The ESI for Org. Biomol. Chem., 2007, 5, 2756-2757 was updated on 11th May 2020. Some of the NMR spectra included in the original submission were inadvertently cropped and have been replaced.

Simple, Efficient Route to O- and S-Linked Carbohydrate Functionalized N-

Carboxyanhydrides (GlycoNCAs)

Matthew I. Gibson1, Gregory J. Hunt, Neil R. Cameron*

IRC in Polymer Science and Technology, Department of Chemistry, University of Durham, Durham, UK, DH1 3LE

n.r.cameron@durham.ac.uk

Table of Contents:

S1 – S2
S2 - S4
S5 – S17
S18 – S27

General Experimental Details

All chemicals were used as supplied unless otherwise stated. *N*-Boc-L-threonine (>98%), *N*-Boc-L-serine (>98%) and *N*-Boc-L-cysteine were purchased from Fluka. Acetobromo- α -D-glucose (>95%), acetobromo- α -D-galactose (>95%), iodine (>99.8%), potassium carbonate (>99%), trifloroacetic acid (>98%), sulfuric acid (95-98%) and potassium carbonate (>99.5%) were purchased from Aldrich. Triphosgene (>96%) and triethylamine (>98%) were purchased from Lancaster synthesis. THF, MeCN and DCM (>99.5%) were purchased from Fischer Scientific and dried by passage through two alumina columns using an Innovative Technology Inc. solvent purification system and stored under N₂. Anhydrous ethyl acetate (>99.8%, <0.005% H₂O) was purchased from Aldrich. Hexane (Fischer Scientific >99%) was dried over

¹ Current address. Laboratoire des Polymers, Ecole Polytechnique Fédérale de Lausanne, Batiment MXD, Station 12, CH-1015, Lausanne

3A molecular sieves. 3A molecular sieves (Aldrich) were activated in an oven at 200_oC before use. Glass-backed silica plates (Biotage) were used for TLC with 5wt % phosphomolybdic acid in ethanol, developed at 80_oC as the visualising agent. Column chromatography was undertaken on a Biotage SP1 flash chromatography unit using UV detectors at 200 and 236nm. Gradients were calculated the on-board software using Rf values from TLC.

NMR spectroscopy (1H, 13C, COSY, NOESY, HSQC) was conducted on a Varian Inova-500 at 500Mhz. Mass spectral analyses were preformed on a Micromass LCT using positive or negative electrospray mode. Infrared spectroscopy was conducted on a Nicolet Nexus FT-IR as a KBr disc. Liquid samples were analysed by direct injection of the reaction medium into a liquid cell with KBr windows. Elemental analyses were conducted on an Exeter Analytical E-440 elemental analyser.

GENERAL EXPERIMENTAL PROCEDURE

The same methodology was applied for the synthesis of each glycosylated NCA, with only the carbohydrate or *N*-Boc amino acid varying in each case. The full procedure for the synthesis of Thr(GluAc) NCA is detailed below.

Synthesisof $N\alpha$ -(Butoxycarbonyl)-3-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)-L-threonine - BocThr(GluAc) - 2(a)

N-Boc-L- threonine (3.0g, 13.6mmol), potassium carbonate (1.41g, 10.2mmol) and acetobromoglucose (2.79g, 6.8mmol) were dissolved in 40 ml of dry acetonitrile under an N₂ atmosphere with stirring for 5 minutes to ensure dissolution. Iodine (2.59g, 10.2mmol) was then added in a single portion against a flow of N₂. The vessel was sealed and allowed to stir at ambient temperature with the exclusion of light for 5 hours.

To the still stirring solution, saturated sodium thiosulfate (aqueous) was added until the deep red colouration had disappeared, to leave a slightly yellow solution. The insoluble components (residual potassium carbonate) were removed by filtration and the filtrate concentrated to ¹/₄ of its original volume under reduced pressure on a rotorary evaporator. 40 mL of dichloromethane were then added, and the solution extracted with sodium bicarbonate (aqueous 5% w/v, 50 mL) followed by brine (2 x 50mL), and the organic layer dried over magnesium sulphate. Separation by column chromatography (Hexane/THF 10:1 \rightarrow 2:8) yielded the title product as a white solid. Isolated yield 2.05 g, 55%.

Synthesis of $3-O-(2,3,4,6-tetra-O-acetyl-\beta-D-glucopyranosyl)-L-threonine acetic acid salt – Thr(GluAc) – 2(b)$

BocThr(GluAc) (1g, 1.82 mmol) was dissolved in 10 mL of DCM under N₂. TFA (0.7 ml, 9.11 mmol) was then added dropwise and allowed to stir at ambient temperature for 90 minutes. The solution was then concentrated under reduced pressure on a rotorary evaporator to give a thick syrup. The syrup was taken up in 20 mL of 10% w/v acetic acid(aq) solution and extracted 3 times with 20 mL portions of n-hexane. The aqueous solution was freeze dried, and re-precipitated from CHCl₃ to di-ethyl ether to afford the title product without further purification as an off-white solid: 0.90 g, 97% yield.

Synthesis of $3-O-(2,3,4,6-tetraacetyl)-\beta-D-glucopyranose)-L-threonine-N-carboxyanhydride - Thr(GluAc)NCA - 2(c)$

Thr(GluAc) (100mg, 0.20 mmol) was taken up in 5 mL of EtOAc under N₂. Triphosgene (39mg, 0.13 mmol) was added in a single portion, against the flow of N₂ followed by α -pinene (0.37mL, 2.34 mmol) and allowed to stir at ambient temperature for 20 hours then concentrated to ½ of its original volume on a rotary evaporator under reduced pressure, with the water bath below 30_oC. The concentrate was precipitated into hexane at -20_oC. This was re-precipitated from ethyl acetate into hexane at -20_oC three times. The white solid was obtained by centrifugation following each precipitation, and finally dried under vacuum to give a white solid. Yield 81mg, 87% yield.

