

**Novel Thiourea-Amine Bifunctional Catalysts for Asymmetric
Conjugate Addition of Ketones/Aldehydes to Nitroalkenes: Rational
Structural Combination for High Catalytic Efficiency**

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Experimental Section

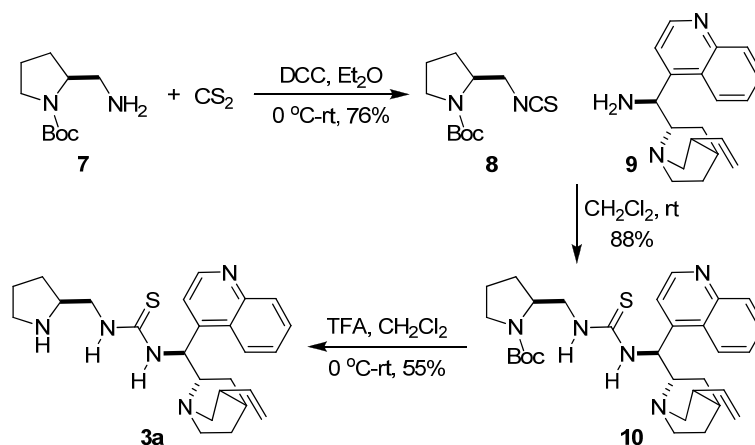
1.0 General Information

Unless otherwise noted, material were purchased from commercial suppliers and used without further purification. Cyclohexanone and dichloromethane were freshly distilled from calcium hydride. n-Hexane, ethyl ether and tetrahydrofuran (THF) were distilled from sodium / benzophenone. Reactions were monitored by thin layer chromatography (TLC), and column chromatography purifications were performed using 200-300 mesh silica gel.

¹H NMR spectra were recorded on 400 MHz or 600 MHz spectrophotometers. Solvent for NMR is CDCl₃, unless the otherwise noted. Chemical shifts are reported in delta (δ) units in parts per million (ppm) relative to the singlet (0 ppm) for tetramethylsilane (TMS). Data are reported as follows: chemical shift, multiplicity (s = single, d = doublet, t = triplet, q = quartet, br = broad, m = multiplet), coupling constants (Hz) and integration. ¹³C NMR spectra were recorded on 100 MHz or 150 MHz. Chemical shifts are reported in ppm relative to the central line of the heptalet at 77.0 ppm for CDCl₃. The ee values determination was carried out using chiral high-performance liquid chromatography (HPLC) with Daicel Chiracel AD-H, OD-H and AS-H column and the dr values were determined by 400 MHz or 600 MHz ¹H NMR. Organocatalys **3-6** were prepared according to the previous procedure¹⁻² from proline-derived amine and cinchona alkaloid-derived amines.³⁻⁴

2.0 Characterization data of catalysts 3-6

General Procedure for Preparing Bifunctional Organocatalysts 3-6. Preparation of 3a is typical.



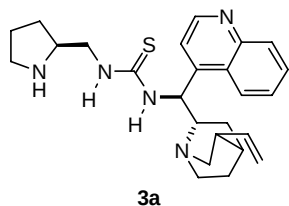
The pure (s)-2-amino-1-N-Boc-pyrrolidine **7** was obtained as a colorless oil according to the reported procedures.^{3a} Carbon bisulfide (6.23 mL, 103.09 mmol) and N,N'-dicyclohexylcarbodiimide (DCC) (3.22 g, 10 mmol) were added to a solution of (s)-2-amino-1-N-Boc-pyrrolidine **7** (3.13 g, 15.62 mmol) in dry ether (50 mL) at 0 °C. The reaction mixture was allowed to warm slowly to room temperature over a period of 3 h and then was stirred for a further 12 h at room temperature. After separation of the precipitated thiourea by filtration, the solvent was removed under reduced pressure. The residue was purified through flash column chromatography on silica gel (eluent, petroleum ether / ethyl acetate = 20:1 to 5:1) to give the corresponding isothiocyante **8** in 76% yield (2.89 g).^{3b} ¹H NMR (400 MHz, CDCl₃, TMS): δ 1.46 (s, 9H), 1.84-1.99 (m, 3H), 2.03-2.14 (m, 1H), 3.37-3.47 (m, 2H), 3.61-3.89 (m, 2H), 3.96-3.97 (m, 1H). ¹³C NMR (100 MHz, CDCl₃): δ 22.8, 23.6, 28.3, 28.6, 29.3, 46.6, 47.0, 47.1, 47.8, 56.3, 79.7, 80.0, 131.0, 153.9, 154.4.

To a solution of the above isothiocyante **8** (2.23 g, 9.2 mmol) in dry dichloromethane (50 mL) was added cinchonidine-derived amine **9** (2.70 g, 9.2 mmol).⁴ The reaction mixture was stirred for 9 h at room temperature

and was concentrated in vacuo. After column chromatography on silica gel (eluent: ethyl acetate / methanol = 10:1 to 5:1 (0.5% v/v Et₃N)), the Boc-protected thiourea-amine **10** as a white solid was isolated in 88 % yield (4.32 g), which was used directly in the next step. The above Boc-protected thiourea-amine **10** was dissolved in a mixture of TFA/CH₂Cl₂ (12 mL/60 mL) and stirred for 12 h at room temperature. The mixture was basified with concentrated ammonia solution and extracted with CH₂Cl₂ (2×40 mL). After the removal of the solvent in vacuo, the residue was first purified through flash column chromatography on silica gel (eluent: methanol / TEA = 10:1), then second purification through a short pad of Na₂SO₄ (eluent: CH₂Cl₂) gave **3a** as a white solid in 55% yield (1.93 g).

