Evaluation of stereochemically dense morpholine-based scaffolds as proline surrogates in β -turn peptides

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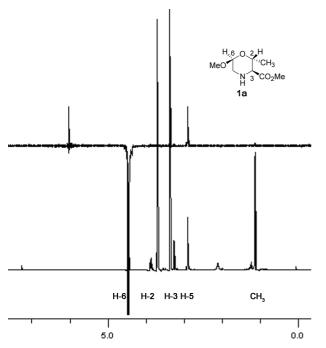


Fig. S1 NOe irradiation of H-6 of compound **1a** at 4.44 ppm (NOESY1D, mixing time of 500 ms): the nOe correlations between H-6 and H-2 are not present.

H-Gly-D-Leu-D-Val-OMe. Peptide H-Gly-D-Leu-D-Val-OMe was prepared using standard protocols for solution phase peptide synthesis using Boc-glycine Boc-D-leucine and D-valine methyl ester hydrochlorides as starting materials. $\delta_{\rm H}$ (400 MHz, DMSO-d₆) 8.47 (d, J = 8.0 Hz, 1H, NH), 8.32 (d, J = 8.0 Hz, 1 H, NH), 8.02 (br, 3 H), 4.49 (q, J = 8.0 Hz, 1 H, D-Leu H- α), 4.11 (t, J = 7.2 Hz, 1 H, D-Val H- α), 3.59 (s, 3 H, OCH₃), 2.60 (m, 2 H, Gly H- α), 2.01 (septet, J = 7.2 Hz, 1 H, D-Val H- β), 1.59 (septet, J = 6.8 Hz, 1 H, D-Leu H- γ), 1.41 (t, J = 6.8 Hz, 2 H, D-Leu H- β), 0.88-0.84 (m, 12 H, D-Leu H- δ and D-Val H- γ).

H-Mor-Gly-D-Leu-D-Val-OMe (9a). Tripeptide H-Gly-D-Leu-D-Val-OMe (125 mg, 0.30 mmol) was suspended in CH₂Cl₂ (3 mL) and DIPEA (177 μ L, 1.10 mmol) was added. The mixture was stirred until a clear solution was obtained, and Fmoc-amino acid **8** (100 mg, 0.27 mmol) and TBTU (96 mg, 0.30 mmol) were sequentially added. The reaction mixture was stirred 4 hours at room temperature and then CH₂Cl₂ was evaporated. The resultant oil was dissolved in EtOAc, washed with 1M HCl, 5% Na₂CO₃, brine and dried over anhydrous Na₂SO₄. The solution was filtered and concentrated to dryness to yield a solid that was treated with a 30% Et₂NH in CH₃CN (3 mL). The Fmoc deprotection was monitored by

TLC. When complete conversion was obtained, volatiles were removed under reduced pressure and the residue was eluted over silica gel (Et₂O/MeOH 30:1 to pure MeOH) to yield amine **9a** as a white solid (110 mg, 95%). (Found: C, 56.18; H, 8.55; N, 12.98. C₂₀H₃₆N₄O₆ requires C, 56.06; H, 8.47; N, 13.07%). $[\alpha]_D^{21} = -44.6$ (c 0.5, CHCl₃); δ_H (400 MHz, CDCl₃) 7.50 (br, 1 H, NH), 6.63 (br, 1 H, NH), 6.45 (d, J = 8.4 Hz, 1 H, NH), 4.53-4.43 (m, 2 H), 3.92 (dd, J = 15.6, 5.6 Hz, 1 H), 3.88-3.79 (m, 2 H), 3.73 (s, 3 H, OCH₃), 3.63 (t, J = 12.0 Hz, 1 H), 3.45 (m, 1 H), 3.17 (d, J = 8.0 Hz, 1 H), 2.97 (t, J = 12.0 Hz, 1 H), 2.1 (m, 1 H, D-Val H- β), 1.83 (br, 1 H, H-4), 1.55 (m, 2 H, D-Leu H- β), 1.53 (t, J = 8.0 Hz, 1 H, D-Leu H- γ); δ_C (60 MHz, CDCl₃) 172.2 (s, CO), 172.1 (s, CO), 171.9 (s, CO), 168.7 (s, <u>CO</u>₂Me), 74.8, (d), 67.3 (t), 65.4 (d, C-2), 57.2 (d), 52.1 (d), 51.9 (q, OCH₃), 44.2 (t), 42.6 (t), 41.2 (t), 31.1 (d), 24.6 (d), 22.8 (q, CH₃), 22.0 (q, CH₃), 18.9 (q, CH₃), 18.3 (q, CH₃), 17.9 (q, CH₃). ESI-MSMS *m/z* 429.47 (M⁺+1, 16), 397.19 (M⁺-OMe, 28), 298.11 [M⁺-(D-Val₂OMe), 100].

H-[(6S)-methoxy]-Mor-Gly-D-Leu-D-Val-OMe (9b). Tripeptide H-Gly-D-Leu-D-Val-OMe (120 mg, 0.29 mmol) was suspended in CH₂Cl₂ (3 mL) and DIPEA (181 µL, 1.06 mmol) was added. The mixture was stirred until a clear solution was obtained, then 4 (105 mg, 0.26 mmol) and TBTU (93 mg, 0.29 mmol) were sequentially added. The reaction mixture was stirred 4 hours at room temperature, then CH₂Cl₂ was evaporated. The resultant oil was dissolved in EtOAc, washed with 1M HCl, 5% Na₂CO₃, brine and dried over anhydrous Na₂SO₄. The solution was filtered and concentrated to dryness to yield a solid that was treated with 30% Et₂NH in CH₃CN (3 mL). The Fmoc deprotection was monitored by TLC, and after complete conversion was obtained, volatiles were removed under reduced pressure and the residue was eluted over silica gel (Et₂O/MeOH 30:1 to pure MeOH) to yield amine **9b** as a white solid (114 mg, 96%). (Found: C, 55.12; H, 8.42; N, 12.13. C₂₁H₃₈N₄O₇ requires C, 55.00; H, 8.35; N, = 8.4 Hz, 1 H, NH), 6.44 (d, J = 8.8 Hz, 1 H, NH), 4.55 (s, 1 H, H-6), 4.50 (dd, J = 8.8, 4.0 Hz, 1 H), 4.46-4.41 (m, 1 H), 3.98 (dd, J = 16.0, 6.0 Hz, 1 H), 3.91 (m, 1 H), 3.84 (dd, J = 16.0, 6.0 Hz, 1 H), 3.73 (s, 3 H, CO₂CH₃), 3.39 (s, 3 H, OCH₃), 3.13 (d, *J* = 9.6 Hz, 1 H), 2.96 (dd, *J* = 13.2, 3.6 Hz, 1 H), 2.89 $(d, J = 13.2 \text{ Hz}, 1 \text{ H}), 2.10 \text{ (m 1 H, D-Val H-}\beta), 1.71 \text{ (br, 1 H, H-}4), 1.69-1.61 \text{ (m, 2 H, D-Leu H-}\beta),$ 1.55-1.50 (m, 1 H, D-Leu H- γ), 1.25 (d, J = 6.4 Hz, 3 H, CH₃-C-2), 0.94-0.88 (m, 12 H, D-Leu H- δ and D-Val H-γ); δ_C (60 MHz, CDCl₃) 172.0 (s, CO), 171.8 (s, CO), 171.4 (s, CO), 168.5 (s, <u>CO</u>₂Me), 95.9 (d, C-6), 65.7 (t), 65.7 (d), 64.3 (d), 57.2 (d), 54.5 (q, OCH₃), 52.1 (d), 51.9 (d), 47.2 (t), 42.8 (t), 41.3

(t), 31.1 (d), 24.7 (d), 22.8 (q), 22.2 (q), 19.0 (q), 18.5 (q), 17.9 (q). ESI-MSMS *m*/*z* 459.46 (M⁺+1, 67), 427.00 (M⁺-OMe, 100), 322.23 [M⁺-(OMe)Mor+Na, 100].

H-[(6R)-methoxy]-Mor-Gly-D-Leu-D-Val-OMe (9c). Tripeptide H-Gly-D-Leu-D-Val-OMe (120 mg, 0.29 mmol) was suspended in CH₂Cl₂ (3 mL) and DIPEA (181 μ L, 1.06 mmol) was added. The mixture was stirred until a clear solution was obtained and Fmoc-amino acid 5 (105 mg, 0.26 mmol) and TBTU (93 mg, 0.29 mmol) were sequentially added. The reaction mixture was stirred 4 hours at room temperature and then CH₂Cl₂ was evaporated. The resultant oil was dissolved in EtOAc, washed with 1M HCl, 5% Na₂CO₃, brine and dried over anhydrous Na₂SO₄. The solution was filtered and concentrated to dryness to yield a solid that was treated with 30% Et₂NH in CH₃CN (3 mL). The Fmoc deprotection was monitored by TLC. When complete conversion was obtained, volatiles were removed under reduced pressure and the residue was eluted over silica gel (Et₂O/MeOH 30:1 to pure MeOH) to yield amine **9c** as a white solid (90 mg, 75%). (Found: C, 55.07; H, 8.39; N, 12.17. C₂₁H₃₈N₄O₇ requires C, 55.00; H, 8.35; N, 12.22%). $[\alpha]_D^{20} = -36.5$ (c 0.84, CH₂Cl₂); δ_H (400 MHz, CDCl₃) 7.34 (t, J = 7.2Hz, 1 H, Gly NH), 6.54 (d, J = 12.4, 1 H, D-Leu NH), 6.39 (d, J = 12.4, 1 H, D-Val NH), 4.51 (dd, J = 8.7, 4.8 Hz, 1 H, D-Val H- α), 4.46-4.43 (m, 2 H, D-Leu H- α + H-6), 4.01 (dd, J = 15.9, 6.1, 1 H, Gly H- α), 3.81 (dd, J = 15.9, 5.5 Hz, 1, Gly H- α), 3.72 (s, 3 H, CO₂CH₃), 3.61 (dq, J = 8.8, 6.2 Hz, 1 H, H-2), 3.50 (s, 3 H, OCH₃), 3.08 (d, J = 8.8 Hz, 1 H, H-3), 3.03 (dd, J = 11.7, 2.4 Hz, 1 H, H-5), 2.61 (dd, J = 11.7, 2.4 Hz, 1 H, H-5), 2.411.6, 8.5, Hz, 1 H, H-5), 2.15 (m, 1 H, D-Val H- β), 1.66 (m, 3 H, D-Leu H- β/γ), 1.29 (d, J = 6.3 Hz, 3 H, CH₃-C-2), 0.95-0-88 (m, 12 H, D-Leu H- δ and D-Val H- γ). δ_{C} (50 MHz, CDCl₃) 172.2 (s, CO), 172.0 (s, CO), 171.4 (s, CO), 168.4 (s, CO₂Me), 100.4 (d, C-6), 74.8, (d), 73.8 (d), 64.1 (d), 57.1 (d), 56.1 (q, OCH₃), 52.2 (d), 51.8 (d), 48.0 (t), 42.6 (t), 41.5 (t), 31.2 (d), 24.7 (d), 22.8 (q, CH₃), 22.2 (q, CH₃), 19.0 (q, CH₃), 18.1 (q, CH₃), 18.0 (q, CH₃). ESI-MSMS m/z 481.85 (M⁺+Na, 14), 449.33 (M⁺-OMe+Na, 93), 322.23 [M⁺-(OMe)Mor+Na, 100].

