

Supporting Information for:

**Molecular recognition of N-protected dipeptides by pseudopeptidic macrocycles: a comparative study of the supramolecular complexes by ESI-MS and NMR**

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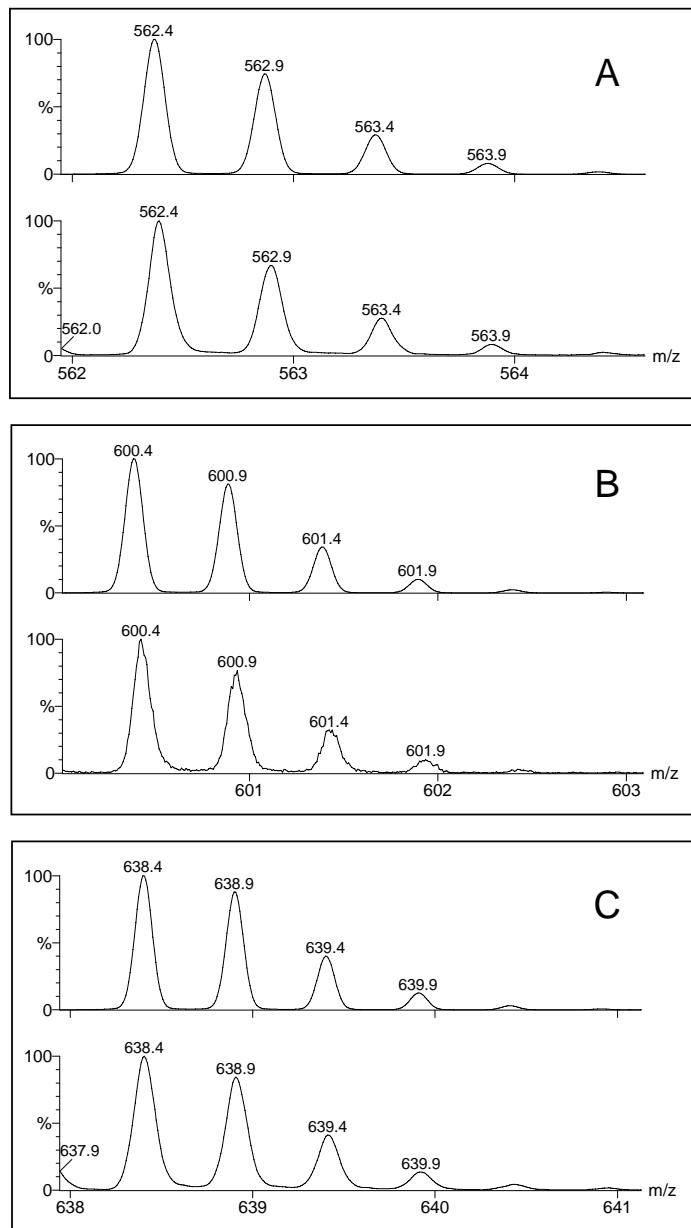
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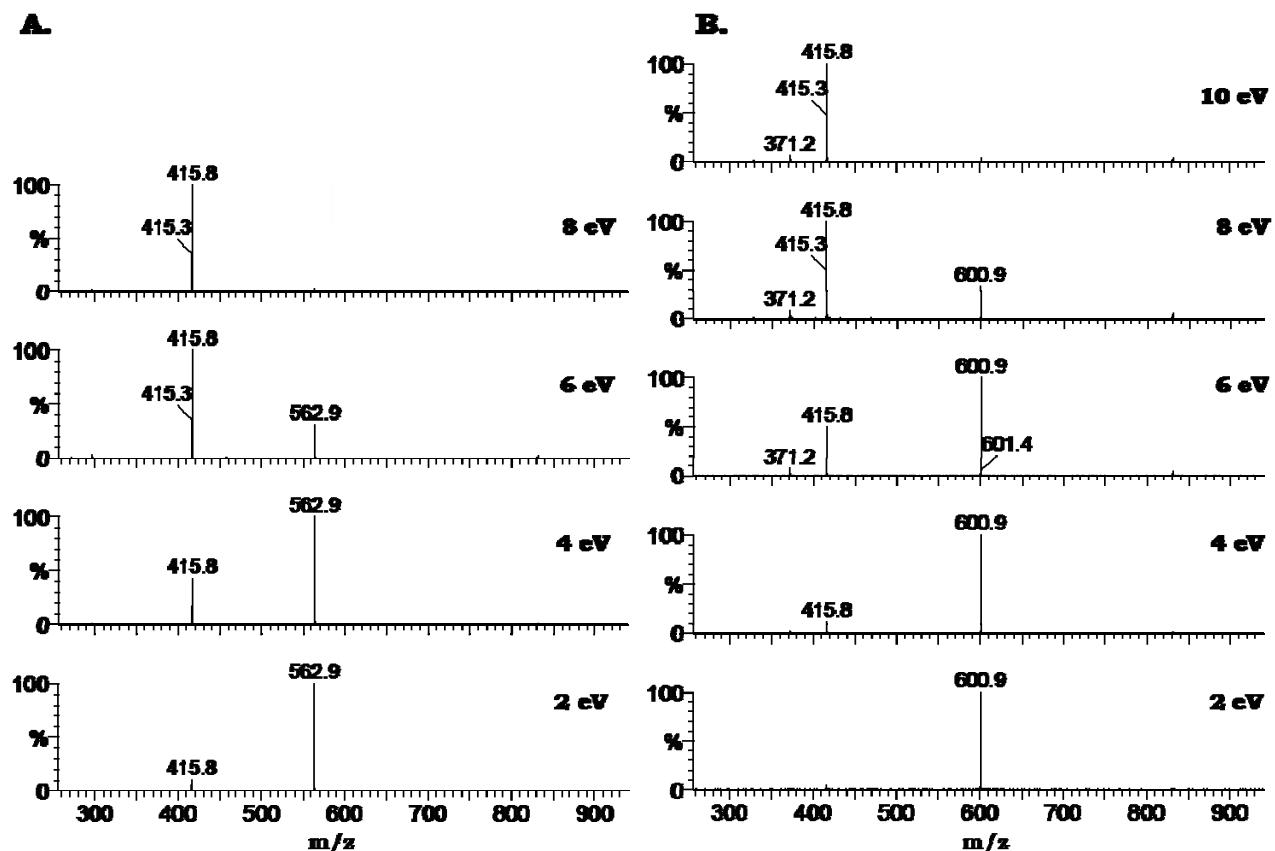
**Table of contents:**

High resolution ESI-TOF MS spectra (Figure S1)	S2
CID spectra (Figure S2)	S3
NMR spectra of dipeptides (Figures S3-S9)	S4
NMR spectra of <b>2a</b> (Figures S10-S14)	S8
NOESY/ROESY spectra of the [ <b>2a</b> · Z-AF-OH] complex (Figures S15-S17)	S10
<sup>1</sup> H NMR Titration procedures (Figures S18-S29)	S12
X ray diffraction data of the crystals of Z-AF-OH (Table S1)	S20
Cartesian coordinates for the minimum structure of [ <b>2a</b> · Z-AF-OH]	S21
Output file from the HYDRONMR program	S22

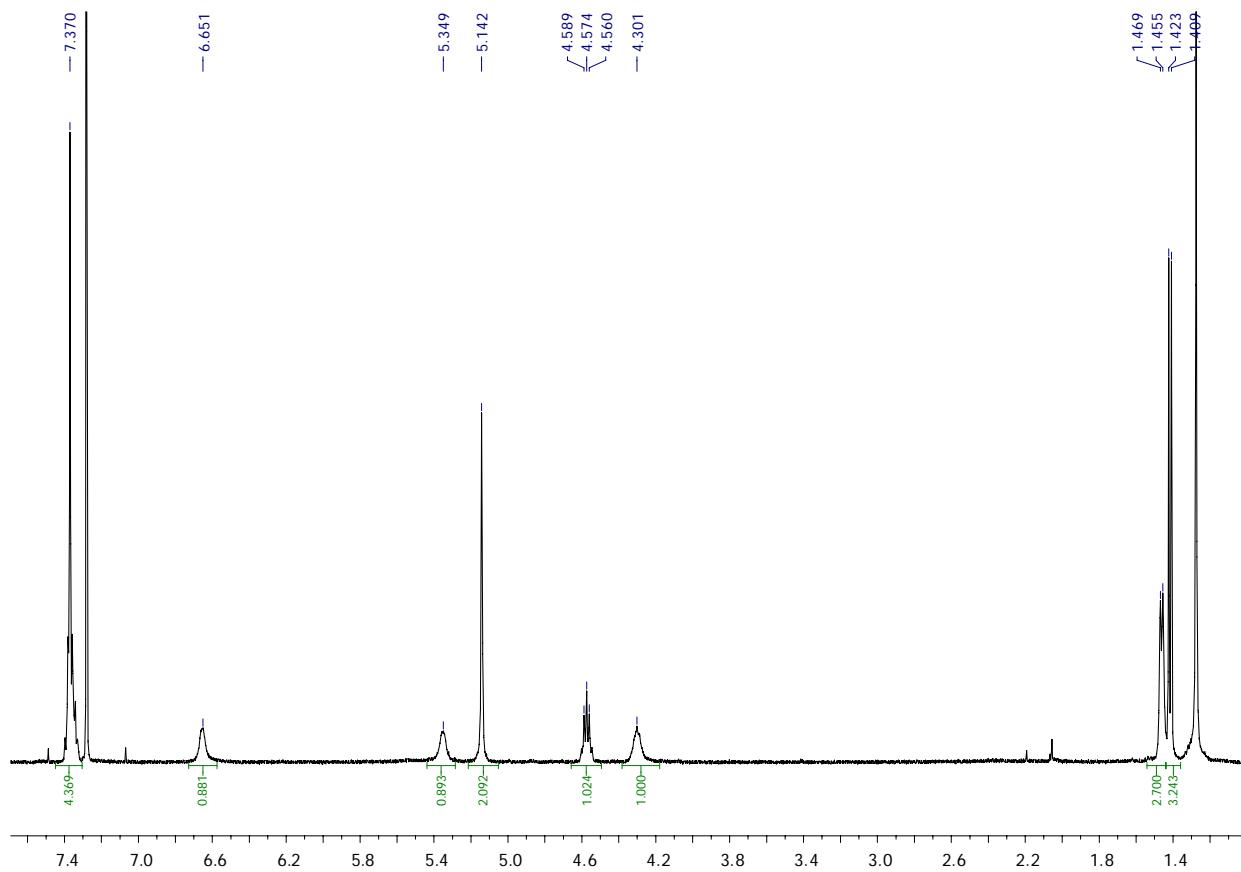
**Figure S1.** Detail of the ESI-TOF spectra for the doubly charged supramolecular complexes formed between **2a** and (a) Z-AA-OH, (b) Z-AF-OH and (c) Z-FF-OH. For every case, the upper and lower traces show the simulated and experimental peaks, respectively.