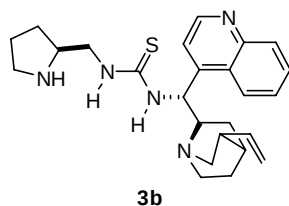
Other thiourea-amine catalysts (**3b-6**) were synthesized according to the above procedures.

Catalyst 3a



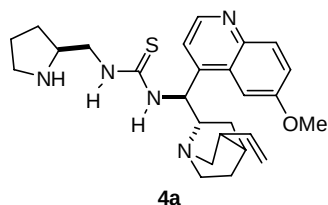
¹H NMR (400 MHz, CDCl₃, TMS): δ 0.81 (s, 1H), 1.14-1.32 (m, 2H), 1.56-1.71 (m, 5H), 2.05 (s, 2H), 2.31 (s, 1H), 2.69-2.91 (m, 4H), 3.15 (s, 1H), 3.25-3.59 (m, 5H), 4.97 (t, J = 14.4 Hz, 2H), 5.69-5.75 (m, 1H), 6.08 (br s, 3H), 7.65 (t, J = 7.8 Hz, 2H), 7.72 (t, J = 7.2 Hz, 1H), 8.10 (d, J = 8.4 Hz, 1H), 8.65 (s, 1H), 8.89 (d, J = 4.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 24.7, 25.4, 27.1, 27.3, 27.9, 39.0, 41.4, 45.1, 45.8, 55.4, 58.9, 114.4, 124.4, 126.6, 127.1, 129.0, 129.8, 140.8, 148.2, 150.0, 182.7; MS (EI) m/z 435 (rel intensity) (M^+); Analysis calculated for C₂₅H₃₃N₅S: C, 68.93, H, 7.64, N, 16.08. Found: C, 68.89; H, 7.61; N, 16.14.

Catalyst 3b



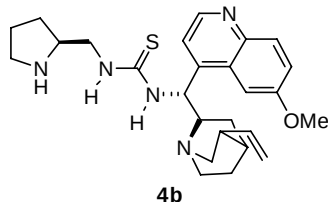
¹H NMR (400 MHz, CDCl₃, TMS): δ 0.75 (s, 1H), 1.09-1.51 (m, 5H), 1.65-1.69 (m, 2H), 1.68-1.89 (m, 1H), 2.22 (q, J = 6.8, 14.8 Hz, 1H), 2.76-2.97 (m, 5H), 3.21-3.79 (m, 4H), 5.15 (t, J = 6.8 Hz, 1H), 5.46 (br s, 2H), 5.80-5.89 (m, 1H), 7.63-7.82 (m, 3H), 8.08 (d, J = 9.6 Hz, 1H), 8.60 (d, J = 8.4 Hz, 1H), 8.90 (d, J = 4.4 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 24.4, 24.7, 25.6, 26.9, 28.3, 39.0, 45.3, 45.9, 46.5, 47.3, 48.8, 59.5, 60.5, 114.9, 119.6, 124.3, 126.5, 127.2, 129.1, 129.7, 147.7, 148.1, 150.0, 183.0; MS (EI) m/z 436 (rel intensity) (M^+ +1); Analysis calculated for C₂₅H₃₃N₅S: C, 68.93, H, 7.64, N, 16.08. Found: C, 68.97; H, 7.69; N, 16.04.

Catalyst 4a



¹H NMR (400 MHz, CDCl₃, TMS): δ 0.74-0.79 (m, 1H), 1.27-1.41 (m, 1H), 1.65-1.78 (m, 7H), 2.33 (s, 1H), 2.80-3.06 (m, 4H), 3.23-3.32 (m, 2H), 3.55-3.87 (m, 4H), 4.03 (s, 3H), 4.90 (d, J = 10.4 Hz, 1H), 4.97 (d, J = 17.2 Hz, 1H), 5.61-5.70 (m, 1H), 6.57 (br s, 3H), 7.37 (dd, J = 2.4, 9.2 Hz, 1H), 7.60 (s, 1H), 7.95 (d, J = 9.2 Hz, 1H), 8.10 (s, 1H), 8.67 (d, J = 3.2 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 24.0, 25.2, 26.9, 27.7, 38.6, 41.8, 45.0, 45.8, 53.7, 55.2, 55.7, 59.1, 60.1, 102.5, 114.8, 119.7, 122.0, 128.1, 131.0, 140.0, 144.4, 145.2, 147.5, 157.7, 182.5; MS (EI) m/z 465 (rel intensity) (M^+); Analysis calculated for C₂₆H₃₅N₅OS: C, 67.06, H, 7.58, N, 15.04. Found: C, 67.09; H, 7.54; N, 15.09.

