuo* [<sup>19</sup>F NMR (400 MHz; CDCl<sub>3</sub>)  $\delta$  –70.2]. The product was dissolved in anhydrous dichloromethane (10 cm<sup>3</sup>) to give a 0.21 M solution of Mosher's acid chloride. The amine (0.2 mmol) is dissolved in dichloromethane (5 cm<sup>3</sup>) and Mosher's acid chloride (0.3

mmol, 1.4 cm<sup>3</sup>, 0.21 M in dichloromethane) is added followed by saturated aqueous sodium carbonate (5 cm<sup>3</sup>). After stirring overnight the phases are separated and the organic phase dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated *in vacuo* to give crude Mosher's amide that is analysed by NMR without further purification.

#### (E)-tert-Butyl 5-phenyl-pent-4-enoate 8

*tert*-Butyl acetate (1.0 g, 8.6 mmol) was dissolved in anhydrous THF (40 cm<sup>3</sup>) and cooled to -78 °C. Freshly prepared LDA (9.0 mmol) was added by cannula to give a red solution. After ½ hour HMPA (1.5 cm<sup>3</sup>, 8.6 mmol) was added. After an additional ½ hour (*E*)-cinnamyl bromide (1.7 g, 8.6 mmol) dissolved in anhydrous THF (10 cm<sup>3</sup>) and cooled to -78 °C was added by cannula. After 4 hours the reaction was quenched by the addition of saturated aqueous NH<sub>4</sub>Cl (20 cm<sup>3</sup>) and allowed to warm to room temperature. The reaction mixture was transferred to a separatory funnel with water (10 cm<sup>3</sup>) and extracted with ethyl acetate (3 × 50 cm<sup>3</sup>). The combined organic phases were washed with water (3 × 50 cm<sup>3</sup>), dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated *in vacuo* to give a yellow liquid. The product was purified by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions; 6 × hexanes; 2.5-20% EtOAc in hexanes (v/v) – 2.5% increments; two fractions of each solvent mixture were collected] to give *tert*-butyl ester **8** (1.92 g, 96%) as a clear colourless liquid. *R*<sub>f</sub> 0.30 (5% EtOAc in hexanes, v/v); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 1722 (C=O) and 1149 (C-O); *m/z* (+ESI) found: MNa<sup>+</sup>, 255.1367. (C<sub>15</sub>H<sub>20</sub>O<sub>2</sub>Na requires *M*, 355.1356); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>)  $\delta$  7.36-7.34 (2H, m, *ortho*-Ph), 7.32-7.29 (2H, m, *meta*-Ph), 7.21 (1H, tt, *J* 7.0 and 1.5. *para*-Ph), 6.44 (1H, d, *J* 16.0, CH=CHPh),

6.22 (1H, dt, *J* 16.0 and 7.0, C*H*=CHPh), 2.53-2.49 (2H, m, C*H*<sub>2</sub>CH=CH), 2.42-2.39 (2H, m, CH<sub>2</sub>C=O) and 1.47 [9H, s, C(CH<sub>3</sub>)]; <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$  172.3 (C1), 137.4 (*ipso*-Ph), 130.7 (C5), 128.7 (C4), 128.4 (*meta*-Ph), 127.0 (*para*-Ph), 126.0 (*ortho*-Ph), 80.2 [*C*(CH<sub>3</sub>)<sub>3</sub>], 35.2 (C2), 28.5 (C3) and 28.1 [C(CH<sub>3</sub>)<sub>3</sub>]; (Found: C, 77.81; H, 8.73. C<sub>15</sub>H<sub>20</sub>O<sub>2</sub> requires C, 77.55; H, 8.68%). Compound **8** has been reported before with no characterisation.<sup>7</sup>

# (4R,5R)-tert-Butyl 4,5-dihydroxy-5-phenyl-pentanoate 9

By method 1 *tert*-butyl ester **8** (3.0 g, 12.9 mmol) after 4 days at 3 °C gave a viscous yellow liquid that was purified by DCVC [id 6 cm; 50 cm<sup>3</sup> fractions; 3 × hexanes; 10-100% EtOAc in hexanes (v/v) – 10% increments; 3 × EtOAc] to give *diol* **9** (2.95 g, 86%) as white needles. e.e. >95% (determined by chiral HPLC); HPLC [ $R_T$  (min), flow rate 1 cm<sup>3</sup>/min, 4% EtOH in *iso*-hexane (v/v)]: 18.7; [ $\alpha$ ]<sub>D</sub><sup>23</sup> –22 (c. 1.7, CHCl<sub>3</sub>); mp 55-56 °C (from EtOAc, hexanes);  $R_f$  0.70 (50% EtOAc in hexanes, v/v); *m/z* (+ESI) found: MNa<sup>+</sup>, 289.1402. (C<sub>15</sub>H<sub>22</sub>O<sub>4</sub>Na requires *M*, 289.1416); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 3386 (br., O-H), 1724 (C=O) and 1148 (C-O); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>)  $\delta$  7.38-7.28 (5H, m, Ph), 4.43 (1H, d, *J* 7.0, PhC*H*OH), 3.68 (1H, ddd, *J* 9.0, 7.0 and 3.5, CH<sub>2</sub>C*H*OH), 3.12 (1H, br s, OH), 3.00 (1H, br s, OH), 2.38 (1H, dt, *J* 16.5 and 7.0, CH<sub>a</sub>H<sub>b</sub>C=O), 1.70-1.58 (2H, m, CH<sub>2</sub>CHOH) and 1.41 [9H, s, C(CH<sub>3</sub>)]; <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$  173.7 (C1), 140.8 (*ipso*-Ph), 128.5 (Ph), 128.1 (*para*-Ph), 126.9 (Ph), 80.7 [*C*(CH<sub>3</sub>)<sub>3</sub>], 77.9 (C5), 75.5 (C4), 32.0 (C2), 28.0 [C(CH<sub>3</sub>)<sub>3</sub>] and 27.7 (C3); (Found: C, 67.66; H, 8.44. C<sub>15</sub>H<sub>22</sub>O<sub>4</sub> requires C, 67.64; H, 8.33%).

# (4RS,5RS)-tert-Butyl 4,5-dihydroxy-5-phenyl-pentanoate (±)-9

By method **2** *tert*-butyl ester **8** (0.20 g, 0.90 mmol) after 24 hours gave *diol* (±)-**9** (0.23 g, 95%) as a clear gum that required no further purification. HPLC [ $R_T$  (min), flow rate 1 cm<sup>3</sup>/min, 4% EtOH in *iso*-hexane (v/v)]: 14.7 and 19.3; All analytical data were identical with that for *tert*-butyl (4R,5R)-**9** reported above.

## (4R,5R)-tert-Butyl 4,5-diphenylphosphinoyloxy-5-phenyl-pentanoate 10

By method **3** diol **9** (0.47 g, 1.76 mmol) after 1 day gave a yellow foam. Purification by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions: 2 × hexanes; 10-100% EtOAc in hexanes (v/v) – 10% increments; 11 × EtOAc] gave *bis-phosphinate* **10** (0.62 g, 52%) as a clear gum.  $[\alpha]_D^{23}$  +18 (c. 1, CHCl<sub>3</sub>);  $R_f$  0.25

(80% EtOAc in hexanes, v/v); m/z (+ESI) found: MH<sup>+</sup>, 667.2398. (C<sub>39</sub>H<sub>41</sub>O<sub>6</sub>P<sub>2</sub> requires *M*, 667.2378); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 1724 (C=O), 1439 (P-Ph) and 1226 (P=O); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>)  $\delta$  7.83-7.75 (4H, m, Ph), 7.65-7.61 (2H, m, Ph), 7.53-7.42 (7H, m, Ph), 7.38-7.32 (4H, m, Ph), 7.24-7.14 (8H, m, Ph), 5.50 (1H, dd, *J* 10.0 and 6.0, PhC*H*), 4.84-4.79 (1H, m, PhCHC*H*), 2.25-2.15 (2H, m, CH<sub>2</sub>C=O), 2.06-1.99 (1H, m, C*H*<sub>a</sub>H<sub>b</sub>CH<sub>2</sub>C=O), 1.69-1.62 (1H, m, CH<sub>a</sub>H<sub>b</sub>CH<sub>2</sub>C=O) and 1.32 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>]; <sup>31</sup>P NMR (162 MHz; CDCl<sub>3</sub>)  $\delta$  32.2 (×2); <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$  172.0 (C1), 136.3 (*ipso*-PhC), 132.1 (d, *J* 2.5), 132.0 (d, *J* 3.0) (2 × *para*-PhP), 131.9-131.5 (m, Ph), 131.8 (d, *J* 138.0), 131.4 (d, *J* 140.5) (2 × *ipso*-Ph), 130.5 (×2), 128.5-128.0 (m) (Ph), 127.6 (*para*-PhC), 80.2 [*C*(CH<sub>3</sub>)<sub>3</sub>], 77.4 (t, *J* 5.5, C5), 76.9 (t, *J* 6.0, C4), 30.6 (C2), 28.0 [C(CH<sub>3</sub>)<sub>3</sub>] and 26.2 (d, *J* 2.5, C3).

