

Total synthesis of Le^A-LacNAc pentasaccharide as a ligand for *Clostridium difficile* toxin A

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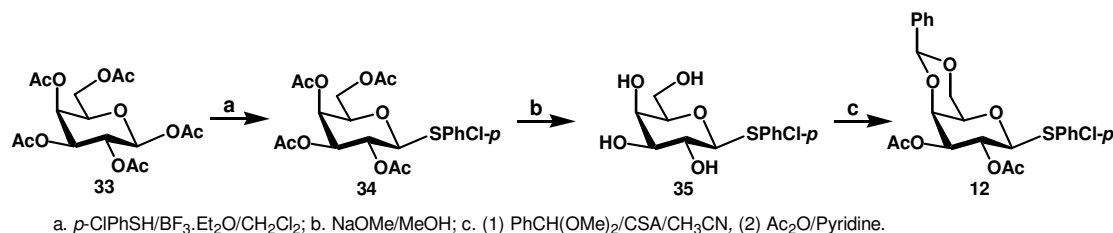
Table of Contents

I. Experimental procedures

Synthesis of compound 12.....	S2-S3
Synthesis of compound 15.....	S3-S4
Synthesis of compound 17.....	S5
Synthesis of compound 18.....	S5
Synthesis of compound 14.....	S6
Synthesis of compound 21.....	S6
Synthesis of compound 25.....	S7
Synthesis of compound 26.....	S7
Synthesis of compound 28.....	S7
Synthesis of compound 29 and 30.....	S8-S9

II. ¹ H and ¹³ C spectra of all synthesized compounds.....	S9-S71
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Synthesis of compounds 12



p-Chlorophenyl 2,3,4,6-tetra-*O*-acetyl-1-thio-β-D-galactopyranoside (34)

To a solution containing 1,2,3,4,6-penta-*O*-acetyl-β-D-galactopyranose **33** (20.0 g, 51.2 mmol) and *p*-chlorophenylthiol (11.1 g, 76.9 mmol, 1.5 equiv.) in anhydrous CH₂Cl₂ (250 mL), was added boron trifluoride etherate (9.7 mL, 76.9 mmol), and the mixture was stirred at room temperature overnight. The mixture was cooled to 0° C, Et₃N (10 mL) was added to quench the reaction. The unreacted thiol was acetylated by adding acetic anhydride (8.0 mL) to the mixture. After stirring at room temperature for 1 hour, the mixture was concentrated under reduced pressure. The residue was dissolved in EtOAc (300 mL), successively washed with H₂O (2 × 200 mL), 2N NaOH (1 × 100 mL), 2N HCl (1 × 100 mL), dried over anhydrous Na₂SO₄, and evaporated. Pure compound **34** was obtained by recrystallization from a mixture of AcOEt – hexane (21.0 g, 86.3% yield) as colorless crystals (Found: C, 50.63; H, 5.05%. C₂₀H₂₃O₉SCl requires C, 50.58; H, 4.88%); [α]_D -1.3° (*c* 0.9, CHCl₃); δ_H (400 MHz, CDCl₃) 7.47 (d, 2H, *J* 8.6 Hz, SPhCl), 7.29 (d, 2H, *J* 8.6 Hz, SPhCl), 5.41 (dd, 1H, *J* 3.2, 0.7 Hz, H-4), 5.20 (1H, dd, *J* 9.9, 9.9 Hz, H-2), 5.05 (dd, 1H, *J* 9.9, 3.3 Hz, H-3), 4.66 (d, 1H, *J* 9.9 Hz, H-1), 4.18 (dd, 1H, *J* 11.4, 7.0 Hz, H-6a), 4.10 (dd, 1H, *J* 11.4, 6.1 Hz, H-6b), 3.93 (ddd, 1H, *J* 7.0, 6.3, 0.7, H-5), 2.11 (s, 3H, Ac), 2.10 (s, 3H, Ac), 2.05 (s, 3H, Ac), 1.98 (s, 3H, Ac); δ_C (100 MHz, CDCl₃) 170.33 (CO), 170.10 (CO), 170.01 (CO), 169.38 (CO), 137.45 C-1_SPhCl), 134.41 (C-3_SPhCl + C-5_SPhCl), 130.33 (C-4_SPhCl), 128.99 (C-2_SPhCl + C-6_SPhCl), 86.06 (C-1), 74.51 (C-5), 71.93 (C-3), 67.15 (C-4 or C-2), 67.12 (C-2 or C-4), 61.58 (C-6), 20.82 (Ac), 20.66 (Ac), 20.59 (Ac), 20.56 (Ac); *m/z* (ESI-HRMS) calcd for [C₂₀H₂₃O₉SCl + Na]⁺ 497.0644, found 497.0645.

p-Chlorophenyl 1-thio-β-D-galactopyranoside (35)

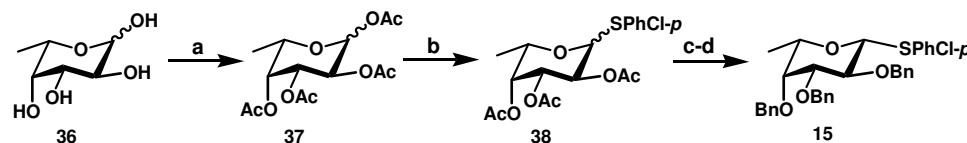
The per-acetate **34** (20.0 g, 42.1 mmol) was dissolved in anhydrous MeOH (170 mL), a solution of NaOMe in MeOH (1.5 M, 3 mL) was added, and the mixture was stirred at room temperature for 30 minutes. After neutralizing the reaction with Amberlite IR-120 (H⁺), the mixture was evaporated under reduced pressure to give pure tetraol **35** (12.9 g, 100% yield) as white solid (Found: C, 46.66; H, 5.28%. C₁₂H₁₅O₅SCl requires C, 46.98; H, 4.93%); [α]_D -46.4° (*c* 1.0, CHCl₃); δ_H (400 MHz, CDCl₃) 7.54 (d, 2H, *J* 8.6 Hz, SPhCl), 7.29 (m, 2H, *J* 8.6 Hz, SPhCl), 4.57 (d, 1H, *J* 9.6 Hz, H-1), 3.90 (dd, 1H, *J* 3.2, 0.6 Hz, H-4), 3.77 (dd, 1H, *J* 11.5, 6.9 Hz, H-6a), 3.71 (dd, 1H, *J* 11.5, 5.1 Hz, H-6b), 3.60 (dd, 1H, *J* 9.3, 9.3 Hz, H-2),

3.57 (ddd, 1H, J 0.9, 5.1, 6.9 Hz, H-5), 3.50 (dd, 1H, J 9.2, 3.3 Hz, H-3); δ_C (100 MHz, CD₃OD) 133.29 (C-1_SPhCl), 132.66 (C-4_SPhCl), 132.33 (C-3_SPhCl + C-5_SPhCl), 128.43 (C-2_SPhCl + C-6_SPhCl), 88.51 (C-1), 79.27 (C-5), 74.88 (C-3), 69.45 (C-2), 69.01 (C-4), 61.25 (C-6); m/z (ESI-HRMS) calcd for [C₁₂H₁₅O₅SCl + Na]⁺ 329.0221, found 329.0224.

***p*-Chlorophenyl 2,3-di-*O*-acetyl-4,6-*O*-benzylidene-1-thio- β -D-galactopyranoside (12)**

To a suspension of tetraol **35** (7.24 g, 23.6 mmol) and benzaldehyde dimethyl acetal (7.09 mL, 47.2 mmol) in anhydrous CH₃CN (100 mL) was added (\pm)-Camphor-10-sulfonic acid (500 mg), and the mixture was stirred for 30 minutes. Et₃N (2.0 mL) was added to quench the reaction. After removing the solvent under reduced pressure, the residue was acetylated with a mixture of acetic anhydride (25 mL) and anhydrous pyridine (30 mL) for 4 hours at room temperature. The reaction mixture was concentrated again and co-evaporated with toluene (3 \times 50 mL). After a column chromatography on silica gel using a gradient of AcOEt – toluene (5% \rightarrow 7%) as the eluent, compound **12** (10.2 g, 90% yield) was obtained in pure form (Found: C, 57.60; H, 5.35%. C₂₃H₂₃O₇SCl requires C, 57.68; H, 4.84%); [α]_D -21.8° (c 0.6, CHCl₃); δ_H (400 MHz, CDCl₃) 7.58 (d, 2H, J 8.4 Hz, SPhCl), 7.44 – 7.32 (m, 5H, Ph), 7.21 (d, 2H, J 8.5, SPhCl), 5.46 (s, 1H, PhCH), 5.29 (dd, 1H, J 9.8, 9.8 Hz, H-2), 5.01 (dd, 1H, J 9.9, 3.3 Hz, H-3), 4.65 (d, 1H, J 9.7 Hz, H-1), 4.38 – 4.29 (m, 2H, H-4 + H-6a), 3.98 (dd, 1H, J 12.1, ~1 Hz, H-6b), 3.56 (m, 1H, H-5), 2.10 (s, 3H, Ac), 2.03 (s, 3H, Ac); δ_C (100 MHz, CDCl₃) 170.53 (CO), 169.03 (CO), 137.44, 135.58, 134.61, 129.25, 128.94, 128.90, 128.23, 126.43, 100.98 (PhCH), 84.22 (C-1), 73.34 (C-4), 73.03 (C-3), 69.66 (C-5), 68.99 (C-6), 66.59 (C-2), 20.89 (Ac), 20.85 (Ac); m/z (ESI-HRMS) calcd for [C₂₃H₂₃O₇SCl + Na]⁺ 501.0745, found 501.0749.

Synthesis of compound 15



a. Ac₂O/pyridine; b. *p*-ClPhSH/BF₃·Et₂O/CH₂Cl₂; c. NaOMe/MeOH; d. BnBr/NaH/DMF;

***p*-Chlorophenyl 2,3,4-tri-*O*-acetyl-1-thio- α,β -L-fucopyranoside (38)**

A solution of L-fucose (**36**, 20.34 g, 0.124 mol) in a mixture of acetic anhydride (70 mL) and anhydrous pyridine (80 mL) was stirred at 50° C overnight. The mixture was evaporated under vacuum and coevaporated with toluene (3 \times 100 mL) to afford the crude 1,2,3,4-tetra-*O*-acetyl- α,β -L-fucopyranose (**37**, 40 g, quantitative). The crude α,β -mixture was dissolved in anhydrous CH₂Cl₂ (200 mL), and *p*-chlorophenylthiol (27.9 g, 192.6 mmol, 1.6 equiv.) was added. After the addition of boron trifluoride etherate (22.7 mL, 180.6 mmol, 1.5

equiv.), the mixture was stirred at room temperature overnight. The mixture was cooled to 0° C, Et₃N (20 mL) was added to quench the reaction. The unreacted thiol was acetylated by adding acetic anhydride (16.0 mL) to the mixture. After stirring at room temperature for 1 hour, the mixture was concentrated under reduced pressure. The residue was dissolved in EtOAc (600 mL), successively washed with H₂O (2 × 300 mL), 2N HCl (1 × 300 mL), and 5% NaHCO₃ (2 × 300 mL), dried over anhydrous Na₂SO₄, and evaporated. Compound **38** (41.15 g, 82% yield) was obtained as an α,β-mixture (1:9) by chromatography on silica gel using a mixture of 5% AcOEt – hexane as eluent (Found: C, 51.95; H, 5.09%. C₁₈H₂₁O₇SCl requires C, 51.86; H, 5.08%); δ_H (400 MHz, CDCl₃) for the β-anomer: 7.49 – 7.45 (m, 2H, SPhCl), 7.32 – 7.27 (m, 2H, SPhCl), 5.26 (dd, 1H, *J* 3.4, 0.8 Hz, H-4), 5.18 (dd, 1H, *J* 9.8, 9.8 Hz, H-2), 5.05 (dd, 1H, *J* 9.9, 3.3 Hz, H-3), 4.65 (d, 1H, *J* 9.8 Hz, H-1), 3.83 (dq, 1H, *J* 6.5, 1.0 Hz, H-5), 2.13 (s, 3H, Ac), 2.09 (s, 3H, Ac), 1.97 (s, 3H, Ac), 1.23 (d, 1H, *J* 6.4 Hz, H-6); δ_C (100 MHz, CDCl₃) 170.43 (CO), 170.01 (CO), 169.35 (CO), 134.40 (Ar), 134.16 (Ar), 130.73 (Ar), 128.91 (Ar), 85.88 (C-1), 73.22 (C-5), 72.35 (C-3), 70.24 (C-4), 67.21 (C-2), 20.79 (Ac), 20.57 (Ac), 20.55 (Ac), 16.40 (C-6); *m/z* (ESI-HRMS) calcd for [C₁₈H₂₁O₇SCl + Na]⁺ 439.0589, found 439.0590.

***p*-Chlorophenyl 2,3,4-tri-*O*-benzyl-1-thio-β-L-fucopyranoside (15)**

The triacetate **38** (1:9 α:β mixture, 21.5 g, 51.6 mmol) was dissolved in anhydrous MeOH (250 mL), and a solution of NaOMe in MeOH (1.5 M, 3.0 mL) was added. After stirring at room temperature for 2 hours, the mixture was neutralized with Amberlite IR-120 (H⁺). The resin was removed by filtration and the organic solution was concentrated under reduced pressure to give a syrup (~15 g). The crude triol was dissolved in anhydrous DMF (130 mL), NaH (60% in mineral oil, 10.0g, 250 mmol) was added portion wise, and the reaction mixture was stirred at room temperature for 1 hour. After cooling to 0° C, benzyl bromide (35 mL, 295.2 mmol) was added dropwise, and the mixture was allowed to warm-up to temperature. After stirring overnight, MeOH (15 mL) was added, and the mixture was concentrated under reduced pressure to remove most of the solvent. The mixture was dissolved in EtOAc (400 mL), washed with H₂O (1 × 200 mL), 2N HCl (1 × 200 mL), 10% NaHCO₃ (1 × 200 mL), dried over anhydrous Na₂SO₄, and evaporated. After a chromatography on silica gel using 3.5% AcOEt – hexane as eluent, the pure β-anomer **15** was obtained (23.6 g, 85.5 yield%) as a white solid (Found: C, 70.70; H, 5.83%. C₃₃H₃₃O₄SCl requires C, 70.63; H, 5.93%); [α]_D +3.8° (*c* 0.7, CHCl₃); δ_H (400 MHz, CDCl₃) 7.54 (d, 2H, *J* 8.6 Hz, SPhCl), 7.47 – 7.28 (m, 15H, 3 × Bn), 7.15 (d, 2H, *J* 8.6 Hz, SPhCl), 5.03 (d, 1H, *J* 11.4 Hz, Bn), 4.77 (s, 4H, Bn), 4.68 (d, 1H, *J* 11.4 Hz, Bn), 4.58 (d, 1H, *J* 9.6 Hz, H-1), 3.92 (dd, 1H, *J* 9.4, 9.4 Hz, H-2), 3.67 (dd, 1H, *J* 2.3, ~1 Hz, H-4), 3.62 (dd, 1H, *J* 2.7, 9.2 Hz, H-3), 3.56 (dq, 1H, *J* 6.3, ~1 Hz, H-5), 1.30 (d, 3H, *J* 6.4 Hz, H-6); δ_C (100 MHz, CDCl₃) 138.62, 138.29, 133.12, 132.94, 132.57, 128.85, 128.44, 128.33, 128.25, 128.22, 127.98, 127.74, 127.71, 127.59, 127.55 (Ar), 87.12 (C-1), 84.50 (C-3), 76.87 (C-2), 76.55 (C-4), 75.55 (Bn), 74.68 (C-5), 74.67 (Bn), 72.82 (Bn), 17.29 (C-6); *m/z* (ESI-HRMS) calcd for [C₃₃H₃₃O₄SCl + Na]⁺ 583.1680, found 583.1678.

***p*-Chlorophenyl 3,4,6-tri-*O*-acetyl-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside (17)**

To a solution of 1,3,4,6-tetra-*O*-acetyl-2-deoxy-2-phthalimido- α,β -glucopyranose **16** (40.0 g, 89.4 mmol) and *p*-chlorophenylthiol (20.6 g, 142.4 mmol) in anhydrous CH₂Cl₂ (200 mL), was added BF₃·Et₂O (16.8 mL, 134 mmol) dropwise. After stirring for 3 days at rt, the reaction was cooled to 0°, and Et₃N (35 mL) was added to quench the reaction. The unreacted thiol was then acetylated by the addition of excess Ac₂O (15 mL). After 1h at rt, the mixture was concentrated under reduced pressure; the residue was dissolved in EtOAc (600 mL), and the organic solution was washed successively with H₂O (2 × 300 mL), a solution of 2 N HCl (1 × 300 mL) and a 5% solution of aqueous NaHCO₃ (1 × 150 mL), dried over anhydrous Na₂SO₄, and concentrated to give a syrup which was recrystallized from a mixture of AcOEt –hexane. The desired thioglycoside **17** (38 g, yield 76%) was obtained in pure form as a white solid (Found: C, 55.47; H, 4.69; N, 2.41%. C₂₆H₂₄NO₉ClS requires C, 55.57; H, 4.30; N, 2.49%); [α]_D +39.7° (*c* 1.2, CHCl₃); δ _H (400 MHz, CDCl₃) 7.88 (m, 2H, Phth), 7.77 (m, 2H, Phth), 7.37 (m, 2H, SPhCl), 7.26 (m, 2H, SPhCl), 5.79 (dd, 1H, *J* 9.7 Hz, H-3), 5.67 (d, 1H, *J* 10.5 Hz, H-1), 5.12 (dd, 1H, *J* 9.7 Hz, H-4), 4.32 (dd, 1H, *J* 10.4, 10.4 Hz, H-2), 4.29 (dd, 1H, *J* 4.9, 12.4 Hz, H-6a), 4.21 (dd, 1H, *J* 2.3, 12.3 Hz, H-6b), 3.90 (ddd, 1H, *J* 2.3, 4.9, 10.2 Hz, H-5), 2.11 (s, 1H, Ac), 2.03 (s, 3H, Ac), 1.84 (s, 3H, Ac); δ _C (100 MHz, CDCl₃) 170.53 (CO), 170.05 (CO), 169.38 (CO), 167.82 (CO), 166.91, (CO), 135.01, 134.99, 129.04, 123.74, 82.70 (C-1), 75.98 (C-5), 71.52 (C-3), 68.59 (C-4), 62.11 (C-6), 53.55 (C-2), 20.75 (Ac), 20.59 (Ac), 20.37 (Ac); *m/z* (ESI-HRMS) calcd for [C₂₆H₂₄NO₉ClS + Na]⁺ 584.07525, found 584.07542.

***p*-Chlorophenyl 2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside (18)**

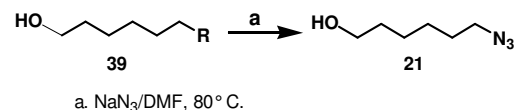
The peracetylated thioglycoside **17** (20.0 g, 35.6 mmol) was added to a solution of ~0.1 M guanidine/guanidinium chloride (400 mL); the starting material was observed to gradually dissolve to give a clear solution followed by the precipitation of desired deacetylated thioglycoside from the reaction mixture. The solid was filtered off under vacuum and washed with MeOH to afford pure **18** (14.53 g); after concentration of the mother liquor, more **18** (0.62 g) was crystallized out from the mixture. The total amount of **18** is 15.15 g (yield: 97.7%) (Found: C, 53.27; H, 4.50; N, 3.04%. C₂₀H₂₀NO₇ClS·H₂O requires C, 52.92; H, 4.44; N, 3.09%); [α]_D +59.5° (*c* 0.34, CHCl₃); δ _H (400 MHz, CDCl₃) 7.93 – 7.81 (m, 4H, Phth), 7.40 (d, 2H, *J* 8.6 Hz, SPhCl), 7.25 (d, 2H, *J* 8.6 Hz, SPhCl), 5.57 (d, 1H, *J* 10.4 Hz, H-1), 4.24 (dd, 1H, *J* 10.2, 8.5 Hz, H-3), 4.08 (dd, 1H, *J* 10.3, 10.3 Hz, H-2), 3.95 (dd, 1H, *J* 12.1, 2.1 Hz, H-6a), 3.75 (dd, 1H, *J* 12.1, 5.7 Hz, H-6b), 3.49 (ddd, 1H, *J* 9.9, 5.5, 2.0 Hz, H-5), 3.43 (dd, 1H, *J* 9.8, 8.6 Hz, H-4); δ _C (100 MHz, CDCl₃) 135.84, 135.79, 135.06, 134.65, 133.12, 130.16, 124.64, 124.34 (Ar), 85.14 (C-1), 82.92 (C-5), 73.96 (C-3), 72.36 (C-4), 62.97 (C-6), 57.84 (C-2); *m/z* (ESI-HRMS) calcd for [C₂₀H₁₈NO₆ClS + Na]⁺ 458.0436, found 458.0435.

***p*-Chlorophenyl 4,6-*O*-benzylidene-2-deoxy-2-phthalimido-1-thio- β -D-glucopyranoside**

(14)

To a suspension containing the triol **18** (3.56 g, 8.2 mmol) and benzaldehyde dimethyl acetal (2.38 mL, 15.8 mmol) in anhydrous CH₃CN (20 mL), was added (±)-camphor-10-sulfonic acid (350 mg); the mixture was stirred at rt for 2 hrs. Et₃N (1.0 mL) was added to quench the reaction, and the mixture was concentrated under reduced pressure. The residue was dissolved in EtOAc (100 mL) and the organic solution was washed with H₂O (2 × 50 mL), dried over anhydrous Na₂SO₄ and evaporated under vacuum. The desired compound **14** (4.0 g, yield: 93%) was obtained by column chromatography on silica gel using 5% EtOAc – toluene as eluent. $[\alpha]_D^{+29.2^\circ}$ (*c* 0.65, CHCl₃); δ_H (400 MHz, CDCl₃) 7.96 – 7.83 (m, 2H, Phth), 7.81 – 7.72 (m, 2H, Phth), 7.53 – 7.45 (m, 2H, Ph), 7.43 – 7.32 (m, 5H, 3 × Ph + 2 × SPhCl), 7.30 – 7.23 (m, 2H, SPhCl), 5.65 (d, 1H, *J* 10.5 Hz, H-1), 5.57 (s, 1H, PhCH), 4.63 (dd, 1H, *J* 9.5, 9.5 Hz, H-3), 4.40 (dd, 1H, *J* 10.3, 4.8 Hz, H-6a), 4.30 (dd, 1H, *J* 10.3, 10.3 Hz, H-2), 3.81 (dd, 1H, *J* 10.2, 10.2 Hz, H-6b), 3.70 (ddd, 1H, *J* 9.6, 9.6, 4.8 Hz, H-5), 3.58 (dd, 1H, *J* 9.2, 9.2 Hz, H-4), 2.57 (s, 1H, OH-3); δ_C (100 MHz, CDCl₃) 168.20 (CO), 167.45 (CO), 136.81, 134.64, 134.42, 134.31, 131.50, 129.79, 129.40, 129.10, 129.01, 128.38, 128.20, 126.28, 101.97 (PhCH), 83.92 (C-1), 81.79 (C-4), 70.33 (C-3), 69.69 (C-5), 68.49 (C-6), 55.47 (C-2); *m/z* (ESI-HRMS) calcd for [C₂₇H₂₂NO₆ClS + Na]⁺ 546.0749, found 546.0751.

Synthesis of compound 21



6-Azido-1-hexanol (21)

Compound was prepared from 6-chloro-1-hexanol (**39**) according to identical procedure published before (93% yield). The NMR data is identical to the literature.¹ δ_H (400 MHz, CDCl₃) 3.63 (t, 2H, *J* 6.5 Hz, H-1), 3.26 (t, 2H, *J* 6.9 Hz, H-6), 1.67 – 1.52 (m, 4H, H-2 + H-5), 1.45 – 1.34 (m, 4H, H-3 + H-4). δ_C (100 MHz, CDCl₃) 62.70, 51.37, 32.52, 28.79, 26.51, 25.32.

Attempt to synthesize *p*-chlorophenyl 2,3,4,6-tetra-*O*-acetyl-β-D-galactopyranosyl-(1→3)-6-*O*-benzyl-2-deoxy-4-*O*-*p*-methoxybenzyl-2-phthalimido-1-thio-β-D-glucopyranoside (6)

A mixture of imidate **10** (63.8 mg, 0.129 mmol, 3.0 eq), alcohol **13** (27.9 mg, 0.043 mmol) and 4 Å molecular sieves (100 mg) in anhydrous CH₂Cl₂ (0.8 mL) was stirred under Ar for 1

h. The mixture was cooled to 0° C, and TMSOTf (3.0 µL) was added. After 1 h, the reaction was neutralized with Et₃N (5 drops). The mixture was filtered off and concentrated. The residue was purified by chromatography on silica gel using a gradient of AcOEt – hexane (7% → 15%) to obtain the recovered acceptor **13** (19 mg) and rearranged imidate **25** (12 mg). No desired disaccharide **6** was formed. Data for **2,3,4,6-Tetra-O-acetyl-1-N-trichloroacetyl-β-D-galactopyranosylamine (25)**: [α]_D +23.7° (*c* 0.5, CHCl₃); δ_H (400 MHz, CDCl₃) 7.47 (d, 1H, *J* 8.6 Hz, NH), 5.48 (dd, 1H, *J* 3.1, 0.9 Hz, H-4), 5.24 (dd, 1H, *J* 10.1, 9.0 Hz, H-2), 5.18 (dd, 1H, *J* 9.9, 3.2 Hz, H-3), 5.14 (dd, 1H, *J* 8.9, 8.9 Hz, H-1), 4.18 – 4.11 (m, 2H, H-6a + H-6b), 4.09 (ddd, 1H, *J* 7.0, 5.7, 0.9 Hz, H-5), 2.18 (s, 3H, Ac), 2.08 (s, 3H, Ac), 2.06 (s, 3H, Ac), 2.02 (s, 3H, Ac); δ_C (100 MHz, CDCl₃) 171.38 (CO), 170.36 (CO), 169.93 (CO), 169.72 (CO), 162.03 (CO), 80.42 (C-1), 72.86 (C-5), 70.43 (C-3), 68.01 (C-2), 66.97 (C-4), 60.95 (C-6), 20.65 (Ac), 20.64 (Ac), 20.59 (Ac), 20.52 (Ac); *m/z* (ESI-MS) calcd for [C₁₆H₂₀NO₁₀Cl₃ + Na]⁺ 514.0, 516.0, 518.0, found 514.2, 516.1, 518.0.

Attempt to synthesize *p*-chlorophenyl 2,3-di-O-acetyl-4,6-O-benzylidene-β-D-galactopyranosyl-(1→3)-6-O-benzyl-2-deoxy-4-O-*p*-methoxybenzyl-2-phthalimido-1-thio-β-D-glucopyranoside (24)

A mixture of imidate **11** (31 mg, 0.062 mmol, 2.0 eq), alcohol **13** (20 mg, 0.031 mmol) and 4 Å molecular sieves (100 mg) in anhydrous CH₂Cl₂ (0.8 mL) was stirred under argon for 1 h. The mixture was cooled to 0° C, and TMSOTf (3.0 µL) was added. After 1 h, the reaction was neutralized with Et₃N (5 drops). The mixture was filtered off and concentrated. The residue was purified by chromatography on silica gel using a gradient of AcOEt – hexane (7% → 15%) to obtain the recovered acceptor **13** (14 mg) and rearranged imidate **26** (21 mg). No desired disaccharide **24** was formed. Data for **2,3-Di-O-acetyl-4,6-O-benzylidene-β-D-galactopyranosylamine trichloroacetate (26)** (Found: C, 46.21; H, 4.43; N, 2.75%. C₁₉H₂₀NO₈Cl₃ requires C, 45.94; H, 4.06; N, 2.82%); [α]_D +50.2° (*c* 0.6, CHCl₃); δ_H (400 MHz, CDCl₃) 7.61 (d, 1H, *J* 8.9 Hz, NH), 7.55 – 7.49 (m, 2H, Ph), 7.43 – 7.34 (m, 3H, Ph), 5.55 (s, 1H, PhCH), 5.44 (dd, 1H, *J* 10.2, 9.4 Hz, H-2), 5.18 (dd, 1H, *J* 9.1, 9.1 Hz, H-1), 5.13 (dd, 1H, *J* 10.3, 3.6 Hz, H-3), 4.50 (dd, 1H, *J* 3.6, 0.8 Hz, H-4), 4.36 (dd, 1H, *J* 12.6, 1.5 Hz, H-6a), 4.09 (dd, 1H, *J* 12.7, 1.8 Hz, H-6b), 3.71 (ddd, 1H, *J* 1.7, 1.7, 1.1 Hz, H-5), 2.13 (s, 3H, Ac), 2.07 (s, 3H, Ac); δ_C (100 MHz, CDCl₃) 171.19 (CO), 170.38 (CO), 162.21 (CO), 137.15, 129.15, 128.26, 126.07 (Ar), 100.75 (PhCH), 80.32 (C-1), 73.21 (C-4), 71.57 (C-3), 68.83 (C-6), 68.29 (C-5), 67.89 (C-2), 20.84 (Ac), 20.66 (Ac); *m/z* (ESI-MS) calcd for [C₁₉H₂₀NO₈Cl₃ + Na]⁺ 518.0, 520.0, 522.0, found 518.2, 520.1, 522.0.

2,3,4-tri-O-acetyl-1-thio-α,β-L-fucopyranosyl trichloroacetimidate (28)

Thioglycoside **15** (5.16 g, 9.20 mmol) was dissolved in a mixture of CH₃CN (30 mL) – H₂O (2.8 mL); *N*-iodosuccinimide (4.36 g, 18.39 mmol) was added and the mixture was stirred for 30 minutes at rt. Et₃N (3 mL) was added to quench the reaction. The mixture was diluted with EtOAc (200 mL), and washed with 10% aqueous Na₂S₂O₃ solution (2 × 100 mL), dried

over anhydrous Na₂SO₄ and evaporated. The residue was purified by chromatography on silica gel using 20% AcOEt – toluene as eluent to afford the hemiacetal **27** (3.29 g, 82.3% yield). Part of the hemiacetal (3.0 g, 6.90 mmol) and CCl₃CN (6.3 mL, 62.8 mmol) were dissolved in anhydrous CH₂Cl₂ (20 mL), anhydrous K₂CO₃ (3.3 g, 23.9 mmol) was added; the mixture was stirred at rt overnight. The reaction was diluted with EtOAc (150 mL), washed with H₂O (2 × 50 mL), and evaporated to dryness. NMR showed that the mixture contained **28**² as a mixture (α/β 1:2) which was pure enough for use in next step, no further purification was performed. δ_H (300 MHz, CDCl₃) for α-anomer: 8.49 (s, 1H, C=NH), 7.44 – 7.11 (m, 15H, Ar), 6.51 (d, 1H, *J* 3.4 Hz, H-1), 5.06 – 4.61 (m, 6H, Bn), 1.14 (d, 3H, *J* 6.0 Hz, H-6). For β-anomer 8.59 (s, 1H, C=NH), 7.44 – 7.11 (m, 15H), 5.71 (d, 1H, *J* 8.1 Hz, H-1), 5.06 – 4.61 (m, 6H, Ar), 1.21 (d, 3H, *J* 6.4 Hz, H-6).

***p*-Chlorophenyl 2,3,4-tri-*O*-benzyl-α-L-fucopyranosyl-(1→3)-6-*O*-benzyl-2-deoxy-2-phthalimido-1-thio-β-D-glucopyranoside (29) and *p*-Chlorophenyl 2,3,4-tri-*O*-benzyl-α-L-fucopyranosyl-(1→4)-6-*O*-benzyl-2-deoxy-2-phthalimido-1-thio-β-D-glucopyranoside (30)**

A mixture of imidate **28** (1.57 g, 2.7 mmol), diol **19** (1.10 g, 2.09 mmol) and 4 Å molecular sieves (1.5 g) in anhydrous CH₂Cl₂ (10 mL) were stirred under Ar for 1 h. The mixture was cooled to -78° C, and TMSOTf (15 µL) was added dropwise. After 30 mins, the reaction was neutralized with Et₃N (0.5 mL). The mixture was diluted with EtOAc (100 mL) and the insoluble molecular sieves were removed by filtration. After concentration, the residue was purified by chromatography on silica gel using a gradient of AcOEt – hexane (15% → 20%) to afford **29** (1.46 g, 52% yield) and **30** (0.760 g, 27% yield).

