

# SUPPORTING INFORMATION

## Lipophilic sugar nucleotide synthesis by structure-based design of nucleotidyltransferase substrates

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## General Methods

Dichloromethane and THF were dried over alumina (Innovative Technology) and stored over 3 Å molecular sieves. All other reagents and solvents were purchased and used without further purification. All reactions were monitored by thin layer chromatography (TLC) using Silicycle precoated silica gel plates (250 µm thickness). The TLC plates were visualized with a potassium permanganate solution (3 g potassium permanganate, 20 g potassium carbonate, 5 mL 5% aqueous sodium hydroxide, 300 mL distilled water), a phenol solution (10 g phenol, 5 mL H<sub>2</sub>SO<sub>4</sub>, 95 mL ethanol), or ultraviolet light ( $\lambda = 254$  nm). Unless otherwise specified, flash chromatographic purification was performed on a Biotage SP1 HPFC. <sup>1</sup>H, <sup>13</sup>C and <sup>31</sup>P NMR spectra were obtained on a Bruker AVANCE-500 NMR Spectrometer operating at frequencies of 500.13 MHz, 125.76 MHz, and 202.45 MHz, respectively. Assignments are based on COSY and HSQC 2D NMR experiments. For chloroform-*d*, <sup>1</sup>H NMR chemical shifts are reported as  $\delta$  in units of parts per million (ppm) downfield from tetramethylsilane ( $\delta$  0.0) and <sup>13</sup>C NMR chemical shifts are reported as  $\delta$  in units of parts per million (ppm) relative to the residual solvent signal of chloroform-*d* ( $\delta$  77.16). For methanol-*d*<sub>4</sub>, <sup>1</sup>H and <sup>13</sup>C NMR chemical shifts are reported as  $\delta$  in units of parts per million (ppm) relative to the residual solvent signal of methanol-*d*<sub>4</sub> ( $\delta$  3.31 and  $\delta$  49.00, respectively). <sup>31</sup>P NMR chemical shifts are reported as  $\delta$  in units of parts per million (ppm) relative to 85% H<sub>3</sub>PO<sub>4</sub>. Enzymatic reactions were monitored by HPLC performed on a

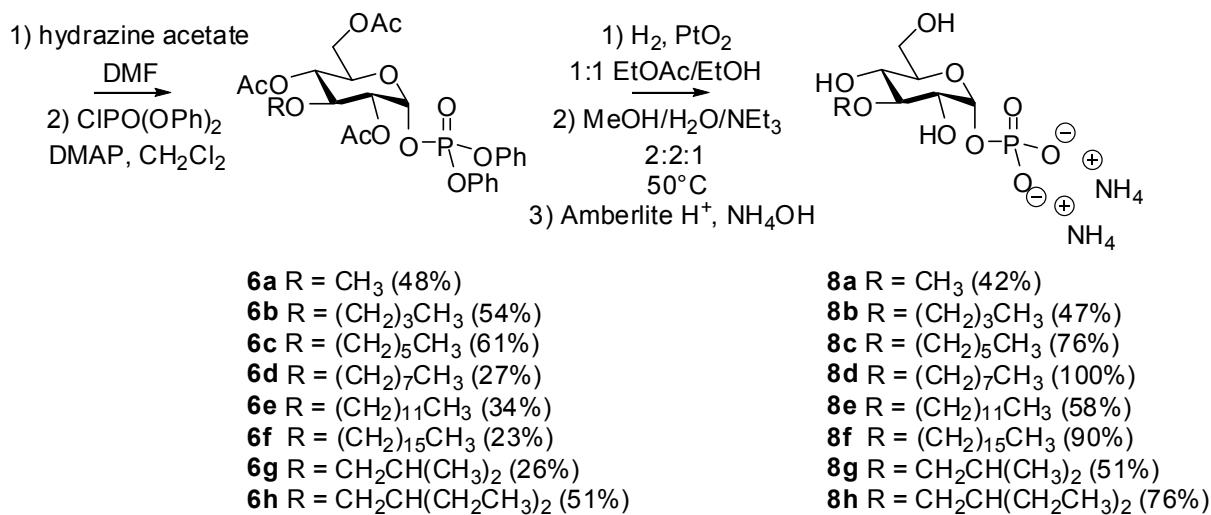
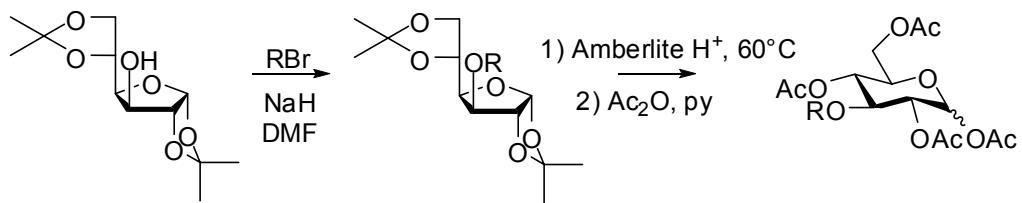
Hewlett Packard Series 1050 instrument with an Agilent Zorbax 5 $\mu$ m Rx-C18 column (150 cm x 4.6 cm).

Compounds with a nucleotide base chromophore were monitored using a UV detector ( $\lambda = 254$  nm). The linear gradient used was 90/10 A/B to 40/60 A/B over 8.0 min, followed by a plateau at 40/60 A/B from 8.0 to 10.0 min at 1.0 mL/min where A is an aqueous buffer containing 12 mM NBu<sub>4</sub>Br, 10 mM KH<sub>2</sub>PO<sub>4</sub>, and 5% HPLC grade CH<sub>3</sub>CN (pH 4.9) and B is HPLC CH<sub>3</sub>CN except for assays containing compound **8f**, whereby the gradient was run to 80% CH<sub>3</sub>CN instead of 60%. High resolution mass spectra were recorded on a micrOTOF instrument (Bruker Dalton) running in negative ion mode (ESI). Low resolution mass spectra were recorded on an LCQDuo ion trap instrument (Thermo Finnigan) running in positive mode (ESI).

An Agilent 1100 LC system was coupled to an Applied Biosystems-MDS SCIEX hybrid triple quadrupole linear ion trap (QTRAP 4000) mass spectrometer equipped with a Turbo V source for electrospray ionization for the ESI-MS/MS experiments. The sample was analyzed by flow-injection analysis in 75:25 (v/v) acetonitrile:de-ionized water using a flow-rate of 200  $\mu$ L/min. Precursor ion scanning of m/z 323 ([UMP – H]-) and m/z 383 ([TDP – H]-) were used initially to selectively detect for uridine- and thymidine-linked sugar nucleotides, respectively, in the sample. ESI-MS/MS analysis in the enhanced product ion (EPI) mode was then performed to confirm the presence of the expected synthetic sugar nucleotide. The mass spectrometer settings for precursor ion scanning were: ionspray voltage 4.5 kV; mass range Q1 m/z 300 – 800; scan time 5 sec, Q1 and Q3 set to unit resolution. For the ESI-MS/MS experiments: ionspray voltage 4.5 kV; Q3 m/z 100–800; scan speed 1000 amu/sec; trap fill time was set to dynamic; Q1 and Q3 set to unit resolution. All acquisitions were made in the negative mode.

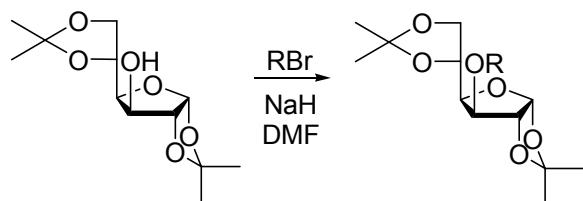
pKa values were measured in the following way: Using an IQ Scientific Instruments IQ150 fitted with an ISFET probe, a 0.01 M solution of the diammonium salt was adjusted to pH 10 with 0.2 M NaOH. Titration was done with 5 uL aliquots of 0.2 M HCl until pH 2, and the pKa values were determined by plotting in GraFit 5.0.4 (Erihacus Software Limited).

## Chemical Synthesis



Overall yields: **8a** (20%), **8b** (14%), **8c** (21%), **8d** (21%), **8e** (3%), **8f** (4%), **8g** (3%), **8h** (6%).

### General Procedure for Alkylation (2b-h)



To a stirring solution of 1,2:5,6-di-*O*-isopropylidene- $\alpha$ -D-glucofuranose (**1**) (1.5 g, 5.76 mmol) in anhydrous DMF (15 mL) was added sodium hydride (0.42 g, 17 mmol) at 0°C. After stirring at 0°C for 10 min, the alkyl bromide (8.64 mmol) was added under a nitrogen atmosphere, and the mixture was stirred for another 30 min at 0°C followed by 30 min at rt. The mixture was diluted with H<sub>2</sub>O (50 mL) and extracted with dichloromethane (2 x 25 mL). The combined organic extracts were washed with brine (10 mL) and dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated. Elution through a silica plug afforded the title compounds (**2b-h**).

### **3-*O*-Butyl-1,2:5,6-di-*O*-isopropylidene- $\alpha$ -D-glucofuranose (**2b**)<sup>1</sup>**

Reaction with 1-bromobutane (0.93 mL, 8.64 mmol) followed by chromatographic purification (hexanes/EtOAc, 95:5) furnished compound **2b** as a colorless liquid (1.12 g, 62 %); R<sub>F</sub> = 0.48 (hexanes/EtOAc, 85:15); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 5.87 (d, 1H, J<sub>1,2</sub> = 3.8 Hz, H-1), 4.52 (d, 1H, J<sub>1,2</sub> = 3.7 Hz, H-2), 4.31 (dd, 1H, J<sub>5,6</sub> = 13.5 Hz, J<sub>4,5</sub> = 6.2 Hz, H-5), 4.13 (dd, 1H, J<sub>4,5</sub> = 7.4 Hz, J<sub>3,4</sub> = 3.0 Hz, H-4), 4.08 (ddd, 1H, J<sub>5,6a</sub> = 8.5 Hz, <sup>2</sup>J<sub>6a,6b</sub> = 6.2 Hz, <sup>4</sup>J<sub>4,6a</sub> = 0.7 Hz, H-6a), 3.98 (ddd, 1H, J<sub>5,6b</sub> = 8.5 Hz, <sup>2</sup>J<sub>6a,6b</sub> = 6.0 Hz, <sup>4</sup>J<sub>4,6b</sub> = 0.6 Hz, H-6b), 3.85 (d, 1H, J<sub>3,4</sub> = 3.0 Hz, H-3), 3.56 (m, 2H, OCH<sub>2</sub>), 1.54 (m, 2H, OCH<sub>2</sub>CH<sub>2</sub>), 1.50, 1.42, 1.35, 1.32 (s, 12H, CH<sub>3</sub> x 4), 1.38 (m, 2H, OCH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 0.91 (t, 3H, J = 7.5 Hz, (CH<sub>2</sub>)<sub>3</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 111.8, 109.0 (C(CH<sub>3</sub>)<sub>2</sub> x 2) 105.4 (C-1), 82.7 (C-2), 82.3 (C-3), 81.4 (C-4), 72.7 (C-5), 70.5 (OCH<sub>2</sub>), 67.4 (C-6), 31.9 (OCH<sub>2</sub>CH<sub>2</sub>), 27.0, 26.9, 26.4, 25.5 (C(CH<sub>3</sub>)<sub>2</sub> x 2), 19.4 (OCH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 14.0 ((CH<sub>2</sub>)<sub>3</sub>CH<sub>3</sub>); LRMS *m/z* calcd for C<sub>16</sub>H<sub>28</sub>O<sub>6</sub> [M+Na]<sup>+</sup>: 339.2. Found 339.2.

### **3-*O*-Hexyl-1,2:5,6-di-*O*-isopropylidene- $\alpha$ -D-glucofuranose (**2c**)<sup>1</sup>**

Reaction with 1-bromohexane (1.21 mL, 8.64 mmol) followed by chromatographic purification (hexanes/EtOAc, 91:9) furnished compound **2c** as a colorless liquid (1.35 g, 68 %); R<sub>F</sub> = 0.22 (hexanes/EtOAc, 95:5); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 5.87 (d, 1H, J<sub>1,2</sub> = 3.8 Hz, H-1), 4.52 (d, 1H, J<sub>1,2</sub> = 3.6 Hz, H-2), 4.31 (dd, 1H, J<sub>5,6</sub> = 13.5 Hz, J<sub>4,5</sub> = 6.1 Hz, H-5), 4.13 (dd, 1H, J<sub>4,5</sub> = 7.4 Hz, J<sub>3,4</sub> = 3.1 Hz, H-4), 4.08 (ddd, 1H, J<sub>5,6a</sub> = 8.4 Hz, <sup>2</sup>J<sub>6a,6b</sub>

<sup>1</sup> Ikekawa, T.; Irinoda, K.; Saze, K.; Katori, T.; Matsuda, H.; Ohkawa, M.; Kosik, M. *Chem. Pharm. Bull.* **1987**, *35*, 2894-2899.

= 6.2 Hz,  $^4J_{4,6a}$  = 0.7 Hz, H-6a), 3.98 (dd, 1H,  $J_{5,6b}$  = 8.7 Hz,  $^2J_{6a,6b}$  = 6.0 Hz, H-6b), 3.85 (d, 1H,  $J_{3,4}$  = 3.1 Hz, H-3), 3.55 (m, 2H, OCH<sub>2</sub>), 1.55 (m, 2H, OCH<sub>2</sub>CH<sub>2</sub>), 1.50, 1.42, 1.35, 1.32 (s, 12H, CH<sub>3</sub> x 4), 1.38-1.24 (m, 6H, OCH<sub>2</sub>CH<sub>2</sub>(CH<sub>2</sub>)<sub>3</sub>CH<sub>3</sub>), 0.89 (t, 3H,  $J$  = 6.7 Hz, (CH<sub>2</sub>)<sub>5</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 111.8, 109.0 (C(CH<sub>3</sub>)<sub>2</sub> x 2) 105.4 (C-1), 82.7 (C-2), 82.3 (C-3), 81.4 (C-4), 72.7 (C-5), 70.9 (OCH<sub>2</sub>), 67.4 (C-6), 31.7 (OCH<sub>2</sub>CH<sub>2</sub>), 29.8 (OCH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 27.0, 26.9, 26.4, 25.5 (C(CH<sub>3</sub>)<sub>2</sub> x 2), 25.9 (O(CH<sub>2</sub>)<sub>3</sub>CH<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 22.7 (O(CH<sub>2</sub>)<sub>4</sub>CH<sub>2</sub>CH<sub>3</sub>), 14.2 ((CH<sub>2</sub>)<sub>5</sub>CH<sub>3</sub>); LRMS *m/z* calcd for C<sub>18</sub>H<sub>32</sub>O<sub>6</sub> [M+Na]<sup>+</sup>: 367.2. Found 367.2.

### **3-*O*-Octyl-1,2:5,6-di-*O*-isopropylidene- $\alpha$ -D-glucofuranose (2d)<sup>1</sup>**

Reaction with 1-bromooctane (1.49 mL, 8.64 mmol) followed by chromatographic purification (hexanes/EtOAc, 93:7) furnished compound **2d** as a colorless liquid (1.64 g, 76 %); R<sub>F</sub> = 0.38 (hexanes/EtOAc, 90:10); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 5.87 (d, 1H,  $J_{1,2}$  = 3.7 Hz, H-1), 4.52 (d, 1H,  $J_{1,2}$  = 3.7 Hz, H-2), 4.31 (dd, 1H,  $J_{5,6}$  = 13.5 Hz,  $J_{4,5}$  = 6.1 Hz, H-5), 4.13 (dd, 1H,  $J_{4,5}$  = 7.5 Hz,  $J_{3,4}$  = 3.1 Hz, H-4), 4.08 (dd, 1H,  $J_{5,6a}$  = 8.5 Hz,  $^2J_{6a,6b}$  = 6.3 Hz, H-6a), 3.98 (dd, 1H,  $J_{5,6b}$  = 8.5 Hz,  $^2J_{6a,6b}$  = 6.0 Hz, H-6b), 3.85 (d, 1H,  $J_{3,4}$  = 3.2 Hz, H-3), 3.55 (m, 2H, OCH<sub>2</sub>), 1.55 (m, 2H, OCH<sub>2</sub>CH<sub>2</sub>), 1.49, 1.42, 1.35, 1.32 (s, 12H, CH<sub>3</sub> x 4), 1.38-1.21 (m, 10H, OCH<sub>2</sub>CH<sub>2</sub>(CH<sub>2</sub>)<sub>5</sub>CH<sub>3</sub>), 0.88 (t, 3H,  $J$  = 7.1 Hz, (CH<sub>2</sub>)<sub>7</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 111.8, 109.0 (C(CH<sub>3</sub>)<sub>2</sub> x 2) 105.4 (C-1), 82.7 (C-2), 82.3 (C-3), 81.4 (C-4), 72.7 (C-5), 70.9 (OCH<sub>2</sub>), 67.4 (C-6), 32.0, 29.5, 29.4, 26.2, 22.8 (OCH<sub>2</sub>CH<sub>2</sub>(CH<sub>2</sub>)<sub>5</sub>CH<sub>3</sub>), 29.9 (OCH<sub>2</sub>CH<sub>2</sub>(CH<sub>2</sub>)<sub>5</sub>CH<sub>3</sub>), 27.0, 26.9, 26.4, 25.5 (C(CH<sub>3</sub>)<sub>2</sub> x 2), 14.2 ((CH<sub>2</sub>)<sub>7</sub>CH<sub>3</sub>); LRMS *m/z* calcd for C<sub>20</sub>H<sub>36</sub>O<sub>6</sub> [M+Na]<sup>+</sup>: 395.2. Found 395.3.

### **3-*O*-Dodecyl-1,2:5,6-di-*O*-isopropylidene- $\alpha$ -D-glucofuranose (2e)<sup>1</sup>**

Reaction with 1-bromododecane (2.07 mL, 8.64 mmol) followed by chromatographic purification (hexanes/EtOAc, 93:7) furnished compound **2e** as a colorless liquid (1.75 g, 71 %); R<sub>F</sub> = 0.19 (hexanes/EtOAc, 95:5); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 5.87 (d, 1H,  $J_{1,2}$  = 3.7 Hz, H-1), 4.52 (d, 1H,  $J_{1,2}$  = 3.7 Hz, H-2), 4.30 (dd, 1H,  $J_{5,6}$  = 13.6 Hz,  $J_{4,5}$  = 6.2 Hz, H-5), 4.12 (dd, 1H,  $J_{4,5}$  = 7.5 Hz,  $J_{3,4}$  = 3.1 Hz, H-4), 4.08 (dd, 1H,  $J_{5,6a}$  = 8.5 Hz,  $^2J_{6a,6b}$  = 6.3 Hz, H-6a), 3.98 (dd, 1H,  $J_{5,6b}$  = 8.5 Hz,  $^2J_{6a,6b}$  = 6.0 Hz, H-6b), 3.85 (d, 1H,  $J_{3,4}$  = 3.1 Hz, H-3), 3.55 (m, 2H, S6

$OCH_2$ ), 1.55 (m, 2H,  $OCH_2CH_2$ ), 1.49, 1.42, 1.35, 1.31 (s, 12H,  $CH_3$  x 4), 1.37-1.21 (m, 18H,  $OCH_2CH_2(CH_2)_9CH_3$ ), 0.88 (t, 3H,  $J = 6.6$  Hz,  $(CH_2)_{11}CH_3$ );  $^{13}C$  NMR ( $CDCl_3$ )  $\delta$  111.8, 109.0 ( $C(CH_3)_2$  x 2) 105.4 (C-1), 82.7 (C-2), 82.3 (C-3), 81.4 (C-4), 72.7 (C-5), 70.9 ( $OCH_2$ ), 67.4 (C-6), 32.0, 29.9, 29.8, 29.8, 29.7, 29.7, 29.5, 29.5, 26.4, 22.8 ( $OCH_2(CH_2)_{10}CH_3$ ), 27.0, 26.9, 26.2, 25.5 ( $C(CH_3)_2$  x 2), 14.2 ( $((CH_2)_{11}CH_3$ ); LRMS  $m/z$  calcd for  $C_{24}H_{44}O_6$  [M+Na] $^+$ : 451.3. Found 451.3.

### **3-O-Hexadecyl-1,2:5,6-di-O-isopropylidene- $\alpha$ -D-glucofuranose (2f)<sup>1</sup>**

Reaction with 1-bromohexadecane (3.52 mL, 11.5 mmol) followed by chromatographic purification (hexanes/EtOAc, 90:10) furnished compound **2f** as a colorless liquid (1.95 g, 70 %);  $R_F = 0.40$  (hexanes/EtOAc, 90:10);  $^1H$  NMR ( $CDCl_3$ )  $\delta$  5.87 (d, 1H,  $J_{1,2} = 3.7$  Hz, H-1), 4.52 (d, 1H,  $J_{1,2} = 3.7$  Hz, H-2), 4.30 (ddd, 1H,  $J_{4,5} = 7.3$  Hz,  $J_{5,6a} = 6.2$  Hz,  $J_{5,6b} = 6.2$  Hz, H-5), 4.13 (dd, 1H,  $J_{4,5} = 7.4$  Hz,  $J_{3,4} = 3.1$  Hz, H-4), 4.08 (dd, 1H,  $J_{5,6a} = 8.5$  Hz,  $J_{6a,6b} = 6.2$  Hz, H-6a), 3.97 (dd, 1H,  $J_{5,6b} = 8.5$  Hz,  $J_{6a,6b} = 6.0$  Hz, H-6b), 3.85 (d, 1H,  $J_{3,4} = 3.1$  Hz, H-3), 3.55 (m, 2H,  $OCH_2$ ), 1.55 (m, 2H,  $OCH_2CH_2$ ), 1.49, 1.42, 1.35, 1.31 (s, 12H,  $CH_3$  x 4), 1.37-1.21 (m, 26H,  $OCH_2CH_2(CH_2)_{13}CH_3$ ), 0.88 (t, 3H,  $J = 7.0$  Hz,  $(CH_2)_{15}CH_3$ );  $^{13}C$  NMR ( $CDCl_3$ )  $\delta$  111.8, 109.0 ( $C(CH_3)_2$  x 2) 105.4 (C-1), 82.7 (C-2), 82.3 (C-3), 81.4 (C-4), 72.7 (C-5), 70.9 ( $OCH_2$ ), 67.4 (C-6), 32.1, 29.9, 29.9, 29.9, 29.8, 29.8, 29.8, 29.8, 29.7, 29.6, 29.5, 26.2, 22.8 ( $OCH_2(CH_2)_{14}CH_3$ ), 27.0, 26.9, 26.4, 25.5 ( $C(CH_3)_2$  x 2), 14.2 ( $((CH_2)_{15}CH_3$ ); LRMS  $m/z$  calcd for  $C_{28}H_{52}O_6$  [M+Na] $^+$ : 507.4. Found 507.4.

### **1,2:5,6-Di-O-Isopropylidene-3-O-(2-methylpropyl)- $\alpha$ -D-glucofuranose (2g)**

Reaction with 1-bromo-2-methylpropane (2.51 mL, 23.0 mmol) followed by chromatographic purification (hexanes/EtOAc, 90:10) furnished compound **2g** as a colorless liquid (0.67 g, 37 %);  $R_F = 0.86$  (hexanes/EtOAc, 66:33);  $^1H$  NMR ( $CDCl_3$ )  $\delta$  5.87 (d, 1H,  $J_{1,2} = 3.5$  Hz, H-1), 4.52 (d, 1H,  $J_{1,2} = 3.8$  Hz, H-2), 4.33 (dd, 1H,  $J_{5,6} = 13.5$  Hz,  $J_{4,5} = 6.2$  Hz, H-5), 4.14 (dd, 1H,  $J_{4,5} = 7.4$  Hz,  $J_{3,4} = 3.1$  Hz, H-4), 4.08 (dd, 1H,  $J_{5,6a} = 8.5$  Hz,  $J_{6a,6b} = 6.2$  Hz, H-6a), 3.97 (dd, 1H,  $J_{5,6b} = 8.5$  Hz,  $J_{6a,6b} = 6.2$  Hz, H-6b), 3.84 (d, 1H,  $J_{3,4} = 3.2$  Hz, H-3), 3.37 (dd, 1H,  $J = 6.5$  Hz,  $^2J = 9.0$  Hz,  $OCH_{2a}$ ), 3.28 (dd, 1H,  $J = 6.5$  Hz,  $^2J = 9.0$  Hz,  $OCH_{2b}$ ), 1.84

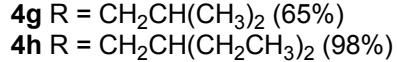
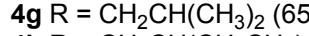
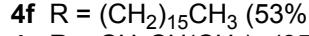
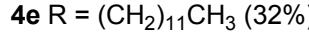
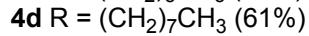
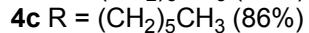
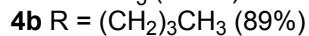
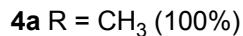
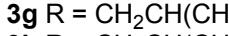
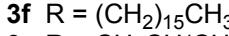
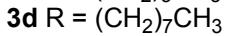
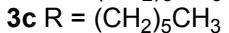
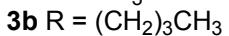
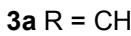
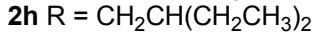
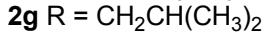
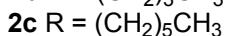
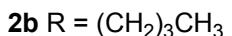
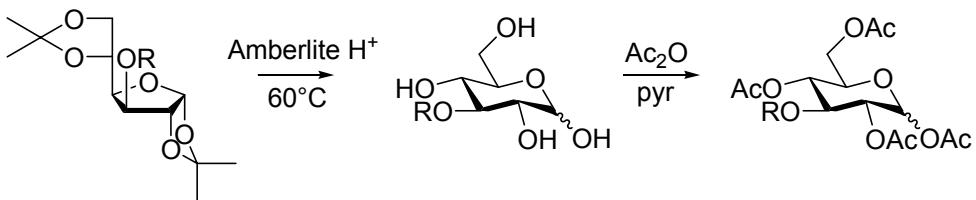
(nonet, 1H,  $J = 6.7$  Hz,  $\text{OCH}_2\text{CH}(\text{CH}_3)_2$ ), 1.50, 1.42, 1.35, 1.32 (s, 12H,  $\text{CH}_3 \times 4$ ), 0.91 (d, 3H,  $J = 3.8$  Hz,  $\text{CH}_{3a}$ ), 0.90 (d, 3H,  $J = 3.8$  Hz,  $\text{CH}_{3b}$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  111.7, 108.9 ( $\text{C}(\text{CH}_3)_2 \times 2$ ) 105.3 (C-1), 82.4 (C-2), 82.3 (C-3), 81.3 (C-4), 77.3 ( $\text{OCH}_2$ ), 72.6 (C-5), 67.3 (C-6), 28.6 ( $\text{OCH}_2\text{CH}$ ), 26.9, 26.8, 26.3, 25.4 ( $\text{C}(\text{CH}_3)_2 \times 2$ ), 19.3, 19.2 ( $\text{OCH}_2\text{CH}(\text{CH}_3)_2$ ); LRMS  $m/z$  calcd for  $\text{C}_{16}\text{H}_{28}\text{O}_6$  [ $\text{M}+\text{Na}]^+$ : 339.2. Found 339.1.

### **1,2:5,6-Di-O-Isopropylidene-3-O-(2-ethylbutyl)- $\alpha$ -D-glucofuranose (2h)**

Reaction with 1-bromo-2-ethylbutane (1.61 mL, 11.5 mmol) followed by chromatographic purification (hexanes/EtOAc, 91:9) furnished compound **2h** as a colorless liquid (0.34 g, 17 %);  $R_F = 0.67$  (hexanes/EtOAc, 80:20);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  5.86 (d, 1H,  $J_{1,2} = 3.7$  Hz, H-1), 4.51 (d, 1H,  $J_{1,2} = 3.7$  Hz, H-2), 4.30 (dd, 1H,  $J_{5,6} = 13.6$  Hz,  $J_{4,5} = 6.2$  Hz, H-5), 4.13 (dd, 1H,  $J_{4,5} = 7.4$  Hz,  $J_{3,4} = 3.1$  Hz, H-4), 4.08 (dd, 1H,  $J_{5,6a} = 8.5$  Hz,  $^2J_{6a,6b} = 6.3$  Hz, H-6a), 3.96 (dd, 1H,  $J_{5,6b} = 8.4$  Hz,  $^2J_{6a,6b} = 6.2$  Hz, H-6b), 3.83 (d, 1H,  $J_{3,4} = 3.1$  Hz, H-3), 3.50 (dd, 1H,  $J = 5.3$  Hz,  $^2J = 9.0$  Hz,  $\text{OCH}_{2a}$ ), 3.40 (dd, 1H,  $J = 5.4$  Hz,  $^2J = 9.0$  Hz,  $\text{OCH}_{2b}$ ), 1.50, 1.42, 1.34, 1.32 (s, 12H,  $\text{CH}_3 \times 4$ ), 1.45-1.28 (m, 5H,  $\text{OCH}_2\text{CH}(\text{CH}_2\text{CH}_3)_2$ ), 0.88 (t, 3H,  $J = 7.4$  Hz,  $\text{CH}_{3a}$ ), 0.90 (t, 3H,  $J = 7.4$  Hz,  $\text{CH}_{3b}$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  111.9, 109.0 ( $\text{C}(\text{CH}_3)_2 \times 2$ ) 105.5 (C-1), 82.5 (C-2), 82.5 (C-3), 81.6 (C-4), 72.8 ( $\text{OCH}_2$ ), 72.7 (C-5), 67.5 (C-6), 41.5 ( $\text{OCH}_2\text{CH}$ ), 27.0, 26.9, 26.4, 25.5 ( $\text{C}(\text{CH}_3)_2 \times 2$ ), 23.5, 23.5 ( $\text{OCH}_2\text{CH}(\text{CH}_2\text{CH}_3)_2$ ), 11.3, 11.2 ( $\text{OCH}_2\text{CH}(\text{CH}_2\text{CH}_3)_2$ ); LRMS  $m/z$  calcd for  $\text{C}_{18}\text{H}_{32}\text{O}_6$  [ $\text{M}+\text{Na}]^+$ : 367.2. Found 367.2.

Found 367.2.

## General Procedure for Hydrolysis and Acetylation (3a-h and 4a-h, respectively)



To a solution of 3-*O*-alkyl-1,2:5,6-di-*O*-isopropylidene- $\alpha$ -D-glucofuranose (**2b-h**) (1.5 g) in 5:1 MeOH/H<sub>2</sub>O (30 mL) was added Amberlite IR-120 PLUS(H) ion exchange resin (45 g) and the reaction was stirred overnight at 60°C. The reaction mixture was then filtered and concentrated to dryness. The hydrolyzed intermediate (**3b-h**) was immediately reacted without further purification. Under a nitrogen atmosphere, a solution of the 3-*O*-alkyl-D-glucopyranose (**3a-h**) in anhydrous pyridine (20 mL) was charged with acetic anhydride (3.0 mL) and stirred overnight at rt. This mixture was then diluted with H<sub>2</sub>O (50 mL) and extracted with dichloromethane (2 x 25 mL). The combined organic extracts were washed with brine (10 mL) and dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated to afford the acetylated product (**4a-h**).

### 1,2,4,6-Tetra-*O*-acetyl-3-*O*-methyl- $\alpha$ -D-glucopyranose (**4a**)

Compound **4a** was obtained as a colorless liquid (2.80 g, 100 %); R<sub>F</sub> = 0.46 (hexanes/EtOAc, 67:33); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 6.29 (d, 1H, J<sub>1,2</sub> = 3.7 Hz, H-1), 5.08 (dd, 1H, J<sub>4,5</sub> = 10.2 Hz, J<sub>3,4</sub> = 9.6 Hz, H-4), 5.00 (dd, 1H, J<sub>2,3</sub> = 9.9 Hz, J<sub>1,2</sub> = 3.6 Hz, H-2), 4.20 (dd, 1H, <sup>2</sup>J<sub>6a,6b</sub> = 12.5 Hz, J<sub>5,6a</sub> = 4.5 Hz, H-6a), 4.07 (dd, 1H, <sup>2</sup>J<sub>6a,6b</sub> = 12.3 Hz, J<sub>5,6b</sub> = 2.3 Hz, H-6b), 4.01 (ddd, 1H, J<sub>4,5</sub> = 10.2 Hz, J<sub>5,6a</sub> = 4.2 Hz, J<sub>5,6b</sub> = 2.2 Hz, H-5), 3.72 (dd, 1H, J<sub>2,3</sub> = 9.8 Hz, J<sub>3,4</sub> = 9.8 Hz, H-3), 3.47 (s, 3H, CH<sub>3</sub>), 2.17, 2.11, 2.09, 2.07 (s, 12H, C(O)CH<sub>3</sub> x 4); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 170.8, 169.7, 169.4, 168.9 (C(O)CH<sub>3</sub> x 4), 89.6 (C-1), 78.3 (C-3), 71.2 (C-2), 70.4 (C-5), 69.1 (C-4), 61.9 (C-6),

60.2 ( $\text{CH}_3$ ), 21.0, 20.9, 20.8, 20.8 ( $\text{C}(\text{O})\text{CH}_3 \times 4$ ); LRMS  $m/z$  calcd for  $\text{C}_{15}\text{H}_{22}\text{O}_{10}$  [ $\text{M}+\text{Na}]^+$ : 385.1. Found 385.1.

### **1,2,4,6-Tetra-*O*-acetyl-3-*O*-butyl- $\beta$ -D-glucopyranose (4b)**

Compound **4b** was obtained as a colorless liquid (1.74 g, 89 %);  $R_F = 0.46$  (hexanes/EtOAc, 67:33);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  5.64 (d, 1H,  $J_{1,2} = 8.2$  Hz, H-1), 5.09 (dd, 1H,  $J_{2,3} = 9.4$  Hz,  $J_{1,2} = 8.3$  Hz, H-2), 5.09 (dd, 1H,  $J_{4,5} = 9.5$  Hz,  $J_{3,4} = 9.4$  Hz, H-4), 4.23 (dd, 1H,  $J_{6a,6b} = 12.5$  Hz,  $J_{5,6a} = 4.9$  Hz, H-6a), 4.09 (dd, 1H,  $J_{6a,6b} = 12.5$  Hz,  $J_{5,6b} = 2.3$  Hz, H-6b), 3.72 (ddd, 1H,  $J_{4,5} = 9.8$  Hz,  $J_{5,6a} = 4.7$  Hz,  $J_{5,6b} = 2.2$  Hz, H-5), 3.56 (dd, 1H,  $J_{2,3} = 9.5$  Hz,  $J_{3,4} = 9.5$  Hz, H-3), 3.54 (m, 2H,  $\text{OCH}_2$ ), 2.11, 2.09, 2.08, 2.08 (s, 12H,  $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 1.46 (m, 2H,  $\text{OCH}_2\text{CH}_2$ ), 1.30 (m, 2H,  $\text{OCH}_2\text{CH}_2\text{CH}_2$ ), 0.88 (t, 3H,  $J = 7.3$  Hz,  $(\text{CH}_2)_3\text{CH}_3$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  170.9, 169.4, 169.4, 169.2 ( $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 92.2 (C-1), 80.4 (C-3), 73.2 (C-5), 72.4 ( $\text{OCH}_2$ ), 71.7 (C-2), 69.2 (C-4), 62.0 (C-6), 32.3 ( $\text{OCH}_2\text{CH}_2$ ), 21.0, 20.9, 20.9, 20.9 ( $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 19.1 ( $\text{OCH}_2\text{CH}_2\text{CH}_2$ ), 14.0 ( $\text{O}(\text{CH}_2)_3\text{CH}_3$ ); LRMS  $m/z$  calcd for  $\text{C}_{18}\text{H}_{28}\text{O}_{10}$  [ $\text{M}+\text{Na}]^+$ : 427.2. Found 427.2.

### **1,2,4,6-Tetra-*O*-acetyl-3-*O*-hexyl- $\beta$ -D-glucopyranose (4c)**

Compound **4c** was obtained as a white solid (1.62 g, 86 %);  $R_F = 0.26$  (hexanes/EtOAc, 80:20);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  5.64 (d, 1H,  $J_{1,2} = 8.2$  Hz, H-1), 5.10 (dd, 1H,  $J_{2,3} = 9.4$  Hz,  $J_{1,2} = 8.3$  Hz, H-2), 5.08 (dd, 1H,  $J_{4,5} = 9.5$  Hz,  $J_{3,4} = 9.6$  Hz, H-4), 4.23 (dd, 1H,  $J_{6a,6b} = 12.5$  Hz,  $J_{5,6a} = 4.9$  Hz, H-6a), 4.09 (dd, 1H,  $J_{6a,6b} = 12.5$  Hz,  $J_{5,6b} = 2.3$  Hz, H-6b), 3.72 (ddd, 1H,  $J_{4,5} = 9.9$  Hz,  $J_{5,6a} = 4.9$  Hz,  $J_{5,6b} = 2.4$  Hz, H-5), 3.57 (dd, 1H,  $J_{2,3} = 9.4$  Hz,  $J_{3,4} = 9.4$  Hz, H-3), 3.53 (m, 2H,  $\text{OCH}_2$ ), 2.11, 2.09, 2.08, 2.08 (s, 12H,  $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 1.46 (p, 2H,  $J = 7.5$  Hz,  $\text{OCH}_2\text{CH}_2$ ), 1.26 (m, 6H,  $\text{OCH}_2\text{CH}_2(\text{CH}_2)_3\text{CH}_3$ ), 0.88 (t, 3H,  $J = 6.9$  Hz,  $\text{O}(\text{CH}_2)_5\text{CH}_3$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  170.9, 169.4, 169.3, 169.2 ( $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 92.2 (C-1), 80.4 (C-3), 73.2 (C-5), 72.8 ( $\text{OCH}_2$ ), 71.6 (C-2), 69.1 (C-4), 61.9 (C-6), 31.7, 25.7, 22.7 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_3\text{CH}_3$ ), 30.3 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_3\text{CH}_3$ ), 21.0, 20.9, 20.9, 20.9 ( $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 14.1 ( $\text{O}(\text{CH}_2)_5\text{CH}_3$ ); LRMS  $m/z$  calcd for  $\text{C}_{20}\text{H}_{32}\text{O}_{10}$  [ $\text{M}+\text{Na}]^+$ : 455.2. Found 455.1.

### **1,2,4,6-Tetra-O-acetyl-3-O-octyl- $\beta$ -D-glucopyranose (4d)<sup>2</sup>**

Diethyl ether was used instead of dichloromethane as an extraction solvent *en route* to compound **4d**, a white solid (1.21 g, 61 %);  $R_F$  = 0.32 (hexanes/EtOAc, 75:25);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  5.64 (d, 1H,  $J_{1,2}$  = 8.3 Hz, H-1), 5.08 (dd, 1H,  $J_{2,3}$  = 9.3 Hz,  $J_{1,2}$  = 8.2 Hz, H-2), 5.08 (dd, 1H,  $J_{4,5}$  = 9.6 Hz,  $J_{3,4}$  = 9.6 Hz, H-4), 4.22 (dd, 1H,  $J_{6a,6b}$  = 12.4 Hz,  $J_{5,6a}$  = 4.9 Hz, H-6a), 4.09 (dd, 1H,  $J_{6a,6b}$  = 12.2 Hz,  $J_{5,6b}$  = 2.2 Hz, H-6b), 3.72 (ddd, 1H,  $J_{4,5}$  = 9.9 Hz,  $J_{5,6a}$  = 4.9 Hz,  $J_{5,6b}$  = 2.2 Hz, H-5), 3.57 (dd, 1H,  $J_{2,3}$  = 9.3 Hz,  $J_{3,4}$  = 9.3 Hz, H-3), 3.53 (m, 2H,  $\text{OCH}_2$ ), 2.10, 2.08, 2.08, 2.07 (s, 12H,  $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 1.46 (m, 2H,  $\text{OCH}_2\text{CH}_2$ ), 1.33-1.20 (m, 10H,  $\text{OCH}_2\text{CH}_2(\text{CH}_2)_5\text{CH}_3$ ), 0.88 (t, 3H,  $J$  = 7.1 Hz,  $(\text{CH}_2)_7\text{CH}_3$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  170.8, 169.4, 169.3, 169.1 ( $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 92.2 (C-1), 80.4 (C-3), 73.2 (C-5), 72.7 ( $\text{OCH}_2$ ), 71.7 (C-2), 69.2 (C-4), 62.0 (C-6), 31.9, 29.5, 29.3, 26.1, 22.7 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_5\text{CH}_3$ ), 30.3 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_5\text{CH}_3$ ), 21.0, 20.9, 20.9, 20.9 ( $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 14.2 ( $\text{O}(\text{CH}_2)_7\text{CH}_3$ ); LRMS  $m/z$  calcd for  $\text{C}_{22}\text{H}_{36}\text{O}_{10}$  [M+Na]<sup>+</sup>: 483.2. Found 483.2.

### **1,2,4,6-Tetra-O-acetyl-3-O-dodecyl- $\beta$ -D-glucopyranose (4e)<sup>2</sup>**

Diethyl ether was used instead of dichloromethane as an extraction solvent and the reaction mixture was purified by column chromatography (hexanes/EtOAc, 75:25). Compound **4e** was obtained as a white solid (0.58 g, 32 %);  $R_F$  = 0.21 (hexanes/EtOAc, 80:20);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  5.64 (d, 1H,  $J_{1,2}$  = 8.3 Hz, H-1), 5.09 (dd, 1H,  $J_{2,3}$  = 9.3 Hz,  $J_{1,2}$  = 9.4 Hz, H-2), 5.07 (dd, 1H,  $J_{4,5}$  = 9.5 Hz,  $J_{3,4}$  = 9.5 Hz, H-4), 4.22 (dd, 1H,  $J_{6a,6b}$  = 12.5 Hz,  $J_{5,6a}$  = 5.1 Hz, H-6a), 4.10 (dd, 1H,  $J_{6a,6b}$  = 12.4 Hz,  $J_{5,6b}$  = 2.1 Hz, H-6b), 3.72 (ddd, 1H,  $J_{4,5}$  = 9.9 Hz,  $J_{5,6a}$  = 4.8 Hz,  $J_{5,6b}$  = 2.3 Hz, H-5), 3.56 (dd, 1H,  $J_{2,3}$  = 9.4 Hz,  $J_{3,4}$  = 9.4 Hz, H-3), 3.53 (m, 2H,  $\text{OCH}_2$ ), 2.10, 2.08, 2.08, 2.07 (s, 12H,  $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 1.46 (m, 2H,  $\text{OCH}_2\text{CH}_2$ ), 1.33-1.19 (m, 18H,  $\text{OCH}_2\text{CH}_2(\text{CH}_2)_9\text{CH}_3$ ), 0.88 (t, 3H,  $J$  = 7.0 Hz,  $(\text{CH}_2)_{11}\text{CH}_3$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  170.9, 169.4, 169.3, 169.1 ( $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 92.2 (C-1), 80.4 (C-3), 73.2 (C-5), 72.8 ( $\text{OCH}_2$ ), 71.7 (C-2), 69.2 (C-4), 62.0 (C-6), 32.1, 29.8, 29.8, 29.8, 29.7, 29.6, 29.5, 26.1, 22.8 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_9\text{CH}_3$ ), 30.3 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_9\text{CH}_3$ ), 21.0, 20.9, 20.9, 20.9 ( $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 14.2 ( $\text{O}(\text{CH}_2)_{11}\text{CH}_3$ ); LRMS  $m/z$  calcd for  $\text{C}_{26}\text{H}_{44}\text{O}_{10}$  [M+Na]<sup>+</sup>: 539.3. Found 539.2.

<sup>2</sup> Miethchen, R.; Gabriel, T.; Peters, D.; Holz, J.; Michalik, M. *Carbohydr. Res.* **1991**, 214, 331-6.

### **1,2,4,6-Tetra-O-acetyl-3-O-hexadecyl- $\beta$ -D-glucopyranose (4f)<sup>2</sup>**

Hydrolysis of the isopropylidene functionality was performed at 85°C and chloroform was used instead of dichloromethane as an extraction solvent for the acetylation. Compound **4f** was obtained as a white solid (0.80 g, 53 %);  $R_F$  = 0.62 (hexanes/EtOAc, 67:33);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  5.64 (d, 1H,  $J_{1,2}$  = 8.2 Hz, H-1), 5.08 (dd, 1H,  $J_{2,3}$  = 9.3 Hz,  $J_{1,2}$  = 9.4 Hz, H-2), 5.07 (dd, 1H,  $J_{4,5}$  = 9.5 Hz,  $J_{3,4}$  = 9.5 Hz, H-4), 4.22 (dd, 1H,  $^2J_{6a,6b}$  = 12.4 Hz,  $J_{5,6a}$  = 4.9 Hz, H-6a), 4.10 (dd, 1H,  $^2J_{6a,6b}$  = 12.5 Hz,  $J_{5,6b}$  = 2.3 Hz, H-6b), 3.72 (ddd, 1H,  $J_{4,5}$  = 9.9 Hz,  $J_{5,6a}$  = 4.9 Hz,  $J_{5,6b}$  = 2.3 Hz, H-5), 3.56 (dd, 1H,  $J_{2,3}$  = 9.3 Hz,  $J_{3,4}$  = 9.3 Hz, H-3), 3.52 (m, 2H,  $\text{OCH}_2$ ), 2.10, 2.08, 2.08, 2.07 (s, 12H,  $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 1.46 (m, 2H,  $\text{OCH}_2\text{CH}_2$ ), 1.33-1.19 (m, 26H,  $\text{OCH}_2\text{CH}_2(\text{CH}_2)_{13}\text{CH}_3$ ), 0.88 (t, 3H,  $J$  = 7.0 Hz,  $(\text{CH}_2)_{15}\text{CH}_3$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  170.9, 169.4, 169.3, 169.1 ( $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 92.2 (C-1), 80.5 (C-3), 73.2 (C-5), 72.8 ( $\text{OCH}_2$ ), 71.7 (C-2), 69.2 (C-4), 62.0 (C-6), 32.1, 29.8, 29.8, 29.8, 29.8, 29.8, 29.8, 29.7, 29.6, 29.5, 26.1, 22.8 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_{13}\text{CH}_3$ ), 30.3 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_9\text{CH}_3$ ), 21.0, 20.9, 20.9, 20.9 ( $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 14.2 ( $\text{O}(\text{CH}_2)_{15}\text{CH}_3$ ); LRMS  $m/z$  calcd for  $\text{C}_{30}\text{H}_{52}\text{O}_{10} [\text{M}+\text{Na}]^+$ : 595.3. Found 595.3.

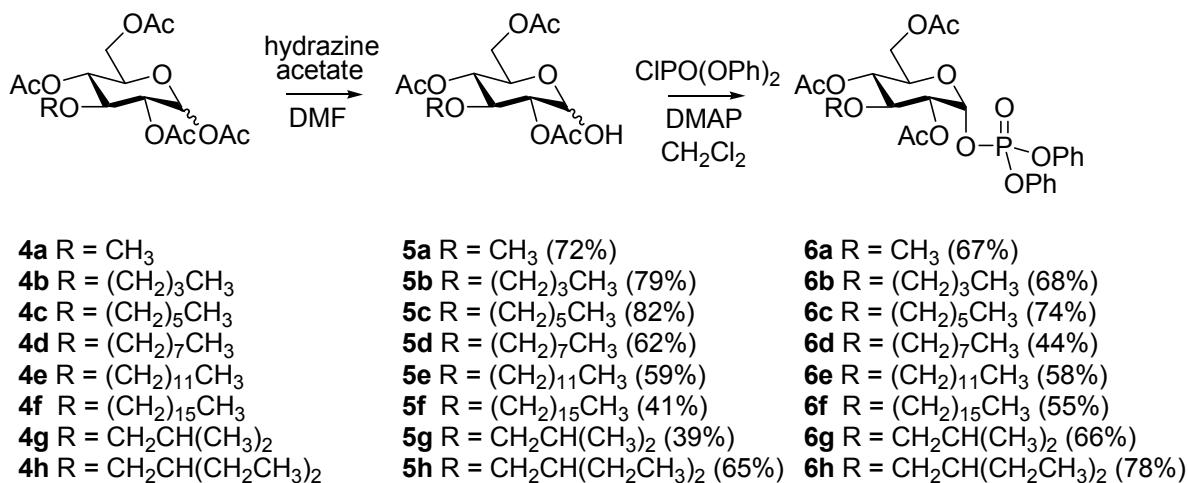
### **1,2,4,6-Tetra-O-acetyl-3-O-(2-methylpropyl)- $\beta$ -D-glucopyranose (4g)**

Compound **4g** was obtained as a colorless liquid (1.25 g, 65 %);  $R_F$  = 0.44 (hexanes/EtOAc, 67:33);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  5.64 (d, 1H,  $J_{1,2}$  = 8.2 Hz, H-1), 5.11 (m, 1H, H-2), 5.09 (m, 1H, H-4), 4.22 (dd, 1H,  $^2J_{6a,6b}$  = 12.4 Hz,  $J_{5,6a}$  = 4.9 Hz, H-6a), 4.09 (dd, 1H,  $^2J_{6a,6b}$  = 12.5 Hz,  $J_{5,6b}$  = 2.3 Hz, H-6b), 3.72 (ddd, 1H,  $J_{4,5}$  = 9.9 Hz,  $J_{5,6a}$  = 4.5 Hz,  $J_{5,6b}$  = 2.4 Hz, H-5), 3.56 (dd, 1H,  $J_{2,3}$  = 9.2 Hz,  $J_{3,4}$  = 9.2 Hz, H-3), 3.30 (dd, 2H,  $J$  = 6.3 Hz,  $^2J$  = 2.2 Hz,  $\text{OCH}_2$ ), 2.10, 2.08, 2.08, 2.07 (s, 12H,  $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 1.74 (nonet, 1H,  $J$  = 6.6 Hz,  $\text{OCH}_2\text{CH}$ ), 0.84 (d, 6H,  $J$  = 6.7 Hz,  $\text{OCH}_2\text{CH}(\text{CH}_3)_2$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  170.7, 169.3, 169.2, 169.0 ( $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 92.0 (C-1), 80.4 (C-3), 79.4 ( $\text{OCH}_2$ ), 73.0 (C-5), 71.5 (C-2), 69.1 (C-4), 61.8 (C-6), 28.9 ( $\text{OCH}_2\text{CH}$ ), 20.9, 20.8, 20.8, 20.8 ( $\text{C}(\text{O})\text{CH}_3 \times 4$ ), 19.1, 19.0 ( $\text{OCH}_2\text{CH}(\text{CH}_3)_2$ ); LRMS  $m/z$  calcd for  $\text{C}_{18}\text{H}_{28}\text{O}_{10} [\text{M}+\text{Na}]^+$ : 427.2. Found 427.2.

### 1,2,4,6-Tetra-O-acetyl-3-O-(2-ethylbutyl)- $\beta$ -D-glucopyranose (4h)

Compound **4h** was obtained as a white solid (0.98 g, 98 %);  $R_F = 0.47$  (hexanes/EtOAc, 67:33);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  5.64 (d, 1H,  $J_{1,2} = 8.3$  Hz, H-1), 5.10 (dd, 1H,  $J_{2,3} = 9.4$  Hz,  $J_{1,2} = 8.3$  Hz, H-2), 5.09 (dd, 1H,  $J_{4,5} = 9.6$  Hz,  $J_{3,4} = 9.6$  Hz, H-4), 4.22 (dd, 1H,  ${}^2J_{6a,6b} = 12.4$  Hz,  $J_{5,6a} = 5.0$  Hz, H-6a), 4.09 (dd, 1H,  ${}^2J_{6a,6b} = 12.5$  Hz,  $J_{5,6b} = 2.3$  Hz, H-6b), 3.72 (ddd, 1H,  $J_{4,5} = 9.9$  Hz,  $J_{5,6a} = 4.9$  Hz,  $J_{5,6b} = 2.3$  Hz, H-5), 3.56 (dd, 1H,  $J_{2,3} = 9.3$  Hz,  $J_{3,4} = 9.3$  Hz, H-3), 3.46 (dd, 2H,  $J = 4.4$  Hz,  ${}^2J = 2.6$  Hz,  $\text{OCH}_2$ ), 2.10, 2.08, 2.08, 2.07 (s, 12H,  $\text{C(O)CH}_3 \times 4$ ), 1.27 (m, 5H,  $\text{OCH}_2\text{CH(CH}_2\text{CH}_3)_2$ ), 0.83 (t, 6H,  $J = 7.3$  Hz,  $\text{OCH}_2\text{CH(CH}_2\text{CH}_3)_2$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  170.9, 169.4, 169.3, 169.1 ( $\text{C(O)CH}_3 \times 4$ ), 92.2 (C-1), 80.4 (C-3), 74.9 ( $\text{OCH}_2$ ), 73.2 (C-5), 71.8 (C-2), 69.4 (C-4), 61.9 (C-6), 42.0 ( $\text{OCH}_2\text{CH}$ ), 23.1, 23.1 ( $\text{OCH}_2\text{CH(CH}_2\text{CH}_3)_2$ ), 21.0, 20.9, 20.9, 20.8 ( $\text{C(O)CH}_3 \times 4$ ), 11.2, 11.2 ( $\text{OCH}_2\text{CH(CH}_2\text{CH}_3)_2$ ); LRMS  $m/z$  calcd for  $\text{C}_{20}\text{H}_{32}\text{O}_{10} [\text{M}+\text{Na}]^+$ : 455.2. Found 455.2.

### General Procedure for Selective Deacetylation and Phosphorylation (5a-h and 6a-h, respectively)



To a stirring solution of the acetylated glucopyranose (**4a-h**) (2.41 mmol) in anhydrous DMF (10 mL) was added hydrazine acetate (0.33 g, 3.62 mmol) and the solution subsequently heated to 60°C for 15 min. The mixture was then stirred for 1 hr at rt before being diluted with H<sub>2</sub>O (20 mL) and extracted with dichloromethane (2 x 20 mL). The combined organic layers were washed with brine (10 mL) and dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated. The major product was purified by flash chromatography to give the selectively deacetylated pyranose (**5a-h**). Under a nitrogen atmosphere, a solution of the selectively

deacetylated product (**5a-h**) (1.49 mmol) and 4-dimethylaminopyridine (438 mg, 3.58 mmol) in anhydrous dichloromethane (8 mL) was stirred for 15 min at rt. The flask was cooled to -10°C and charged with diphenyl chlorophosphate (464 µL, 2.24 mmol). After 1.5 hr, the mixture was diluted with H<sub>2</sub>O (20 mL) and extracted with dichloromethane (2 x 15 mL). The combined organic extracts were washed with brine (10 mL) and dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated. Purification by flash chromatography afforded the title compounds (**6a-g**).

### Diphenyl 2,4,6-tri-*O*-acetyl-3-*O*-methyl- $\alpha$ -D-glucopyranosyl-1-phosphate (**6a**)

Compound **5a**<sup>3</sup> ( $R_F$  = 0.28 (hexanes/EtOAc, 60:40)), a colorless liquid, was provided by silica gel chromatography (hexanes/EtOAc, 60:40) (556 mg, 72%) and immediately carried onto the subsequent phosphorylation. Chromatographic purification (hexanes/EtOAc, 72:28) furnished compound **6a** as a colorless liquid (552 mg, 67%);  $R_F$  = 0.36 (hexanes/EtOAc, 67:33); <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ 7.38-7.20 (m, 10H, Ph x 2), 6.06 (dd, 1H, <sup>1</sup>J<sub>C,H</sub> = 180 Hz, J<sub>1,P</sub> = 6.5 Hz, J<sub>1,2</sub> = 3.3 Hz, H-1), 5.07 (dd, 1H, J<sub>4,5</sub> = 10.2 Hz, J<sub>3,4</sub> = 9.6 Hz, H-4), 4.90 (ddd, 1H, J<sub>2,3</sub> = 10.0 Hz, J<sub>1,2</sub> = 3.1 Hz, <sup>4</sup>J<sub>1,P</sub> = 3.1 Hz, H-2), 4.14 (dd, 1H, <sup>2</sup>J<sub>6a,6b</sub> = 12.5 Hz, J<sub>5,6a</sub> = 4.3 Hz, H-6a), 4.02 (ddd, 1H, J<sub>4,5</sub> = 10.4 Hz, J<sub>5,6a</sub> = 4.2 Hz, J<sub>5,6b</sub> = 2.1 Hz, H-5), 3.90 (dd, 1H, J<sub>6a,6b</sub> = 12.2 Hz, J<sub>5,6b</sub> = 2.2 Hz, H-6b), 3.73 (dd, 1H, J<sub>2,3</sub> = 9.7 Hz, J<sub>3,4</sub> = 9.7 Hz, H-3), 3.45 (s, 3H, CH<sub>3</sub>), 2.10, 2.00, 1.94 (s, 9H, C(O)CH<sub>3</sub> x 3); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 170.8, 170.0, 169.4, (C(O)CH<sub>3</sub> x 3), 150.6-120.1 (12C, Ph x 2), 95.6 (d, <sup>2</sup>J<sub>1,P</sub> = 6.3 Hz, C-1), 77.9 (C-3), 71.9 (d, <sup>3</sup>J<sub>1,P</sub> = 7.1 Hz, C-2), 70.3 (C-5), 68.7 (C-4), 61.5 (C-6), 60.6 (CH<sub>3</sub>), 20.9, 20.8, 20.7 (C(O)CH<sub>3</sub> x 3); <sup>31</sup>P NMR (CDCl<sub>3</sub>) δ -13.98 (s, 1P, P-1); LRMS *m/z* calcd for C<sub>25</sub>H<sub>29</sub>O<sub>12</sub>P [M+Na]<sup>+</sup>: 575.1. Found 575.0.