CHARACTERISATION

As indicated in the main text, NCA polymerisation is often complicated by residual amino acid or acidic impurities remaining from the synthetic procedure1. Here acidic impurities were removed by the HCl scavenger α -pinene. In the 1H NMR spectra of the NCAs, trace amounts of amino acid residues (appearing slightly shifted relative to the main peaks) can be seen. Additional purification is possible (by an aqueous extraction) but this should only be conducted prior to polymerisation, as further degradation reactions will occur rapidly negating any increase in purity2. This purification was not undertaken, allowing comparison with other published glycoNCA synthetic procedures.

$N\alpha$ -(Butoxycarbonyl)-3-O-(2,3,4,6-tetra-O-acetyl- β -D-galatopyranosyl)-L-

threonine - BocThr(GalAc) - 1(a)

1**H NMR** (CDCl₃) δ_{ppm} ; 5.73 (1H, d, $J_{1-2} = 8.26$ Hz, H₁), 5.42 (1H, dd, $J_{4-3} = 3.41$ Hz, $J_{4-5} = 0.94$, H₄), 5.33 (1H, dd, $J_{2-1} = 8.26$ Hz, $J_{2-3} = 10.44$ Hz, H₂), 5.08 (1H, dd, $J_{3-4} = 3.41$ Hz, $J_{3-2} = 10.44$ Hz, H₃), 4.22 (1H, d, $J_{7-16} = 9.2$ Hz, H₇), 4.35 (1H, m, H₈), 4.20 (1H, dd, $J_{6a-5} = 6.40$ Hz, $J_{6a-6b} = 10.86$ Hz, H_{6a}), 4.07 (1H, dd, $J_{6b-5} = 6.14$ Hz, $J_{6b-6a} = 10.89$ Hz, H_{6b}), 4.04 (1H, td, $J_{5-4} = 0.74$ Hz, $J_{5-6a} = J_{5-6b} = 6.47$ Hz, H₅), 1.99 – 2.06 (12H, m, H₁₅), 1.26 (3H, d, $J_{16-7} = 6.38$ Hz, H₁₆).

13**C NMR** (CDCl₃) δ_{ppm}: 170.6 (C₉), 169.9-170.1 (4x C₁₄), 156.0 (C₁₁), 92.9 (C₁), 80.1 (C₁₂), 72.3, (C₅), 71.4 (C₃), 67.9 (C₂), 67.2 (C₄), 62.3 (C₈), 60.8 (C₆), 59.0 (C₇), 28.2 (C₁₃), 20.5-20.7 (4xC₁₅), 19.9 (C₁₆),

IR (KBr disc) cm-1: 3447 (br., N-H), 2981 (C-H), 1757 (C=O Ac), 1718 (C=O Boc). **MS** (ES+) m/z = 572.2 [M+Na]+ 100%.

Elemental (Found %) C, 50.48; H, 6.54; N, 2.36 (Expected %) C, 50.27; H, 6.42; N, 2.55.



 $N\alpha$ -(Butoxycarbonyl)-3-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)-L-

threenine BocThr(GluAc) - 2(a)

1**H NMR** (CDCl₃) δ_{ppm} ; 5.75 (1H, d, *J*₁₋₂ = 8.17Hz, H₁), 5.30 (1H, d, *J*₈₋₇ = 9.6Hz, H₈), 5.26 (1H, t, *J* = 9.47Hz, H₃), 5.15 (1H, dd, *J*₂₋₁ = 8.18Hz, *J*₂₋₃ = 9.60Hz, H₂), 5.10 (1H, t, *J* = 9.70Hz, H₄), 4.33 (1H, m, H₈), 4.30 (1H, dd, *J*_{6b-5} = 5.02Hz, *J*_{6b-6a} = 12.44Hz, H_{6b}), 4.23 (1H, dd, *J*₇₋₉ = 6.36Hz, *J*₇₋₈ = 9.7Hz, H₇), 4.09 (1H, dd, *J*_{6a-5} = 2.19Hz, *J*_{6a-6b} = 12.36Hz, H_{6a}), 3.82 (1H, ddd, *J*_{5-6a} = 2.20Hz, *J*_{5-6b} = 4.96Hz, *J*₅₋₄ = 10.07Hz, H₅), 2.01 – 2.06 (12H, m, H₁₅), 1.25 (3H, d, *J*₁₆₋₇ = 6.39Hz, H₁₆).

13**C NMR** (CDCl₃) δ_{ppm}; 170.9 (C₉), 169.4-167.0 (4xCl₄), 156.0 (Cl₁), 92.4 (Cl), 80.1 (Cl₂), 72.9 (C₅), 72.5 (C₃), 70.0 (C₂), 67.8 (C₄), 67.5 (C₈), 61.3 (C₆), 58.9 (C₇), 28.2 (Cl₆), 20.5 - 20.7 (4xCl₅), 19.9 (Cl₆).

IR (KBr disc) cm-1: 3447 (br., N-H), 2980 (C-H), 1757 (C=O Ac), 1718 (C=O Boc). **MS** (ES+) m/z = 572.2 [M+Na]+ 100%.

