Facile synthesis of dicyanovinyl-di(*meso*-aryl)dipyrromethenes via a dipyrromethene-DDQ adduct

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Experimental and NMR data

General

All chemicals were purchased from commercial suppliers and used without further purification. Column chromatography of all products was carried out using silica gel (particle size: 0.040-0.063 mm, 230-400 mesh). ¹H and ¹³C NMR spectra were recorded with Bruker Avance 300 and 400 NMR spectrometers in CD₂Cl₂. Mass spectra were determined on EI or ESI mass spectrometers. Optical spectra were recorded with a Cary 5000 UV-vis spectrophotometer using a 1 cm cell. The crystals were grown by diffusion of hexane into CH₂Cl₂ solutions. Data were collected and integrated using the Bruker SAINT software package and corrected for absorption effects using the multi-scan technique (SADABS). The structures were solved by direct methods and all refinements were performed using the SHELXTL crystallographic software package from Bruker-AXS.

Procedure

Dipyrromethane 1:

Meso-2,6-dichlorophenyldipyrromethane was prepared following the reference method.^{S1} Column chromatography on silica gel using a mixed eluent of CH_2Cl_2 and hexane (3:1), followed by recrystallization from CH_2Cl_2 and hexane yielded an off-white solid in 60% yield.

Compounds 2-4:

DDQ (2.29 g, 10.09 mmol) was added into a CH_2Cl_2 (150 mL) solution of *meso*-2,6dichlorophenyldipyrromethane (1.11g, 3.85 mmol). After stirring for 2 days, the reaction mixture was concentrated and column chromatographed on silica gel using CH_2Cl_2 . Three fractions were collected (first red (3), second yellow (2), and third blue) and each was recrystallized from CH_2Cl_2 and hexane (yields: 30% (330 mg) and 8% (124 mg) for 2, and 3, respectively). A small amount of compound 3 was lost during the isolation due to its poor solubility. Compound 4 was prepared by the addition of Et_3N (0.5 mL) into a CH_2Cl_2 (75 mL) solution of 3 (30 mg, 37.4 µmol) and purified by column chromatography on silica gel using CH_2Cl_2 as eluent (80% yield).

Spectral data of **2** : ¹H NMR (400 MHz, CD₂Cl₂) δ = 12.36 (bs, 1H, NH), 7.66 (s, 2H, α H), 7.47 (dd, J = 8.6, ^{dd}J = 1.2, 2H, meta-H), 7.39 (t, J = 7.2, 1H, para-H), 6.39 (m, 4H, β H); ¹³C NMR (100 MHz, CD₂Cl₂) δ = 145.11 (α C), 140.45, 136.02, 135.49, 135.25, 130.93 (para-C), 128.60 (meta-C), 127.29 (β C), 118.96 (β C); m/z HREIMS found 288.02273 (100%), calcd. 288.02210 for C₁₅H₁₀N₂³⁵Cl₂ (M⁺); λ_{max} (nm, CH₂Cl₂, log ε) 431 (4.49).

Spectral data of **3** : ¹H NMR (400 MHz, CD₂Cl₂) δ = 11.18 (bs, 2H, NH), 7.64 (m, 2H, α H), 7.52-7.40 (m, 6H, Aryl-H), 6.79 (d, J = 4.3, 2H, β H of inner pyrrole), 6.73 (d, J = 4.3, 2H, β H of inner pyrrole), 6.42 (m, 4H, β H of terminal pyrrole); ¹³C NMR (100 MHz, CD₂Cl₂) δ = 176.67, 156.04, 146.84, 142.83, 141.78, 136.71 (β C), 135.96, 135.66, 134.93 (α C), 134.19, 131.56 (*para*-C), 131.50, 128.82 (*meta*-C), 128.68 (*meta*-C), 126.19 (β C), 121.95 (β C), 114.57 (β C), 114.27, 30.28 (sp³C); *m*/*z* HRESIMS found 802.9671, calcd. 802.9691 for C₃₈H₁₉N₆O₂³⁵Cl₅³⁷Cl ([M+H]⁺); λ_{max} (nm, CH₂Cl₂, log ε) 500 (4.64).