### Diphenyl 2,4,6-tri-*O*-acetyl-3-*O*-butyl- $\alpha$ -D-glucopyranosyl-1-phosphate (**6b**)

Compound **5b** ( $R_F$  = 0.43 (hexanes/EtOAc, 67:33)), a colorless liquid, was provided by silica gel chromatography (hexanes/EtOAc, 72:28) (690 mg, 79%) and immediately carried onto the subsequent phosphorylation. Chromatographic purification (hexanes/EtOAc, 78:22) furnished compound **6b** as a colorless

<sup>3</sup> Han, X. B.; Jiang, H.; Schmidt, R. R. *Liebigs Ann. Chem.* **1993**, 8, 853-8.

liquid (602 mg, 68%);  $R_F$  = 0.26 (hexanes/EtOAc, 75:25);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  7.38-7.20 (m, 10H, Ph x 2), 6.06 (dd, 1H,  $^1J_{\text{C},\text{H}} = 183$  Hz,  $J_{1,\text{P}} = 6.6$  Hz,  $J_{1,2} = 3.3$  Hz, H-1), 5.08 (dd, 1H,  $J_{4,5} = 10.0$  Hz,  $J_{3,4} = 10.0$  Hz, H-4), 4.90 (ddd, 1H,  $J_{2,3} = 10.0$  Hz,  $J_{1,2} = 3.2$  Hz,  $^4J_{1,\text{P}} = 3.2$  Hz, H-2), 4.14 (dd, 1H,  $^2J_{6\text{a},6\text{b}} = 12.5$  Hz,  $J_{5,6\text{a}} = 4.2$  Hz, H-6a), 4.01 (ddd, 1H,  $J_{4,5} = 10.3$  Hz,  $J_{5,6\text{a}} = 4.1$  Hz,  $J_{5,6\text{b}} = 2.1$  Hz, H-5), 3.89 (dd, 1H,  $J_{6\text{a},6\text{b}} = 12.5$  Hz,  $J_{5,6\text{b}} = 2.2$  Hz, H-6b), 3.79 (dd, 1H,  $J_{2,3} = 9.8$  Hz,  $J_{3,4} = 9.8$  Hz, H-3), 3.56 (m, 2H,  $\text{OCH}_2$ ), 2.08, 2.00, 1.92 (s, 9H,  $\text{C(O)CH}_3$  x 3), 1.46 (m, 2H,  $\text{OCH}_2\text{CH}_2$ ), 1.30 (m, 2H,  $\text{OCH}_2\text{CH}_2\text{CH}_2$ ), 0.89 (t, 3H,  $J = 7.5$  Hz,  $(\text{CH}_2)_3\text{CH}_3$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  170.7, 169.9, 169.3,  $(\text{C(O)CH}_3$  x 3), 150.5-120.1 (12C, Ph x 2), 95.7 (d,  $^2J_{1,\text{P}} = 6.0$  Hz, C-1), 76.5 (C-3), 73.0 ( $\text{OCH}_2$ ), 72.1 (d,  $^3J_{1,\text{P}} = 7.1$  Hz, C-2), 70.3 (C-5), 68.9 (C-4), 61.5 (C-6), 32.3 ( $\text{OCH}_2\text{CH}_2$ ), 20.8, 20.7, 20.6 ( $\text{C(O)CH}_3$  x 3), 19.1 ( $\text{OCH}_2\text{CH}_2\text{CH}_2$ ), 13.9  $((\text{CH}_2)_3\text{CH}_3)$ ;  $^{31}\text{P}$  NMR ( $\text{CDCl}_3$ )  $\delta$  -14.00 (s, 1P, P-1); LRMS  $m/z$  calcd for  $\text{C}_{28}\text{H}_{35}\text{O}_{12}\text{P} [\text{M}+\text{Na}]^+$ : 617.2. Found 617.2.

### Diphenyl 2,4,6-tri-O-acetyl-3-O-hexyl- $\alpha$ -D-glucopyranosyl-1-phosphate (6c)

Diethyl ether was used instead of dichloromethane as an extraction solvent *en route* to compound **5c** ( $R_F$  = 0.30 (hexanes/EtOAc, 70:30)), a colorless liquid, which was provided by silica gel chromatography (hexanes/EtOAc, 63:37) (772 mg, 82%) and immediately carried onto the subsequent phosphorylation. Diethyl ether was again used instead of dichloromethane as an extraction solvent for compound **6c**, a colorless liquid, which was isolated by chromatographic purification (hexanes/EtOAc, 75:25) (538 mg, 58%);  $R_F$  = 0.29 (hexanes/EtOAc, 75:25);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  7.38-7.20 (m, 10H, Ph x 2), 6.06 (dd, 1H,  $^1J_{\text{C},\text{H}} = 180$  Hz,  $J_{1,\text{P}} = 6.5$  Hz,  $J_{1,2} = 3.4$  Hz, H-1), 5.08 (dd, 1H,  $J_{4,5} = 10.0$  Hz,  $J_{3,4} = 10.0$  Hz, H-4), 4.90 (ddd, 1H,  $J_{2,3} = 9.9$  Hz,  $J_{1,2} = 3.0$  Hz,  $^4J_{1,\text{P}} = 3.0$  Hz, H-2), 4.14 (dd, 1H,  $^2J_{6\text{a},6\text{b}} = 12.5$  Hz,  $J_{5,6\text{a}} = 4.2$  Hz, H-6a), 4.01 (ddd, 1H,  $J_{4,5} = 10.4$  Hz,  $J_{5,6\text{a}} = 4.1$  Hz,  $J_{5,6\text{b}} = 2.0$  Hz, H-5), 3.89 (dd, 1H,  $J_{6\text{a},6\text{b}} = 12.6$  Hz,  $J_{5,6\text{b}} = 2.1$  Hz, H-6b), 3.79 (dd, 1H,  $J_{2,3} = 9.7$  Hz,  $J_{3,4} = 9.7$  Hz, H-3), 3.55 (m, 2H,  $\text{OCH}_2$ ), 2.08, 2.00, 1.92 (s, 9H,  $\text{C(O)CH}_3$  x 3), 1.47 (m, 2H,  $\text{OCH}_2\text{CH}_2$ ), 1.28 (m, 6H,  $\text{OCH}_2\text{CH}_2(\text{CH}_2)_3\text{CH}_3$ ), 0.88 (t, 3H,  $J = 6.8$  Hz,  $\text{O}(\text{CH}_2)_5\text{CH}_3$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  170.8, 169.9, 169.3,  $(\text{C(O)CH}_3$  x 3), 150.5-120.1 (12C, Ph x 2), 95.7 (d,  $^2J_{1,\text{P}} = 6.0$  Hz, C-1), 76.5 (C-3), 73.4 ( $\text{OCH}_2$ ), 72.1 (d,  $^3J_{1,\text{P}} = 7.3$  Hz, C-2), 70.4 (C-5), 68.9 (C-4), 61.5 (C-6), 31.7, 25.8, 22.7 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_3\text{CH}_3$ ), 30.3 ( $\text{OCH}_2\text{CH}_2$ ),

20.9, 20.8, 20.6 ( $\text{C}(\text{O})\text{CH}_3$  x 3), 14.2 ( $\text{O}(\text{CH}_2)_5\text{CH}_3$ );  $^{31}\text{P}$  NMR ( $\text{CDCl}_3$ )  $\delta$  -14.00 (s, 1P, P-1); LRMS  $m/z$  calcd for  $\text{C}_{30}\text{H}_{39}\text{O}_{12}\text{P}$  [M+Na] $^+$ : 645.2. Found 645.1.

### Diphenyl 2,4,6-tri-O-acetyl-3-O-octyl- $\alpha$ -D-glucopyranosyl-1-phosphate (6d)

Diethyl ether was used instead of dichloromethane as an extraction solvent *en route* to compound **5d** ( $R_F = 0.27$  (hexanes/EtOAc, 67:33)), a colorless liquid which was provided by silica gel chromatography (hexanes/EtOAc, 63:37) (625 mg, 62%) and immediately carried onto the subsequent phosphorylation. Chromatographic purification (hexanes/EtOAc, 81:19) furnished compound **6d** as a colorless liquid (717 mg, 74%);  $R_F = 0.45$  (hexanes/EtOAc, 67:33);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  7.38-7.20 (m, 10H, Ph x 2), 6.06 (dd, 1H,  $^1J_{\text{C},\text{H}} = 181$  Hz,  $J_{1,\text{P}} = 6.6$  Hz,  $J_{1,2} = 3.4$  Hz, H-1), 5.08 (dd, 1H,  $J_{4,5} = 10.2$  Hz,  $J_{3,4} = 10.2$  Hz, H-4), 4.90 (ddd, 1H,  $J_{2,3} = 10.1$  Hz,  $J_{1,2} = 3.2$  Hz,  $^4J_{1,\text{P}} = 3.2$  Hz, H-2), 4.14 (dd, 1H,  $^2J_{6\text{a},6\text{b}} = 12.5$  Hz,  $J_{5,6\text{a}} = 4.2$  Hz, H-6a), 4.01 (ddd, 1H,  $J_{4,5} = 10.2$  Hz,  $J_{5,6\text{a}} = 3.9$  Hz,  $J_{5,6\text{b}} = 2.0$  Hz, H-5), 3.89 (dd, 1H,  $J_{6\text{a},6\text{b}} = 12.6$  Hz,  $J_{5,6\text{b}} = 2.1$  Hz, H-6b), 3.78 (dd, 1H,  $J_{2,3} = 9.8$  Hz,  $J_{3,4} = 9.8$  Hz, H-3), 3.55 (m, 2H,  $\text{OCH}_2$ ), 2.08, 2.00, 1.92 (s, 9H,  $\text{C}(\text{O})\text{CH}_3$  x 3), 1.47 (m, 2H,  $\text{OCH}_2\text{CH}_2$ ), 1.34-1.19 (m, 10H,  $\text{OCH}_2\text{CH}_2(\text{CH}_2)_5\text{CH}_3$ ), 0.88 (t, 3H,  $J = 7.1$  Hz,  $(\text{CH}_2)_7\text{CH}_3$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  170.8, 169.9, 169.3, ( $\text{C}(\text{O})\text{CH}_3$  x 3), 150.6-120.1 (12C, Ph x 2), 95.7 (d,  $^2J_{1,\text{P}} = 6.1$  Hz, C-1), 76.5 (C-3), 73.5 ( $\text{OCH}_2$ ), 72.1 (d,  $^3J_{1,\text{P}} = 7.2$  Hz, C-2), 70.4 (C-5), 68.9 (C-4), 61.5 (C-6), 32.0, 29.5, 29.4, 26.1, 22.8 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_5\text{CH}_3$ ), 30.3 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_5\text{CH}_3$ ), 20.9, 20.8, 20.6 ( $\text{C}(\text{O})\text{CH}_3$  x 3), 14.2 ( $(\text{CH}_2)_7\text{CH}_3$ );  $^{31}\text{P}$  NMR ( $\text{CDCl}_3$ )  $\delta$  -14.00 (s, 1P, P-1); LRMS  $m/z$  calcd for  $\text{C}_{32}\text{H}_{43}\text{O}_{12}\text{P}$  [M+Na] $^+$ : 673.2. Found 673.3.

### Diphenyl 2,4,6-tri-O-acetyl-3-O-dodecyl- $\alpha$ -D-glucopyranosyl-1-phosphate (6e)

Diethyl ether was used instead of dichloromethane as an extraction solvent *en route* to compound **5e**<sup>4</sup> ( $R_F = 0.29$  (hexanes/EtOAc, 67:33)), a colorless liquid, which was not purified by silica gel chromatography (675 mg, 59%). For the phosphorylation, diethyl ether was again used instead of dichloromethane as an extraction solvent, which was isolated by chromatographic purification (hexanes/EtOAc, 80:20) to furnish compound **6e**

<sup>4</sup> Miethchen, R.; Holz, J.; Prade, H.; Peters, D. *Ger. (East)* **1991**, 3 pp.

as a colorless liquid (463 mg, 44%);  $R_F = 0.38$  (hexanes/EtOAc, 75:25);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  7.38-7.20 (m, 10H, Ph x 2), 6.05 (dd, 1H,  $^1J_{\text{C},\text{H}} = 180$  Hz,  $J_{1,\text{P}} = 6.6$  Hz,  $J_{1,2} = 3.4$  Hz, H-1), 5.08 (dd, 1H,  $J_{4,5} = 10.3$  Hz,  $J_{3,4} = 9.7$  Hz, H-4), 4.90 (ddd, 1H,  $J_{2,3} = 10.0$  Hz,  $J_{1,2} = 3.1$  Hz,  $^4J_{1,\text{P}} = 3.1$  Hz, H-2), 4.14 (dd, 1H,  $^2J_{6\text{a},6\text{b}} = 12.5$  Hz,  $J_{5,6\text{a}} = 4.2$  Hz, H-6a), 4.01 (ddd, 1H,  $J_{4,5} = 10.4$  Hz,  $J_{5,6\text{a}} = 4.1$  Hz,  $J_{5,6\text{b}} = 2.1$  Hz, H-5), 3.89 (dd, 1H,  $J_{6\text{a},6\text{b}} = 12.5$  Hz,  $J_{5,6\text{b}} = 2.1$  Hz, H-6b), 3.79 (dd, 1H,  $J_{2,3} = 9.8$  Hz,  $J_{3,4} = 9.8$  Hz, H-3), 3.55 (m, 2H,  $\text{OCH}_2$ ), 2.08, 2.00, 1.92 (s, 9H,  $\text{C(O)CH}_3$  x 3), 1.47 (m, 2H,  $\text{OCH}_2\text{CH}_2$ ), 1.33-1.19 (m, 18H,  $\text{OCH}_2\text{CH}_2(\text{CH}_2)_9\text{CH}_3$ ), 0.88 (t, 3H,  $J = 6.7$  Hz,  $\text{O}(\text{CH}_2)_{11}\text{CH}_3$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  170.8, 169.9, 169.3, ( $\text{C(O)CH}_3$  x 3), 150.6-120.1 (12C, Ph x 2), 95.7 (d,  $^2J_{1,\text{P}} = 6.1$  Hz, C-1), 76.6 (C-3), 73.5 ( $\text{OCH}_2$ ), 72.1 (d,  $^3J_{1,\text{P}} = 7.3$  Hz, C-2), 70.4 (C-5), 68.9 (C-4), 61.5 (C-6), 32.1, 29.8, 29.8, 29.8, 29.6, 29.5, 26.1, 22.8 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_9\text{CH}_3$ ), 30.4 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_9\text{CH}_3$ ), 20.9, 20.8, 20.7 ( $\text{C(O)CH}_3$  x 3), 14.2 ( $\text{O}(\text{CH}_2)_{11}\text{CH}_3$ );  $^{31}\text{P}$  NMR ( $\text{CDCl}_3$ )  $\delta$  -14.00 (s, 1P, P-1); LRMS  $m/z$  calcd for  $\text{C}_{36}\text{H}_{51}\text{O}_{12}\text{P} [\text{M}+\text{Na}]^+$ : 729.3. Found 729.1.

### Diphenyl 2,4,6-tri-O-acetyl-3-O-hexadecyl- $\alpha$ -D-glucopyranosyl-1-phosphate (6f)

Chloroform was used instead of dichloromethane as an extraction solvent *en route* to compound **5f**<sup>4</sup> ( $R_F = 0.21$  (hexanes/EtOAc, 67:33)), a white solid, which was provided by silica gel chromatography (hexanes/EtOAc, 67:33) (524 mg, 41%) and immediately carried onto the subsequent phosphorylation. Chromatographic purification (hexanes/EtOAc, 80:20) furnished compound **6f** as a colorless liquid (625 mg, 55%);  $R_F = 0.25$  (hexanes/EtOAc, 80:20);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  7.39-7.18 (m, 10H, Ph x 2), 6.06 (dd, 1H,  $^1J_{\text{C},\text{H}} = 180$  Hz,  $J_{1,\text{P}} = 6.6$  Hz,  $J_{1,2} = 3.3$  Hz, H-1), 5.08 (dd, 1H,  $J_{4,5} = 10.0$  Hz,  $J_{3,4} = 10.0$  Hz, H-4), 4.90 (ddd, 1H,  $J_{2,3} = 10.0$  Hz,  $J_{1,2} = 3.0$  Hz,  $^4J_{1,\text{P}} = 3.0$  Hz, H-2), 4.14 (dd, 1H,  $^2J_{6\text{a},6\text{b}} = 12.6$  Hz,  $J_{5,6\text{a}} = 4.3$  Hz, H-6a), 4.01 (ddd, 1H,  $J_{4,5} = 10.3$  Hz,  $J_{5,6\text{a}} = 3.9$  Hz,  $J_{5,6\text{b}} = 2.1$  Hz, H-5), 3.89 (dd, 1H,  $J_{6\text{a},6\text{b}} = 12.5$  Hz,  $J_{5,6\text{b}} = 2.2$  Hz, H-6b), 3.79 (dd, 1H,  $J_{2,3} = 9.8$  Hz,  $J_{3,4} = 9.8$  Hz, H-3), 3.54 (m, 2H,  $\text{OCH}_2$ ), 2.08, 2.00, 1.92 (s, 9H,  $\text{C(O)CH}_3$  x 3), 1.47 (m, 2H,  $\text{OCH}_2\text{CH}_2$ ), 1.33-1.19 (m, 26H,  $\text{OCH}_2\text{CH}_2(\text{CH}_2)_{13}\text{CH}_3$ ), 0.88 (t, 3H,  $J = 7.1$  Hz,  $\text{O}(\text{CH}_2)_{15}\text{CH}_3$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  170.8, 169.9, 169.3, ( $\text{C(O)CH}_3$  x 3), 150.6-120.1 (12C, Ph x 2), 95.7 (d,  $^2J_{1,\text{P}} = 6.2$  Hz, C-1), 76.5 (C-3), 73.4 ( $\text{OCH}_2$ ), 72.1 (d,  $^3J_{1,\text{P}} = 7.2$  Hz, C-2), 70.4 (C-5), 68.9 (C-4), 61.5 (C-6), 32.0, 29.8, 29.8, 29.8, 29.8,

29.8, 29.8, 29.8, 29.7, 29.6, 29.5, 26.1, 22.8 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_{13}\text{CH}_3$ ), 30.3 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_{13}\text{CH}_3$ ), 20.9, 20.7, 20.6 ( $\text{C}(\text{O})\text{CH}_3 \times 3$ ), 14.2 ( $\text{O}(\text{CH}_2)_{15}\text{CH}_3$ );  $^{31}\text{P}$  NMR ( $\text{CDCl}_3$ )  $\delta$  -14.00 (s, 1P, P-1); LRMS  $m/z$  calcd for  $\text{C}_{40}\text{H}_{59}\text{O}_{12}\text{P} [\text{M}+\text{Na}]^+$ : 785.4. Found 785.4.

### Diphenyl 2,4,6-tri-*O*-acetyl-3-*O*-(2-methylpropyl)- $\alpha$ -D-glucopyranosyl-1-phosphate (6g)

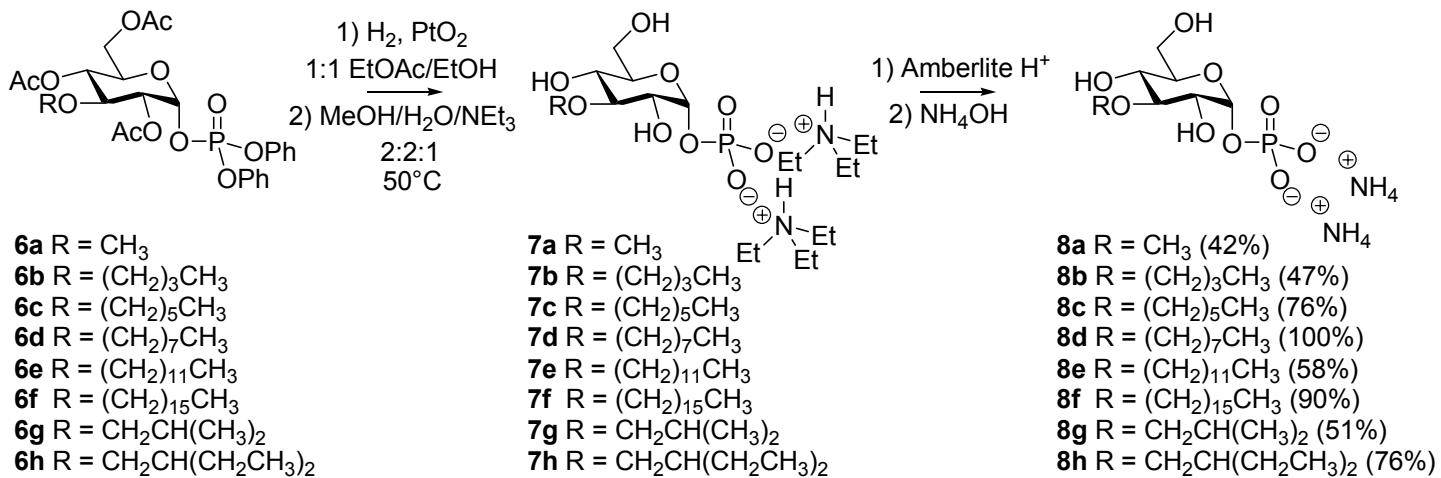
Compound **5g** ( $R_F = 0.23$  (hexanes/EtOAc, 67:33)), a colorless liquid, was provided by silica gel chromatography (hexanes/EtOAc, 60:40) (341 mg, 39%) and immediately carried onto the subsequent phosphorylation. Chromatographic purification (hexanes/EtOAc, 71:29) furnished compound **6g** as a colorless liquid (585 mg, 66%);  $R_F = 0.36$  (hexanes/EtOAc, 66:33);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  7.39-7.18 (m, 10H, Ph x 2), 6.05 (dd, 1H,  $^1J_{\text{C},\text{H}} = 180$  Hz,  $J_{1,\text{P}} = 6.5$  Hz,  $J_{1,2} = 3.2$  Hz, H-1), 5.10 (dd, 1H,  $J_{4,5} = 10.0$  Hz,  $J_{3,4} = 10.0$  Hz, H-4), 4.92 (ddd, 1H,  $J_{2,3} = 9.9$  Hz,  $J_{1,2} = 3.0$  Hz,  $^4J_{1,\text{P}} = 3.0$  Hz, H-2), 4.14 (dd, 1H,  $^2J_{6\text{a},6\text{b}} = 12.6$  Hz,  $J_{5,6\text{a}} = 4.2$  Hz, H-6a), 4.01 (ddd, 1H,  $J_{4,5} = 10.4$  Hz,  $J_{5,6\text{a}} = 4.2$  Hz,  $J_{5,6\text{b}} = 2.3$  Hz, H-5), 3.90 (dd, 1H,  $J_{6\text{a},6\text{b}} = 12.5$  Hz,  $J_{5,6\text{b}} = 2.2$  Hz, H-6b), 3.78 (dd, 1H,  $J_{2,3} = 9.8$  Hz,  $J_{3,4} = 9.8$  Hz, H-3), 3.37 (dd, 1H,  $J = 6.2$  Hz,  $^2J = 8.4$  Hz,  $\text{OCH}_{2a}$ ), 3.28 (dd, 1H,  $J = 6.4$  Hz,  $^2J = 8.8$  Hz,  $\text{OCH}_{2b}$ ), 2.08, 2.00, 1.92 (s, 9H,  $\text{C}(\text{O})\text{CH}_3 \times 3$ ), 1.75 (nonet, 1H,  $J = 6.6$  Hz,  $\text{OCH}_2\text{CH}$ ), 0.84 (d, 6H,  $J = 6.6$  Hz,  $\text{OCH}_2\text{CH}(\text{CH}_3)_2$ );  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ )  $\delta$  170.8, 170.0, 169.3, ( $\text{C}(\text{O})\text{CH}_3 \times 3$ ), 150.6-120.2 (12C, Ph x 2), 95.8 (d,  $^2J_{1,\text{P}} = 6.1$  Hz, C-1), 80.3 ( $\text{OCH}_2$ ), 76.7 (C-3), 72.1 (d,  $^3J_{1,\text{P}} = 7.2$  Hz, C-2), 70.4 (C-5), 68.9 (C-4), 61.5 (C-6), 29.0 ( $\text{OCH}_2\text{CH}$ ), 20.9, 20.8, 20.6 ( $\text{C}(\text{O})\text{CH}_3 \times 3$ ), 19.2, 19.2 ( $\text{OCH}_2\text{CH}(\text{CH}_3)_2$ );  $^{31}\text{P}$  NMR ( $\text{CDCl}_3$ )  $\delta$  -14.00 (s, 1P, P-1); LRMS  $m/z$  calcd for  $\text{C}_{28}\text{H}_{35}\text{O}_{12}\text{P} [\text{M}+\text{Na}]^+$ : 617.2. Found 617.2.

### Diphenyl 2,4,6-tri-*O*-acetyl-3-*O*-(2-ethylbutyl)- $\alpha$ -D-glucopyranosyl-1-phosphate (6h)

Compound **5h** ( $R_F = 0.23$  (hexanes/EtOAc, 67:33)), a white solid, was provided by silica gel chromatography (hexanes/EtOAc, 61:39) (612 mg, 65%) and immediately carried onto the subsequent phosphorylation. Chromatographic purification (hexanes/EtOAc, 78:22) furnished compound **6h** as a colorless liquid (724 mg, 78%);  $R_F = 0.24$  (hexanes/EtOAc, 78:22);  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  7.39-7.17 (m, 10H, Ph x 2), 6.05 (dd, 1H,  $^1J_{\text{C},\text{H}} =$

180 Hz,  $J_{1,P} = 6.6$  Hz,  $J_{1,2} = 3.4$  Hz, H-1), 5.09 (dd, 1H,  $J_{4,5} = 10.2$  Hz,  $J_{3,4} = 9.5$  Hz, H-4), 4.91 (ddd, 1H,  $J_{2,3} = 10.0$  Hz,  $J_{1,2} = 3.2$  Hz,  $J_{1,P} = 3.2$  Hz, H-2), 4.13 (dd, 1H,  $J_{6a,6b} = 12.6$  Hz,  $J_{5,6a} = 4.4$  Hz, H-6a), 4.01 (ddd, 1H,  $J_{4,5} = 10.4$  Hz,  $J_{5,6a} = 4.2$  Hz,  $J_{5,6b} = 2.1$  Hz, H-5), 3.89 (dd, 1H,  $J_{6a,6b} = 12.5$  Hz,  $J_{5,6b} = 2.2$  Hz, H-6b), 3.78 (dd, 1H,  $J_{2,3} = 9.7$  Hz,  $J_{3,4} = 9.7$  Hz, H-3), 3.52 (dd, 1H,  $J = 4.6$  Hz,  $^2J = 9.0$  Hz, OCH<sub>2a</sub>), 3.42 (dd, 1H,  $J = 4.4$  Hz,  $^2J = 8.8$  Hz, OCH<sub>2b</sub>), 2.08, 2.00, 1.92 (s, 9H, C(O)CH<sub>3</sub> x 3), 1.28 (m, 5H, OCH<sub>2</sub>CH(CH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 0.83 (t, 6H,  $J = 7.0$  Hz, OCH<sub>2</sub>CH(CH<sub>3</sub>)<sub>2</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ 170.8, 170.0, 169.3, (C(O)CH<sub>3</sub> x 3), 150.5-120.2 (12C, Ph x 2), 95.7 (d,  $^2J_{1,P} = 6.1$  Hz, C-1), 76.6 (C-3), 75.4 (OCH<sub>2</sub>), 72.3 (d,  $^3J_{1,P} = 7.3$  Hz, C-2), 70.4 (C-5), 69.0 (C-4), 61.5 (C-6), 42.0 (OCH<sub>2</sub>CH), 23.2, 23.2 (OCH<sub>2</sub>CH(CH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>), 20.9, 20.8, 20.6 (C(O)CH<sub>3</sub> x 3), 11.3, 11.3 (OCH<sub>2</sub>CH(CH<sub>2</sub>CH<sub>3</sub>)<sub>2</sub>); <sup>31</sup>P NMR (CDCl<sub>3</sub>) δ -13.99 (s, 1P, P-1); LRMS *m/z* calcd for C<sub>30</sub>H<sub>39</sub>O<sub>12</sub>P [M+Na]<sup>+</sup>: 645.2. Found 645.2.

### General Procedure for Deprotection and Ion Exchange (7a-h and 8a-h, respectively)



A solution of phosphorylated pyranoside (0.384 mmol) and platinum (IV) oxide (0.242 mmol) in 1:1 EtOH/EtOAc (4 mL) was shaken under H<sub>2</sub> in a Parr apparatus at 54 PSI for 2 h. The reaction mixture was filtered through Celite and concentrated. The residue was taken up in 2:2:1 MeOH/H<sub>2</sub>O/NEt<sub>3</sub> (10 mL) and stirred at 50°C for 36 h. Again, the mixture was concentrated and the residue was partitioned between H<sub>2</sub>O (10 mL) and EtOAc (10 mL). The aqueous layer containing the triethyl ammonium salt (7a-h) was passed through Amberlite IR-120 PLUS(H) ion exchange resin. The resulting acidic aqueous fraction was immediately

adjusted to pH 8 with NH<sub>4</sub>OH 0.1 M, concentrated to 5 mL, and lyophilized. Titration with NH<sub>4</sub>OH 0.1 M and lyophilization was repeated twice more to remove NH<sub>4</sub>OAc and isolate the ammonium salt (**8a-h**).

### **3-O-Methyl- $\alpha$ -D-glucopyranosyl phosphate diammonium salt (8a)<sup>5</sup>**

Deacetylation was performed at rt and compound **8a** was isolated as a colorless foam (50 mg, 42%); <sup>1</sup>H NMR (D<sub>2</sub>O) δ 5.35 (dd, 1H, <sup>1</sup>J<sub>C,H</sub> = 176 Hz, J<sub>1,P</sub> = 7.2 Hz, J<sub>1,2</sub> = 3.0 Hz, H-1), 3.80 (ddd, 1H, J<sub>4,5</sub> = 9.6 Hz, J<sub>5,6b</sub> = 4.7 Hz, J<sub>5,6a</sub> = 2.0 Hz, H-5), 3.75 (dd, 1H, <sup>2</sup>J<sub>6a,6b</sub> = 12.2 Hz, J<sub>5,6a</sub> = 2.0 Hz, H-6a), 3.65 (dd, 1H, J<sub>6a,6b</sub> = 12.4 Hz, J<sub>5,6b</sub> = 4.9 Hz, H-6b), 3.52 (s, 3H, CH<sub>3</sub>), 3.47 (m, 1H, H-2), 3.44 (m, 1H, H-3), 3.39 (m, 1H, H-4); <sup>13</sup>C NMR (D<sub>2</sub>O) δ 94.2 (d, <sup>2</sup>J<sub>1,P</sub> = 5.7 Hz, C-1), 82.8 (C-3), 72.3 (C-5), 71.4 (d, <sup>3</sup>J<sub>1,P</sub> = 7.5 Hz, C-2), 69.0 (C-4), 60.5 (C-6), 60.0 (CH<sub>3</sub>); <sup>31</sup>P NMR (D<sub>2</sub>O) δ 0.77 (s, 1P, P-1); HRMS *m/z* calcd for C<sub>7</sub>H<sub>13</sub>O<sub>9</sub>P<sup>2-</sup> [M+H]<sup>-</sup>: 273.0386. Found 273.0390.

### **3-O-Butyl- $\alpha$ -D-glucopyranosyl phosphate diammonium salt (8b)**

Compound **8b** was obtained as a colorless foam (63 mg, 47%); <sup>1</sup>H NMR (D<sub>2</sub>O) δ 5.33 (dd, 1H, <sup>1</sup>J<sub>C,H</sub> = 172 Hz, J<sub>1,P</sub> = 7.4 Hz, J<sub>1,2</sub> = 3.4 Hz, H-1), 3.79 (ddd, 1H, J<sub>4,5</sub> = 10.1 Hz, J<sub>5,6b</sub> = 5.0 Hz, J<sub>5,6a</sub> = 2.2 Hz, H-5), 3.75 (dd, 1H, <sup>2</sup>J<sub>6a,6b</sub> = 12.5 Hz, J<sub>5,6a</sub> = 2.2 Hz, H-6a), 3.72 (t, 2H, *J* = 6.7 Hz, OCH<sub>2</sub>), 3.63 (dd, 1H, J<sub>6a,6b</sub> = 12.5 Hz, J<sub>5,6b</sub> = 5.2 Hz, H-6b), 3.50 (dd, 1H, J<sub>2,3</sub> = 9.5 Hz, J<sub>3,4</sub> = 9.2 Hz, H-3), 3.42 (ddd, 1H, J<sub>2,3</sub> = 9.6 Hz, J<sub>1,2</sub> = 3.3 Hz, <sup>4</sup>J<sub>2,P</sub> = 2.0 Hz, H-2), 3.33 (dd, 1H, J<sub>3,4</sub> = 9.6 Hz, J<sub>4,5</sub> = 9.6 Hz, H-4), 1.49 (p, 2H, *J* = 7.0 Hz, OCH<sub>2</sub>CH<sub>2</sub>), 1.26 (sextet, 2H, *J* = 7.5 Hz, O(CH<sub>2</sub>)<sub>2</sub>CH<sub>2</sub>CH<sub>3</sub>), 0.79 (t, 3H, *J* = 7.8 Hz, O(CH<sub>2</sub>)<sub>3</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (D<sub>2</sub>O) δ 94.1 (d, <sup>2</sup>J<sub>1,P</sub> = 5.5 Hz, C-1), 81.5 (C-3), 73.1 (OCH<sub>2</sub>), 72.3 (C-5), 71.8 (d, <sup>3</sup>J<sub>1,P</sub> = 7.4 Hz, C-2), 69.3 (C-4), 60.7 (C-6), 31.5 (OCH<sub>2</sub>CH<sub>2</sub>), 18.5 (OCH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>), 13.1 (O(CH<sub>2</sub>)<sub>3</sub>CH<sub>3</sub>); <sup>31</sup>P NMR (D<sub>2</sub>O) δ 1.32 (s, 1P, P-1); HRMS *m/z* calcd for C<sub>10</sub>H<sub>19</sub>O<sub>9</sub>P<sup>2-</sup> [M+H]<sup>-</sup>: 315.0856. Found 315.0856.

### **3-O-Hexyl- $\alpha$ -D-glucopyranosyl phosphate diammonium salt (8c)**

<sup>5</sup> Jiang, J.; Albermann, C.; Thorson, J. *ChemBioChem*. **2003**, *4*, 443-446.

Compound **8c** was obtained as a colorless foam (110 mg, 76%);  $^1\text{H}$  NMR ( $\text{D}_2\text{O}$ )  $\delta$  5.34 (dd, 1H,  $^1J_{\text{C},\text{H}} = 173$  Hz,  $J_{1,\text{P}} = 6.9$  Hz,  $J_{1,2} = 3.0$  Hz, H-1), 3.78 (m, 1H, H-5), 3.74 (m, 1H, H-6a), 3.70 (t, 2H,  $J = 6.6$  Hz,  $\text{OCH}_2$ ), 3.64 (dd, 1H,  $J_{6\text{a},6\text{b}} = 12.0$  Hz,  $J_{5,6\text{b}} = 4.5$  Hz, H-6b), 3.49 (dd, 1H,  $J_{2,3} = 9.3$  Hz,  $J_{3,4} = 9.3$  Hz, H-3), 3.44 (ddd, 1H,  $J_{2,3} = 9.7$  Hz, H-2), 3.35 (dd, 1H,  $J_{3,4} = 9.5$  Hz,  $J_{4,5} = 9.5$  Hz, H-4), 1.50 (p, 2H,  $J = 7.0$  Hz,  $\text{OCH}_2\text{CH}_2$ ), 1.22 (m, 6H,  $\text{OCH}_2\text{CH}_2(\text{CH}_2)_3\text{CH}_3$ ), 0.76 (m, 3H,  $\text{O}(\text{CH}_2)_5\text{CH}_3$ );  $^{13}\text{C}$  NMR ( $\text{D}_2\text{O}$ )  $\delta$  94.4 (d,  $^2J_{1,\text{P}} = 5.7$  Hz, C-1), 81.4 (C-3), 73.4 ( $\text{OCH}_2$ ), 72.4 (C-5), 71.6 (d,  $^3J_{1,\text{P}} = 7.7$  Hz, C-2), 69.1 (C-4), 60.6 (C-6), 30.9, 24.8, 22.0 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_3\text{CH}_3$ ), 29.2 ( $\text{OCH}_2\text{CH}_2$ ), 13.4 ( $\text{O}(\text{CH}_2)_5\text{CH}_3$ );  $^{31}\text{P}$  NMR ( $\text{D}_2\text{O}$ )  $\delta$  0.41 (s, 1P, P-1); HRMS  $m/z$  calcd for  $\text{C}_{12}\text{H}_{23}\text{O}_9\text{P}^{2-}$   $[\text{M}+\text{H}]^-$ : 343.1169. Found 343.1176.

### **3-O-Octyl- $\alpha$ -D-glucopyranosyl phosphate diammonium salt (8d)**

Diethyl ether was used instead of ethyl acetate as an extraction solvent *en route* to compound **8d**, a colorless foam (66 mg, 42%);  $^1\text{H}$  NMR ( $\text{D}_2\text{O}$ )  $\delta$  5.36 (dd, 1H,  $^1J_{\text{C},\text{H}} = 173$  Hz,  $J_{1,\text{P}} = 7.2$  Hz,  $J_{1,2} = 3.2$  Hz, H-1), 3.78 (ddd, 1H,  $J_{4,5} = 9.8$  Hz,  $J_{5,6\text{b}} = 5.0$  Hz,  $J_{5,6\text{a}} = 2.1$  Hz, H-5), 3.75 (m, 1H, H-6a), 3.72 (t, 2H,  $J = 7.0$  Hz,  $\text{OCH}_2$ ), 3.66 (dd, 1H,  $J_{6\text{a},6\text{b}} = 12.3$  Hz,  $J_{5,6\text{b}} = 4.9$  Hz, H-6b), 3.50 (m, 1H, H-3), 3.45 (m, 1H, H-2), 3.37 (dd, 1H,  $J_{3,4} = 9.4$  Hz,  $J_{4,5} = 9.4$  Hz, H-4), 1.50 (p, 2H,  $J = 7.2$  Hz,  $\text{OCH}_2\text{CH}_2$ ), 1.30-1.12 (m, 10H,  $\text{OCH}_2\text{CH}_2(\text{CH}_2)_5\text{CH}_3$ ), 0.77 (t, 3H,  $J = 7.0$  Hz,  $(\text{CH}_2)_7\text{CH}_3$ );  $^{13}\text{C}$  NMR ( $\text{D}_2\text{O}$ )  $\delta$  94.6 (d,  $^2J_{1,\text{P}} = 5.9$  Hz, C-1), 81.4 (C-3), 73.5 ( $\text{OCH}_2$ ), 72.5 (C-5), 71.5 (d,  $^3J_{1,\text{P}} = 7.7$  Hz, C-2), 69.1 (C-4), 60.5 (C-6), 31.1, 28.5, 28.4, 25.1, 22.0 ( $\text{OCH}_2\text{CH}_2(\text{CH}_2)_5\text{CH}_3$ ), 29.3 ( $\text{OCH}_2\text{CH}_2$ ), 13.4 ( $\text{O}(\text{CH}_2)_7\text{CH}_3$ );  $^{31}\text{P}$  NMR ( $\text{D}_2\text{O}$ )  $\delta$  0.12 (s, 1P, P-1); HRMS  $m/z$  calcd for  $\text{C}_{14}\text{H}_{28}\text{O}_9\text{P}^{2-}$   $[\text{M}+\text{H}]^-$ : 371.1482. Found 371.1490.

### **3-O-Dodecyl- $\alpha$ -D-glucopyranosyl phosphate diammonium salt (8e)**

Deacetylation was performed at 70°C instead of 50°C and diethyl ether was used instead of ethyl acetate as an extraction solvent. Triethylamine (1 mL) was added before rotary evaporation to suppress the surfactant-like behavior of the salt. After the third  $\text{NH}_4\text{OAc}$  sublimation, the residue was dissolved in water and filtered through a short pad of Bio-Rad Chelex 100. The solution was then passed through Amberlite IR-120 PLUS(H) ion exchange resin, titrated with  $\text{NH}_4\text{OH}$  0.1 M, and lyophilized to forge compound **8e** as a colorless foam (103

mg, 58%);  $^1\text{H}$  NMR (MeOD)  $\delta$  5.48 (dd, 1H,  $^1J_{\text{C},\text{H}} = 173$  Hz,  $J_{1,\text{P}} = 6.9$  Hz,  $J_{1,2} = 3.3$  Hz, H-1), 3.85 (ddd, 1H,  $J_{4,5} = 9.8$  Hz,  $J_{5,6\text{b}} = 5.7$  Hz,  $J_{5,6\text{a}} = 2.1$  Hz, H-5), 3.81 (m, 2H, OCH<sub>2</sub>), 3.80 (m, 1H, H-6a), 3.63 (dd, 1H,  $J_{6\text{a},6\text{b}} = 11.6$  Hz,  $J_{5,6\text{b}} = 5.5$  Hz, H-6b), 3.47 (dd, 1H,  $J_{2,3} = 9.0$  Hz,  $J_{3,4} = 9.0$  Hz, H-3), 3.42 (ddd, 1H,  $J_{2,3} = 9.6$  Hz,  $J_{1,2} = 3.0$  Hz,  $^4J_{1,\text{P}} = 3.0$  Hz, H-2), 3.32 (m, 1H, H-4), 1.62 (p, 2H,  $J = 7.3$  Hz, OCH<sub>2</sub>CH<sub>2</sub>), 1.41-1.23 (m, 18H, OCH<sub>2</sub>CH<sub>2</sub>(CH<sub>2</sub>)<sub>9</sub>CH<sub>3</sub>), 0.90 (t, 3H,  $J = 7.0$  Hz, (CH<sub>2</sub>)<sub>11</sub>CH<sub>3</sub>);  $^{13}\text{C}$  NMR (MeOD)  $\delta$  96.6 (d,  $^2J_{1,\text{P}} = 6.2$  Hz, C-1), 83.6 (C-3), 74.6 (OCH<sub>2</sub>), 74.5 (C-5), 74.0 (d,  $^3J_{1,\text{P}} = 7.5$  Hz, C-2), 71.4 (C-4), 62.8 (C-6), 33.1, 30.8, 30.8, 30.8, 30.7, 30.5, 27.1, 23.7 (OCH<sub>2</sub>CH<sub>2</sub>(CH<sub>2</sub>)<sub>9</sub>CH<sub>3</sub>), 31.4 (OCH<sub>2</sub>CH<sub>2</sub>), 14.4 (O(CH<sub>2</sub>)<sub>11</sub>CH<sub>3</sub>);  $^{31}\text{P}$  NMR (MeOD)  $\delta$  -1.86 (s, 1P, P-1); HRMS  $m/z$  calcd for C<sub>18</sub>H<sub>35</sub>O<sub>9</sub>P<sup>2-</sup> [M+H]<sup>-</sup>: 427.2108. Found 427.2115.

### **3-O-Hexadecyl- $\alpha$ -D-glucopyranosyl phosphate diammonium salt (8f)**

Deacetylation was performed at 70°C instead of 50°C and diethyl ether was used instead of ethyl acetate as an extraction solvent. Triethylamine (1 mL) was added before rotary evaporation to suppress the surfactant-like behavior of the salt. After the third NH<sub>4</sub>OAc sublimation, the residue was dissolved in water and filtered through a short pad of Bio-Rad Chelex 100. The solution was then passed through Amberlite IR-120 PLUS(H) ion exchange resin, titrated with NH<sub>4</sub>OH 0.1 M, and lyophilized to forge compound **8f** as a colorless foam (179 mg, 90%). Coupling constants are omitted from the  $^1\text{H}$  NMR spectrum because of the compound's inherent insolubility in a variety of NMR solvents led to significant line-broadening;  $^1\text{H}$  NMR (D<sub>2</sub>O)  $\delta$  5.44 (d, 1H,  $^1J_{\text{C},\text{H}} = 174$  Hz, H-1), 3.86 (m, 1H, H-5), 3.86 (m, 1H, H-6a), 3.74 (m, 2H, OCH<sub>2</sub>), 3.69 (m, 1H, H-6b), 3.56 (m, 1H, H-2), 3.52 (m, 1H, H-3), 3.39 (m, 1H, H-4), 1.58 (m, 2H, OCH<sub>2</sub>CH<sub>2</sub>), 1.42-1.15 (m, 26H, OCH<sub>2</sub>CH<sub>2</sub>(CH<sub>2</sub>)<sub>13</sub>CH<sub>3</sub>), 0.84 (m, 3H, (CH<sub>2</sub>)<sub>15</sub>CH<sub>3</sub>);  $^{13}\text{C}$  NMR (D<sub>2</sub>O)  $\delta$  94.9 (d,  $^2J_{1,\text{P}} = 5.1$  Hz, C-1), 81.0 (C-3), 72.7 (C-5), 72.2 (OCH<sub>2</sub>), 71.1 (d,  $^3J_{1,\text{P}} = 7.3$  Hz, C-2), 69.4 (C-4), 61.0 (C-6), 32.1, 30.3, 30.3, 30.3, 30.3, 30.3, 30.3, 30.1, 30.1, 30.0, 29.7, 26.2, 22.7 (OCH<sub>2</sub>(CH<sub>2</sub>)<sub>14</sub>CH<sub>3</sub>), 13.8 (O(CH<sub>2</sub>)<sub>15</sub>CH<sub>3</sub>);  $^{31}\text{P}$  NMR (D<sub>2</sub>O)  $\delta$  -0.57 (s, 1P, P-1); HRMS  $m/z$  calcd for C<sub>22</sub>H<sub>43</sub>O<sub>9</sub>P<sup>2-</sup> [M+H]<sup>-</sup>: 483.2734. Found 483.2708.

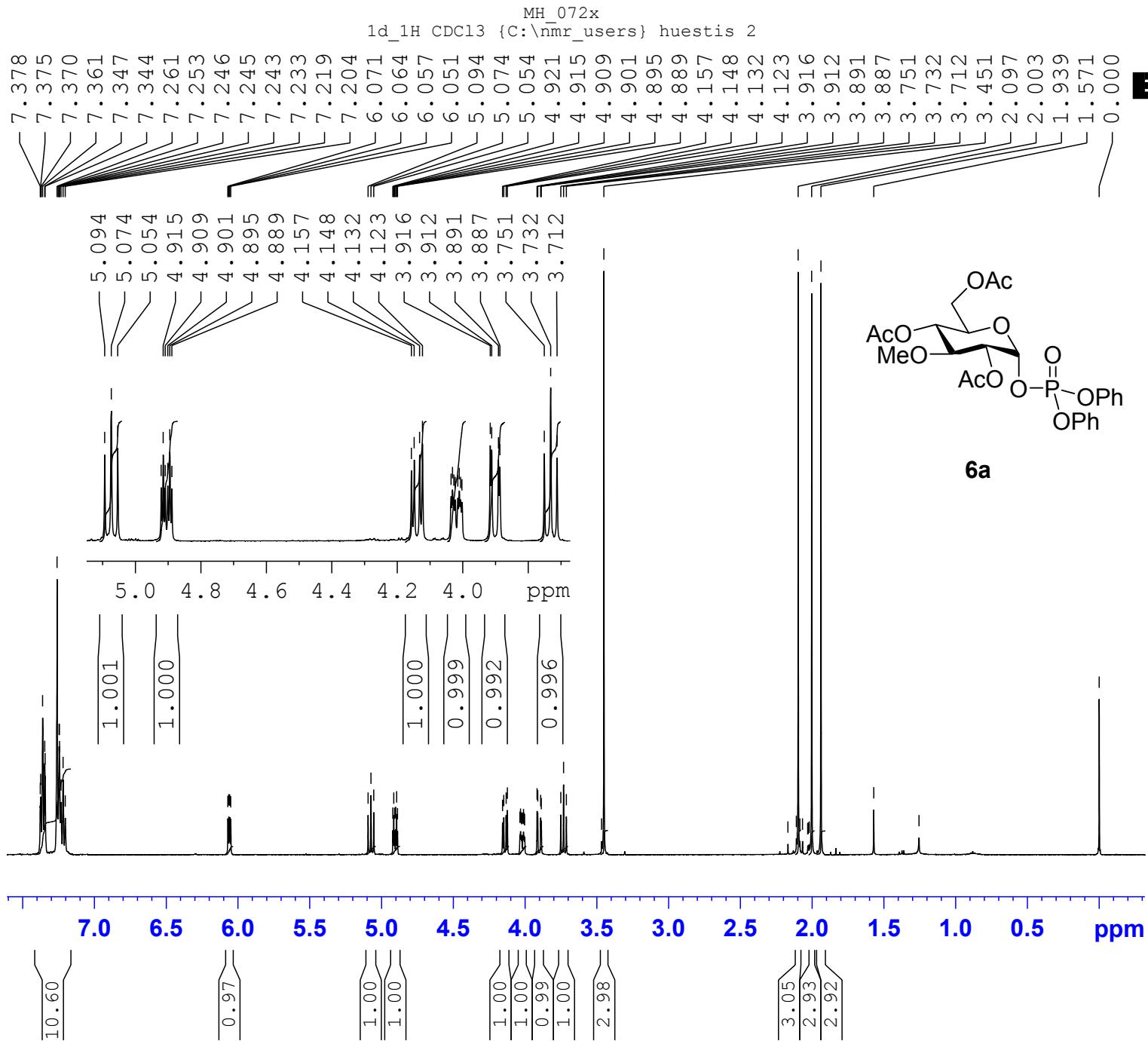
### **3-O-(2-Methylpropyl)- $\alpha$ -D-glucopyranosyl phosphate diammonium salt (8g)**

Compound **8g** was obtained as a colorless foam (69 mg, 51%);  $^1\text{H}$  NMR ( $\text{D}_2\text{O}$ )  $\delta$  5.29 (dd, 1H,  $^1J_{\text{C},\text{H}} = 171$  Hz,  $J_{1,\text{P}} = 7.3$  Hz,  $J_{1,2} = 3.3$  Hz, H-1), 3.76 (ddd, 1H,  $J_{4,5} = 10.0$  Hz,  $J_{5,6\text{b}} = 4.8$  Hz,  $J_{5,6\text{a}} = 1.8$  Hz, H-5), 3.72 (dd, 1H,  $J_{6\text{a},6\text{b}} = 12.3$  Hz,  $J_{5,6\text{a}} = 1.9$  Hz, H-6a), 3.60 (dd, 1H,  $J_{6\text{a},6\text{b}} = 12.1$  Hz,  $J_{5,6\text{b}} = 5.2$  Hz, H-6b), 3.47 (dd, 1H,  $J_{2,3} = 9.8$  Hz,  $J_{3,4} = 9.4$  Hz, H-3), 3.46 (d, 2H,  $J = 6.1$  Hz,  $\text{OCH}_2$ ), 3.41 (m, 1H, H-2), 3.31 (dd, 1H,  $J_{3,4} = 9.5$  Hz,  $J_{4,5} = 9.5$  Hz, H-4), 1.74 (nonet, 1H,  $J = 6.5$  Hz,  $\text{OCH}_2\text{CH}$ ), 0.81, 0.81 (d, 3H,  $J = 6.7$  Hz,  $\text{OCH}_2\text{CH}(\text{CH}_3)_2$ );  $^{13}\text{C}$  NMR ( $\text{D}_2\text{O}$ )  $\delta$  94.0 (d,  $^2J_{1,\text{P}} = 5.6$  Hz, C-1), 81.7 (C-3), 80.1 ( $\text{OCH}_2$ ), 72.2 (C-5), 71.8 (d,  $^3J_{1,\text{P}} = 7.2$  Hz, C-2), 69.3 (C-4), 60.7 (C-6), 28.2 ( $\text{OCH}_2\text{CH}$ ), 18.6, 18.6 ( $\text{OCH}_2\text{CH}(\text{CH}_3)_2$ );  $^{31}\text{P}$  NMR ( $\text{D}_2\text{O}$ )  $\delta$  1.76 (s, 1P, P-1); HRMS  $m/z$  calcd for  $\text{C}_{10}\text{H}_{19}\text{O}_9\text{P}^{2-}$  [M+H] $^-$ : 315.0856. Found 315.0869.

### **3-O-(2-Ethylbutyl)- $\alpha$ -D-glucopyranosyl phosphate diammonium salt (8h)**

Compound **8h** was obtained as a colorless foam (145 mg, 76%);  $^1\text{H}$  NMR ( $\text{D}_2\text{O}$ )  $\delta$  5.32 (dd, 1H,  $^1J_{\text{C},\text{H}} = 172$  Hz,  $J_{1,\text{P}} = 4.6$  Hz,  $J_{1,2} = 3.0$  Hz, H-1), 3.79 (m, 1H, H-5), 3.75 (d, 1H,  $^2J_{6\text{a},6\text{b}} = 12.6$  Hz, H-6a), 3.63 (m, 1H, H-6b), 3.61 (m, 2H,  $\text{OCH}_2$ ), 3.49 (dd, 1H,  $J_{2,3} = 9.2$  Hz,  $J_{3,4} = 9.2$  Hz, H-3), 3.42 (d, 1H,  $J_{2,3} = 9.6$  Hz, H-2), 3.33 (dd, 1H,  $J_{3,4} = 9.2$  Hz,  $J_{4,5} = 9.2$  Hz, H-4), 1.39 (m, 1H,  $\text{OCH}_2\text{CH}(\text{CH}_2\text{CH}_3)_2$ ), 1.25 (m, 4H,  $\text{OCH}_2\text{CH}(\text{CH}_2\text{CH}_3)_2$ ), 0.76 (t, 6H,  $J = 6.7$  Hz,  $\text{OCH}_2\text{CH}(\text{CH}_2\text{CH}_3)_2$ );  $^{13}\text{C}$  NMR ( $\text{D}_2\text{O}$ )  $\delta$  94.1 (d,  $^2J_{1,\text{P}} = 5.6$  Hz, C-1), 81.7 (C-3), 75.7 ( $\text{OCH}_2$ ), 72.3 (C-5), 71.8 (d,  $^3J_{1,\text{P}} = 7.3$  Hz, C-2), 69.3 (C-4), 60.7 (C-6), 41.0 ( $\text{OCH}_2\text{CH}$ ), 22.5, 22.4 ( $\text{OCH}_2\text{CH}(\text{CH}_2\text{CH}_3)_2$ ), 10.2, 10.2 ( $\text{OCH}_2\text{CH}(\text{CH}_2\text{CH}_3)_2$ );  $^{31}\text{P}$  NMR ( $\text{D}_2\text{O}$ )  $\delta$  1.62 (s, 1P, P-1); HRMS  $m/z$  calcd for  $\text{C}_{12}\text{H}_{23}\text{O}_9\text{P}^{2-}$  [M+H] $^-$ : 343.1169. Found 343.1159.

### **$^1\text{H}$ , $^{13}\text{C}$ and $^{31}\text{P}$ NMR Spectra of compounds 6a-h and 8a-h**



The Bruker logo consists of the word "BRUKER" in a bold, black, sans-serif font. Behind the text is a stylized blue atom model with three elliptical orbits intersecting at two points where small blue spheres are located.