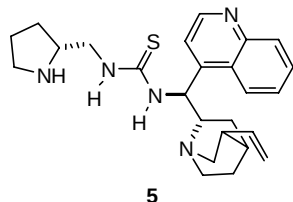
Catalyst 4b



¹H NMR (400 MHz, CDCl₃, TMS): δ 0.81-0.87 (m, 1H), 1.12-1.50 (m, 4H), 1.62-1.79 (m, 2H), 1.86-1.96 (m, 1H), 2.21-2.26 (m, 1H), 2.72-2.95 (m, 5H), 3.33-3.82 (m, 4H), 4.03 (s, 3H), 5.11 (d, J = 10.4 Hz, 1H), 5.19 (d, J = 17.2

Hz, 1H), 5.80-5.89 (m, 1H), 6.57 (br s, 1H), 7.34 (dd, $J = 2.4, 9.2$ Hz, 1H), 7.80 (s, 1H), 7.93 (d, $J = 9.2$ Hz, 1H), 8.03 (s, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 23.8, 24.7, 25.2, 26.5, 27.8, 38.4, 44.8, 45.5, 47.0, 48.6, 52.9, 55.4, 59.9, 102.1, 114.6, 119.6, 122.1, 128.0, 130.6, 139.5, 144.2, 145.5, 147.3, 157.5, 182.7; MS (EI) m/z 465 (rel intensity) (M^+); Analysis calculated for $\text{C}_{26}\text{H}_{35}\text{N}_5\text{OS}$: C, 67.06, H, 7.58, N, 15.04. Found: C, 67.10; H, 7.53; N, 15.08.

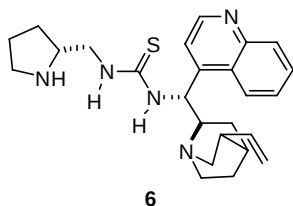
Catalyst 5



^1H NMR (400 MHz, CDCl_3 , TMS): δ 0.85-0.89 (m, 1H), 1.31-1.47 (m, 2H), 1.63-1.76 (m, 5H), 1.90-1.92 (m, 1H), 2.34 (s, 1H), 2.84-2.93 (m, 4H), 3.23-3.27 (m, 1H), 3.54-3.75 (m, 5H), 4.93 (d, $J = 6.8$ Hz, 1H), 4.98 (d, $J = 11.2$ Hz, 1H), 5.60-5.65 (m, 1H), 6.35 (br s, 3H), 7.65 (t, $J = 5.2$ Hz, 1H), 7.72 (t, $J = 4.8$ Hz, 2H), 8.10 (d, $J = 5.6$ Hz, 1H), 8.64 (d, $J = 5.2$ Hz, 1H), 8.90 (d, $J = 3.2$ Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 23.9, 24.5, 24.9, 26.8, 27.8, 38.6, 41.3, 44.8, 45.4,

45.9, 55.0, 59.2, 60.3, 114.5, 119.6, 124.2, 126.4, 127.0, 128.9, 129.6, 140.0, 147.2, 147.9, 149.9, 182.6; MS (EI) m/z 435 (rel intensity) (M^+); Analysis calculated for $\text{C}_{25}\text{H}_{33}\text{N}_5\text{S}$: C, 68.93, H, 7.64, N, 16.08. Found: C, 68.96; H, 7.62; N, 16.03.

Catalyst 6

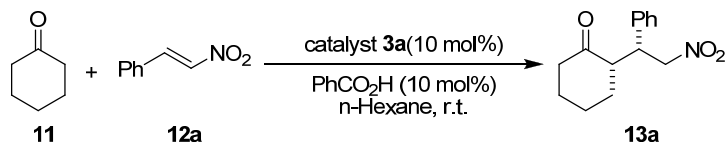


^1H NMR (400 MHz, CDCl_3 , TMS): δ 0.78-0.82 (m, 1H), 1.09-1.20 (m, 2H), 1.42-1.58 (m, 7H), 2.24-2.30 (m, 1H), 2.57-3.15 (m, 10H), 3.91 (br s, 1H), 5.16 (dd, $J = 6.0, 7.2$ Hz, 2H), 5.84-5.93 (m, 1H), 7.48 (s, 1H), 7.63 (t, $J = 7.6$ Hz, 1H), 7.72 (t, $J = 8.0$ Hz, 1H), 8.09 (d, $J = 8.0$ Hz, 1H), 8.53 (s, 1H), 8.82 (s, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 24.6, 25.8, 26.8, 28.1, 38.7, 45.4, 46.8, 48.5, 57.7, 60.2, 114.4, 123.9, 126.1, 126.8, 128.7, 129.4, 139.6, 147.7, 149.6, 182.2; MS (EI)

m/z 435 (rel intensity) (M^+); Analysis calculated for $\text{C}_{25}\text{H}_{33}\text{N}_5\text{S}$: C, 68.93, H, 7.64, N, 16.08. Found: C, 68.99; H, 7.67; N, 16.04.

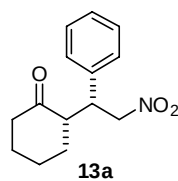
3.0 Characterization data of Michael products 13a-o

Representative procedure for the Asymmetric Michael Reaction of Nitroolefin 12a with Cyclohexanone 11 (Table 3, entry 1).



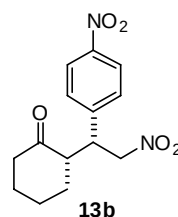
The organocatalyst **3a** (22.0 mg, 0.05 mmol), PhCO₂H (6.1 mg, 0.05 mmol) and cyclohexanone **11** (0.53 mL, 5.0 mmol) were stirred in 1 mL of n-hexane for 10 min at room temperature. *trans*-β-Styrene **12a** (75.0 mg, 0.5 mmol) was then added and the reaction mixture was stirred for 5 h. After the complete consumption of **12a** (as monitored by TLC). The reaction mixture was concentrated under reduced pressure, and the residue was purified by flash column chromatography on silica gel (petroleum ether / ethyl acetate (4:1 to 2:1)) to give the corresponding pure Michael product **13a** (110.0 mg, 89%) as a white solid. Relative and absolute configurations of the products were determined by comparison of ¹H NMR, ¹³C NMR spectra and HPLC data with the known literature. Compounds **13a-o** and **15** are known.^{1-2,5}

2-(2-nitro-1-phenylethyl)cyclohexanone (**13a**), (Table 3, entry 1).