Spectral data of 4: ¹H NMR (400 MHz, CD₂Cl₂) δ = 12.94 (bs, 2H, NH), 7.59 (m, 2H, α H), 7.54-7.42 (m, 6H, Aryl-H), 7.16 (d, J = 4.5, 2H, β H of inner pyrrole), 6.72 (d, J = 4.3, 2H, β H of inner pyrrole), 6.47 (dd, 2H, J =

4.1, ${}^{dd}J = 1.2$, β H of terminal pyrrole), 6.42 (dd, 2H, J = 4.3, ${}^{dd}J = 2.2$, β H of terminal pyrrole); 13 C NMR (100 MHz, CD₂Cl₂) $\delta = 140.04$, 136.82 (α C), 135.95, 134.58, 133.75, 133.57 (β C), 131.47, 128.77 (Aryl-C), 126.19 (β C), 125.77 (β C), 119.57, 115.38 (β C), 114.09; m/z HREIMS found 650.03271 (100%), calcd. 650.03471 for C₃₄H₁₈N₆³⁵Cl₄ (M⁺); λ_{max} (nm, CH₂Cl₂, log ε) 335 (4.18), 492 (3.59), 680 (4.31), 735 (4.50).

Compound 5:

AlCl₃ (300 mg) in MeOH (15 mL) was added into a THF (35 mL) solution of **3** (54 mg, 67.3 μ mol). After refluxing for 2 days, the reaction mixture was poured into water and extracted with CH₂Cl₂. The result solution was evaporated and the residue was column chromatographed on silica gel using CH₂Cl₂. The fastest pink fraction was collected and recrystallized from CH₂Cl₂ and hexane (10 mg, 30%)

Spectral data of **5** : ¹H NMR (400 MHz, CD₂Cl₂) δ = 12.43 (bs, 2H, NH), 7.54-7.35 (m, 6H, Aryl-H), 6.99 (dd, *J* = 4.0, ^{dd}*J* = 2.4, 2H, βH), 6.56 (d, *J* = 4.8, 2H, βH), 6.33 (d, 2H, *J* = 4.8, βH), 6.05 (dd, 2H, *J* = 4.4, ^{dd}*J* = 1.6, βH), 4.29 (s, 6H, OMe); ¹³C NMR (100 MHz, CD₂Cl₂) δ = 178.39, 160.22, 149.84, 144.99, 140.59, 138.79, 137.14 (βC), 136.90, 134.20, 131.24 (Aryl-C), 129.45, 128.79 (Aryl-C), 124.18, 122.67 (βC), 122.00, 120.70 (βC), 118.04 (βC), 113.99, 91.93, 58.36; *m/z* HRESIMS found 505.9636 (100%), calcd. 505.9633 for C₂₂H₁₂N₃³⁵Cl₄O₃ [(M+H)⁺]; λ_{max} (nm, CH₂Cl₂, log ε) 332 (4.51), 420 (3.94), 542 (4.48), 735 (4.50).



NMR spectra of 2, 3 and 4.

Figure S1. ¹H NMR (400 MHz) spectrum of 2 in CD_2Cl_2 .



Figure S2. ¹³C NMR (100 MHz) spectrum of 2 in CD₂Cl₂.



Figure S3. HMQC NMR (F1 = 100 MHz and F2 = 400 MHz) spectrum of 2 in CD_2Cl_2 .



Figure S4. ¹H NMR (400 MHz) spectrum of 3 in CD₂Cl₂.



Figure S5. HH COSY NMR (F1 = F2 = 400 MHz) spectrum of 3 in CD_2Cl_2 .



Figure S6. 13 C NMR (100 MHz) spectrum of 3 in CD₂Cl₂.





Figure S8. ¹H NMR (400 MHz) spectrum of 4 in CD₂Cl₂.



Figure S9. HH COSY NMR (F1 = F2 = 400 MHz) spectrum of 4 in CD_2Cl_2 .



Figure S10. ¹³C NMR (100 MHz) spectrum of 4 in CD₂Cl₂.