Current	Data	Parameters
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PROCNO		1

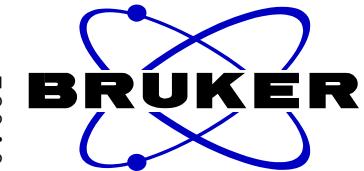
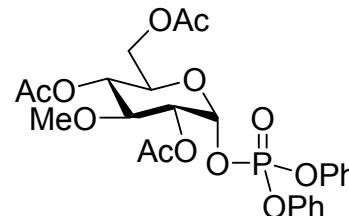
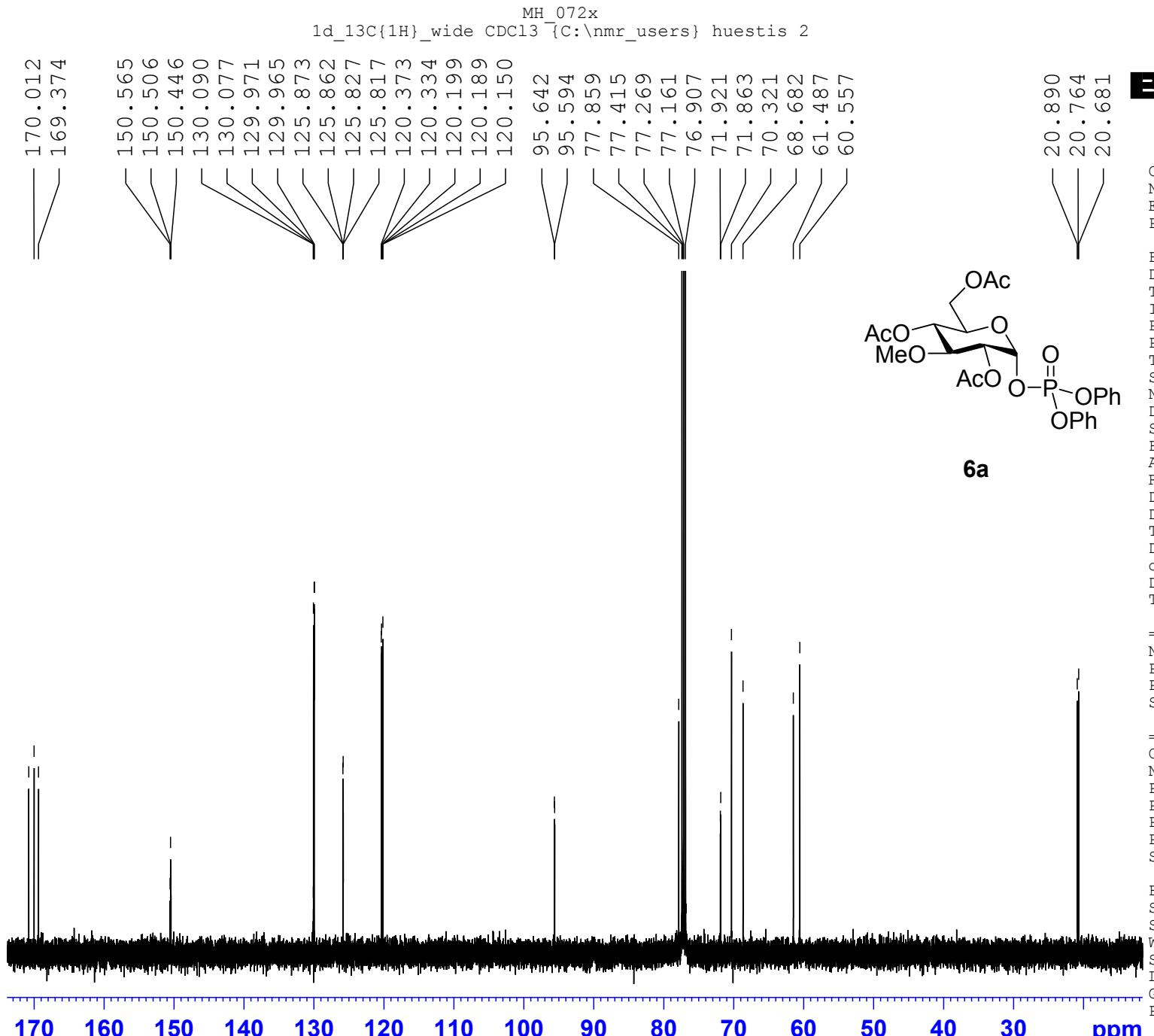
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D1
TD0             1.00000000 sec

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```
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PL1         -2.00 dB  
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Current Data Parameters  
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EXPNO 11  
PROCNO 1

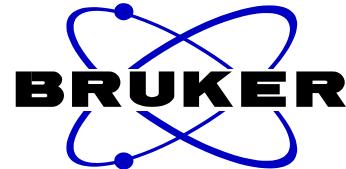
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AQ 0.9699976 sec  
RG 645.1  
DW 14.800 usec  
DE 6.00 usec  
TE 300.4 K  
D1 1.0000000 sec  
d11 0.0300000 sec  
DELTA 0.8999998 sec  
TD0 4

===== CHANNEL f1 ======  
NUC1 13C  
P1 7.90 usec  
PL1 0.00 dB  
SFO1 125.7716270 MHz

===== CHANNEL f2 ======  
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PCPD2 80.00 usec  
PL2 -2.00 dB  
PL12 15.72 dB  
PL13 19.37 dB  
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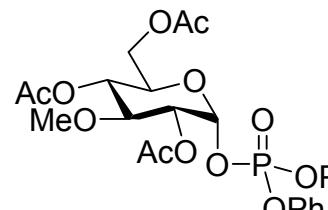
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MH\_072x  
1d\_31P{1H} CDC13 {C:\nmr\_users} huestis 2



Current Data Parameters  
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EXPNO 14  
PROCNO 1

F2 - Acquisition Parameters  
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Time 14.59  
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PULPROG zgpg30  
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NS 128  
DS 0  
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FIDRES 1.495960 Hz  
AQ 0.3342887 sec  
RG 8192  
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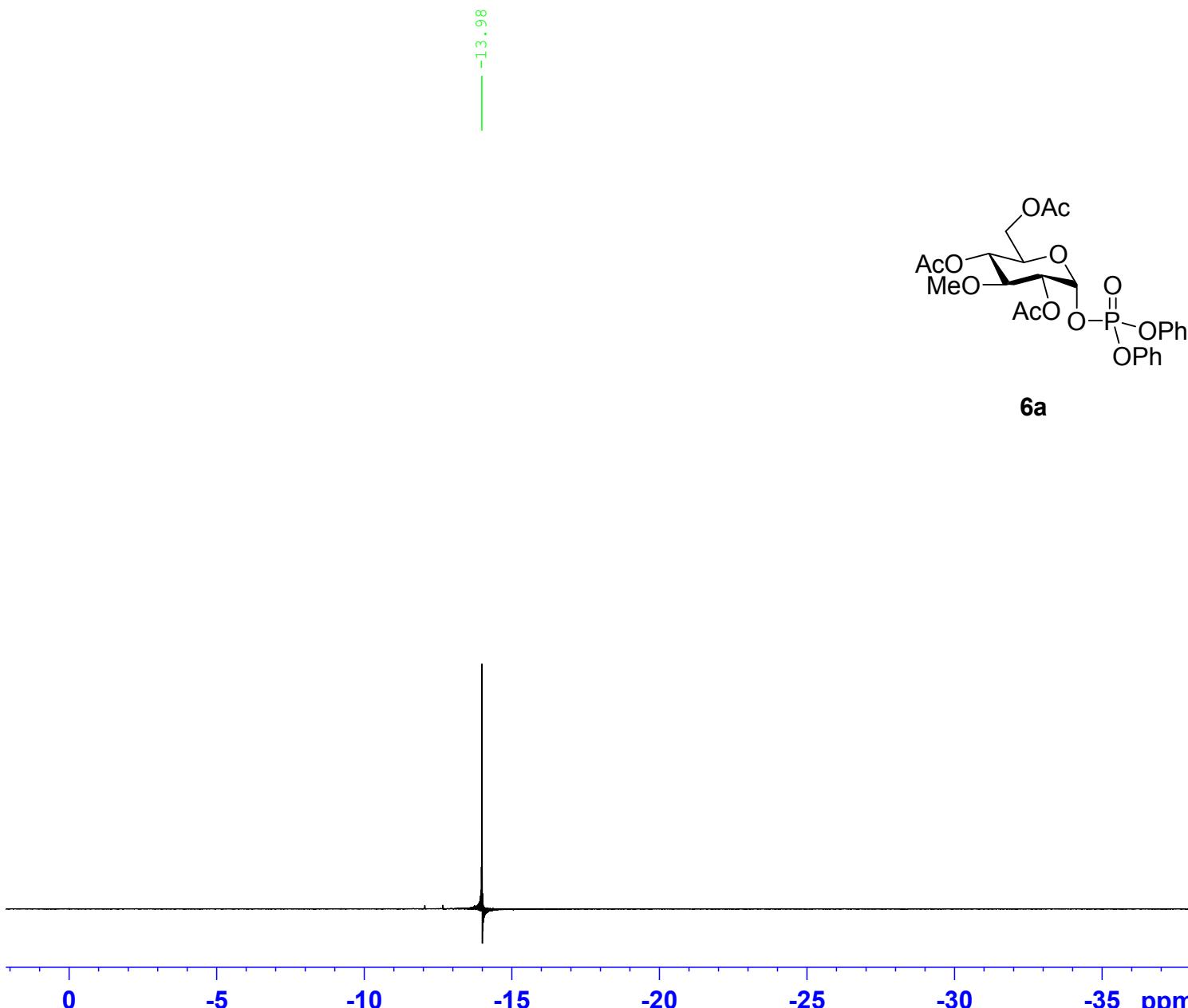


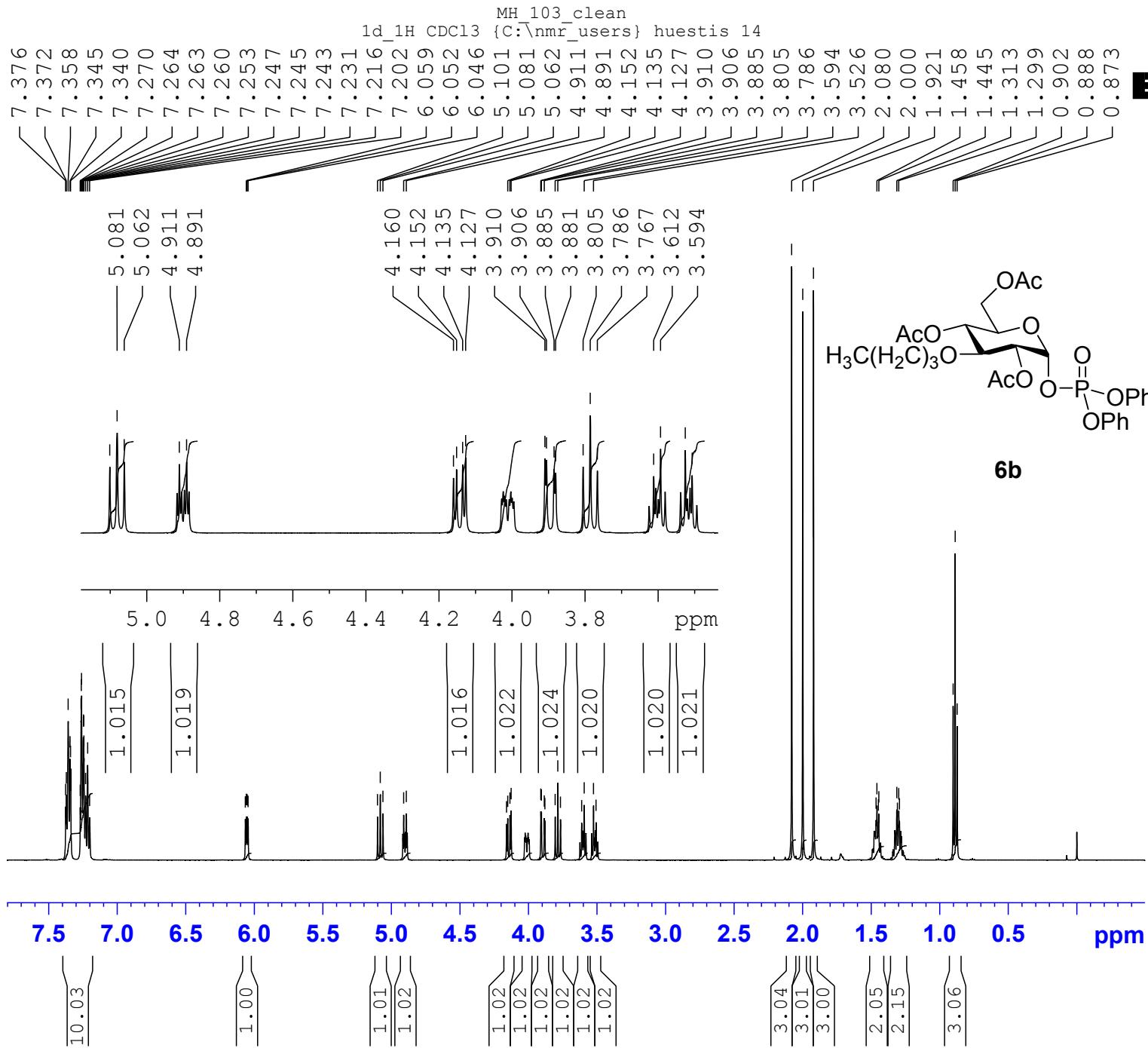
6a

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PL1 0.50 dB  
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===== CHANNEL f2 =====  
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NUC2 1H  
PCPD2 80.00 usec  
PL2 -2.00 dB  
PL12 15.72 dB  
PL13 19.37 dB  
SFO2 500.1322696 MHz

F2 - Processing parameters  
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GB 0  
PC 1.00





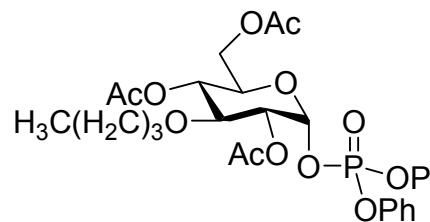
**BRUKER**

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PROCNO		1

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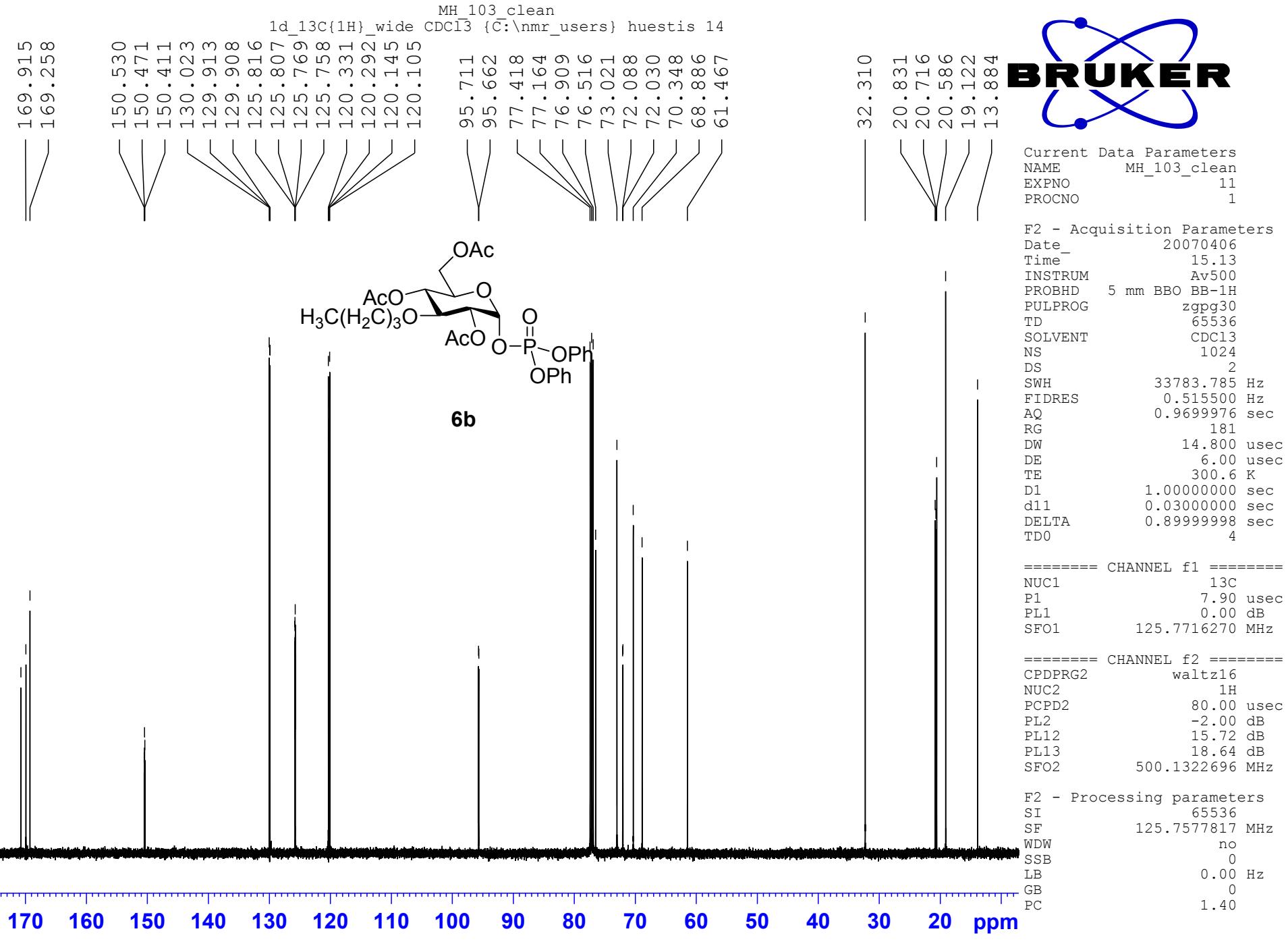
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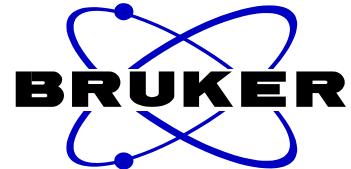
6b

```
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PL1          -2.00 dB  
SFO1      500.1332698 MHz
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F2 - Processing parameters  
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SF 500.1300326 MHz  
WDW no  
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PC 1.00



MH\_103\_clean  
1d\_31P{1H} CDC13 {C:\nmr\_users} huestis 14



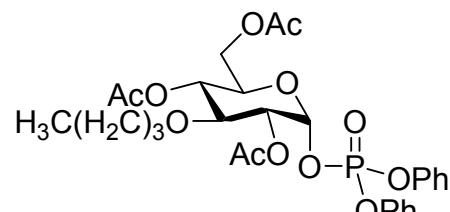
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SOLVENT CDCl3  
NS 512  
DS 0  
SWH 98039.219 Hz  
FIDRES 1.495960 Hz  
AQ 0.3342887 sec  
RG 8192  
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TE 300.4 K  
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d11 0.03000000 sec  
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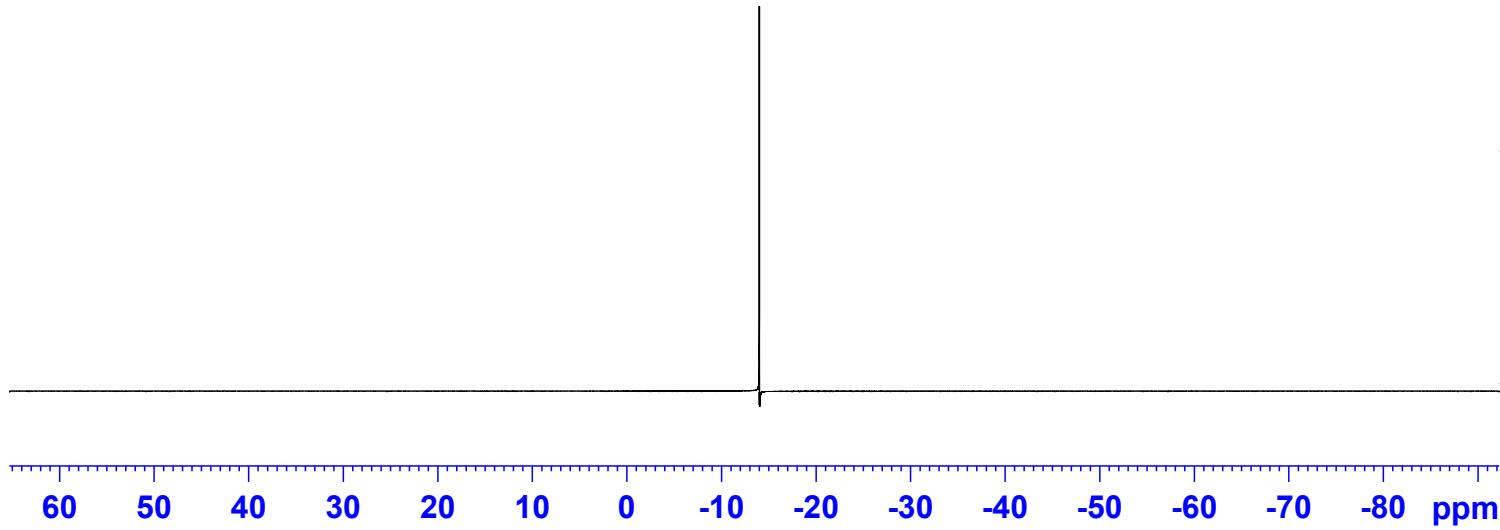
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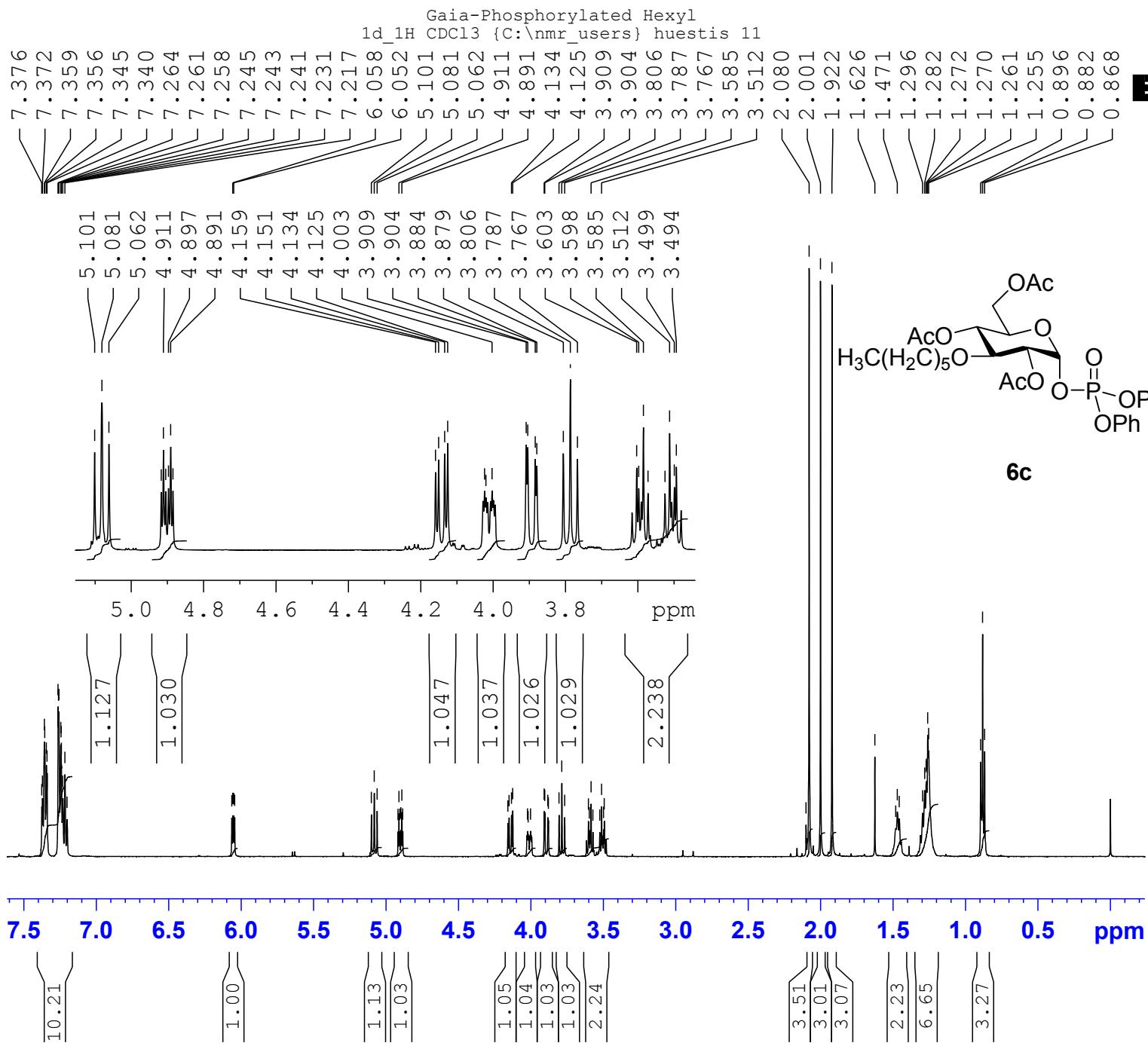
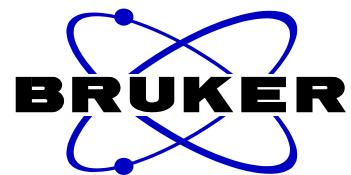
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PL12 15.72 dB  
PL13 18.64 dB  
SFO2 500.1322696 MHz

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SF 202.4563430 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 1.00



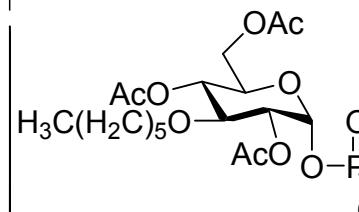
6b





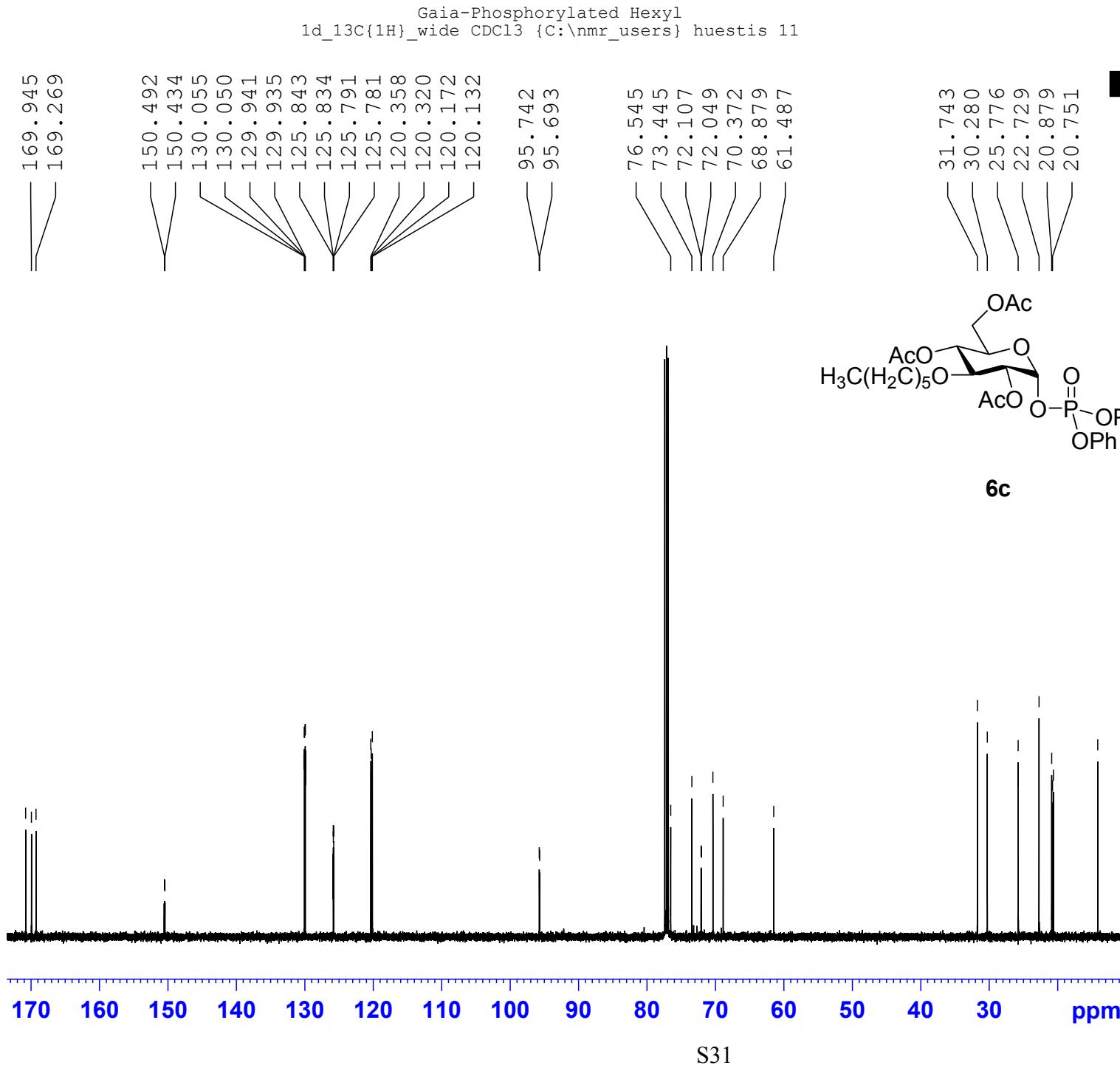
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PROCNO 1

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SOLVENT CDC13  
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DS 2  
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DW 50.000 usec  
DE 6.00 usec  
TE 300.0 K  
D1 1.0000000 sec  
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===== CHANNEL f1 =====  
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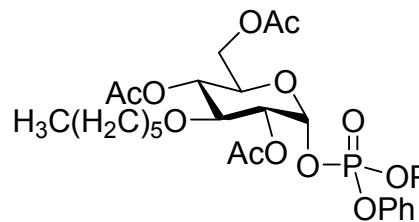
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GB 0  
PC 1.00



**BRUKER**

Current Data Parameters  
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 EXPNO 11  
 PROCNO 1

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 Date 20070529  
 Time 11.50  
 INSTRUM Av500  
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 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDC13  
 NS 1024  
 DS 2  
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 FIDRES 0.515500 Hz  
 AQ 0.9699976 sec  
 TE 574.7  
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 DELTA 0.89999998 sec  
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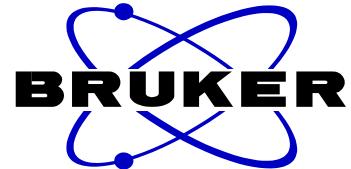


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 PL1 0.00 dB  
 SFO1 125.7716270 MHz

===== CHANNEL f2 ======  
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 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 -2.00 dB  
 PL12 15.72 dB  
 PL13 18.64 dB  
 SFO2 500.1322696 MHz

F2 - Processing parameters  
 SI 65536  
 SF 125.7577788 MHz  
 WDW no  
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 PC 1.40

Gaia-Phosphorylated Hexyl  
1d\_31P{1H} CDC13 {C:\nmr\_users} huestis 11



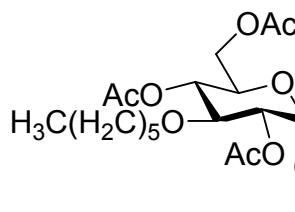
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FIDRES 1.495960 Hz  
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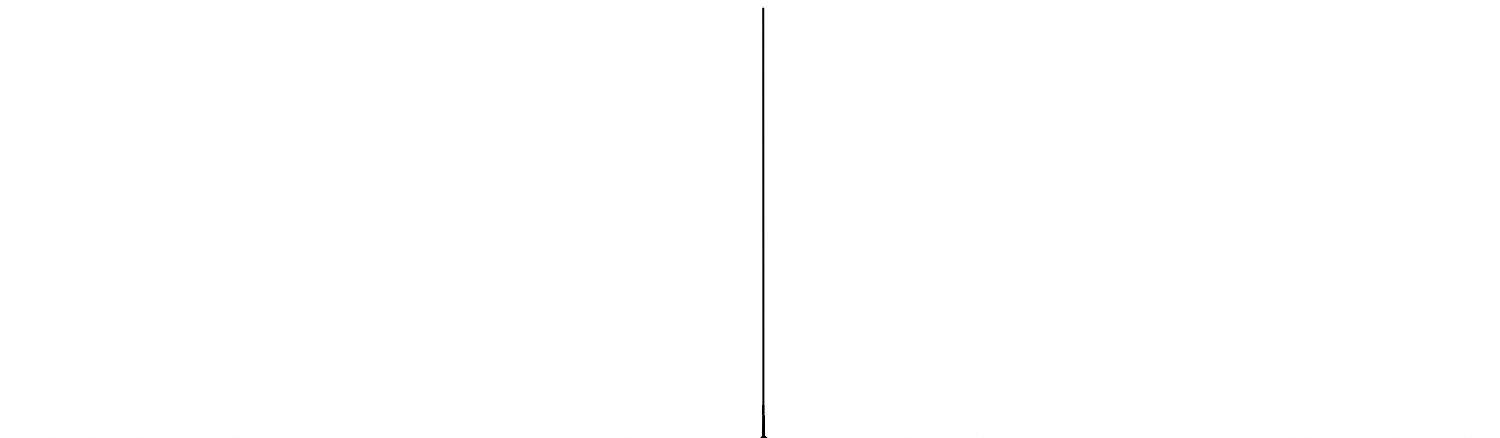
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SFO1 202.4664658 MHz

===== CHANNEL f2 =====  
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NUC2 1H  
PCPD2 80.00 usec  
PL2 -2.00 dB  
PL12 15.72 dB  
PL13 18.64 dB  
SFO2 500.1322696 MHz

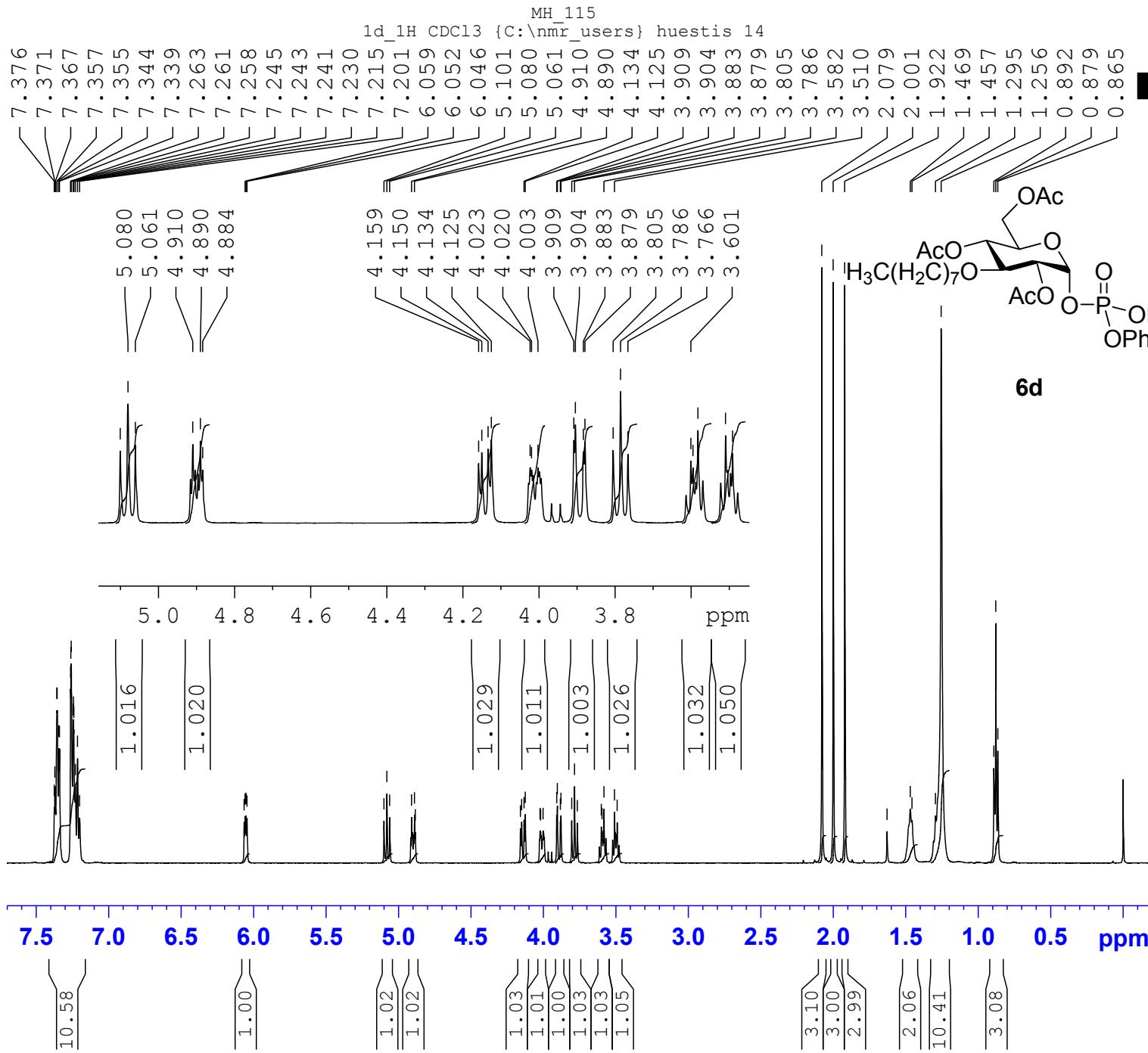
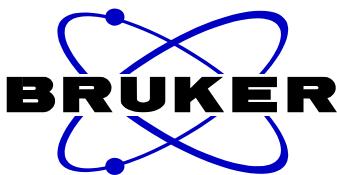
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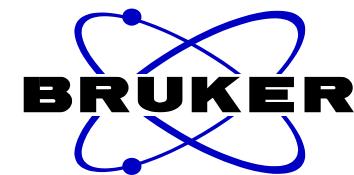
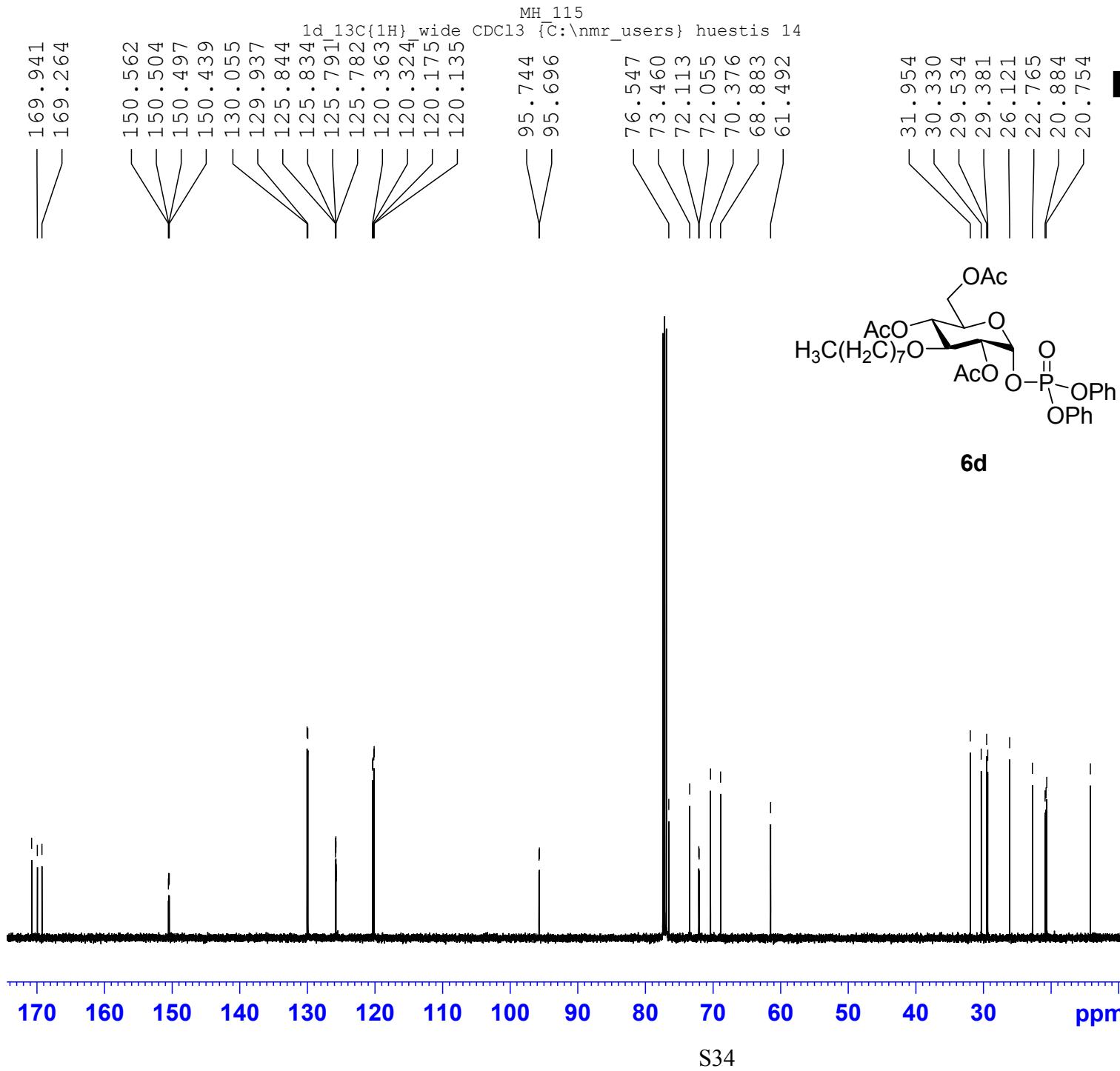


**6c**



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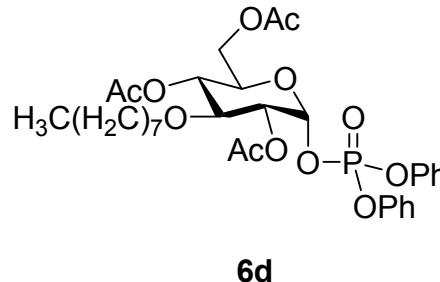
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 TD 65536  
 SOLVENT CDC13  
 NS 1024  
 DS 2  
 SWH 33783.785 Hz  
 FIDRES 0.515500 Hz  
 AQ 0.9699976 sec  
 RG 161.3  
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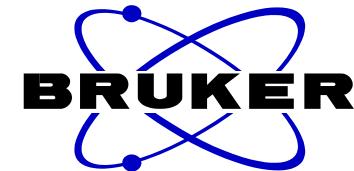
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 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 -2.00 dB  
 PL12 15.72 dB  
 PL13 18.64 dB  
 SFO2 500.1322696 MHz

F2 - Processing parameters  
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 SF 125.7577785 MHz  
 WDW no  
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 GB 0  
 PC 1.40



MH\_115  
1d\_31P{1H} CDC13 {C:\nmr\_users} huestis 14



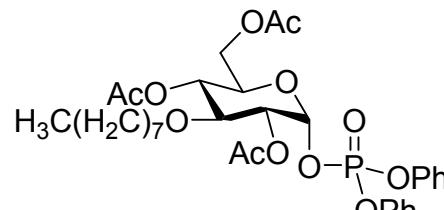
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NS 512  
DS 0  
SWH 98039.219 Hz  
FIDRES 1.495960 Hz  
AQ 0.3342887 sec  
RG 10321.3  
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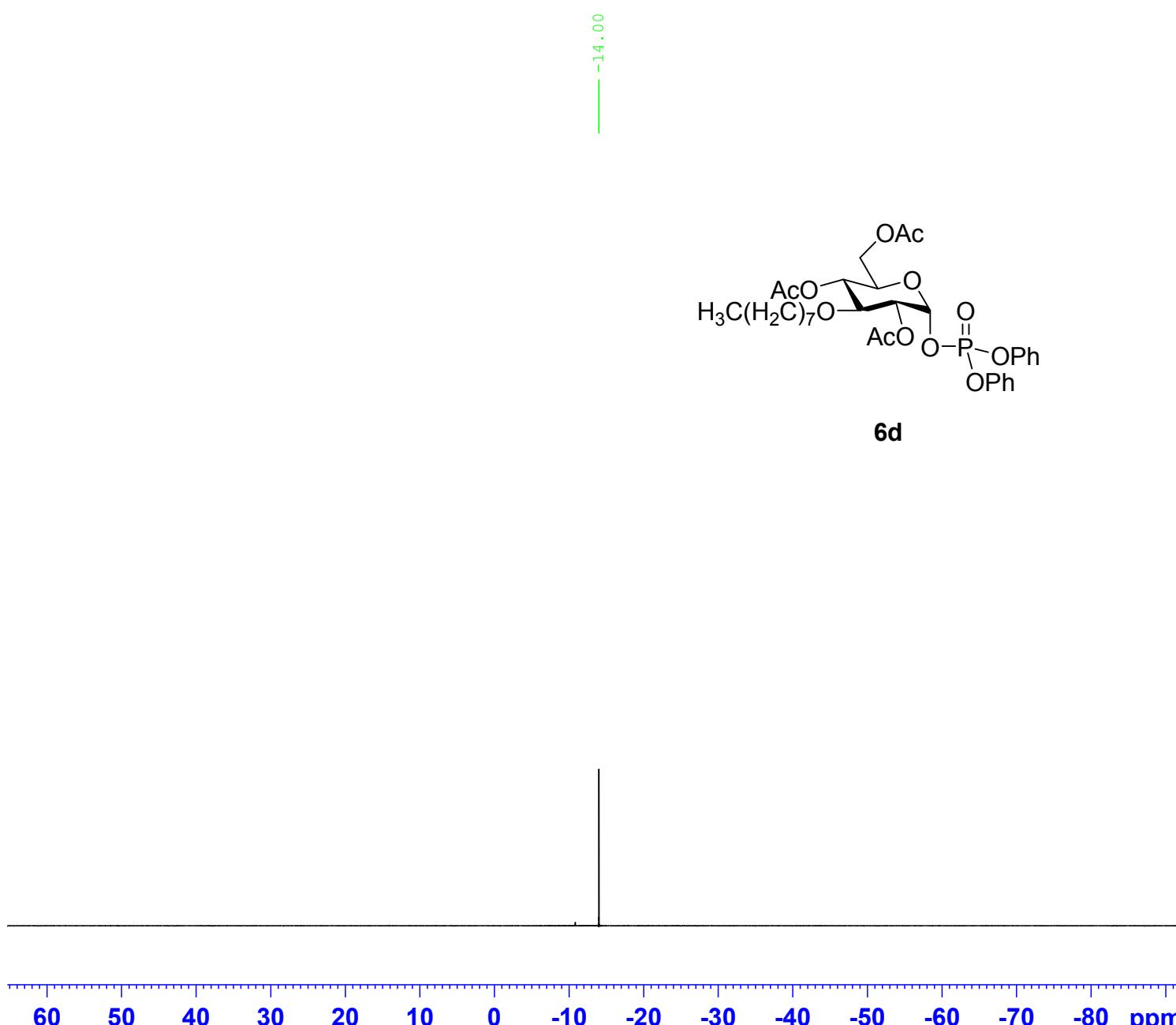
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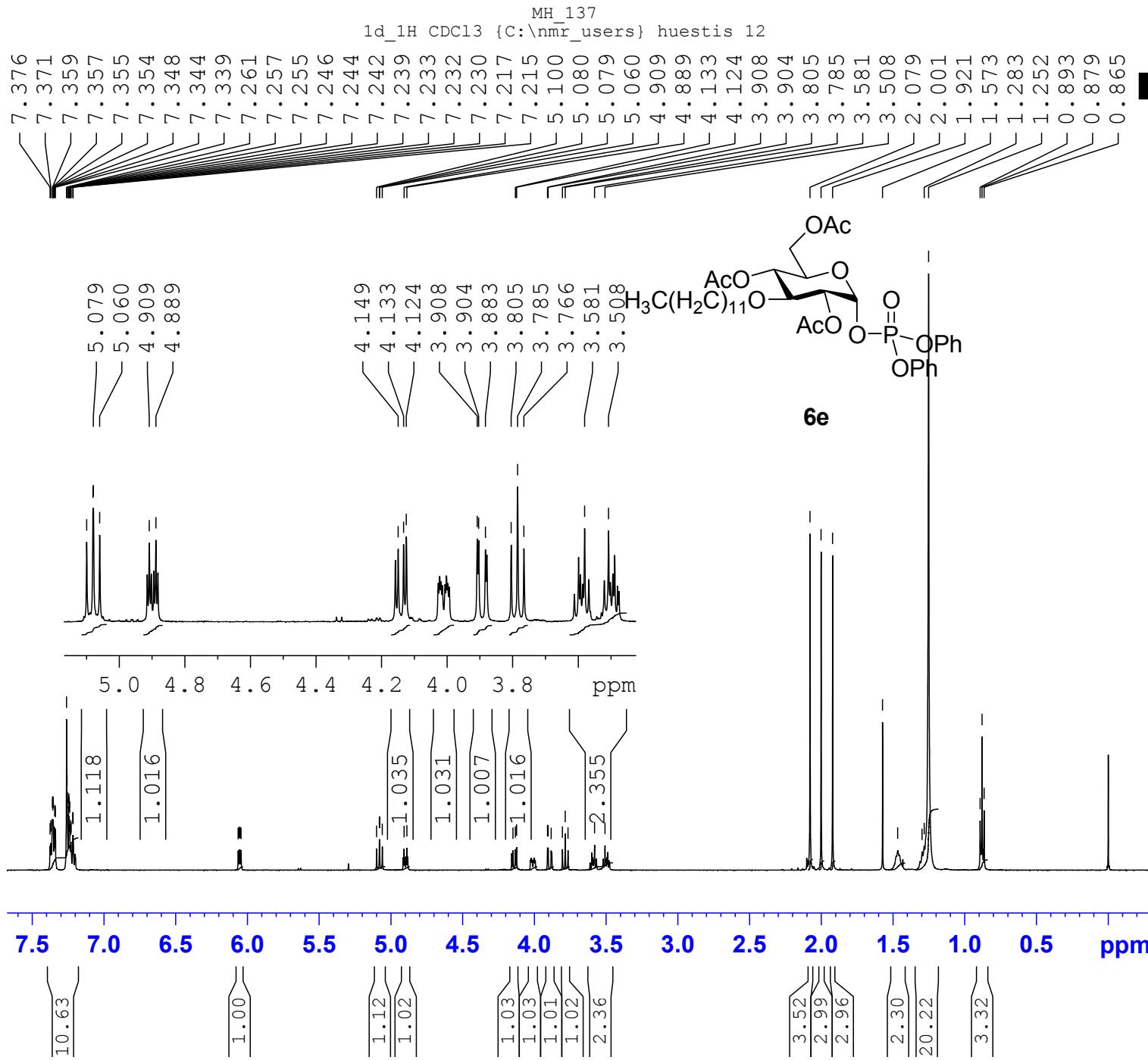
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PCPD2 80.00 usec  
PL2 -2.00 dB  
PL12 15.72 dB  
PL13 18.64 dB  
SFO2 500.1322696 MHz

F2 - Processing parameters  
SI 65536  
SF 202.4563430 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 1.00



6d







**BRUKER**

Current Data Parameters  
NAME MH\_137  
EXPNO 10  
PROCNO 1

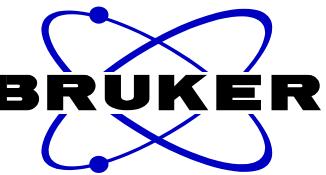
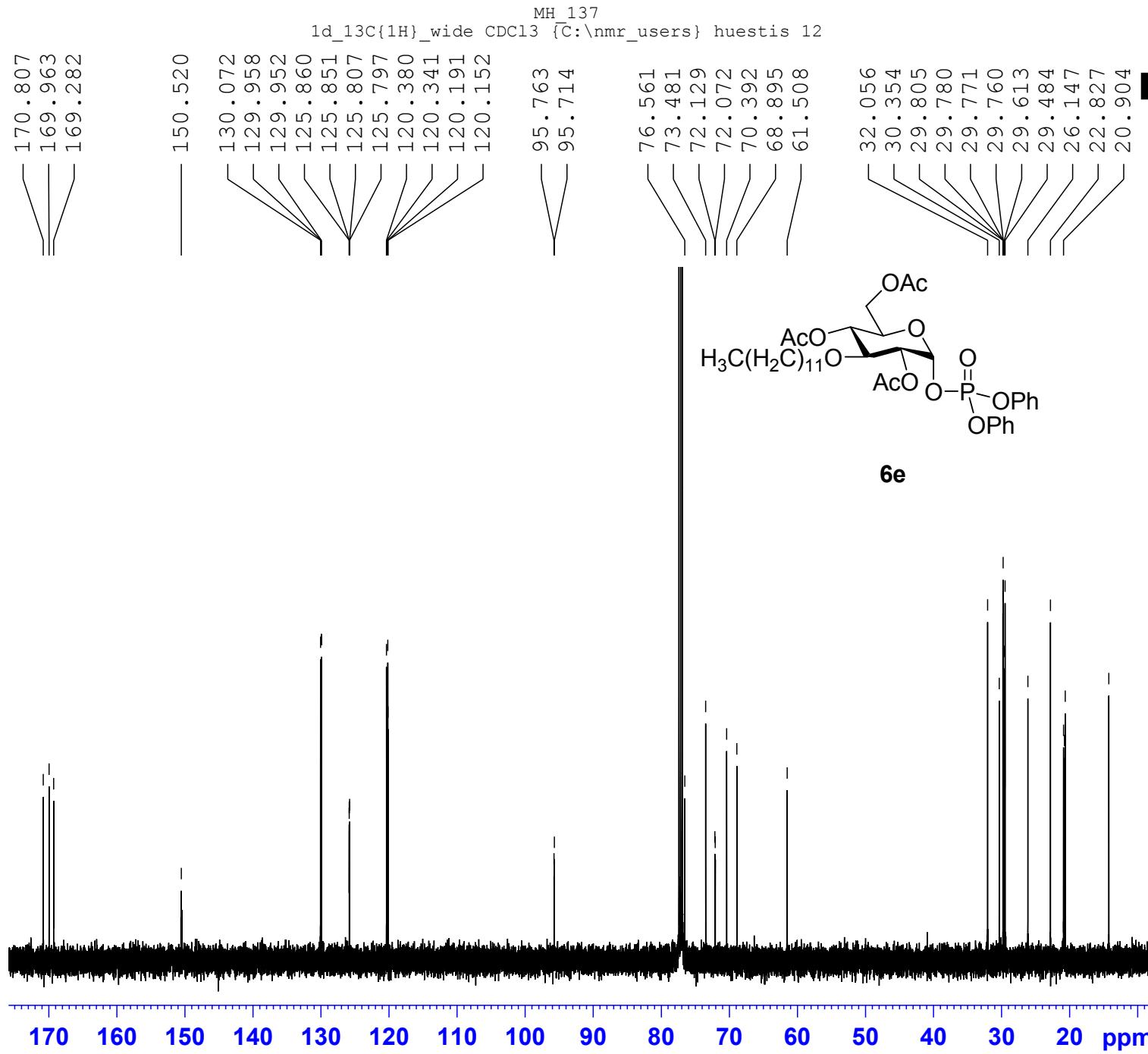
```