H-D-Val-Mor-Gly-D-Leu-D-Val-OMe (10a). 2,6-Lutidine (112 μ L, 0.96 mmol) was added to a solution of peptide 9a (135 mg, 0.32 mmol) in CH₂Cl₂ (3 mL). Solid Fmoc-D-Val-Cl (114 mg, 0.32 mmol) was added in small portions. The reaction mixture was stirred at room temperature for 3 hours and dichloromethane was then removed under reduced pressure. The resultant oil was dissolved in EtOAc, washed with 1M HCl, 5% Na₂CO₃, brine and dried over anhydrous Na₂SO₄. The solution was filtered and concentrated to dryness to yield a solid that was treated with 30% Et₂NH in CH₃CN (3 mL). The Fmoc deprotection was monitored by TLC, until complete conversion was obtained, then volatiles were removed under reduced pressure and the residue was eluted over silica gel (Et₂O/MeOH 30:1 to

pure MeOH) to yield amine **10a** as a white solid (140 mg, 83%). (Found: C, 56.99; H, 8.56; N, 13.21. $C_{25}H_{45}N_5O_7$ requires C, 56.91; H, 8.60; N, 13.27%). $[\alpha]_D^{21} = +10.5$ (c 1.00, CHCl₃); δ_H (400 MHz, CDCl₃) 7.15 (d, J = 7.6 Hz, 1H, NH), 7.06 (br, 1 H, NH), 6.55 (d, J = 8.4 Hz, 1 H, NH), 4.85 (m, 2 H), 4.34 (m, 1 H), 4.28 (d, J = 5.6 Hz, 1 H), 3.97 (d, J = 6.0 Hz, 2 H), 4.0-3.82 (m, 1 H), 3.80-3.62 (m, 2 H), 3.73 (s, 3H, CO₂CH₃), 3.63-3.57 (m, 1 H), 3.50 (d, J = 6.4 Hz, 1 H), 2.17-2.04 (m, 1 H), 1.97-1.82 (m, 4 H), 1.68-1.65 (m, 2 H), 1.32 (d, J = 6.4 Hz, 3 H), 1.02-0.88 (m, 18 H, D-Leu H- δ and D-Val H- γ). δ_C (60 MHz, CDCl₃) δ 176.3 (s, CO), 171.9 (s, CO), 171.8 (s, CO), 169.6 (s, CO), 169.1 (s, <u>CO₂Me)</u>, 69.3 (d), 61.1 (d), 59.9 (d), 57.3 (d), 56.6 (q, OCH₃), 20.0 (q, CH₃), 19.0 (q, CH₃), 17.9 (q, CH₃), 17.1 (q, CH₃). ESI-MSMS *m*/*z* 527.52 (M⁺, 18), 496 (18, M⁺-OMe), 397.07 (M⁺-D-Val₂OMe, 78), 269.25 [M⁺-(D-Val₂OMe)-(C=O), 35], 284.15 [M⁺-(D-Leu-D-Val₂OMe), 4], 227.04 [M⁺-(Gly-D-Leu-D-Val₂OMe), 100].

H-D-Val-[(6S)-methoxy]-Mor-Gly-D-Leu-D-Val-OMe (10b). 2,6-Lutidine (85 µL, 0.73 mmol) was added to a solution of peptide 9b (118 mg, 0.24 mmol) in CH₂Cl₂ (3 mL). Solid Fmoc-D-Val-Cl (86 mg, 0.24 mmol) was added in small portions. The reaction mixture was stirred at room temperature for 4 hours and dichloromethane was then removed under reduced pressure. The resultant oil was dissolved in EtOAc, washed with 1M HCl, 5% Na₂CO₃, brine and dried over anhydrous Na₂SO₄. The solution was filtered and concentrated to dryness to yield a solid that was treated with 30% Et₂NH in CH₃CN (3 mL). The Fmoc deprotection was monitored by TLC. When complete conversion was obtained, volatiles were removed under reduced pressure and the residue was eluted over silica gel (Et₂O/MeOH 30:1 to pure MeOH) to yield amine 10b as a white solid (114 mg, 85%). (Found: C, 56.21; H, 8.61; N, 12.45. $C_{26}H_{47}N_5O_8$ requires C, 56.00; H, 8.49; N, 12.56%). $[\alpha]_D^{21} = +7.5$ (c 5.50, CH₃CN); δ_H (400 MHz, CDCl₃) 7.11 (d, J = 7.6 Hz, 2 H, NH), 6.56 (d, J = 8.8 Hz, 1 H, NH), 4.81 (dd, J = 7.6, 5.6 Hz, 1 H, H-6), 4.42 (m, 2 H), 4.31 (dt, J = 16.0, 6.4 Hz, 1 H), 4.05 (dd, J = 18.0, 9.6 Hz, 2 H), 3.98 (dd, J = 14.0, 5.6 Hz, 1 H), 3.85 (dd, J = 17.2, 6.0 Hz, 1 H), 3.71 (s, 3 H, CO₂CH₃), 3.53 (d, J = 5.6 Hz, 1 H), 3.42 (dd, J = 17.2, 6.0 Hz, 1 14.4, 8.4 Hz, 1 H), 3.41 (s, 3 H, CO_2CH_3), 2.07 (septet, J = 6.8 Hz, 1 H), 1.98 (br, 2 H, NH_2), 1.89 (septet, J = 6.4 Hz, 1 H), 1.67 (d, J = 6.4, 2 H, D-Leu H- β), 1.35 (d, J = 6.4 Hz, 3 H, CH₃-C-2), 0.99-0.86 (m, 18 H, D-Leu H- δ and D-Val H- γ). δ_{C} (60 MHz, CDCl₃) 175.3 (s, CO), 171.8 (s, CO), 171.7 (s, CO), 169.4 (s, CO), 169.0 (s, CO₂Me), 97.2 (d, C-6), 63.9 (d), 63.0 (d), 57.3 (d), 56.4 (d), 55.0 (q, OCH₃), 52.0 (d), 51.7 (d), 44.1 (t), 42.9 (t), 40.0 (t), 31.8 (q, CH₃), 30.9 (d), 24.6 (d), 23.0 (q, CH₃), 22.0

(q, CH₃), 19.9 (q, CH₃), 19.0 (q, CH₃), 17.8 (q, CH₃), 17.0 (q, CH₃). ESI-MSMS *m*/*z* 557.92 (M⁺+1, 10), 526.23 (M⁺-OMe+1, 100).

H-D-Val-[(6R)-methoxy]-Mor-Gly-D-Leu-D-Val-OMe (10c). 2,6-Lutidine (85 µL, 0.57 mmol) was added to a solution of peptide 9c (80 mg, 0.16 mmol) in CH₂Cl₂ (3 mL). Solid Fmoc-D-Val-Cl (57 mg, 0.16 mmol) was added in small portions. The reaction mixture was stirred at room temperature for 4 hours and dichloromethane was then removed under reduced pressure. The resultant oil was dissolved in EtOAc, washed with 1M HCl, 5% Na₂CO₃, brine and dried over anhydrous Na₂SO₄. The solution was filtered and concentrated to dryness to yield a solid that was treated with 30% Et₂NH in CH₃CN (3 mL). The Fmoc deprotection was monitored by TLC. When complete conversion was obtained, volatiles were removed under reduced pressure and the residue was eluted over silica gel (Et₂O/MeOH 30:1 to pure MeOH) to yield amine 10c as a white solid (87 mg, 98%). (Found: C, 56.13; H, 8.53; N, 12.51. $C_{26}H_{47}N_5O_8$ requires C, 56.00; H, 8.49; N, 12.56%). $[\alpha]_D^{21} = +2.3$ (c 1.0, CH₃CN); ¹H-NMR (400 MHz, CDCl₃) δ 7.49 (d, *J* = 7.2 Hz, 1 H, NH), 6.71 (br, 1 H, NH), 4.80 (t, *J* = 2.8 Hz, 1 H, H-6), 4.45-4.39 (m, 2 H), 4.2 (q, J = 6.4 Hz, 1 H), 4.12 (d, J = 8.4 Hz, 1 H), 4.08 (d, J = 7.2 Hz, 1 H), 3.80 (dd, J = 17.2, 5.2Hz, 1 H), 3.75-3.67 (m, 2 H), 3.71 (s, 3 H, CO₂CH₃), 3.42 (s, 3 H, CO₂CH₃), 3.37 (d, J = 6.4 Hz, 1 H), 2.14 (septet, J = 6.8 Hz, 1 H), 2.04 (septet, J = 6.0 Hz, 1 H), 1.71 (br, 2 H, NH₂), 1.32 (d, J = 6.0 Hz, 3 H, CH₃-C-2), 0.96-0.80 (m, 18 H, D-Leu H- δ and D-Val H- γ). δ_{Γ} (60 MHz, CDCl₃) 176.5 (s, CO), 171.8 (s, CO), 171.7 (s, CO), 169.7 (s, CO), 169.2 (s, CO₂Me), 97.1 (d, C-6), 68.1 (d), 61.9 (d), 57.2 (d), 56.7 (d), 55.2 (q, OCH₃), 52.0 (d), 51.8 (t), 45.2 (t), 43.0 (t), 39.7 (q, CH₃), 31.0 (d), 24.6 (d), 23.0 (q, CH₃), 21.9 (q, CH₃), 20.1 (q, CH₃), 19.3 (q, CH₃), 19.0 (q, CH₃), 17.8 (q, CH₃), 16.7 (q, CH₃). ESI-MSMS *m/z*. 558.40 (M⁺+1, 15), 526.10 (M⁺-OMe+1, 100).

