

Supplementary Material

Synthesis and Biological Evaluation of Functionalised Tetrahydro- β -carboline Analogues as Inhibitors of *Toxoplasma gondii* Invasion.

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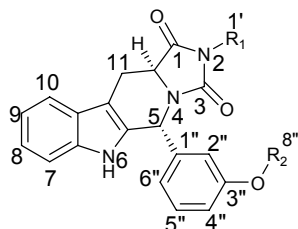
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Numbering scheme used for the β -carboline ring system

The following numbering scheme was used throughout the experimental section of the paper.



NOSEY spectrum of *cis*-6b and *trans*-7a

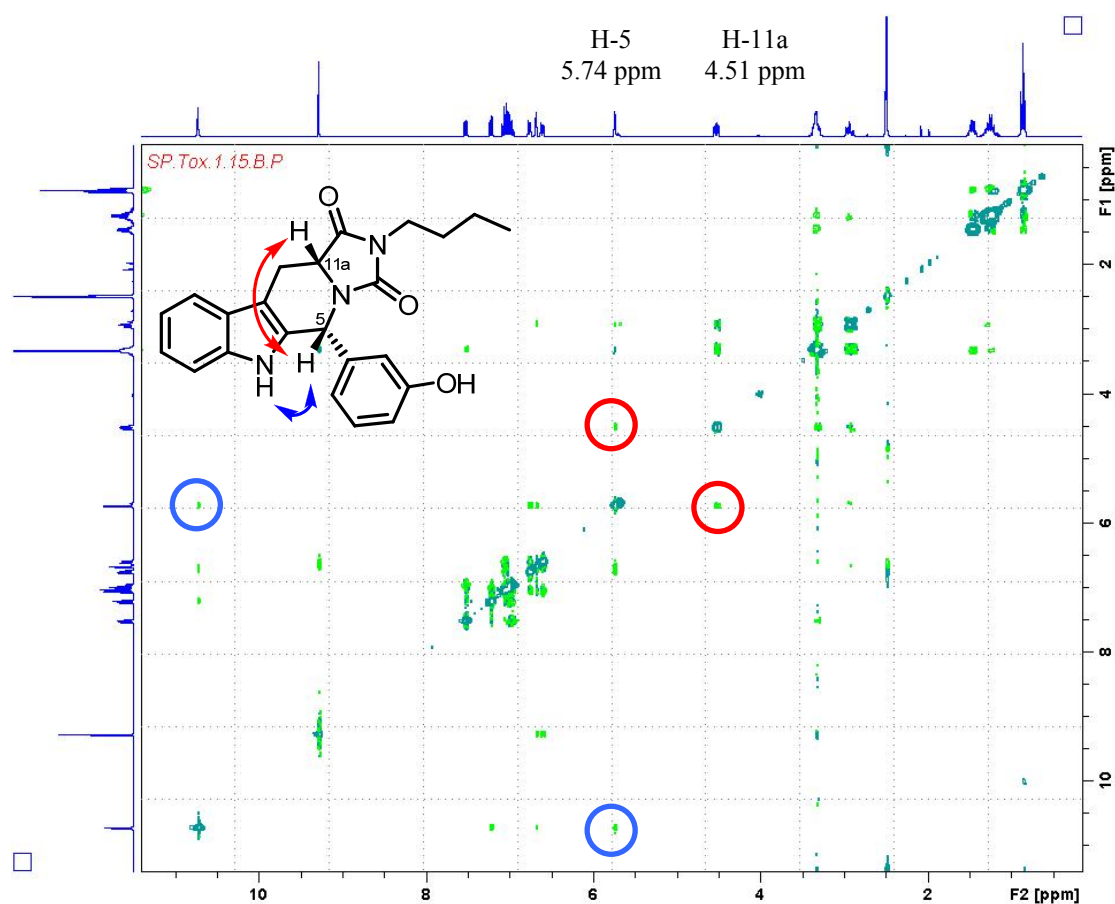


Figure S1. NOESY spectrum of *cis*-6b.

Whilst not conclusive, the observed lack of an nOe correlation between the C11a and C5-hydrogens in **7a** was also consistent with these two hydrogens being *trans*-oriented.

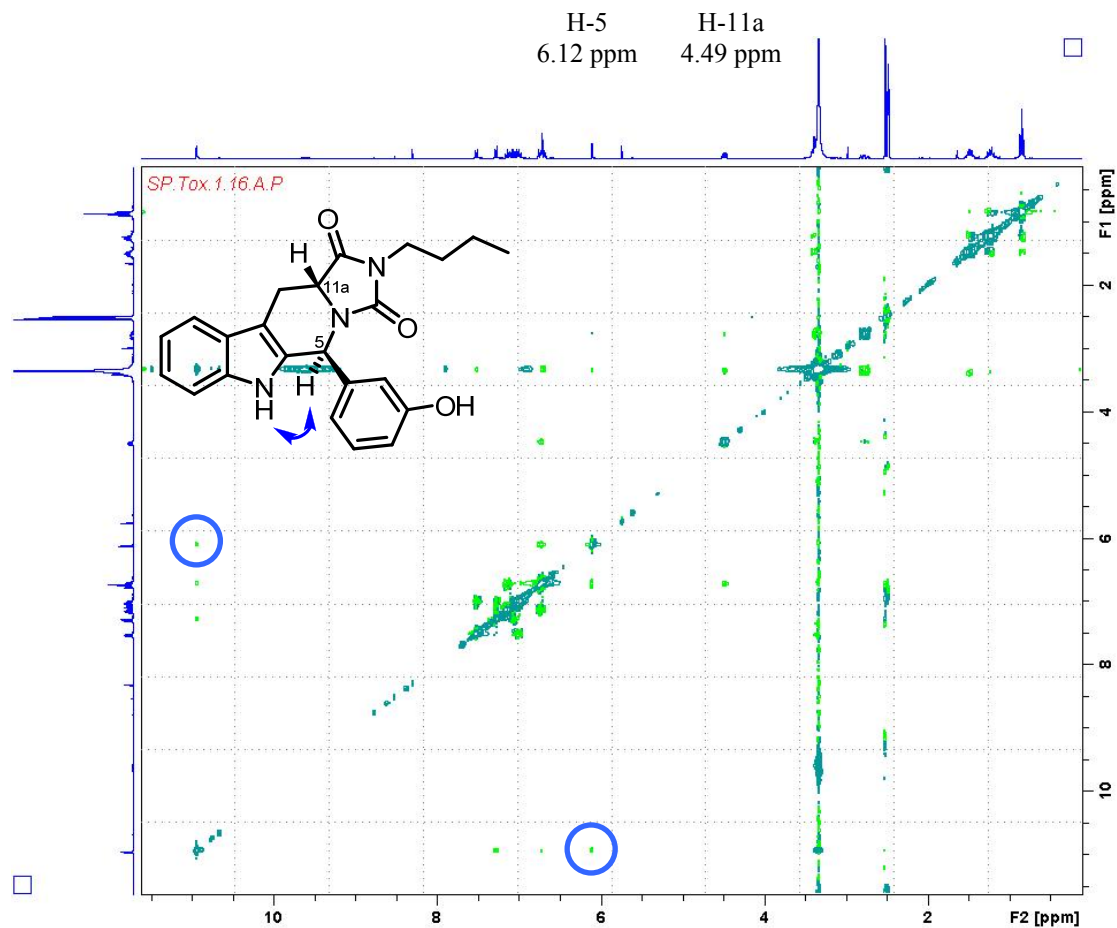


Figure S2. NOESY spectrum of *trans*-**7a**.

Bioinformatic analysis of *T.gondii* kinesins

	Inhibitor binding (Y125-E145)		Inhibitor Binding (I202-L227)
SpoCut7	SDGILSEG-----AGLI PRALYQLFSS 129		ESYIKNAGDGLRLLREGSHRRQVAATKCN 209
AnBimC	TLGILSDN-----AGIIPRVLYSLFAK 81		FTYIDSATAGIKLLQQGSHKQVAATKCN 160
ScKip1	NI-LLGEH-----AGIIPRVLDLDFKE 91		EIFINSAHEGLNLLMQGSLKRKVAATKCN 179
ScCin8	YNGELSDA-----AGIIPRVLLKLFDT 87		EFHITNAMEGLNLLQKGLKHRQVASTKMN 270
NtKRP125	KSGPNGEL-----PQEAGVIPRAVKQVFD 85		EIVTSANEIIFTLLEKSAKRRTAETLLN 171
XlEg5	EEFTWEQ-----DPLAGIIPRTLHQIFEK 85		EISVHNKDEVYHILERGAARKKTASTLMN 166
XlEg52	EEFTWEQ-----DPLAGIIPRTLHQIFEK 85		EISVHNKDEVYQILERGAARKKTASTLMN 166
HsKSP	EEYTWEE-----DPLAGIIPRTLHQIFEK 85		EITVHNKDEVYQILEKGAARKTTAATLMN 166
CeF23B12	KSSIDDP-----TTGIIPRAVEDIFEQ 75		EVPVRNRSDVFKLLQLGAEKRRTAATLMN 157
DmKlp61F	LKSSWED-----DSDIGIIPRALSHLFDE 82		EIPVHSKDDVYKLLKKGKERRKTATLMN 161
BmKRP	-FTTWQK-----DPLAGIIPRALSQLFDE 81		EITVYNKKEVFRIMAQQERKKVASTLMN 161
113.m00758	GESIYSANDEISENADCGLVGRSVQRIFST 316		EKEVKSEREV LALLRAAAPRRSFAASSAN 482
20.m05941	QEKVDEERGDRKLRCDGLLPRMINGIFTW 370		DKLVREPHEAFSVLEKAIKARRAATFMN 453
42.m03387	LLSGEQSRDYVVYKQAKALRVQTIINRLIQH 398		IFEVKSTDELIHLLMEGTFNRAFRAATKQN 462
641.m01532	R-----ER-----REERCVLPRALNYLFSK 175		RLPVFDEAQLQIFAQGVKNRRTAETRLN 329

Figure S3 Alignments comparing the two regions of human Eg5 shown to interact with **1** with other family 5 kinesins and the four candidate family 5 kinesins identified in *T. gondii*. The sequences of known family 5 kinesins used in the alignment are; *S. pombe* Cut7, *A. nidulans* BimC, *S. cerevisiae* Kip1 and Cin8, *N. tabacum* KRP125, *X. laevis* Eg5 and Eg52, *H. sapiens* KSP (Eg5), *C. elegans* F23B12, *D. melanogaster* Klp61F, and *B. mori* KRP. The *T. gondii* proteins that were aligned are; 113.m00758, 20.m05941, 42.m03387, and 641.m01532. The regions of human Eg5 that have been shown to interact with 5118793, Tyr125-Glu145 (left) and Ile202-Leu227 (right), and the corresponding regions of the other proteins are shown.

The yeast-3-hybrid approach: Y3H is based on the more familiar yeast two-hybrid (Y2H) assay that allows the detection of protein-protein interactions. In the Y2H assay the two functional domains of a transcriptional activator, the DNA-binding domain (DBD) and the activation domain (AD), are independently fused to 1) a protein of interest, the "bait" and 2) a cDNA library encoding potential interacting proteins, the "prey". A transcriptional read-out is obtained when the bait protein interacts with a prey protein reconstituting the transcriptional activator by bringing the DBD and AD into close proximity. The analogous Y3H assay allows the detection of small molecule-protein interactions by making the transcriptional read-out dependent on the interaction of the bait and prey proteins through a "dimerizing" hybrid small molecule, also known as a chemical inducer of dimerization (CID) (i.e. the bait and prey proteins themselves do not interact) (Fig. S4).

The Y3H system used in this study was developed by the laboratory of Virginia Cornish^{S1}. The Cornish Y3H system utilizes two transcriptional reporters of interaction, *LEU2* and *lacZ*, each preceded by an array of LexA operator sites. (Figure S4, modified from reference S1). The system is composed of a DHFR-LexA DBD fusion protein and a B42 AD fusion with a cDNA library of putative targets of interest. It relies on DHFR's high affinity interaction with methotrexate (MTX) to anchor a MTX-small molecule CID to the DHFR-LexA DBD fusion (Fig. S4). In this system transcriptional activation is dependent upon binding of the portion of the CID containing the small molecule of interest (the bait), to its putative target-B42 AD fusion.

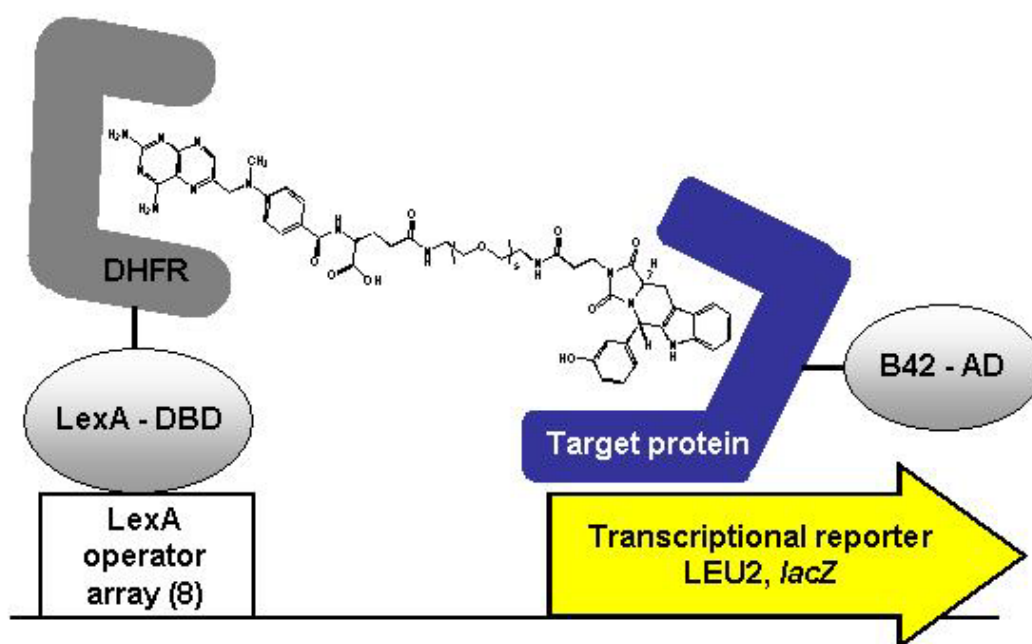


Figure S4. Schematic representation of the yeast-3-hybrid approach adapted from figure in Reference S1.

➤ **Literature compounds with improved characterisation.**

General procedure for the Pictet-Spengler reaction

D-(5a) or **L-Tryptophan (5b)** (29 mmol) was suspended in 0.05 M H₂SO₄ (42 mL) and the desired aldehyde (2.0 equiv, 59 mmol) was added. The reaction mixture was stirred at reflux for 4-6 hours and the solid product collected by filtration and thoroughly rinsed with diethyl ether. Whilst **2a** and **2b** are known compounds,¹² optical rotations and melting points have not been previously reported. Previous reports of the synthesis of **2d**²⁵ described the formation of a diastereomeric mixture only and provided no spectroscopic data.

(1R,3R)-1-(3-Hydroxy-phenyl)-2,3,4,9-tetrahydro-1H-β-carboline-3-carboxylic

acid (2a)¹² Yield 70%, yellow solid. m.p. 265-266 °C; ν_{\max} (KBr) 3450, 1636, 1560, 1394, 1298 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 7.44 (1H, d, *J* 8.0, H5), 7.15-7.26 (2H, m, H5', H8), 7.03-7.01 (1H, m, H6), 7.01-6.90 (1H, m, H7), 6.86 (1H, d, *J* 7.6, H6'), 6.81-6.73 (2H, m, H4', H2'), 5.29 (1H, s, H1), 3.75 (1H, dd, *J* 11.4, 4.2, H3), 3.10 (1H, dd, *J* 13.6, 4.2, H4_{syn}), 2.84 (1H, dd, *J* 13.6, 12.5, H4_{anti}); ¹³C NMR (100 MHz, DMSO-d₆) δ 172.2 (C=O), 157.3 (C), 140.9 (C), 136.5 (C), 133.4 (C), 129.3 (CH), 126.3 (C), 120.8 (CH), 119.7 (CH), 118.4 (CH), 117.6 (CH), 115.9 (CH), 115.3 (CH), 111.3 (CH), 107.6 (C), 57.6 (CH), 56.8 (CH) 24.5 (CH₂); LRMS (ES⁻) *m/z*: 307.20 [M-H]; $[\alpha]_{\text{D}}^{20} = +147.2$ (*c* = 0.0075, MeOH).

(1S,3S)-1-(3-Hydroxy-phenyl)-2,3,4,9-tetrahydro-1H-β-carboline-3-carboxylic

acid (2b)¹² Yield 50%, yellow solid. m.p. 262-263°C °C; ν_{\max} (KBr) 3327 (m), 1638 (s), 1606 (m), 1465 (m) 1396 (m), 1298 cm⁻¹ (w); ¹H NMR (500 MHz, DMSO-d₆) δ 10.45 (1H, s, NH), 7.46 (1H, d, *J* 8.0, H5), 7.25-7.15 (2H, m, H5', H8), 7.05-7.02 (1H, m, H6), 7.00-6.96 (1H, m, H7), 6.88 (1H, d, *J* 7.5, H6'), 6.80-6.77 (2H, m, H4', H2'), 5.31 (1H, s, H1), 3.77 (1H, dd, *J* 11.5, 4.5, H3), 3.14-3.10 (1H, m, H4_{syn}), 2.89-2.83 (1H, m, H4_{anti}); ¹³C NMR (125 MHz, DMSO-d₆) δ 172.2 (C=O), 157.3 (C), 141.0 (C), 136.5 (C), 133.4 (C), 129.3 (CH), 126.3 (C), 120.9 (CH), 119.8 (CH), 118.5 (CH), 117.7 (CH), 115.9 (CH), 115.3 (CH), 111.3 (CH), 107.6 (C), 57.6 (CH), 56.8 (CH) 24.5 (CH₂); LRMS (ES⁺) *m/z*: 331.02 [M+Na]⁺; HRMS (ES⁺) [M+Na]⁺ *m/z* expected for C₁₈H₁₆N₂O₃Na 331.1059 found 331.1059; $[\alpha]_{\text{D}}^{20} = -139.5$ (*c* = 0.005, MeOH).

