# Dendritic effects in catalysis by Pd complexes of bidentate phosphines on a dendronized support: Heck vs. carbonylation reactions

Amal Mansour, Tzofit Kehat and Moshe Portnoy\*

Supporting Information

# General

All reactions were conducted under an atmosphere of nitrogen in oven-dried glassware with magnetic stirring. Solvents were dried prior to use. Dry NMP was purchased from Sigma-Aldrich. Reagents were obtained from Sigma-Aldrich, Fluka, Strem Chemicals or Merck at the highest available purity and used as received. Pd(dba)<sub>2</sub> was prepared according to known procedure.<sup>1</sup> All resins used are 1% crosslinked divinylbenzene-styrene copolymer, 100-200 mesh, with loading 0.77-1.30 mmol/g and were purchased from Novabiochem. HPLC grade acetonitrile and water were purchased from Bio-Lab and Merck respectively and used after filtration. HPLC experiments were carried out using an Inertsil ODS-3v column on a Jasco chromatograph equipped with a UV/Vis detector with acetonitrile and water as the eluting solvents.

<sup>1</sup>H NMR (200, 400 MHz), <sup>13</sup>C NMR (100.8 MHz) and <sup> $\overline{31}$ </sup>P NMR (162.6 MHz) spectra were recorded on Bruker AVANCE-200 and AVANCE-400 spectrometers, in CDCl<sub>3</sub> or CDCl<sub>3</sub>/TFA (1:1) using residual CHCl<sub>3</sub> (<sup>1</sup>H, 7.26 ppm), or CDCl<sub>3</sub> (<sup>13</sup>C, 77.0 ppm) as an internal standard. Gel-phase <sup>13</sup>C NMR (100.8 MHz) and <sup>31</sup>P NMR (162.6 MHz) spectra were recorded in benzene-*d*<sub>6</sub>, on a Bruker AVANCE-400 instrument using the solvent (<sup>13</sup>C, 126.0 ppm) as an internal standard or H<sub>3</sub>PO<sub>4</sub>, 85% (<sup><sup>31</sup>P</sup>, 0.0 ppm) as an external standard. The yields of the polymerbound ligands and complexes were determined using gel-phase <sup><sup>31</sup>P NMR with an internal phosphorus-containing resin standard.<sup>2</sup></sup>

#### Characterization of Gn(CO<sub>2</sub>Me) resins

# $G1(CO_2Me)$

Prepared from Wang Bromo PS (0.76 mmol/g).

Yield >99%, purity >99%, loading 0.69 mmol/g.

Gel-phase <sup>13</sup>C NMR (100.8 MHz, C<sub>6</sub>D<sub>6</sub>):  $\delta$  165.8, 159.3, 158.0, 132.4, 129.5, 123.3, 120.4, 115.0, 70.0, 51.9. Following TFA-induced cleavage: <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>/TFA 1:1):  $\delta$  8.35 (t, *J* = 1.3 Hz, 1H), 7.86 (d, *J* = 1.3 Hz, 2H), 4.06 (s, 6H). <sup>13</sup>C NMR (100.8 MHz, CDCl<sub>3</sub>/TFA 1:1):  $\delta$  168.4, 154.8, 130.9, 123.8, 121.5, 53.1.

# $G2(CO_2Me)$

Prepared from G1(Cl) (0.50 mmol/g).

Yield >99%, purity >95%, loading 0.42 mmol/g.

Partial Gel-phase <sup>13</sup>C NMR (100.8 MHz,  $C_6D_6$ ):  $\delta$  165.7, 158.9, 138.0, 132.3, 123.4, 120.2, 114.1, 69.8, 51.9. Following TFA-induced cleavage: <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>/TFA 1:1):  $\delta$  8.35 (s, 2H), 7.93 (s, 4H), 7.20 (s, 1H), 7.05 (s, 2H), 5.20 (s, 4H), 4.06 (s, 12H). <sup>13</sup>C NMR (100.8 MHz, CDCl<sub>3</sub>/TFA 1:1):  $\delta$  168.6, 158.4, 157.0, 138.1, 130.7, 123.6, 121.0, 119.5, 114.0, 69.6, 53.1.

#### $G3(CO_2Me)$

Prepared from G2(Cl) (0.30 mmol/g).

Yield >99%, purity >95%, loading 0.26 mmol/g.

Partial gel-phase <sup>13</sup>C NMR (100.8 MHz,  $C_6D_6$ ):  $\delta$  165.5, 158.9, 138.5, 132.2, 123.2, 120.0, 113.6, 69.7, 51.9. Following TFA-induced cleavage: <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>/TFA 1:1):  $\delta$  8.33 (s, 4H), 7.91 (s, 8H), 6.98-7.21 (m, 9H), 5.16 (s, 12H), 4.04 (s, 24H). <sup>13</sup>C NMR (100.8 MHz, CDCl<sub>3</sub>/TFA 1:1):  $\delta$  168.5, 158.5, 158.4, 138.5, 137.8, 130.7, 123.6, 120.9, 119.7, 119.4, 114.0, 113.8, 69.9, 69.7, 53.1.

