

**Supplementary Information**

**[<sup>18</sup>F]- and [<sup>11</sup>C]N-benzyl-isatin sulfonamide analogues as PET tracers for Apoptosis: Synthesis, radiolabeling mechanism, and *in vivo* imaging study of Apoptosis in Fas-treated mice using [<sup>11</sup>C]WC-98**

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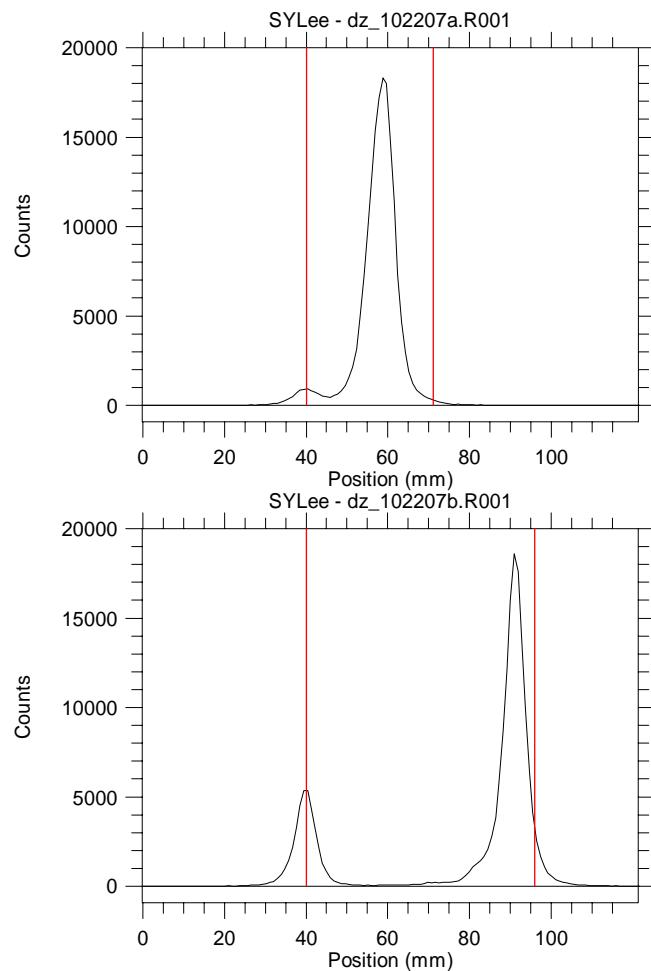
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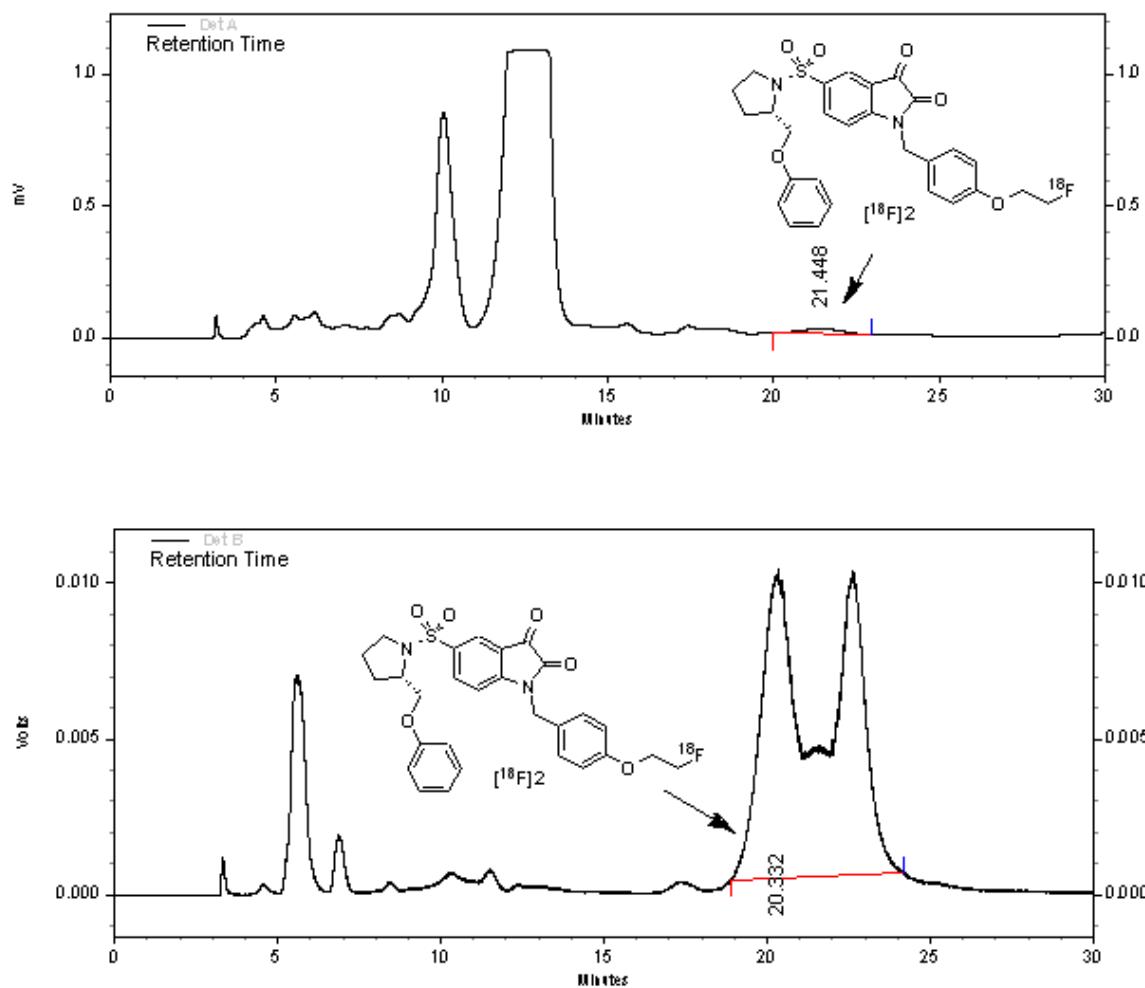


**Figure S1.** RadioTLCs of the reaction mixture of radiosynthesis of [ $^{18}\text{F}$ ] 2.

Above: Precursor **7a**, [ $^{18}\text{F}$ ]fluoride, K<sub>222</sub>, K<sub>2</sub>CO<sub>3</sub>, DMSO, Microwave;

Bottom: 1 N HCl, Microwave.

Silica Gel TLC place; solvent: 20% MeOH, 80% CH<sub>2</sub>Cl<sub>2</sub>.

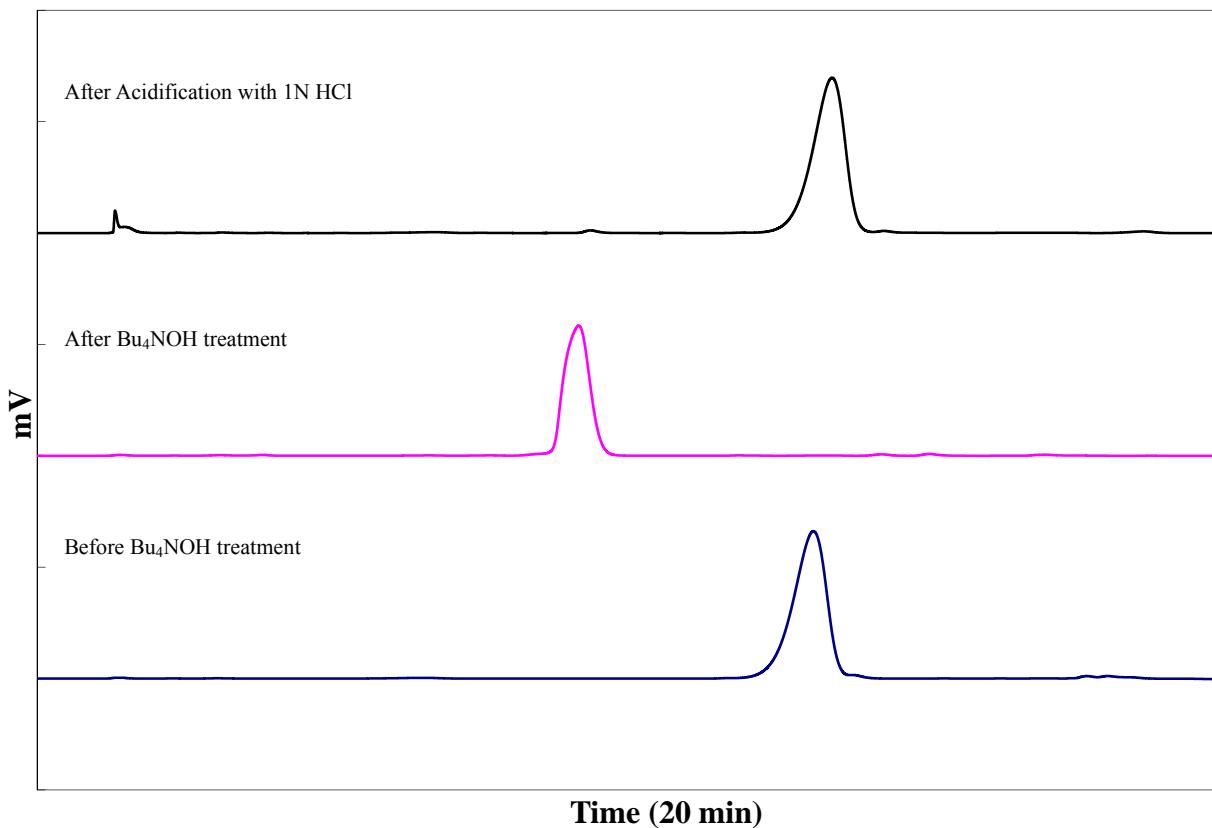


**Figure S2.** Typical HPLC chromatograms for  $[^{18}\text{F}]2$  purification

Above: UV absorbance at 251 nm;

Bottom: radioactivity (The “rabbit” ears are due to saturation of the radioactive detector)

Alltech Econosil 250×10 mm, 10 $\mu$ ; 4.0 mL/min, 251 nm, 800 psi; 24% Acetonitrile, 44% methanol, 32% Ammonium formate buffer (pH = 4.5)