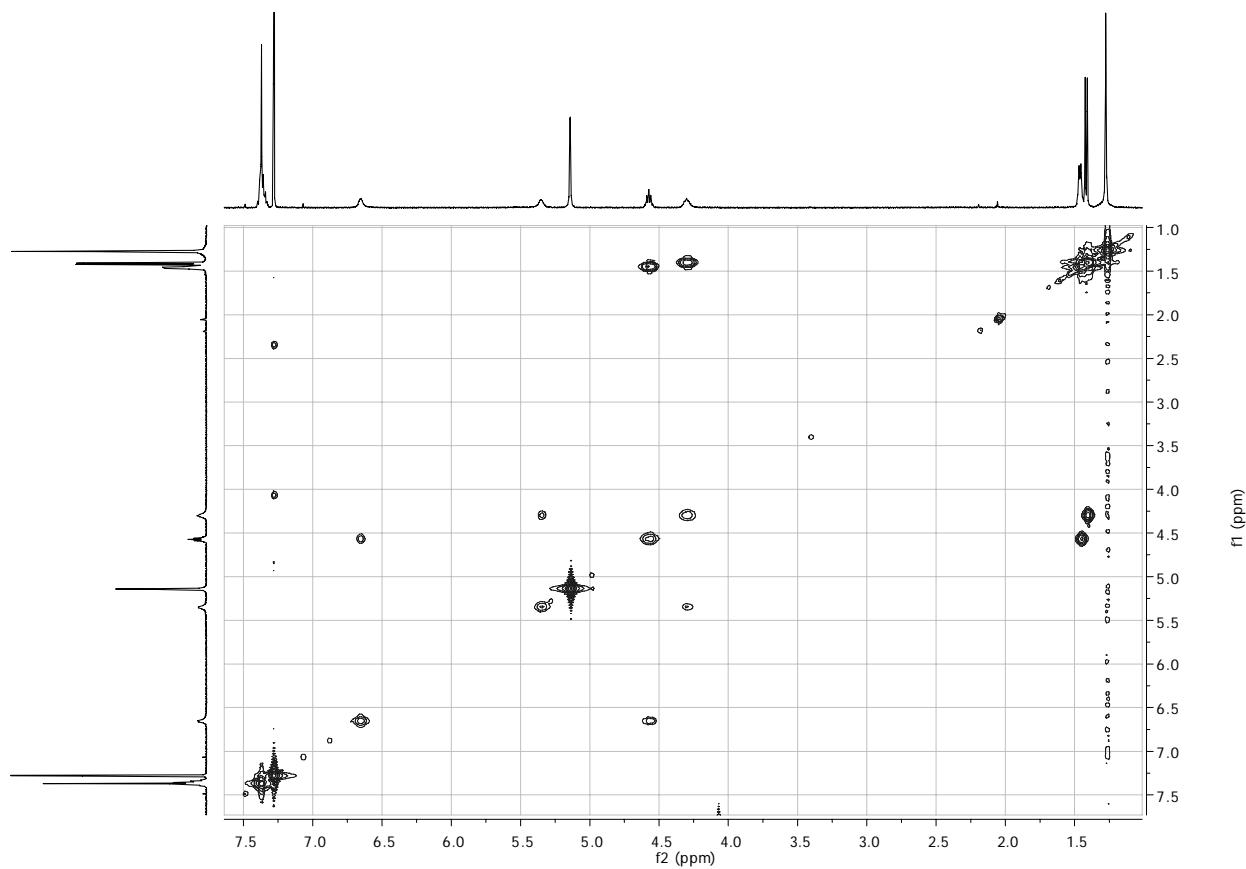
**Figure S2.** Collision Induced Dissociation (CID) spectra of the mass-selected (A)  $[2\mathbf{a} + \text{Z-AA-OH} + 2\text{H}]^{2+}$  and (B)  $[2\mathbf{a} + \text{Z-AF-OH} + 2\text{H}]^{2+}$  supramolecular complexes at increasing collision energies in the  $E_{\text{laboratory}} = 2\text{-}12\text{ eV}$  range.



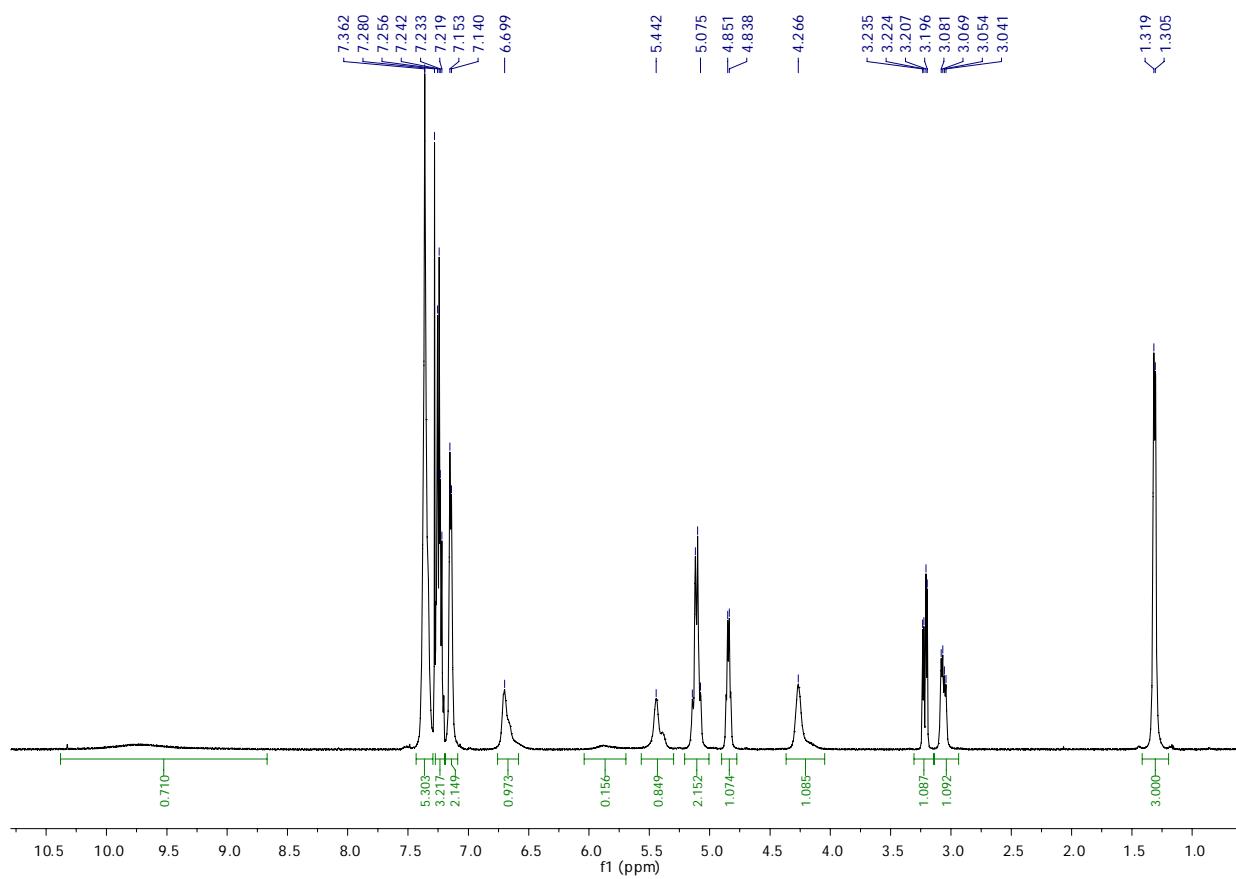
**Figure S3.**  $^1\text{H}$  NMR spectrum (500 MHz,  $\text{CDCl}_3$ ) of Z-AA-OH.



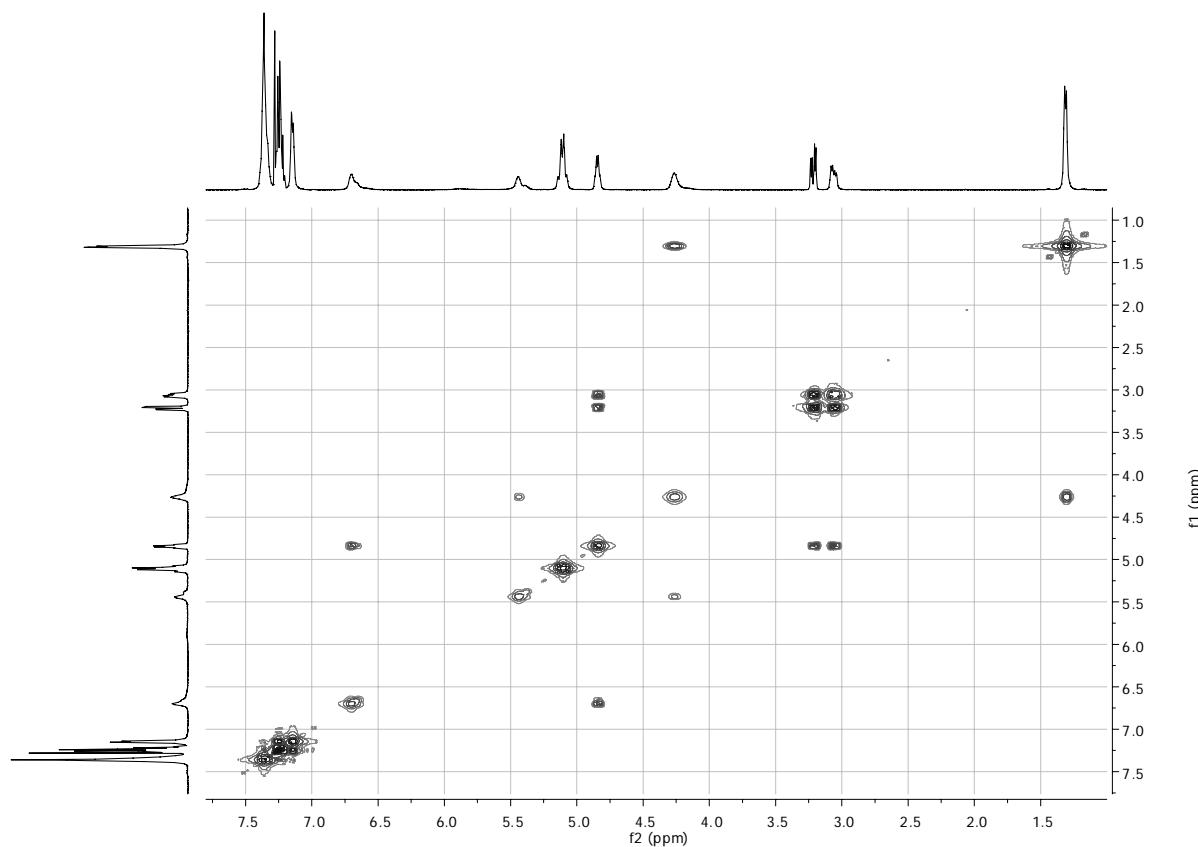
**Figure S4.** gCOSY NMR spectrum (500 MHz,  $\text{CDCl}_3$ ) of Z-AA-OH.



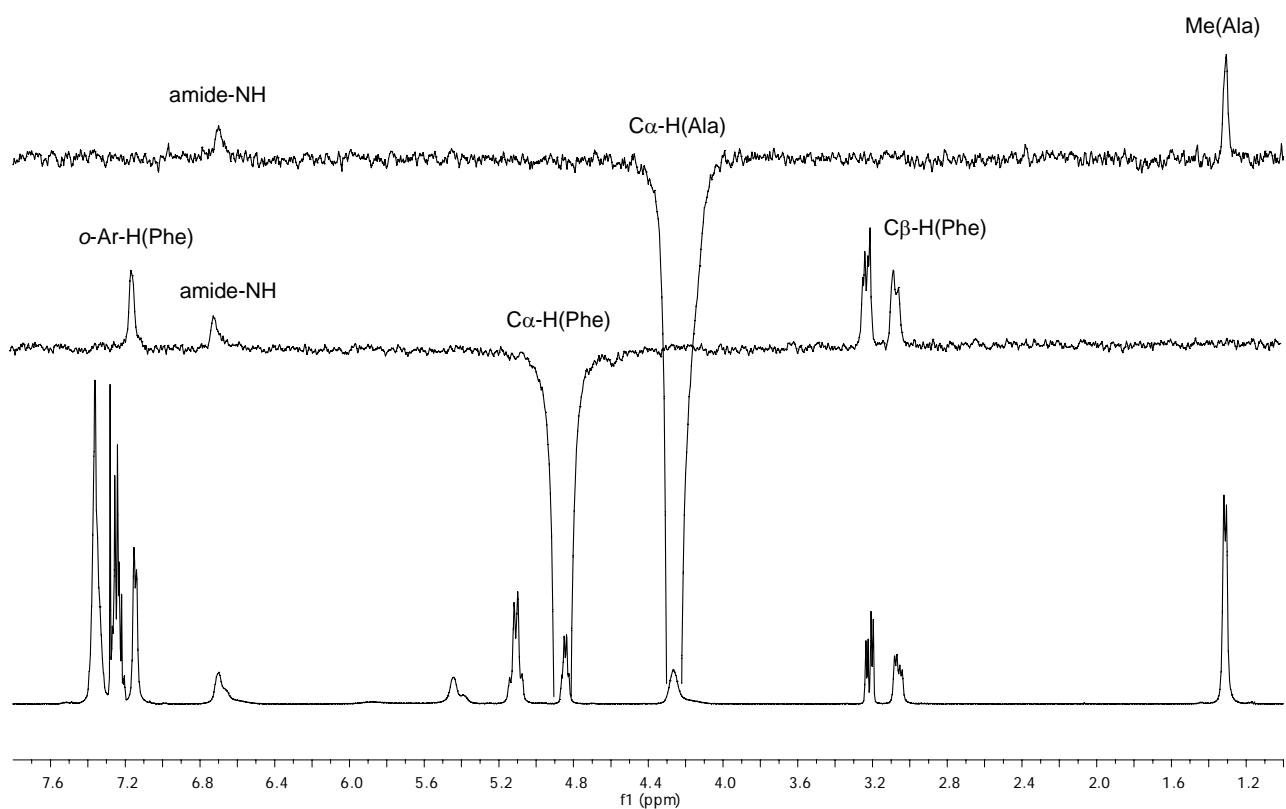
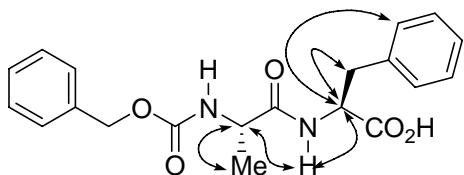
**Figure S5.**  $^1\text{H}$  NMR spectrum (500 MHz,  $\text{CDCl}_3$ ) of Z-AF-OH.