F2 - Acquisition Parameters
Date_           20070529
Time_          16.22
INSTRUM        Av500
PROBHD        5 mm BBO BB-1H
PULPROG       zg30
TD             65536
SOLVENT        CDCl3
NS              32
DS               2
SWH            10000.000 Hz
FIDRES        0.152588 Hz
AQ             3.2769001 sec
RG              228.1
DW             50.000 usec
DE              6.00 usec
TE              300.0 K
D1             1.00000000 sec
TD0                  1

```

===== CHANNEL f1 =====  
NUC1 1H  
P1 9.80 usec  
PL1 -2.00 dB  
SFO1 500.1332698 MHz

F2 - Processing parameters  
SI 65536  
SF 500.1300373 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 1.00



Current Data Parameters  
 NAME MH\_137  
 EXPNO 11  
 PROCNO 1

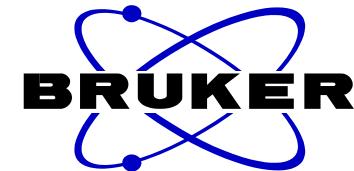
F2 - Acquisition Parameters  
 Date 20070529  
 Time 16.31  
 INSTRUM Av500  
 PROBHD 5 mm BBO BB-1H  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDC13  
 NS 1024  
 DS 2  
 SWH 33783.785 Hz  
 FIDRES 0.515500 Hz  
 AQ 0.9699976 sec  
 RG 1149.4  
 DW 14.800 usec  
 DE 6.00 usec  
 TE 300.6 K  
 D1 1.0000000 sec  
 d11 0.03000000 sec  
 DELTA 0.89999998 sec  
 TD0 4

===== CHANNEL f1 =====  
 NUC1 <sup>13</sup>C  
 P1 7.90 usec  
 PL1 0.00 dB  
 SFO1 125.7716270 MHz

===== CHANNEL f2 =====  
 CPDPRG2 waltz16  
 NUC2 <sup>1</sup>H  
 PCPD2 80.00 usec  
 PL2 -2.00 dB  
 PL12 15.72 dB  
 PL13 18.64 dB  
 SFO2 500.1322696 MHz

F2 - Processing parameters  
 SI 65536  
 SF 125.7577766 MHz  
 WDW no  
 SSB 0  
 LB 0.00 Hz  
 GB 0  
 PC 1.40

MH\_137  
1d\_31P{1H} CDC13 {C:\nmr\_users} huestis 12



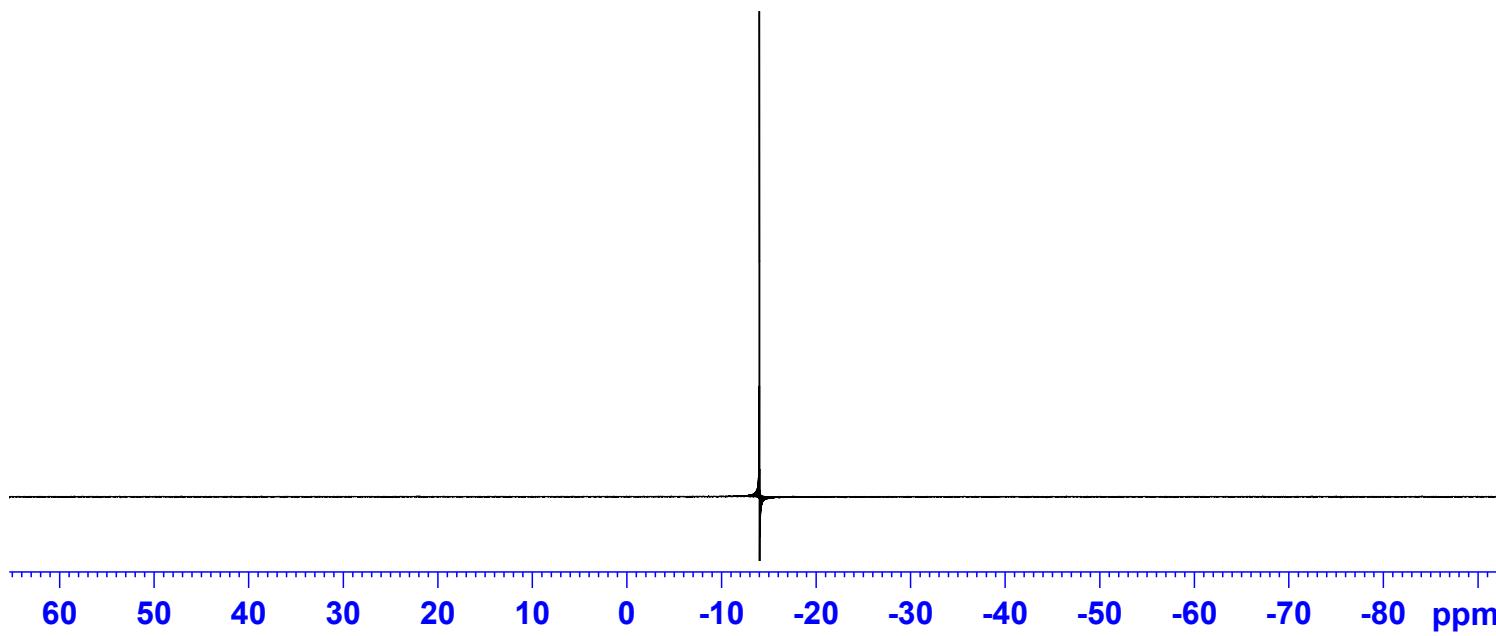
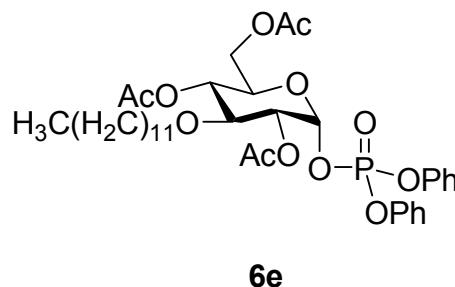
Current Data Parameters  
NAME MH\_137  
EXPNO 12  
PROCNO 1

F2 - Acquisition Parameters  
Date 20070529  
Time 17.05  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 512  
DS 0  
SWH 98039.219 Hz  
FIDRES 1.495960 Hz  
AQ 0.3342887 sec  
RG 7298.2  
DW 5.100 usec  
DE 6.00 usec  
TE 300.4 K  
D1 1.00000000 sec  
d11 0.03000000 sec  
DELTA 0.89999998 sec  
TD0 2

===== CHANNEL f1 =====  
NUC1 31P  
P1 8.41 usec  
PL1 0.50 dB  
SFO1 202.4664658 MHz

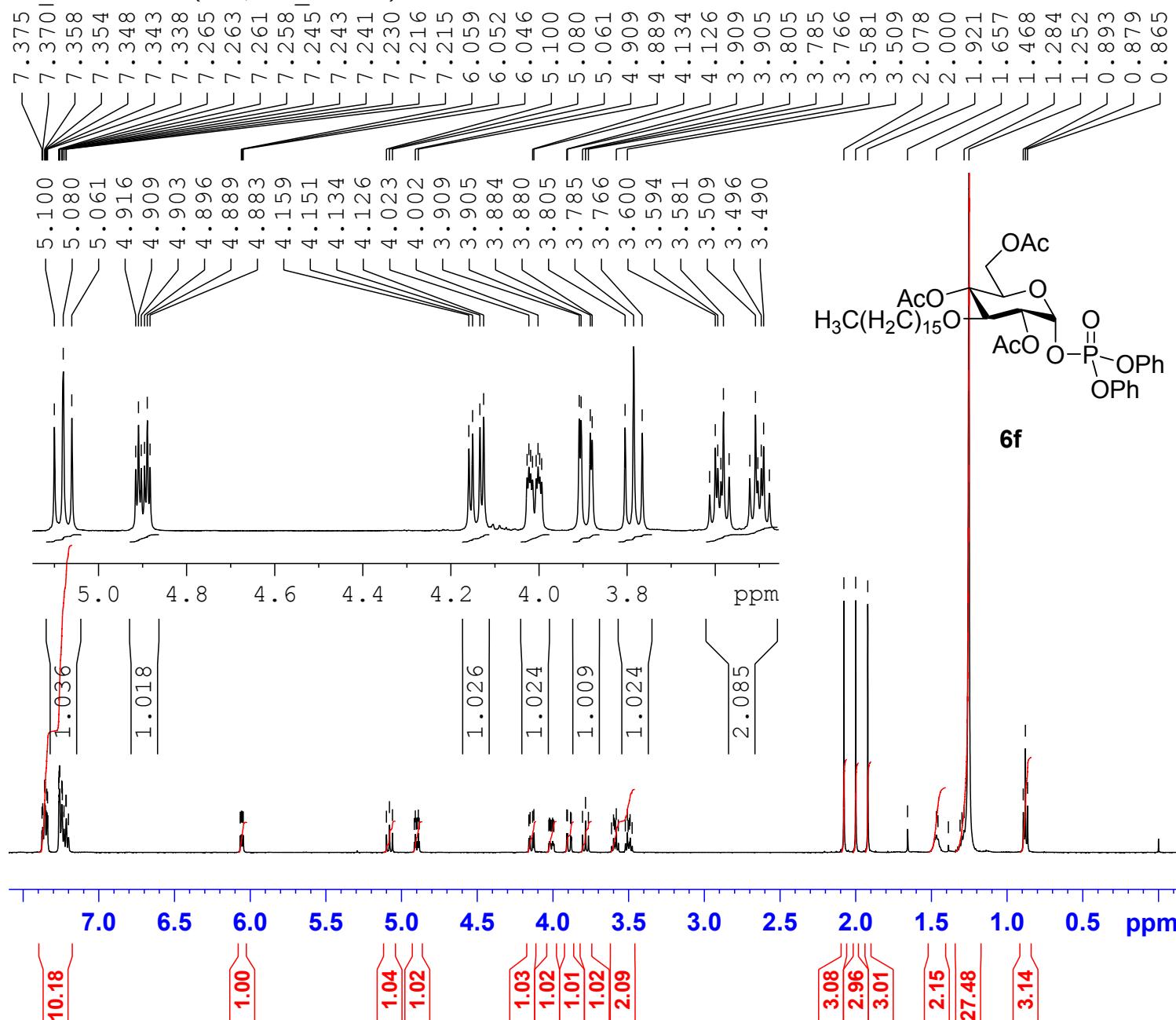
===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -2.00 dB  
PL12 15.72 dB  
PL13 18.64 dB  
SFO2 500.1322696 MHz

F2 - Processing parameters  
SI 65536  
SF 202.4563430 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 1.00



Gaia-hexadecyl phos\_2

1d\_1H CDC13 {C:\nmr\_users} huestis 27



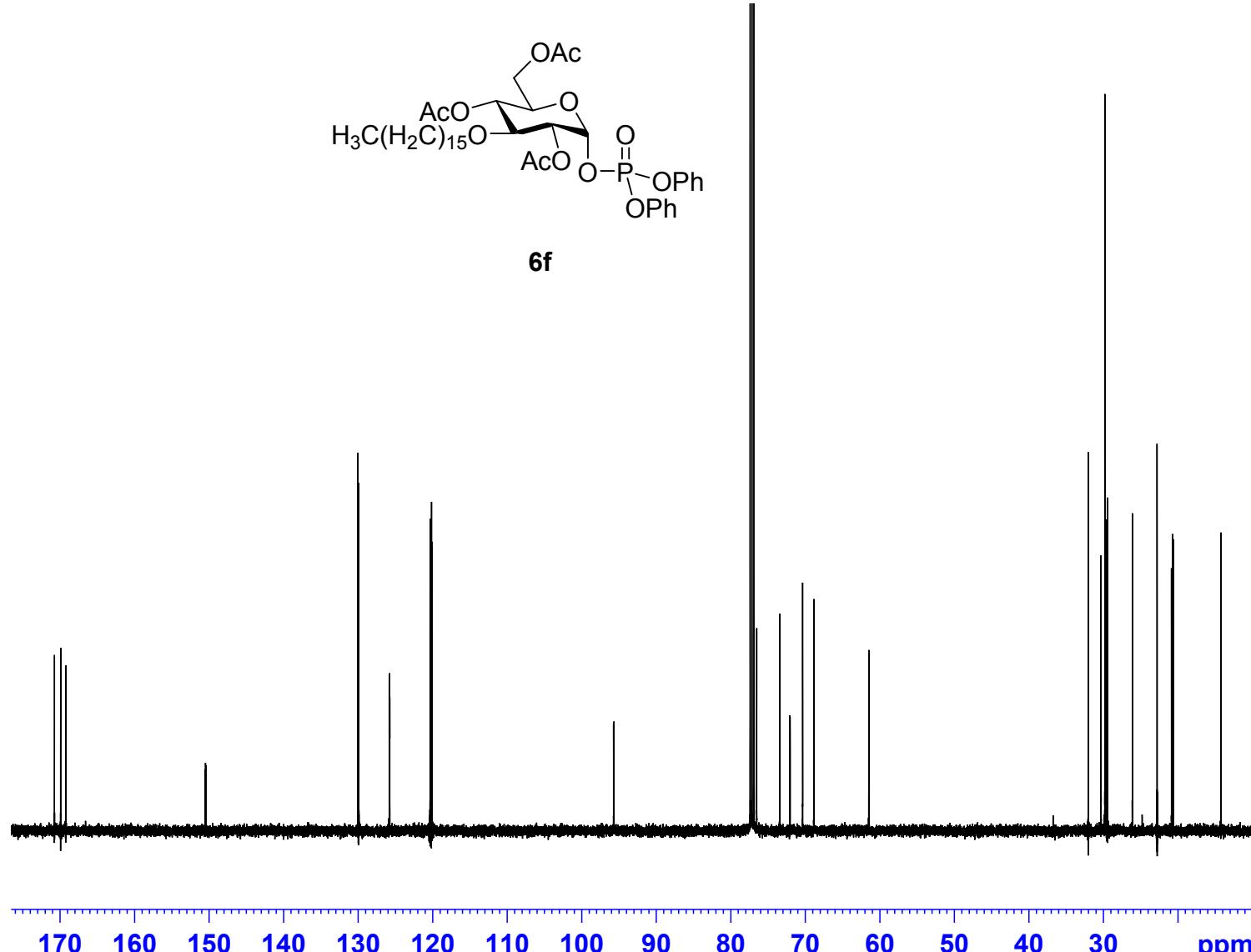
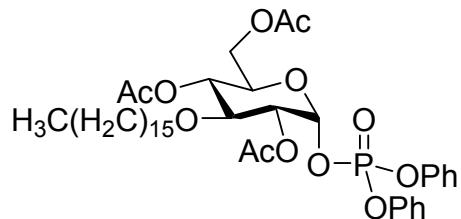
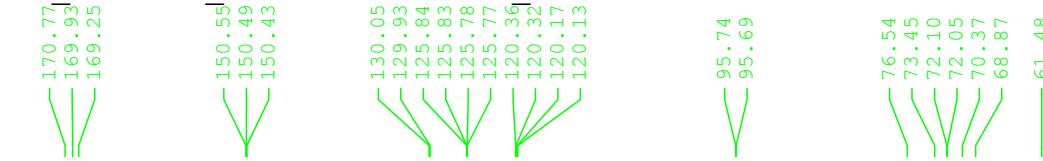
Current Data Parameters  
NAME Gaia-hexadecyl phos\_2  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date 20070704  
Time 10.19  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zg30  
TD 65536  
SOLVENT CDC13  
NS 32  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.152588 Hz  
AQ 3.2769001 sec  
RG 80.6  
DW 50.000 usec  
DE 6.00 usec  
TE 300.0 K  
D1 1.00000000 sec  
TD0 1

===== CHANNEL f1 =====  
NUC1 1H  
P1 9.80 usec  
PL1 -2.00 dB  
SFO1 500.1332698 MHz

F2 - Processing parameters  
SI 65536  
SF 500.1300353 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 0.75

Gaia-hexadecyl phos\_2  
1d\_13C{1H}\_d CDC13 {C:\nmr\_users} huestis 27



Current Data Parameters  
NAME Gaia-hexadecyl phos\_2  
EXPNO 11  
PROCNO 1

F2 - Acquisition Parameters  
Date 20070704  
Time 10.27  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 54048  
SOLVENT CDCl3  
NS 1024  
DS 4  
SWH 33783.785 Hz  
FIDRES 0.625070 Hz  
AQ 0.7999752 sec  
RG 10321.3  
DW 14.800 usec  
DE 21.14 usec  
TE 300.4 K  
D1 1.00000000 sec  
d11 0.03000000 sec  
DELTA 0.89999998 sec  
TDO 4

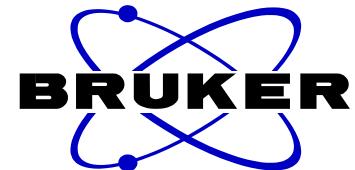
===== CHANNEL f1 ======  
NUC1 13C  
P1 7.90 usec  
PL1 0.00 dB  
SFO1 125.7716273 MHz

===== CHANNEL f2 ======  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -2.00 dB  
PL12 15.72 dB  
PL13 18.64 dB  
SFO2 500.1322696 MHz

F2 - Processing parameters  
SI 65536  
SF 125.7577794 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 0.75

170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 ppm

Gaia-hexadecyl phos\_2  
1d\_31P{1H}\_d CDC13 {C:\nmr\_users} huestis 27



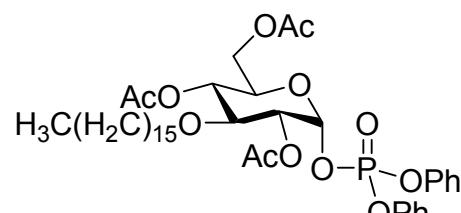
Current Data Parameters  
NAME Gaia-hexadecyl phos\_2  
EXPNO 14  
PROCNO 1

F2 - Acquisition Parameters  
Date 20070705  
Time 7.54  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT CDC13  
NS 512  
DS 4  
SWH 98039.219 Hz  
FIDRES 1.495960 Hz  
AQ 0.3342887 sec  
RG 9195.2  
DW 5.100 usec  
DE 7.29 usec  
TE 300.6 K  
D1 1.00000000 sec  
d11 0.03000000 sec  
DELTA 0.89999998 sec  
TDO 2

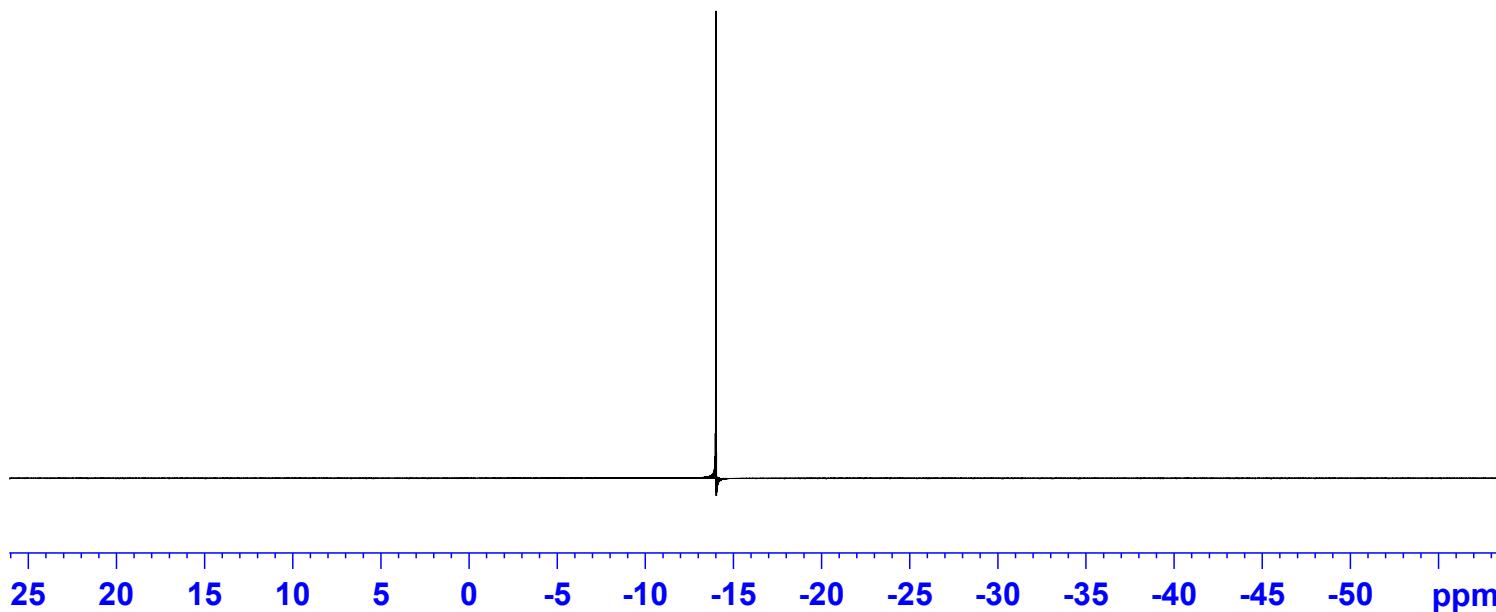
===== CHANNEL f1 =====  
NUC1 31P  
P1 8.41 usec  
PL1 0.50 dB  
SFO1 202.4664658 MHz

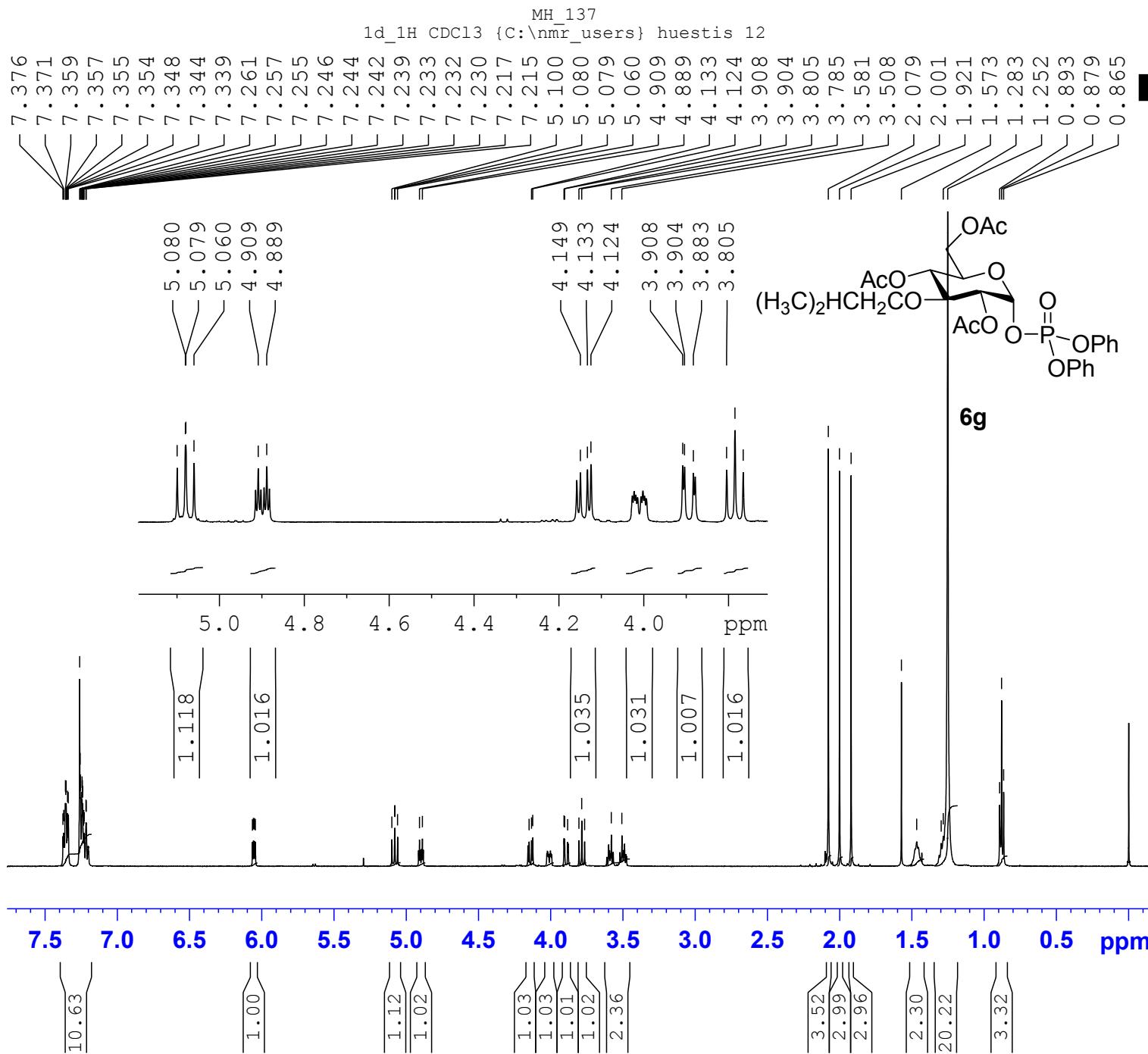
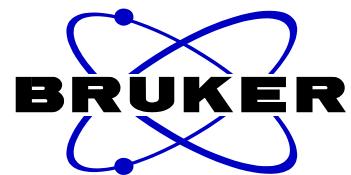
===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -2.00 dB  
PL12 15.72 dB  
PL13 18.64 dB  
SFO2 500.1322696 MHz

F2 - Processing parameters  
SI 65536  
SF 202.4563430 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 0.75



**6f**



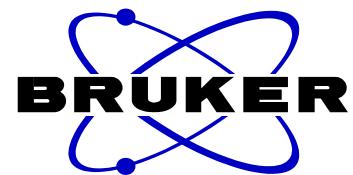
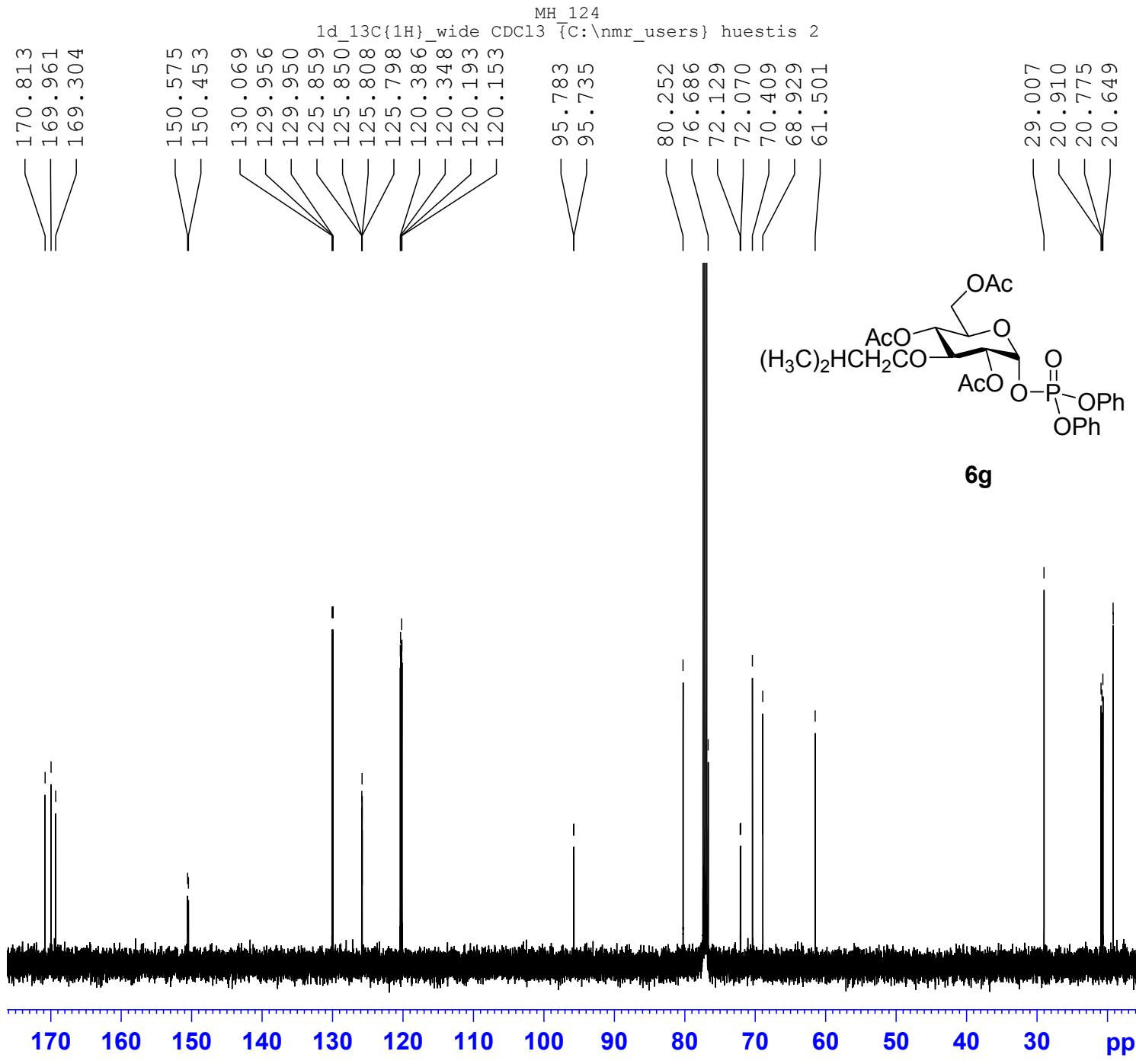


Current Data Parameters  
 NAME MH\_133  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters  
 Date 20070529  
 Time 16.22  
 INSTRUM Av500  
 PROBHD 5 mm BBO BB-1H  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 32  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.152588 Hz  
 AQ 3.2769001 sec  
 RG 228.1  
 DW 50.000 usec  
 DE 6.00 usec  
 TE 300.0 K  
 D1 1.00000000 sec  
 TD0 1

===== CHANNEL f1 ======  
 NUC1 1H  
 P1 9.80 usec  
 PL1 -2.00 dB  
 SFO1 500.1332698 MHz

F2 - Processing parameters  
 SI 65536  
 SF 500.1300373 MHz  
 WDW no  
 SSB 0  
 LB 0.00 Hz  
 GB 0  
 PC 1.00



Current Data Parameters  
 NAME MH\_124  
 EXPNO 11  
 PROCNO 1

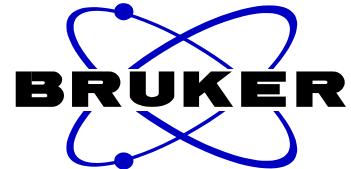
F2 - Acquisition Parameters  
 Date 20070429  
 Time 16.13  
 INSTRUM Av500  
 PROBHD 5 mm BBO BB-1H  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT CDCl3  
 NS 1024  
 DS 2  
 SWH 33783.785 Hz  
 FIDRES 0.515500 Hz  
 AQ 0.9699976 sec  
 RG 143.7  
 DW 14.800 usec  
 DE 6.00 usec  
 TE 300.6 K  
 D1 1.0000000 sec  
 d11 0.03000000 sec  
 DELTA 0.89999998 sec  
 TD0 4

===== CHANNEL f1 ======  
 NUC1 13C  
 P1 7.90 usec  
 PL1 0.00 dB  
 SFO1 125.7716270 MHz

===== CHANNEL f2 ======  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 -2.00 dB  
 PL12 15.72 dB  
 PL13 19.37 dB  
 SFO2 500.1322696 MHz

F2 - Processing parameters  
 SI 65536  
 SF 125.7577769 MHz  
 WDW no  
 SSB 0  
 LB 0.00 Hz  
 GB 0  
 PC 1.40

MH\_124  
1d\_31P{1H} CDC13 {C:\nmr\_users} huestis 2



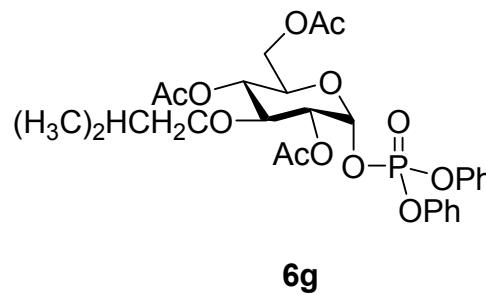
Current Data Parameters  
NAME MH\_124  
EXPNO 12  
PROCNO 1

F2 - Acquisition Parameters  
Date 20070429  
Time 16.47  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 512  
DS 0  
SWH 98039.219 Hz  
FIDRES 1.495960 Hz  
AQ 0.3342887 sec  
RG 7298.2  
DW 5.100 usec  
DE 6.00 usec  
TE 300.4 K  
D1 1.0000000 sec  
d11 0.0300000 sec  
DELTA 0.8999998 sec  
TD0 2

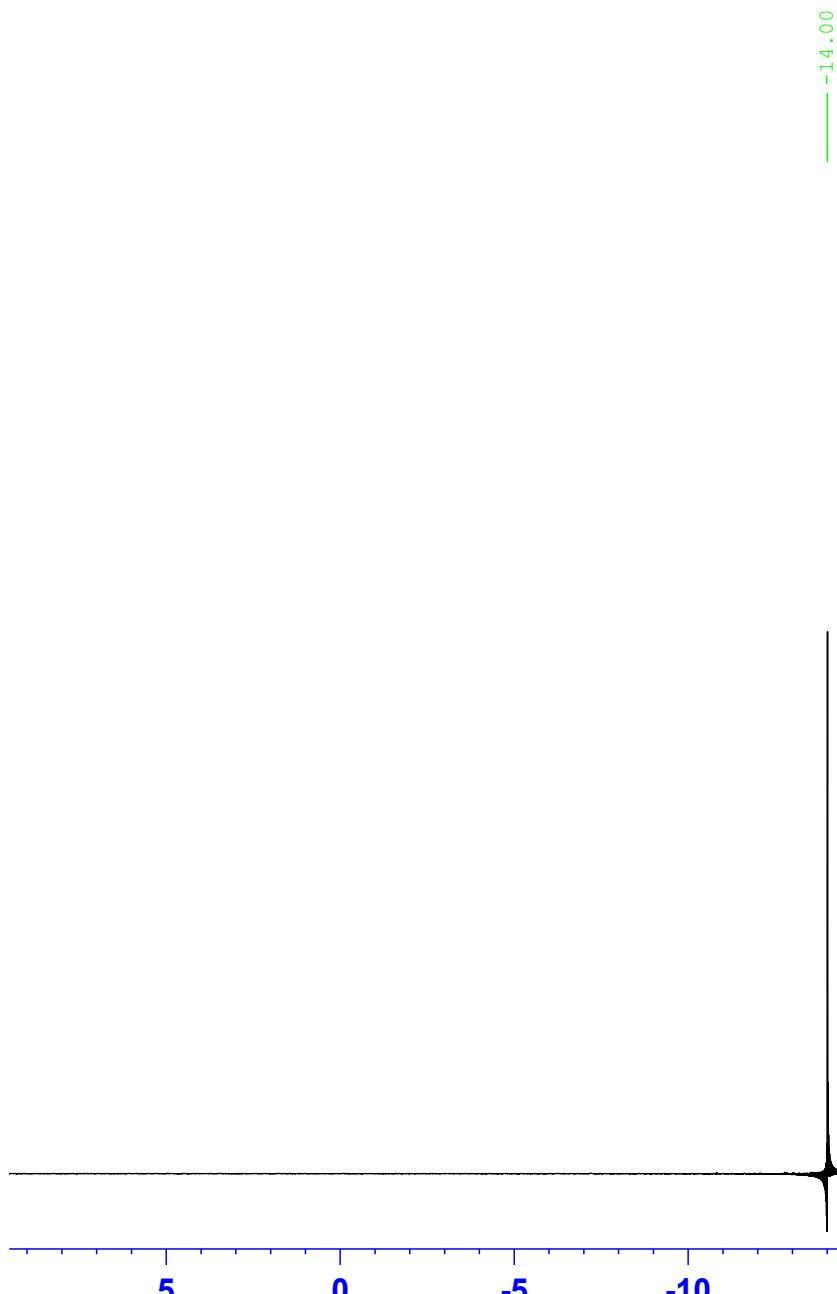
===== CHANNEL f1 =====  
NUC1 31P  
P1 8.41 usec  
PL1 0.50 dB  
SFO1 202.4664658 MHz

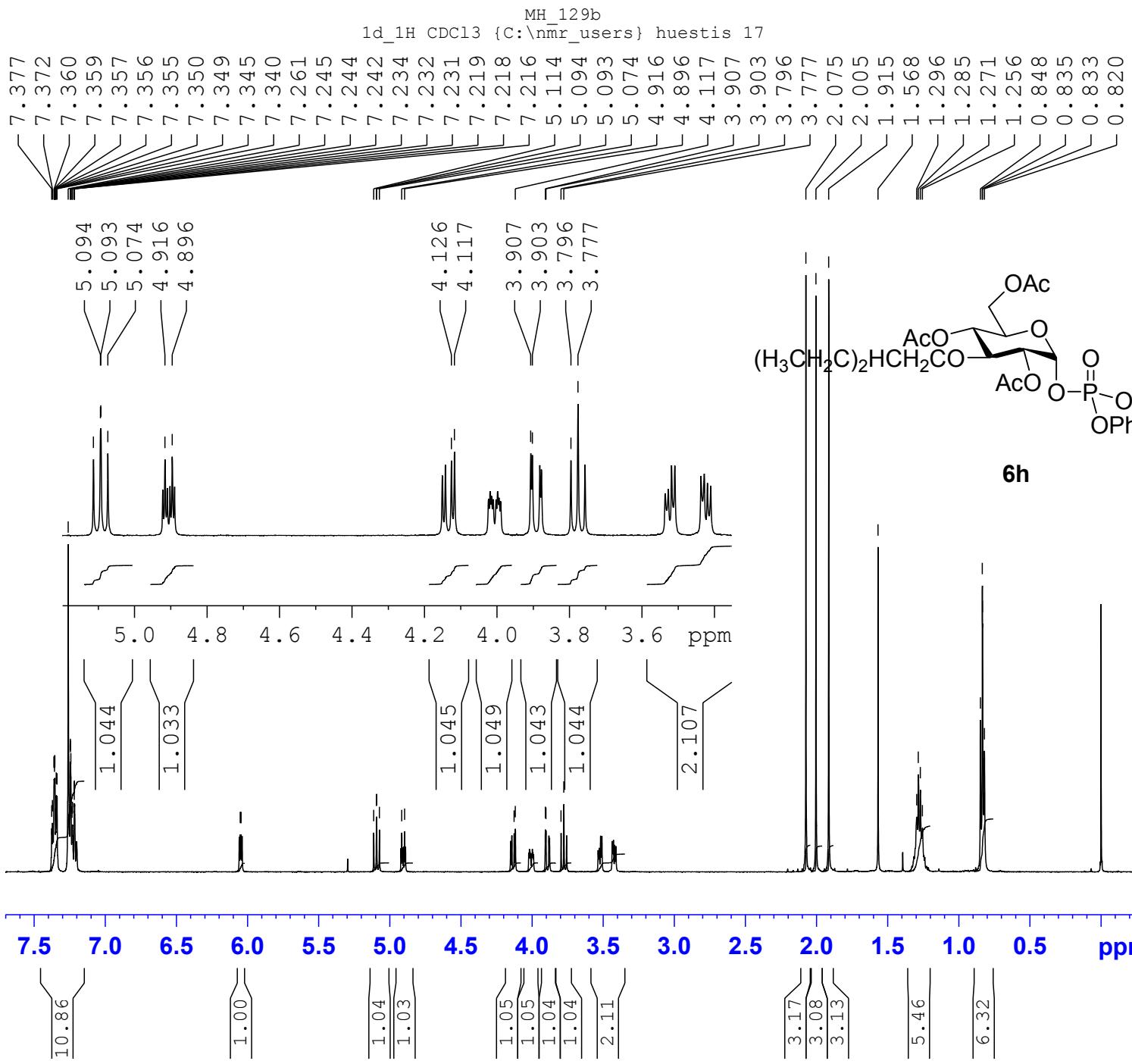
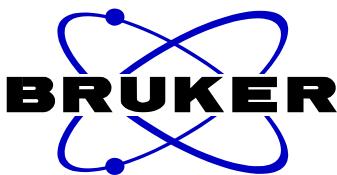
===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -2.00 dB  
PL12 15.72 dB  
PL13 19.37 dB  
SFO2 500.1322696 MHz

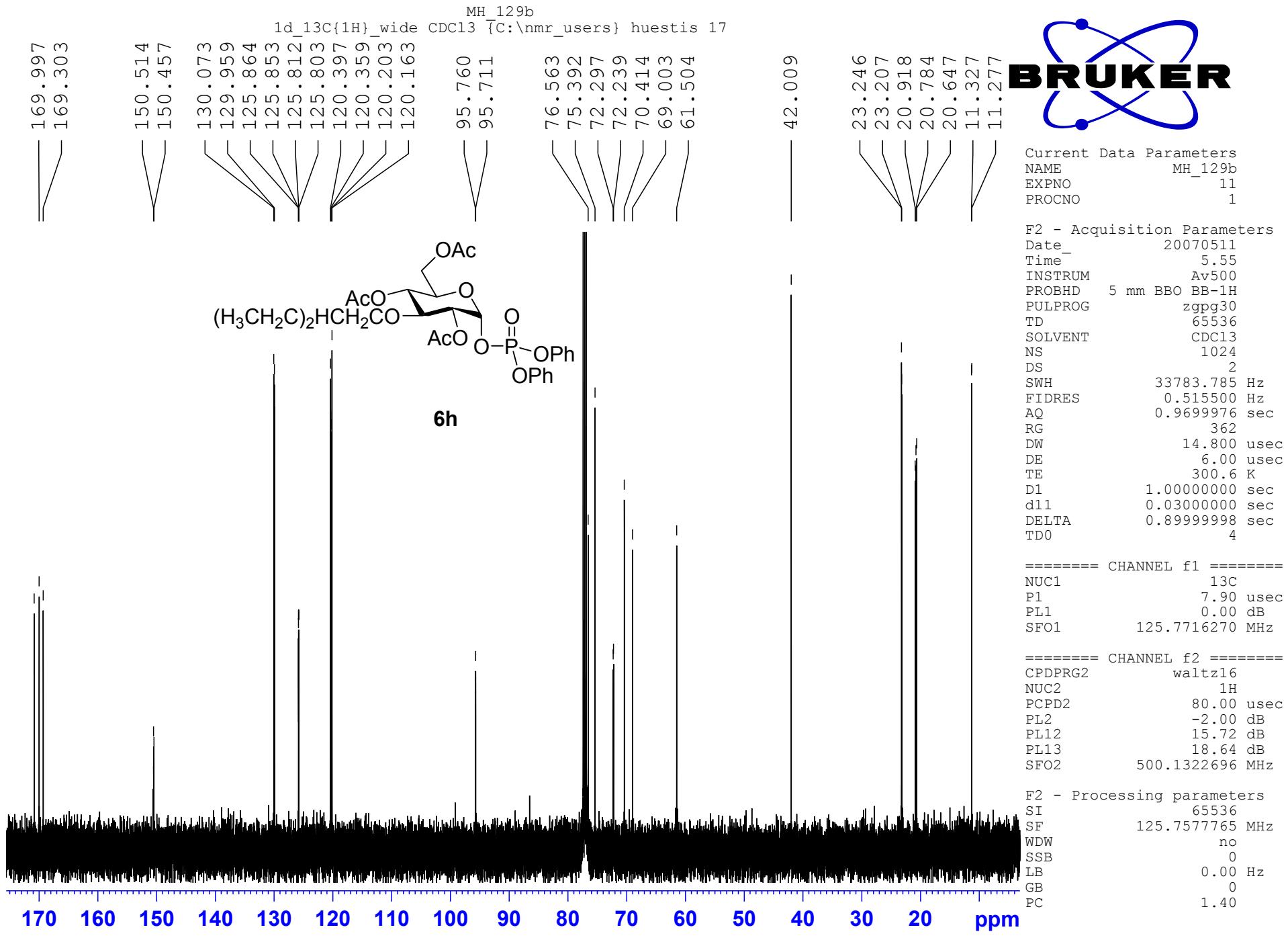
F2 - Processing parameters  
SI 65536  
SF 202.4563430 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 1.00



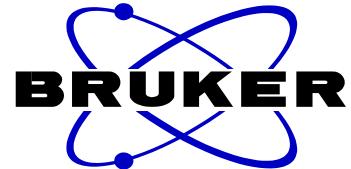
**6g**







MH\_129b  
1d\_31P{1H} CDC13 {C:\nmr\_users} huestis 17



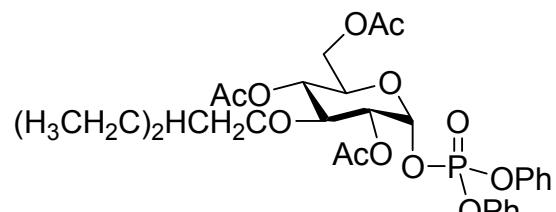
Current Data Parameters  
NAME MH\_129b  
EXPNO 12  
PROCNO 1

F2 - Acquisition Parameters  
Date 20070511  
Time 6.28  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT CDCl3  
NS 512  
DS 0  
SWH 98039.219 Hz  
FIDRES 1.495960 Hz  
AQ 0.3342887 sec  
RG 6502  
DW 5.100 usec  
DE 6.00 usec  
TE 300.6 K  
D1 1.00000000 sec  
d11 0.03000000 sec  
DELTA 0.89999998 sec  
TD0 2

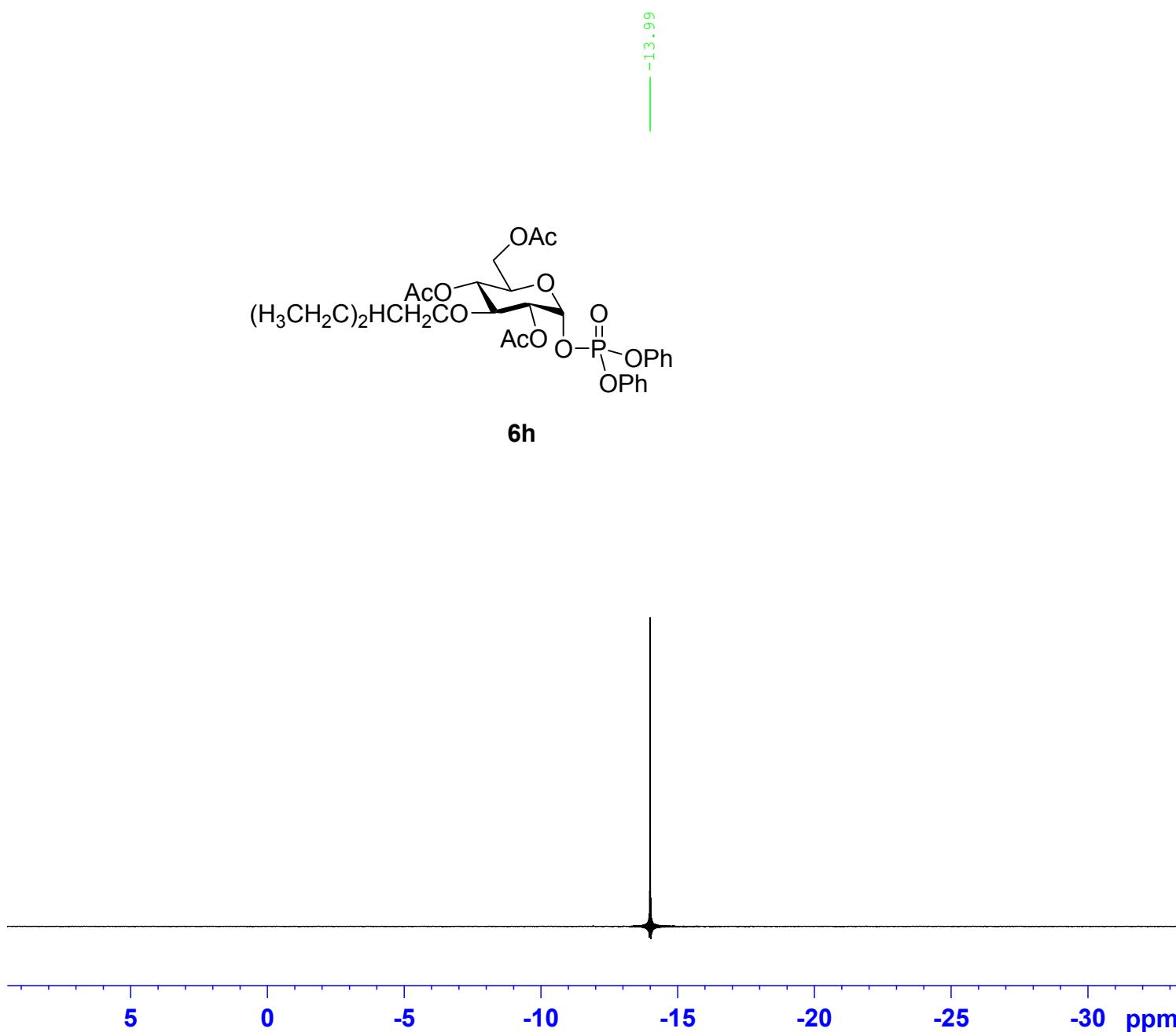
===== CHANNEL f1 =====  
NUC1 31P  
P1 8.41 usec  
PL1 0.50 dB  
SFO1 202.4664658 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -2.00 dB  
PL12 15.72 dB  
PL13 18.64 dB  
SFO2 500.1322696 MHz

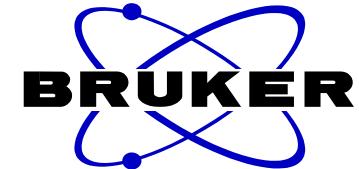
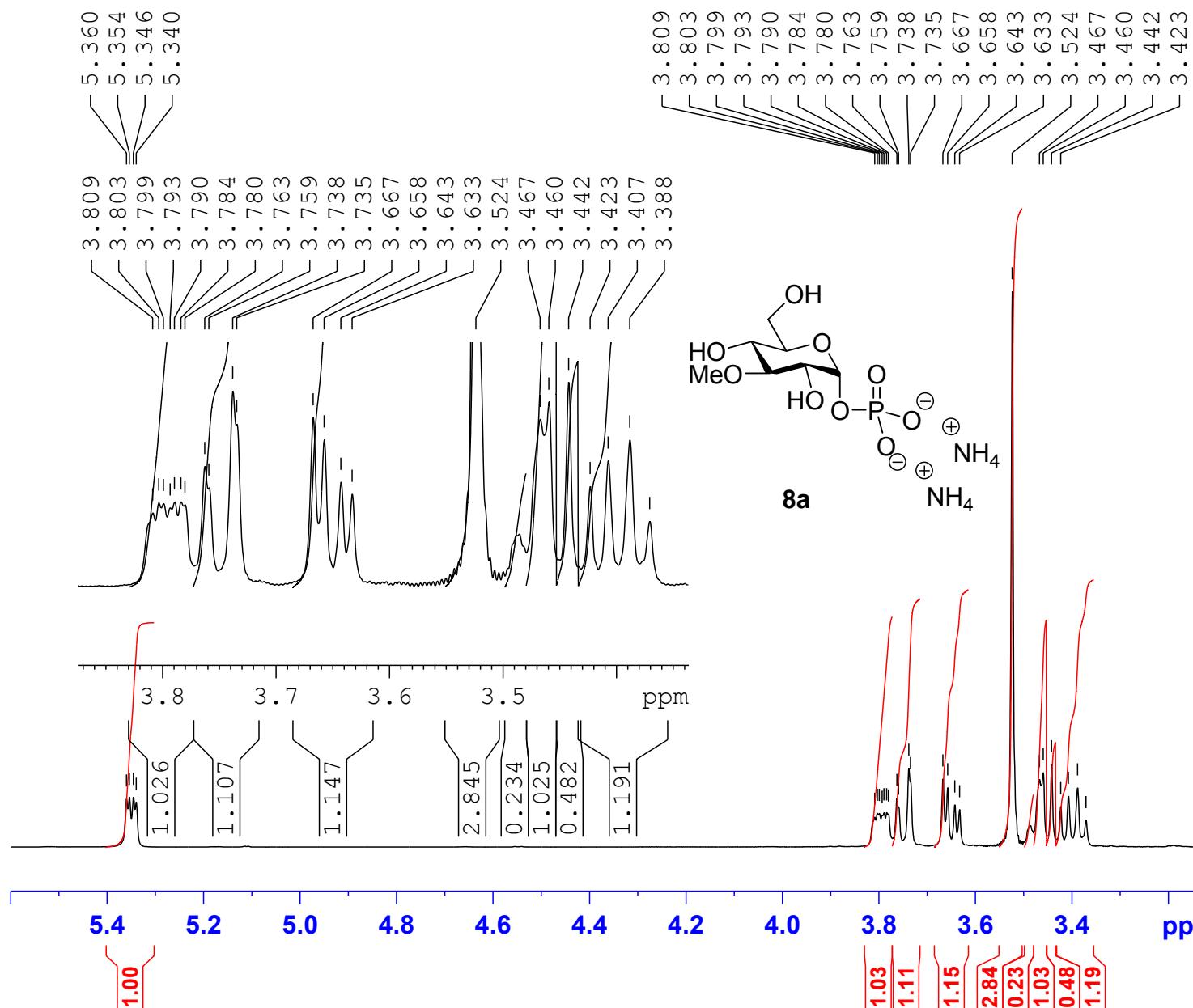
F2 - Processing parameters  
SI 65536  
SF 202.4563430 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 1.00



6h



Clean NH<sub>4</sub> OMe Salt  
1d\_water D2O {C:\nmr\_users} huestis 12



Current Data Parameters  
NAME Clean NH<sub>4</sub> OMe Salt  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20070330  
Time 7.37  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zgcpgrppr  
TD 8192  
SOLVENT D2O  
NS 32  
DS 2  
SWH 7507.507 Hz  
FIDRES 0.916444 Hz  
AQ 0.5457038 sec  
RG 161.3  
DW 66.600 usec  
DE 6.00 usec  
TE 305.0 K  
D1 2.00000000 sec  
d12 0.00002000 sec  
D16 0.00020000 sec  
TD0 1

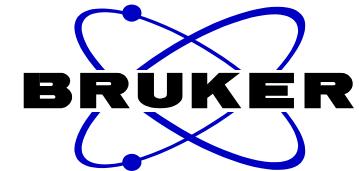
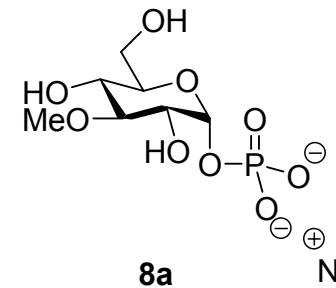
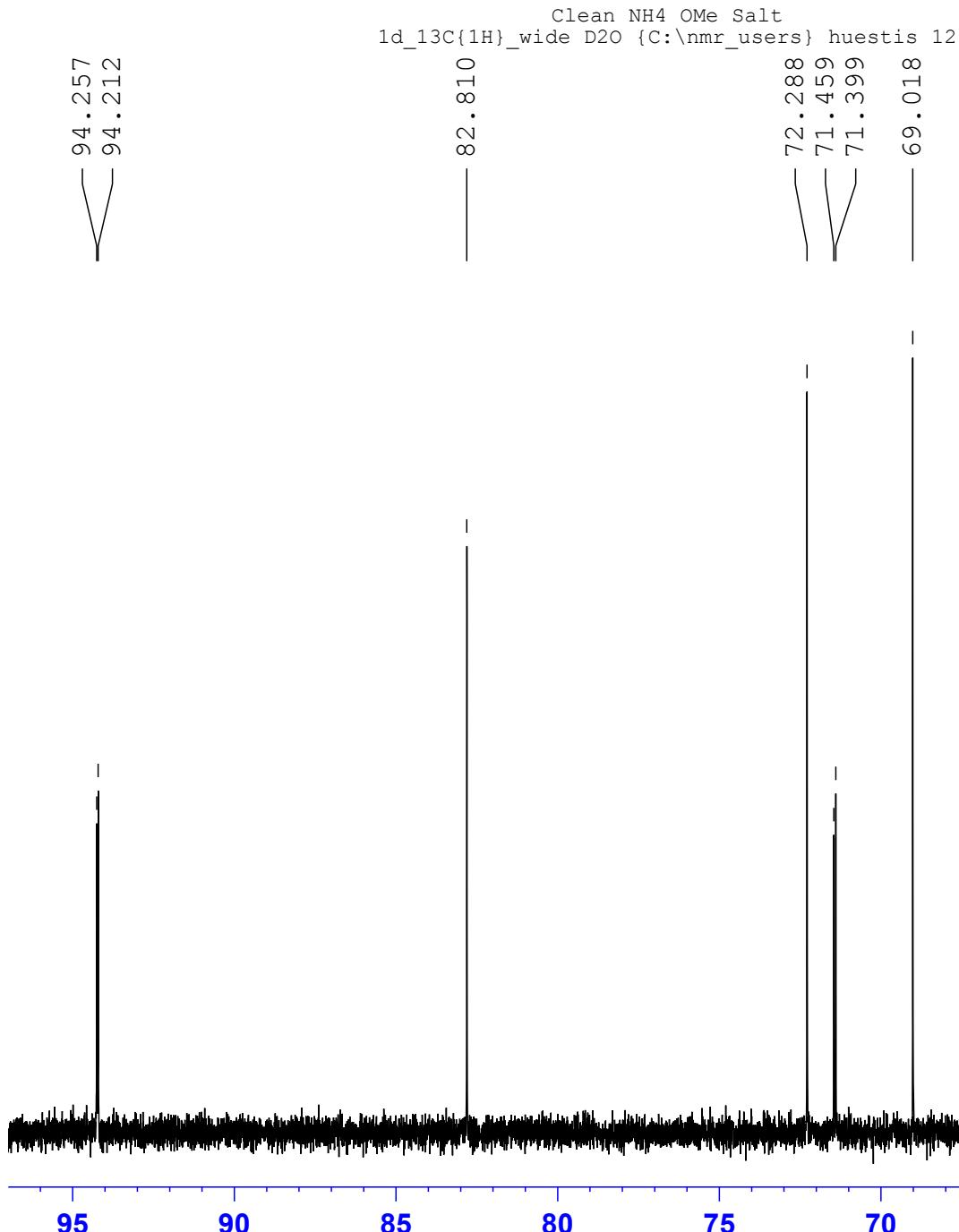
===== CHANNEL f1 ======

NUC1	1H
P1	11.10 usec
PL1	-1.40 dB
PL9	51.67 dB
SFO1	500.1323692 MHz

===== GRADIENT CHANNEL =====

GPNAM1	sine.100
GPZ1	50.00 %
P16	1000.00 usec

F2 - Processing parameters  
SI 32768  
SF 500.1300390 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
 NAME Clean NH<sub>4</sub> OMe Salt  
 EXPNO 11  
 PROCNO 1

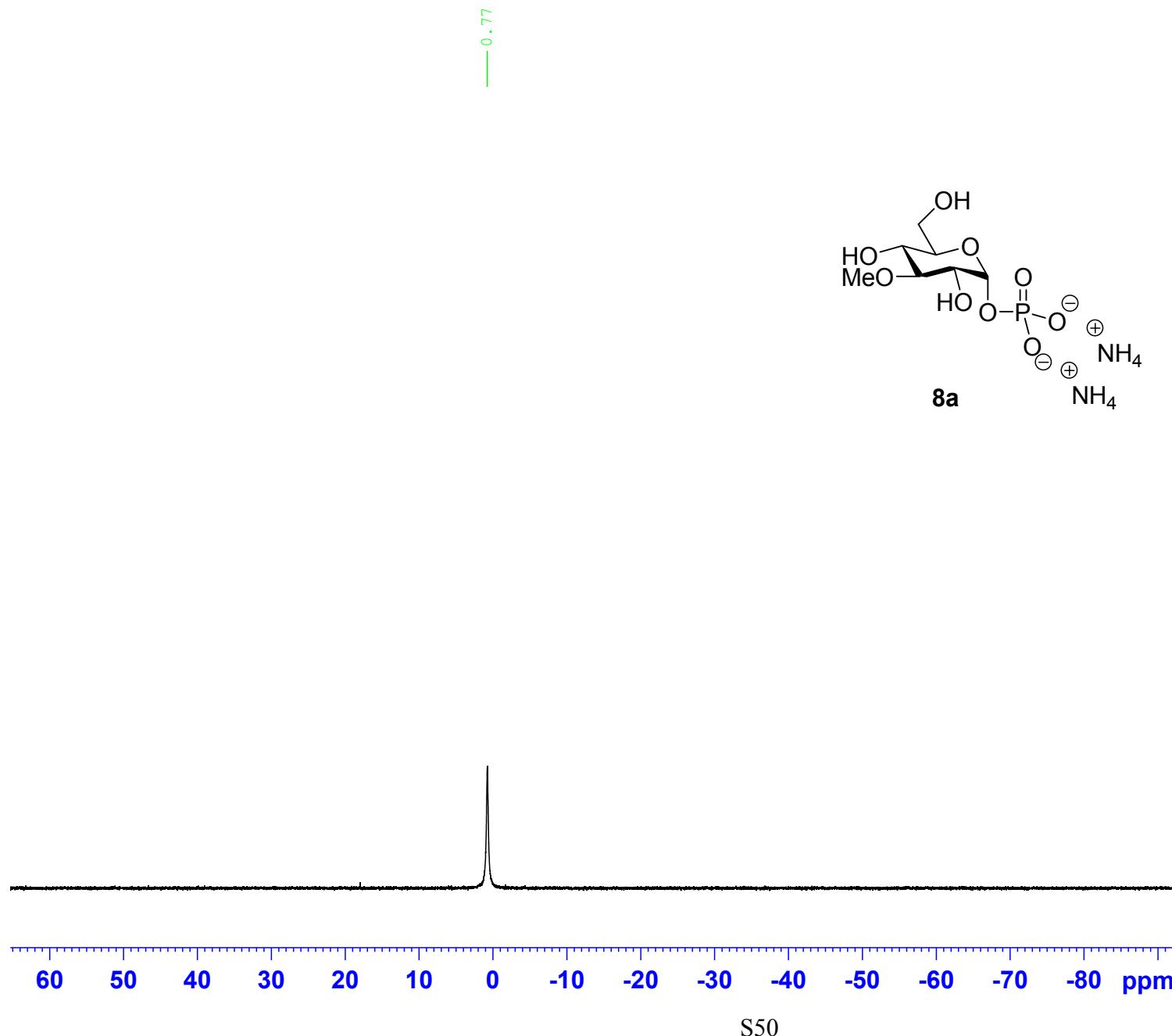
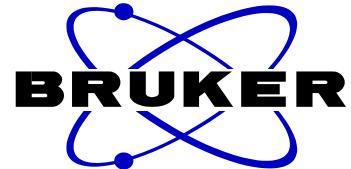
F2 - Acquisition Parameters  
 Date 20070330  
 Time 7.47  
 INSTRUM Av500  
 PROBHD 5 mm BBO BB-1H  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT D2O  
 NS 1024  
 DS 2  
 SWH 33783.785 Hz  
 FIDRES 0.515500 Hz  
 AQ 0.9699976 sec  
 RG 912.3  
 DW 14.800 usec  
 DE 6.00 usec  
 TE 300.6 K  
 D1 1.0000000 sec  
 d11 0.0300000 sec  
 DELTA 0.8999998 sec  
 TD0 4

===== CHANNEL f1 ======  
 NUC1 13C  
 P1 7.90 usec  
 PL1 0.00 dB  
 SFO1 125.7716270 MHz

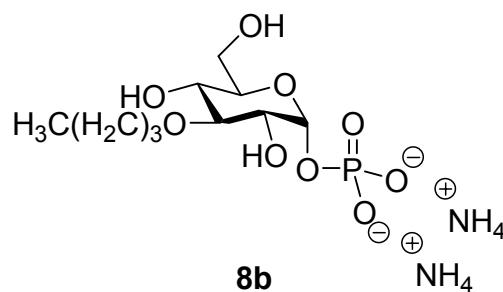
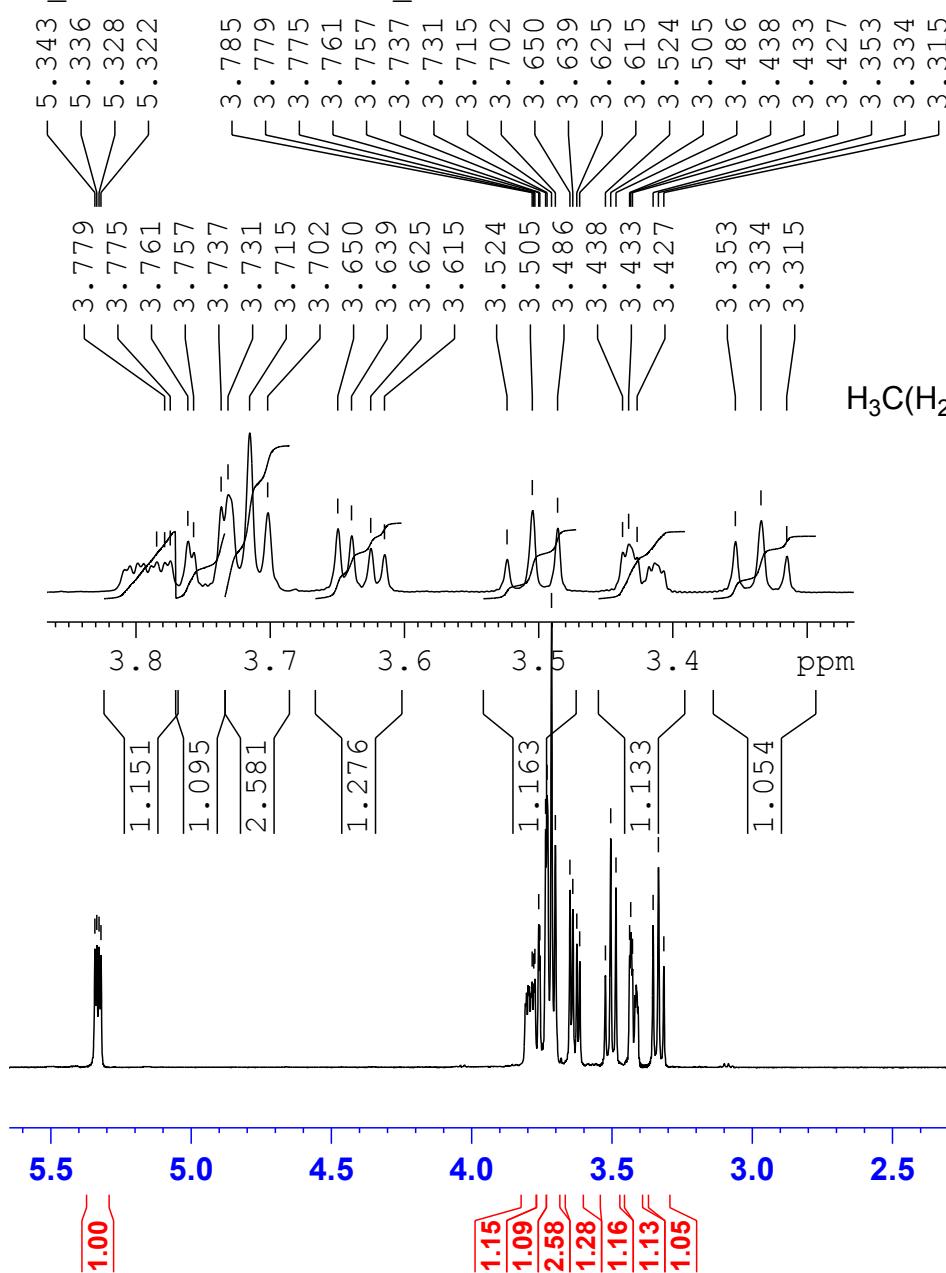
===== CHANNEL f2 ======  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 -1.40 dB  
 PL12 15.83 dB  
 PL13 19.37 dB  
 SFO2 500.1322696 MHz

F2 - Processing parameters  
 SI 65536  
 SF 125.7577930 MHz  
 WDW no  
 SSB 0  
 LB 0.00 Hz  
 GB 0  
 PC 1.40

Clean NH4 OMe Salt  
1d\_31P{1H} D2O {C:\nmr\_users} huestis 12



MH\_104  
1d\_water D2O {C:\nmr users} huestis 2



8b

**BRUKER**

Current Data Parameters  
NAME MH\_104  
EXPNO 10  
PROCNO 1

