

Supporting Information

A Straightforward Microwave Method for Rapid Synthesis of

N-1, C-6 Functionalized 3,5-Dichloro-2(1H)-Pyrazinones

*Johan Gising, Pernilla Örtqvist, Anja Sandström and Mats Larhed**

Organic Pharmaceutical Chemistry, Department of Medicinal Chemistry, Uppsala Biomedical Centre,
Uppsala University, Box 574, SE-751 23 Uppsala, Sweden.

mats@orgfarm.uu.se

General section	S2
General One-Pot, Two-Step Procedure for Preparation of 4a-n . Method I.	S3
One-Pot, Two-Step Procedure for Preparation of 4j . Method II.	S3
5 mmol, One-Pot, Two Step Procedure for Preparation of 4m . Method III.	S4
General Procedure for Preparation of α -Aminonitrile 3e*,i-k . Method IV.	S4
General Procedure for Preparation of 4i,o-s from pure α -Aminonitriles. Method V.	S5
General Procedure for Determination of Diastereomeric Ratio of Compounds 4j-n . Method VI.	S5
Experimental Procedures and Spectroscopic Data for Compounds 4a-n .	S5-12
Experimental Procedures and Spectroscopic Data for Compounds 3e*,i-k .	S12-14
Experimental Procedures and Spectroscopic Data for Compounds 4o-s .	S14-16
Experimental Procedures and Spectroscopic Data for Compounds 5, 6, 7a, 7b .	S16-19
References	S19
Spectra of compounds	S20-77

General Section

All reactions were performed in reaction vials dedicated for microwave processing. The microwave experiments were carried out using a Smith Synthesizer™ or Emrys Initiator™ single mode cavity, equipped with magnetic stirring and producing controlled irradiation at 2450 MHz. The temperature of the reaction mixture was measured using a built-in, on-line infrared temperature sensor. To avoid over pressurizations or explosions, sealed reactions should always be performed in dedicated equipment. For flash chromatography commercially available silica gel 60 (particle size: 0.040-0.063 mm) was used. Analytical thin-layer chromatography was performed on silica gel 60 F-254 plates (E. Merck) and visualized with UV light. ¹H NMR spectra were recorded at 399.9 MHz and ¹³C NMR spectra at 100.5 MHz. The chemical shifts for ¹H NMR and ¹³C NMR are referenced to TMS via residual solvent signals (¹H, CDCl₃ at 7.26 ppm, CD₃OD at 3.31 ppm; ¹³C, CDCl₃ at 77.16 ppm, CD₃OD at 49.0 ppm). Optical rotations were obtained on a Perkin-Elmer 241 polarimeter and the concentration (*c*) is given as g/100 mL in the specified solvent. Analytical RPLC-MS analysis of reaction mixtures and pure products were performed using a Gilson HPLC system with a Chromolith SpeedROD RP-18e column (50×4.6 mm) and a Finning AQA quadropole mass spectrometer using a 4 mL/min CH₃CN/H₂O gradient (0.05% aqueous HCOOH) and detection by UV (DAD) or ELSD and MS (ESI+). Preparative RP-HPLC was performed using a Zorbax SB-C8 column (5 μm, 21.2 × 150 mm) with gradients of MeCN/H₂O (0.1% TFA) at a flow rate of 5 mL/min and UV detection at 230 nm. The purity of pseudo peptides **7a** and **7b** were determined on an ACE 5 C18 column (5 μm, 4.6 × 50 mm) using the same buffer system at a flow rate of 2 mL/min and with UV detection at 220 nm. Analytische Laboratorien, Lindlar, Germany performed elemental analyses. Compounds **3a** and **3e** were purchased from Acros. Compounds **3e***,¹⁻³ **3i**,⁴ **3k**,⁵ **4f**,⁶ **4g**,⁷ **4i**,⁸ **4p**,⁹ **4q**,⁴ **4r**,¹⁰ **4s**,¹⁰ (1*R*,2*S*)-

ethyl 1-amino-2-vinylcyclopropanecarboxylate hydrochloride are all known compounds. Spectral data were in agreement with the proposed structures.

Biochemical evaluation: Compounds **7a** and **7b** were evaluated against the hepatitis C virus NS3 protease in vitro, using the full-length NS3 protein, the central part of the co-factor NS4A and an internally quenched fluorescent peptide substrate as described elsewhere.¹¹

General One-Pot, Two-Step Procedure for Preparation of 2(1*H*)-Pyrazinones 4a-n.

Method I.

A 2.0-5.0 mL Smith microwave vial was charged with amine (**1a-h**, 1.0 mmol) and was dissolved in 3.0 mL DME. The aldehyde (**2a-e**, 1.2 mmol) was added and the mixture was stirred for 30 s before adding trimethylsilyl cyanide (1.1 mmol, 138 μ L). The vial was sealed and irradiated with microwaves for 10 min at stated temperatures (step 1, Table 1). The solvent was removed under a stream of nitrogen gas and the residue was dissolved in 5.0 mL diethylether. HCl gas was bubbled through the reaction mixture for 5 min followed by evaporation under a stream of nitrogen gas. DME (3.0 mL) and oxalyl chloride (2.5 mmol, 214 μ L) were added and the vial was sealed. After 30 s of stirring, the overpressure was released with a needle before irradiation with microwaves for 10 min at 170 °C (creating a pressure of 10-17 bar). Purification by flash chromatography yielded pure products **4a-n** (>95% by LC-ELSD).

One-Pot, Two-Step Procedure for Preparation of Benzyl 2-(3,5-dichloro-6-methyl-2-oxopyrazin-1(2*H*)-yl)-3-methylbutanoate 4j. Method II.

The title compound was prepared following method I with amine **1f** (2.04 mmol, 0.423 g), acetaldehyde **1a** (2.46 mmol, 0.137 mL), trimethylsilyl cyanide (2.25 mmol, 0.282 mL),

oxalyl chloride (5.11 mmol, 0.438 mL), using 10 min of microwave heating at 80 °C in step 1. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 5:95 to 15:85, yielded pure product **4j** as colorless oil, 0.397 g (53%).

5 mmol, One-Pot, Two-Step Procedure for Preparation of Ethyl 2-(3,5-dichloro-6-methyl-2-oxopyrazin-1(2*H*)-yl)-3-phenylpropanoate 4m. Method III.

A 10.0-20.0 mL Smith microwave vial was charged with amine **1h** (5.0 mmol, 0.965 g) and 10.0 mL DME. Acetaldehyde **2a** (6.0 mmol, 0.335 mL) was added and the mixture was stirred for 30 s before adding trimethylsilyl cyanide (5.5 mmol, 0.710 mL). The vial was sealed and irradiated with microwaves at 80 °C for 10 min (step 1). The solvent was removed under a stream of nitrogen gas and the residue was dissolved in 15 mL diethylether. HCl gas was bubbled through the reaction mixture for 5 min followed by evaporation under a stream of nitrogen gas. DME (10.0 mL) and oxalyl chloride (12.5 mmol, 1.09 mL) were added and the vial was sealed. After 30 s stirring the overpressure was released with a needle before irradiation with microwaves for 25 min at 145 °C (creating a pressure of 20 bar). Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 10:90 to 20:80, yielded pure product **4m** as pale yellow solid, 0.973 g (55%).

General Procedure for Preparation of α -Aminonitrile 3e*,i-k. Method IV.

A 2.0-5.0 mL Smith microwave vial was charged with 1 equiv amine **1e,i-j** and 3.0 mL DME. (Diisopropylamine (1.2 equiv) was added if the amine was a HCl salt). Addition of 1.2 equiv aldehyde **2a,b** or phenylacetaldehyde (**2f**) followed by 30 s stirring before addition of 1.1 equiv trimethylsilyl cyanide. The vial was sealed and irradiated with microwaves for 10

minutes at stated temperatures. The solvent was evaporated in vacuum and the crude residue purified by flash chromatography yielding pure products **3e*,i-k**.

General Procedure for Preparation of 2(1*H*)-Pyrazinones **4i,o-s** from Pure α -Aminonitriles. Method V.

A 2.0-5.0 mL Smith microwave vial was loaded with 1 mmol α -aminonitrile **3a,e*,e,i-k** and 5.0 mL diethylether. HCl gas was bubbled through the reaction mixture for 5 min followed by removal of the diethylether under a stream of nitrogen gas. DME (3.0 mL) and oxalyl chloride (2.5 mmol, 214 μ L) were added and the vial was sealed. After 30 s stirring the overpressure was released and the reaction irradiated by microwaves for 10 min at 170 $^{\circ}$ C. Purification by flash chromatography yielded pure products **4i,o-s**.

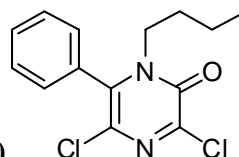
General Procedure for Determination of Diastereomeric Ratio of compounds **4j-n**. Method VI.

A 0.5-2.0 mL Smith microwave vial was loaded with 2(1*H*)-pyrazinones **4j-n** (0.05-0.15 mmol), (1*S*,2*R*)-*cis*-1-amino-2-indanol (3 equiv.) and acetonitrile (2.0 mL). The reaction was then irradiated with microwaves: **4j** at 150 $^{\circ}$ C for 1.5 h; **4k** at 140 $^{\circ}$ C for 2 h; **4l** at 140 $^{\circ}$ C for 0.5 h; **4m** at 160 $^{\circ}$ C for 2 h; **4n** at 160 $^{\circ}$ C for 1.5 h. The diastereomeric ratios were determined by 1 H-NMR or by HPLC separation with ELSD and ESI-MS detection.



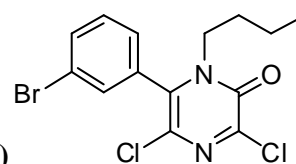
The compound was prepared according to method I, using 60 $^{\circ}$ C temperature in step 1. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 10:90 to 20:80. Pale

yellow oil, 72% yield. ^1H NMR (CDCl_3) δ 4.06 (m, 2H), 2.49 (s, 3H), 1.66 (m, 2H), 1.43 (qm, $J=7.4$ Hz, 2H), 0.97 (t, $J=7.4$ Hz, 3H); ^{13}C NMR (CDCl_3) δ 152.7, 143.5, 135.6, 123.8, 47.4, 29.9, 20.2, 16.6, 13.7. ESI-MS (m/z) 235 ($\text{M} + \text{H}^+$), 471 ($2\text{M} + \text{H}^+$). Anal. Calcd for $\text{C}_9\text{H}_{12}\text{Cl}_2\text{N}_2\text{O}$: C, 45.98; H, 5.14; N, 11.91. Found: C, 46.14; H, 5.25; N, 12.04.



1-Butyl-3,5-dichloro-6-phenylpyrazin-2(1H)-one (4b)

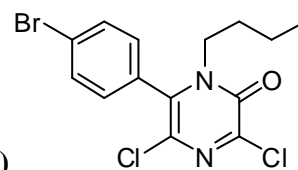
The compound was prepared according to method I, using 60 °C temperature in step 1. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 5:95 to 15:85. Pale yellow oil, 58% yield. ^1H NMR (CDCl_3) δ 7.49-7.44 (m, 3H), 7.22 (m, 2H), 3.66 (m, 2H), 1.43 (m, 2H), 1.03 (qm, $J = 7.4$ Hz, 2H), 0.62 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (CDCl_3) δ 152.4, 145.9, 138.6, 130.7, 130.5, 129.4, 129.1, 124.1, 48.6, 30.0, 19.9, 13.4. ESI-MS (m/z) 297 ($\text{M} + \text{H}^+$), 595 ($2\text{M} + \text{H}^+$). Anal. Calcd for $\text{C}_{14}\text{H}_{14}\text{Cl}_2\text{N}_2\text{O}$: C, 56.58; H, 4.75; N, 9.43. Found: C, 56.68; H, 4.82; N, 9.35.



6-(3-Bromophenyl)-1-butyl-3,5-dichloropyrazin-2(1H)-one (4c)

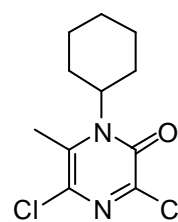
The compound was prepared according to method I, using 60 °C temperature in step 1. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 5:95 to 15:85. Colourless oil, 57% yield. ^1H NMR (CDCl_3) δ 7.71 (dm, $J=8.1$ Hz, 1H), 7.49 (m, 1H), 7.45 (ddm, $J = 8.8, 8.1$ Hz, 1H), 7.27 (dm, $J=8.8$ Hz, 1H), 3.75 (m, 2H), 1.54 (m, 2H), 1.17 (qm, $J=7.3$ Hz, 2H), 0.76 (t, $J=7.3$ Hz, 3H); ^{13}C NMR (CDCl_3) δ 151.2, 145.6, 135.9, 132.9, 131.3, 131.1, 130.0, 126.9, 123.1, 122.4, 47.7, 29.1, 18.9, 12.4. ESI-MS (m/z) 377 ($\text{M} + \text{H}^+$), 753

(2M + H⁺). Anal. Calcd for C₁₄H₁₃BrCl₂N₂O: C, 44.71; H, 3.48; N, 7.45. Found: C, 44.84; H, 3.61; N, 7.40.



6-(4-Bromophenyl)-1-butyl-3,5-dichloropyrazin-2(1H)-one (4d)

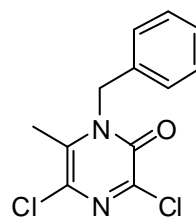
The compound was prepared according to method I, using 80 °C temperature in step 1. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 5:95 to 15:85. White solid, 43% yield. ¹H NMR (CDCl₃) δ 7.70 (m, 2H), 7.20 (m, 2H), 3.75 (m, 2H), 1.51 (m, 2H), 1.15 (qm, *J*=7.3 Hz, 2H), 0.76 (t, *J*=7.3 Hz, 3H); ¹³C NMR (CDCl₃) δ 152.2, 146.3, 137.4, 132.8, 130.8, 129.3, 125.3, 124.1, 48.6, 30.1, 20.0, 13.4. ESI-MS (*m/z*) 377 (M + H⁺), 753 (2M + H⁺). Anal. Calcd for C₁₄H₁₃BrCl₂N₂O: C, 44.71; H, 3.48; N, 7.45. Found: C, 44.88; H, 3.63; N, 7.44.



3,5-Dichloro-1-cyclohexyl-6-methylpyrazin-2(1H)-one (4e)

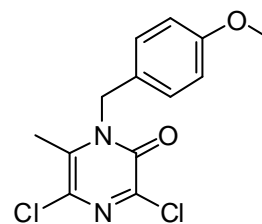
The compound was prepared according to method I, using 60 °C temperature in step 1. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 5:95 to 15:85. White solid, 79% yield. ¹H NMR (CDCl₃) δ 4.00 (m, 1H), 2.64 (m, 2H), 2.46 (s, 3H), 1.85 (m, 2H), 1.67-1.54 (m, 3H), 1.33-1.16 (m, 3H); ¹³C NMR (CDCl₃) δ 152.6, 145.1, 135.4, 123.9, 63.5, 27.2, 26.0, 24.8, 17.3. ESI-MS (*m/z*) 261 (M + H⁺), 523 (2M + H⁺). Anal. Calcd for C₁₁H₁₄Cl₂N₂O: C, 50.59; H, 5.40; N, 10.73. Found: C, 50.62; H, 5.49; N, 10.59.

1-Benzyl-3,5-dichloro-6-methylpyrazin-2(1H)-one (4f)⁶



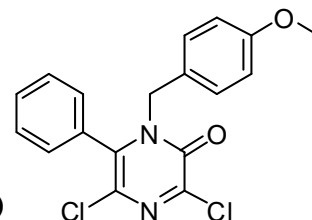
The compound was prepared according to method I, using 60 °C temperature in step 1. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 10:90 to 20:80. Colourless oil, 67% yield.

3,5-Dichloro-1-(4-methoxybenzyl)-6-methylpyrazin-2(1H)-one (4g)⁷

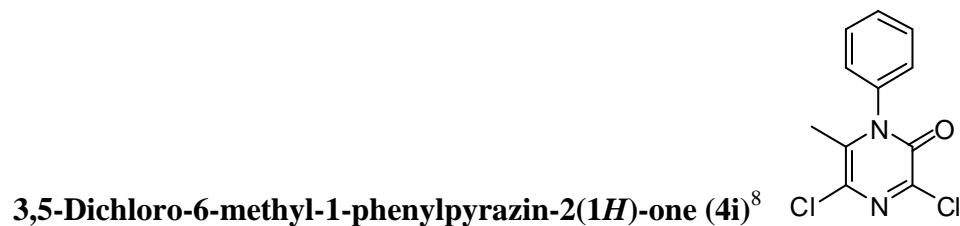


The compound was prepared according to method I, using 80 °C temperature in step 1. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 10:90 to 25:75. Colourless oil, 55% yield.

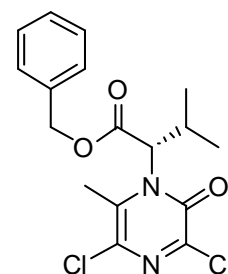
3,5-Dichloro-1-(4-methoxybenzyl)-6-phenylpyrazin-2(1H)-one (4h)



The compound was prepared according to method I, using 80 °C temperature in step 1. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 10:90 to 20:80. Pale yellow oil, 38% yield. ¹H NMR (CDCl₃) δ 7.57-7.44 (m, 3H), 7.12 (m, 2H), 6.79-6.68 (m, 4H), 5.03 (br s, 2H), 3.76 (s, 3H); ¹³C NMR (CDCl₃) δ 159.7, 153.0, 146.3, 138.6, 130.7, 130.4, 129.7, 129.6, 129.3, 127.0, 124.5, 114.0, 55.4, 50.9. ESI-MS (*m/z*) 361 (M + H⁺), 723 (2M + H⁺). Anal. Calcd for C₁₈H₁₄Cl₂N₂O₂: C, 59.85; H, 3.91; N, 7.76. Found: C, 59.80; H, 3.92; N, 7.65.

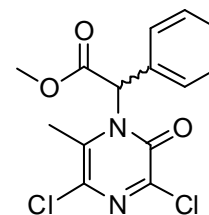


The compound was prepared according to method I using 100 °C temperature in step 1 or from **3i** utilizing method V. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 10:90 to 20:80. White solid, 29% yield utilizing method I, 70% yield utilizing method V. ¹³C NMR (CDCl₃) δ 152.9, 144.7, 137.2, 136.0, 130.5, 130.3, 127.0, 123.5, 18.3.



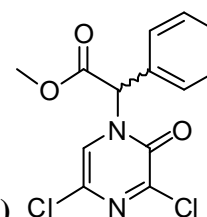
Benzyl 2-(3,5-dichloro-6-methyl-2-oxopyrazin-1(2H)-yl)-3-methylbutanoate (4j)

The compound was prepared according to method I, using 100 °C temperature in step 1 or by method II, using 80 °C temperature in step 1. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 5:95 to 15:85. Colourless oil, 51% yield from method I, 53% yield from method II. The product **4j** was isolated in >98:2 diastereomeric ratio according to ¹H-NMR using method VI. ¹H NMR (CD₃OD at 50 °C) δ 7.34-7.21 (m, 5H), 5.22 (d, *J*=12.2 Hz, 1H), 5.06 (d, *J*=12.2 Hz, 1H), 4.90 (m, 1H), 2.88 (m, 1H), 2.45 (s, 3H), 1.28 (d, *J*=6.5 Hz, 3H), 0.71 (d, *J*=6.5 Hz, 3H); ¹³C NMR (CD₃OD at 50 °C) δ 168.8, 154.0, 144.2, 138.2, 136.7, 129.6, 129.4, 129.3, 125.4, 68.5, 68.0, 28.4, 22.0, 19.2, 17.7. ESI-MS (*m/z*) 369 (M + H⁺), 739 (2M + H⁺). [α]_D²¹ -53.5 ° (*c* 0.95, CH₂Cl₂). Anal. Calcd for C₁₇H₁₈Cl₂N₂O₃: C, 55.30; H, 4.91; N, 7.59. Found: C, 55.46; H, 5.01; N, 7.50.



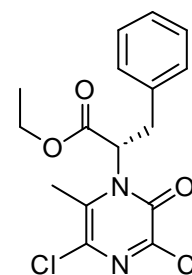
Methyl 2-(3,5-dichloro-6-methyl-2-oxopyrazin-1(2H)-yl)-2-phenylacetate (4k)

The compound was prepared according to method I, using 80 °C temperature in step 1. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 15:85 to 25:75. Pale yellow solid, 44% yield. The product **4k** was isolated in a 47:54 diastereomeric ratio according to HPLC separation with ELSD and ESI-MS detection using method VI. ¹H NMR (CD₃OD) δ 7.43-7.35 (m, 5H), 6.47 (br s, 1H), 3.77 (s, 3H), 2.48 (s, 3H); ¹³C NMR (CD₃OD) δ 168.7, 154.4, 144.7, 138.2, 133.7, 129.9, 129.72, 129.69, 125.7, 65.4, 53.5, 17.5. EIS-MS (*m/z*) 327 (M + H⁺), 655 (2M + H⁺). [α]_D²¹ -57.5 ° (*c* 0.92, CH₂Cl₂). Anal. Calcd for C₁₄H₁₂Cl₂N₂O₃: C, 51.40; H, 3.70; N, 8.56. Found: C, 51.13; H, 3.64; N, 8.46.



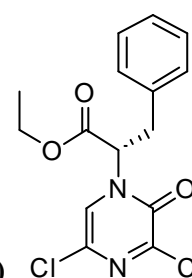
Methyl 2-(3,5-dichloro-2-oxopyrazin-1(2H)-yl)-2-phenylacetate (4l)

The compound was prepared according to method I, using 170 °C temperature in step 1. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 15:85. White semi solid, 38% yield. The product **4l** was isolated in a 44:56 diastereomeric ratio according to HPLC separation with ELSD and ESI-MS detection using method VI. ¹H NMR (CDCl₃) δ 7.54-7.48 (m, 3H), 7.31 (m, 2H), 6.94 (s, 1H), 6.51 (br s, 1H), 3.85 (s, 3H); ¹³C NMR (CDCl₃) δ 168.1, 152.0, 146.8, 130.9, 130.8, 130.2, 129.5, 124.5, 124.3, 63.4, 53.6. ESI-MS (*m/z*) 313 (M + H⁺), 627 (2M + H⁺). [α]_D²⁴ -0.3 ° (*c* 1.05, CH₂Cl₂). Anal. Calcd for C₁₃H₁₀Cl₂N₂O₃: C, 49.86; H, 3.22; N, 8.95. Found: C, 49.46; H, 3.30; N, 8.72.



Ethyl 2-(3,5-dichloro-6-methyl-2-oxopyrazin-1(2H)-yl)-3-phenylpropanoate (4m)

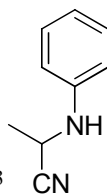
The compound was prepared according to method I, using 100 °C temperature in step 1 or by method III, using 80 °C temperature in step 1. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 10:90 to 20:80. Pale yellow solid; 63% yield from method I, 55% yield from method III. The product **4m** was isolated in a 97:3 diastereomeric ratio according to HPLC separation with ELSD and ESI-MS detection using method VI. ¹H NMR (CD₃OD at 50 °C) δ 7.28-7.21 (m, 3H), 7.05 (m, 2H), 5.36 (ddm, *J*=10.4, 4.5 Hz, 1H), 4.25 (qm, *J*=7.1 Hz, 2H), 3.59 (dd, *J*=14.1, 4.5 Hz, 1H), 3.49 (dd, *J*=14.1, 10.4 Hz, 1H), 1.97 (s, 3H), 1.25 (t, *J*=7.1 Hz, 3H); ¹³C NMR (CD₃OD at 50 °C) δ 168.8, 153.8, 144.0, 138.5, 137.8, 130.3, 129.9, 128.5, 125.0, 64.7, 63.2, 34.1, 17.1, 14.3. ESI-MS (*m/z*) 355 (M + H⁺), 711 (2M + H⁺). [α]²¹_D -328.0° (*c* 0.99, CH₂Cl₂). Anal. Calcd for C₁₆H₁₆Cl₂N₂O₃: C, 54.10; H, 4.54; N, 7.89. Found: C, 54.28; H, 4.96; N, 7.92.



Ethyl 2-(3,5-dichloro-2-oxopyrazin-1(2H)-yl)-3-phenylpropanoate (4n)

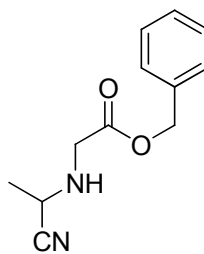
The compound was prepared according to method I, using 170 °C temperature in step 1. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 10:90 to 20:80. White solid, 54% yield. The product **4n** was isolated in >98:2 diastereomeric ratio according to ¹H-NMR using method VI. ¹H NMR (CD₃OD) δ 7.56 (s, 1H), 7.29-7.18 (m, 3H), 7.12 (m, 2H), 5.43 (dd, *J*=10.8, 5.5 Hz, 1H), 4.25 (q, *J*=7.2 Hz, 2H), 3.57 (dd, *J*=14.4, 5.5 Hz, 1H), 3.39 (dd,

$J=14.4, 10.8$ Hz, 1H), 1.25 (t, $J=7.2$ Hz, 3H); ^{13}C NMR (CD_3OD) δ 167.7, 151.4, 146.9, 134.3, 129.1, 128.8, 127.8, 125.4, 123.7, 62.7, 61.7, 36.3, 14.1. ESI-MS (m/z) 341 ($\text{M} + \text{H}^+$), 683 ($2\text{M} + \text{H}^+$). HRMS calcd for $\text{C}_{15}\text{H}_{14}\text{Cl}_2\text{N}_2\text{O}_3$ ($\text{M} + \text{H}^+$) 341.0460; found: 341.0447. $[\alpha]_{\text{D}}^{21}$ -25.8 ° (c 1.05, CH_2Cl_2).



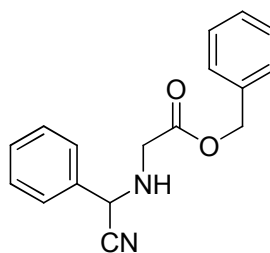
2-(Phenylamino)propanenitrile (3e*)¹⁻³

The compound was prepared according to method IV, using aniline **1e** (2.6 mmol, 0.239 g), acetaldehyde **2a** (3.1 mmol, 172 μL), trimethylsilyl cyanide (2.8 mmol, 354 μL) and a temperature of 100 °C. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 10:90 to 15:85. White solid, 42% yield. ^1H NMR (CD_3OD) δ 7.19 (m, 2H), 6.80-6.73 (m, 3H), 4.42 (m, 1H), 1.60 (m, 3H); ^{13}C NMR (CD_3OD) δ 147.5, 130.2, 122.0, 120.0, 115.0, 41.7, 19.4. ESI-MS (m/z) 147 ($\text{M} + \text{H}^+$).



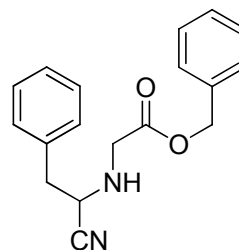
Benzyl 2-(1-cyanoethylamino)acetate (3i)⁴

The compound was prepared according to method IV, using $\text{HCl} \times \text{HGlyOBn}$ (2.0 mmol, 0.402 g), diisopropylamine (2.3 mmol, 400 μL), acetaldehyde **2a** (2.5 mmol, 140 μL), trimethylsilyl cyanide (2.2 mmol, 280 μL) and a temperature of 100 °C. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 30:70 to 75:25. Colorless oil, 68% yield. ^{13}C -NMR (CD_3OD) δ 172.5, 137.3, 129.7, 129.5 (two peaks), 121.7, 67.8, 49.2, 45.8, 19.6.



Benzyl 2-(cyano(phenyl)methylamino)acetate (3j)

The compound was prepared according to method IV, using HCl×HGlyOBn (2.0 mmol, 0.403 g), diisopropylamine (2.4 mmol, 400 μ L), benzaldehyde **2b** (2.5 mmol, 250 μ L), trimethylsilyl cyanide (2.2 mmol, 280 μ L) and a temperature of 100 °C. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 30:70 to 75:25. Colorless oil, 83% yield. $^1\text{H-NMR}$ (CD_3OD) δ 7.52-7.49 (m, 2H), 7.42-7.28 (m, 8H), 5.19 (d, $J=12.3$ Hz, 1H), 5.15 (d, $J=12.3$ Hz, 1H), 5.05 (s, 1H), 3.58 (d, $J=17.5$ Hz, 1H), 3.51 (d, $J=17.5$ Hz, 1H). $^{13}\text{C-NMR}$ (CD_3OD) δ 172.5, 137.2, 136.1, 130.2, 130.1, 129.7, 129.5 (two peaks), 128.8, 67.8, 54.3, 48.7. ESI-MS (m/z) 281 ($\text{M} + \text{H}^+$), 561 ($2\text{M} + \text{H}^+$). Anal. calcd for $\text{C}_{17}\text{H}_{16}\text{N}_2\text{O}_2$: C, 72.83; H, 5.75; N, 10.00. Found: C, 72.63; H, 5.86; N, 9.83.

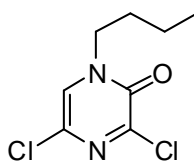


Benzyl 2-(1-cyano-2-phenylethylamino)acetate (3k)⁵

The compound was prepared according to method IV, using HCl×HGlyOBn (2.0 mmol, 0.402 g), diisopropylamine (2.4 mmol, 400 μ L), 2-phenylacetaldehyde (2.5 mmol, 310 μ L), trimethylsilyl cyanide (2.2 mmol, 280 μ L) and a temperature of 100 °C. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 30:70 to 75:25. Colorless oil, 88% yield. $^1\text{H-NMR}$ (CD_3OD) δ 7.37-7.28 (m, 10H), 5.19 (d, $J=12.3$ Hz, 1H), 5.16 (d, $J=12.3$ Hz, 1H), 4.01 (dd, $J=8.7, 5.9$ Hz, 1H), 3.62 (d, $J=17.4$ Hz, 1H), 3.54 (d, $J=17.4$ Hz, 1H), 3.09 (dd, $J=13.5, 5.9$ Hz, 1H), 2.99 (dd, $J=13.5, 8.7$ Hz, 1H). $^{13}\text{C-NMR}$ (CD_3OD) δ 172.5, 137.3, 137.1, 130.6,

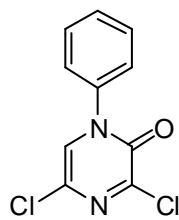
129.8, 129.7, 129.5 (two peaks), 128.5, 67.8, 52.8, 49.3, 40.2. ESI-MS (m/z) 295 ($M + H^+$), 589 ($2M + H^+$).

1-Butyl-3,5-dichloropyrazin-2(1H)-one (4o)



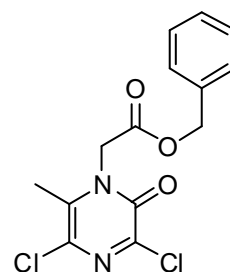
The compound was prepared according to method V. The α -aminonitrile **3a** was bought. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 10:90 to 20:80. White solid, 64% yield. ^1H NMR (CDCl_3) δ 7.21 (s, 1H), 3.93 (m, 2H), 1.75 (m, 2H), 1.38 (qm, $J=7.3$ Hz, 2H), 0.96 (t, $J=7.3$ Hz, 3H); ^{13}C NMR (CDCl_3) δ 151.9, 147.5, 126.9, 123.9, 51.3, 30.7, 19.9, 13.6. ESI-MS (m/z) 221 ($M + H^+$), 443 ($2M + H^+$). Anal. Calcd for $\text{C}_8\text{H}_{10}\text{Cl}_2\text{N}_2\text{O}$: C, 43.46; H, 4.56; N, 12.67. Found: C, 43.51; H, 4.58; N, 12.63.

3,5-Dichloro-1-phenylpyrazin-2(1H)-one (4p)⁹

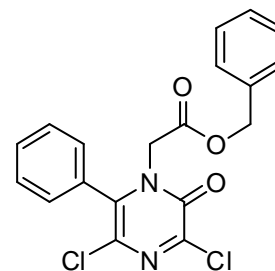


The compound was prepared according to method V. The α -aminonitrile **3e** was bought. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 10:90 to 20:80. White solid, 80% yield.

Benzyl 2-(3,5-dichloro-6-methyl-2-oxopyrazin-1(2H)-yl)acetate (4q)⁴

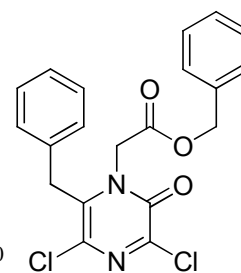


The compound was prepared according to method V. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 20:80 to 35:65. Pale yellow solid, 86% yield. ^{13}C -NMR (CDCl_3) δ 166.1, 152.7, 143.8, 135.2, 134.6, 129.1, 129.0, 128.7, 124.0, 68.4, 47.6, 16.9. ESI-MS (m/z) 327 ($\text{M} + \text{H}^+$), 655 ($2\text{M} + \text{H}^+$).



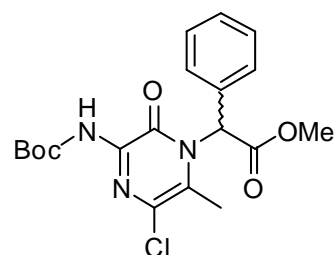
Benzyl 2-(3,5-dichloro-2-oxo-6-phenylpyrazin-1(2H)-yl)acetate (4r)¹⁰

The compound was prepared according to method V. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 20:80 to 35:65. Pale yellow solid, 65% yield. ^1H -NMR (CD_3OD) δ 7.54 (m, 1H), 7.47 (m, 2H), 7.35 (m, 3H), 7.30-7.26 (m, 4H), 5.15 (s, 2H), 4.54 (s, 2H). ^{13}C -NMR (CD_3OD) δ 168.1, 154.0, 146.3, 140.2, 136.7, 132.1, 131.5, 130.7, 130.3, 129.8 (two peaks), 129.7, 125.3, 68.9, 50.7. ESI-MS (m/z) 389 ($\text{M} + \text{H}^+$), 779 ($2\text{M} + \text{H}^+$).



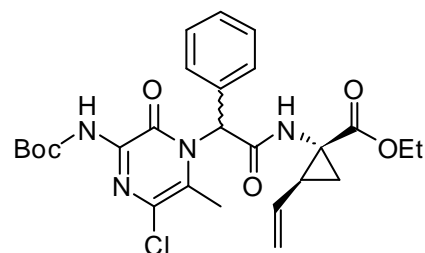
Benzyl 2-(6-benzyl-3,5-dichloro-2-oxopyrazin-1(2H)-yl)acetate (4s)¹⁰

The compound was prepared according to method V. Purification by flash chromatography, eluent ethyl acetate:*iso*-hexane 20:80 to 35:65. Pale yellow solid, 56% yield. ^1H -NMR (CD_3OD) δ 7.38-7.25 (m, 8H), 7.17 (m, 2H), 4.96 (s, 2H), 4.82 (s, 2H), 4.25 (s, 2H). ^{13}C -NMR (CD_3OD) δ 167.3, 154.6, 145.3, 139.3, 136.8, 135.4, 130.4, 129.8, 129.7, 129.4 (two peaks), 128.8, 126.3, 68.8, 49.5, 36.5. ESI-MS (m/z) 403 ($\text{M} + \text{H}^+$), 807 ($2\text{M} + \text{H}^+$).



Methyl 2-(3-(tert-butoxycarbonylamino)-5-chloro-6-methyl-2-oxopyrazin-1(2H)-yl)-2-phenylacetate (5)

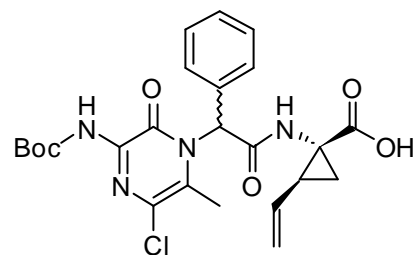
A 2.0-5.0 mL Smith microwave vial was charged with **4k** (0.196 g, 0.601 mmol), *tert*-butyl carbamate (0.353 g, 3.01 mmol), Pd(OAc)₂ (0.007 g, 0.052 mmol), Xantphos (0.028 g, 0.048 mmol) and Cs₂CO₃ (0.392 g, 1.20 mmol). The vial was sealed and irradiated with microwaves for 30 min at 100 °C. The solvent was removed and the residue purified by flash chromatography, eluent DCM:ethyl acetate 100:2 to 100:4 yielding the sought product **5** in 51% yield, 0.090 g as a white solid. ¹H NMR (CDCl₃ with two drops of D₂O) δ 7.41-7.29 (m, 5H), 6.25 (br s, 1H), 3.81 (s, 3H), 2.33 (s, 3H), 1.52 (s, 9H); ¹³C NMR (CDCl₃) δ 167.4, 151.2, 149.5, 143.3, 132.5, 129.1, 129.0, 128.3, 126.5, 126.0, 82.4, 62.7, 53.4, 28.3, 17.0. ESI-MS (*m/z*) 408 (M + H⁺), 815 (2M + H⁺). [α]_D²¹ -0.2 ° (*c* 1.09, CH₂Cl₂). Anal. Calcd for C₁₉H₂₂ClN₃O₅·0.25H₂O: C, 55.34; H, 5.50; N, 10.19. Found: C, 55.48; H, 5.26; N, 10.14.



(1R,2S)-Ethyl 1-(2-(3-(tert-butoxycarbonylamino)-5-chloro-6-methyl-2-oxopyrazin-1(2H)-yl)-2-phenylacetamido)-2-vinylcyclopropanecarboxylate (6)

Compound **5** (0.37 mmol, 0.150 g) was dissolved in THF (7.0 mL). A solution of LiOH (3.8 mmol, 0.092 g) and H₂O (4.0 mL) was added and the reaction was stirred at room temperature for 7 h. Water (20 mL) was added and the THF was removed in vacuo. The pH

was adjusted to ≈ 2 using 1 M HCl (aq). The aqueous phase was extracted twice with ethyl acetate, the organic phases were combined and the solvent was removed in vacuo. The corresponding acid was used in the subsequent coupling reaction without further purification. The crude acid (0.051 g), (1*R*,2*S*)-ethyl 1-amino-2-vinylcyclopropanecarboxylate hydrochloride (0.20 mmol, 0.039 g), HATU (0.16 mmol, 0.061 g) and DIEA (1.2 mmol, 0.20 mL) was dissolved in DMF (1.3 mL). The pH of the solution was controlled and found to be >10 . The reaction was stirred at room temperature over night, after which another portion of (1*R*,2*S*)-ethyl 1-amino-2-vinylcyclopropanecarboxylate hydrochloride (0.13 mmol, 0.025 g), HATU (0.16 mmol, 0.062 g) and DIEA (0.6 mmol, 100 μ L) were added. The pH was stable >10 . Stirring was continued for a total of five days. Ethyl acetate (10 mL) was added, and the solution was washed with 0.1 M NaHSO₄ (3 \times 5 mL) and NaHCO₃ (sat) (2 \times 5 mL) and the solvent was removed in vacuo. The crude product **6** was purified by column chromatography (DCM:methanol 95:5) to give (0.057 g, 29% over two steps) as a diastereomeric mixture (approximately 1:1 as determined by NMR). ¹H-NMR (CDCl₃) δ 8.16 (s, 1H), 8.15 (s, 1H), 7.41 (m, 10 H), 6.65 (s, 1H), 6.56 (s, 1H), 6.36 (s, 1H), 6.14 (s, 1H), 5.74 (m, 2H), 5.28 (m, 2H), 5.12 (m, 2H), 4.16 (m, 4H), 2.37 (s, 3H), 2.34 (s, 3H), 2.24 (m, 1H), 2.10 (m, 1H), 1.90 (dd, $J=8.2, 5.5$ Hz, 1H), 1.85 (dd, $J=8.2, 5.5$ Hz, 1H), 1.67 (dd, $J=9.7, 5.5$ Hz, 1H), 1.54 (dd, $J=9.7, 5.5$ Hz, 1H), 1.52 (s, 18 H), 1.26 (t, $J=7.1$ Hz, 3H), 1.26 (t, $J=7.1$ Hz, 3H). ¹³C-NMR (CDCl₃) δ 169.8 (two peaks), 167.0 (two peaks), 151.5, 151.4, 149.6, 149.5, 143.4, 143.3, 133.3, (two peaks), 133.1, 132.9, 129.7, 129.6 (two peaks), 129.4, 128.4, 128.3, 127.4, 127.3, 126.2 (broad signal), 118.5, 118.4, 82.3 (two peaks), 65.4, 64.7, 61.8 (two peaks), 40.5, 40.4, 34.2, 34.1, 28.3, 23.0 (two peaks), 17.5, 17.2, 14.4. Anal. Calcd for C₂₆H₃₁ClN₄O₆: C, 58.81; H, 5.88; N, 10.55. Found: C, 58.92; H, 5.80; N, 10.49.