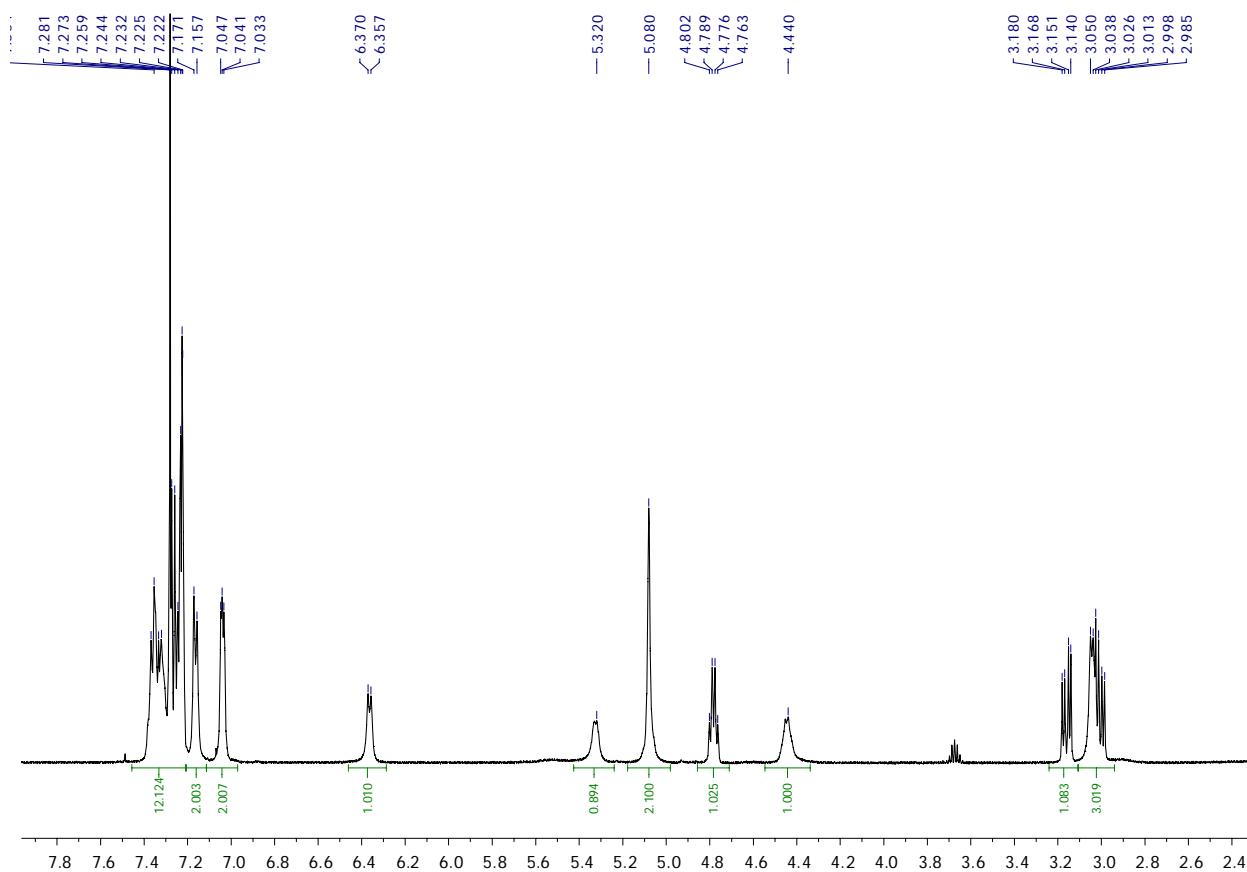
**Figure S6.** gCOSY NMR spectrum (500 MHz,  $\text{CDCl}_3$ ) of Z-AF-OH.



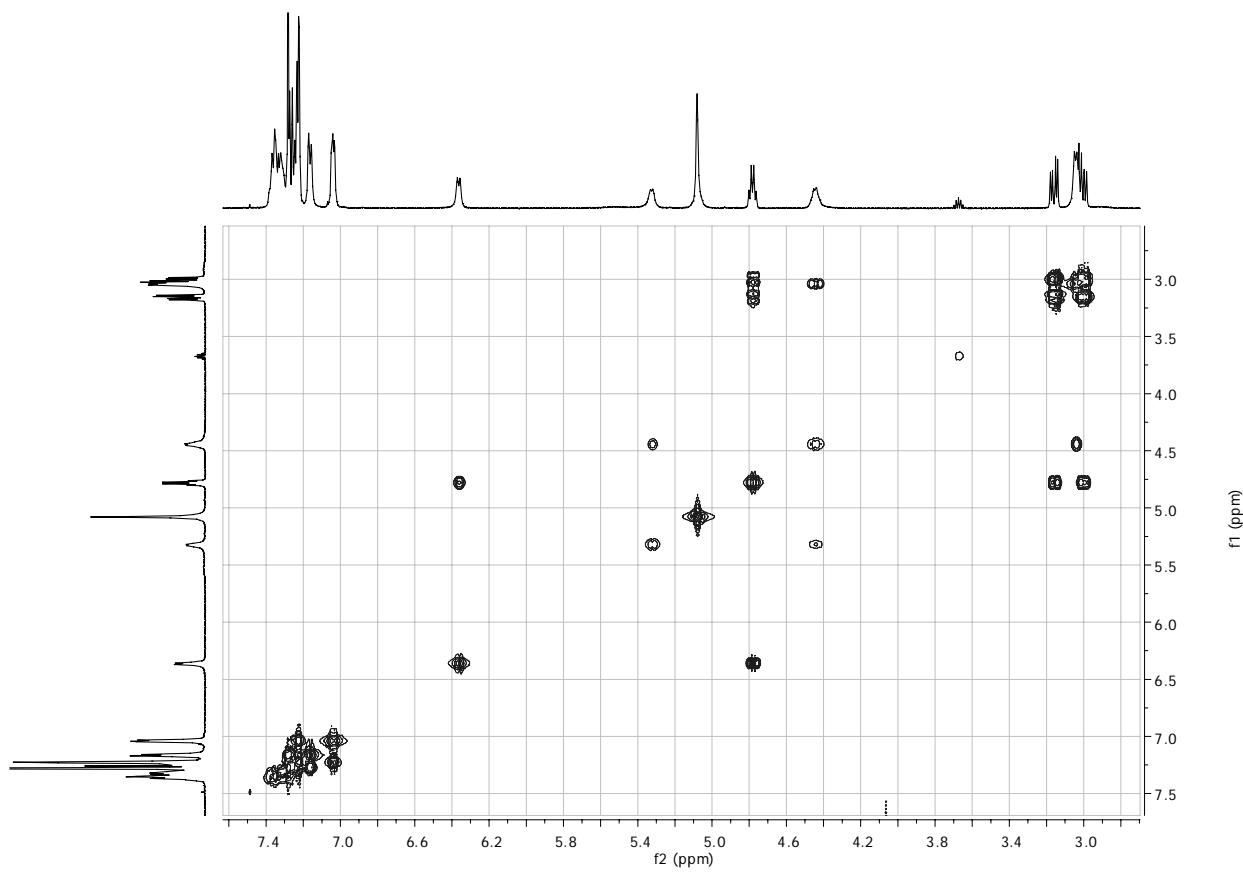
**Figure S7.**  $^1\text{H}$  NMR (lower trace) and 1D NOESY spectra (500 MHz,  $\text{CDCl}_3$ ) of Z-AF-OH upon irradiation of Ala (upper trace) and Phe (middle trace)  $\text{C}\alpha$  protons. The concentration of the sample was 30 mM, and positive nOEs were clearly observed (in antiphase with respect to the corresponding irradiated signal).



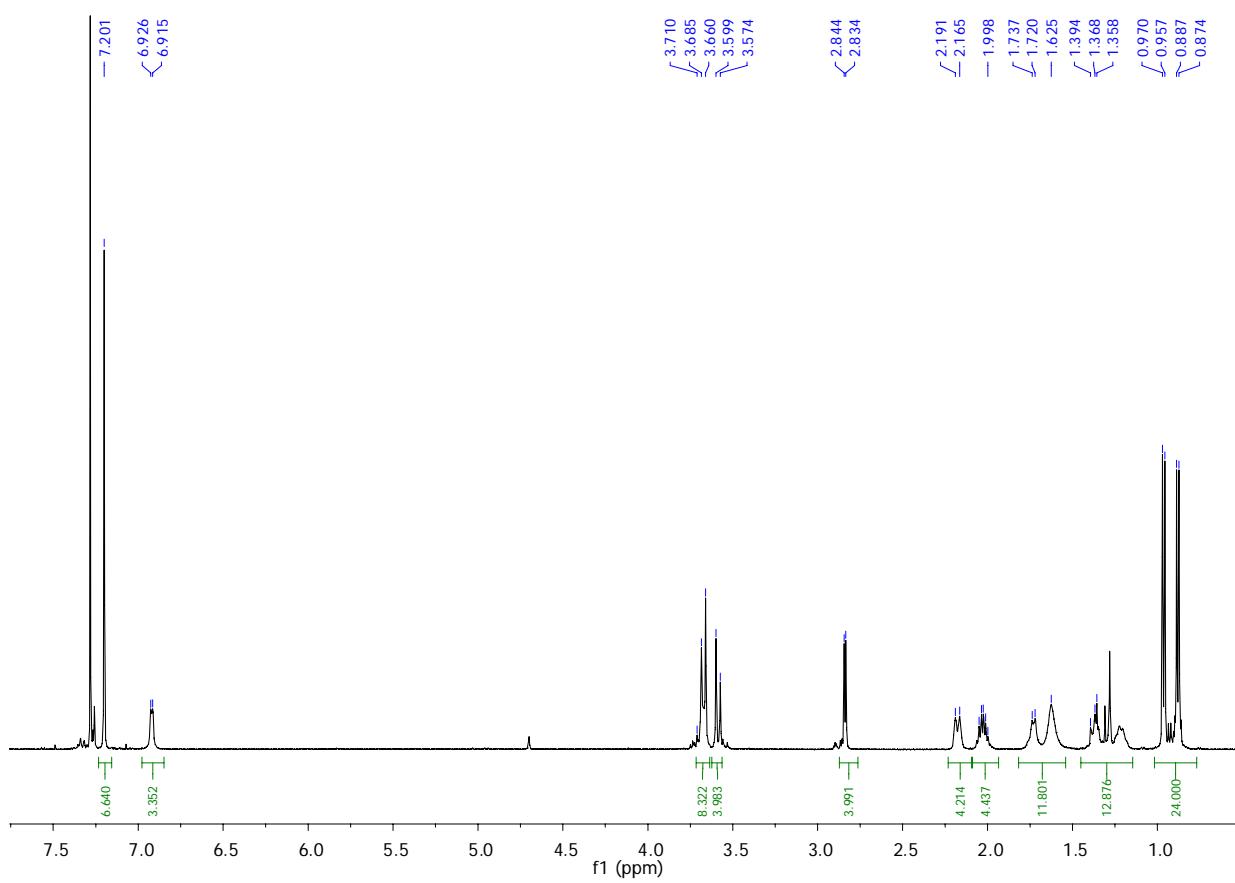
**Figure S8.**  $^1\text{H}$  NMR spectrum (500 MHz,  $\text{CDCl}_3$ ) of Z-FF-OH.