Figure S11. HMQC NMR (F1 = 100 MHz and F2 = 400 MHz) spectrum of 4 in CD_2Cl_2 .



Figure S12. ¹H NMR spectral changes of $CD_2Cl_2(1 \text{ mL})$ solution of **3** (1 µmole) by adding Et₃N (5 ~ 60 µL) followed by D₂O and H₂O; for the comparison, the ¹H NMR spectrum of **4** in CD_2Cl_2 has been added as the last spectrum.



Figure S13. ¹H NMR (400 MHz) spectra of 5 in CD₂Cl₂.





 184
 176
 168
 160
 152
 144
 136
 128
 120
 112
 104
 96
 88
 80
 72
 64
 56
 48

 Chemical Shift (ppm)

 Figure S15. ¹³C NMR (100 MHz) spectrum of 5 in CD₂Cl₂.



56 48 128 120 112 Chemical Shift (ppm) Figure S16. 45 deg APT (up) and 13 C NMR (down) spectra (100 MHz) of 5 in CD₂Cl₂.



Figure S17. HMQC (red) and HMBC (blue) NMR (F1 = 400 MHz, F2 = 100 MHz) spectra of 5 in CD₂Cl₂.

Crystallographic data

Crystallographic data of 3

A red crystal of $C_{38}H_{18}N_6O_2Cl_6CH_2Cl_2$ having approximate dimensions of $0.04 \times 0.22 \times 0.50$ mm was mounted on a glass fiber. All measurements were made on a Bruker X8 APEX II diffractometer with graphite monochromated Mo-K α radiation. The data were collected at a temperature of $-100.0 \pm 0.1^{\circ}C$ to a maximum 20 value of 56.0°. Data were collected in a series of ϕ and ω scans in 0.50° oscillations with 20.0 second exposures. The crystal-to-detector distance was 36.00 mm.

Of the 33043 reflections that were collected, 9356 were unique ($R_{int} = 0.032$); equivalent reflections were merged. Data were collected and integrated using the Bruker SAINT^{S2} software package. The linear absorption coefficient, μ , for Mo-K α radiation is 6.19 cm⁻¹. Data were corrected for absorption effects using the multi-scan technique (SADABS^{S3}), with minimum and maximum transmission coefficients of 0.892 and 0.976, respectively. The data were corrected for Lorentz and polarization effects.

The structure was solved by direct methods^{S4}. The molecule crystallizes with CH_2Cl_2 in the lattice. One solvent molecule was modelled, however a second was disordered in such a way that it was not possible to achieve a reasonable model. As a result the PLATON/SQUEEZE^{S5} program was used to generate a data set free of any electron density in the region of the disordered solvent. All non-hydrogen atoms were refined anisotropically. All hydrogen atoms were placed in calculated positions but were not refined. The final cycle of full-matrix least-squares refinement^{S6} on F² was based on 9356 reflections and 504 variable parameters and converged (largest parameter shift was 0.00 times its esd with unweighted and weighted agreement factors (R1 = 0.077 and wR2 = 0.174).

The standard deviation of an observation of unit weight⁸⁷ was 1.17. The weighting scheme was based on counting statistics. The maximum and minimum peaks on the final difference Fourier map corresponded to 0.95 and -1.38 $e/Å^3$, respectively.

Neutral atom scattering factors were taken from Cromer and Waber^{S8}. Anomalous dispersion effects were included in Fcalc^{S9}; the values for $\Delta f'$ and $\Delta f''$ were those of Creagh and McAuley^{S10}. The values for the mass

attenuation coefficients are those of Creagh and Hubbell^{S11}. All refinements were performed using the SHELXTL^{S12} crystallographic software package from Bruker-AXS.



Figure S18. ORTEP structure of 3. Thermal ellipsoids are scaled to the 50% probability level.

Table S1. Parameters of the hydrogen-bonds for 3.