(1R,2S)-1-(2-(3-(*Tert*-butoxycarbonylamino)-5-chloro-6-methyl-2-oxopyrazin-1(2H)-yl)-2-phenylacetamido)-2-vinylcyclopropanecarboxylic acid (7a and 7b)

Compounds **7a** and **7b** was prepared from diastereomeric mixture of **6** (0.11 mmol, 0.057 g), LiOH (1.2 mmol, 0.028 g), THF (3.0 mL) and H₂O (1.0 mL). After two days stirring at room temperature, the pH was adjusted to 2 using 1 M HCl (aq). The aqueous phase was extracted with ethyl acetate (3×10 mL), the organic phases were combined and the solvent was removed in vacuum. The crude product was purified by RP-HPLC (Sorbax SB-C8 column, MeCN:H₂O gradient with 0.1% TFA) to give the pure diastereomers **7a** (0.0029 g, yield 5.3 %) and **7b** (0.0035 g, yield 6.3%) of the title compound.

7a: ¹H-NMR (CDCl₃) δ 8.22 (s, 1H), 7.44 (m, 3H), 7.35 (m, 2H), 6.41 (s, 1H), 5.99 (s, 1H), 5.80 (ddd, *J*=17.1, 10.4, 8.4 Hz, 1H), 5.31 (dd, *J*=17.1, 1.9 Hz, 1H), 5.17 (dd, *J*=10.3, 1.9 Hz, 1H), 2.38 (s, 3H), 2.23 (m, 1H), 1.89 (dd, *J*=8.4, 5.7 Hz, 1H), 1.54 (s, 9H), 1.51 (dd, *J*=9.6, 5.7 Hz, 1H). ¹³C-NMR (CDCl₃) δ 172.3, 168.2, 151.4, 149.8, 143.5, 133.0, 132.6, 130.2, (two peaks) 128.5, 127.3, 126.8, 119.0, 66.0, 40.4, 35.0, 28.4, 23.9, 17.1. HRMS calcd for C₂₄H₂₇ClN₄O₆ (M+H⁺) 503.1697; found 503.1707. RP-HPLC purity, column A (ACE 5 C8-A3071, MeCN:H₂O linear gradient with 0.1% TFA, UV detection at 220 nm): 97.5%.

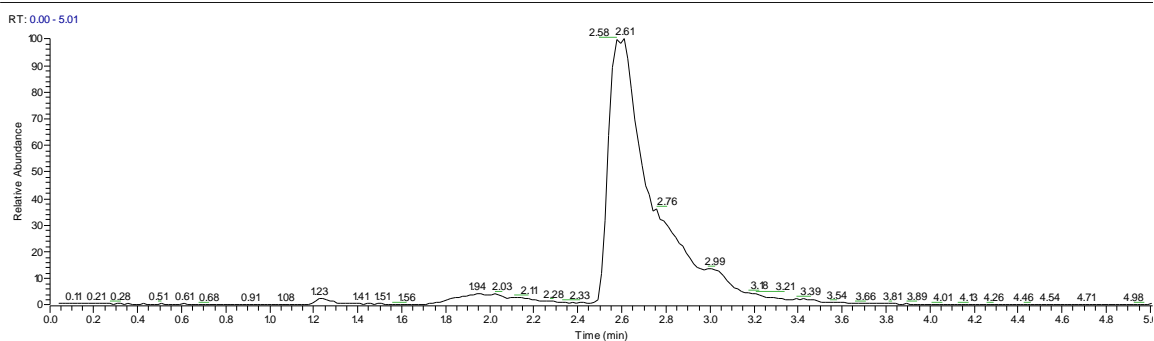
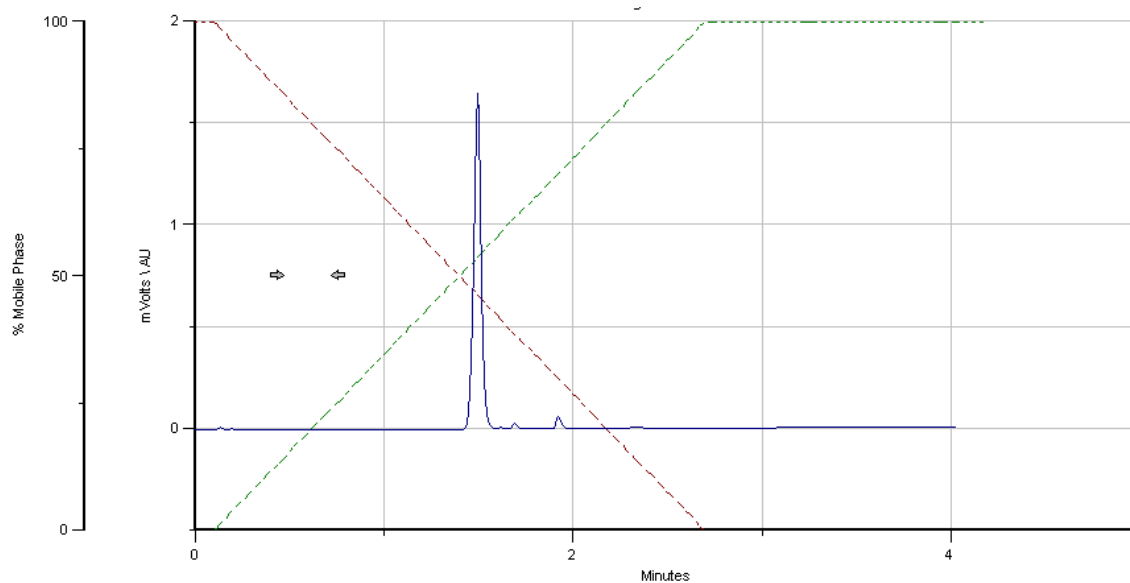
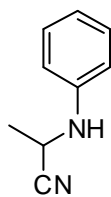
7b: ¹H-NMR (CDCl₃) δ 8.18 (s, 1H), 7.46 (m, 3H), 7.37 (m, 2H), 6.38 (brs, 1H), 5.92 (s, 1H), 5.78 (ddd, *J*=17.1, 10.3, 8.7 Hz, 1H), 5.31 (dd, *J*=17.1, 2.0 Hz, 1H), 5.17 (dd, *J*=10.3, 2.0 Hz, 1H), 2.39 (s, 3H), 2.20 (m, 1H), 1.90 (dd, *J*=8.4, 5.6 Hz, 1H), 1.57-1.53 (m, 10H). ¹³C-NMR (CDCl₃) δ 172.3, 168.1, 151.4, 149.8, 143.6, 132.9, 132.2, 130.4, 130.3, 128.5, 127.2, 126.8, 119.1, 82.8, 66.3, 40.4, 35.5, 28.4, 23.7, 17.0. HRMS calcd for C₂₄H₂₇ClN₄O₆ (M+H⁺)

503.1697; found 503.1699. RP-HPLC purity, column A (ACE 5 C8-A3071, MeCN:H₂O linear gradient with 0.1% TFA, UV detection at 220 nm): 98.6%.

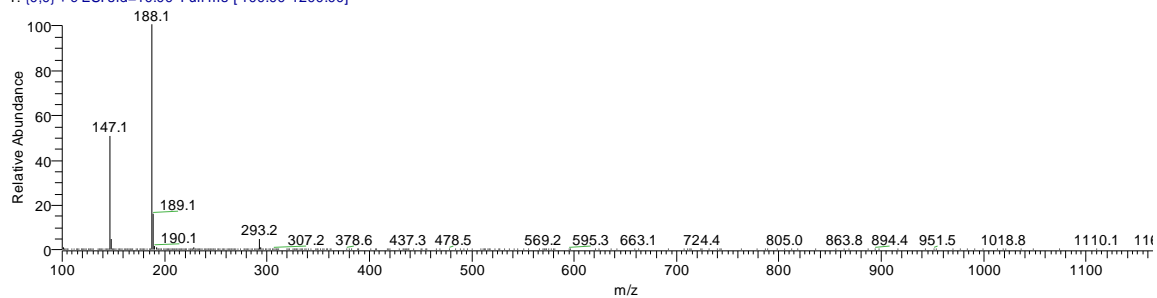
References

- (1) von Walther, R.; Hubner, R. *J. Prakt. Chem.* **1916**, *93*, 119-136.
- (2) Fourneau, J. P. *Bull. Soc. Chim. Fr.* **1944**, *11*, 141-148.
- (3) Ainley, A. D.; Sexton, W. A. *Biochem. J.* **1948**, *43*, 468-474.
- (4) Sanderson, P. E. J.; Lyle, T. A.; Cutrona, K. J.; Dyer, D. L.; Dorsey, B. D.; McDonough, C. M.; Naylor-Olsen, A. M.; Chen, I. W.; Chen, Z. G.; Cook, J. J.; Cooper, C. M.; Gardell, S. J.; Hare, T. R.; Krueger, J. A.; Lewis, S. D.; Lin, J. H.; Lucas, B. J.; Lyle, E. A.; Lynch, J. J.; Stranieri, M. T.; Vastag, K.; Yan, Y. W.; Shafer, J. A.; Vacca, J. P. *J. Med. Chem.* **1998**, *41*, 4466-4474.
- (5) Morley, J. S. *J. Chem. Soc. C.* **1969**, 809-813.
- (6) Buysens, K. J.; Vandenberghe, D. M.; Toppet, S. M.; Hoornaert, G. J. *Tetrahedron* **1995**, *51*, 12463-12478.
- (7) Rombouts, F. J. R.; De Borggraeve, W. M.; Delaere, D.; Froeyen, M.; Toppet, S. M.; Compennolle, F.; Hoornaert, G. J. *Eur. J. Org. Chem.* **2003**, 1868-1878.
- (8) Vandenberghe, S. M.; Buysens, K. J.; Meerpoel, L.; Loosen, P. K.; Toppet, S. M.; Hoornaert, G. J. *J. Org. Chem.* **1996**, *61*, 304-308.
- (9) Vekemans, J.; Pollerswieers, C.; Hoornaert, G. J. *Heterocycl. Chem.* **1983**, *20*, 919-923.
- (10) South, M. S.; Case, B. L.; Wood, R. S.; Jones, D. E.; Hayes, M. J.; Girard, T. J.; Lachance, R. M.; Nicholson, N. S.; Clare, M.; Stevens, A. M.; Stegeman, R. A.; Stallings, W. C.; Kurumbail, R. G.; Parlow, J. J. *Bioorg. Med. Chem. Lett.* **2003**, *13*, 2319-2325.
- (11) Poliakov, A.; Hubatsch, I.; Shuman, C. F.; Stenberg, G.; Danielson, U. H. *Protein Expr. Purif.* **2002**, *25*, 363-371.

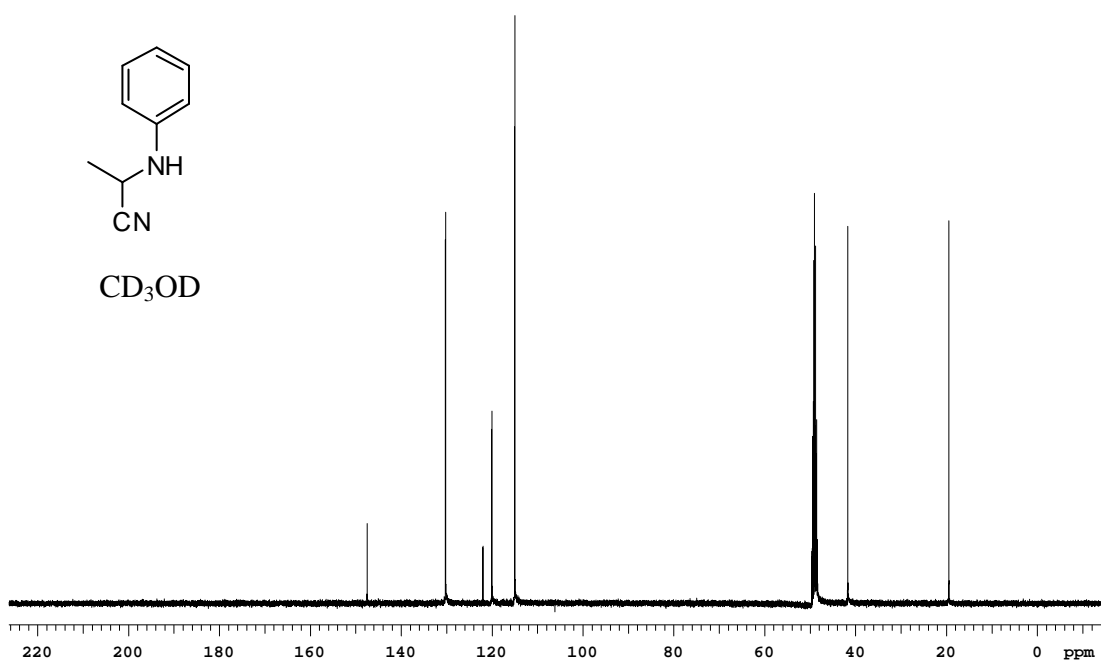
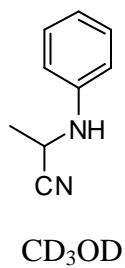
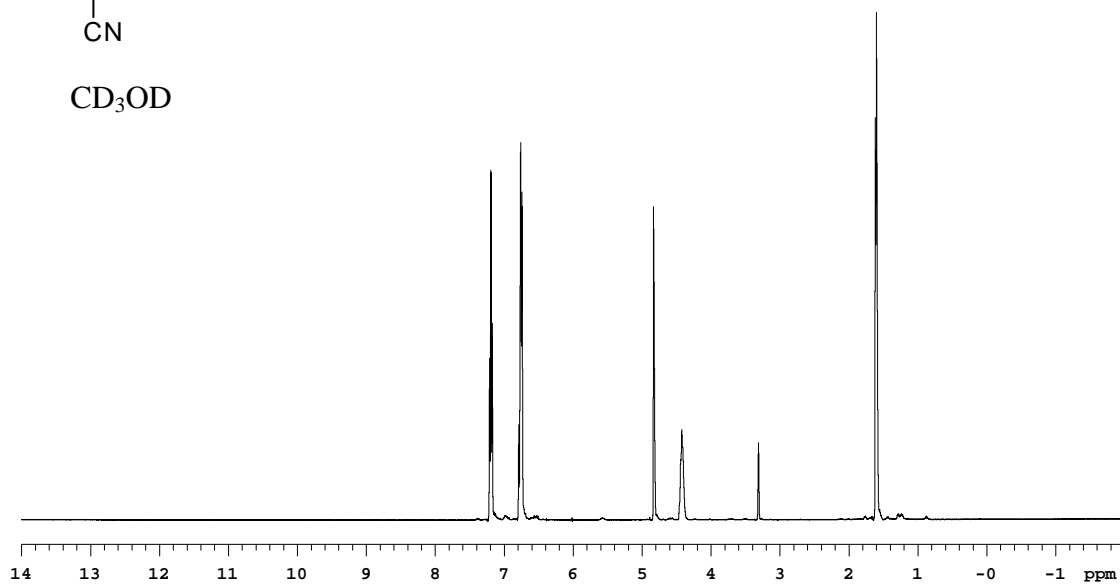
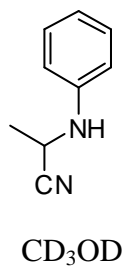
3e*



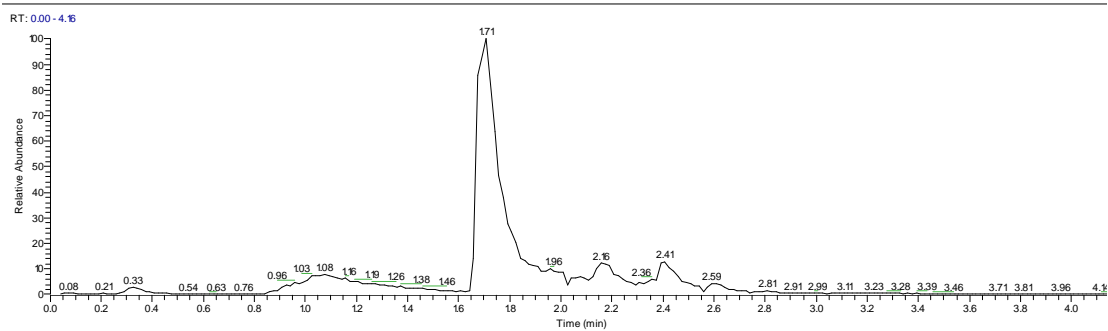
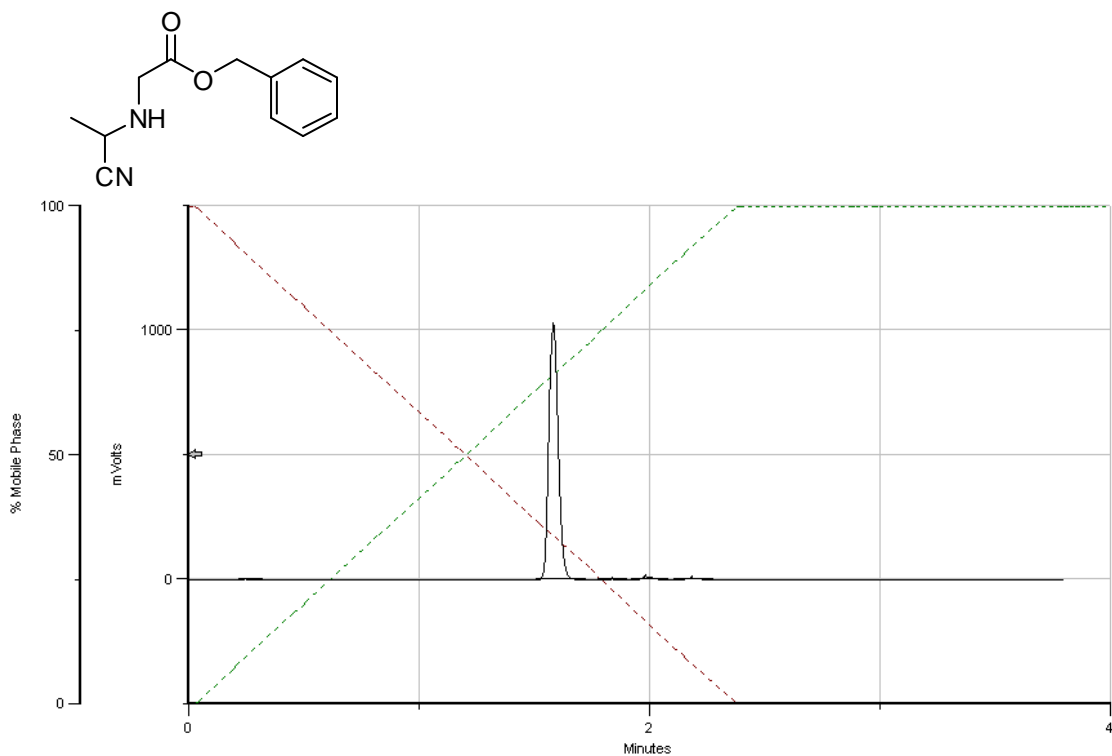
SQ_gis007644 #154 RT: 2.59 AV: 1 NL: 2.32E6
T: (0,0) + c ESI sid=10.00 Full ms [100.00-1200.00]



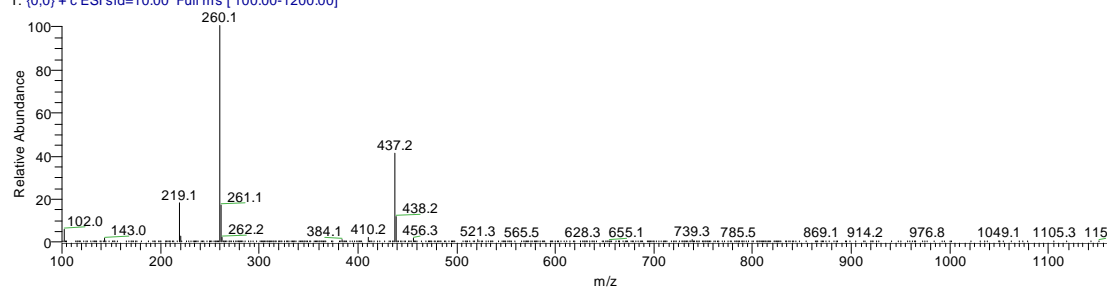
3e*



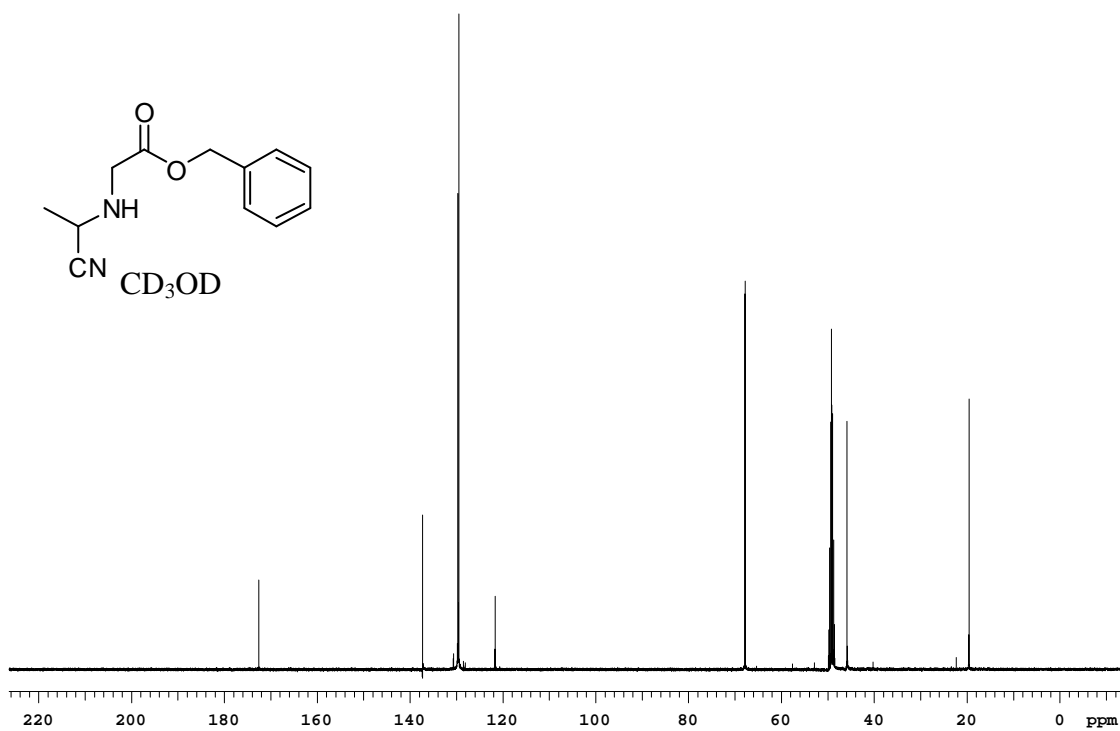
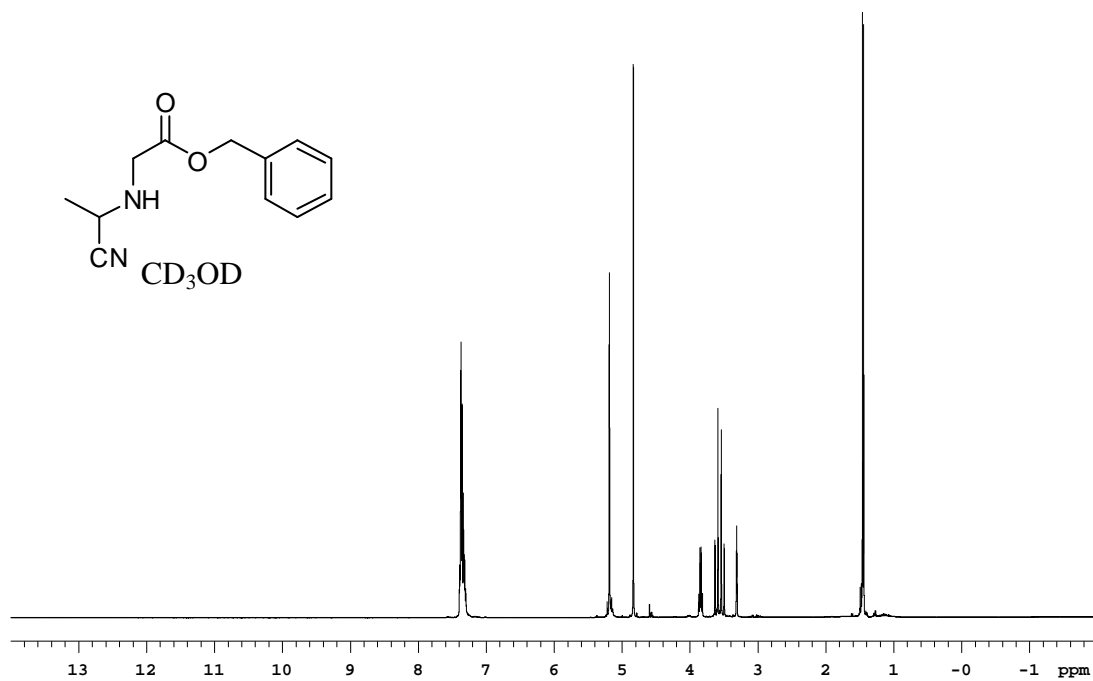
3i



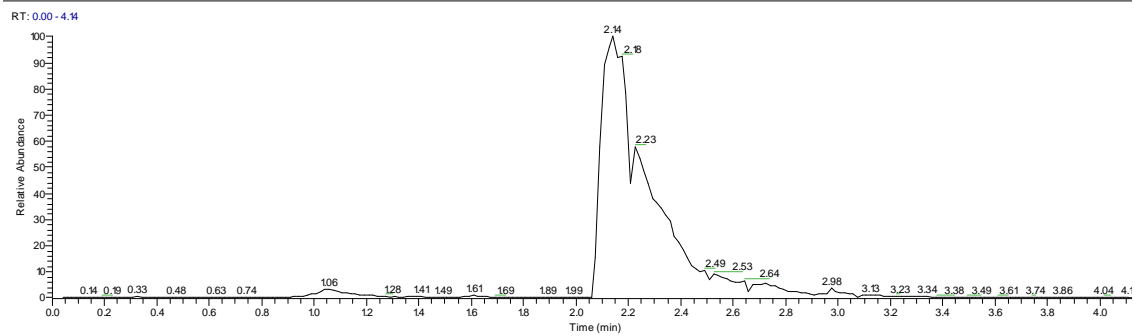
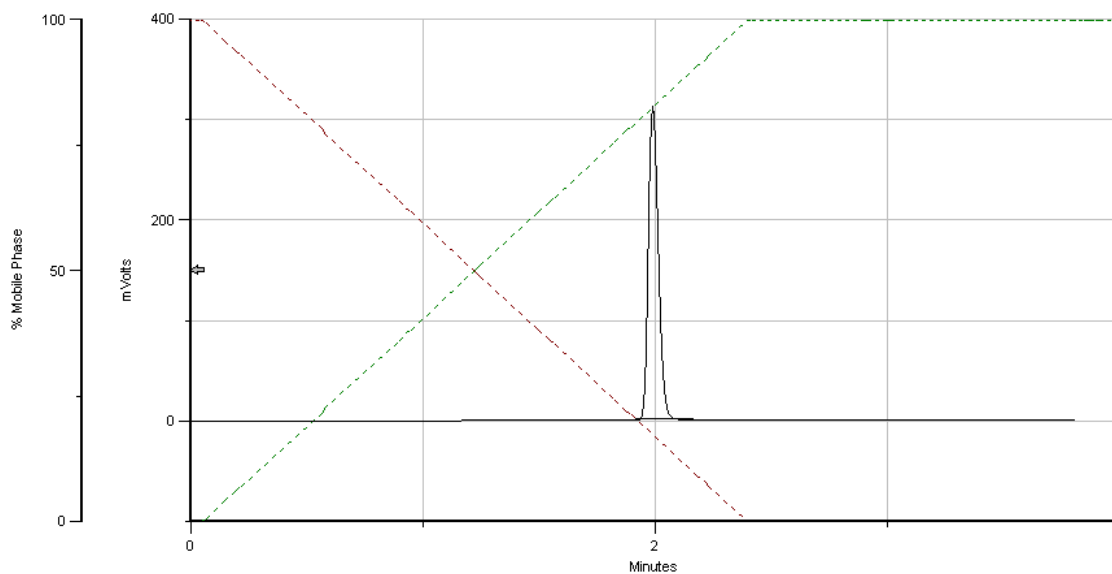
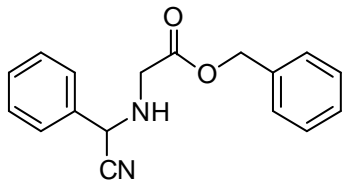
SQ_gis_3i #101 RT: 1.71 AV: 1 NL: 2.37E6
T: (0,0) + c ESI sid=10.00 Full ms [100.00-1200.00]



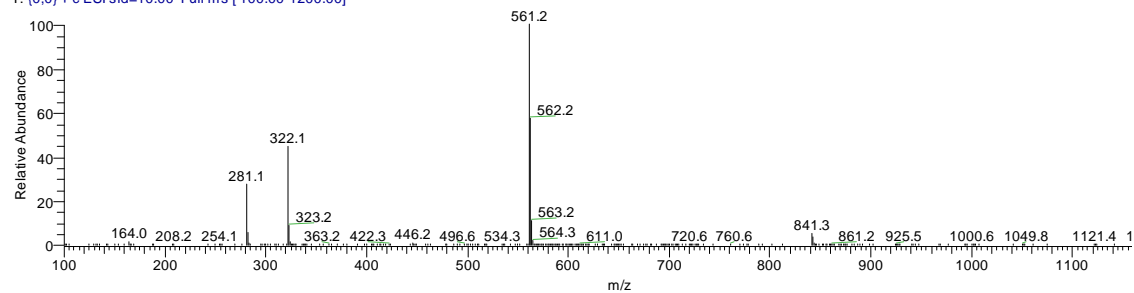
3i



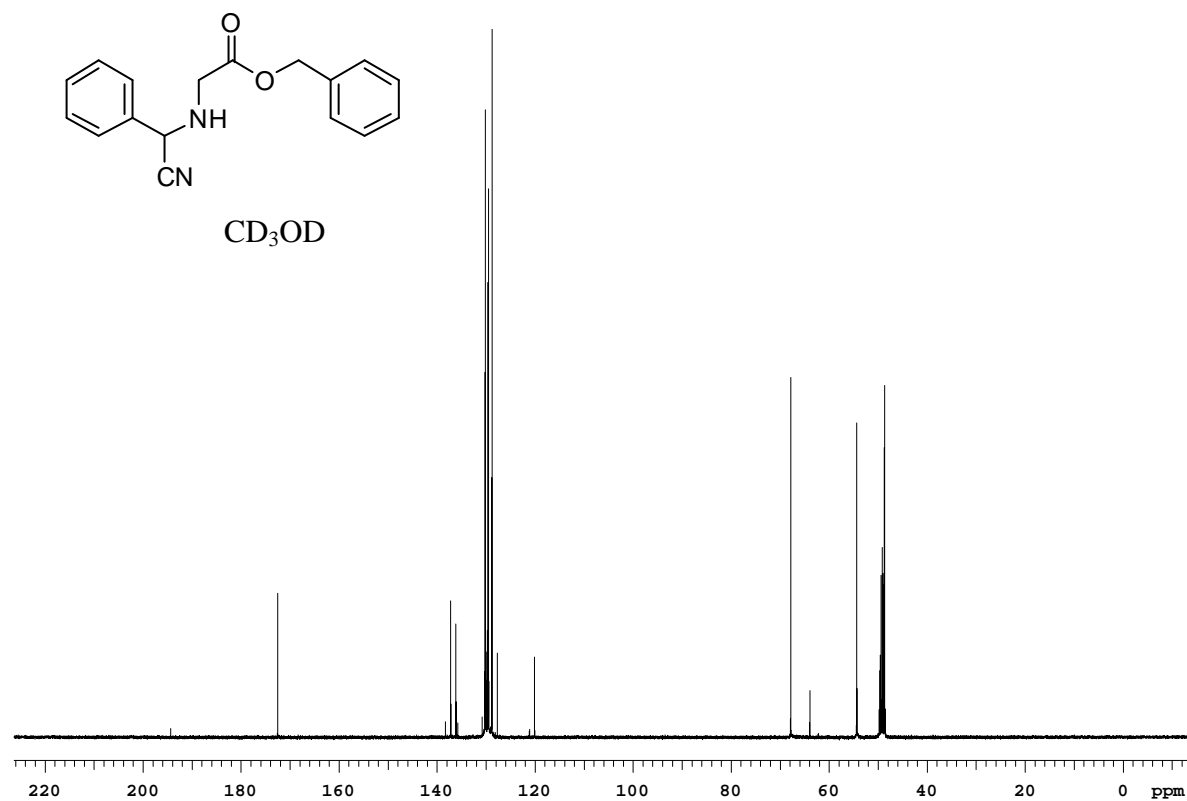
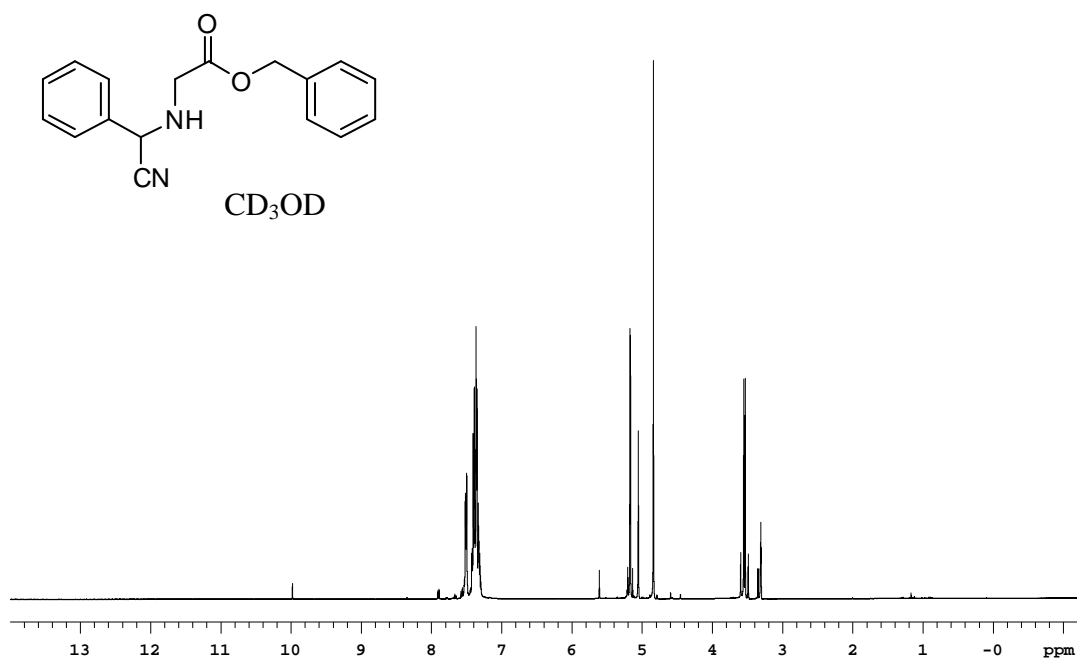
3j



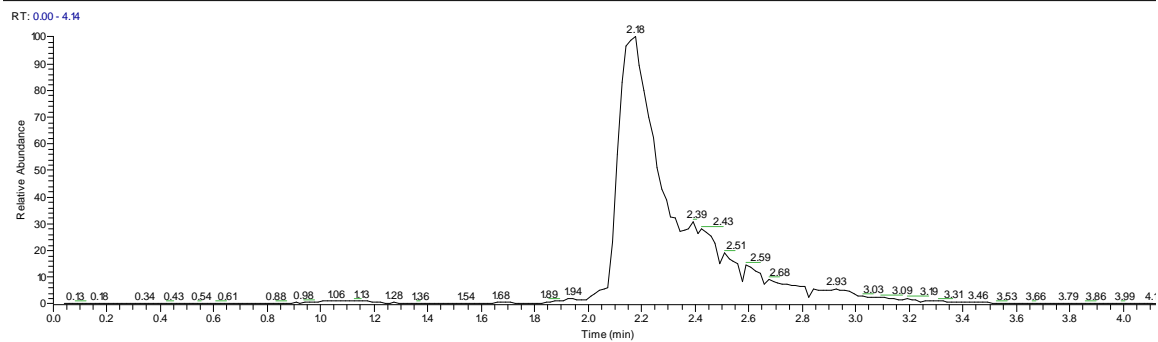
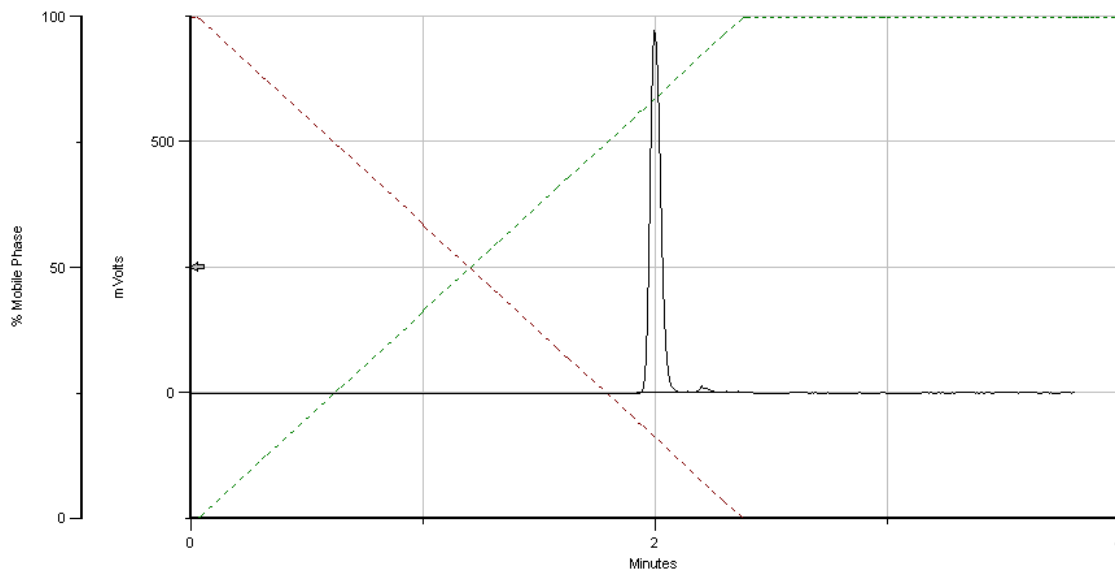
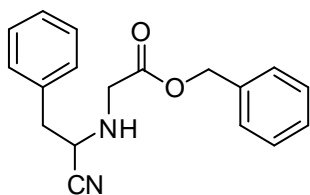
SQ_gis_3j #127 RT: 2.14 AV: 1 NL: 3.54E6
T: (0.0) + c ESI sid=10.00 Full ms [100.00-1200.00]