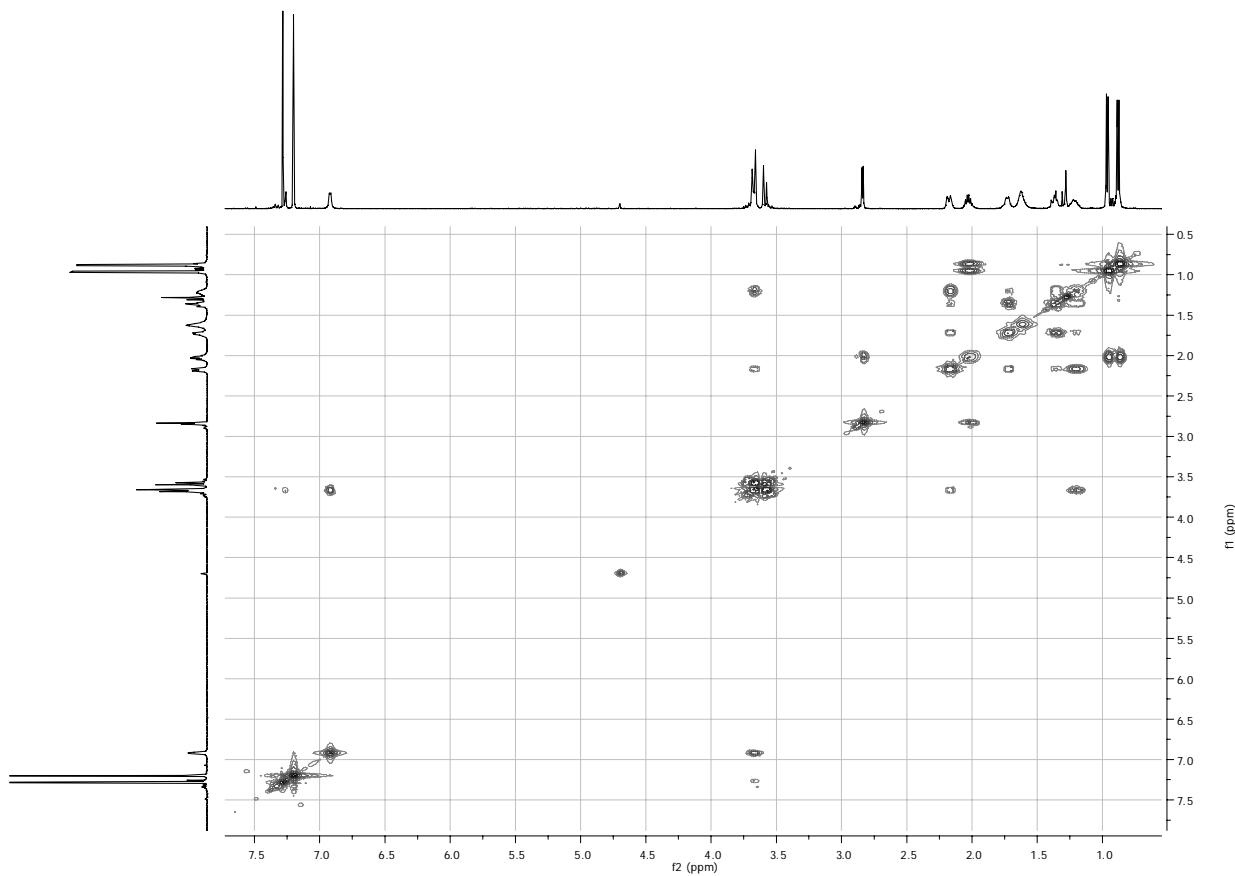
**Figure S9.** gCOSY NMR spectrum (500 MHz,  $\text{CDCl}_3$ ) of Z-FF-OH.



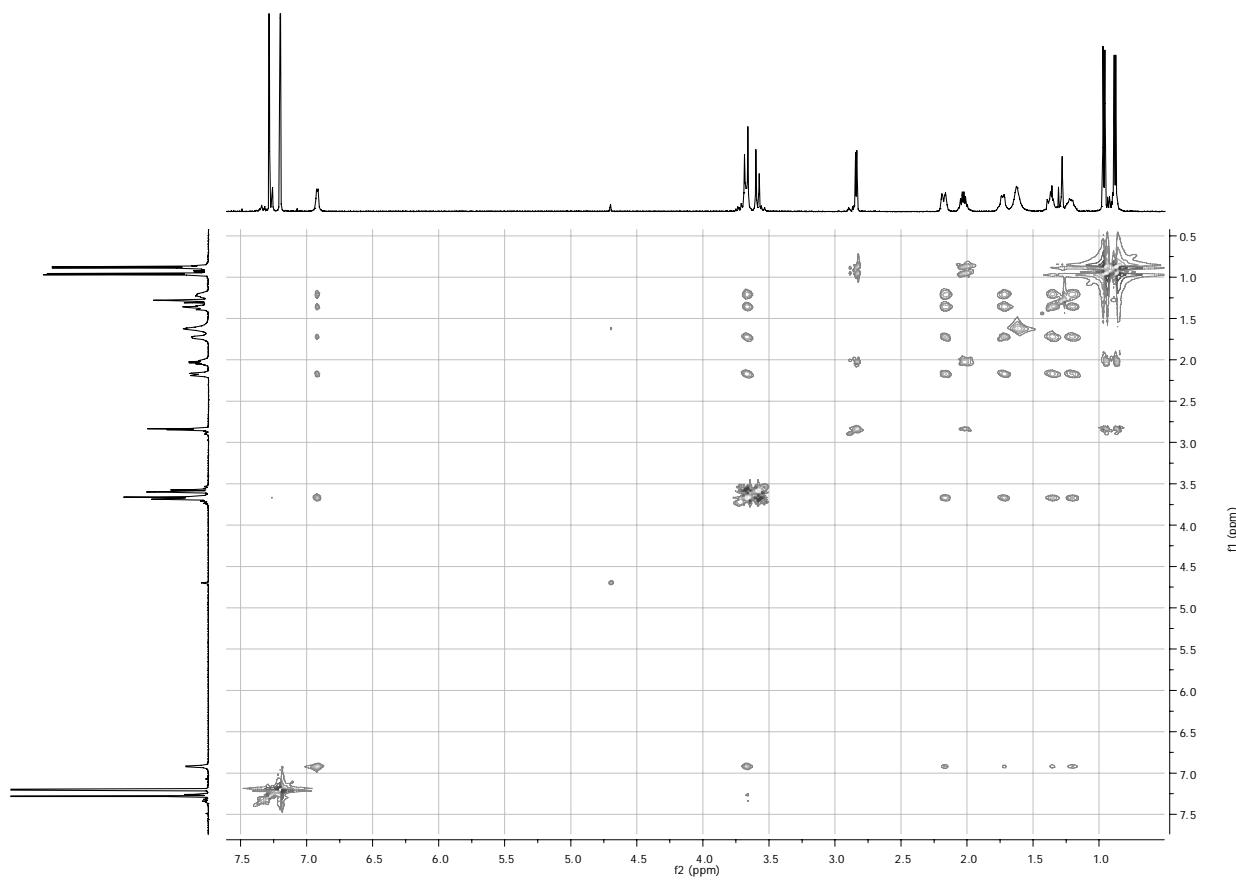
**Figure S10.**  $^1\text{H}$  NMR spectrum (500 MHz,  $\text{CDCl}_3$ ) of **2a**.



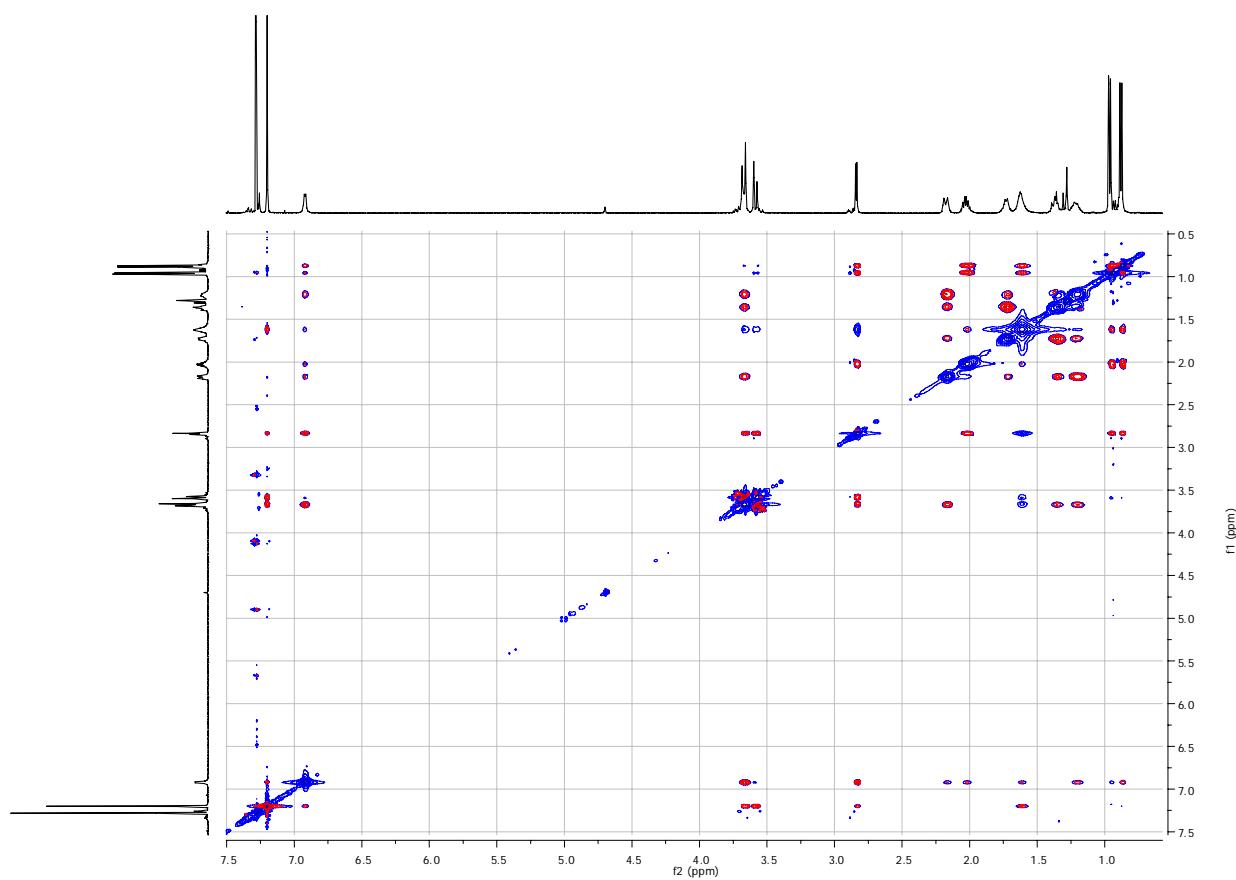
**Figure S11.** gCOSY NMR spectrum (500 MHz, CDCl<sub>3</sub>) of **2a**.



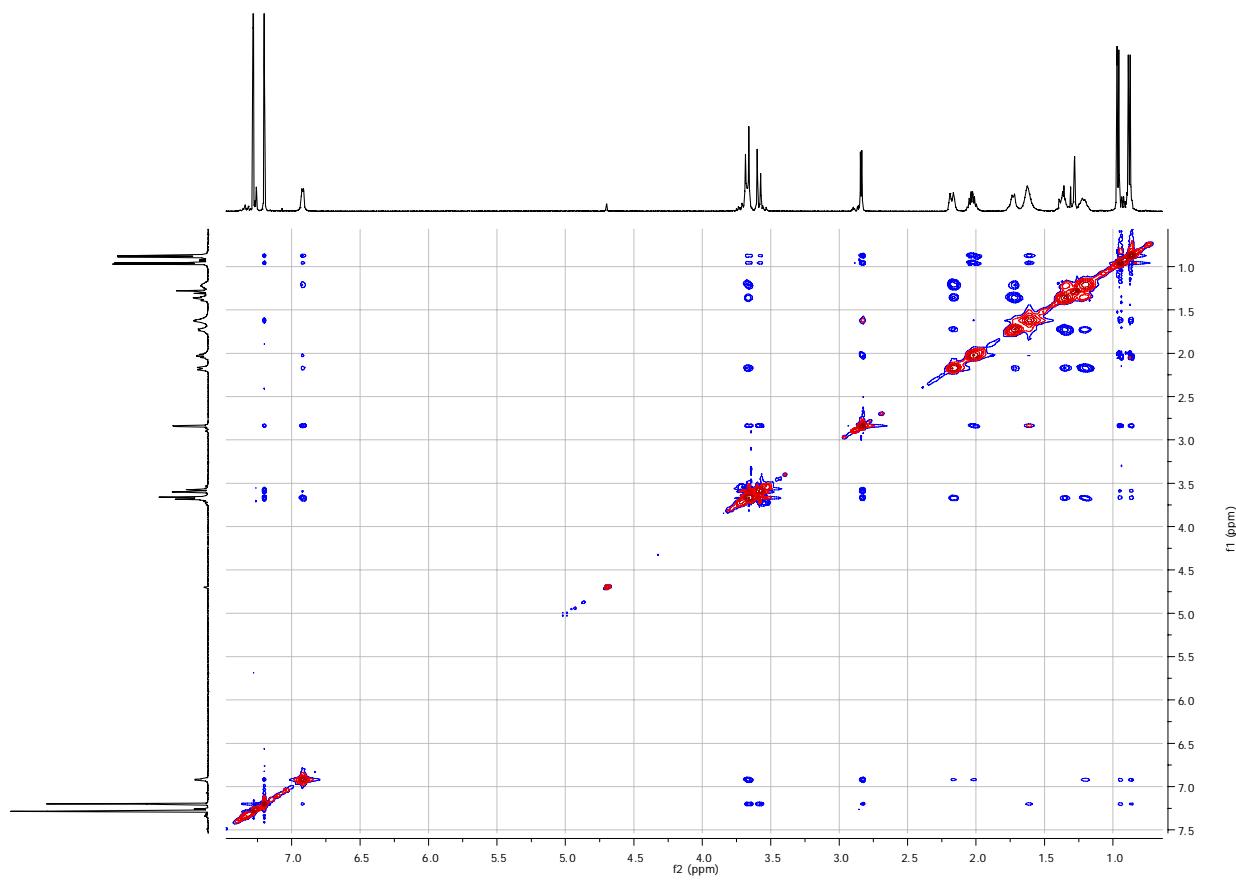
**Figure S12.** gTOCSY NMR spectrum (500 MHz,  $\text{CDCl}_3$ ) of **2a**.