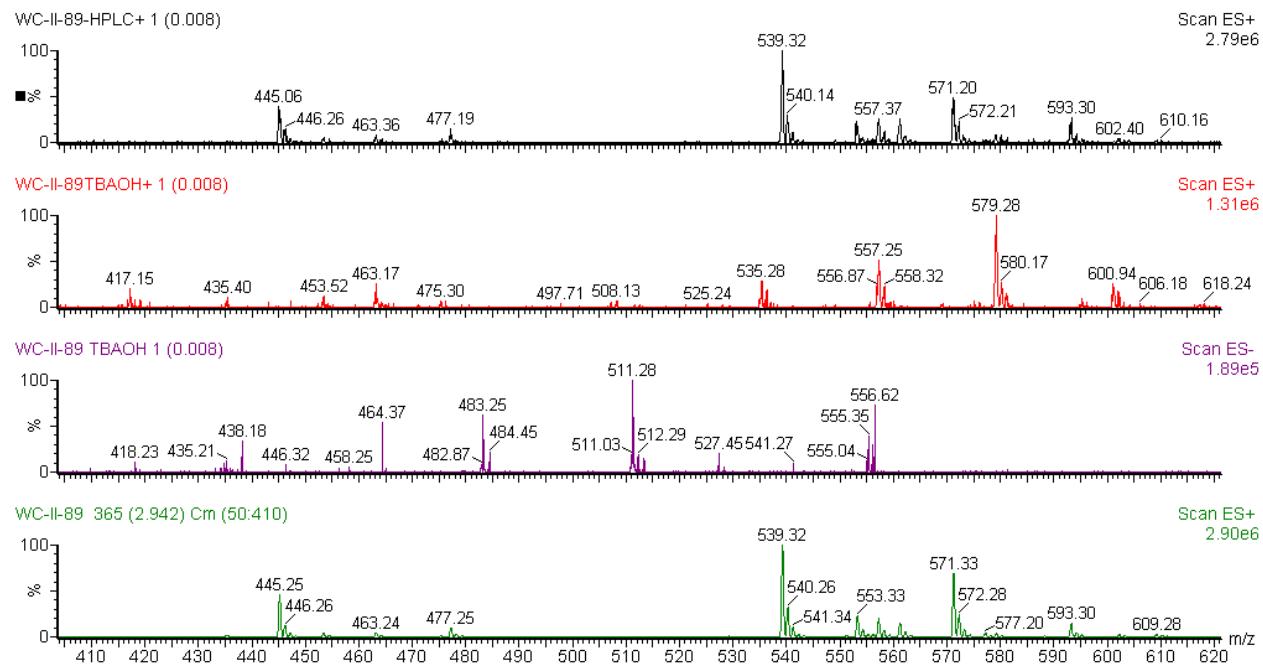
**Figure S3.** HPLC chromatograms of isatin analogue **2** (bottom), isatinate **17** (middle) and isatin **2** recyclized from isatinate **17** (top).

HPLC condition: Alltech Altima C18  $250 \times 4.6$  mm  $10\mu$ , gradient: 25% Acetonitrile, 40% ammonium formate buffer ( $\text{pH} = 4.5$ ), 35% Methanol to 35% Acetonitrile, 20% ammonium formate buffer ( $\text{pH} = 4.5$ ), 45% Methanol over 15 min, 1.0 mL/min, 251 nm.

Before  $\text{Bu}_4\text{NOH}$  treatment: 20  $\mu\text{L}$  1000 ppm **2** injected

$\text{Bu}_4\text{NOH}$  treatment: 0.1 mg **2** in 100  $\mu\text{L}$  acetonitrile treated with 2  $\mu\text{L}$  1 M  $\text{Bu}_4\text{NOH}/\text{H}_2\text{O}$ , 20  $\mu\text{L}$  1000 ppm injected

1 N HCl treatment: 100  $\mu\text{L}$  1 N HCl, 1 hour at ambient temperature, 40  $\mu\text{L}$  500 ppm injected.



**Figure S4.** ESI/MS of isatin analogue **2** (bottom), isatinate **17** (middle, negative and positive) and isatin **2** recyclized from isatinate **17** (top).

Isatinate and isatin recyclized from isatinate were purified by HPLC (see above)

19F OBSERVE  
STANDARD PARAMETERS

Pulse Sequence: s2pul  
Solvent: DMSO  
Ambient temperature  
File: isatin\_TBAF\_F  
Mercury-300 "m300"  
  
PULSE SEQUENCE  
Relax. delay 4.000 sec  
Pulse 19.5 degrees  
Acq. time 0.300 sec  
Width 50000.0 Hz  
64 repetitions  
OBSERVE F19, 282.3941399 MHz  
DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 32768  
Total time 1 hr, 16 min, 2 sec

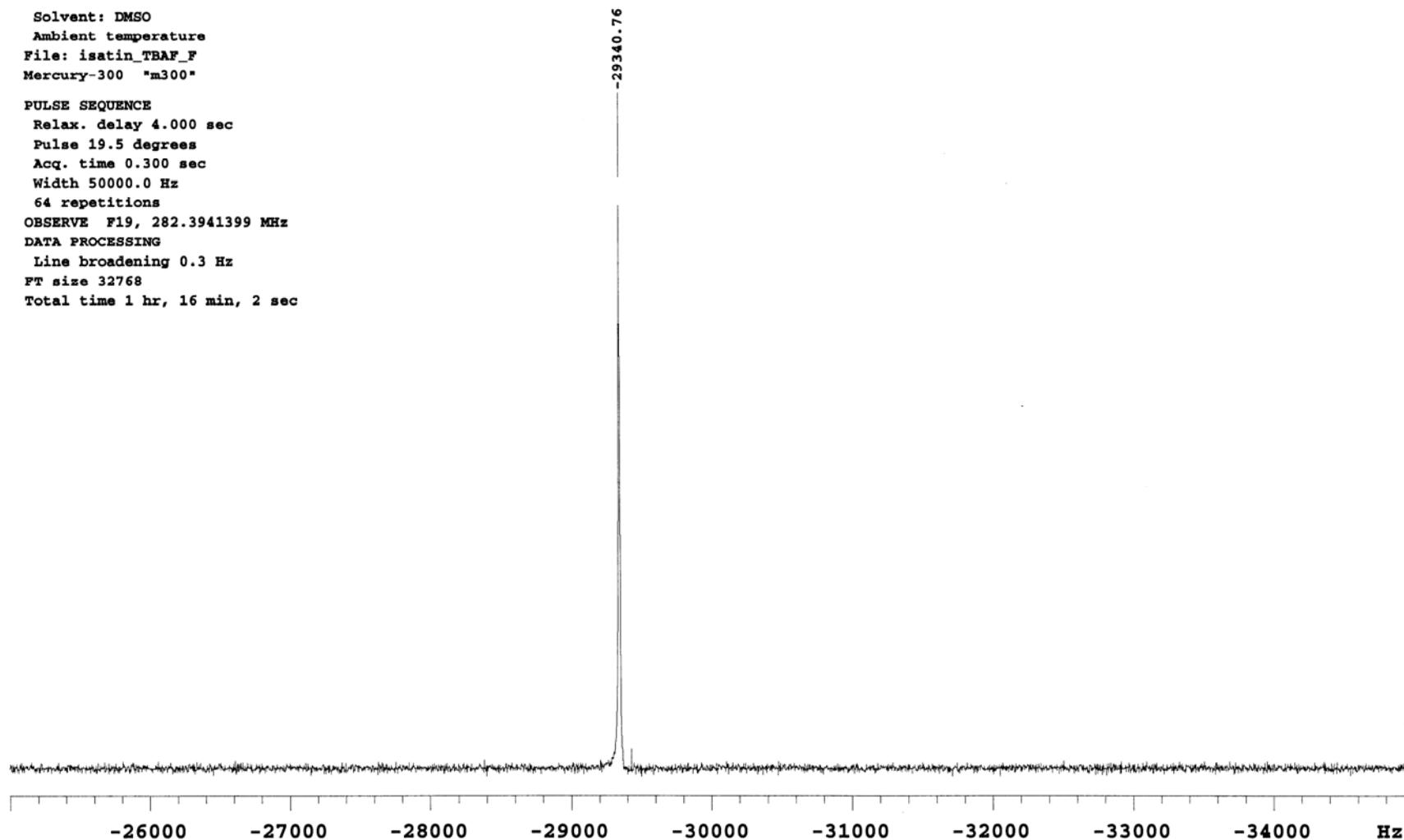


Figure S5. 282 MHz <sup>19</sup>F NMR spectrum (DMSO-d6) of Bu<sub>4</sub>NF

19F OBSERVE  
STANDARD PARAMETERS

Pulse Sequence: s2pul  
Solvent: DMSO  
Ambient temperature  
File: isatin\_plusTBAF300ul\_F  
Mercury-300 "m300"

PULSE SEQUENCE  
Relax. delay 4.000 sec  
Pulse 19.5 degrees  
Acq. time 0.300 sec  
Width 50000.0 Hz  
64 repetitions  
OBSERVE F19, 282.3941399 MHz

DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 32768  
Total time 1 hr, 16 min, 2 sec