#### (4RS,5RS)-tert-Butyl 4,5-diphenylphosphinoyloxy-5-phenyl-pentanoate (±)-10

By method **3** diol (±)-9 (0.20 g, 0.75 mmol) after 14 hours gave a yellow gum. Purification by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions; 2 × hexanes; 10-100% EtOAc in hexanes (v/v) – 10% increments; 10 × EtOAc] gave *bis-phosphinate* (±)-10 (0.27 g, 54%) as a clear gum. All analytical data were identical with that for (4R,5R)-10 reported above.

# (1'*R*,2'*R*,1''*S*)-*tert*-Butyl 2'-(1''-diphenylphosphinoyloxy-1''-phenyl-methyl)-cyclopropane carboxylate 11

By method **7b** *bis*-phosphinate **10** (0.27 g, 0.41 mmol) gave a yellow gum. The product was purified by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions; 2 × hexanes; 10-100% EtOAc in hexanes (v/v) – 10% increments; 5 × EtOAc] to give *cyclopropane* **11** (75 mg, 41%) as a white amorphous solid. e.e. >95% (determined by chiral HPLC); HPLC [ $R_T$  (min), flow rate 1 cm<sup>3</sup>/min, 5% *iso*-propanol in *iso*-hexane (v/v)]: 57.1; [ $\alpha$ ]<sub>D</sub><sup>23</sup> –29 (c. 0.7, CHCl<sub>3</sub>);  $R_f$  0.50 (60% EtOAc in hexanes, v/v); *m/z* (+ESI) found: MNa<sup>+</sup>, 471.1684. (C<sub>27</sub>H<sub>29</sub>O<sub>4</sub>PNa requires *M*, 471.1701); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 1717 (C=O), 1439 (PPh), 1220 (P=O) and 1151 (C-O); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>)  $\delta$ 7.85-7.81 (2H, m, *ortho*-PPh), 7.59-7.54 (2H, m, *ortho*-PPh), 7.53-7.49 (1H, m, *para*-PhP), 7.46-7.42 (2H, m, *meta*-PhP), 7.40-7.37 (1H, m, *para*-PhP), 7.29-7.24 (7H, m, *meta*-PhP and PhC), 5.09 (1H, dd, *J* 9.5 and 7.5, PhCH), 1.89 (1H, dddd, *J* 8.5, 7.5, 6.5 and 4.5, PhCHC*H*), 1.62 (1H, ddd, *J* 8.5, 5.0 and 4.5, C*H*C=O), 1.35 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>] and 1.08-1.02 (2H, m, CH<sub>2</sub>); <sup>31</sup>P NMR (162 MHz; CDCl<sub>3</sub>)  $\delta$  32.3; <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$ 172.3 (C1), 139.4 (d, *J* 4.0, *ipso*-PhC), 132.4 (d, *J* 93.0, *ipso*-PhP),

132.1 (d, *J* 2.5, *para*-PhP), 131.9 (d, *J* 3.0, *para*-PhP), 131.6 (d, *J* 10.5, *ortho*-PhP), 131.5 (d, *J* 10.5, *ortho*-PhP), 128.4 (d, *J* 13.0, *meta*-PhP), 128.3 (PhC), 128.2 (PhC), 128.1 (d, *J* 13.5, *meta*-PhP), 126.7 (PhC), 80.4 [*C*(CH<sub>3</sub>)<sub>3</sub>], 78.4 (d, *J* 6.0, C1<sup>''</sup>), 28.0 [*C*(CH<sub>3</sub>)<sub>3</sub>], 27.2 (C2<sup>'</sup>), 19.6 (C1<sup>'</sup>) and 13.6 (C3<sup>'</sup>).

# (1'*RS*,2'*RS*,1''*SR*)-*tert*-Butyl 2'-(1''-diphenylphosphinoyloxy-1''-phenyl-methyl)-cyclopropane carboxylate (±)-11

By method **7b** *bis*-phosphinate (±)-10 (0.27 g, 0.44 mmol) gave a yellow residue. The product was purified by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions; 2 × hexanes; 10-100% EtOAc in hexanes (v/v) – 10% increments; 5 × EtOAc] to give cyclopropane (±)-11 (25 mg, 14%) as a white amorphous solid. HPLC [ $R_T$  (min), flow rate 1 cm<sup>3</sup>/min, 5% *iso*-propanol in *iso*-hexane (v/v)]: 39.5 and 58.5; All analytical data were identical with that for (1'R,2'R,1''S)-11 reported above.

# (1'RS,4'R,5'R)-tert-Butyl 3-(1'-Oxo-3'-phenyl-[2',5',1']dioxathiolan-4'-yl)-propanoate 15

By method **4** diol **9** (0.60 g, 2.25 mmol), after 4 hours, gave a yellow gum. The product was purified by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions;  $3 \times$  hexanes; 5-50% EtOAc in hexanes (v/v) – 5% increments; 1-5% MeOH in EtOAc (v/v) – 1% increments] to give *cyclic sulfite* **15** (0.60 g, 85%) as a viscous yellow liquid. d.r. = 56:44 (<sup>1</sup>H NMR, epimers at S);  $R_f$  0.25 (10% EtOAc in hexanes, v/v); m/z (+ESI) found: MNa<sup>+</sup>, 335.0921. (C<sub>15</sub>H<sub>20</sub>O<sub>5</sub>SNa requires *M*, 335.0929); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 1724 (C=O), 1207 (S=O) and 1150 (C-O); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>) Two diastereoisomers A: major isomer, B: minor isomer.  $\delta$  7.48-7.37 (m, 5H, Ph A,B), 5.51 (d, 1H, *J* 9.5, *CH*Ph A), 4.95 (d, 1H, *J* 9.5, *CH*Ph B), 4.75 (dt, 1H, *J* 9.5 and 3.0, CH<sub>2</sub>*CH* B), 4.38 (dt, 1H, *J* 9.0 and 6.0, CH<sub>2</sub>*CH* A), 2.55-2.31 (m, 4H, 2 × CH<sub>2</sub>C=O A,B), 2.16-1.97 (m, 4H, 2 × CH<sub>2</sub>CH A,B) and 1.39 [s, 18H, 2 × C(CH<sub>3</sub>)<sub>3</sub>]; <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$  171.4, 171.3 (2 × C1 A,B), 133.8, 133.1 (2 × *ipso*-Ph A,B), 129.8, 129.4, 129.1 (×2), 127.7, 127.2 (6 × Ph A,B), 89.6 (C5 B), 88.3 (C4 A), 84.3 (C5 A), 83.9 (C4 B), 80.9, 83.9 [2 × *C*(CH<sub>3</sub>)<sub>3</sub> A,B], 31.6, 31.5 (2 × C2 A,B), 28.0 [2 × C(CH<sub>3</sub>)<sub>3</sub> A,B], 27.5 and 25.3 (2 × C3 A,B); (Found: C, 57.96; H, 6.43. C<sub>15</sub>H<sub>20</sub>O<sub>5</sub>S requires C, 57.67; H, 6.45%).

# (4R,5R)-tert-Butyl 4,5-methanesulfonyloxy-5-phenyl-pentanoate 16

By method **5** diol **9** (0.52 g, 1.95 mmol) after 20 hours gave a yellow liquid. Purification by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions:  $2 \times$  hexanes; 0-50% EtOAc in hexanes (v/v) – 5% increments; 50-100%

EtOAc in hexanes (v/v) – 10% increments; 5 × EtOAc] gave *mesylate* **16** (0.61 g, 73%) as a white amorphous solid.  $[\alpha]_D^{23}$  –35 (c. 0.2, CHCl<sub>3</sub>); mp 52-54 °C (EtOAc, hexanes);  $R_f$  0.55 (50% EtOAc in hexanes, v/v); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 1723 (C=O), 1349 (SO<sub>2</sub>) and 1168 (SO<sub>2</sub>); <sup>1</sup>H NMR (400 MHz; CDCl<sub>3</sub>)  $\delta$  7.50-7.41 (5H, m, Ph), 5.52 (1H, d, *J* 8.0, PhC*H*), 5.11 (1H, dt, *J* 8.0 and 6.5, PhCHC*H*), 3.09 (3H, s, CH<sub>3</sub>S), 2.62 (3H, s, CH<sub>3</sub>S), 2.40 (2H, t, *J* 7.5, CH<sub>2</sub>C=O), 1.73-1.67 (2H, m, CH<sub>2</sub>CH<sub>2</sub>C=O) and 1.40 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>]; <sup>13</sup>C NMR (100 MHz; CDCl<sub>3</sub>)  $\delta$  171.4 (C1), 133.7 (*ipso*-Ph), 130.4 (*para*-Ph), 129.5, 127.8 (*ortho-* and *meta*-Ph), 84.3 (C5), 81.6 (C4), 80.9 [*C*(CH<sub>3</sub>)<sub>3</sub>], 39.5, 39.1 (2 × CH<sub>3</sub>S), 30.4 (C2), 28.0 [C(CH<sub>3</sub>)<sub>3</sub>] and 26.4 (C3); (Found: C, 48.79; H, 6.10. C<sub>17</sub>H<sub>26</sub>O<sub>8</sub>S<sub>2</sub> requires C, 48.33; H, 6.20%).