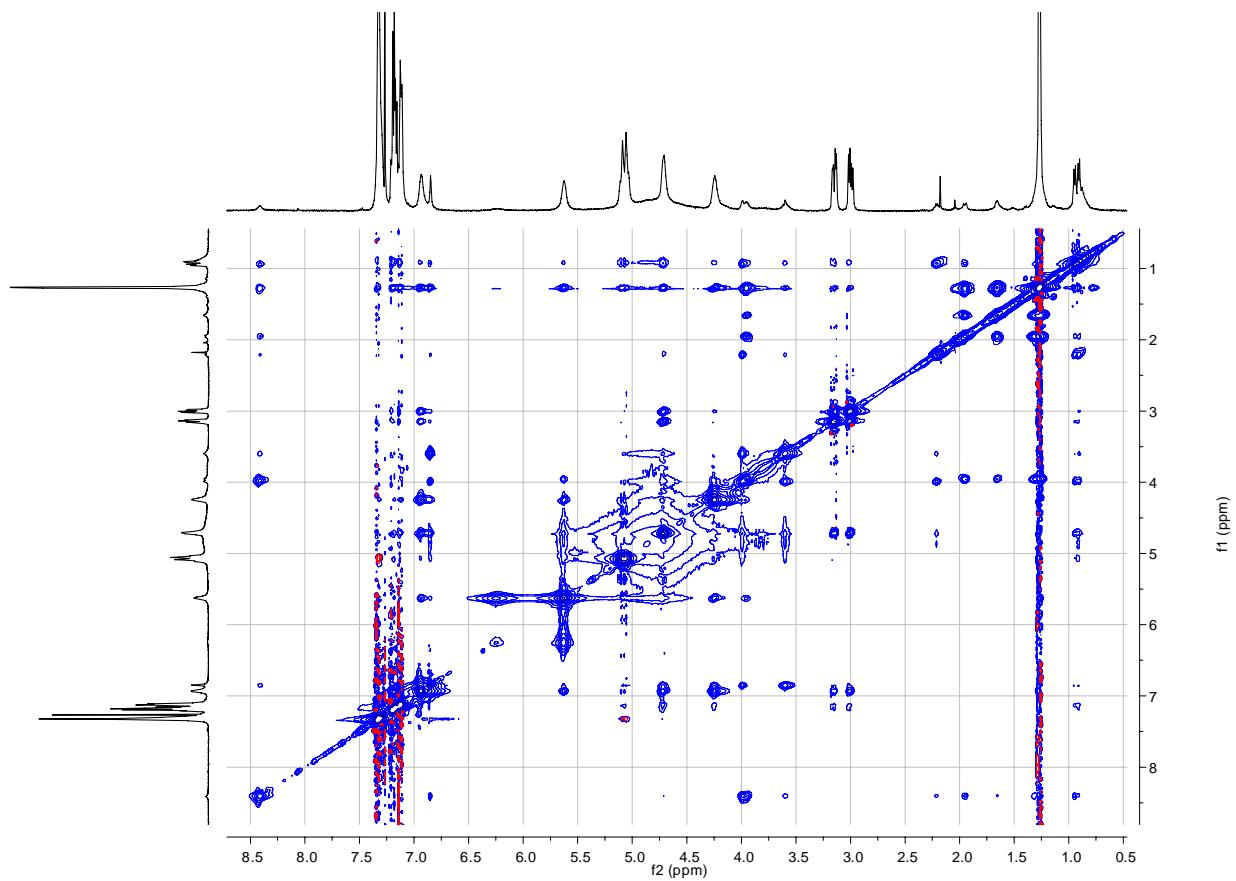
**Figure S13.** NOESY NMR spectrum (500 MHz,  $\text{CDCl}_3$ ) of **2a**.



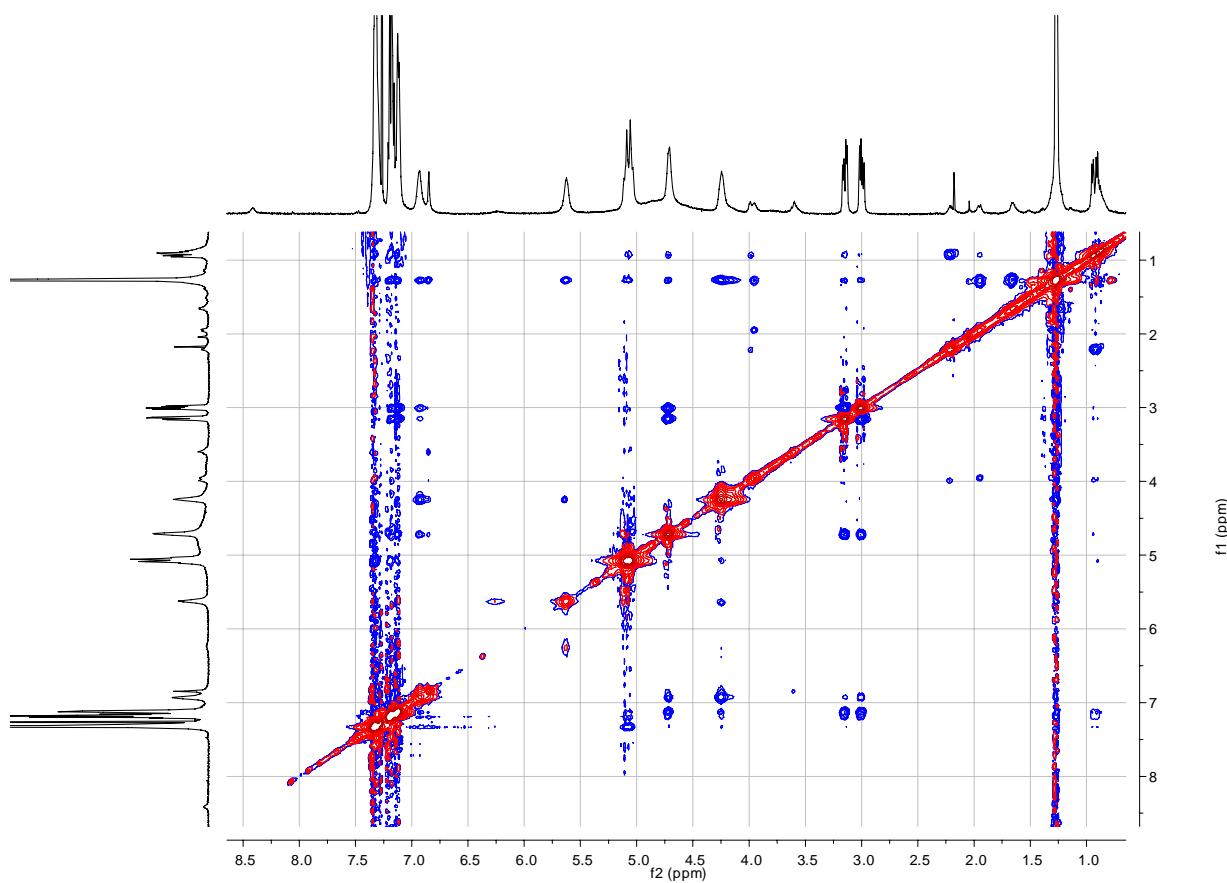
**Figure S14.** ROESY NMR spectrum (500 MHz,  $\text{CDCl}_3$ ) of **2a**.



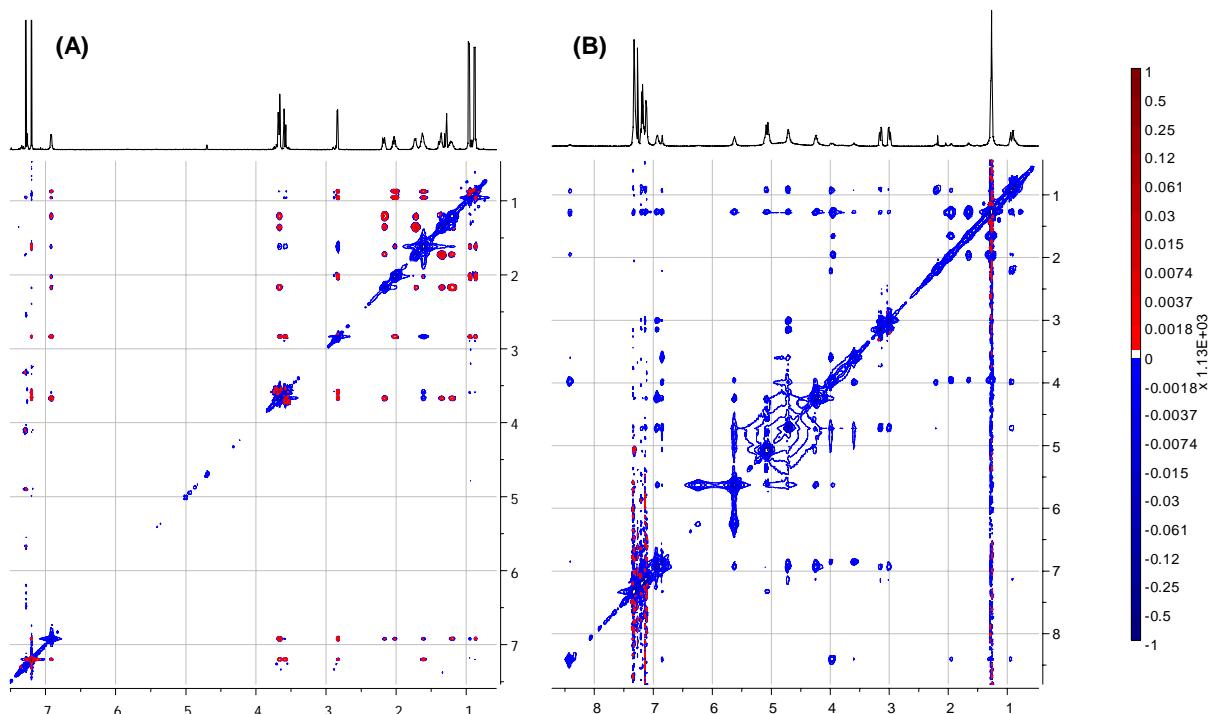
**Figure S15.** NOESY NMR spectrum (500 MHz,  $\text{CDCl}_3$ ) of **2a** saturated with Z-AF-OH



**Figure S16.** ROESY NMR spectrum (500 MHz,  $\text{CDCl}_3$ ) of **2a** saturated with Z-AF-OH



**Figure S17.** Comparison of the NOESY NMR spectra (500 MHz,  $\text{CDCl}_3$ ) of **2a** alone (A) and saturated with Z-AF-OH (B)



## <sup>1</sup>H NMR Titration procedures

All the NMR titrations were performed at 303 K in a Varian INOVA 500 spectrometer operating at 500 MHz for proton. The non-linear least squares fitting calculations were performed with KaleidaGraph versio 4.03 by Synergy Software.

**Titration procedure for Z-F-OH:** We prepared a 3.3 mM stock solution of **2a** in dry, acid free CDCl<sub>3</sub> containing 1% MeOH. For the titrant solutions, weighted amounts of Z-F-OH were dissolved in the stock solution of the receptor (0.5-1.0 M final concentration of Z-F-OH) and thus, the solution of the titrant also contained a 3.3 mM concentration of **2a** in order to keep the concentration of the receptor constant during the titration experiment. Then, 0.5 mL of the stock solution of the receptor was placed in a NMR tube and the <sup>1</sup>H NMR spectrum was acquired. Afterwards, the receptor was carefully titrated by addition of small amounts of titrant solutions of the substrate. The <sup>1</sup>H NMR spectra were acquired (500 MHz, 303 K) after each addition, using 16-32 scans and a relaxation delay of 5 s. Different <sup>1</sup>H NMR signals of **2a** were monitored during the experiment, although the most suitable for quantitative analysis was that corresponding to the receptor amide NH protons.