(1R,3R)-1-(4-Hydroxy-phenyl)-2,3,4,9-tetrahydro-1H- β -carboline-3-carboxylic acid (2c)²⁵ Yield 34%, yellow solid. m.p. 263-264°C; ν_{\max} (KBr) 3417 (m), 1652 (s), 1600 (m), 1400 (m), 1278 cm^{-1} (m); ^1H NMR (500 MHz, DMSO- d_6) δ 10.52 (1H, s, NH), 7.47 (1H, d, J 8.0, H5), 7.26-7.23 (3H, m, H8, H2'), 7.07-7.04 (1H, m, H7), 7.01-6.98 (1H, m, H6), 6.84-6.81 (2H, AA'BB', H3'), 5.45 (1H, s, H1), 3.80 (1H, dd, J 12.0, 4.5, H3), 3.19 (1H, dd, J 15.5, 4.5, H4_{syn}), 2.96-2.91 (1H, m, H4_{anti}); ^{13}C NMR (125 MHz, DMSO- d_6) δ 171.1 (C=O), 158.1 (C), 136.6 (C), 132.2 (C), 131.0 (CH), 127.4 (C), 126.2 (C), 121.1 (CH), 118.6 (CH), 117.9 (CH), 115.1 (CH), 111.4 (CH), 108.0 (C), 57.5 (CH), 57.2 (CH), 23.7 (CH₂); LRMS (ES⁺) m/z : 309.06 [M+H]⁺; HRMS (ES⁺) [M+H]⁺ m/z expected for C₁₈H₁₇N₂O₃ 309.1239 found 309.1239; $[\alpha]_{\text{D}}^{20} = +141$ ($c = 0.1$, MeOH).

(1S,3S)-1-(4-Hydroxy-phenyl)-2,3,4,9-tetrahydro-1H- β -carboline-3-carboxylic acid (2d)²⁵ Yield 52%, off white solid. m.p. 263-264°C; ν_{\max} (KBr) 3417 (m), 1653 (s), 1617 (m), 1401 (s), 1278 cm^{-1} (m); ^1H NMR (300 MHz, DMSO- d_6) δ 10.52 (1H, s, NH), 7.54 (1H, d, J 7.2, H5), 7.31-7.26 (3H, m, H2' & H8), 7.13-7.02 (2H, m, H6 & H7), 6.87-6.84 (2H, AA'BB', H3'), 5.44 (1H, s, H1), 3.82 (1H, dd, J 11.5, 4.0, H3), 3.22 (1H, dd, J 15.6, 4.0, H4_{syn}), 3.00-2.91 (1H, m, H4_{anti}); ^{13}C NMR (75 MHz, DMSO- d_6) δ 171.1 (C=O), 158.0 (C), 136.5 (C), 132.4 (C), 130.9 (CH), 127.8 (C), 126.2 (C), 121.1 (CH), 118.6 (CH), 117.8 (CH), 115.1 (CH), 111.3 (CH), 108.0 (C), 57.4 (CH), 57.1 (CH), 23.8 (CH₂); LRMS (ES⁺) m/z : 309.06 [M+H]⁺; HRMS (ES⁺) [M+H]⁺ m/z expected for C₁₈H₁₇N₂O₃ 309.1239 found 309.1239; $[\alpha]_{\text{D}}^{20} = -141$ ($c = 0.1$, MeOH).

General procedure for the synthesis of the *cis*-hydantoins (6a-6d)

To a solution of the desired carboxylic acid **2** (13 mmol) in a mixture of anhydrous acetone (60 mL) and anhydrous DMSO (24 mL) was added the corresponding isocyanate (1.0 eq, 13 mmol). The solution was stirred at reflux for 4 hrs then poured onto H₂O (200 mL). The aqueous phase was extracted with DCM (3 x 100 mL) and the combined organic layers washed with brine (200 mL), dried (Na₂SO₄) and reduced *in vacuo*. Purification of the residue by chromatography on silica gel (EtOAc:Hex 20:80) afforded the desired product. **6a** and **6b** have been reported,

however, the melting points previously associated with these compounds are confusing.¹²

(5R,11aR)-2-Butyl-5-(3-hydroxyphenyl)-6H-1,2,3,5,11,11a-hexahydro-imidazo[1,5-b]- β -carboline-1,3-dione (6a)¹² Yield 65%, yellow solid. m.p. 186-187 °C (lit. 134° C)¹²; ν_{\max} (KBr) 3433, 2956, 1765, 1701 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 10.74 (1H, s, NH), 9.28 (1H, s, OH), 7.53 (1H, d, *J* 7.9, H10), 7.30 (1H, d, *J* 7.9, H7), 7.17-7.15 (1H, m, H5''), 7.10-7.08 (1H, m, H9), 7.02-7.01 (1H, m, H8), 6.77 (1H, d, *J* 7.7 H6''), 6.70 (1H, s, H2''), 6.62 (1H, d, *J* 8.0, H4''), 6.13 (1H, s, H5), 4.53 (1H, dd, *J* 11.7, 4.3, H11a), 3.41-3.30 (3H, m, 2 x H1', H11_{syn}), 2.90-2.80 (1H, m, H11_{anti}), 1.50-1.45 (2H, m, H2'), 1.20-1.10 (2H, m, H3'), 0.86 (3H, t, H4'); ¹³C NMR (100 MHz, DMSO-d₆) δ 173.3 (C=O), 158.9 (C), 154.9 (C=O), 142.0 (C), 137.4 (C), 132.0 (C), 130.4 (CH), 126.5 (C), 122.3 (CH), 119.5 (CH), 118.9 (CH), 118.7 (CH), 115.9 (CH), 115.6 (CH), 112.1 (CH), 106.6 (C), 53.5 (CH), 52.1 (CH), 38.4 (CH₂), 32.9 (CH₂), 23.5 (CH₂), 20.1 (CH₂), 14.2 (CH₃); LRMS (ES⁺) *m/z*: 390.40 [M+H]⁺; [α]_D²⁰ = +58.7 (*c* = 0.002, EtOAc).

(5S,11aS)-2-Butyl-5-(3-hydroxyphenyl)-6H-1,2,3,5,11,11a-hexahydro-imidazo[1,5-b]- β -carboline-1,3-dione (6b)¹² Yield 92%, yellow solid. m.p. 191-192 °C (lit. 225° C)¹² ν_{\max} (KBr) 3340 (m), 2952 (w), 1763 (m), 1694 (s), 1457 cm⁻¹ (m); ¹H NMR (500 MHz, DMSO-d₆) δ 10.75 (1H, s, NH), 9.31 (1H, s, OH), 7.54 (1H, d, *J* 7.5, H10), 7.24 (1H, d, *J* 7.5, H7), 7.09-7.03 (2H, m, H5' & H8), 7.01-6.98 (1H, m, H9), 6.78 (1H, d, *J* 7.5 H6''), 6.70-6.69 (1H, m, H2''), 6.63 (1H, ddd, *J* 8.0, 2.5, 1.0, H4''), 5.76 (1H, s, H5), 4.54 (1H, dd, *J* 11.5, 4.5, H11a), 3.41-3.31 (3H, m, H11_{syn} & H1'), 2.98-2.92 (1H, m, H11_{anti}), 1.51-1.45 (2H, m, H2'), 1.30-1.22 (2H, m, H3'), 0.87 (3H, t, *J* 7.5, H4'); ¹³C NMR (125 MHz, DMSO-d₆) δ 171.7 (C=O), 157.2 (C), 154.1 (C=O), 142.1 (C), 136.6 (C), 134.8 (C), 129.1 (CH), 125.8 (C), 121.3 (CH), 118.8 (CH), 118.1 (CH), 118.08 (CH), 114.4 (CH), 114.1 (CH), 111.3 (CH), 104.8 (C), 57.5 (CH), 55.6 (CH), 37.4 (CH₂), 29.6 (CH₂), 21.8 (CH₂), 19.3 (CH₂), 13.5 (CH₃); LRMS (ES⁺) *m/z*: 412.10 [M+Na]⁺; HRMS (ES⁺) [M+Na]⁺ *m/z* expected for C₂₃H₂₃N₃O₃Na 412.1637 found 412.1637; [α]_D²⁰ = -55.9 (*c* = 0.0054, EtOAc).

General procedure for epimerisation to the *trans*-hydantoin (7a-7m)

A mixture of the desired *cis*-hydantoin **6** (8.00 mmol) and K₂CO₃ (1.3 equiv, 10.4 mmol) in anhydrous MeCN (120 mL) was stirred at reflux for 4 hrs. The solids were removed *via* filtration and the solvent reduced *in vacuo*. Purification of the residue by chromatography on silica gel (EtOAc:Hex 20:80) afforded the desired product. **7a** and **7b** have been reported, however, the m.p. previously associated with these compounds are confusing.¹² For spectroscopic details associated with **7e-7j** see supplementary material.¹⁰

(5R,11aS)-2-Butyl-5-(3-hydroxyphenyl)-6H-1,2,3,5,11,11a-hexahydro-imidazo[1,5-b]-β-carboline-1,3-dione (7a)¹² Yield 80%, yellow solid. m.p. 187-189 °C (lit. 107 °C)¹²; ν_{\max} (KBr) 3433, 2956, 1765, 1701 cm⁻¹; ¹H NMR (400 MHz, DMSO-d₆) δ 10.93 (1H, s, NH), 9.46 (1H, s, OH), 7.54 (1H, d, *J* 7.8, H10), 7.30 (1H, d, *J* 8.0, H7), 7.20-6.95 (3H, m, H5'', H9, H8), 6.77 (1H, d, *J* 7.7 H6''), 6.72-6.70 (2H, m, H4'', H2''), 6.13 (1H, s, H5), 4.49 (1H, dd, *J* 10.9, 5.7, H11a), 3.40 (2H, t, *J* 7.8, H1'), 3.39-3.30 (1H, m, H11_{syn}), 2.80 (1H, dd, *J* 15.1, 10.9, H11_{anti}), 1.45-1.31 (2H, m, H2'), 1.29-1.10 (2H, m, H3'), 0.86 (3H, t, *J* 7.3, H4'); ¹³C NMR (100 MHz, DMSO-d₆) δ 173.4 (C=O), 158.9 (C), 155.0 (C=O), 142.06 (C), 137.40 (C), 132.0 (C), 130.4 (CH), 126.5 (C), 122.4 (CH), 119.5 (CH), 118.8 (CH), 118.7 (CH), 116.0 (CH), 115.6 (CH), 112.1 (CH), 106.7 (C), 53.5 (CH), 52.1 (CH), 38.4 (CH₂), 32.9 (CH₂), 23.5 (CH₂), 20.1 (CH₂), 14.2 (CH₃); LRMS (ES⁺) *m/z*: 412.2 [M+Na]⁺; [α]_D²⁰ = -215.5 (*c* = 0.006, EtOAc).

(5S,11aR)-2-Butyl-5-(3-hydroxyphenyl)-6H-1,2,3,5,11,11a-hexahydro-imidazo[1,5-b]-β-carboline-1,3-dione (7b)¹² Yield 99%, yellow solid. m.p 185-186°C (lit. 118 °C)¹²; ν_{\max} (KBr) 3338 (m), 2958 (w), 1763 (m), 1700 (s), 1457 cm⁻¹ (s); ¹H NMR (500 MHz, DMSO-d₆) δ 11.00 (1H, s, NH), 7.59 (1H, d, *J* 8.0, H10), 7.36 (1H, d, *J* 8.0, H7), 7.20 (1H, dd, *J* 8.0, 8.0, H5''), 7.17-7.13 (1H, m, H8), 7.09-7.06 (1H, m, H9), 6.81-6.76 (3H, m, H2'', H4'' & H6''), 6.18 (1H, s, H5), 4.56 (1H, dd, *J* 10.5, 5.5, H11a), 3.48-3.41 (3H, m, H1' & H11_{syn}), 2.88-2.85 (1H, m, H11_{anti}), 1.59-1.53 (2H, m, H2'), 1.34-1.27 (2H, m, H3'), 0.92 (3H, t, *J* 7.3, H4'); ¹³C NMR (125 MHz, DMSO-d₆) δ 172.6 (C=O), 158.0 (C), 154.3 (C=O), 141.3 (C), 136.7 (C), 131.3 (C), 129.7 (CH), 125.8 (C), 121.6 (CH), 118.8 (CH), 118.2 (CH), 118.0 (CH), 115.1 (CH), 114.8 (CH), 111.4 (CH), 105.9 (C), 52.7 (CH), 51.4 (CH), 37.7 (CH₂),

29.6 (CH₂), 22.8 (CH₂), 19.4 (CH₂), 13.5 (CH₃); LRMS (ES⁺) *m/z*: 412.11 [M+Na]⁺; HRMS (ES⁺) [M+Na]⁺ *m/z* expected for C₂₃H₂₃N₃O₃Na 412.1637 found 412.1637; [α]_D²⁰ = +204.8 (*c* = 0.007, EtOAc).

General procedure for the synthesis of the *trans*-3-thioxohydantoins (8a & 8b)

To a solution of the desired carboxylic acid **2a** or **2b** (1.0 mmol) in a mixture of anhydrous acetone (3.5 mL) and anhydrous DMSO (1.5 mL) was added the *n*-butylisothiocyanate (1.0 eq, 1.0 mmol). The solution was stirred at reflux for 2 hrs, then poured onto H₂O (200 mL). Extracted with DCM (3 x 100 mL) and the combined organic layers washed with brine (200 mL), dried (Na₂SO₄) and reduced *in vacuo*. Purification of the residue by chromatography on silica gel (EtOAc:Hex 20:80) afforded the desired product.

(5*R*,11*aS*)-2-Butyl-5-(3-hydroxyphenyl)-3-thioxo-6*H*-1,2,3,5,11,11*a*-hexahydroimidazo[1,5-*b*]-β-carboline-1-on *trans* (8a) Yield 56%, orange glass. m.p. 54-55°C; *v*_{max} (KBr) 3338 (m), 1742 (m), 1599(w), 1458 cm⁻¹ (s); ¹H NMR (500 MHz, DMSO-*d*₆) δ 11.05 (1H, s, NH), 9.51 (1H, s, OH), 7.56 (1H, d, *J* 8.0, H10), 7.31 (1H, d, *J* 8.0, H7), 7.17 (1H, dd, *J* 8.0, 8.0, H5''), 7.13-7.10 (1H, m, H8), 7.06-7.03 (1H, m, H9), 6.92-6.90 (1H, m, H6''), 6.88-6.87 (1H, m, H2''), 6.82 (1H, s, H5), 6.74 (1H, ddd, *J* 8.0, 2.5, 0.5, H4''), 4.79 (1H, dd, *J* 10.5, 6.0, H11*a*), 3.77-3.71 (2H, m, H1'), 3.47 (1H, dd, *J* 15.0, 6.0, H11_{syn}), 2.94-2.91 (1H, m, H11_{anti}), 1.61-1.55 (2H, m, H2'), 1.32-1.25 (2H, m, H3'), 0.89 (3H, t, *J* 7.5, H4'); ¹³C NMR (75 MHz, CDCl₃) δ 180.5 (C=S), 173.5 (C=O), 156.9 (C), 139.7 (C), 136.7 (C), 131.0 (C), 130.2 (CH), 125.9 (C), 122.8 (CH), 120.2 (CH), 120.1 (CH), 118.3 (CH), 116.2 (CH), 115.5 (CH), 111.3 (CH), 107.0 (C), 55.8 (CH), 55.1 (CH), 41.4 (CH₂), 29.8 (CH₂), 23.8 (CH₂), 20.0 (CH₂), 13.7 (CH₃); LRMS (ES⁺) *m/z* 406 [M+H]⁺; [α]_D²⁰ = -216 (*c* = 0.1, DCM).

(5*S*,11*aR*)-2-Butyl-5-(3-hydroxyphenyl)-3-thioxo-6*H*-1,2,3,5,11,11*a*-hexahydroimidazo[1,5-*b*]-β-carboline-1-on *trans* (8b) Yield 50%, yellow solid. m.p. 57-59°C; *v*_{max} (KBr) 3398 (m), 1726 (m), 1599(w), 1459 cm⁻¹ (s); ¹H NMR (500 MHz, DMSO-*d*₆) δ 11.04 (1H, s, NH), 9.51 (1H, s, OH), 7.56 (1H, d, *J* 8.0, H10), 7.31 (1H, d, *J* 8.0, H7), 7.17 (1H, dd, *J* 8.0, 8.0, H5''), 7.13-7.10 (1H, m, H8), 7.06-7.03 (1H, m, H9), 6.92-6.90 (1H, m, H6''), 6.87-6.86 (1H, m, H2''), 6.81 (1H, s, H5), 6.74-6.72 (1H, m, H4''), 4.79 (1H, dd, *J* 10.5, 6.0, H11*a*), 3.79-3.69 (2H, m, H1'), 3.47 (1H, dd, *J* 15.0,

6.0, H11_{syn}), 2.94-2.89 (1H, m, H11_{anti}), 1.62-1.54 (2H, m, H2'), 1.32-1.25 (2H, m, H3'), 0.89 (3H, t, *J* 7.5, H4'); ¹³C NMR (125 MHz, DMSO-d6) δ 180.6 (C=S), 173.4 (C=O), 156.1 (C), 139.8 (C), 136.7 (C), 130.6 (C), 130.4 (CH), 125.9 (C), 123.0 (CH), 121.0 (CH), 120.3 (CH), 118.5 (CH), 116.2 (CH), 115.5 (CH), 111.3 (CH), 107.5 (C), 55.7 (CH), 55.0 (CH), 41.5 (CH₂), 29.8 (CH₂), 23.8 (CH₂), 20.1 (CH₂), 13.7 (CH₃); LRMS (ES⁺) *m/z* 406 [M+H]⁺; [α]_D²⁰ = +216 (*c* = 0.1, DCM).

(5*R*,11*aS*)-2-Butyl-5-(3-methoxyphenyl)-6*H*-1,2,3,5,11,11*a*-hexahydro-imidazo[1,5-*b*]-β-carboline-1,3-dione (9i)

Diethyl azidocarboxylate (60.16 μL, 0.38 mmol) was added dropwise to a solution of triphenylphosphine (101 mg, 0.38 mmol) in anhydrous THF (10 mL) and stirred for 30 minutes. A solution of **7a** (100 mg, 0.25 mmol) and anhydrous MeOH (104.1 μL, 2.5 mmol) in anhydrous THF (10 mL) was added, and the mixture stirred for 48 hrs. The solvent was reduced, and the residue partitioned between EtOAc (20 mL) and H₂O (20 mL). The aqueous layer was extracted with EtOAc (2 x 10 mL) and the combined organics were dried over Na₂SO₄ and reduced. Purification of the residue by chromatography on silica gel (EtOAc:Hex 1:9) afforded **9i** as a white solid (85 mg, 0.21 mmol, 85 %). m.p. 115.1-115.5 °C; *v*_{max} (KBr) 2960, 1713, 1490, 1222 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.49 (1H, d, *J* 7.8, H10), 7.25-7.07 (4H, 4H, H8, 9, 5'', 7), 6.87-6.78 (3H, m, H4'', 2'', 6''), 6.19 (1H, s, H5), 4.25 (1H, dd, *J* 11.0, 5.58, H11*a*), 3.69 (3H, s, H8''), 3.52-3.38 (3H, m, H1', H11_{syn}), 2.81 (1H, dd, *J* 15.4, 11.0, H11_{anti}), 1.56-1.49 (2H, m, H2'), 1.29-1.21 (2H, m, H3'), 0.88 (3H, t, *J* 7.6, H4'); ¹³C NMR (100 MHz, CDCl₃) δ 172.8 (C=O), 160.2 (C), 155.0 (C=O), 140.7 (C), 136.6 (C), 130.2 (CH), 130.4 (C), 126.2 (C), 122.9 (CH), 120.3 (CH), 120.2 (CH), 118.4 (CH), 114.1 (CH), 114.0 (CH), 111.2 (CH), 108.1 (C), 55.3 (CH₃), 53.4 (CH), 51.9 (CH), 38.6 (CH₂), 30.2 (CH₂), 23.5 (CH₂), 20.0 (CH₂), 13.6 (CH₃); LRMS (ES⁺) *m/z*: 404.31 [M+H]⁺; HRMS (ES⁺) [M+H]⁺ *m/z* expected for C₂₄H₂₆N₃O₃ 404.1896 obtained 404.1906; [α]_D²⁰ = -207.5 (*c* = 0.00342, DCM).