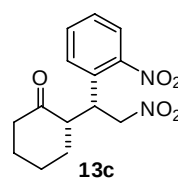
The ee was determined by HPLC (Chiralpak AS column, hexane/*i*-PrOH 75:25, flow rate 1.0 mL/min, λ = 210 nm, 20 °C). *t_R* (minor) = 9.3 min; *t_R* (major) = 13.3 min. *syn/anti* = 97/3, *syn*: ee = 96%. ¹H NMR (400 MHz, CDCl₃, TMS): δ 1.18-1.28 (m, 1H), 1.51-1.61 (m, 1H), 1.66-1.81 (m, 3H), 2.05-2.11 (m, 1H), 2.35-2.43 (m, 1H), 2.45-2.50 (m, 1H), 2.65-2.72 (m, 1H), 3.73-3.79 (m, 1H), 4.62 (dd, *J* = 10.0, 12.4 Hz, 1H), 4.95 (dd, *J* = 4.4, 12.4 Hz, 1H), 7.16-7.18 (m, 2H), 7.24-7.34 (m, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 24.8, 28.3, 33.0, 42.5, 43.7, 52.2, 78.7, 127.5, 127.9, 128.7, 137.5, 211.9.

2-(2-nitro-1-(4-nitrophenyl)ethyl)cyclohexanone (**13b**), (Table 3, entry 2).



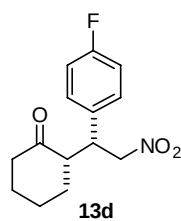
The ee was determined by HPLC (Chiralpak AD column, hexane/*i*-PrOH 75:25, flow rate 1.0 mL/min, λ = 254 nm, 20 °C). *t_R* (minor) = 13.1 min; *t_R* (major) = 28.9 min, *syn/anti* = 93/7, *syn*: ee = 94%. ¹H NMR (600 MHz, CDCl₃, TMS): δ 1.24-1.30 (m, 1H), 1.57-1.71 (m, 4H), 1.81-1.84 (m, 1H), 2.11-2.13 (m, 1H), 2.36-2.42 (m, 1H), 2.48-2.50 (m, 1H), 2.70-2.75 (m, 1H), 3.91-3.95 (m, 1H), 4.70 (dd, *J* = 10.8, 12.6 Hz, 1H), 5.00 (dd, *J* = 4.2, 13.2 Hz, 1H), 7.40 (d, *J* = 8.4 Hz, 2H), 8.20 (d, *J* = 8.4 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 24.9, 28.1, 33.0, 42.5, 43.6, 52.0, 77.9, 123.9, 129.2, 145.5, 147.2, 210.8.

2-(2-nitro-1-(2-nitrophenyl)ethyl)cyclohexanone (**13c**), (Table 3, entry 3).



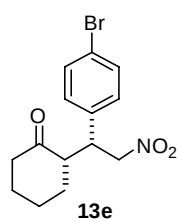
The ee was determined by HPLC (Chiralpak AS column, hexane/*i*-PrOH 80:20, flow rate 1.0 mL/min, λ = 254 nm, 20 °C). *t_R* (minor) = 23.2 min; *t_R* (major) = 29.8 min, *syn/anti* = 98/2, *syn*: ee = 95%. ¹H NMR (600 MHz, CDCl₃, TMS): δ 1.45-1.51 (m, 1H), 1.60-1.73 (m, 2H), 1.80-1.85 (m, 2H), 2.10-2.13 (m, 1H), 2.34-2.41 (m, 1H), 2.46-2.49 (m, 1H), 2.92-2.96 (m, 1H), 4.31-3.35 (m, 1H), 4.92 (d, *J* = 6.6 Hz, 2H), 7.42-7.47 (m, 2H), 7.59 (t, *J* = 7.8 Hz, 1H), 7.84 (d, *J* = 8.4 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 25.2, 28.2, 33.0, 41.8, 42.7, 52.1, 77.6, 124.7, 128.4, 129.0, 132.7, 133.0, 150.7, 211.0.

2-(1-(4-fluorophenyl)-2-nitroethyl)cyclohexanone (13d) , (Table 3, entry 4).



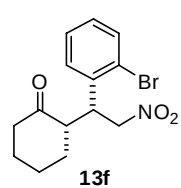
The ee was determined by HPLC (Chiralpak OD column, hexane/*i*-PrOH 95:5, flow rate 1.0 mL/min, λ = 210 nm, 20 °C). t_R (minor) = 18.8 min; t_R (major) = 20.8 min, *syn/anti* = 97/3, *syn*: ee = 94%. ^1H NMR (600 MHz, CDCl_3 , TMS): δ 1.20-1.26 (m, 1H), 1.55-1.73 (m, 2H), 1.79-1.87 (m, 1H), 2.07-2.10 (m, 1H), 2.33-2.41 (m, 1H), 2.46-2.48 (m, 1H), 2.63-2.67 (m, 1H), 3.75-3.79 (m, 1H), 4.60 (t, J = 10.8 Hz, 1H), 4.93 (dd, J = 4.2, 12.0 Hz, 1H), 7.01 (t, J = 8.4 Hz, 2H), 7.14-7.16 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 24.9, 28.3, 33.0, 42.6, 43.1, 52.4, 78.7, 115.6, 115.8, 129.6, 129.7, 133.4, 133.5, 160.7, 163.2, 211.6.