		I		i for peptides I – IV in C II		III		IV	
	CDCl ₃	CD ₃ CN	CDCl ₃	CD ₃ CN	CDCl ₃	CD ₃ CN	CDCl ₃	CD ₃ CN	
Boc	1.48	1.42	1.48	1.42	1.46	1.42	1.49	1.43	
D-Ala NH	5.32	5.71	5.29	5.63	5.32	5.67	5.27	5.73	
D-Ala H-α	4.24	4.13	4.30	4.16	4.26	4.13	4.12	4.15	
D-Ala H-β	1.32	1.23	1.31	1.20	1.32	1.20	1.32	1.22	
D-Val ₁ NH	7.65	7.37	7.79	7.60	7.68	7.55	7.50	7.23	
$D-Val_1$ H- α	4.48	4.56	4.49	4.58	4.56	4.55	4.38	4.62	
$D-Val_1 H-\beta$	2.16	2.06	2.16	2.07	2.19	2.14	2.15	2.12	
$D-Val_1 H-\gamma$	1.03	0.96	0.97	0.87	0.98	0.88	0.97	0.97	
H-2	4.42	3.87-3.66	4.35-4.14	4.15-4.01	4.27	4.18-4.13	5.13-4.49	5.00-4.97	
CH ₃ -2	1.33	1.25	1.35	1.30	1.33	1.27	1.32	1.23	
H-3	4.35	3.86-3.67	4.31	4.18	4.15	4.18-4.13	4.87	4.63	
H-5	3.78-3.75	3.62-3.59	4.16-3.37	4.20-3.24	3.96-3.76	3.84-3.76	5.96	5.94	
H-6	3.90-3.74	3.60	4.85	4.86	4.83	4.84	6.44	6.46	
OCH ₃ -6	-	-	3.43	3.36	3.45	3.40	-	-	
Gly NH	7.08	7.35	6.79	7.30	6.72	7.25	7.50	7.47	
Gly H-α	3.99-3.91	3.76	4.15-3.72	3.85,3.55	4.07-3.78	3.80,3.64	3.72	3.78	
D-Leu NH	7.11	7.13	7.16	7.28	7.40	7.40	6.53	6.61	
D-Leu H-α	4.43	4.44	4.42	4.46	4.44	4.48	4.39	4.36	
D-Leu H-β	1.69	1.68-1.53	1.70	1.73-1.60	1.72	1.72-1.59	1.70-1.59	1.55	
D-Leu H-γ	1.69	1.68-1.53	1.70	1.73-1.58	1.72	1.71	1.59	1.41	
D-Leu H-δ	0.98	0.92	0.95	0.92	0.96	0.90	0.94	0.91	
D-Val ₂ NH	6.69	7.01	6.71	7.02	6.62	7.03	6.74	6.92	
$D-Val_2$ H- α	4.49	4.27	4.49	4.25	4.49	4.25	4.49	4.27	
$D-Val_2 H-\beta$	2.18	2.10	2.18	2.07	2.31	2.09	2.19	2.10	
$D-Val_2$ H- γ	0.94	0.90	0.94	0.90	0.94	0.90	0.94	0.91	
CO ₂ CH ₃	3.76	3.67	3.76	3.67	3.76	3.68	3.76	3.67	

Table S1. ¹H chemical shift data for peptides I - IV in CDCl₃ and CD₃CN

Supplementary Material (ESI) for Organic and Biomolecular Chemistry This journal is The Royal Society of Chemistry 2009

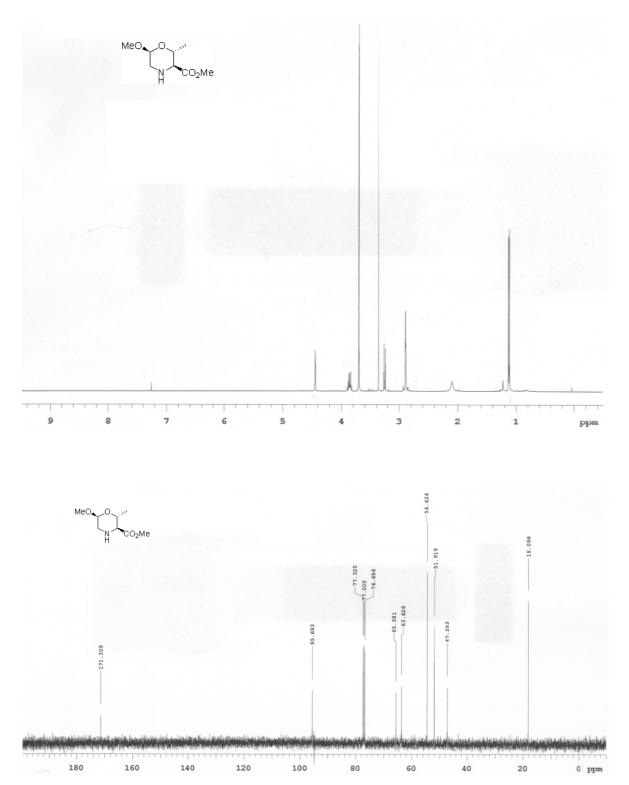
Table S2.	¹³ C chei	mical shi	ft data f	or peptid	es I – IV i	in CDCl ₃	and CD ₃ C	CN
		Ι		II	II	I	IV	7
	CDCl ₃	CD ₃ CN	CDCl ₃	CD ₃ CN	CDCl ₃	CD ₃ CN	CDCl ₃	CD ₃ CN
Boc	28.2	27.5	27.8	27.5	27.8	27.6	28.5	27.5
D-Ala NH	-	-	-	-	-	-	-	-
D-Ala C-α	50.1	49.8	50.1	49.7	49.8	49.8	49.4	49.8
D-Ala C-β	17.9	16.9	19.2	17.5	18.5	17.6	16.5	16.6
D-Val ₁ NH	-	-	-	-	-	-	-	-
$D-Val_1 C-\alpha$	55.1	54.1	54.7	53.7	54.4	54.1	55.1	54.7
$D-Val_1 C-\beta$	30.8	30.1	30.8	30.1	30.2	29.8	29.5	29.8
$D-Val_1 C-\gamma$	18.2	18.5	18.6	18.7	22.5	17.9	21.5	22.2
C-2	68.7	60.4	62.7	63.0	67.3	67.3	57.7	69.7
CH ₃ -2	17.8	17.0	18.9	17.8	19.2	18.4	16.8	16.7
C-3	60.1	42.5	64.0	63.7	61.7	62.0	56.4	56.4
C-5	42.5	51.5	44.1	43.8	45.1	42.5	102.5	103.9
C-6	60.1	42.4	97.2	97.2	97.2	97.2	129.4	130.5
OCH ₃	-	-	55.1	54.1	54.7	54.4		-
Gly NH	-	-	-	-	-	-	-	-
Gly C-α	43.1	42.5	43.1	42.1	42.8	42.5	42.7	42.8
D-Leu NH	-	-	-	-	-	-	-	-
D-Leu C-α	51.8	51.1	51.7	50.7	51.7	50.8	51.7	51.1
D-Leu C-β	39.8	24.2	39.8	40.1	39.4	40.1	39.8	40.5
D-Leu C-y	24.5	17.5	24.5	24.2	24.2	24.2	24.2	27.5
D-Leu C-δ	19.2	17.3	19.5	22.2	17.9,17.5	20.5	18.5,17.5	17.9
D-Val ₂ NH	-	-	-	-	-	-	-	-
$D-Val_2 C-\alpha$	57.4	57.7	57.7	57.7	56.7	57.7	56.7	57.4
$D-Val_2 C-\beta$	30.8	29.8	30.8	30.1	30.2	29.8	30.5	29.8
$D-Val_2 C-\gamma$	22.8	20.9	22.8	20.9	21.5	20.6	21.5	20.5
CO ₂ CH ₃	52.1	51.4	52.1	51.1	51.7	51.4	52.1	51.4