Elemental (Found %) C, 50.16; H, 6.38; N, 2.35 (Expected %) C, 50.27; H, 6.42; N, 2.55.



 $N\alpha$ -(Butoxycarbonyl)-3-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)-L-serine - BocSer(GalAc) - 3(a)

1**H NMR** (CDCl₃) δ_{ppm} ; 5.73 (1H, d, $J_{1-2} = 8.21$ Hz, H₁), 5.46 (1H, dd, $J_{4-5} = 1.08$ Hz, $J_{4-3} = 3.35$ Hz, H₄), 5.31(1H, dd, $J_{2-1} = 8.28$ Hz, $J_{2-3} = 10.43$ Hz, H₂), 5.08 (1H, dd, $J_{3-4} = 3.45$ Hz, $J_{3-2} = 10.45$ Hz, H₃), 1.99 – 2.06 (12H, m, H₁₅), 1.43 (9H, s, H₁₃).

13**C NMR** (CDCl₃) δ_{ppm}; 169.2 – 170.5 (4x C₁₄ and C₉), 155.0 (C₁₁), 89.8 (C₁), 80.3 (C₁₂), 72.1 (C₄), 70.8 (C₂), 67.5 (C₃), 66.7 (C₅), 62.0 (C₈), 61.0 (C₆), 57.7 (C₇), 29.2 (C₁₃), 20.4 – 20.7 (4x C₁₅).

IR (KBr disc) cm-1: 3411 (br., N-H), 2976 (C-H), 1760 (C=O Ac), 1714 (C=O Boc). **MS** (ES+) m/z = 558.2 (85%) [M+Na]+, 599.2 (100%) [M+Na+CH₃CN]+.

Elemental (Found %) C, 49.12; H, 6.30; N, 2.30 (Expected %) C, 49.34; H, 6.21; N, 2.62.



 $N\alpha$ -(Butoxycarbonyl)-3-O-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)-L-serine - (BocSer(GluAc) - 4(a)

1**H NMR** (CDCl₃) δ_{ppm} ; 5.75 (1H, d, $J_{1-2} = 8.13$ Hz, H₁), 5.40 (1H, dd, $J_{8-7a} = 2.79$ Hz, $J_{8-7b} = 6.67$ Hz, H₈), 5.26 (1H, t, J = 9.38Hz, H₃), 5.14 (1H, dd, $J_{2-1} = 8.15$ Hz, $J_{2-3} = 9.47$ Hz, H₂), 5.11 (1H, t, J = 9.90Hz, H₄), 4.35 (1H, m, H_{7b}), 4.28 (1H, dd, $J_{6b-5} = 4.81$ Hz, $J_{6b-6a} = 12.45$ Hz, H_{6b}), 4.10 (1H, dd, $J_{6a-5} = 2.20$ Hz, $J_{6a-6b} = 12.48$ Hz, H_{6a}), 4.02 (1H, m, H_{7a}), 3.83 (1H, ddd, $J_{5-6a} = 2.15$ Hz, $J_{5-6b} = 4.65$ Hz, $J_{5-4} = 9.95$ Hz, H₅), 1.99 – 2.07 (12H, m, H₁₅), 1.43 (9H, s, H₁₃).

13**C NMR** (CDCl₃) δ_{ppm}: 170.8 (C₉), 169.2 – 170.0 (4x C₁₄), 155.1 (C₁₁), 92.4 (C₁), 72.8 (C₅), 72.4 (C₃), 71.1 (C₁₂), 70.0 (C₂), 67.6 (C₄), 62.3 (C₈), 61.0 (C₆), 56.2 (C₇), 28.0 (C₁₃), 20.5 – 20.7 (4xC₁₅).

IR (KBr disc) cm-1: 3447 (br., N-H), 2979 (C-H), 1753 (C=O Ac), 1717 (C=O Boc).**MS** (ES+) m/z = 558.2 (100%) [M+Na]+.

Elemental (Found %) C, 49.52; H, 6.25; N, 2.10 (Expected %) C, 49.34; H, 6.21; N, 2.62.



Nα-(Butoxycarbonyl)-3-*O*-(2,3,4,6-tetraacetyl-β-D-galactopyranosyl-(1-4)-1,2,3,4,6-tetra-*O*-acetyl-β-D-glucopyranose)-L-serine - BocSer(LacAc) – 5(a) 1H NMR (CDCl₃) $\delta_{ppm:}$ 5.77 (1H, d, *J*₁₋₂ 7.50Hz, H₁), 5.72 (1H, d, *J*_{1'-2'} = 9.00Hz, H_{1'}), 5.54 (1H, t, *J* = 9.68Hz, H_{4'}), 5.41 (1H, t, *J* = 3.5Hz, H₄), 5.30 (1H, d, *J*_{3'-2'} = 9.52 Hz, H_{3'}), 5.25 (1H, t, *J* = 9.00 Hz, H₃), 5.07-5.15 (2H, m, H_{2'},H₂), 4.24-4.31 (1H, m, H_{6b}), 4.01-4.17 (4H, m, H_{6'a}, H_{6'b}, H_{6a}, H_{5'}), 3.88 (1H, td *J*₅₋₆ = 6.0Hz, *J*₅₋₄ = 1.5Hz, H₅), 3.72 (1H, t, *J*₃₋₄ = 6.00 Hz, H₃), 1.96 - 2.16 (21H, m, H₁₅), 1.41 - 1.44 (9H, s(b), H₁₃). 13**C NMR** (CDCl₃) δ_{ppm}: 171.1 (C₉), 166.5-170.3 (4 x C₁₄), 157 (C₁₁), 94.0 (C₁[']), 92.6 (C₁), 73.1 (C₄), 72.6 (C₅), 72.1 (C₃[']), 71.4 (C₅[']), 70.7 (C₂[']), 70.3 (C₃), 68.3 (C₂[']), 66.9 (C₄[']), 61.8 (C₆), 61.1 (C₆[']), 30.2 (C₁₃), 20.7-20.9 (4xC₁₅).