Diamino PEG Linker (16)

To a solution of **18** (1.17 g, 3.07 mmol) in anhydrous DCM (15 mL) at 0 °C was added DIPEA (1.10 mL, 6.13 mmol) and methane sulfonyl chloride (0.31 mL, 3.98 mmol). After stirring at 0 °C for 20 minutes, the reaction mixture was allowed to

warm to room temperature and stirred overnight. The mixture was washed with H₂O (30 mL), sat NaHCO₃ (30 mL), dried (Na₂SO₄) and reduced *in vacuo*. Purification of the residue by chromatography on silica gel (DCM:MeOH 97:3) afforded the mesylate as a yellow oil (1.18g, 2.30 mmol, 75%). ν_{\max} (Nujol) 2873, 1700, 1517, 1391, 1170 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 4.18 (2H, t, *J* 4.6, CH₂-OH), 3.76-3.59 (20H, m, CH₂-O), 3.48-3.33 range (2H, m, CH₂-NH), 2.93 (3H, s, CH₃-S), 1.45 (9H, s, (CH₃)C); ¹³C NMR (75 MHz CDCl₃), δ 157.5 (C=O), 70.6 (C), 71.5-62.2 (CH₂ x10), 45.3 (CH₂), 33.9 (CH₃), 28.7 (CH₃); LRMS (ES⁺) *m/z*: 460.35 [M+H]⁺.

The above mesylate (1.12 g, 2.18 mmol) and sodium azide (0.24 g, 3.27 mmol) were dissolved in anhydrous DMF (20 mL) and heated at 110°C for 4 h. Upon cooling the solvent was reduced *in vacuo*. Purification of the residue by chromatography on silica gel (DCM: MeOH 97: 3) afforded the azide as a yellow oil (0.76 g, 1.94 mmol, 89%). ν_{\max} (Nujol) 2922, 2113, 1720, 1301, 1151 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 3.79-3.61 (20H, m, CH₂-O), 3.42 (2H, t, *J* 7.3, CH₂-N₃), 3.45-3.33 (2H, m, CH₂-NH), 1.45 (9H, s, (CH₃)C); ¹³C NMR (75 MHz CDCl₃) δ 157.5 (C=O), 71.4-70.0 (CH₂ x10), 70.5 (C), 50.7 (CH₂), 40.4 (CH₂), 28.4 (CH₃); LRMS (ES⁺) *m/z*: 429.32 [M+Na]⁺.

The azide (0.70 g 1.7 mmol) was dissolved in anhydrous THF (5 mL) and cooled to 0°C, PS-PPh₃ (2g, 1.83 mmol/g) was added and mixture was allowed to warm to room temperature overnight. H₂O (0.5 mL) was added into mixture stirred for 4 hours. The reaction was filtered and the solvent was reduced *in vacuo* to afford the desired product without the need for purification (0.64 g, 1.6 mmol, 99%). ¹H NMR (300 MHz, CDCl₃) δ 3.72-3.50 (20H, m, (CH₂)x10), 3.43 (2H, t, *J* 8.0, CH₂-OH), 2.82 (2H, t, *J* 7.5, CH₂-NH), 1.45 (9H, s, (CH₃)C) ¹³C NMR (75 MHz, CDCl₃) δ 157.5 (C=O), 71.4-70.0 (CH₂ x10), 70.5 (C) 41.78 (CH₂), 40.33 (CH₂), 28.41 (CH₃); LRMS (ES⁺) *m/z*: 381.29 [M+H]⁺.

➤ Unknown enantiomers synthesised using well established literature route.

(5*R*,11*aR*)-2-Butyl-5-(4-hydroxyphenyl)-6*H*-1,2,3,5,11,11*a*-hexahydro-imidazo[1,5-*b*]- β -carboline-1,3-dione (6c) Yield 74%, white solid. m.p. 272-273°C; ν_{\max} (KBr) 3401 (m), 2960 (w), 1765 (m), 1699 (s), 1454 cm⁻¹ (m); ¹H NMR (300 MHz, DMSO-d₆) δ 10.67 (1H, s, NH), 9.33 (1H, s, OH), 7.53 (1H, d, *J* 7.1, H10), 7.22 (1H, d, *J* 7.3, H7), 7.11-6.96 (4H, m, H8, H9, H2''), 6.67-6.64 (2H, AA'BB', H3''), 5.76 (1H, s, H5), 4.52 (1H, dd, *J* 11.4, 4.3, H11*a*), 3.33-3.27 (3H, m, H11_{syn} &

H1'), 3.00-2.91 (1H, m, H11^{anti}), 1.52-1.42 (2H, m, H2'), 1.31-1.18 (2H, m, H3'), 0.86 (3H, t, *J* 7.2, H4'); ¹³C NMR (75 MHz, DMSO-d₆) δ 171.7 (C=O), 156.7 (C), 154.0 (C=O), 136.6 (C), 135.2 (C), 130.9 (C), 128.6 (CH), 125.9 (C), 121.2 (CH), 118.7 (CH), 118.1 (CH), 114.9 (CH), 111.2 (CH), 104.8 (C), 57.6 (CH), 55.3 (CH), 37.3 (CH₂), 29.6 (CH₂), 21.8 (CH₂), 19.3 (CH₂), 13.5 (CH₃); LRMS (ES⁺) *m/z* 390 [M+H]⁺; [α]_D²⁰ = +54 (*c* = 0.1, MeOH).

(5S,11aS)-2-Butyl-5-(4-hydroxyphenyl)-6H-1,2,3,5,11,11a-hexahydro-imidazo[1,5-*b*]-β-carboline-1,3-dione (6d) Yield 64%, white solid. m.p. 274-275°C; *v*_{max} (KBr) 3324 (m), 2957 (w), 1760 (m), 1694 (s), 1458 cm⁻¹ (m); ¹H NMR (500 MHz, DMSO-d₆) δ 10.68 (1H, s, NH), 9.34 (1H, s, OH), 7.53 (1H, d, *J* 7.5, H10), 7.23 (1H, d, *J* 8.0, H7), 7.11-7.09 (2H, AA'BB', H2''), 7.06-7.03 (1H, m, H8), 7.01-6.98 (1H, m, H9), 6.68-6.65 (2H, AA'BB', H3''), 5.77 (1H, s, H5), 4.53 (1H, dd, *J* 11.5, 4.5, H11a), 3.38-3.29 (3H, m, H11_{syn} & H1'), 2.99-2.93 (1H, m, H11^{anti}), 1.50-1.44 (2H, m, H2'), 1.29-1.21 (2H, m, H3'), 0.87 (3H, t, *J* 7.0, H4'); ¹³C NMR (125 MHz, DMSO-d₆) δ 171.8 (C=O), 156.8 (C), 154.1 (C=O), 136.7 (C), 135.3 (C), 130.9 (C), 128.7 (CH), 126.0 (C), 121.3 (CH), 118.8 (CH), 118.2 (CH), 115.0 (CH), 111.3 (CH), 104.9 (C), 57.7 (CH), 53.4 (CH), 37.4 (CH₂), 29.7 (CH₂), 21.9 (CH₂), 19.4 (CH₂), 13.6 (CH₃); LRMS (ES⁺) *m/z* 296 (10) [M-C₆H₅O]⁺, 390 (100) [M+H]⁺; [α]_D²⁰ = -54 (*c* = 0.1, MeOH).

(5R,11aS)-2-Butyl-5-(4-hydroxyphenyl)-6H-1,2,3,5,11,11a-hexahydro-imidazo[1,5-*b*]-β-carboline-1,3-dione (7c) Yield 53%, off-white solid. m.p. 145-146°C; *v*_{max} (KBr) 3399 (m), 2958 (w), 1762 (m), 1700 (s), 1457 cm⁻¹ (s); ¹H NMR (500 MHz, DMSO-d₆) δ 10.90 (1H, s, NH), 9.52 (1H, s, OH), 7.54 (1H, d, *J* 8.0, H10), 7.29 (1H, d, *J* 8.0, H7), 7.15-7.13 (2H, AA'BB', H2''), 7.11-7.08 (1H, m, H8), 7.04-7.01 (1H, m, H9), 6.76-6.74 (2H, AA'BB', H3''), 6.12 (1H, s, H5), 4.53 (1H, dd, *J* 11.0, 5.5, H11a), 3.44-3.38 (3H, m, H11_{syn} & H1'), 2.81-2.76 (1H, m, H11^{anti}), 1.53-1.48 (2H, m, H2'), 1.29-1.21 (2H, m, H3), 0.87 (3H, t, *J* 7.3, H4); ¹³C NMR (125 MHz, DMSO-d₆) δ 172.7 (C=O), 157.3 (C), 154.1 (C=O), 136.6 (C), 131.7 (C), 130.5 (C), 129.1 (CH), 125.8 (C), 121.6 (CH), 118.7 (CH), 118.1 (CH), 115.3 (CH), 111.3 (CH), 105.8 (C), 52.6 (CH), 51.0 (CH), 37.6 (CH₂), 29.6 (CH₂), 22.8 (CH₂), 19.4 (CH₂), 13.5 (CH₃); LRMS (ES⁺) *m/z*: 412.11 [M+Na]⁺; HRMS (ES⁺) [M+Na]⁺ *m/z* expected for C₂₃H₂₃N₃O₃Na 412.1637 found 412.1637; [α]_D²⁰ = -207 (*c* = 0.2, DCM).

(5*S*,11*aR*)-2-Butyl-5-(4-hydroxyphenyl)-6*H*-1,2,3,5,11,11*a*-hexahydro-imidazo[1,5-*b*]- β -carboline-1,3-dione (7d) Yield 77%, white solid. m.p. 142-143°C; ν_{\max} (KBr) 3344 (m), 2959 (w), 1761 (m), 1699 (s), 1457 cm^{-1} (s); ^1H NMR (500 MHz, DMSO-*d*₆) δ 10.89 (1H, s, NH), 9.52 (1H, s, OH), 7.54 (1H, d, *J* 8.0, H10), 7.29 (1H, d, *J* 8.0, H7), 7.15-7.13 (2H, AA'BB', H2''), 7.11-7.08 (1H, m, H8), 7.04-7.01 (1H, m, H9), 6.76-6.73 (2H, AA'BB', H3''), 6.13 (1H, s, H5), 4.53 (1H, dd, *J* 11.0, 5.5, H11*a*), 3.44-3.38 (3H, m, H11_{*syn*} & H1'), 2.81-2.76 (1H, m, H11_{*anti*}), 1.54-1.48 (2H, m, H2'), 1.29-1.21 (2H, m, H3'), 0.89 (3H, t, *J* 7.0, H4'); ^{13}C NMR (75 MHz, CDCl₃) δ 173.5 (C=O), 156.5 (C), 155.1 (C=O), 136.7 (C), 131.0 (C), 130.7 (C), 129.6 (CH), 126.0 (C), 122.7 (CH), 120.0 (CH), 118.4 (CH), 116.0 (CH), 111.3 (CH), 107.6 (C), 53.2 (CH), 51.6 (CH), 38.7 (CH₂), 30.2 (CH₂), 23.3 (CH₂), 20.0 (CH₂), 13.6 (CH₃); LRMS (ES⁺) *m/z*: 412.11 [M+Na]⁺; HRMS (ES⁺) [M+Na]⁺ *m/z* expected for C₂₃H₂₃N₃O₃Na 412.1637 found 412.1637; $[\alpha]_{\text{D}}^{20} = +207$ (*c* = 0.2, DCM).

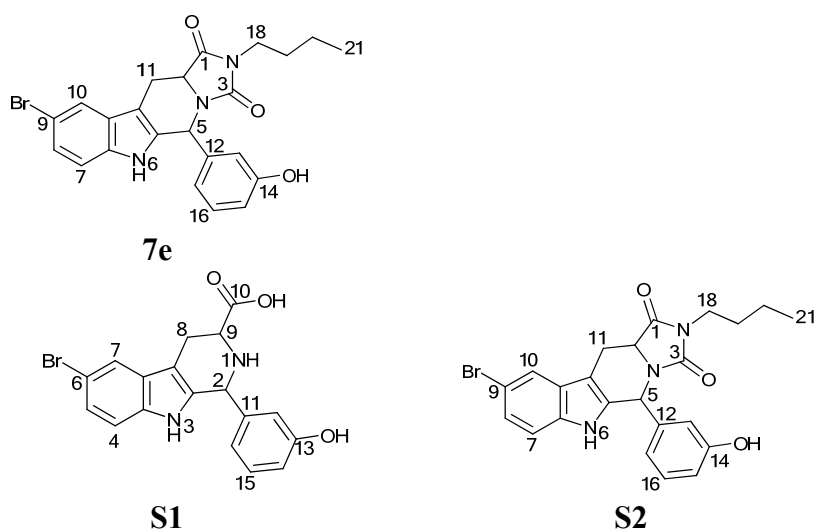
(5*R*,11*aS*)-2-Octyl-5-(3-hydroxyphenyl)-6*H*-1,2,3,5,11,11*a*-hexahydro-imidazo[1,5-*b*]- β -carboline-1,3-dione (7k) Yield 87%, yellow solid. m.p. 93.5-95 °C; ν_{\max} (KBr) 3344 (m), 2927 (m), 1764 (m), 1699 (s), 1457 cm^{-1} (s); ^1H NMR (300 MHz, DMSO-*d*₆) δ 11.0 (1H, s, NH), 9.52 (1H, s, OH), 7.60 (1H, d, *J* 7.7, H10), 7.36 (1H, d, *J* 7.9, H7), 7.24-7.05 (3H, m, H8, H9, H5''), 6.84-6.77 (3H, m, H2', H4', H6'), 6.19 (1H, s, H4), 4.54 (1H, dd, *J* 10.8, 5.7, H11*a*), 3.47-3.40 (3H, m, H11_{*syn*} & H1'), 2.89-2.81 (1H, m, H11_{*anti*}), 1.62-1.53 (2H, m, H2'), 1.32-1.24 (10H, m, H3'-H7'), 0.88 (3H, t, *J* 6.7, H8'); ^{13}C NMR (75 MHz, DMSO-*d*₆) δ 172.5 (C=O), 157.6 (C), 154.2 (C=O), 141.3 (C), 136.6 (C), 131.1 (C), 129.7 (CH), 125.7 (C), 121.6 (CH), 118.8 (CH), 118.4 (CH), 118.1 (CH), 115.0 (CH), 114.7 (CH), 111.3 (CH), 105.9 (C), 52.7 (CH), 51.3 (CH), 37.9 (CH₂), 31.1 (CH₂), 28.5 (CH₂), 28.4 (CH₂), 27.4 (CH₂), 26.0 (CH₂), 22.7 (CH₂), 22.0 (CH₂), 13.9 (CH₃); LRMS (ES⁺) *m/z* 446 [M+H]⁺; $[\alpha]_{\text{D}}^{20} = -244$ (*c* = 0.05, DCM).

(5*R*,11*aS*)-2-(3-Propionic acid ethyl ester)-5-(3-hydroxyphenyl)-6*H*-1,2,3,5,11,11*a*-hexahydro-imidazo[1,5-*b*]- β -carboline-1,3-dione (7l) Yield 82%, yellow solid. m.p. 128-130 °C; ν_{\max} (KBr) 3398 (m), 1707 (s), 1458 cm^{-1} (m); ^1H NMR (500 MHz, DMSO-*d*₆) δ 11.06 (1H, s, NH), 9.59 (1H, s, OH), 7.66 (1H, d, *J*

7.5, H10), 7.41 (1H, d, J 8.0, H7), 7.29-7.26 (1H, m, H5''), 7.22-7.19 (1H, m, H8), 7.15-7.12 (1H, m, H9), 6.69-6.82 (3H, m, H2'', H4'' & H6''), 6.23 (1H, s, H5), 4.59 (1H, dd, J 11, 5.5, H11a), 4.13-4.08 (2H, m, OCH₂CH₃), 3.79-3.76 (2H, m, H1'), 3.5-3.48 (1H, m, H11_{syn}), 2.93 (1H, dd, J 14, 11, H11_{anti}), 2.73-2.70 (2H, m, H2'), 1.21 (3H, t, J 7.0, OCH₂CH₃); ¹³C NMR (75 MHz, CDCl₃) δ 172.7 (C=O), 171.2 (C=O), 156.6 (C), 154.6 (C=O), 140.5 (C), 136.6 (C), 130.3 (CH), 130.2 (C), 126.0 (C), 122.8 (CH), 120.0 (CH), 119.6 (CH), 118.4 (CH), 116.1 (CH), 115.3 (CH), 111.2 (CH), 107.6 (C), 61.0 (CH₂), 53.4 (CH), 52.0 (CH), 34.6 (CH₂), 32.5 (CH₂), 23.2 (CH₂), 14.0 (CH₃); LRMS (ES⁺) m/z : 456.06 [M+Na]⁺; HRMS (ES⁺) [M+Na]⁺ m/z expected for C₂₄H₂₃N₃O₅Na 456.1535 found 456.1535; [α]_D²⁰ = -94 (c = 0.1, DCM).