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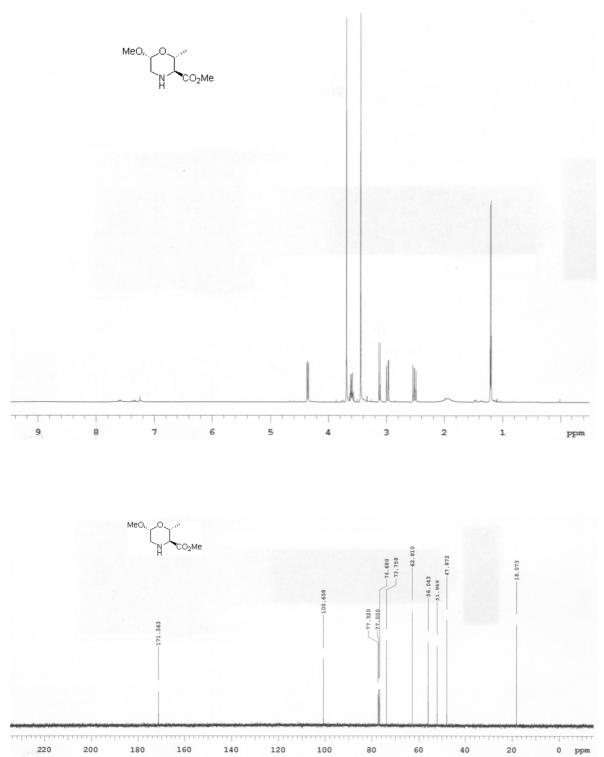
ROESY cross-peaks	Ι	II	III	IV
$D-Val_1 H-\alpha / H-5$	4.56 / 3.62-3.59	4.58 / 4.20	4.55 / 3.84	-
$D-Val_1 NH / D-Ala H-\alpha$	7.37 / 4.13	7.60 / 4.16	7.55 / 4.13	7.23 / 4.15
D -Val ₁ NH / D-Ala H- β	-	7.60 / 1.20	7.55 / 1.20	-
H-2 / H-6	-	-	4.16 / 4.84	-
Gly NH / H-2	7.35 / 3.87	7.30 / 4.07	7.25 / 4.15	7.47 / 4.98
Gly NH / D Leu H- α	-	7.30 / 4.44	-	-
Gly H-a / H-3	-	-	3.81 / 4.18	-
Gly NH / CH ₃ -2	7.35 / 1.25	-	7.25 / 1.27	-
D-Leu NH / Gly H-α	-	7.28 / 3.57	7.40 / 3.80-3.65	6.61 / 3.78
D-Leu NH / Gly NH	-	-	7.40 / 7.25	6.61 / 7.47
D -Val ₂ NH / D-Leu H- α	7.01 / 4.44	7.02 / 4.46	7.03 / 4.48	6.92 / 4.36

Table S3. ROESY data in CD₃CN for compounds I-IV.