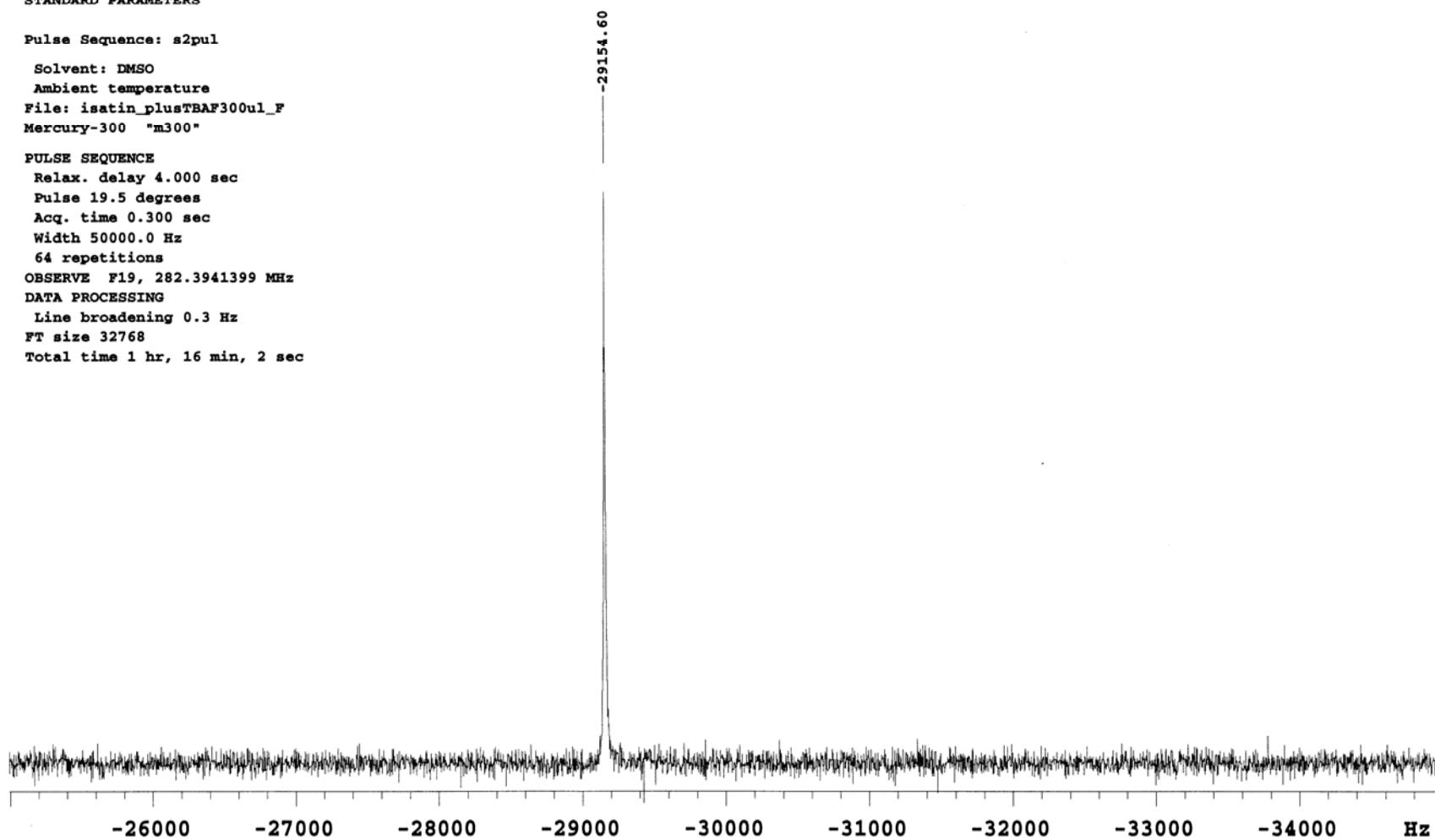
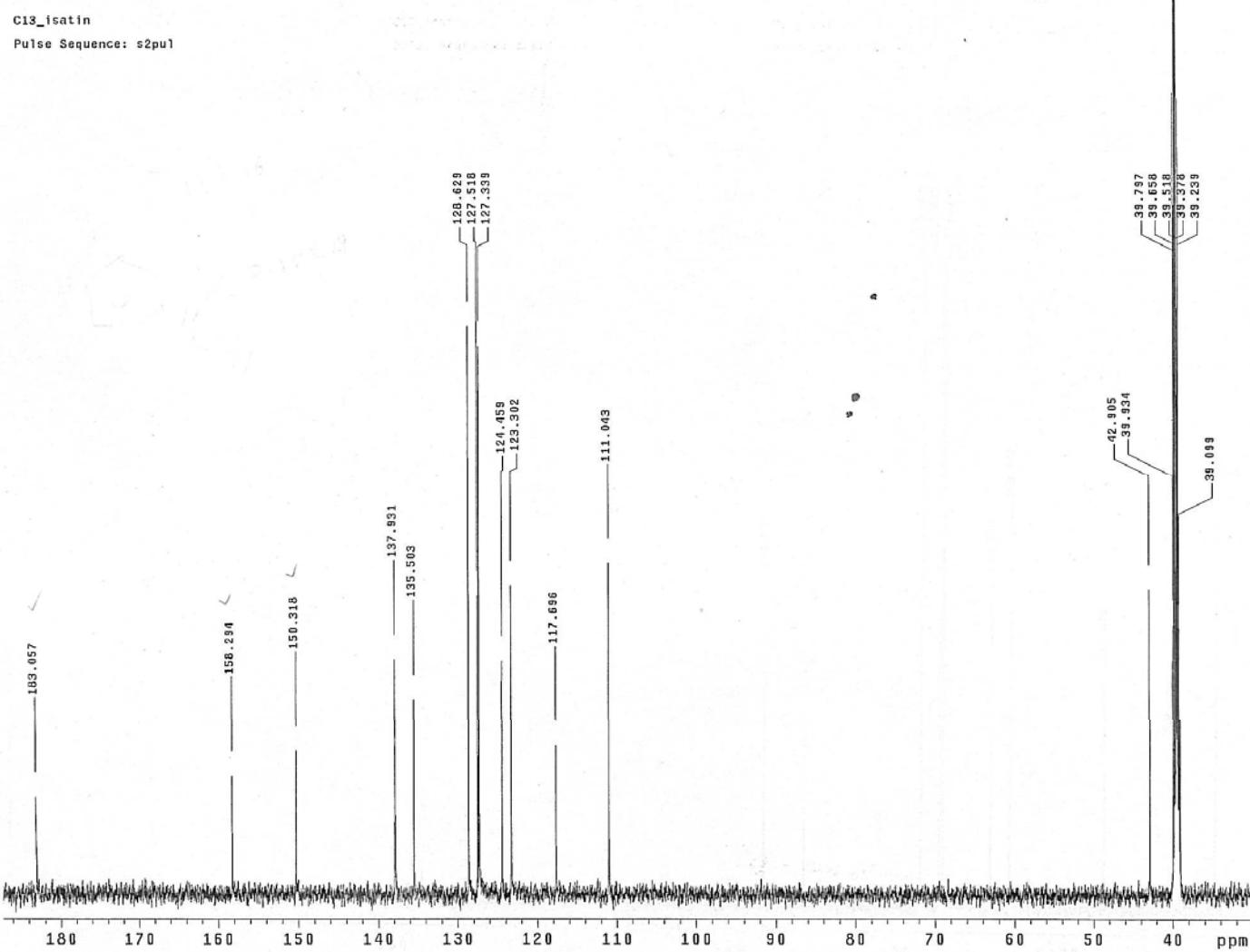
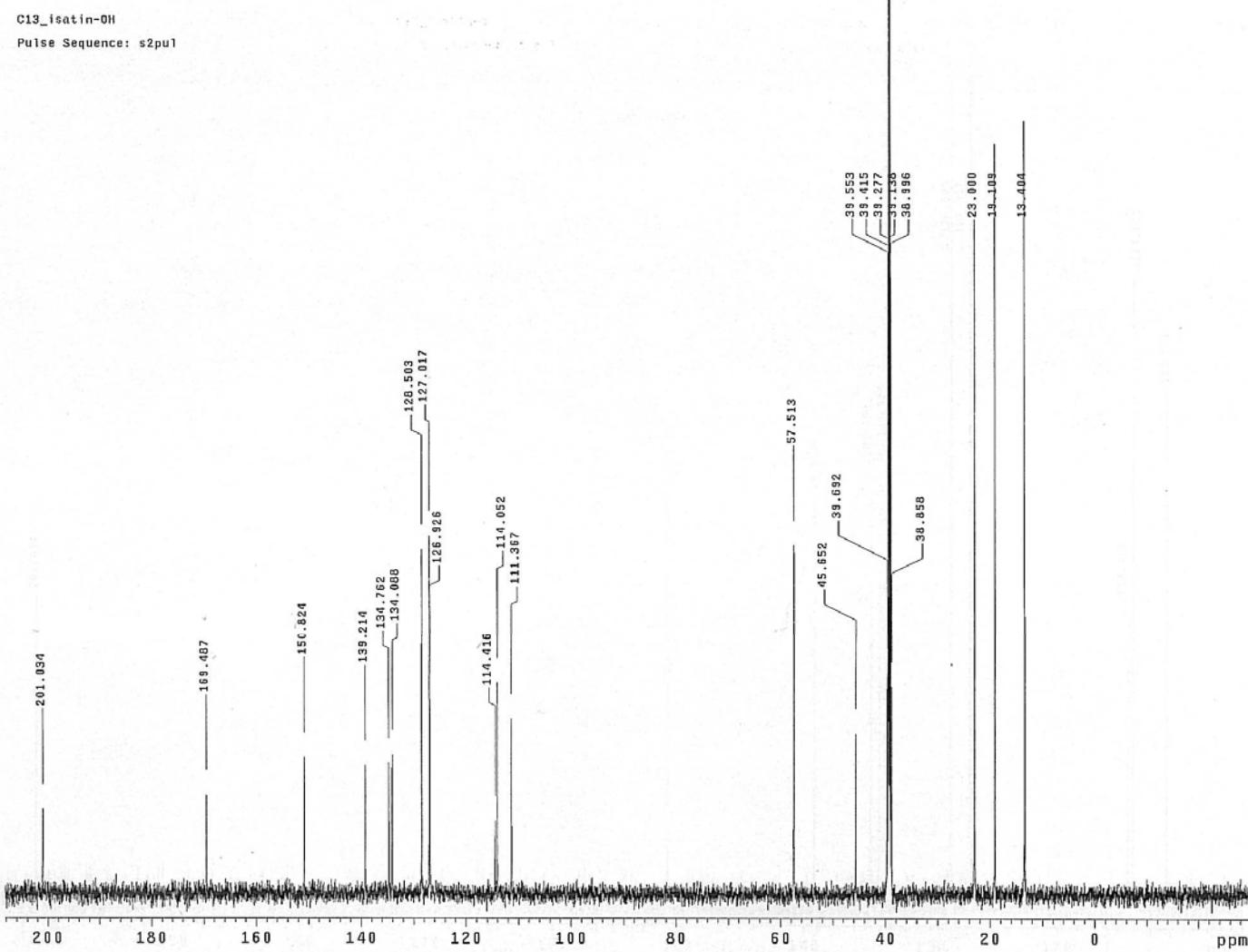