Synthesis of C9-substituted analogues 7e-7j

Synthesis of 7e via S1 and S2



To a suspension of DL-5-bromotryptophan (**5c**) (1.02 g, 3.60 mmol) in 0.1 M H₂SO₄ (10 ml) was added 3-hydroxybenzaldehyde (0.88 g, 7.20 mmol). The mixture was heated at reflux for 6 hours, cooled to ambient temperature at which point a solid crashed out. The solid was collected by filtration and then washed with water (3 x 25ml) and ether (4 x 50 ml) to give the carboxylic acid **S1** as a white solid (1.08 g, 77%). ¹H NMR (400 MHz, (CD₃)₂SO) δ 7.68 (s, 1H, H7), 7.12-7.33 (m, 3H, H5, H4, H15), 6.89 (d, 1H, H16, ³ J = 7.5) 6.72-6.85 (m, 2H, H14, H12), 5.32 (s, 1H, H2), 3.80 (dd, 1H, H9, ³ J = 4.0, 12.5), 3.11 (dd, 1H, H8_{syn}, ³ J = 13.5, 4.0), 2.86 (dd, 1H, H8_{anti}, ³ J = 13.5, 12.5). ¹³C NMR (100 MHz, (CD₃)₂SO), δ 172.4(C10), 157.4(C13), 140.9(C11), 135.5(C2a), 135.1(C3a), 129.3(C15), 128.1(C7a), 120.1(C7), 119.7(C16), 115.8(C12), 115.5(C4), 115.3(C14), 113.2(C5), 111.1(C6), 107.5(C7b), 57.5(C2), 56.6(C9), 24.4(C8); LRMS m/z , ES- 385.15 (100%) [M-H]⁻. To a suspension of **s1** (1.05 g, 2.70 mmol) in anhydrous DMSO (4 ml) was added *n*-

butylisocyanate (0.268 g, 2.70 mmol) and anhydrous acetone (10 ml). The mixture was heated to reflux for 4 hours before cooling. Solvent was removed *in vacuo* to give an orange that was purified by column chromatography (eluting with EtOAc: petrol ether 30-100%) to give **cis-S2** as an off white solid (0.820 g, 65%). ¹H NMR (400 MHz, (CD₃)₂SO) δ 10.98 (s, 1H, NH), 9.35 (s, 1H, OH), 7.79 (s, 1H, H10), 7.24 (d, 1H, H7, ³J= 8.5), 7.11 (dd, 1H, H16, ³J= 7.50, 8.0), 6.80 (d, 1H, H17, ³J= 7.5), 6.73 (s, 1H, H13), 6.67 (d, 1H, H15, ³J= 8.0), 5.80 (s, 1H, H5), 4.56 (dd, 1H, H11a, ³J= 4.5, 11.5), 3.32-3.48 (m, 3H, H18, H11_{syn}), 2.95 (m, 1H, H11_{anti}), 1.46-1.56 (m, 2H, H19), 1.23-1.34 (m, 2H, H20), 0.90 (t, 3H, H21, ³J= 7.5). ¹³C NMR (100 MHz, (CD₃)₂SO), δ 171.5(C1), 157.2(C14), 154.0(C3), 141.7(C12), 136.4(C6a), 135.3(C5a), 129.2(C16), 127.6(C10a), 123.8(C7), 120.6(C10), 118.0(C15), 114.5(C17), 114.1(C13), 113.2(C8), 111.4(C9), 104.8(C10b), 57.3(C11a), 55.5(C5), 37.4(C18), 29.6(C19), 29.6(C19), 21.7(C11), 19.3(C20), 13.4(C21), *m/z*, ES-466.44 (100%) [M-H]⁻. K₂CO₃ (0.117 g, 0.85 mmol) and **cis-S2** (0.263 g, 0.56 mmol) were then suspended in anhydrous MeCN (3 ml) and refluxed under N₂ for 4 hrs. The crude reaction mixture was filtered and the solvent removed *in vacuo* to give a yellow solid. This solid was purified by column chromatography (eluting with EtOAc: petrol ether 30-100%) to yield the desired product **7e** as a pale yellow solid (0.261 g, 0.56 mmol, 99%). m.p: 236-237 °C (dec). ¹H NMR (400 MHz, (CD₃)₂SO) δ 11.20 (s, 1H, H6), 9.53 (s, 1H, OH), 7.80 (s, 1H, H10), 7.30 (d, 1H, ³J= 8.5, H7), 7.24 (d, 1H, ³J= 8.5, H8), 7.20 (m, 1H, H16), 6.74-6.80 (m, 3H, H15, H13, H17), 6.18 (s, 1H, H5), 4.53 (dd, 1H, ³J= 5.5, 15.5, H11a), 3.40-3.47 (m, 3H, H18, H11_{syn}), 2.80 (dd, 1H, H11_{anti}), 1.54 (m, 2H, H19), 1.29 (m, 2H, H10), 0.90 (t, 3H, ³J= 7.5, H21). ¹³C NMR (100 MHz, (CD₃)₂SO), δ 172.4 (C1), 157.6(C14), 154.2(C3), 141.1(C12), 135.3(C6a), 132.9(C5a), 129.8(C16), 127.5(C10a), 124.1(C15), 120.6(C10), 118.3(C17), 115.1(C13), 114.7(C7), 113.3(C8), 111.4(C9), 105.9(C10b), 52.6(C11a), 51.2(C5), 37.6(C18), 29.6(C19), 22.5(C11), 19.3(C2), 13.4(C21), *m/z*, ES-466.01 (100%) [M-1]⁻.

Conversion of **7e** to **7g-7j**

General procedure for Suzuki coupling: sodium carbonate (0.048 g, 0.45 mmol), the required boronic acid (0.23 mmol) and Pd(PPh₃)₄ (0.012 g, 7 mol%) were added to a sealed tube under argon. Water (1ml) was added, followed by **7e** (0.070 g, 0.15mmol) as a solution in DME (3ml). The reaction mixture was then heated at 70 °C for 24 hrs. The crude reaction mixture was concentrated *in vacuo* and the residue filtered through a silica column (eluting with 40% EtOAc/petrol ester) to yield the desired compound **7g-7j**.

7g (0.039 g, 55%). Mp: 150 °C (dec) ¹H NMR (400 MHz, (CD₃)₂SO) δ 11.00 (s, 1H, H6), 9.52 (s, 1H, OH), 7.91 (s, 1H, H10), 7.76 (d, 2H, ³J= 8.5, H24), 7.53 (d, 2H, ³J= 8.5, H23), 7.40-7.47 (m, 2H, H7, H8), 7.19-7.24 (m, 1H, H16), 6.74-6.85 (m, 3H, H15, H13, H17), 6.19 (s, 1H, H5), 4.54 (dd, 1H, ³J= 5.5, 11.0, H11a), 3.43-3.53 (m, 3H, H18, H11_{syn}), 2.86 (dd, 1H, ³J= 5.5, 15.5, H11_{anti}), 1.54 (m, 2H, H19), 1.29 (m, 2H, H20), 0.90 (t, 3H, ³J= 7.5, H21). ¹³C NMR (100 MHz, (CD₃)₂SO), δ 172.5(C1), 157.6(C14), 154.2(C3), 141.2(C12), 140.5(C9), 136.4(C6a), 132.2(C5a), 131.0(C10a) 129.9(C22), 129.8(C16), 128.6(C24), 128.3(C23), 126.4(C25), 120.7(C15), 118.4(C17), 116.5(C13), 115.1(C10), 114.7(C8), 111.8(C7), 106.6(C10b), 52.6(C11a), 51.3(C5), 37.6(C18), 29.6(C19), 22.7(C11), 19.3(C20), 13.4(C21), *m/z*, ES- 498.13 (100%) [M-1]⁻.

7h (0.029 g, 41%). m.p.: 135 °C (dec). ^1H NMR (400 MHz, $(\text{CD}_3)_2\text{SO}$) δ 11.00 (s, 1H, H6), 9.52 (s, 1H, OH), 7.85 (s, 1H, H10), 7.62 (d, 2H, $^3J = 8.0$, H23), 7.37-7.45 (m, 2H, H7, H8), 7.29 (d, 2H, $^3J = 8.0$, H24), 7.21 (m, 1H, H16), 6.73-6.85 (m, 3H, H13, H15, H17), 6.19 (s, 1H, H5), 4.54 (dd, 1H, $^3J = 5.5$, 11.0, H11a), 3.40-3.53 (m, 3H, H18, H11_{syn}), 2.86 (dd, 1H, $^3J = 5.5$, 15.5, H11_{anti}), 2.38 (s, 3H, H26), 1.54 (m, 2H, H19), 1.29 (m, 2H, H20), 0.90 (t, 3H, $^3J = 7.5$, H21). ^{13}C NMR (100 MHz, $(\text{CD}_3)_2\text{SO}$), δ 172.5(C1), 157.6(C14), 154.2(C3), 141.3(C12), 138.8(C22), 136.1(C6a), 135.3(C25), 131.9(C5a), 131.3(C10a), 129.7(C24), 129.3(C16), 126.4(C23), 126.3(C9), 120.8(C15), 118.4(C17), 116.0(C10), 115.0(C13), 114.7(C8), 111.7(C7), 106.4(C10b), 52.6(C11a), 51.3(C5), 37.6(C18), 29.6(C19), 22.7(C11), 20.6(C26), 19.3(C20), 13.4(C21), *m/z*, ES- 478.18 (100%) [M-1].

7i (21 mg, 84%) m.p.: 215 °C (dec) ^1H NMR (400 MHz, $(\text{CD}_3)_2\text{SO}$) δ 11.00 (s, 1H, H6), 9.52 (s, 1H, OH), 7.81 (s, 1H, H10), 7.65 (d, 2H, $^3J = 9.0$, H24), 7.40 (m, 2H, H7, H8), 7.21 (m, 1H, H16), 7.05 (d, 2H, $^3J = 9.0$, H23), 6.72-6.84 (m, 3H, H15, H13, H17), 6.18 (s, 1H, H5), 4.54 (dd, 1H, $^3J = 5.5$, 11.0, H11a), 3.8 (s, 3H, H26), 3.43-3.53 (m, 3H, H18, H11_{syn}), 2.86 (dd, 1H, $^3J = 15.5$, 5.5, H11_{anti}), 1.54 (m, 2H, H19), 1.29 (m, 2H, H20), 0.90 (t, 3H, $^3J = 7.5$, H21). ^{13}C NMR (100 MHz, $(\text{CD}_3)_2\text{SO}$), δ 172.6(C1), 158.0(C25), 157.6(C14), 154.2(C3), 141.3(C12), 135.8(C6a), 134.1(C5a), 131.8(C16), 131.1(C10a), 129.7(C15), 127.6(C24), 126.3(C9), 120.7(C10), 118.4(C17), 115.8(C13), 115.0(C7), 114.7(C8), 114.2(C23), 111.6(C22), 106.4(C10b), 55.1(C26), 52.7(C11a), 51.3(C5), 37.6(C18), 29.6(C19), 22.7(C11), 19.3(C20), 13.4(C21), *m/z*, ES- 494.18 (100%) [M-1].

7j (44mg, 63%) m.p.: 135 °C (dec) ^1H NMR (400 MHz, $(\text{CD}_3)_2\text{SO}$) δ 11.12 (s, 1H, H6), 9.52 (s, 1H, OH), 8.02 (m, 2H, H10, H23), 7.75 (d, 1H, $^3J = 8.5$, H27), 7.72 (d, 1H, $^3J = 8.5$, H26), 7.50 (d, 1H, $^3J = 9.0$, H7), 7.42 (d, 1H, $^3J = 9.0$, H8), 7.21 (m, 1H, H16), 6.74-6.85 (m, 3H, H15, H13, H17), 6.19 (s, 1H, H5), 4.54 (dd, 1H, $^3J = 5.5$, 11.0, H11a), 3.40-3.53 (m, 3H, H18, H11_{syn}), 2.86 (dd, 1H, $^3J = 5.5$, 15.5, H11_{anti}), 1.54 (m, 2H, H19), 1.29 (m, 2H, H20), 0.90 (t, 3H, $^3J = 7.5$, H21). ^{13}C NMR (100 MHz, $(\text{CD}_3)_2\text{SO}$), δ 172.5(C1), 157.6(C14), 154.2(C3), 142.3(C12), 141.2(C9), 136.6(C6a), 132.3(C5a), 131.5(C10a), 130.8(C26), 129.8(C16), 128.7(C24), 128.4(C25), 128.1(C23), 126.6(C27), 126.4(C22), 120.7(C15), 118.4(C17), 116.9(C13), 115.1(C10), 114.7(C7), 111.9(C8), 106.8(C10b), 52.6(C11a), 51.30(C5), 37.6(C18), 29.6(C19), 22.8(C11), 19.3(C20), 13.4(C21), *m/z*, ES- 532.08 (100%) [M-1].

Synthesis of analogues 9ii, 9iv, 9x-xvii

During the course of this work additional analogues of **9** were prepared. The spectroscopic analysis and biological data relating to these additional are given below (see Table S1). In addition spectroscopic analysis for compounds included in Table 2 of the paper but not discussed specifically in the text of the paper are included here to reduce the size of the experimental section.

Representative procedure for the alkylation of 7a

To a solution of **7a** (50 mg) in anhydrous MeCN (2 mL) was added K_2CO_3 (27 mg) and alkyl bromide (1.05 eq). After stirring at 50 °C for 48 hours, the reaction mixture diluted with water (3 mL). The aqueous layer was extracted with DCM (2 x 2 mL) and the combined extracts were dried and concentrated *in vacuo*. Purification of the residue by filtration through a plug of silica (EtOAc:Hex 1:9) afforded the desired product.

(5R, 11aS)-2-Butyl-5-(3-ethoxyphenyl)-6H-1,2,3,5,11,11a-hexahydro-imidazo[1,5-b]- β -carboline-1,3-dione (9ii) Yield 71%, yellow oil; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.85 (1H, s, NH), 7.56 (1H, d, J 7.5, H10), 7.31-7.15 (4H, m, H8, 9, 5'', 7), 6.91 (1H, d, J 7.7, H6''), 6.88-6.85 (2H, m, H4'', 2''), 6.26 (1H, d, J 1.3, H5), 4.32 (1H, dd, J 11.0, 5.5, H11a), 3.98 (2H, q, J 7.0, H8''), 3.57-3.48 (3H, m, H1', 11_{syn}), 2.88 (1H, ddd, J 15.3, 11.0, 1.8, H11_{anti}), 1.65-1.57 (2H, m, H2'), 1.39-1.30 (5H, m, H9'', H3'), 0.92 (3H, t, J 7.4 Hz, H4'); LRMS (ES^+) m/z : 440.20 (100%) [$\text{M}+\text{Na}$] $^+$.

(5R, 11aS)-2-Butyl-5-(3-butoxyphenyl)-6H-1,2,3,5,11,11a-hexahydro-imidazo[1,5-b]- β -carboline-1,3-dione (9iv) Yield 89%, yellow oil, $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.76 (1H, s, NH), 7.56 (1H, d, J 7.5, H10), 7.31-7.16 (4H, m, H8, 9, 5'', 7), 6.91-6.86 (3H, m, H4'', 2'', 6''), 6.27 (1H, d, J 1.2, H5), 4.33 (1H, dd, J 11.1, 5.4, H11a), 3.88 (2H, t, J 6.5, H8''), 3.57-3.49 (3H, m, H1', 11_{syn}), 2.88 (1H, ddd, J 15.2, 11.1, 1.8, H11_{anti}), 1.81-1.73 (2H, m, H9''), 1.65-1.58 (2H, m, H2'), 1.38-1.28 (2H, m, H3'), 1.26-1.25 (2H, m, H12''), 1.00 (3H, t, J 7.5, H11''), 0.93 (3H, t, J 7.4, H4'); LRMS (ES^+) m/z : 468.18 (100%) [$\text{M}+\text{Na}$] $^+$.

(5R,11aS)-2-Butyl-5-(3-(4-methoxy-benzyloxy)phenyl)-6H-1,2,3,5,11,11a-hexahydro-imidazo[1,5-b]- β -carboline-1,3-dione (9x) Yield 91%, yellow solid, $^1\text{H NMR}$ (300 MHz, CDCl_3) δ 8.00 (1H, s, NH), 7.55 (1H, d, J 7.5, H10), 7.28-7.14 (6H, m, H8, 9, 5'', 7, 10''), 6.94-6.85 (5H, m, H6'', 4'', 2'', 11''), 6.22 (1H, s, H5), 4.88 (2H, s, H8''), 4.23 (1H, dd, J 11.0, 5.5, H11a), 3.80 (3H, s, H14''), 3.57-3.42 (3H, m, H1', 11_{syn}), 2.86 (1H, ddd, J 15.3, 11.0, 1.8, H11_{anti}), 1.66-1.56 (2H, m, H2'), 1.40-1.26 (2H, m, H3'), 0.92 (3H, t, J 7.4, H4'); LRMS (ES^+) m/z : 532.22 (100%) [$\text{M}+\text{Na}$] $^+$.

(5R,11aS)-2-Butyl-5-(3-(3,4-Dichloro-benzyloxy)phenyl)-6H-1,2,3,5,11,11a-hexahydro-imidazo[1,5-b]- β -carboline-1,3-dione (9xi) Yield 87%, yellow solid, $^1\text{H NMR}$ (300 MHz, CDCl_3) δ 7.88 (1H, s, NH), 7.57 (1H, d, J 7.5, H10), 7.48 (1H, d, J 2.0, H10''), 7.39 (1H, d, J 8.2, H12''), 7.30-7.16 (5H, m, H8, 9, 5'', 7, 13''), 6.95-6.89 (3H, m, H6'', 4'', 2''), 6.26 (1H, d, J 1.0, H5), 4.94 (2H, s, H8''), 4.28 (1H, dd, J 11.0, 5.5, H11a), 3.58-3.46 (3H, m, H1', 11_{syn}), 2.88 (1H, ddd, J 15.3, 11.0, 1.8, H11_{anti}), 1.66-1.55 (2H, m, H2'), 1.40-1.24 (2H, m, H3'), 0.93 (3H, t, J 7.4, H4'); LRMS (ES^+) m/z : 570.13 [$\text{M}+\text{Na}$] $^+$.

(5R,11aS)-2-Butyl-5-(3-carbamoylmethoxyphenyl)-6H-1,2,3,5,11,11a-hexahydro-imidazo[1,5-b]- β -carboline-1,3-dione (9xii) Yield 55%, yellow oil, $^1\text{H NMR}$ (300 MHz, CDCl_3) δ 8.01 (1H, s, NH), 7.58 (1H, d, J 7.5, H10), 7.34-7.16 (4H, m, H8, 9, 5'', 7), 7.01 (1H, d, J 7.9, H6''), 6.96-6.95 (1H, m, H2''), 6.86 (1H, dd, J 8.1, 2.5, H4''), 6.57-6.47 (1H, br s, NH), 6.29 (1H, d, J 1.4, H5), 5.72-5.59 (1H, br s, NH), 4.41 (2H, s, H8''), 4.30 (1H, dd, J 11.0, 5.5, H11a), 3.56-3.48 (3H, m, H1', 11_{syn}), 2.89 (1H, ddd, J 15.3, 11.0, 1.8, H11_{anti}), 1.66-1.56 (2H, m, H2'), 1.39-1.25 (2H, m, H3'), 0.92 (3H, t, J 7.4, H4'); LRMS (ES^+) m/z : 469.18 (100%) [$\text{M}+\text{Na}$] $^+$.

(5R,11aS)-2-Butyl-5-(3-cyanomethoxyphenyl)-6H-1,2,3,5,11,11a-hexahydro-imidazo[1,5-b]- β -carboline-1,3-dione (9xiii) Yield 54%, yellow oil, $^1\text{H NMR}$ (300 MHz, CDCl_3) δ 7.90 (1H, s, NH), 7.57 (1H, d, J 7.5, H10), 7.37-7.16 (4H, m, H8, 9, 5'', 7), 7.06-6.95 (3H, m, H6'', 4'', 2''), 6.31 (1H, d, J 1.4, H5), 4.74 (2H, s, H8''), 4.30 (1H, dd, J 11.0, 5.5, H11a), 3.58-3.46 (3H, m, H1', 11_{syn}), 2.88 (1H, ddd, J 15.3, 11.0, 1.8, H11_{anti}), 1.66-1.56 (2H, m, H2'), 1.40-1.25 (2H, m, H3'), 0.93 (3H, t, J 7.4, H4'); LRMS (ES^+) m/z : 451.18 (100%) [$\text{M}+\text{Na}$] $^+$.