# (4R,5R)-tert-Butyl 4,5-bis-(4'-methyl-phenylsulfonyloxy)-5-phenyl-pentanoate 17

Diol 9 (0.25 g, 0.94 mmol) was dissolved in anhydrous pyridine (5 cm<sup>3</sup>) and *para*-phenylsulfonyl chloride (1.14 g, 6.0 mmol) was added. After 20 hours the reaction was quenched with saturated aqueous NH<sub>4</sub>Cl (15 cm<sup>3</sup>) and extracted with ethyl acetate (3  $\times$  25 cm<sup>3</sup>). The combined organic phases were dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated *in vacuo* to give a clear gum that was purified by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions;  $2 \times$  hexanes; 5-90% EtOAc in hexanes (v/v) – 5% increments] to give *bis-tosylate* 17 (0.39 g, 72%) as a white amorphous solid.  $[\alpha]_{D}^{23}$  +6.38 (c. 1.27, CHCl<sub>3</sub>);  $R_f 0.30$  (30% EtOAc in hexanes, v/v); m/z (+ESI) found: MNa<sup>+</sup>, 597.1609. (C<sub>29</sub>H<sub>34</sub>O<sub>8</sub>S<sub>2</sub>Na requires M, 597.1593); IR  $v_{max}$  (CHCl<sub>3</sub>)/cm<sup>-1</sup> 1711 (C=O), 1364 (SO<sub>2</sub>) and 1171 (SO<sub>2</sub>); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>) & 7.73-7.71 (2H, m, ortho-Ts), 7.54-7.52 (2H, m, ortho-Ts), 7.31-7.30 (2H, m, meta-Ts), 7.21 (1H, tt, J 7.5 and 1.5, para-Ph), 7.16-7.12 (4H, m, meta-Ts and meta-Ph), 7.08-7.06 (2H, m, ortho-Ph), 5.49 (1H, d, J 5.5, CHPh), 4.88 (1H, ddd, J 9.1, 9.0 and 5.5, CHCHPh), 2.45 (3H, s, CH<sub>3</sub>Ar), 2.35 (3H, s, CH<sub>3</sub>Ar), 2.23-2.10 (2H, m, CH<sub>2</sub>C=O), 1.93-1.86 (1H, m, CH<sub>a</sub>H<sub>b</sub>CH), 1.61-1.52 (1H, m, CH<sub>a</sub>H<sub>b</sub>CH) and 1.39 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>]; <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$  171.3 (C1), 144.9, 144.6 (2 × para-Ts), 133.3 (×2), 133.1 (2 × ipso-Ts and ipso-Ph), 129.8, 129.4 (2 × meta-Ts), 128.8 (para-Ph), 128.3 (meta-Ph), 128.0, 127.9 (2 × ortho-Ts), 127.3 (ortho-Ph), 81.7 (C5), 80.9 (C4), 80.6 [ $C(CH_3)_3$ ], 30.3 (C2), 28.0 [ $C(CH_3)_3$ ], 25.3 (C3), 21.7 and 21.6 ( $2 \times CH_3Ar$ ); (Found: C, 57.30; H, 5.56. C<sub>29</sub>H<sub>34</sub>O<sub>8</sub>S<sub>2</sub>·0.15 EtOAc requires C, 57.18; H, 5.69%).

# (Z)-tert-Butyl 5-methanesulfonyloxy-5-phenyl-pent-4-enoate 18

By method **7b** *bis*-mesylate **16** (126 mg, 0.30 mmol) gave a yellow gum. The product was purified by DCVC [id 1 cm; 9 cm<sup>3</sup> fractions; 4 × hexanes; 5-70% EtOAc in hexanes (v/v) – 5% increments; two fractions of each solvent mixture were collected] to give *olefin* **18** (30 mg, 30%) as a clear liquid.  $R_f$  0.35 (30% EtOAc in hexanes, v/v); *m/z* (+ESI) found: MNa<sup>+</sup>, 349.1080. (C<sub>16</sub>H<sub>22</sub>O<sub>5</sub>SNa requires *M*, 349.1086); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 1721 (C=O), 1364 (SO<sub>2</sub>) and 1174 (SO<sub>2</sub>); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>)  $\delta$  7.49-7.47 (2H, m, *ortho*-Ph), 7.41-7.31 (3H, m, Ph), 5.84 (1H, t, *J* 7.5, PhC=C*H*), 2.98 (3H, s, CH<sub>3</sub>S), 2.65 (2H, q, CHC*H*<sub>2</sub>), 2.42 (2H, t, *J* 7.5, C*H*<sub>2</sub>C=O) and 1.45 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>]; <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$  171.9 (C1), 146.4 (C5), 134.5 (*ipso*-Ph), 129.0 (*para*-Ph), 128.7 (*meta*-Ph), 125.7 (*ortho*-Ph), 121.3 (C4), 80.5 [*C*(CH<sub>3</sub>)<sub>3</sub>], 39.4 (CH<sub>3</sub>S), 34.5 (C2), 28.1 [C(CH<sub>3</sub>)<sub>3</sub>] and 22.6 (C3).



No NOE was observed between the C4 proton and the mesyl group and a strong NOE between the C4 proton and the *ortho*-protons on the aromatic ring was observed indicating that the compound has the (Z)-geometry as shown.

# (Z)-tert-Butyl 5-(4-methyl-phenylsulfonyloxy)-5-phenyl-pent-4-enoate 19

According to method **7b** *bis*-tosylate **17** (0.12 g, 0.21 mmol) gave a grey gum. The product was purified by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions; 2 × hexanes; 10-100% EtOAc in hexanes (v/v) – 10% increments; 4 × EtOAc] to give starting material **17** (47 mg, 39%) and olefin **19** (8 mg, 10%) as a clear film.  $R_f$  0.30 (20% EtOAc in hexanes, v/v); m/z (+ESI) found: MNa<sup>+</sup>, 425.1386. (C<sub>22</sub>H<sub>26</sub>O<sub>5</sub>SNa requires *M*, 425.1399); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 1726 (C=O), 1368 (SO<sub>2</sub>), 1177 (SO<sub>2</sub>) and 1152 (C-O); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>)  $\delta$  7.67 (2H, dt, *J* 8.5 and 2.0, *ortho*-Ts), 7.27-7.25 (3H, m, Ar), 7.21-7.17 (4H, m, Ar), 2.43 (2H, q, *J* 7.0, CHC*H*<sub>2</sub>), 2.39 (3H, s, *CH*<sub>3</sub>Ar), 2.29 (2H, t, *J* 7.0, *CH*<sub>2</sub>C=O) and 1.44 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>]; <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$  171.9 (C1), 146.9, 144.9, 134.6, 133.6 (C5, *ipso*-Ph, *ipso*-Ts and *para*-Ts), 129.5, 128.3 (×2), 128.1, 125.7 (Ph and Ts), 120.8 (C4), 80.4 [*C*(CH<sub>3</sub>)<sub>3</sub>], 34.5 (C2), 28.1 [C(*C*H<sub>3</sub>)<sub>3</sub>], 22.3 (C3) and 21.6 (*C*H<sub>3</sub>Ar).