Figure S6. 282 MHz <sup>19</sup>F NMR spectrum (DMSO-d<sub>6</sub>) of Bu<sub>4</sub>NF with addition of N-benzyl isatin **21**

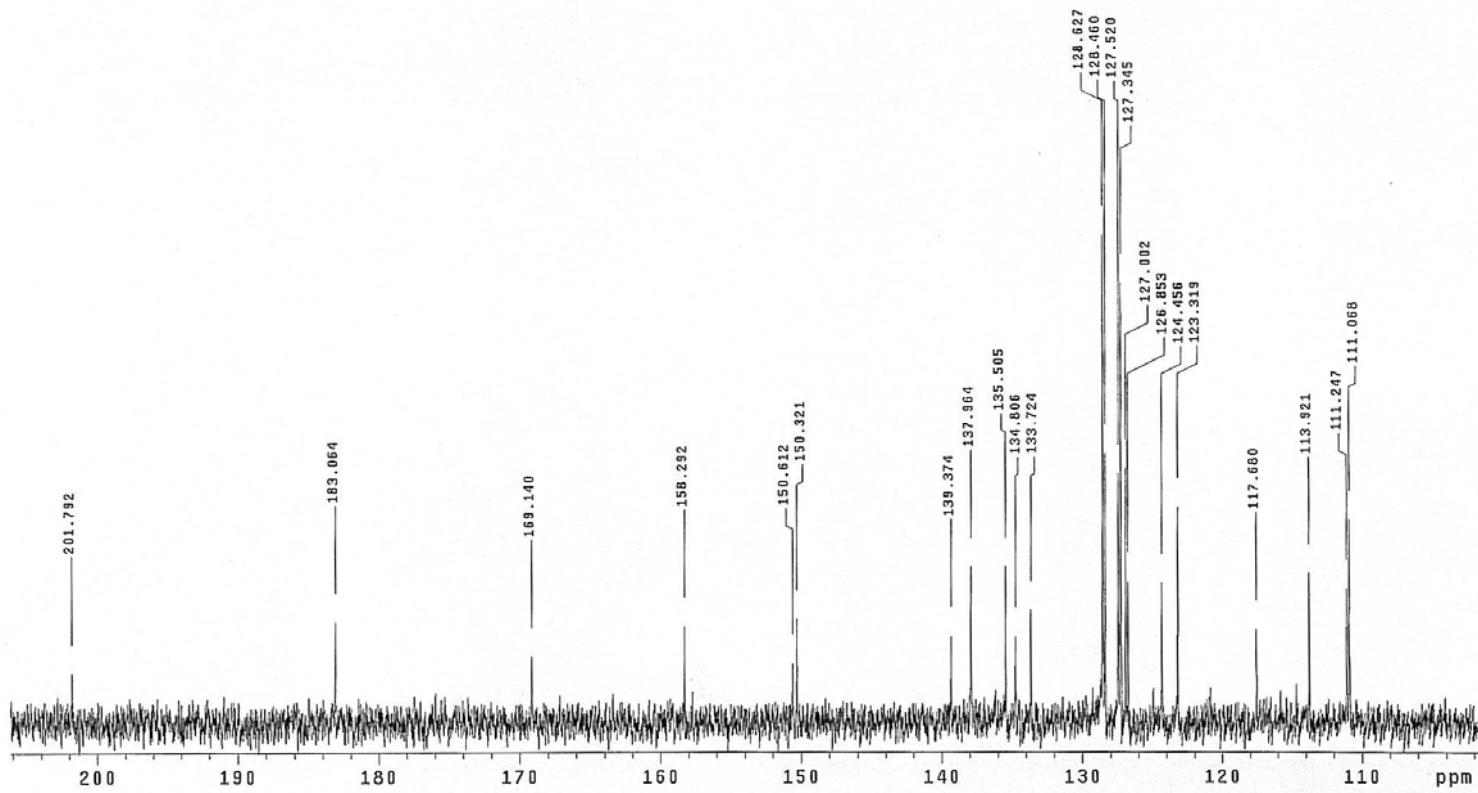


**Figure S7.** 150 MHz  $^{13}\text{C}$  NMR (DMSO-d<sub>6</sub>) Spectrum of N-benzyl isatin **21**.

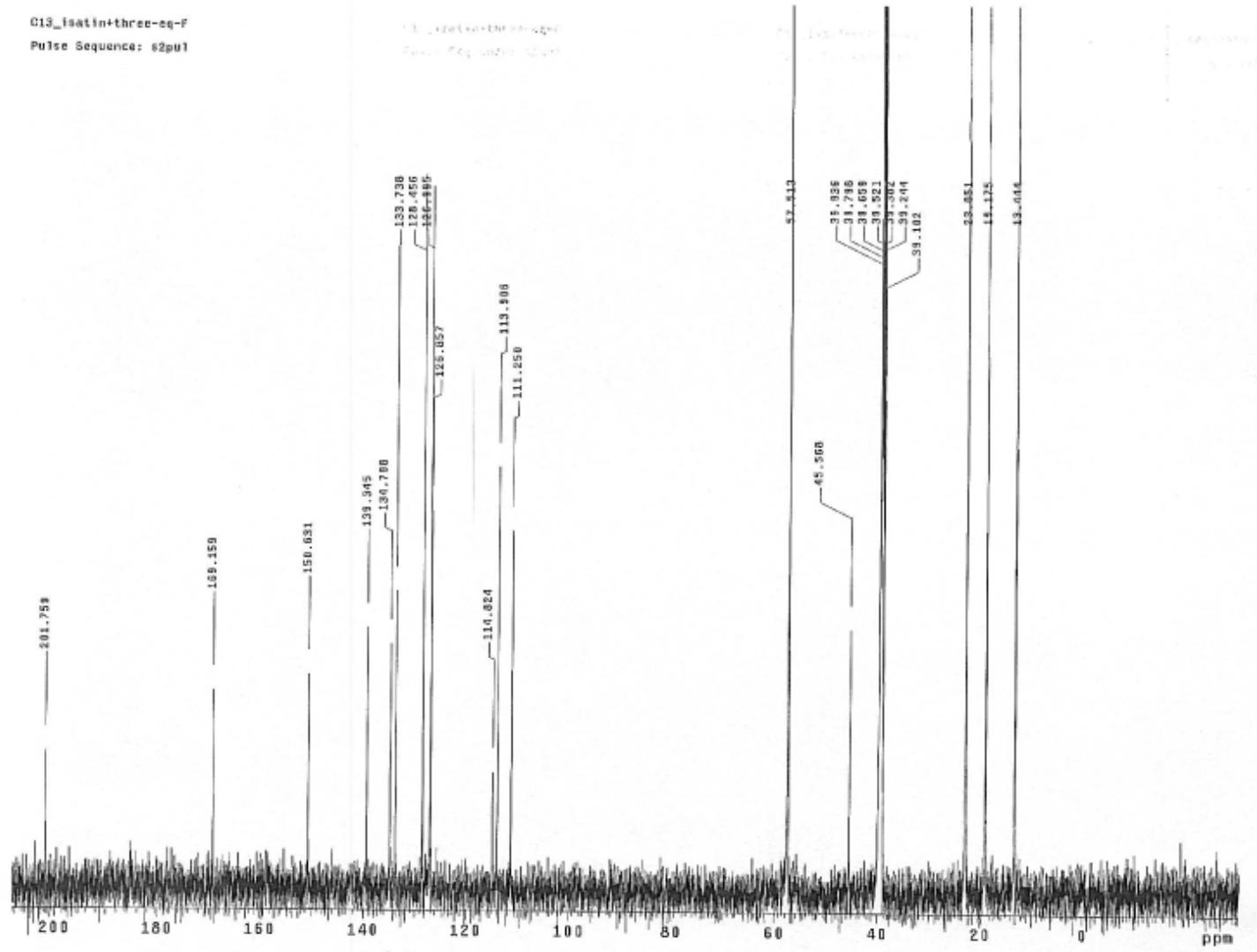


**Figure S8.** 150 MHz  $^{13}\text{C}$  NMR Spectrum (DMSO-d<sub>6</sub>) of N-benzyl isatin **21** treated with 1M Bu<sub>4</sub>NOH/H<sub>2</sub>O (10 mg **21** in 0.6 mL DMSO-d<sub>6</sub>)