```

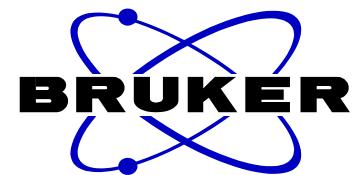
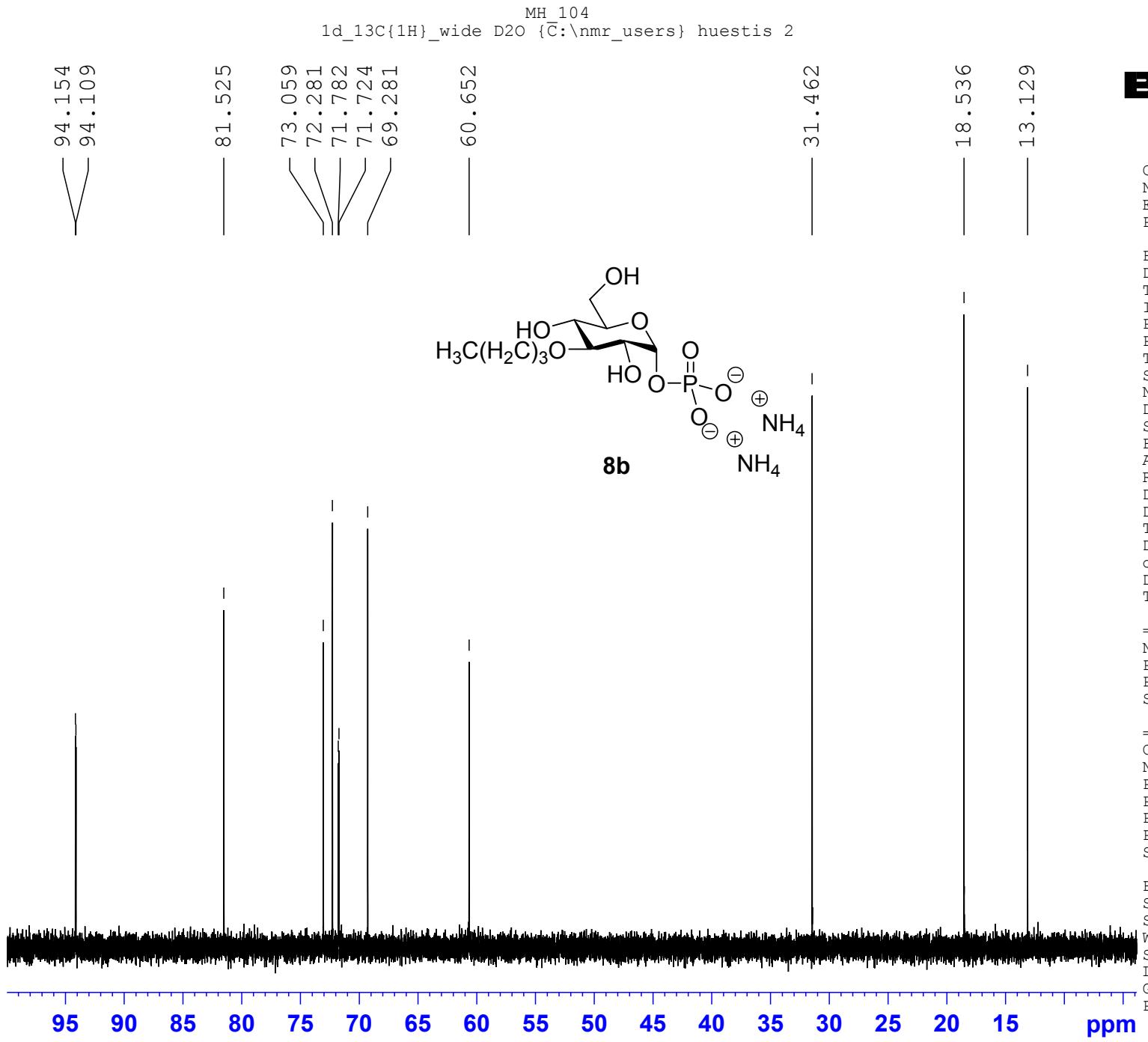
F2 - Acquisition Parameters
Date_           20070405
Time            11.23
INSTRUM         Av500
PROBHD          5 mm BBO BB-1H
PULPROG         zgcpgr
TD              8192
SOLVENT          D2O
NS              32
DS              2
SWH             7507.507 Hz
FIDRES         0.916444 Hz
AQ              0.5457038 sec
RG              128
DW              66.600 usec
DE              6.00 usec
TE              305.0 K
D1              2.00000000 sec
d12             0.00002000 sec
D16             0.00020000 sec
TD0                  1

```

```
===== CHANNEL f1 =====  
NUC1          1H  
P1           11.10 usec  
PL1          -1.40 dB  
PL9           51.67 dB  
SFO1        500.1323698 MHz
```

===== GRADIENT CHANNEL =====  
GPNAME sine.100  
GPZ1 50.00 %  
P16 1000.00 usec

F2 - Processing parameters  
SI 32768  
SF 500.1300390 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
m PC 1.00



Current Data Parameters  
 NAME MH\_104  
 EXPNO 11  
 PROCNO 1

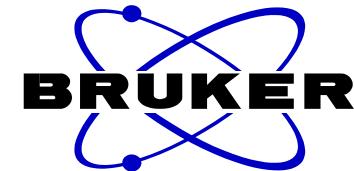
F2 - Acquisition Parameters  
 Date 20070405  
 Time 11.32  
 INSTRUM Av500  
 PROBHD 5 mm BBO BB-1H  
 PULPROG zgpg30  
 TD 65536  
 SOLVENT D2O  
 NS 1024  
 DS 2  
 SWH 33783.785 Hz  
 FIDRES 0.515500 Hz  
 AQ 0.9699976 sec  
 RG 1024  
 DW 14.800 usec  
 DE 6.00 usec  
 TE 300.6 K  
 D1 1.0000000 sec  
 d11 0.0300000 sec  
 DELTA 0.8999998 sec  
 TD0 4

===== CHANNEL f1 ======  
 NUC1 13C  
 P1 7.90 usec  
 PL1 0.00 dB  
 SFO1 125.7716270 MHz

===== CHANNEL f2 ======  
 CPDPRG2 waltz16  
 NUC2 1H  
 PCPD2 80.00 usec  
 PL2 -1.40 dB  
 PL12 15.83 dB  
 PL13 19.37 dB  
 SFO2 500.1322696 MHz

F2 - Processing parameters  
 SI 65536  
 SF 125.7577930 MHz  
 WDW no  
 SSB 0  
 LB 0.00 Hz  
 GB 0  
 PC 1.40

MH\_104  
1d\_31P{1H} D2O {C:\nmr\_users} huestis 2



Current Data Parameters  
NAME MH\_104  
EXPNO 12  
PROCNO 1

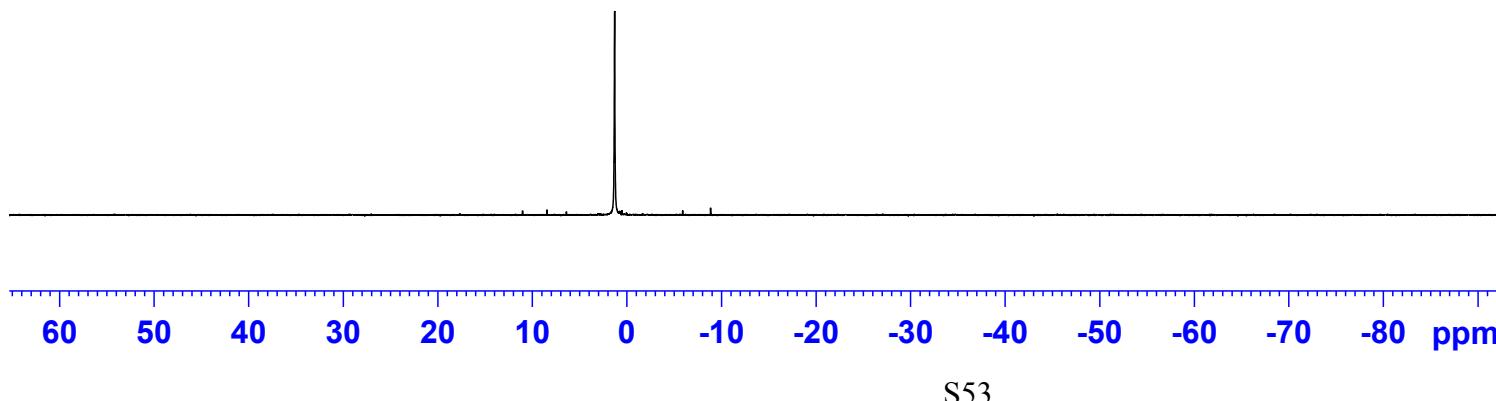
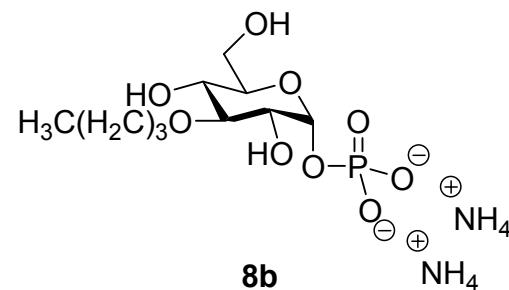
F2 - Acquisition Parameters  
Date 20070405  
Time 12.06  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT D2O  
NS 512  
DS 0  
SWH 98039.219 Hz  
FIDRES 1.495960 Hz  
AQ 0.3342887 sec  
RG 10321.3  
DW 5.100 usec  
DE 6.00 usec  
TE 300.6 K  
D1 1.00000000 sec  
d11 0.03000000 sec  
DELTA 0.89999998 sec  
TD0 2

===== CHANNEL f1 =====  
NUC1 31P  
P1 8.41 usec  
PL1 0.50 dB  
SFO1 202.4664658 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -1.40 dB  
PL12 15.83 dB  
PL13 19.37 dB  
SFO2 500.1322696 MHz

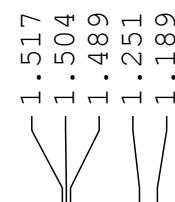
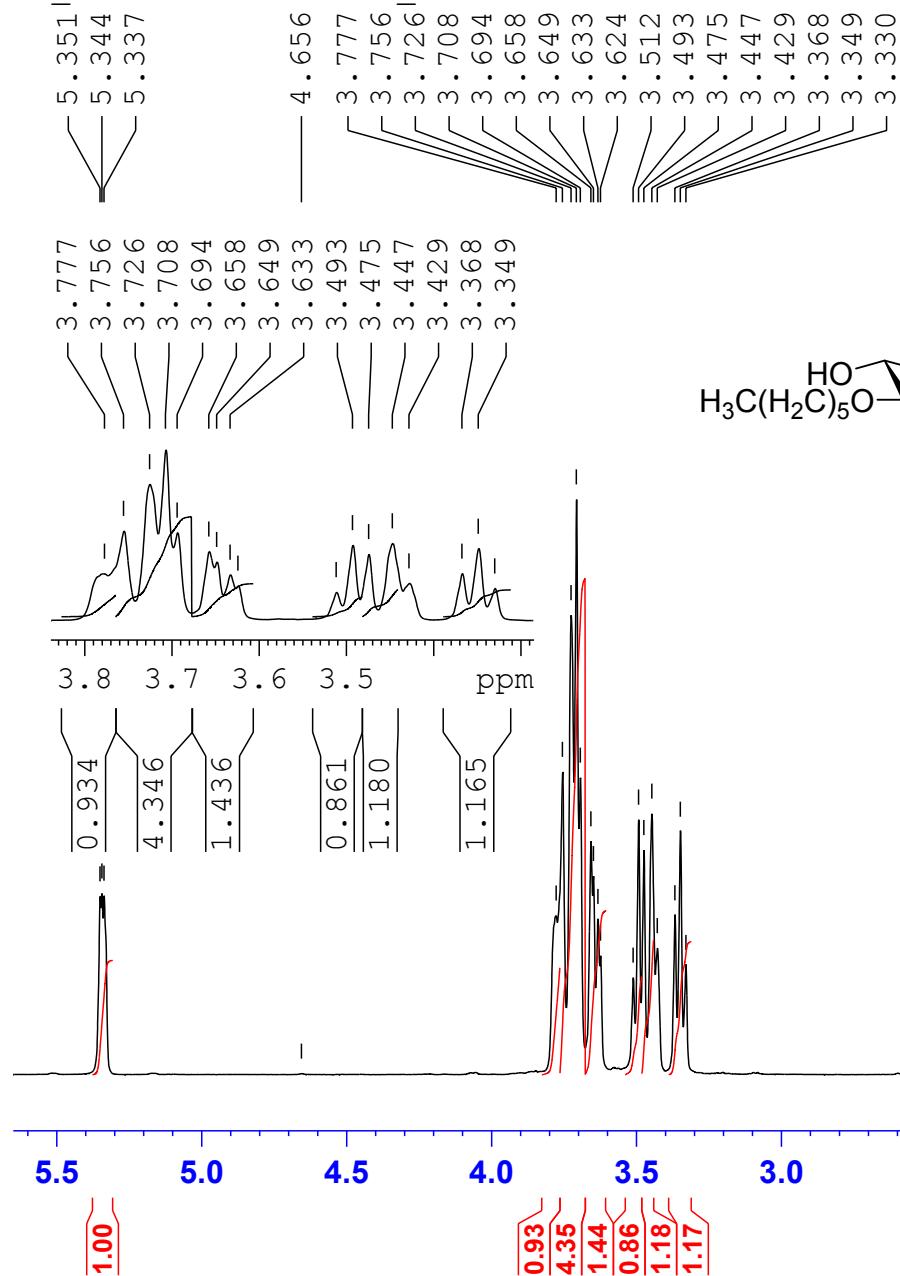
F2 - Processing parameters  
SI 65536  
SF 202.4563430 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 1.00

1.32



S53

hexyl\_NH4  
1d\_water D2O {C:\nmr\_users} huestis 2



Current Data Parameters  
NAME hexyl\_NH4  
EXPNO 10  
PROCNO 1

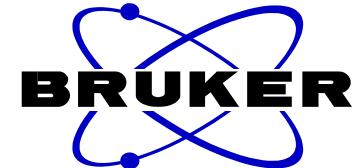
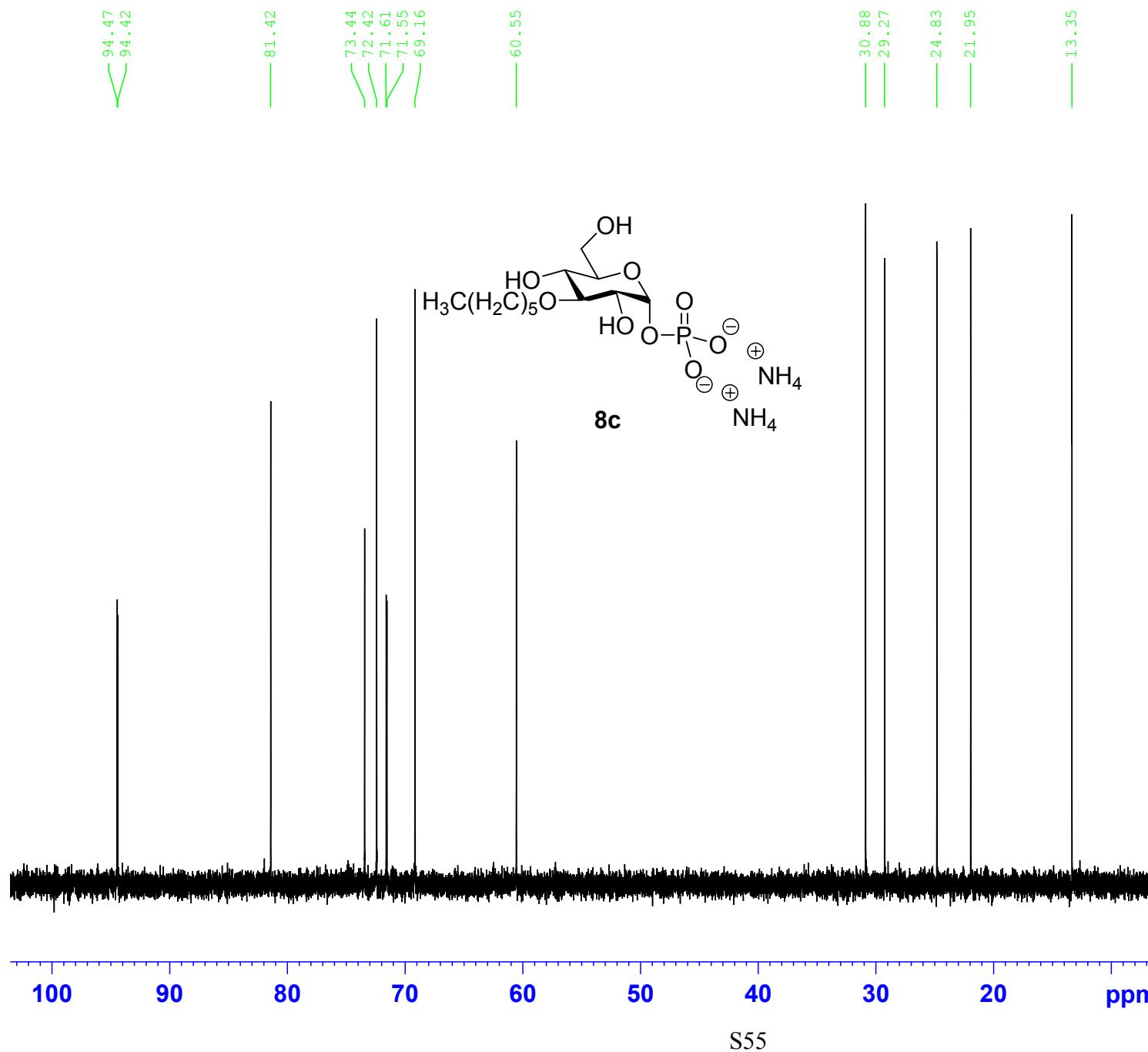
F2 - Acquisition Parameters  
Date\_ 20070618  
Time 21.23  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zgcpgrppr  
TD 8192  
SOLVENT D2O  
NS 32  
DS 2  
SWH 7507.507 Hz  
FIDRES 0.916444 Hz  
AQ 0.5457038 sec  
RG 90.5  
DW 66.600 usec  
DE 6.00 usec  
TE 305.0 K  
D1 2.00000000 sec  
d12 0.00002000 sec  
D16 0.00020000 sec  
TD0 1

===== CHANNEL f1 =====  
NUC1 1H  
P1 11.10 usec  
PL1 -1.40 dB  
PL9 51.67 dB  
SF01 500.1323675 MHz

===== GRADIENT CHANNEL =====  
GPNAME sine.100  
GPZ1 50.00 %  
P16 1000.00 usec

F2 - Processing parameters  
SI 32768  
SF 500.1300390 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 0.75

hexyl\_NH4  
1d\_13C{1H}\_n D2O {C:\nmr\_users} huestis 2



Current Data Parameters  
NAME hexyl\_NH4  
EXPNO 12  
PROCNO 1

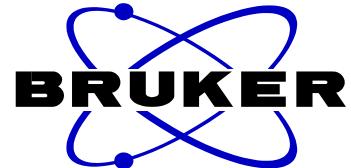
F2 - Acquisition Parameters  
Date 20070618  
Time 21.37  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 54048  
SOLVENT D2O  
NS 1024  
DS 4  
SWH 33783.785 Hz  
FIDRES 0.625070 Hz  
AQ 0.7999752 sec  
RG 9195.2  
DW 14.800 usec  
DE 21.14 usec  
TE 300.4 K  
D1 1.00000000 sec  
d11 0.03000000 sec  
DELTA 0.89999998 sec  
TD0 4

===== CHANNEL f1 =====  
NUC1 13C  
P1 7.90 usec  
PL1 0.00 dB  
SFO1 125.7716273 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -1.40 dB  
PL12 15.83 dB  
PL13 19.37 dB  
SFO2 500.1322696 MHz

F2 - Processing parameters  
SI 65536  
SF 125.7577930 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 0.75

hexyl\_NH4  
1d\_31P{1H}\_d D2O {C:\nmr\_users} huestis 2



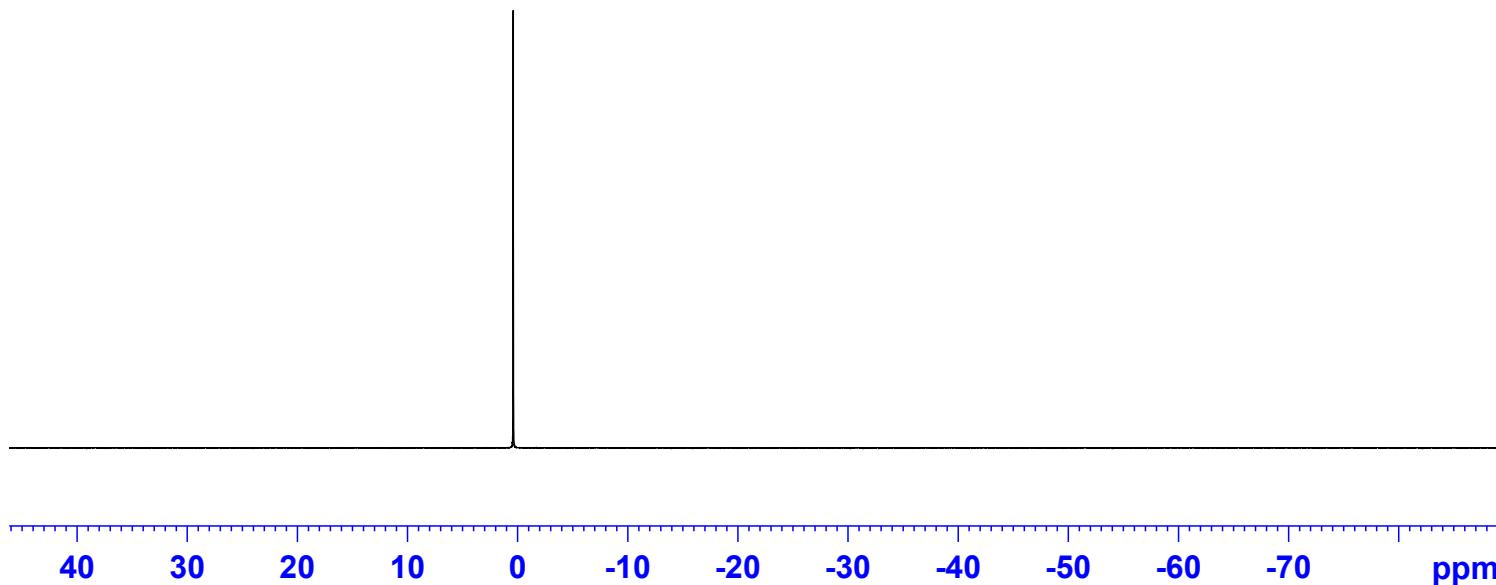
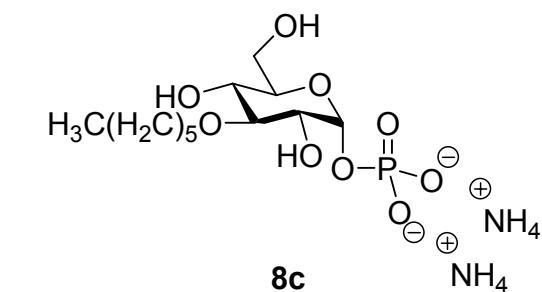
Current Data Parameters  
NAME hexyl\_NH4  
EXPNO 13  
PROCNO 1

F2 - Acquisition Parameters  
Date 20070618  
Time 22.09  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT D2O  
NS 512  
DS 4  
SWH 98039.219 Hz  
FIDRES 1.495960 Hz  
AQ 0.3342887 sec  
RG 9195.2  
DW 5.100 usec  
DE 7.29 usec  
TE 300.4 K  
D1 1.0000000 sec  
d11 0.0300000 sec  
DELTA 0.8999998 sec  
TD0 2

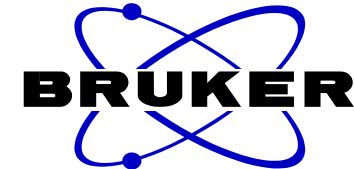
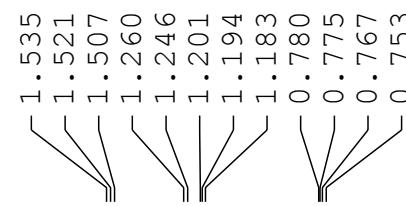
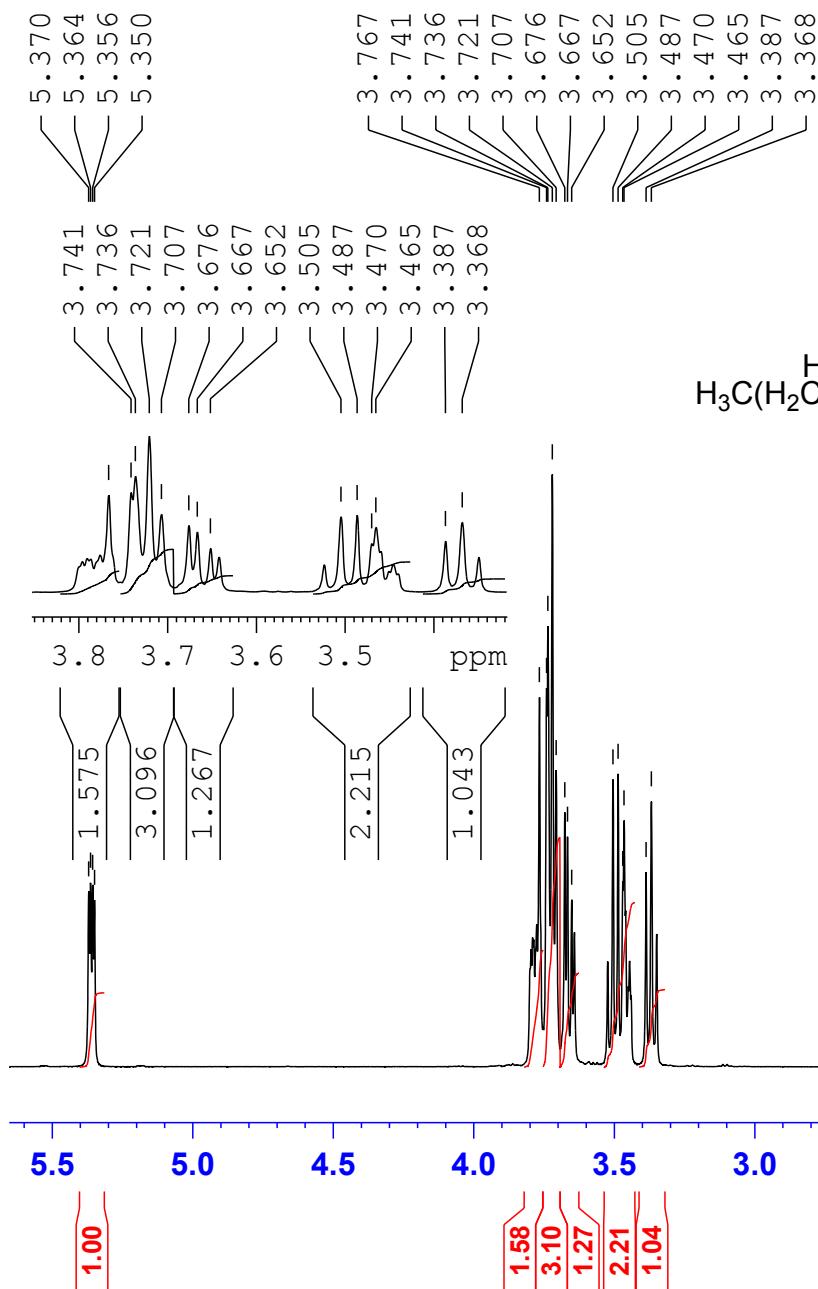
===== CHANNEL f1 =====  
NUC1 31P  
P1 8.41 usec  
PL1 0.50 dB  
SFO1 202.4664658 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -1.40 dB  
PL12 15.83 dB  
PL13 19.37 dB  
SFO2 500.1322696 MHz

F2 - Processing parameters  
SI 65536  
SF 202.4563430 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 0.75



MH\_116  
1d\_water D2O {C:\nmr\_users} huestis 7



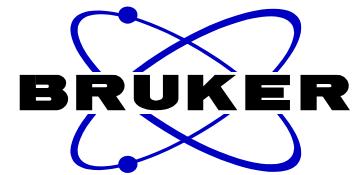
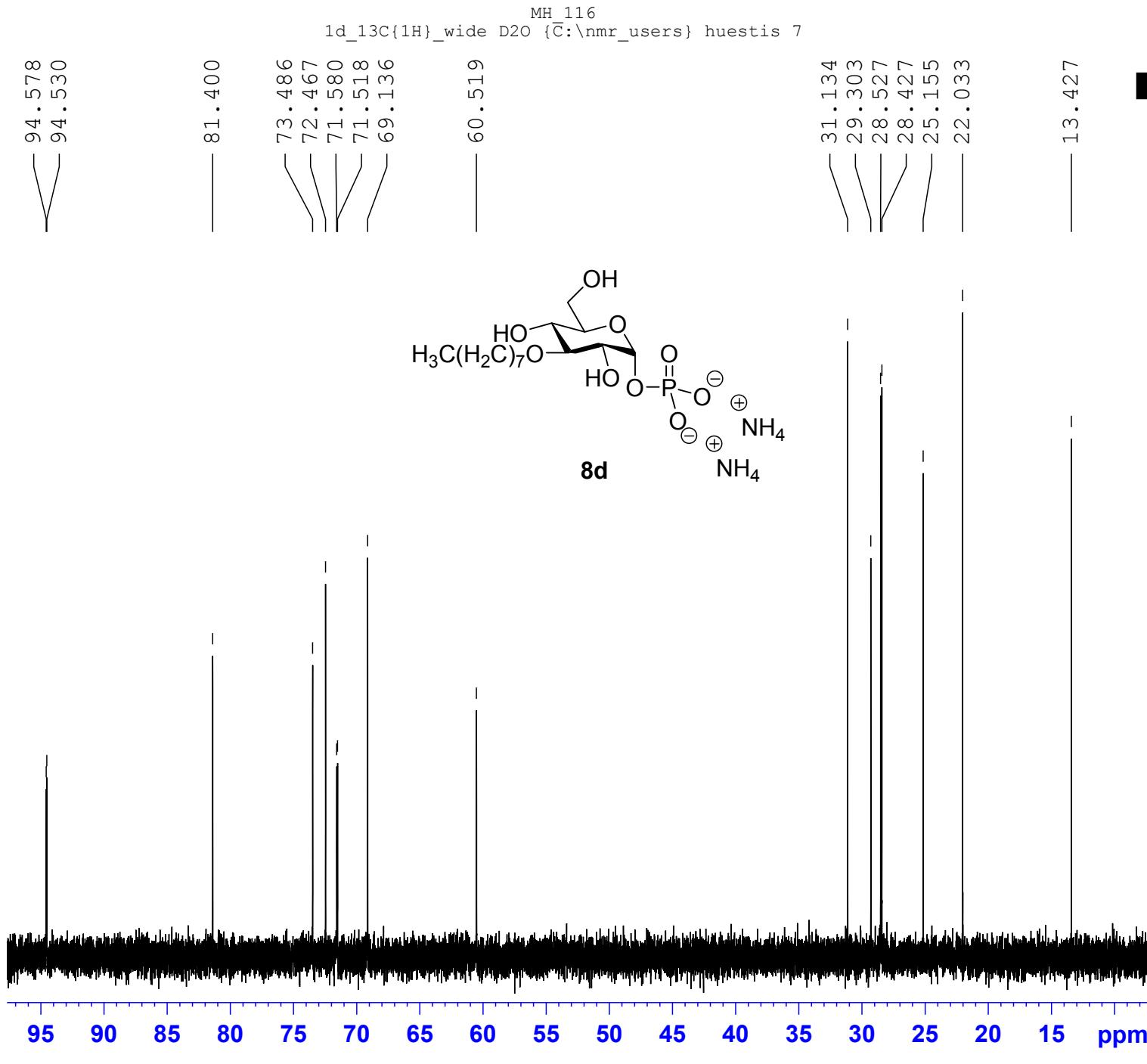
Current Data Parameters  
NAME MH\_116  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20070420  
Time 14.04  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zgcpgrppr  
TD 8192  
SOLVENT D2O  
NS 32  
DS 2  
SWH 7507.507 Hz  
FIDRES 0.916444 Hz  
AQ 0.5457038 sec  
RG 128  
DW 66.600 usec  
DE 6.00 usec  
TE 305.0 K  
D1 2.00000000 sec  
d12 0.00002000 sec  
D16 0.00020000 sec  
TD0 1

===== CHANNEL f1 ======  
NUC1 1H  
P1 11.10 usec  
PL1 -1.40 dB  
PL9 51.67 dB  
SF01 500.1323707 MHz

===== GRADIENT CHANNEL =====  
GPNAME sine.100  
GPZ1 50.00 %  
P16 1000.00 usec

F2 - Processing parameters  
SI 32768  
SF 500.1300390 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00



Current Data Parameters  
NAME MH\_116  
EXPNO 11  
PROCNO 1

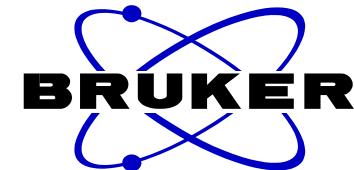
F2 - Acquisition Parameters  
Date 20070420  
Time 14.14  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT D2O  
NS 1024  
DS 2  
SWH 33783.785 Hz  
FIDRES 0.515500 Hz  
AQ 0.9699976 sec  
RG 1024  
DW 14.800 usec  
DE 6.00 usec  
TE 300.4 K  
D1 1.0000000 sec  
d11 0.0300000 sec  
DELTA 0.8999998 sec  
TD0 4

===== CHANNEL f1 ======  
NUC1 13C  
P1 7.90 usec  
PL1 0.00 dB  
SFO1 125.7716270 MHz

===== CHANNEL f2 ======  
CPDPG2 waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -1.40 dB  
PL12 15.83 dB  
PL13 19.37 dB  
SFO2 500.1322696 MHz

F2 - Processing parameters  
SI 65536  
SF 125.7577930 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 1.40

MH\_116  
1d\_31P{1H} D2O {C:\nmr\_users} huestis 7



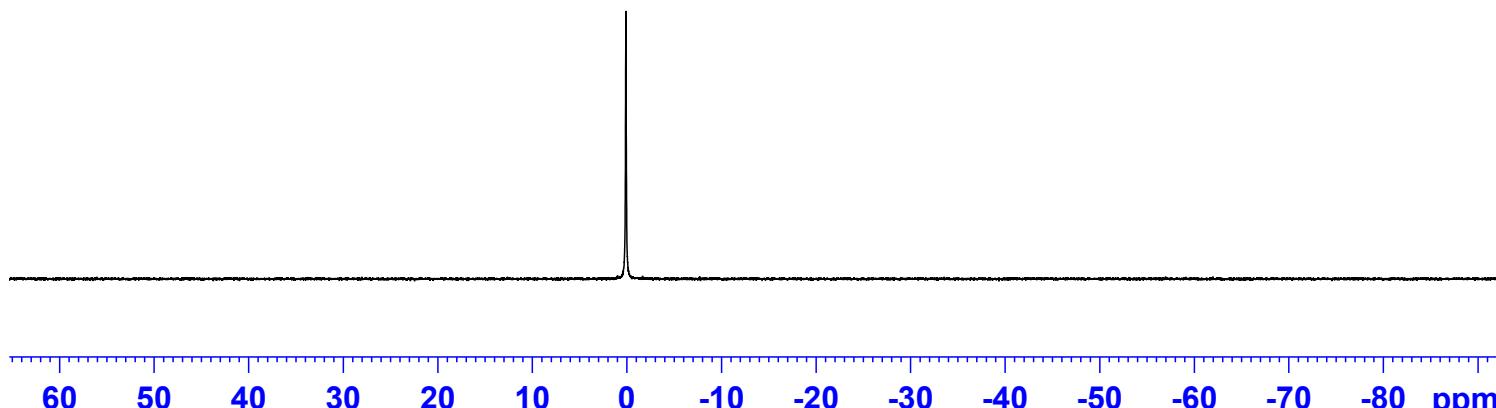
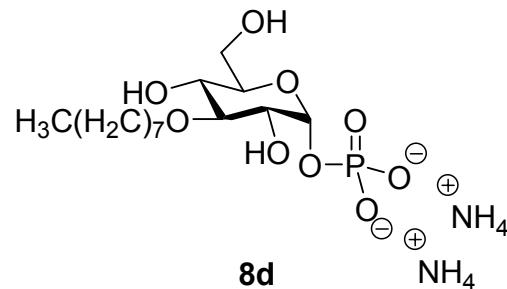
Current Data Parameters  
NAME MH\_116  
EXPNO 12  
PROCNO 1

F2 - Acquisition Parameters  
Date 20070420  
Time 14.48  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT D2O  
NS 512  
DS 0  
SWH 98039.219 Hz  
FIDRES 1.495960 Hz  
AQ 0.3342887 sec  
RG 10321.3  
DW 5.100 usec  
DE 6.00 usec  
TE 300.6 K  
D1 1.00000000 sec  
d11 0.03000000 sec  
DELTA 0.89999998 sec  
TD0 2

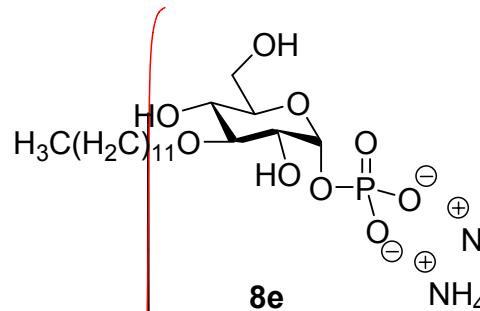
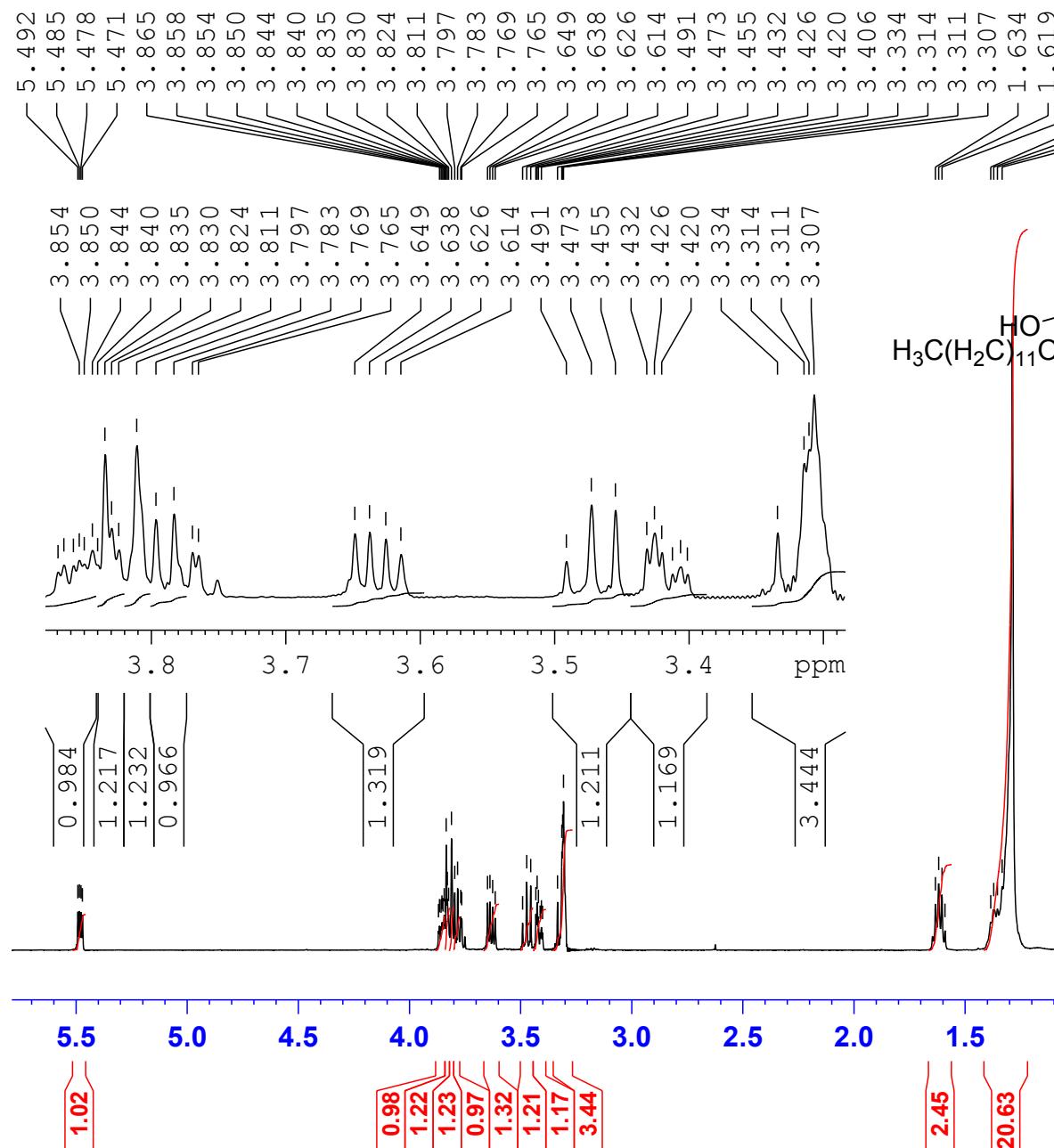
===== CHANNEL f1 =====  
NUC1 31P  
P1 8.41 usec  
PL1 0.50 dB  
SFO1 202.4664658 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -1.40 dB  
PL12 15.83 dB  
PL13 19.37 dB  
SFO2 500.1322696 MHz

F2 - Processing parameters  
SI 65536  
SF 202.4563430 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 1.00



MH\_137\_NH4  
1d\_water MeOD {C:\nmr users} huestis 1



Current	Data	Parameters
NAME	MH_137_NH4	
EXPNO		10
PROCNO		1