(5R,11aS)-2-Butyl-5-(3-but-2-enyloxyphenyl)-6H-1,2,3,5,11,11a-hexahydro-imidazo[1,5-b]- β -carboline-1,3-dione (9xiv) Yield 64%, yellow oil, ^1H NMR (300 MHz, CDCl_3) δ 7.83 (1H, s, NH), 7.56 (1H, d, J 7.5, H10), 7.31-7.14 (4H, m, H8, 9, 5'', 7), 6.92 (1H, d, J 7.7, H6''), 6.90-6.87 (2H, m, H4'', 2''), 6.27 (1H, s, H5), 5.88-5.76 (1H, m, H10''), 5.71-5.61 (1H, m, H9''), 4.40 (2H, d, J 6.0, H8''), 4.31 (1H, dd, J 11.0, 5.5, H11a), 3.56-3.47 (3H, m, H1', 11_{syn}), 2.88 (1H, ddd, J 15.3, 11.0, 1.8 Hz, H11_{anti}), 1.72 (3H, dd, J 6.3, 1.3, H11''), 1.68-1.59 (2H, m, H2'), 1.40-1.27 (2H, m, H3'), 0.92 (3H, t, J 7.4, H4'); LRMS (ES^+) m/z : 466.28 (100%) [$\text{M}+\text{Na}$] $^+$.

(5R,11aS)-2-Butyl-5-(3-but-3-enyloxyphenyl)-6H-1,2,3,5,11,11a-hexahydro-imidazo[1,5-b]- β -carboline-1,3-dione (9xv) Yield 72%, yellow oil, ^1H NMR (300 MHz, CDCl_3) δ 7.90 (1H, s, NH), 7.55 (1H, d, J 7.5, H10), 7.28-7.13 (4H, m, H8, 9, 5'', 7), 6.90-6.79 (3H, m, H6'', 4'', 2''), 6.23 (1H, d, J 1.3, H5), 5.88-5.78 (1H, m, H10''), 5.14-5.04 (2H, m, H11''), 4.30 (1H, dd, J 11.0, 5.5, H11a), 3.93 (2H, pseudo t, J 8.7, H8''), 3.53-3.43 (3H, m, H1', 11_{syn}), 2.88 (1H, ddd, J 15.3, 11.0, 1.8 Hz, H11_{anti}), 2.50-2.44 (2H, m, H9''), 1.65-1.55 (2H, m, H2'), 1.40-1.27 (2H, m, H3'), 0.92 (3H, t, J 7.4, H4'); LRMS (ES^+) m/z : 466.21 (100%) [$\text{M}+\text{Na}$] $^+$.

(5R,11aS)-2-Butyl-5-(3-(3-Methyl-but-2-enyloxy)phenyl)-6H-1,2,3,5,11,11a-hexahydro-imidazo[1,5-b]- β -carboline-1,3-dione (9xvi) Yield 51%, yellow oil, ^1H NMR (300 MHz, CDCl_3) δ 7.82 (1H, s, NH), 7.55 (1H, d, J 7.5, H10), 7.30-7.14 (4H, m, H8, 9, 5'', 7), 6.95 (1H, d, J 7.7, H6''), 6.93-6.84 (2H, m, H4'', 2''), 6.27 (1H, d, J 1.3, H5), 5.44-5.38 (1H, m, H9''), 4.45 (2H, d, J 6.8, H8''), 4.31 (1H, dd, J 11.0, 5.5, H11a), 3.56-3.46 (3H, m, H1', 11_{syn}), 2.88 (1H, ddd, J 15.3, 11.0, 1.8, H11_{anti}), 1.74 (3H, s, H11''), 1.67 (3H, s, H12''), 1.64-1.56 (2H, m, H2'), 1.40-1.25 (2H, m, H3'), 0.92 (3H, t, J 7.4, H4'); LRMS (ES^+) m/z : 480.23 (100%) [$\text{M}+\text{Na}$] $^+$.

(5R,11aS)-2-Butyl-5-(3-cyclopentyloxyphenyl)-6H-1,2,3,5,11,11a-hexahydro-imidazo[1,5-b]- β -carboline-1,3-dione (9xvii) Yield 72%, yellow oil, ^1H NMR (400 MHz, MeOD) δ 7.50 (1H, d, J = 7.7 Hz, H10), 7.25 (1H, d, J = 7.9 Hz, H7), 7.20 (1H, dd, J 7.9, H5'), 7.11-7.07 (1H, m, H9), 7.04-7.00 (1H, m, H8), 6.88-6.85 (1H, m, H6''), 6.82-6.78 (2H, m, H4'', 2''), 6.23 (1H, s, H5), 4.74-4.70 (1H, m, H8''), 4.39 (1H, dd, J 11.1, 5.5, H11a), 3.49 (2H, t, J 7.2, H1'), 3.42 (1H, dd, J 15.1, 5.5, 11_{syn}), 2.79 (1H, ddd, J 15.1, 11.1, 1.7, H11_{anti}), 1.89-1.70 (6H, m, H9'', H2'), 1.60-1.53 (4H, m, H10''), 1.35-1.26 (2H, m, H3'), 0.91 (3H, t, J 7.4, H4'); LRMS (ES^+) m/z : 480.23 (100%) [$\text{M}+\text{Na}$] $^+$.

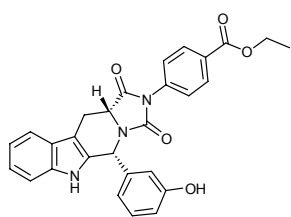
Entry #	Cpd #	C5	C11a	R ¹	R ³	R ⁴	MIC (μM)
1	9i	R	S	OMe	ⁿ Butyl	H	12.5
2	9ii	R	S	Oet	ⁿ Butyl	H	6.25
3	9iii	R	S	O ⁿ Pr	ⁿ Butyl	H	12.5
4	9iv	R	S	O ⁿ Bu	ⁿ Butyl	H	6.25
5	9v	R	S	Oallyl	ⁿ Butyl	H	12.5
6	9vi	R	S	O ⁱ Pr	ⁿ Butyl	H	1.6
7	9vii	R	S	OCH ₂ C(CH ₃)=CH ₂	ⁿ Butyl	H	3.1
8	9viii	R	S	Opropargyl	ⁿ Butyl	H	3.1
9	9ix	R	S	OBn	ⁿ Butyl	H	12.5
10	9x	R	S	OCH ₂ <i>p</i> -C ₆ H ₄ -OMe	ⁿ Butyl	H	100
11	9xi	R	S	OCH ₂ <i>m</i> , <i>p</i> -C ₆ H ₃ -Cl ₂	ⁿ Butyl	H	100
12	9xii	R	S	CH ₂ CONH ₂	ⁿ Butyl	H	25

13	9xiii	<i>R</i>	<i>S</i>	OCH ₂ CN	ⁿ Butyl	H	6.25
14	9xiv	<i>R</i>	<i>S</i>	OCH ₂ CH=CHCH ₃	ⁿ Butyl	H	3.1
15	9xv	<i>R</i>	<i>S</i>	OCH ₂ CH ₂ CH=CH ₂	ⁿ Butyl	H	1.6
16	9xvi	<i>R</i>	<i>S</i>	OCH ₂ CH=C(CH ₃) ₂	ⁿ Butyl	H	3.1
17	9xvii	<i>R</i>	<i>S</i>	O ^o Pentyl	ⁿ Butyl	H	1.6

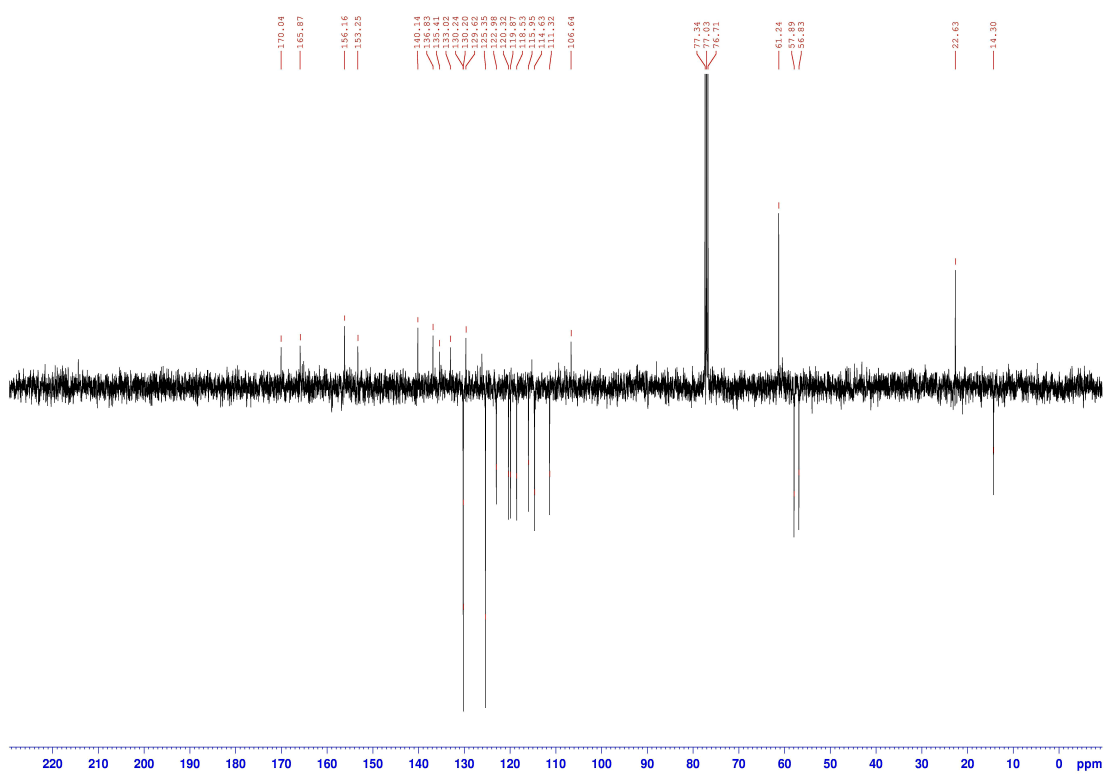
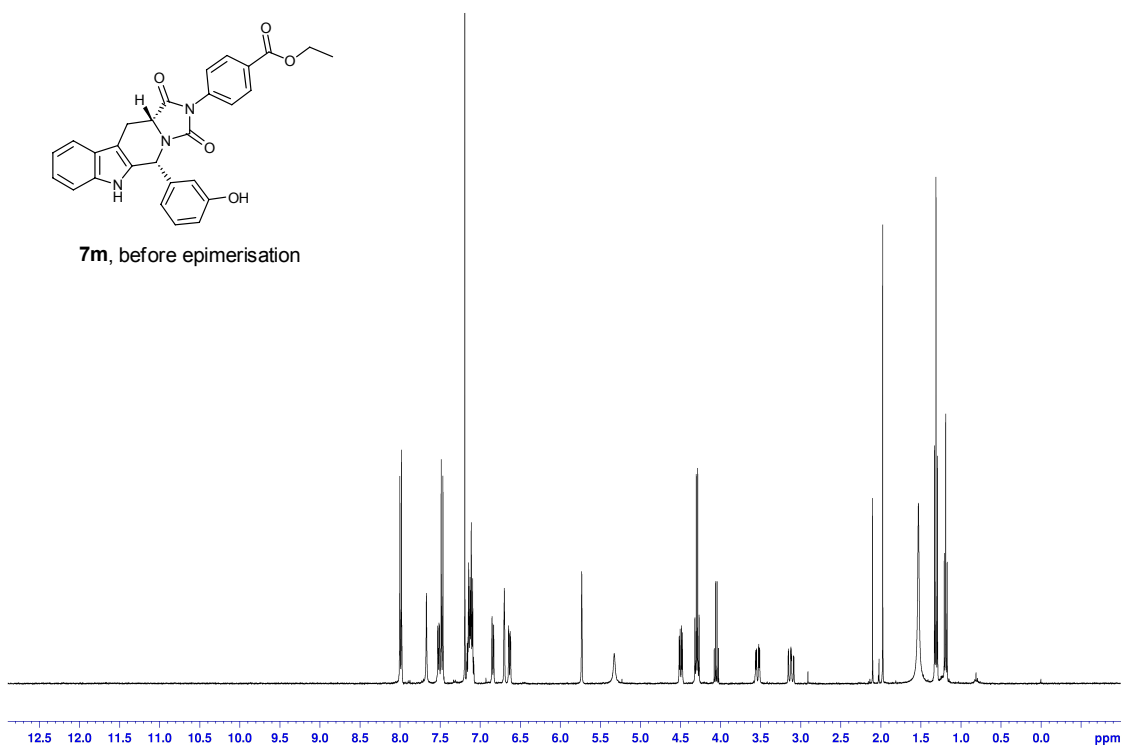
Table S1. Minimum inhibitory concentrations (MIC) in the *T. gondii* cell invasion assay for analogues of **1**.

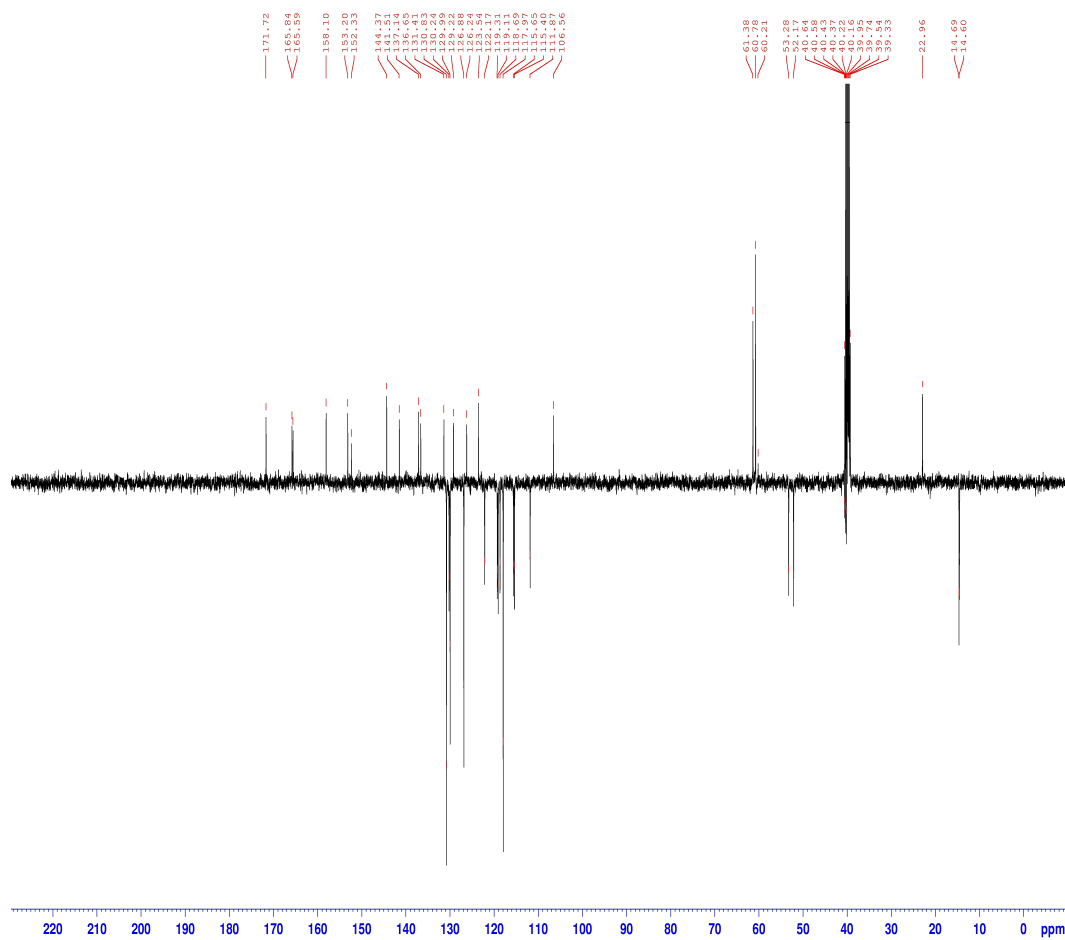
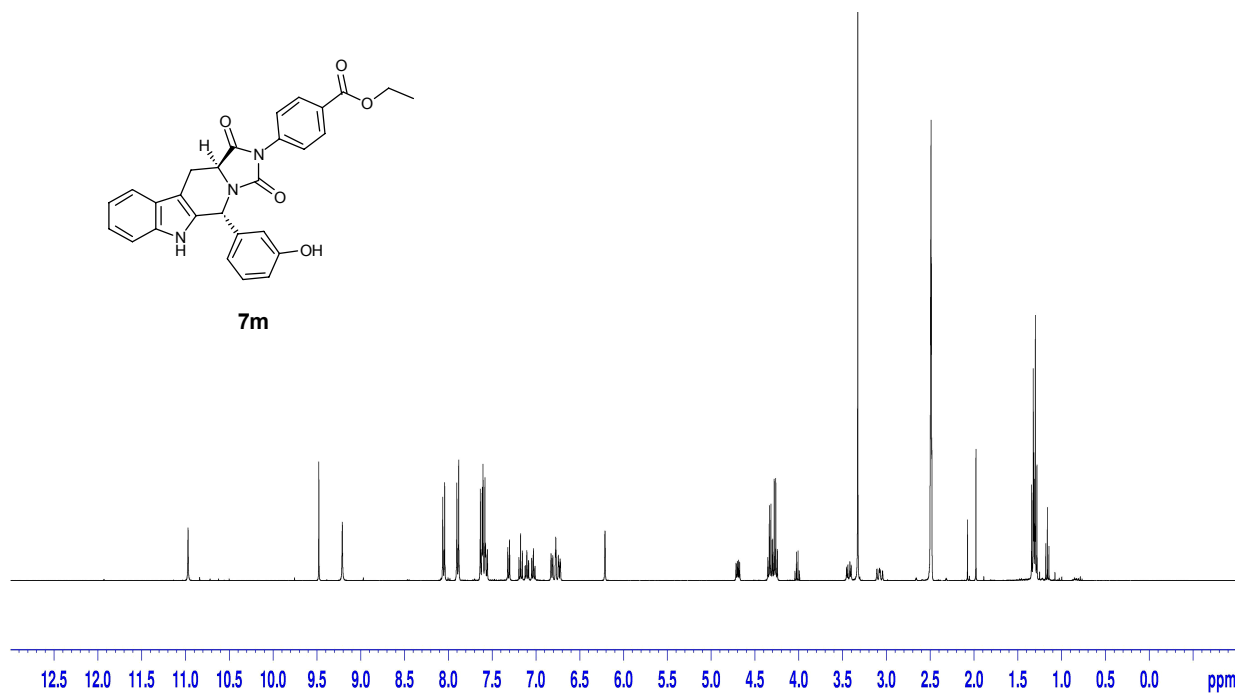
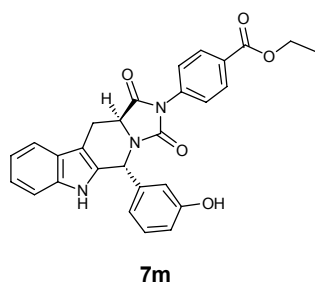
References