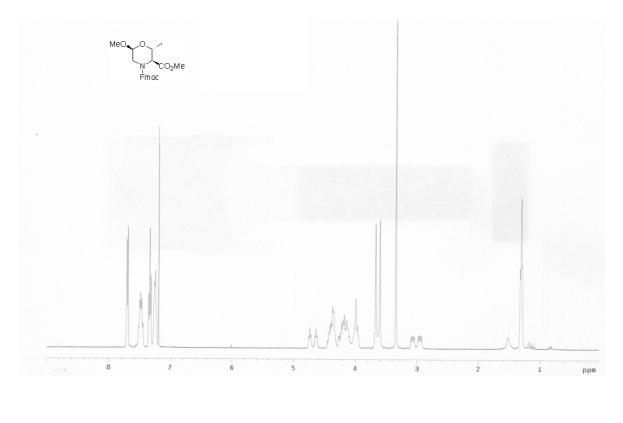
Compound 1a

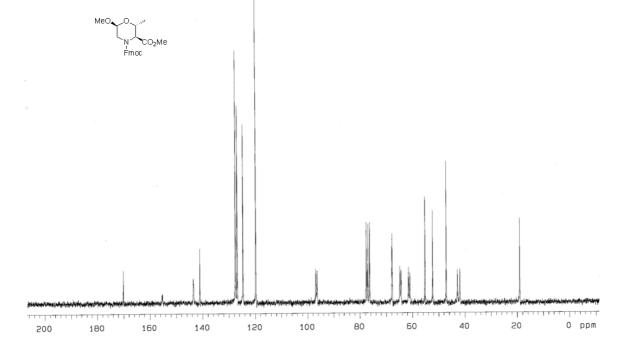


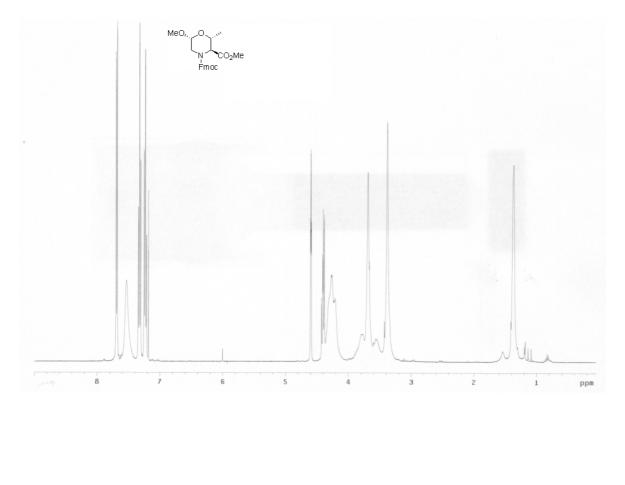
Compound 1b

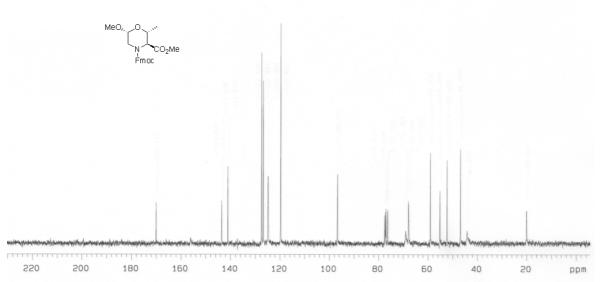


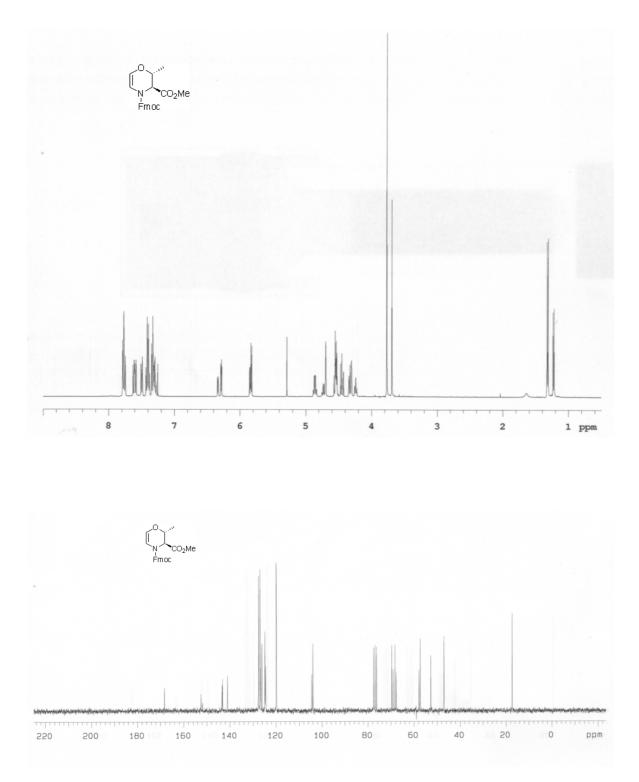
S11

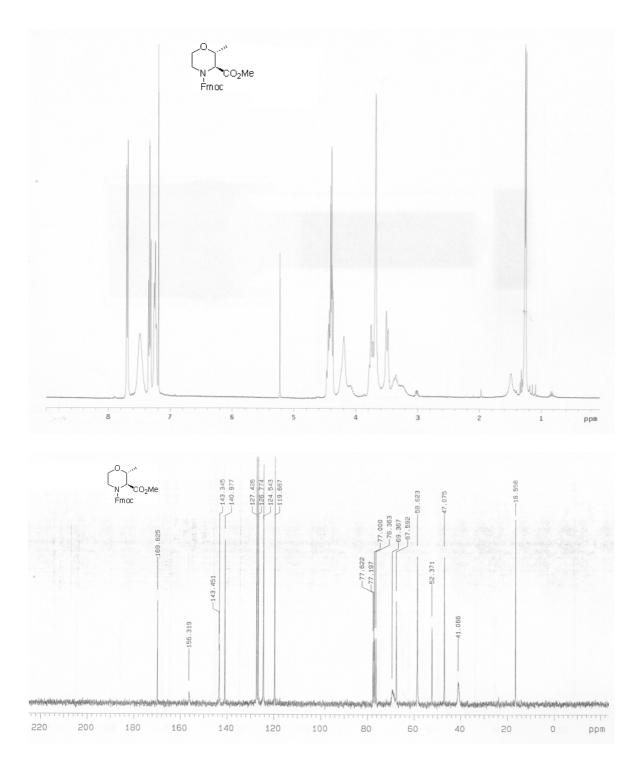


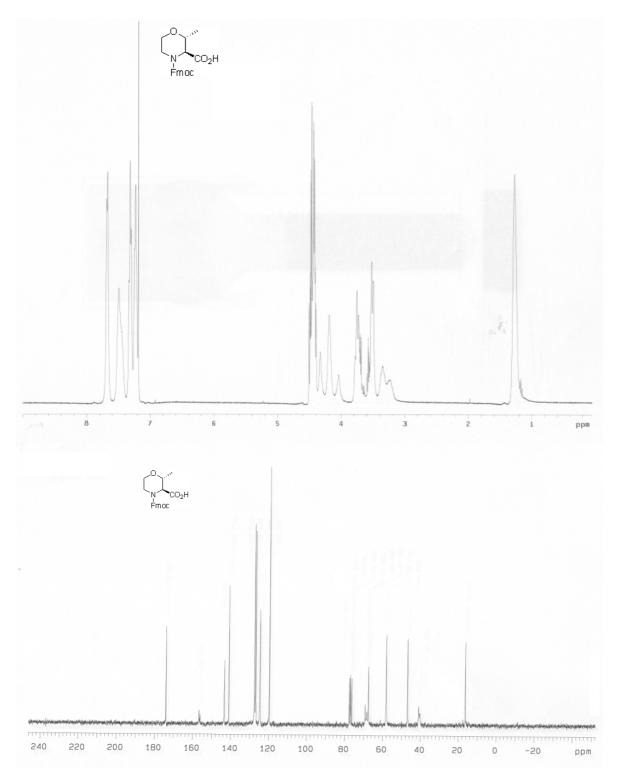


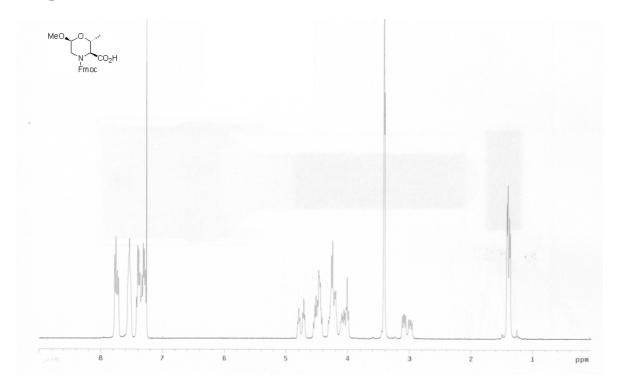


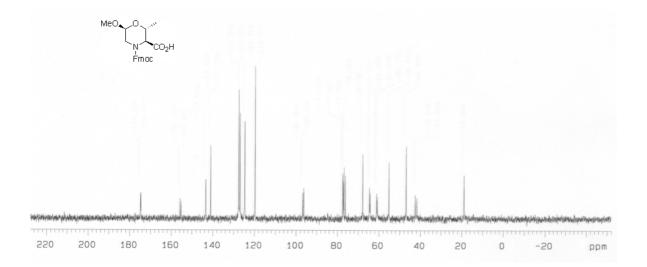


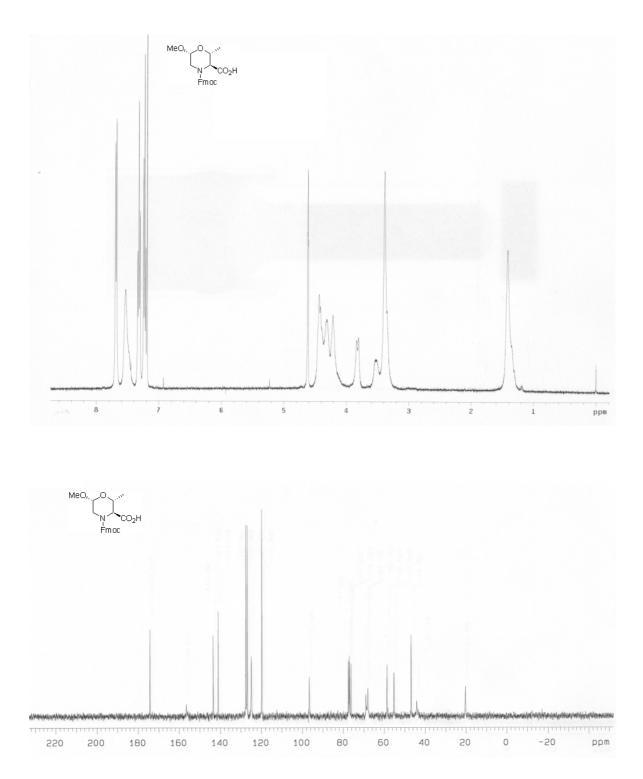




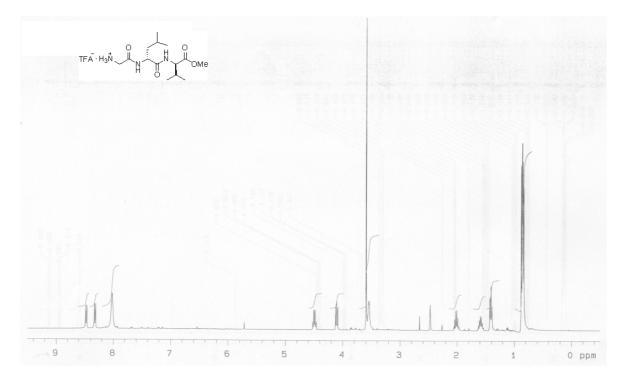




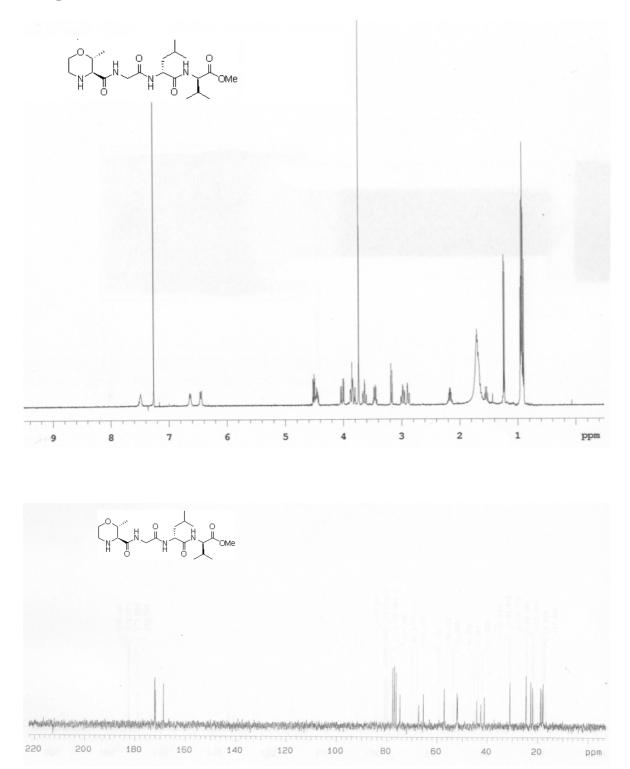




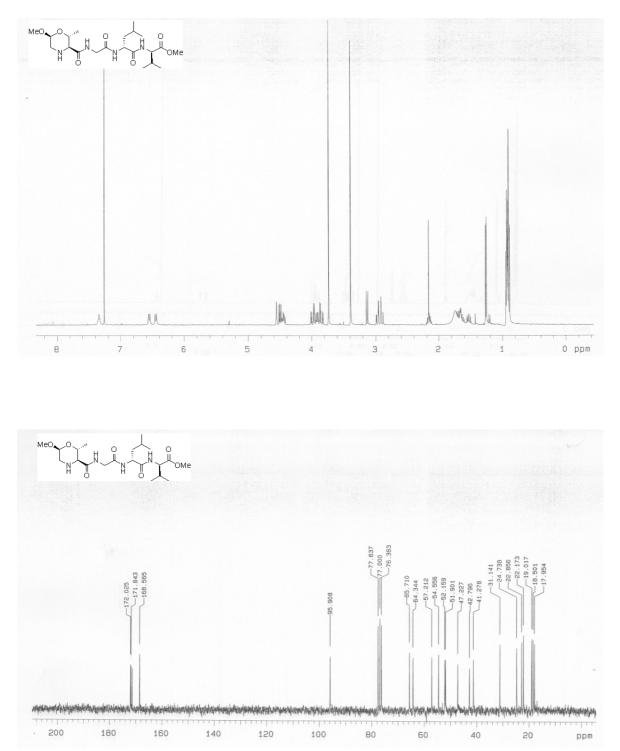
Peptide Gly-D-Leu-D-Val-OMe