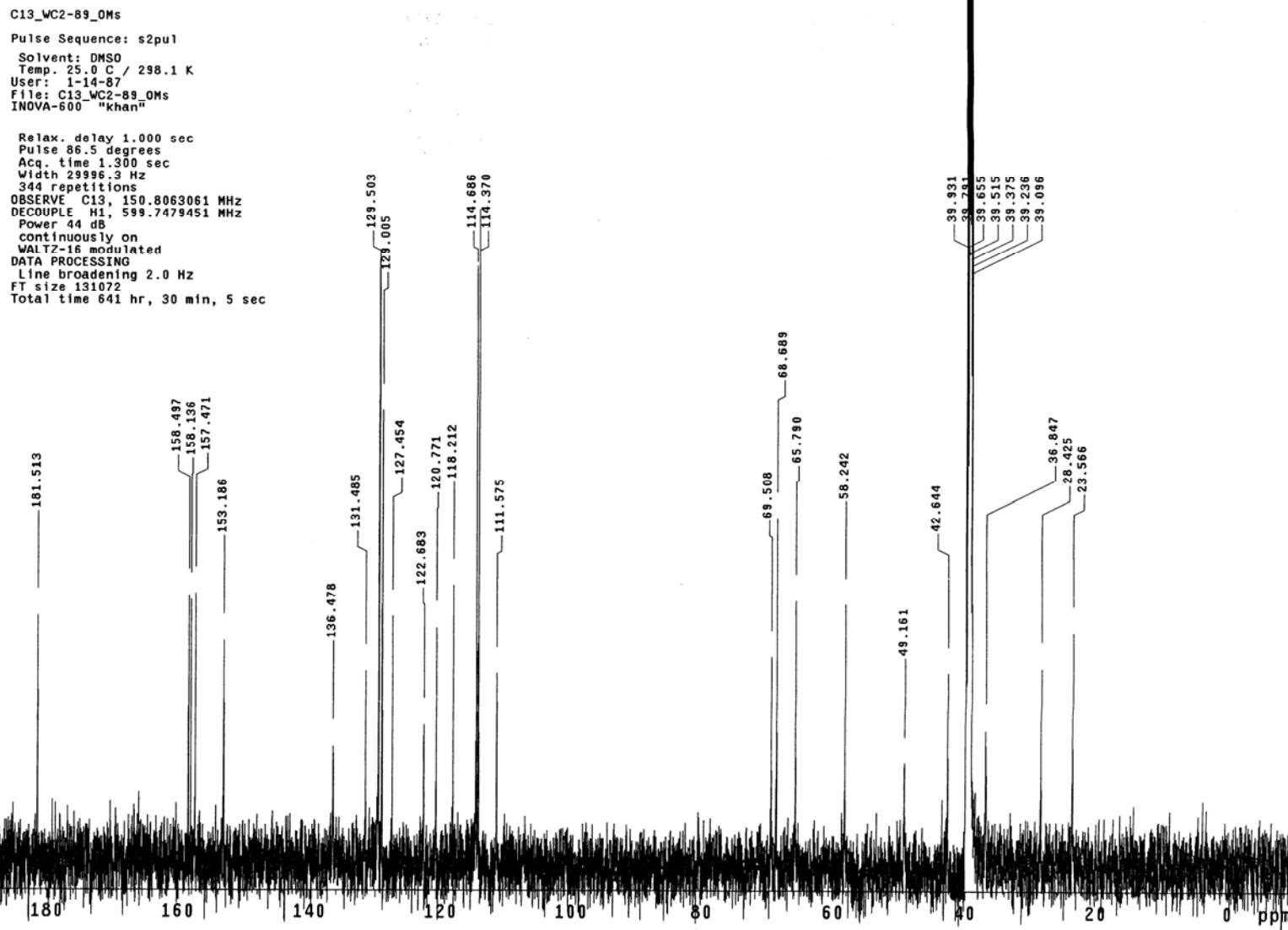
C13\_isatin+one-eq-F  
Pulse Sequence: s2pul



**Figure S9.** 150 MHz <sup>13</sup>C NMR Spectrum (DMSO-d<sub>6</sub>) of N-benzyl isatin **21** treated with 1 equivalent Bu<sub>4</sub>NF/H<sub>2</sub>O (10 mg **21** in 0.6 mL DMSO-d<sub>6</sub>)



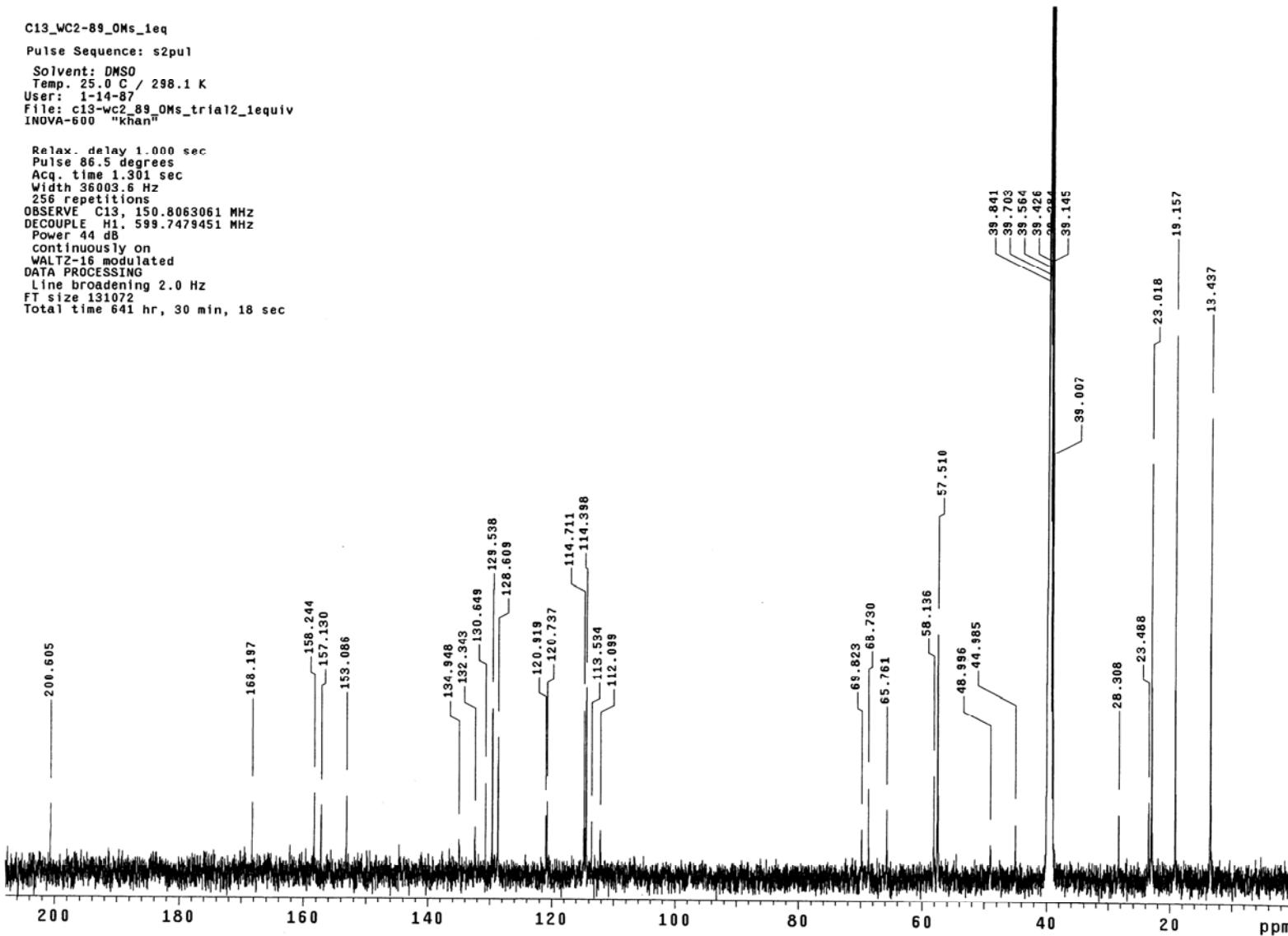
**Figure S10.** 150 MHz  $^{13}\text{C}$  NMR Spectrum (DMSO-d<sub>6</sub>) of N-benzyl isatin **21** treated with 3 equivalents Bu<sub>4</sub>NF/H<sub>2</sub>O (10 mg **21** in 0.6 mL DMSO-d<sub>6</sub>)



**Figure S11.** 150 MHz  $^{13}\text{C}$  NMR spectrum (DMSO-d6) of **7a** (10 mg **7a** in 0.6 mL DMSO-d6)

C13\_WC2-89\_0Ms\_1eq  
Pulse Sequence: s2pu1  
Solvent: DMSO  
Temp. 25.0 C / 298.1 K  
User: 1-14-87  
File: c13-wc2-89\_0Ms\_trial2\_1equiv  
INOVA-600 "khann"

Relax. delay 1.000 sec  
Pulse 86.5 degrees  
Acq. time 1.301 sec  
Width 36003.6 Hz  
256 repetitions  
OBSERVE C13, 150.8063061 MHz  
DECOUPLE H1, 599.7479451 MHz  
Power 44 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 2.0 Hz  
FT size 131072  
Total time 641 hr, 30 min, 18 sec



**Figure S12.** 150 MHz  $^{13}\text{C}$  NMR spectrum (DMSO-d6) of ring-opened **7a** (10 mg **7a** in 0.6 mL DMSO-d6 treated with 20  $\mu\text{L}$  1M  $\text{Bu}_4\text{NOH}/\text{H}_2\text{O}$ )

C13\_WC2-89

Pulse Sequence: s2pul

Solvent: DMSO

Temp. 25.0 C / 298.1 K

User: 1-14-87

File: C13\_WC2-89

INOVA-600 "khan"