```

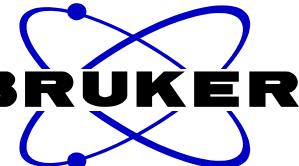
F2 - Acquisition Parameters
Date_           20070618
Time            15.53
INSTRUM         Av500
PROBHD         5 mm BBO BB-1H
PULPROG        zgcpgrpr
TD              8192
SOLVENT         MeOD
NS              32
DS              2
SWH             7507.507 Hz
FIDRES         0.916444 Hz
AQ              0.5457038 sec
RG              181
DW              66.600 usec
DE              6.00 usec
TE              305.0 K
D1              2.00000000 sec
d12             0.00002000 sec
D16             0.00020000 sec
TDO              1

```

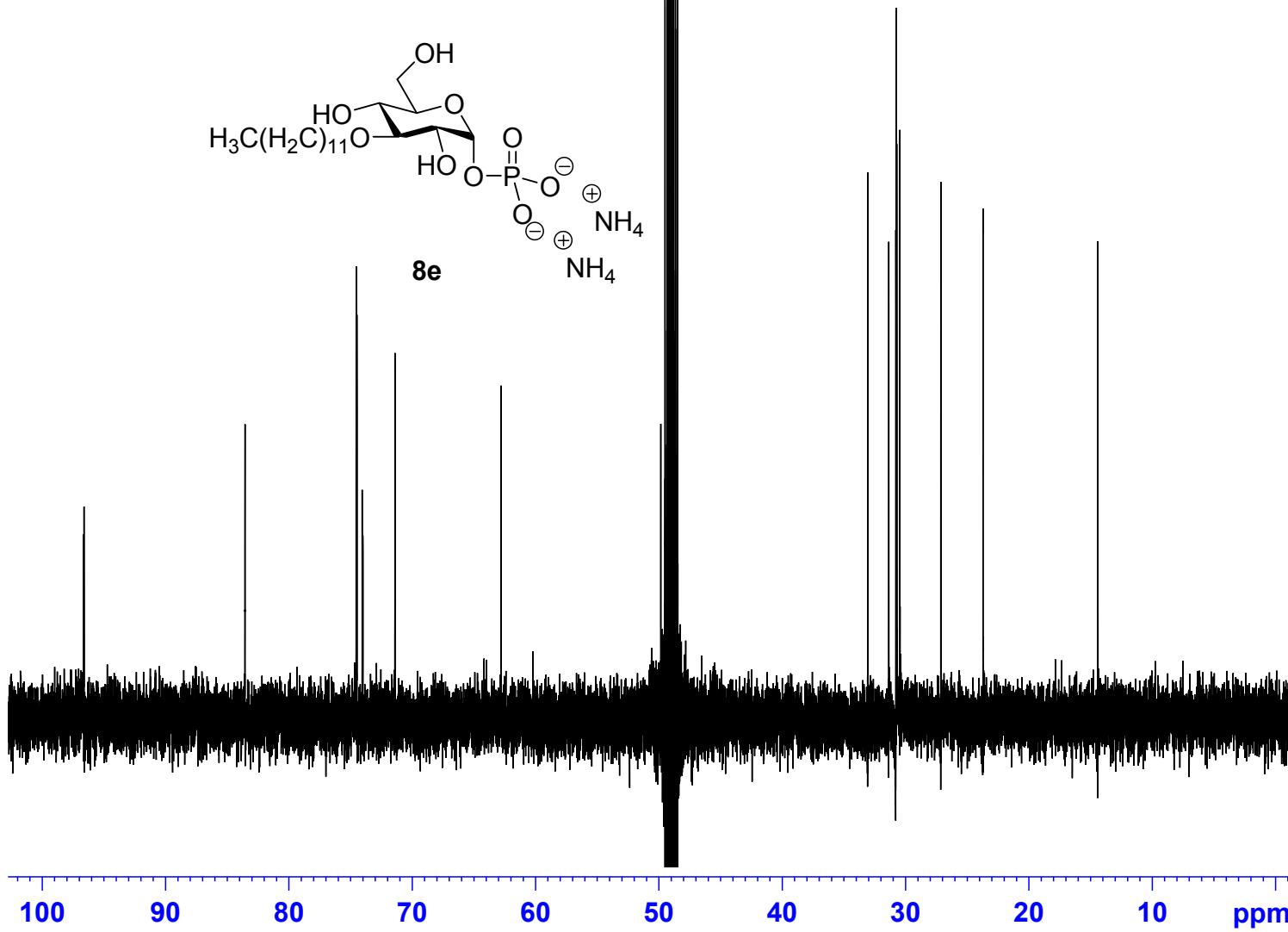
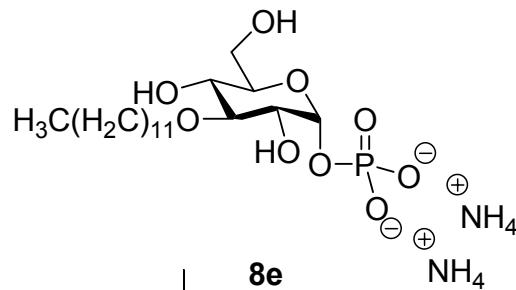
```
===== CHANNEL f1 =====  
NUC1          1H  
P1           9.80  usec  
PL1         -2.00  dB  
PL9         52.15  dB  
SFO1      500 1317007 MHZ
```

===== GRADIENT CHANNEL =====  
GPNAME sine.100  
GPZ1 50.00 %  
P16 1000.00 usec

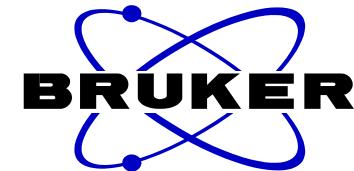
F2 - Processing parameters  
SI 32768  
SF 500.1292742 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
mPC 0.75



Current	Data	Parameters
NAME	MH_137	NH4
EXPNO		11
PROCNO		1



MH\_137\_NH4  
1d\_31P{1H}\_d MeOD {C:\nmr\_users} huestis 1



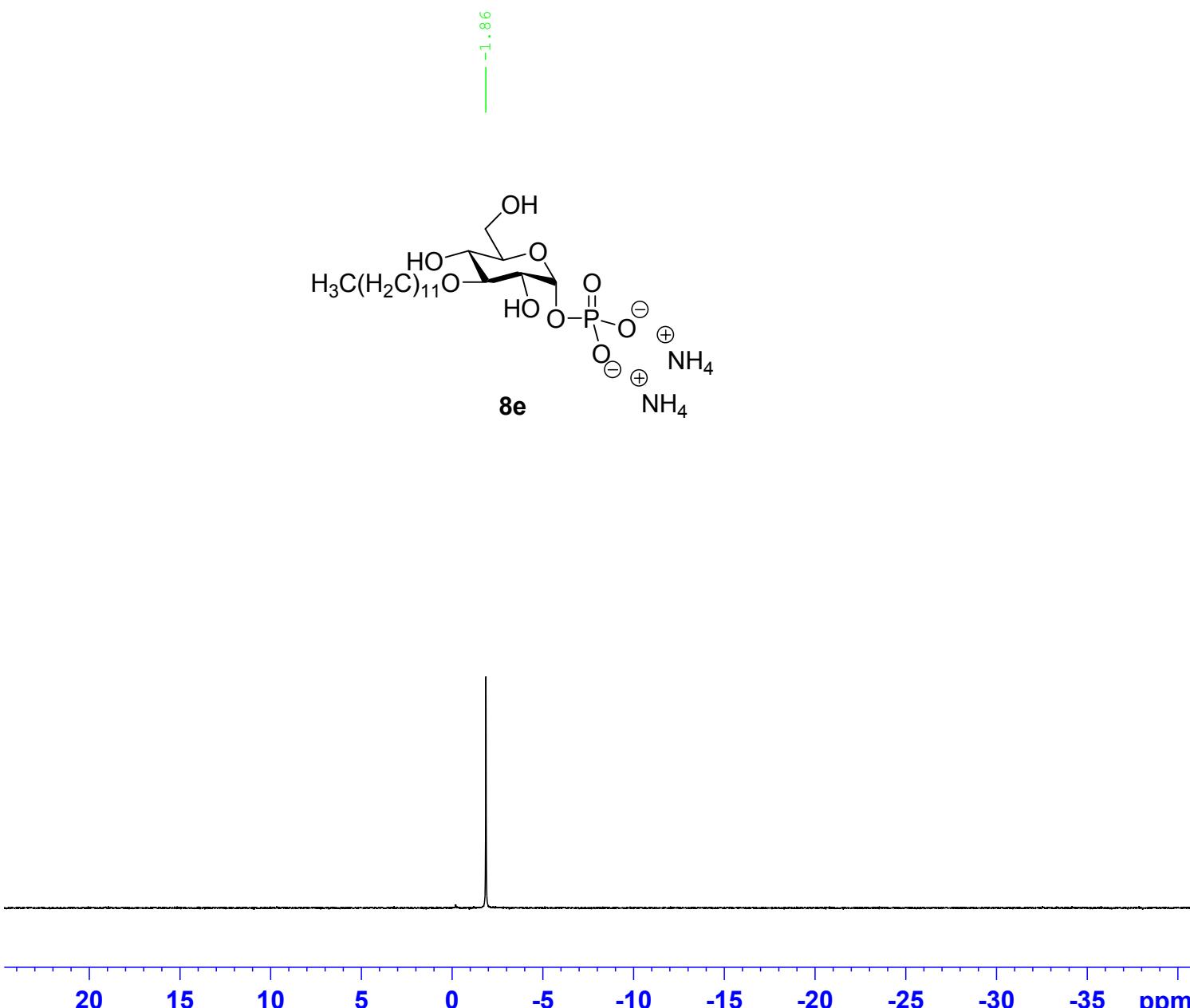
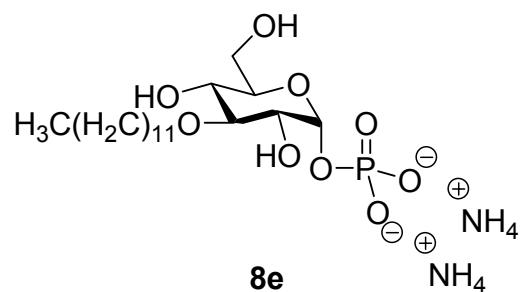
Current Data Parameters  
NAME MH\_137\_NH4  
EXPNO 12  
PROCNO 1

F2 - Acquisition Parameters  
Date 20070618  
Time 16.01  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT MeOD  
NS 512  
DS 4  
SWH 98039.219 Hz  
FIDRES 1.495960 Hz  
AQ 0.3342887 sec  
RG 9195.2  
DW 5.100 usec  
DE 7.29 usec  
TE 300.4 K  
D1 1.0000000 sec  
d11 0.0300000 sec  
DELTA 0.8999998 sec  
TD0 2

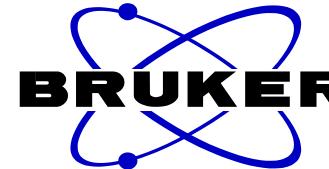
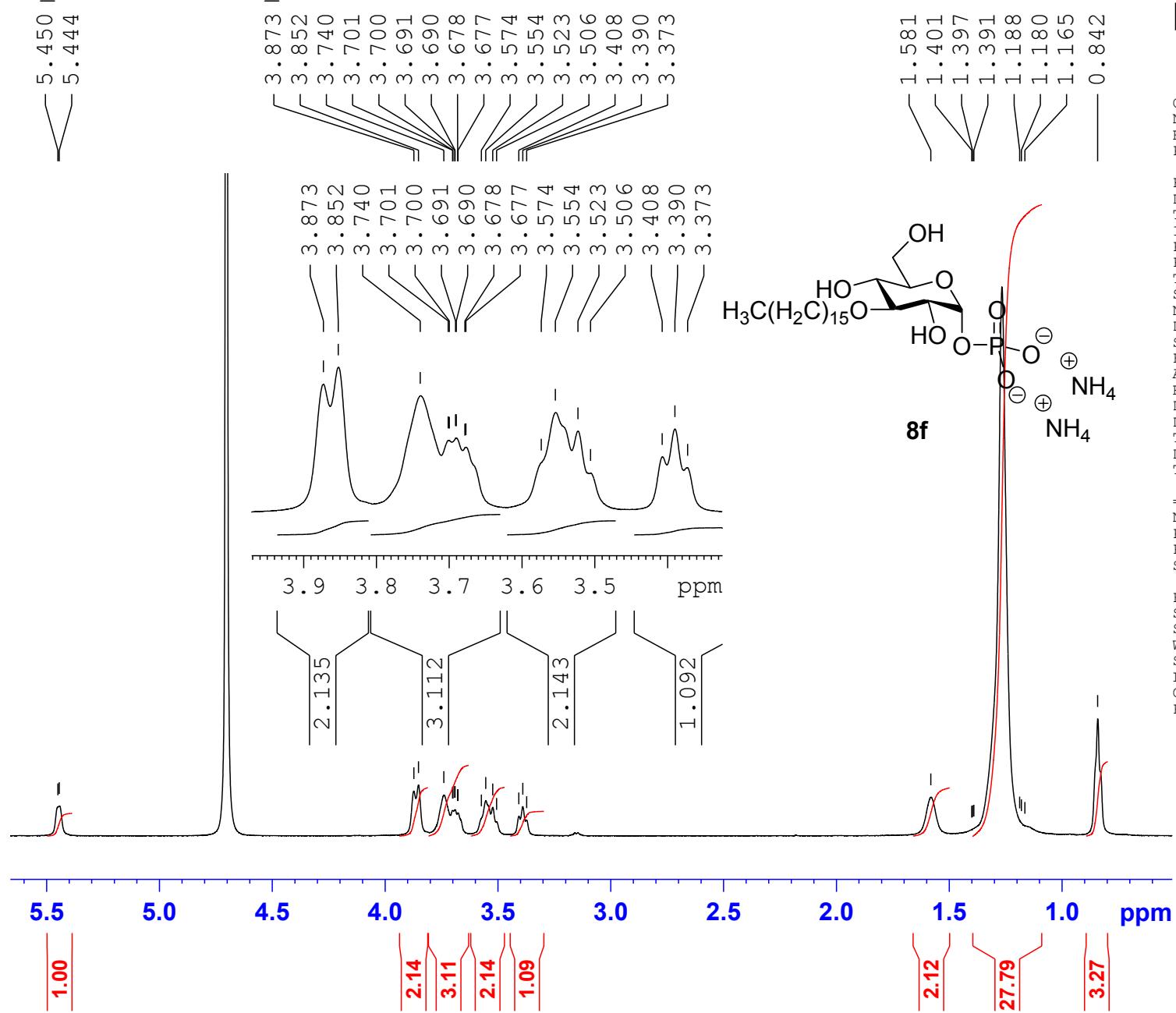
===== CHANNEL f1 =====  
NUC1 31P  
P1 8.41 usec  
PL1 0.50 dB  
SFO1 202.4664658 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -2.00 dB  
PL12 15.72 dB  
PL13 19.37 dB  
SFO2 500.1322696 MHz

F2 - Processing parameters  
SI 65536  
SF 202.4563430 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 0.75



Gaia-4, 6-benzylidene  
1d\_1H D2O {C:\nmr\_users} huestis 4



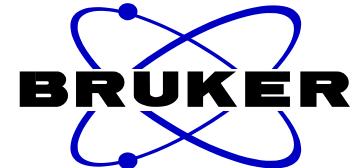
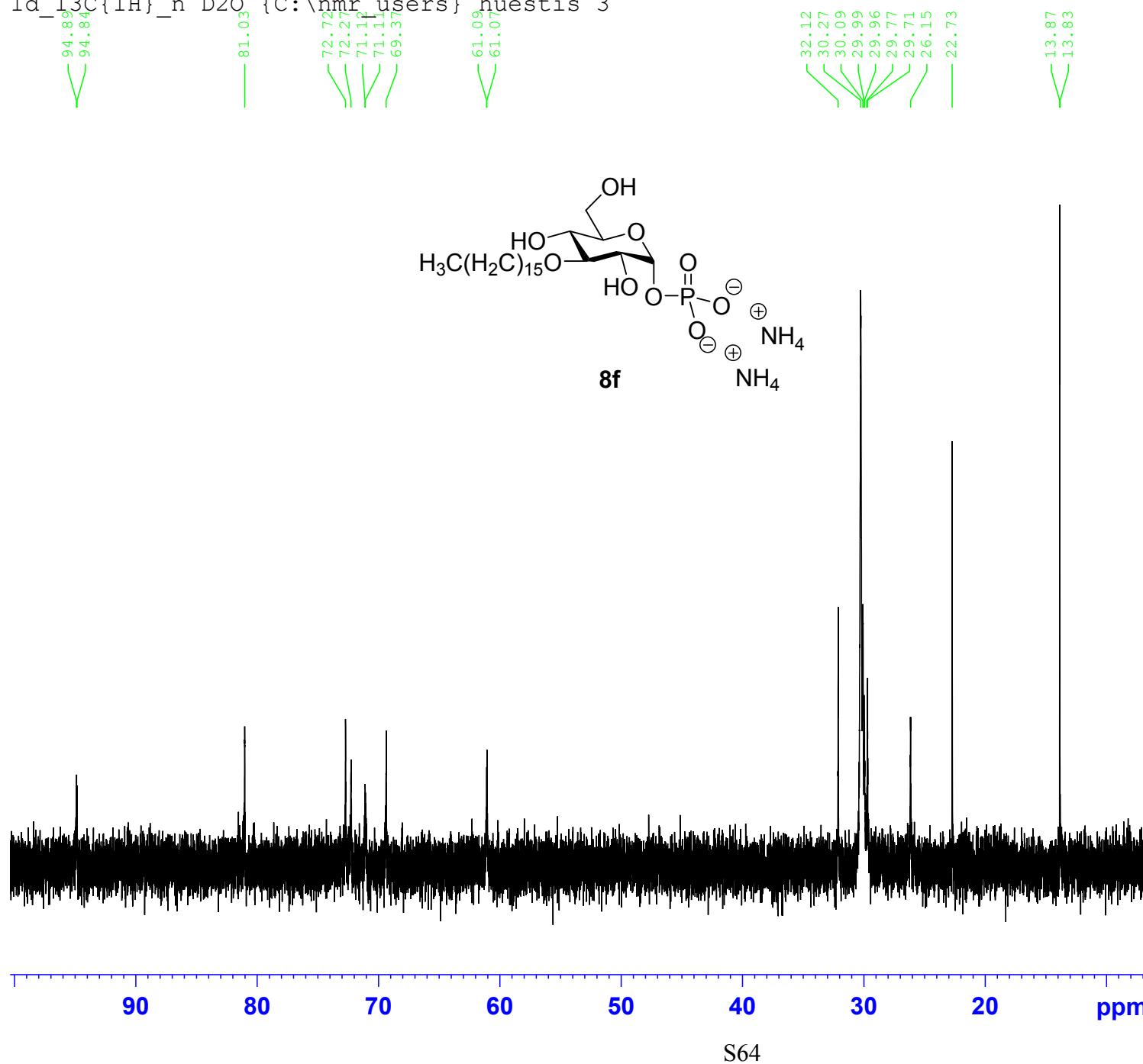
Current Data Parameters  
NAME Hexadecyl deprotected 3  
EXPNO 11  
PROCNO 1

F2 - Acquisition Parameters  
Date 20070719  
Time 2.00  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zg30  
TD 65536  
SOLVENT D2O  
NS 128  
DS 2  
SWH 10000.000 Hz  
FIDRES 0.152588 Hz  
AQ 3.2769001 sec  
RG 228.1  
DW 50.000 usec  
DE 6.00 usec  
TE 300.0 K  
D1 1.0000000 sec  
TD0 1

===== CHANNEL f1 =====  
NUC1 1H  
P1 9.80 usec  
PL1 -2.00 dB  
SFO1 500.1332698 MHz

F2 - Processing parameters  
SI 65536  
SF 500.1300190 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 0.75

hexadecyl deprotected carbon  
 1d\_13C{1H}\_n D2O {C:\nmr\_users} huestis 3



Current Data Parameters  
 NAME hexadecyl deprotected carbon  
 EXPNO 10  
 PROCNO 1

F2 - Acquisition Parameters

Date 20070724  
 Time 18.18  
 INSTRUM Av500  
 PROBHD 5 mm BBO BB-1H  
 PULPROG zgpg30  
 TD 54048  
 SOLVENT D2O  
 NS 8192  
 DS 4  
 SWH 33783.785 Hz  
 FIDRES 0.625070 Hz  
 AQ 0.7999752 sec  
 RG 10321.3  
 DW 14.800 usec  
 DE 21.14 usec  
 TE 296.6 K  
 D1 1.0000000 sec  
 d11 0.0300000 sec  
 DELTA 0.8999998 sec  
 TDO 32

===== CHANNEL f1 =====

NUC1 <sup>13</sup>C  
 P1 7.90 usec  
 PL1 0.00 dB  
 SFO1 125.7716273 MHz

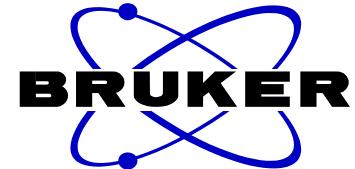
===== CHANNEL f2 =====

CPDPRG2 waltz16  
 NUC2 <sup>1</sup>H  
 PCPD2 80.00 usec  
 PL2 -2.00 dB  
 PL12 15.72 dB  
 PL13 18.64 dB  
 SFO2 500.1322696 MHz

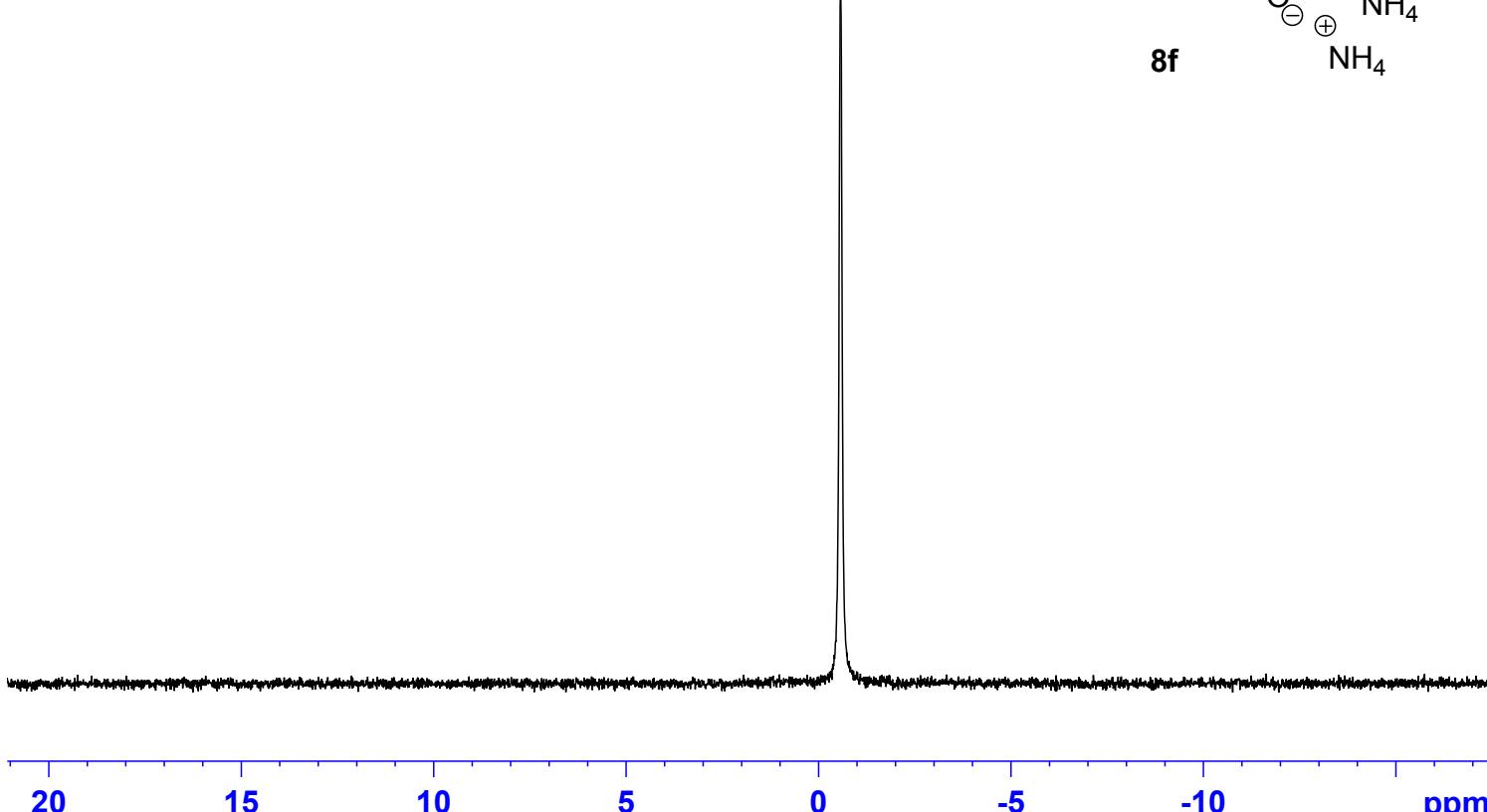
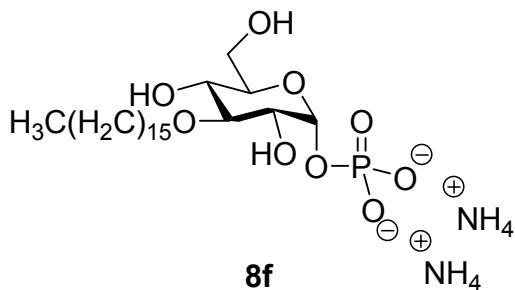
F2 - Processing parameters

SI 65536  
 SF 125.7577930 MHz  
 WDW no  
 SSB 0  
 LB 0.00 Hz  
 GB 0  
 PC 0.75

Gaia-4, 6-benzylidene  
1d\_31P\_d D2O {C:\nmr\_users} huestis 4



-0.57



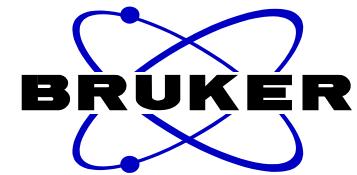
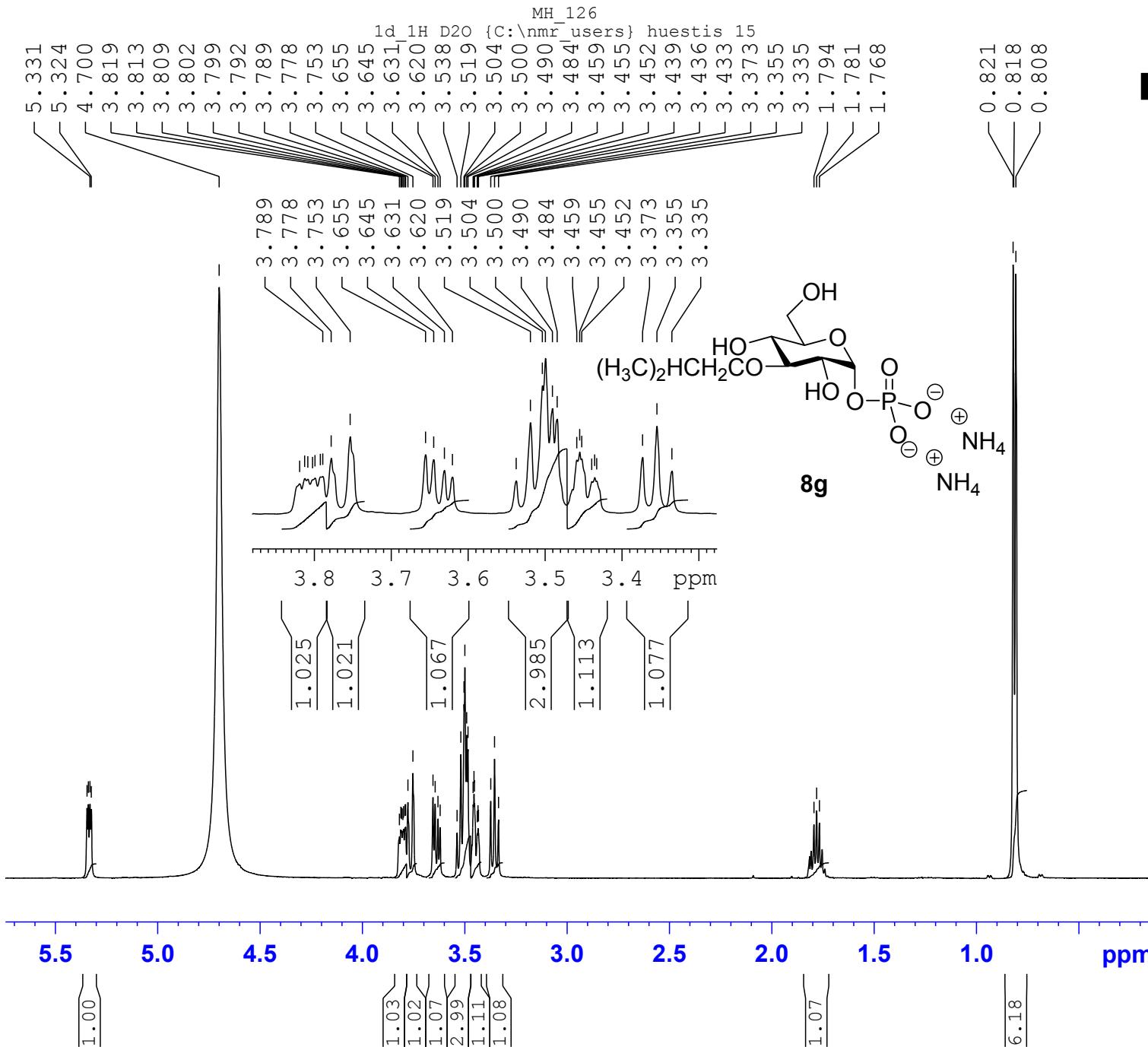
Current Data Parameters  
NAME Hexadecyl deprotected 3  
EXPNO 10  
PROCNO 1

F2 - Acquisition Parameters  
Date 20070719  
Time 1.33  
INSTRUM Av500  
PROBHD 5 mm BBO BB-1H  
PULPROG zgdd30  
TD 65536  
SOLVENT D2O  
NS 1024  
DS 2  
SWH 98039.219 Hz  
FIDRES 1.495960 Hz  
AQ 0.3342887 sec  
RG 7298.2  
DW 5.100 usec  
DE 7.29 usec  
TE 300.6 K  
D1 1.0000000 sec  
d11 0.03000000 sec  
TD0 4

===== CHANNEL f1 =====  
NUC1 31P  
P1 8.41 usec  
PL1 0.50 dB  
SFO1 202.4664652 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -2.00 dB  
PL12 15.72 dB  
SFO2 500.1322696 MHz

F2 - Processing parameters  
SI 65536  
SF 202.4563430 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 0.75

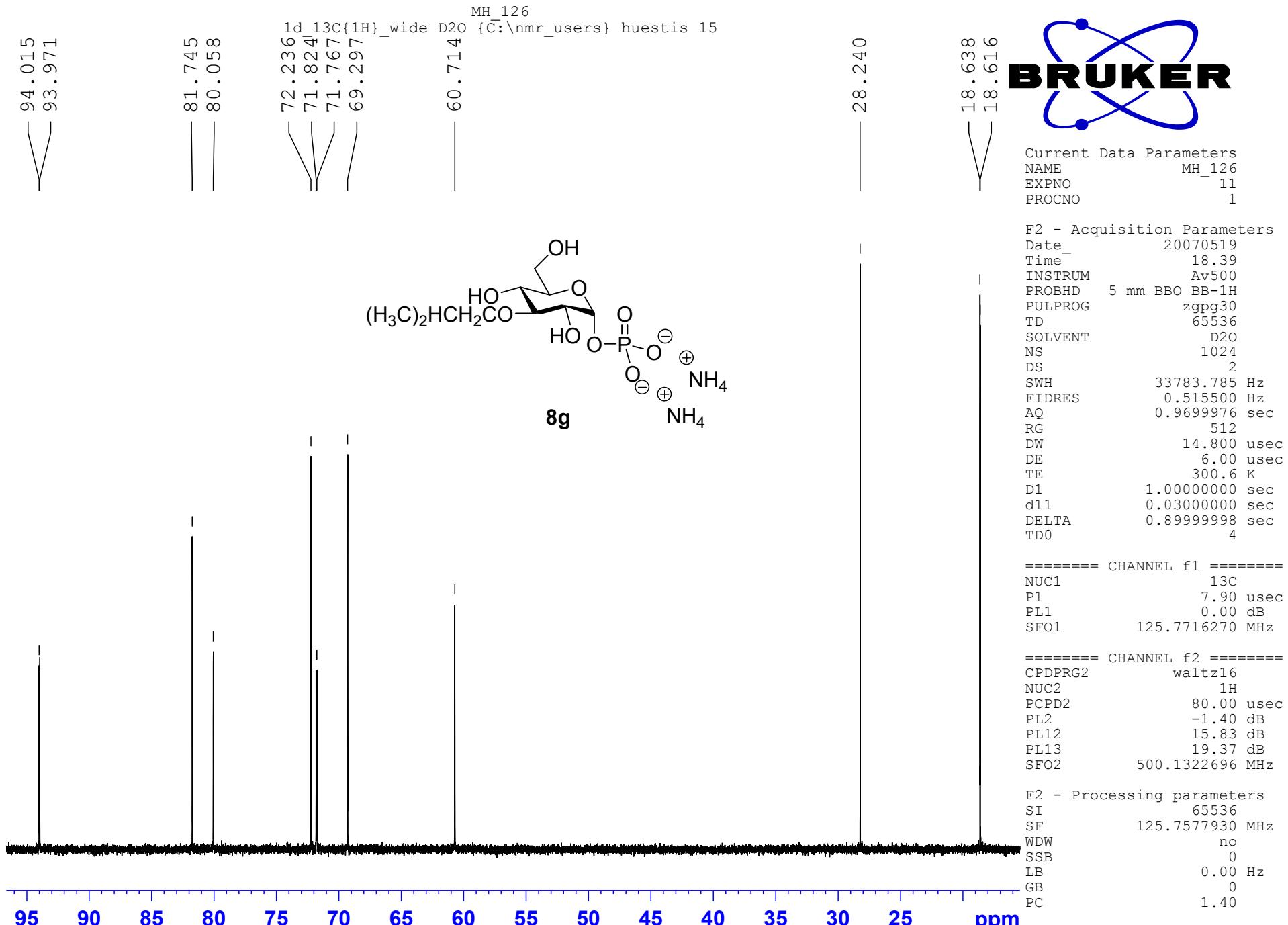


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 PROCNO 1

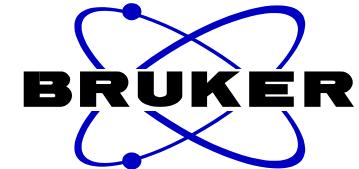
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 PULPROG zg30  
 TD 65536  
 SOLVENT D2O  
 NS 64  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.152588 Hz  
 AQ 3.2769001 sec  
 RG 71.8  
 DW 50.000 usec  
 DE 6.00 usec  
 TE 300.0 K  
 D1 1.00000000 sec  
 TD0 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 11.10 usec  
 PL1 -1.40 dB  
 SFO1 500.1332698 MHz

F2 - Processing parameters  
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 SF 500.1300190 MHz  
 WDW no  
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 GB 0  
 PC 1.00



MH\_126  
1d\_31P{1H} D2O {C:\nmr\_users} huestis 15



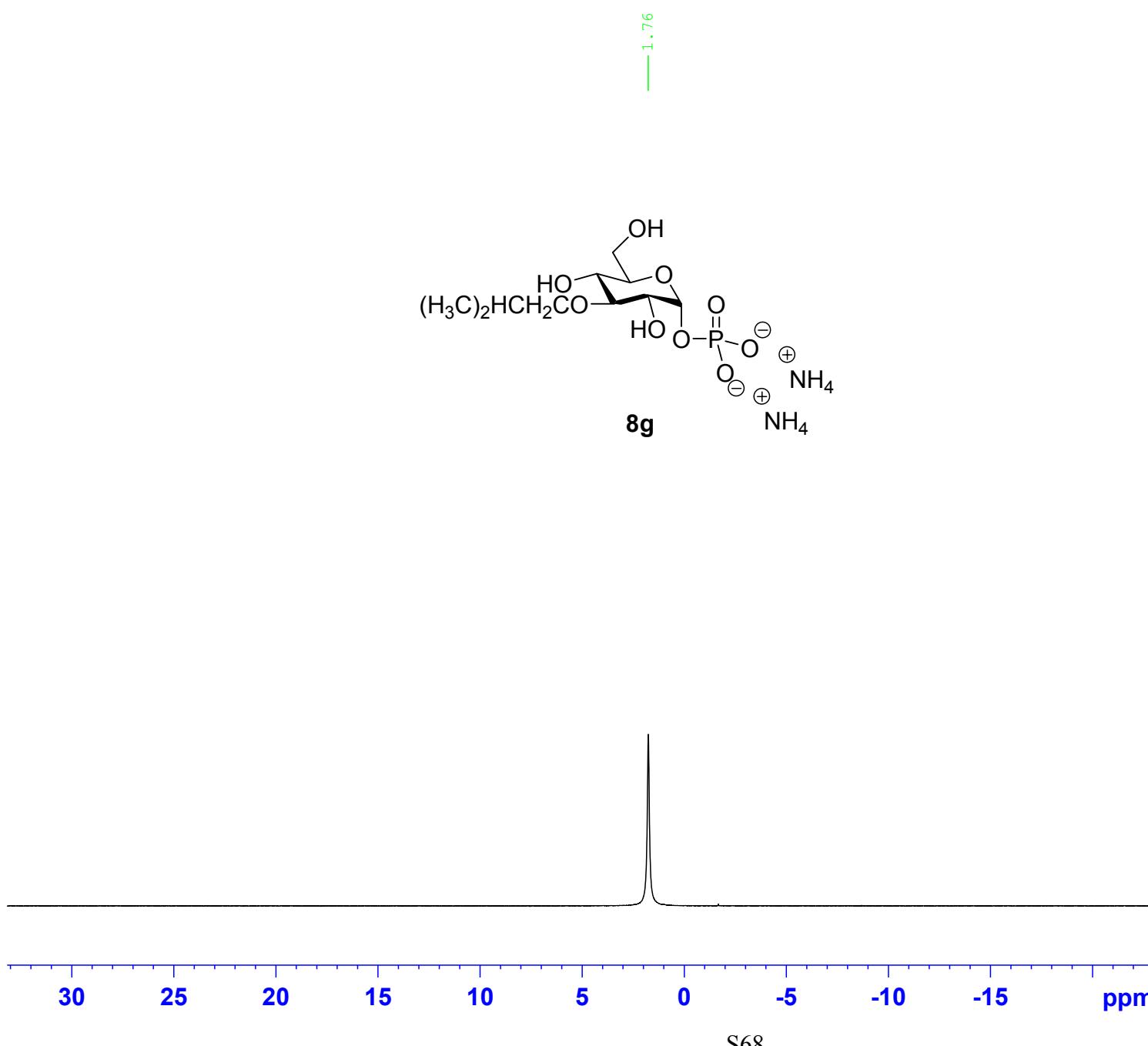
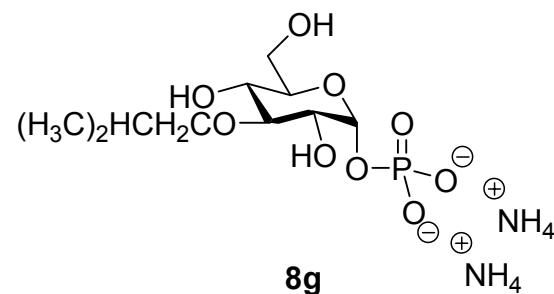
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EXPNO 12  
PROCNO 1

F2 - Acquisition Parameters  
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Time 19.13  
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PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT D2O  