We used the simplest 1 : 1 binding model to fit the data, using the non-linear least squares fitting of the experimental values to the following equation:<sup>1</sup>

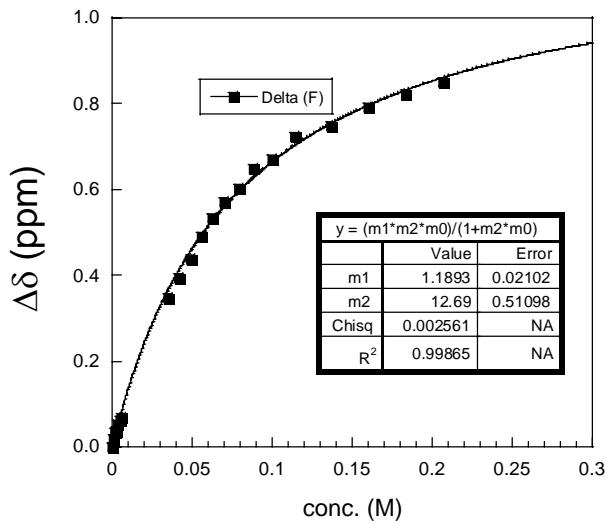
$$\Delta\delta = \frac{\Delta\delta_{\max} K_{\text{as}} [L]_{\text{tot}}}{1 + K_{\text{as}} [L]_{\text{tot}}}$$

Where:  $\Delta\delta$  (y, dependent variable) is the observed variation of the chemical shift ( $\delta_{\text{obs}} - \delta_0$ ) being  $\delta_{\text{obs}}$  the chemical shift at each titration point and  $\delta_0$  the initial chemical shift (host alone)  
 $[L]_{\text{tot}}$  is the overall concentration of the guest titrant (m0, independent variable)  
 $\Delta\delta_{\max}$  is the maximum variation of the observed chemical shift (m1 in the fitting)  
 $K_{\text{as}}$  is the host-guest binding constant (m2 in the fitting)

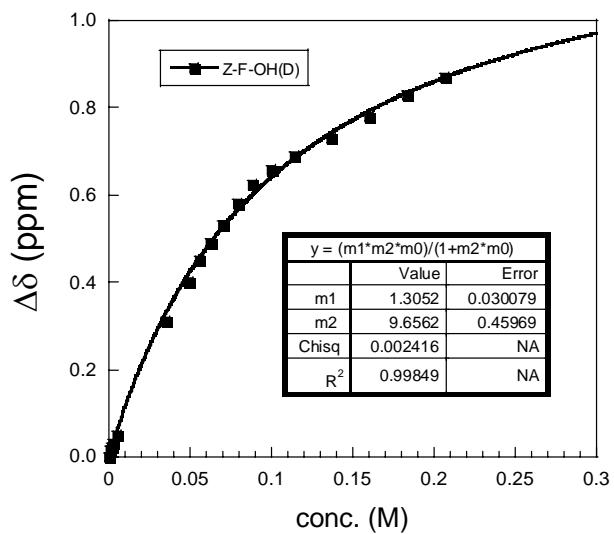
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<sup>1</sup> Connors, K. A. *Binding Constants: The Measurement of Molecular Complex Stability*, John Wiley & Sons, **1987**.

**Figure S18.** Titration of Z-F-OH(L) (500 MHz, 303 K, 1% MeOH in CDCl<sub>3</sub>): the symbols correspond to experimental data and the line to the non-linear least squares fitting. The final values for the best fitting are given in the figure.



**Figure S19.** Titration of Z-F-OH(D) (500 MHz, 303 K, 1% MeOH in CDCl<sub>3</sub>): the symbols correspond to experimental data and the line to the non-linear least squares fitting. The final values for the best fitting are given in the figure.



**Self-assembling of dipeptides:** Solutions of the dipeptides in  $\text{CDCl}_3$  containing 1% of MeOH were prepared at different final concentrations (0.1-90 mM, depending on peptide solubility). The  $^1\text{H}$  NMR spectra were acquired (500 MHz, 303 K) for every sample, using 32-64 scans and a relaxation delay of 5 s. Different  $^1\text{H}$  NMR signals were monitored during the experiment, although the most suitable for quantitative analysis were those corresponding to the amide NH protons. We used the simplest monomer-dimer equilibrium model to successfully fit the experimental data using the non-linear least squares fitting to the following equation:<sup>2</sup>

$$\delta_{\text{obs}} = \delta_m + \frac{1 + 4C_{\text{tot}}K_{\text{dim}} - \sqrt{1 + 8C_{\text{tot}}K_{\text{dim}}}}{4C_{\text{tot}}K_{\text{dim}}} \times (\delta_d - \delta_m)$$

Where:  $\delta_{\text{obs}}$  is the observed chemical shift at each titration point (y, dependent variable)

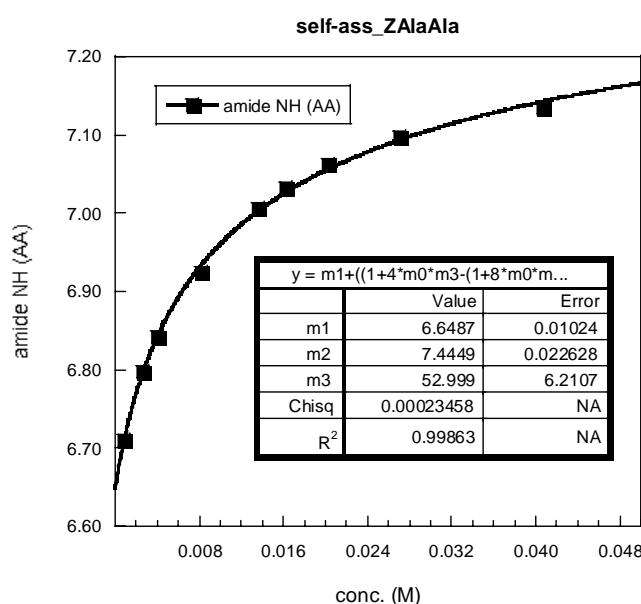
$C_{\text{tot}}$  is the overall concentration of the compound ( $m_0$ , independent variable)

$\delta_m$  is the chemical shift of the monomer ( $m_1$  in the fitting)

$\delta_d$  is the chemical shift of the dimer ( $m_2$  in the fitting)

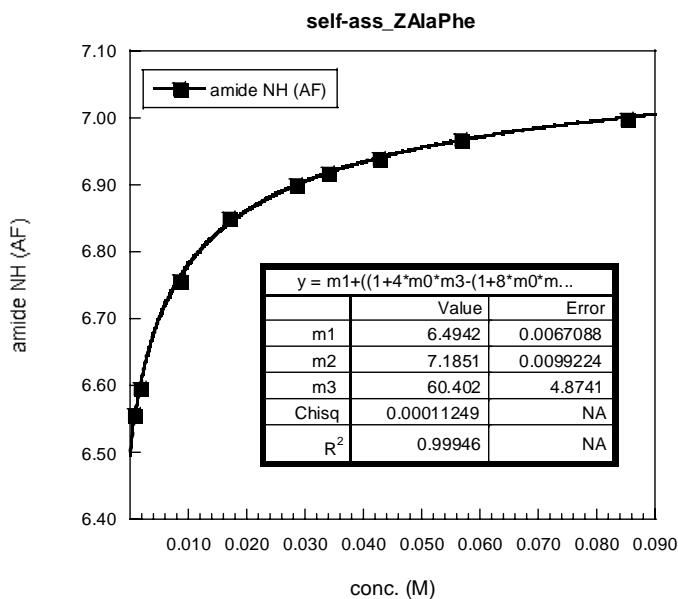
$K_{\text{dim}}$  is the dimerization equilibrium constant ( $m_3$  in the fitting)

**Figure S20.** Dilution titration of Z-AA-OH (500 MHz, 303 K, 1% MeOH in  $\text{CDCl}_3$ ): the symbols correspond to experimental data and the line to the non-linear least squares fitting. The final values for the best fitting are given in the figure.

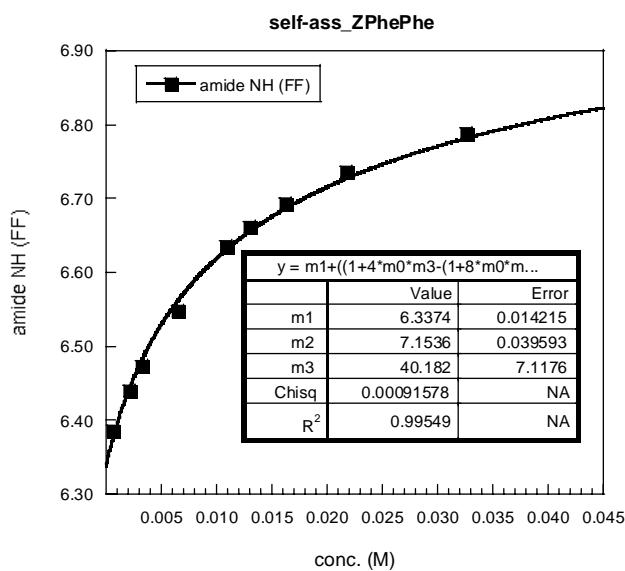


<sup>2</sup> (a) Schmuck, C.; Rehm, T.; Geiger, L.; Schäfer, M. *J. Org. Chem.* **2007**, 72, 6162. (b) Martin, R. B. *Chem. Rev.* **1996**, 96, 3043. (c) Davis, J. C. Jr.; Deb, K. K. *Adv. Magn. Reson.* **1970**, 4, 201. (d) Bangerter, B. W.; Chan, S. I. *J. Am. Chem. Soc.* **1969**, 91, 3910.

**Figure S21.** Dilution titration of Z-AF-OH (500 MHz, 303 K, 1% MeOH in CDCl<sub>3</sub>): the symbols correspond to experimental data and the line to the non-linear least squares fitting. The final values for the best fitting are given in the figure.



**Figure S22.** Dilution titration of Z-FF-OH (500 MHz, 303 K, 1% MeOH in CDCl<sub>3</sub>): the symbols correspond to experimental data and the line to the non-linear least squares fitting. The final values for the best fitting are given in the figure.



**Host-guest binding by dilution titration:** Samples of exactly equimolecular amounts of **2a** or **1a** and a given substrate were prepared in  $\text{CDCl}_3$  containing 1% of MeOH and at different final global concentrations (1-20 mM). The  $^1\text{H}$  NMR spectra were acquired (500 MHz, 303 K) for every sample, using 32-64 scans and a relaxation delay of 5 s. Different  $^1\text{H}$  NMR signals were monitored during the experiment, although the most suitable for quantitative analysis were those corresponding to the dipeptide amide NH protons.

We used the simplest 1 : 1 binding model to fit the data, using the non-linear least squares fitting of the experimental values to the following equation:<sup>3</sup>

$$\delta_{\text{obs}} = \delta_o - \frac{(\delta_o - \delta_c)}{2C_{\text{tot}}K_{\text{as}}} \times \left[ 2C_{\text{tot}}K_{\text{as}} + 1 - \sqrt{4C_{\text{tot}}K_{\text{as}} + 1} \right]$$

Where:  $\delta_{\text{obs}}$  is the observed chemical shift at each titration point (y, dependent variable)

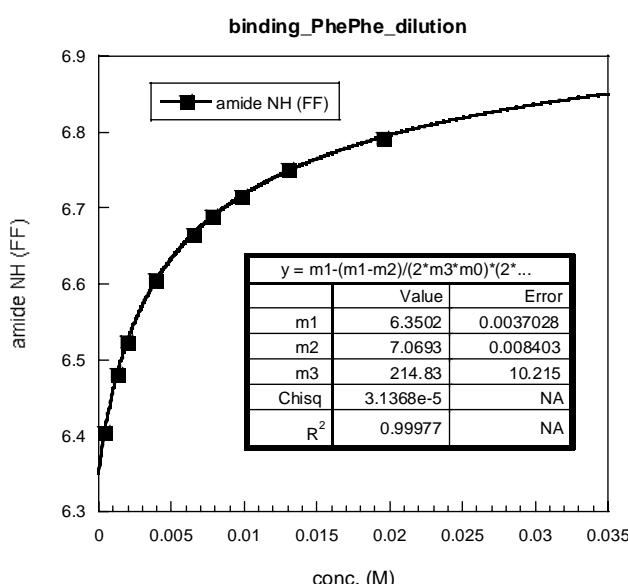
$C_{\text{tot}}$  is the overall concentration of both components (m0, independent variable)

$\delta_o$  is the chemical shift of the unbound species (m1 in the fitting)

$\delta_c$  is the chemical shift of the complex species (m2 in the fitting)

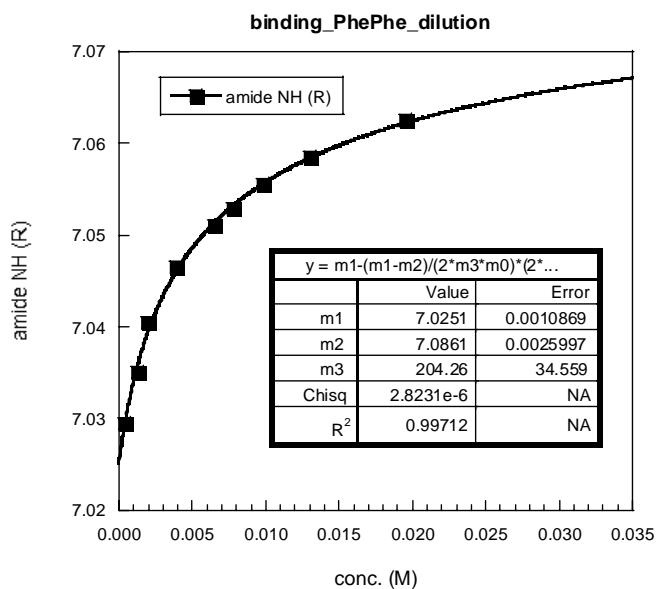
$K_{\text{as}}$  is the host-guest binding constant (m3 in the fitting)

**Figure S23.** Dilution titration of a 1 : 1 mixture of **2a** : Z-FF-OH (500 MHz, 303 K, 1% MeOH in  $\text{CDCl}_3$ ) using the dipeptide amide NH proton signal. The symbols correspond to experimental data and the line to the non-linear least squares fitting. The final values for the best fitting are given in the figure.

