

## **Synthesis and characterization of the 5-methyl-2'-deoxycytidine glycol–dioxoosmium–bipyridine ternary complex in DNA**

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### **Electronic Supplementary Information**

#### **Contents**

1. Experimental information-----	S2 – S3
2. RP HPLC profiles of thymine glycol–dioxoosmium(VI)–2,2'-bipyridine ternary complexes-----	S4
3. NOESY profile of (5 <i>R</i> ,6 <i>S</i> )-5-methyl-2'-deoxycytidine glycol–dioxoosmium(VI)–2,2'-bipyridine ternary complex-----	S5
4. NOESY profile of (5 <i>S</i> ,6 <i>R</i> )-5-methyl-2'-deoxycytidine glycol–dioxoosmium(VI)–2,2'-bipyridine ternary complex-----	S6

## Experimental information

### Preparation of DNA-Os-Bpy complexes and its digestion procedure

Oligonucleotide (5  $\mu$ M) was treated with 2,2'-bipyridine (100 mM), potassium osmate (5 mM) and potassium hexacyanoferrate(III) (100 mM) in a buffer solution [100 mM Tris-HCl buffer (pH 7.7)–acetonitrile, 9:1(v/v)] for 1 h at 50 °C. The excess reactants were removed with Micro Bio-Spin®. The eluate was fully digested with calf intestine alkaline phosphatase (50 U/mL), snake venom phosphodiesterase (0.15 U/mL), and P1 nuclease (50 U/mL) at 37 °C for 3 h. The digested product was analyzed by RP-HPLC on tandem 5-ODS-H columns (10  $\times$  150 mm  $\times$  2) with an eluent [0.1 M triethylammonium acetate (TEAA, pH 7.0)–acetonitrile, 93:7 (v/v)] at a flow rate 3.0 mL/min. This condition was used for all RP-HPLC experiments.

**Synthesis of M-Os-Bpy.** An aqueous solution of 2'-deoxy-5-methylcytidine (100  $\mu$ L, 1 mM), a solution of 2,2'-bipyridine in acetonitrile (100  $\mu$ L, 1 M), an aqueous solution of potassium osmate (200  $\mu$ L, 25 mM), a freshly prepared aqueous solution of potassium hexacyanoferrate(III) (100  $\mu$ L, 1 M), and 1 M Tris-HCl buffer (100  $\mu$ L, pH 7.7) were mixed and measured up to 1 mL with water. After 1 h at 50 °C, the reaction mixture was analyzed by RP-HPLC. Two diastereomers were purified using RP-HPLC, and then the elution buffer was removed by repeating lyophilization.

**(5R,6S)-M-Os-Bpy (major isomer).**  $^1\text{H}$  NMR (400 MHz,  $\text{D}_2\text{O}$ )  $\delta_{\text{H}}$  9.18 (1H, d,  $J$  = 4.5, H–C(3-bpy)), 8.95 (1H, d,  $J$  = 4.4 H–C(3'-bpy)), 8.52 (2H, m, H–C(6,6'-bpy)), 8.36–8.30 (2H, m, H–C(4,4'-bpy)), 7.87–7.77 (2H, m, H–C(5,5'-bpy)), 6.32 (1H, dd,  $J$  = 8.8, 5.9, H–C(1')) 5.37 (1H, s, H–C(6)), 4.28–4.25 (1H, m, H–C(3')), 3.82–3.79 (1H, m, H–C(4')), 3.58–3.49 (2H, m, H–C(5')), 2.50–2.42, 2.22–2.16 (2H, m, H–C(2',2'')), 1.62 (3H, s,  $\text{H}_3\text{C}(5)$ ); FAB-HRMS ( $m/z$ ): found 654.1236 ( $[\text{M} + \text{H}]^+$ ,  $\text{C}_{20}\text{H}_{24}\text{N}_5\text{O}_8\text{Os}$  calc. 654.1240).