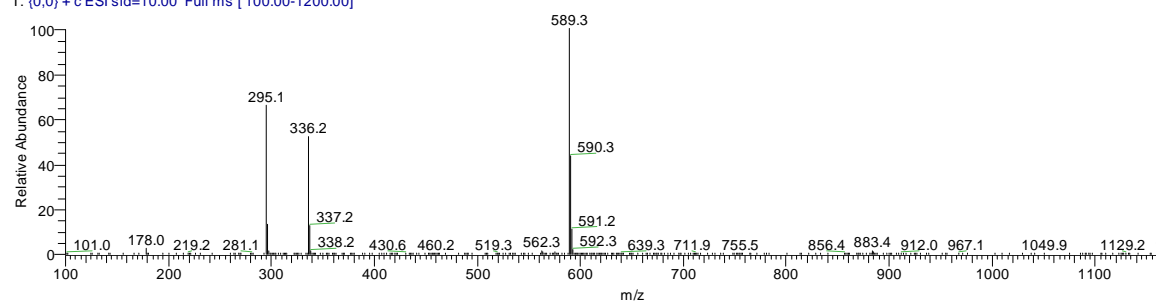
3j



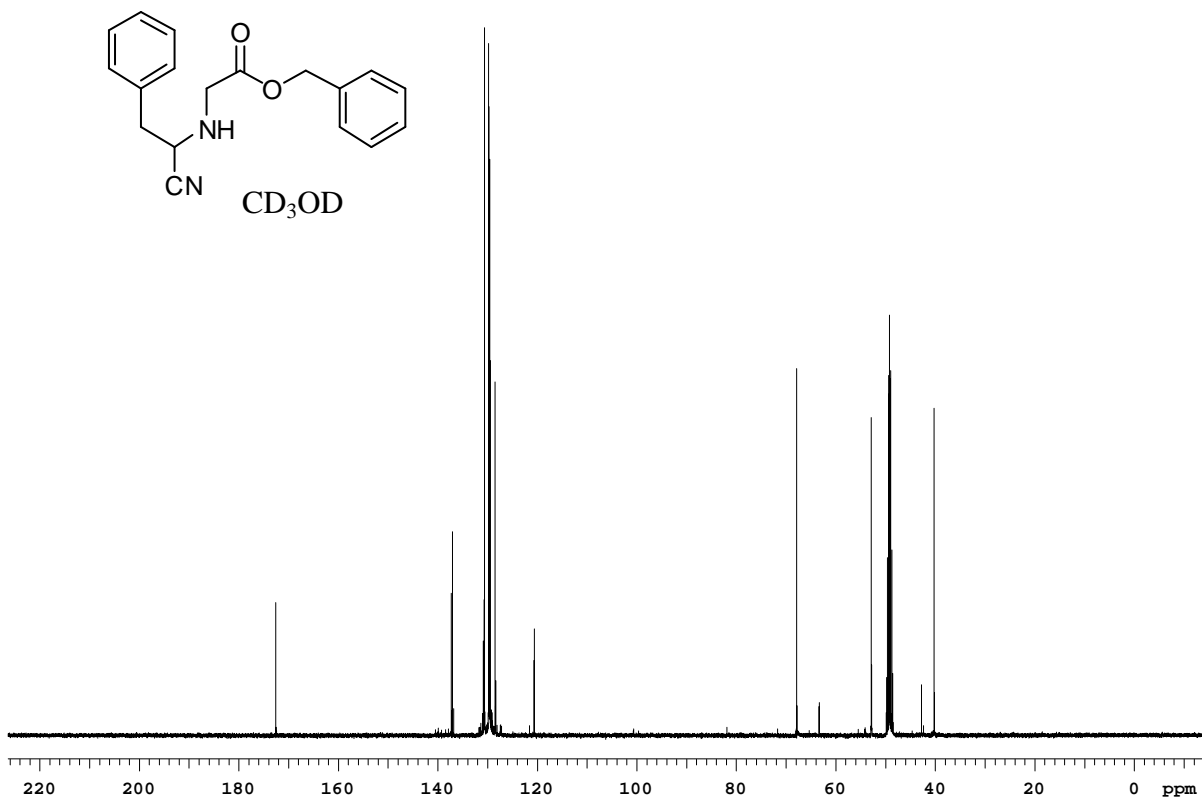
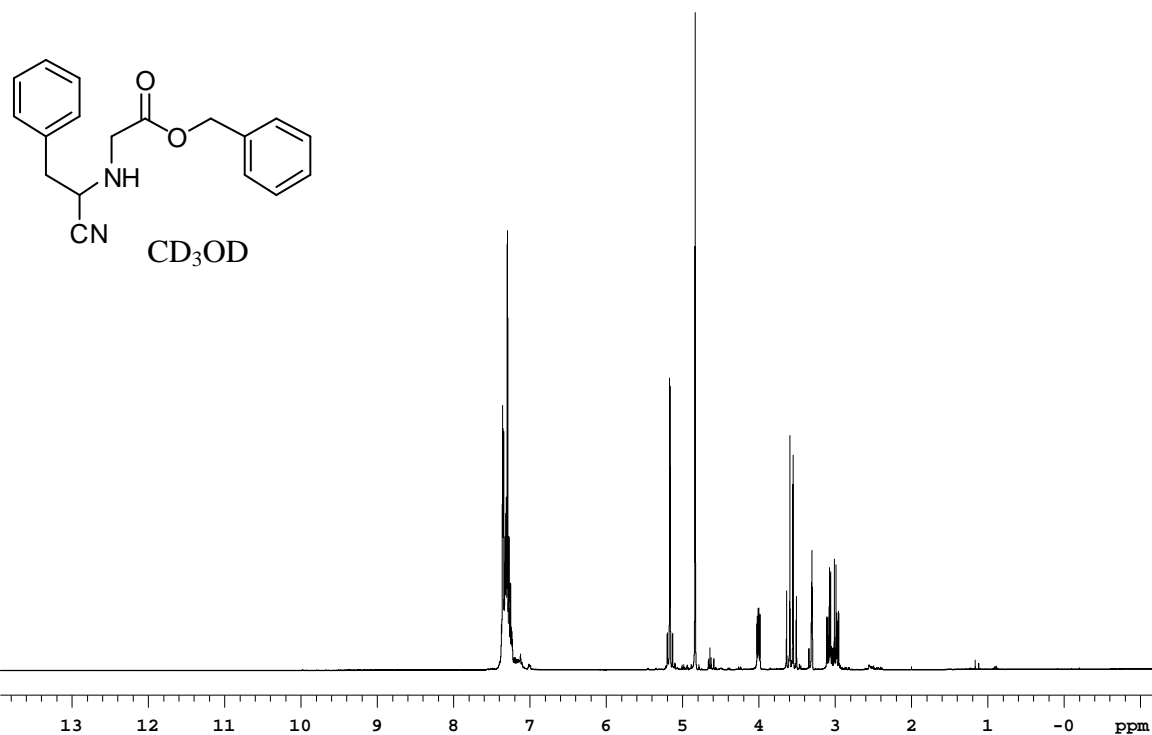
3k



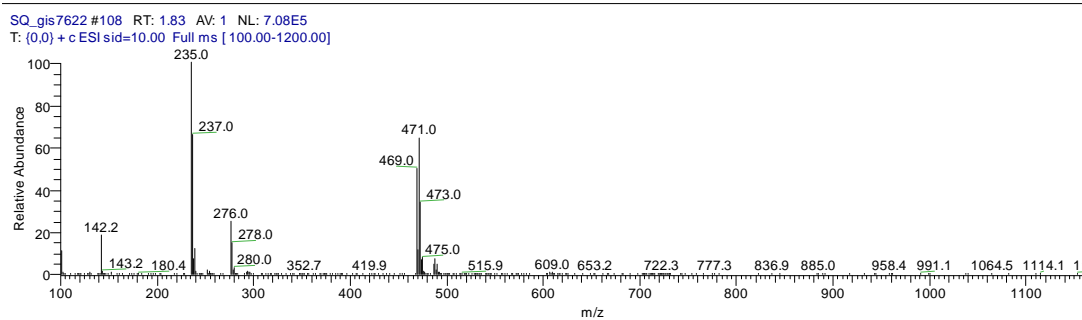
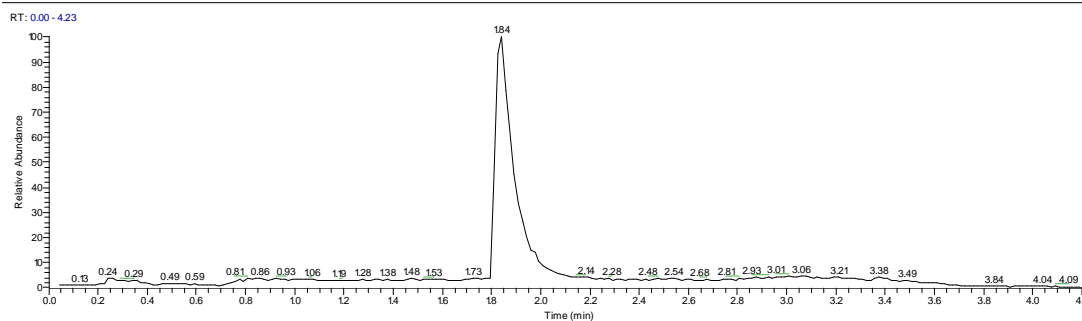
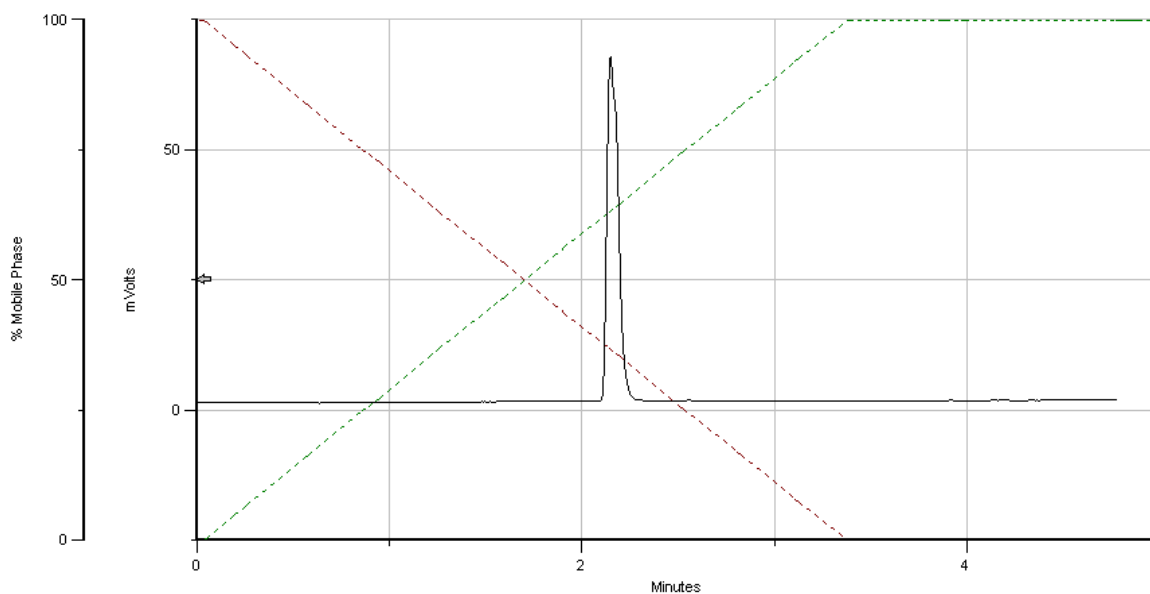
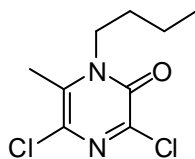
SQ_gis_3k #128 RT: 2.16 AV: 1 NL: 2.66E6
T: (0,0) + c ESI sid=10.00 Full ms [100.00-1200.00]



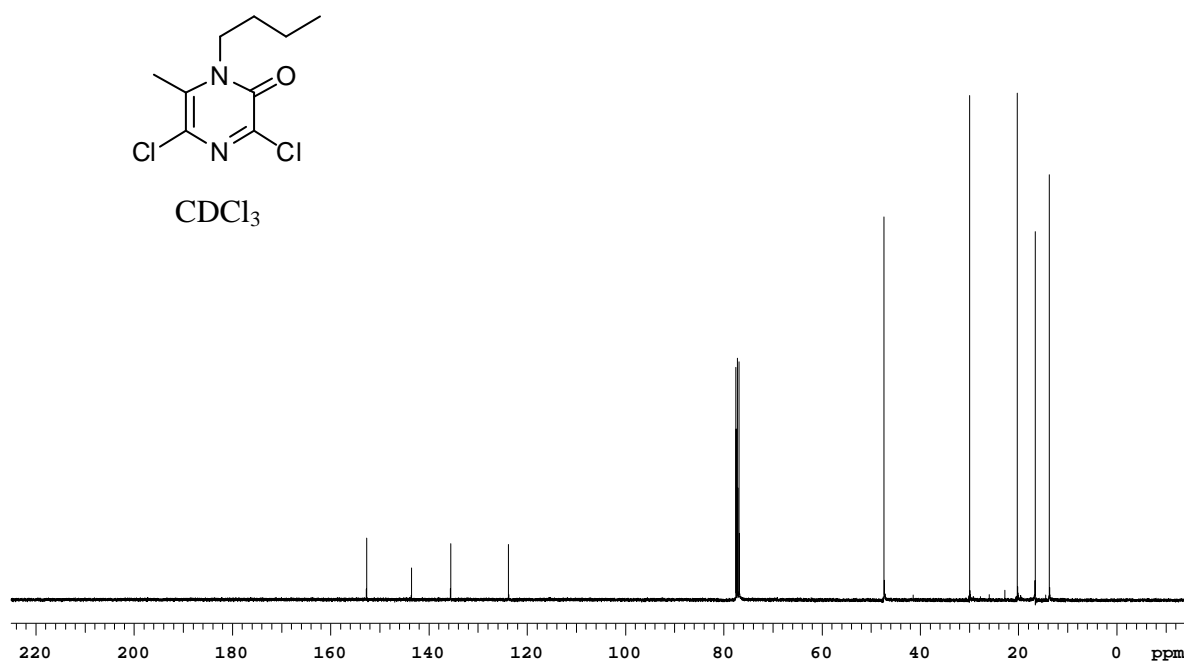
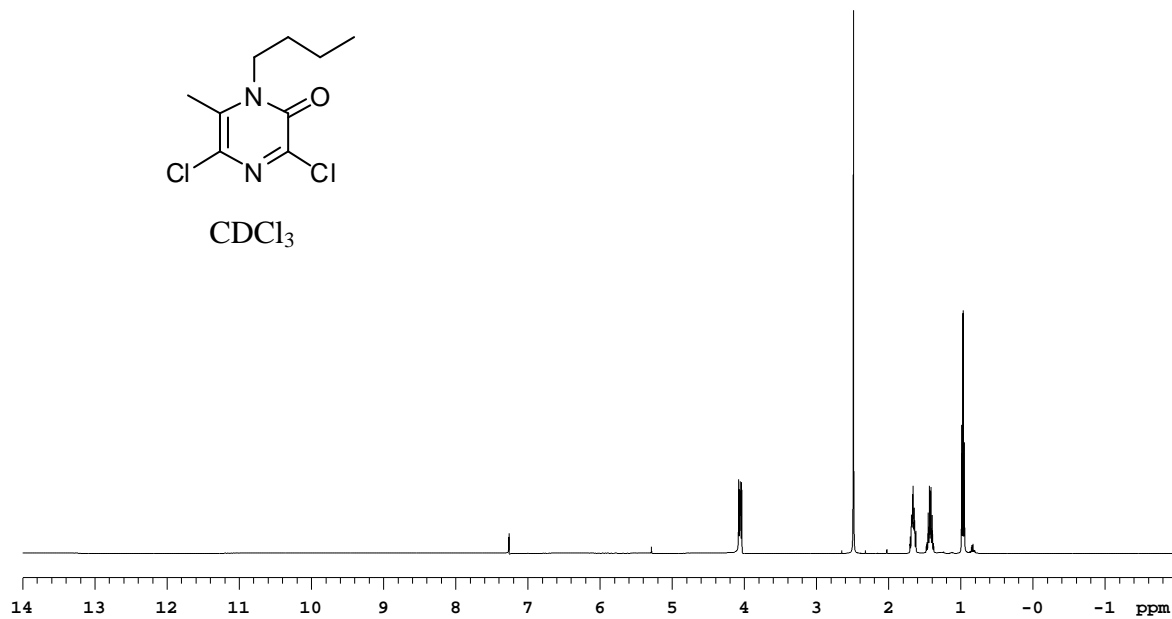
3k



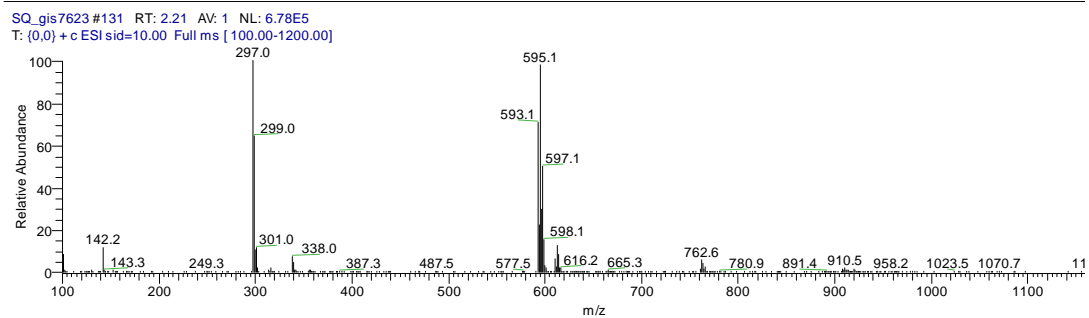
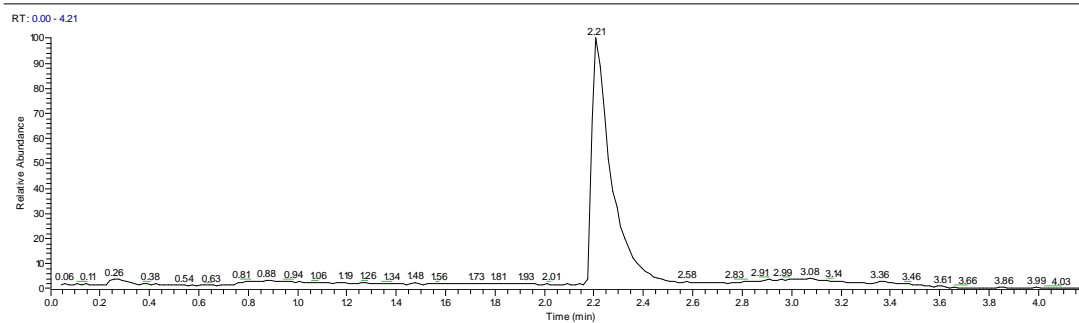
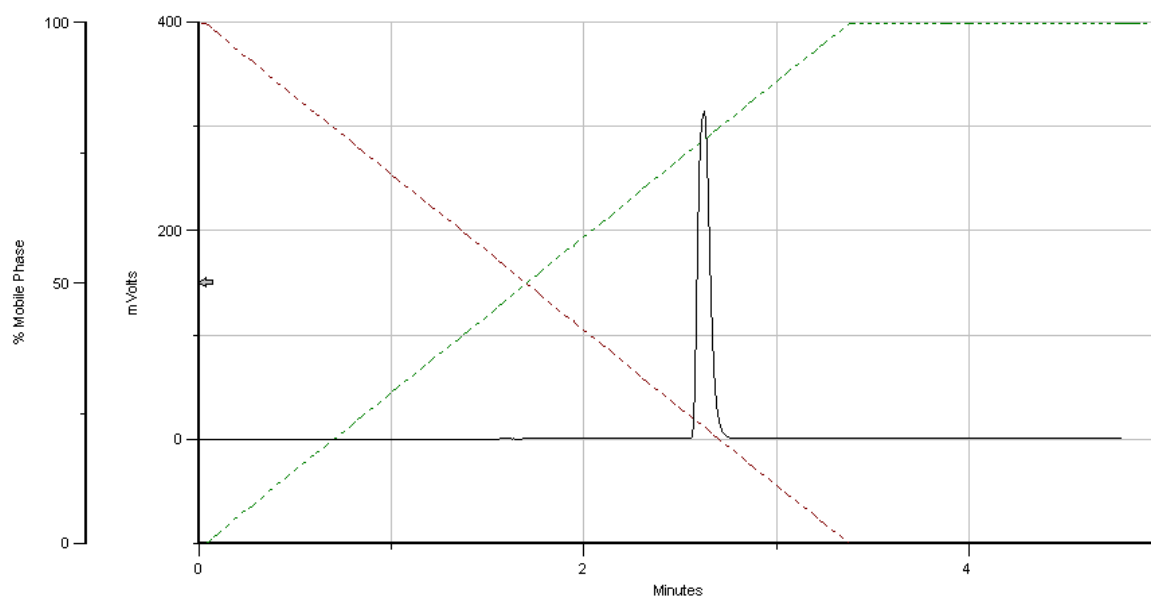
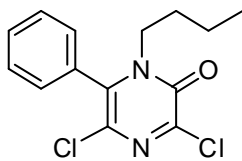
4a



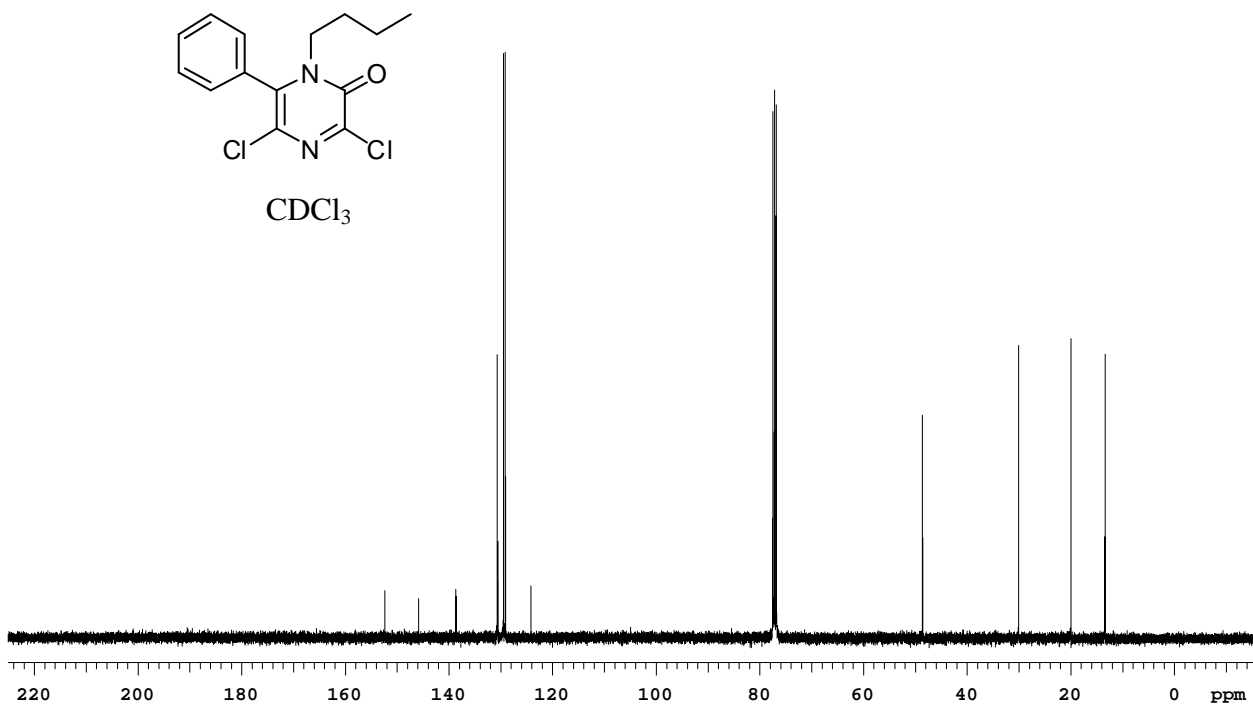
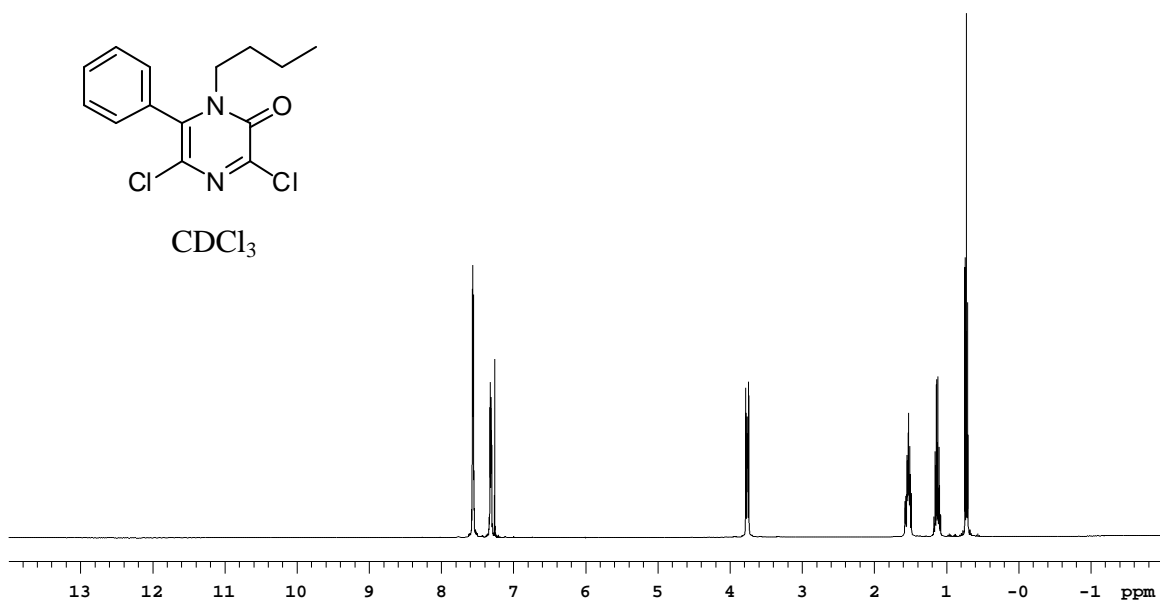
4a



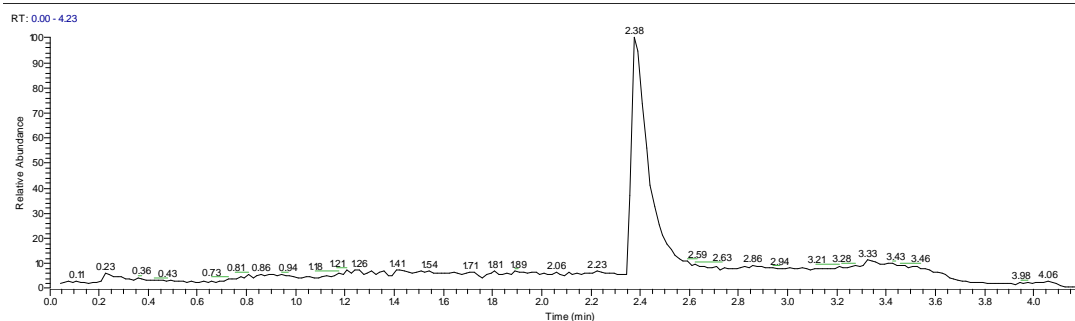
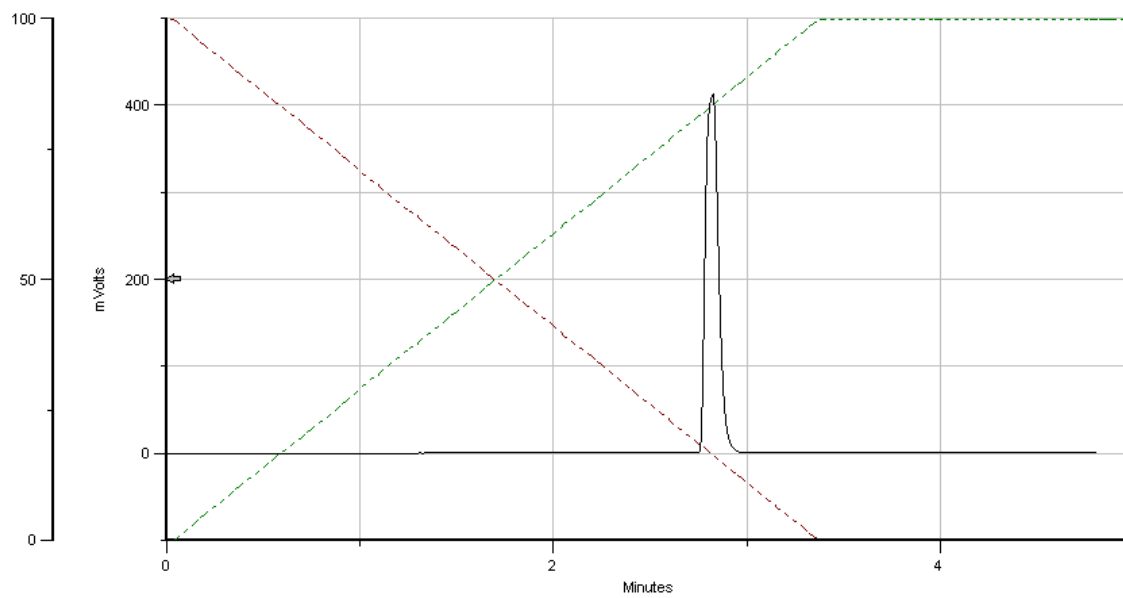
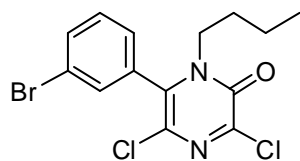
4b



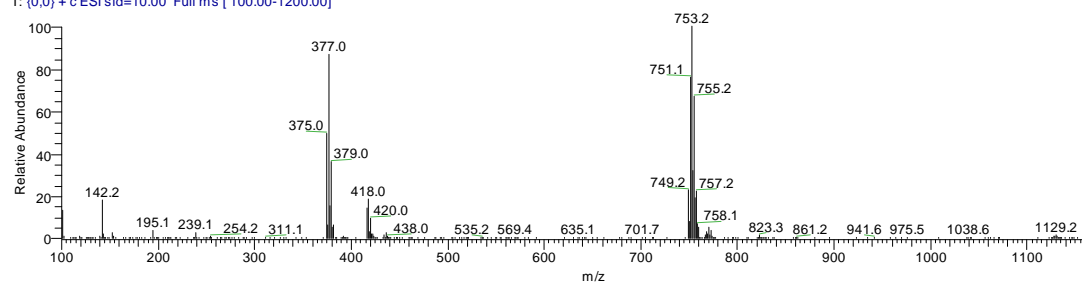
4b



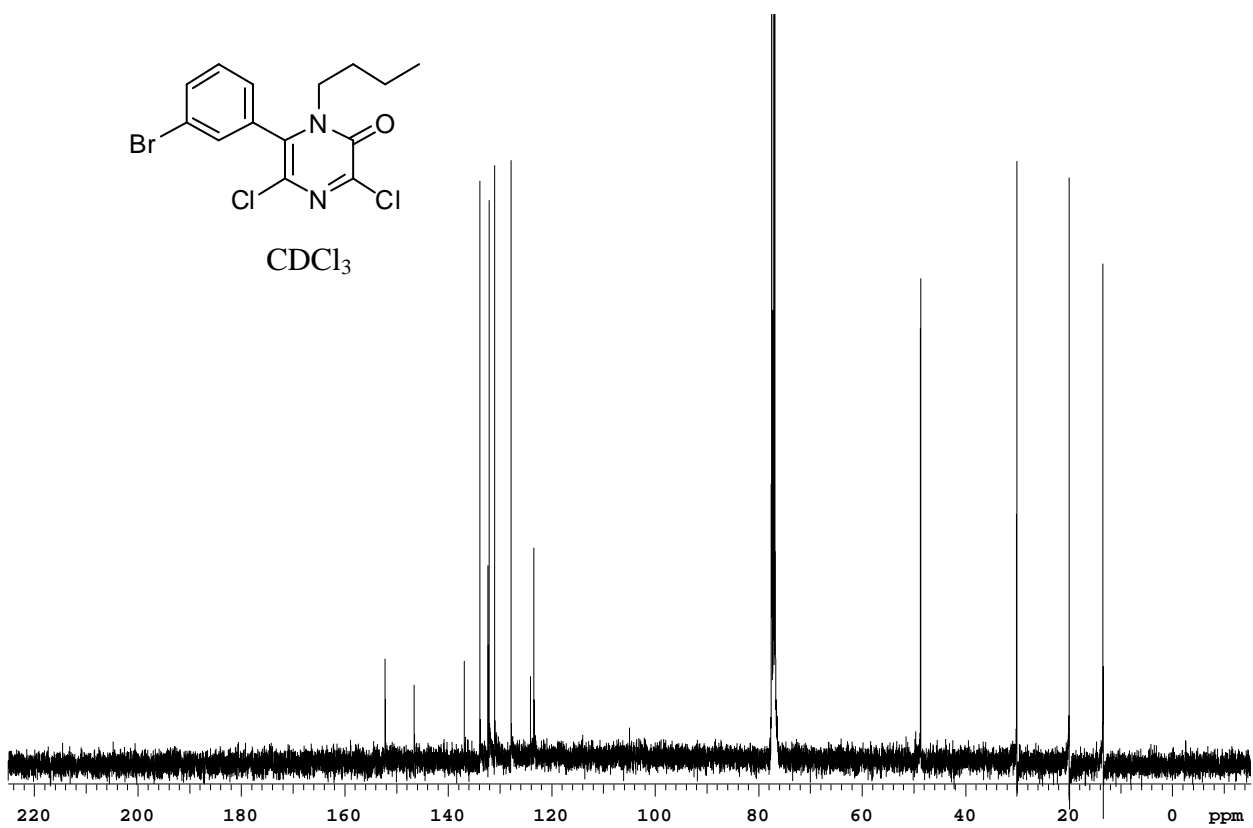
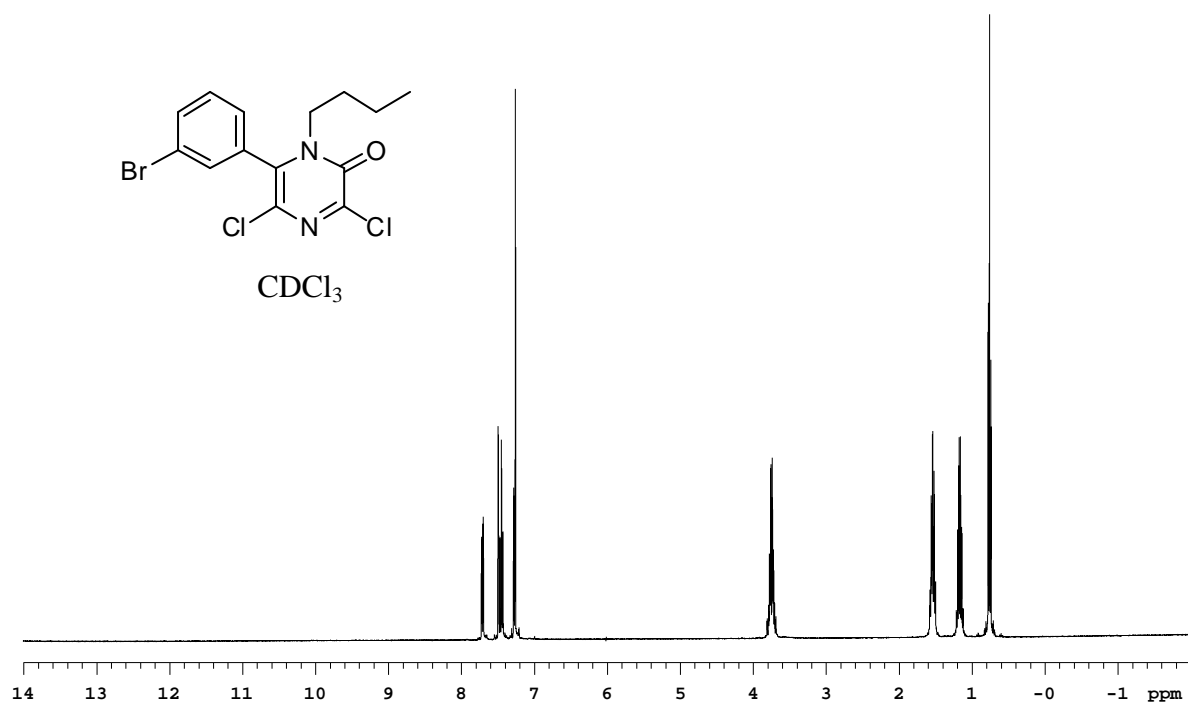
4c



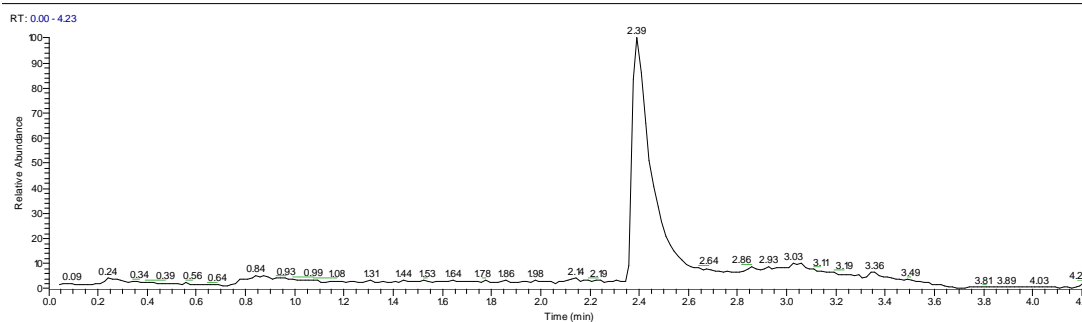
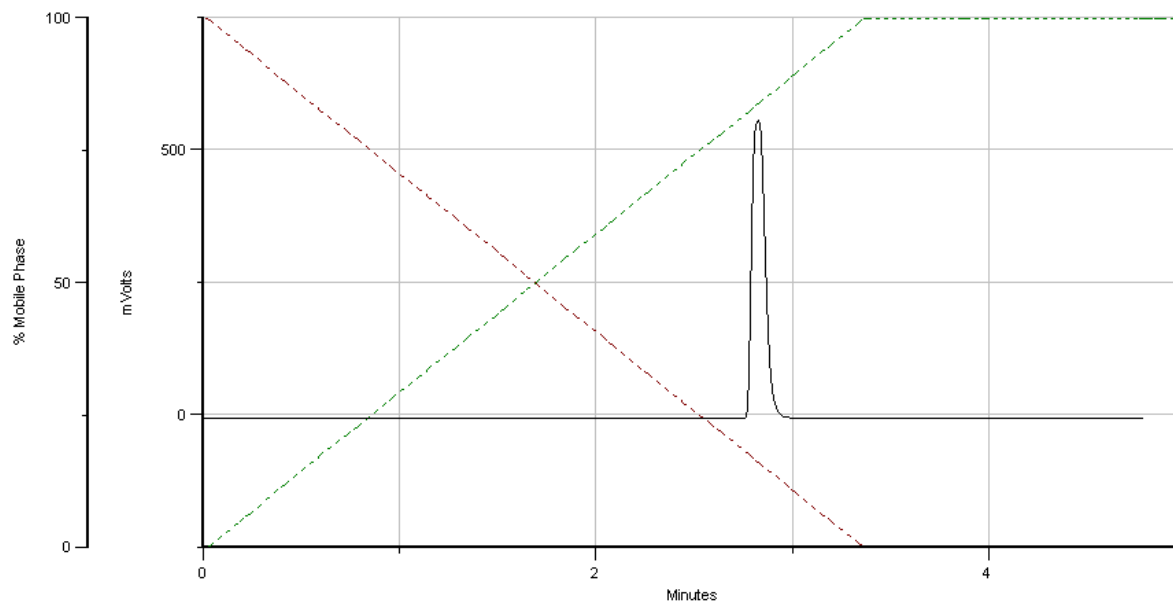
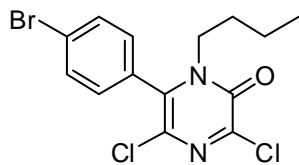
SQ_gis7627 #141 RT: 2.38 AV: 1 NL: 3.61E5
T: (0,0) + c ESI sid=10.00 Full ms [100.00-1200.00]