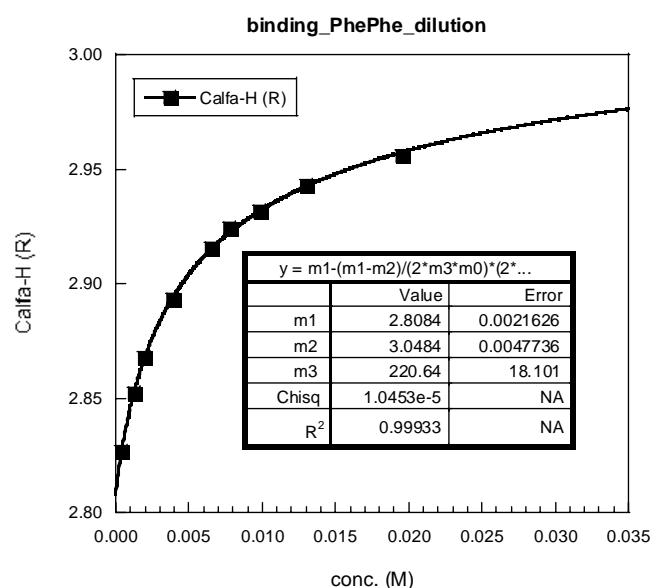


<sup>3</sup> (a) Peters, L.; Fröhlich, R; Boyd, A. S. F.; Kraft, A. *J. Org. Chem.* **2001**, *66*, 3291. (b) Macomber, R. S. *J. Chem. Educ.* **1992**, *69*, 375. (c) Schneider, H. J.; Kramer, R.; Simova, S.; Schneider, U. *J. Am. Chem. Soc.* **1988**, *110*, 6442.

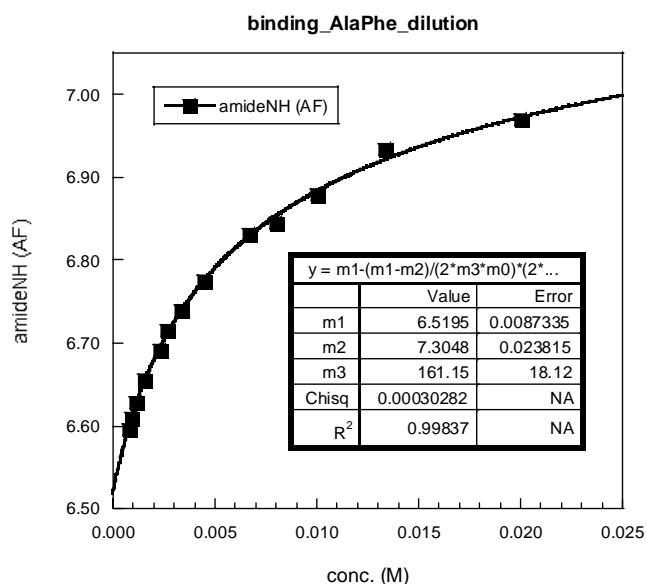
**Figure S24.** Dilution titration of a 1 : 1 mixture of **2a** : Z-FF-OH (500 MHz, 303 K, 1% MeOH in CDCl<sub>3</sub>) using the receptor amide NH proton signal. The symbols correspond to experimental data and the line to the non-linear least squares fitting. The final values for the best fitting are given in the figure.



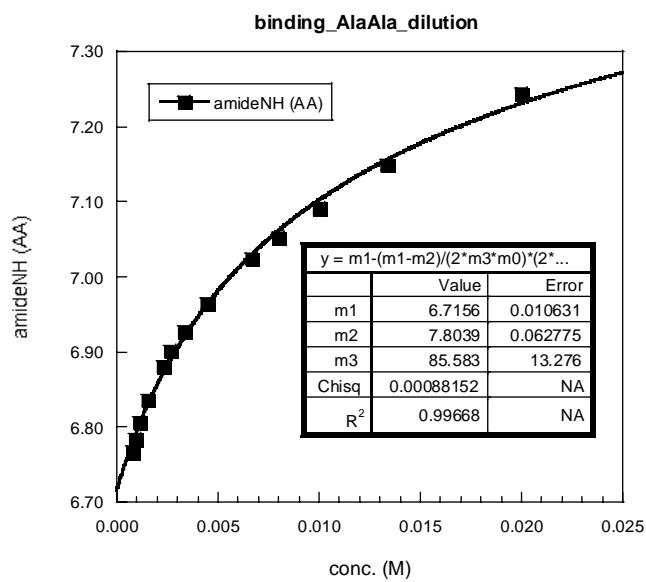
**Figure S25.** Dilution titration of a 1 : 1 mixture of **2a** : Z-FF-OH (500 MHz, 303 K, 1% MeOH in CDCl<sub>3</sub>) using the receptor C $\alpha$  proton signal. The symbols correspond to experimental data and the line to the non-linear squares least fitting. The final values for the best fitting are given in the figure.



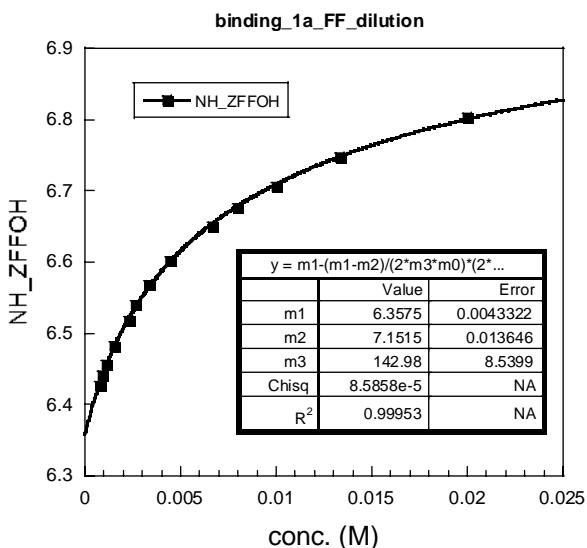
**Figure S26.** Dilution titration of a 1 : 1 mixture of **2a** : Z-AF-OH (500 MHz, 303 K, 1% MeOH in CDCl<sub>3</sub>) using the dipeptide amide NH proton signal. The symbols correspond to experimental data and the line to the non-linear least squares fitting. The final values for the best fitting are given in the figure.



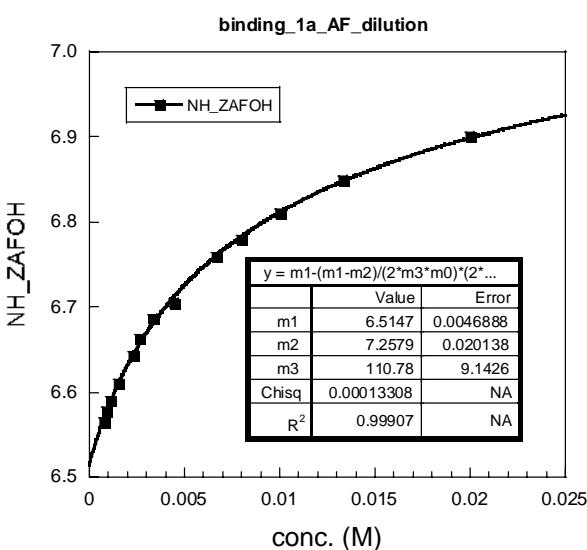
**Figure S27.** Dilution titration of a 1 : 1 mixture of **2a** : Z-AA-OH (500 MHz, 303 K, 1% MeOH in CDCl<sub>3</sub>) using the dipeptide amide NH proton signal. The symbols correspond to experimental data and the line to the non-linear least squares fitting. The final values for the best fitting are given in the figure.

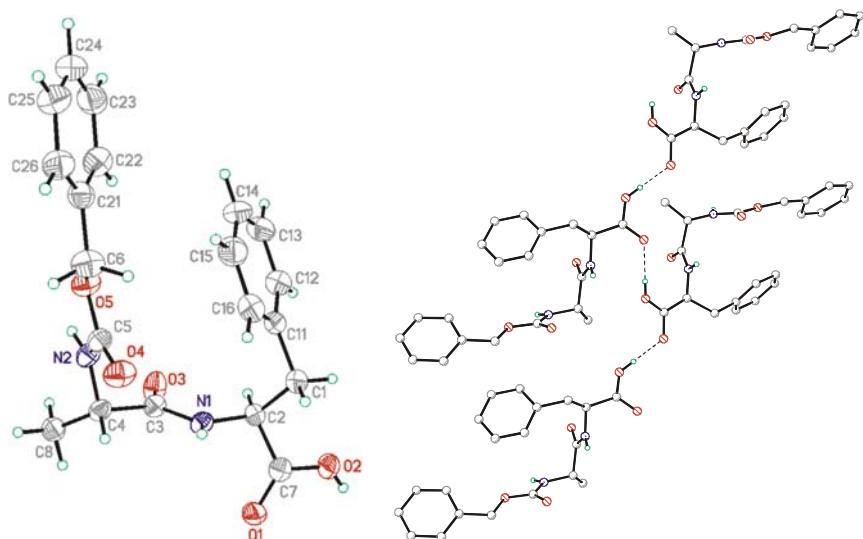


**Figure S28.** Dilution titration of a 1 : 1 mixture of **1a** : Z-FF-OH (500 MHz, 303 K, 1% MeOH in CDCl<sub>3</sub>) using the receptor C $\alpha$  proton signal. The symbols correspond to experimental data and the line to the non-linear squares least fitting. The final values for the best fitting are given in the figure.



**Figure S28.** Dilution titration of a 1 : 1 mixture of **1a** : Z-AF-OH (500 MHz, 303 K, 1% MeOH in CDCl<sub>3</sub>) using the receptor C $\alpha$  proton signal. The symbols correspond to experimental data and the line to the non-linear squares least fitting. The final values for the best fitting are given in the figure.