2-(1-(4-bromophenyl)-2-nitroethyl)cyclohexanone (13e) , (Table 3, entry 5).



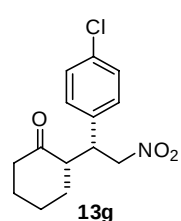
The ee was determined by HPLC (Chiralpak AD column, hexane/*i*-PrOH 90:10, flow rate 1.0 mL/min, λ = 210 nm, 20 °C). t_R (minor) = 14.0 min; t_R (major) = 22.9 min, *syn/anti* = 94/6, *syn*: ee = 94%. ^1H NMR (600 MHz, CDCl_3 , TMS): δ 1.19-1.26 (m, 1H), 1.56-1.73 (m, 3H), 1.78-1.81 (m, 1H), 2.07-2.10 (m, 1H), 2.34-2.40 (m, 1H), 2.46-2.48 (m, 1H), 2.62-2.67 (m, 1H), 3.72-3.76 (m, 1H), 4.60 (dd, J = 10.2, 12.6 Hz, 1H), 4.93 (dd, J = 4.2, 12.6 Hz, 1H), 7.06 (d, J = 8.4 Hz, 2H), 7.45 (d, J = 8.4 Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 24.9, 28.3, 33.0, 42.6, 43.3, 52.2, 78.4, 121.6, 129.8, 131.9, 136.7, 211.4.

2-(1-(2-bromophenyl)-2-nitroethyl)cyclohexanone (13f) , (Table 3, entry 6).



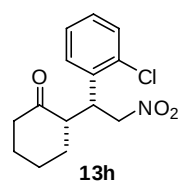
The ee was determined by HPLC (Chiralpak AD column, hexane/*i*-PrOH 85:15, flow rate 1.0 mL/min, λ = 210 nm, 20 °C). t_R (minor) = 8.9 min; t_R (major) = 14.3 min, *syn/anti* = 98/2, *syn*: ee = 90%. ^1H NMR (600 MHz, CDCl_3 , TMS): δ 1.37-1.39 (m, 1H), 1.61-1.76 (m, 3H), 1.81-1.87 (m, 1H), 2.09-2.12 (m, 1H), 2.35-2.41 (m, 1H), 2.46-2.48 (m, 1H), 2.90 (s, 1H), 4.30 (s, 1H), 4.86-4.94 (m, 2H), 7.13 (t, J = 7.2 Hz, 1H), 7.21 (d, J = 7.2 Hz, 1H), 7.29 (t, J = 7.2 Hz, 1H), 7.57 (d, J = 7.8 Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 25.0, 28.3, 32.7, 41.7, 42.5, 52.0, 77.2, 127.8, 128.8, 133.3, 137.1, 211.3.

2-(1-(4-chlorophenyl)-2-nitroethyl)cyclohexanone (13g) , (Table 3, entry 7).



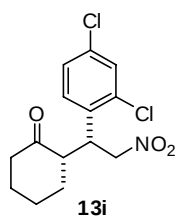
The ee was determined by HPLC (Chiralpak AD column, hexane/*i*-PrOH 90:10, flow rate 1.0 mL/min, λ = 210 nm, 20 °C). t_R (minor) = 13.2 min; t_R (major) = 20.0 min, *syn/anti* = 93/7, *syn*: ee = 94%. ^1H NMR (600 MHz, CDCl_3 , TMS): δ 1.19-1.26 (m, 1H), 1.53-1.59 (m, 1H), 1.63-1.73 (m, 2H), 1.78-1.81 (m, 1H), 2.07-2.11 (m, 1H), 2.32-2.40 (m, 1H), 2.45-2.48 (m, 1H), 2.62-2.67 (m, 1H), 3.74-3.78 (m, 1H), 4.60 (dd, J = 9.6, 12.0 Hz, 1H), 4.93 (dd, J = 4.2, 12.6 Hz, 1H), 7.11 (d, J = 7.8 Hz, 2H), 7.30 (d, J = 8.4 Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 24.9, 28.3, 33.0, 42.6, 43.2, 52.2, 78.5, 129.0, 129.4, 133.4, 136.2, 211.4.

2-(1-(2-chlorophenyl)-2-nitroethyl)cyclohexanone (13h) , (Table 3, entry 8).



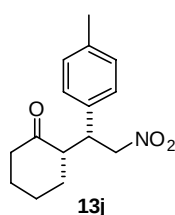
The ee was determined by HPLC (Chiralpak AD column, hexane/*i*-PrOH 95:5, flow rate 1.0 mL/min, λ = 210 nm, 20 °C). t_R (minor) = 13.8 min; t_R (major) = 24.3 min, *syn/anti* = 97/3, *syn*: ee = 94%. ^1H NMR (600 MHz, CDCl_3 , TMS): δ 1.30-1.36 (m, 1H), 1.57-1.73 (m, 3H), 1.80-1.87 (m, 1H), 2.08-2.11 (m, 1H), 2.32-2.41 (m, 1H), 2.46-2.48 (m, 1H), 2.91 (s, 1H), 4.29 (d, J = 7.8 Hz, 1H), 4.89 (d, J = 6.6 Hz, 2H), 7.19-7.25 (m, 3H), 7.37 (d, J = 7.8 Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 25.0, 28.3, 32.7, 40.7, 42.5, 51.5, 77.1, 127.2, 128.6, 129.2, 130.0, 134.3, 135.3, 211.5.