Data for **29** (Found: C, 68.51; H, 5.56; N, 1.54%. C₅₄H₅₂NO₁₀SCl requires C, 68.81; H, 5.56; N, 1.49%): [α]_D +32.1° (*c* 0.5, CHCl₃); δ_H (400 MHz, CDCl₃) 7.88 (dd, 1H, *J* 7.4, 1.0 Hz, Phth), 7.73 – 7.66 (m, 2H, 2 × Phth), 7.58 (ddd, 1H, *J* 7.4, 7.4, ~1 Hz, Phth), 7.47 – 7.15 (m, 22H, Ar), 6.99 – 6.93 (m, 2H, Ar), 5.68 (d, 1H, *J* 10.3 Hz, H-1_GlcN), 4.86 (d, 1H, *J* 11.4 Hz, Bn), 4.65 (d, 1H, *J* 12.0, Bn), 4.61 (d, 1H, *J* 12.1 Hz, Bn, overlapped), 4.61 (d, 1H, *J* 3.5 Hz, H-1_Fuc), 4.61 (d, 1H, *J* 12.1 Hz, Bn, overlapped) 4.54 (d, 1H, *J* 11.4 Hz, Bn), 4.53 (d, 1H, *J* 10.9 Hz, Bn), 4.30 (dd, 1H, *J* 10.3, 10.3 Hz, H-2_GlcN), 4.26 (d, 1H, *J* 1.2, OH-4_GlcN), 4.21 (dd, 1H, *J* 10.3, 8.2 Hz, H-3_GlcN), 4.10 (d, 1H, *J* 13.4 Hz, Bn), 4.07 (dq, 1H, *J* 6.6, ~1 Hz, H-5_Fuc), 3.93 (dd, 1H, *J* 10.6, 1.4 Hz, H-6a_GlcN), 3.82 – 3.68 (m, 4H, H-3_Fuc + H-5_GlcN + H-2_Fuc + H-6b_GlcN), 3.57 – 3.49 (m, 2H, H-4_GlcN + H-4_Fuc), 3.39 (d, 1H, *J* 13.0 Hz, Bn), 1.07 (d, 3H, *J* 6.5 Hz, H-6_Fuc); δ_C (100 MHz, CDCl₃) 168.61 (CO), 167.64 (CO), 138.70, 138.43, 138.26, 138.03, 134.33, 134.24, 133.77, 132.38, 131.95, 130.27, 128.95, 128.42, 128.34, 128.25, 128.15, 127.61, 127.59, 127.54, 123.19, 123.00, 100.87 (C-1_Fuc), 84.11 (C-3_GlcN), 82.82 (C-1_GlcN), 79.35 (C-5_GlcN or C-3_Fuc), 78.90 (C-3_Fuc or C-5_GlcN), 77.86 (C-4_Fuc), 74.74 (Bn), 73.85 (C-2_Fuc), 73.42 (CH₂Ph), 73.41 (CH₂Ph), 72.41 (CH₂Ph), 70.99 (C-4_GlcN), 69.37 (C-6_GlcN), 68.52 (C-5_Fuc), 53.52 (C-2_GlcN), 16.43 (C-6_Fuc); *m/z* (ESI-HRMS) calcd for [C₅₄H₅₂NO₁₀SCl + Na]⁺ 964.2893, found 964.2891.

Data for **30** (Found: C, 68.61; H, 5.59; N, 1.52%. $C_{54}H_{52}NO_{10}SCl$ requires C, 68.81; H, 5.56; N, 1.49%): $[\alpha]_D -14.1^\circ$ (c 0.5, $CHCl_3$); δ_H (400 MHz, $CDCl_3$) 7.92 (m, 1H, Phth), 7.83 (m, 1H, Phth), 7.80 – 7.69 (m, 2H, Phth), 7.45 – 7.20 (m, 22H, $5 \times Ph + 2 \times SPhCl$), 7.13 (d, 2H, J 8.3, $SPhCl$), 5.55 (d, 1H, J 10.3, H-1_GlcN), 4.96 (d, 1H, J 3.9 Hz, H-1_Fuc), 4.96 (d, 1H, J 11.4 Hz, Bn), 4.84 (d, 1H, J 12.1 Hz, Bn), 4.80 (d, 1H, J 11.7 Hz, Bn), 4.74 (d, 1H, J 11.8 Hz, Bn), 4.66 (d, 1H, J 11.6 Hz, Bn), 4.61 (d, 1H, J 11.5 Hz, Bn), 4.38 (d, 1H, J 11.6 Hz, Hz), 4.35 (d, 1H, J 11.9 Hz, Bn), 4.31 (m, 1H, H-3_GlcN), 4.22 (dd, 1H, J 9.9, 9.9 Hz, H-2_GlcN), 4.07 (dd, 1H, J 10.2, 3.7 Hz, H-2_Fuc), 4.02 (dq, 1H, J 6.4, ~ 1 Hz, H-5_Fuc), 3.95 (high order m, 1H, H-6a_GlcN), 3.87 (dd, 1H, J 10.0, 2.9 Hz, H-3_Fuc), 3.81 – 3.72 (m, 2H, H-5_GlcN + H-6b_GlcN), 3.65 (br, 1H, H-4_Fuc), 3.49 (t, 2H, J 8.7 Hz, CH_2N_3), 1.57 (s, 1H, OH-3_GlcN), 1.07 (d, 3H, J 6.5 Hz, H-6_Fuc); m/z (ESI-HRMS) calcd for $[C_{54}H_{52}NO_{10}SCl + Na]^+$ 964.2893, found 964.2900.

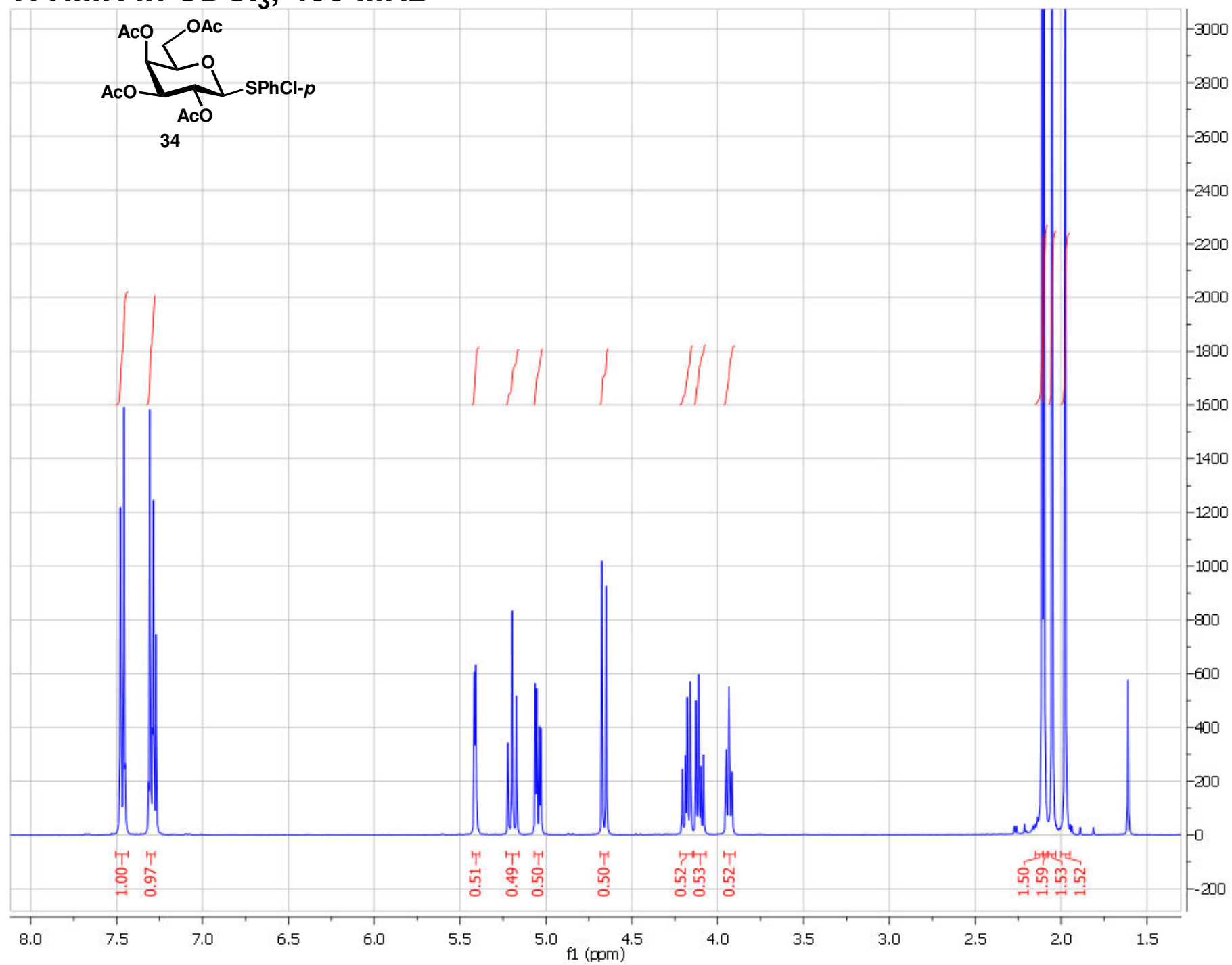
II. 1H and ^{13}C spectra of all synthesized compounds

The 1H and ^{13}C spectra of all synthesized compounds and the 1H - 1H GCOSY and 1H - ^{13}C GHSQC of the final products.

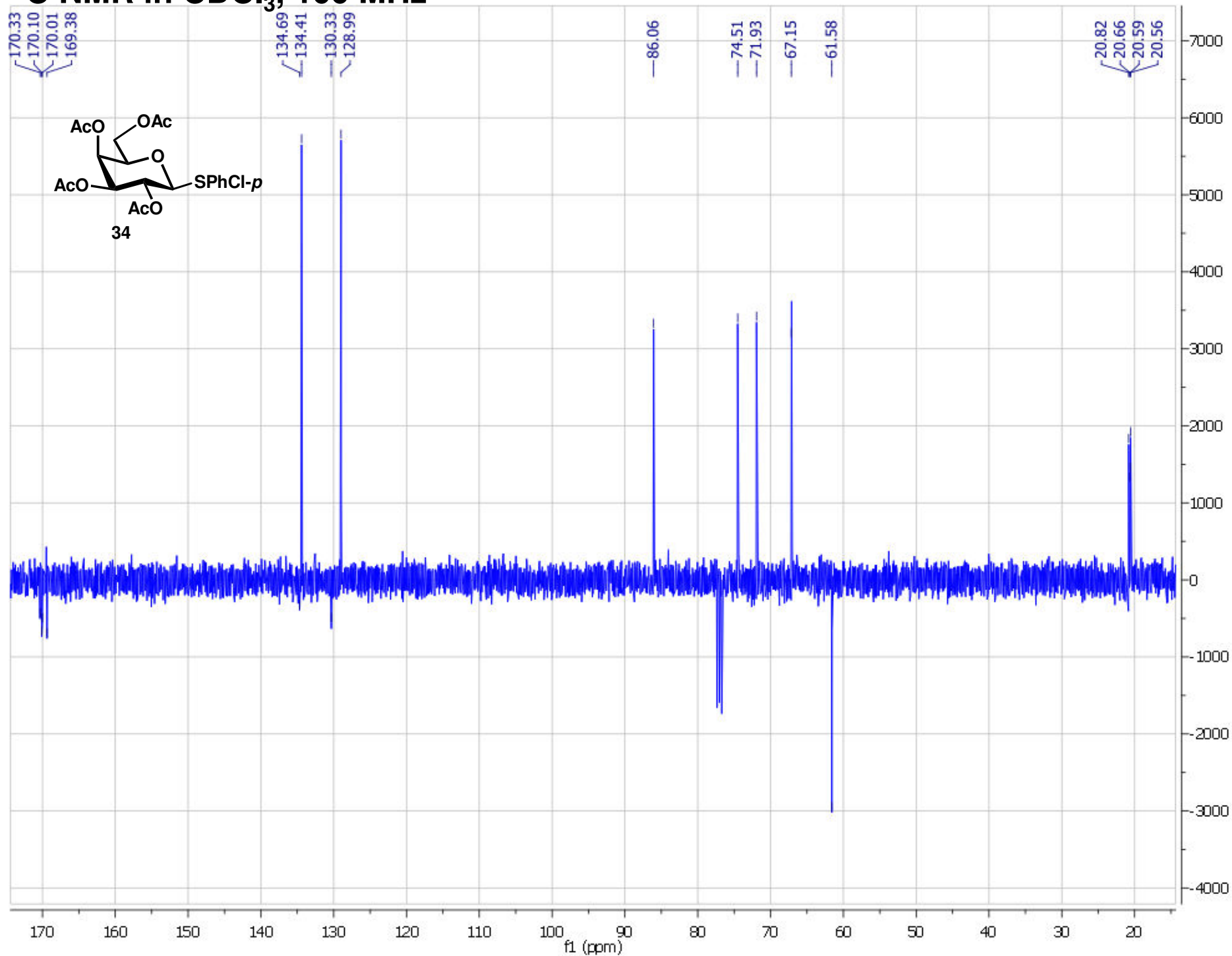
¹ M. Malkoch, K. Schleicher, E. Drockenmuller, C. J. Hawker, T. P. Russel, P. Wu and V. V. Fokin, *Macromolecules*, 2005, **38**, 3663-3678.

² (a) H. Rathore, A. H. L. From, K. Ahmed and D. S. Fullerton, *J. Med. Chem.*, 1986, **29**, 1945-1952. (b) D. P. Larson, C. H. Heathcock and H. Clayton, *J. Org. Chem.*, 1997, **62**, 8406-8418.