Relax. delay 1.000 sec

Pulse 86.5 degrees

Acq. time 1.301 sec

Width 36003.6 Hz

112 repetitions

OBSERVE C13, 150.8063061 MHz

DECOPPLE H1, 599.7479451 MHz

Power 44 dB

continuously on

WALTZ-16 modulated

DATA PROCESSING

Line broadening 2.0 Hz

FT size 131072

Total time 641 hr, 30 min, 18 sec

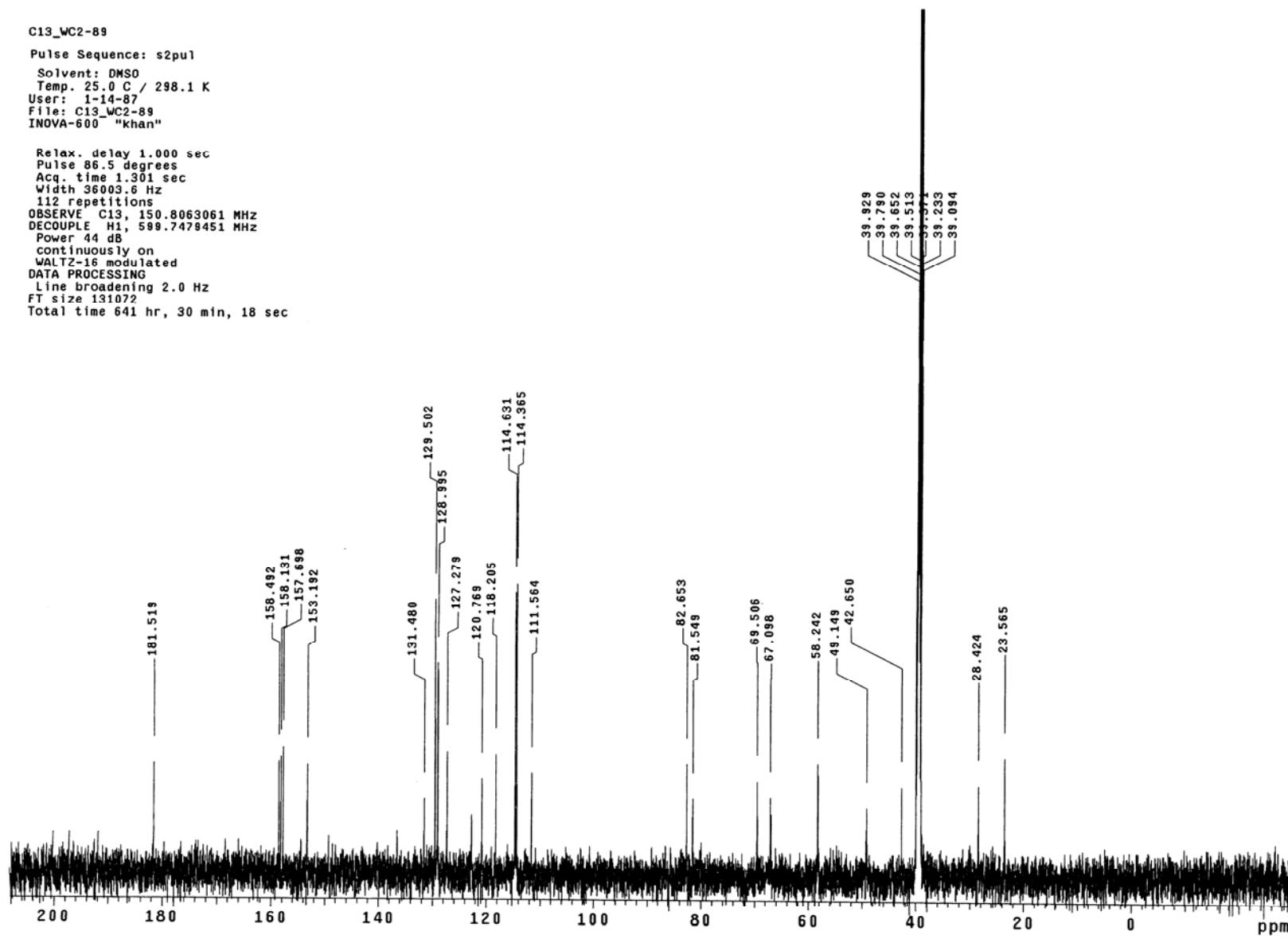
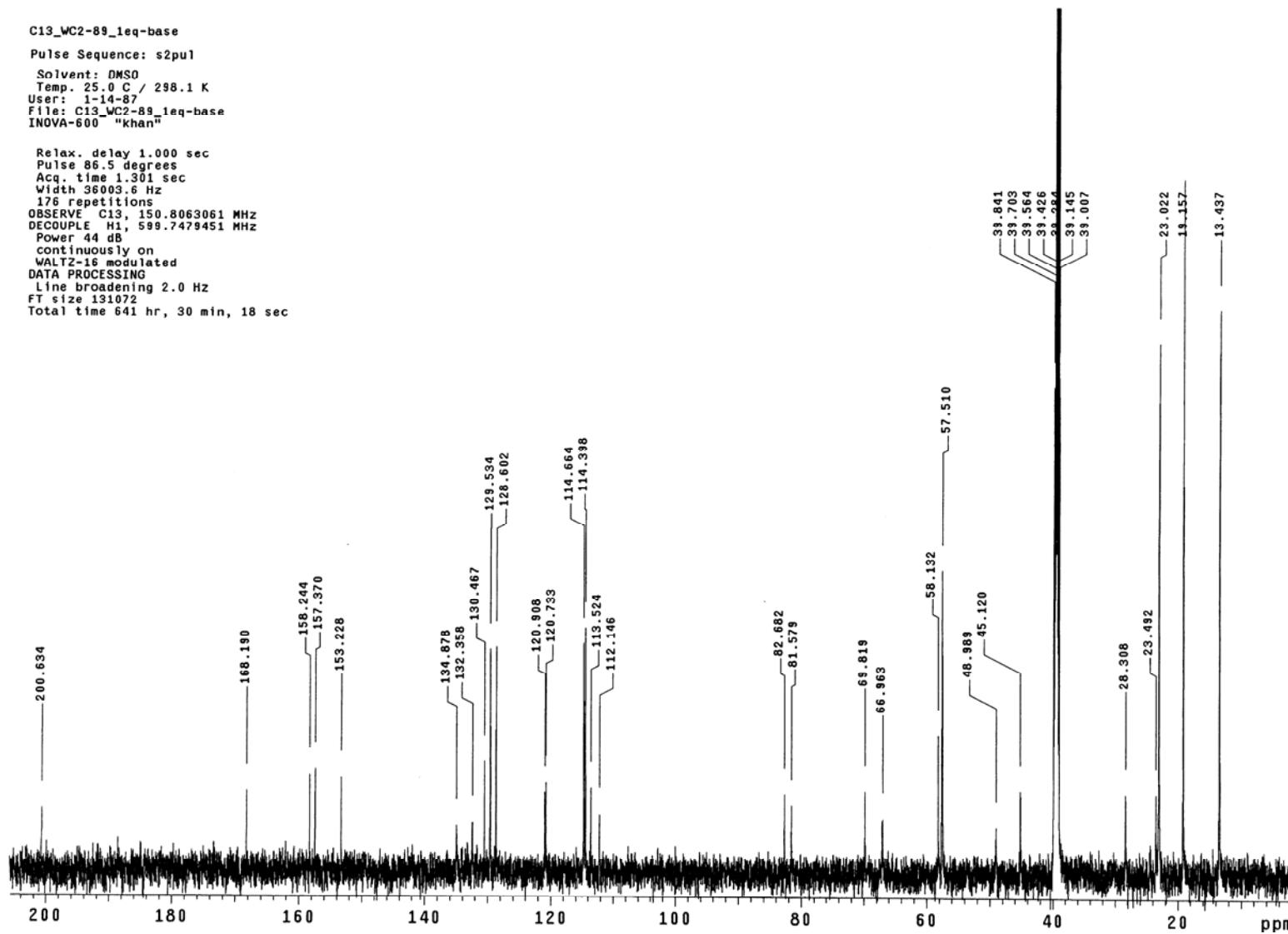


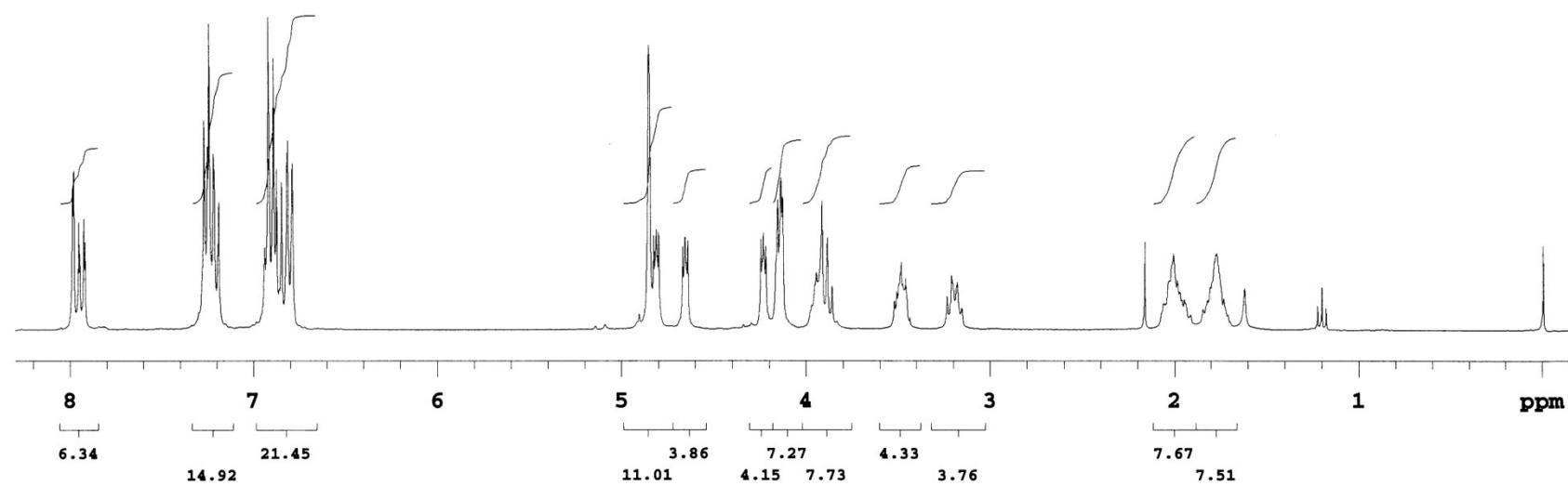
Figure S13. 150 MHz  $^{13}\text{C}$  NMR spectrum (DMSO-d6) of **2** (10 mg **2** in 0.6 mL DMSO-d6)

C13\_WC2-89\_1eq-base  
Pulse Sequence: s2pul  
Solvent: DMSO  
Temp. 25.0 C / 298.1 K  
User: 1-14-87  
File: C13\_WC2-89\_1eq-base  
INOVA-600 "khan"