#### Characterization of Gn(serinol-OH) resins

#### G1(serinol-OH)

Prepared from **G1(Cl**) (0.50 mmol/g). Yield >99%, purity >99%, loading 0.47 mmol/g. Following TFA-induced cleavage: <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>/TFA 1:1): δ 7.19 ( m, 3H), 4.73 (m, 2H), 4.43 ( s, 4H), 4.29-4.04 ( m, 8H). <sup>13</sup>C NMR (100.8 MHz, CDCl<sub>3</sub>/TFA 1:1): δ 156.7, 132.2, 131.8, 119.1, 62.9, 58.1, 55.8.

### G2(serinol-OH)

Prepared from **G2(Cl)** (0.30 mmol/g). Yield 90%, purity >95%, loading 0.25 mmol/g. Following TFA-induced cleavage: <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>/TFA 1:1): δ 7.56 (br, NH), 7.21-6.62 (m, 9H), 4.92 (m, 4H), 4.74 (m, 4H), 4.45 (m, 8H), 4.11 (m, 16H-xH), 3.70 (m, xH) . Partial  $^{13}C$  NMR (100.8 MHz, CDCl<sub>3</sub>/TFA 1:1):  $\delta$  132.0, 117.9, 114.2, 112.3, 69.5, 62.9, 58.0, 55.6.

### G3(serinol-OH)

Prepared from G3(Cl) (0.26 mmol/g).

Yield >99%, purity >95%, loading 0.23 mmol/g.

Following TFA-induced cleavage: <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>/TFA 1:1):  $\delta$  7.6 (br, NH), 7.24-7.07 (m, 21H), 5.08 (m, 8H), 4.78 (m, 8H), 4.50 (m, 16H), 4.20-3.96 (m, 32H). Partial <sup>13</sup>C NMR (100.8 MHz, CDCl<sub>3</sub>/TFA 1:1):  $\delta$  132.0, 117.9, 69.6, 63.3, 58.0, 55.7.

#### Characterization of Gn(serinol-Cl) resins

#### G1(serinol-Cl)

Prepared from G1(serinol-OH) (0.47 mmol/g).

Yield >99 %, purity >99%, loading 0.44 mmol/g.

Following TFA-induced cleavage: <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>/TFA 1:1):  $\delta$  8.04 ( br, NH), 7.25 (m, 3H), 4.50 (s, 4H), 4.03 (s, 10H). <sup>13</sup>C NMR (100.8 MHz, CDCl<sub>3</sub>/TFA 1:1):  $\delta$  156.5, 131.5, 124.3, 119.2, 59.9, 50.1, 39.3. MS: found (m/z), 373.1; calcd for C<sub>14</sub>H<sub>22</sub><sup>35</sup>Cl<sub>4</sub>N<sub>2</sub>O (MH<sup>+</sup>) 373.1.

# G2(serinol-Cl)

Prepared from G2(serinol-OH) (0.25 mmol/g).

Yield >99 %, purity >95%, loading 0.23 mmol/g.

Following TFA-induced cleavage: <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>/TFA 1:1):  $\delta$  8.09 (br, NH), 7.20-6.70 (m, 9H), 5.20 (m, 4H), 4.46 (m, 8H), 4.01 (m, 20H). <sup>13</sup>C NMR (100.8 MHz, CDCl<sub>3</sub>/TFA 1:1):  $\delta$  159.8, 138.1, 134.0, 124.4, 119.3, 118.7, 114.3, 69.8, 60.0, 52.9, 39.6.

#### G3(serinol-Cl)

Prepared from G3(serinol-OH) (0.23 mmol/g).

Yield 53 %, purity >95%, loading 0.11 mmol/g.

Following TFA-induced cleavage: <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>/TFA 1:1):  $\delta$  8.12 (br, NH), 7.13-6.99 (m, 21H), 5.09 (m, 12H), 4.50 (m, 16H), 4.01 (m, 40H). Partial <sup>13</sup>C NMR (100.8 MHz, CDCl<sub>3</sub>/TFA 1:1):  $\delta$  159.9, 132.3, 132.1, 131.5, 129.3, 129.1, 69.9, 59.8, 50.3, 39.5.

#### References

1. L. S. Hegedus in Organometallics in synthesis, ed. M. Schlosser, Wiley, 1994.

2. B. Ben-Aroya Bar-Nir and M. Portnoy, Tetrahedron, 2002, 58, 5147.