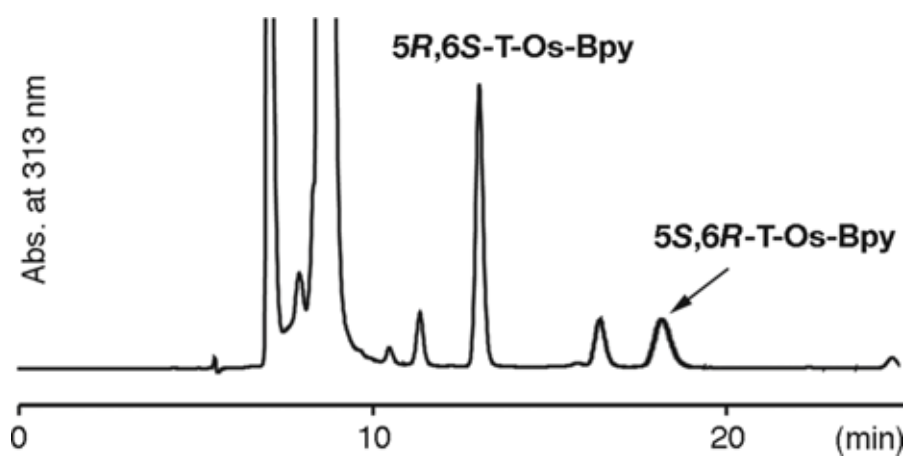
**(5S,6R)-M-Os-Bpy (minor isomer).**  $^1\text{H}$  NMR (400 MHz,  $\text{D}_2\text{O}$ )  $\delta_{\text{H}}$  9.19 (1H, d  $J$  = 4.4, H–C(3-bpy)), 9.10 (1H, d  $J$  = 4.4, H–C(3'-bpy)), 8.58 (2H, m, H–C(6,6'-bpy)), 8.41–8.34 (2H, m, H–C(4,4'-bpy)), 7.88–7.80 (2H, m, H–C(5,5'-bpy)), 6.22 (1H, q,  $J$  = 6.9, H–C(1')), 5.27 (1H, s, H–C(6)), 4.57–4.31 (1H, m, H–C(3')), 3.87–3.86 (1H, m, H–C(4')), 3.69–3.65, 3.57–3.53 (2H, m,  $\text{H}_2\text{C}(5')$ ), 2.37–2.21 (2H, m, H–C(2',2'')),

1.65(3H, s, H<sub>3</sub>C(5)); FAB-HRMS (*m/z*): found 654.1235 ([M + H]<sup>+</sup>, C<sub>20</sub>H<sub>24</sub>N<sub>5</sub>O<sub>8</sub>Os calc. 654.1240).