MS (ES+) $m/z = 887.7 [M+Na+CH_3CN]_+$.

Elemental Found (%) C 49.39, H 6.33, N 2.22, Expected (%)C 49.57; H 6.00; N 1.70 IR (KBr disc) cm-1: 3450 (br., N-H), 2970 (CH), 1755 (C=O Ac), 1717 (C=O Boc).



 N_{α} -(Butoxycarbonyl)-3-S-(2,3,4,6-tetra-O-acetyl- β -D-glucopyranosyl)-L-cysteine - BocCys(GluAc) - 6(a)

1**H NMR** (CDCl₃) $\delta_{ppm:}$ 6.25 (1H, d, J₁₋₂ = 9.11Hz, H₁), 5.49 (1H, d, J₄₋₃ = 3.50Hz, H₄), 5.29 (1H, t, J₂₋₁ = 9.00 Hz, H₂), 5.10 -5.20 (4H, m, H_{7a}, H_{7b}, H₈, H₃), 4.93 (1H, dd, J₅₋₄ = 3.5Hz, J₅₋₆ = 6.50Hz, H₅), 4.88 (1H, dd, J_{6a-5} = 8.00Hz, J_{6a-6b} 2.00Hz, H_{6a}3.91 (1H, dd, J_{6a-6b} = 4.00Hz, J_{6b-5}, 9.50Hz, H_{6b}), 2.04 - 2.11 (12H, m, H₁₅), 1.36 -1.38 (9H, s(b), H₁₃).

13C NMR (CDCl₃) δ_{ppm}: 171.5 (C9), 169.5-170.5 (4 x C14), 158 (C11), 90.1 (C1), 70.1 (C5), 69.8 (C3), 68.7 (C2), 67.7 (C4), 67.3 (C8), 62.2 (C6), 61.8 (C7), 32.9 (C13), 20.8 - 21.2 (4xC15).

MS (ES+) $m/z = 615.4 [M+Na+CH_3CN] + 100\%$.

IR (KBr disc) cm-1: 3450 (br., N-H), 2980 (C-H), 1757 (br., C=O Ac), 1715 (C=O Boc).



 $N\alpha$ -(Butoxycarbonyl)-3-S-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)-Lcysteine - BocCys(GalAc) - 7(a) 1**H NMR** (CDCl₃) δ_{ppm} : 6.04 (1H, d, J₁₋₂ = 6.00Hz, H₁), 5.46 (1H, dd, J₄₋₃ = 2.50Hz, J₄₋₅ = 6.00Hz, H₄), 5.15-5.23 (2H, m, H₂, H₃), 4.99 (1H, dd, J_{6b-6a} = 3.50Hz, J_{6b-5} = 4.50Hz, H_{6b}) 4.69 (1H, dd, J_{6a-6b} = 2.50Hz, J_{6a-5} = 7.50Hz, H_{6a}), 4.39 (1H, d, J₄₋₅ = 6.50Hz, H₄), 4.14 (1H, ddd, J_{5-6a} = 3.00Hz, J_{5-6b} 3.60Hz, J₅₋₄ = 7.06Hz, H₅), 2.05 - 2.15 (12H, m, H₁₅), 1.42 - 1.44 (9H, s(b), H₁₃).

13C NMR (CDCl₃) δ_{ppm}; 171.1 (C₉), 169.2 -170.4 (4 x C₁₄), 154.0 (C₁₁), 91.1 (C₁), 70.1 (C₅), 69.6 (C₃), 68.8 (C₂), 68.3 (C₄), 67.3 (C₈), 62.2 (C₆), 61.7 (C₇), 33.2 (C₁₃), 20.7 - 21.1 (4xC₁₅).

IR (KBr disc) cm-1: 3450 (br., N-H), 2985 (C-H), 1754 (br., C=O Ac), 1713 (C=O Boc). **MS** (ES+) m/z = 574.9 [M+Na]+.

 $3-O-(2,3,4,6-tetra-O-acetyl-\beta-D-galactopyranosyl)-L-threonine acetic acid salt - Thr(GalAc) - 1(b)$

1**H NMR** (CDCl₃) δ_{ppm} ; 5.96 (1H, d, J₁₋₂ = 8.07Hz, H₁), 5.45 (1H, d, J₄₋₃ = 2.89Hz, H₄), 5.31 (1H, dd, J₂₋₁ = 8.25Hz, J₂₋₃ = 10.10Hz, H₂), 5.22 (1H, dd, J₃₋₂ = 10.44Hz, J₃₋₄ = 3.04Hz, H₃), 4.39 (1H, q, J₇₋₁₅ = J₇₋₈ = 6.27Hz, H₇), 4.19 (1H, dd, J_{6b-6a} = 10.31Hz, J_{6b-5} = 5.91Hz, H_{6b}), (1H, dd, J_{6a-6b} = 10.84Hz, J_{6a-5} = 5.92Hz, H_{6a}), 4.08 (1H, dd, J_{5-6a} = 6.62Hz, J_{5-6b} = 7.88Hz, H₅), 3.97 (1H, m, H₈), 1.95 – 2.06 (15H, m, H₁₁ + H₁₃), 1.41 (1H, d, J₁₅₋₇ = 5.88, H₁₅).