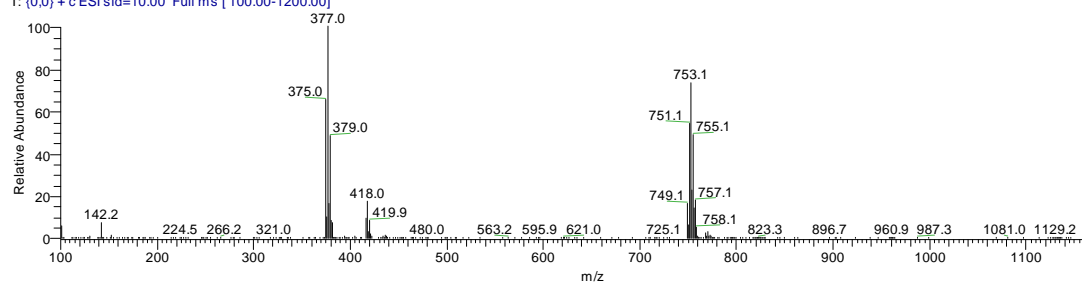
4c



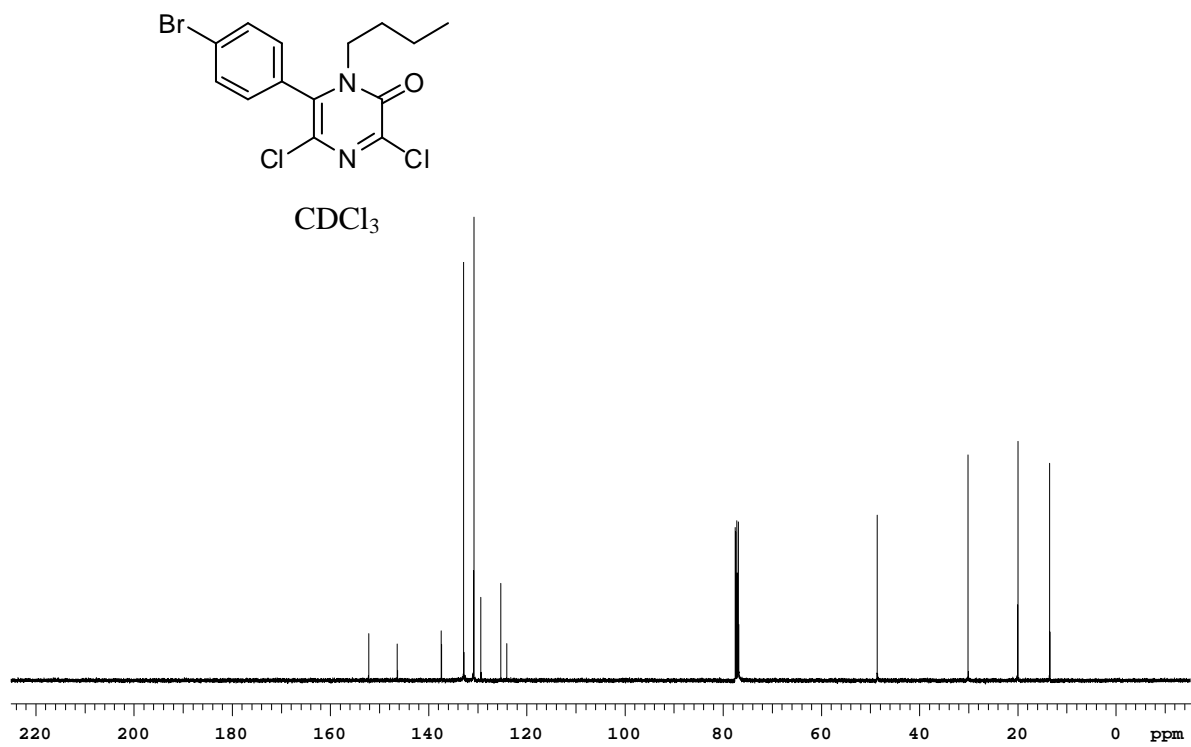
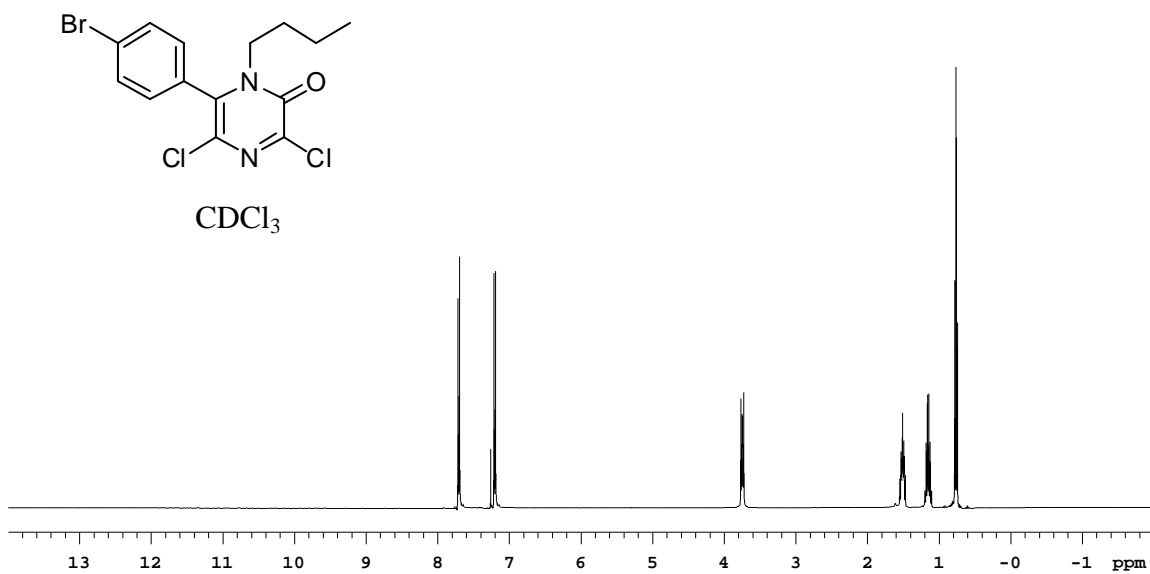
4d



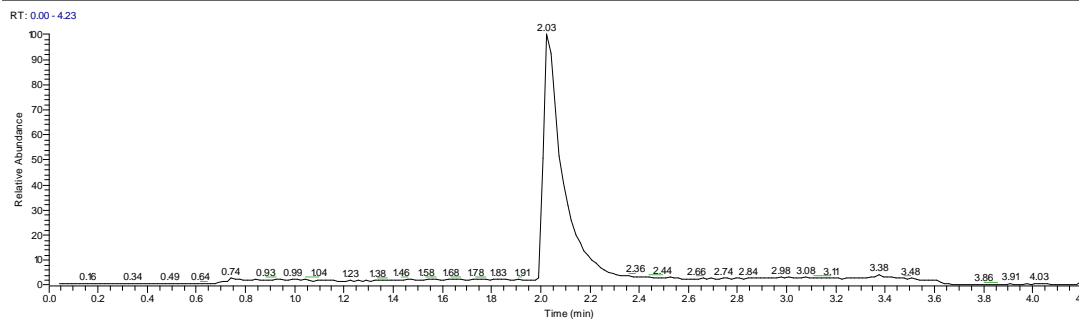
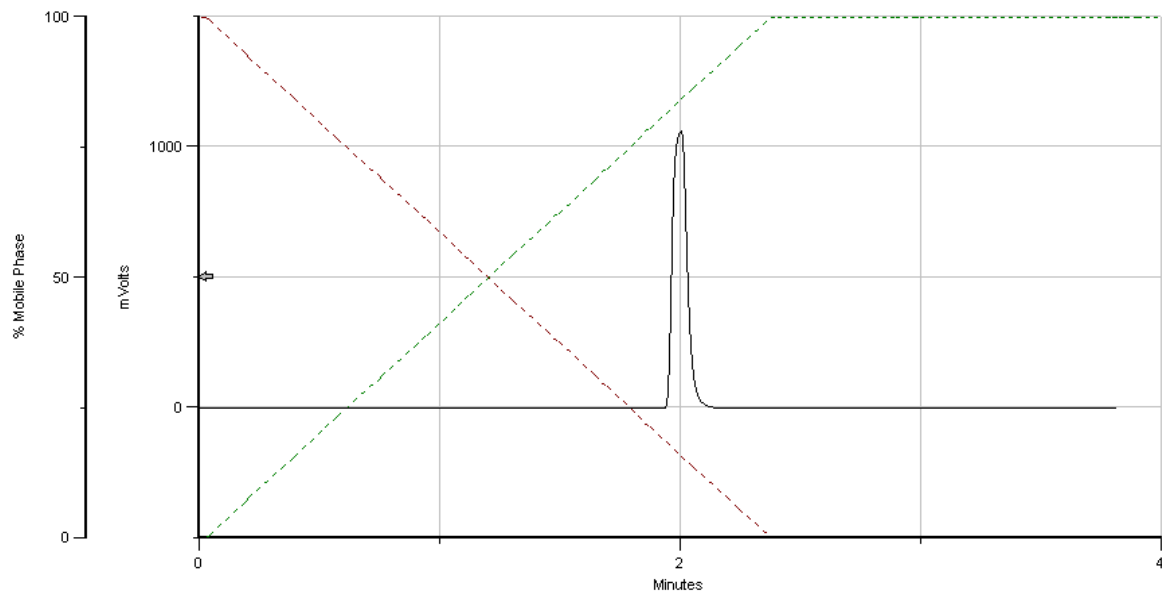
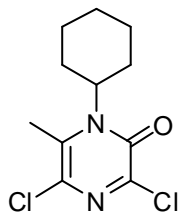
SQ_gis7628 #142 RT: 2.39 AV: 1 NL: 3.82E5
T: (0,0) + c ESI sid=10.00 Full ms [100.00-1200.00]



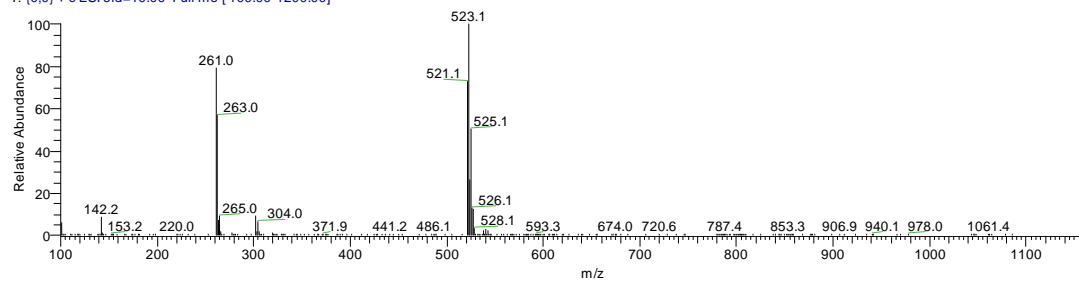
4d



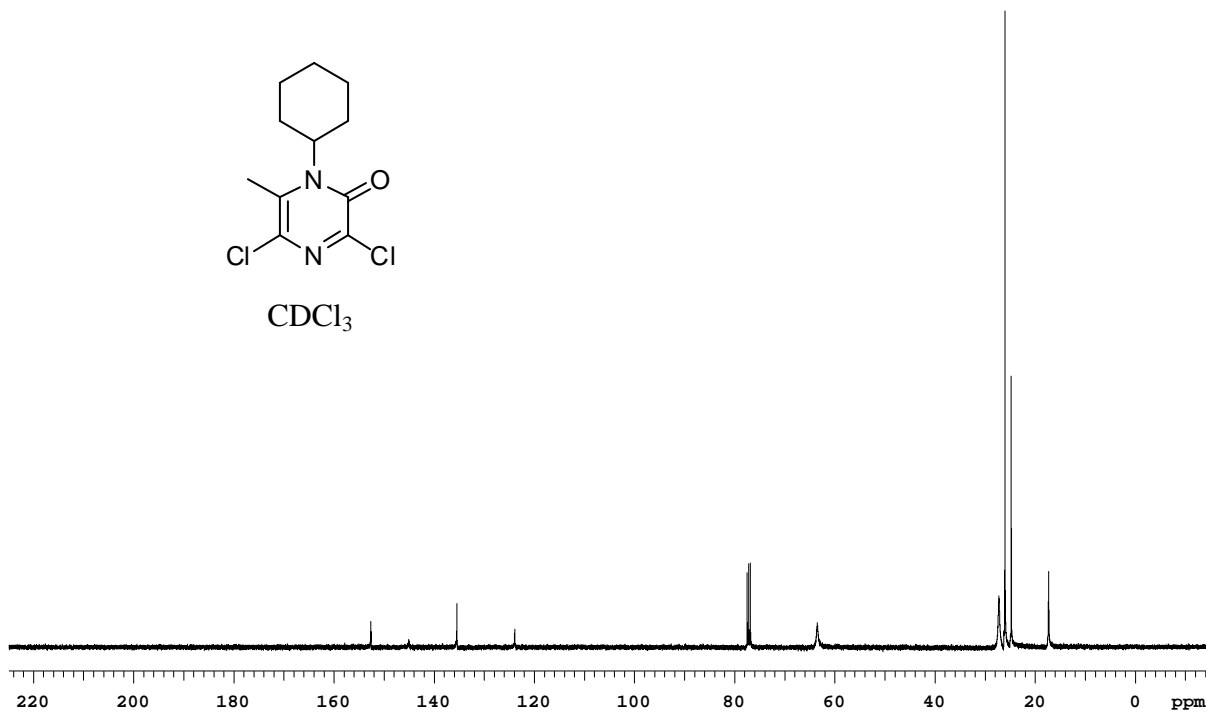
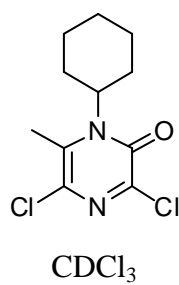
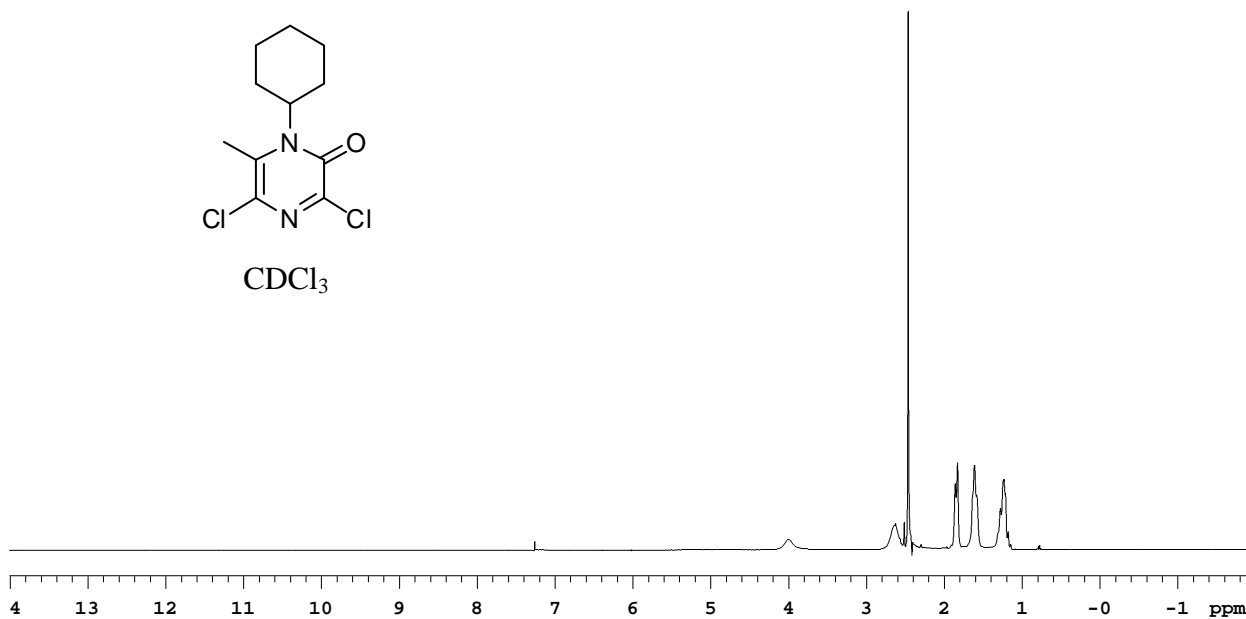
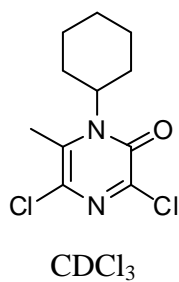
4e



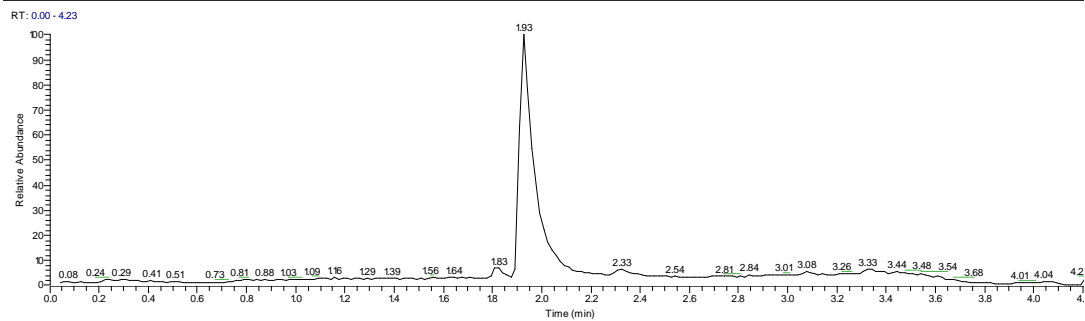
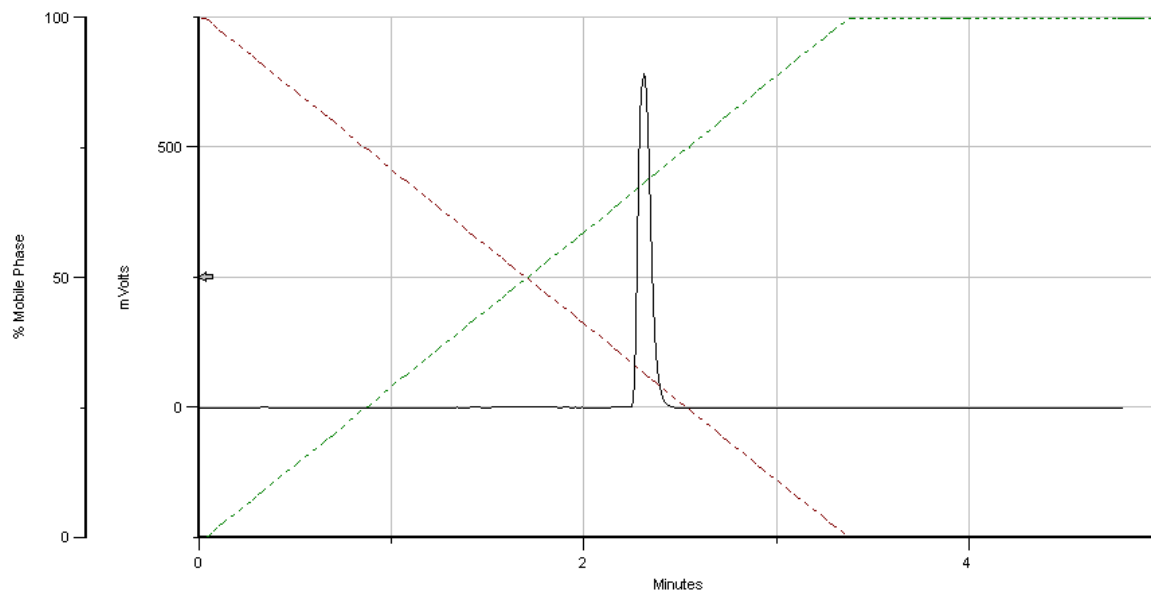
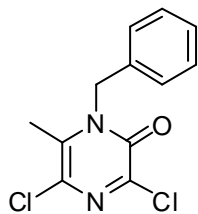
SQ_gis7630 #120 RT: 2.03 Av: 1 NL: 1.22E6
T: (0.0) + c ESI: sid=10.00 Full ms [100.00-1200.00]



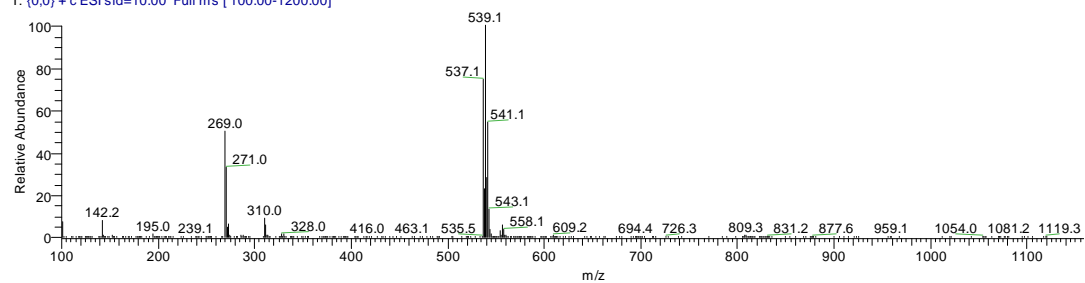
4e



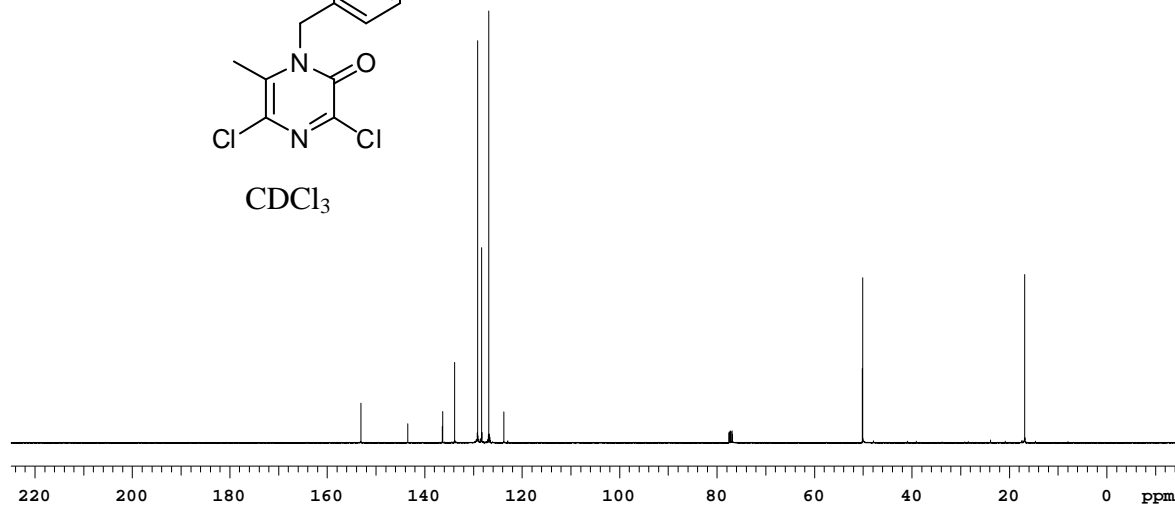
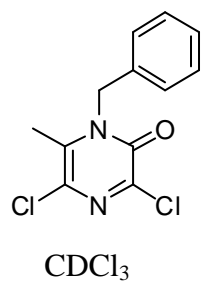
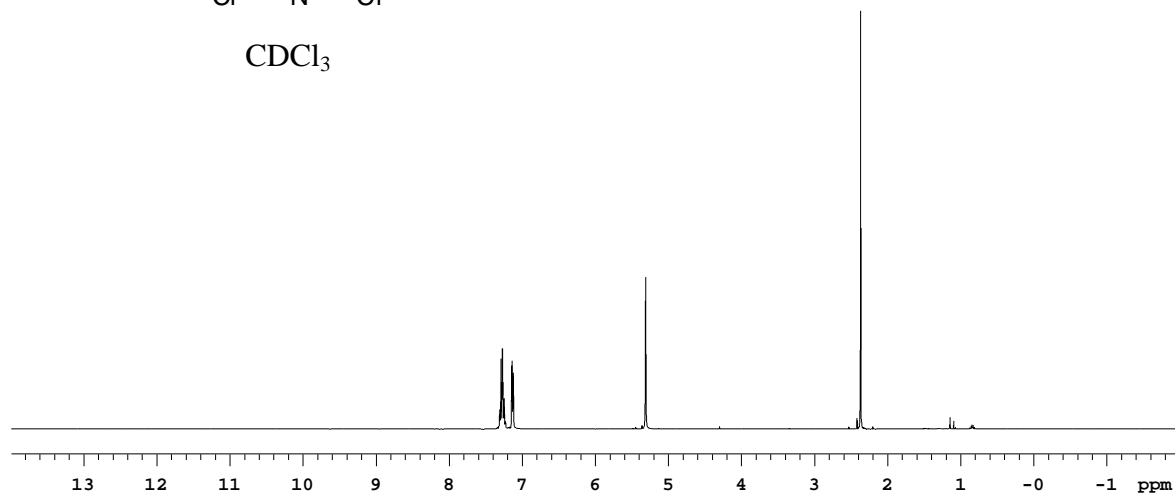
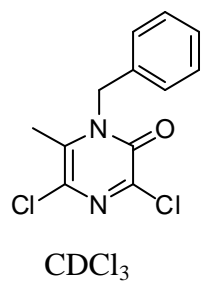
4f



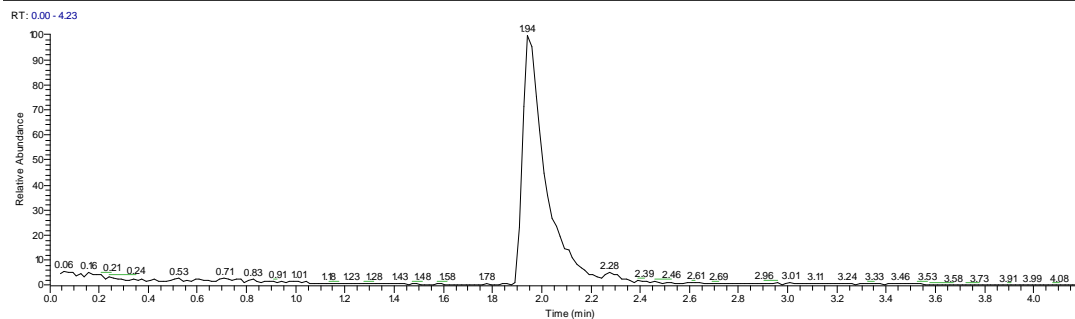
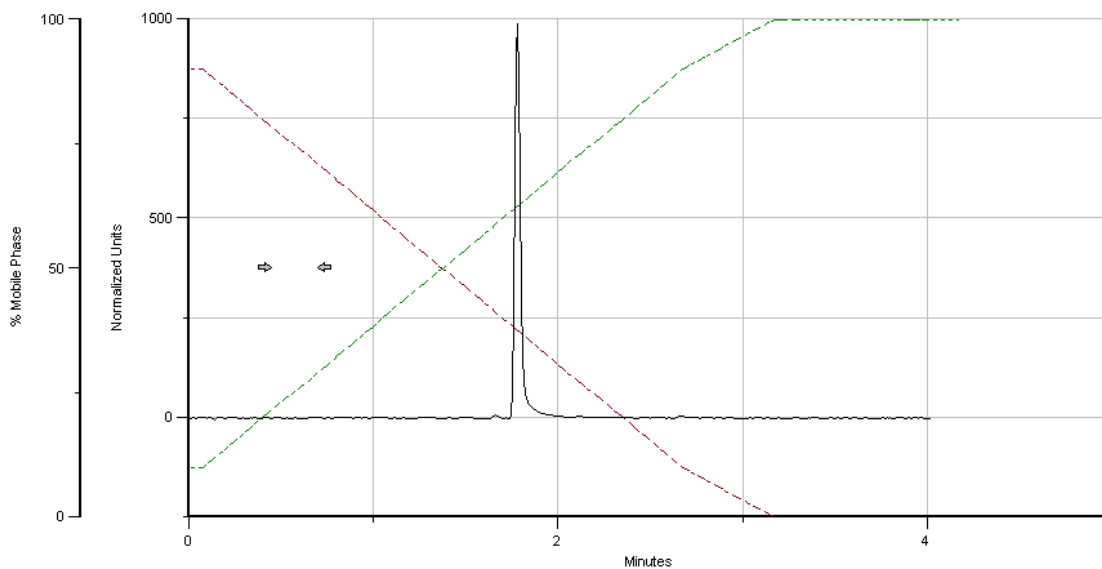
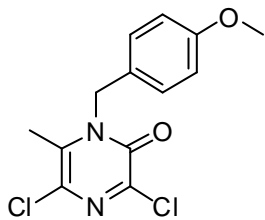
SQ_gis7629 #114 RT: 1.93 AV: 1 NL: 1.00E6
T: (0,0) + c ESI: sid=10.00 Full ms [100.00-1200.00]



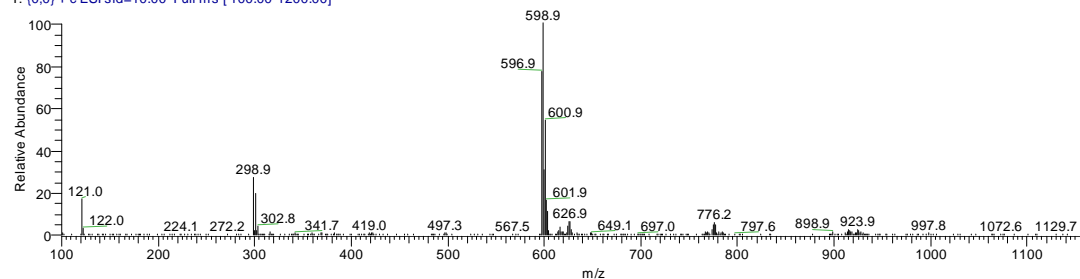
4f



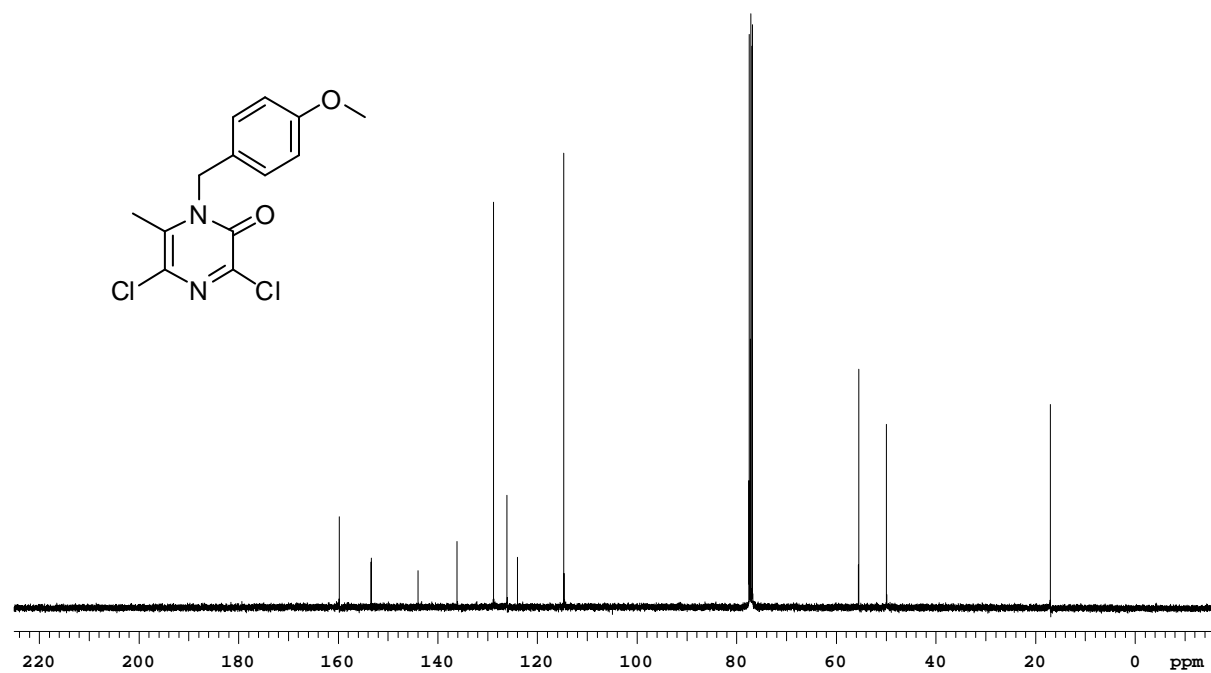
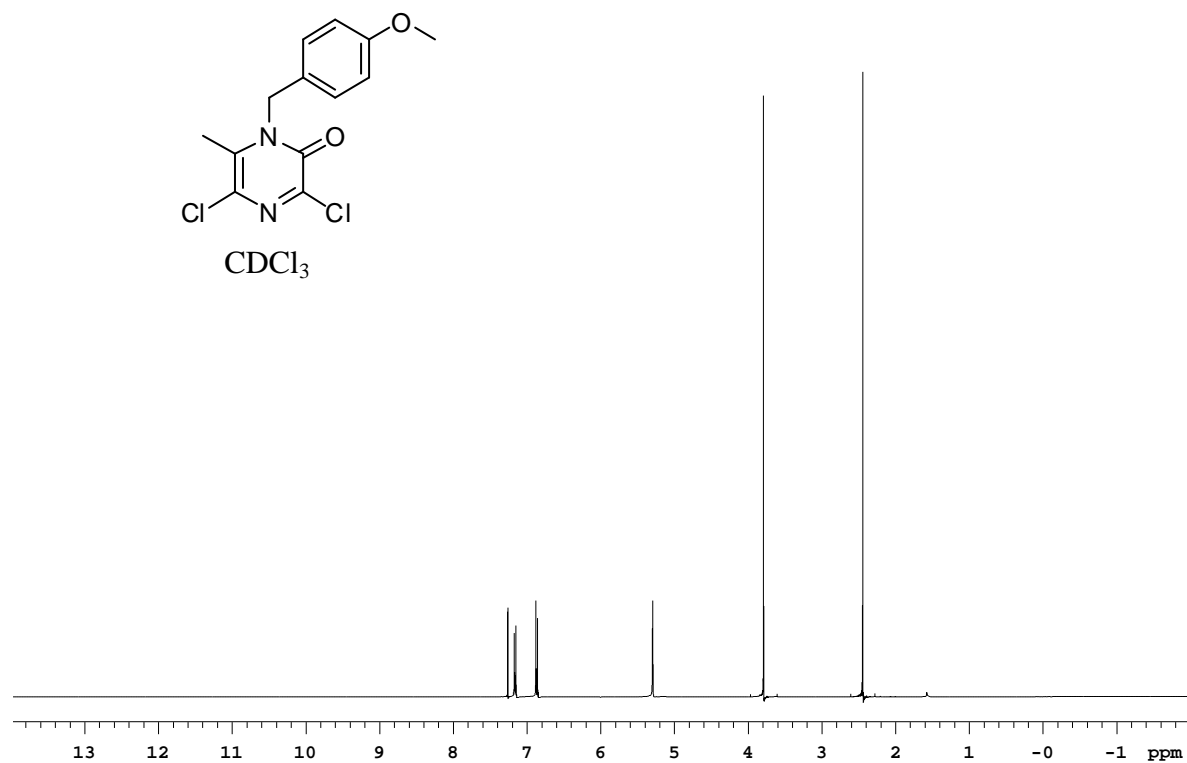
4g



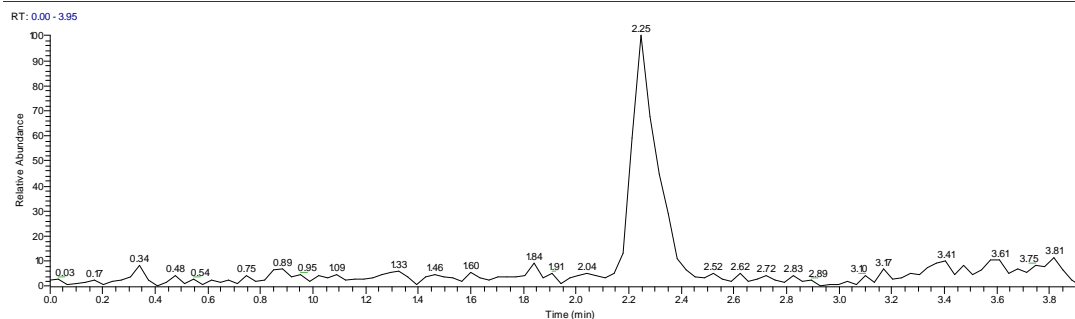
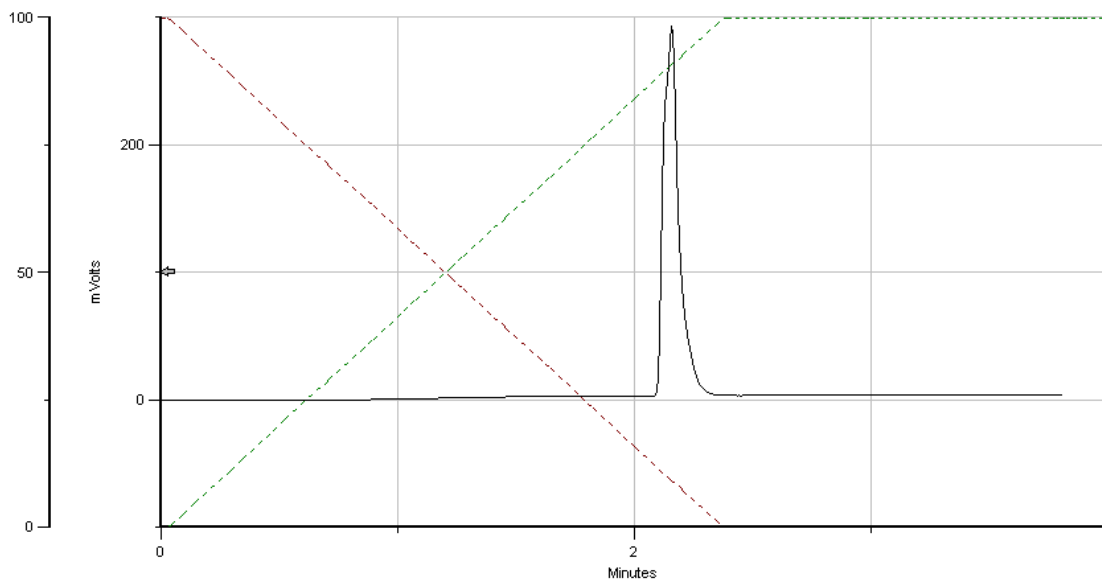
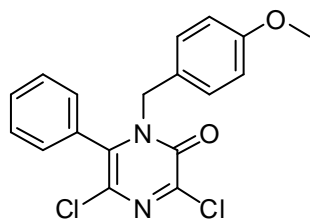
SQ_gis187_iso1 #115 RT: 1.94 AV: 1 NL: 3.15E5
T: (0.0) + c ESI sid=10.00 Full ms [100.00-1200.00]



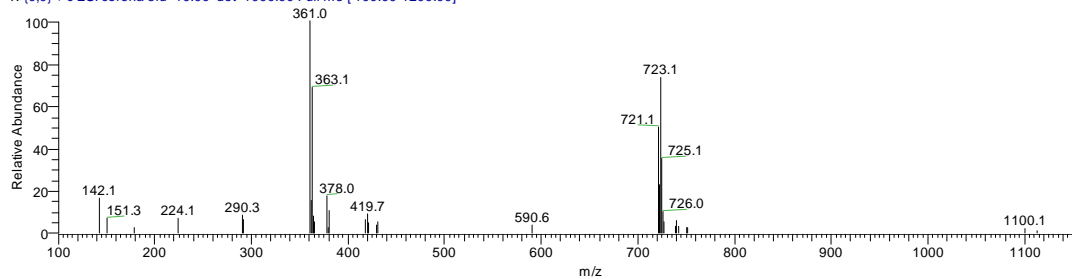
4g



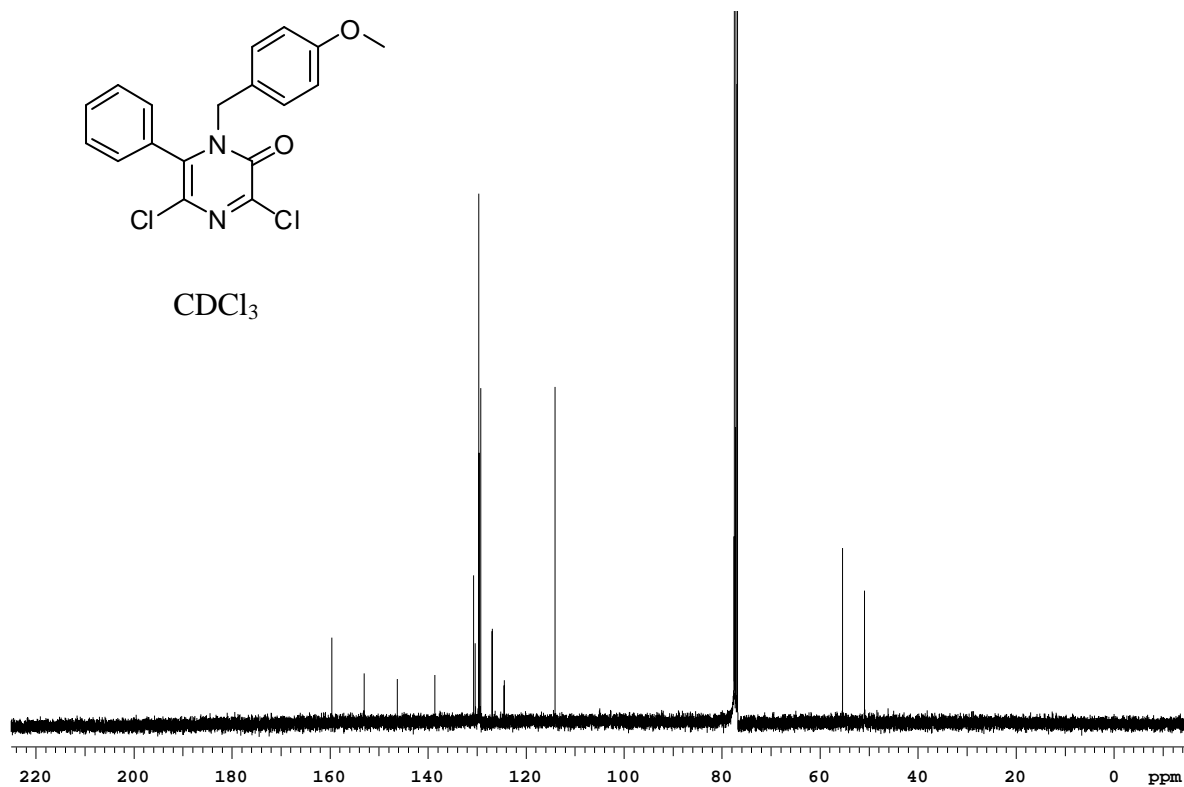
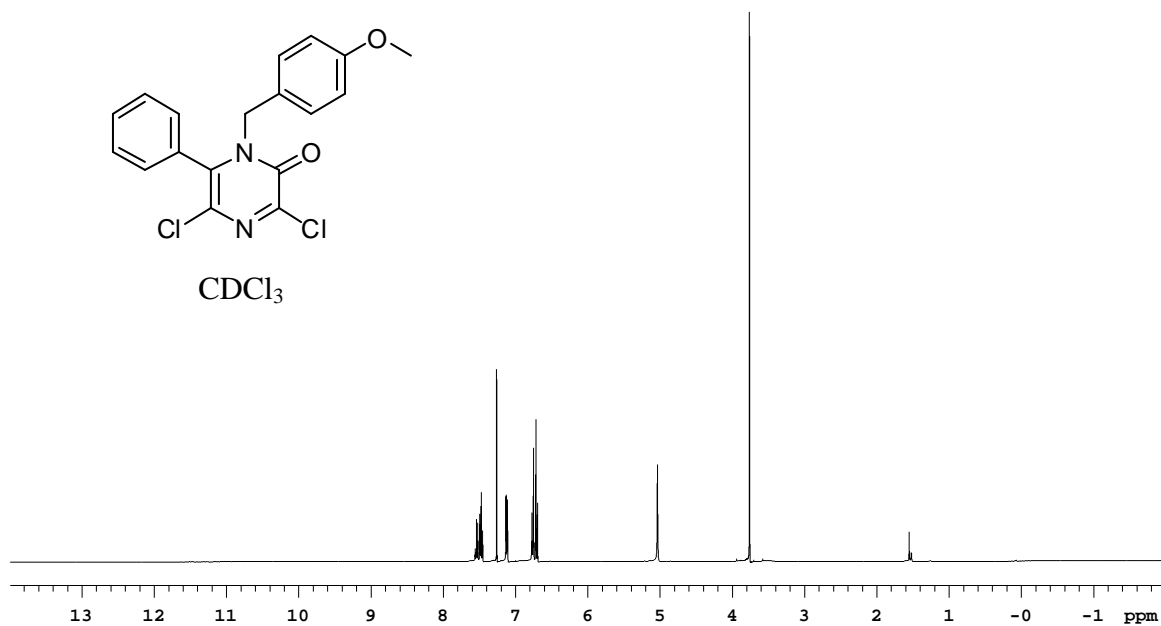
4h



