Supplementary information

The neomycin biosynthetic gene cluster of *Streptomyces fradiae* NCIMB 8233: genetic and biochemical evidence for the roles of two glycosyltransferases and a deacetylase

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Figure S1. LC-MS analysis of the Δ neo8 mutant strain and its verification. (a) LC-MS analysis of neomycin from purified *Streptomyces fradiae* wild-type supernatant, as a control. (b) PCR screening for Δ neo8 mutant: M, Marker; Lane 1-9, verification of mutant candidates (band at 865 bp); Lane 10, plasmid pQZ27, as positive control; Lane 11, plasmid pQZ25, as negative control. (c) Restoration of neomycin production in the pQZ60-complemented Δ neo8 mutant. (d) Restoration of neomycin production upon feeding ribostamycin. The peak at 6 min corresponds to diphosphorylated ribostamycin. This arises likely from the action of *neo1*, the phosphotransferase resistance gene. (e) Production of paromomycin upon feeding paromamine.



Figure S2. LC-MS analysis of the Δ neo15 mutant strain, its verification and Neo15 protein expression. (a) Accumulation of ribostamycin in the purified supernatant of Δ neo15. (b) Restoration of neomycin production in the pQZ62-complemented Δ neo15 mutant. (c) PCR screening for Δ neo15 mutant: M, Marker; Lane 1, plasmid pQZ36, as positive control; Lane 2, plasmid pQZ35, as negative control; Lane 3-6, verification of mutant candidates (band at 486 bp); Lane 7, wild-type genomic DNA control. (d) SDS-PAGE analysis of purified Neo15 protein.



Figure S3. LC-MS analysis of the Δ neo16 mutant strain, its verification and Neo16 protein expression. (a) PCR screening for Δ neo16 mutant: Lane 1-4, verification of mutant candidates (band at 1986 bp); M, Marker; Lane 5, plasmid pQZ22, as positive control; Lane 6, plasmid pQZ15, as negative control; Lane 7, wild-type genomic DNA control. (b) SDS-PAGE analysis of purified Neo16 protein. (c) Southern blot analysis of Δ neo16 with *Age*l digestion and *Sac*l digestion respectively using the *Age*l-digested fragments

(2606 bp and 2040 bp) as probe: Lane 1, plasmid pQZ15, as negative control; Lane 2, plasmid pQZ16, as positive control; Lane 3, wild-type genomic DNA control; Lane 4-7, verification of mutant candidates (band at 2606 bp and 2040 bp for Agel digestion; band at 5670 bp for Sacl digestion. (d) Restoration of neomycin production upon feeding neamine. (e) Production of paromomycin upon feeding paromamine. (f) Production of neomycin upon feeding ribostamycin. Starred peaks (*) are not aminoglycoside-related compounds as judged by MS/MS.



of

N-acetylglucosaminylribostamycin to form glucosaminylribostamycin.

Sample ID	Culture	Compound fed to the culture	Yield of neomycin / mg L ⁻¹	Average yield of neomycin/ mg L ⁻¹	Yield of neomycin as a percentage of wild-type neomycin production / %	Yield of paromomycin / mg L ⁻¹	Average yield of paromomycin / mg L ⁻¹	Yield of paromomycin as a percentage of wild-type neomycin production / %
WT1	wild-type	Nil	0.689	0.666	100	_		· ·
WT2	wild-type	Nil	0.644	0.000	100			
WT4	wild-type	paromamine	0.200		30	0.258		39
WT3	wild-type	ribostamycin	0.255		38	-		
N8-1	∆neo8	neamine	0.232	0.470	07			
N8-2	$\Delta neo8$	neamine	0.125	0.179	27	-		
N8-3	∆neo8	naromamine	0 172			0.368		
N8-4	∆neo8	paromamine	0.370	0.275	41	0.982	0.772	116
N8-5	∆neo8	paromamine	0.282			0.966		
N8 6	Aneo8	ribostamycin	0.214					
N8-7	∆neo8	ribostamycin	0.214	0.216	32	-		
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N16-1	∆neo16	neamine	0.096					
N16-2	∆neo16	neamine	0.032	0.088	13	-		
N16-3	∆neo16	neamine	0.137					
N16-4	∆neo16	paromamine	0.004	0.000	0	0.400	0.454	<u></u>
N16-5	$\Delta neo16$, paromamine	0	0.002	0	0.509	0.454	68
N16-6	Aneo16	ribostamycin	0.010					
N16-7	Aneo16	ribostamycin	0.010					
N16-8		ribostamycin	0.000					
N16-0		ribostamycin	0.001	0 004	1	_		
N16-10		ribostamycin	0.001	0.004	1			
N16_11		rihostamycin	0.000					
N16-12	Aneo16	ribostamycin	0.002					
1110 12		nsostantyon	0.004					

Table S1. Neomycin and paromomycin production of wild-type and mutant cultures supplemented with different intermediates of the biosynthetic pathway