13**C NMR** (CDCl₃) δ_{ppm}; 175.6 (C₁₄), 179.8 – 171.0 (4xC₁₂), 166.6 (C₉), 93.7 (C₁), 71.6 (C₅), 70.3 (C₃), 67.5 (C₂), 66.6 (C₄), 65.8 (C₇), 61.9 (C₅), 60.6 (C₆), 59.2 (C₈), 19.5 – 20.9 (4xC₁₁, C₁₃, C₁₅).

MS (ES+) $m/z = 449.9 [M]_+ (100\%)$.



 $3-O-(2,3,4,6-tetra-O-acetyl-\beta-D-glucopyranosyl)-L-threonine acetic acid salt - Thr(GluAc) - 2(b)$

1**H NMR** (CDCl₃) δ_{ppm} ; 6.00 (1H, d, J₁₋₂ = 8.12Hz, H₁). 5.35 (1H, t, J = 9.41Hz, H₃), 5.12 (2H, t, 9.24Hz, H₂ + H₄), 4.35 (1H, q, J₇₋₈ = J₇₋₁₅ = 6.14Hz, H₇), 4.25 (1H, dd, J_{6a-6b} = 12.37Hz, J_{6a-5} = 2.74Hz, H_{6a}), 4.15 – 4.22 (2H, m, H₈ + H_{6b}), 4.06 (1H, d(b), J= 9.95Hz, H₅), 1.99 – 2.14 (12H, m, H₁₁ + H₁₃), 1.49 (3H, d, J₁₅₋₇ = 6.08Hz, H₁₅). **MS** (ES+) m/z = 449.9 [M]+ (100%).



3-O-((2,3,4,6-tetra-O-acetyl)-β-D-glucopyranosyl)-L-serine acetic acid salt -Ser(GluAc) – 3(b)

1H NMR (CDCl₃) δ_{ppm} ; 5.90 (1H, d, J₁₋₂ = 8.58Hz, H₁). 5.33 (1H, td, J₂₋₃ = 2.00Hz, J₂₋₁ = 8.5Hz, H₂), 5.29 (1H, t, J₃₋₄ = J₃₋₂ = 5.00Hz, H₃), 5.21 (1H, dd, J₄₋₅ = 7.45Hz, J₄₋₃ = 3.50Hz, H₄), 4.22 (1H, t, J₈₋₇ = 4.50Hz, H₈), 4.02-4.14 (4H, m, H_{6a}, H_{6b}, H_{7b}, H₅), 3.86 (1H, dd, J_{7a-7b} = 3.63Hz, J_{7a-8} = 4.43Hz, H_{7a}), 1.98- 2.18 (15H, m, H₁₁, H₁₃) **13C NMR** (CDCl₃) δ_{ppm} ; 166.6 - 171 (C₉, 4xC₁₂, C₁₄), 98.9 (C₁), 72.2 (C₅), 71.0 (C₃), 69.8 (C₂), 67.4 (C₄), 66.2 (C₈), 60.3 (C₈), 59.1 (C₆), 58.9 (C₇), 18.4 - 21.4 (4xC₁₁, C₁₃). **MS** (ES+) m/z = 435.0[M+] 80%, 458.7[M+Na]+ 20%.



3-O-((2,3,4,6-tetra-O-acetyl)-β-D-galactopyranosyl))-L-serine acetic acid salt -Ser(GalAc) – 4(b)

1H NMR (CDCl₃) δ_{ppm} ; 6.04 (1H, d, J₁₋₂ = 6.00Hz, H₁), 5.05 (1H, dd, J₃₋₄ = 3.5Hz, J₃₋₂ = 4.5Hz, H₃), 5.46 (1H, m, H_{6b}), 4.91 (1H, t, J = 4.50Hz, H₂), 4.80 (1H, m, H_{6a}), 4.69 (1H, dd, J_{7a-7b} = 2.50Hz, J_{7a-8} = 7.5Hz, H_{7a}), 4.39 (1H, t, J₄₋₅ = J₄₋₃ = 6.5Hz, H₄), 4.14 (1H, ddd, J_{5-6a} = 3.00Hz, J_{5-6b} = 3.60Hz J₅₋₄ = 7.06Hz, H₅), 2.05 - 2.15 (12H, m, H₁₁). **13C NMR** (CDCl₃) δ_{ppm} ; 174.8 (Cl₄), 168.2-170.6 (C₉, 4xCl₂), 99.1 (Cl₁), 72.6 (C₅), 72.3 (C₄), 71.0 (C₃), 69.1 (C₂), 65.2 (C₈), 60.2 (C₈), 59.3 (C₆), 59.0 (C₇), 20.5 - 21.2 (4xCl₁). **MS** (ES+): m/z = 435.1[M+] 100%.