Donor HAcceptor	D - H	HA	DA	D - HA
N(1)H(1n)O(1)	0.77(4)	2.55(3)	3.155(3)	137(3)
N(1)H(1n)N(2)	0.77(4)	2.25(3)	2.765(3)	125(3)
N(6)H(6n)O(2)	0.80(4)	2.50(3)	3.158(3)	140(3)
N(6)H(6n)N(5)	0.80(4)	2.19(3)	2.758(3)	128(3)

Crystallographic data of 4

An irregular green crystal of $C_{34}H_{18}N_6Cl_4$ having approximate dimensions of $0.17 \times 0.20 \times 0.30$ mm was mounted on a glass fiber. All measurements were made on a Bruker X8 APEX II diffractometer with graphite monochromated Mo-K α radiation. The data were collected at a temperature of $-100.0 \pm 0.1^{\circ}C$ to a maximum 20 value of 56.0°. Data were collected in a series of ϕ and ω scans in 0.50° oscillations with 20.0 second exposures. The crystal-to-detector distance was 36.00 mm.

Of the 63004 reflections that were collected, 7054 were unique ($R_{int} = 0.038$); equivalent reflections were merged. Data were collected and integrated using the Bruker SAINT^{S2} software package. The linear absorption coefficient, μ , for Mo-K α radiation is 4.39 cm⁻¹. Data were corrected for absorption effects using the multi-scan technique (SADABS^{S3}), with minimum and maximum transmission coefficients of 0.868 and 0.928, respectively. The data were corrected for Lorentz and polarization effects.

The structure was solved by direct methods^{S4}. All non-hydrogen atoms were refined anisotropically. All N-H hydrogen atoms were located in difference maps and refined isotropically. All other hydrogen atoms were placed in calculated positions but were not refined. The final cycle of full-matrix least-squares refinement^{S6} on F^2 was based on 7054 reflections and 405 variable parameters and converged (largest parameter shift was 0.00 times its esd) with unweighted and weighted agreement factors (R1 = 0.053 and wR2 = 0.082).

The standard deviation of an observation of unit weight⁸⁷ was 1.02. The weighting scheme was based on counting statistics. The maximum and minimum peaks on the final difference Fourier map corresponded to 0.32 and $-0.33 \text{ e}^2/\text{Å}^3$, respectively.

Neutral atom scattering factors were taken from Cromer and Waber^{S8}. Anomalous dispersion effects were included in Fcalc^{S9}; the values for Δf and $\Delta f''$ were those of Creagh and McAuley^{S10}. The values for the mass attenuation coefficients are those of Creagh and Hubbell^{S11}. All refinements were performed using the SHELXTL^{S12} crystallographic software package from Bruker-AXS.



Figure S19. ORTEP structure of 4. Thermal ellipsoids are scaled to the 50% probability level.



Figure S21. C2 symmetric axis for crystal structure of 4.

Table S2. Parameters of the hydrogen-bonds for 4.

Donor HAcceptor	D - H	HA	DA	D - HA
N(1)H(1n)N(2)	0.83(2)	2.16(2)	2.7338(19)	126.3(17)
N(1)H(1n)N(4)	0.83(2)	2.31(2)	3.008(2)	142.9(18)
N(6)H(6n)N(3)	0.87(2)	2.52(2)	3.264(2)	144.0(17)
N(6) $H(6n)$ $N(5)$	0.87(2)	2.13(2)	2.7542(19)	127.7(17)

Crystallographic data of 5

An irregular violet crystal of $C_{22}H_{11}N_3O_3Cl_4$ having approximate dimensions of $0.12 \times 0.18 \times 0.40$ mm was mounted on a glass fiber. All measurements were made on a Bruker X8 APEX II diffractometer with graphite monochromated Mo-K α radiation. The data were collected at a temperature of $-100.0 \pm 0.1^{\circ}C$ to a maximum 2 θ value of 56.0°. Data were collected in a series of ϕ and ω scans in 0.50° oscillations with 20.0 second exposures. The crystal-to-detector distance was 36.00 mm.

Of the 36940 reflections that were collected, 5085 were unique ($R_{int} = 0.029$); equivalent reflections were merged. Data were collected and integrated using the Bruker SAINT^{S2} software package. The linear absorption coefficient, μ , for Mo-K α radiation is 5.95 cm⁻¹. Data were corrected for absorption effects using the multi-scan technique (SADABS^{S3}), with minimum and maximum transmission coefficients of 0.864 and 0.931, respectively. The data were corrected for Lorentz and polarization effects.