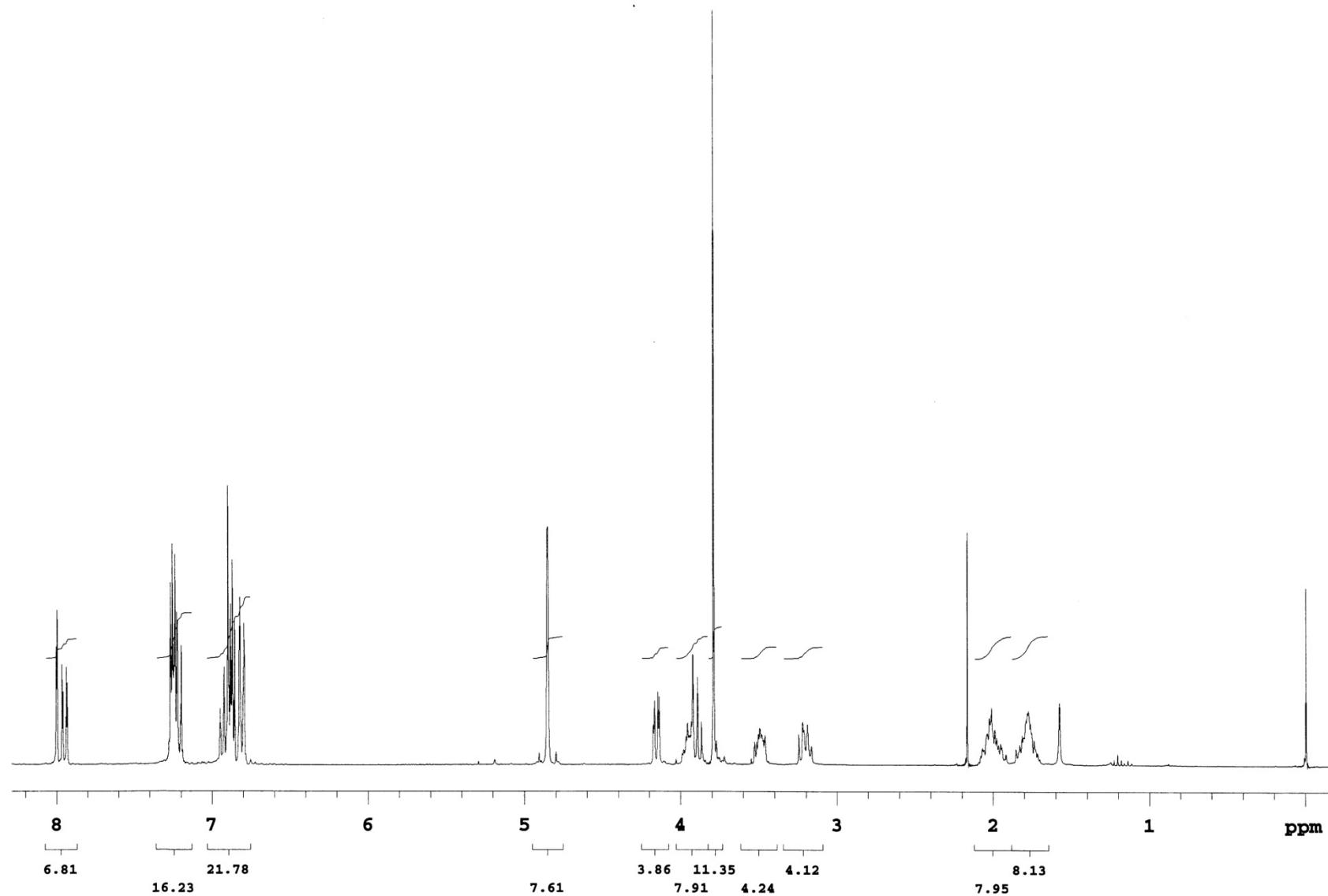
Relax. delay 1.000 sec  
Pulse 86.5 degrees  
Acq. time 1.301 sec  
Width 36003.6 Hz  
176 repetitions  
OBSERVE C13, 150.8063061 MHz  
DECOUPLE H1, 599.7479451 MHz  
Power 44 dB  
continuously on  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 2.0 Hz  
FT size 131072  
Total time 641 hr, 30 min, 18 sec



**Figure S14.** 150 MHz <sup>13</sup>C NMR spectrum (DMSO-d6) of ring-opened **2**(10 mg **2** in 0.6 mL DMSO-d6 treated with 20  $\mu$ L 1M Bu<sub>4</sub>NOH/H<sub>2</sub>O)



**Figure S15.** 300 MHz  ${}^1\text{H}$  NMR spectrum ( $\text{CDCl}_3$ ) of **2**



**Figure S16.** 300 MHz  ${}^1\text{H}$  NMR spectrum ( $\text{CDCl}_3$ ) of **4**

## STANDARD 1H OBSERVE

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exp1 std1h
      SAMPLE           DEC. & VT
date  Dec 14 2005 dfreq    299.945
solvent   CDCl3 dn      H1
file      exp dppr     30
      ACQUISITION dof      0
sfrq    299.945 dm      nnn
tn       H1 dppm     c
at      3.744 dmf     200
np      28952 dseq
sw      4000.0 dres    1.0
fb      2200 homo     n
bs       16 PROCESSING
tpwr     55 wtfille
pw       7.0 proc      ft
di       0 fn      not used
tof      0 math     f
nt       16
ct       16 werr
alock    n wexp
gain    not used wbs
      FLAGS wnt
il       n
in       n
dp       y
hs       nn
      DISPLAY
sp      -40.4
wp      2497.7
vs      500
sc       0
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hzmm    10.41
is      500.00
rf1     500.1
rfp      0
th      12
ins    100.000
nm cdc ph
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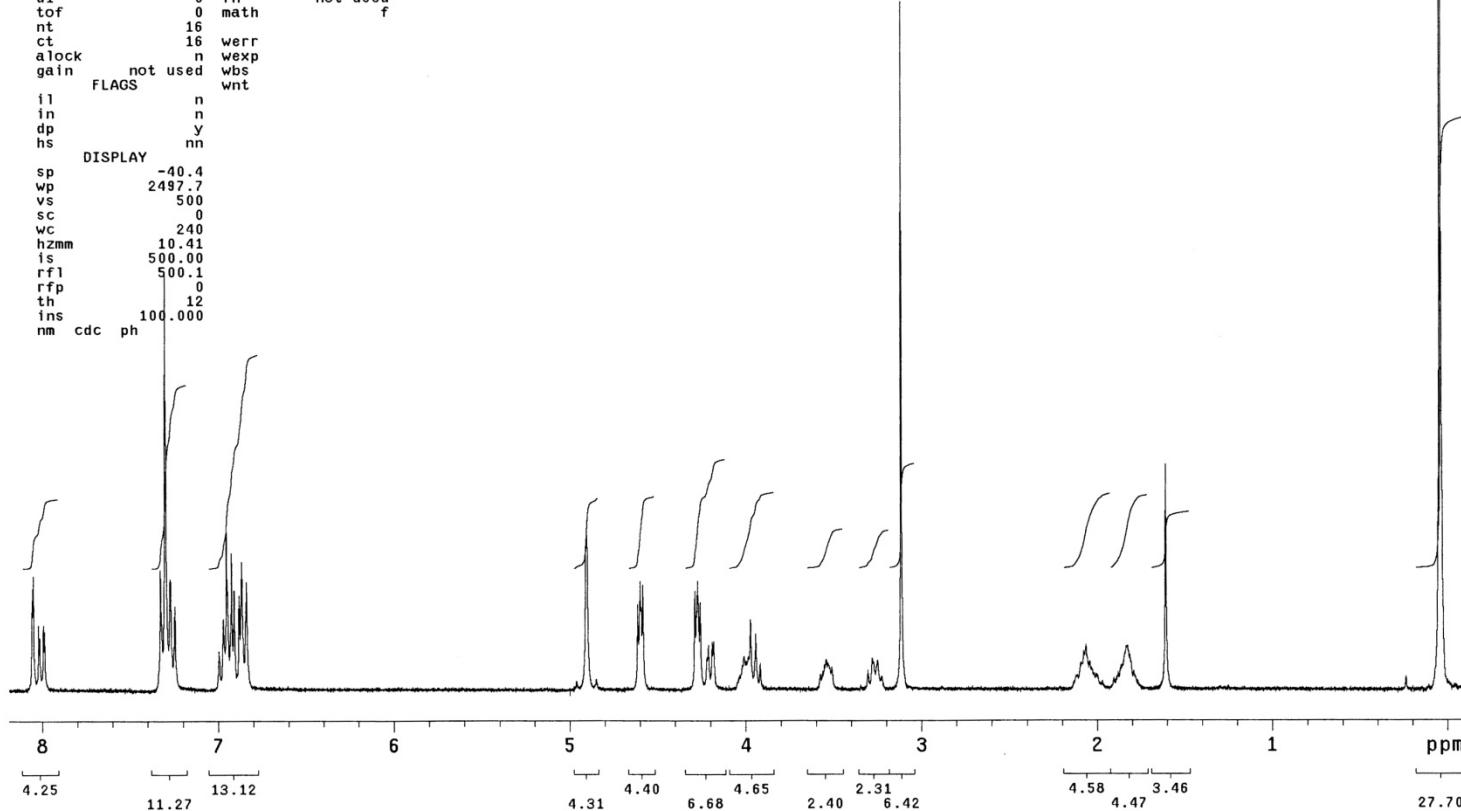
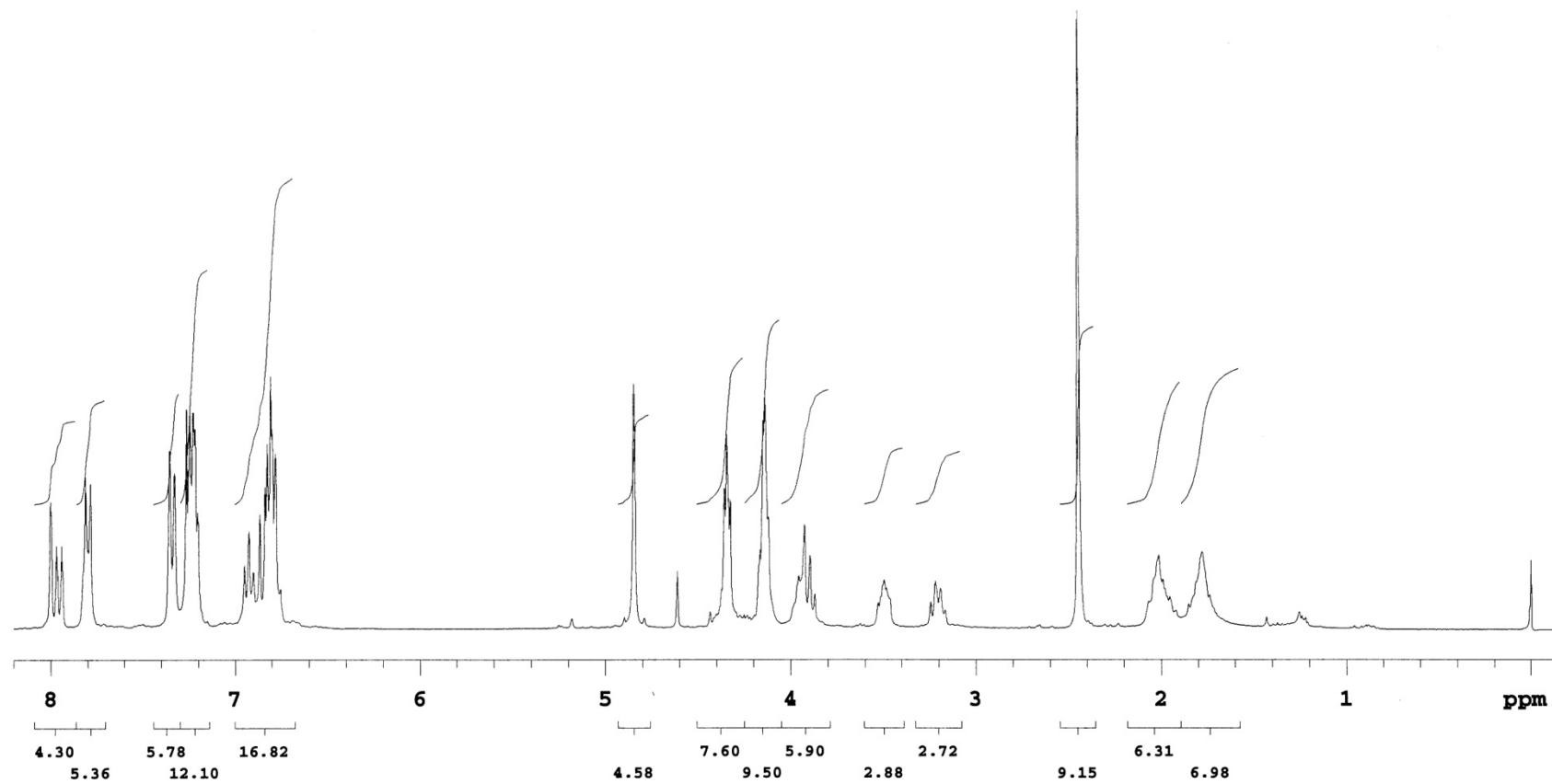