2-(1-(2,4-dichlorophenyl)-2-nitroethyl)cyclohexanone (13i) , (Table 3, entry 9).



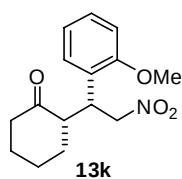
The ee was determined by HPLC (Chiralpak AD column, hexane/*i*-PrOH 90:10, flow rate 1.0 mL/min, λ = 210 nm, 20 °C). t_R (minor) = 10.1 min; t_R (major) = 14.6 min, *syn/anti* = 95/5, *syn*: ee = 95%. ^1H NMR (600 MHz, CDCl_3 , TMS): δ 1.30-1.36 (m, 1H), 1.59-1.75 (m, 3H), 1.82-1.87 (m, 1H), 2.09-2.12 (m, 1H), 2.37-2.40 (m, 1H), 2.46-2.48 (m, 1H), 2.87 (s, 1H), 4.24 (dd, J = 7.2, 15.6 Hz, 1H), 4.88 (d, J = 6.6 Hz, 2H), 7.18 (d, J = 8.4 Hz, 1H), 7.23 (d, J = 8.4 Hz, 1H), 7.40 (s, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 25.2, 27.0, 28.4, 32.9, 40.5, 42.7, 51.6, 77.4, 127.7, 130.0, 133.9, 134.2, 135.2, 211.3.

2-(2-nitro-1-*p*-tolylethyl)cyclohexanone (13j) , (Table 3, entry 10).



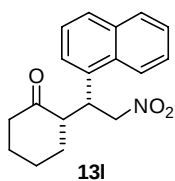
The ee was determined by HPLC (Chiralpak AS column, hexane/*i*-PrOH 90:10, flow rate 0.8 mL/min; λ = 210 nm, 20 °C). t_R (minor) = 15.9 min; t_R (major) = 28.1 min, *syn/anti* = 94/6, *syn*: ee = 92%. ^1H NMR (600 MHz, CDCl_3 , TMS): δ 1.19-1.26 (m, 1H), 1.53-1.59 (m, 1H), 1.64-1.79 (m, 3H), 2.05-2.09 (m, 1H), 2.31 (s, 3H), 2.35-2.40 (m, 1H), 2.45-2.48 (m, 1H), 2.64-2.68 (m, 1H), 3.69-3.73 (m, 1H), 4.60 (dd, J = 10.2, 12.0 Hz, 1H), 4.92 (dd, J = 4.8, 12.6 Hz, 1H), 7.04 (d, J = 8.4 Hz, 2H), 7.12 (d, J = 7.8 Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 20.9, 24.9, 28.4, 33.0, 42.6, 43.4, 52.4, 78.9, 127.9, 129.4, 134.5, 137.2, 211.9.

2-(1-(2-methoxyphenyl)-2-nitroethyl)cyclohexanone (13k) , (Table 3, entry 11).



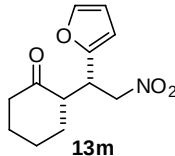
The ee was determined by HPLC (Chiralpak AS column, hexane/*i*-PrOH 90:10, flow rate 1.0 mL/min, λ = 210 nm, 20 °C). t_R (minor) = 15.3 min; t_R (major) = 17.7 min, *syn/anti* = 95/5, *syn*: ee = 92%. ^1H NMR (400 MHz, CDCl_3 , TMS): δ 1.15-1.25 (m, 1H), 1.51-1.52 (m, 1H), 1.66-1.87 (m, 3H), 2.03-2.09 (m, 1H), 2.31-2.42 (m, 1H), 2.44-2.48 (m, 1H), 2.94-3.01 (m, 1H), 3.84 (s, 3H), 3.93-3.99 (m, 1H), 4.82 (dd, J = 2.4, 5.2 Hz, 2H), 6.88 (t, J = 7.6 Hz, 2H), 7.08 (dd, J = 1.6, 7.6 Hz, 1H), 7.22-7.26 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 24.9, 28.3, 33.1, 41.1, 42.5, 50.3, 55.1, 110.8, 120.6, 125.1, 128.7, 130.7, 157.4, 212.3.

2-(1-(naphthalen-1-yl)-2-nitroethyl)cyclohexanone (13l) , (Table 3, entry 12).



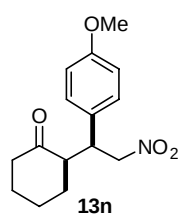
The ee was determined by HPLC (Chiralpak AS column, hexane/*i*-PrOH 50:50, flow rate 0.7 mL/min, λ = 254 nm, 20 °C). t_R (minor) = 12.0 min; t_R (major) = 17.2 min, *syn/anti* = 93/7, *syn*: ee = 92%. ^1H NMR (400 MHz, CDCl_3 , TMS): δ 1.20-1.30 (m, 1H), 1.47-1.56 (m, 1H), 1.62-1.71 (m, 3H), 2.06-2.11 (m, 1H), 2.38-2.53 (m, 1H), 2.85 (s, 1H), 4.77 (s, 1H), 4.91 (t, J = 10.0 Hz, 1H), 5.08 (dd, J = 4.4, 12.8 Hz, 1H), 7.37-7.58 (m, 4H), 7.78 (d, J = 8.4 Hz, 1H), 7.86 (d, J = 8.0 Hz, 1H), 8.18 (d, J = 7.2 Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 25.0, 28.5, 33.1, 36.6, 42.7, 53.6, 78.6, 122.6, 123.4, 125.2, 125.7, 126.4, 127.9, 128.8, 132.3, 133.8, 134.5, 212.1.