**Table S1. Crystal data and structure refinement for Z-AF-OH.**

Identification code	nacho1
Empirical formula	C <sub>20</sub> H <sub>22</sub> N <sub>2</sub> O <sub>5</sub>
Formula weight	370.40
Temperature	173(2) K
Wavelength	0.71073 Å
Crystal system	Monoclinic
Space group	C 2
Unit cell dimensions	a = 43.977(3) Å      α = 90°. b = 5.0960(3) Å      β = 95.848(6)°. c = 8.3187(6) Å      γ = 90°.
Volume	1854.6(2) Å <sup>3</sup>
Z	4
Density (calculated)	1.327 Mg/m <sup>3</sup>
Absorption coefficient	0.096 mm <sup>-1</sup>
F(000)	784
Crystal size	0.19 x 0.17 x 0.14 mm <sup>3</sup>
Theta range for data collection	1.86 to 25.00°.
Index ranges	-48<=h<=52, -5<=k<=6, -9<=l<=9
Reflections collected	7601
Independent reflections	1833 [R(int) = 0.0762]
Completeness to theta = 25.00°	99.8 %
Absorption correction	None
Max. and min. transmission	0.9867 and 0.9820
Refinement method	Full-matrix least-squares on F <sup>2</sup>
Data / restraints / parameters	1833 / 1 / 251
Goodness-of-fit on F <sup>2</sup>	1.151
Final R indices [I>2sigma(I)]	R1 = 0.0533, wR2 = 0.1587
R indices (all data)	R1 = 0.0580, wR2 = 0.1607
Absolute structure parameter	1(3)
Extinction coefficient	0.014(3)
Largest diff. peak and hole	0.370 and -0.219 e.Å <sup>-3</sup>

Cartesian coordinates for the minimum structure of [2a · Z-AF-OH]

188		C 2.6930 -1.3430 -6.8940	H -1.0980 1.2620 4.7220
	X Y Z	H 3.2370 -2.0030 -6.2080	N 2.5290 5.0710 3.7990
C -2.3200 5.7600 7.3190		C 1.2120 -1.8440 -7.0330	H 3.5210 4.8700 3.9730
C -1.6110 8.2680 7.0590		H 0.5650 -1.0050 -7.3190	C 1.4990 1.6440 4.7310
C -3.3090 7.6510 8.7590		O 0.0330 -0.5430 -4.7890	O 0.4630 0.9110 4.6910
C -2.9310 8.6640 7.6940		O 3.0780 -0.4590 -4.2180	H 2.0420 4.1490 3.9490
C -3.5330 6.2960 8.1150		C 1.0410 -2.9700 -8.0830	O 1.5470 2.8900 4.4840
C -1.6000 6.8480 6.4450		H 1.3960 -3.9270 -7.6740	C 2.8460 0.9700 5.0230
H -0.8350 8.3600 7.8330		C 1.7840 -2.7260 -9.3840	H 3.5100 1.2280 4.1860
H -2.5240 7.5830 9.5220		H 1.3720 -1.8470 -9.8940	N 2.6630 -0.5110 5.0430
H -3.7140 8.7300 6.9290		C 3.2630 -2.5330 -9.1020	H 2.3540 -0.9250 5.9310
H -4.4120 6.3870 7.4600		H 3.8050 -2.4110 -10.0460	C 2.7750 -1.2600 3.9430
H -2.0650 6.8990 5.4530		C 3.4640 -1.3040 -8.2360	C 2.5040 -2.7480 3.8520
H -1.6120 5.3100 8.0280		C 4.5760 2.5630 -5.6110	H 1.4250 -2.8590 3.7360
H -1.3480 9.0010 6.2850		H 4.1830 2.4720 -6.6320	O 3.0670 -0.6810 2.8530
H -4.2250 7.9730 9.2660		C 5.9600 1.9050 -5.6040	N 3.1860 -3.3270 2.6480
H -2.8380 9.6580 8.1460		H 5.8950 0.8300 -5.8000	H 4.1940 -3.2210 2.5020
H -3.8120 5.5690 8.8900		H 6.4780 2.0490 -4.6510	C 2.8420 -3.5240 5.1030
N -2.9130 4.6080 6.4950		H 6.5910 2.3410 -6.3870	H 3.8470 -3.3100 5.4800
H -3.9410 4.5980 6.4190		C 4.7160 4.0690 -5.3560	H 2.7420 -4.6050 4.9620
C -2.2310 3.5570 6.0140		H 3.7350 4.5510 -5.3110	H 2.0940 -3.2090 5.8460
C -2.8970 2.3160 5.3480		H 5.2800 4.5400 -6.1690	C 3.5080 1.4880 6.2940
H -3.7420 2.6890 4.7560		H 5.2510 4.2840 -4.4260	H 3.4510 2.5840 6.3100
O -0.9760 3.5810 6.1510		C -1.6610 -3.7600 -3.9050	H 4.5810 1.2550 6.2530
C -2.4910 1.3610 0.6190		H -2.4070 -3.9000 -3.1130	C 2.9330 0.9150 7.5630
C -1.4480 -1.1830 0.7930		C -0.7610 -4.9960 -3.8000	C 1.8690 -0.2600 9.8740
C -2.6040 0.4860 -0.4780		H -0.0230 -5.0470 -4.6020	C 3.6830 -0.0130 8.3030
C -1.8700 0.9850 1.8340		H -1.3630 -5.9100 -3.8530	C 1.6530 1.2670 8.0130
C -1.3050 -0.3030 1.8990		H -0.2310 -5.0050 -2.8360	C 1.1250 0.6780 9.1630
C -2.0890 -0.8090 -0.4090		C -2.4510 -3.7450 -5.2140	C 3.1490 -0.5990 9.4500
H -3.1740 0.8250 -1.3450		H -3.1150 -4.6140 -5.2650	H 4.7110 -0.2660 8.0400
H -0.7860 -0.6780 2.8010		H -1.8010 -3.7880 -6.0930	H 1.0620 2.0240 7.5080
N -0.1070 6.5650 6.2560		H -3.0740 -2.8490 -5.2890	H 0.1480 0.9640 9.5480
C 2.0010 5.9900 4.9260		C -3.4570 1.3700 6.4530	H 3.7460 -1.2910 10.0480
C 0.4610 6.0890 5.1340		H -3.9780 1.9890 7.1960	H 1.4740 -0.6960 10.7940
O -0.3100 5.6800 4.2170		C -2.3690 0.5930 7.2020	C 2.5230 -3.7590 1.5780
H 2.3530 5.4550 5.8200		H -1.6340 1.2720 7.6450	O 1.2750 -3.5950 1.5500
C 2.6980 7.3810 4.9460		H -1.8400 -0.1070 6.5480	O 3.2170 -4.0320 0.4260
H 2.4720 7.8250 5.9270		H -2.8090 0.0110 8.0190	C 2.4940 -4.9080 -0.4580
C 2.1790 8.3870 3.9140		C -4.5070 0.3950 5.9050	H 1.4160 -4.6910 -0.4990
C 4.2260 7.2850 4.8780		H -4.0720 -0.3560 5.2400	H 2.6500 -5.9370 -0.1140
H 4.5750 6.9610 3.8930		H -4.9940 -0.1400 6.7280	C 2.9870 -4.7650 -1.8860
H 4.6060 6.5820 5.6260		H -5.2870 0.9320 5.3540	C 3.9160 -4.6620 -4.5440
H 4.6790 8.2610 5.0810		H -1.1150 -2.2150 0.8910	C 3.8000 -3.7070 -2.3190
H 2.6070 9.3770 4.1070		H -2.9520 2.3480 0.5170	C 2.6280 -5.7540 -2.8170
H 1.0910 8.4940 3.9650		H 1.6440 -3.5820 -10.0520	C 3.0880 -5.7030 -4.1350
H 2.4470 8.1210 2.8900		H -0.0260 -3.1110 -8.2990	C 4.2710 -3.6650 -3.6370
C 2.6320 4.3490 1.3180		H 3.6770 -3.4180 -8.6040	H 4.1600 -2.9550 -1.6240
C 3.0890 2.5110 -0.8510		H 4.5360 -1.1740 -8.0380	H 2.0460 -6.6270 -2.5100
C 3.1290 3.0530 1.5560		H 3.1580 -0.4280 -8.8260	H 2.8780 -6.5330 -4.8140
C 2.3740 4.6980 -0.0210		H 0.5110 6.8880 7.0140	H 4.9990 -2.9120 -3.9320
C 2.5910 3.8040 -1.0790		H -0.1220 -2.6300 -2.8380	H 4.3660 -4.6960 -5.5360
C 3.3350 2.1300 0.4910		C -2.3470 -1.8380 -1.5000	N -1.8810 -1.4580 -2.9020
H 3.3790 2.7360 2.5710		H -1.8480 -2.7780 -1.2440	H -2.7430 -1.3750 -3.4730
H 2.0070 5.7000 -0.2660		H -3.4230 -2.0350 -1.5730	N 4.0990 2.0640 -3.1700
H 2.3530 4.1580 -2.0830		C 3.2540 1.5610 -2.0250	H 5.0410 1.6470 -3.0790
H 3.6920 1.1130 0.7470		H 2.2560 1.3400 -2.4220	H 4.2880 3.0680 -3.0070
C 3.5620 1.8850 -4.6410		H 3.7130 0.6200 -1.7010	H -1.5010 -0.5020 -2.8890
H 2.6260 2.4580 -4.6440		C 2.3890 5.4400 2.3620	
C -0.8750 -2.4490 -3.6160		H 3.1190 6.2320 2.1770	
C 3.1770 0.4370 -5.1070		H 1.3750 5.8300 2.2330	
C -0.0760 -1.8060 -4.8050		C -1.8490 2.0100 2.9660	
N 0.6060 -2.4550 -5.7590		H -0.8960 2.5480 2.9660	
H 0.6010 -3.4840 -5.7540		H -2.6480 2.7410 2.7990	
N 2.8440 0.0890 -6.3630		N -2.0620 1.4720 4.3460	
H 2.8540 0.8170 -7.0930		H -2.4970 0.5470 4.2470	

## Output file from the HYDRONMR program

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 HYDRONMR Version 7.C

J. García de la Torre, M. L. Huertas and B. Carrasco,  
 "HYDRONMR: prediction of NMR relaxation of globular proteins  
 from atomic-level structures and hydrodynamic calculations"  
 J. Magnetic Reson. 147, 138-146 (2000).

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 SUMMARY OF DATA AND RESULTS

This file: comp\_AF1.res

Case: comp\_AF1

Structural file: comp\_AF1.pdb

Temperature: 298.0 K

Solvent viscosity: 0.00537 poise

Radius of atomic elements: 2.5 Angs

Translational diffusion coefficient: 4.543E-06 cm<sup>2</sup>/s

Center of diffusion (x): 8.049E-08 cm

Center of diffusion (y): 9.481E-08 cm

Center of diffusion (z): 1.295E-07 cm

Generalized (6x6) diffusion matrix: (Dtt Dtr)  
 (Drt Drr)

4.335E-06	-7.138E-09	-1.327E-07	1.299E-01	-1.848E-01	-2.874E-01
-7.023E-09	4.538E-06	1.781E-07	-1.845E-01	-4.769E-01	-1.174E-01
-1.328E-07	1.782E-07	4.754E-06	-2.874E-01	-1.177E-01	4.969E-01
1.299E-01	-1.845E-01	-2.874E-01	3.232E+08	-2.214E+07	-3.601E+07
-1.848E-01	-4.769E-01	-1.177E-01	-2.214E+07	3.740E+08	8.775E+07
-2.874E-01	-1.174E-01	4.969E-01	-3.601E+07	8.775E+07	4.956E+08

ROTATIONAL DIFFUSION TENSOR

3.232E+08-2.214E+07-3.601E+07

-2.214E+07 3.740E+08 8.775E+07

-3.601E+07 8.775E+07 4.956E+08

Anisotropic rotational diffusion

Eigenvalue (s <sup>-1</sup> )	-- Eigenvector -----
1 3.147E+08	0.9612 0.2690 0.0608
2 3.286E+08	-0.2062 0.8472 -0.4896
3 5.494E+08	-0.1832 0.4581 0.8698

Anisotropic rotational diffusion

Dx, Dy, Dz (s <sup>-1</sup> )	-- Eigenvector -----
Dz 5.494E+08	-0.1832 0.4581 0.8698
Dy 3.147E+08	0.9612 0.2690 0.0608
Dx 3.286E+08	-0.2062 0.8472 -0.4896

Rotational diffusion coefficient: 3.976E+08 s<sup>-1</sup>

Rotational diffusion anisotropy: 2.281E+08 s<sup>-1</sup>

Relaxation time (1): 5.183E-10 s

Relaxation time (2): 4.680E-10 s

Relaxation time (3): 4.590E-10 s

Relaxation time (4): 3.520E-10 s

Relaxation time (5): 3.519E-10 s

Harm. mean relax. (correlation) time: 4.192E-10 s

I flag= 1 N-H bonds, implicit hydrogens (not given in pdb)

----- Calculation for 11.74 Teslas -----  
 NMR DATA AND CONSTANTS

Magnetic field: 11.74 Teslas

Gyromagnetic ratio of 1H : 2.675E+04 rad.s<sup>-1</sup>.G<sup>-1</sup>

Gyromagnetic ratio of X : -2.713E+03 rad.s<sup>-1</sup>.G<sup>-1</sup>

Angular resonance frequency of 1H : 3.140E+09 rad.s<sup>-1</sup>

Resonance frequency of 1H : 500. MHz

Angular resonance frequency of X : 3.185E+08 rad.s<sup>-1</sup>

Internuclear H-X distance : 1.020E-08 cm

Chemical shielding anisotropy : -1.600E-04

Residue-specific T1, T2, RNOE, etc, have been calculated for 0 residues (out of 1), Mean <T1/T2>= NaN NO. IND-CHAIN AMIN T1/T2 NABLA

T1 T2 RNOE TMEAN TINI 1 1 UNK

TABLE OF SPECTRAL DENSITIES

NO.	IND-CHAIN	AMIN	0	WX	WH-WX	WH	WH+WX	0.86WH
			0.000E+00	3.185E+08	2.822E+09	3.140E+09	3.459E+09	2.732E+09
	J(WH+WX)	J(0.86WH)	1	1	UNK	J(WX)	J(WH-WX)	J(WH)