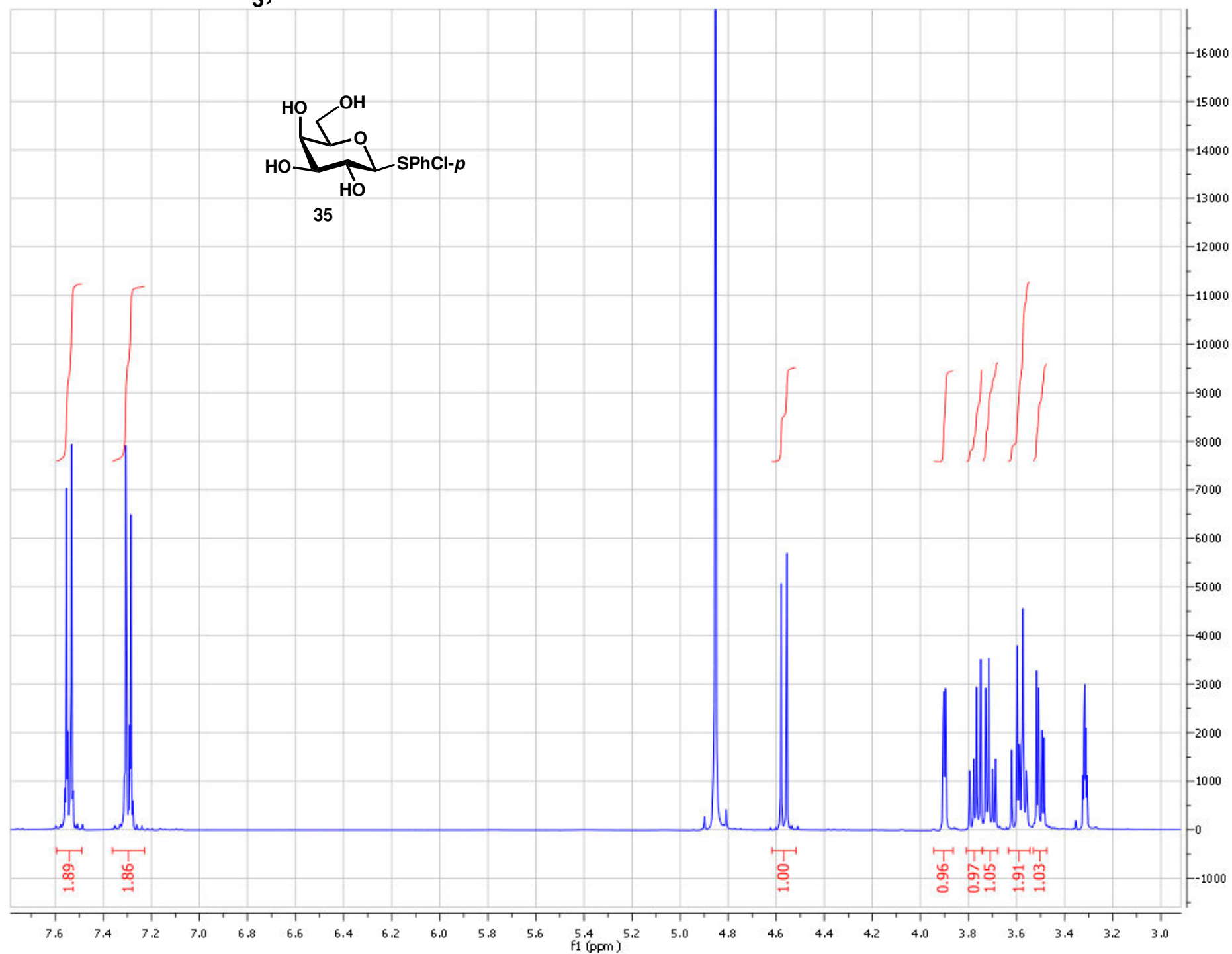
^1H NMR in CDCl_3 , 400 MHz



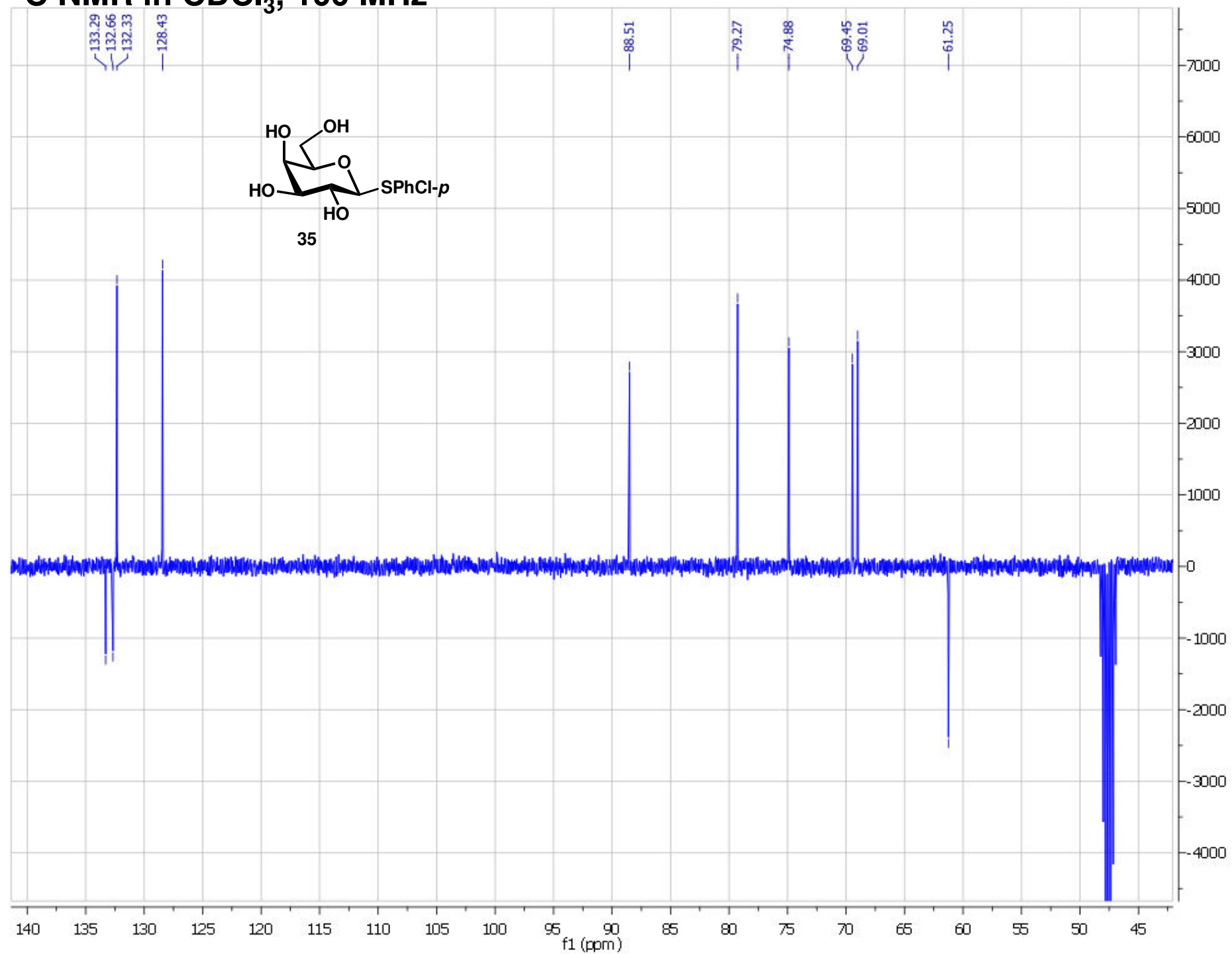
¹³C NMR in CDCl₃, 100 MHz



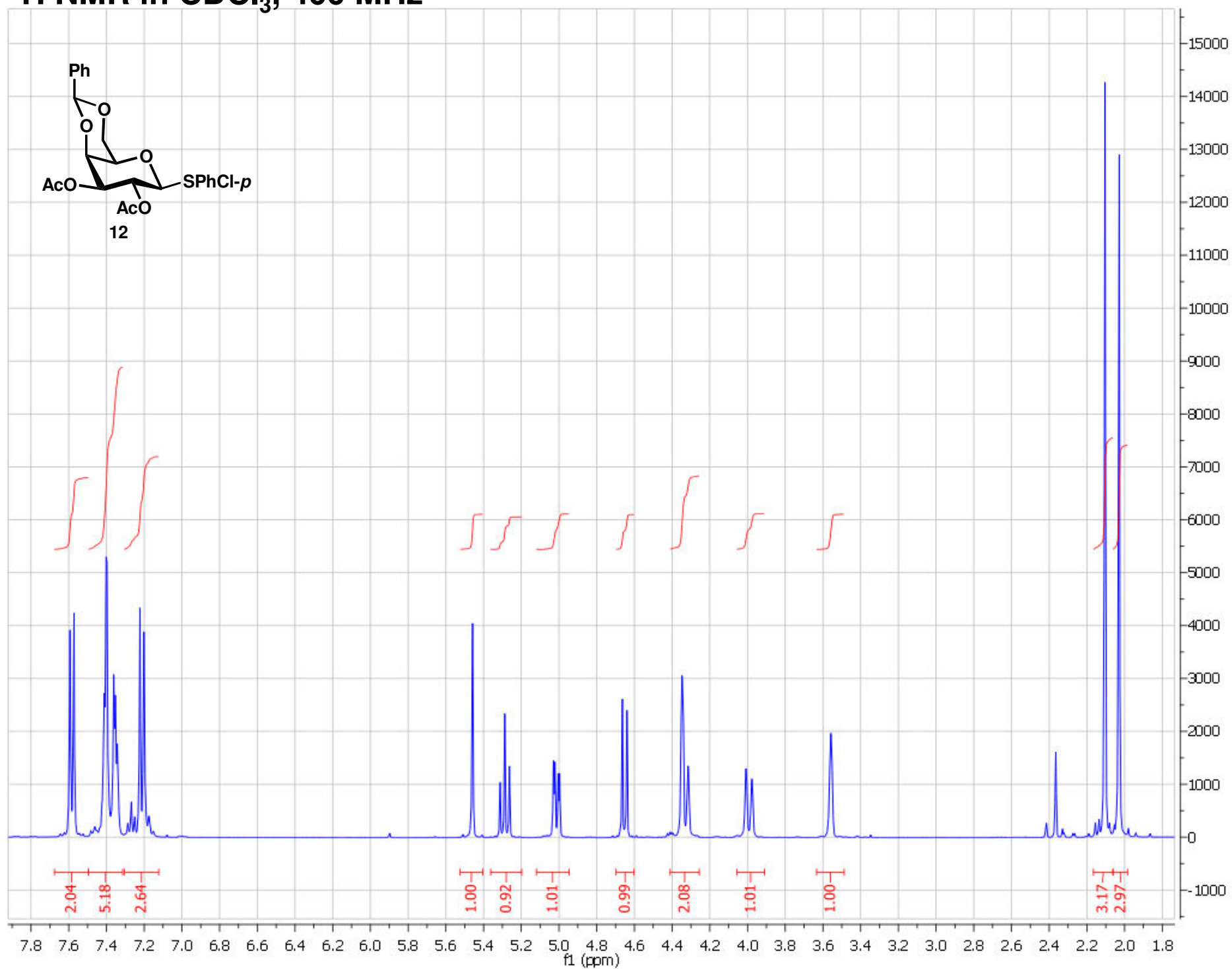
^1H NMR in CDCl_3 , 400 MHz



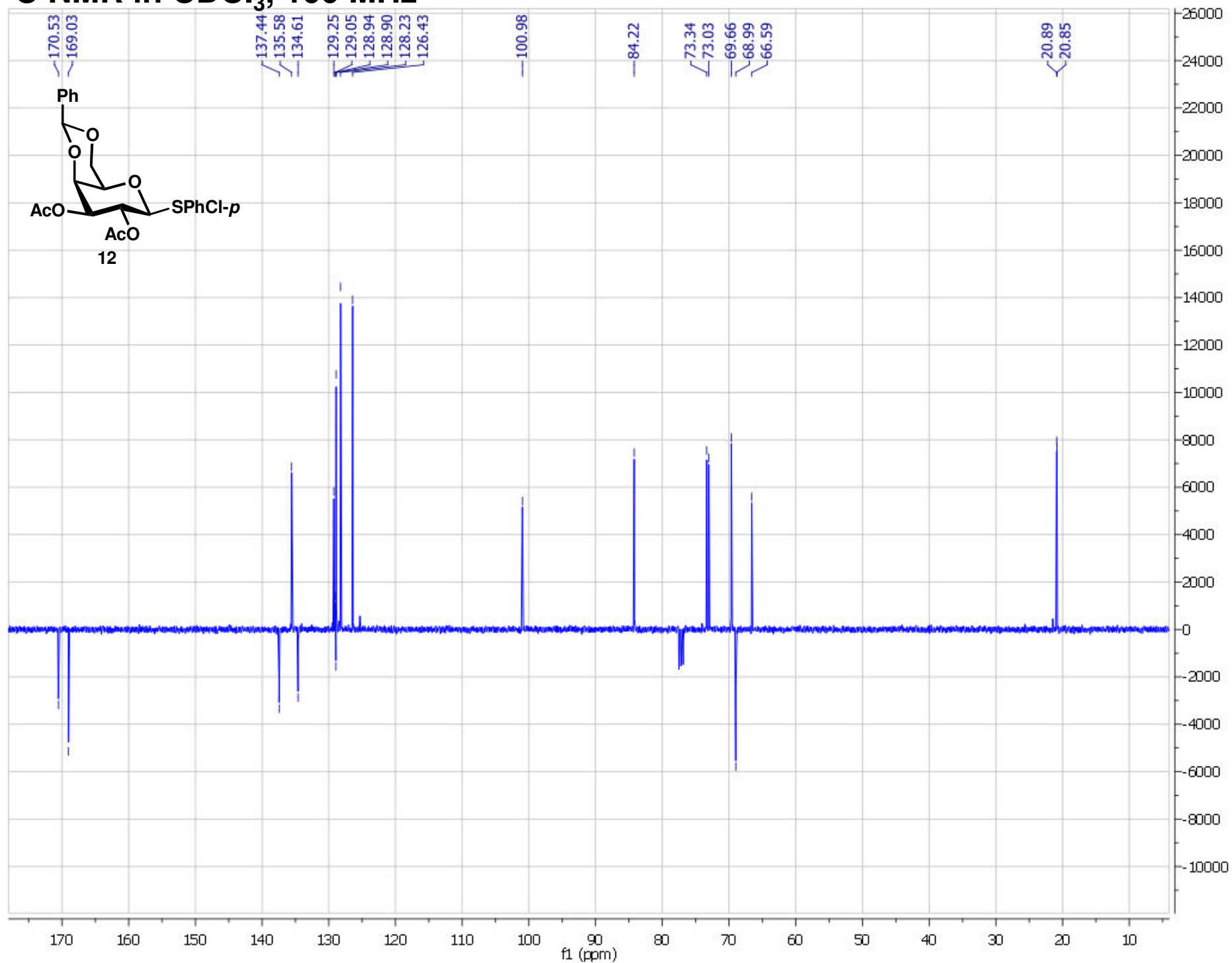
^{13}C NMR in CDCl_3 , 100 MHz



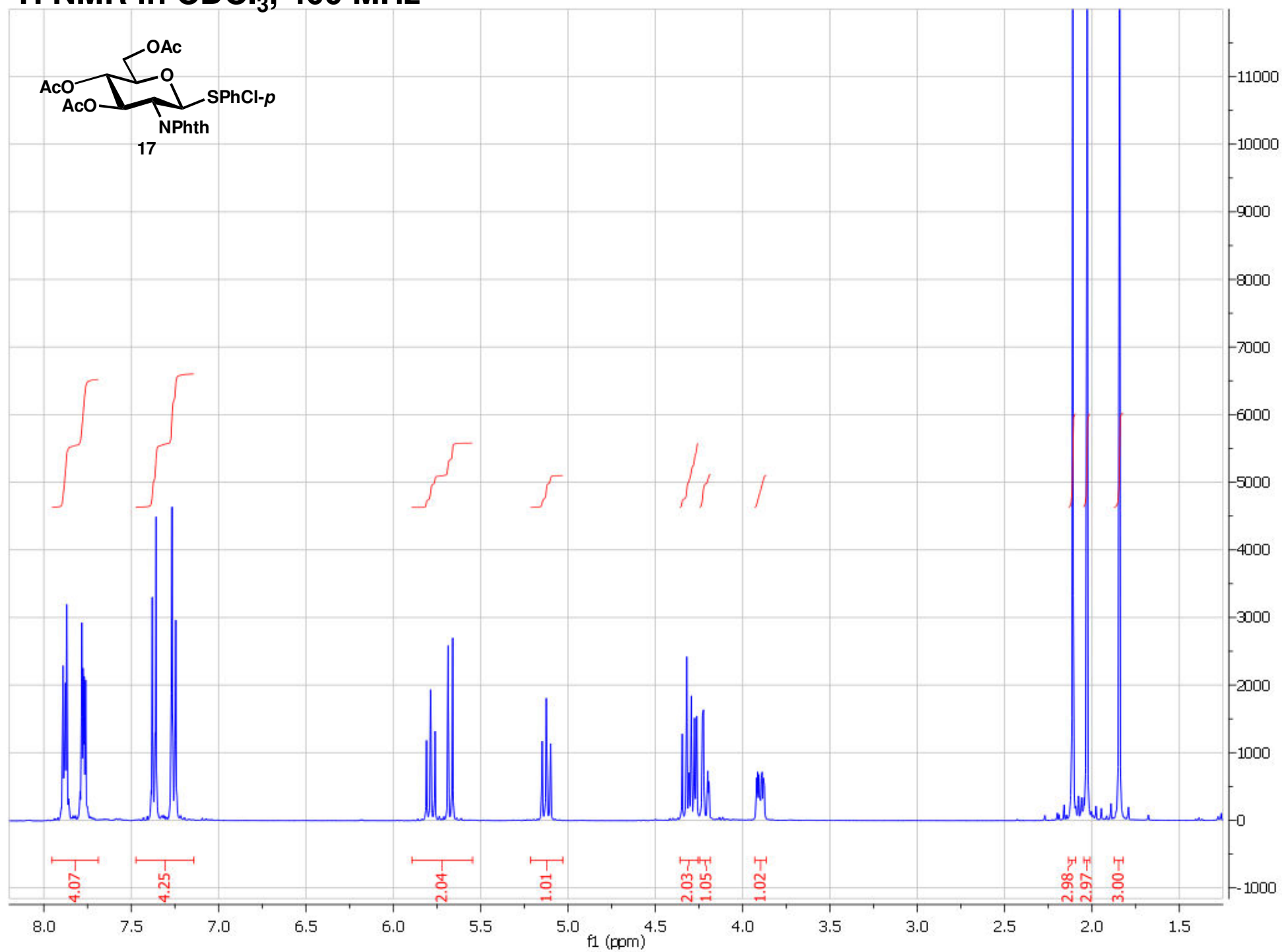
^1H NMR in CDCl_3 , 400 MHz



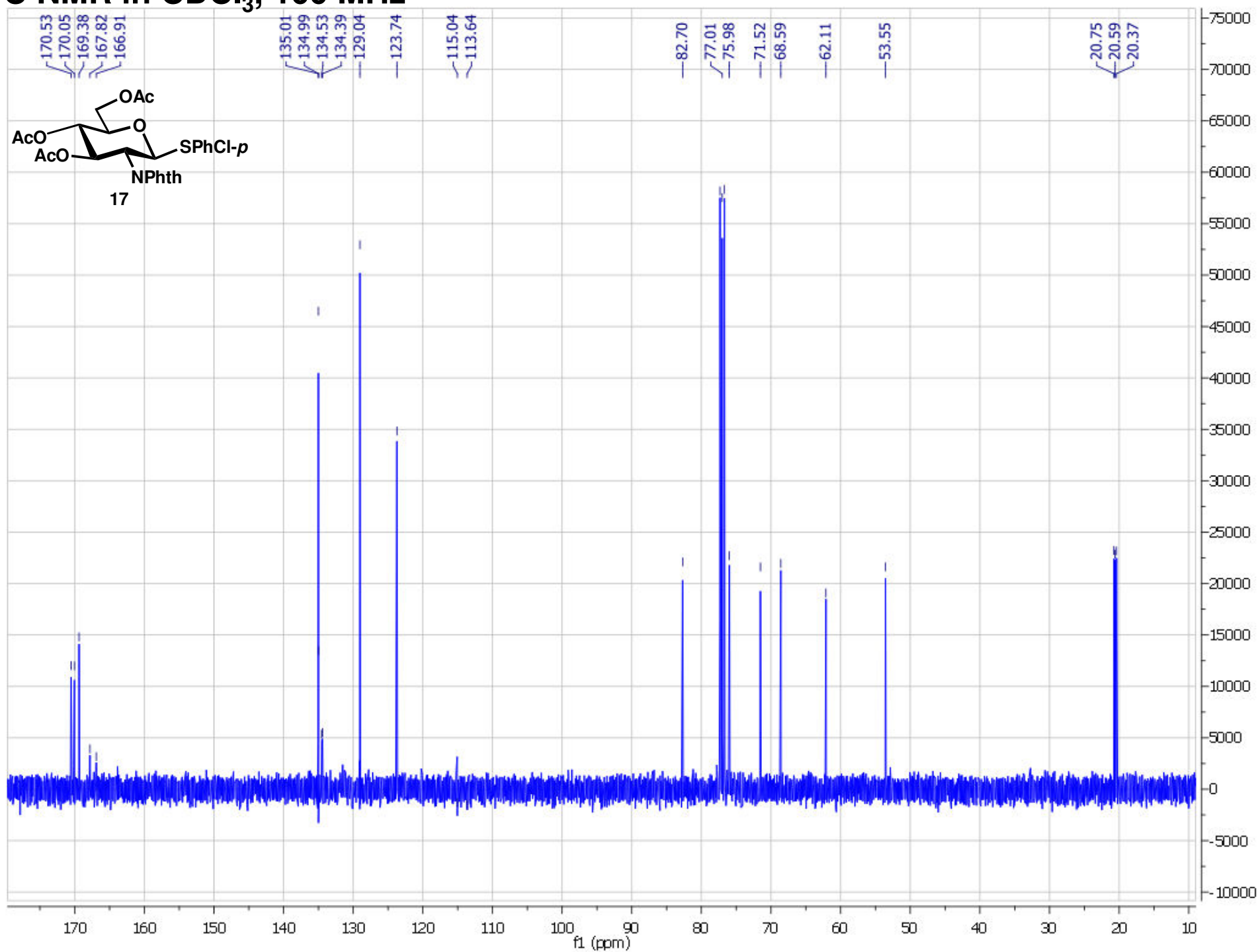
^{13}C NMR in CDCl_3 , 100 MHz



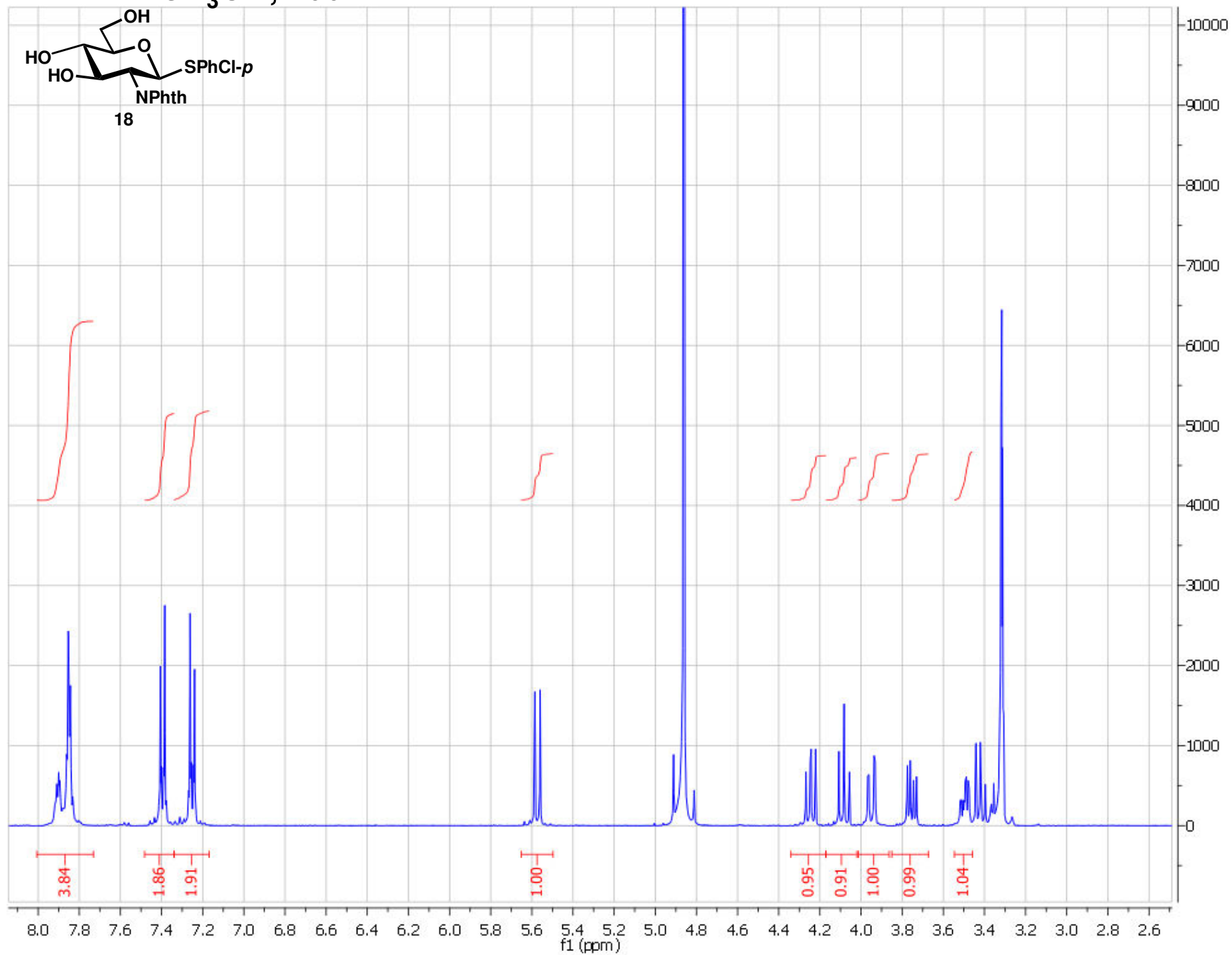
^1H NMR in CDCl_3 , 400 MHz



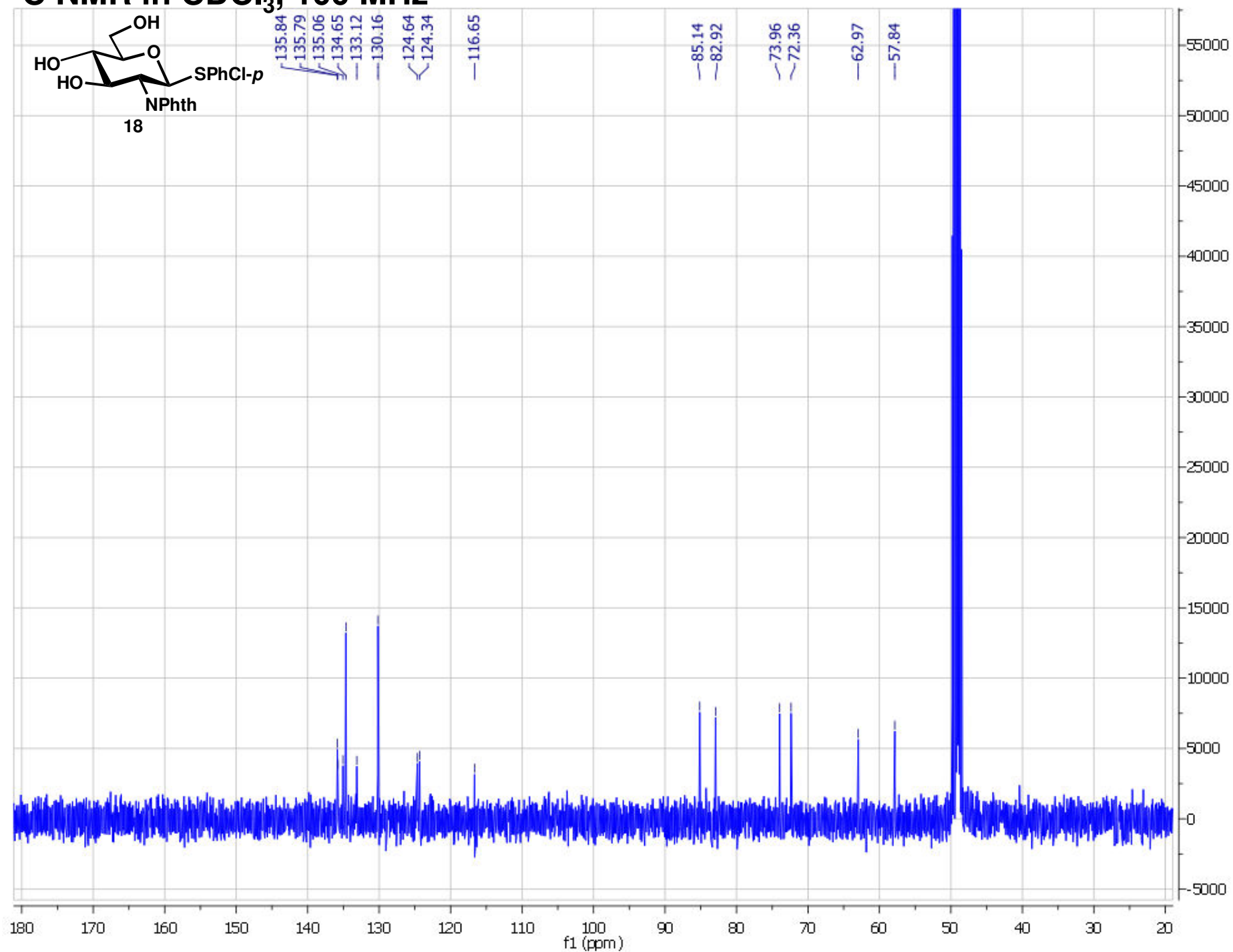
^{13}C NMR in CDCl_3 , 100 MHz



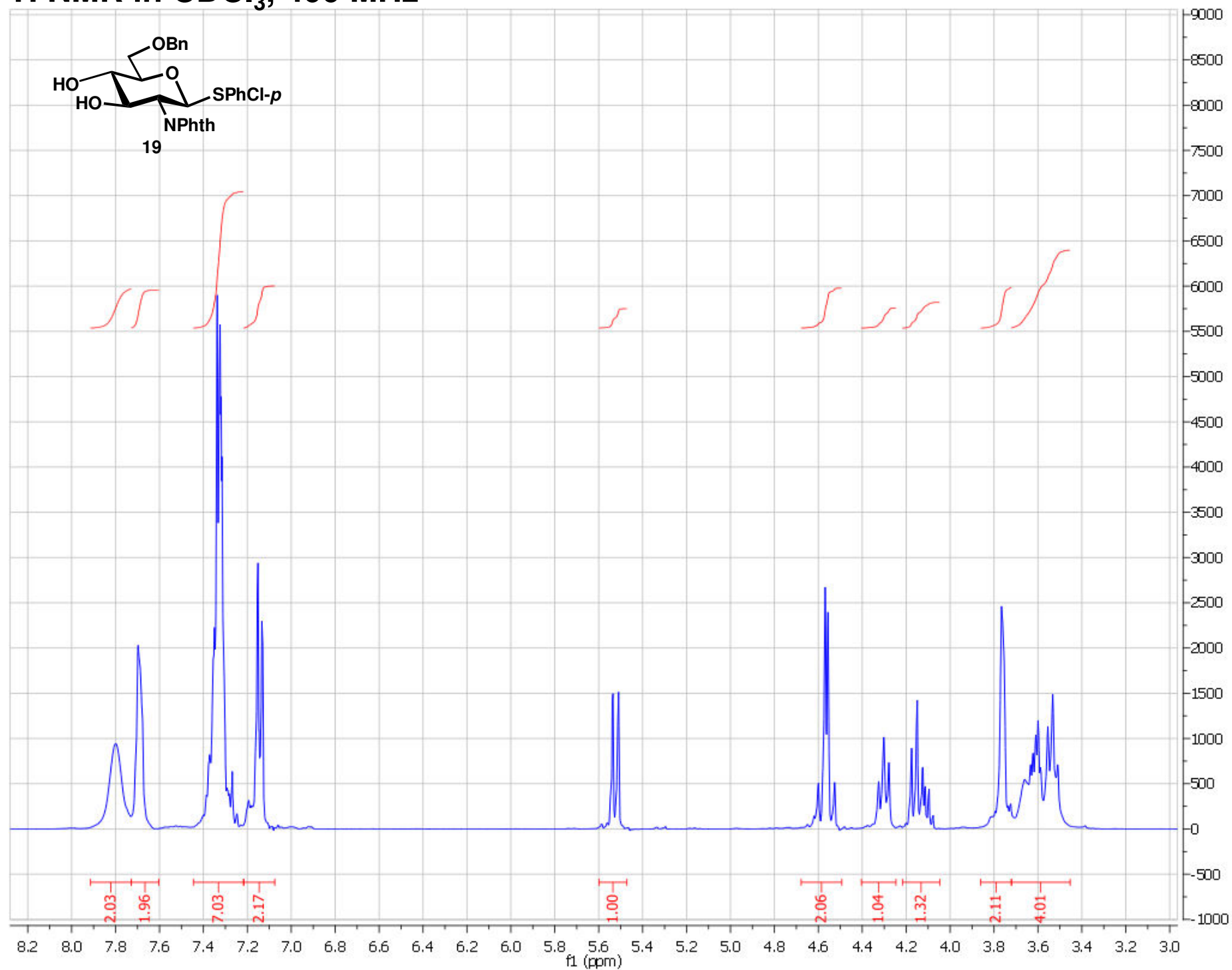
^1H NMR in CD_3OD , 400 MHz



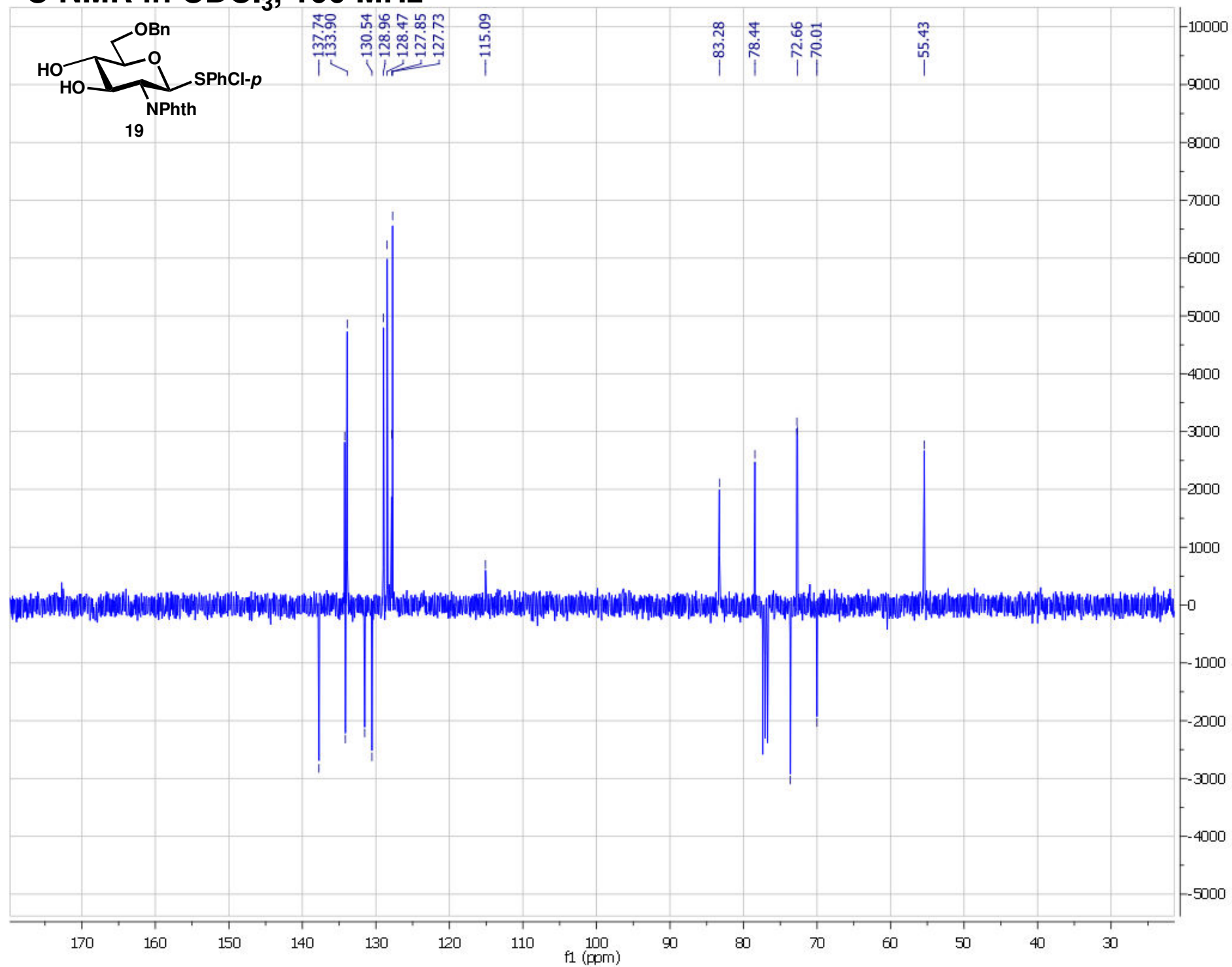
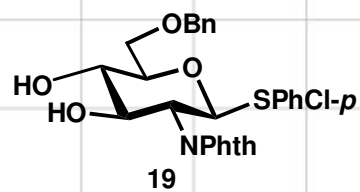
^{13}C NMR in CDCl_3 , 100 MHz



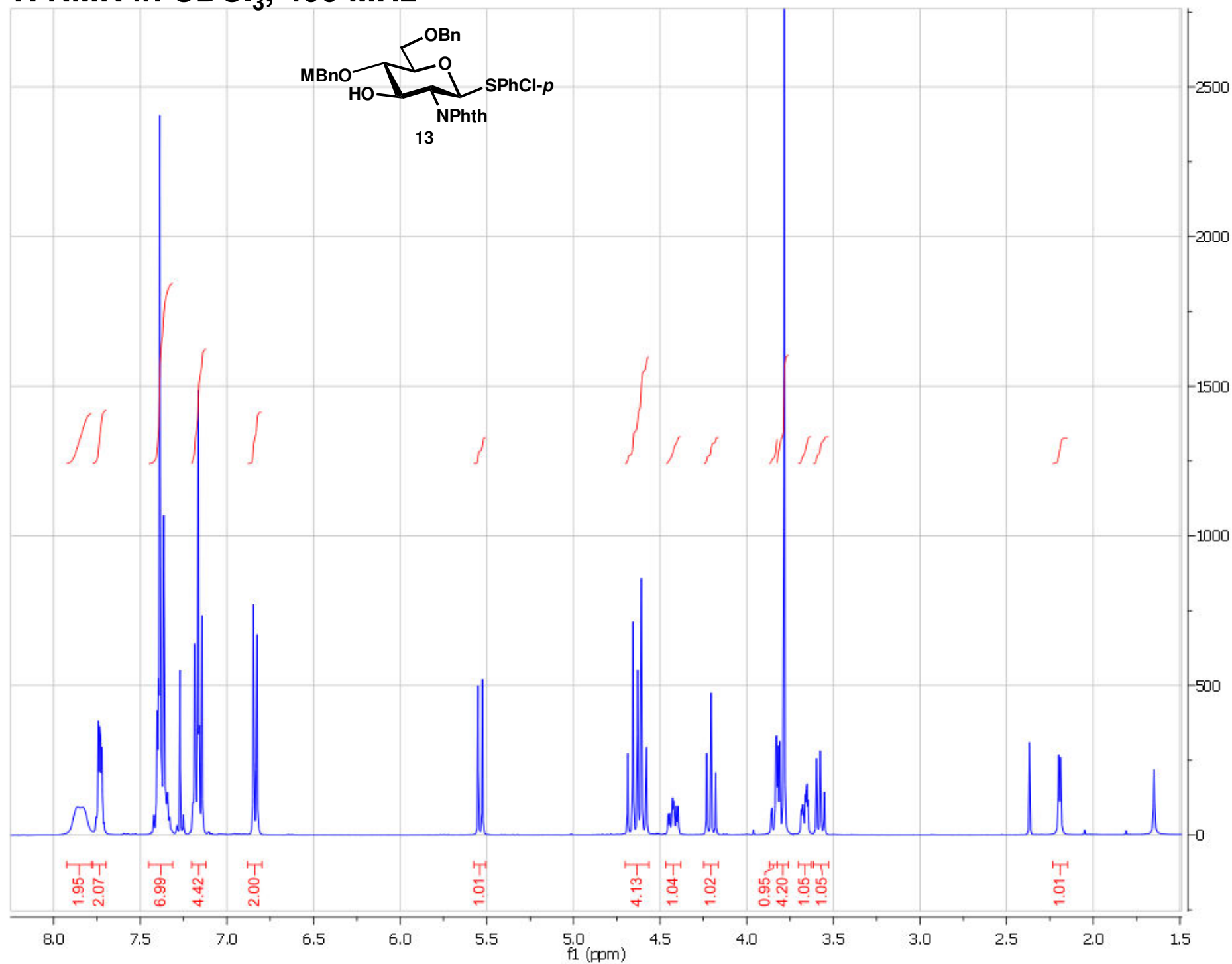
^1H NMR in CDCl_3 , 400 MHz



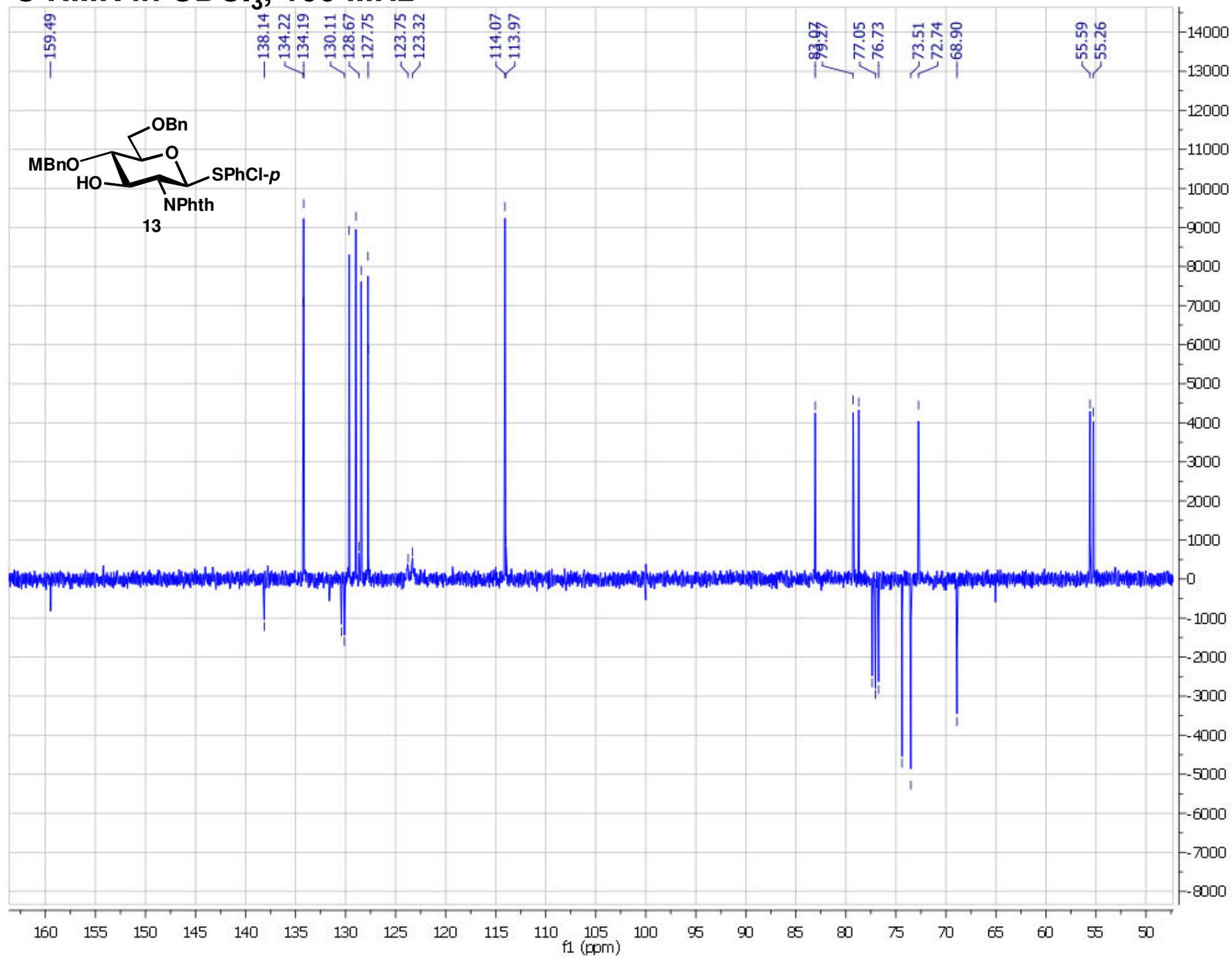
¹³C NMR in CDCl₃, 100 MHz



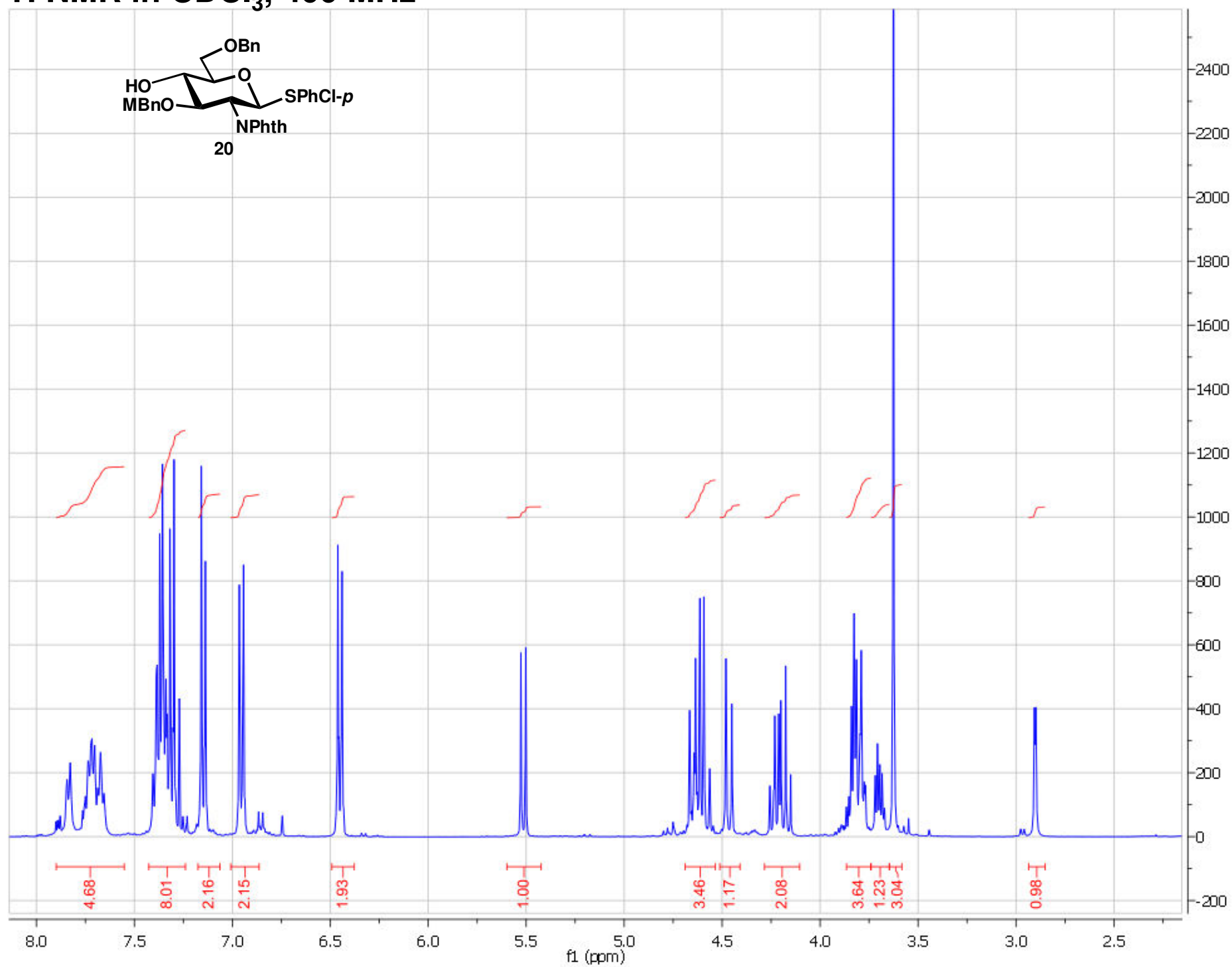
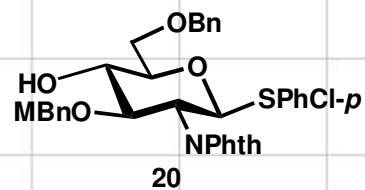
^1H NMR in CDCl_3 , 400 MHz