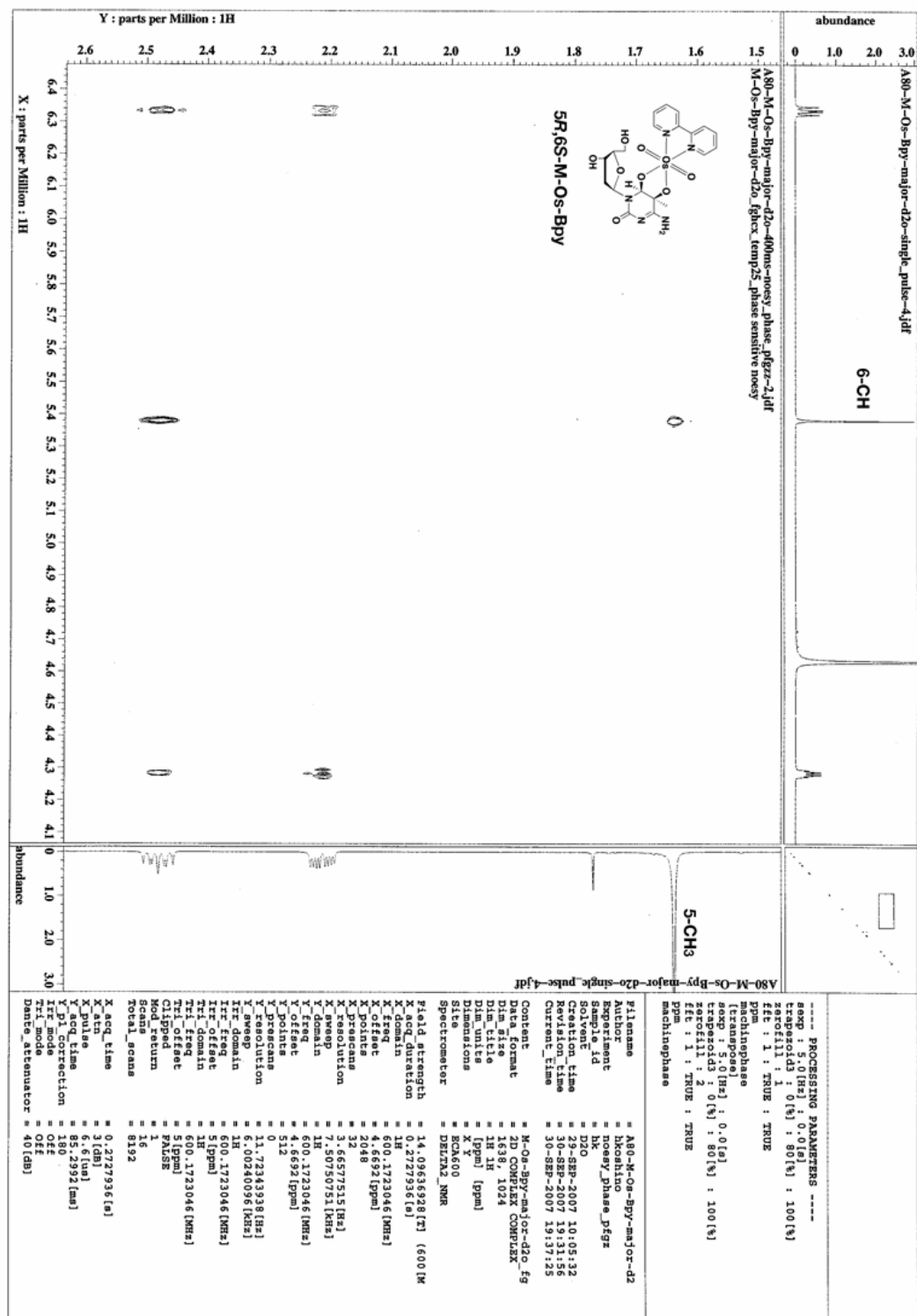
**(5R,6S)-T-Os-Bpy (major isomer).** <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ<sub>H</sub> 10.23 (1H, br s, H–N(3)), 9.16 (1H, d, *J* = 4.4, H–C(3-bpy)), 9.05 (1H, d, *J* = 4.4 H–C(3'-bpy)), 8.96–8.93 (2H, m, H–C(6,6'-bpy)), 8.57–8.51 (2H, m, H–C(4,4'-bpy)), 8.04–8.02 (1H, m, H–C(5-bpy)), 7.98–7.96 (1H, m, H–C(5'-bpy)), 6.23 (1H, dd, *J* = 8.1, 5.9, H–C(1')), 5.25 (1H, s, H–C(6)), 5.14 (1H, br d, *J* = 3.7, H–O(3')), 4.72–4.70 (1H, m, H–O(5')), 4.16–4.15 (1H, m, H–C(3')), 3.67–3.64 (1H, m, H–C(4')), 3.41–3.35 (2H, m, H<sub>2</sub>C(5')), 2.45–2.40, 2.09–2.05 (2H, m, H<sub>2</sub>C(2',2'')), 1.53 (3H, s, H<sub>3</sub>C(5)); FAB-HRMS (*m/z*): found 655.1078 ([M + H]<sup>+</sup>, C<sub>20</sub>H<sub>23</sub>N<sub>4</sub>O<sub>9</sub>Os calc. 655.1080).

**(5S,6R)-T-Os-Bpy (minor isomer).** <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ<sub>H</sub> 10.25 (1H, br s, H–N(3)), 9.17 (1H, d *J* = 5.0, H–C(3-bpy)), 9.12 (1H, d, *J* = 5.0, H–C(3'-bpy)), 8.96–8.93 (2H, m, H–C(6,6'-bpy)), 8.57–8.51 (2H, m, (4,4'-bpy)), 8.05–8.03 (1H, m, H–C(5-bpy)), 7.98–7.96 (1H, m, H–C(5'-bpy)), 6.09 (1H, q, *J* = 6.6, H–C(1')), 5.22 (1H, s, H–C(6)), 5.15 (1H, d, *J* = 4.6, H–O(3')), 4.62–4.60 (1H, m, H–O(5')), 4.21–4.18 (1H, m, H–C(3')), 3.73–3.71 (1H, m, H–C(4')), 3.54–3.47 (2H, m, H–C(5')), 2.33–2.29, 2.17–2.12 (2H, m, H–C(2',2'')), 1.56 (3H, s, H<sub>3</sub>C(5)); FAB-HRMS (*m/z*): found 655.1084 ([M + H]<sup>+</sup>, C<sub>20</sub>H<sub>23</sub>N<sub>4</sub>O<sub>9</sub>Os calc. 655.1080).