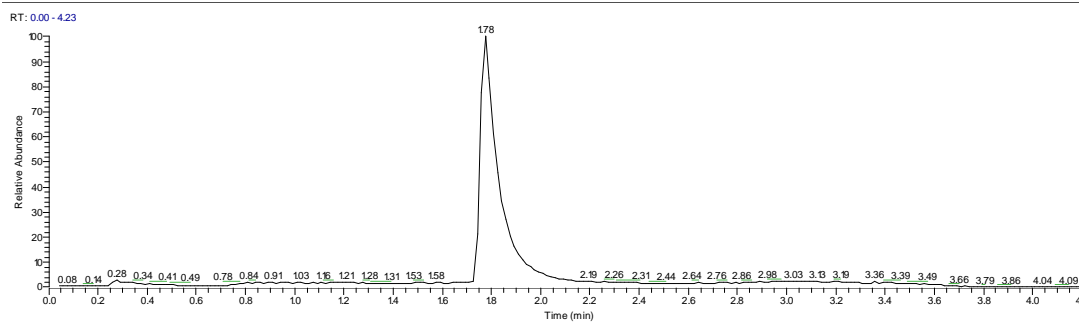
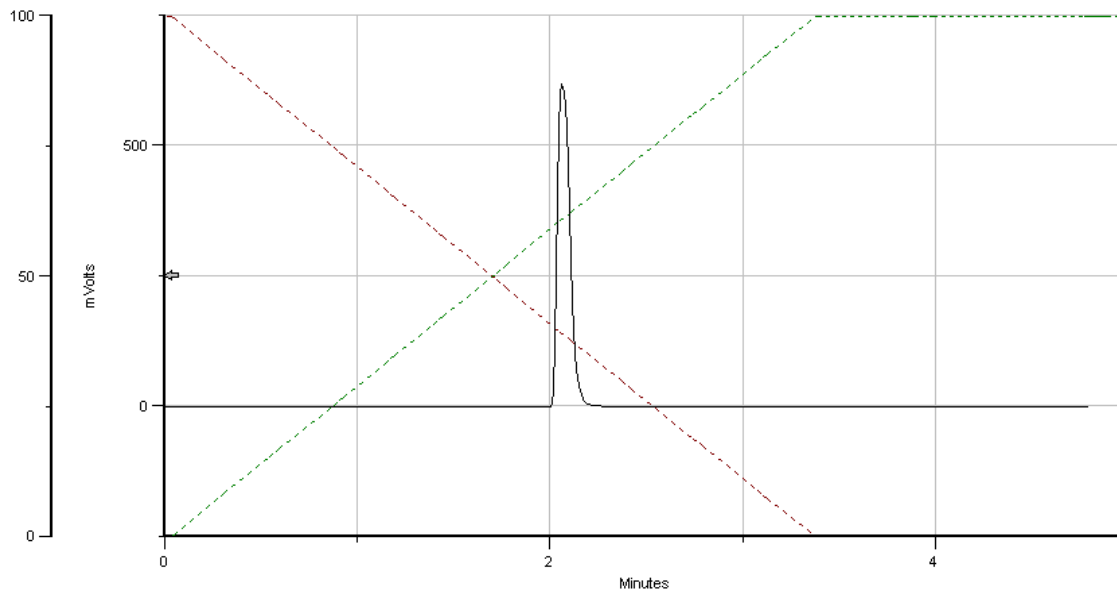
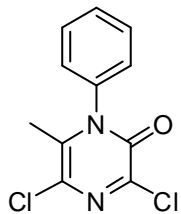
SQ_gis000186_iso #67 RT: 2.25 AV: 1 NL: 2.33E4
T: (0,0) + c ESI corona sid=10.00 det=1000.00 Full ms [100.00-1200.00]



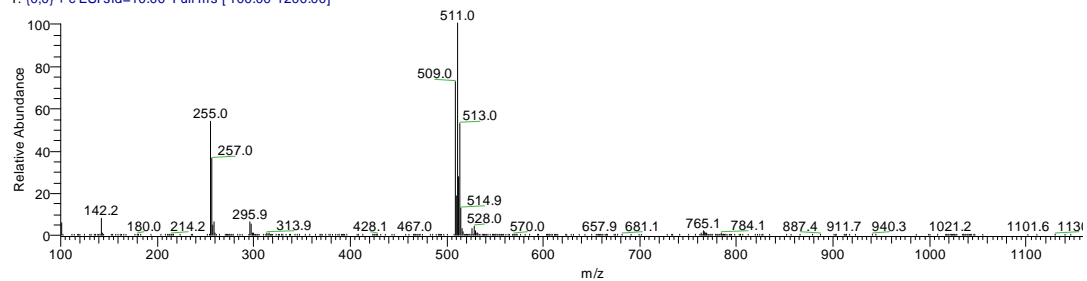
4h



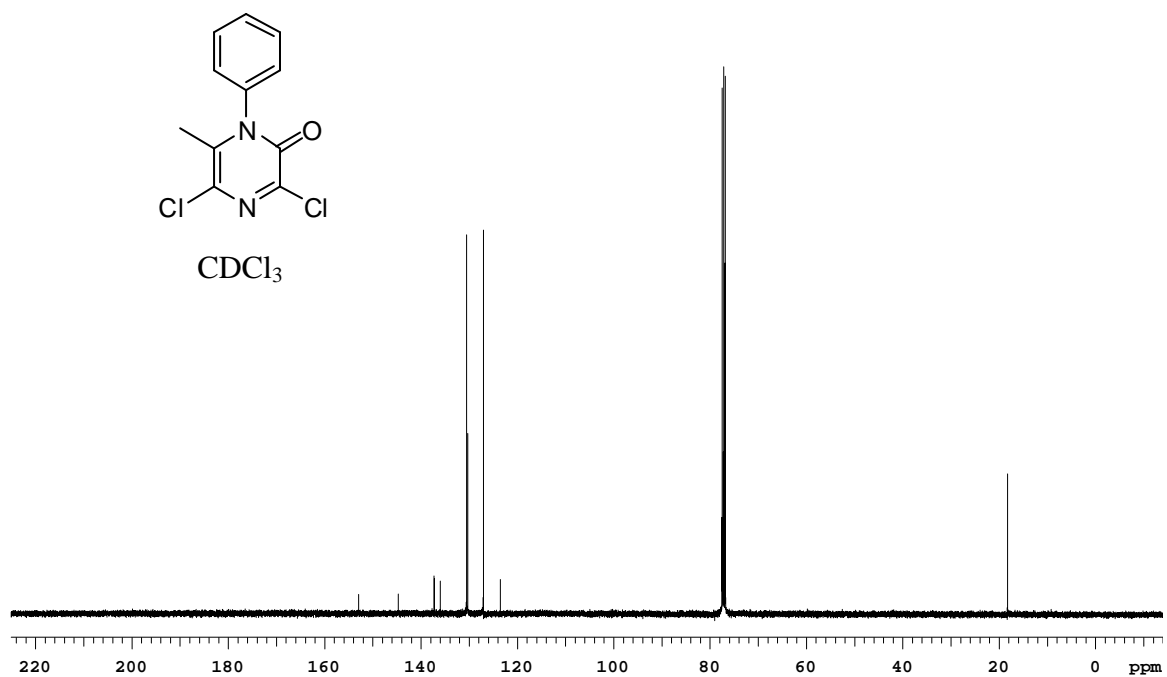
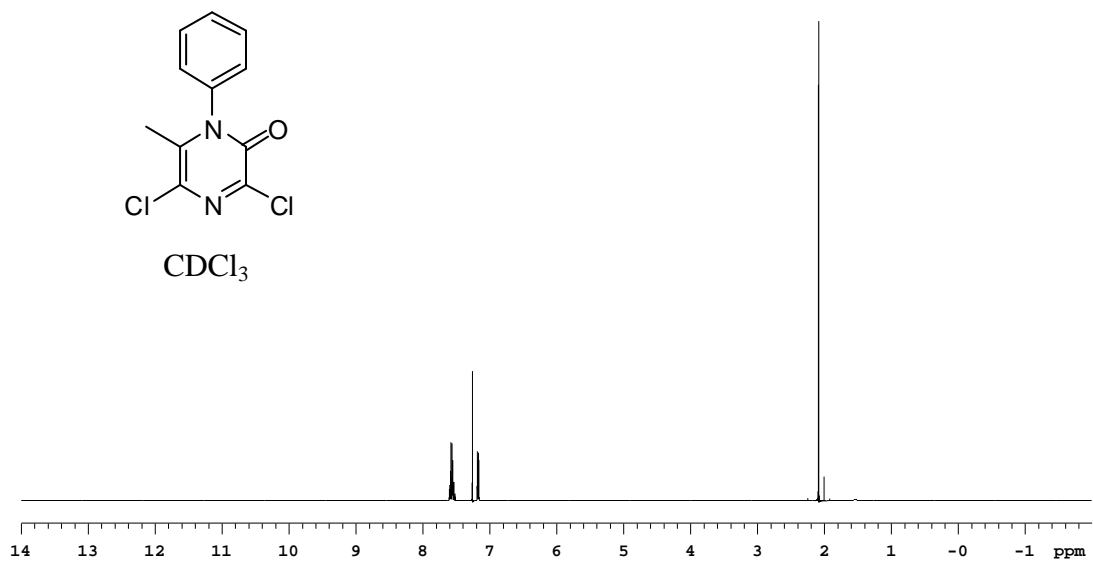
4i



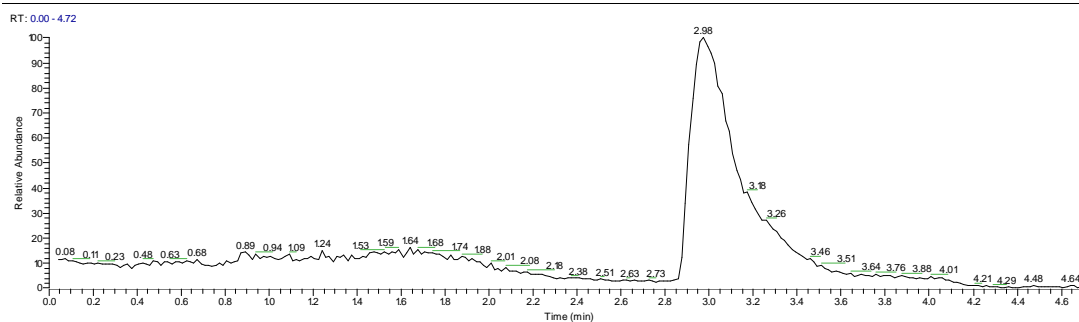
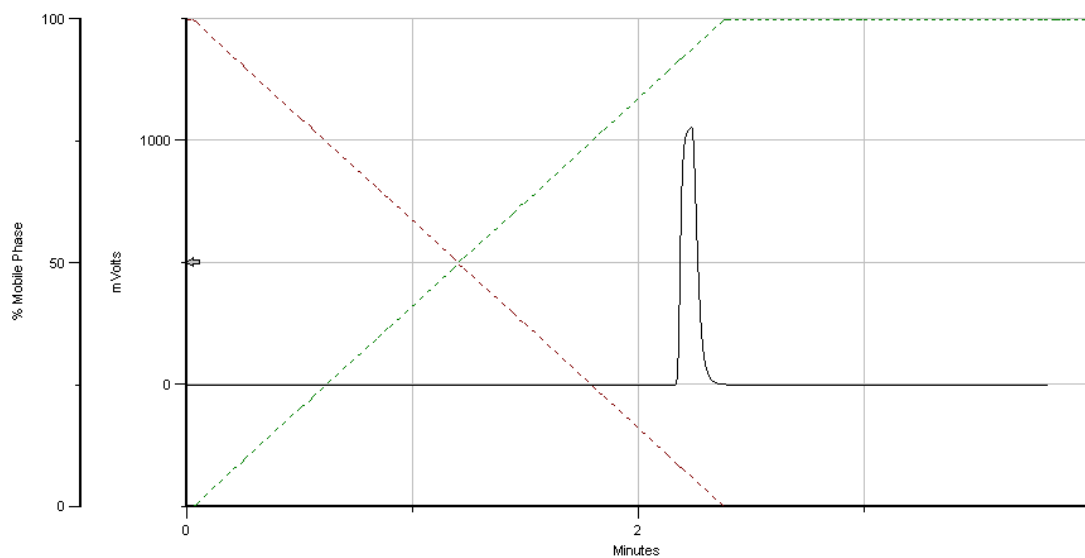
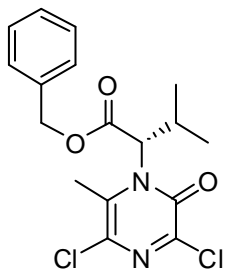
SQ_gis7619 #105 RT: 1.78 AV: 1 NL: 1.53E6
T: (0,0) + c ESI sid=10.00 Full ms [100.00-1200.00]



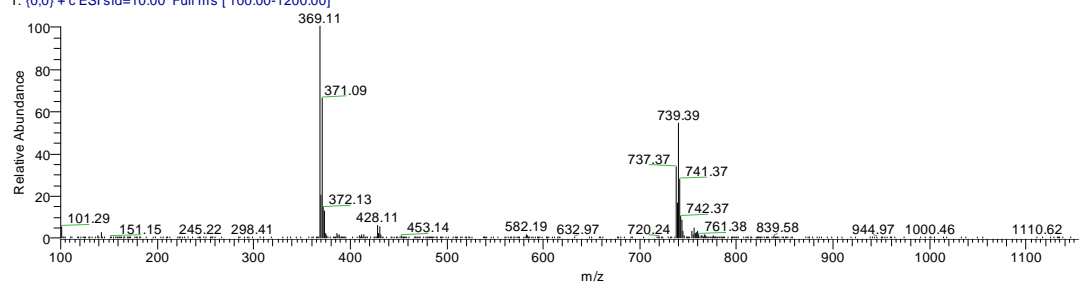
4i



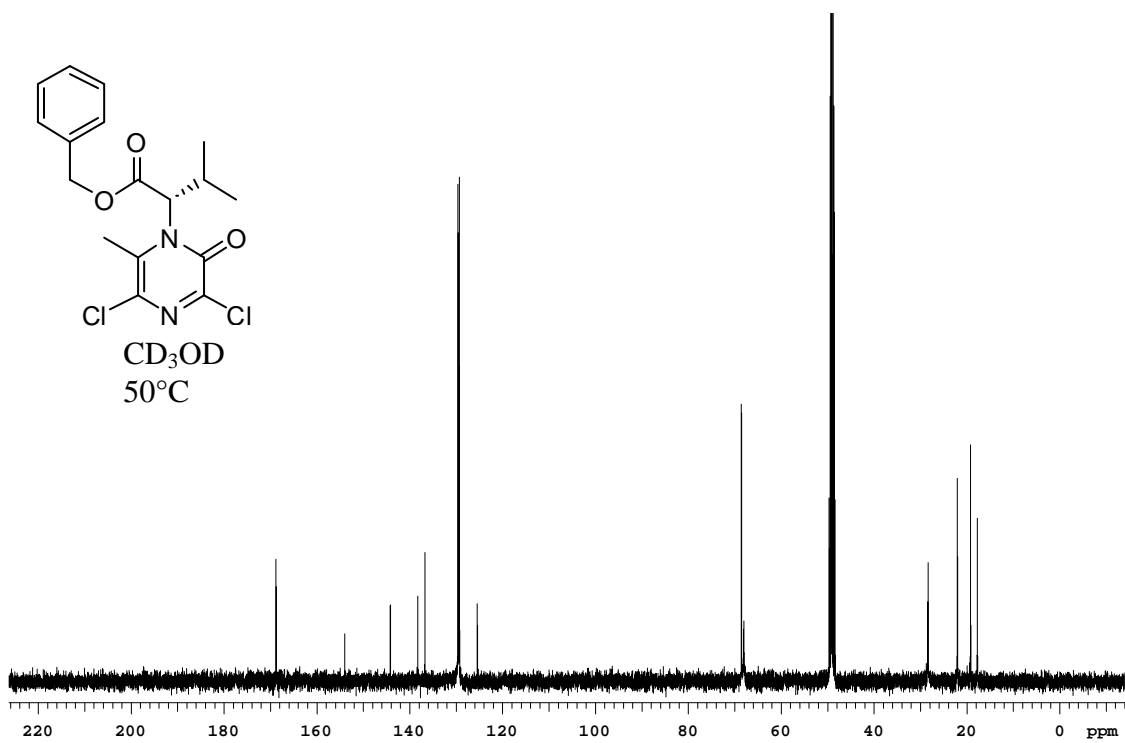
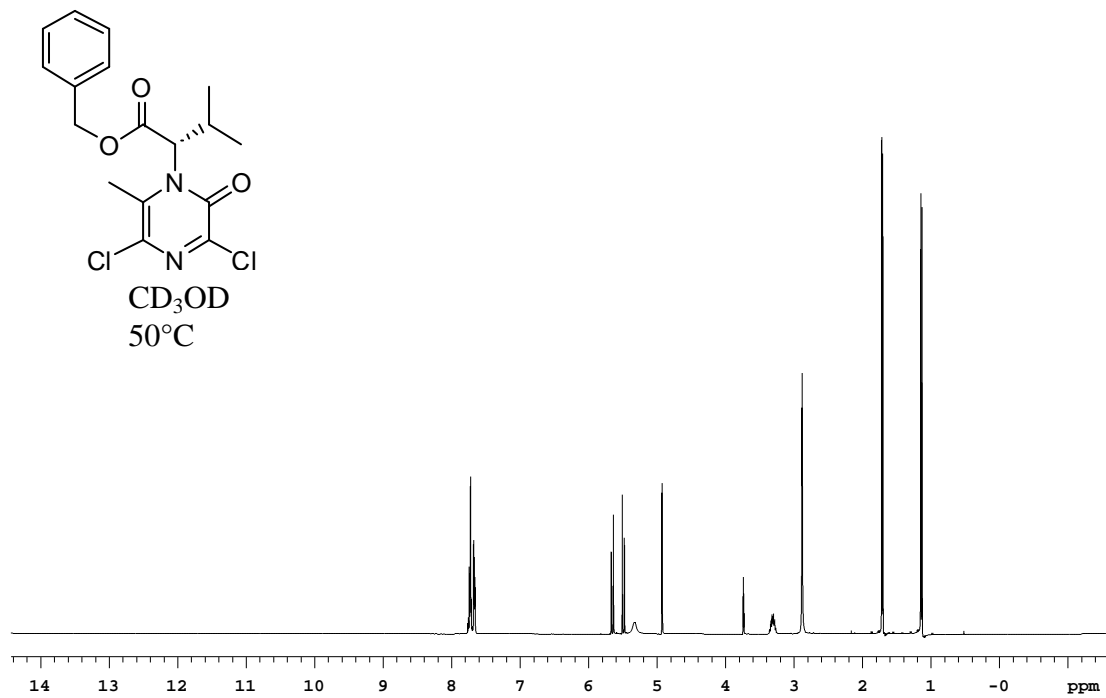
4j



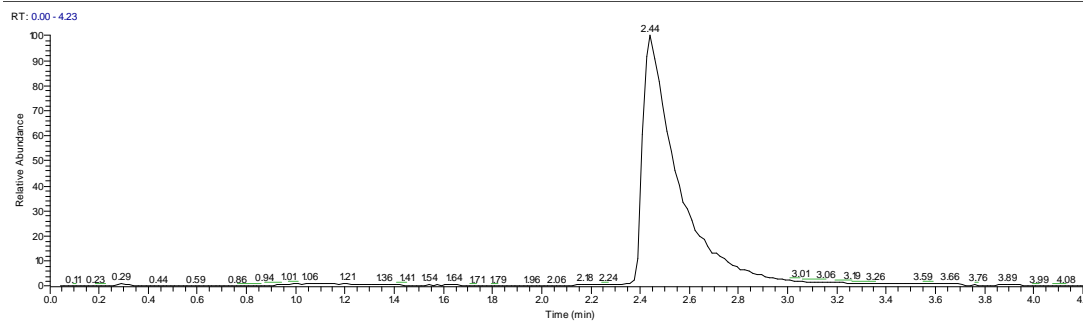
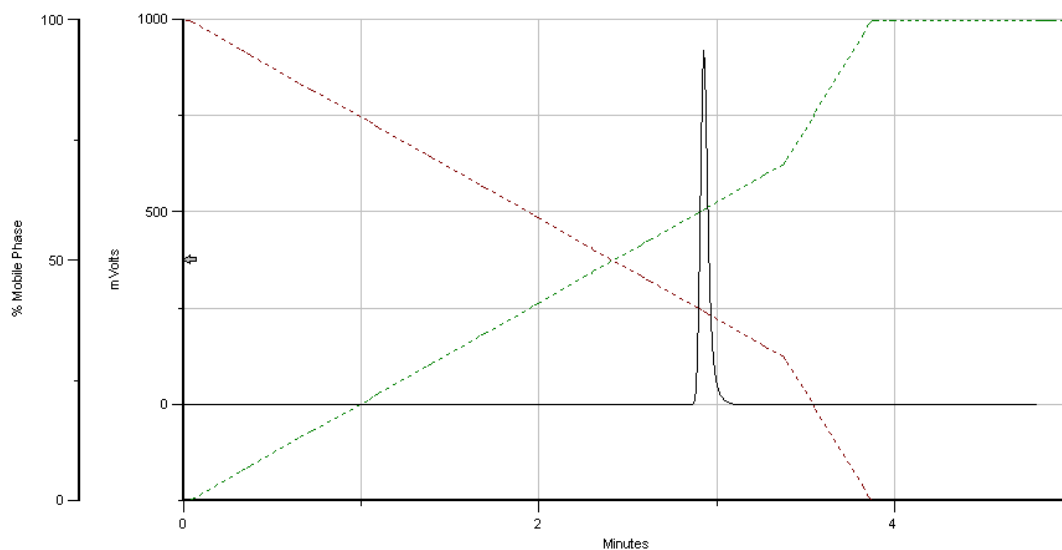
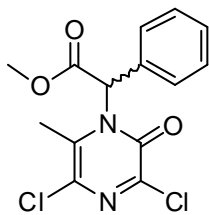
SQ_gis007606 #177 RT: 2.98 AV: 1 NL: 4.13E5
T: (0,0) + c ESI sid=10.00 Full ms [100.00-1200.00]



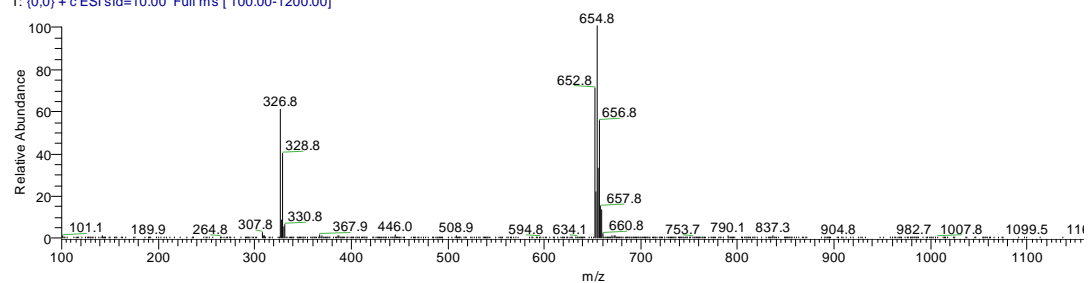
4j



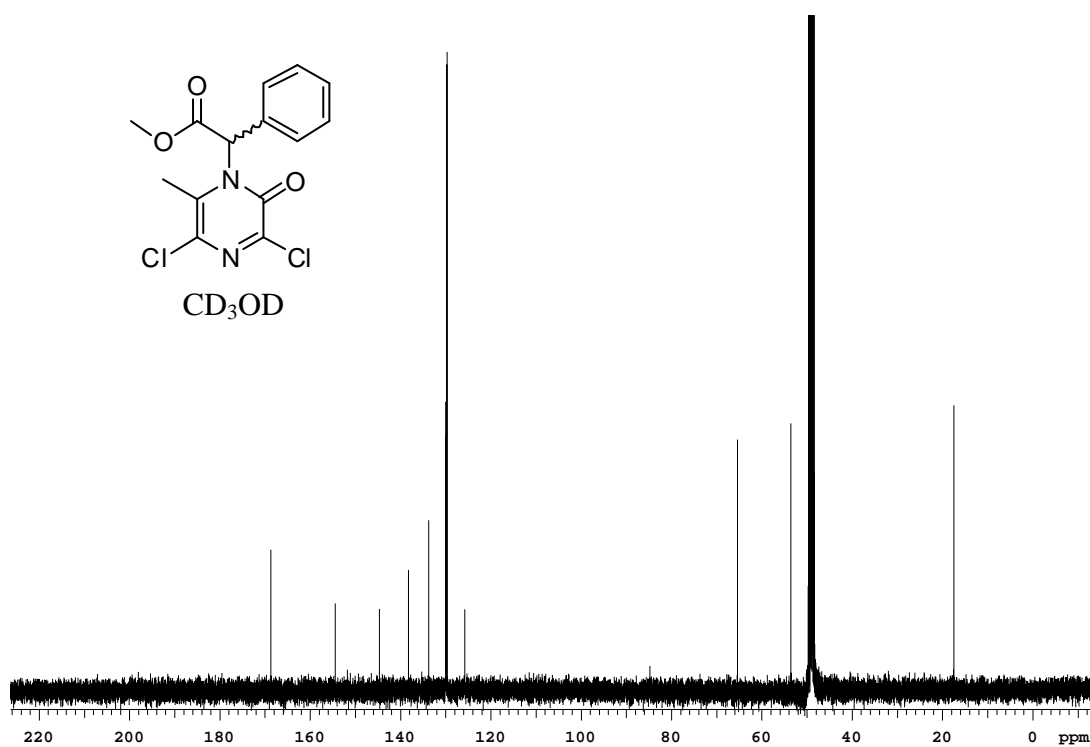
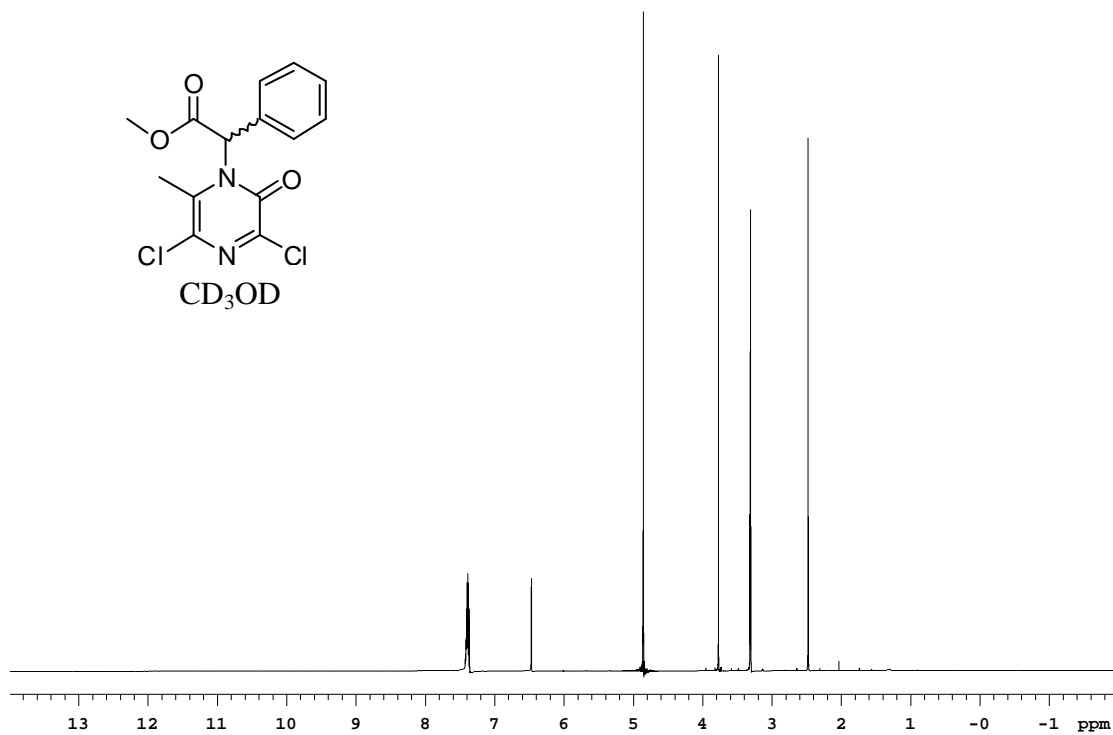
4k



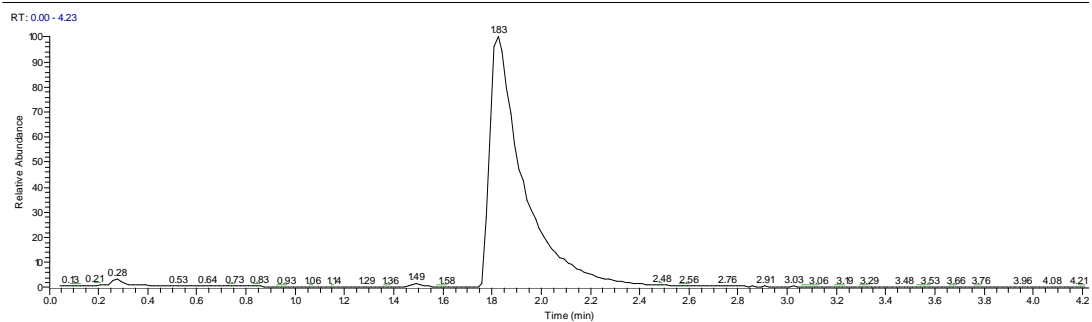
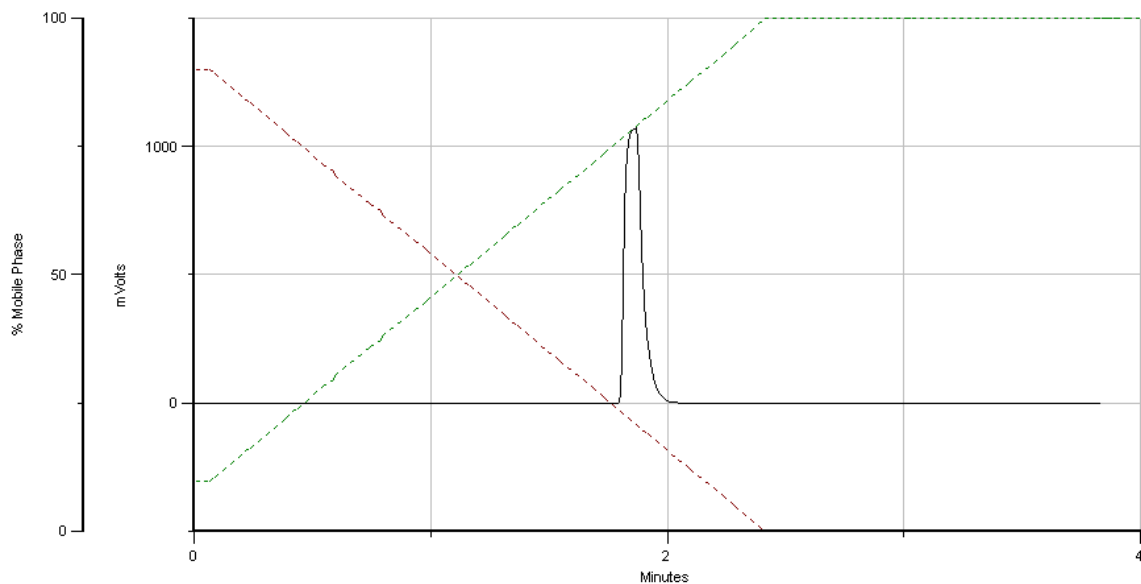
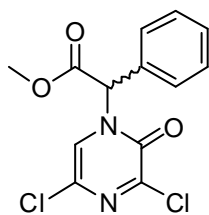
SQ_gis000166 #145 RT: 2.44 AV: 1 NL: 2.09E6
T: (0,0) + c ESI sid=10.00 Full ms [100.00-1200.00]



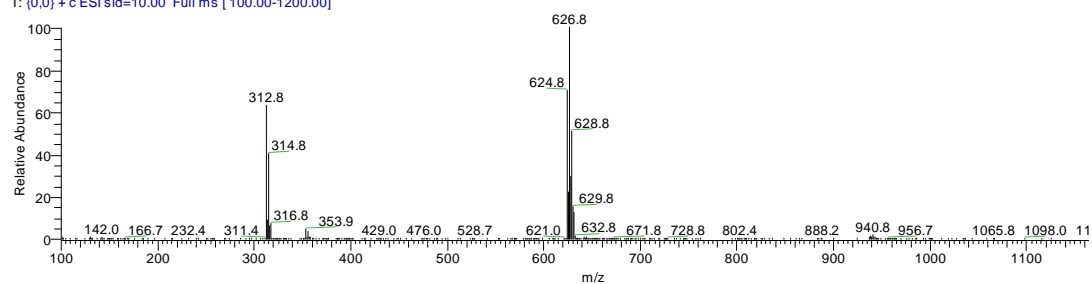
4k



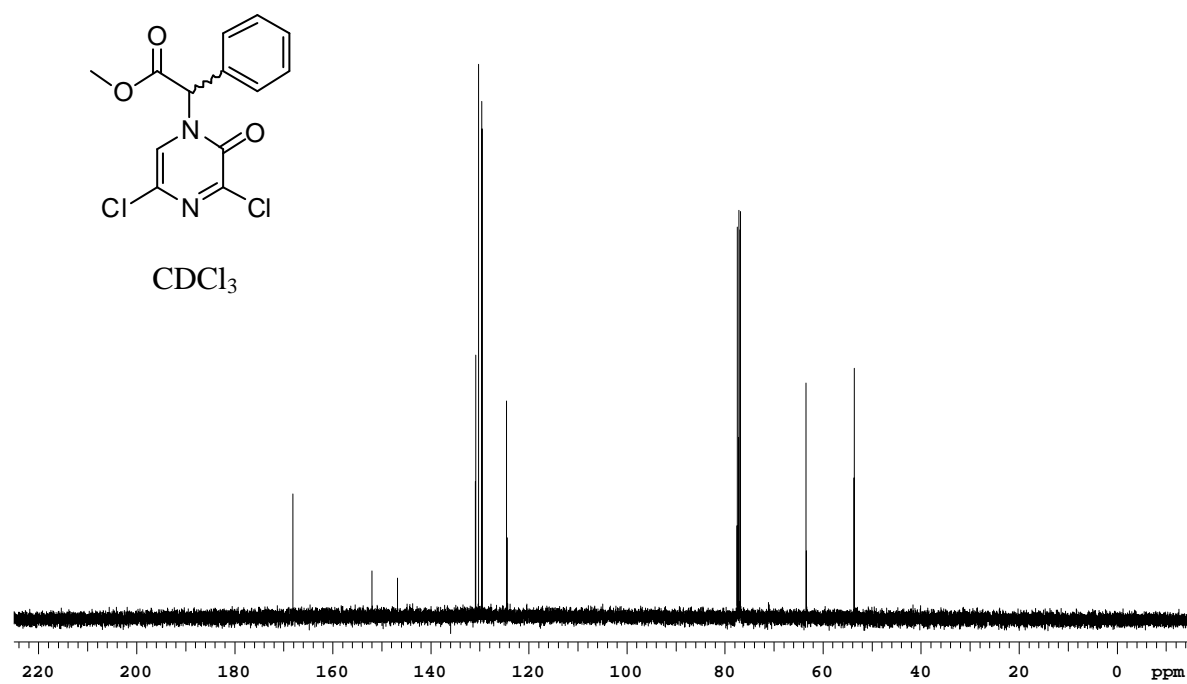
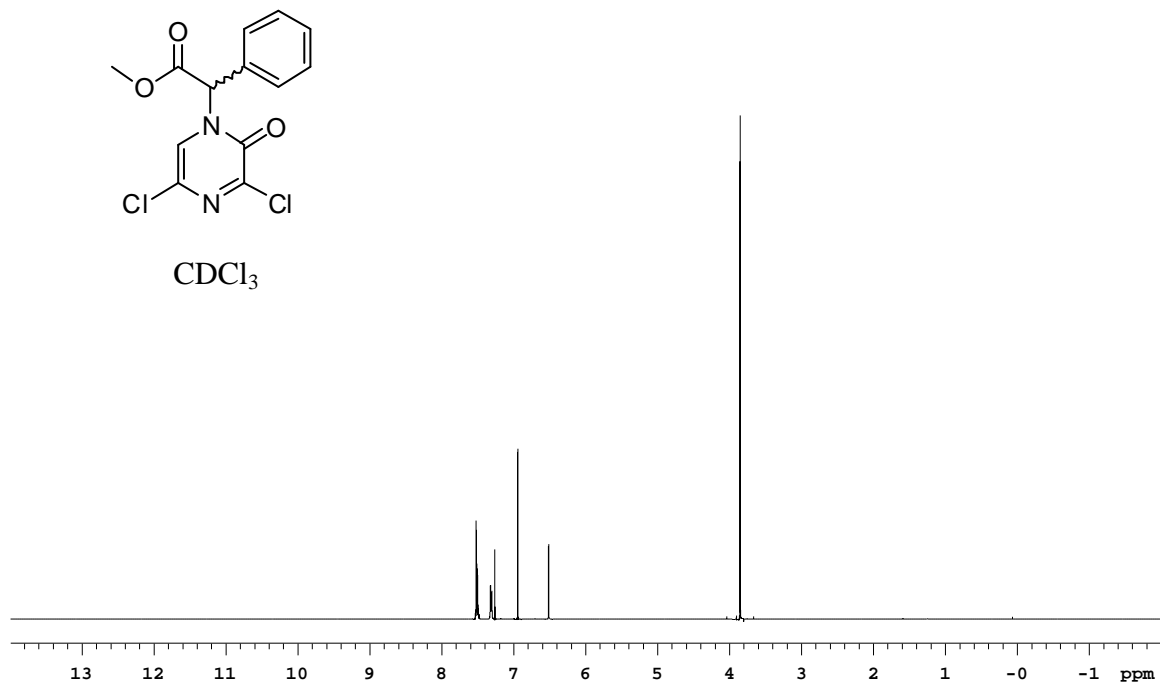
41