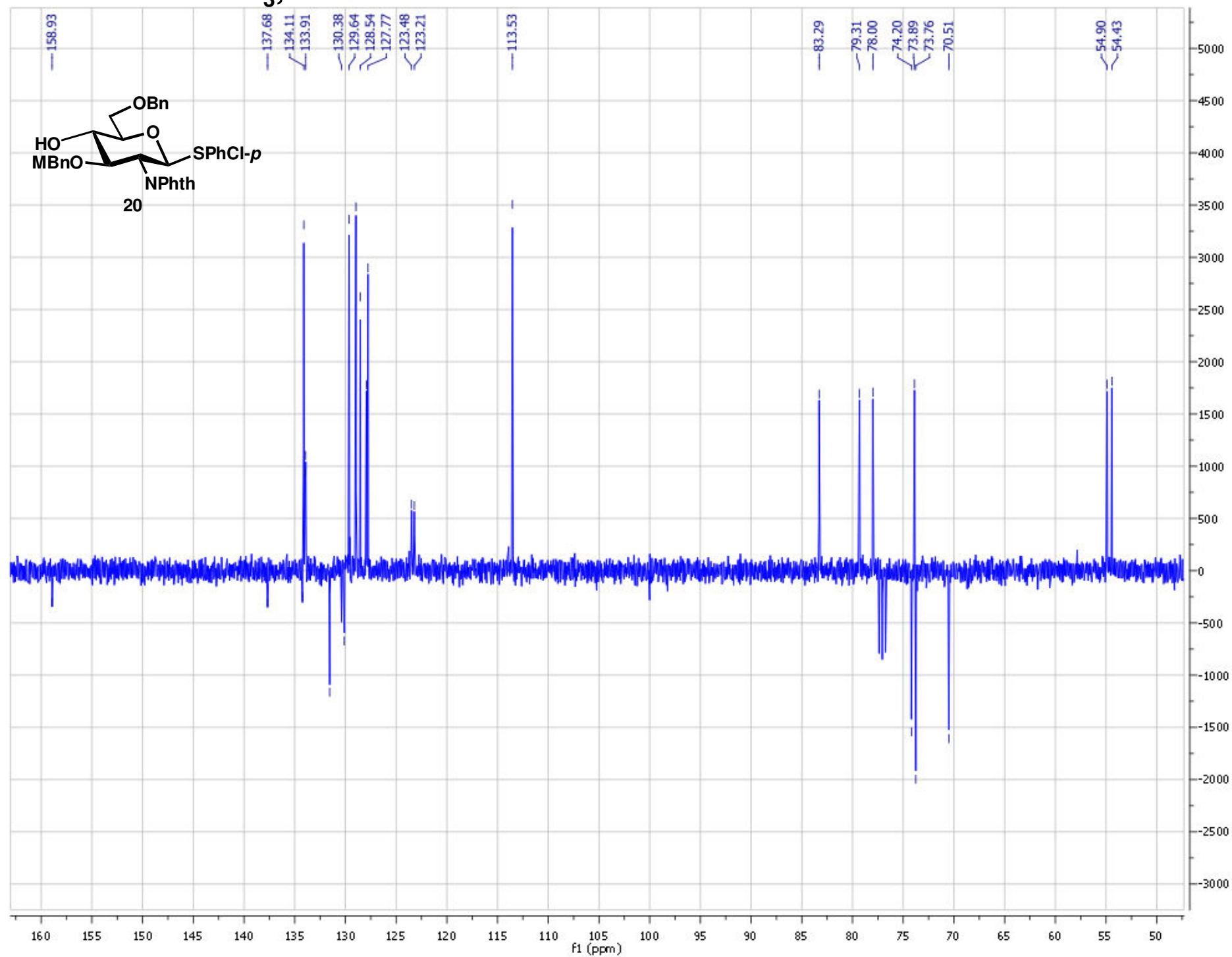
^{13}C NMR in CDCl_3 , 100 MHz



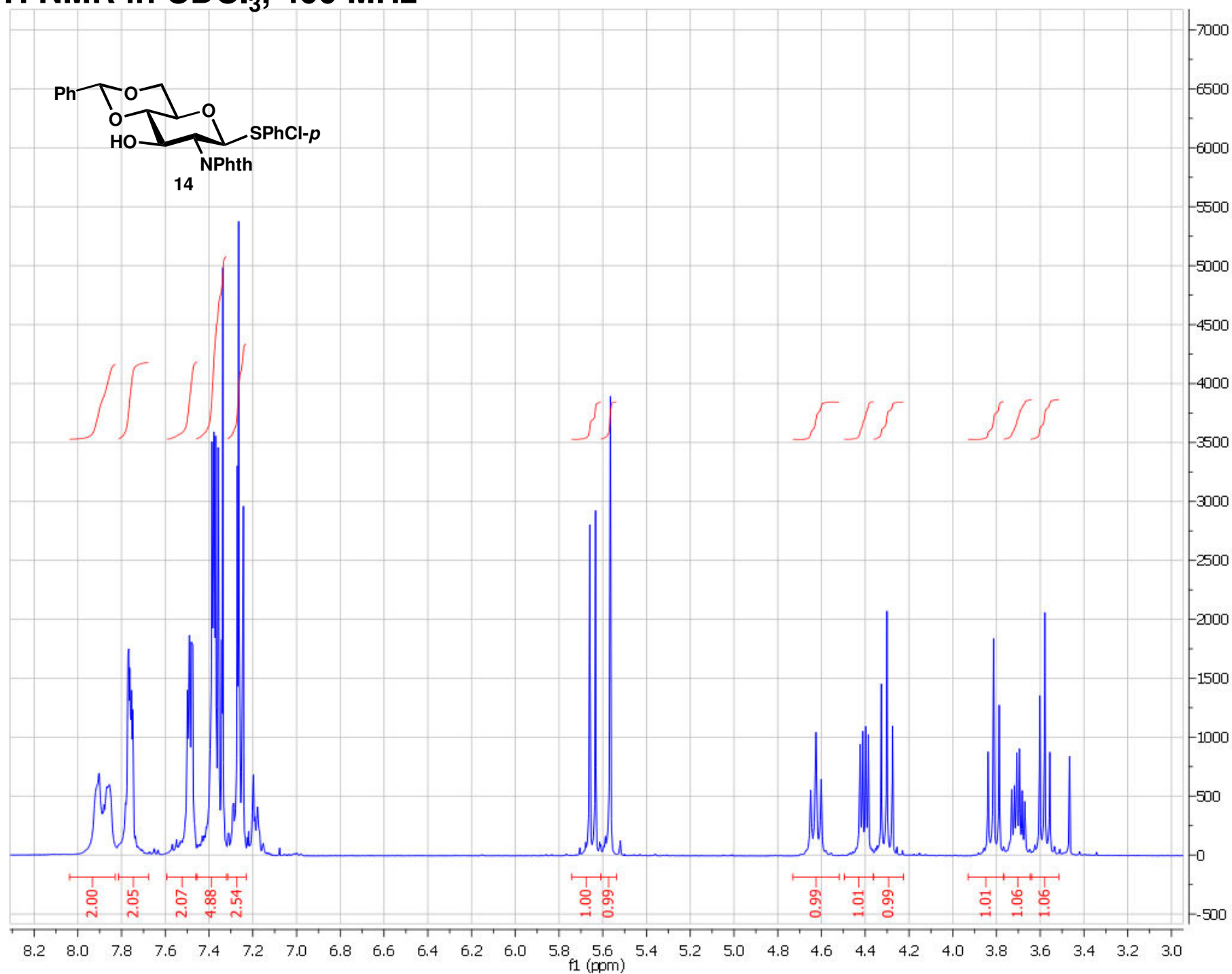
^1H NMR in CDCl_3 , 400 MHz



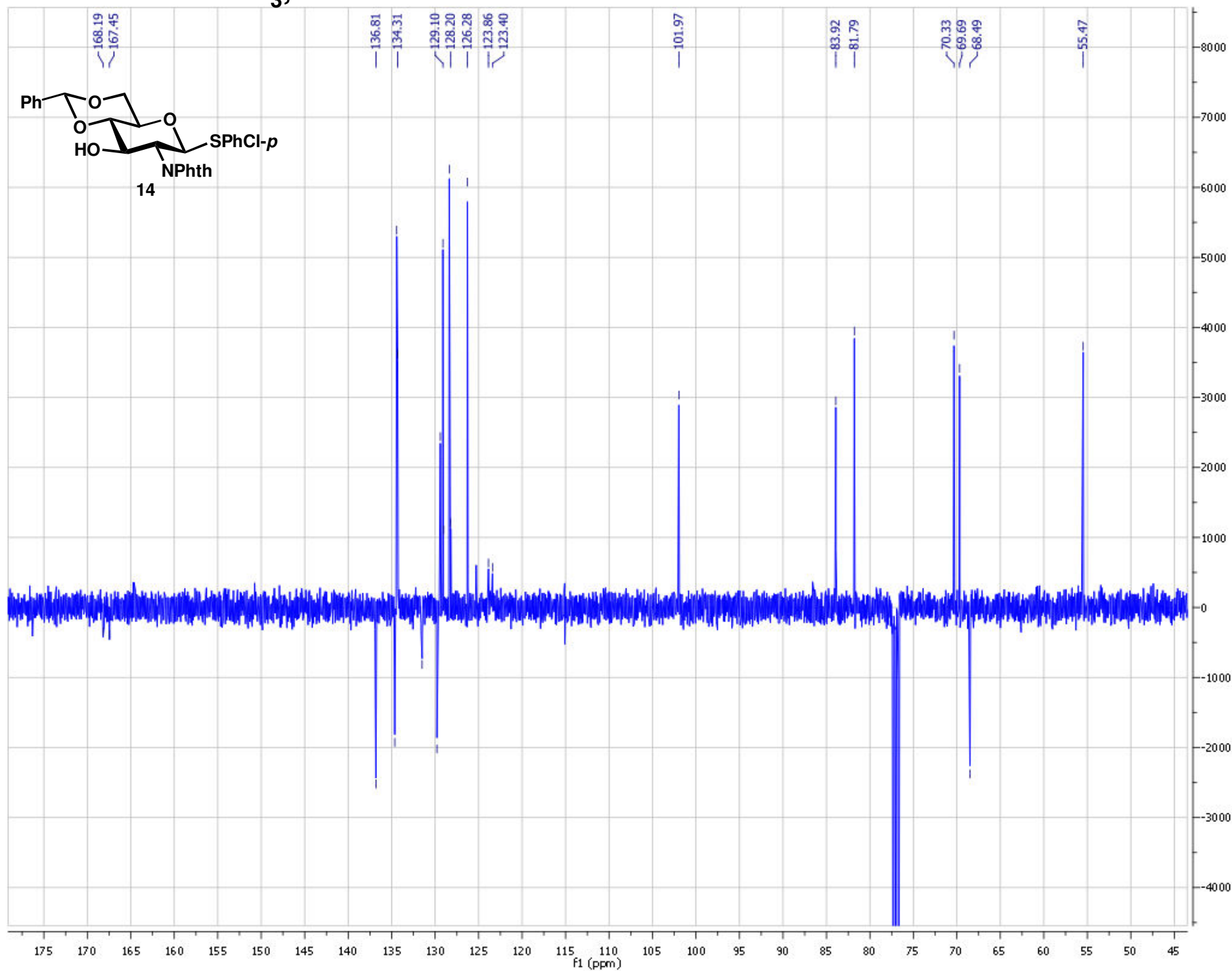
^{13}C NMR in CDCl_3 , 100 MHz



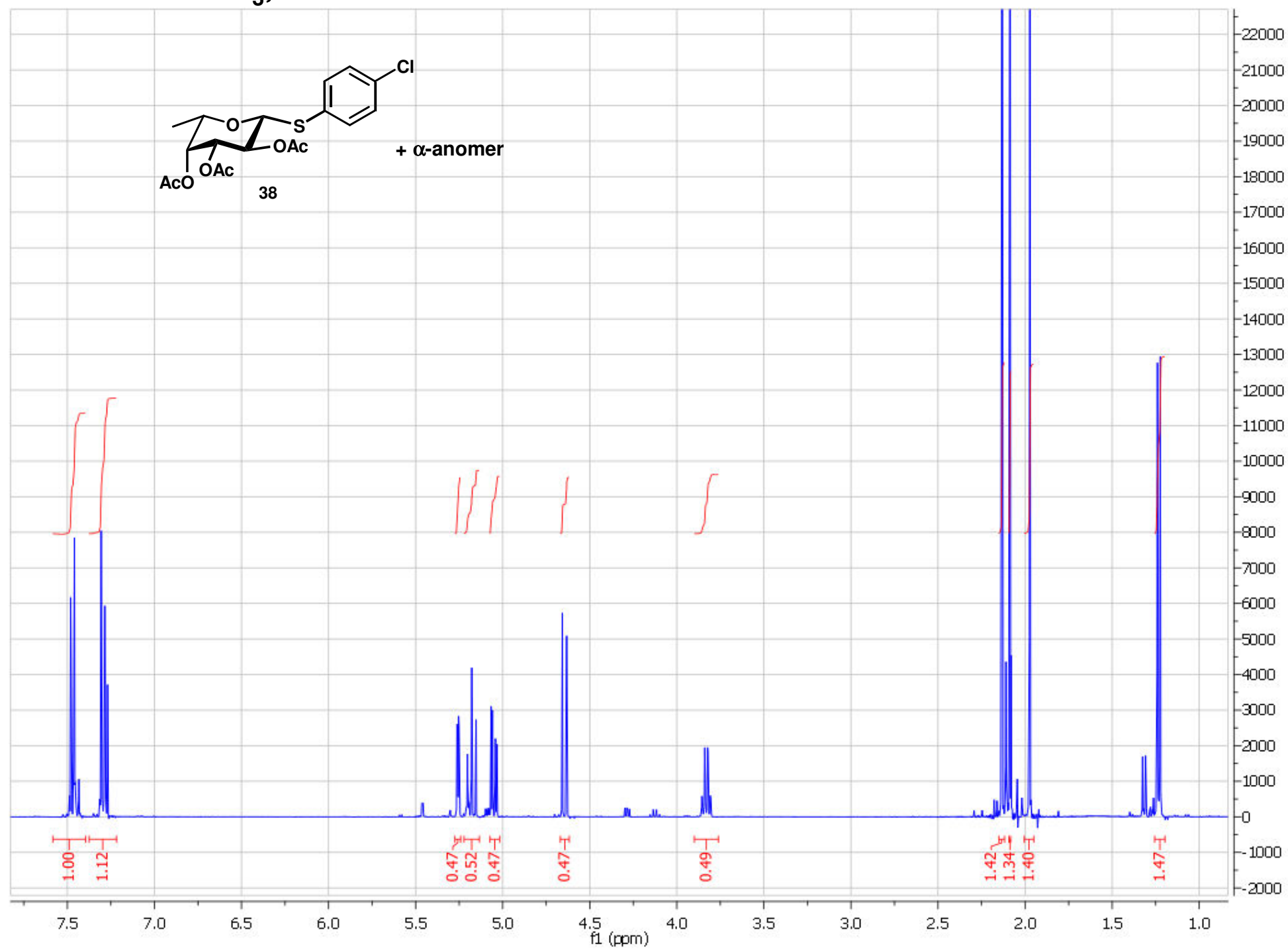
^1H NMR in CDCl_3 , 400 MHz



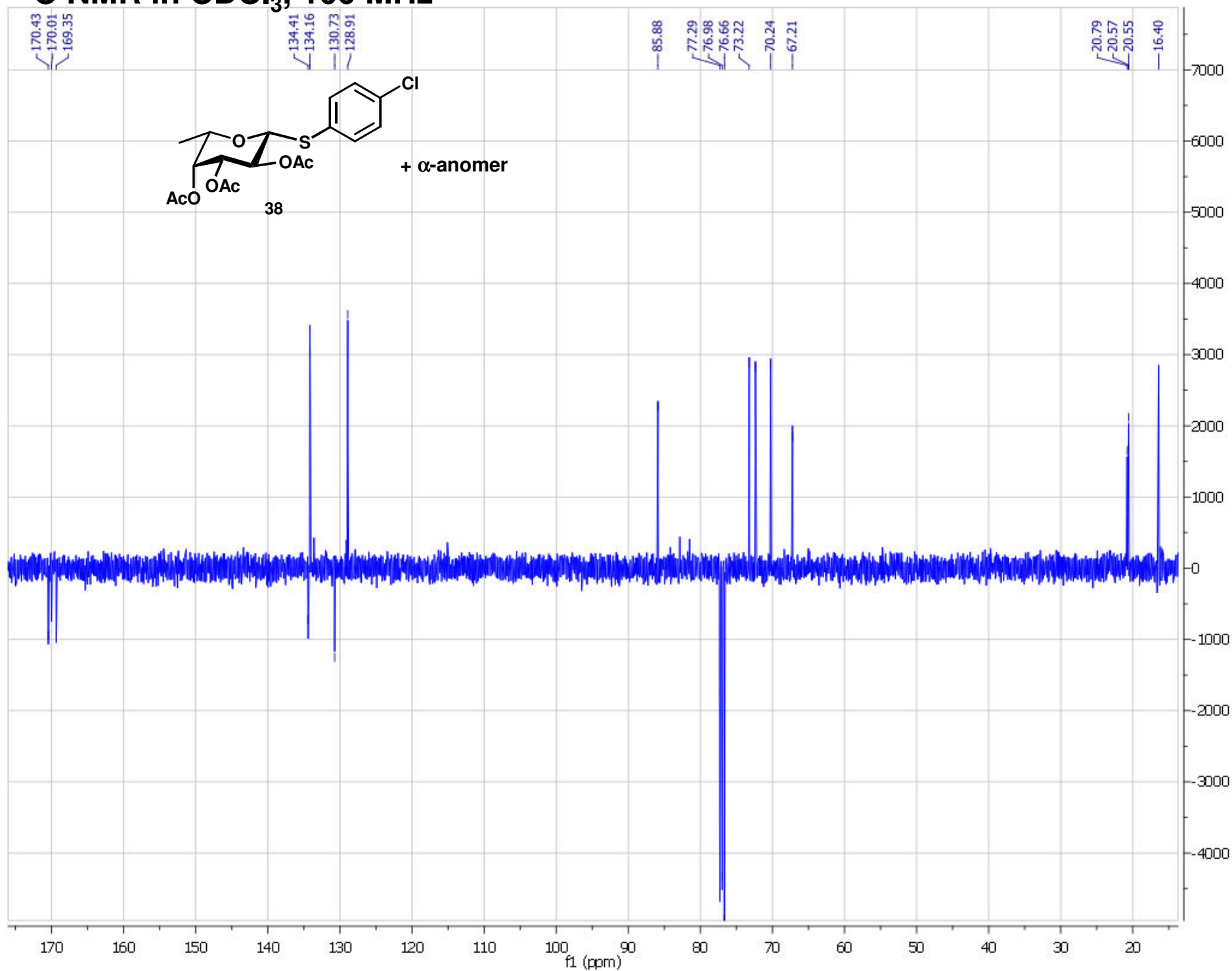
^{13}C NMR in CDCl_3 , 100 MHz



^1H NMR in CDCl_3 , 400 MHz



¹³C NMR in CDCl₃, 100 MHz

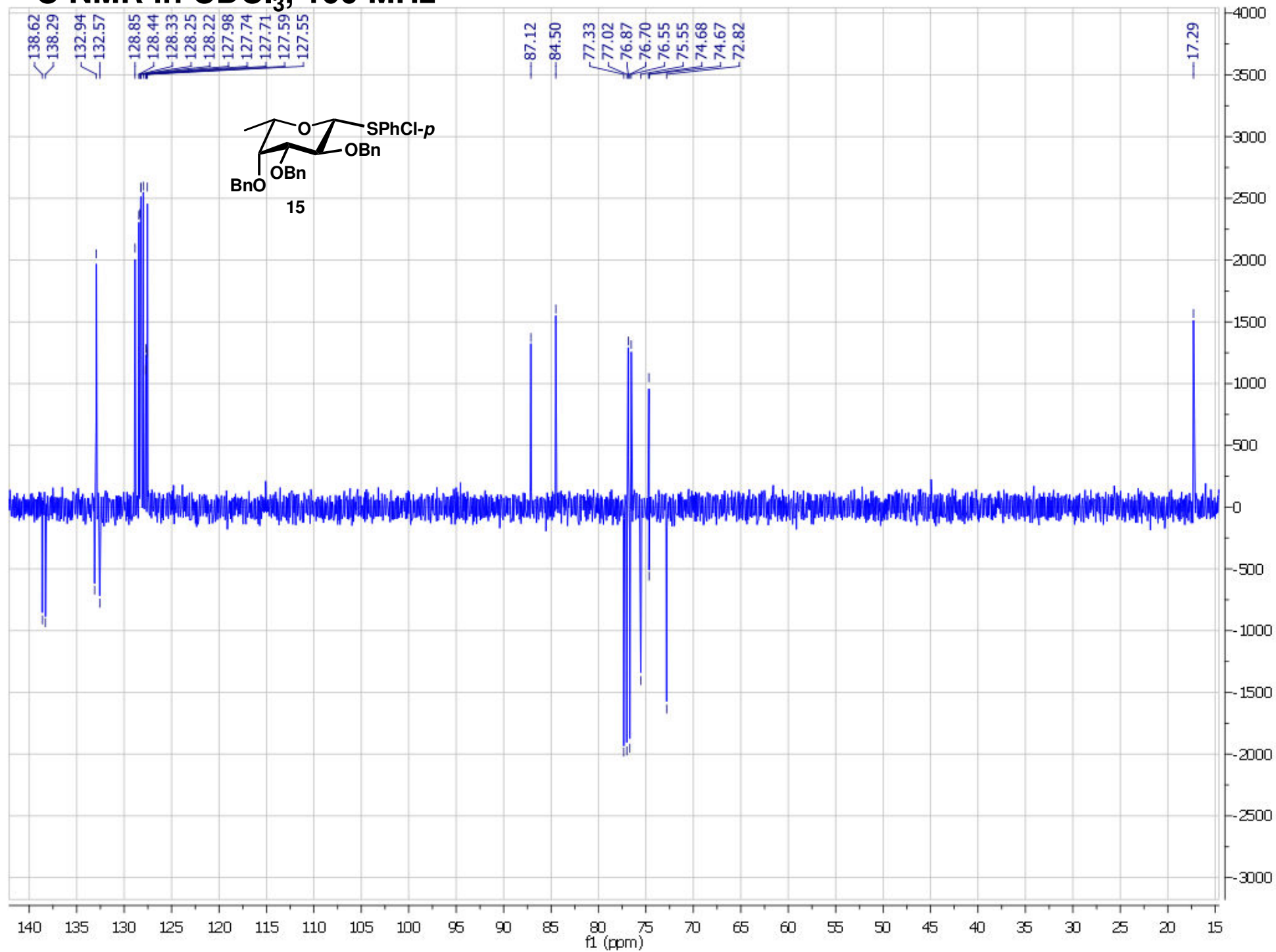


Chemical structure of compound 15 is shown in the top left. The structure is a cyclohexane ring with an oxygen atom at position 1, a BnO group at position 2, an OBn group at position 3, and an SPhCl-p group at position 4.

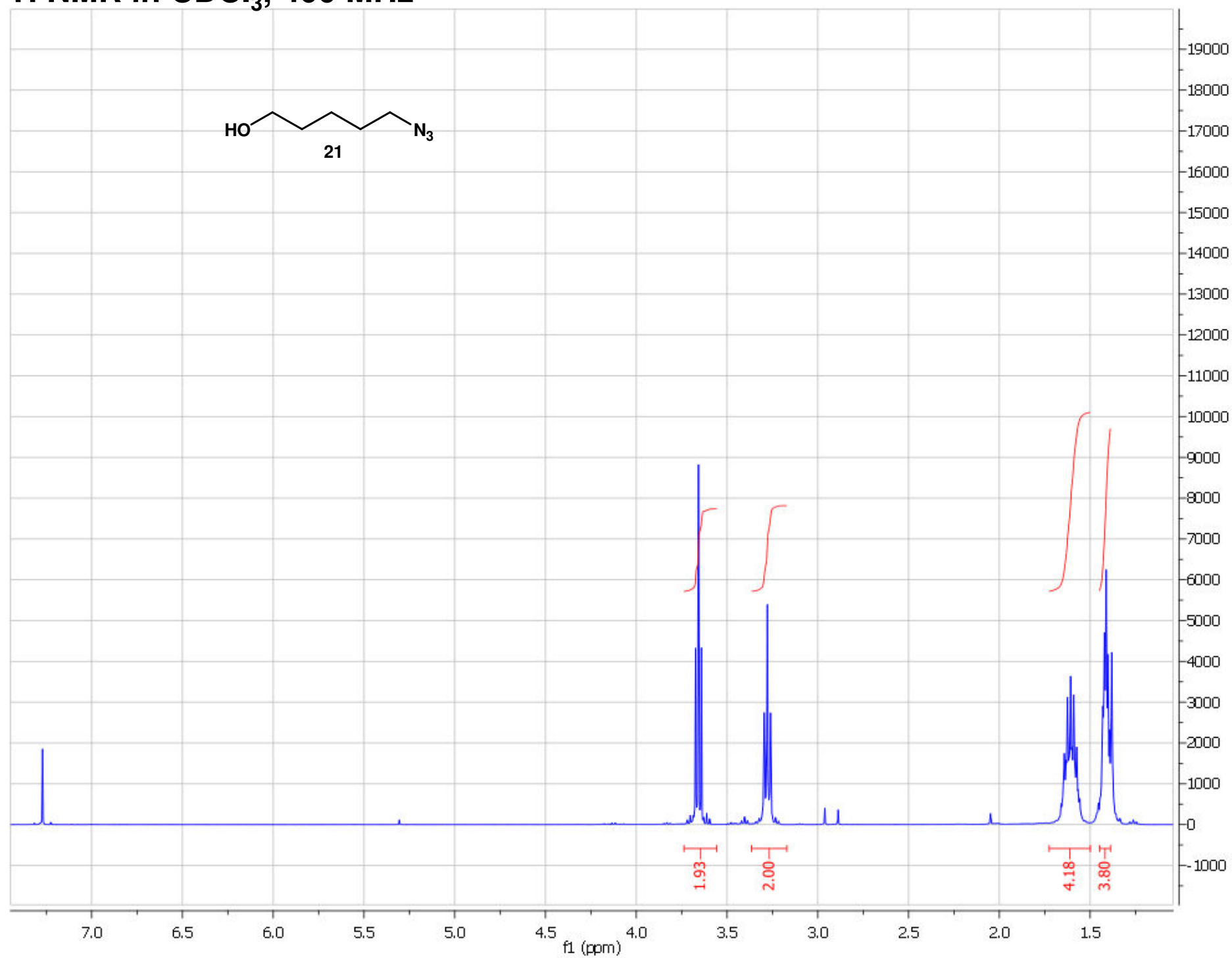
The ^1H NMR spectrum (400 MHz, CDCl_3) shows the following peaks (ppm) and integrations:

- 7.55, 7.53, 7.35, 7.27, 7.16, 7.14 (aromatic protons, integration 1.89, 13.26, 1.91)
- 5.05, 5.02, 4.77, 4.70, 4.67, 4.60, 4.57 (anomeric protons, integration 0.90, 3.76, 1.00, 0.96)
- 3.95, 3.92, 3.90, 3.67, 3.67, 3.64, 3.63, 3.61, 3.58, 3.56, 3.55, 3.53 (benzylic protons, integration 0.90, 0.93, 0.94, 0.93)
- 2.38, 2.28, 2.21 (methyl protons, integration 2.94)
- 1.31, 1.29 (methyl protons, integration 2.94)