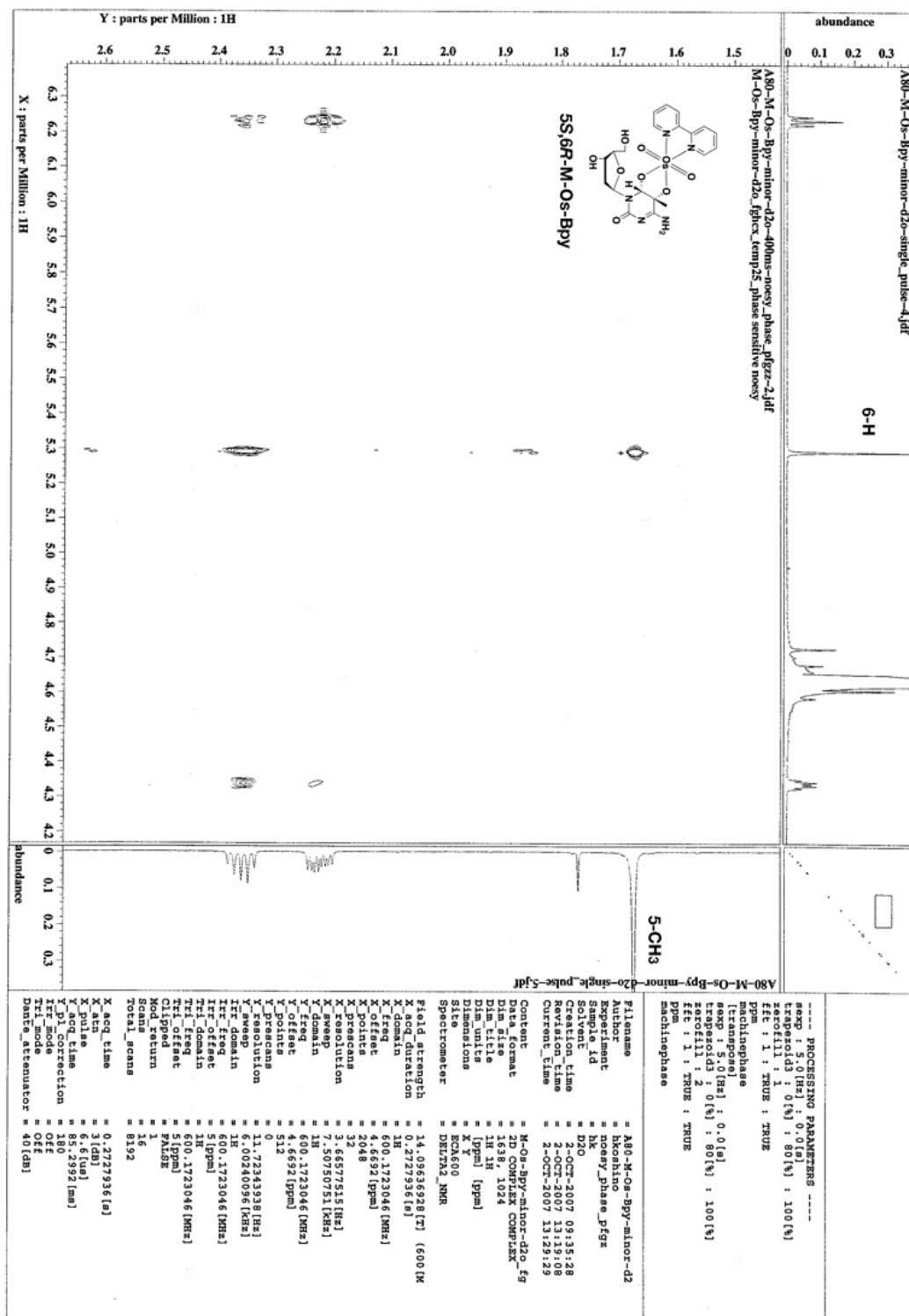
**Monitoring the deamination of 5-methylcytosine glycol-dioxoosmium-2,2'-bipyridine complexes.** A 1 mM solution of M-Os-Bpy in water was incubated at 50 °C. The progress of deamination was monitored using RP-HPLC analysis.



**Fig. S1** HPLC analysis of the crude mixture of chemically synthesized T-Os-Bpy.



**Fig. S2** NOESY profile of (5R,6S)-5-methyl-2'-deoxycytidine glycol-dioxo-osmium(VI)-2,2'-bipyridine ternary complex.



**Fig. S3** NOESY profile of (5*S*,6*R*)-5-methyl-2'-deoxycytidine glycol-dioxo-osmium(VI)-2,2'-bipyridine ternary complex.