 $\label{eq:solution} \begin{array}{l} 3-O-((2,3,4,6-tetra-O-acetyl)-\beta-D-galactopyranosyl-(1-4)-(2,3,4,6-tetra-O-acetyl)-\beta-D-glucopyranose)-L-serine acetic acid salt - Ser(LacAc) - 5(b) \end{array}$

1**H NMR** (CDCl₃) δ_{ppm} : 5.77 (1H, d, J₁₋₂ 7.50Hz, H₁), 5.72 (1H, d, J_{1'-2'} 9.00Hz, H_{1'}), 5.54 (1H, t, J = 9.68Hz, H_{4'}), 5.41 (1H, t, J = 3.5Hz, H₄), 5.30 (1H, t, J_{2',3'} = 9.52Hz, H_{3'}), 5.25 (1H, t, J₂₋₃ = 9.00Hz, H₃), 5.07-5.15 (2H, m, H_{2'}, H₂), 4.24 - 4.31 (1H, m, H_{6b}), 4.01-4.17 (4H, m, H_{6'a}, H_{6'b}, H_{6a}, H_{5'}), 3.88 (1H, td, J₄₋₅ = 6.0Hz, J₅₋₆ 1.5Hz, H₅), 3.72 (1H, t, J₄₋₅ = 6.00Hz, H₄), 1.96 - 2.16 (24H, m, H₁₁).

13**C** NMR (CDCl₃) δ_{ppm} ; 166.5 - 171.1 (C₉, 4 x C₁₂, C₁₄), 94.0 (C₁'), 92.6 (C₁), 73.1 (C₄), 72.6 (C₅), 72.1 (C₃'), 71.4 (C₅), 70.7 (C₂'), 70.3 (C₃), 68.3 (C₂'), 66.9 (C₄'), 61.6 (C₆), 61.1 (C₆'), 20.7-20.9 (4 x C₁₁, C₁₃). **MS** (ES+) m/z = 723.6 (100%), [M]+.



3-S-(2,3,4,6-tetra-O-acetyl-β-D-glucopyranosyl)-L-cysteine acetic acid salt -Cys(GluAc) – 6(b)

1**H NMR** (CDCl₃) δ_{ppm} : 6.12 (1H, d, J₁₋₂ = 6Hz, H₁), 5.38 (1H, d, J₂₋₃ = 3.5Hz, H₂), 5.29 (1H, t, J₄₋₅ = 9.0Hz, H₄) 5.29 (1H, t, J₄₋₅ = 9.0Hz, H₄) 5.06 (4H, m, H_{6a}, H_{6b}, H_{7a}, H₈), 4.90 (1H, dd, J_{7b-7a} = 3.5Hz, J_{7b-8} 6.5Hz, H_{7b}), 4.79 (1H, dd, J₁₋₂ 8.00Hz, J₃₋₄ = 2.0Hz, H₃), 3.93 (1H, dd, J₅₋₆ = 4.00Hz, J₄₋₅ = 9.5Hz, H₅), 1.99 - 2.10 (15H, m, H₁₁, H₁₃). 13C NMR (CDCl₃) δ_{ppm} : 169.5-171.5 (4xC₁₂, C₁₄), 166.4 (C₉), 90.1 (C₁), 70.1 (C₅), 69.8 (C₃), 68.7 (C₂), 67.7 (C₄), 67.3 (C₈), 62.2 (C₆), 61.8 (C₇), 20.8 - 21.2 (4xC₁₁, C₁₃). MS (ES+) m/z = 451.1 [M]+.



 $3-S-((2,3,4,6-tetra-O-acetyl)-\beta-D-galactopyranosyl)-L-cysteine acetic acid salt - Cys(GalAc) - 7(b)$

1**H NMR** (CDCl₃) δ_{ppm}: 6.02 (1H, d, J₁₋₂ = 6.00Hz, H₁), 5.33 (1H, dd, J₂₋₁ = 6.10Hz, J₂₋₃ = 2.50Hz, H₂), 5.05 (1H, dd, J₃₋₄ 3.50Hz, J₂₋₃ = 4.50Hz, H₃), 4.61- 4.81 (3H, m, H_{6a}, H_{6b}, H₈), 4.55 (1H, dd, J_{7a-7b} = 2.50Hz, J₇₋₈ = 7.50Hz, H_{7a}), 4.38 (1H, t, J₃₋₄ = 6.5Hz, H₄), 4.12 (1H, ddd, J_{5-6a} = 3.00Hz, J_{5-6b} = 3.60Hz, J₅₋₄ = 7.06Hz, H₅), 1.99 - 2.15 (15H, m, H₁₁, H₁₃).

13C NMR (CDCl₃) δ_{ppm}: 169.2 -171.1 (4xC₁₂, C₁₄), 166.9 (C₉), 91.1 (C₁), 70.1 (C₅), 69.6 (C₃), 68.8 (C₂), 68.3 (C₄), 67.3 (C₈), 62.5 (C₆), 62.2 (C₇), 18.2 - 21.1 (4xC₁₁,C₁₃). **MS** (ES+) m/z = 451.0 [M + Na]+



 $\label{eq:2.1} 3-O-((2,3,4,6-tetra-O-acetyl)-\beta-D-galactopyranose))-L-threonine-N-carboxyanhydride - (Thr(GalAc)NCA - 1(c)$

1**H NMR** (CDCl₃) δ_{ppm} ; 6.16 (1H, s, H₉), 5.68 (1H, d, J₁₋₂ = 8.18Hz, H₁), 5.43 (1H, d, J₄₋₃ = 2.69 Hz, H₄), 5.29 (1H, dd, J₂₋₃ = 10.40Hz, J₂₋₁ = 8.22Hz, H₂), 5.11 (1H, dd, J₃₋₂ = 10.45Hz, J₃₋₄ = 3.38Hz, H₃), 4.80 (1H, m, H₇), 4.00 (1H, d, J₈₋₇ = 4.83Hz, H₈), 1.96 – 2.19 (12H, m, H₁₂), 1.55 (3H, d, J₁₄₋₇ = 6.32Hz, H₁₄).