2-(1-(furan-2-yl)-2-nitroethyl)cyclohexanone (13m) , (Table 3, entry 13).



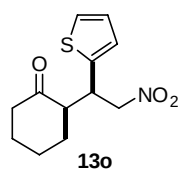
The ee was determined by HPLC (Chiralpak AD column, hexane/*i*-PrOH 90:10, flow rate 1.0 mL/min, λ = 210 nm, 20 °C). t_R (major) = 11.1 min; t_R (minor) = 13.5 min, *syn/anti* = 92/8, *syn*: ee = 85%. ^1H NMR (600 MHz, CDCl_3 , TMS): δ 1.25-1.32 (m, 1H), 1.62-1.66 (m, 2H), 1.74-1.77 (m, 1H), 1.83-1.85 (m, 1H), 2.09-2.11 (m, 1H), 2.34-2.39 (m, 1H), 2.45-2.47 (m, 1H), 2.73-2.77 (m, 1H), 3.95-3.99 (m, 1H), 4.67 (dd, J = 9.6, 12.6 Hz, 1H), 4.79 (dd, J = 4.8, 12.6 Hz, 1H), 6.18 (d, J = 3.0 Hz, 1H), 6.28 (dd, J = 1.8, 3.0 Hz, 1H), 7.34 (s, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 24.9, 28.0, 32.3, 37.4, 42.3, 50.9, 76.5, 108.7, 110.1, 142.1, 150.8, 210.8.

2-(1-(4-methoxyphenyl)-2-nitroethyl)cyclohexanone (13n) , (Table 3, entry 25).



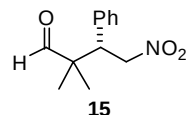
The ee was determined by HPLC (Chiralpak AD column, hexane/*i*-PrOH 75:25, flow rate 0.7 mL/min, λ = 210 nm, 20 °C). t_R (major) = 11.8 min; t_R (minor) = 14.3 min, *syn/anti* = 89/11, *syn*: ee = -85%. ^1H NMR (400 MHz, CDCl_3 , TMS): δ 1.17-1.18 (m, 1H), 1.51-1.58 (m, 1H), 1.65-1.81 (m, 3H), 2.04-2.11 (m, 1H), 2.33-2.41 (m, 1H), 2.44-2.49 (m, 1H), 2.61-2.67 (m, 1H), 3.68-3.74 (m, 1H), 3.77 (s, 3H), 4.58 (dd, J = 10.0, 12.4 Hz, 1H), 4.90 (dd, J = 4.4, 12.0 Hz, 1H), 6.84 (d, J = 8.8 Hz, 2H), 7.08 (d, J = 8.4 Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3): δ 24.8, 28.4, 33.0, 42.5, 43.0, 52.5, 55.0, 78.9, 114.1, 129.0, 129.4, 158.8, 211.9.

2-(2-nitro-1-(thiophen-2-yl)ethyl)cyclohexanone (13o) , (Table 3, entry 26).



The ee was determined by HPLC (Chiralpak AS column, hexane/*i*-PrOH 90:10, flow rate 1.0 mL/min, λ = 210 nm, 20 °C). t_R (major) = 21.0 min; t_R (minor) = 29.2 min, *syn/anti* = 85/15, *syn*: ee = -82%. ^1H NMR (400 MHz, CDCl_3 , TMS): δ 1.27-1.43 (m, 1H), 1.60-1.73 (m, 2H), 1.83-1.93 (m, 2H), 2.04-2.12 (m, 1H), 2.32-2.39 (m, 1H), 2.42-2.48 (m, 1H), 2.64-2.71 (m, 1H), 4.10-4.16 (m, 1H), 4.65 (dd, J = 9.2, 12.4 Hz, 1H), 4.89 (dd, J = 4.8, 12.8 Hz, 1H), 6.87 (d, J = 2.8 Hz, 1H), 6.93 (dd, J = 3.6, 5.2 Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 24.9, 28.1, 32.6, 39.2, 42.4, 53.1, 79.1, 124.8, 126.4, 126.7, 140.3, 211.1.

(S)-2,2-dimethyl-4-nitro-3-phenylbutanal (15) , (Eq 1).



The ee was determined by HPLC (Chiralpak OD column, hexane/*i*-PrOH 80:20, flow rate 0.8 mL/min, λ = 210 nm, 20 °C). t_R (major) = 17.6 min; t_R (minor) = 25.8 min, ee = 85%. ^1H NMR (600 MHz, CDCl_3 , TMS): δ 1.01 (s, 3H), 1.14 (s, 3H), 3.78 (dd, J = 3.0, 10.8 Hz, 1H), 4.69 (dd, J = 3.6, 13.2 Hz, 1H), 4.85 (t, J = 12.6 Hz, 1H), 7.20 (d, J = 7.2 Hz, 2H), 7.29-7.34 (m, 3H), 9.53 (s, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 18.5, 21.4, 48.1, 76.1, 127.9, 128.5, 128.9, 135.1, 204.2.