NS 512  
DS 0  
SWH 98039.219 Hz  
FIDRES 1.495960 Hz  
AQ 0.3342887 sec  
RG 6502  
DW 5.100 usec  
DE 6.00 usec  
TE 300.6 K  
D1 1.0000000 sec  
d11 0.0300000 sec  
DELTA 0.8999998 sec  
TD0 2

===== CHANNEL f1 =====  
NUC1 31P  
P1 8.41 usec  
PL1 0.50 dB  
SFO1 202.4664658 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -1.40 dB  
PL12 15.83 dB  
PL13 19.37 dB  
SFO2 500.1322696 MHz

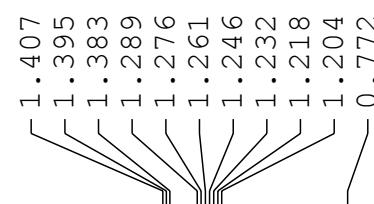
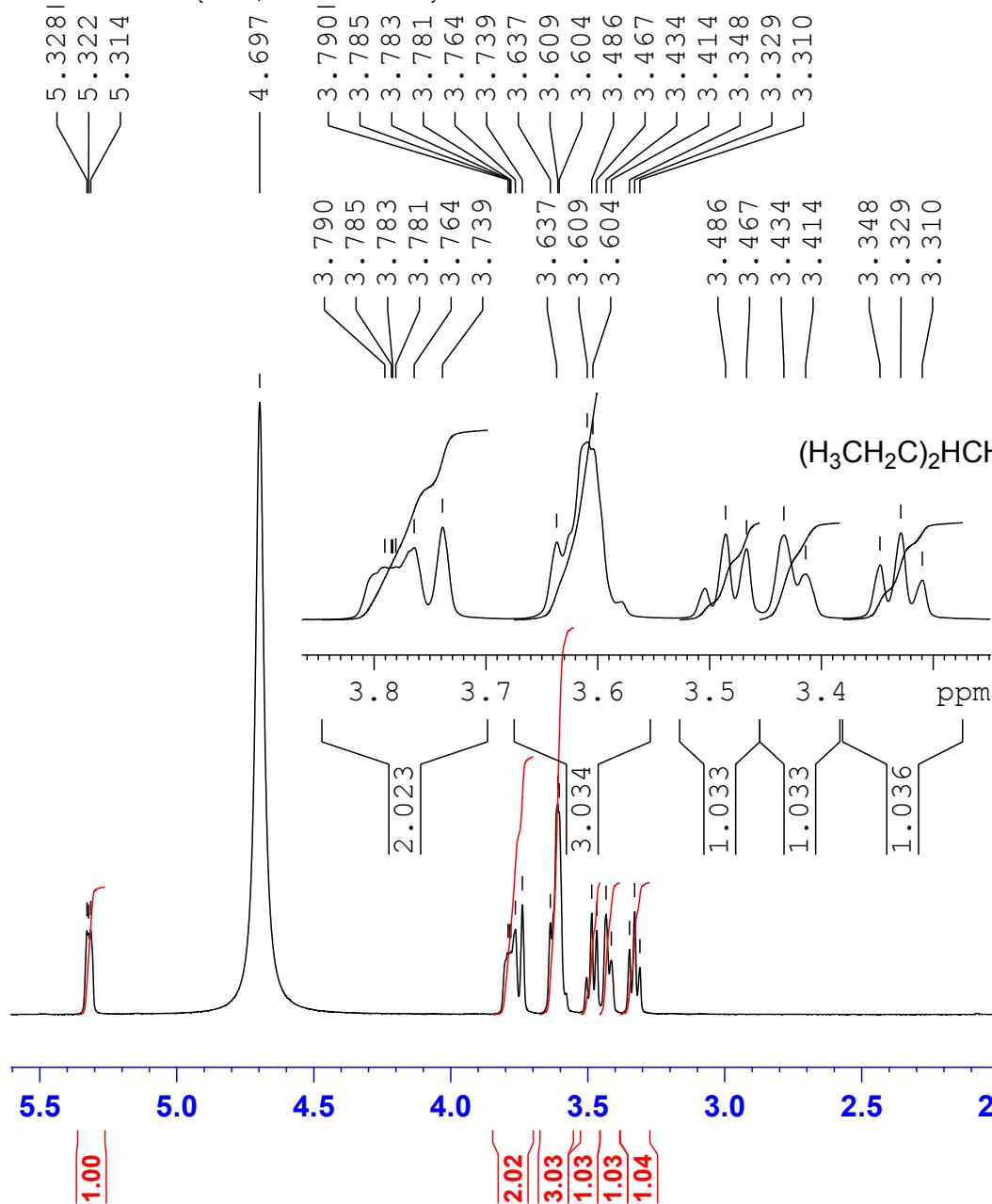
F2 - Processing parameters  
SI 65536  
SF 202.4563430 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 1.00



MH

130

1d\_1H D2O {C:\nmr\_users} huestis 14

5.328  
5.322  
5.314

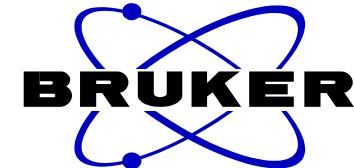
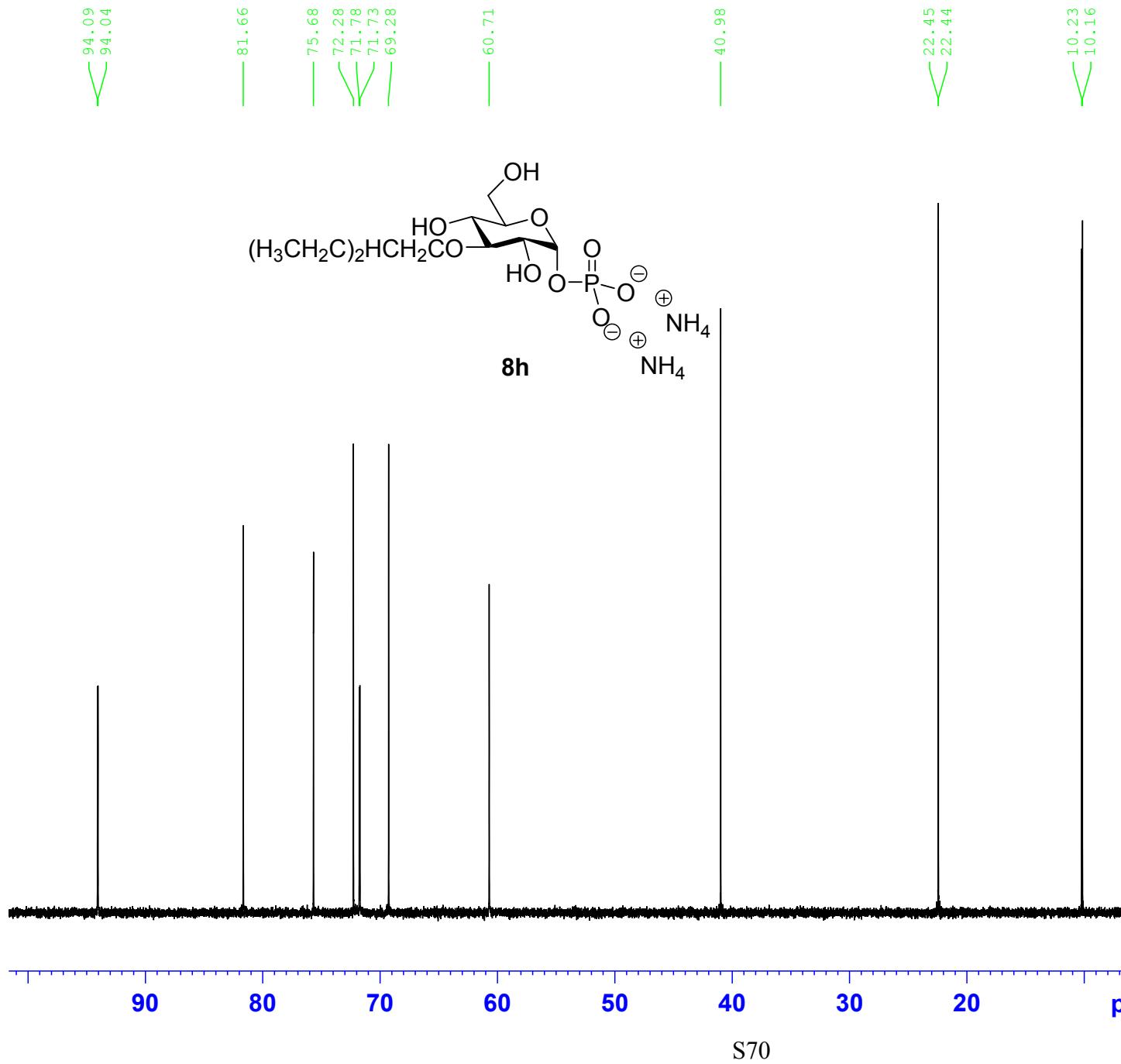
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 PULPROG zg30  
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 SOLVENT D2O  
 NS 32  
 DS 2  
 SWH 10000.000 Hz  
 FIDRES 0.152588 Hz  
 AQ 3.2769001 sec  
 RG 57  
 DW 50.000 usec  
 DE 6.00 usec  
 TE 300.0 K  
 D1 1.00000000 sec  
 TDO 1

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 9.80 usec  
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 SFO1 500.1332698 MHz

F2 - Processing parameters  
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 SF 500.1300190 MHz  
 WDW no  
 SSB 0  
 LB 0.00 Hz  
 GB 0  
 PC 0.75

MH\_130  
1d\_13C{1H}\_n D2O {C:\nmr\_users} huestis 14



Current Data Parameters  
NAME MH\_130  
EXPNO 11  
PROCNO 1

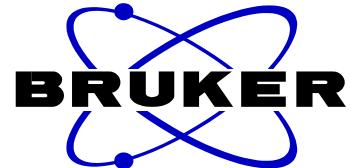
F2 - Acquisition Parameters  
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PULPROG zgpg30  
TD 54048  
SOLVENT D2O  
NS 1024  
DS 4  
SWH 33783.785 Hz  
FIDRES 0.625070 Hz  
AQ 0.7999752 sec  
RG 9195.2  
DW 14.800 usec  
DE 21.14 usec  
TE 300.4 K  
D1 1.00000000 sec  
d11 0.03000000 sec  
DELTA 0.89999998 sec  
TD0 4

===== CHANNEL f1 =====  
NUC1 <sup>13</sup>C  
P1 7.90 usec  
PL1 0.00 dB  
SFO1 125.7716273 MHz

===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -2.00 dB  
PL12 15.72 dB  
PL13 18.64 dB  
SFO2 500.1322696 MHz

F2 - Processing parameters  
SI 65536  
SF 125.7577930 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 0.75

MH\_130  
1d\_31P{1H} D2O {C:\nmr\_users} huestis 14



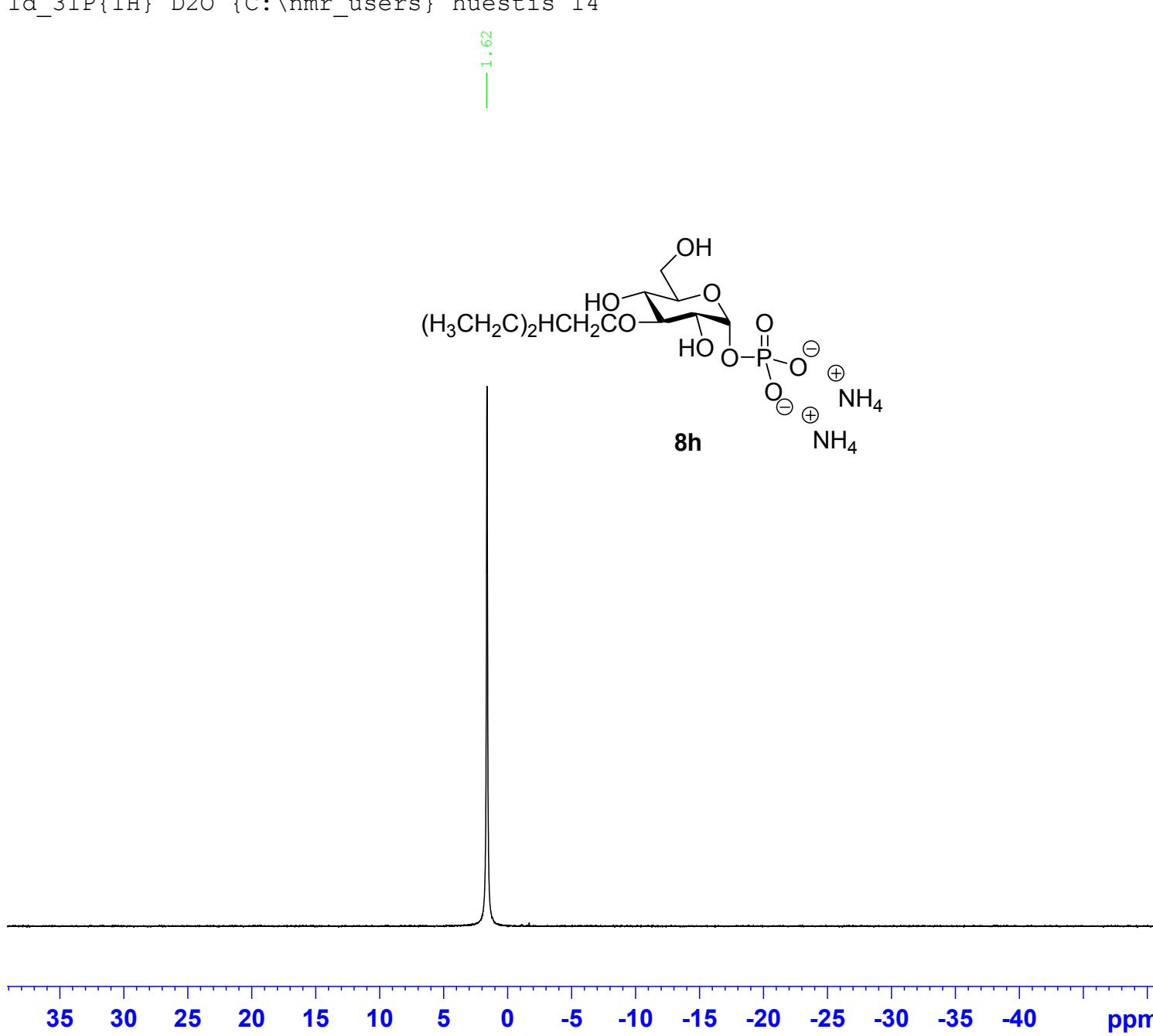
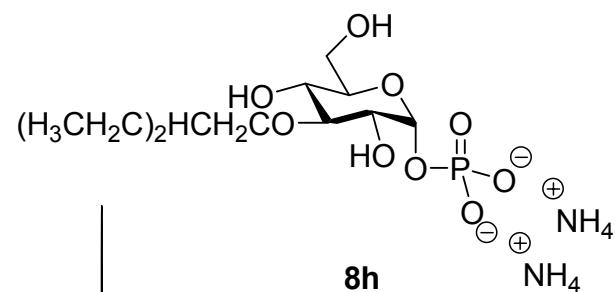
Current Data Parameters  
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PROCNO 1

F2 - Acquisition Parameters  
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Time 2.33  
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PROBHD 5 mm BBO BB-1H  
PULPROG zgpg30  
TD 65536  
SOLVENT D2O  
NS 512  
DS 4  
SWH 98039.219 Hz  
FIDRES 1.495960 Hz  
AQ 0.3342887 sec  
RG 8192  
DW 5.100 usec  
DE 7.29 usec  
TE 300.4 K  
D1 1.0000000 sec  
d11 0.0300000 sec  
DELTA 0.8999998 sec  
TD0 2

===== CHANNEL f1 =====  
NUC1 31P  
P1 8.41 usec  
PL1 0.50 dB  
SFO1 202.4664658 MHz

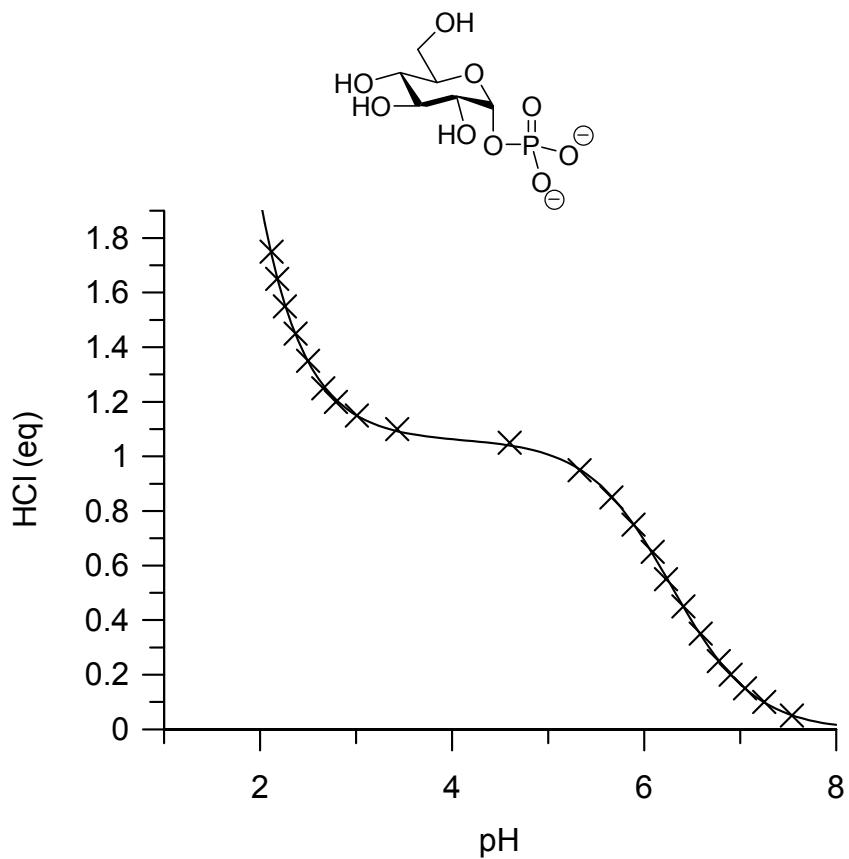
===== CHANNEL f2 =====  
CPDPRG2 waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -2.00 dB  
PL12 15.72 dB  
PL13 18.64 dB  
SFO2 500.1322696 MHz

F2 - Processing parameters  
SI 65536  
SF 202.4563430 MHz  
WDW no  
SSB 0  
LB 0.00 Hz  
GB 0  
PC 0.75



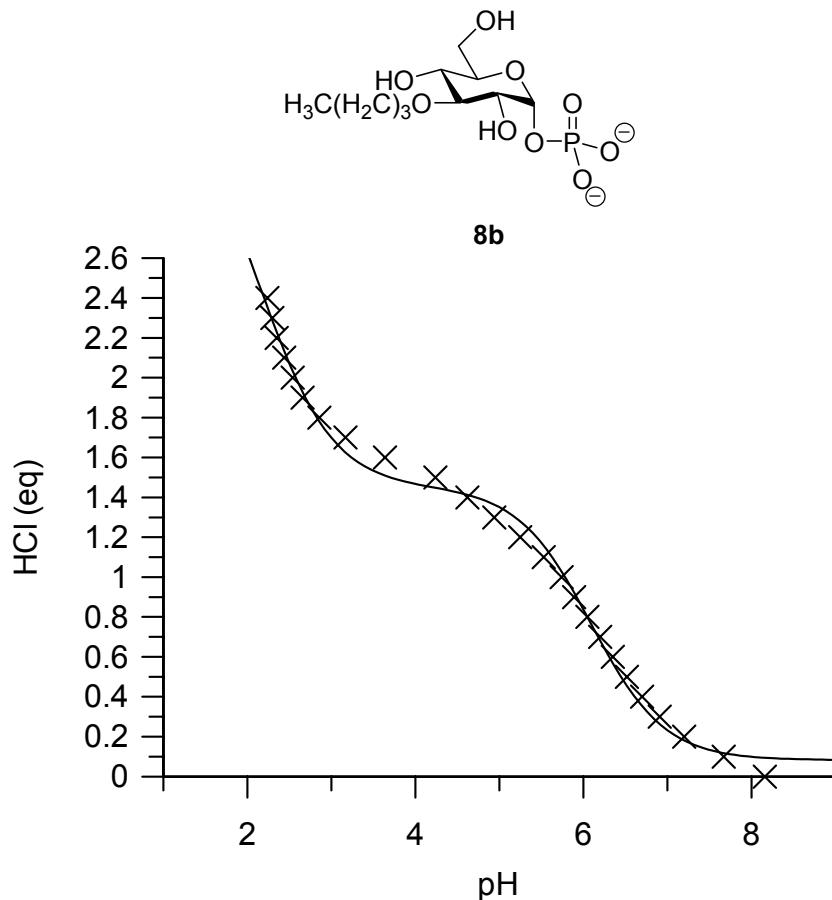
## Determination of pKa2 for $\alpha$ -D-glucopyranosyl phosphate and compounds 8b, 8d, 8e and 8g

pKa determination of  
 $\alpha$ -D-glucopyranosyl phosphate



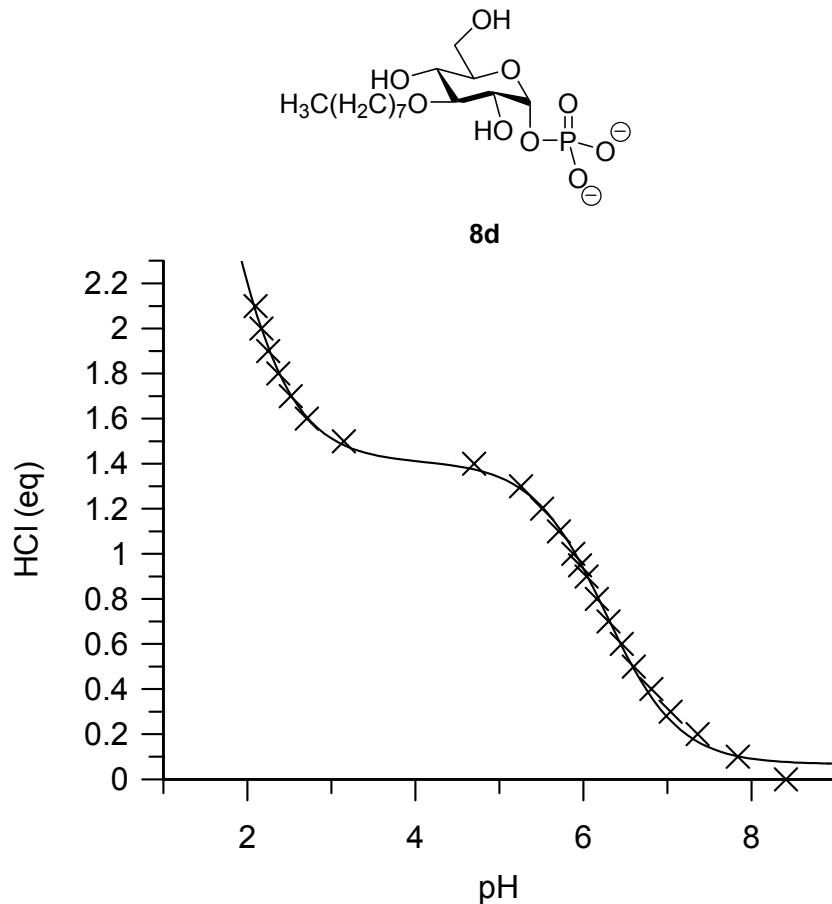
Parameter	Value	Std. Error
pKa 1	0.6783	0.0547
Lower Limit	20.5268	2.2855
Middle Limit	1.0593	0.0031
Upper Limit	-0.0042	0.0048
pKa 2	6.2798	0.0081

pKa determination of  
**3-O-Butyl- $\alpha$ -D-glucopyranosyl phosphate**



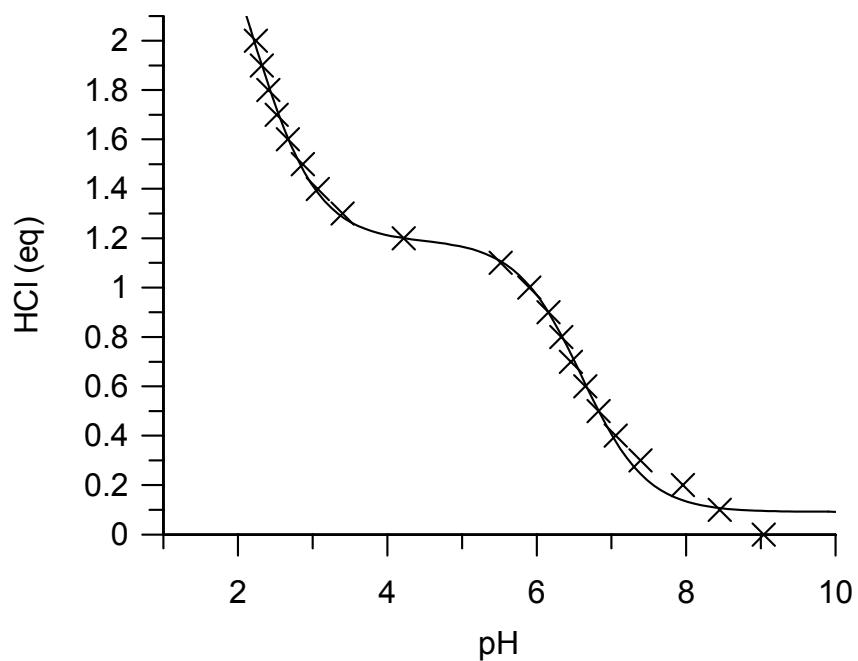
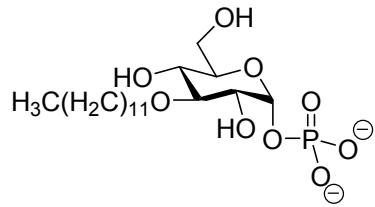
Parameter	Value	Std. Error
pKa 1	2.1514	0.1808
Lower Limit	3.4621	0.4900
Middle Limit	1.4506	0.0281
Upper Limit	0.0822	0.0304
pKa 2	6.0911	0.0461

pKa determination of  
3-O-Octyl- $\alpha$ -D-glucopyranosyl phosphate



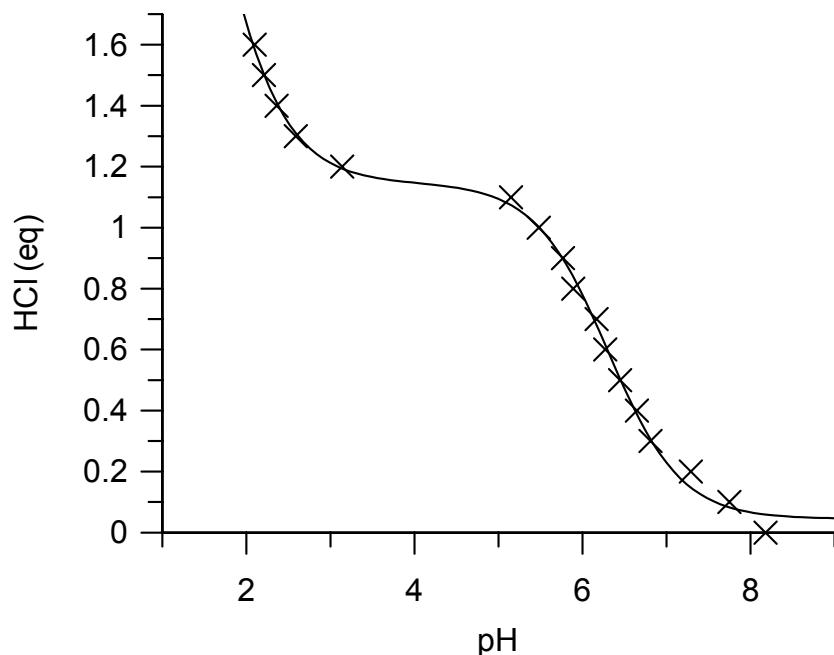
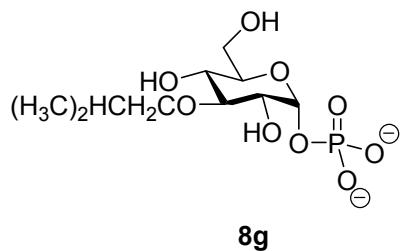
Parameter	Value	Std. Error
pKa 1	1.5658	0.0856
Lower Limit	4.3730	0.4395
Middle Limit	1.4079	0.0152
Upper Limit	0.0668	0.0156
pKa 2	6.2651	0.0232

pKa determination of  
**3-O-Dodecyl- $\alpha$ -D-glucopyranosyl phosphate**



Parameter	Value	Std. Error
pKa 1	2.1883	0.1443
Lower Limit	2.8675	0.3209
Middle Limit	1.1894	0.0234
Upper Limit	0.0913	0.0202
pKa 2	6.6034	0.0411

*pKa* determination of  
**3-O-(2-methylpropyl)- $\alpha$ -D-glucopyranosyl  
phosphate**

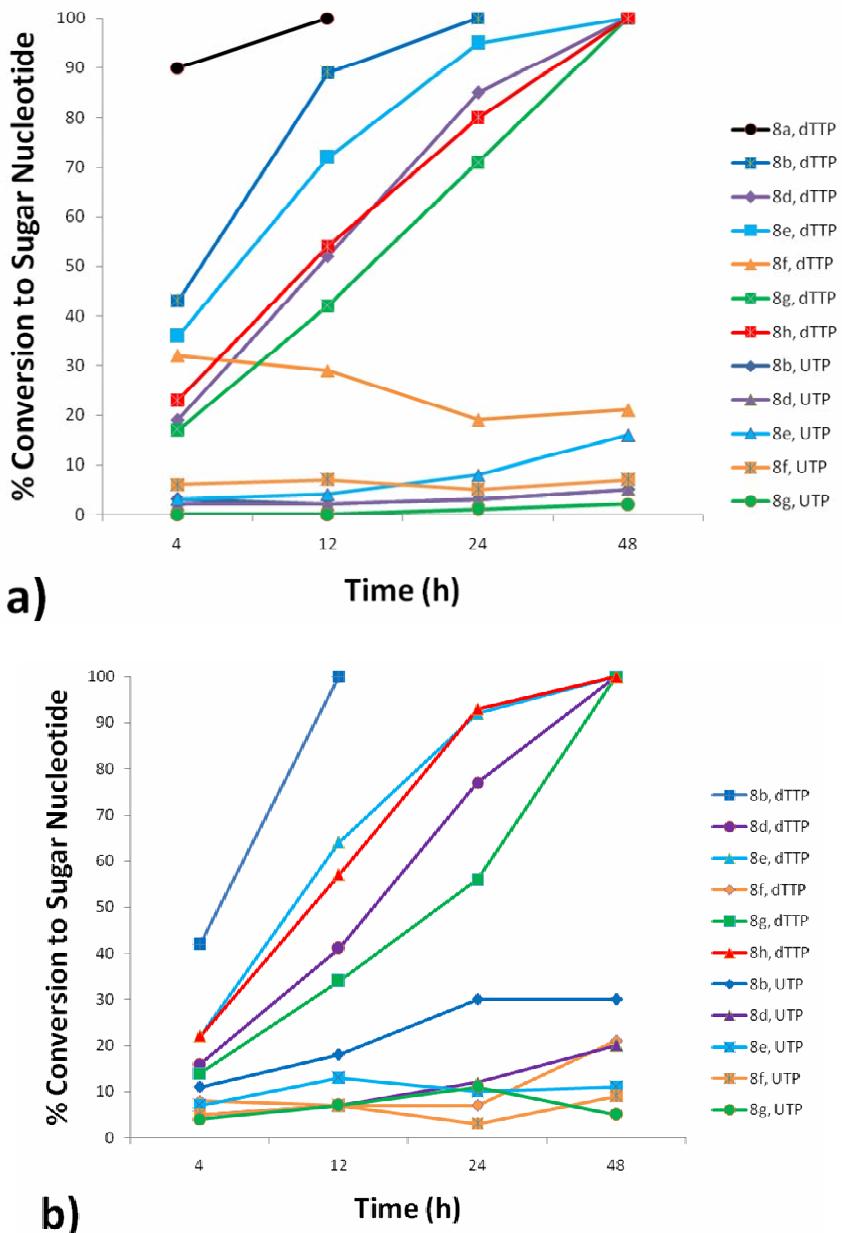


Parameter	Value	Std. Error
pKa 1	1.4930	0.1630
Lower Limit	3.3633	0.6249
Middle Limit	1.1457	0.0193
Upper Limit	0.0446	0.0186
pKa 2	6.3039	0.0349

## Enzymatic Synthesis

### General Procedure for nucleotidyltransferase-catalyzed synthesis of sugar nucleotides (9a-h, 10a-h)

Enzymatic reactions were performed by the method of Timmons ( $\text{MgCl}_2$ , 1.0 mM NTP, 2.0 mM S1P, 0.5EU inorganic pyrophosphatase, 2EU nucleotidyltransferase in Tris-HCl buffer – total 50  $\mu\text{L}$ )<sup>6</sup> except that  $\text{MgCl}_2$  was used at a final concentration of 5.5 mM and incubations were performed at 41°C.



**Figure S1. a)** Selected data for nucleotidyltransferase-catalyzed conversion of sugar-1-phosphates (8a-h) to sugar nucleotides (9a-h, 10a-h) using a) Cps2L b) RmlA3.

<sup>6</sup> Timmons, S. C.; Mosher, R. H.; Knowles, S. A.; Jakeman, D. L. *Org. Lett.* **2007**, 9, 857-860.

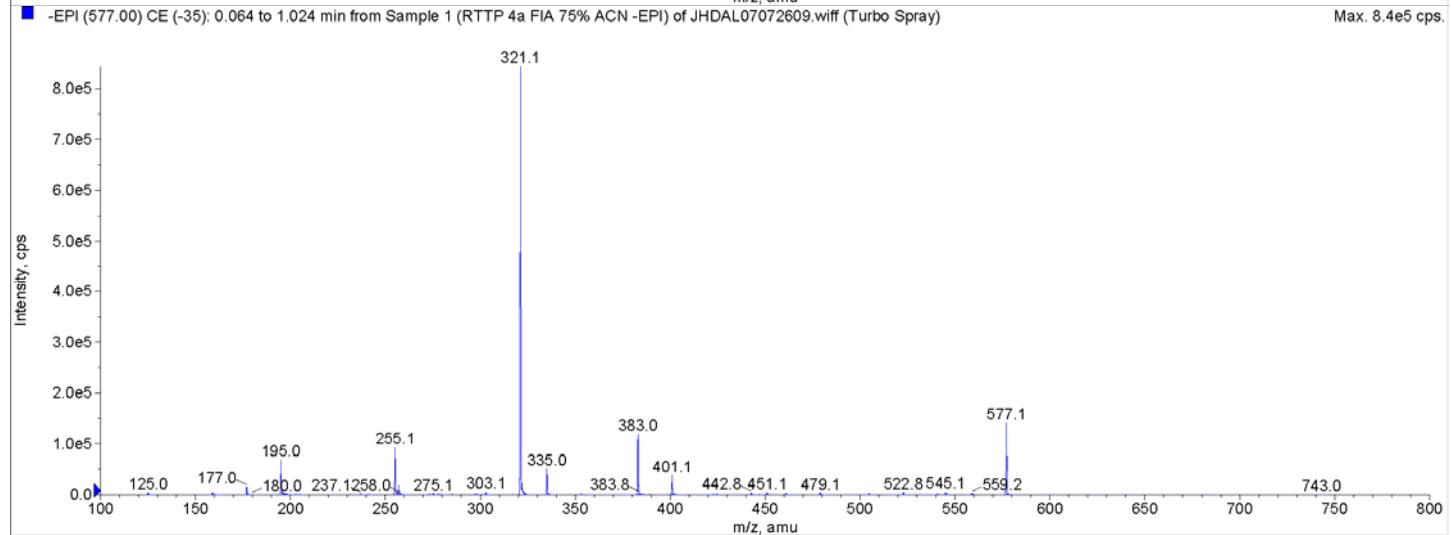
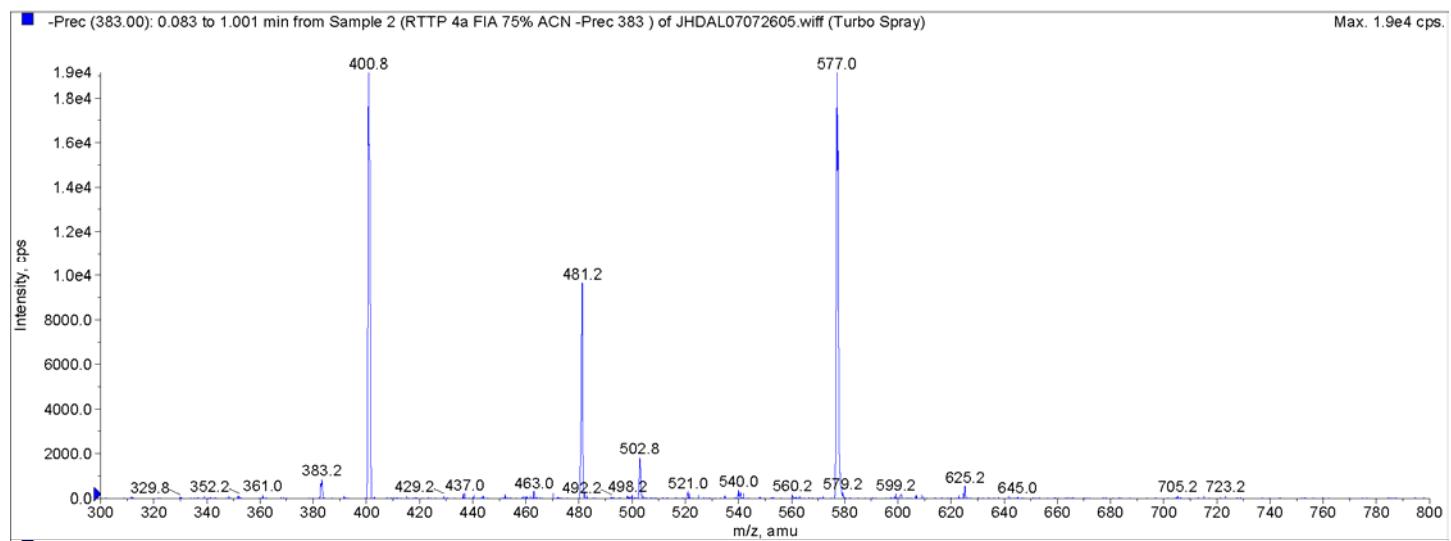
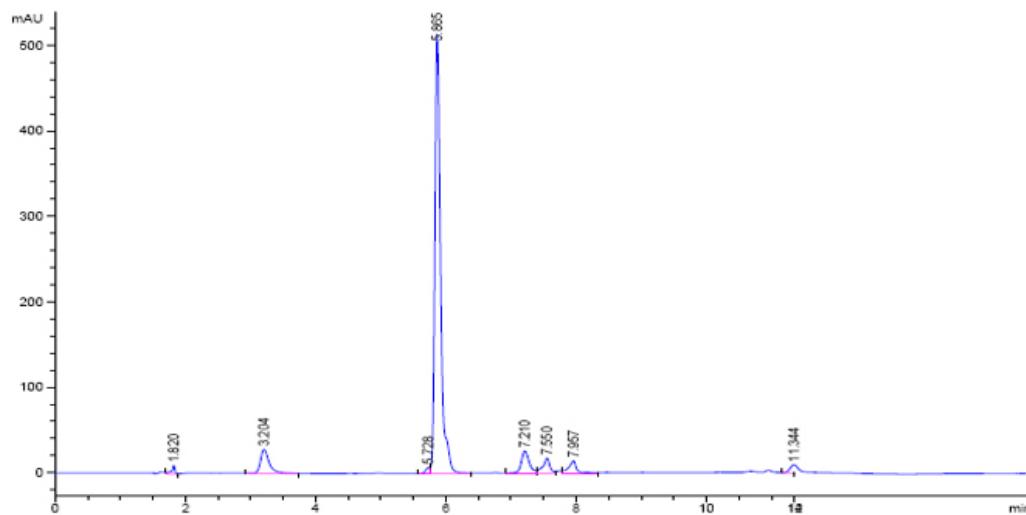
Enzyme	Sugar-1-Phosphate	NTP	% Conv. (4 hr)	% Conv. (12 hr)	% Conv. (24 hr)	% Conv (48 hr)	Sugar Nucleotide retention time (min)
Cps2L	<b>8a</b>	dTTP	90	100			5.82