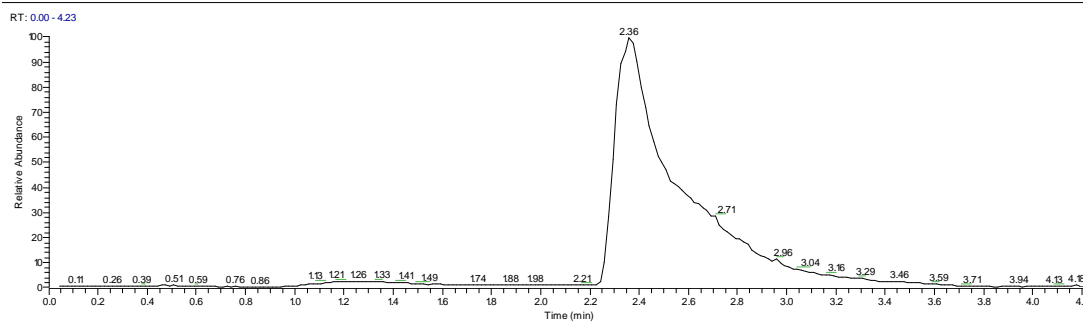
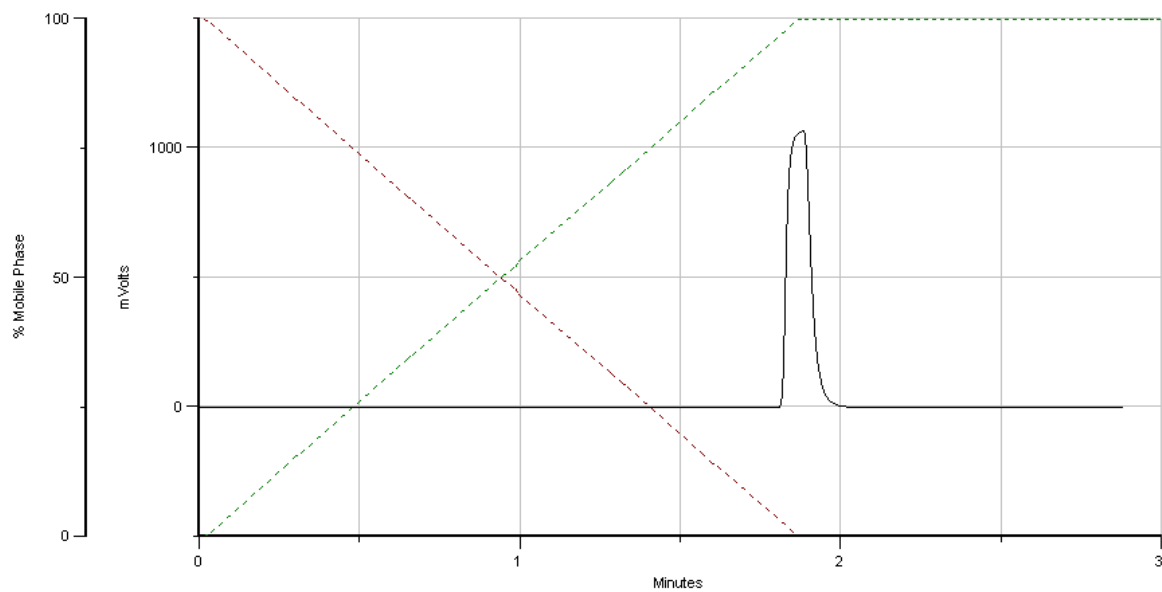
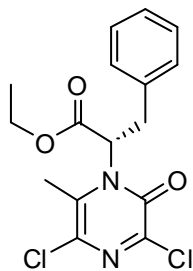
SQ_gis000174 #108 RT: 1.83 AV: 1 NL: 2.01E6
T: (0,0) +c ESI sid=10.00 Full ms [100.00-1200.00]



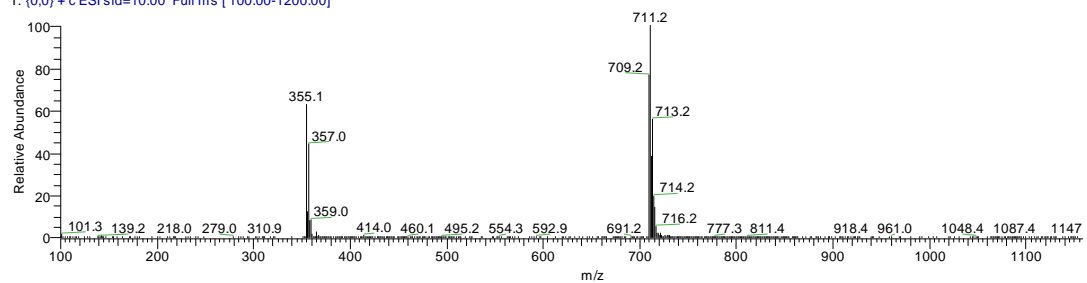
41



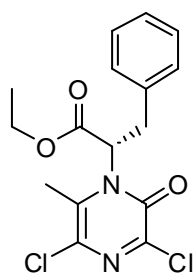
4m



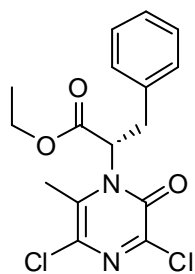
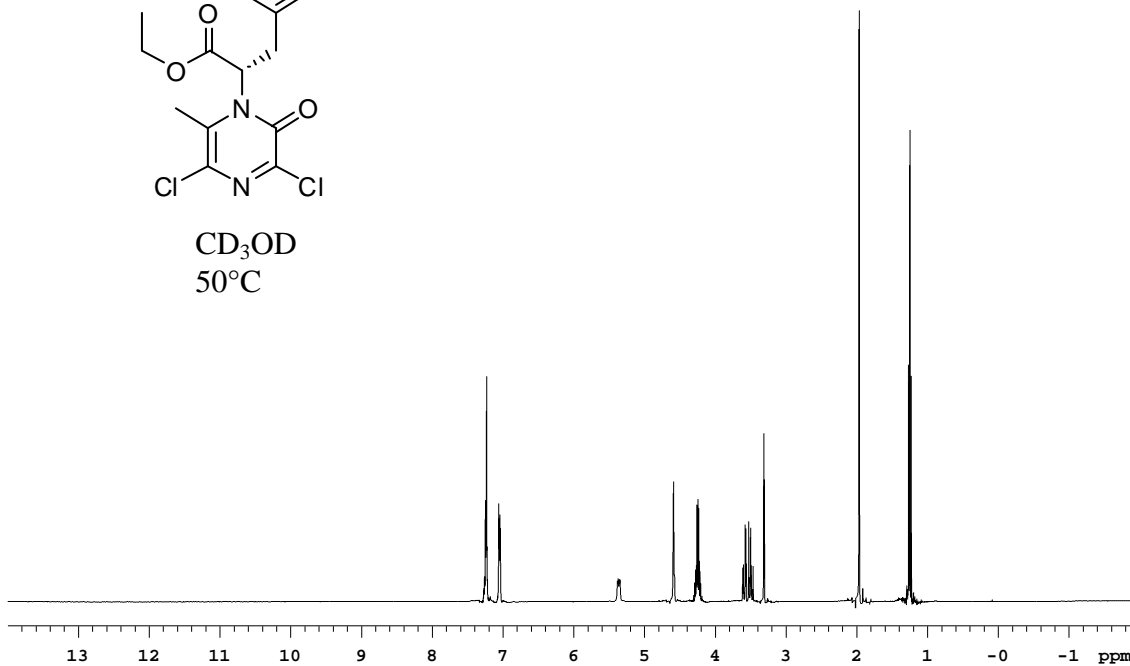
SQ_gis000197 #140 RT: 2.36 AV: 1 NL: 2.76E6
T: (0.0) + c ESI sid=10.00 Full ms [100.00-1200.00]



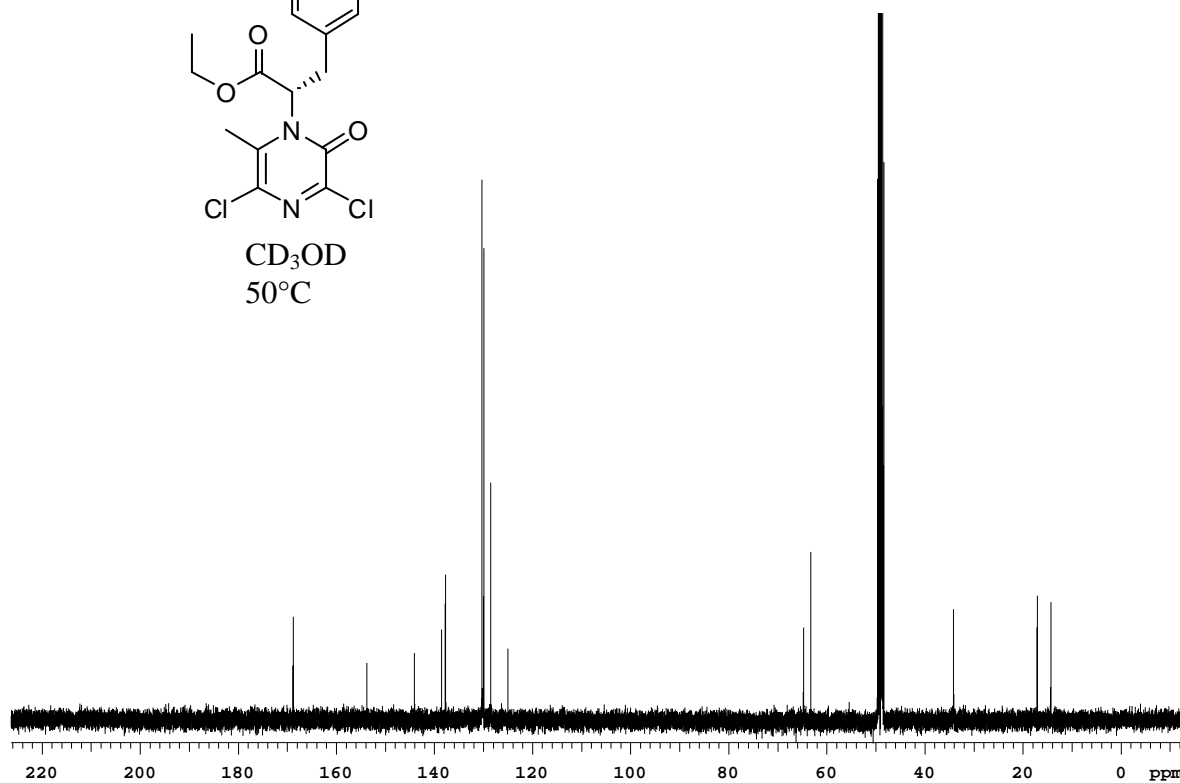
4m



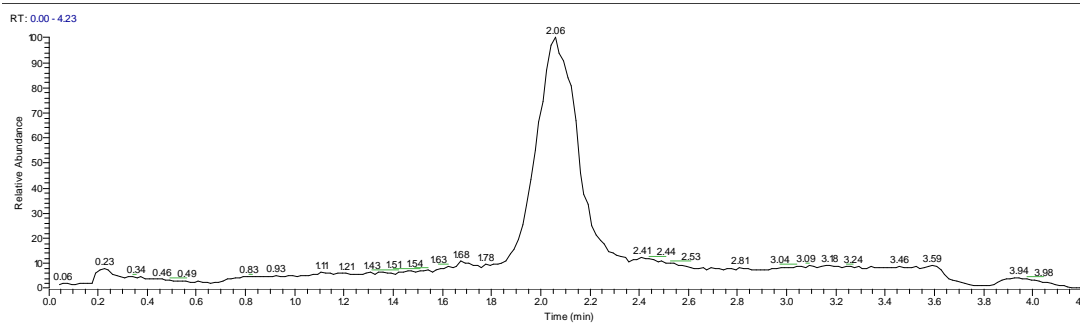
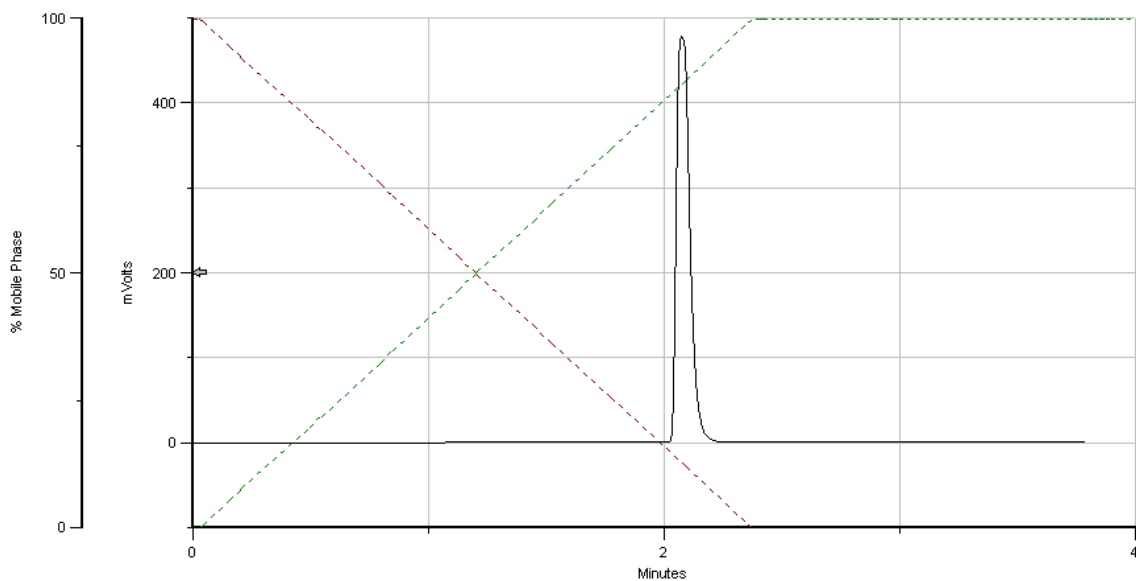
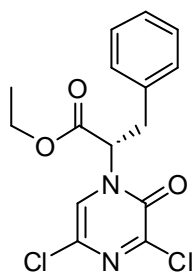
CD₃OD
50°C



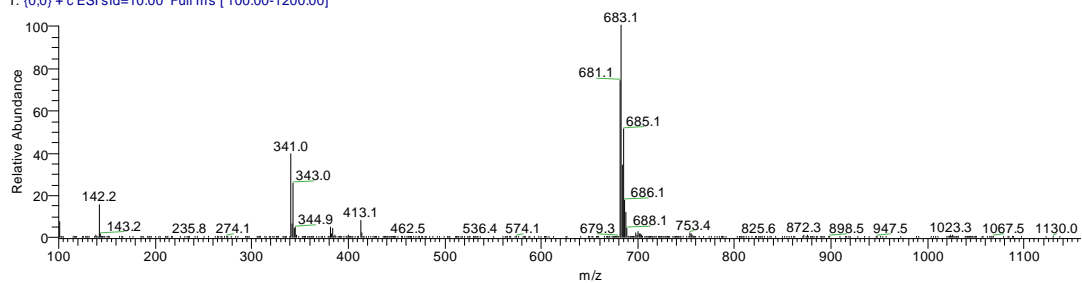
CD₃OD
50°C



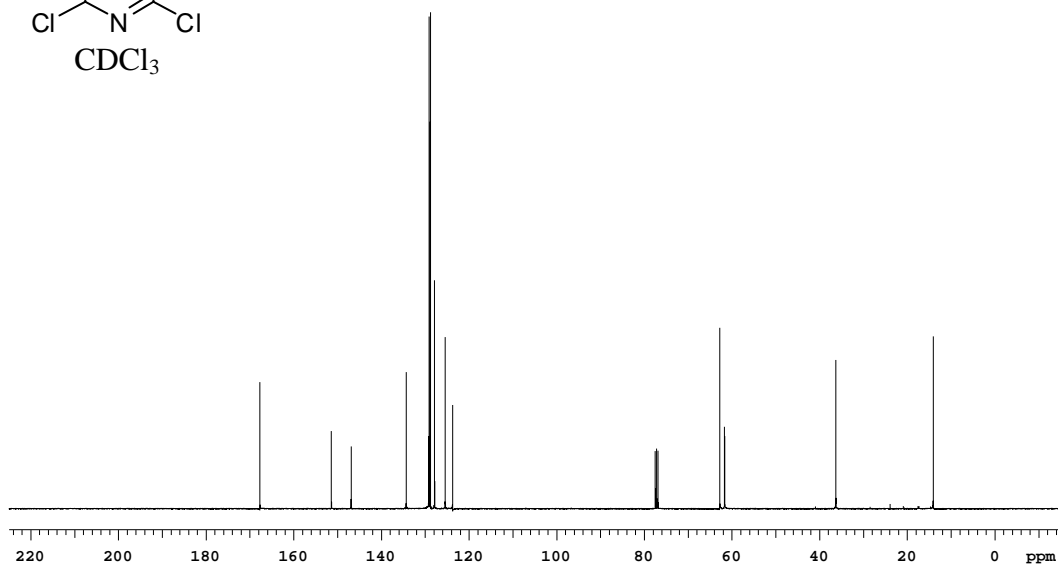
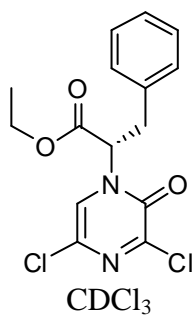
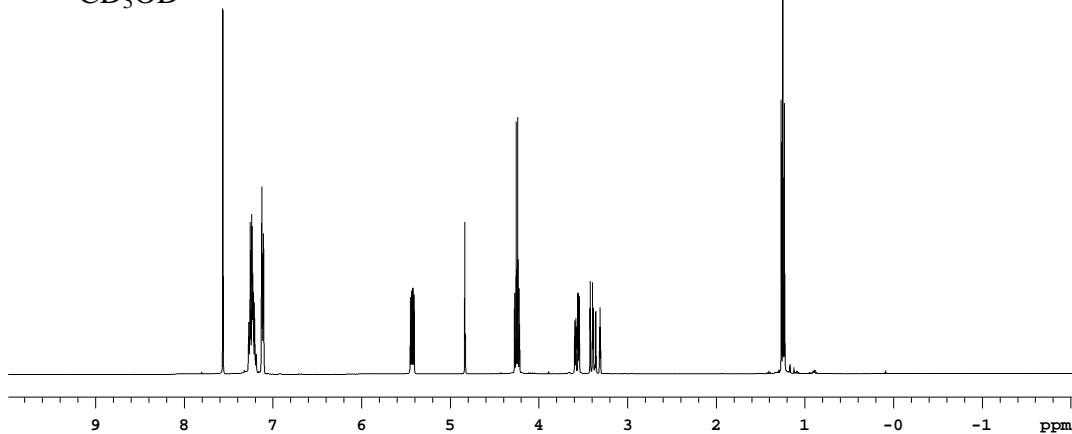
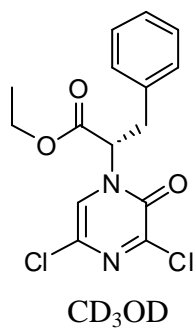
4n



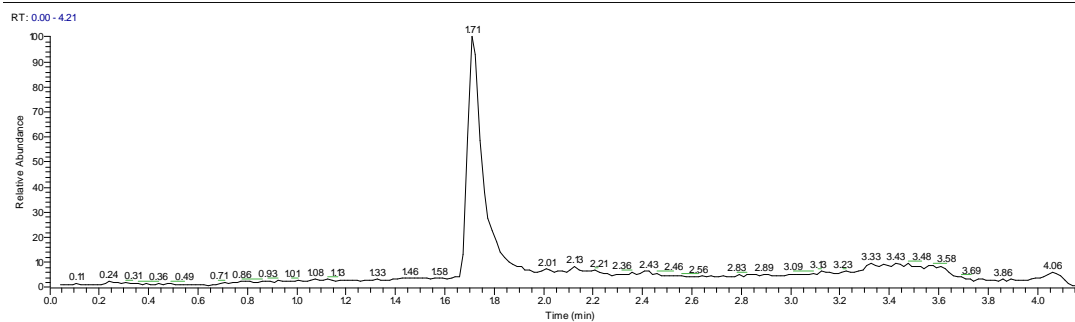
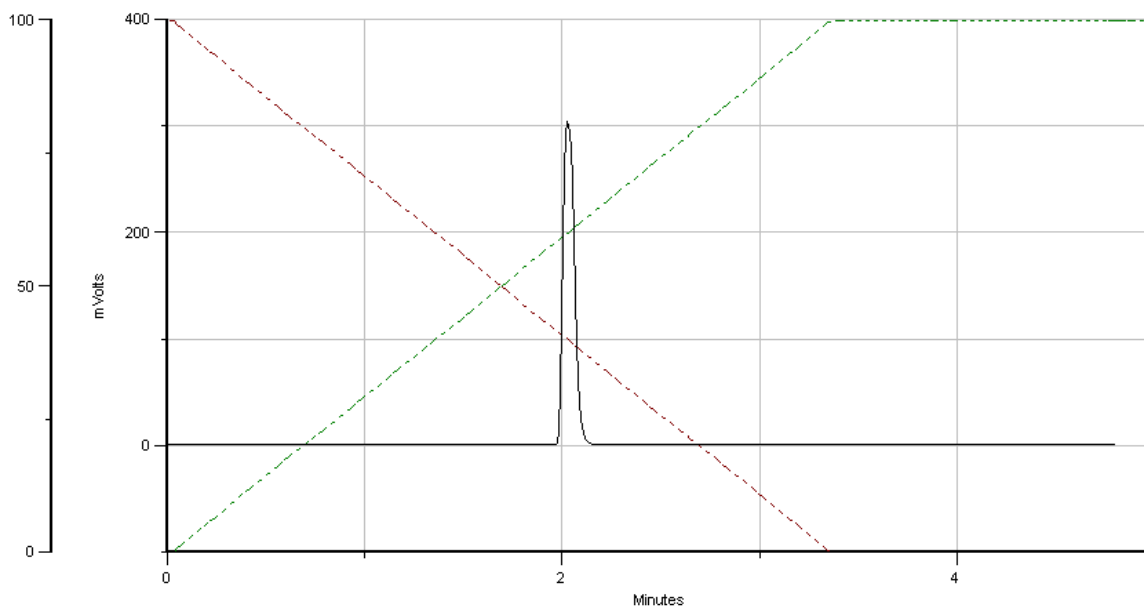
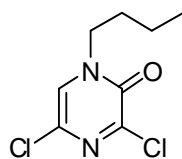
SQ_gis007613 #122 RT: 2.06 AV: 1 NL: 1.33E6
T: (0,0) + c ESI sid=10.00 Full ms [100.00-1200.00]



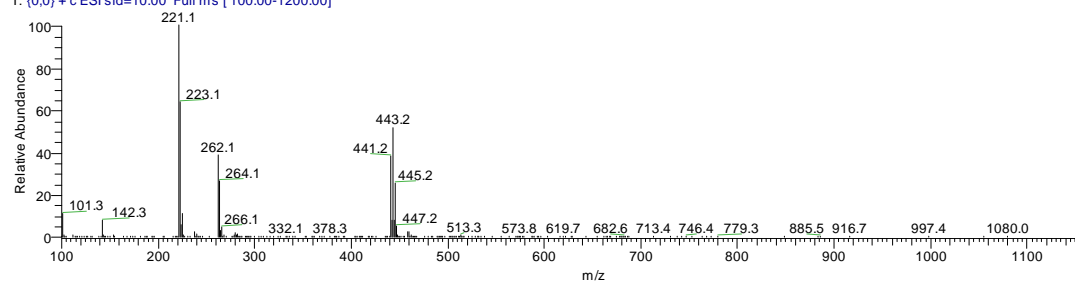
4n



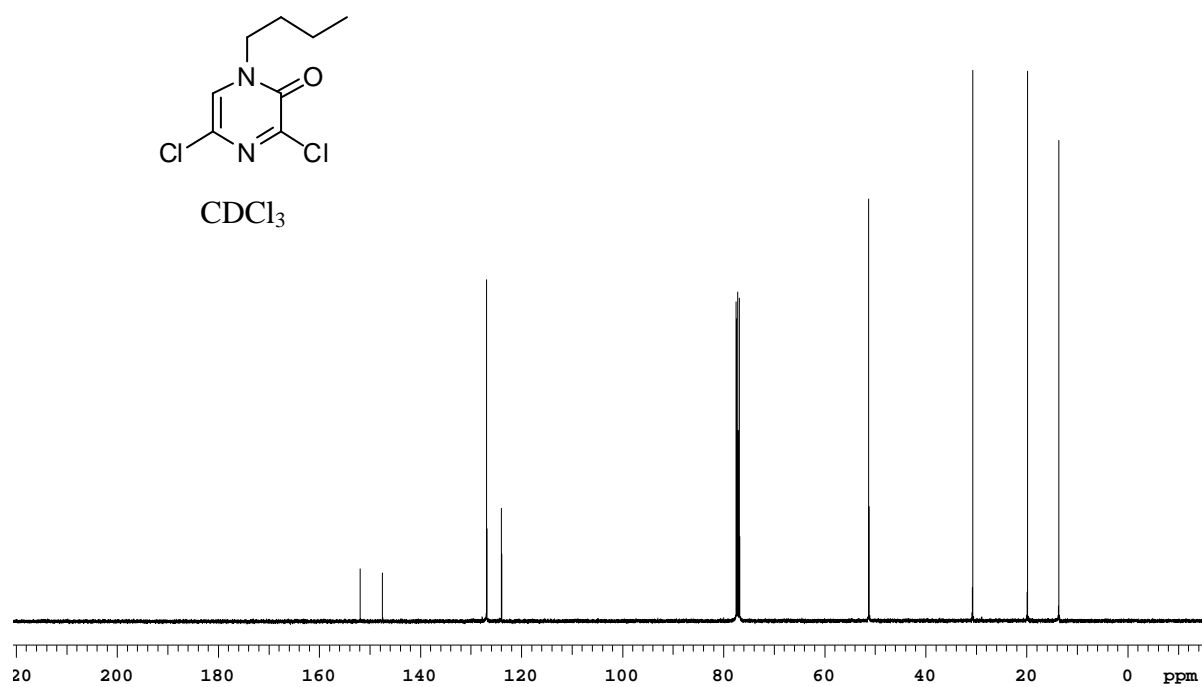
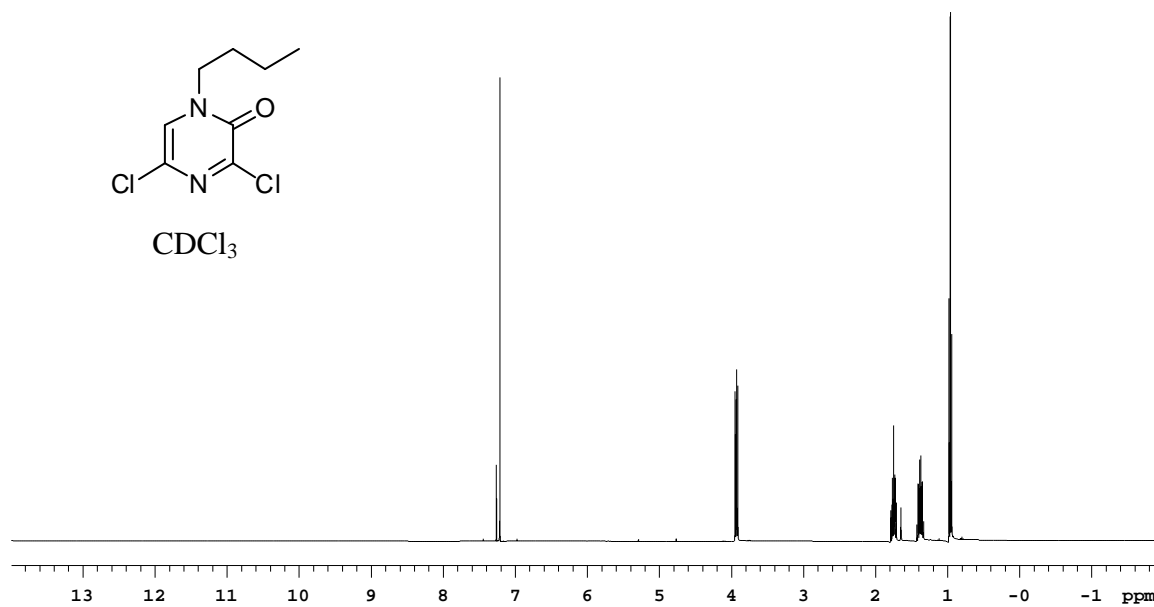
40



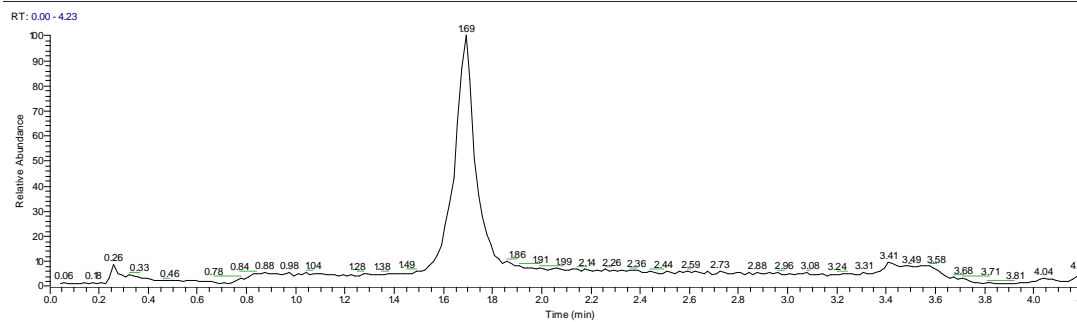
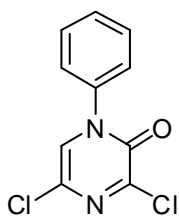
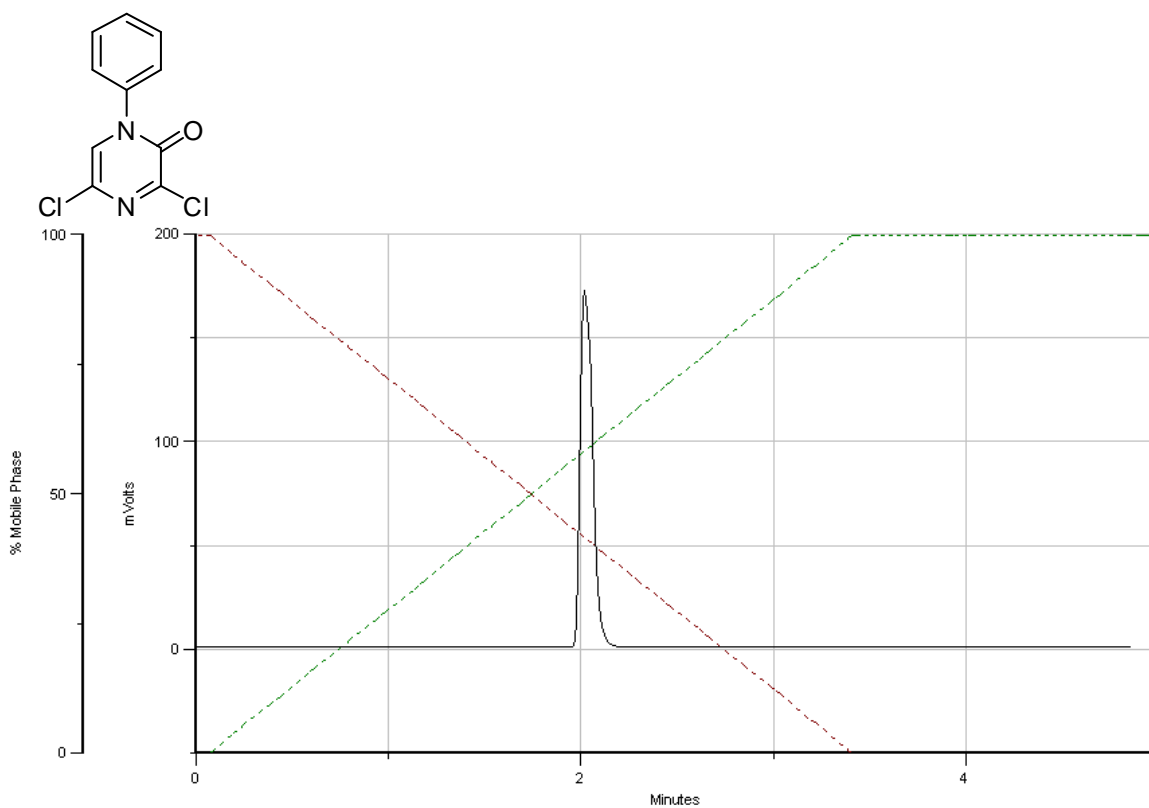
SQ_gis7637 #101 RT: 1.71 AV: 1 NL: 4.86E5
T: (0.0) + c ESI sid=10.00 Full ms [100.00-1200.00]



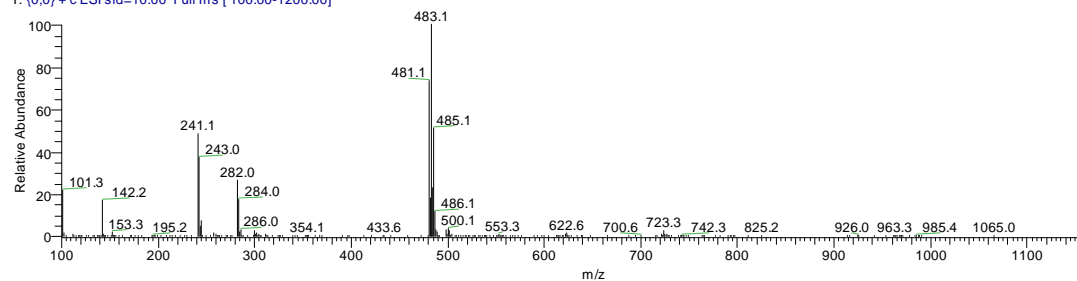
40



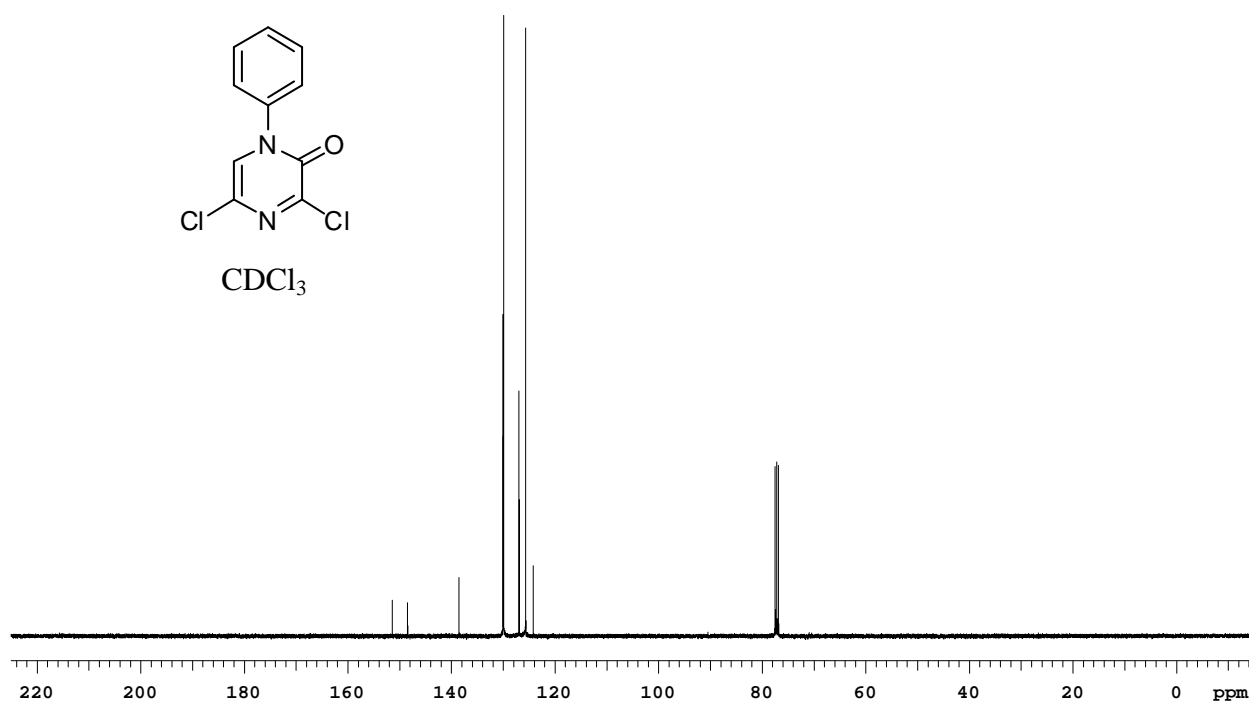
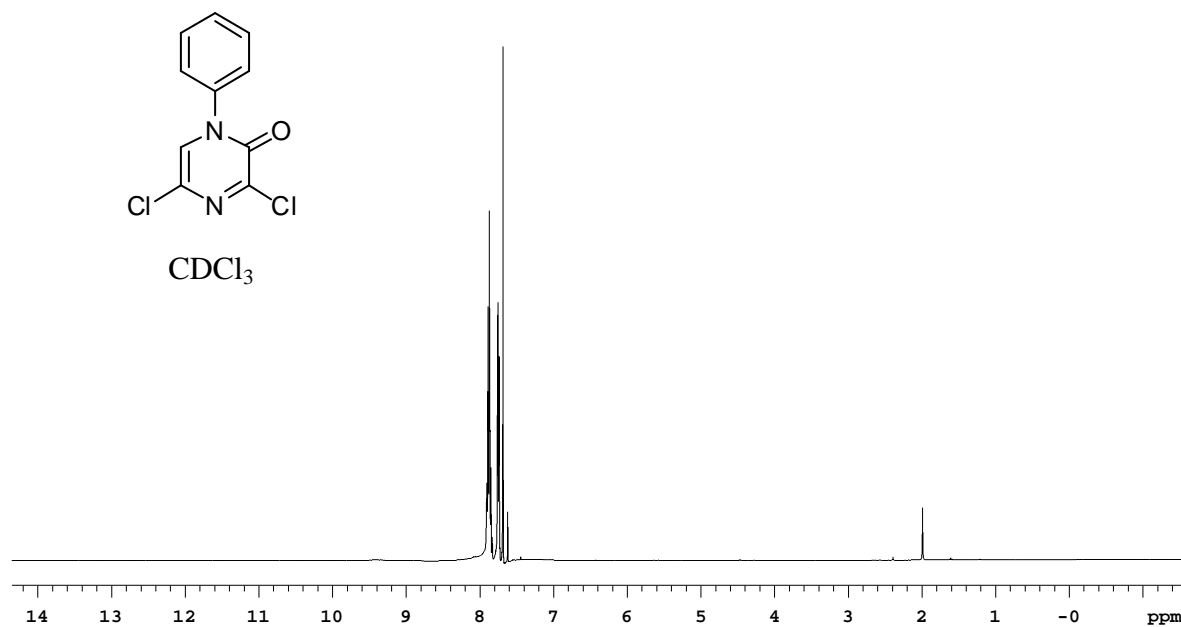
4p