13C NMR (CDCl₃) δ_{ppm}; 169.8 – 170.4 (4 x C₁₃), 168.0 (C₁₁), 157.9 (C₁₀), 93.3 (C₁), 75.4 (C₇), 72.2 (C₅), 70.5 (C₃), 67.9 (C₂), 66.8 (C₄), 61.1 (C₆), 60.2 (C₈), 20.5 – 21.0 (4 x C₁₂, C₁₄).

HRMS (ES+) m/z = 498.1230 [M+Na]+; expected 498.1218



 $\label{eq:2.1} 3-O-((2,3,4,6-tetra-O-acetyl)-\beta-D-glucopyranose))-L-threonine-N-carboxyanhydride - Thr(GluAc)NCA - 2(c)$

1**H NMR** (CDCl₃) δ_{ppm} ; 6.30 (1H, s, H₉), 5.72 (1H, d, J₁₋₂ = 8.14Hz, H₁), 5.26 (1H, t, J = 9.41Hz, H₃), 5.12 (2H, m, H₂ + H₄), 4.76 (1H, m, H₇), 4.28 (1H, dd, J_{6b-6a} = 12.58Hz, J_{6b-5} = 4.52Hz, H_{6b}), 4.09 (1H, dd, J_{6a-6b} = 12.30Hz, J_{6a-5} = 1.14Hz, H_{6a}), 4.00 (1H, d, J₈₋₇ = 4.85Hz, H₈), 3.87 (1H, dd, J₅₋₄ = 10.09Hz, J₅₋₆ = 2.33Hz, H₅), 1.98 – 2.10 (12H, m, H₁₂), 1.54 (3H, d, J₁₄₋₇ = 6.24Hz, H₁₄).

13C NMR (CDCl₃) δ_{ppm}; 169.3 – 170.5 (4xC₁₃), 168.8 (C₁₁), 157.7 (C₁₀), 92.8 (C₁), 75.0 (C₇), 72.9 (C₅), 72.2 (C₃), 69.9 (C₂), 67.6 (C₄), 61.3 (C₆), 59.9 (C₈), 20.5 – 21.0 (4xC₁₂, C₁₄).

HRMS (ES+) m/z = 476.1396 [M+H]+; Expected 476.1399.



((2,3,4,6-tetra-*O*-acetyl)-β-D-galactopyranosyl)-L-serine-*N*-carboxyanhydride Ser(GalAc)NCA – 3(c)

1**H NMR** (CDCl₃) δ_{ppm}; 6.71 (1H, s, H₉), 6.04 (1H, d, J_{1,2} = 6Hz, H₁), 5.45 (1H, dd, J₃-2 2.00Hz, J₃₋₄ = 3.50Hz, H₃), 5.07 (1H, dd, J₂₋₃ = 3.50Hz, J₁₋₂ = 4.50Hz, H₂), 4.35 - 4.48 (4H, m, H₄, H_{6a}, H_{6b} H₈), 4.00 (1H, ddd, J_{5-6a} = 3.00Hz, J_{5-6b} = 3.60Hz, J₅₋₄ = 7.06Hz, H₅), 2.02 - 2.14 (H₁₃).

13C NMR (CDCl₃) δ_{ppm} ; 167.2-170.1 (4xCl₃), 164.7 (Cl₁), 155.8 (Cl₀), 99.1 (Cl₁), 72.6 (Cs), 72.2 (C4), 71.0 (C3), 69.2 (C2), 60.4 (C8), 59.3 (C6), 59.0 (C7), 20.5 - 21.2 (4xCl₃). HRMS (ES+) m/z = 484.1061 [M+Na]+; expected 484.1062



 $3-O-((2,3,4,6-tetra-O-acetyl)-\beta-D-glucopyranosyl)-L-serine-N-carboxyanhydride - Ser(GluAc)NCA - 4(c)$

1**H NMR** (CDCl₃) δ_{ppm} ; 6.07 (1H, s, H₉), 5.75 (1H, d, J₁₋₂ 8.00Hz, H₁), 5.25 (1H, td, J₂₋₃ = 2.00Hz, J₂₋₁ = 7.7Hz, H₂), 5.11 (1H, dd, J₄₋₅ = 7.50Hz, J₄₋₃ = 3.00Hz, H₄), 4.77 (1H, m, H₅), 4.21 (1H, t, J₃₋₄ = 4.50Hz, H₃), 4.06-4.19 (4H, m, H_{6a}, H_{6b}, H_{7ba}, H₈), 3.70 (1H, dd, J_{7a-7b} = 3.50Hz, J_{7a-8} = 4.00Hz, H_{7a}), 1.99 - 2.19 (12H, m, H₁₃).

13C NMR (CDCl₃) δ_{ppm}; 168.7 -171.0 (4xC₁₃), 165.3(C₁₁), 154.1(C₁₀), 98.6 (C₁), 72.2 (C₅), 71.0 (C₃), 69.8 (C₂), 66.2, 67.4 (C₄), 60.1 (C₈), 59.1 (C₆), 58.9 (C₇), 20.7 - 21.8 (4xC₁₂).

HRMS (ES+) m/z = 484.1066 [M+Na]+; expected 484.1062.