# (4R,5S)-tert-Butyl 5-azido-4-hydroxy-5-phenyl-pentanoate 26

By method 4 diol 9 (0.74 g, 2.80 mmol) gave a dark brown gum. According to method 8 the crude cyclic sulfite produced a brown gum that was purified by DCVC [id 4 cm, 20 cm<sup>3</sup> fractions; 2 × hexanes; 0-100% EtOAc in hexanes (v/v) – 10% increments; 5 × EtOAc] to give *azide* 26 (0.53 g,

65%) as a yellow gum.  $[α]_D^{23}$  +113 (c. 1, CHCl<sub>3</sub>);  $R_f$  0.50 (30% EtOAc in hexanes, v/v); *m/z* (+ESI) found: MNa<sup>+</sup>, 314.1475. (C<sub>15</sub>H<sub>21</sub>N<sub>3</sub>O<sub>3</sub>Na requires *M*, 314.1480); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 3443 (br., O-H), 2101 (N<sub>3</sub>), 1724 (C=O) and 1148 (C-O); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>) δ 7.42-7.38 (2H, m, Ph), 7.36-7.33 (3H, m, Ph), 4.52 (1H, d, *J* 6.0, *CH*Ph), 3.84-3.78 (1H, m, *CH*CHPh), 2.54 (1H, br d, *J* 4.0, OH), 2.39 (2H, t, *J* 7.0, CH<sub>2</sub>C=O), 1.85 (1H, dtd, *J* 10.0, 7.0 and 2.5, *CH*<sub>a</sub>H<sub>b</sub>CH<sub>2</sub>C=O), 1.64 (1H, tdd, *J* 14.5, 10.0 and 7.0, CH<sub>a</sub>H<sub>b</sub>CH<sub>2</sub>C=O) and 1.43 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>]; <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>) δ 173.5 (C1), 136.2 (*ipso*-Ph), 128.8 (Ph), 128.5 (*para*-Ph), 127.8 (Ph), 80.7 [*C*(CH<sub>3</sub>)<sub>3</sub>], 73.9 (C4), 70.4 (C5), 31.9 (C2), 28.0 [C(*C*H<sub>3</sub>)<sub>3</sub>] and 27.3 (C3); (Found: C, 61.78; H, 7.25; N, 14.20. C<sub>15</sub>H<sub>21</sub>N<sub>3</sub>O<sub>3</sub> requires C, 61.84; H, 7.27; N, 14.42%).

#### (4R,5S)-tert-Butyl 5-azido-4-diphenylphosphinoyloxy-5-phenyl-pentanoate 27

By method **3** alcohol **26** (0.38 g, 1.3 mmol) after 42 hours gave a yellow gum. Purification by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions; 2 × hexanes; 0-100% EtOAc in hexanes (v/v) – 10% increments; 4 × EtOAc] gave *phosphinate* **27** (0.43 g, 67%) as a yellow gum.  $[\alpha]_D^{23}$  +11.8 (c. 1.0, CHCl<sub>3</sub>);  $R_f$  0.55 (50% EtOAc in hexanes, v/v); *m/z* (+ESI) found: MNa<sup>+</sup>, 514.1878. (C<sub>27</sub>H<sub>30</sub>N<sub>3</sub>O<sub>4</sub>PNa requires *M*, 514.1872); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 2106 (N<sub>3</sub>), 1727 (C=O) 1440 (P-Ph), 1231 (P=O) and 1153 (C-O); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>)  $\delta$ 7.84-7.74 (4H, m, *ortho*-PhP), 7.55-7.50 (2H, m, *para*-PhP), 7.47-7.42 (4H, m, *meta*-PhP), 7.31-7.24 (3H, m, *meta*- and *para*-PhC), 7.19-7.17 (2H, m, *ortho*-PhC), 4.89 (1H, d, *J* 4.0, PhCH), 4.62 (1H, tdd, *J* 9.0, 3.5 and 3.3, PhCHC*H*), 2.91 (1H, ddd, *J* 17.5, 9.5 and 5.0, *CH*<sub>a</sub>H<sub>b</sub>C=O), 2.78 (1H, ddd, *J* 17.5, 9.0 and 6.5, *CH*<sub>a</sub>H<sub>b</sub>C=O), 2.08-2.0 (1H, m, *CH*<sub>a</sub>H<sub>b</sub>CH<sub>2</sub>C=O), 1.80-1.73 (1H, m, CH<sub>a</sub>H<sub>b</sub>CH<sub>2</sub>C=O) and 1.32 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>]; <sup>31</sup>P NMR (162 MHz; CDCl<sub>3</sub>)  $\delta$  172.0 (C1), 135.6 (*ipso*-PhC), 132.3 (×2) (2 × *para*-PhP), 131.6 (d, *J* 10.5, 2 × *ortho*-PhP), 128.3 (*meta*-PhC), 127.1 (*para*-PhC), 80.3 [*C*(CH<sub>3</sub>)<sub>3</sub>], 78.4 (d, *J* 6.5, C4), 68.6 (d, *J* 3.0, C5), 31.1 (C2), 27.9 [C(CH<sub>3</sub>)<sub>3</sub>] and 24.6 (d, *J* 4.0, C3); (Found: C, 65.69; H, 6.45. C<sub>27</sub>H<sub>30</sub>N<sub>3</sub>O<sub>4</sub>P requires C, 65.98; H, 6.15%).

## (4R,5S)-tert-Butyl 5-azido-4-methanesulfonyloxy-5-phenyl-pentanoate 28

According to method **5** alcohol **26** (0.40 g, 1.37 mmol) after 26 hours gave a yellow oil. Purification by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions; 3 × hexanes; 5-100% EtOAc in hexanes (v/v) – 5% increments] gave *mesylate* **28** (0.48 g, 94%) as yellow gum.  $[\alpha]_D^{23}$  +105 (c. 1, CHCl<sub>3</sub>);  $R_f$  0.45 (30% EtOAc in hexanes, v/v); *m/z* (+ESI) found: MNa<sup>+</sup>, 392.1251. (C<sub>16</sub>H<sub>23</sub>N<sub>3</sub>O<sub>5</sub>SNa requires *M*, 392.1256); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 2108 (N<sub>3</sub>), 1726 (C=O) 1366 (SO<sub>2</sub>) and 1174 (SO<sub>2</sub>); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>)  $\delta$ 7.43-7.35 (5H, m, Ph), 4.92 (1H, d, *J* 5.0, PhC*H*), 4.89 (1H, ddd, *J* 8.5, 5.0 and 4.0, C*H*CHPh), 2.82 (3H, s, CH<sub>3</sub>S), 2.39 (1H, ddd, *J* 17.0, 7.5 and 6.0, C*H*<sub>a</sub>H<sub>b</sub>C=O), 2.32 (1H, dt, *J* 17.0 and 8.0, CH<sub>a</sub>H<sub>b</sub>C=O), 1.98-1.87 (2H, m, CH<sub>2</sub>CH<sub>2</sub>C=O) and 1.40 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>]; <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$ 171.6 (C1), 135.0 (*ipso*-Ph), 129.0 (*para*-Ph), 129.0, 127.7 (*ortho-* and *meta*-Ph), 83.1 (C4), 80.8 [*C*(CH<sub>3</sub>)<sub>3</sub>], 68.0 (C5), 38.3 (CH<sub>3</sub>S), 30.8 (C2), 28.0 [C(CH<sub>3</sub>)<sub>3</sub>] and 25.3 (C3); (Found: C, 52.27; H, 6.42; N, 11.62. C<sub>16</sub>H<sub>23</sub>N<sub>3</sub>O<sub>5</sub>S requires C, 52.02; H, 6.28; N, 11.37%).

## (4R,5S)-tert-Butyl 5-azido-4-(4'-methyl-phenylsulfonyloxy)-5-phenyl-pentanoate 29

According to method **6** alcohol **26** (0.22 g, 0.76 mmol) after 48 hours gave a brown gum. Purification by DCVC [id 4 cm; 25 cm<sup>3</sup> fractions; 2 × hexanes; 5-40% EtOAc in hexanes (v/v) – 5% increments; two fractions of each solvent mixture were collected) gave *tosylate* **29** (0.25 g, 73%) as a clear gum and starting material **26** (45 mg, 6%).  $[\alpha]_D^{23}$  +110 (c. 1, CHCl<sub>3</sub>);  $R_f$  0.60 (30% EtOAc in hexanes, v/v); *m/z* (+ESI) found: MH<sup>+</sup>, 446.1762. (C<sub>22</sub>H<sub>28</sub>N<sub>3</sub>O<sub>5</sub>S requires *M*, 446.1750); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 2107 (N<sub>3</sub>), 1727 (C=O) 1367 (SO<sub>2</sub>) and 1176 (SO<sub>2</sub>); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>)  $\delta$  7.79-7.78 (2H, *ortho*-Ts), 7.36-7.29 (5H, m, Ph and *meta*-Ts), 7.25-7.22 (2H, m, Ph), 4.87 (1H, d, *J* 4.0, PhC*H*), 4.84 (1H, ddd, *J* 9.5, 3.5 and 3.0, C*H*CHPh), 2.27 (1H, ddd, *J* 16.5, 8.5 and 5.5, C*H*<sub>a</sub>H<sub>b</sub>CH<sub>2</sub>C=O) and 1.75 (1H, dddd, *J* 15.0, 8.5, 7.5 and 3.0, CH<sub>a</sub>H<sub>b</sub>CH<sub>2</sub>C=O); <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$  171.6 (C1), 145.1 (*para*-Ts), 134.9, 133.7 (*ipso*-Ph and *ipso*-Ts), 129.9, 128.9 (Ph and *meta*-Ts), 128.6 (*para*-Ph), 127.8 (*ortho*-Ts), 127.2 (Ph), 83.6 (C4), 80.6 [*C*(CH<sub>3</sub>)<sub>3</sub>], 67.7 (C5), 30.7 (C2), 28.0 [C(CH<sub>3</sub>)<sub>3</sub>], 23.8 (C3) and 21.7 (PhCH<sub>3</sub>).