Cps2L	<b>8b</b>	dTTP	43	89	100		6.73
Cps2L	<b>8c</b>	dTTP	c	c	c	c	7.50
Cps2L	<b>8d</b>	dTTP	19	52	85	100	8.28
Cps2L	<b>8e</b>	dTTP	36	72	95	100	9.77
Cps2L	<b>8f</b>	dTTP	32	29	19	21	9.25 <sup>a</sup>
Cps2L	<b>8g</b>	dTTP	17	42	71	100	6.75
Cps2L	<b>8h</b>	dTTP	23	54	80	100	7.35
Cps2L	<b>8a</b>	UTP	b	b	b	b	5.75
Cps2L	<b>8b</b>	UTP	3	2	3	5	6.69
Cps2L	<b>8c</b>	UTP	c	c	6	10	7.48
Cps2L	<b>8d</b>	UTP	2	2	3	5	8.26
Cps2L	<b>8e</b>	UTP	3	4	8	16	9.77
Cps2L	<b>8f</b>	UTP	6	7	5	7	9.27 <sup>a</sup>
Cps2L	<b>8g</b>	UTP	0	0	1	2	6.66
Cps2L	<b>8h</b>	UTP	1	c	c	c	7.32
RmlA3	<b>8a</b>	dTTP	28	b	b	b	5.82
RmlA3	<b>8b</b>	dTTP	42	100			6.73
RmlA3	<b>8c</b>	dTTP	c	c	c	c	7.50
RmlA3	<b>8d</b>	dTTP	16	41	77	100	8.28
RmlA3	<b>8e</b>	dTTP	22	64	92	100	9.77
RmlA3	<b>8f</b>	dTTP	8	7	7	21	9.25 <sup>a</sup>
RmlA3	<b>8g</b>	dTTP	14	34	56	100	6.75
RmlA3	<b>8h</b>	dTTP	22	57	93	100	7.35
RmlA3	<b>8a</b>	UTP	b	b	7	b	5.75
RmlA3	<b>8b</b>	UTP	11	18	30	breakdown	6.69
RmlA3	<b>8c</b>	UTP	c	9	13	14	7.48
RmlA3	<b>8d</b>	UTP	5	7	12	20	8.26
RmlA3	<b>8e</b>	UTP	7	13	10	11	9.77
RmlA3	<b>8f</b>	UTP	5	7	3	9	9.27 <sup>a</sup>
RmlA3	<b>8g</b>	UTP	4	7	11	5	6.66
RmlA3	<b>8h</b>	UTP	5	c	c	c	7.32

**Table S1.** Nucleotidyltransferase-catalyzed conversion of sugar-1-phosphates (**8a-h**) to sugar nucleotides (**9a-h**, **10a-h**).

<sup>a</sup>alternate buffer gradient used (see General Methods). <sup>b</sup>inaccurate integration due to co-elution with NDP. <sup>c</sup>inaccurate integration due to co-elution with NTP.

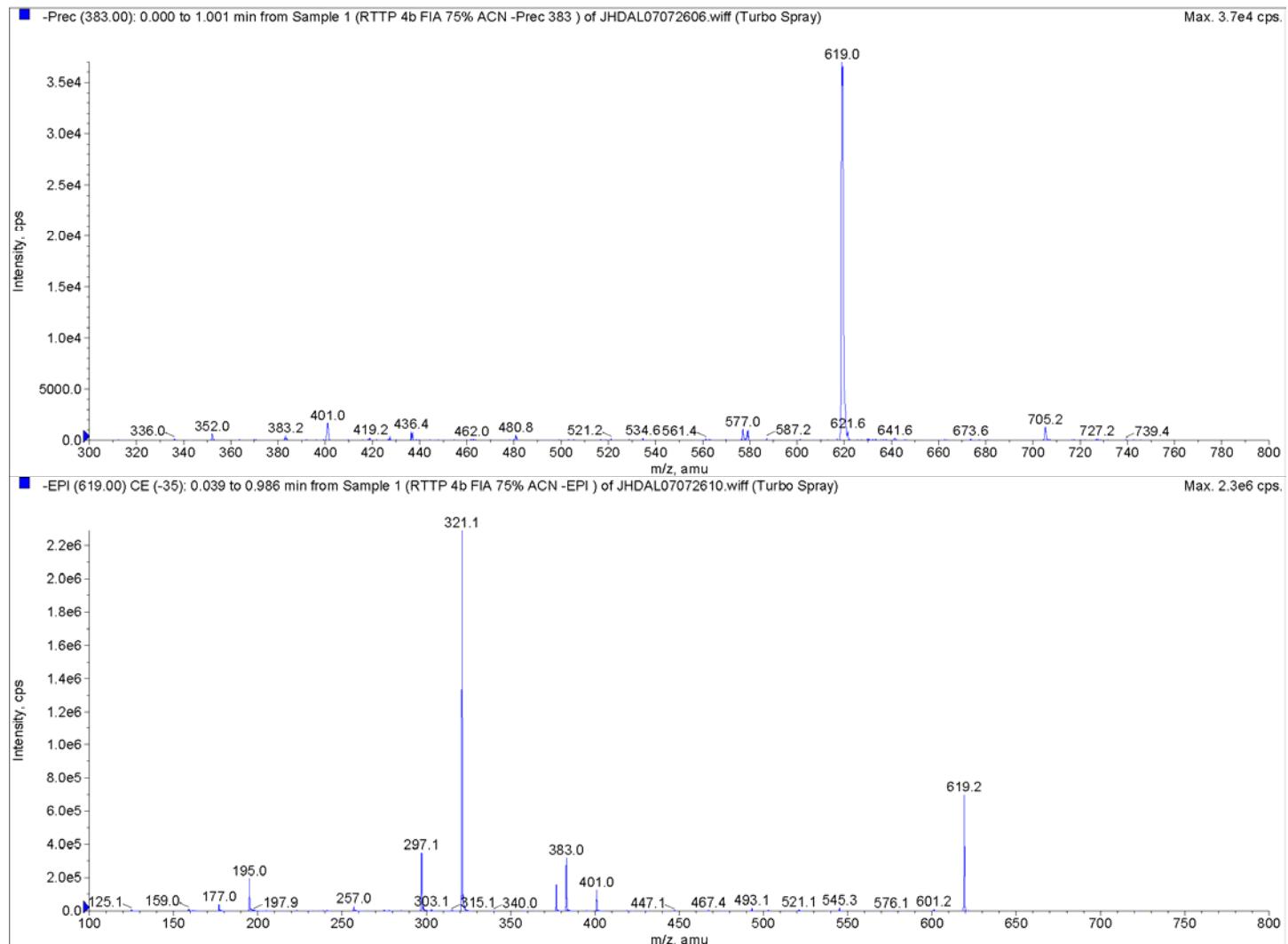
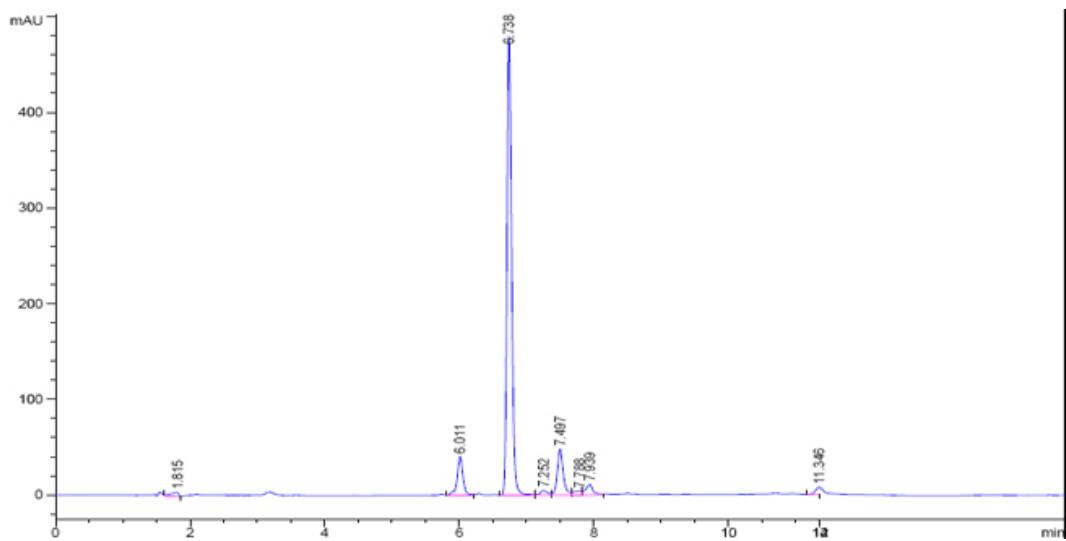
### 3-O-Methyl-dTDP- $\alpha$ -D-glucose (9a)

LRMS  $m/z$  calcd for  $C_{17}H_{26}N_2O_{16}P_2^{2-}$  [M+H] $^-$ : 577.1. Found 577.0. EPI fragments: 321.1.



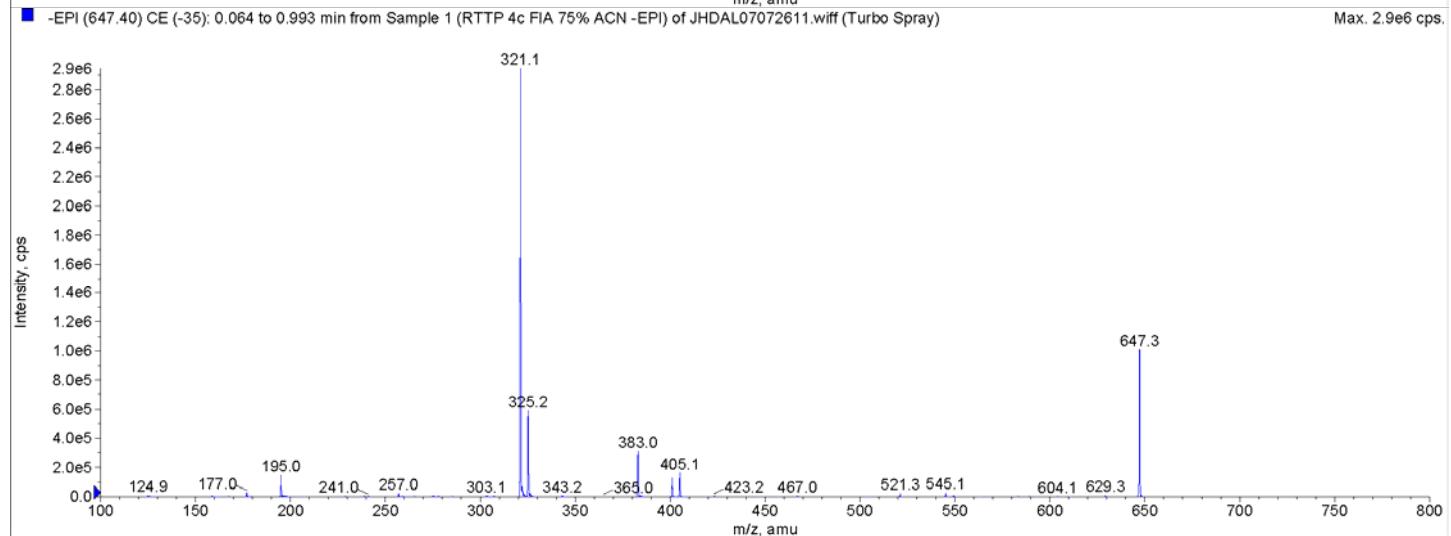
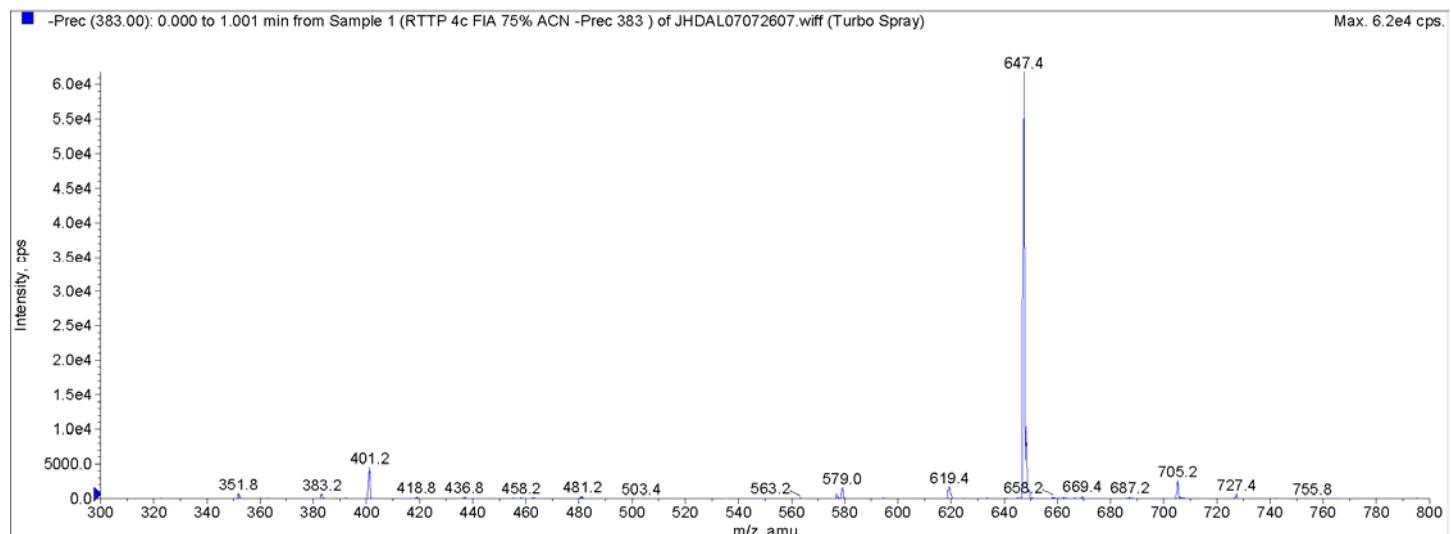
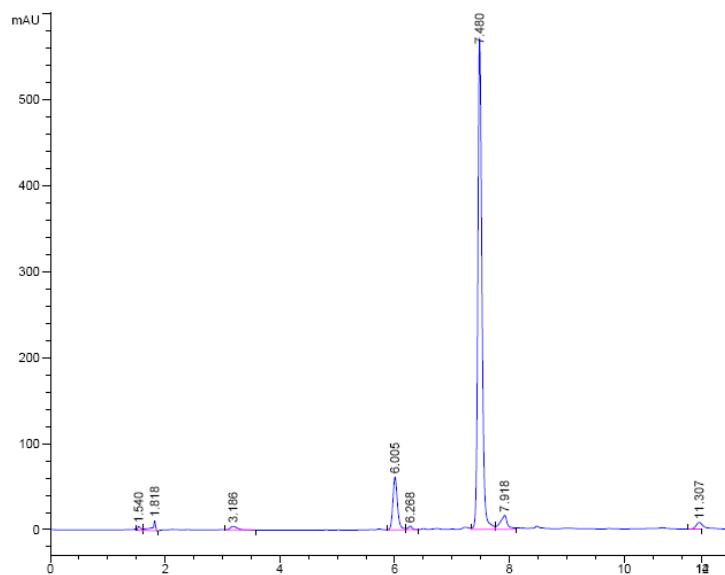
### 3-O-Butyl-dTDP- $\alpha$ -D-glucose (9b)

LRMS  $m/z$  calcd for  $C_{20}H_{32}N_2O_{16}P_2^{2-}$  [M+H] $^-$ : 619.1. Found 619.0. EPI fragments: 321.1.



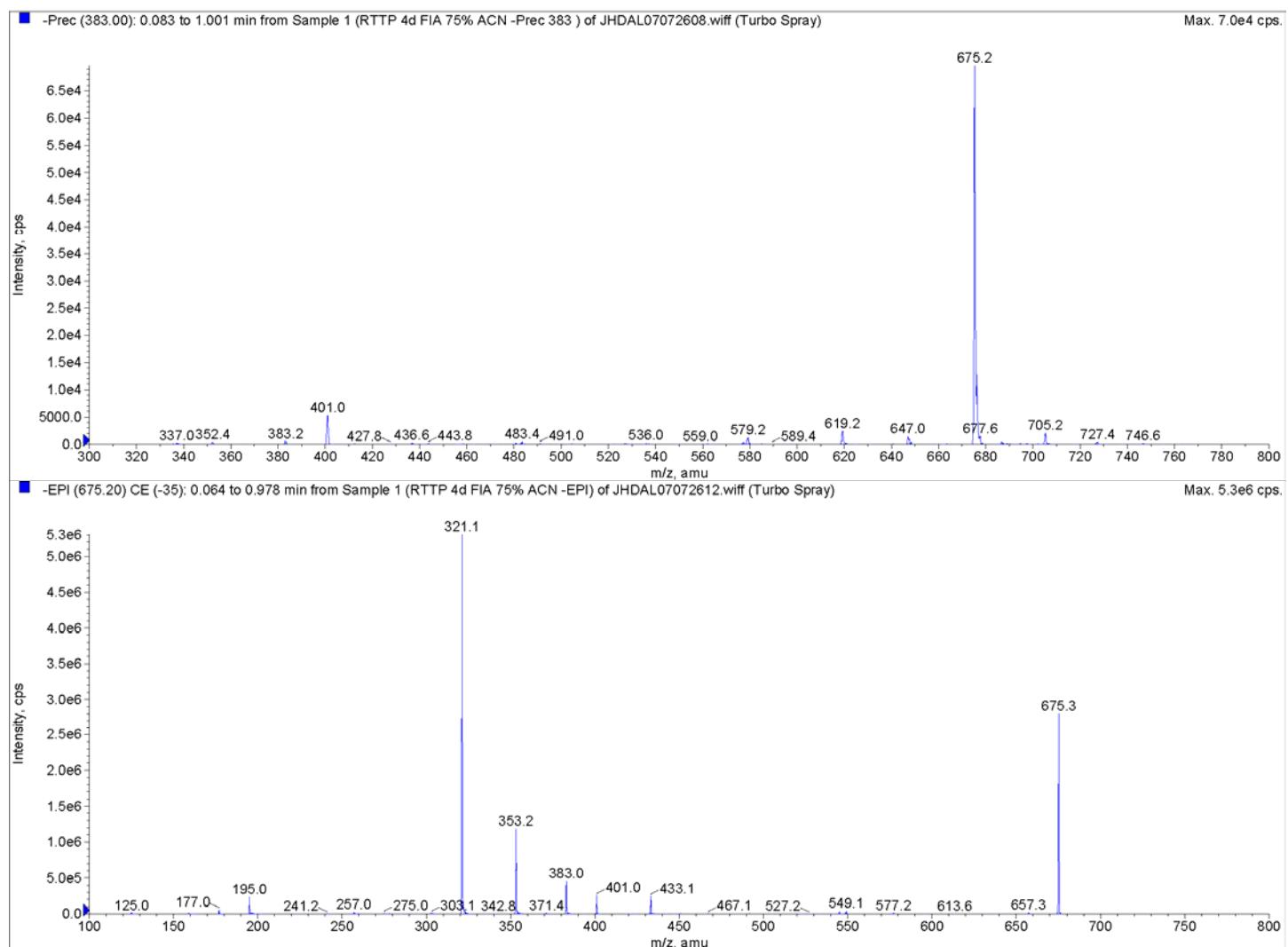
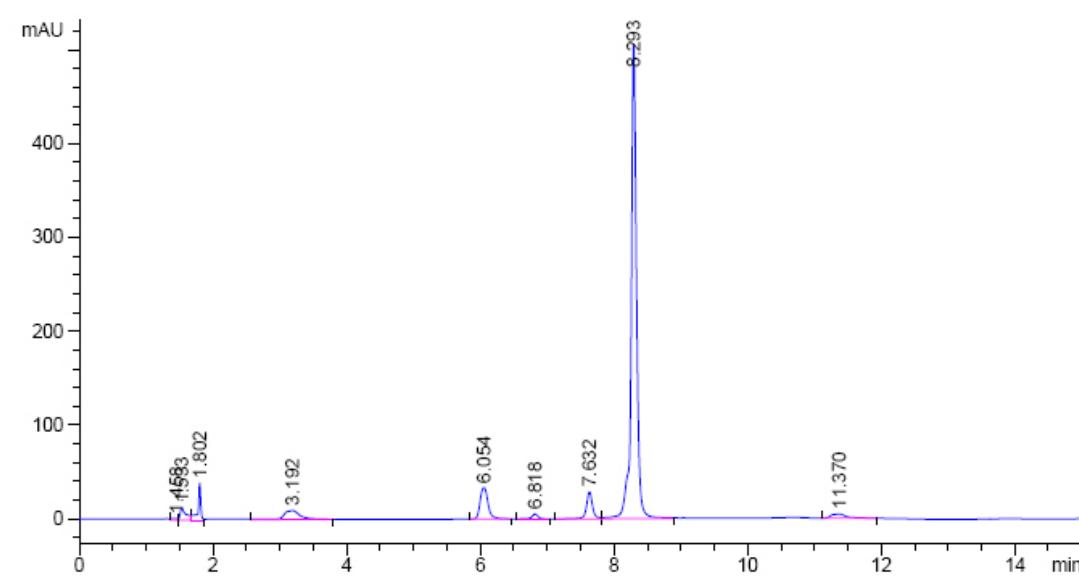
### 3-O-Hexyl-dTDP- $\alpha$ -D-glucose (9c)

LRMS  $m/z$  calcd for  $C_{22}H_{36}N_2O_{16}P_2^{2-}$  [M+H] $^-$ : 647.2. Found 647.4. EPI fragments: 321.1.



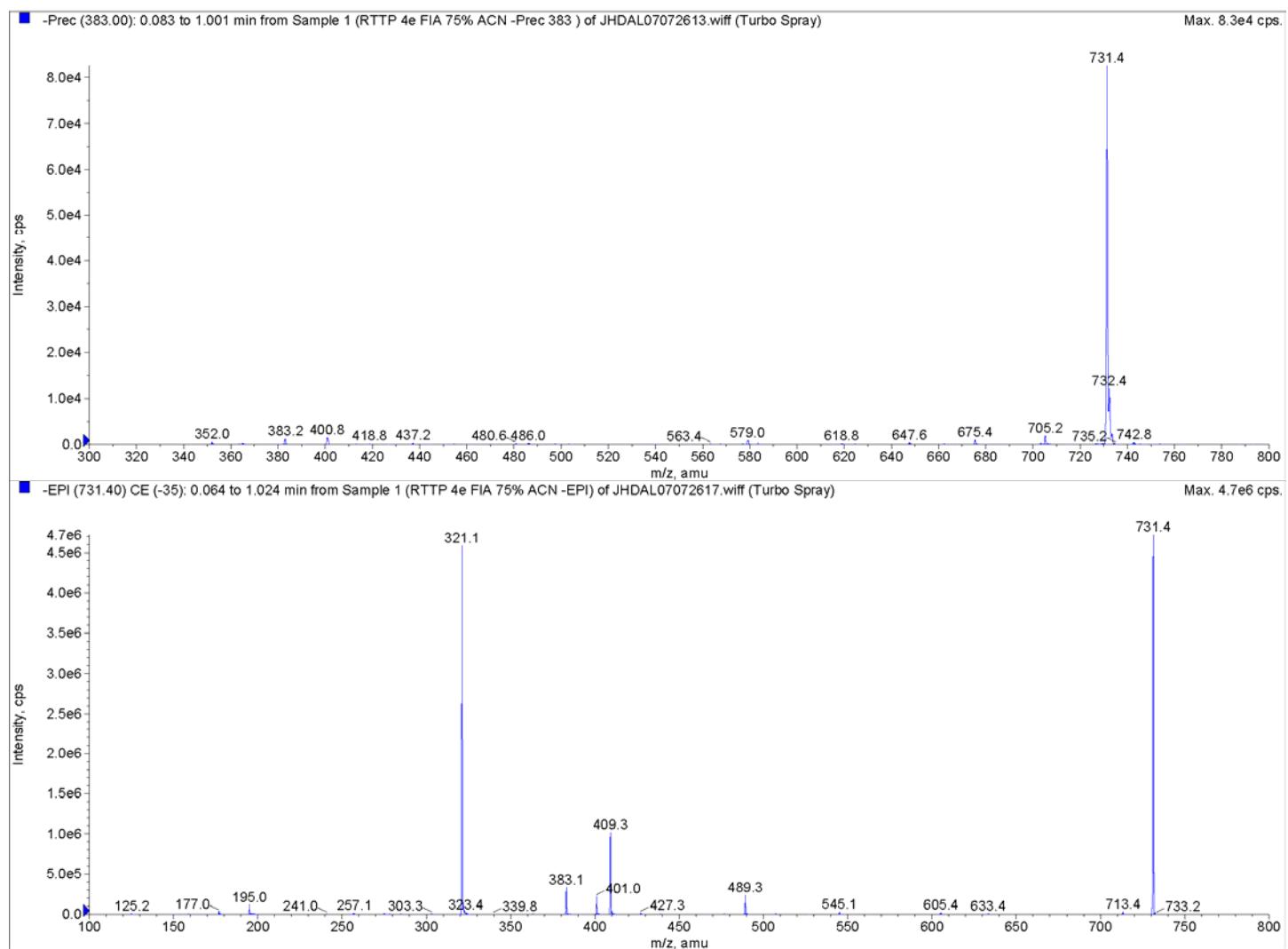
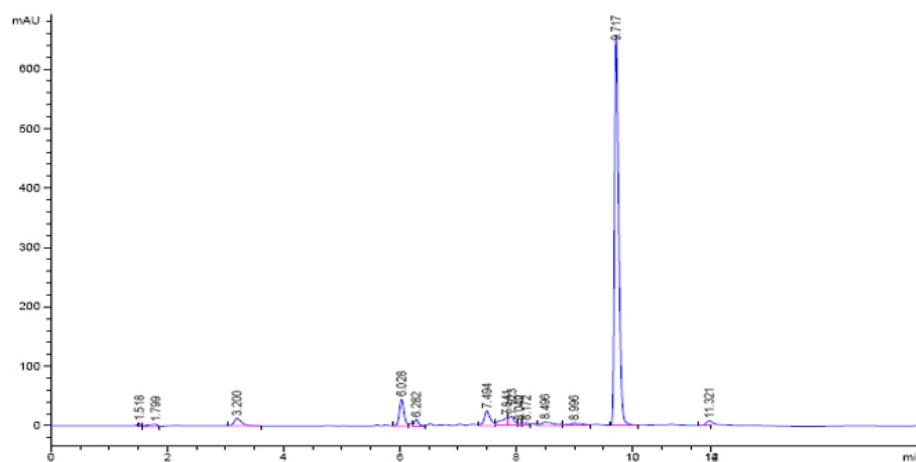
### 3-O-Octyl-dTDP- $\alpha$ -D-glucose (9d)

LRMS  $m/z$  calcd for  $C_{24}H_{40}N_2O_{16}P_2^{2-}$  [M+H] $^-$ : 675.2. Found 675.2. EPI fragments: 321.1.



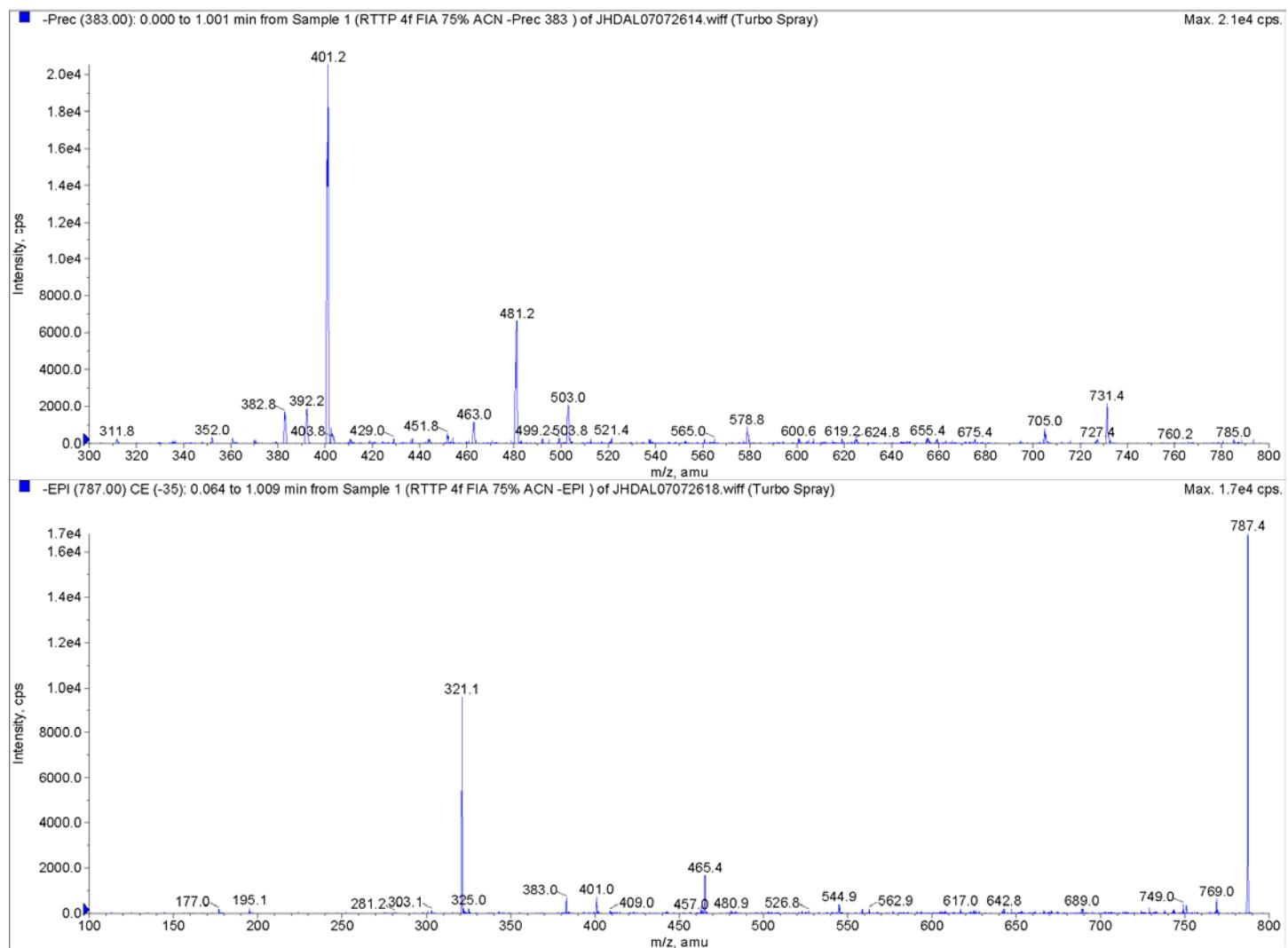
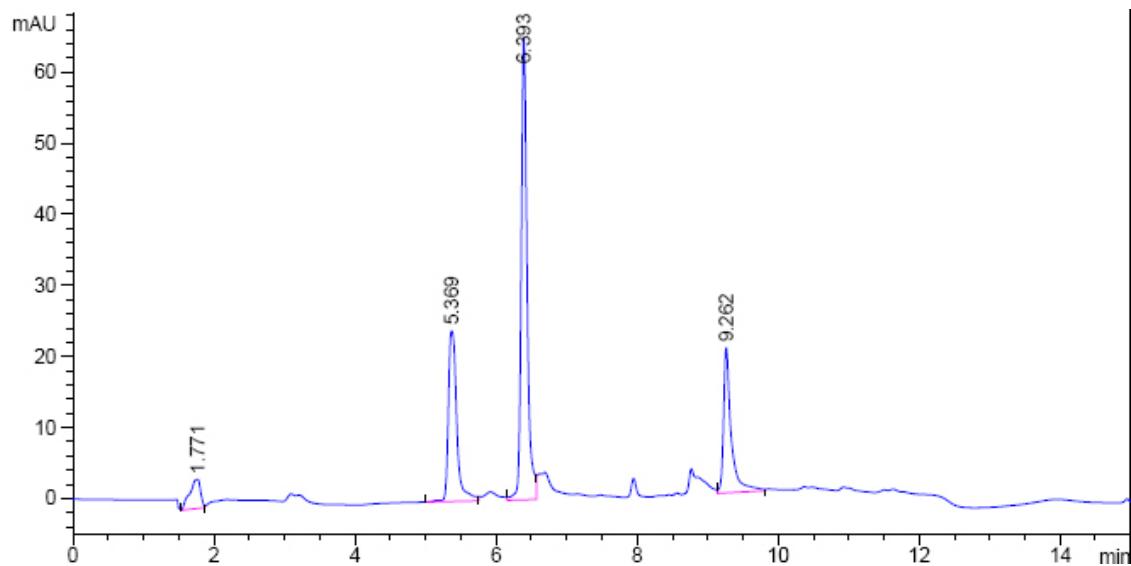
### 3-O-Dodecyl-dTDP- $\alpha$ -D-glucose (9e)

LRMS  $m/z$  calcd for  $C_{28}H_{48}N_2O_{16}P_2^{2-}$  [M+H] $^-$ : 731.3. Found 731.4. EPI fragments: 321.1.



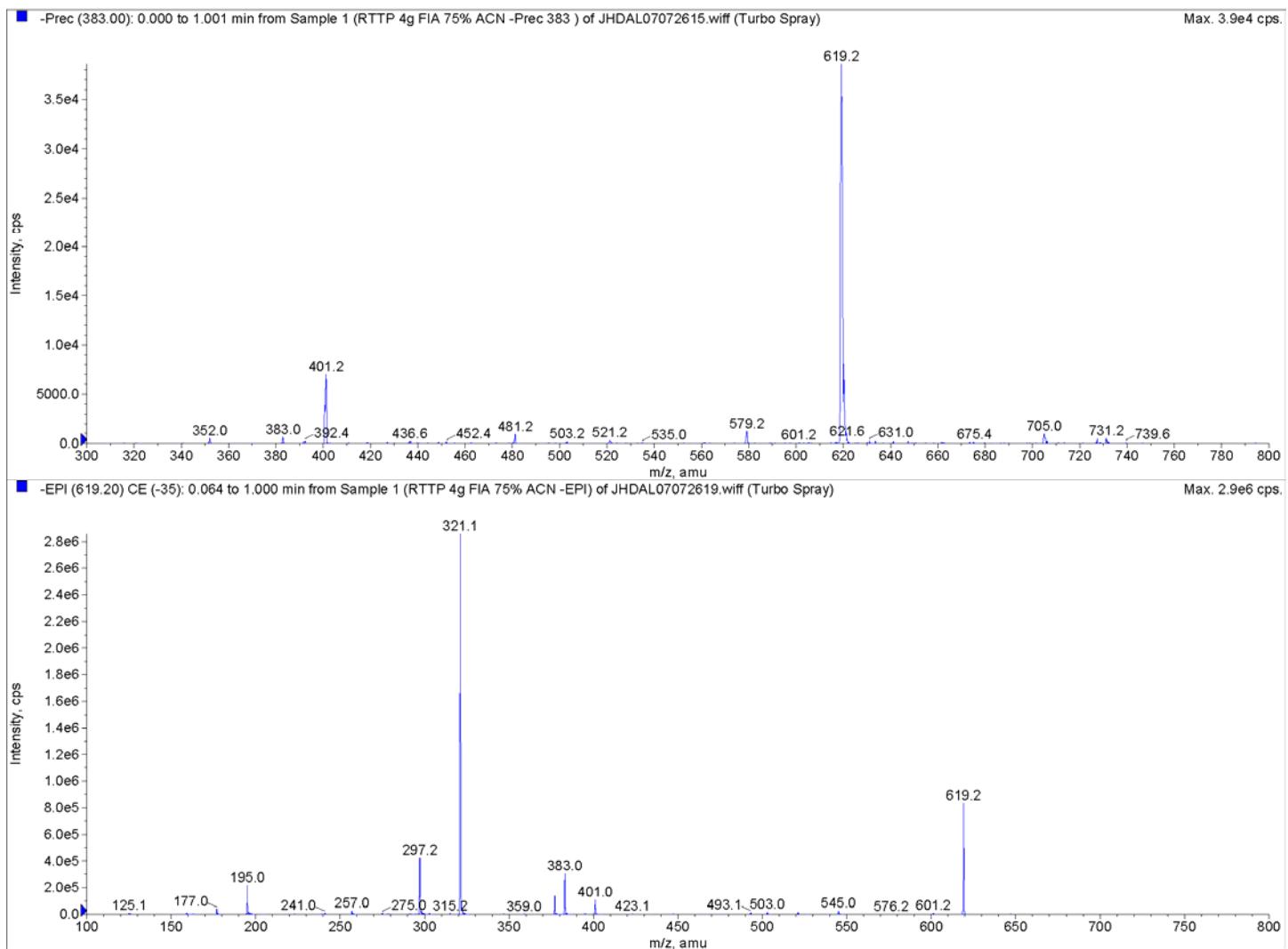
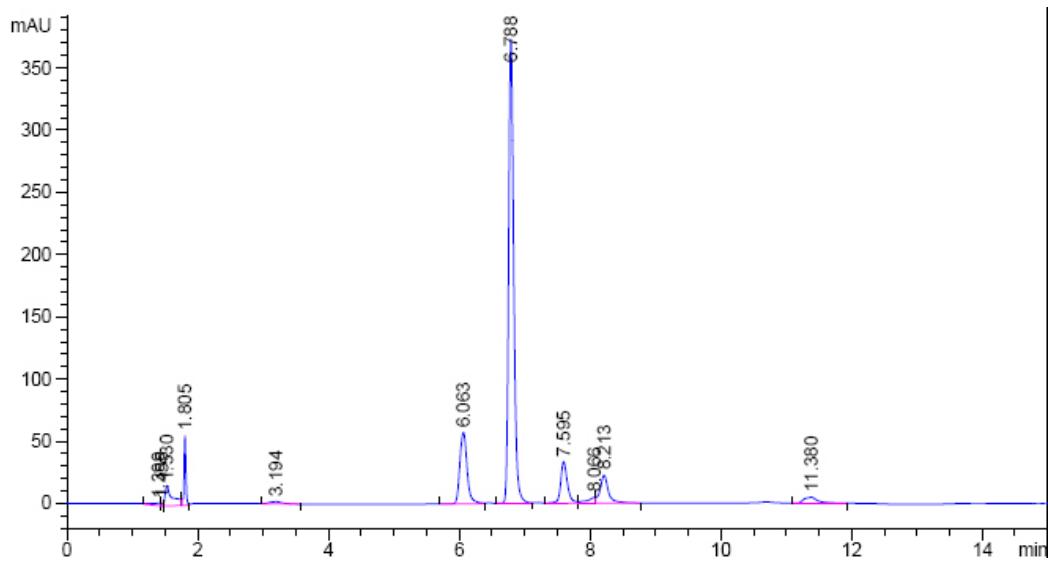
### 3-O-Hexadecyl-dTDP- $\alpha$ -D-glucose (9f)

LRMS  $m/z$  calcd for  $C_{32}H_{56}N_2O_{16}P_2^{2-}$  [M+H] $^-$ : 787.3. Found 787.4. EPI fragments: 321.1.



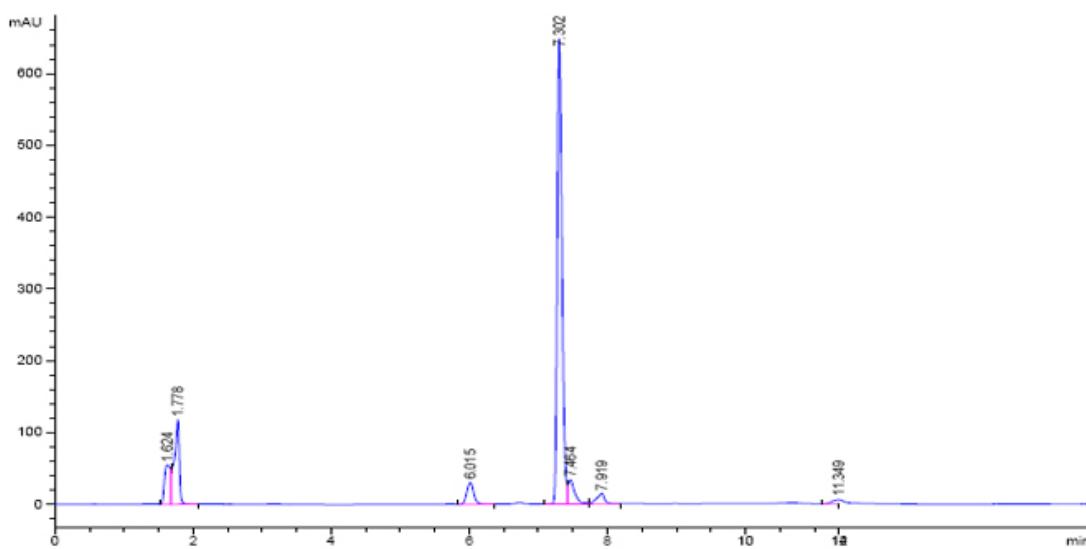
### 3-O-(2-Methylpropyl)-dTDP- $\alpha$ -D-glucose (9g)

LRMS  $m/z$  calcd for  $C_{20}H_{32}N_2O_{16}P_2^{2-}$  [M+H] $^-$ : 619.1. Found 619.2. EPI fragments: 321.1.

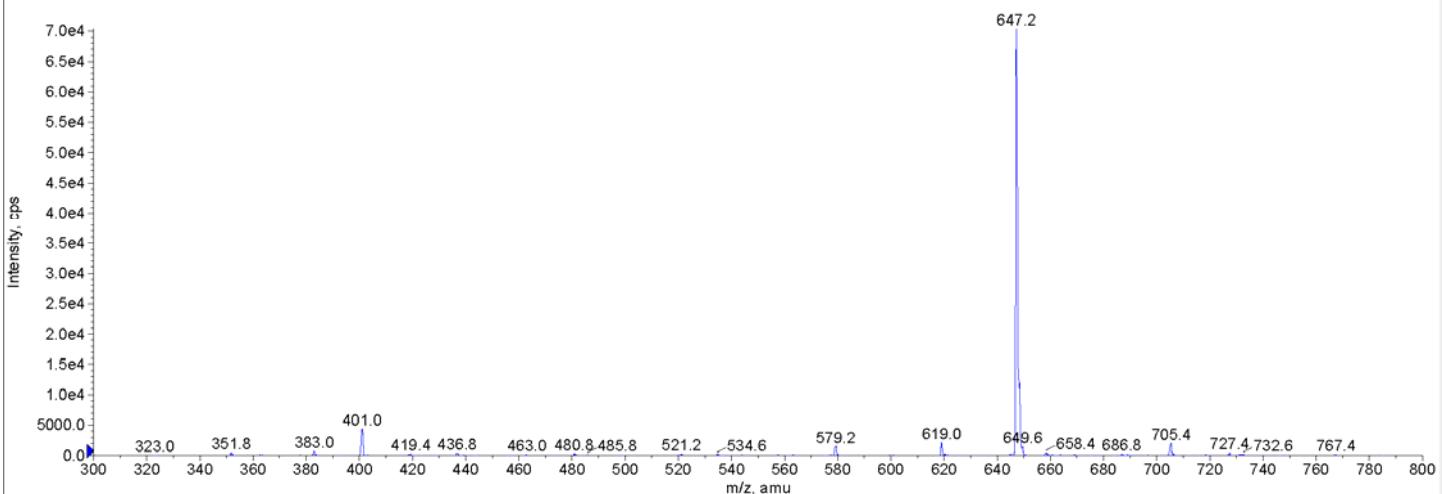


### 3-O-(2-Ethylbutyl)-dTDP- $\alpha$ -D-glucose (9h)

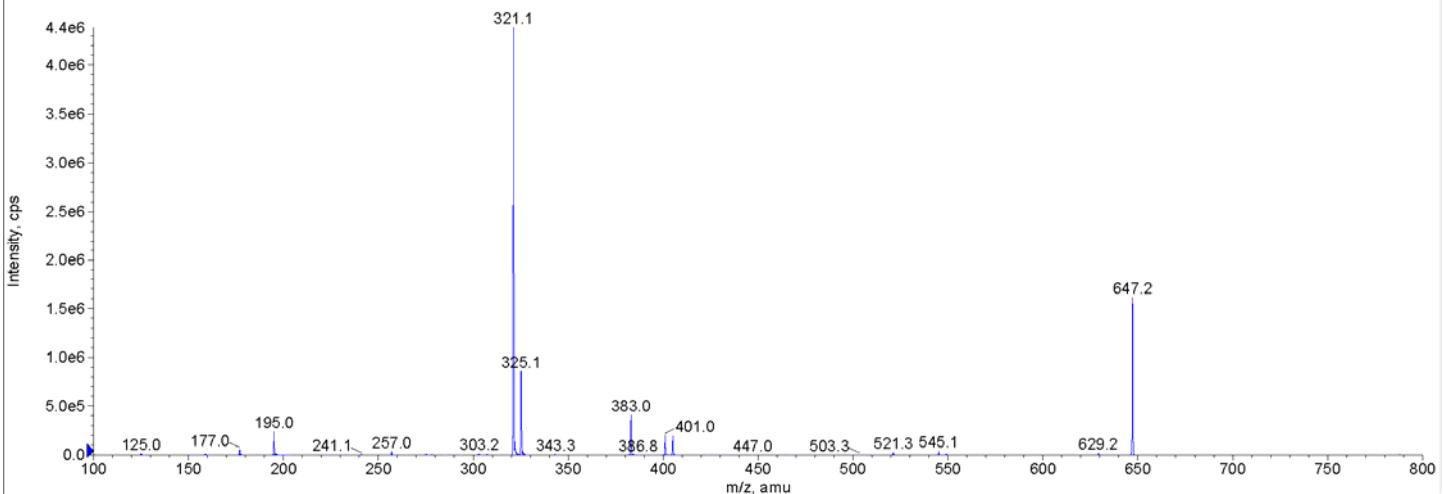
LRMS  $m/z$  calcd for  $C_{22}H_{36}N_2O_{16}P_2^{2-}$  [M+H] $^-$ : 647.2. Found 647.2. EPI fragments: 321.1.



-Prec (383.00): 0.083 to 1.001 min from Sample 1 (RTTP 4h FIA 75% ACN -Prec 383 ) of JHDAL07072616.wiff (Turbo Spray) Max. 7.0e4 cps.

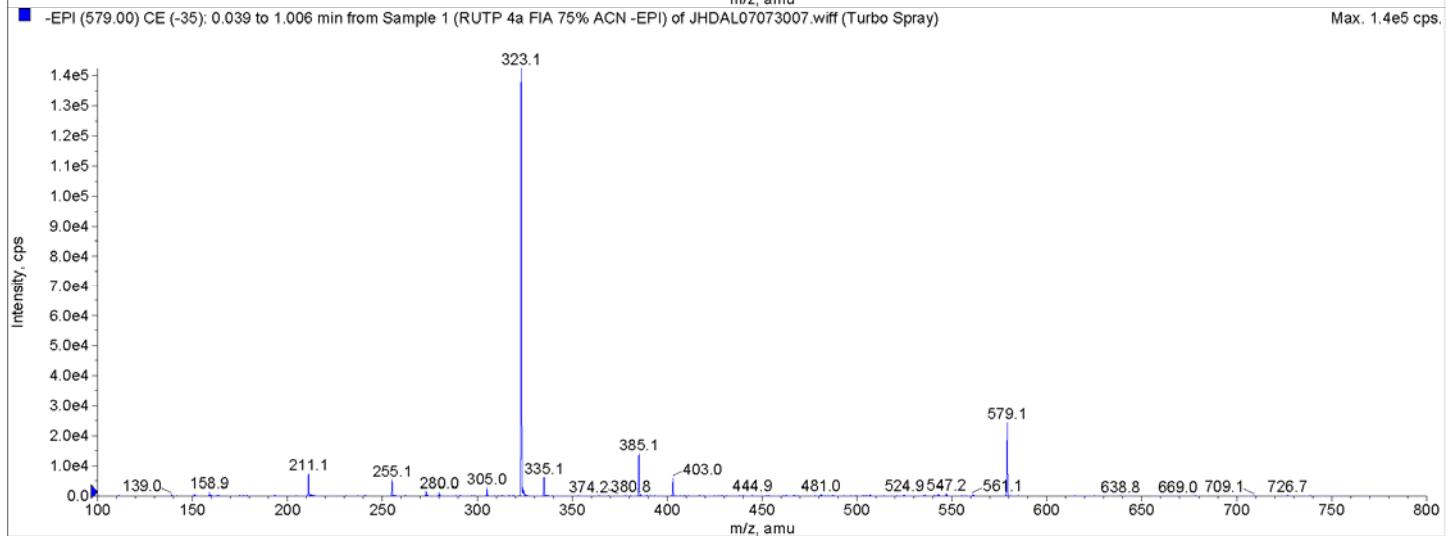
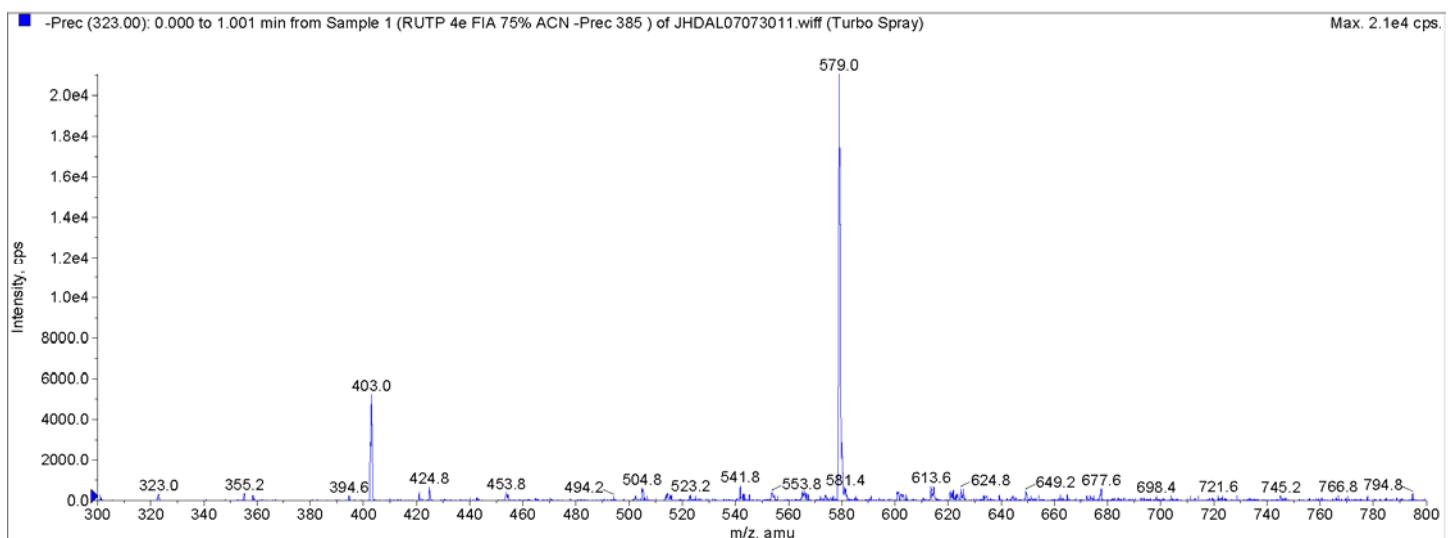
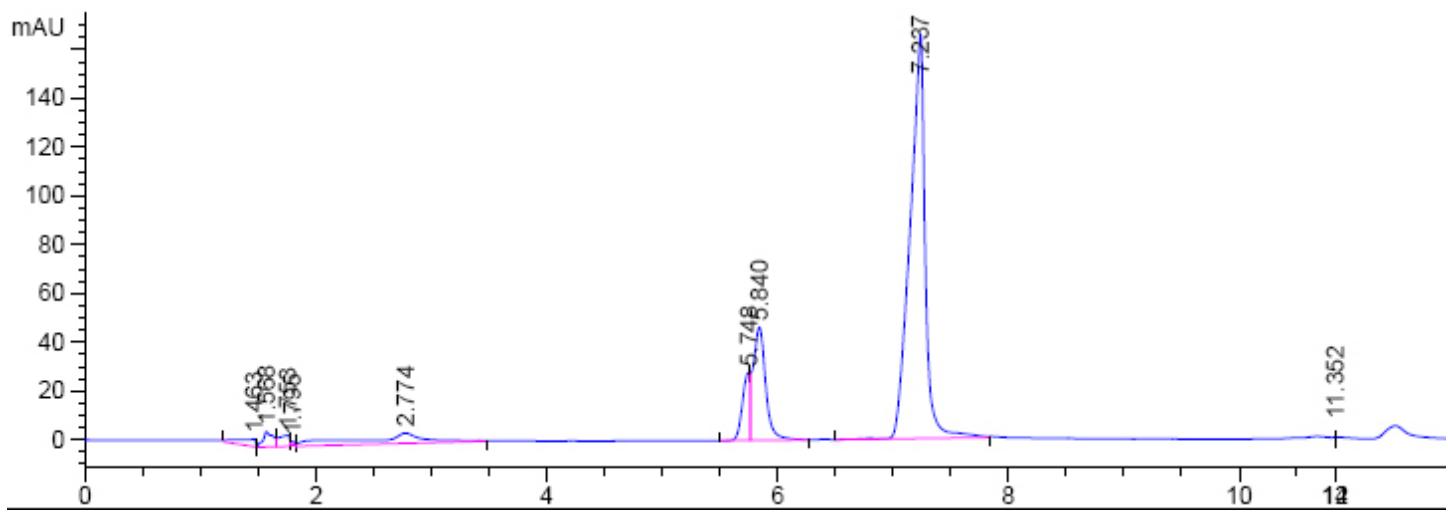


-EPI (647.20) CE (-35): 0.064 to 0.994 min from Sample 1 (RTTP 4h FIA 75% ACN -EPI) of JHDAL07072620.wiff (Turbo Spray) Max. 4.4e6 cps.



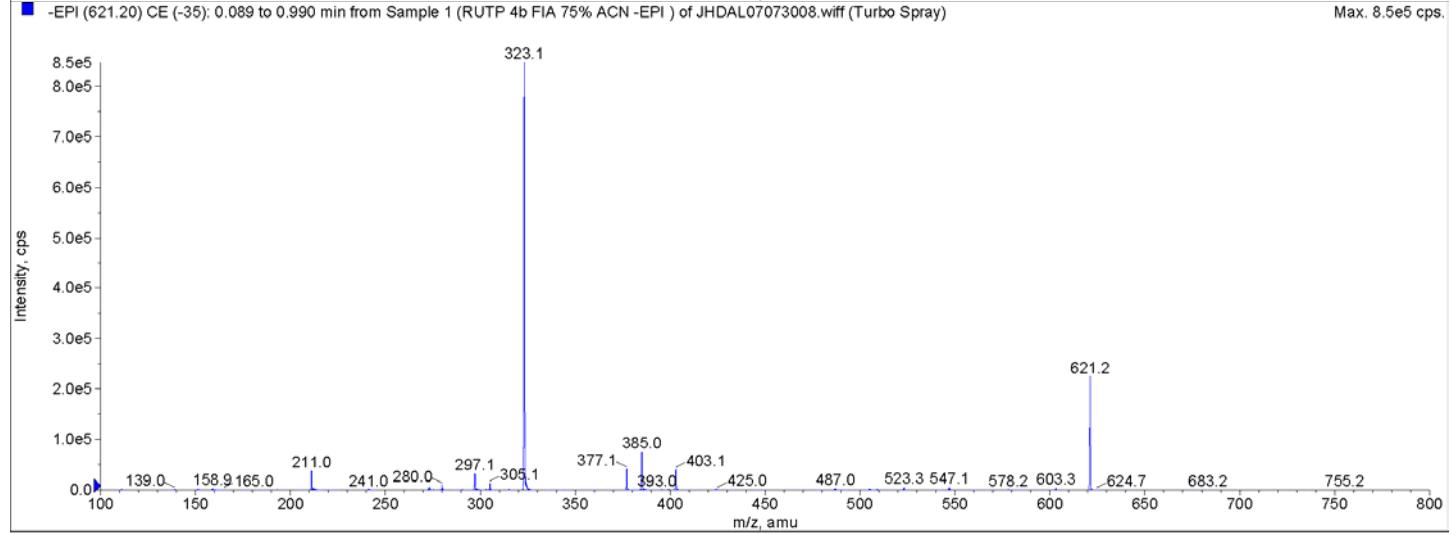
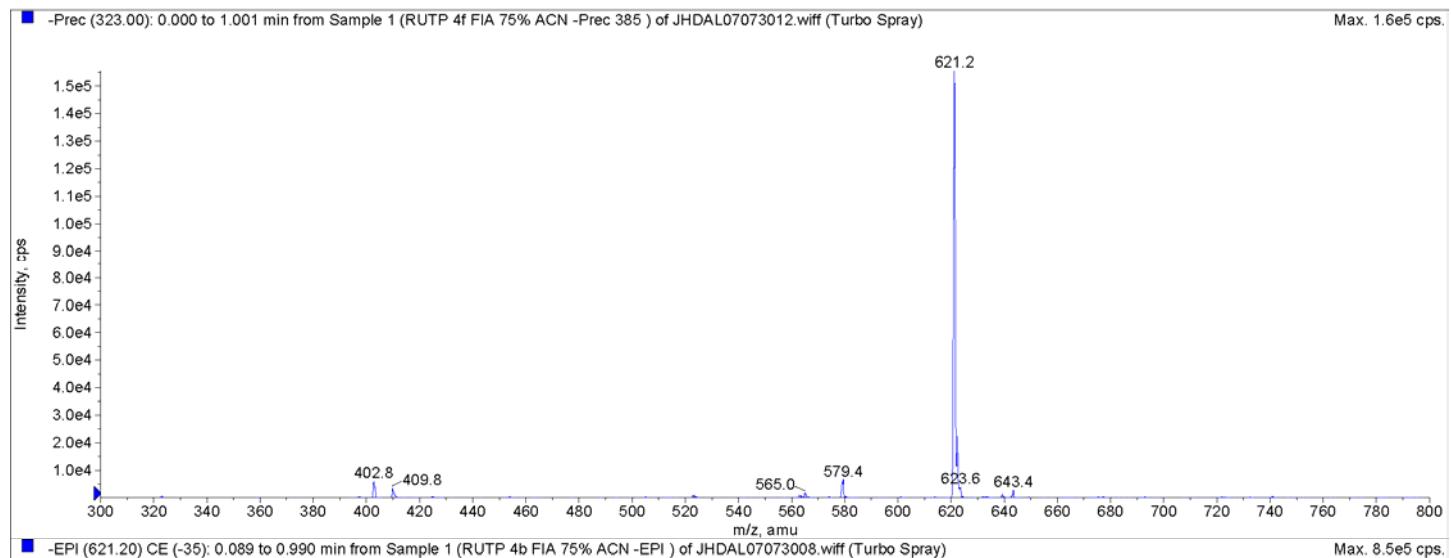
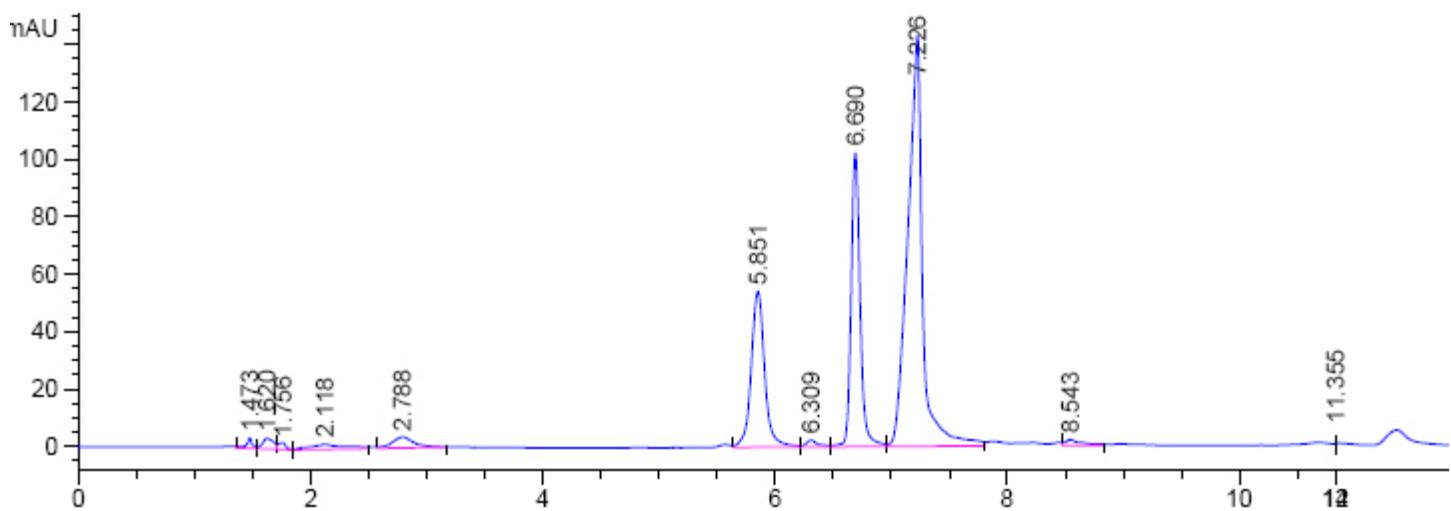
### 3-O-Methyl-UDP- $\alpha$ -D-glucose (10a)

LRMS  $m/z$  calcd for  $C_{16}H_{24}N_2O_{17}P_2^{2-}$  [M+H] $^-$ : 579.1. Found 579.1. EPI fragments: 323.1.



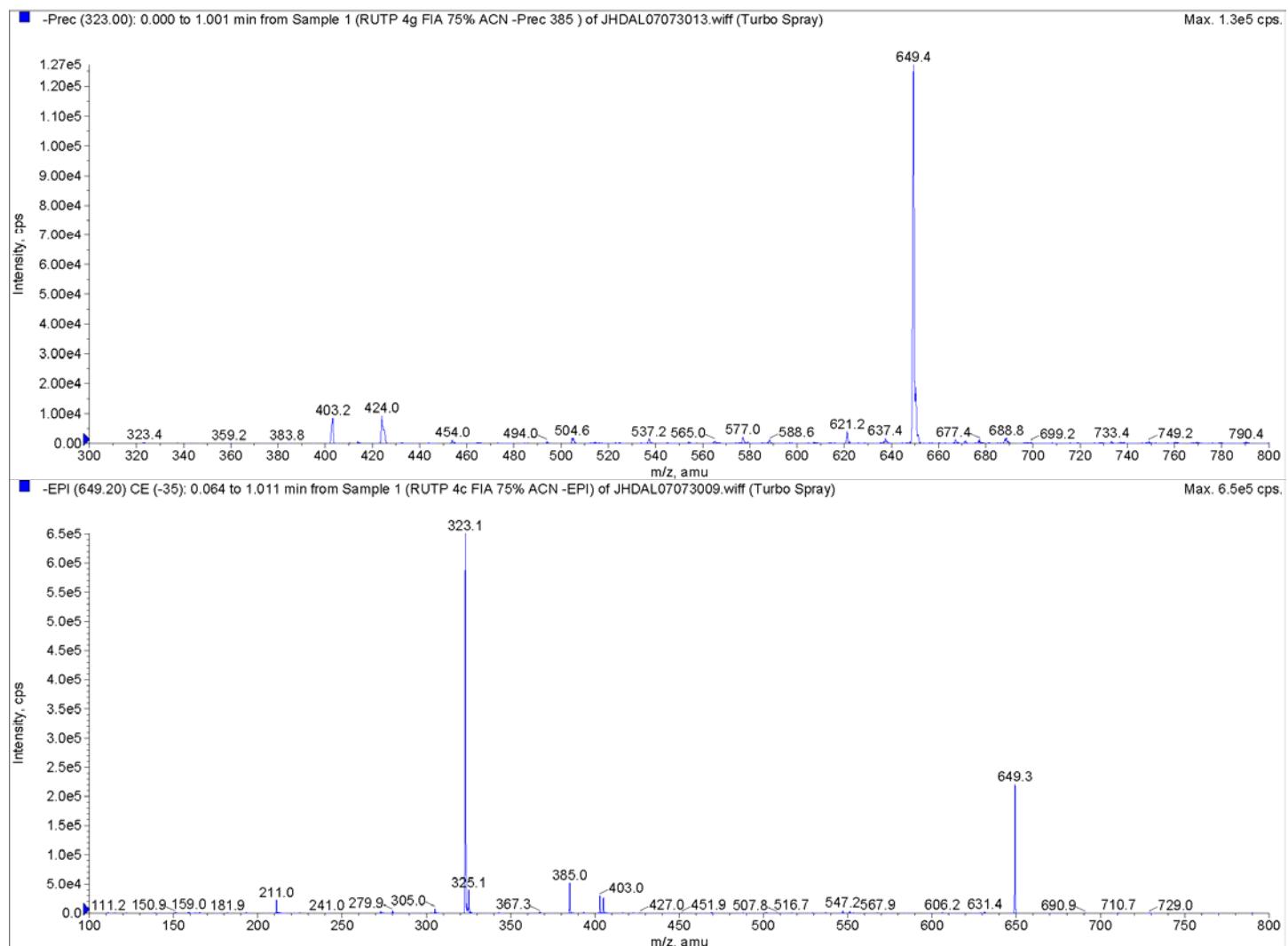
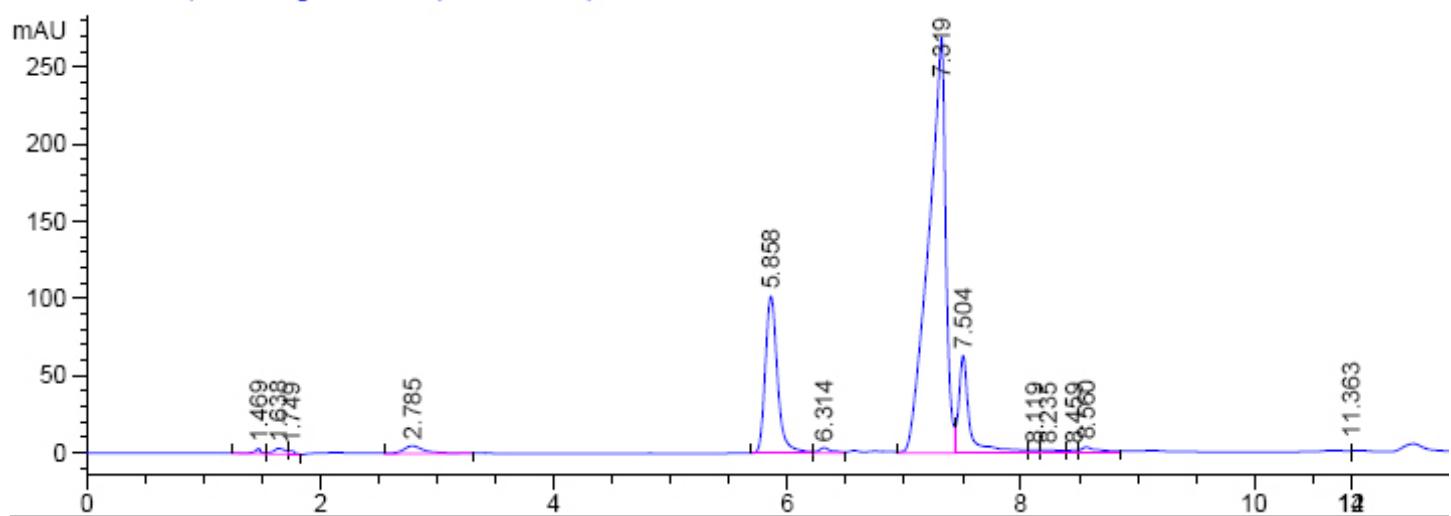
### 3-O-Butyl-UDP- $\alpha$ -D-glucose (10b)

LRMS  $m/z$  calcd for  $C_{19}H_{30}N_2O_{17}P_2^{2-}$  [M+H] $^-$ : 621.1. Found 621.2. EPI fragments: 323.1.



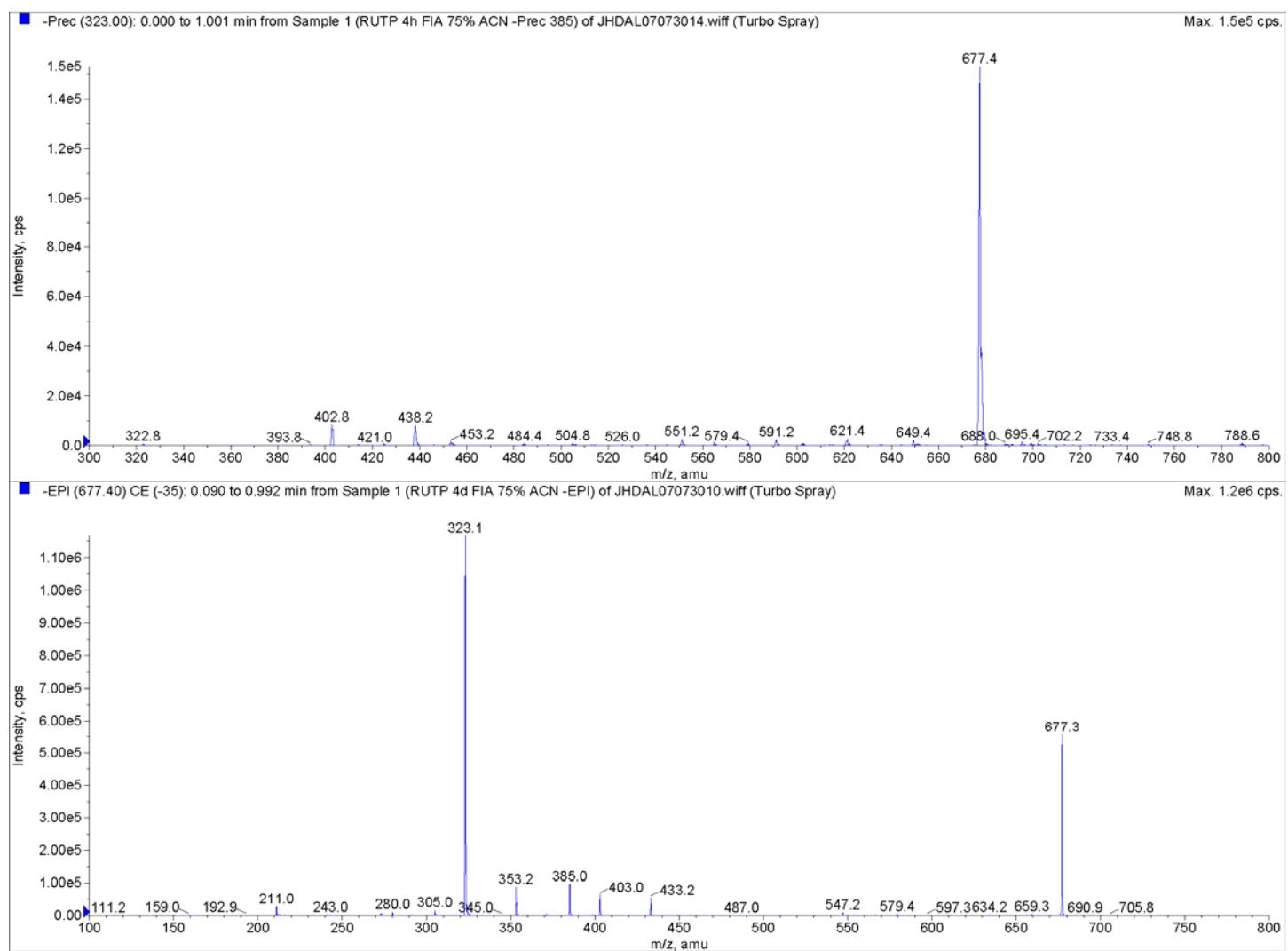
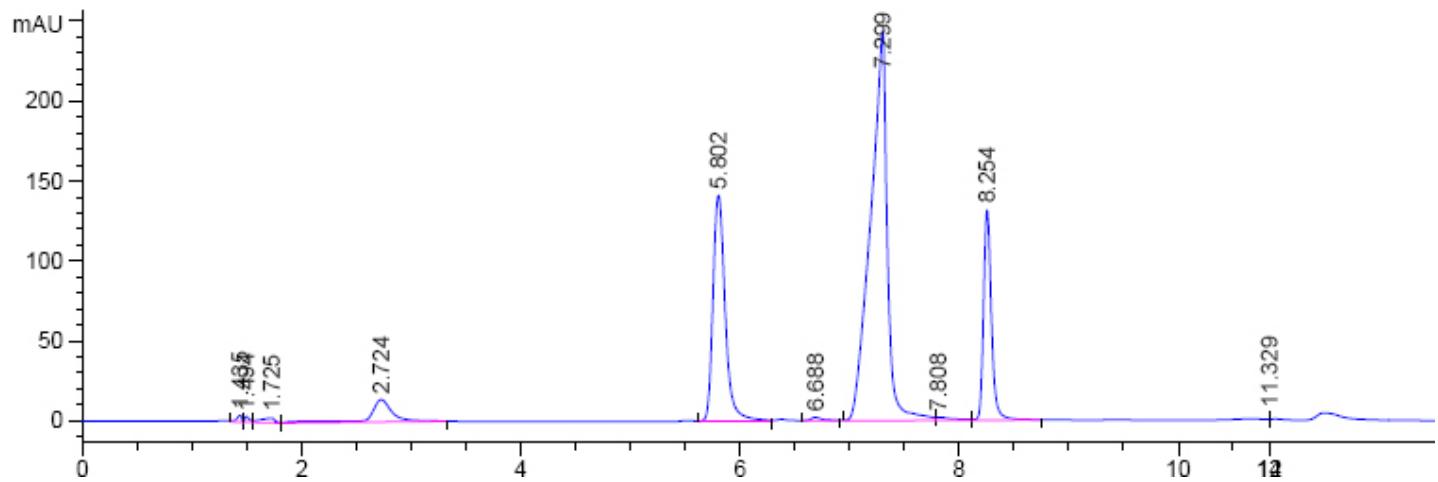
### 3-O-Hexyl-UDP- $\alpha$ -D-glucose (10c)

LRMS  $m/z$  calcd for  $C_{21}H_{34}N_2O_{17}P_2^{2-}$  [M+H] $^-$ : 649.1. Found 649.3. EPI fragments: 323.1.



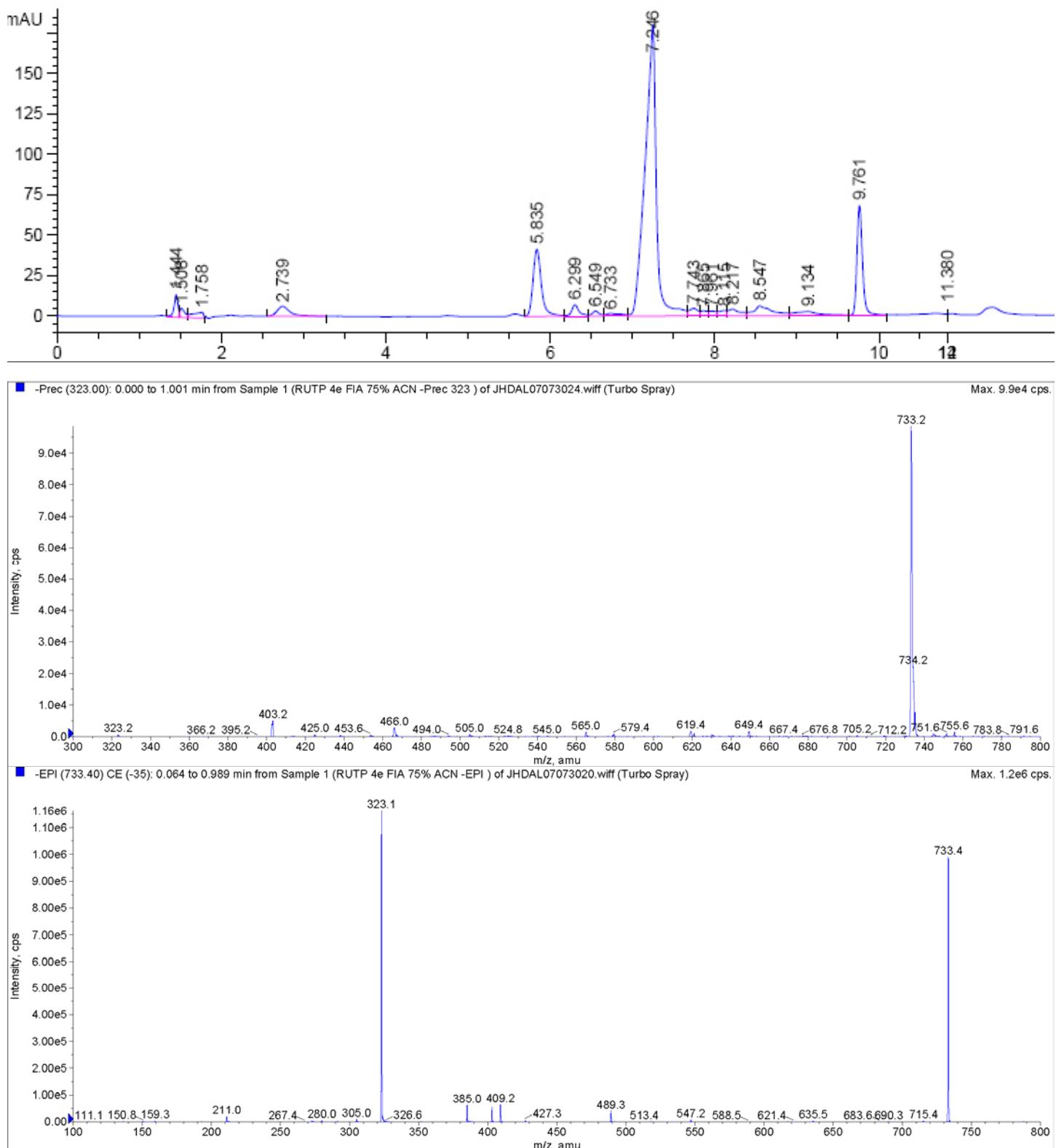
### 3-O-Octyl-UDP- $\alpha$ -D-glucose (10d)

LRMS  $m/z$  calcd for  $C_{23}H_{38}N_2O_{17}P_2^{2-}$  [M+H] $^-$ : 677.2. Found 677.3. EPI fragments: 323.1.



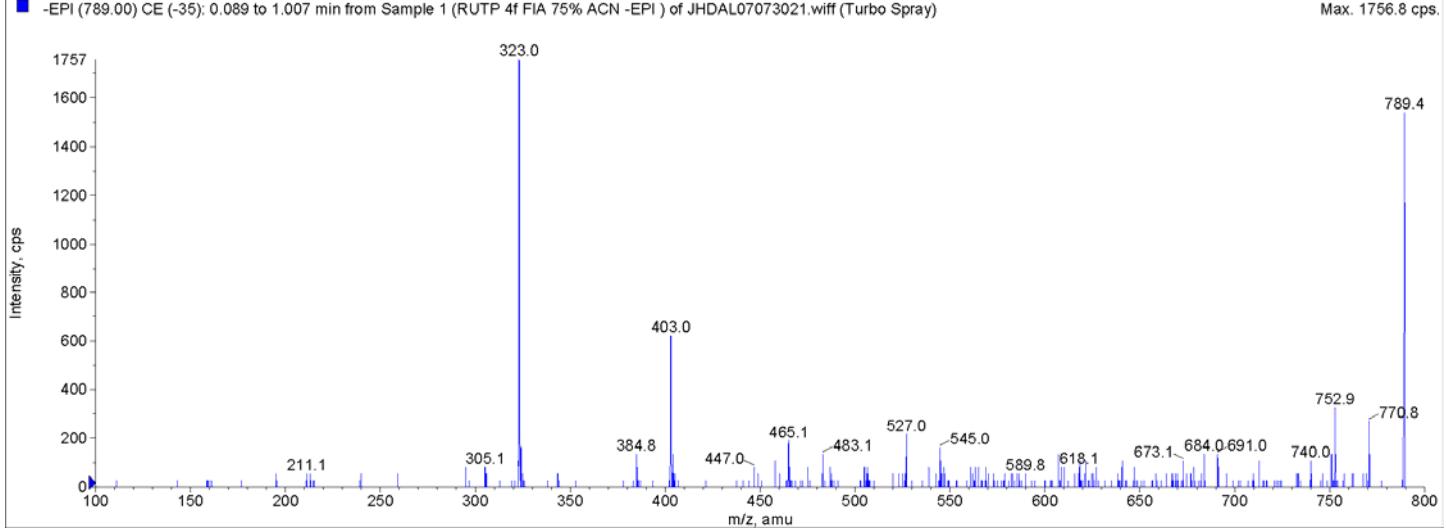
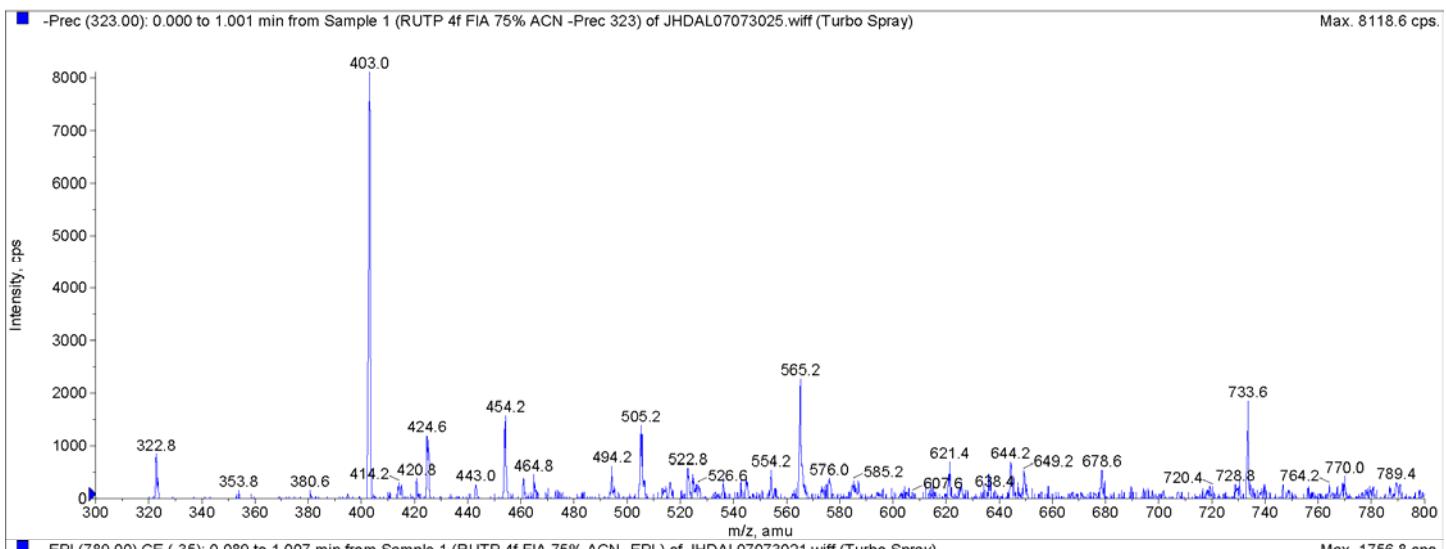
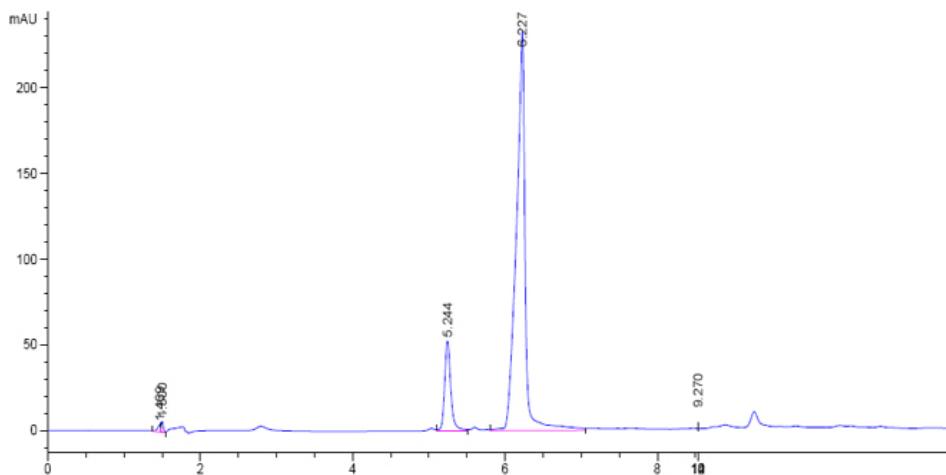
### 3-O-Dodecyl-UDP- $\alpha$ -D-glucose (10e)

LRMS  $m/z$  calcd for  $C_{27}H_{46}N_2O_{17}P_2^{2-}$  [M+H] $^-$ : 733.2. Found 733.2. EPI fragments: 323.1.



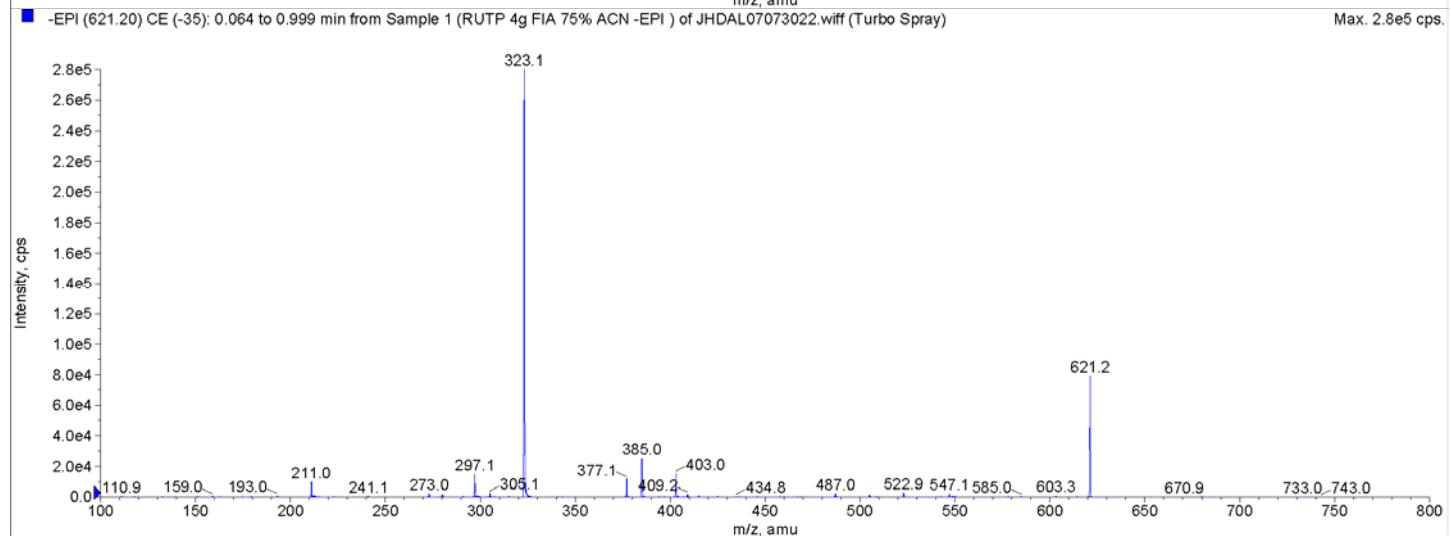
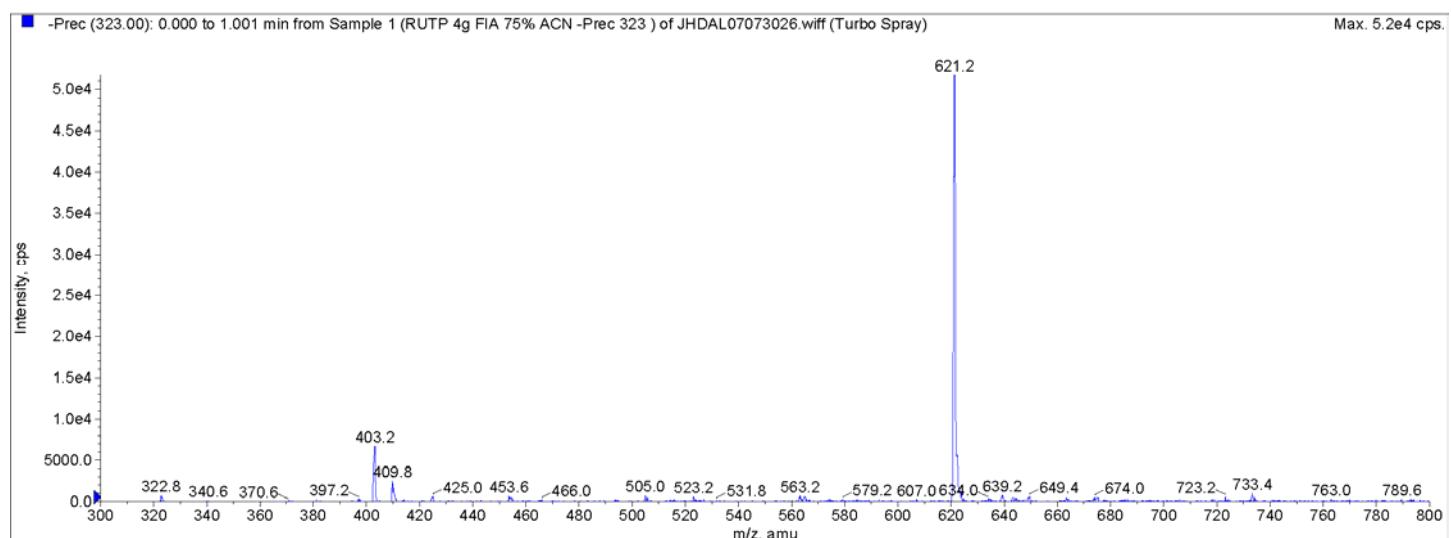
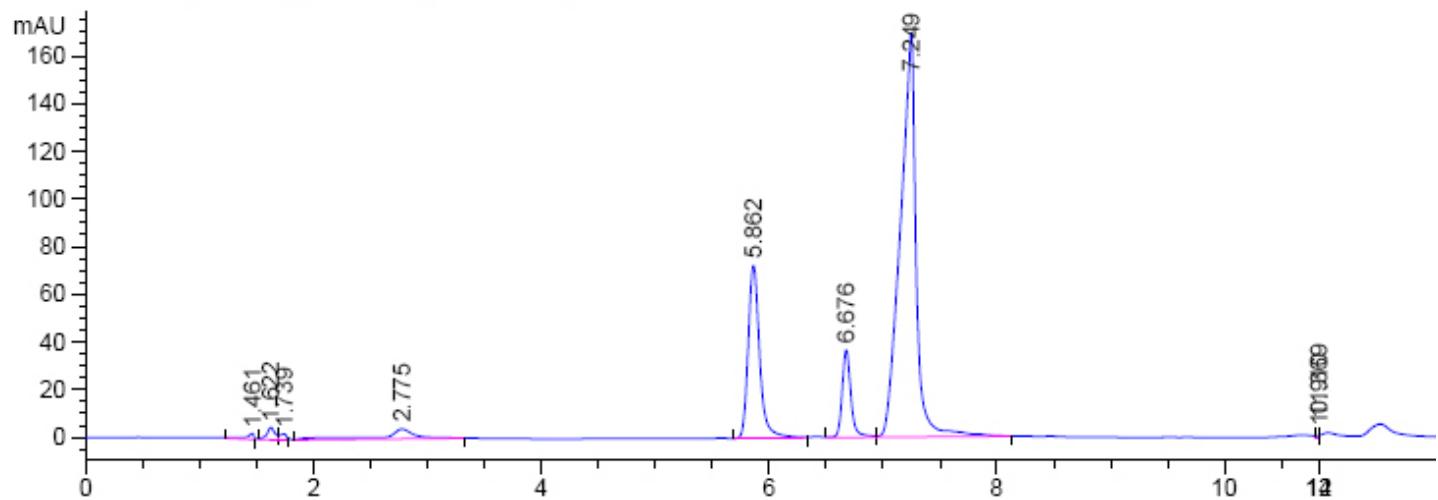
### 3-O-Hexadecyl-UDP- $\alpha$ -D-glucose (10f)

LRMS  $m/z$  calcd for  $C_{31}H_{54}N_2O_{17}P_2^{2-}$  [M+H] $^-$ : 789.3. Found 789.4. EPI fragments: 323.1.



### 3-O-(2-Methylpropyl)-UDP- $\alpha$ -D-glucose (10g)

LRMS  $m/z$  calcd for  $C_{19}H_{30}N_2O_{17}P_2^{2-}$  [M+H] $^-$ : 621.1. Found 621.2. EPI fragments: 323.1.



### 3-O-(2-Ethylbutyl)-UDP- $\alpha$ -D-glucose (10h)

LRMS  $m/z$  calcd for  $C_{21}H_{34}N_2O_{17}P_2^{2-}$  [M+H] $^-$ : 649.1. Found 649.3. EPI fragments: 323.1.