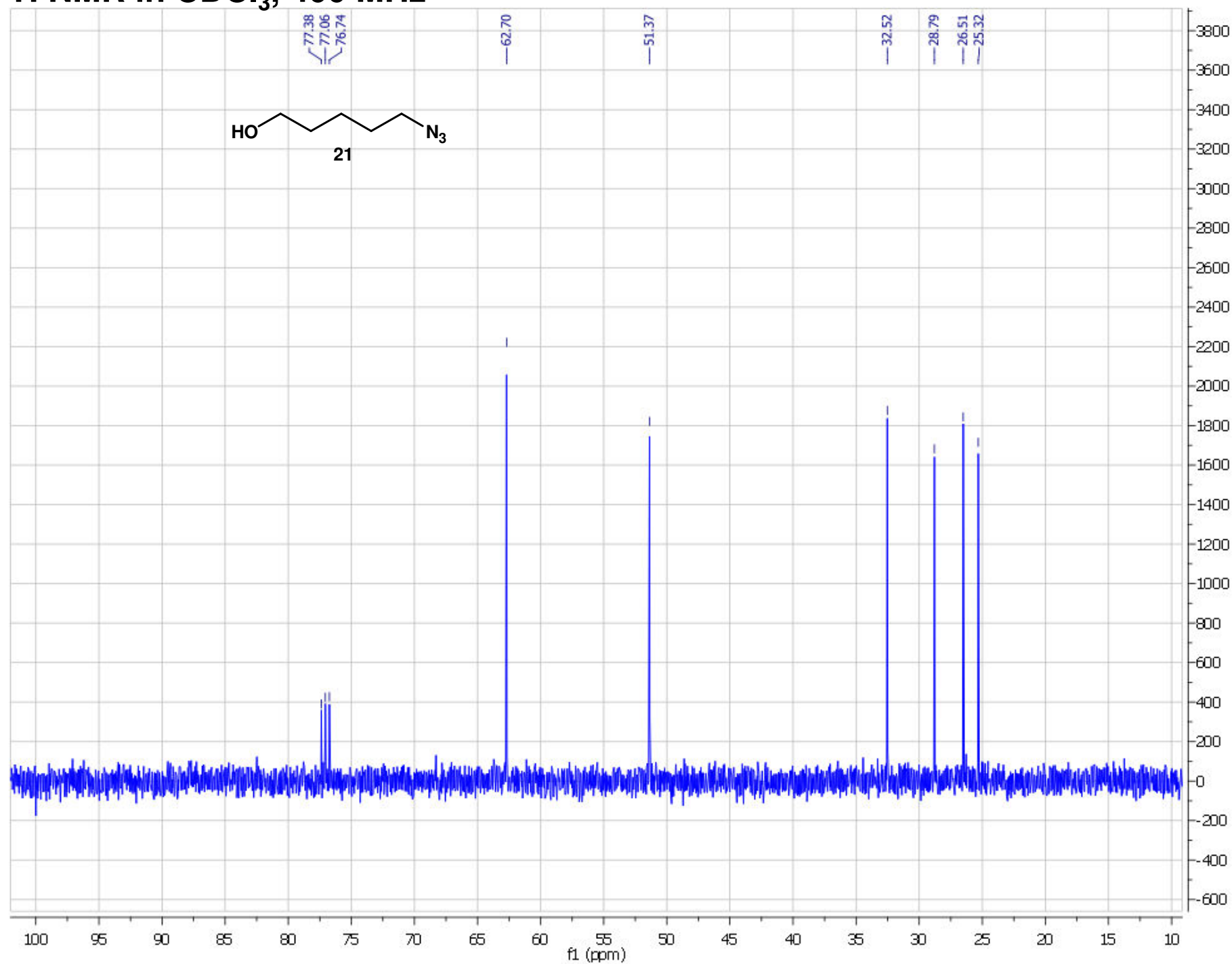
^{13}C NMR in CDCl_3 , 100 MHz



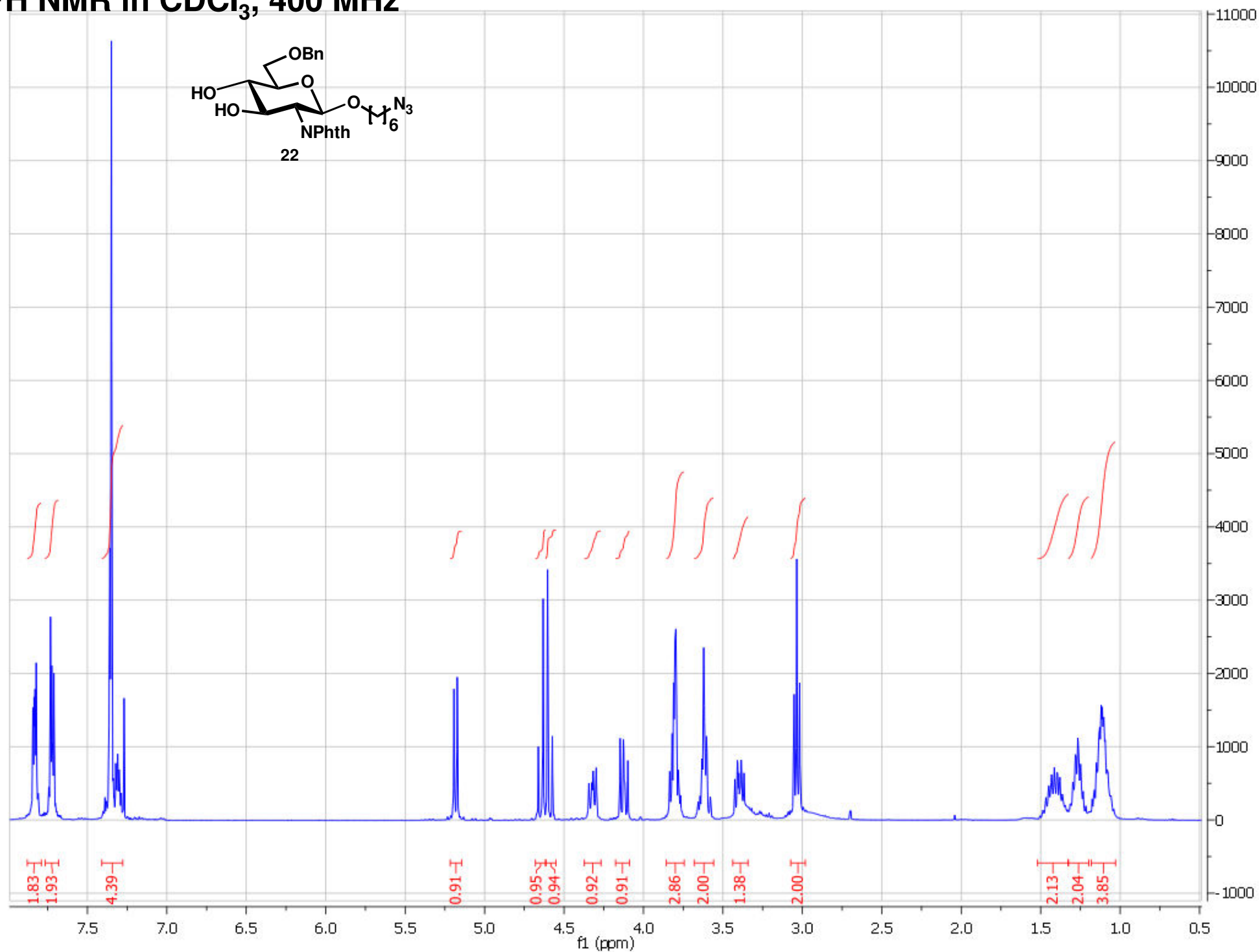
^1H NMR in CDCl_3 , 400 MHz



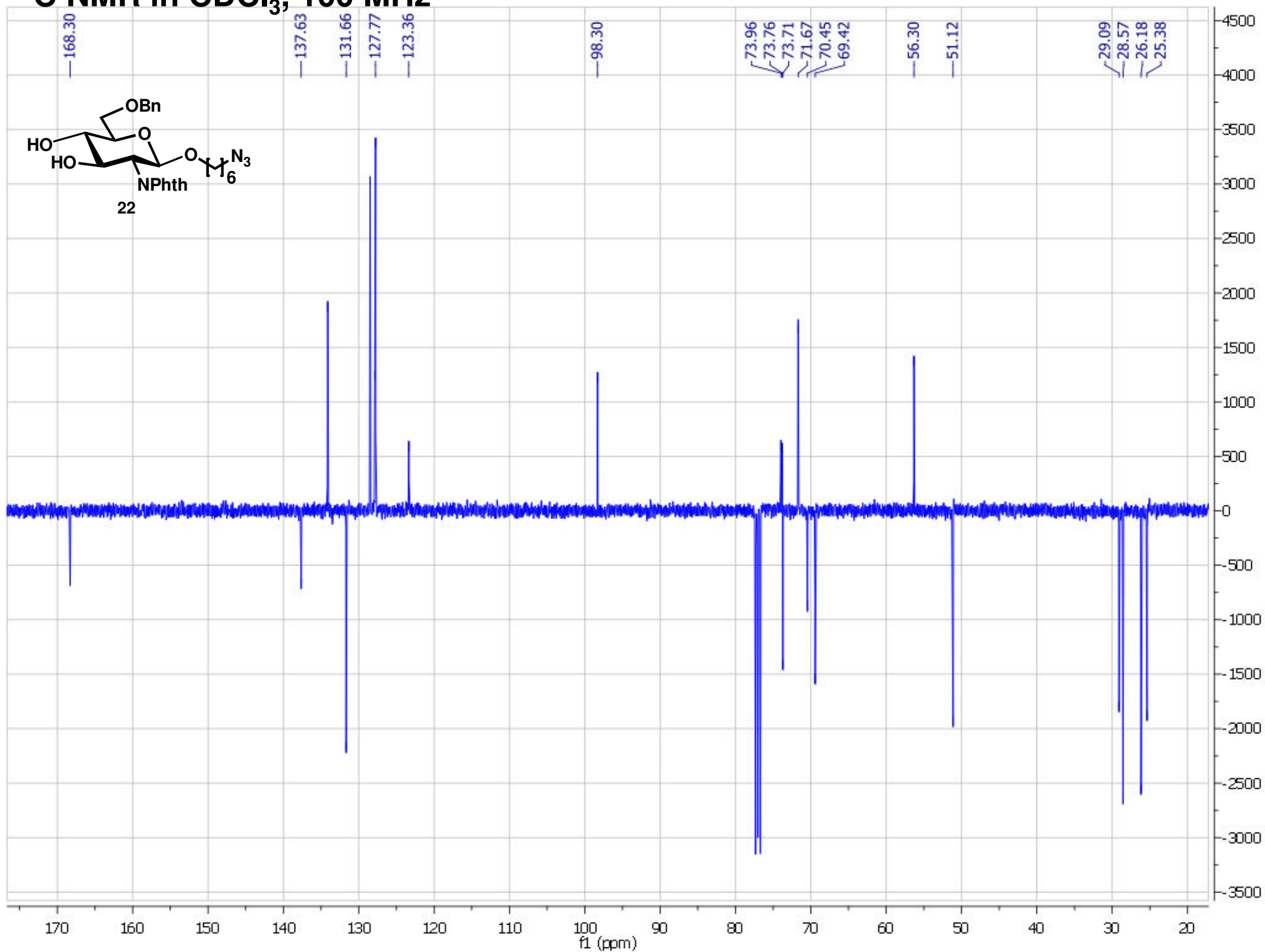
^1H NMR in CDCl_3 , 400 MHz



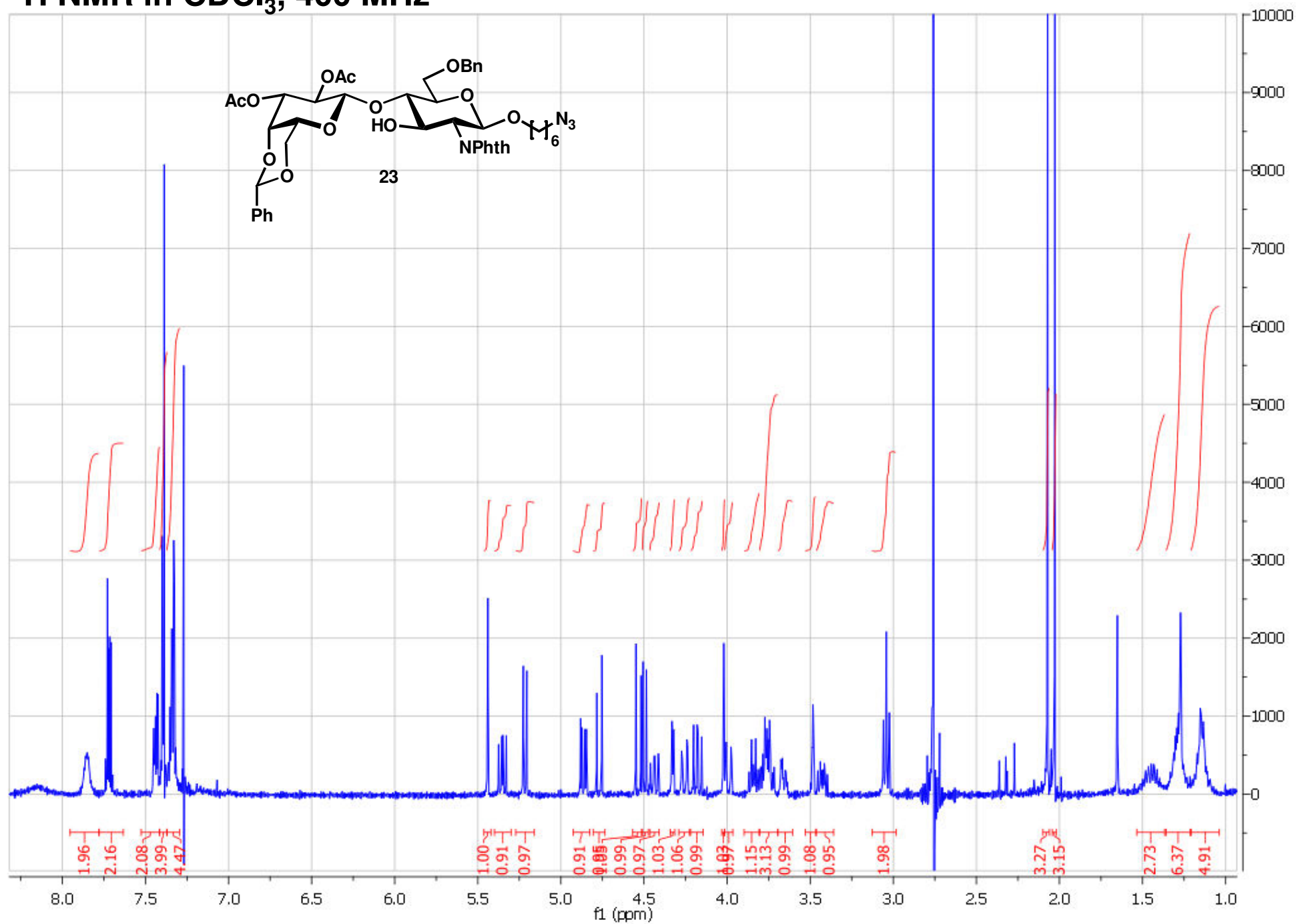
^1H NMR in CDCl_3 , 400 MHz



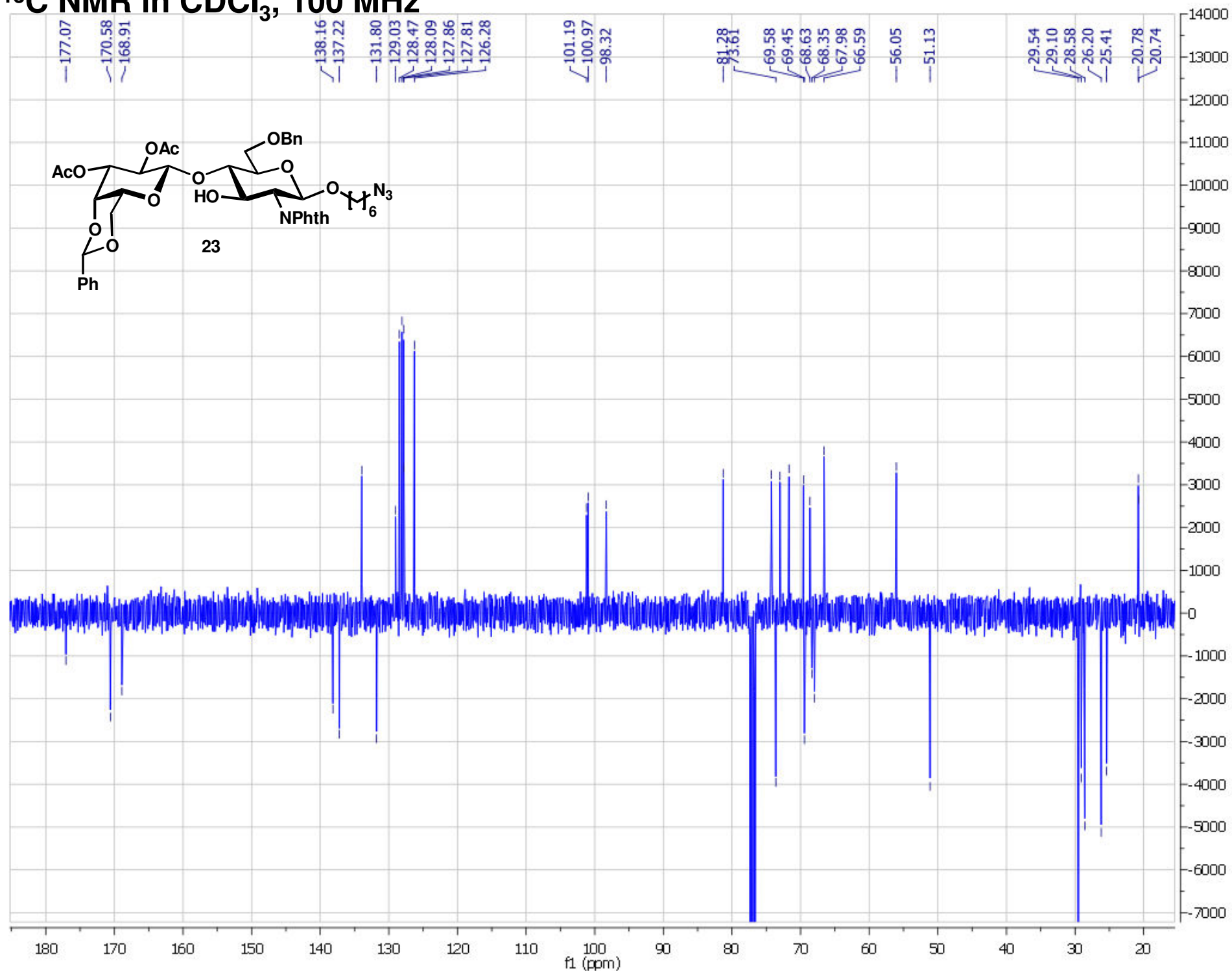
¹³C NMR in CDCl₃, 100 MHz



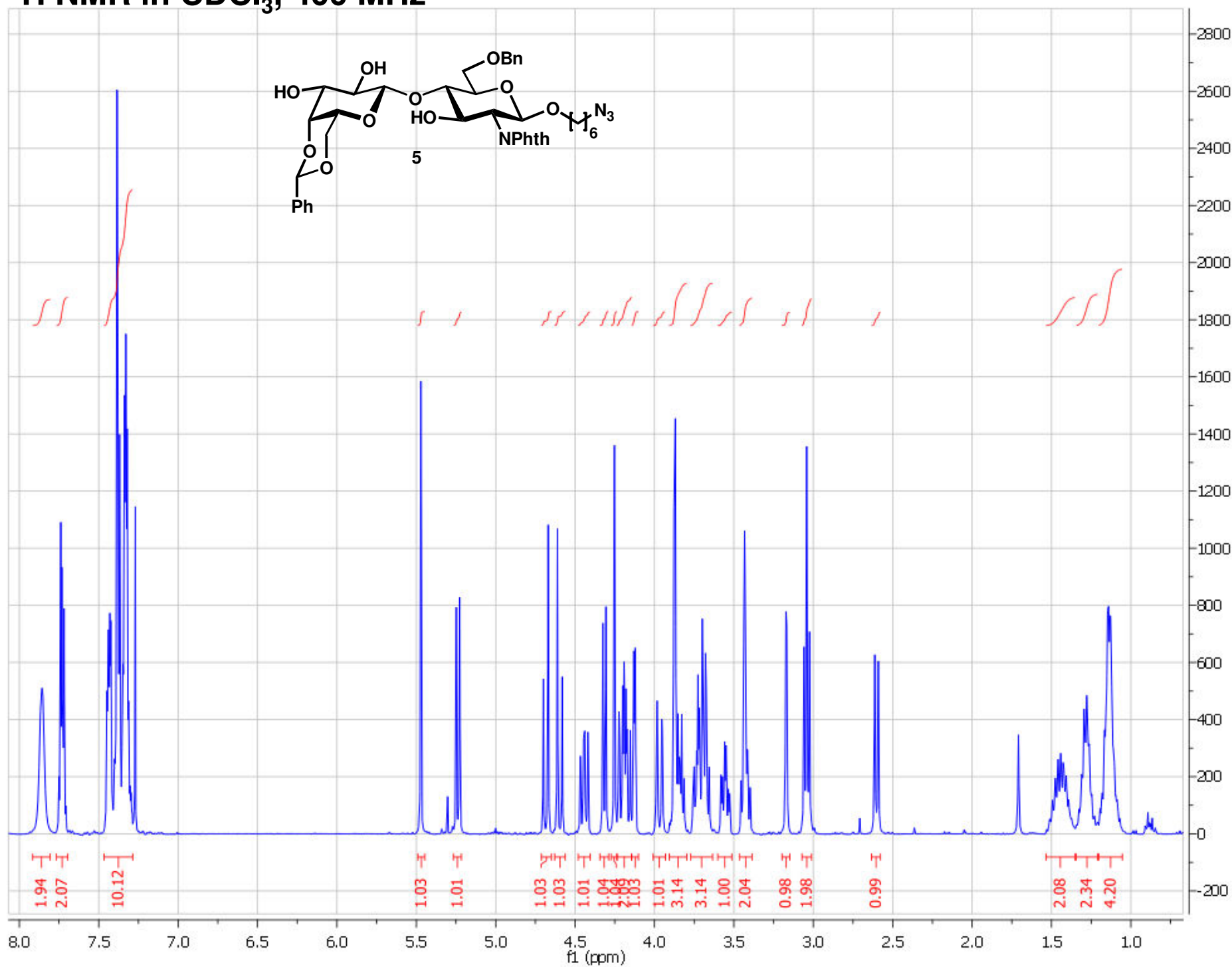
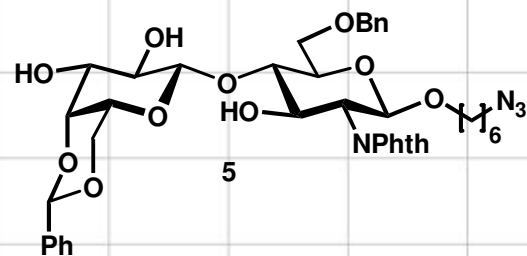
^1H NMR in CDCl_3 , 400 MHz



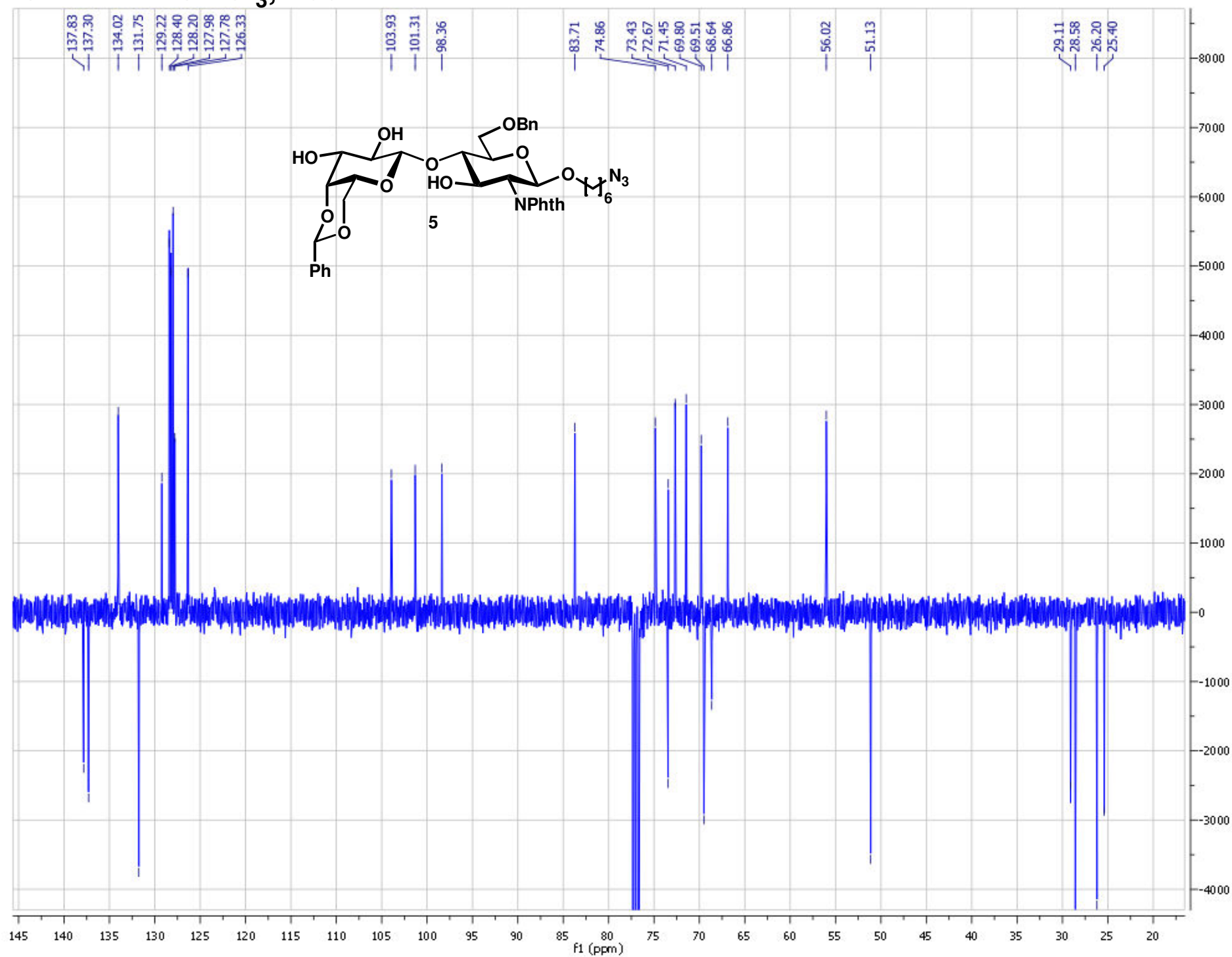
^{13}C NMR in CDCl_3 , 100 MHz



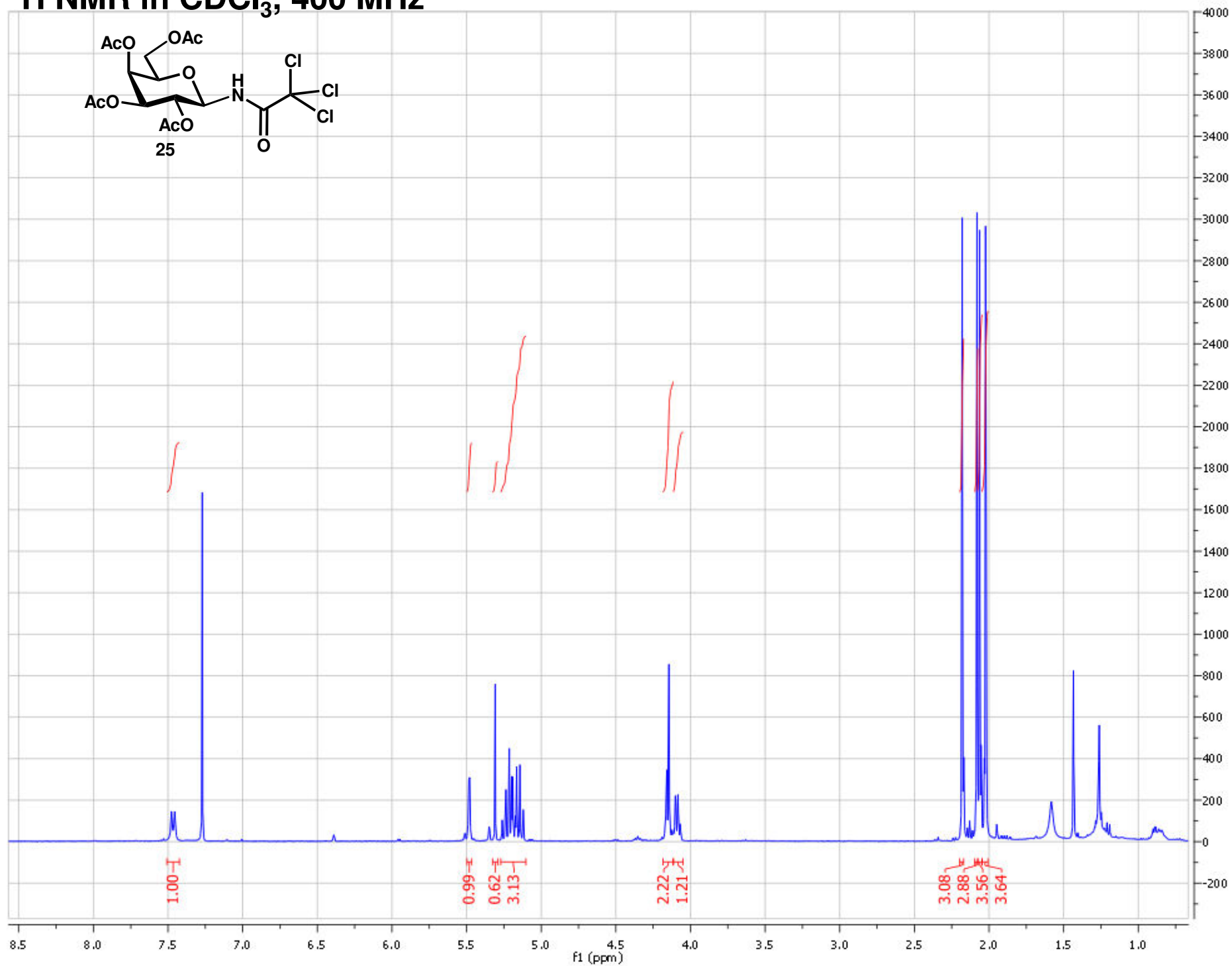
^1H NMR in CDCl_3 , 400 MHz



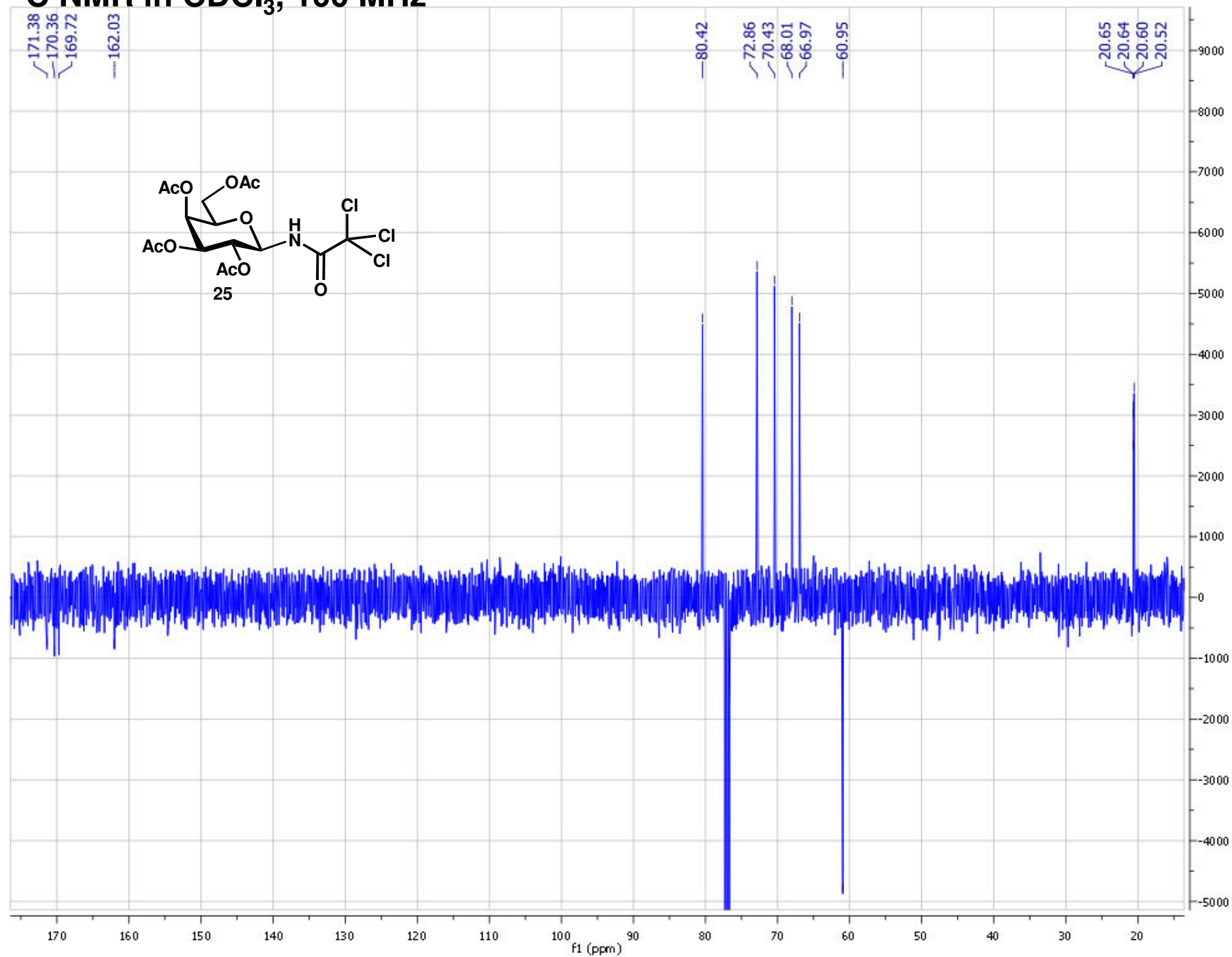
^{13}C NMR in CDCl_3 , 100 MHz



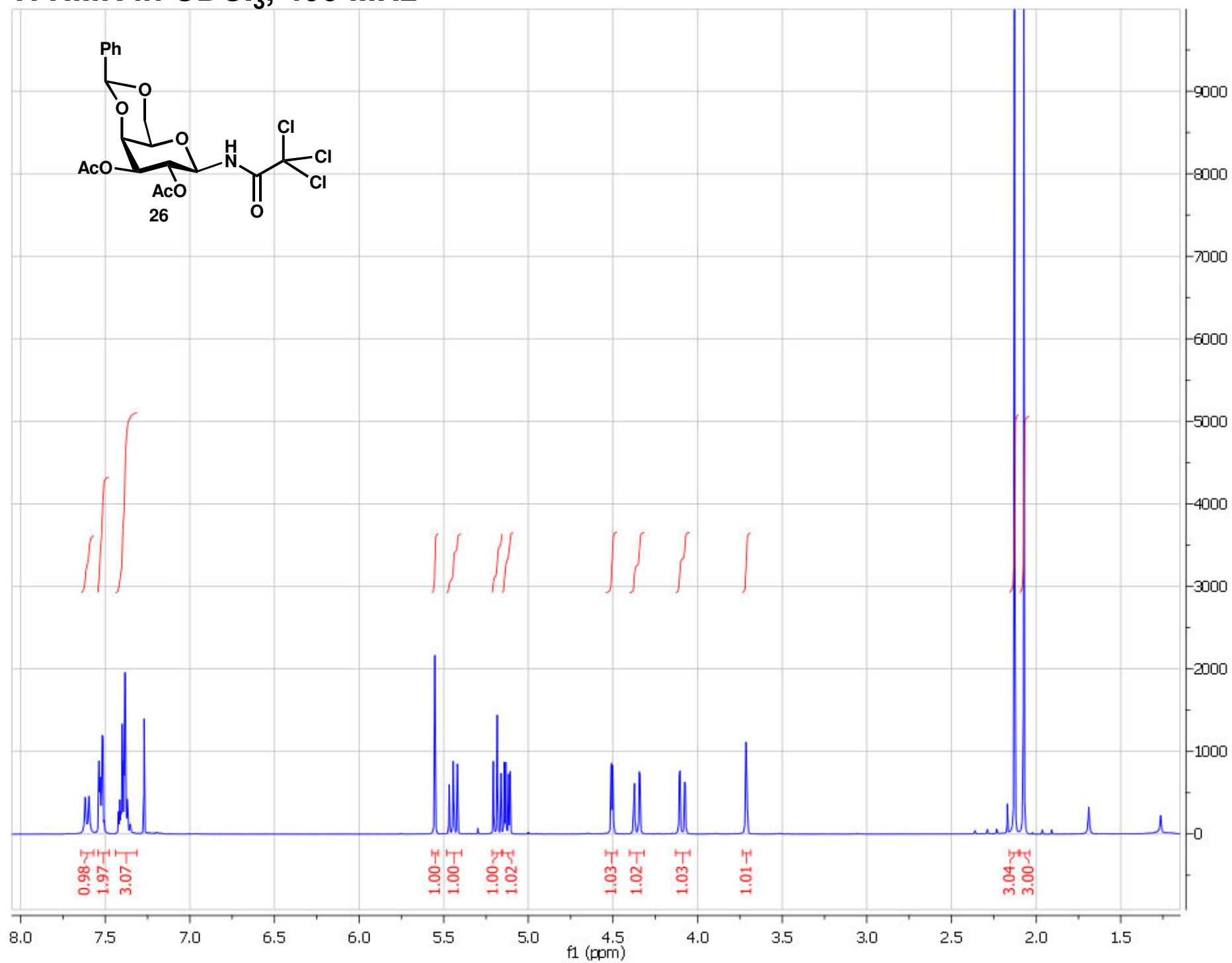
^1H NMR in CDCl_3 , 400 MHz



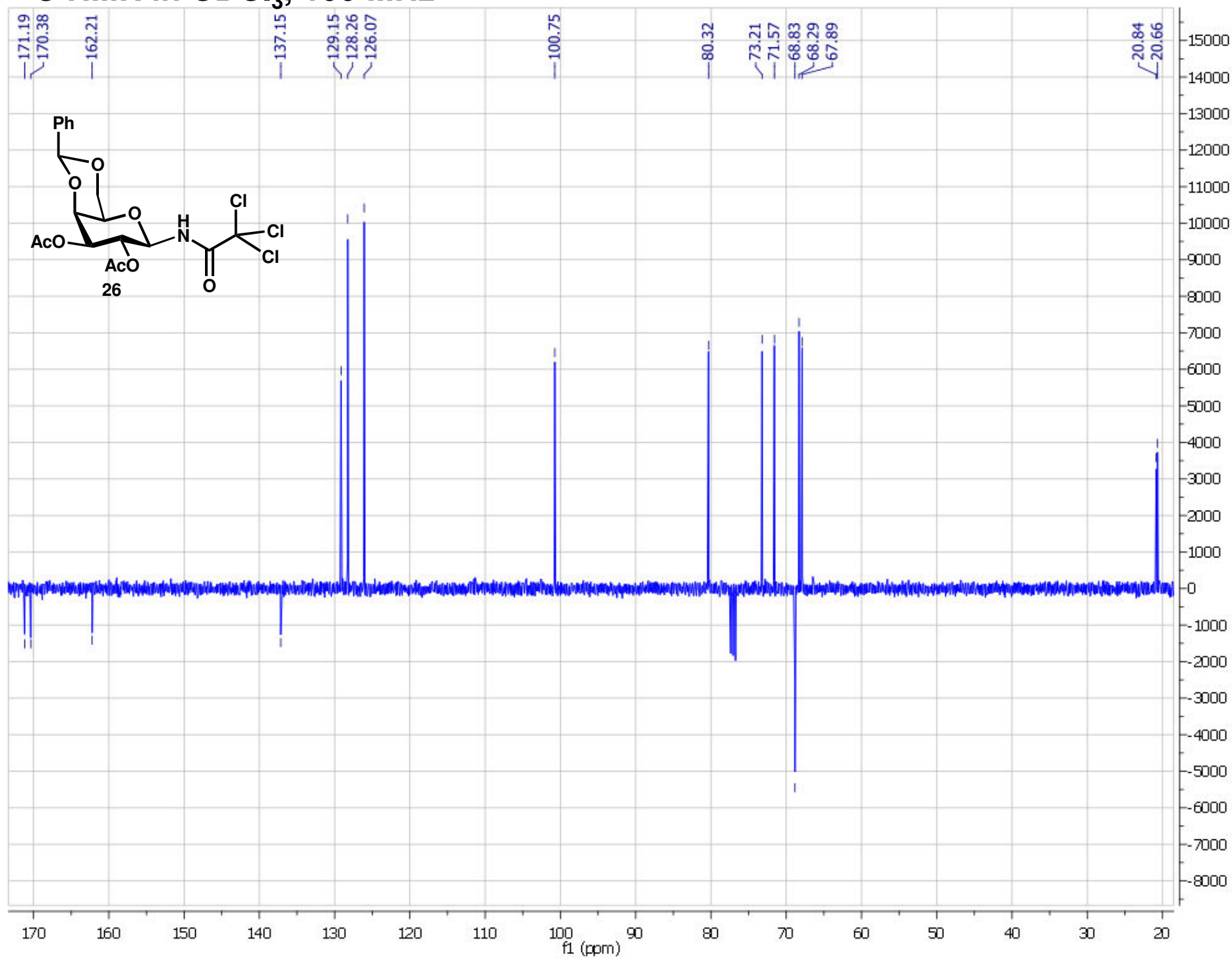
^{13}C NMR in CDCl_3 , 100 MHz



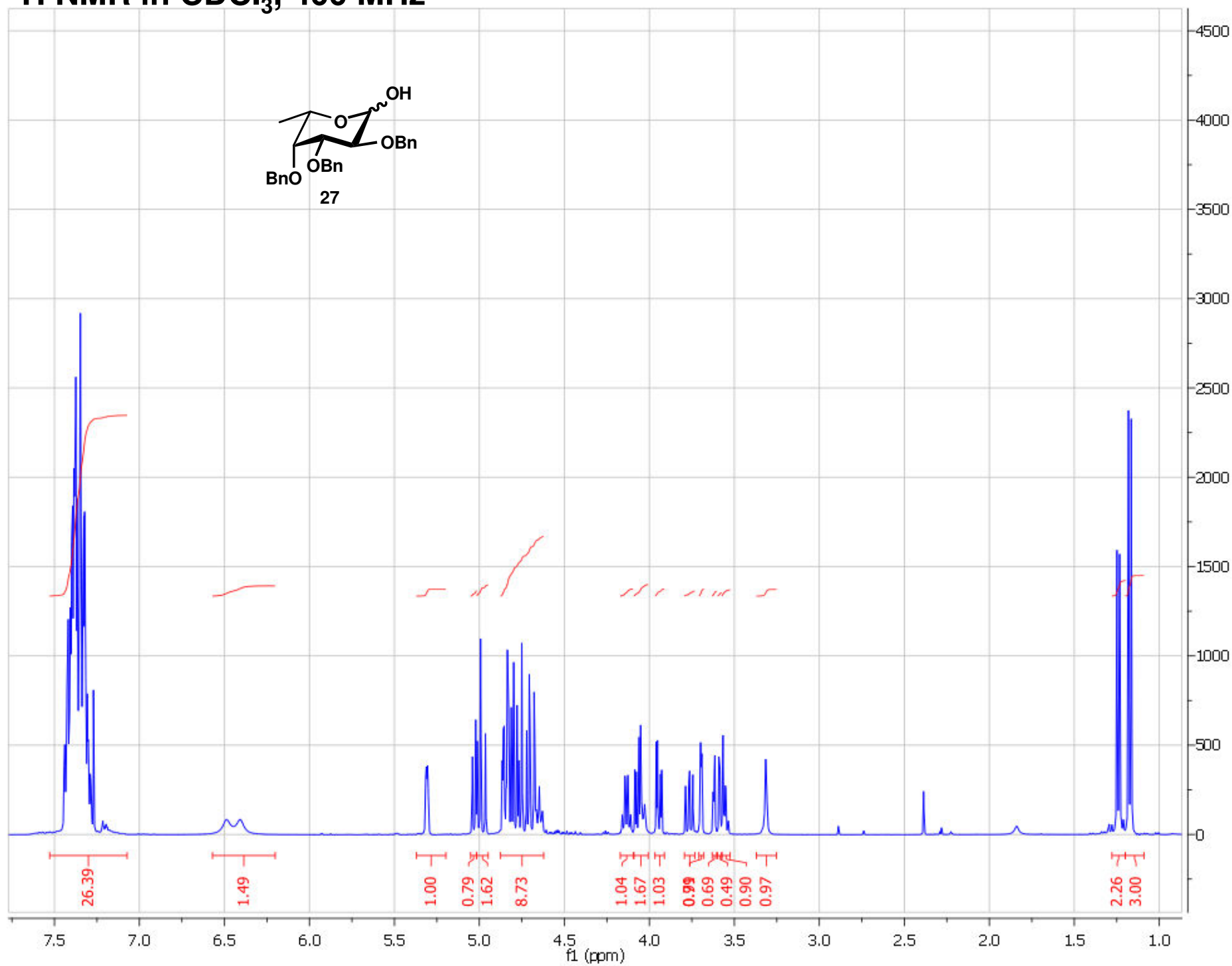
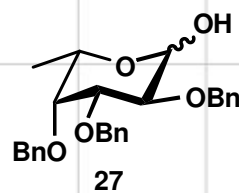
^1H NMR in CDCl_3 , 400 MHz