# (1R,2R,1'R)-tert-Butyl 2-(1'-azido-1'-phenyl-methyl)-cyclopropane carboxylate 30

By a modified method  $7a^{\$}$  mesylate **28** (1.00 g, 2.73 mmol) gave a yellow gum. The product was purified by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions; 2 × hexanes; 0-50% EtOAc in hexanes (v/v) – 10% increments – two fractions of each solvent mixture were collected] to give starting material **28** (225 mg, 22%) and cyclopropane **30** (103 mg, 13%) as a clear oil. <sup>§</sup> The temperature was maintained at –78 °C for 24 hours and then raised to room temperature for an additional 24 hours.  $[\alpha]_D^{23}$  +14 (c. 1, CHCl<sub>3</sub>); *R*<sub>f</sub> 0.25 (5% EtOAc in hexanes, v/v); *m/z* (+ESI) found: MNa<sup>+</sup>, 296.1370. (C<sub>15</sub>H<sub>19</sub>N<sub>3</sub>O<sub>2</sub>Na

requires *M*, 296.1375); IR  $v_{max}$ (CHCl<sub>3</sub>)/cm<sup>-1</sup> 2094 (N<sub>3</sub>), 1717 (C=O) and 1150 (C-O); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>)  $\delta$ 7.41-7.33 (5H, m, Ph), 4.05 (1H, d, *J* 8.0, PhC*H*), 1.83 (1H, dddd, *J* 9.0, 8.0, 6.0 and 4.0, PhCHC*H*), 1.76 (1H, dt, *J* 9.0 and 4.0, C*H*C=O), 1.46 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>], 1.17 (1H, dt, *J* 9.0 and 5.0, C*H*<sub>a</sub>H<sub>b</sub>) and 0.87 (1H, ddd, *J* 8.5, 6.0 and 4.5, CH<sub>a</sub>H<sub>b</sub>); <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$ 172.2 (C1), 138.5 (*ipso*-Ph), 128.8 (Ph), 128.5 (*para*-Ph), 127.0 (Ph), 80.7 [*C*(CH<sub>3</sub>)<sub>3</sub>], 67.3 (C1<sup>''</sup>), 28.1 [C(CH<sub>3</sub>)<sub>3</sub>], 26.1 (C2<sup>'</sup>), 20.6 (C1<sup>'</sup>) and 12.0 (C3<sup>'</sup>).

# (1'R,2'R,1''R)-tert-Butyl 2'-(1''-azido-1''-phenyl-methyl)-cyclopropane carboxylate 30

By method **7b** mesylate **28** (126 mg, 0.34 mmol) gave a yellow gum. The product was purified by DCVC [id 1 cm; 9 cm<sup>3</sup> fractions;  $6 \times$  hexanes; 1-20% EtOAc in hexanes (v/v) – 1% increments; two fractions of each solvent mixture were collected] to give *cyclopropane* **30** (60 mg, 64%) as a clear oil. All analytical data were identical to that reported above.

# (1'R,2'R,1''R)-tert-Butyl 2'-(1''-azido-1''-phenyl-methyl)-cyclopropane carboxylate 30

According to method **7b** tosylate **29** (0.13 g, 0.30 mmol) gave a yellow gum. The product was purified by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions;  $6 \times$  hexanes; 2-20% EtOAc in hexanes (v/v) – 2% increments; two fractions of each solvent mixture were collected] to give *cyclopropane* **30** (42 mg, 52%) as a clear gum. All analytical data were identical with that reported above.

#### (1'R,2'R,1''R)-tert-Butyl 2'-(1''-amino-1''-phenyl-methyl)-cyclopropane carboxylate 34

Azide **30** (169 mg, 0.62 mmol) was dissolved in methanol (10 cm<sup>3</sup>) and the flask was flushed with argon. Pd(OH)<sub>2</sub> on carbon (32 mg; 20wt% dry basis) was added and the flask was flushed with hydrogen, fitted with a hydrogen balloon and stirred vigorously. After stirring overnight (23 hours) the reaction mixture was filtered through a plug of celite and washed with boiling methanol (2 × 25 cm<sup>3</sup>). The combined organic phases were concentrated *in vacuo* to give a yellow solid that was triturated with dichloromethane and dried under reduced pressure to give amine **34** (140 mg, 91%) as a white amorphous powder. e.e. >92% determined by NMR of Mosher's amide derivatives. <sup>19</sup>F NMR (400 MHz; CDCl<sub>3</sub>): Derivative made from R-(+)-Mosher's acid chloride  $\delta$ -68.90. Derivative made from racemic Mosher's acid chloride  $\delta$  –68.78 and –68.90;  $[\alpha]_D^{23}$  –50 (c. 0.3, MeOH); *m/z* (+ESI) found: MH<sup>+</sup>, 248.1645. (C<sub>15</sub>H<sub>22</sub>NO<sub>2</sub> requires *M*, 248.1651); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 2900 (v br NH<sub>2</sub> and CH), 1706 (C=O) and 1155 (C-O); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>)  $\delta$  8.69 (2H, br s, NH<sub>2</sub>),

7.54-7.52 (2H, m, *ortho*-Ph), 7.44 (2H, br tt, *J* 7.5 and 1.5, *meta*-Ph), 7.39 (1H, tt, *J* 7.5 and 1.5, *para*-Ph), 3.80 (1H, d, *J* 9.5, PhC*H*), 1.88-1.80 (2H, m, PhCHC*H* and C*H*C=O), 1.42 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>], 0.96 (1H, ddd, *J* 8.5, 6.0 and 4.5,  $CH_aH_b$ ) and 0.90 (1H, dt, *J* 9.0 and 4.5,  $CH_aH_b$ ); <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$  171.4 (C1), 134.5 (*ipso*-Ph), 128.7 (*meta*-Ph), 128.5 (*para*-Ph), 127.1 (*ortho*-Ph), 80.1 [*C*(CH<sub>3</sub>)<sub>3</sub>], 56.2 (C1''), 27.7 [C(*C*H<sub>3</sub>)<sub>3</sub>], 24.6 (C2'), 20.9 (C1') and 12.8 (C3').

#### (E)-tert-Butyl hept-4-enoate 35

tert-Butyl acetate (1.0 g, 8.6 mmol) was dissolved in anhydrous THF (40 cm<sup>3</sup>) and cooled to -78 <sup>o</sup>C. Freshly prepared LDA (2.76 M, 9.0 mmol) was added by cannula to give a red solution. After <sup>1</sup>/<sub>2</sub> hour HMPA (1.5 cm<sup>3</sup>, 8.6 mmol) was added. After an additional <sup>1</sup>/<sub>2</sub> hour (*E*)-cinnamyl bromide (1.7 g, 8.6 mmol) dissolved in anhydrous THF (10 cm<sup>3</sup>) and cooled to -78 °C was added by cannula. After 6 hours the reaction was quenched by the addition of saturated aqueous NH<sub>4</sub>Cl (20 cm<sup>3</sup>) and allowed to warm to room temperature. The reaction mixture was transferred to a separatory funnel with water (10 cm<sup>3</sup>) and extracted with ethyl acetate ( $3 \times 50$  cm<sup>3</sup>). The combined organic phases were washed with water  $(3 \times 50 \text{ cm}^3)$ , dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated *in vacuo* to give *tert-butyl ester* **35** (1.45 g, 91%) as a light brown liquid that required no further purification.  $R_{\rm f}$  0.35 (5% EtOAc in hexanes, v/v); m/z (+ESI) found: M<sup>+</sup>, 184.1457. (C<sub>11</sub>H<sub>20</sub>O<sub>2</sub> requires M, 184.1463); IR ν<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 1731 (C=O) and 1148 (C-O); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>) δ 5.48 (1H, dt, J 15.5 and 6.0, EtCH=CH), 5.39-5.34 (1H, m, EtCH=CH), 2.25 (2H, br s), 2.24 (2H, br s) (CH<sub>2</sub>CH<sub>2</sub>C=O), 1.97 (2H, quintet, J 7.5, CH<sub>3</sub>CH<sub>2</sub>), 1.42 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>] and 0.93 (3H, t, J 7.5, CH<sub>3</sub>); <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$  172.6 (C1), 133.0 (C5), 127.1 (C4), 80.0 [C(CH<sub>3</sub>)<sub>3</sub>], 35.5 (C2), 28.1 [C3 and C(CH<sub>3</sub>)<sub>3</sub>], 25.5 (C6) and 13.8 (C7); (Found: C, 72.02; H, 11.07. C<sub>11</sub>H<sub>20</sub>O<sub>2</sub> requires C, 71.70; H, 10.94%).