The structure was solved by direct methods^{S4}. All non-hydrogen atoms were refined anisotropically. All C-H hydrogen atoms were placed in calculated positions but were not refined. The N-H hydrogen atom was located in a difference map and refined isotropically. The final cycle of full-matrix least-squares refinement^{S6} on F^2 was based on 5085 reflections and 294 variable parameters and converged (largest parameter shift was 0.00 times its esd) with unweighted and weighted agreement factors (R1 = 0.053 and wR2 = 0.082).

The standard deviation of an observation of unit weight^{\$7} was 1.03. The weighting scheme was based on counting statistics. The maximum and minimum peaks on the final difference Fourier map corresponded to 0.37 and -0.35 e⁷/Å³, respectively.

Neutral atom scattering factors were taken from Cromer and Waber^{S8}. Anomalous dispersion effects were included in Fcalc^{S9}; the values for Δf and $\Delta f''$ were those of Creagh and McAuley^{S10}. The values for the mass attenuation coefficients are those of Creagh and Hubbell^{S11}. All refinements were performed using the SHELXTL^{S12} crystallographic software package from Bruker-AXS.



Figure S22. ORTEP structure of 6. Thermal ellipsoids are scaled to the 50% probability level.

Table S3. Parameters of the hydrogen-bonds for 5.

Donor HAcceptor	D - H	HA	DA	D - HA
N(1)H(1n)O(2)	0.839(19)	2.360(19)	2.7781(16)	111.4(14)
N(1)H(1n)N(2)	0.839(19)	2.140(17)	2.7383(18)	128.1(16)

	3	4	5
Formula	$C_{39}H_{20}N_6O_2Cl_8$	C34H18N6Cl4	C ₂₂ H ₁₁ N ₃ Cl ₄ O ₃
Mw	888.21	652.34	507.14
cryst syst	triclinic	monoclinic	triclinic
Space group	P-1 (# 2)	$P 2_1/c (\#14)$	<i>P</i> -1 (# 2)
a/Å	11.2796(18)	14.1530(16)	7.8137(8)
b/Å	12.066(2)	10.3574(11)	9.9057(11)
$c/\text{\AA}$	14.734(2)	20.116(2)	14.0847(16)
α/deg	83.100(8)	90.0	84.895(5)
β/deg	79.807(8)	92.535(5)	77.368(5)
γ/deg	89.782(8)	90.0	81.828(2)
$V/\text{\AA}^3$	1959.0(6)	2945.8(6)	1051.0(2)
Ζ	2	4	2
$D_{\rm c}/{\rm g~cm^{-3}}$	1.506	1.471	1.603
μ (MoK α)cm ⁻¹	6.19	4.39	5.95
No. of obsd data $(I > 0.00\sigma(I))$	9356	7054	5085
Reflection / Parameter Ratio	18.56	17.42	17.30
$R1^{\rm a}$; $wR2^{\rm b}$	0.077; 0.174	0.053; 0.082	0.038; 0.079
GOF	1.17	1.02	1.03
No. of obsd data ($I \ge 2\sigma(I)$)	6694	5367	4365
(R1; wR2) °	(0.053; 0.163)	(0.033; 0.073)	(0.030; 0.074)
$R1 = \Sigma \omega Fo - Fc / \Sigma \omega Fo ,$	b wR2 = $\sqrt{\Sigma}$ { ω (F	$\overline{\text{Fo}^2 - \text{Fc}^2}$ / $\Sigma \omega (\text{Fo}^2)^2$	^c refined on F, I> $2\sigma(I)$

Table S4. Crystallographic Data of 3, 4 and 5.

Reference

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- S6. Least Squares function minimized: $\Sigma w (F_o^2 F_c^2)^2$
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 - Where: N_0 = number of observations
 - N_v = number of variables
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• Perfluorophenyl substituted dipyrromethane formed a DDQ adduct in higher yield (~ 20%). The (a) 1 H and (b) 19 F NMR spectra (in CD₂Cl₂) are included below.