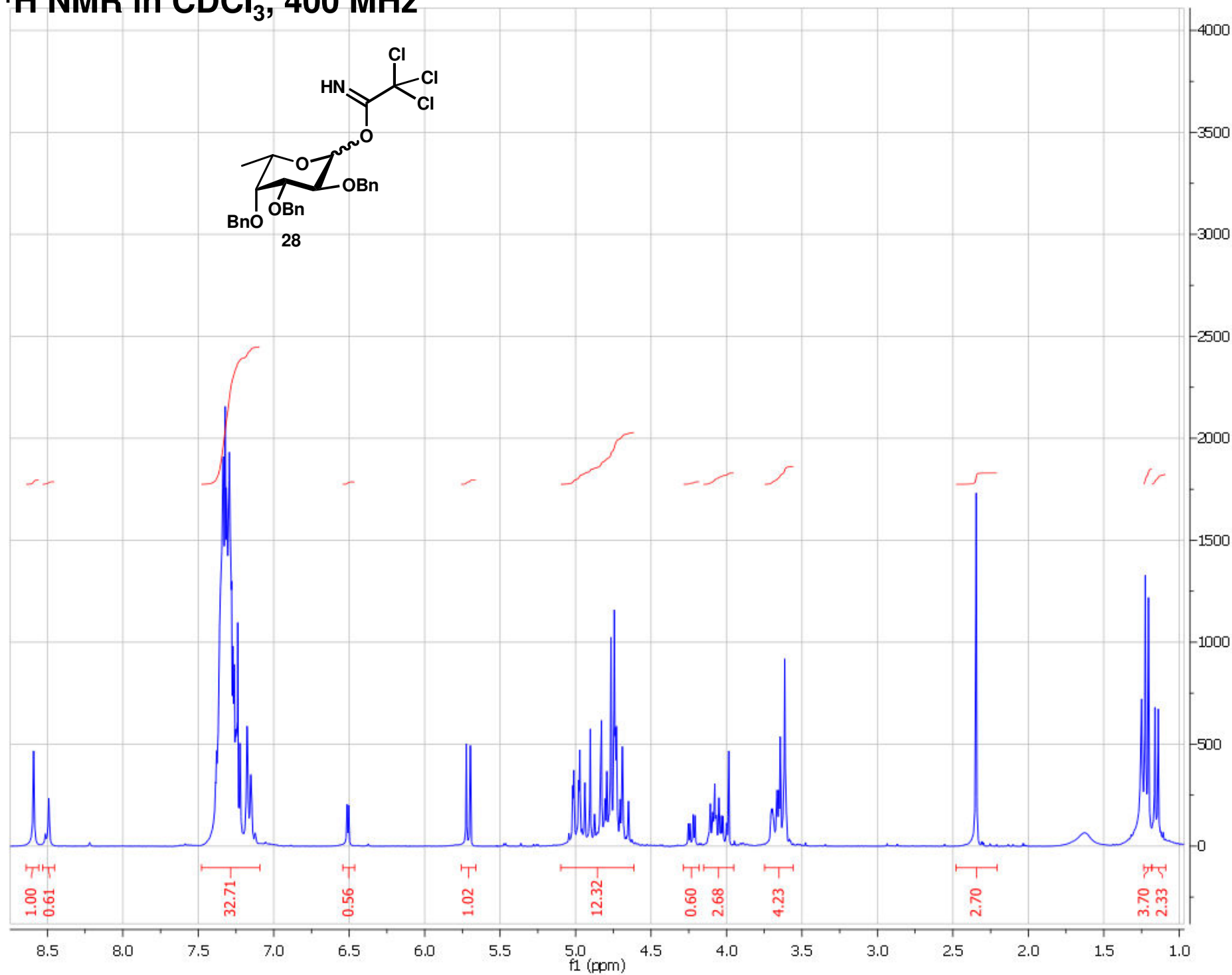
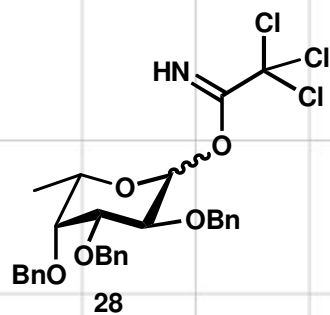
^{13}C NMR in CDCl_3 , 100 MHz