#### (4R,5R)-tert-Butyl 4,5-dihydroxy-heptanoate 36

By method **1** olefin **35** (0.97 g, 5.3 mmol) after 8 hours at 0 °C gave *diol* **36** (0.94 g, 81%) as a clear gum that required no further purification.  $[\alpha]_D^{23}$  +10.3 (c. 2.8, CHCl<sub>3</sub>); *m/z* (+ESI) found: MNa<sup>+</sup>, 241.1416. (C<sub>11</sub>H<sub>22</sub>O<sub>4</sub>Na requires *M*, 241.1411); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 3435 (br., O-H), 1729 (C=O) and 1153 (C-O); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>)  $\delta$  3.43 (1H, ddd, *J* 13.0, 5.0 and 4.0, EtCHC*H*), 3.31 (1H, br dt, *J* 8.5 and 4.5, EtC*H*), 2.44-2.34 (2H, m, CH<sub>2</sub>C=O), 1.83-1.68 (2H, m, CH<sub>2</sub>CHOH), 1.62-1.53 (1H, m, CH<sub>a</sub>H<sub>b</sub>Me), 1.50-1.40 (1H, m, CH<sub>a</sub>H<sub>b</sub>Me), 1.44 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>] and 0.97 (3H, t, *J* 

7.5, CH<sub>3</sub>); <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>) *δ* 173.9 (C1), 80.7 [*C*(CH<sub>3</sub>)<sub>3</sub>], 75.7 (C5), 73.5 (C4), 32.0 (C2), 28.6 (C3), 28.0 [C(*C*H<sub>3</sub>)<sub>3</sub>], 26.3 (C6) and 10.0 (C7).

#### (4RS,5RS)-tert-Butyl 4,5-dihydroxy-heptanoate (±)-36

By method 2 olefin 35 (0.11 g, 0.60 mmol) after 1 day gave *diol* ( $\pm$ )-36 (0.11 g, 87%) as a clear gum that required no further purification. All analytical data for ( $\pm$ )-36 were identical with that for (4*R*,5*R*)-36 reported above.

# (4R,5R)-tert-Butyl 4,5-diphenylphosphinoyloxy-heptanoate 37

By method **3** diol **36** (0.57 g, 2.6 mmol) after 16 hours gave a yellow gum. Purification by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions; 2 × hexanes; 0-100% EtOAc in hexanes (v/v) – 10% increments] gave *bis-phosphinate* **37** (1.20 g, 74%) as a clear gum. e.e. >85% (determined by chiral HPLC); HPLC [ $R_T$  (min), flow rate 1 cm<sup>3</sup>/min, 20% *iso*-propanol in *iso*-hexane, v/v]: 24.8;  $[\alpha]_D^{23}$  +18.5 (c. 1, CHCl<sub>3</sub>);  $R_f$  0.55 (EtOAc); m/z (+ESI) found: MH<sup>+</sup>, 619.2285. (C<sub>35</sub>H<sub>41</sub>O<sub>6</sub>P<sub>2</sub> requires *M*, 619.2378); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 1723 (C=O), 1439 (P-Ph) and 1225 (P=O); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>)  $\delta$  7.67-7.59 (8H, m, *ortho*-Ph), 7.42-7.36 (4H, m, *para*-Ph), 7.34-7.26 (8H, m, *meta*-Ph), 4.55 (1H, tt, *J* 8.0 and 4.0, EtCHC*H*), 4.38 (1H, m, EtC*H*CH), 2.25 (1H, ddd, *J* 15.5, 9.5 and 5.5, C*H*<sub>a</sub>H<sub>b</sub>C=O), 2.14-2.03 (2H, m, CH<sub>a</sub>H<sub>b</sub>C=O and CH<sub>a</sub>H<sub>b</sub>CH<sub>2</sub>C=O), 1.96 (1H, m, CH<sub>a</sub>H<sub>b</sub>CH<sub>2</sub>C=O), 1.84-1.76 (1H, m, CH<sub>a</sub>H<sub>b</sub>Me), 1.73-1.64 (1H, m, CH<sub>a</sub>H<sub>b</sub>Me), 1.36 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>] and 0.76 (3H, t, *J* 7.5, CH<sub>3</sub>); <sup>31</sup>P NMR (162 MHz; CDCl<sub>3</sub>)  $\delta$  31.5 and 31.4; <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$  172.1 (C1), 132.3, 132.2-132.1 (m), 131.7, 131.6, 131.5, 131.2, 131.1, 131.0, 128.5-128.3 (m) (16 × Ph), 80.2 [C(CH<sub>3</sub>)<sub>3</sub>], 77.6 (dd, *J* 6.5 and 3.5, C5), 75.3 (dd, *J* 6.5 and 4.0, C4), 31.3 (C2), 28.0 [C(CH<sub>3</sub>)<sub>3</sub>], 25.9 (d, *J* 3.5, C6), 23.5 (d, *J* 3.5, C3) and 9.9 (C7); (Found: C, 66.43; H, 6.61. C<sub>35</sub>H<sub>40</sub>O<sub>6</sub>P<sub>2</sub>·0.05 EtOAc requires C, 66.68; H, 6.42%).

#### (4RS,5RS)-tert-Butyl 4,5-diphenylphosphinoyloxy-heptanoate (±)-37

By method **3** diol (±)-36 (82 mg, 0.38 mmol) after 24 hours gave a white gum. Purification by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions; 2 × hexanes; 10-100% EtOAc in hexanes (v/v) – 10% increments; 6 × EtOAc] gave *bis-phosphinate* (±)-37 (0.11 g, 48%) as a clear gum. HPLC [ $R_T$  (min), flow rate 1 cm<sup>3</sup>/min, 20% *iso*-propanol in *iso*-hexane (v/v)]: 24.8 and 33.2; All analytical data were identical with that for (4R,5R)-37 reported above.

#### (4R,5R)-tert-Butyl 4,5-methanesulfonyloxy-heptanoate 38

By method **5** diol **36** (0.52 g, 2.38 mmol) after 20 hours gave a yellow liquid. Purification by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions: 2 × hexanes; 0-100% EtOAc in hexanes (v/v) – 10% increments;  $4 \times \text{EtOAc}$ ] gave *bis-mesylate* **38** (0.60 g, 67%) as a clear gum. [ $\alpha$ ]<sub>D</sub><sup>23</sup> +15 (c. 1, CHCl<sub>3</sub>);  $R_f$  0.50 (50% EtOAc in hexanes, v/v); *m/z* (+ESI) found: MNa<sup>+</sup>, 397.0961. (C<sub>13</sub>H<sub>26</sub>O<sub>8</sub>S<sub>2</sub>Na requires *M*, 397.0968); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 1722 (C=O), 1333 (SO<sub>2</sub>) and 1172 (SO<sub>2</sub>); <sup>1</sup>H NMR (400 MHz; CDCl<sub>3</sub>)  $\delta$  4.88 (1H, ddd, *J* 9.0, 5.0 and 4.0, CH<sub>2</sub>CH<sub>2</sub>C*H*), 4.71 (1H, dt, *J* 7.5 and 5.0, CH<sub>3</sub>CH<sub>2</sub>C*H*), 3.10 (3H, s, CH<sub>3</sub>S), 3.09 (3H, s, CH<sub>3</sub>S), 2.49-2.36 (2H, m, CH<sub>2</sub>C=O), 2.09 (1H, dtd, *J* 15.5, 7.5 and 4.0, *CH*<sub>a</sub>H<sub>b</sub>CH<sub>3</sub>CH<sub>3</sub>), 1.44 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>] and 1.06 (3H, t, *J* 7.5, CH<sub>3</sub>); <sup>13</sup>C NMR (100 MHz; CDCl<sub>3</sub>)  $\delta$  171.5 (C1), 82.0 (C5), 81.1 [*C*(CH<sub>3</sub>)<sub>3</sub>], 79.3 (C4), 38.8, 38.6 (2 × CH<sub>3</sub>S), 30.3 (C2), 28.0 [C(*C*H<sub>3</sub>)<sub>3</sub>], 25.7 (C3), 23.5 (C6) and 9.1 (C7); (Found: C, 36.20; H, 6.17. C<sub>13</sub>H<sub>26</sub>O<sub>8</sub>S<sub>2</sub>·0.3 EtOAc requires C, 35.82; H, 6.08%).