Figure S17. 300 MHz  $^1\text{H}$  NMR spectrum ( $\text{CDCl}_3$ ) of **7a**



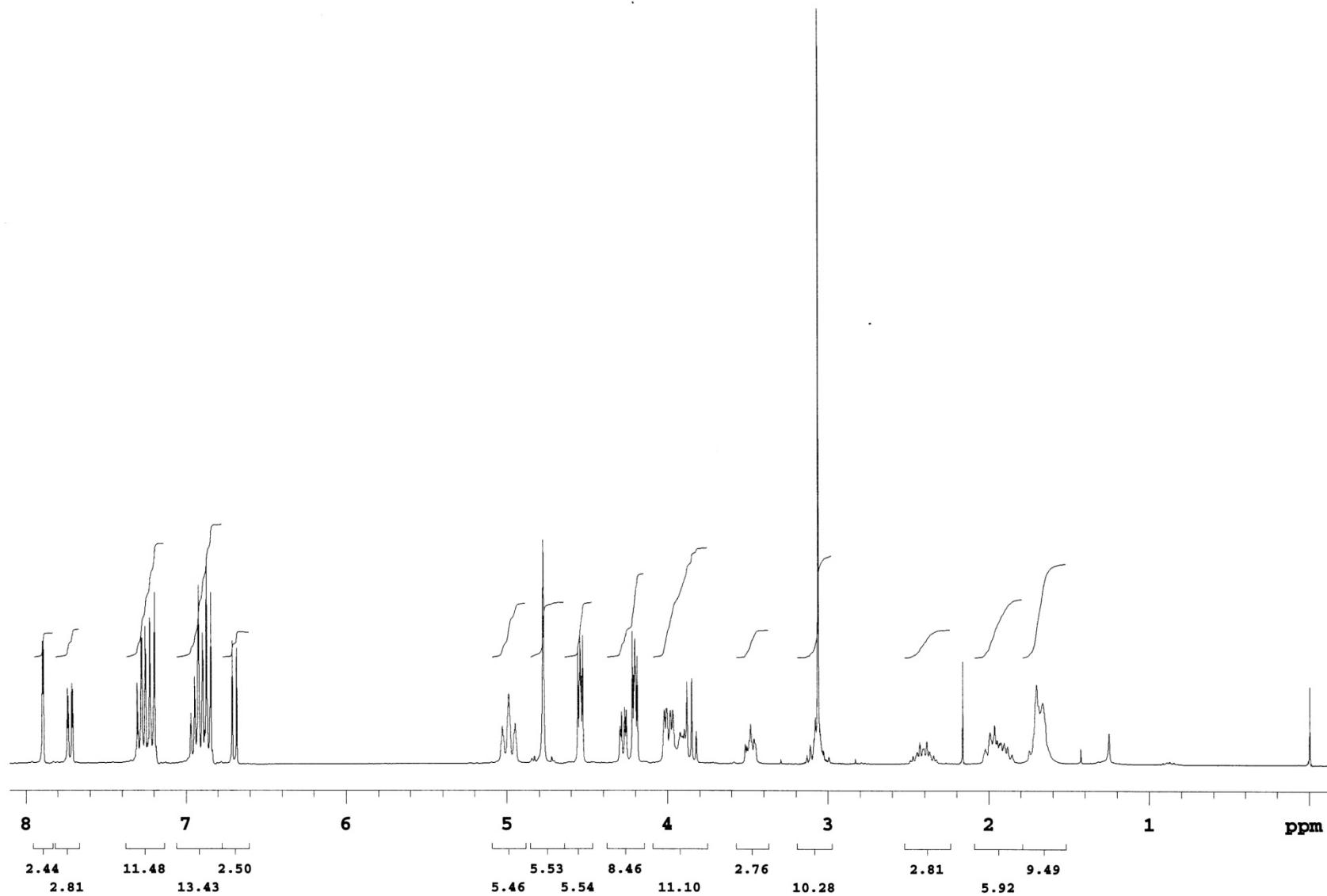
**Figure S18.** 300 MHz  $^1\text{H}$  NMR spectrum ( $\text{CDCl}_3$ ) of **7b**

STANDARD 1H OBSERVE

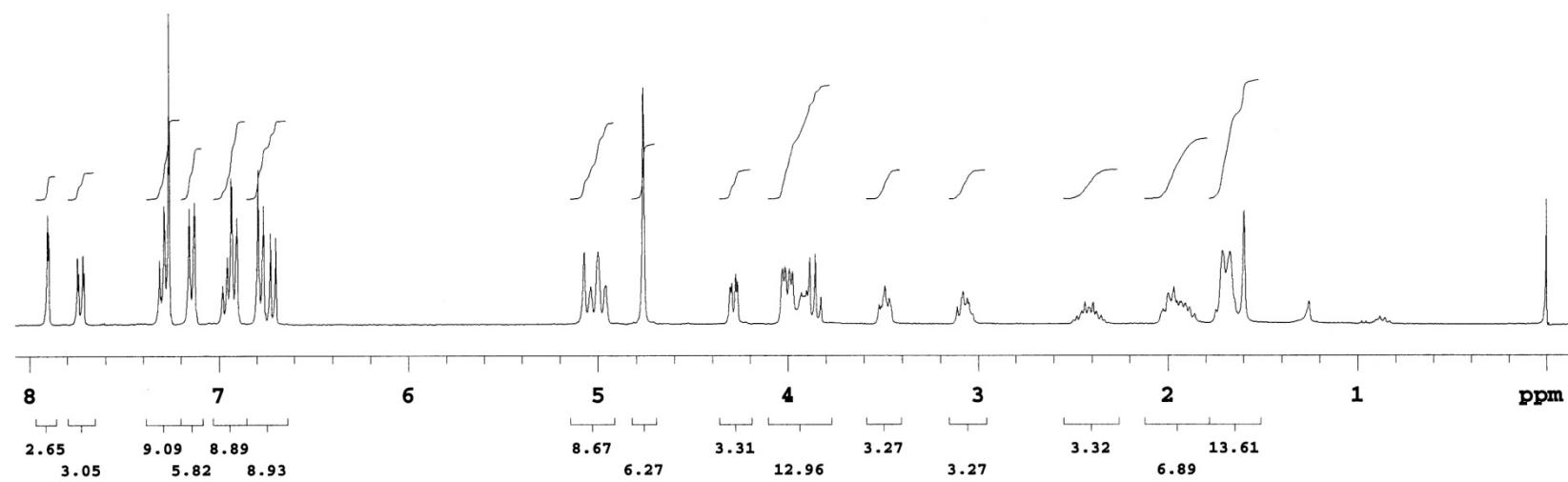
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date Apr 22 2007 dfrq 300.119  
solvent CDCl<sub>3</sub> dn H1  
file exp dpwr 43  
ACQUISITION dof 0  
sfrq 300.119 dm nnn  
tn H1 dmm c  
at 1.998 dmf 11500  
np 17984 PROCESSING  
sw 4500.5 wfile  
fb 2600 proc ft  
bs 16 fn not used  
ss 2  
tpwr 55 werr  
pw 7.0 wexp  
d1 1.000 wbs  
tof 0 wnt wft f ds vp=12~  
nt 16 dscale vsadj aph  
ct 16  
alock n  
gain not used  
FLAGS  
il n  
in n  
dp y  
DISPLAY  
sp -36.2  
wp 2490.2  
vs 50  
sc 0  
wc 250  
tmmn 9.96  
is 240.82  
rf1 795.5  
rfp 0  
th 3  
ins 100.000  
g m cdc ph 7 6 5 4 3 2 1 ppm  
6.82 22.08 3.30 7.86 8.02 4.16 8.47  
17.94 4.06 4.12 8.23 4.93

Figure S19. 300 MHz <sup>1</sup>H NMR spectrum (CDCl<sub>3</sub>) of **8**



**Figure S20.** 300 MHz  ${}^1\text{H}$  NMR spectrum ( $\text{CDCl}_3$ ) of **12**



**Figure S21.** 300 MHz  $^1\text{H}$  NMR spectrum ( $\text{CDCl}_3$ ) of **13**