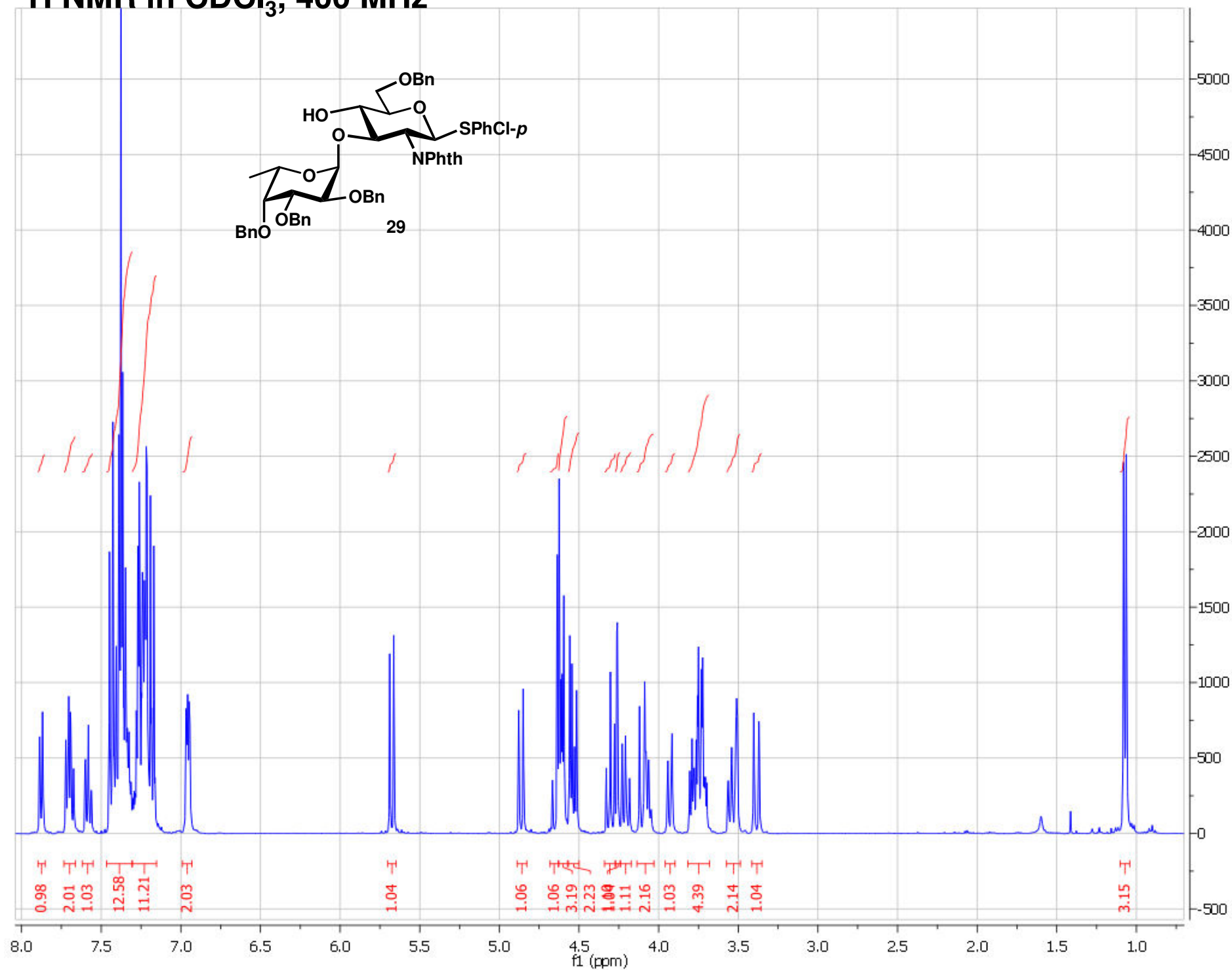
^1H NMR in CDCl_3 , 400 MHz



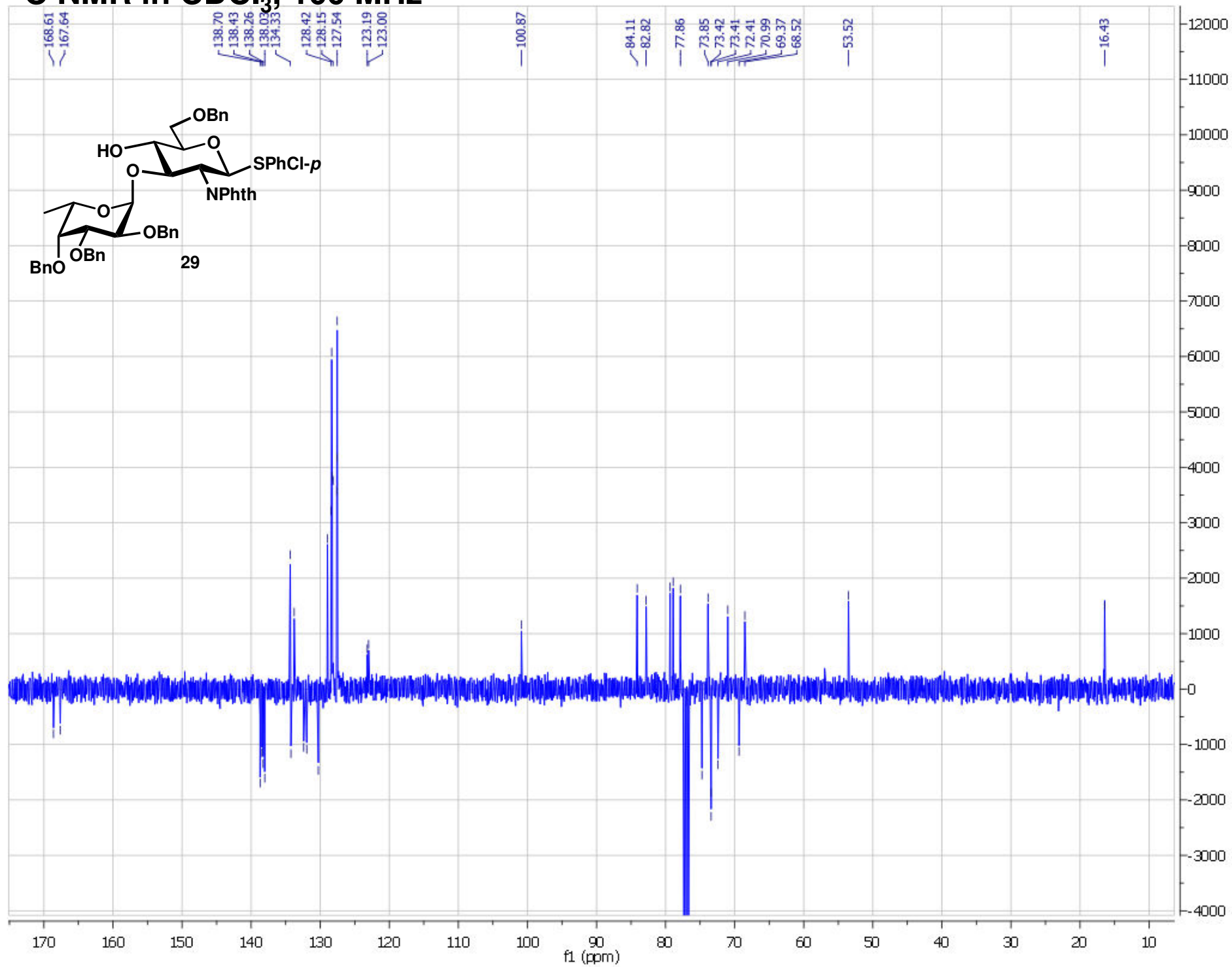
^1H NMR in CDCl_3 , 400 MHz



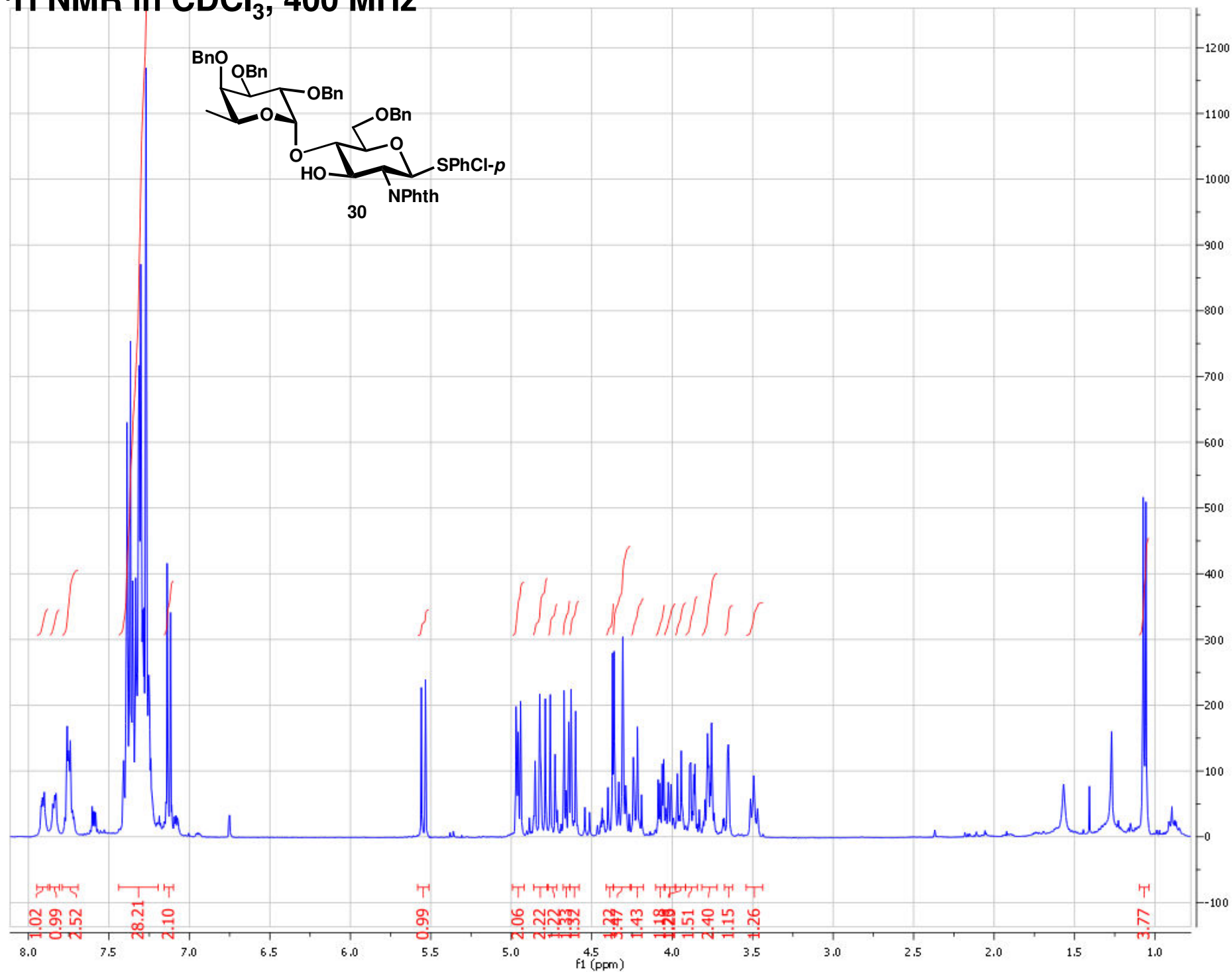
^1H NMR in CDCl_3 , 400 MHz



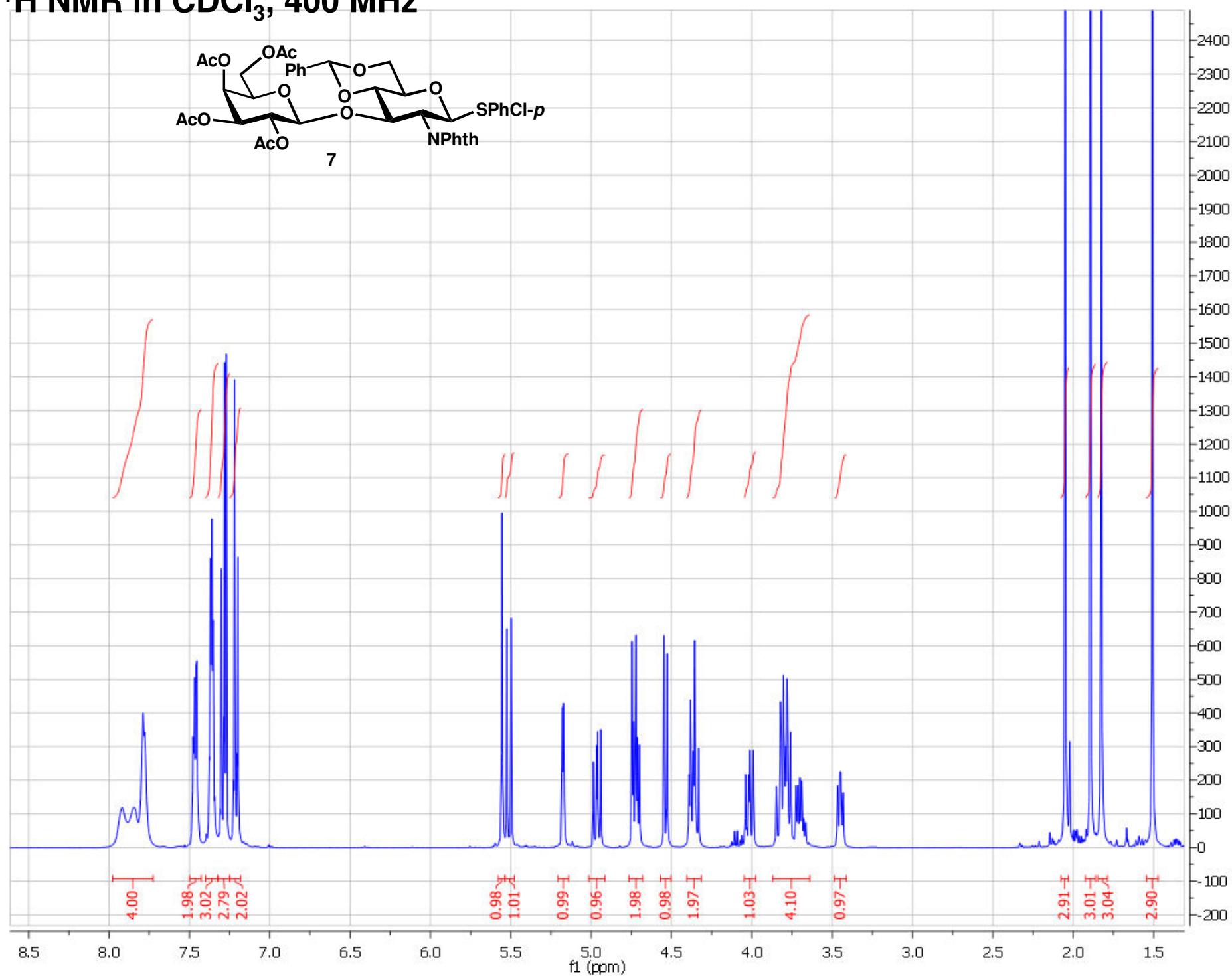
^{13}C NMR in CDCl_3 , 100 MHz



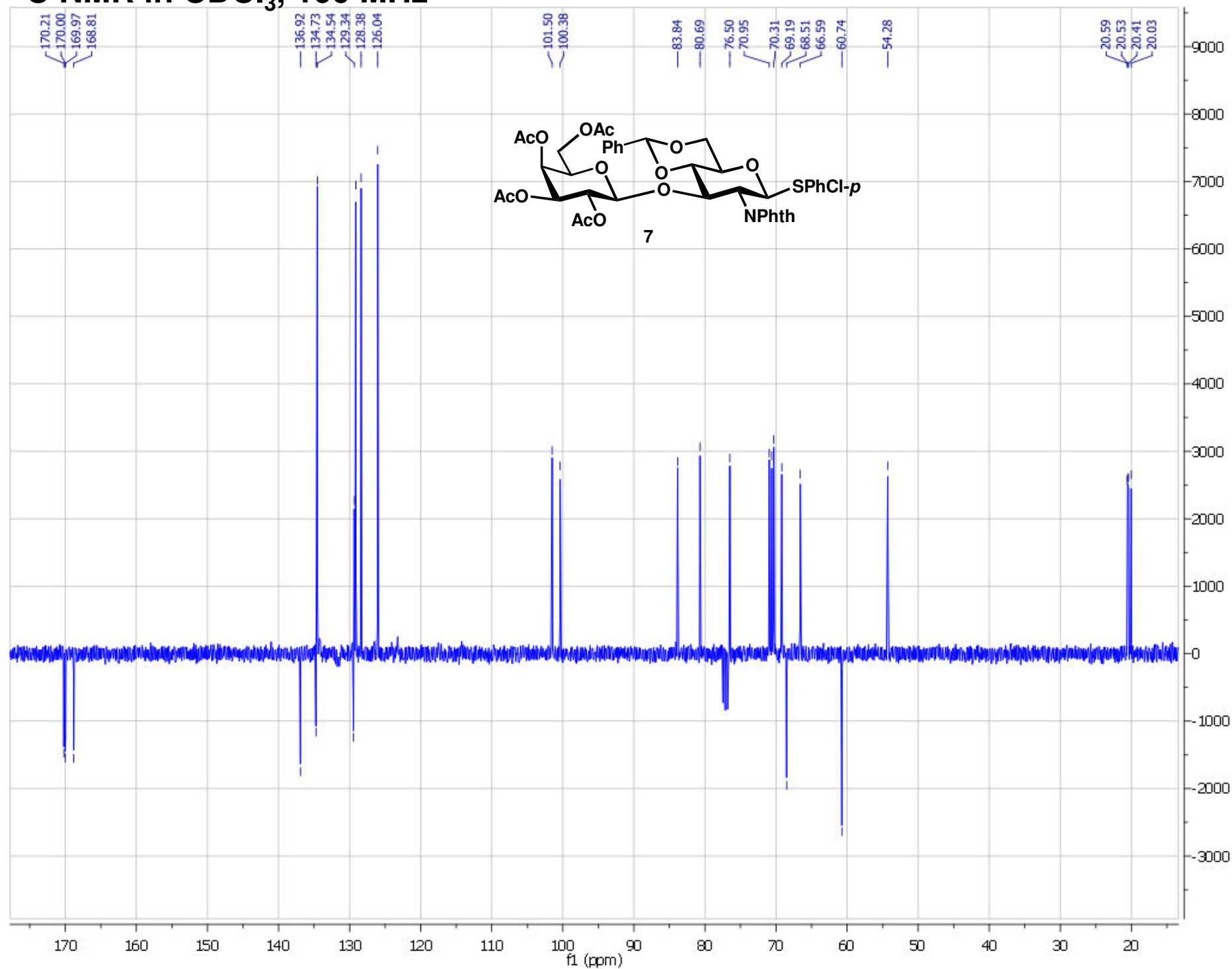
^1H NMR in CDCl_3 , 400 MHz



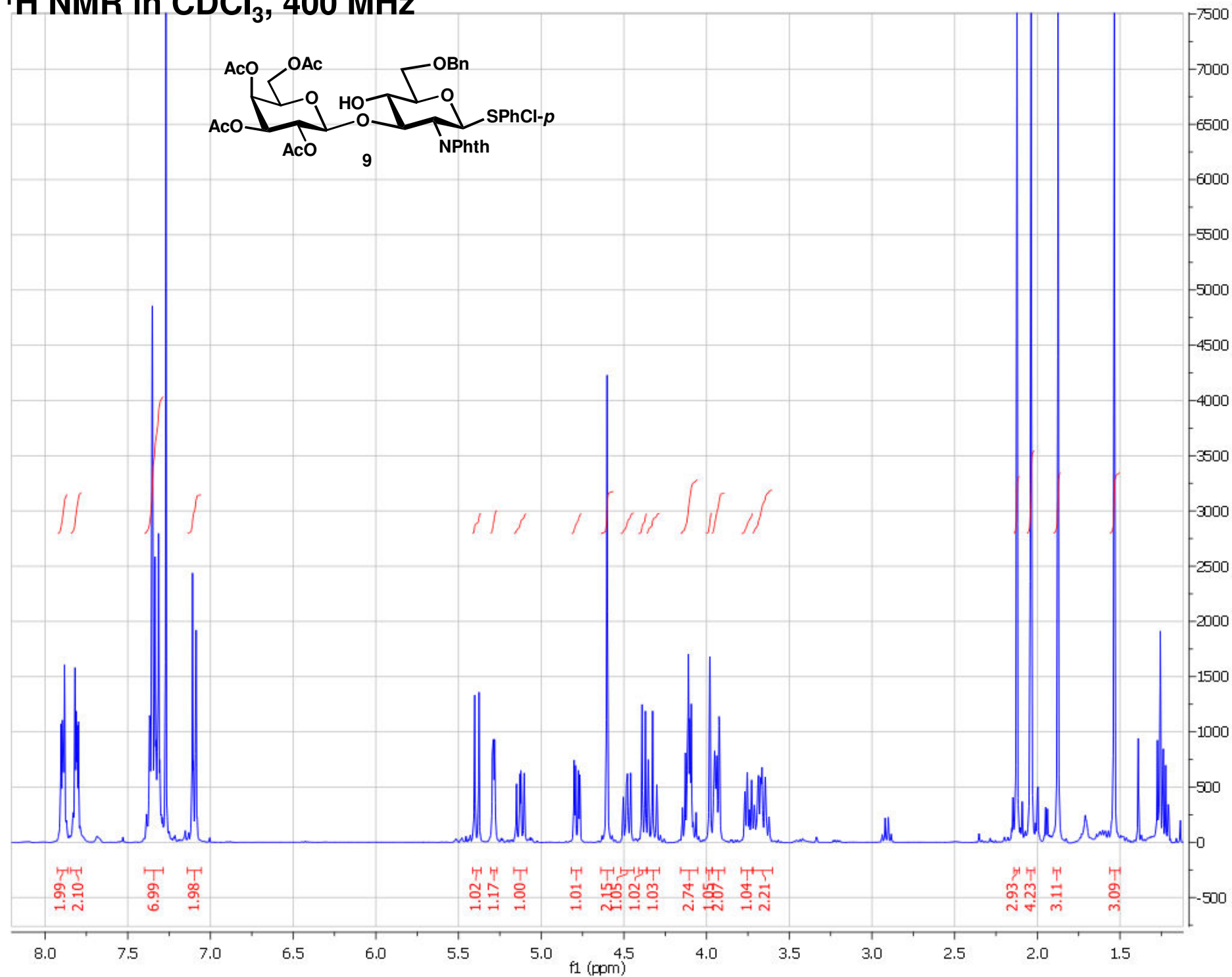
^1H NMR in CDCl_3 , 400 MHz



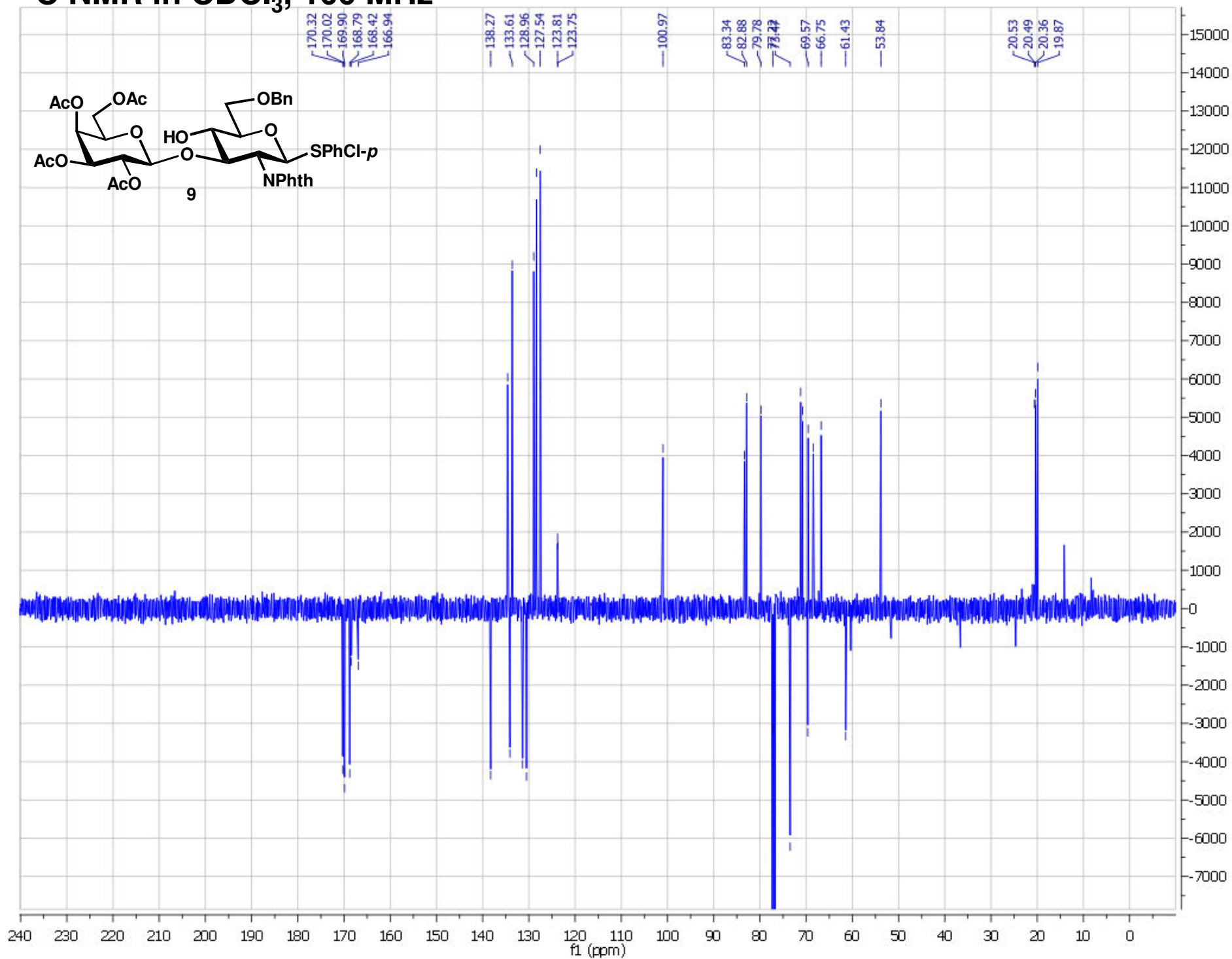
^{13}C NMR in CDCl_3 , 100 MHz



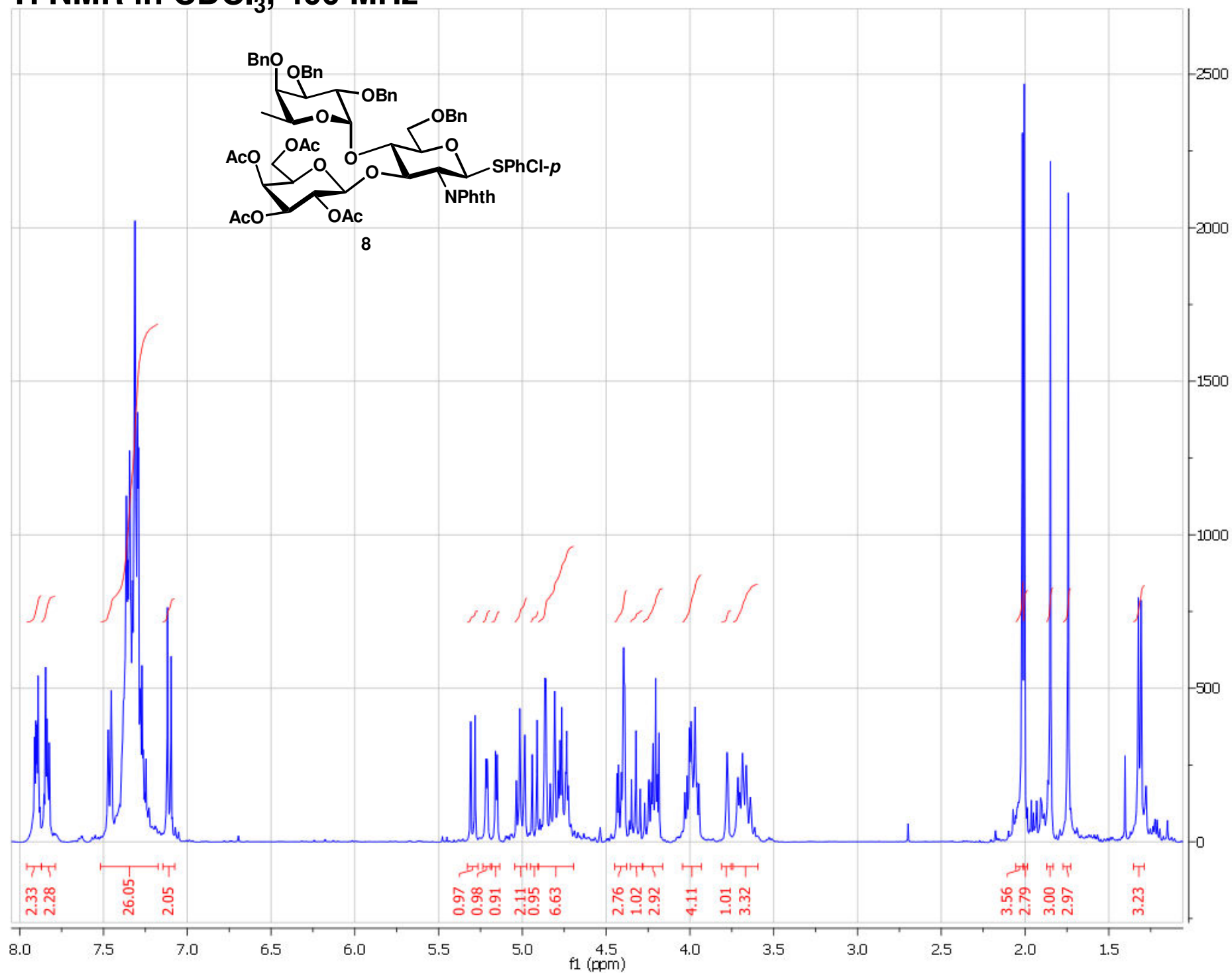
^1H NMR in CDCl_3 , 400 MHz



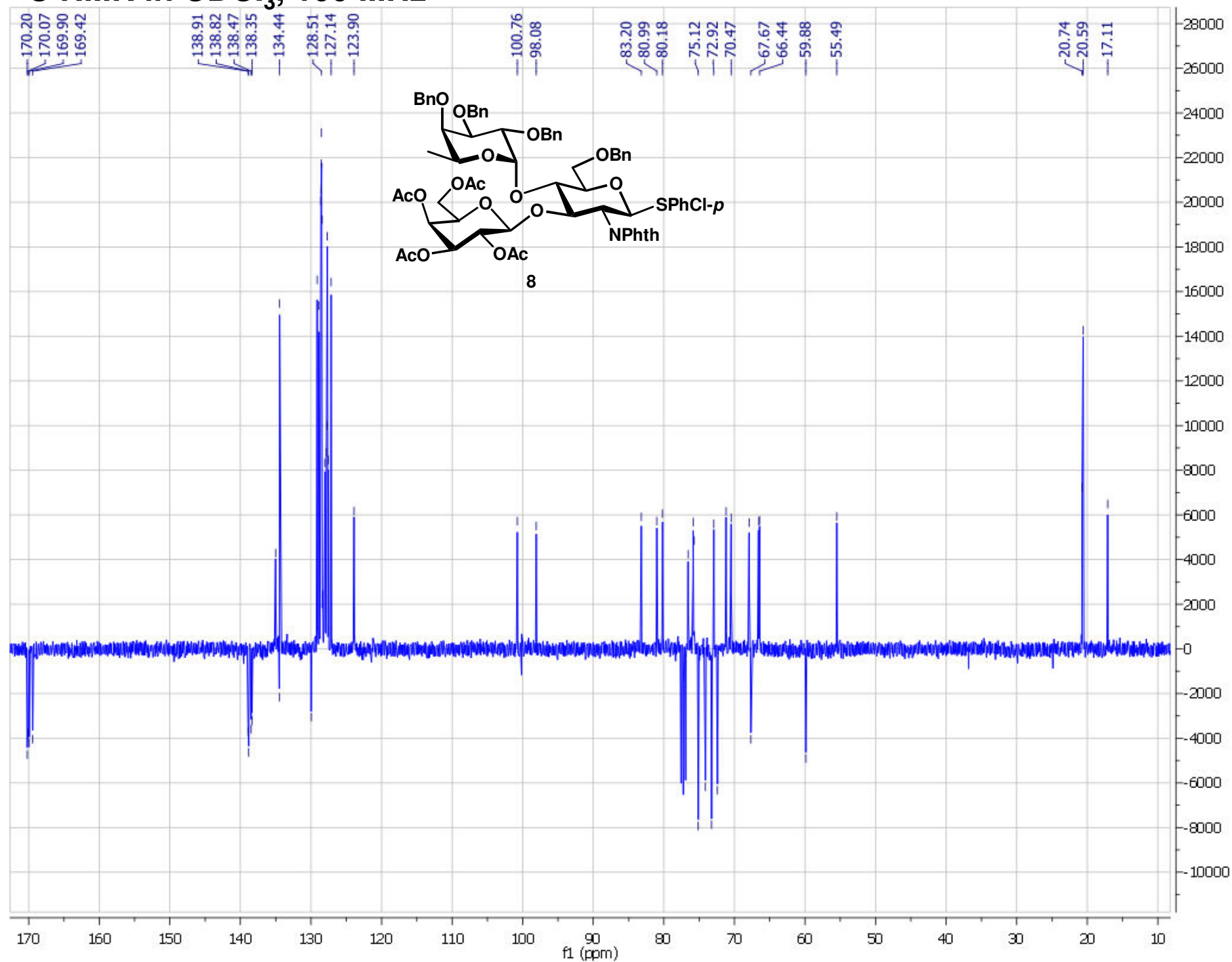
^{13}C NMR in CDCl_3 , 100 MHz



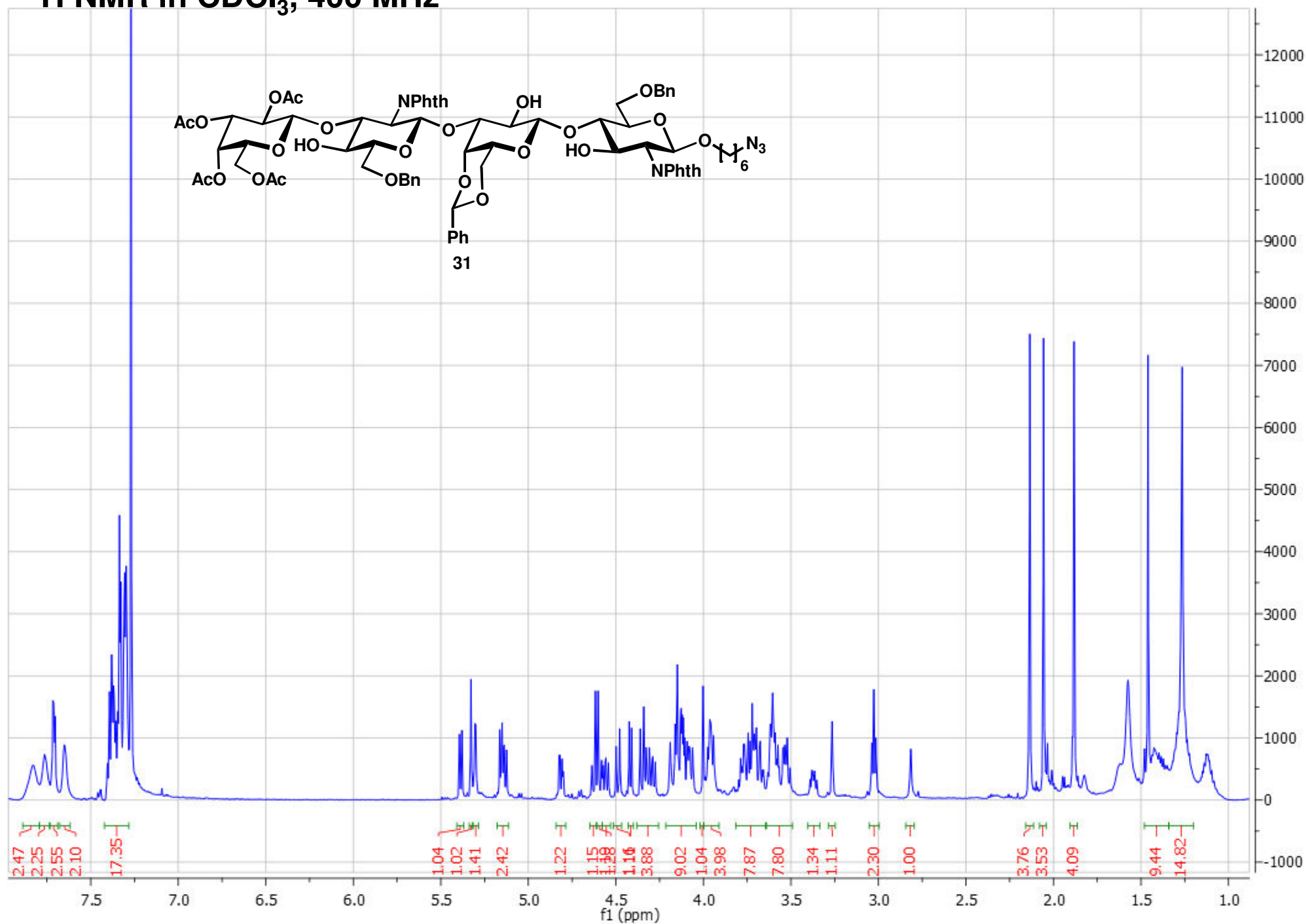
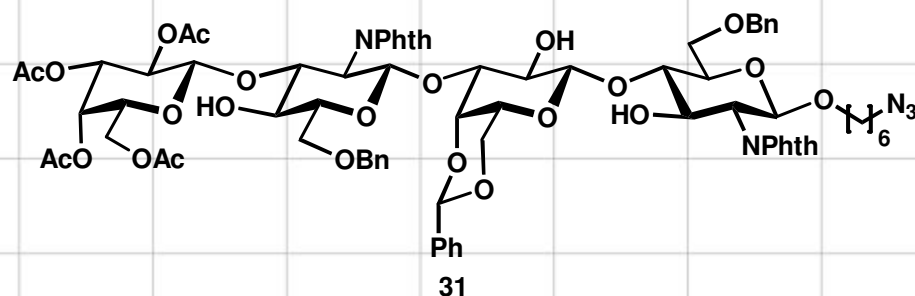
^1H NMR in CDCl_3 , 400 MHz



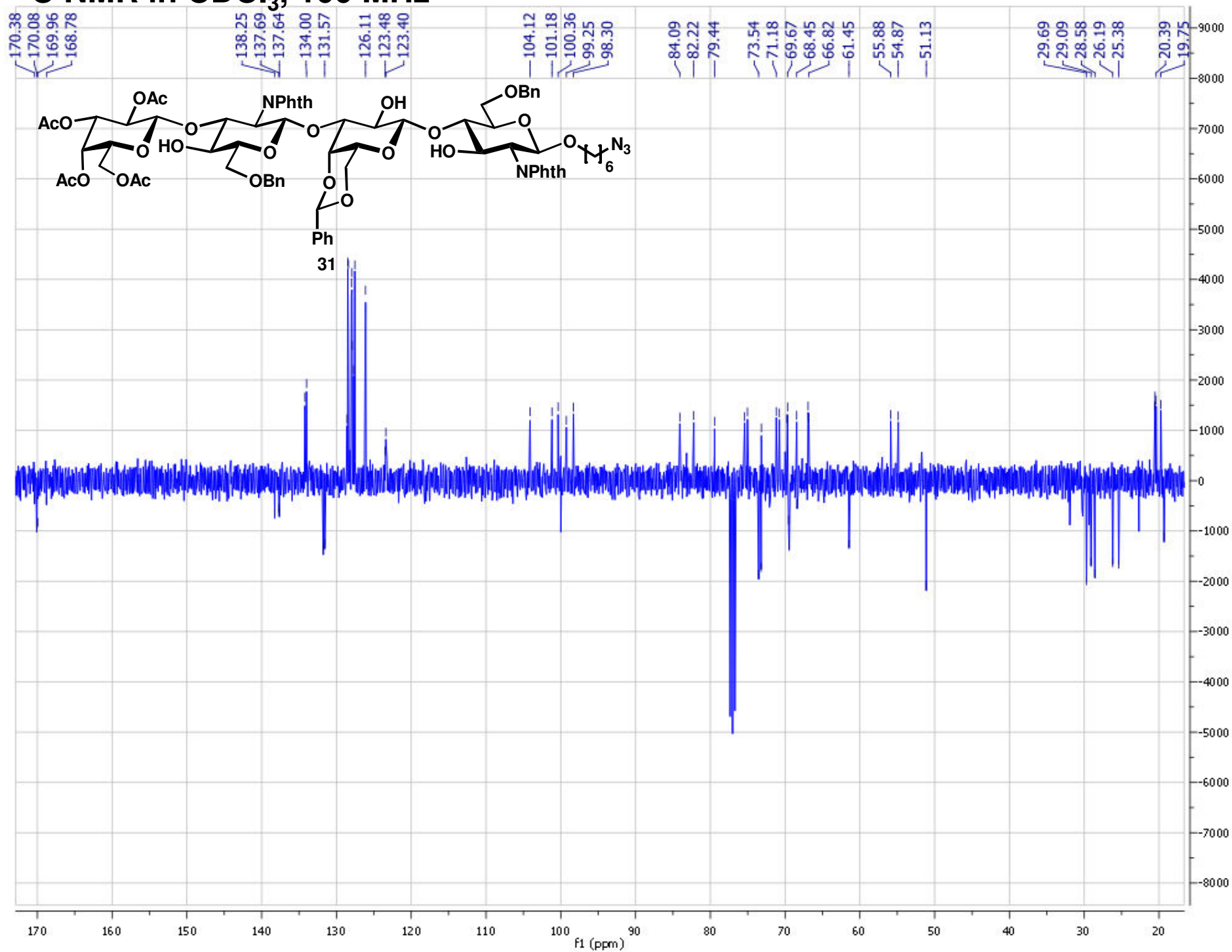
^{13}C NMR in CDCl_3 , 100 MHz



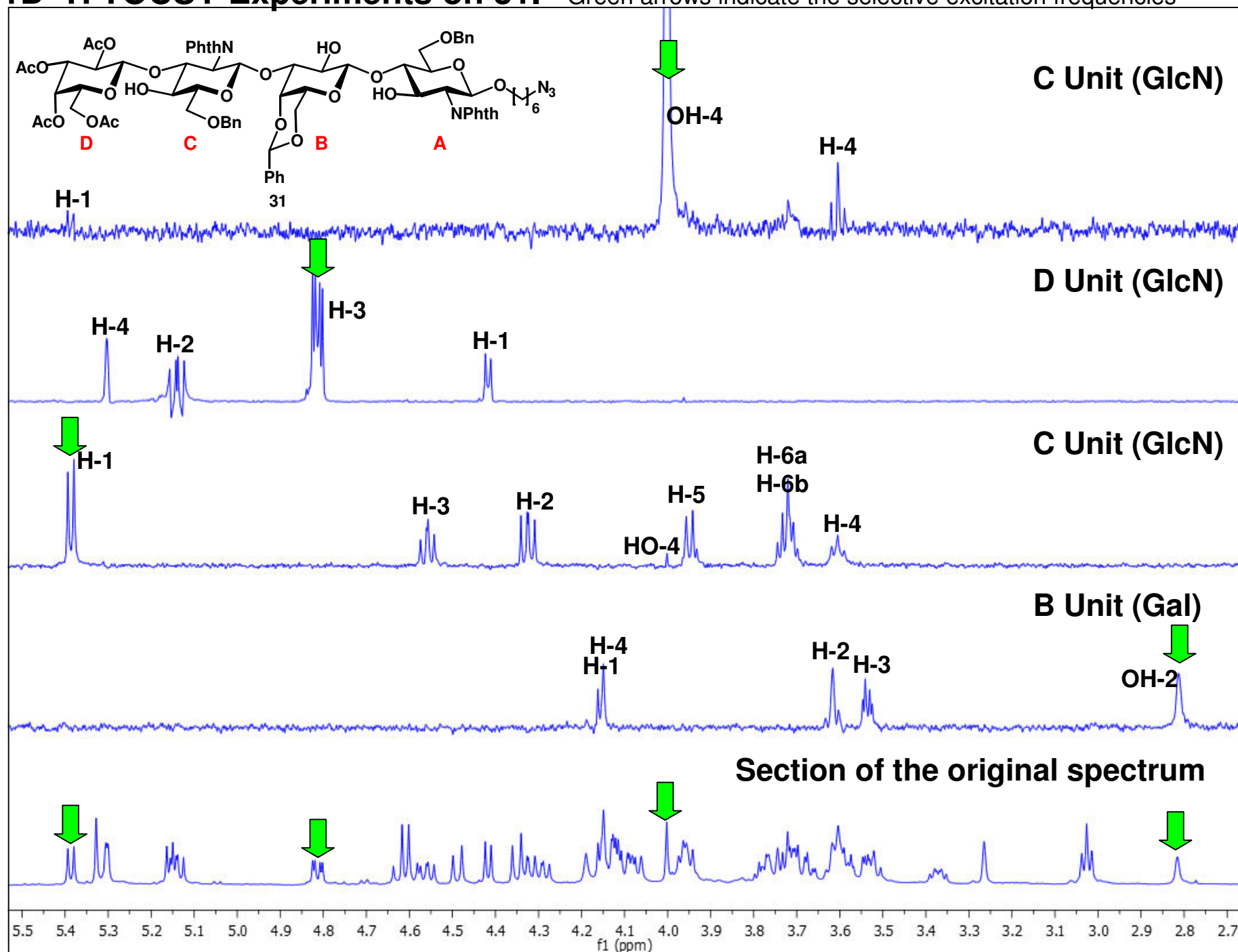
^1H NMR in CDCl_3 , 400 MHz

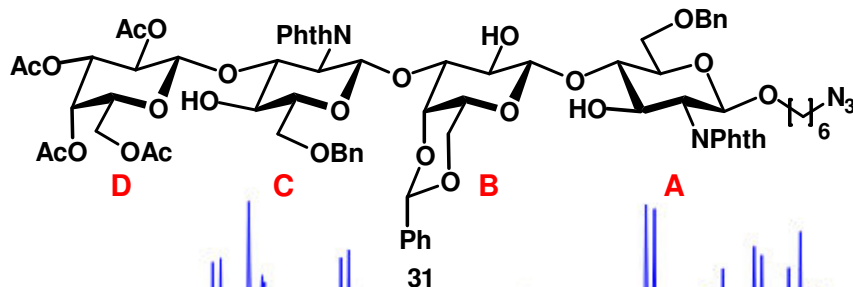


¹³C NMR in CDCl₃, 100 MHz

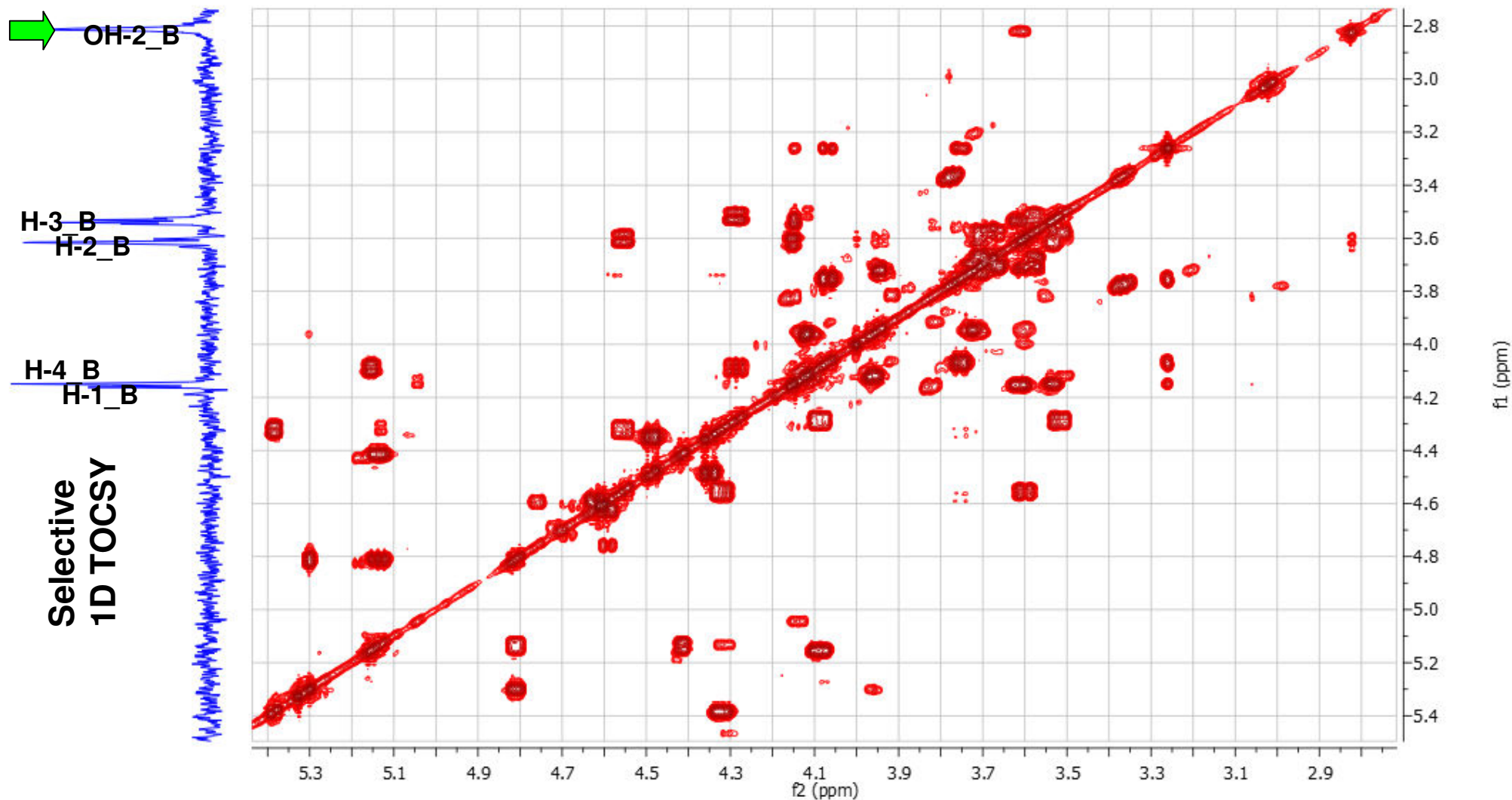


1D ^1H TOCSY Experiments on 31. – Green arrows indicate the selective excitation frequencies

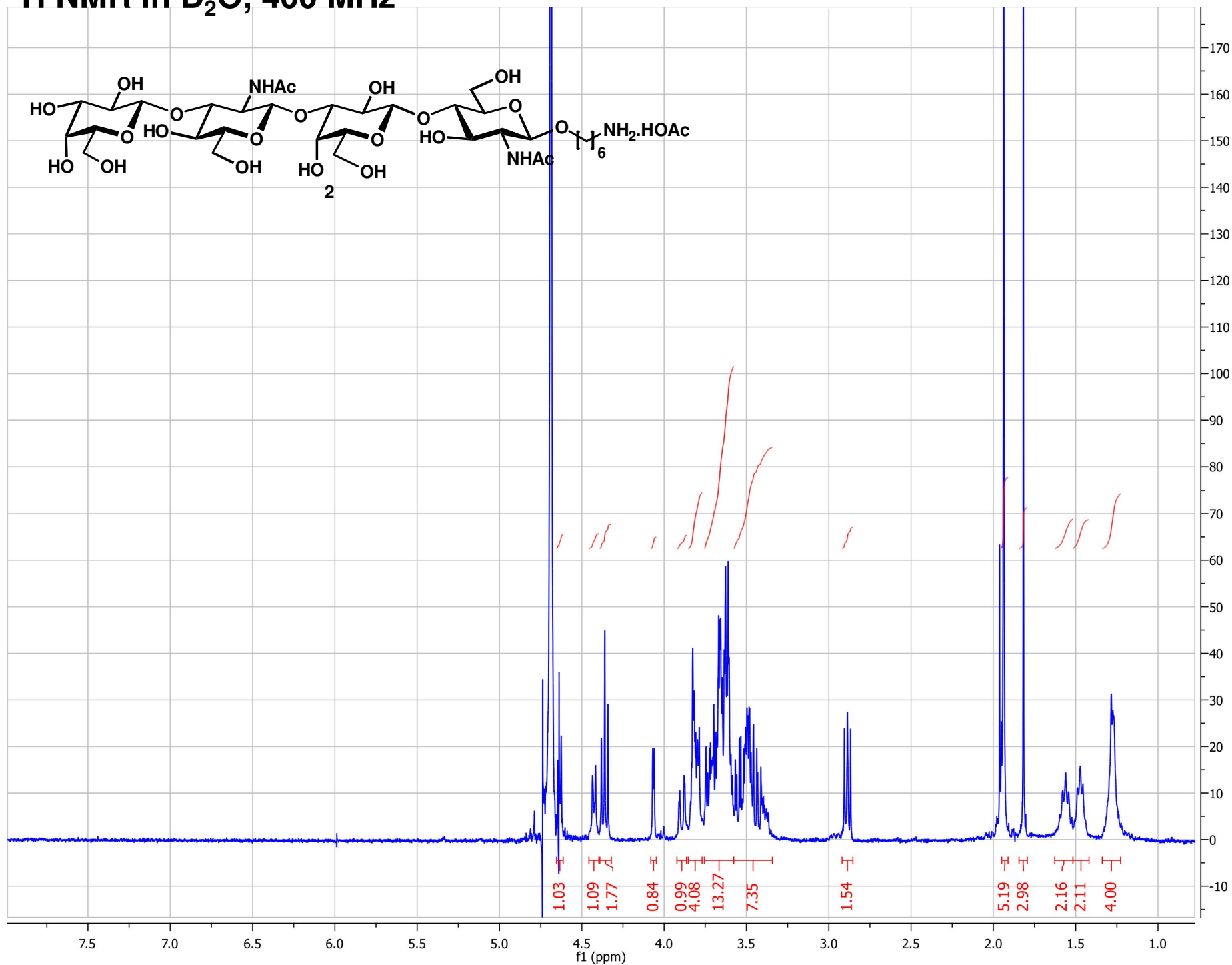




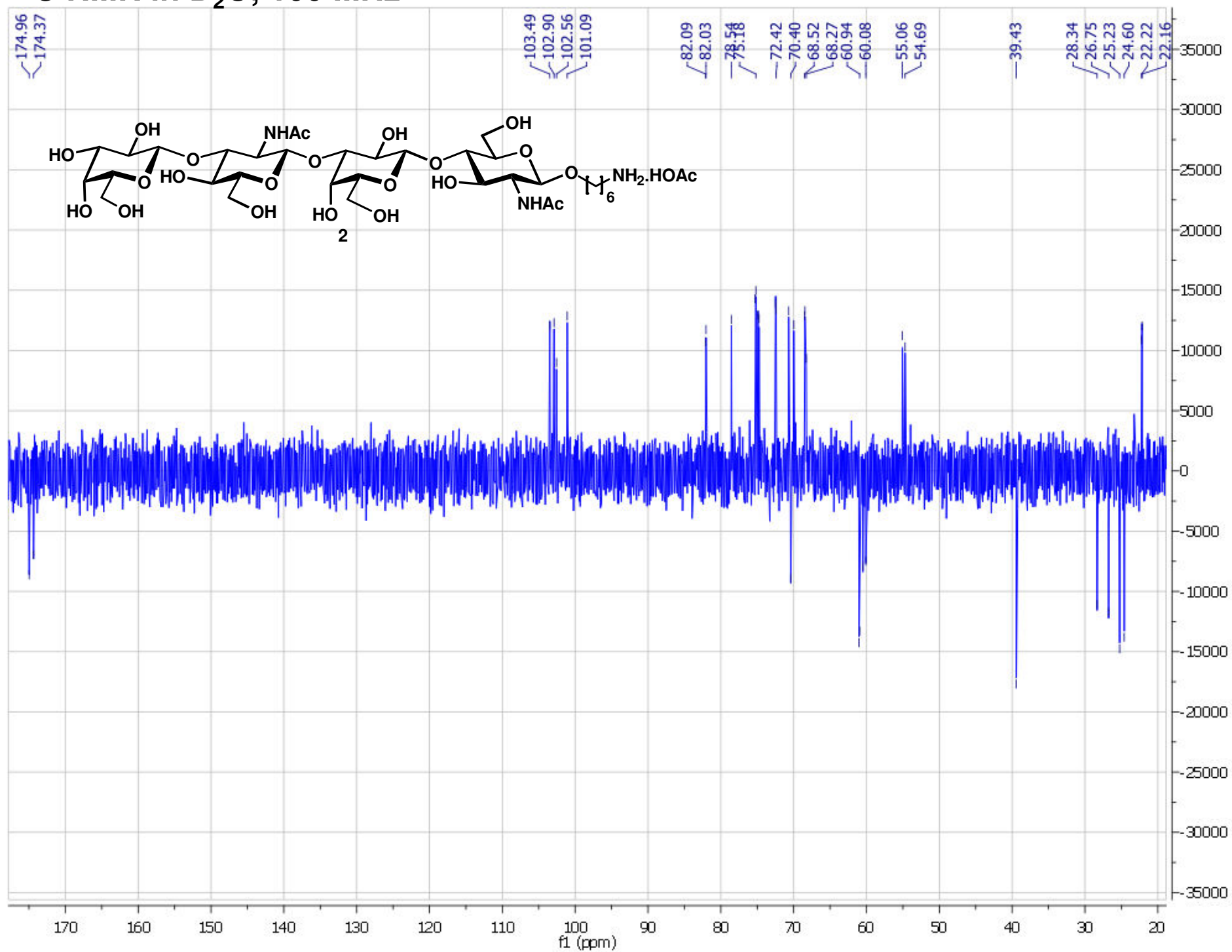
^1H - ^1H COSY NMR (CDCl₃, 600 MHz)
 combined with a 1D TOCSY trace (OH-2' was excited)