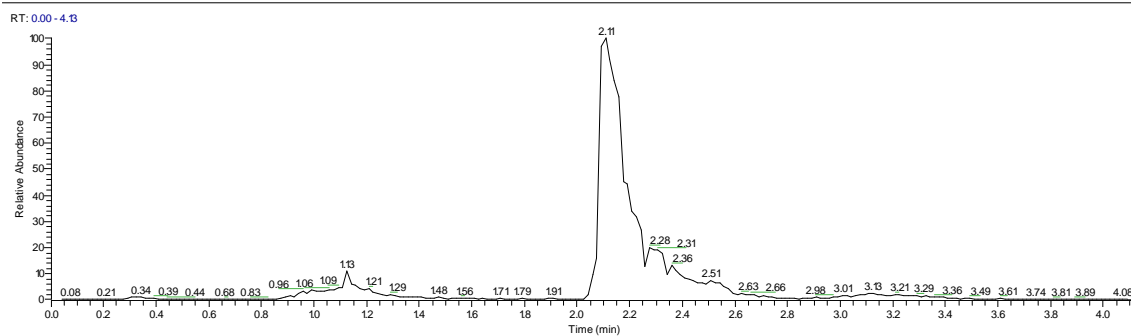
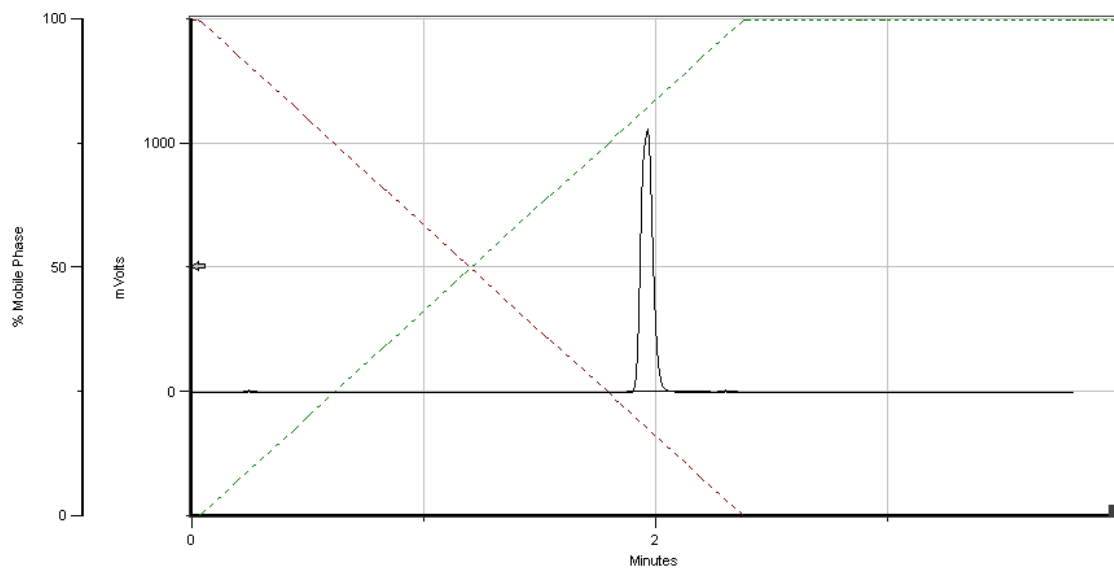
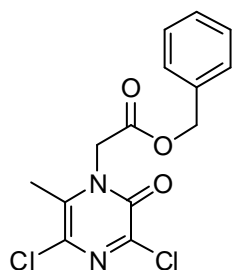
SQ_gis7638 #100 RT: 1.69 AV: 1 NL: 3.84E5
T: (0,0) + c ESI sid=10.00 Full ms [100.00-1200.00]



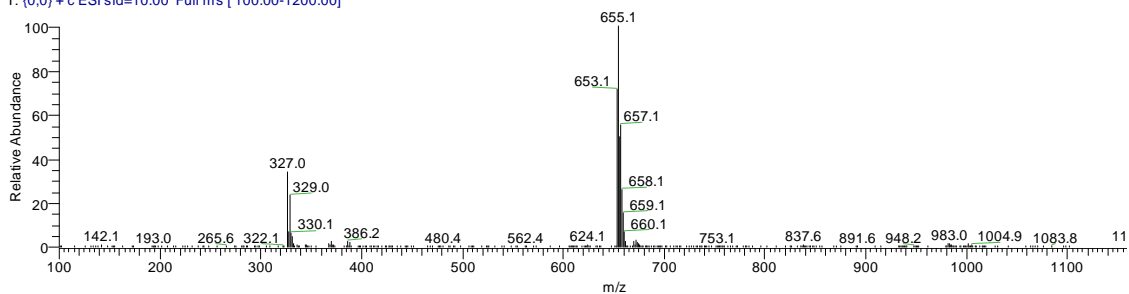
4p



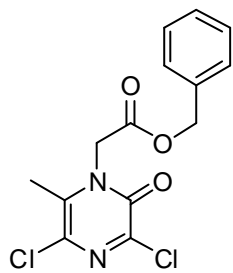
4q



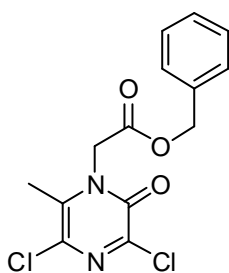
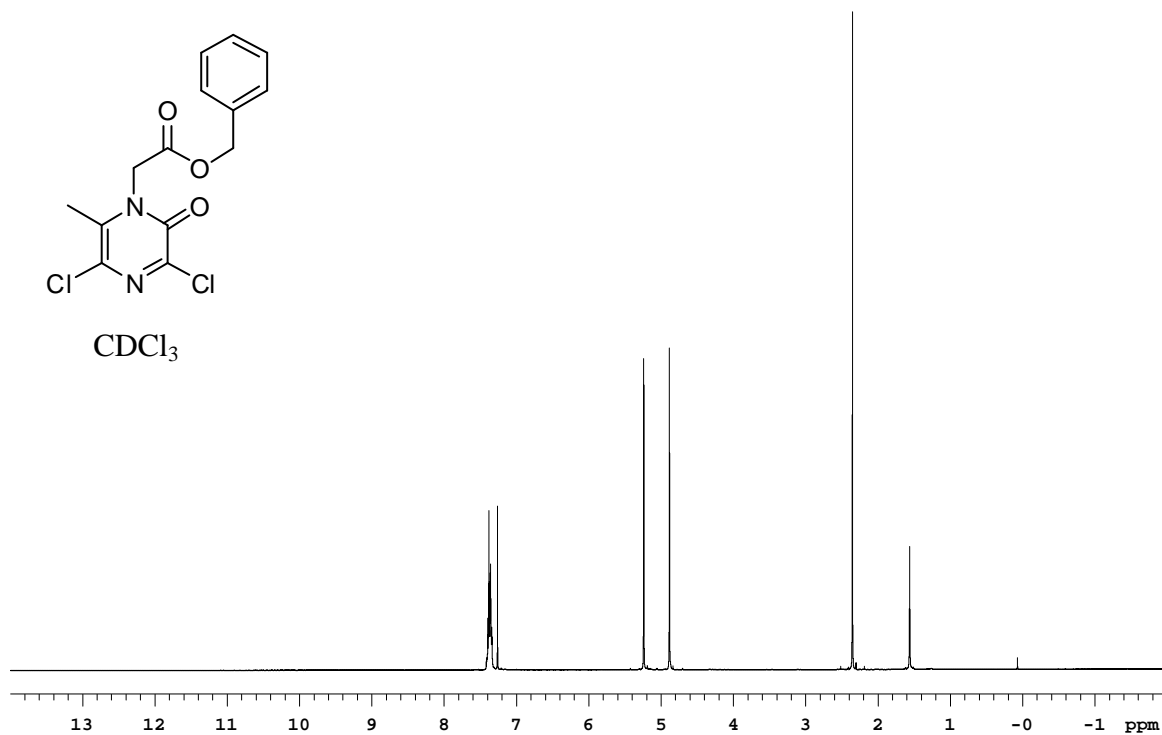
SQ_gis_4q #125 RT: 2.11 AV: 1 NL: 4.18E5
T: (0,0) + c ESI: sid=10.00 Full ms [100.00-1200.00]



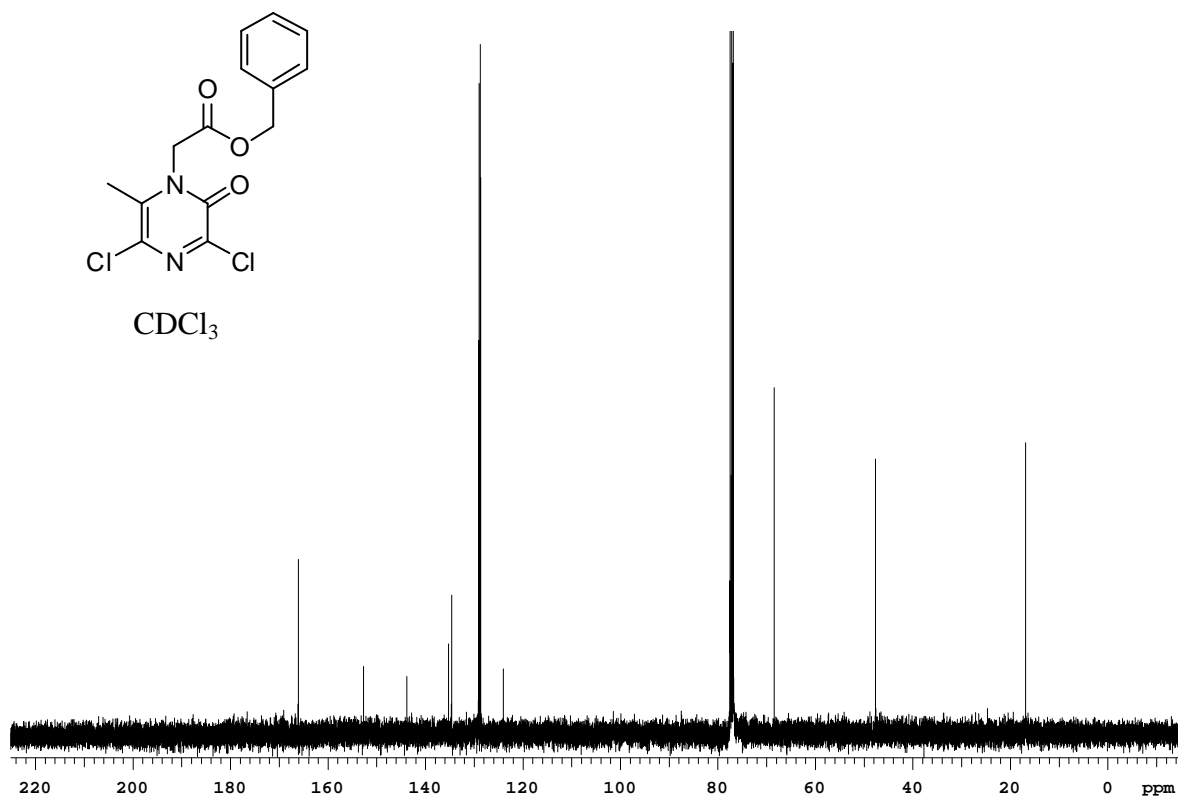
4q



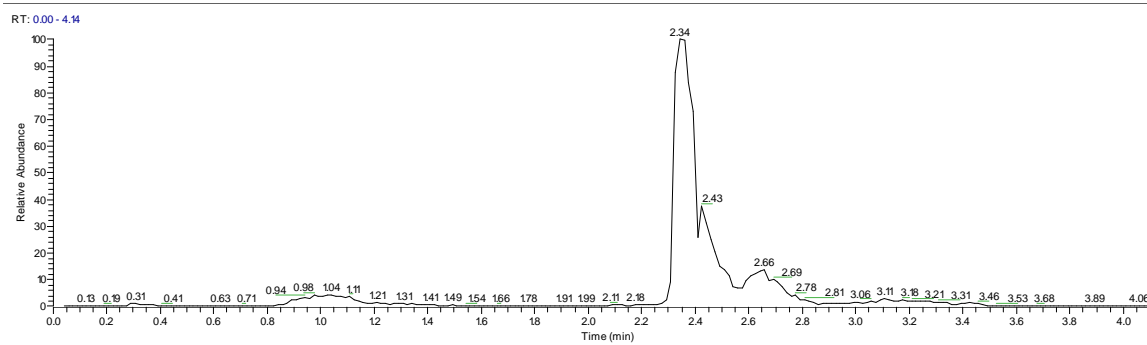
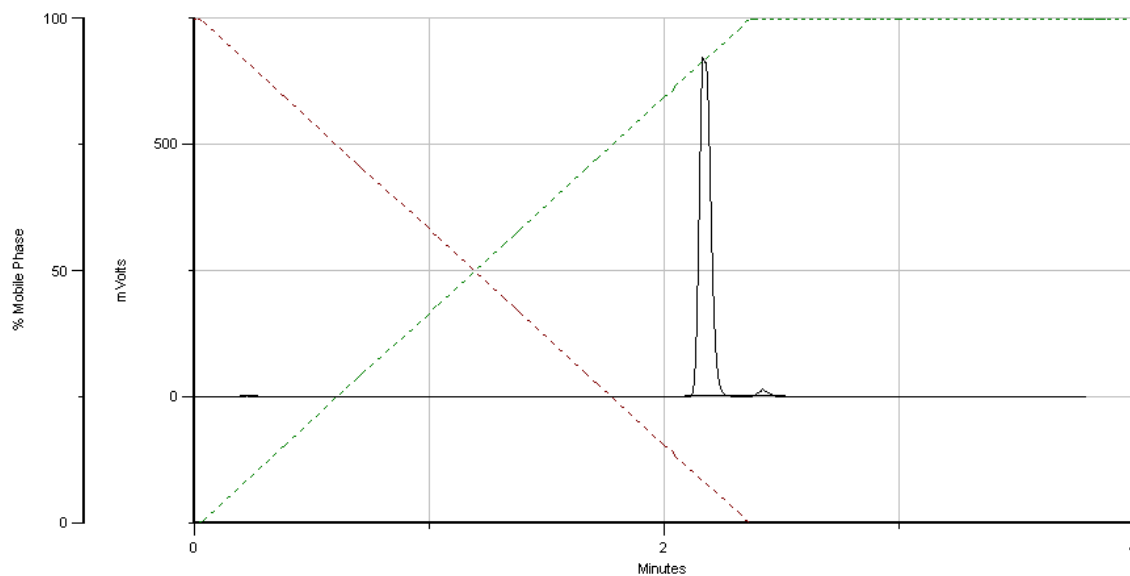
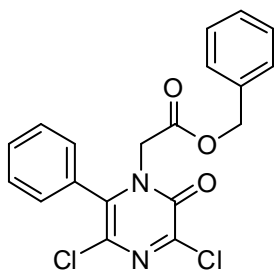
CDCl₃



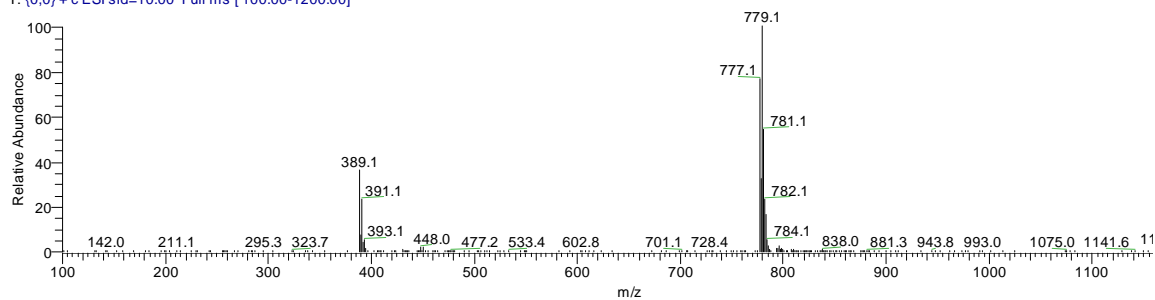
CDCl₃



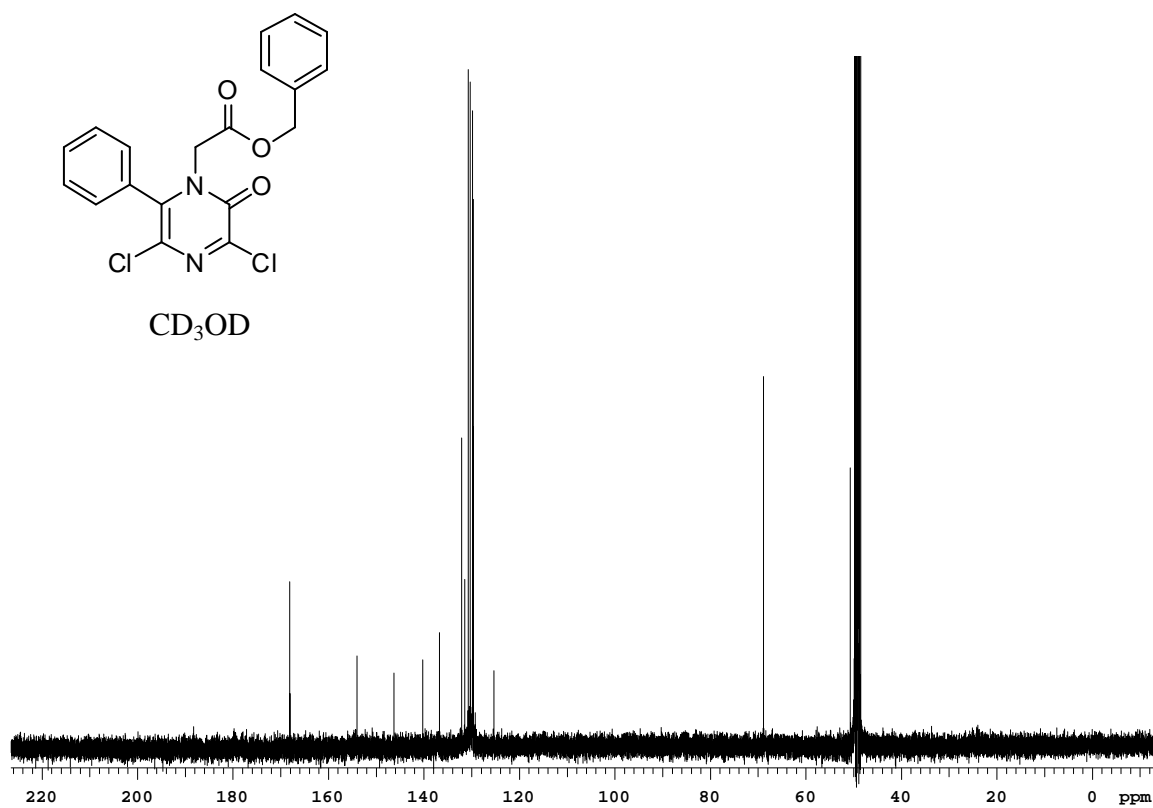
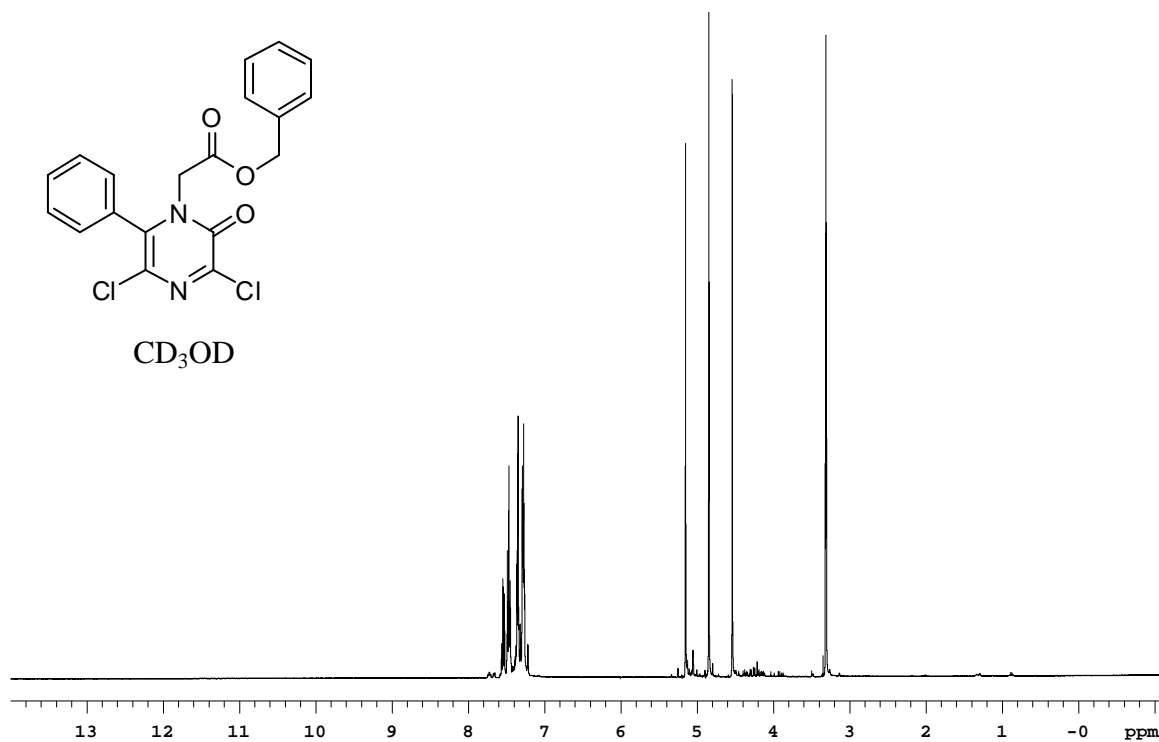
4r



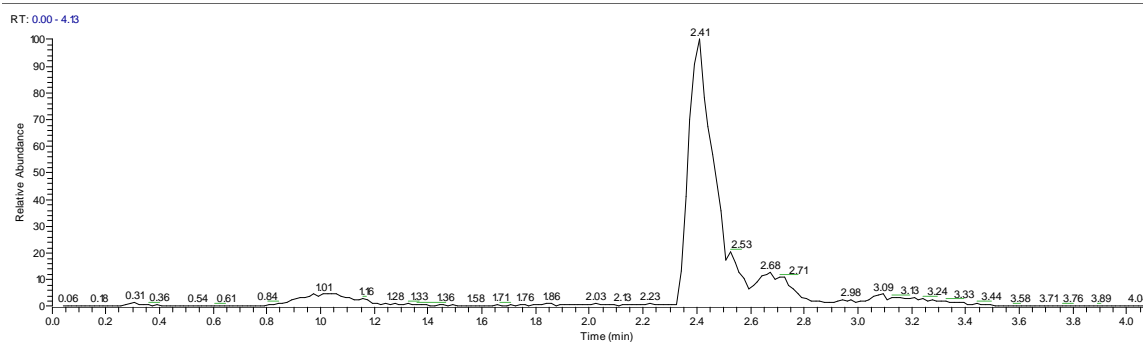
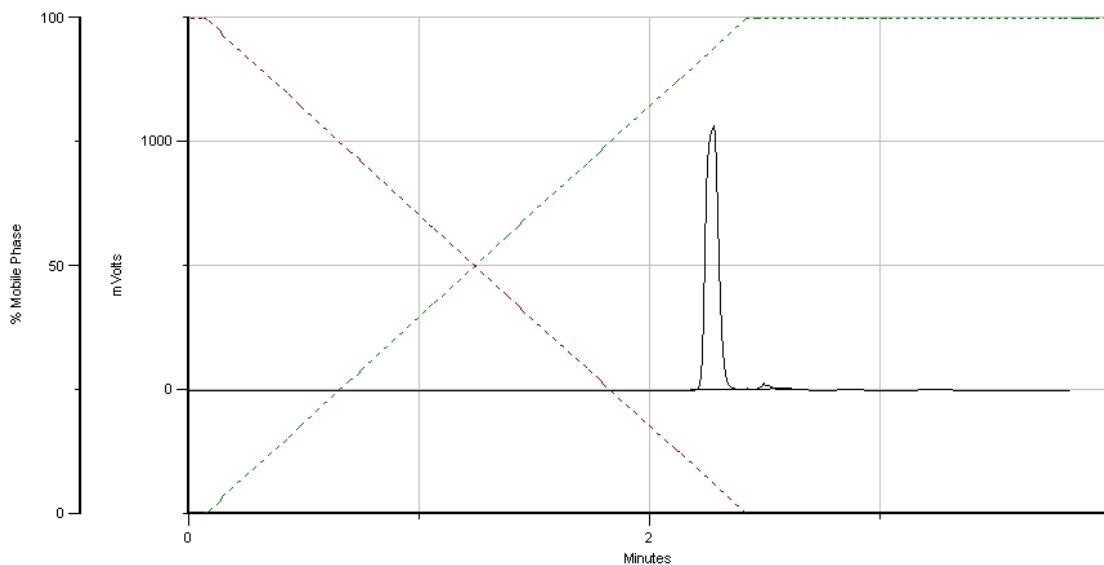
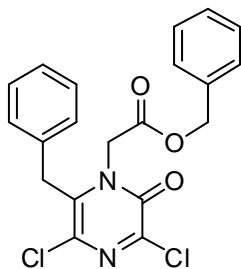
SQ_gis_4r#140 RT: 2.36 AV: 1 NL: 4.09E5
T: (0,0) + c ESI sid=10.00 Full ms [100.00-1200.00]



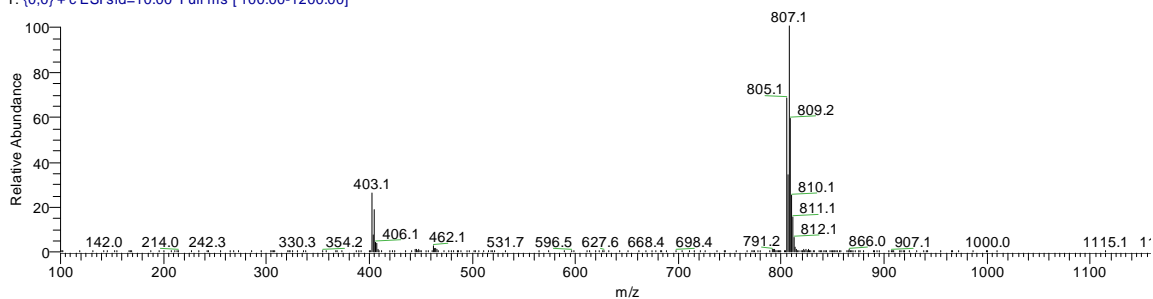
4r



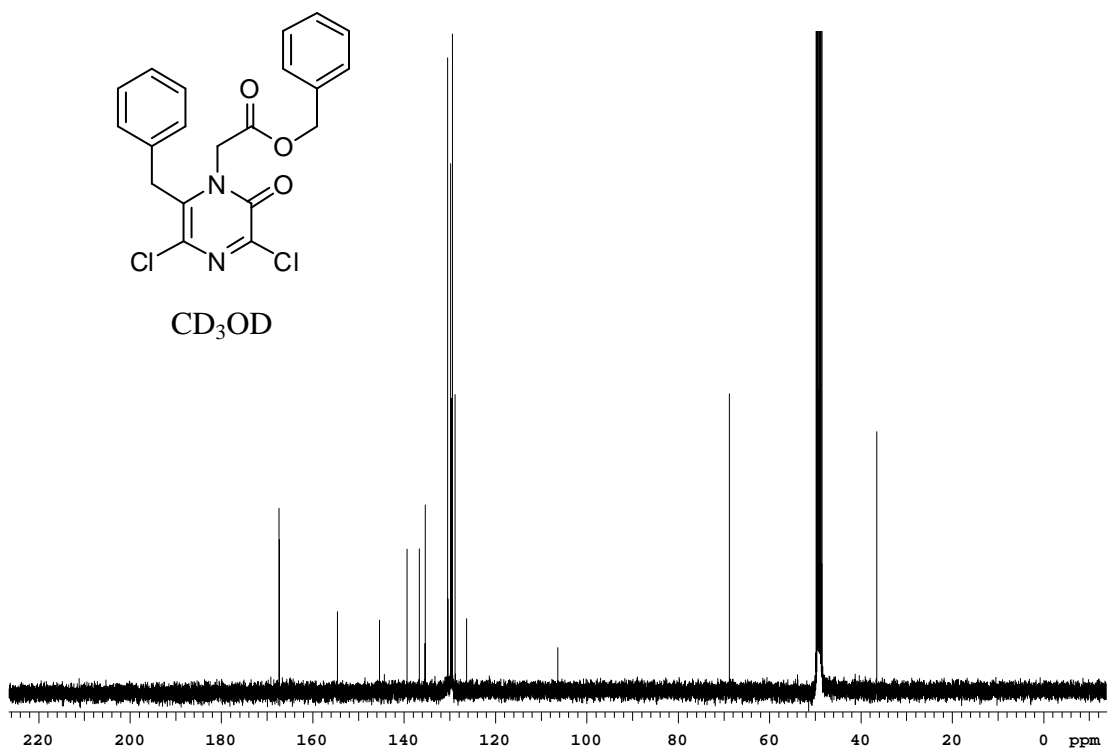
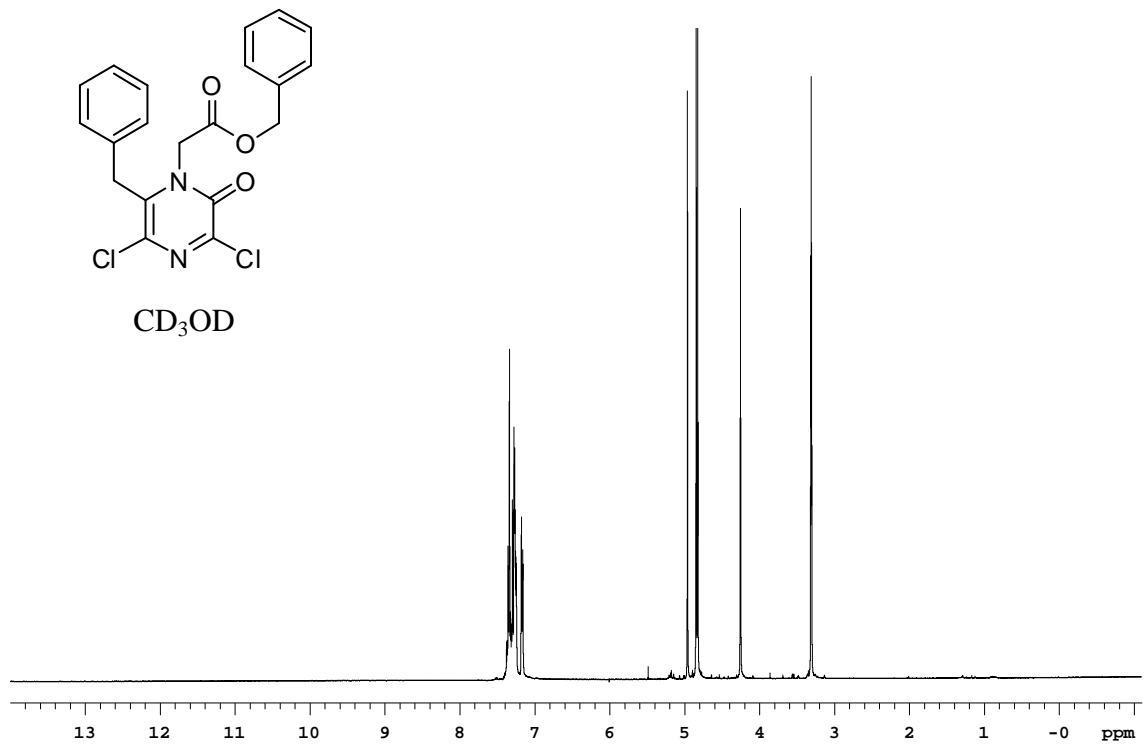
4s



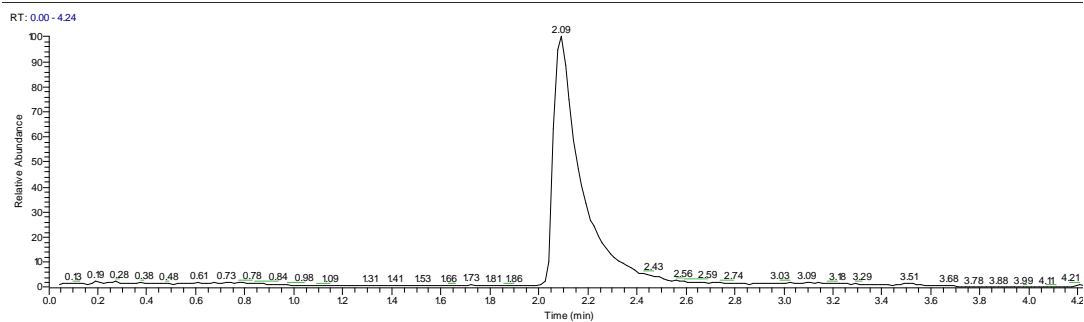
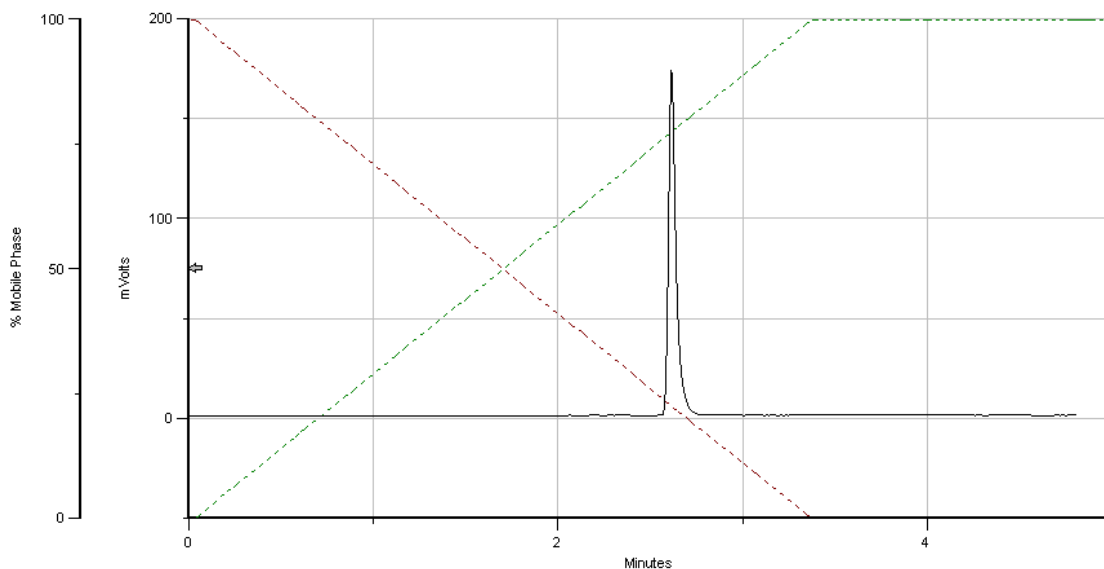
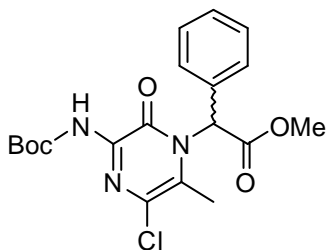
SQ_gis_4s #143 RT: 2.41 AV: 1 NL: 4.15E5
T: (0,0) + c ESI sid=10.00 Full ms [100.00-1200.00]



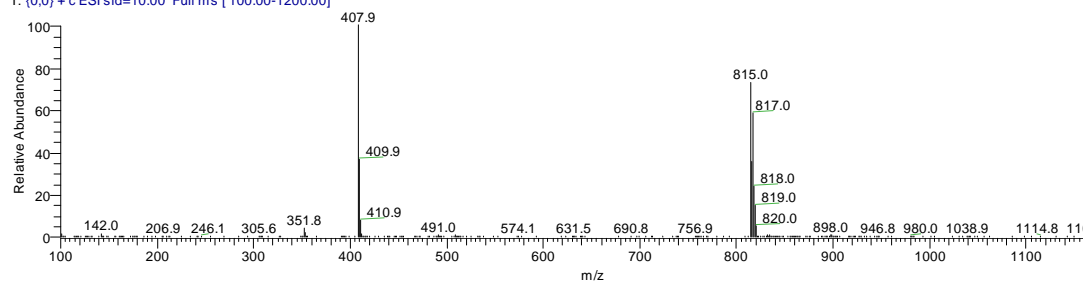
4s



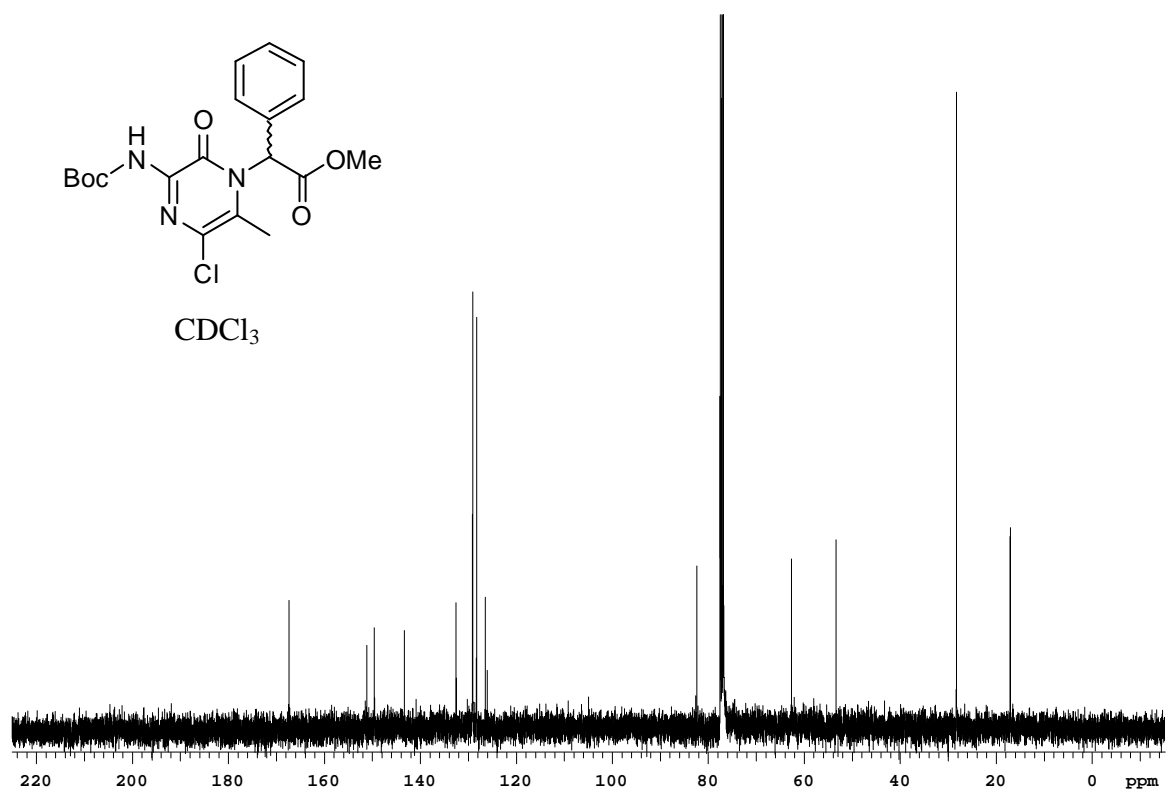
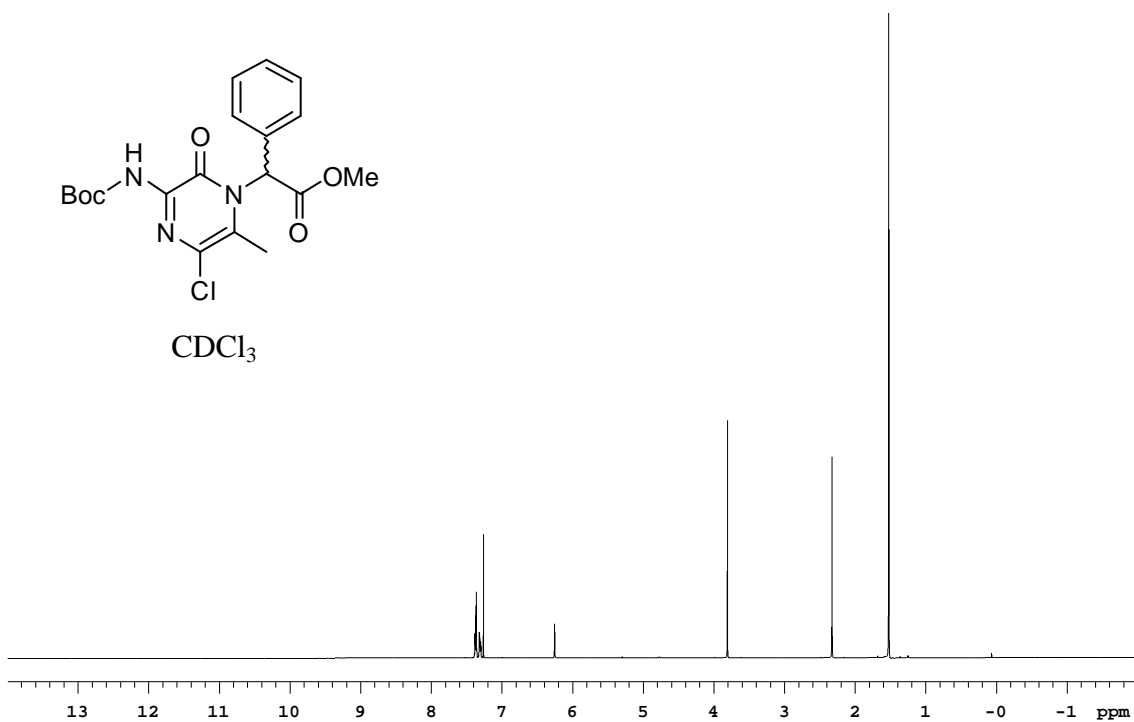
5