#### (4R,5R)-tert-Butyl 4,5-bis-(4'-methyl-phenylsulfonyloxy)-heptanoate 39

By method **6** diol **36** (0.46 g, 2.11 mmol) after 4 days gave a brown gum. Purification by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions: 2 × hexanes; 5-50% EtOAc in hexanes (v/v) – 5% increments; two fractions of each solvent mixture were collected] gave *bis-tosylate* **39** (0.72 g, 64%) as a clear gum.  $[\alpha]_D^{23}$  +41 (c. 1, CHCl<sub>3</sub>);  $R_f$  0.45 (30% EtOAc in hexanes, v/v); *m/z* (+ESI) found: MNa<sup>+</sup>, 549.1606. (C<sub>25</sub>H<sub>34</sub>O<sub>8</sub>S<sub>2</sub>Na requires *M*, 549.1593); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 1726 (C=O), 1366 (S=O) and 1175 (C-O); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>)  $\delta$  7.81 (2H, d, *J* 8.5, *ortho*-Ar), 7.79 (2H, d, *J* 8.5, *ortho*-Ar), 7.37 (2H, d, *J* 8.5, *meta*-Ar), 7.35 (2H, d, *J* 8.5, *meta*-Ar), 4.69 (1H, dt, *J* 9.5 and 3.5, CH<sub>2</sub>CH<sub>2</sub>C*H*), 4.47 (1H, dt, *J* 9.0 and 4.0, CH<sub>3</sub>CH<sub>2</sub>C*H*), 2.47 (3H, s, CH<sub>3</sub>Ar), 2.46 (3H, s, CH<sub>3</sub>Ar), 2.12 (1H, ddd, CH<sub>a</sub>H<sub>b</sub>C=O), 1.99-1.90 (2H, m, CH<sub>a</sub>H<sub>b</sub>C=O and CH<sub>a</sub>H<sub>b</sub>CH<sub>3</sub>), 1.42 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>] and 0.64 (3H, t, *J* 7.5, CH<sub>3</sub>); <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$  171.3 (C1), 145.2, 145.1 (2 × *para*-Ar), 133.1, 133.0 (2 × *ipso*-Ar), 129.9, 129.8 (2 × *meta*-Ar), 128.2, 128.1 (2 × *ortho*-Ar), 82.1 (C5), 80.7 [C(CH<sub>3</sub>)<sub>3</sub>], 79.8 (C4), 30.8, (C2), 28.0 [C(CH<sub>3</sub>)<sub>3</sub>], 24.1 (C3), 21.7 (br. C6 and 2 × CH<sub>3</sub>Ar) and 9.6 (C7); (Found: C, 56.91; H, 6.62. C<sub>25</sub>H<sub>34</sub>O<sub>8</sub>S<sub>2</sub> requires C, 57.01; H, 6.51%).

# (1'*R*,2'*R*,1''*R*)-*tert*-Butyl 2'-(1''-Diphenylphosphinoyloxy-propyl)-cyclopropane carboxylate 40

By method 7a bis-phosphinate 37 (0.80 g, 1.29 mmol) gave a yellow gum. The product was purified by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions;  $2 \times$  hexanes; 10-100% EtOAc in hexanes (v/v) – 10% increments;  $5 \times \text{EtOAc}$ ; to give cyclopropane 40 (0.39 g, 75%) as a white amorphous solid. e.e. >96% (determined by chiral HPLC); HPLC [ $R_T$  (min), flow rate 1 cm<sup>3</sup>/min, 5% iso-propanol in isohexane (v/v)]: 24.4;  $[\alpha]_{D}^{23}$  -25 (c. 1, CHCl<sub>3</sub>); mp 91-92 °C (EtOAc, hexanes);  $R_{\rm f}$  0.60 (80% EtOAc in hexanes, v/v); m/z (+ESI) found: MNa<sup>+</sup>, 423.1721. (C<sub>23</sub>H<sub>29</sub>O<sub>4</sub>PNa requires M, 423.1701); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 1718 (C=O), 1439 (P-Ph), 1217 (P=O) and 1151 (C-O); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>) *S*7.84-7.75 (4H, m, ortho-Ph), 7.53-7.48 (2H, m, para-Ph), 7.46-7.41 (4H, m, meta-Ph), 3.91 (1H, tt, J 8.5 and 6.0, EtCH), 1.84-1.78 (2H, m, CH<sub>2</sub>CH<sub>3</sub>), 1.55 (1H, ddt, J 8.5, 6.5 and 4.0, CHCHEt), 1.44-1.41 [10H, m, CHC=O and C(CH<sub>3</sub>)<sub>3</sub>], 0.97 (3H, t, J 7.5, CH<sub>3</sub>), 0.87 (1H, dt, J 8.5 and 5.0, ring-CH<sub>a</sub>H<sub>b</sub>) and 0.75 (1H, ddd, J 8.5, 6.5 and 4.5, ring-CH<sub>a</sub>H<sub>b</sub>); <sup>31</sup>P NMR (162 MHz; CDCl<sub>3</sub>)  $\delta$  31.0; <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$  172.5 (C1), 132.6 (×2) (d, J 138.5, *ipso*-Ph and d, J 136.0, ipso-Ph), 132.0 (d, J 3.0, para-Ph), 131.9 (d, J 2.5, para-Ph), 131.7 (d, J 10.0, ortho-Ph), 131.4 (d, J 10.5, ortho-Ph), 128.4 (×2) (d, J 13.5, para-Ph d, J 13.0, para-Ph), 80.5 [C(CH<sub>3</sub>)<sub>3</sub>], 80.1 (d, J 6.4, C1''), 29.4 (d, J 4.0, C2''), 28.1 [C(CH<sub>3</sub>)<sub>3</sub>], 25.6 (d, J 3.5, C2'], 20.1 (C1'), 13.6 (C3') and 9.5 (C3"); (Found: C, 68.79; H, 7.34. C<sub>23</sub>H<sub>29</sub>O<sub>4</sub>P requires C, 68.98; H, 7.30%).

# (1'*RS*,2'*RS*,1''*RS*)-*tert*-Butyl 2-(1-Diphenylphosphinoyloxy-propyl)-cyclopropane carboxylate (±)-40

By method **7a** *bis*-phosphinate (±)-**37** (97 mg, 0.16 mmol) gave a white solid. The product was purified by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions; 2 × hexanes; 10-100% EtOAc in hexanes (v/v) – 10% increments; 6 × EtOAc] to give cyclopropane (±)-**40** (10 mg, 16%) as a clear gum. HPLC [ $R_T$  (min), flow rate 1 cm<sup>3</sup>/min, 5% *iso*-propanol in *iso*-hexane (v/v)]: 20.3 and 24.1; All analytical data were identical with that for (1'R,2'R,1''R)-**40** reported above.

#### (1'R,2'R,1''R)-Methyl 2'-(1''-hydroxy-propyl)-cyclopropane carboxylate 44

Cyclopropane **40** (0.12 g, 0.30 mmol) was dissolved in anhydrous methanol (5 cm<sup>3</sup>) and sodium methoxide (81 mg, 1.50 mmol) was added. The reaction mixture was heated to reflux for 4 hours, allowed to cool to room temperature and transferred to a separatory funnel with aqueous sulfate

buffer (25 cm<sup>3</sup>) and extracted with ethyl acetate (3 × 20 cm<sup>3</sup>). The combined organic phases were dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and carefully concentrated *in vacuo* to give a white gum. The product was purified by DCVC [id 4 cm; 20 cm<sup>3</sup> fractions; 2 × hexanes; 10-90% EtOAc in hexanes (v/v) – 10% increments; 5 × EtOAc;] to give *cyclopropane* **44** (34 mg, 72%) as a clear liquid.  $[\alpha]_D^{23}$  –78 (c. 0.5, CHCl<sub>3</sub>);  $R_f$  0.40 (50% EtOAc in hexanes, v/v); m/z (+ESI) found: MNa<sup>+</sup>, 181.0835. (C<sub>8</sub>H<sub>14</sub>O<sub>3</sub>Na requires *M*, 181.0841); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 3450 (br., O-H), 1730 (C=O) and 1174 (C-O); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>)  $\delta$  3.67 (3H, s, OCH<sub>3</sub>), 3.17 (1H, td, *J* 7.0 and 5.5, CHOH), 1.68-1.52 (5H, m, OH, CHCHC=O and CH<sub>2</sub>CH<sub>3</sub>), 1.16 (1H, dt, *J* 9.0 and 4.5, ring-CH<sub>a</sub>H<sub>b</sub>), 0.97 (3H, t, *J* 7.5, CH<sub>2</sub>CH<sub>3</sub>) and 0.95 (1H, ddd, *J* 8.5, 6.5 and 4.5, ring-CH<sub>a</sub>H<sub>b</sub>); <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$  174.3 (C1), 74.2 (C1<sup>''</sup>), 51.8 (OCH<sub>3</sub>), 30.1 (C2<sup>''</sup>), 27.7 (C2<sup>'</sup>), 17.7 (C1<sup>'</sup>), 11.9 (C3<sup>'</sup>) and 9.9 (C3<sup>''</sup>).