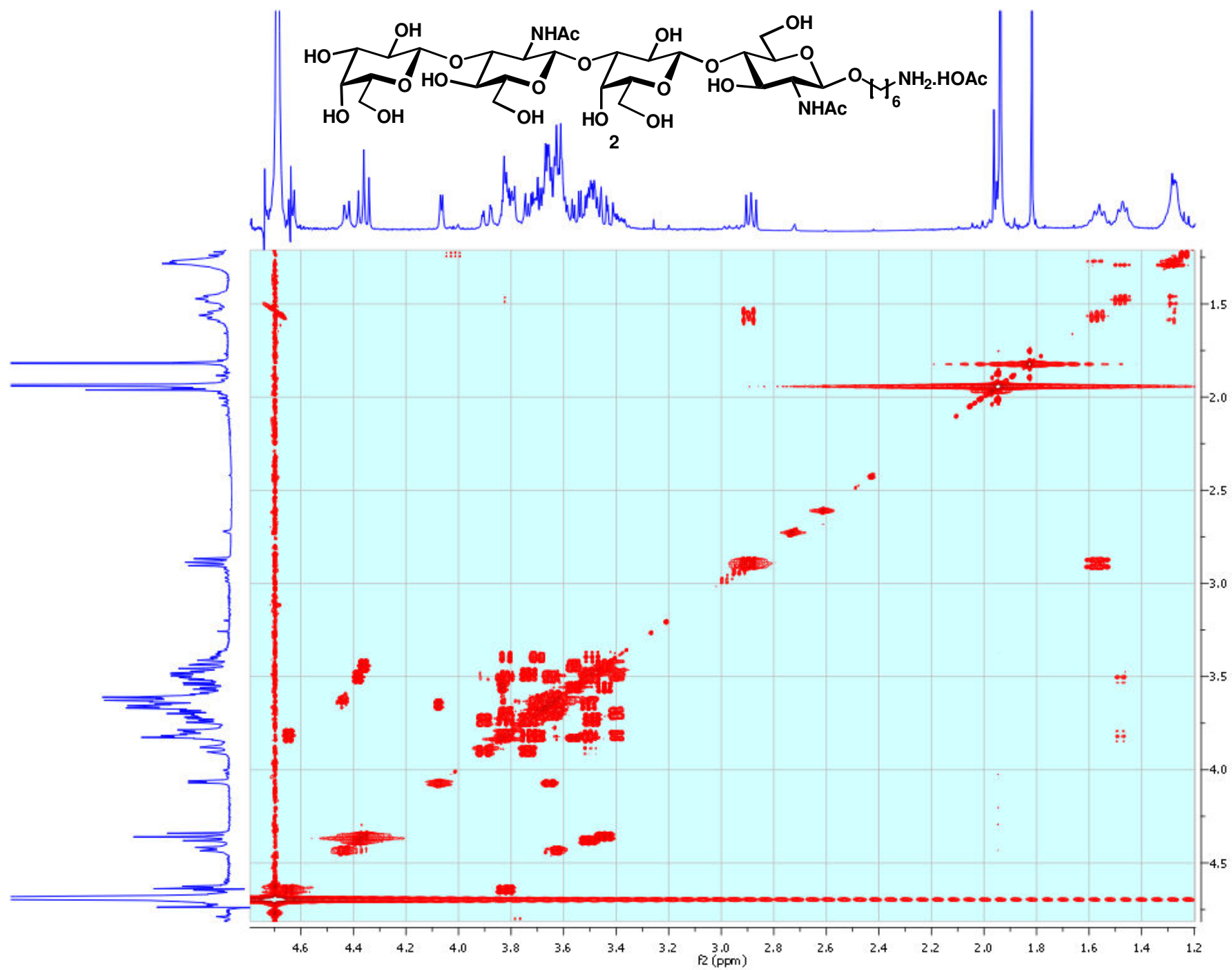
^1H NMR in D_2O , 400 MHz



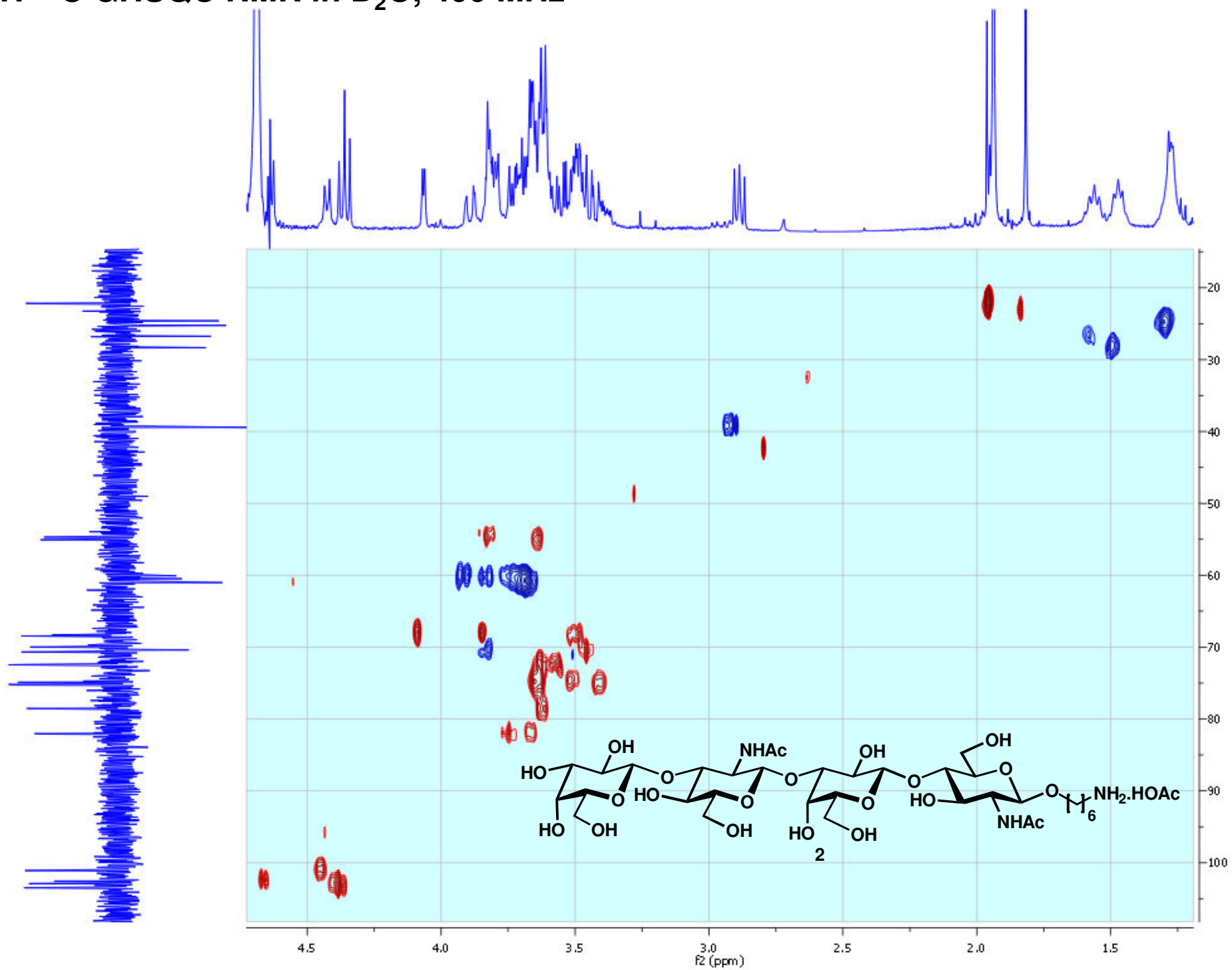
^{13}C NMR in D_2O , 100 MHz



^1H - ^1H 2D GCOSY NMR in D_2O , 400 MHz

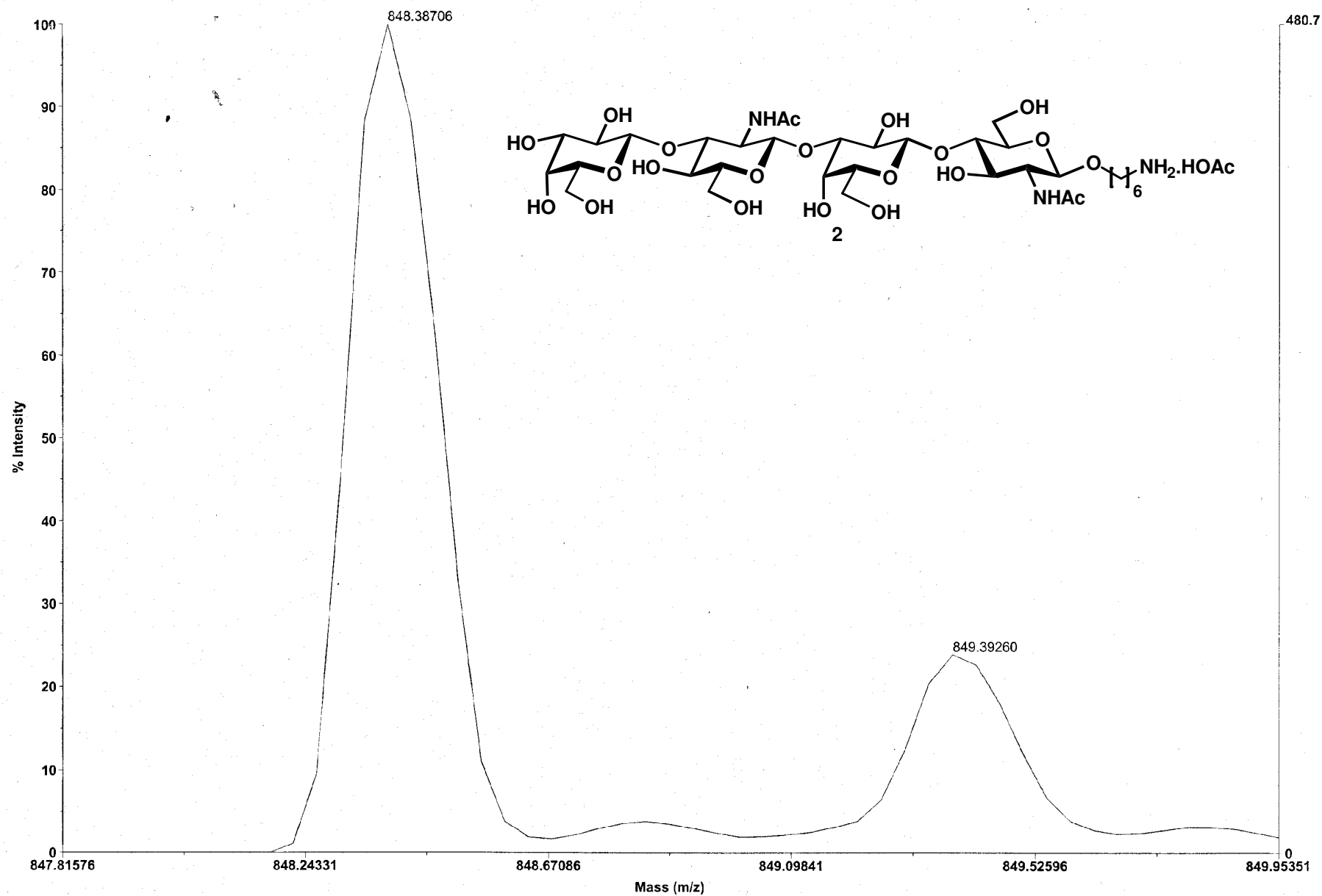


¹H-¹³C GHSQC NMR in D₂O, 400 MHz



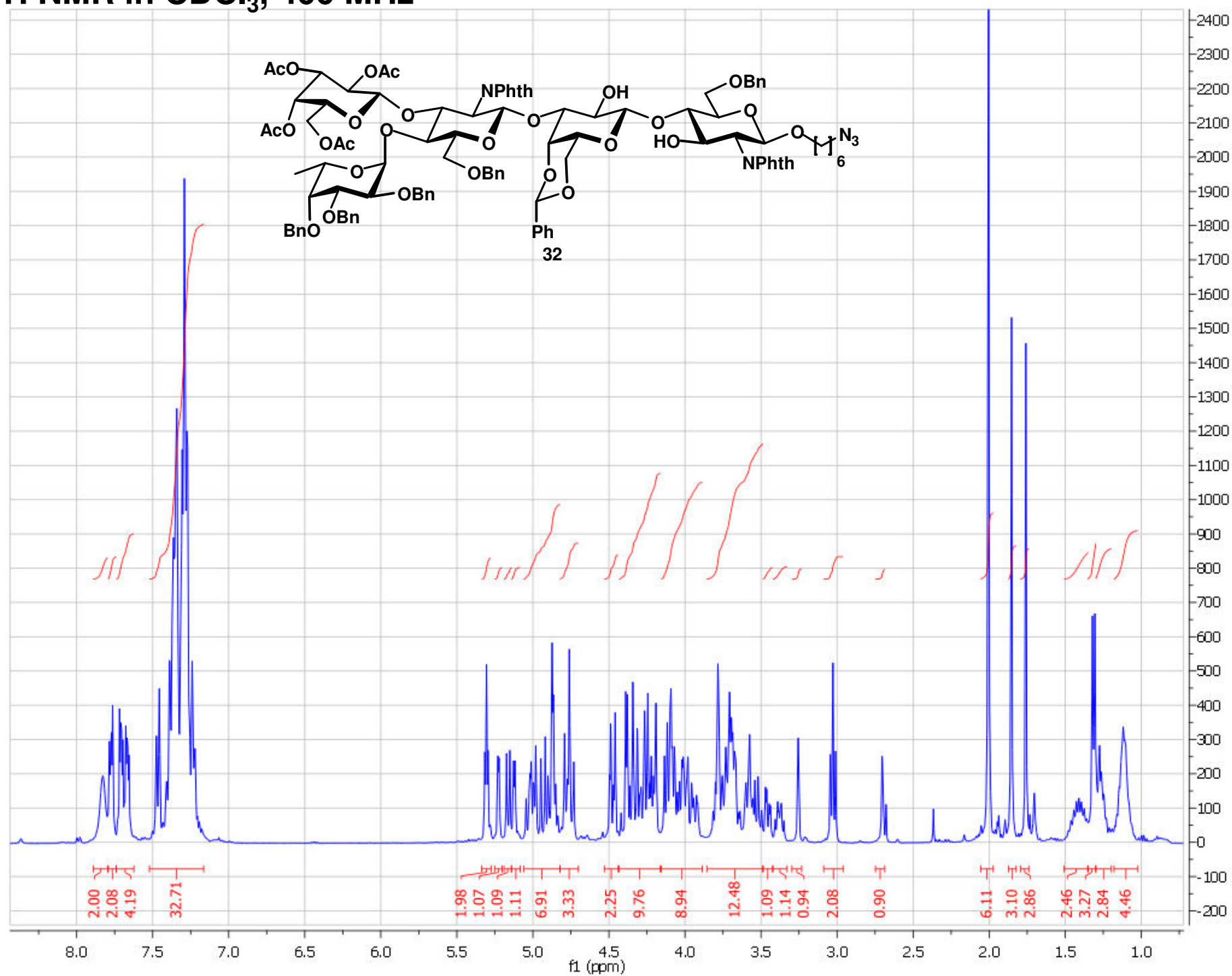
High Resolution ESI Mass Spectra

Mariner Spec /5:9 ASC=>SM5[BP = 848.4, 481]

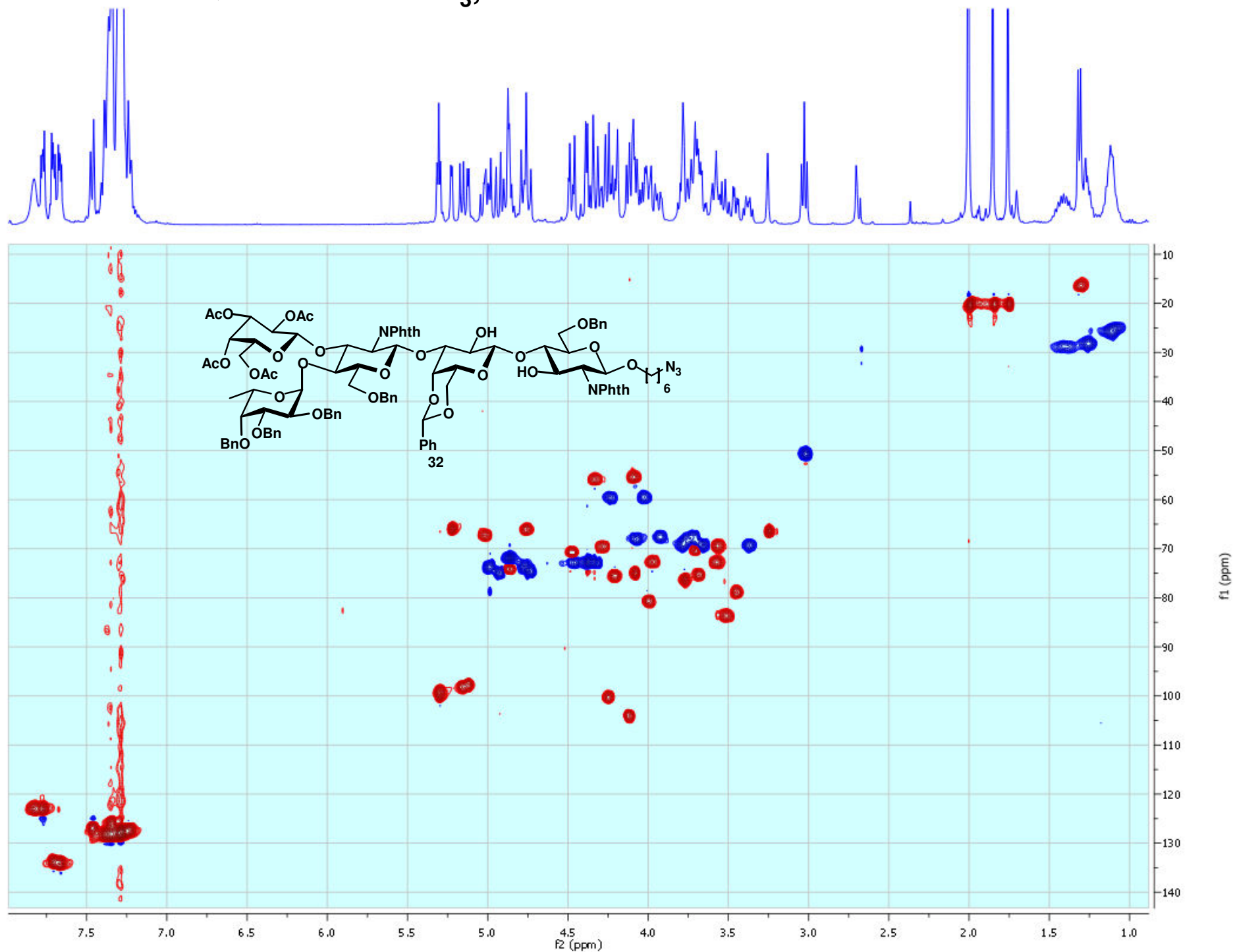


P. Zhang pzi.142 pos es
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Acquired: 15:47, January 06, 2009

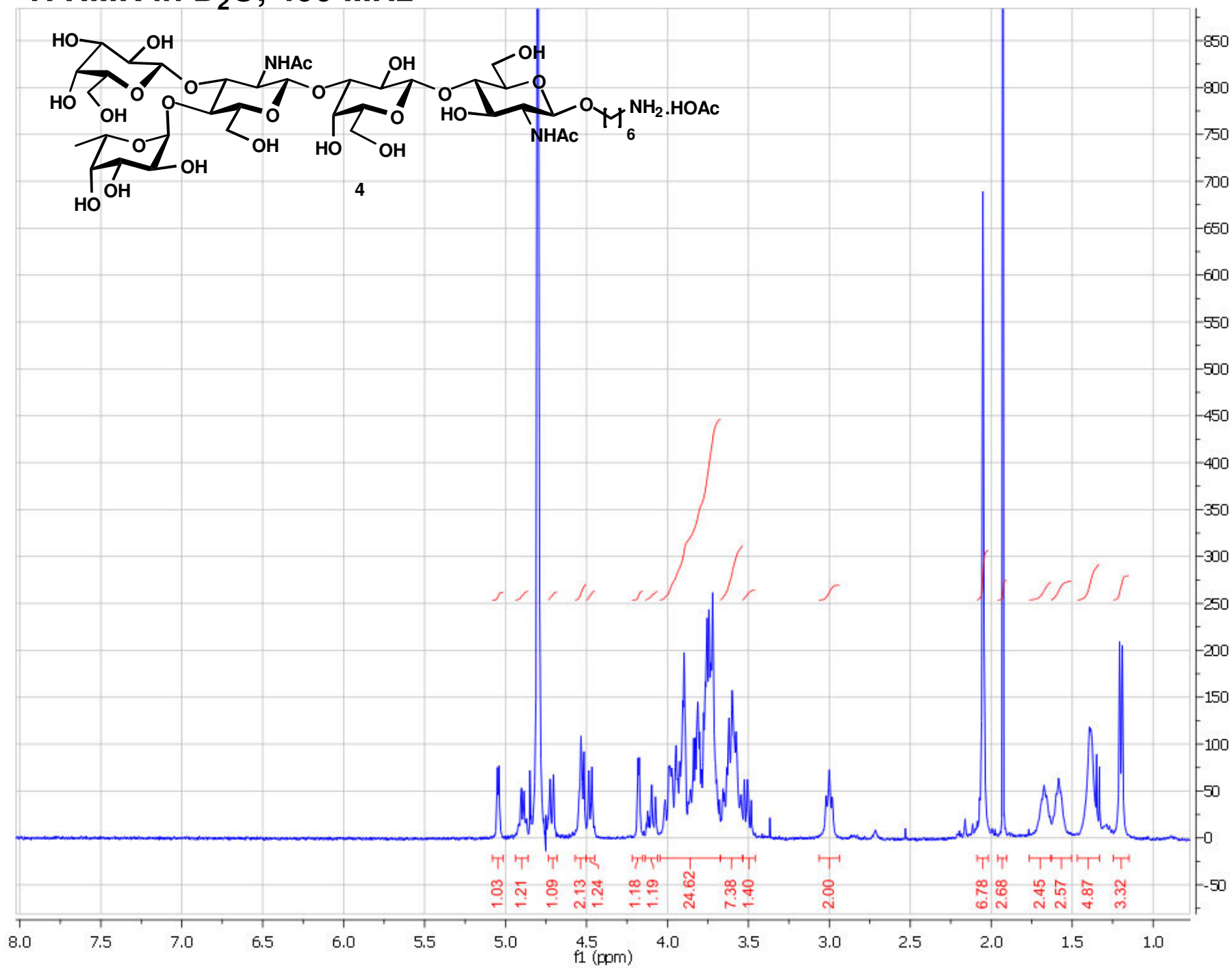
^1H NMR in CDCl_3 , 400 MHz



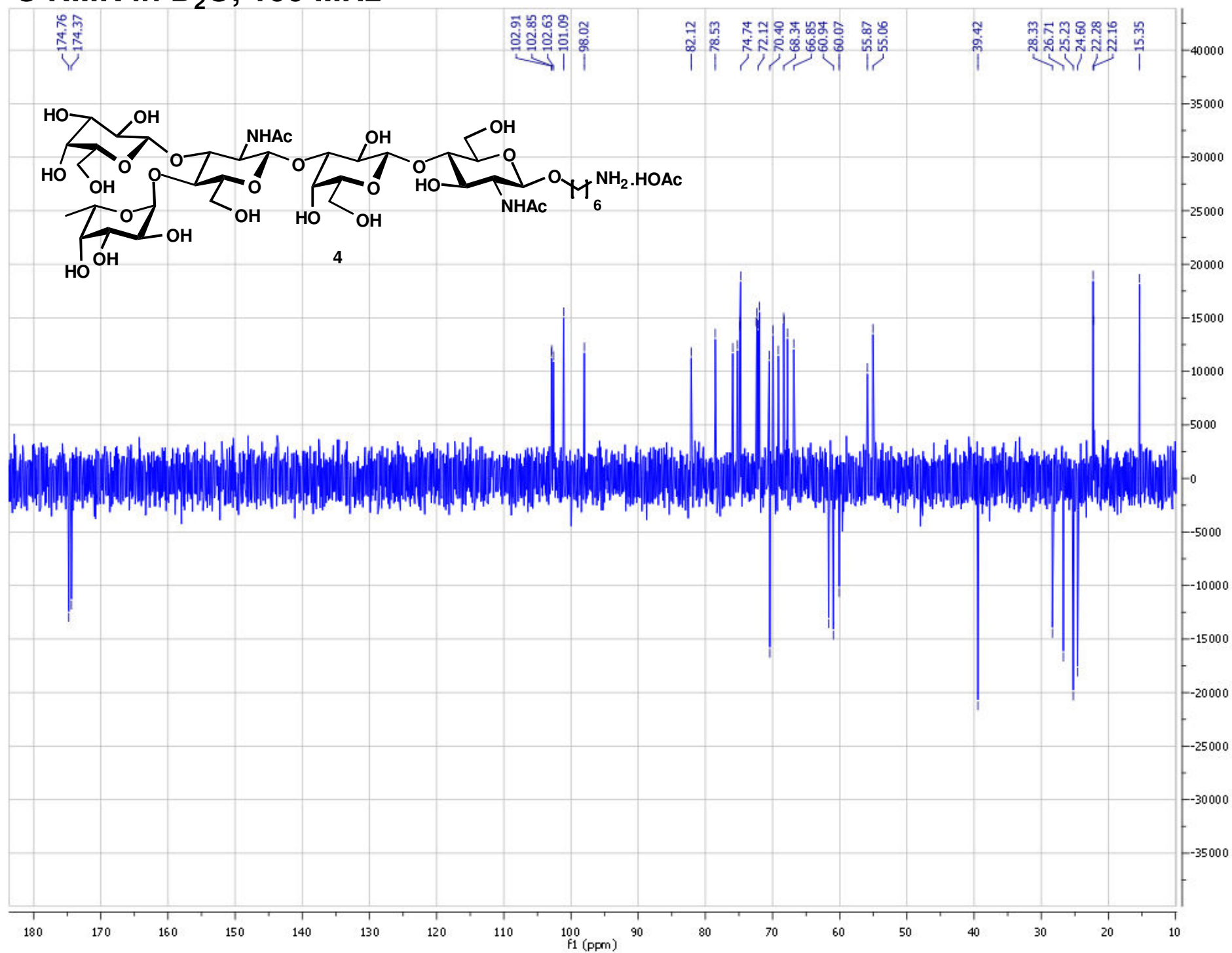
^1H - ^{13}C GHSQC NMR in CDCl_3 , 400 MHz



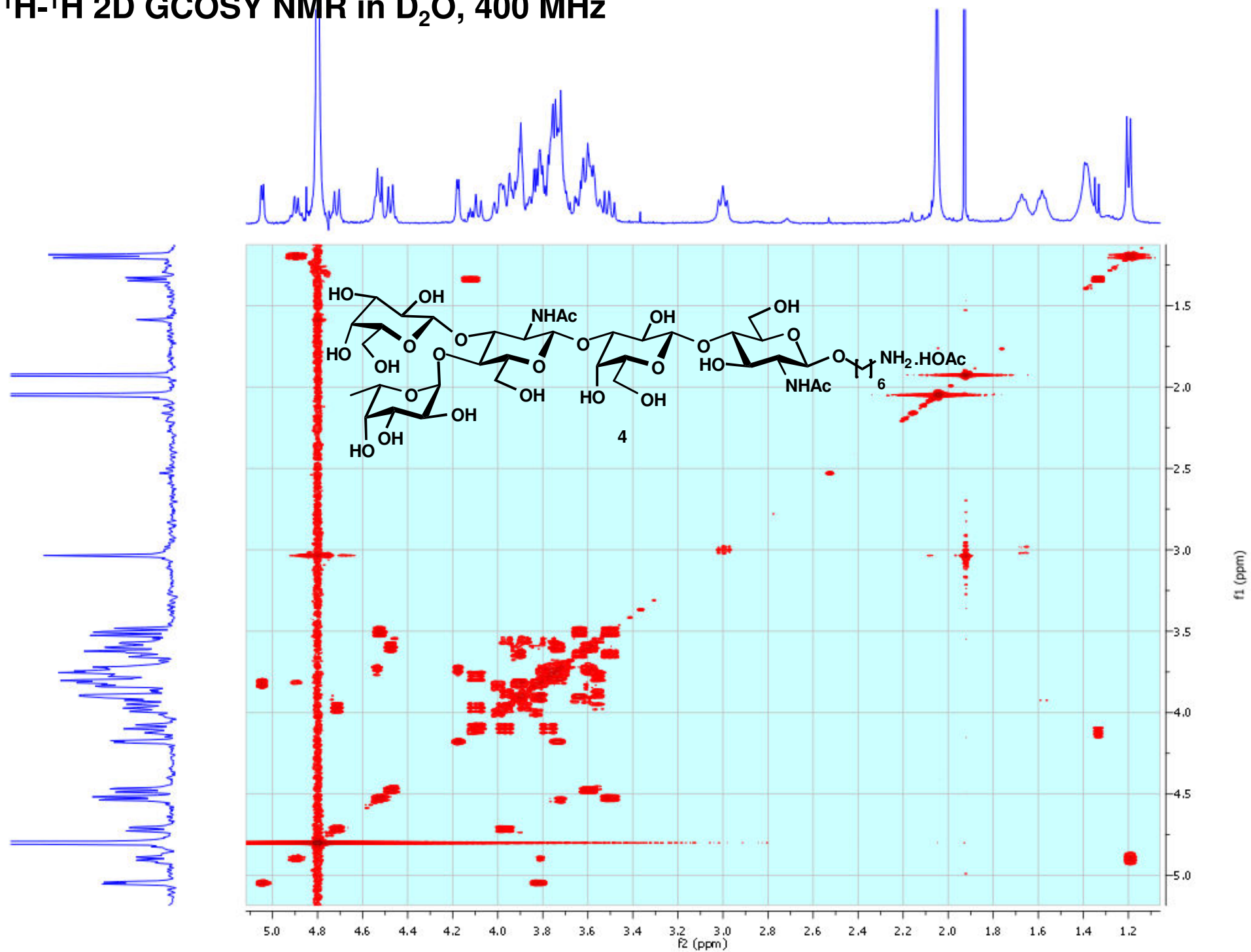
^1H NMR in D_2O , 400 MHz



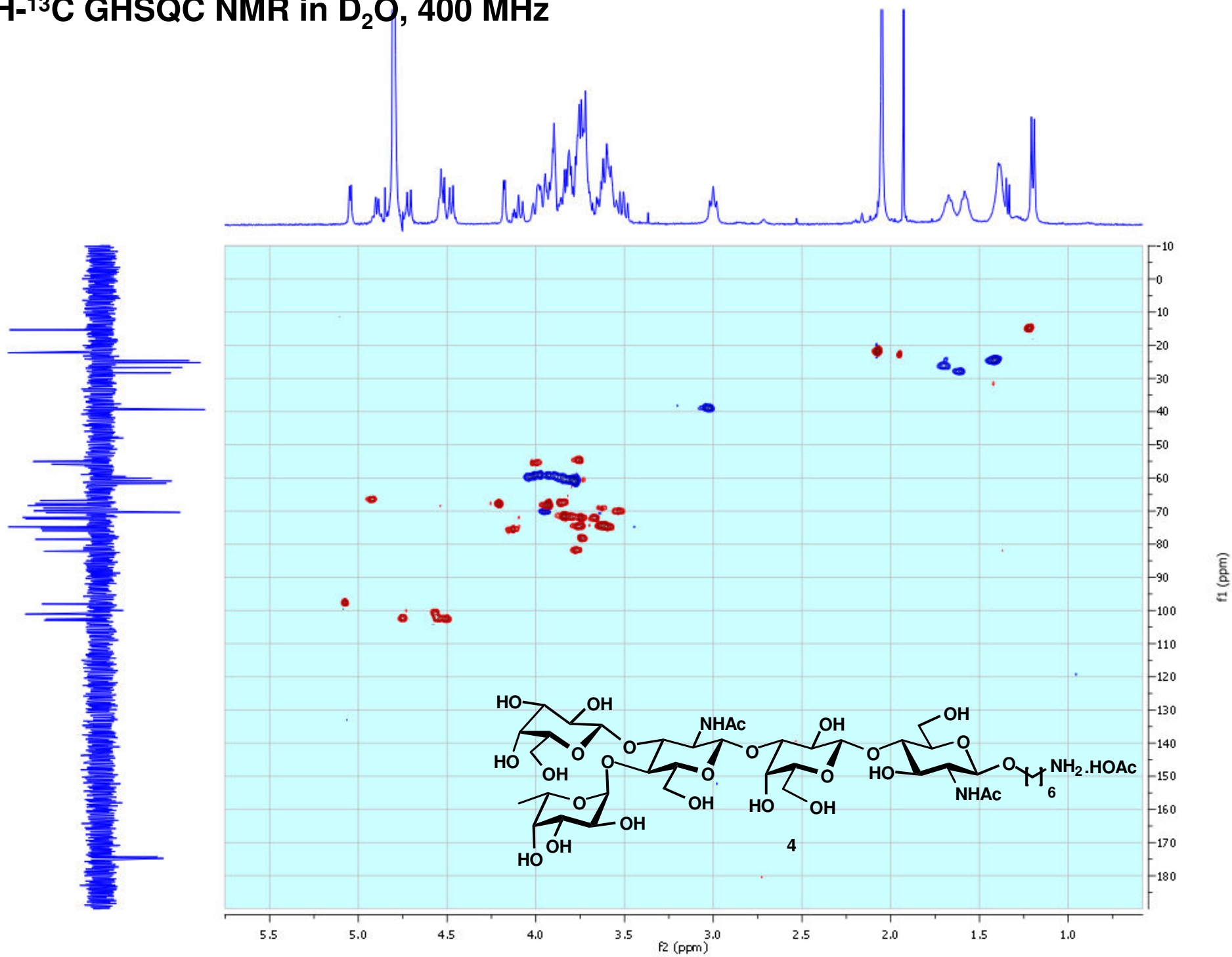
^{13}C NMR in D_2O , 100 MHz



^1H - ^1H 2D GCOSY NMR in D_2O , 400 MHz

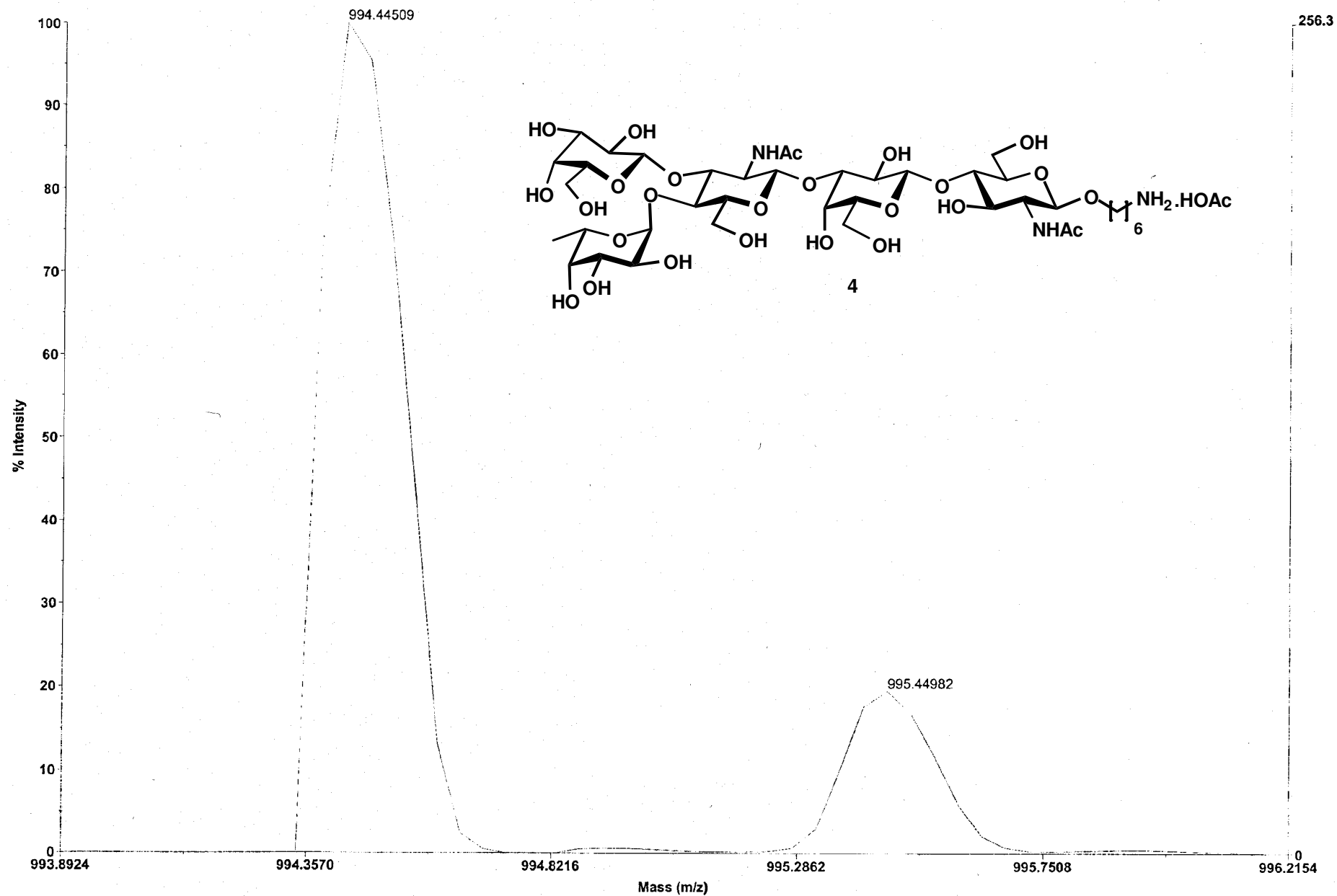


^1H - ^{13}C GHSQC NMR in D_2O , 400 MHz



High Resolution ESI Mass Spectra

Mariner Spec /7:11 ASC MC=>SM5[BP = 994.4, 256]



P. Zhang pz.i.92 pos es
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Acquired: 18:53, June 05, 2008