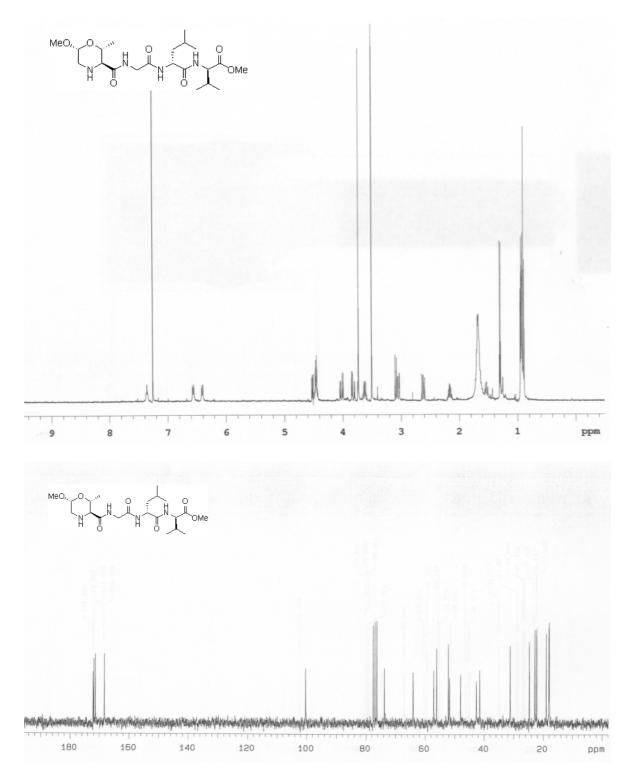
Compound 9a



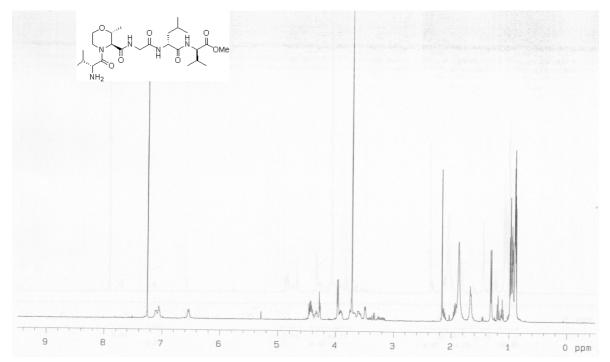
Compound 9b

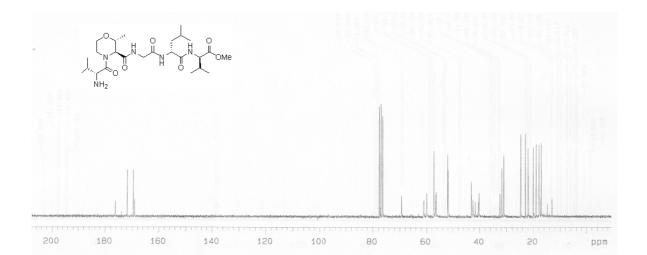


Compound 9c

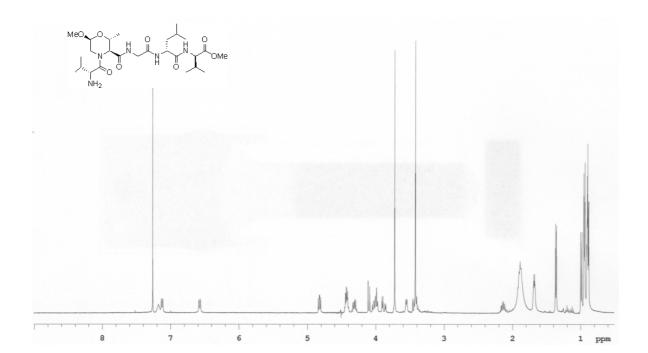


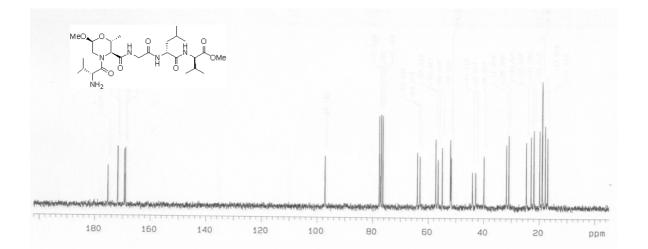
Compound 10a



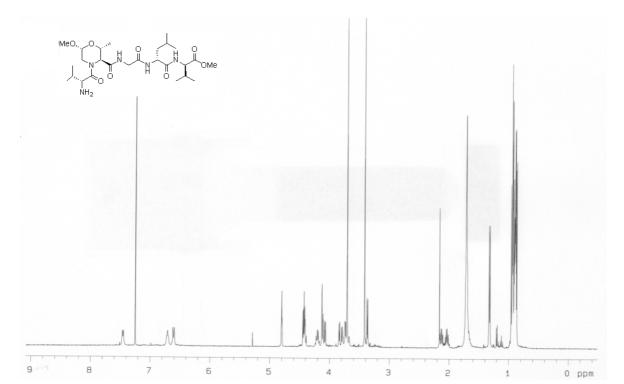


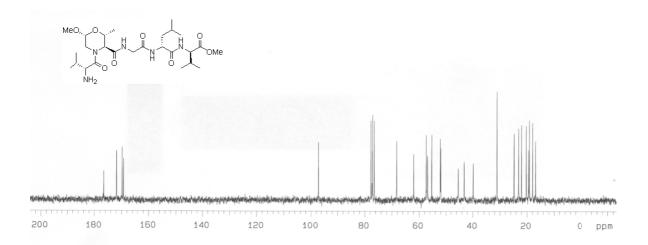
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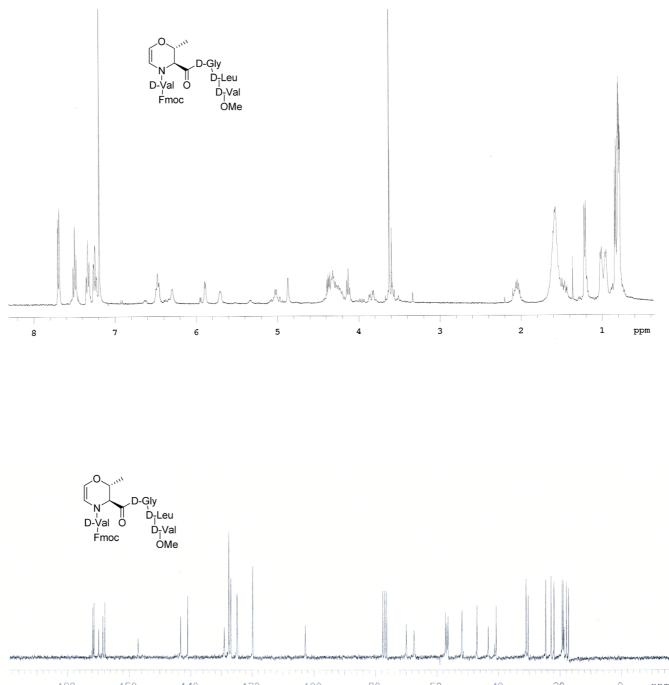




Compound 10c



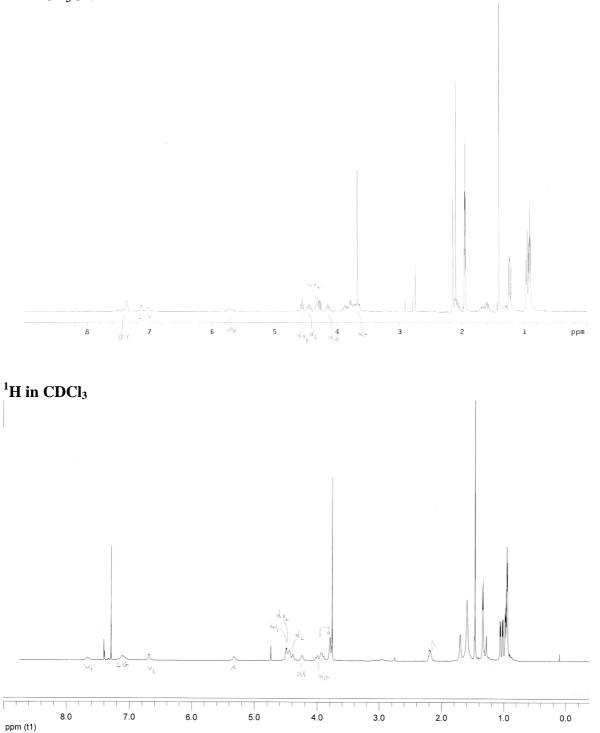




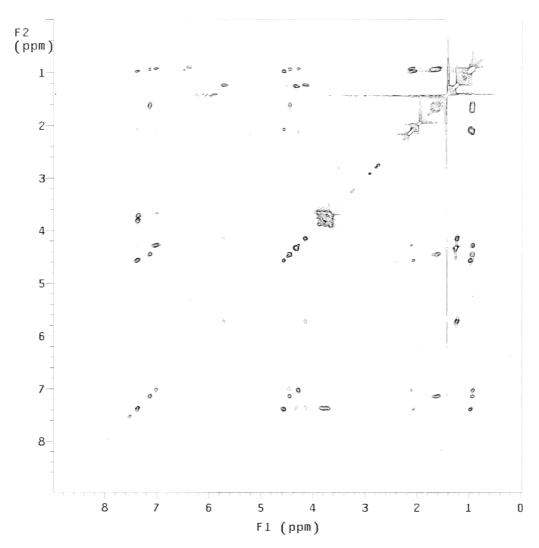
180 160 140 120 100 80 60 40 2**0** 0 ppm