# (7R,1'R)-7-(1'-Methanesulfonyloxy-propyl)-2,2,4-trioxo-[1,2]oxathiepane 45

By method **7b** *bis*-mesylate **41** (112 mg, 0.30 mmol) and NaHMDS (0.17 cm<sup>3</sup>, 0.33 mmol) gave a yellow gum. The product was purified by DCVC [id 1 cm; 9 cm<sup>3</sup> fractions; 4 × hexanes; 10-100% EtOAc in hexanes (v/v) – 10% increments – two fractions of each solvent mixture were collected] to give *oxathiepane* **45** (10 mg, 11%) as a yellow oil.  $[\alpha]_D^{23}$  –53.8 (c. 0.5, CHCl<sub>3</sub>); *R*<sub>f</sub> 0.55 (80% EtOAc in hexanes, v/v); *m/z* (+ESI) found: MNa<sup>+</sup>, 323.0230. (C<sub>9</sub>H<sub>16</sub>O<sub>7</sub>S<sub>2</sub>Na requires *M*, 323.0235; IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 1717 (C=O), 1359 (SO<sub>2</sub>) and 1166 (SO<sub>2</sub>); <sup>1</sup>H NMR (400 MHz; CDCl<sub>3</sub>)  $\delta$  4.98 (1H, ddd, *J* 11.0, 4.5 and 2.0, C7-CH), 4.70 (1H, ddd, *J* 7.5, 6.0 and 4.5, C2<sup>2</sup>-CH), 4.35 (1H, d, *J* 16.0, SCH<sub>a</sub>H<sub>b</sub>), 4.27 (1H, d, *J* 16.0, SCH<sub>a</sub>H<sub>b</sub>), 3.16 (1H, ddd, *J* 15.0, 12.5 and 3.0, CH<sub>2</sub>CH<sub>a</sub>H<sub>b</sub>C=O), 3.11 (3H, s, Ms), 2.81 (1H, ddd, *J* 15.0, 7.0 and 2.0, CH<sub>2</sub>CH<sub>a</sub>H<sub>b</sub>C=O), 2.30 (1H, dddd, *J* 15.0, 13.0, 11.0 and 2.0, CH<sub>a</sub>H<sub>b</sub>CH<sub>2</sub>C=O), 2.15 (1H, ddt, *J* 15.5, 7.0 and 2.5, CH<sub>a</sub>H<sub>b</sub>CH<sub>2</sub>C=O), 1.96-1.78 (2H, m, CH<sub>2</sub>Me) and 1.07 (3H, t, *J* 7.5, CH<sub>3</sub>); <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$  196.4 (C4), 84.2, 82.4 (C7 and C1'), 63.6 (C3), 40.5 (C5), 39.0 (CH<sub>3</sub>S), 28.4, 24.3 (C6 and C2') and 9.3 (C3').

# (1'R,2'R,1''S)-tert-Butyl 2'-(1''-azido-propyl)-cyclopropanoate 43

According to method **7b** *bis*-tosylate **42** (0.27 g, 0.51 mmol) gave a white gum. The crude product was dissolved in anhydrous DMF (5 cm<sup>3</sup>), sodium azide (40 mg, 0.62 mmol) was added and the reaction mixture heated to 50 °C overnight (19 hours). The reaction mixture was transferred to a separatory funnel with water (20 cm<sup>3</sup>) and extracted with ethyl acetate ( $3 \times 20$  cm<sup>3</sup>). The combined organic phases were washed with 3 M aqueous HCl ( $2 \times 20$  cm<sup>3</sup>), saturated aqueous NaHCO<sub>3</sub> (20

cm<sup>3</sup>), dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated *in vacuo* to give a clear gum. The product was purified by DCVC [id 1 cm; 7 cm<sup>3</sup> fractions; 4 × hexanes; 2-6% EtOAc in hexanes (v/v) – 2% increments; seven fractions of each solvent mixture were collected] to give *cyclopropane* **43** (75 mg, 62%) as a clear gum. e.e. >94% determined by NMR of Mosher's amide derivatives. <sup>19</sup>F NMR (400 MHz; CDCl<sub>3</sub>): Derivative made from R-(+)-Mosher's acid chloride  $\delta$  –69.33. Derivative made from racemic Mosher's acid chloride  $\delta$  –69.17 and –69.32; [ $\alpha$ ]<sub>D</sub><sup>23</sup> –51 (c. 1.3, CHCl<sub>3</sub>); *R*<sub>f</sub> 0.45 (10% EtOAc in hexanes, v/v); *m/z* (+ESI) found: MNa<sup>+</sup>, 248.1370. C<sub>11</sub>H<sub>19</sub>N<sub>3</sub>O<sub>2</sub>Na requires *M*, 248.1370); IR v<sub>max</sub>(CHCl<sub>3</sub>)/cm<sup>-1</sup> 2095 (N<sub>3</sub>), 1721 (C=O) and 1154 (C-O); <sup>1</sup>H NMR (500 MHz; CDCl<sub>3</sub>)  $\delta$ 2.76 (1H, ddd, *J* 8.5, 7.5 and 6.0, *CH*N<sub>3</sub>), 1.66-1.58 (3H, m, *CH*<sub>2</sub>CH<sub>3</sub> and CHC=O), 1.49 (1H, tdd, *J* 8.5, 6.0 and 4.0, *CH*CHC=O), 1.43 [9H, s, C(CH<sub>3</sub>)<sub>3</sub>], 1.16 (1H, dt, *J* 9.0 and 4.5, ring-*CH*<sub>a</sub>H<sub>b</sub>), 0.98 (3H, t, *J* 7.5, *CH*<sub>3</sub>CH<sub>2</sub>) and 0.76 (1H, ddd, *J* 8.5, 6.0 and 4.5, ring-*CH*<sub>a</sub>H<sub>b</sub>); <sup>13</sup>C NMR (126 MHz; CDCl<sub>3</sub>)  $\delta$ 172.4 (C1), 80.6 [*C*(CH<sub>3</sub>)<sub>3</sub>], 66.1 (C1<sup>\*\*</sup>), 28.0 [C(*C*H<sub>3</sub>)<sub>3</sub>], 27.8 (C2<sup>\*\*</sup>), 24.9 (C2<sup>\*</sup>), 19.6 (C1<sup>\*</sup>), 12.0 (C3<sup>\*</sup>) and 10.3 (C3<sup>\*\*</sup>); (Found: C, 58.45; H, 8.55. C<sub>11</sub>H<sub>19</sub>N<sub>3</sub>O<sub>2</sub> requires C, 58.64; H, 8.50%).

Crystal data for cyclopropane **40**: C<sub>23</sub>H<sub>29</sub>O<sub>4</sub>P, M = 400.43, Triclinic, P1, a = 5.8939(2), b = 8.5144(3), c = 11.3355(4) Å, a = 82.108(2),  $\beta = 87.448(2)$ ,  $\gamma = 82.337(2)^{\circ}$ , U = 558.23(3) Å<sup>3</sup>, Z = 1,  $\mu$ (Mo-K $\alpha$ ) = 0.147 mm<sup>-1</sup>, 5699 reflections collected at 180(2) K using an Oxford Cryosystems Cryostream cooling apparatus, 3689 unique ( $R_{int} = 0.025$ ); R1 = 0.035, wR2 = 0.096 [ $I > 2\sigma(I$ ]], Absolute structure parameter -0.08(8).

The structures were solved with SHELXS-97,<sup>8</sup> and refined with SHELXL-97.<sup>8</sup>

CCDC reference number 600428. See http://www.rsc.org/suppdata/ for crystallographic data in .cif or other electronic format.

#### References

- 1. W. G. Kofron and L. M. Baclawski, J.Org. Chem., 1976, 41, 1879.
- 2. D. Sejer Pedersen and C. Rosenbohm, Synthesis, 2001, 2431.
- 3. Mestre-C software, ver. 4.5.6, www.mestrec.com.
- 4. H. Kolb, M. S. VanNiewenhze, and K. B. Sharpless, Chem. Rev., 1994, 94, 2483.
- 5. J. Eames, H. J. Mitchell, A. Nelson, P. O'Brien, S. Warren, and P. Wyatt, *Tetrahedron Lett.*, 1995, **36**, 1719.
- 6. J. Eames, H. J. Mitchell, A. Nelson, P. O'Brien, S. Warren, and P. Wyatt, *J. Chem.Soc., Perkin Trans. 1*, 1999, 1095.
- 7. U. Jahn, Chem. Commun., 2001, 1600.
- 8. Sheldrick, G. M., University of Göttingen, Germany, 1997, ver. SHELXS-97/SHELXL-97.