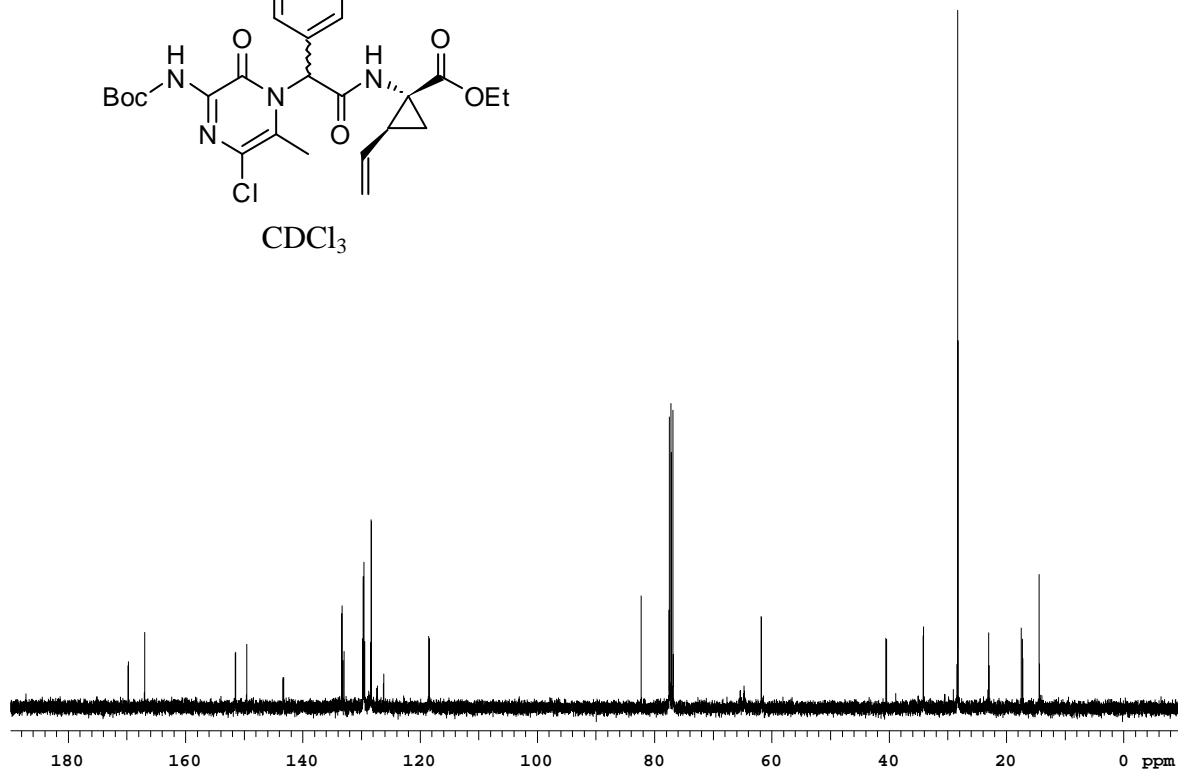
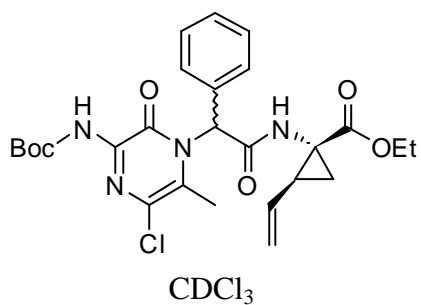
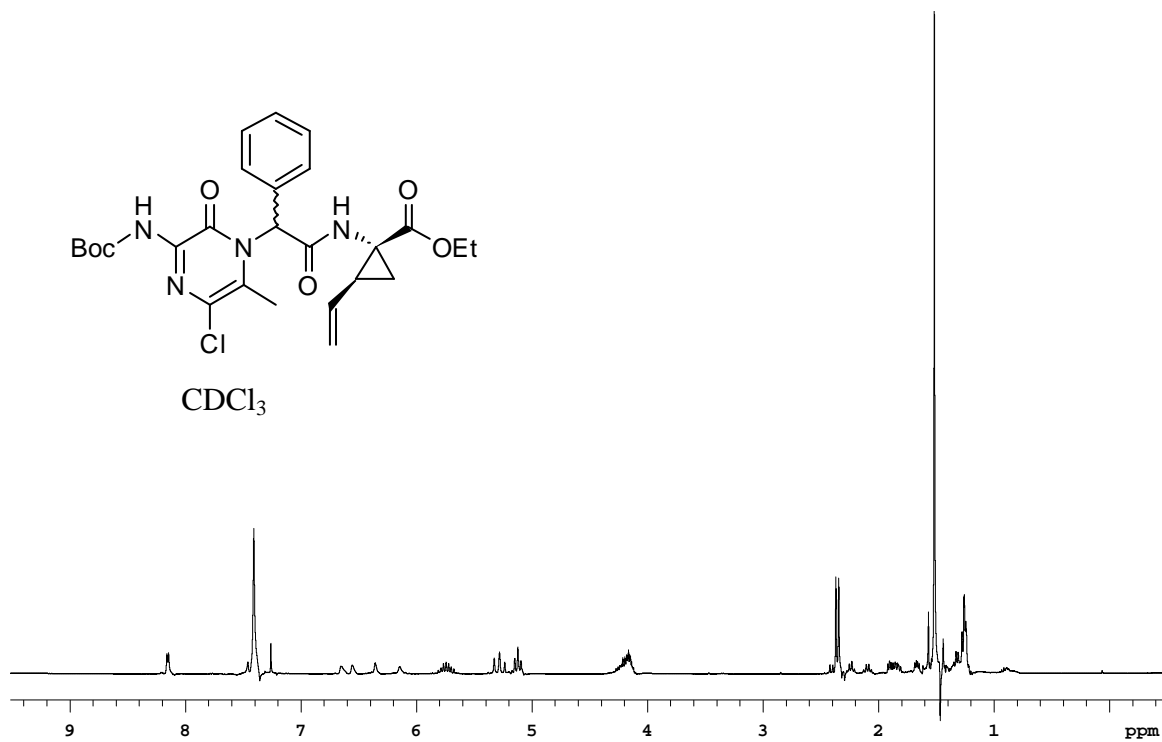
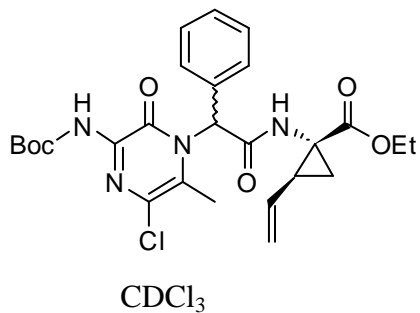
SQ_gis000169 #124 RT: 2.09 AV: 1 NL: 1.23E6
T: (0.0) + c ESI sid=10.00 Full ms [100.00-1200.00]



5

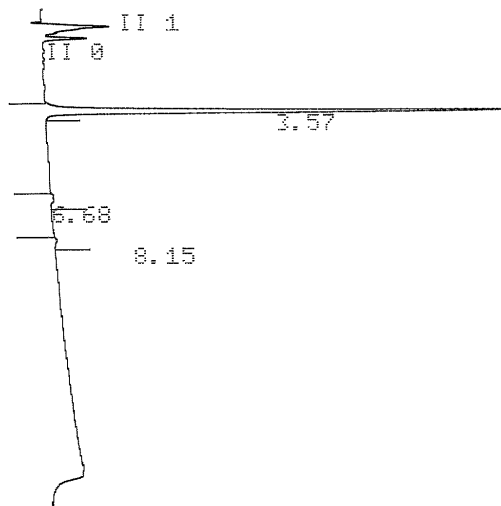


6



7a

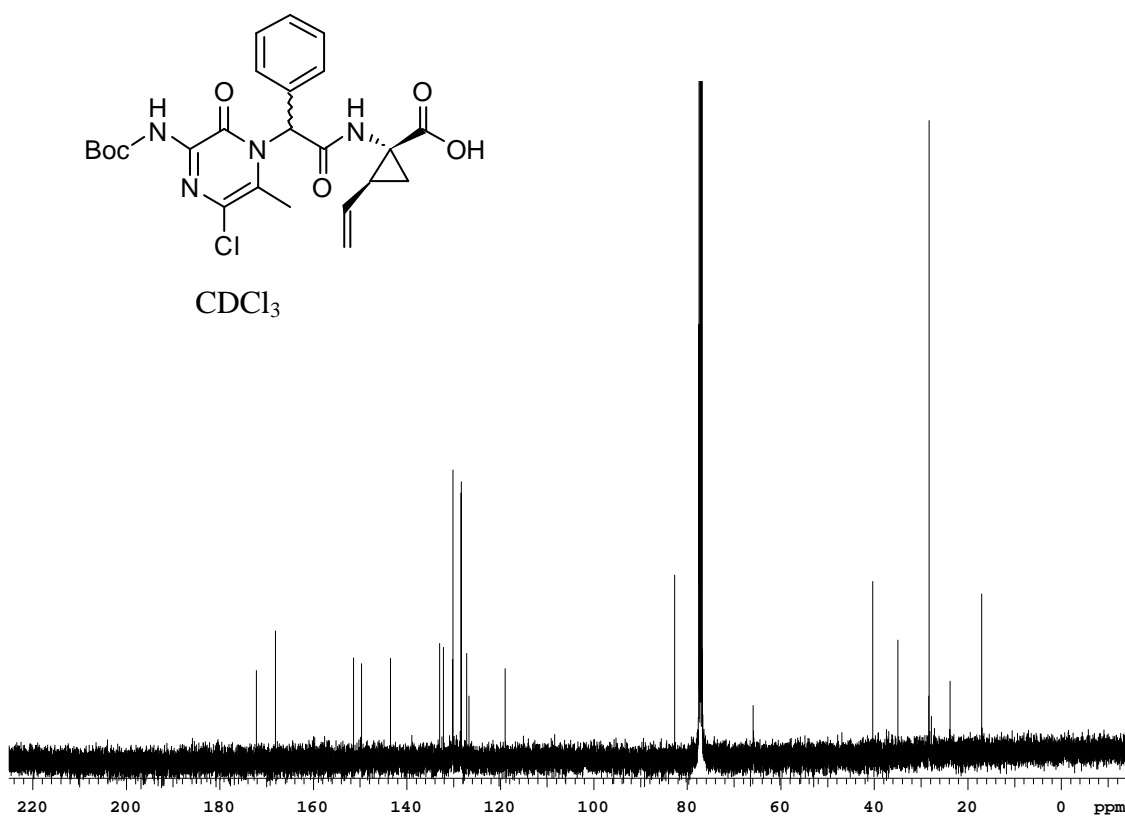
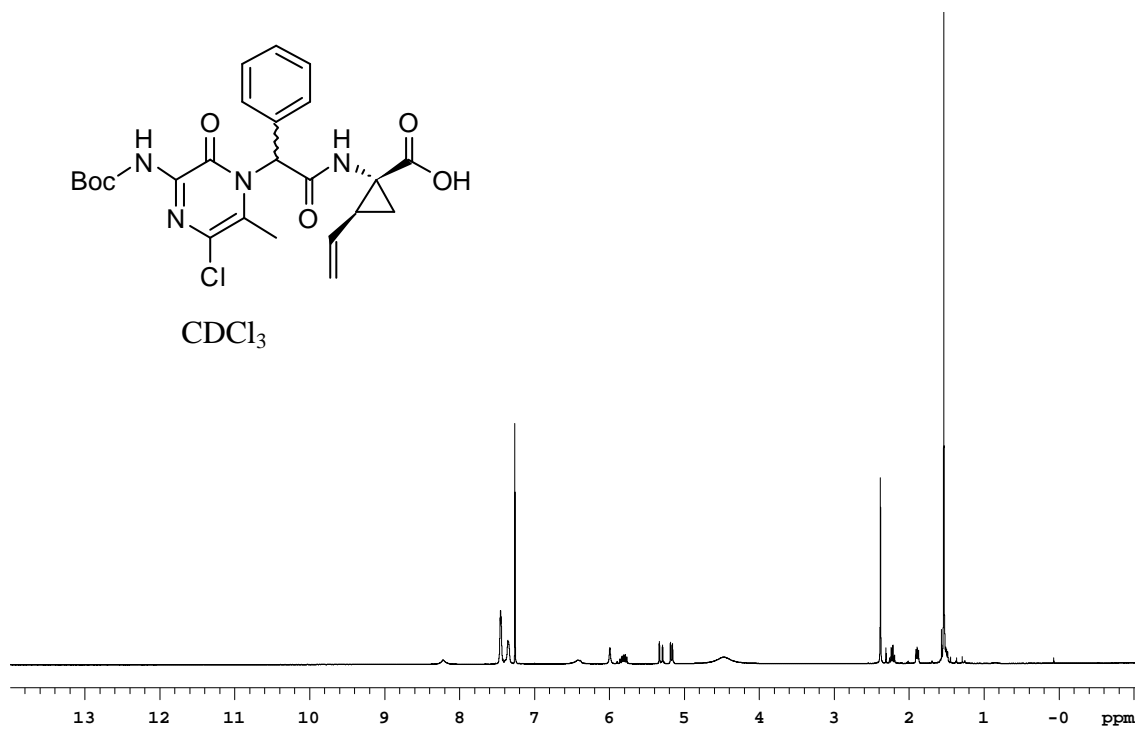
CHANNEL A INJECT 20-02-08 09:08:50



20-02-08 09:08:50 CH= "A" PS= 1.

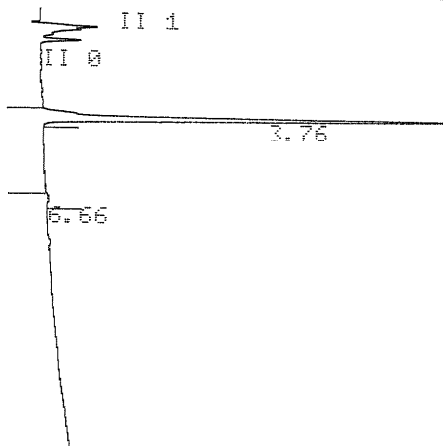
FILE	1.	METHOD	0.	RUN	9	INDEX	9
PEAK#		AREA%		RT		AREA	BC
1		97.53		3.57		406563	01
2		1.509		6.68		6289	01
3		0.961		8.15		4007	01
TOTAL		100.				416859	

7a



7b

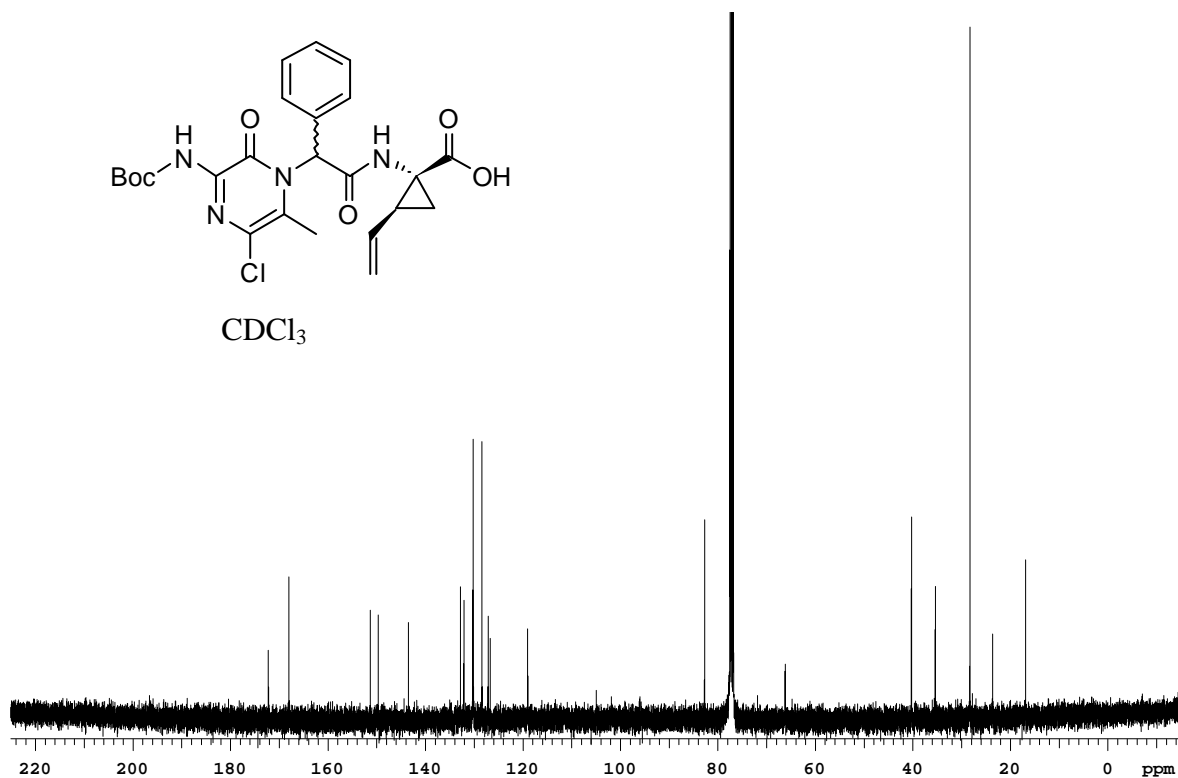
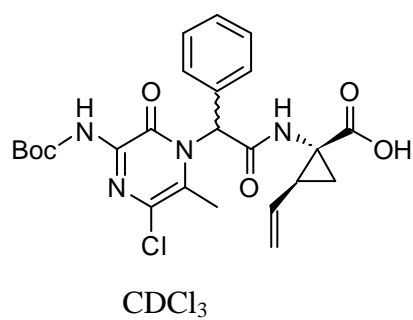
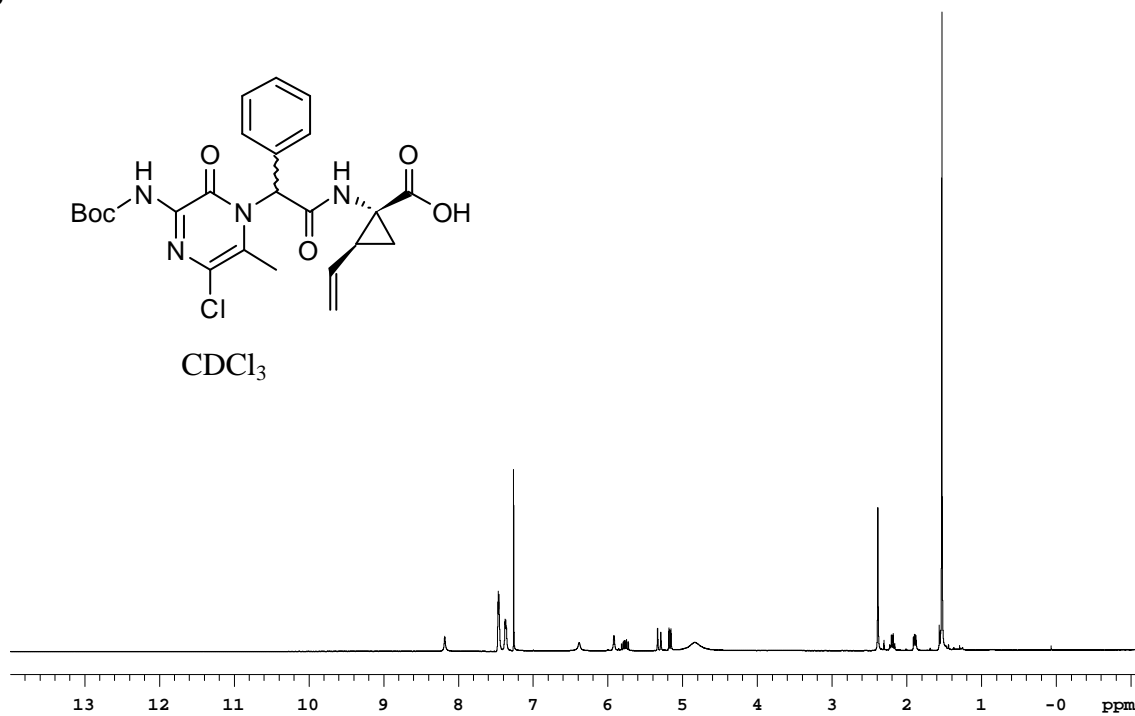
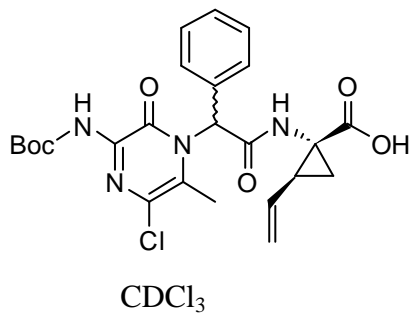
CHANNEL A INJECT 20-02-08 09:30:57



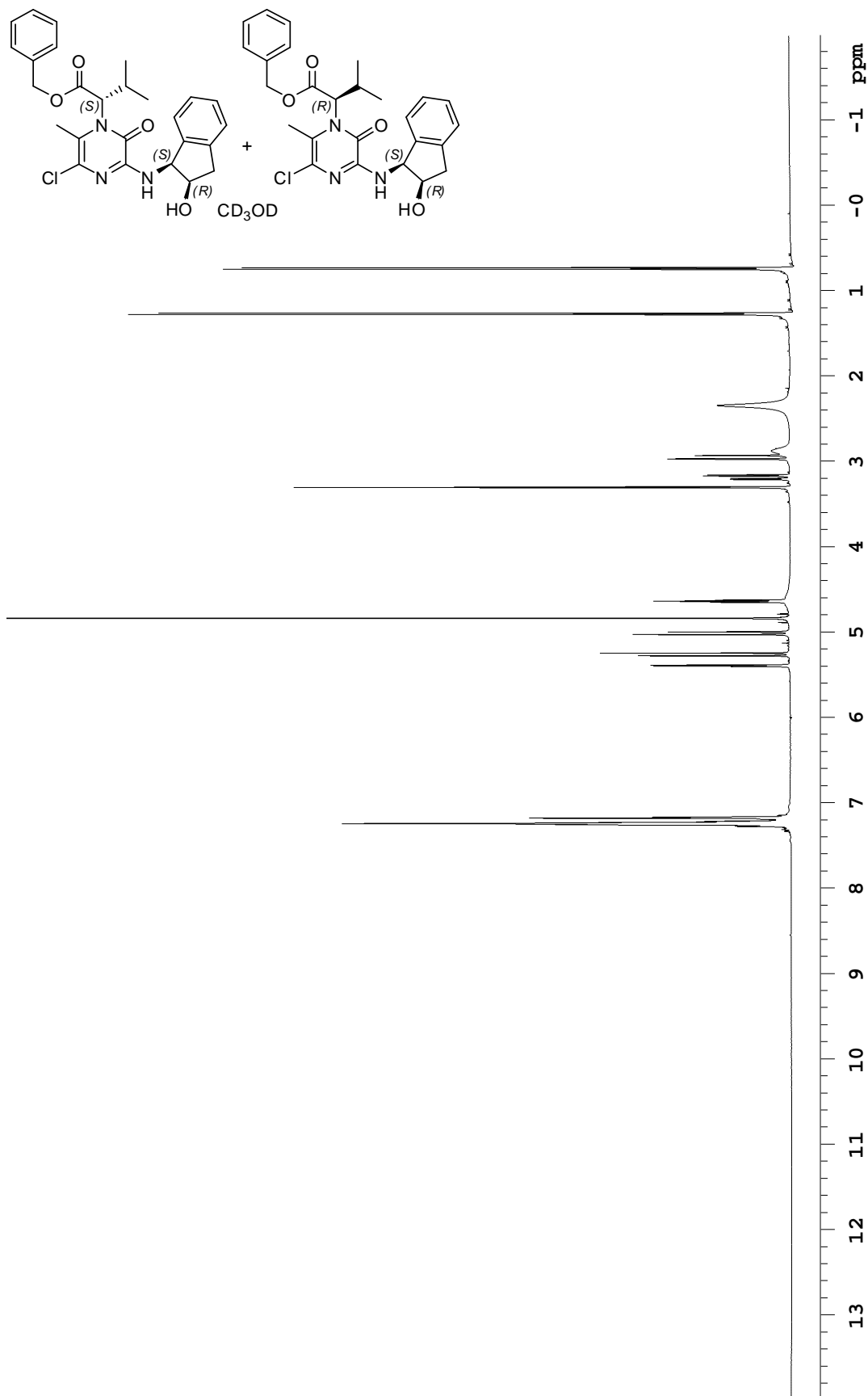
20-02-08 09:30:57 CH= "A" PS= 1.

FILE	METHOD	RT	AREA	BC
1.	0.	10	10	
PEAK#	AREA%	RT	AREA	BC
1	98.633	3.76	392885	01
2	1.367	6.66	5444	01
TOTAL	100.		398329	

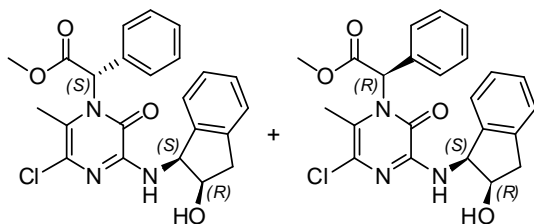
7b



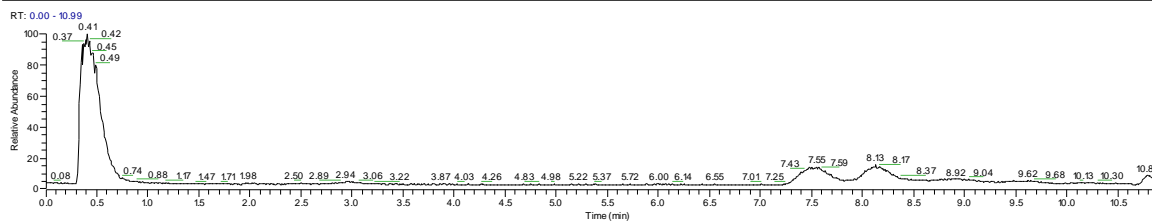
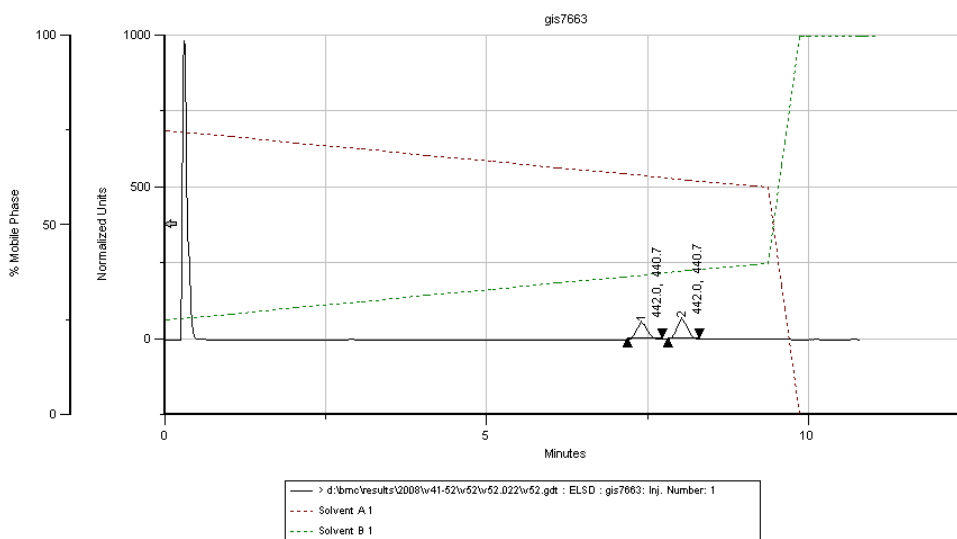
(1*S*,2*R*)-*cis*-1-amino-2-indanol derivitized **4j**



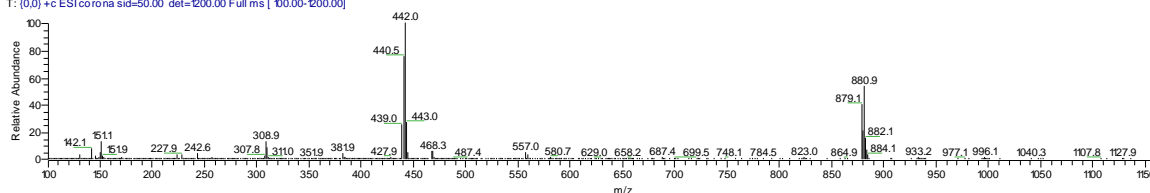
(1*S*,2*R*)-*cis*-1-amino-2-indanol derivitized **4k**



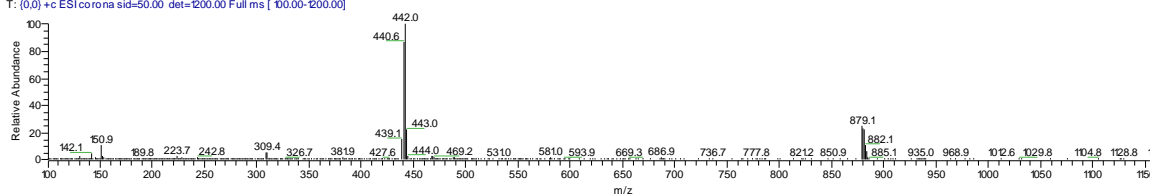
Sample Descrip.	Peak Name	R. Time	Area %	Area
1	gis7663	7.42	46.50	1149471.00
2	gis7663	8.04	53.50	1322533.25



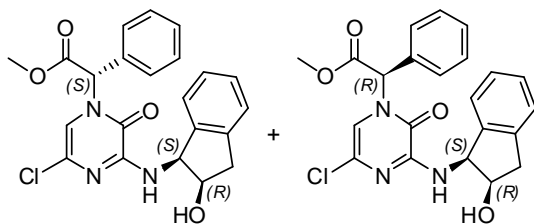
SQ_gis7663 #852 RT: 7.51 AV: 1 NL: 175E6
T: [0.0] +c ESI corona sid=50.00 del=200.00 Full ms [100.0-200.00]



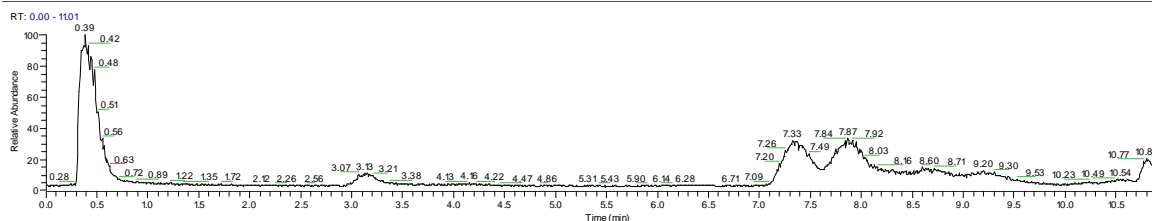
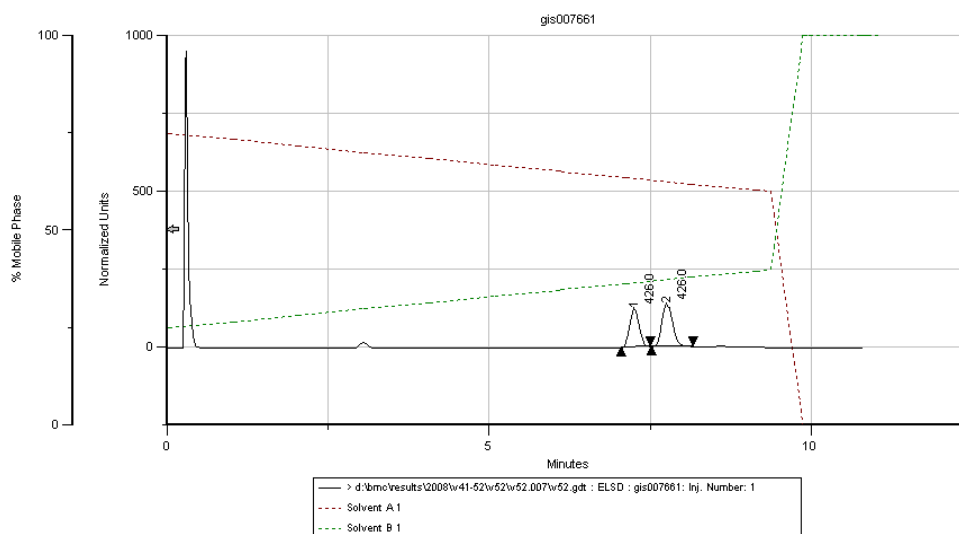
SQ_gis7663 #922 RT: 8.13 AV: 1 NL: 3.08E6
T: [0.0] +c ESI corona sid=50.00 del=200.00 Full ms [100.0-200.00]



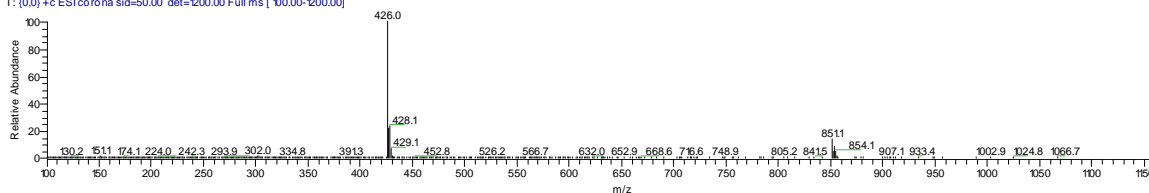
(1*S*,2*R*)-*cis*-1-amino-2-indanol derivitized **4I**



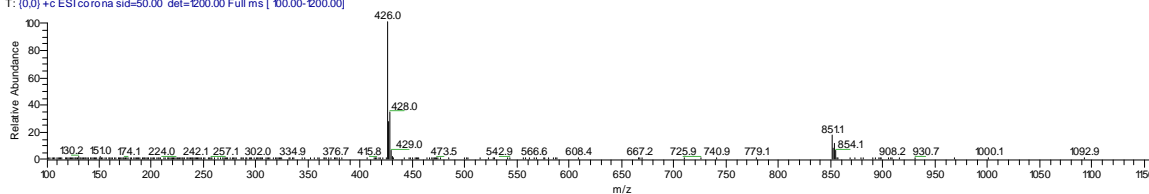
Sample Descrip.	Peak Name	R. Time	Area %	Area
gis007661	1	7.26	43.92	2162772.00
gis007661	2	7.76	56.08	2761818.25



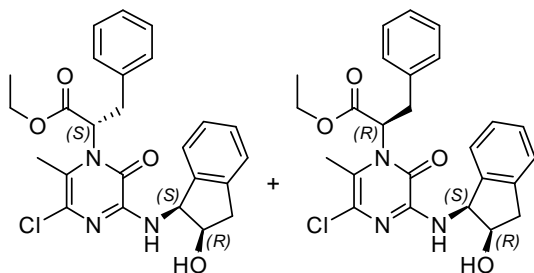
SQ_gis007661#833 RT: 7.35 AV: 1 NL: 5.12E6
T: (0.0) +c ESI corona sid=50.00 del=200.00 Full ms [10.00-200.00]



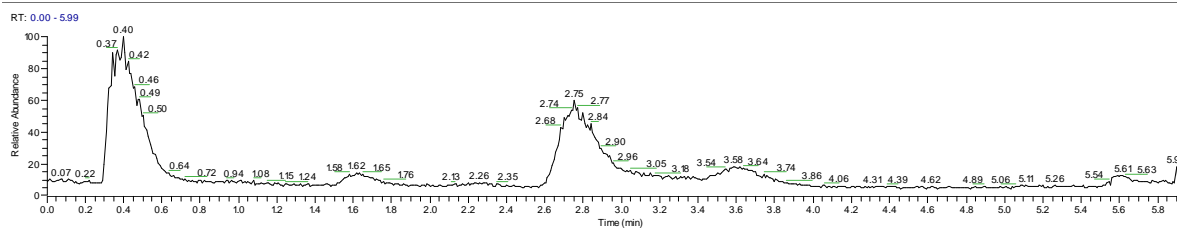
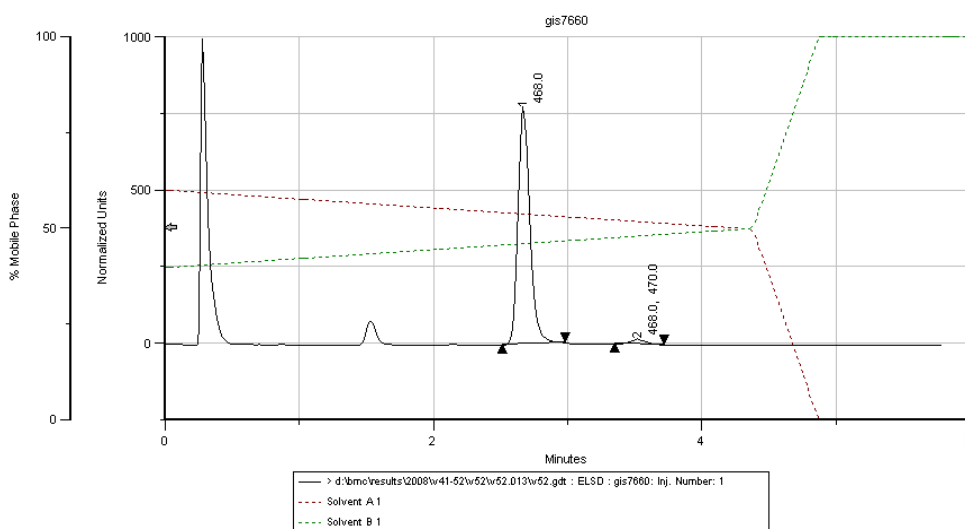
SQ_gis007661#893 RT: 7.87 AV: 1 NL: 5.06E6
T: (0.0) +c ESI corona sid=50.00 del=200.00 Full ms [10.00-200.00]



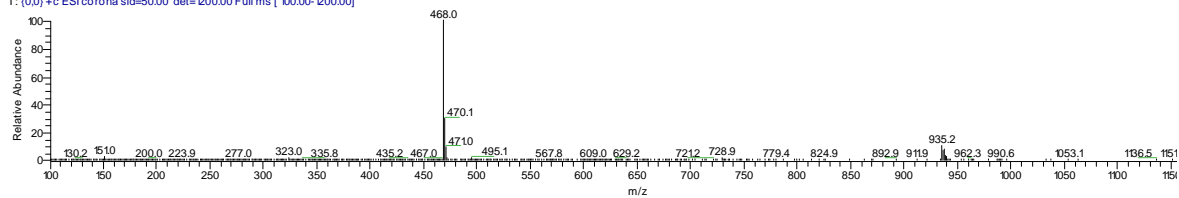
(1*S*,2*R*)-*cis*-1-amino-2-indanol derivitized **4m**



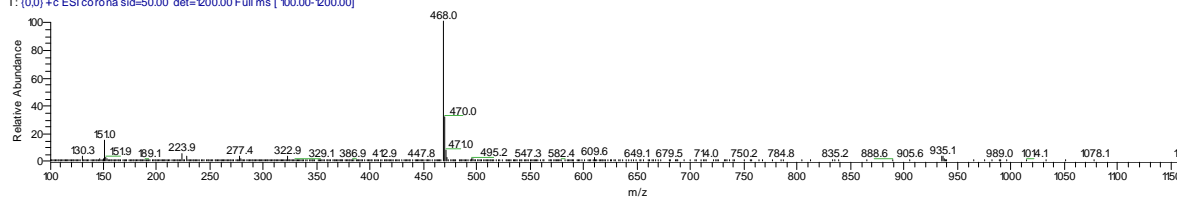
Sample Descrip.	Peak Name	R. Time	Area %	Area	
1	gis7660	*1	2.67	97.10	2307714.75
2	gis7660	*2	3.52	2.90	68963.41



SQ_gis7660 #312 RT: 2.75 AV: 1 NL: 5.85E6
T: (0,0)+c ESI corona sid=50.00 det=200.00 Full ms [100.00-200.00]



SQ_gis7660 #406 RT: 3.58 AV: 1 NL: 153E6
T: (0,0)+c ESI corona sid=50.00 det=200.00 Full ms [100.00-200.00]



(1*S*,2*R*)-*cis*-1-amino-2-indanol derivitized **4n**