Compound I

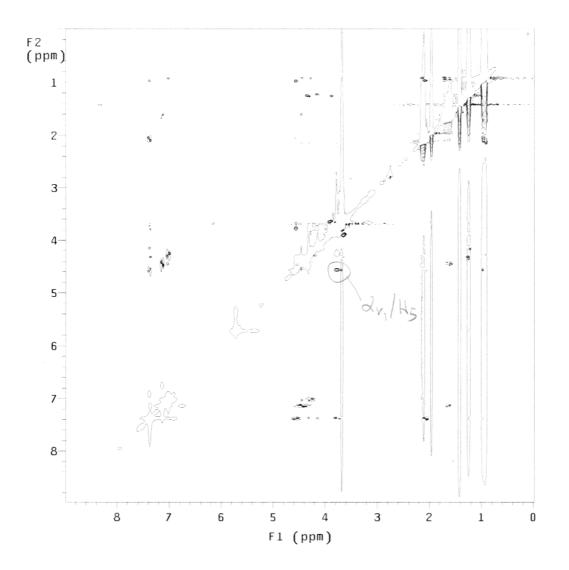




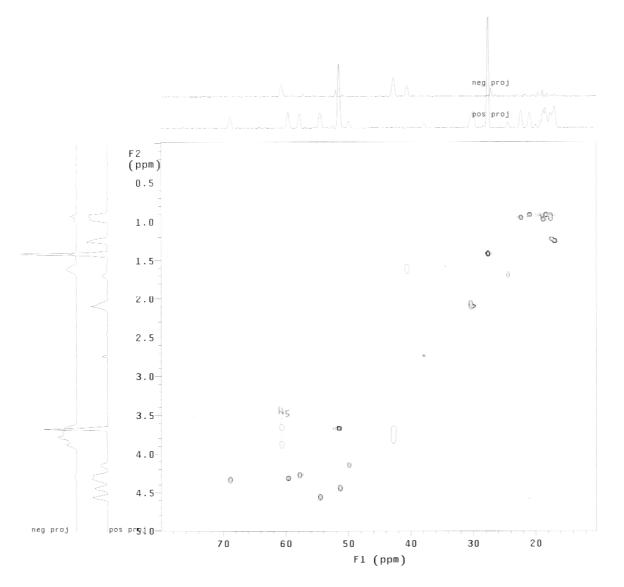
TOCSY



ROESY

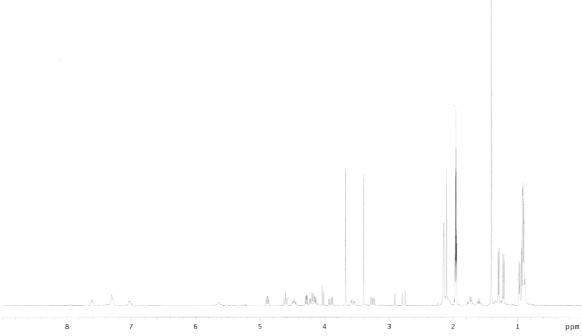


gHSQC

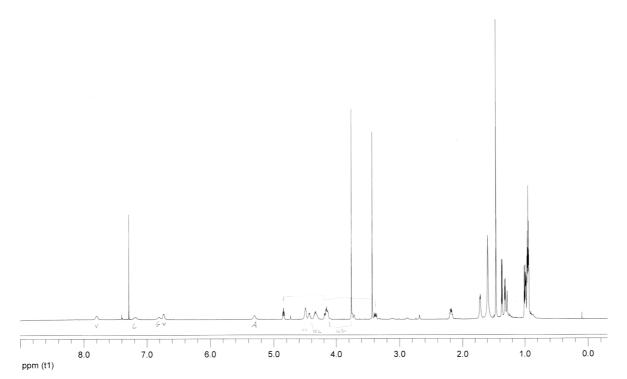


Compound II

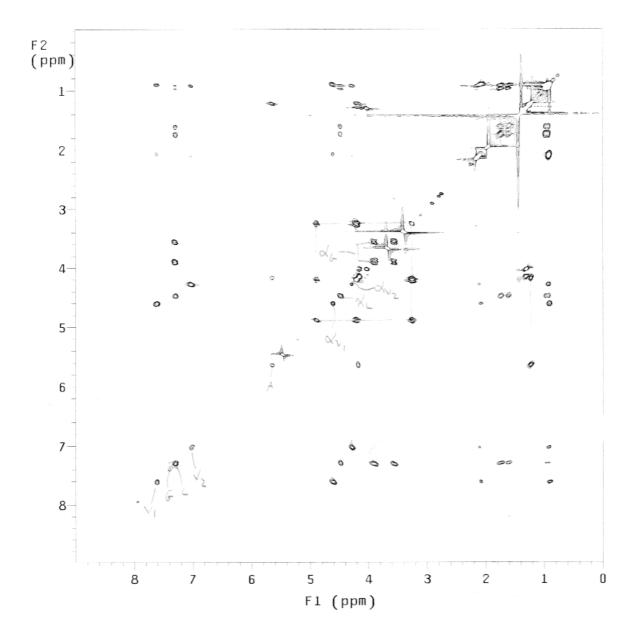




¹H in CDCl₃

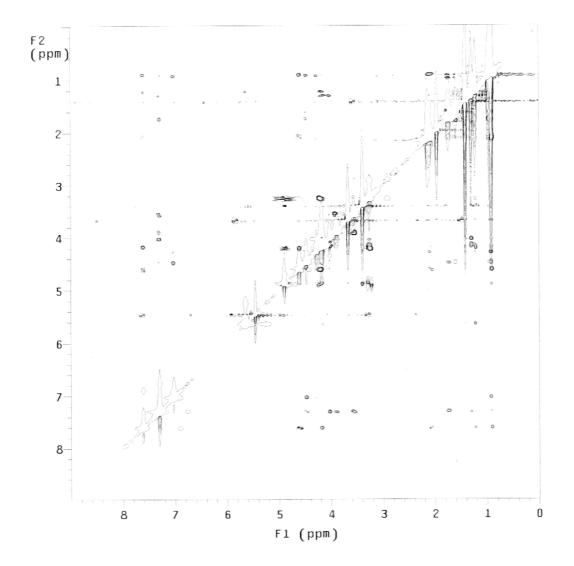


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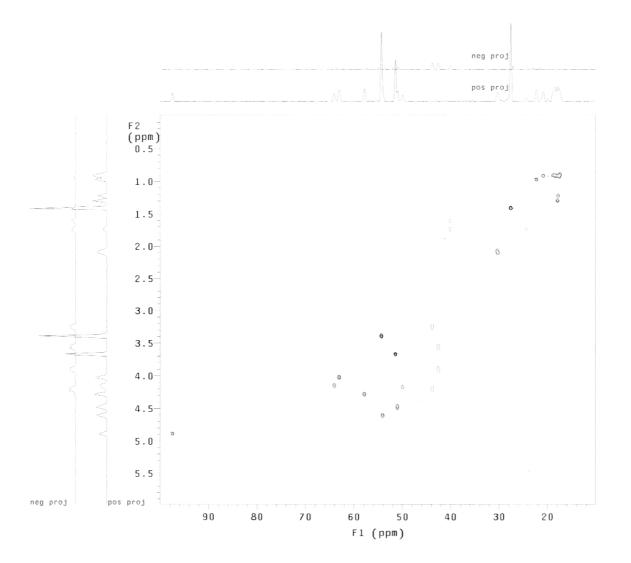


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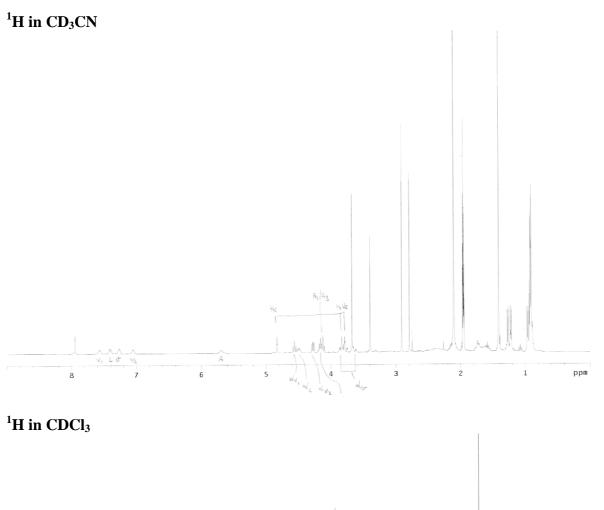
ROESY

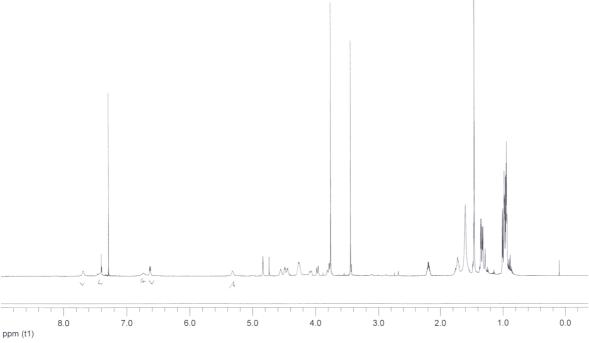


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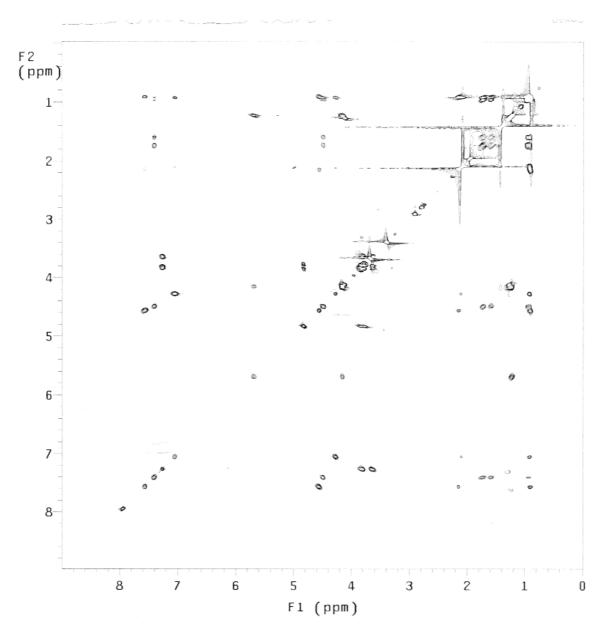


Compound III

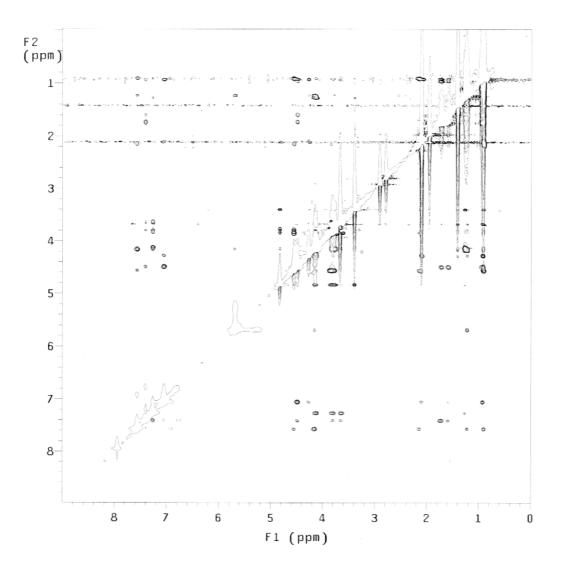




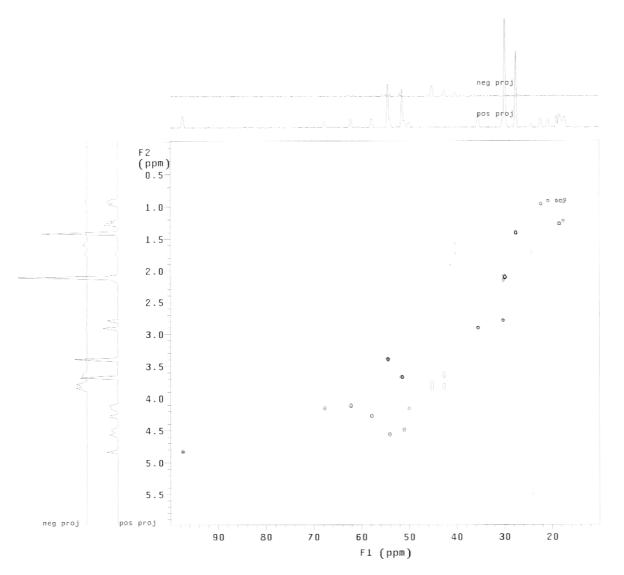
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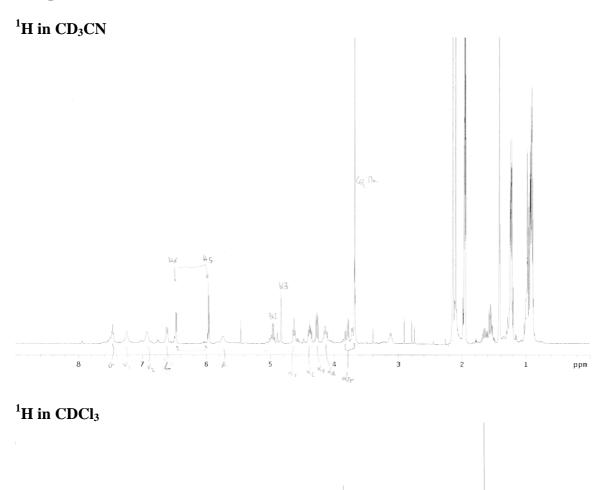
ROESY