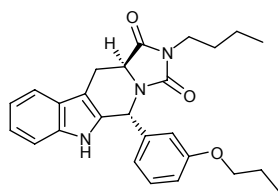
- S1 Baker K.; Sengupta D.; Salazar-Jimenez G.; Cornish V.W. *Anal Biochem.* **2003**, *315(1)*, 124-137



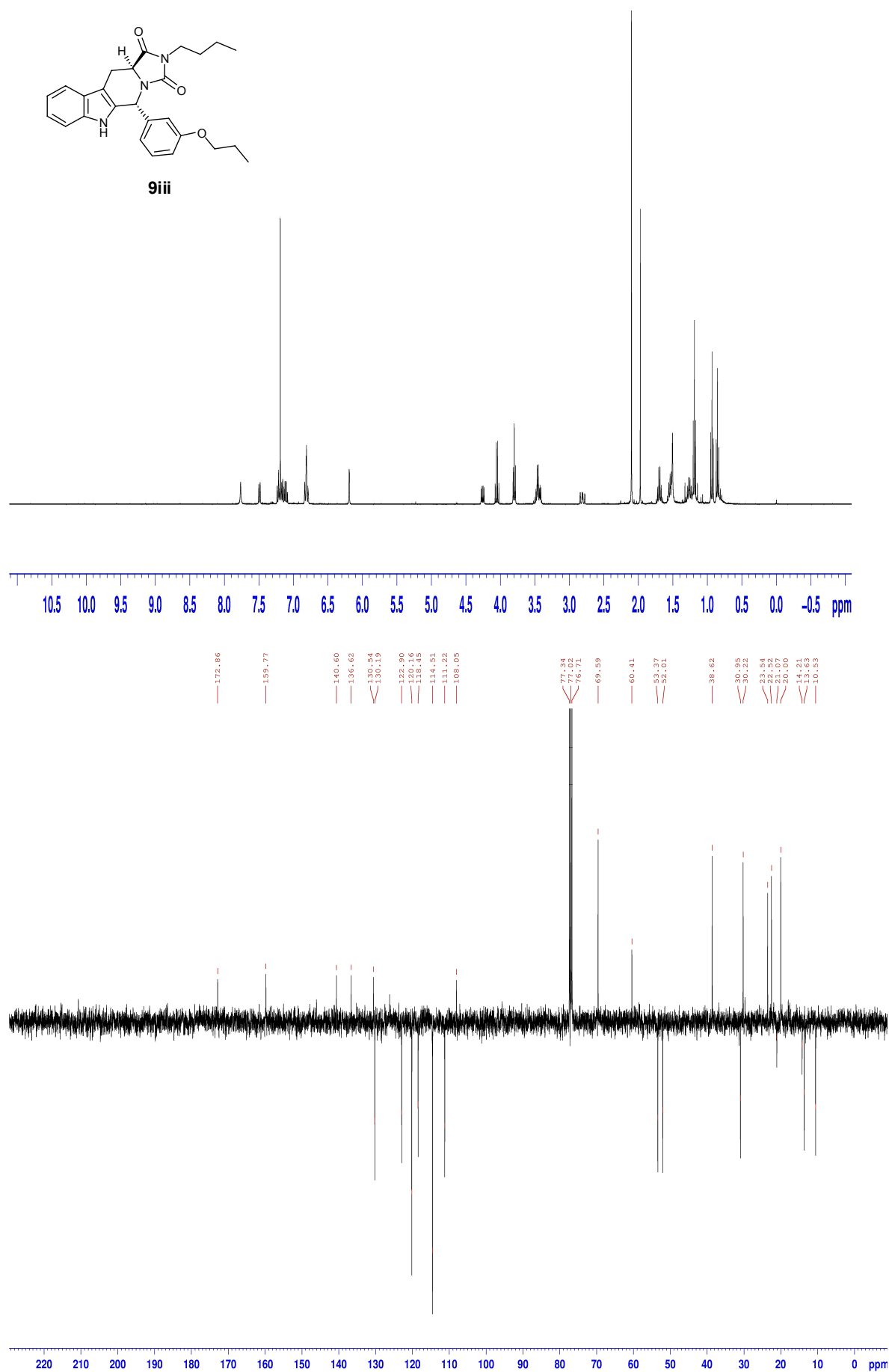
7m, before epimerisation

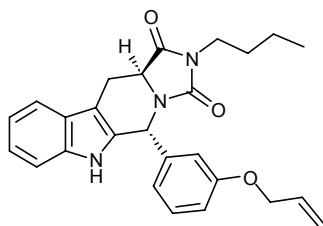




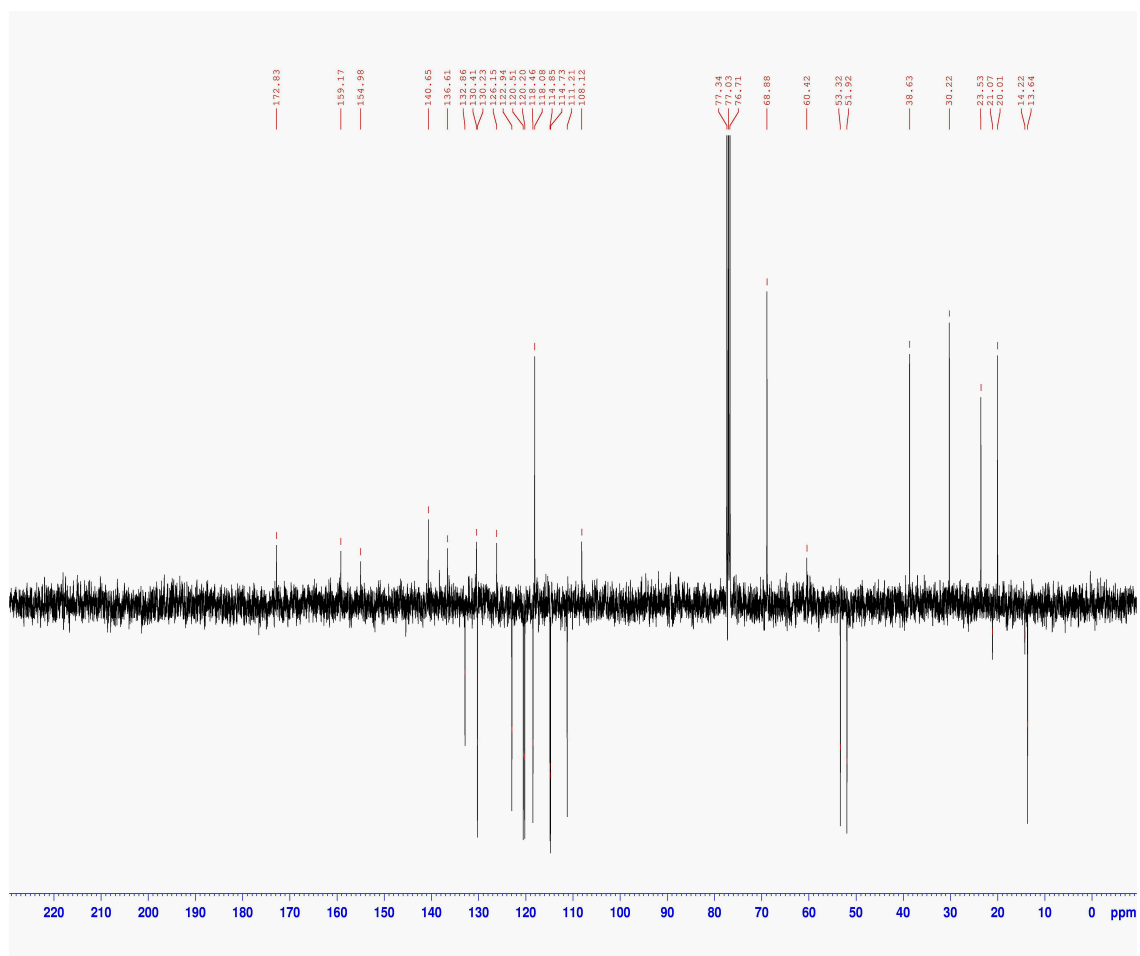
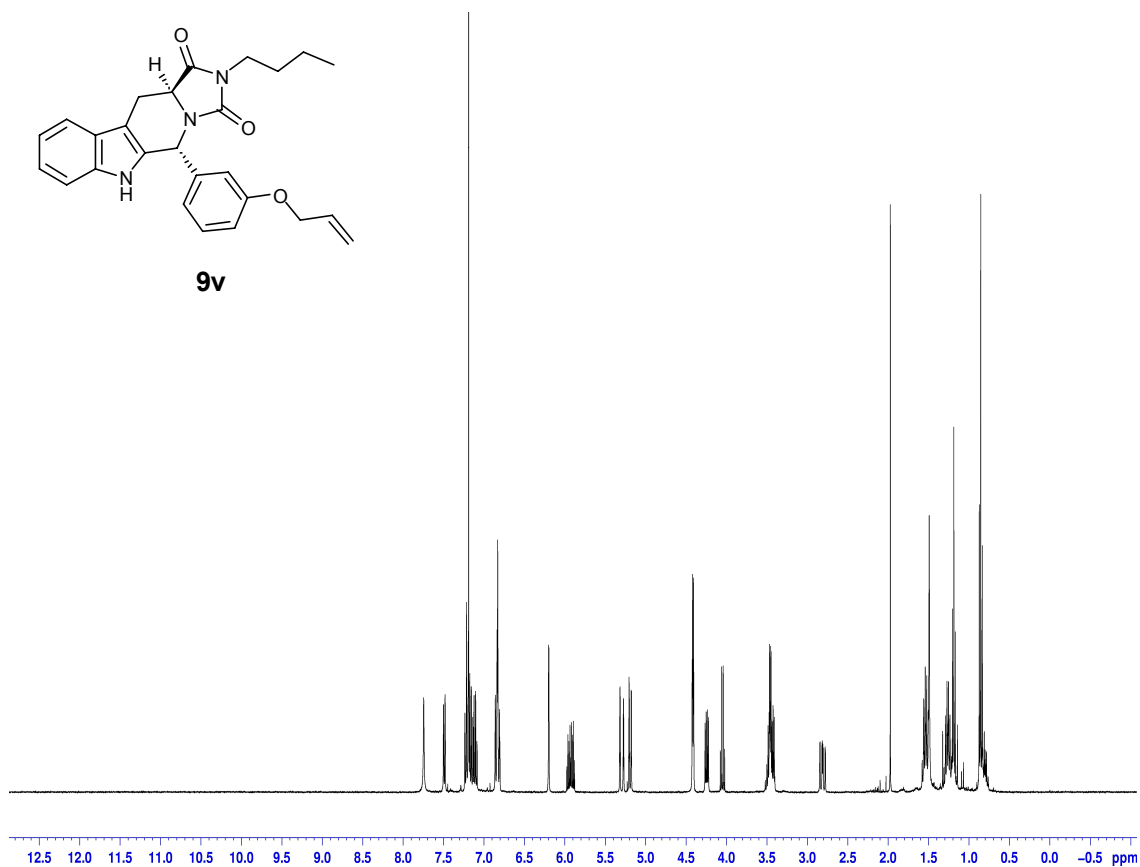


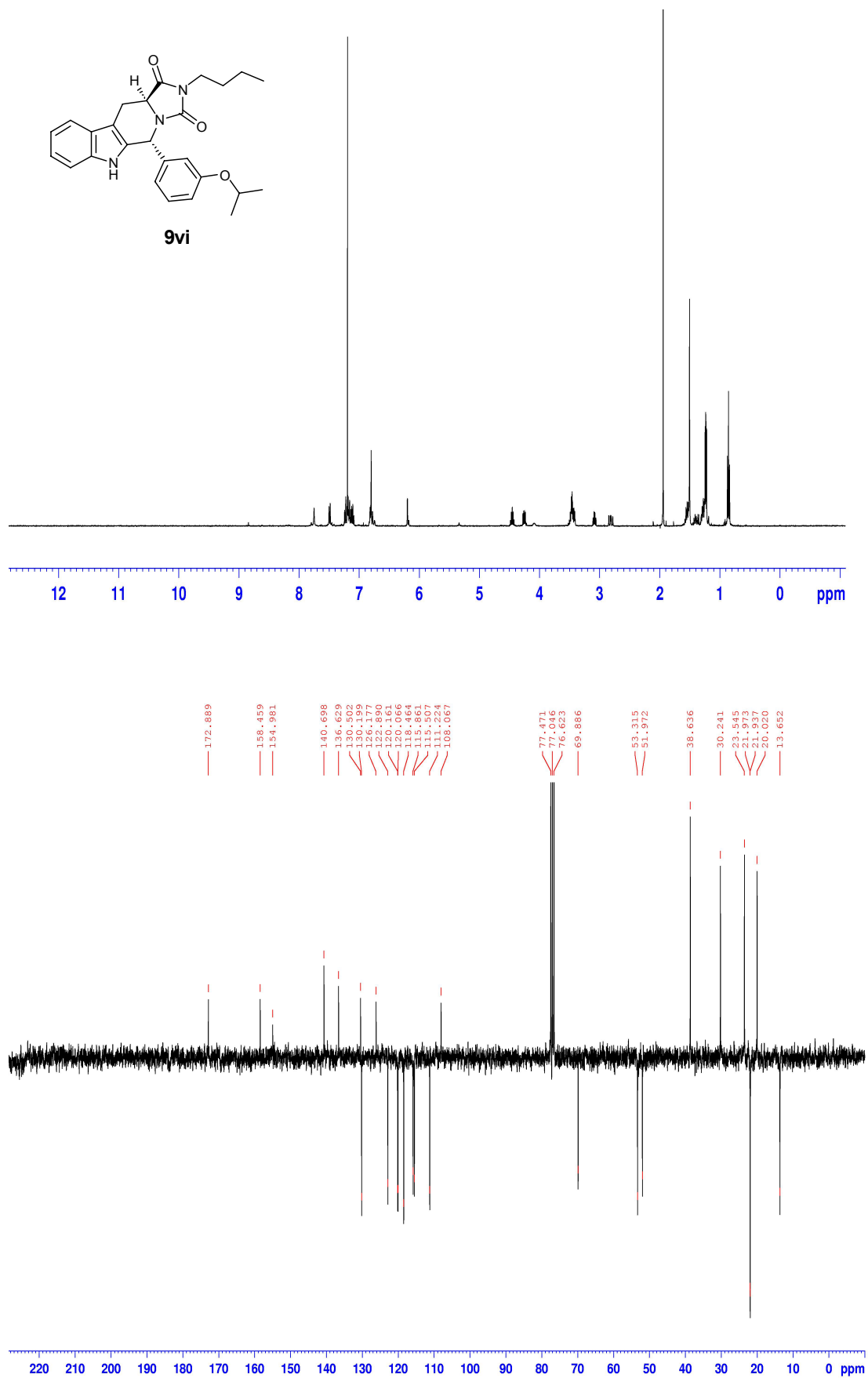
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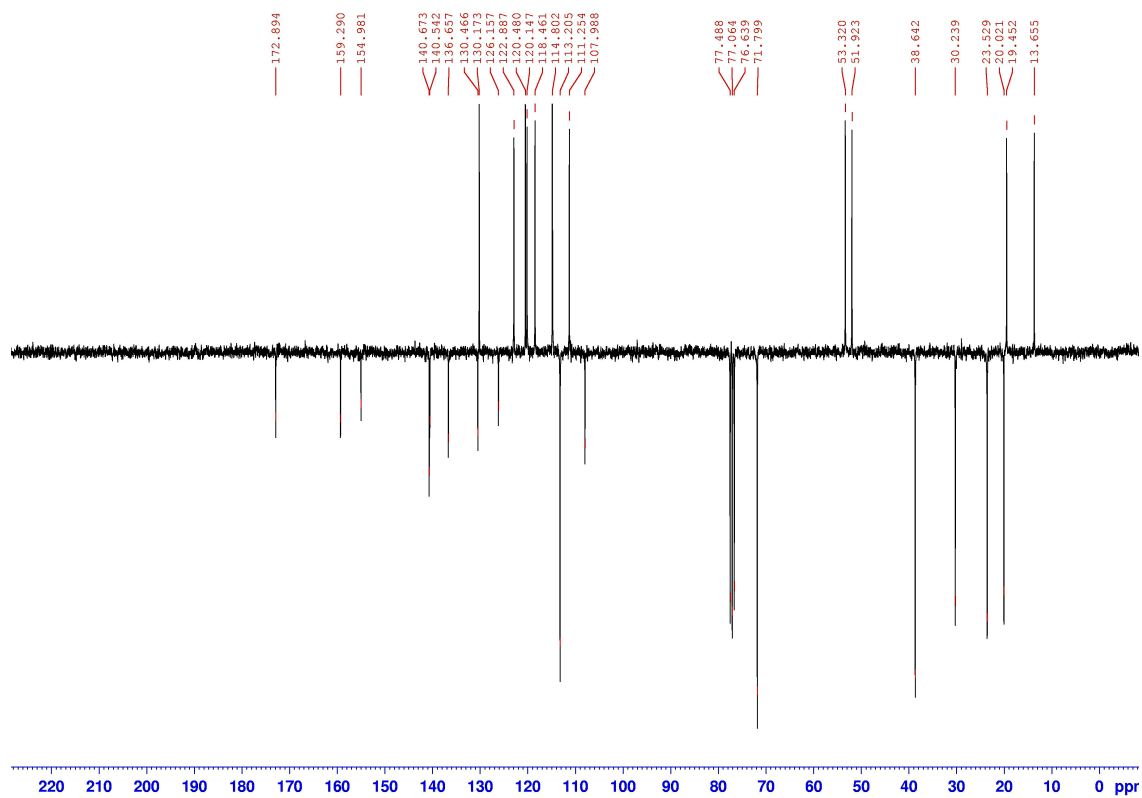
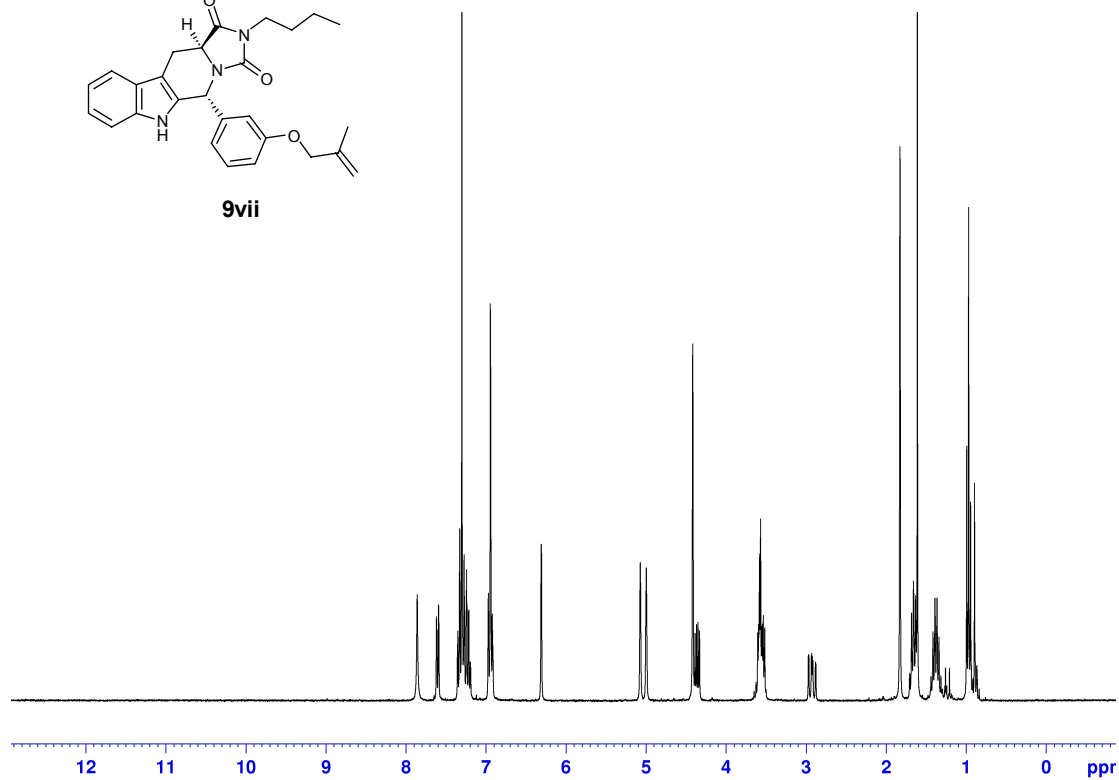
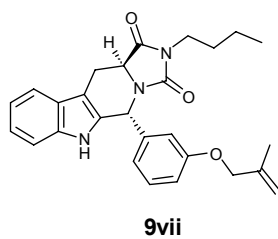