3-*O*-((2,3,4,6-tetra-*O*-acetyl)-β-D-galactopyranosyl-(1-4)-2,3,4,6-tetra-*O*-acetyl-β-D-glucopyranose)-L-serine–*N*-carboxyanhydride - Ser(LacAc)NCA – 5(c) 1H NMR (CDCl₃) δ_{ppm}: 7.67 (1H, s, H₉), 5.82 (1H, d, J₁-2 = 7.50Hz, H₁), 5.76 (1H, d, J₁'-2' = 9.00Hz, H₁'), 5.44 (1H, t, J₄-5 = 9.68Hz, H₄'), 5.22 – 5.37 (3H, m H₃, H₃', H₄), 5.07-5.15 (2H, m, H₂', H₂), 4.24 - 4.31 (1H, m, H₆b), 4.01-4.17 (4H, m, H₆'a, H₆'b, H_{76a}, H₅'), 3.70-3.82 (2H, m, H₄, H₅), 1.97-2.15 (21H, m, H₁₂). **MS** (ES+) m/z =: 749.7 [M]+.



3-S-((2,3,4,6-tetra-O-acetyl)-β-D-glucopyranosyl)-L-cysteine-N-carboxyanhydride - Cys(GluAc) NCA – 6(c)

1**H NMR** (CDCl₃) δ_{ppm} : 6.32 (1H, d, J_{1,2} = 6Hz, H₉), 6.05 (1H, d, J₁₋₂ = 6.2Hz), 5.38 (1H, d, J₂₋₃ = 3.5Hz, H₂), 5.29 (1H, t, J₃₋₄ = 9.0Hz, H₃), 5.06 (4H, m, H_{6a}, H_{6b}, H_{7b}, H₈), 4.90 (1H, dd, J_{7a-7b} = 3.5Hz, J₇₋₈ = 6.5Hz, H_{7a}), 4.79 (1H, dd, J₄₋₅ = 8.00Hz, J₃₋₄ = 2.0Hz, H₄), 3.93 (1H, dd, J₅₋₆ = 4.00Hz, J₄₋₅ = 9.5Hz, H₅), 2.04 - 2.11 (12H, m, H₁₂).

13**C NMR** (CDCl₃) δ_{ppm}: 169.4 -171.5 (4 x C₁₃), 163.9 (C₁₁), 153.6 (C₁₀), 94.1(C₁), 70.1 (C₅), 69.9 (C₃), 68.7 (C₂), 67.7 (C₄), 67.3 (C₈), 62.2 (C₆), 61.8 (C₇), 20.8 - 21.2 (4 x C₁₂).

MS (ES+) $m/z = 477.2 [M]_+$.



 $3-S-((2,3,4,6-tetra-O-acetyl)-\beta-D-galactopyranosyl)-L-cysteine-N-carboxyanhydride - Cys(GalAc)NCA - 7Cc)$

1**H NMR** (CDCl₃) δ_{ppm} : 6.92 (1H, s, H₉), 6.02 (1H, d, J₁₋₂ = 6Hz, H₁), 5.41 (1H, d, J₂₋₁ = 3.5Hz, H₂), 5.33 (1H, dd, J_{6b-6a} =1Hz, J_{6b-5} = 2.5Hz, H_{6b}), 5.29 (1H, t, J₃₋₄ = 9.0Hz, H₃), 5.05 (1H, dd, J_{6a-6b} = 3.5Hz, J_{6a-5} = 4.5Hz, H_{6a}), 4.44 (1H, dd, J_{7a-7b} = 2.5Hz, J₇₋₈ = 7.5Hz, H_{7a}), 4.17-4.29 (2H, m, H₄ + H₅), 2.03 - 2.14 (12H, m, H₁₂).

13C NMR (CDCl₃) δ_{ppm}: 169.0 -171.1 (4xC₁₃), 164.1 (C₁₁), 155.0 (C₁₀), 91.7 (C₁), 70.1 (C₅), 69.6 (C₃), 69.3 (C₂), 68.9 (C₄), 66.7 (C₈), 61.5 (C₆), 60.2 (C₇), 21.0 - 21.4 (4xC₁₂). **MS** (ES+) m/z = 477.4 (100%) [M]+.

Elemental (Found %) C, 45.37; H, 4.66; N, 2.11 (Expected %) C, 45.28 H, 4.86 N, 2.53.



NMR SPECTRA

In this section 1H and 13C spectra for all new compounds are shown. (SerGalAc NCA and SerGluAc NCA have been described previously3 and are not included here. The intermediate glycosylated amino acids, 1 (b) \rightarrow 7 (b), were used without purification and were thus not 'analytically' pure and not included here. A single example, Thr(GluAc) - 1 (b) is shown for completeness.

BocThr(GalAc) - 1(a)



BocThr(GluAc) - 2(a)



BocCys(GalAc) - 7 (a)



BocCys(GluAc) – 6(b)



ThrGal(Ac) – 1 (b)









ThrGluAc NCA – 2(c)





SerLacAc NCA – 5(c)





CysGluAc NCA – 6(c)





CysGalAc NCA – 7(c)



• Indicates pinene residues

References

- 1. H. R. Kricheldorf, in *Alpha amino acid-N-CarboxyAnhydrides and Related Materials*, Springer-Verlag, New York, Editon edn., 1987, pp. 59-157.
- 2. D. Poche, M. J. Moore and J. L. Bowles, Syn. Comm., 1999, 29, 843-854.
- Rude E, Westphal O, Hurwitz E, Fuchs S and Sela M, *Immuno Chem.*, 1966,
 3, 137-151; Rude.E and Meyer-Delius.M, *Car.Res*, 1968, **8**, 219-232; Aoi.K, Tsutsumiuchi.K and Okada.M, *Macromolecules*, 1994, **27**, 875-877.