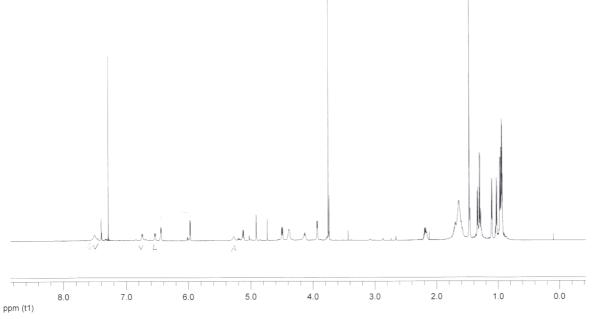


gHSQC

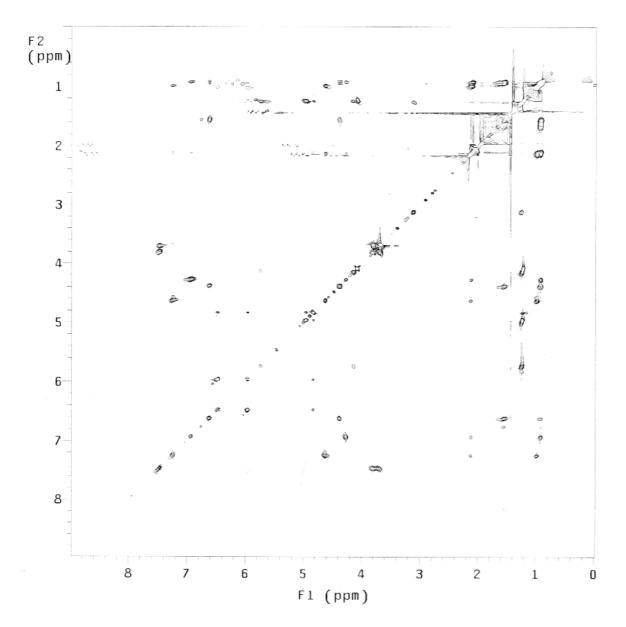


Compound IV

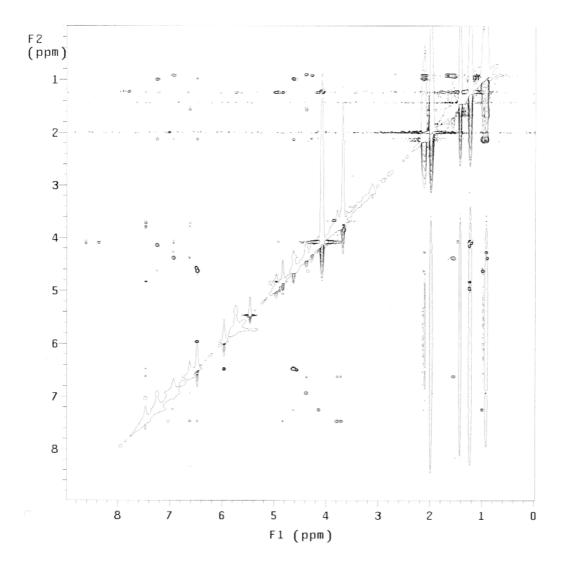




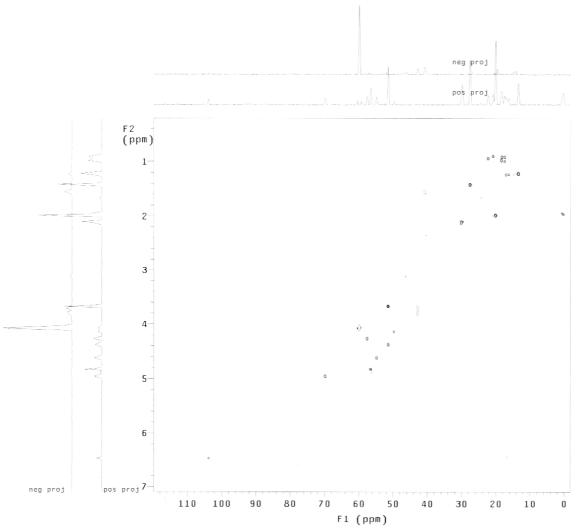
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ROESY



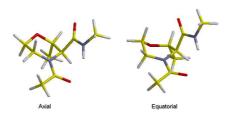
gHSQC



Computational data

Ab initio single point calculations of the electronic properties of the most abundant minimum energy conformer at the $6-31G^*/HF$ level of quantum chemical theory. The axial structure is referred to the conformation having the substituents at C-2 and C-3 in axial orientation (1 a.u. = 627.5 kcal/mol).

4-Acetyl-2-Me-Mor-3-methylamide (scaffold I)

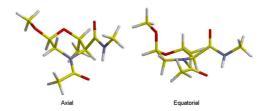


Axial

Run type: Single point energy Model: RHF/3-21G(*) Number of shells: 74 46 S shells 28 SP shells Number of basis functions: 158 Number of electrons: 108 Number of heavy atoms: 14 Number of hydrogens: 16 Use of molecular symmetry disabled Molecular charge: 0 Spin multiplicity: 1 Memory model: direct 9.8 Mb Point Group = C1 Order = 1 Nsymop = 1This system has 84 degrees of freedom E(HF) = -679.8130698 a.u.

Equatorial Run type: Single point energy Model: RHF/3-21G(*) Number of shells: 74 46 S shells 28 SP shells Number of basis functions: 158 Number of electrons: 108 Number of heavy atoms: 14 Number of hydrogens: 16 Use of molecular symmetry disabled Molecular charge: 0 Spin multiplicity: 1 Memory model: direct 9.8 Mb Point Group = C1 Order = 1 Nsymop = 1This system has 84 degrees of freedom E(HF) = -679.8147176 a.u.

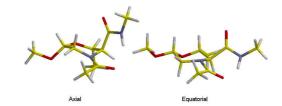
4-Acetyl-2-Me-(6S)-OMe-Mor-3-methylamide (scaffold II)



Axial Run type: Single point energy Model: RHF/3-21G(*) Number of shells: 84 52 S shells 32 SP shells Number of basis functions: 180 Number of electrons: 124 Number of heavy atoms: 16 Number of hydrogens: 18 Use of molecular symmetry disabled Molecular charge: 0 Spin multiplicity: 1 Memory model: direct 11.8 Mb Point Group = C1 Order = 1 Nsymop = 1This system has 96 degrees of freedom E(HF) = -793.0866731 a.u.

Equatorial Run type: Single point energy Model: RHF/3-21G(*) Number of shells: 84 52 S shells 32 SP shells Number of basis functions: 180 Number of electrons: 124 Number of heavy atoms: 16 Number of hydrogens: 18 Use of molecular symmetry disabled Molecular charge: 0 Spin multiplicity: 1 Memory model: direct 11.8 Mb Point Group = C1 Order = 1 Nsymop = 1This system has 96 degrees of freedom E(HF) = -793.0767991 a.u.

4-Acetyl-2-Me-(6R)-OMe-Mor-3-methylamide (scaffold III)

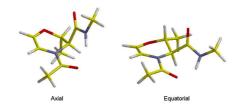


Axial

Run type: Single point energy Model: RHF/3-21G(*) Number of shells: 84 52 S shells 32 SP shells Number of basis functions: 180 Number of electrons: 124 Number of heavy atoms: 16 Number of hydrogens: 18 Use of molecular symmetry disabled Molecular charge: 0 Spin multiplicity: 1 Memory model: direct 11.8 Mb Point Group = C1 Order = 1 Nsymop = 1This system has 96 degrees of freedom E(HF) = -793.0667033 a.u.

Equatorial Run type: Single point energy Model: RHF/3-21G(*) Number of shells: 84 52 S shells 32 SP shells Number of basis functions: 180 Number of electrons: 124 Number of heavy atoms: 16 Number of hydrogens: 18 Use of molecular symmetry disabled Molecular charge: 0 Spin multiplicity: 1 Memory model: direct 11.8 Mb Point Group = C1 Order = 1 Nsymop = 1This system has 96 degrees of freedom E(HF) = -793.0771600 a.u.

4-Acetyl-2-Me-3,4-dihydro-2*H*-[1,4]oxazine-3-methylamide (scaffold IV)



Axial Run type: Single point energy Model: RHF/3-21G(*) Number of shells: 70 42 S shells 28 SP shells Number of basis functions: 154 Number of electrons: 106 Number of heavy atoms: 14 Number of hydrogens: 14 Use of molecular symmetry disabled Molecular charge: 0 Spin multiplicity: 1 Memory model: direct 9.5 Mb Point Group = C1 Order = 1 Nsymop = 1This system has 78 degrees of freedom E(HF) = -678.6528640 a.u.

Equatorial Run type: Single point energy Model: RHF/3-21G(*) Number of shells: 70 42 S shells 28 SP shells Number of basis functions: 154 Number of electrons: 106 Number of heavy atoms: 14 Number of hydrogens: 14 Use of molecular symmetry disabled Molecular charge: 0 Spin multiplicity: 1 Memory model: direct 9.5 Mb Point Group = C1 Order = 1 Nsymop = 1This system has 78 degrees of freedom E(HF) = -678.6417848 a.u.