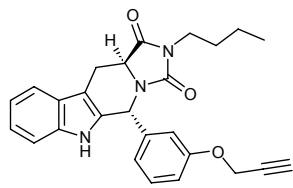


9v

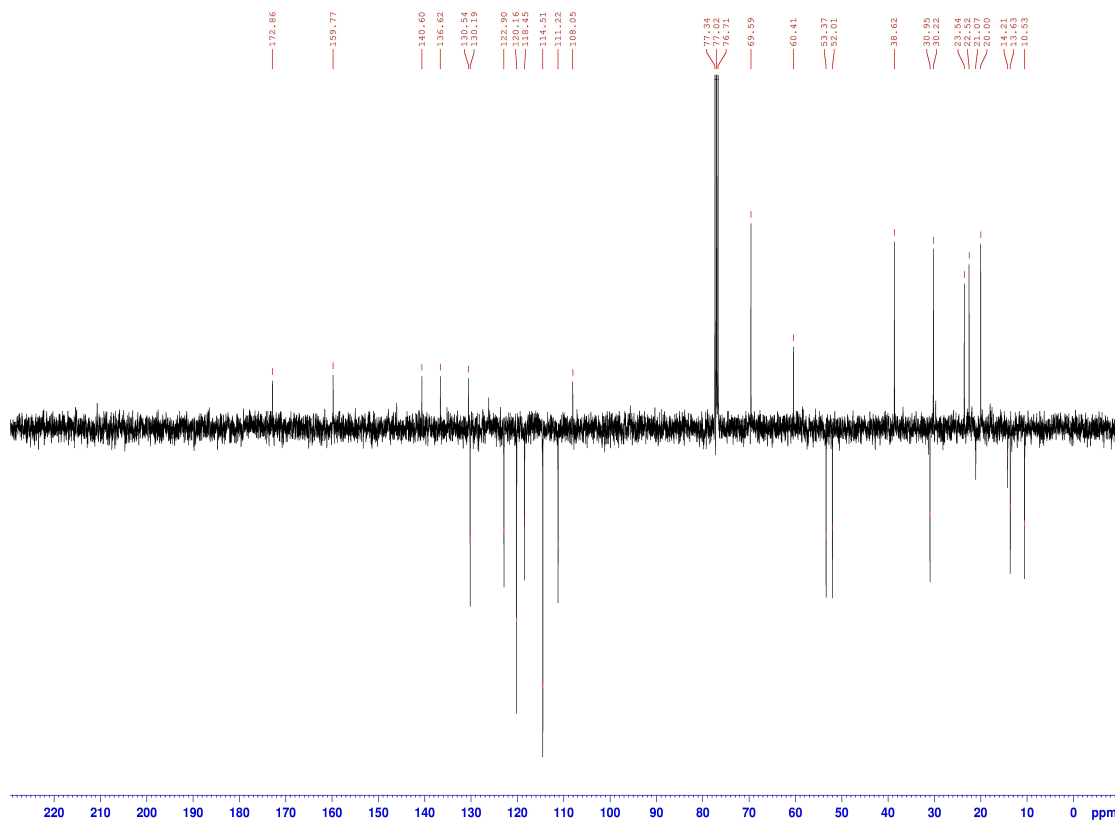
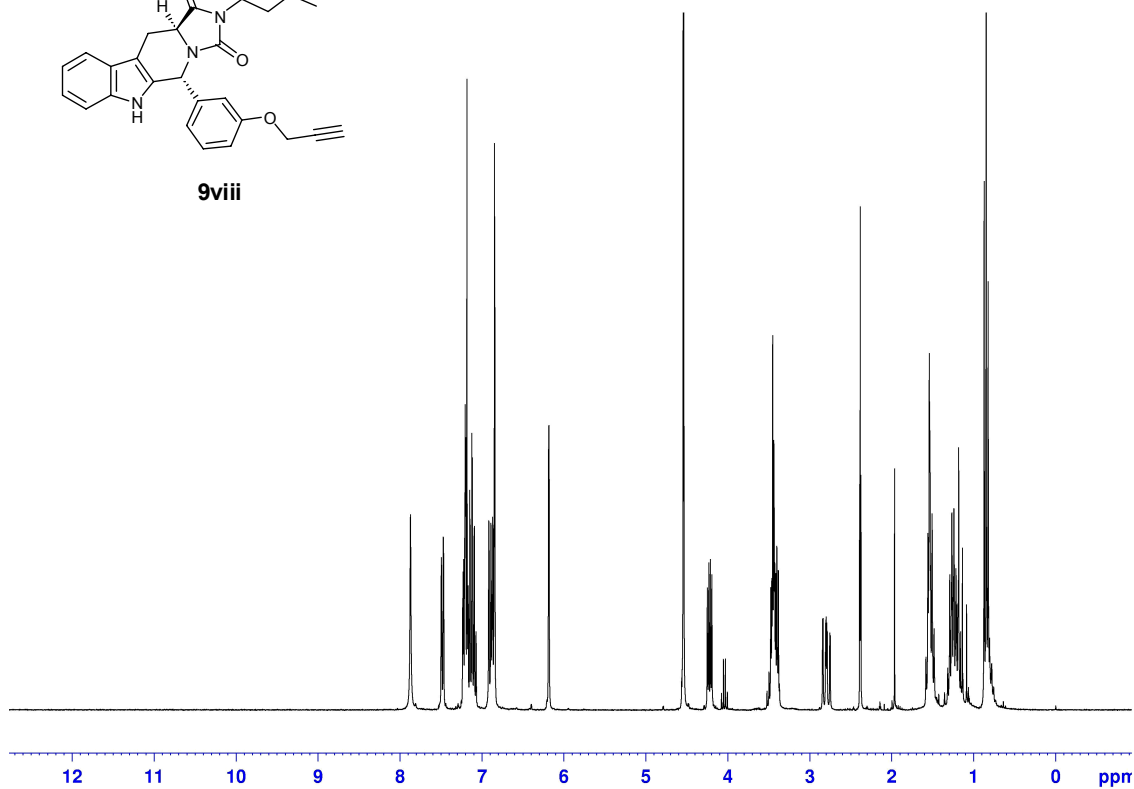


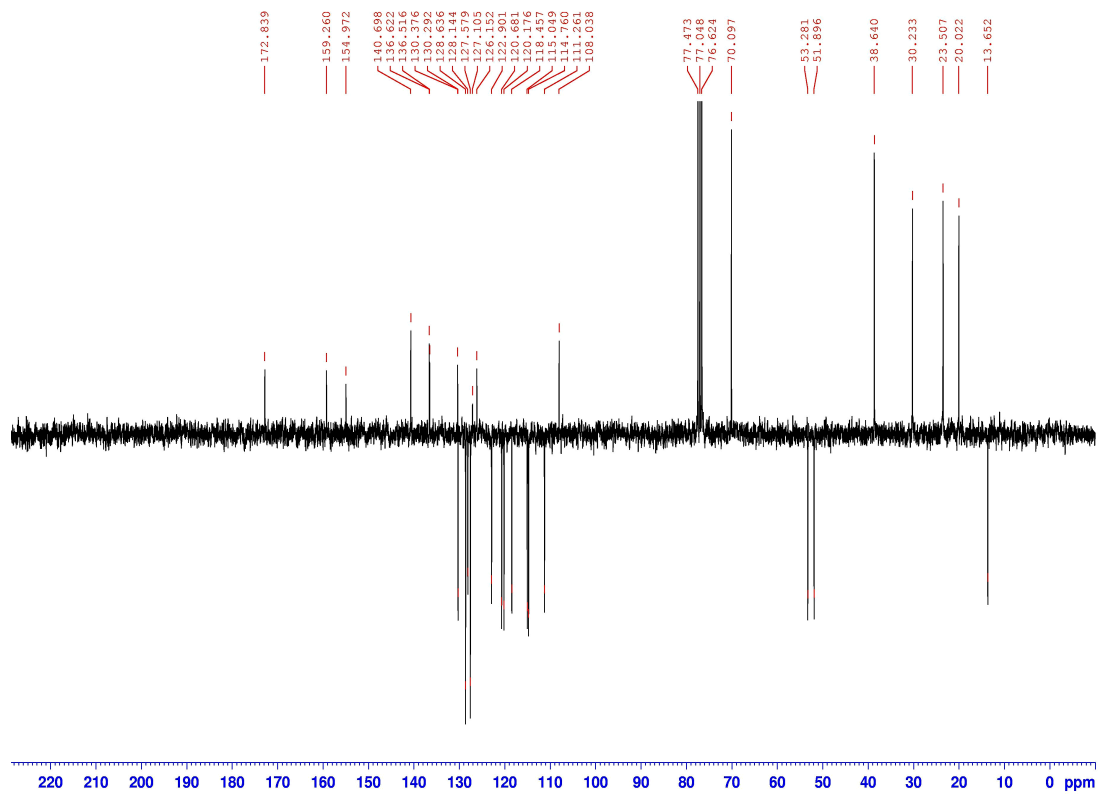
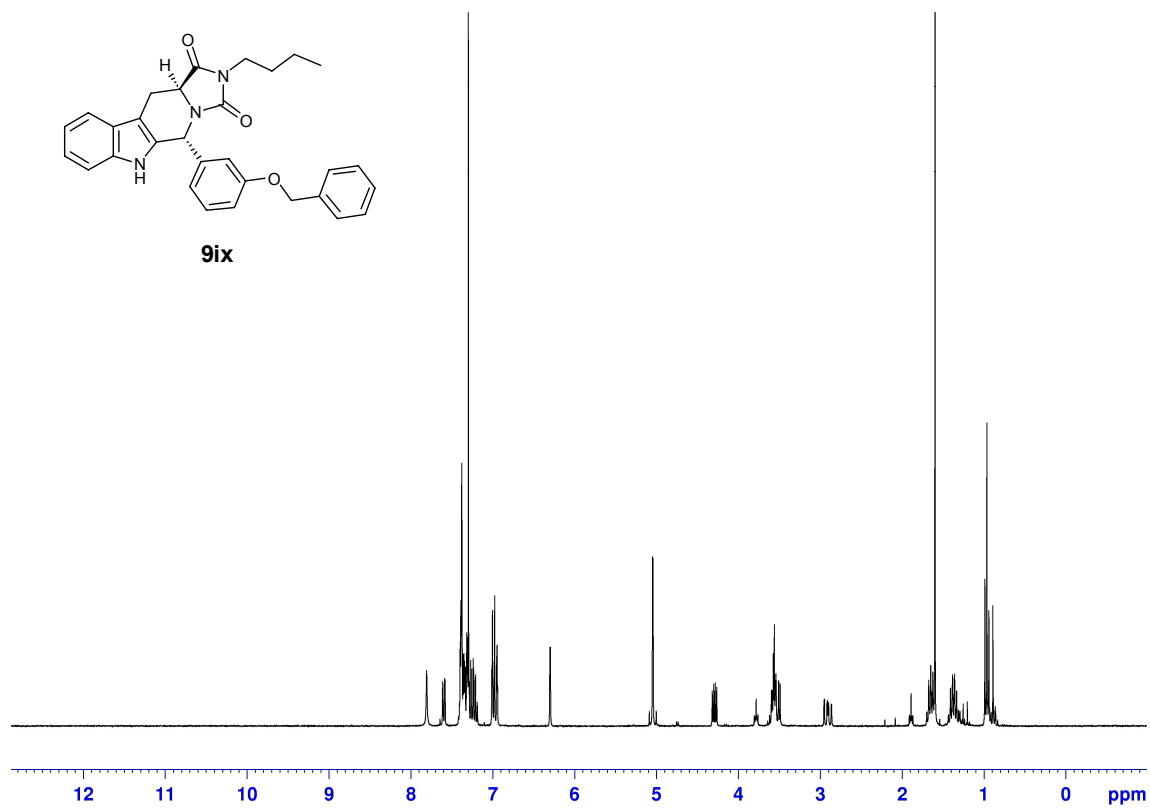


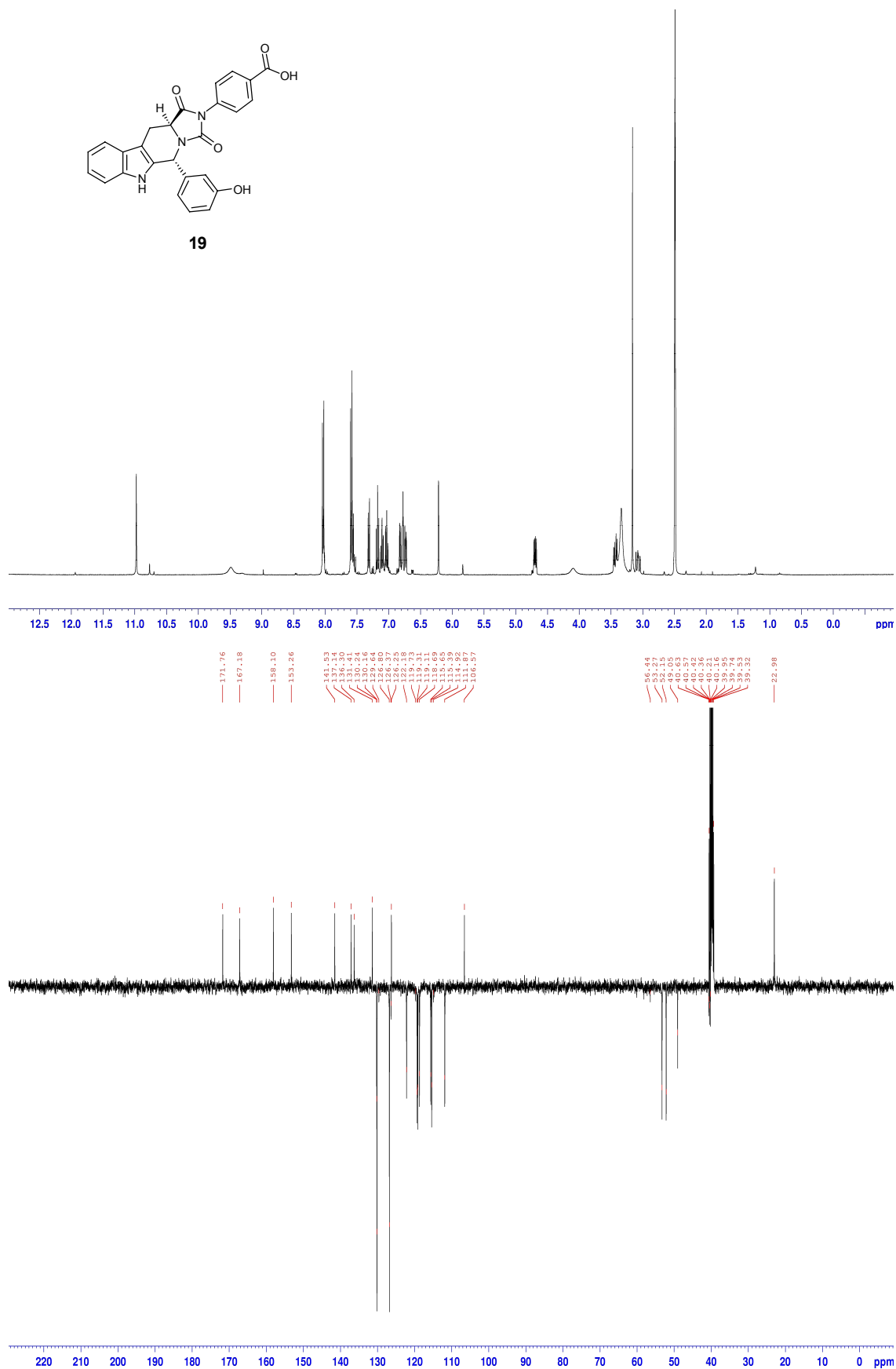
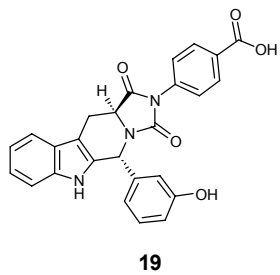


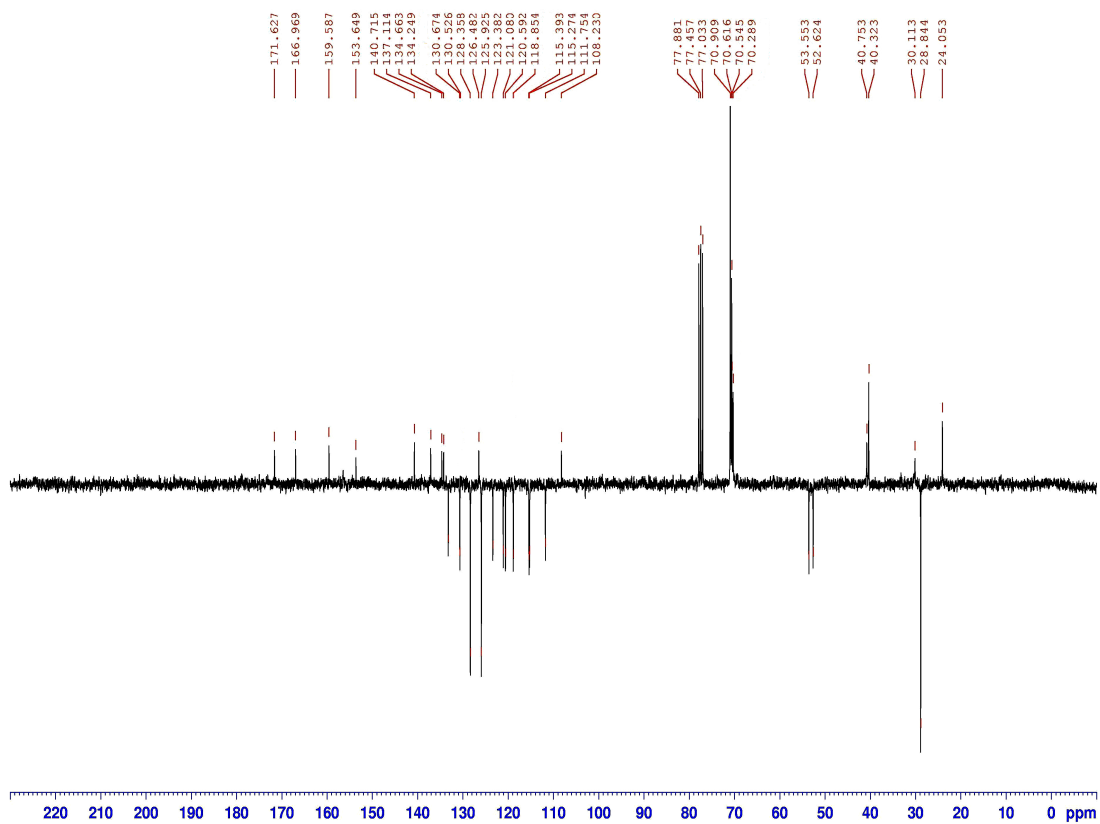
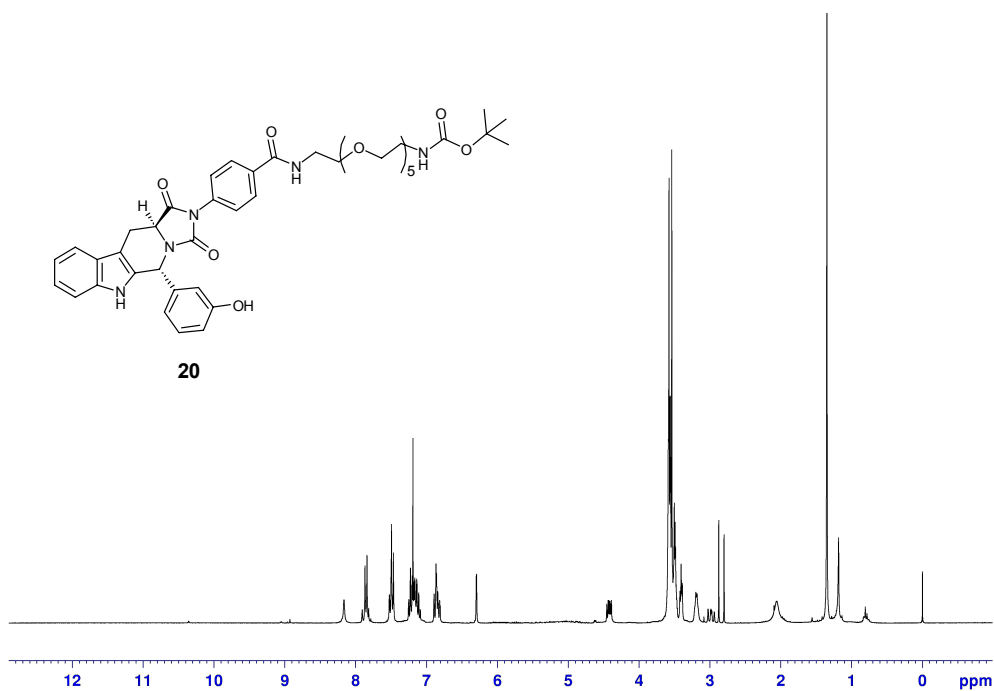
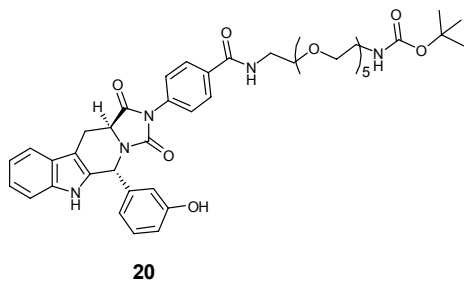


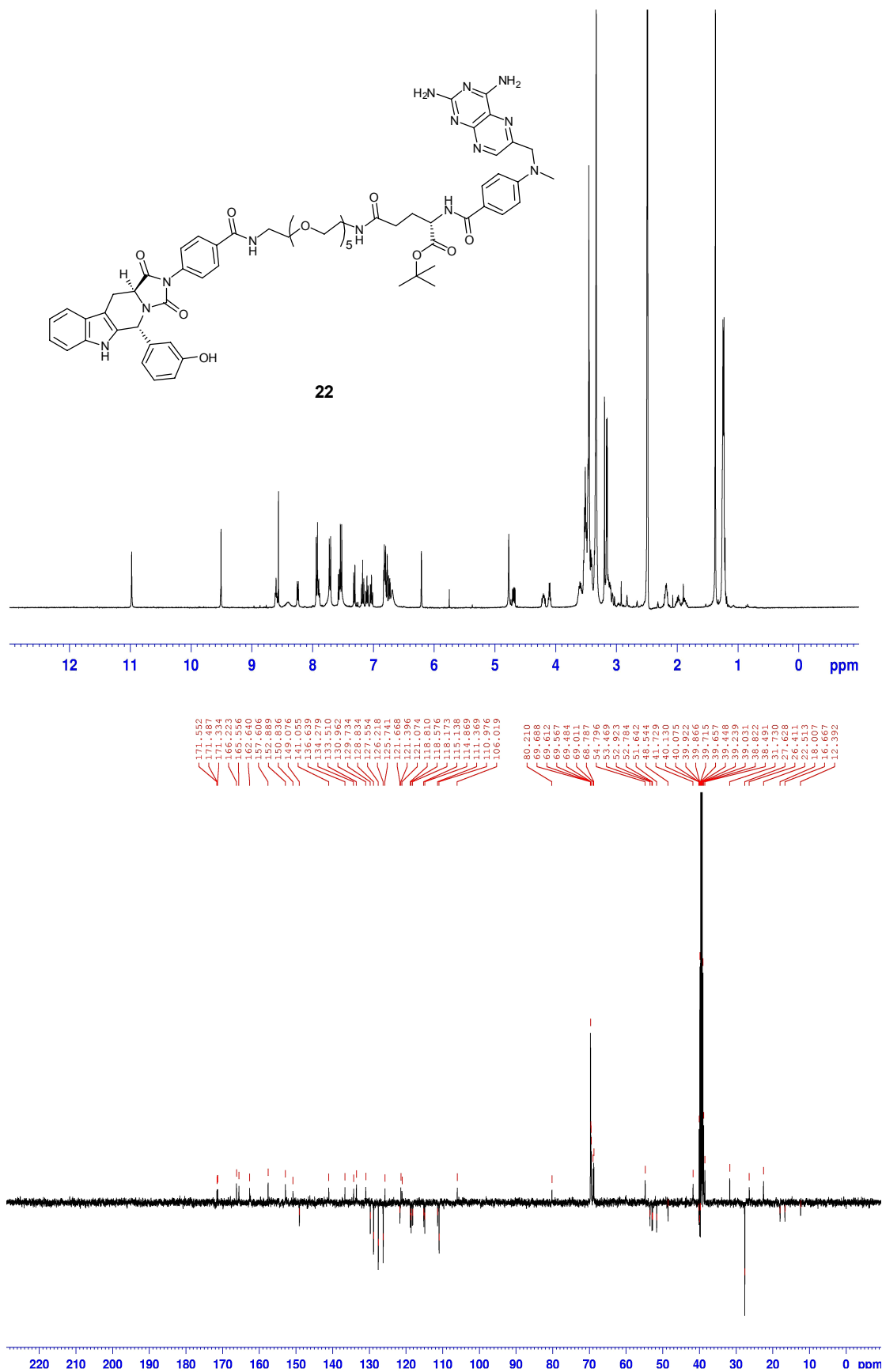
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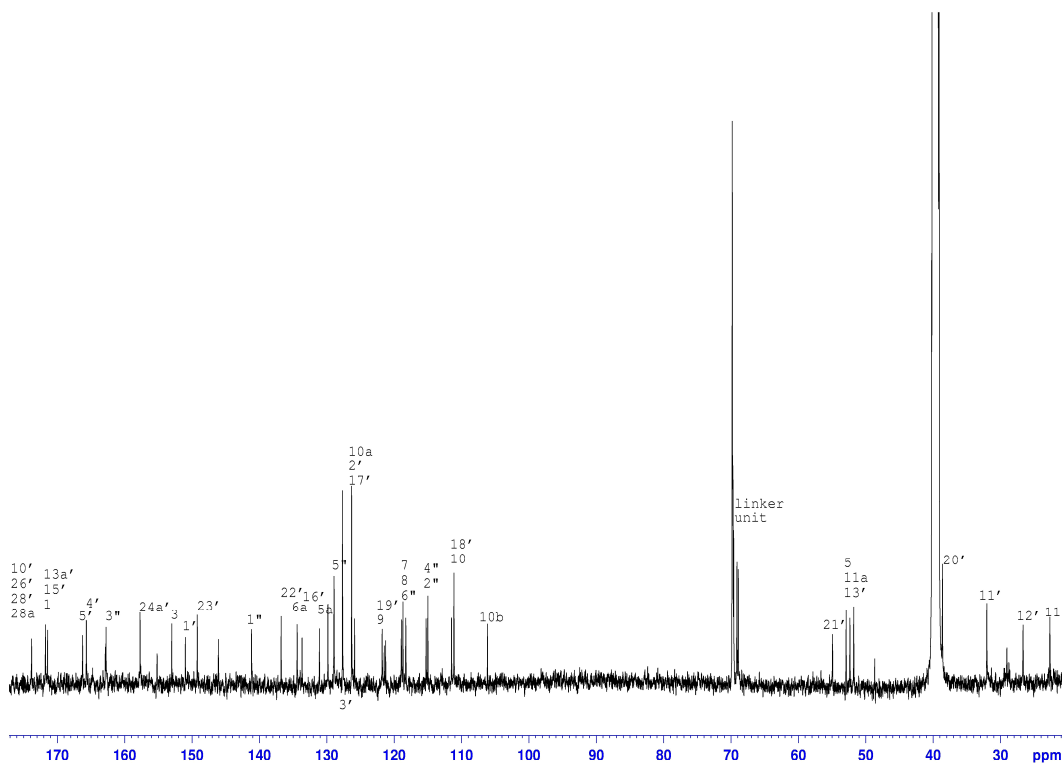
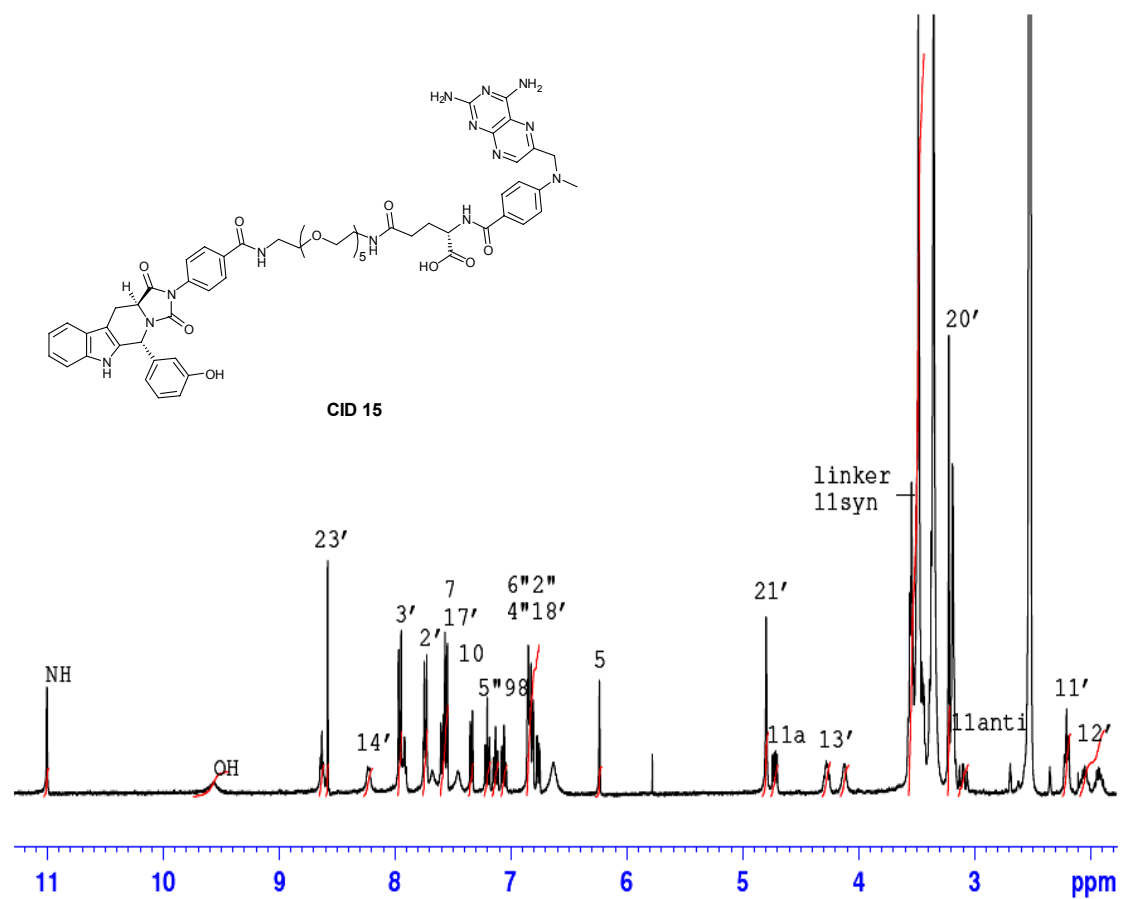


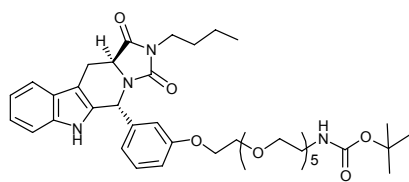




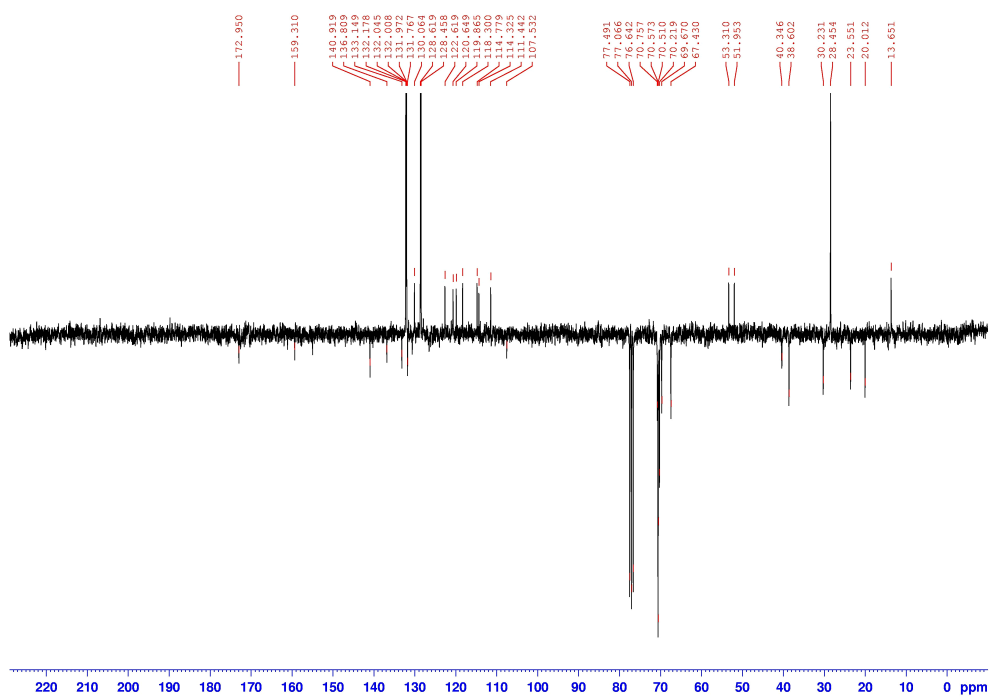
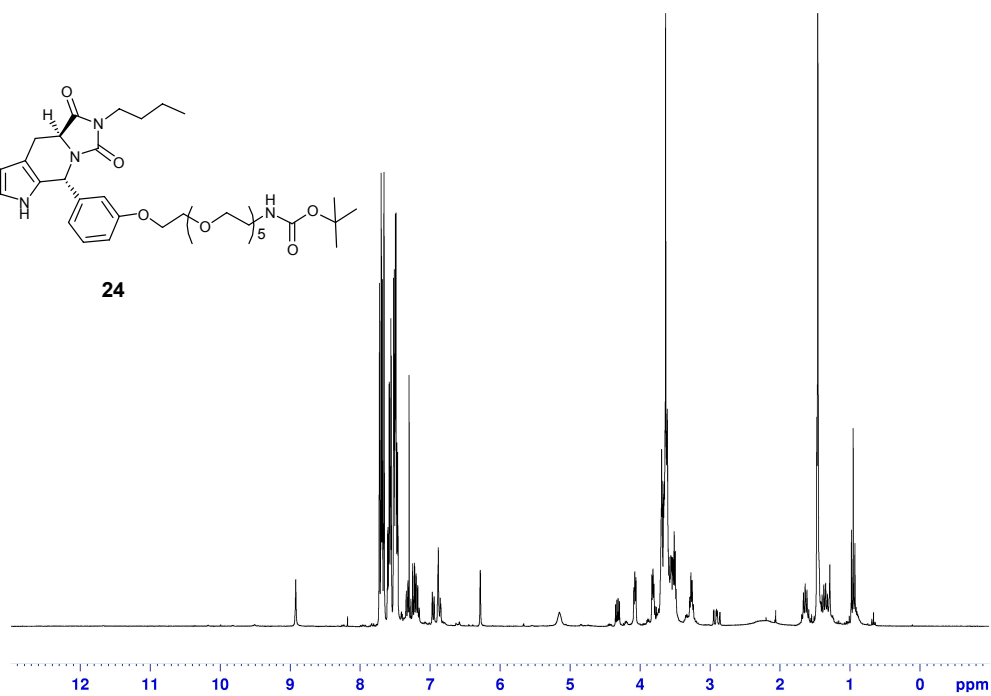


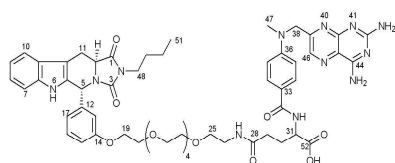






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