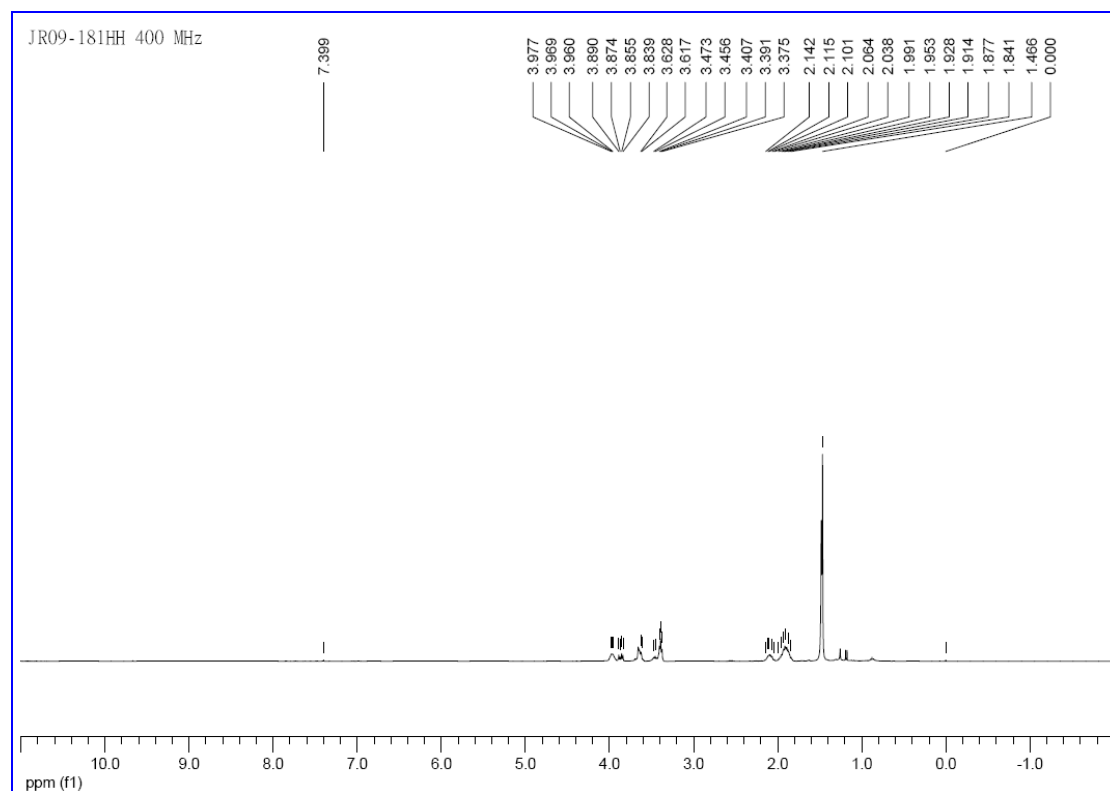
4.0 References

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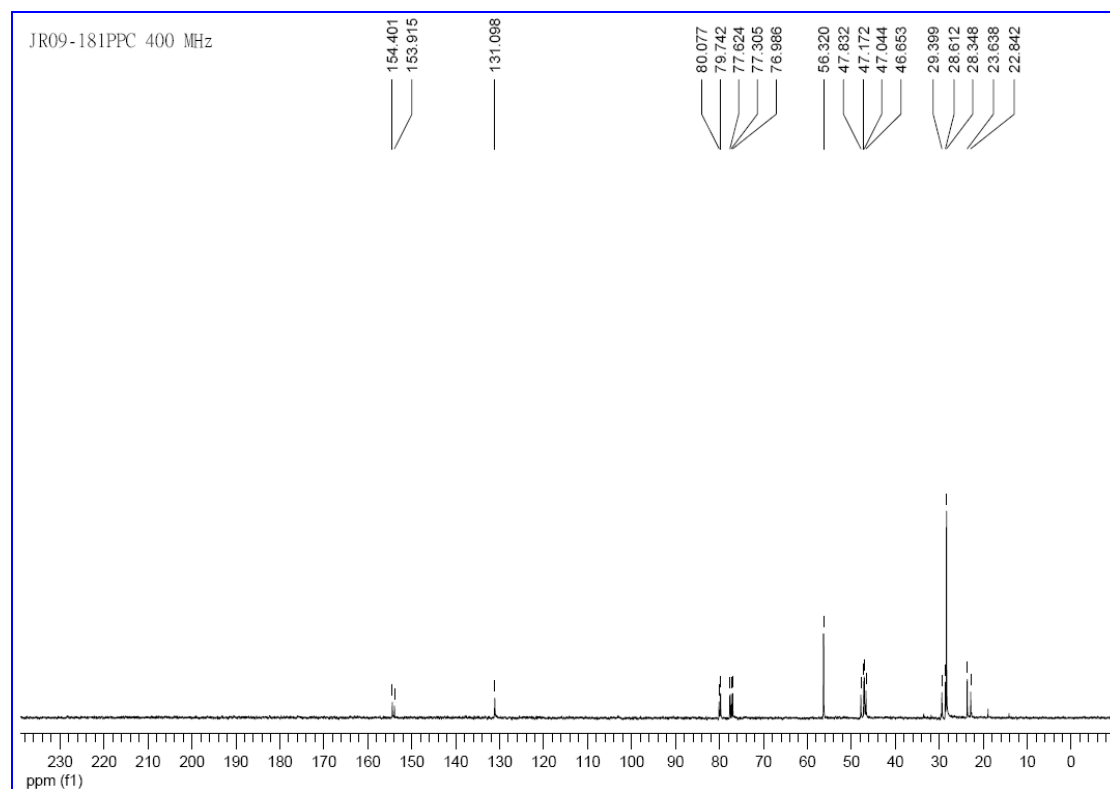
5.0 NMR Spectra

Catalyst NMR Spectra

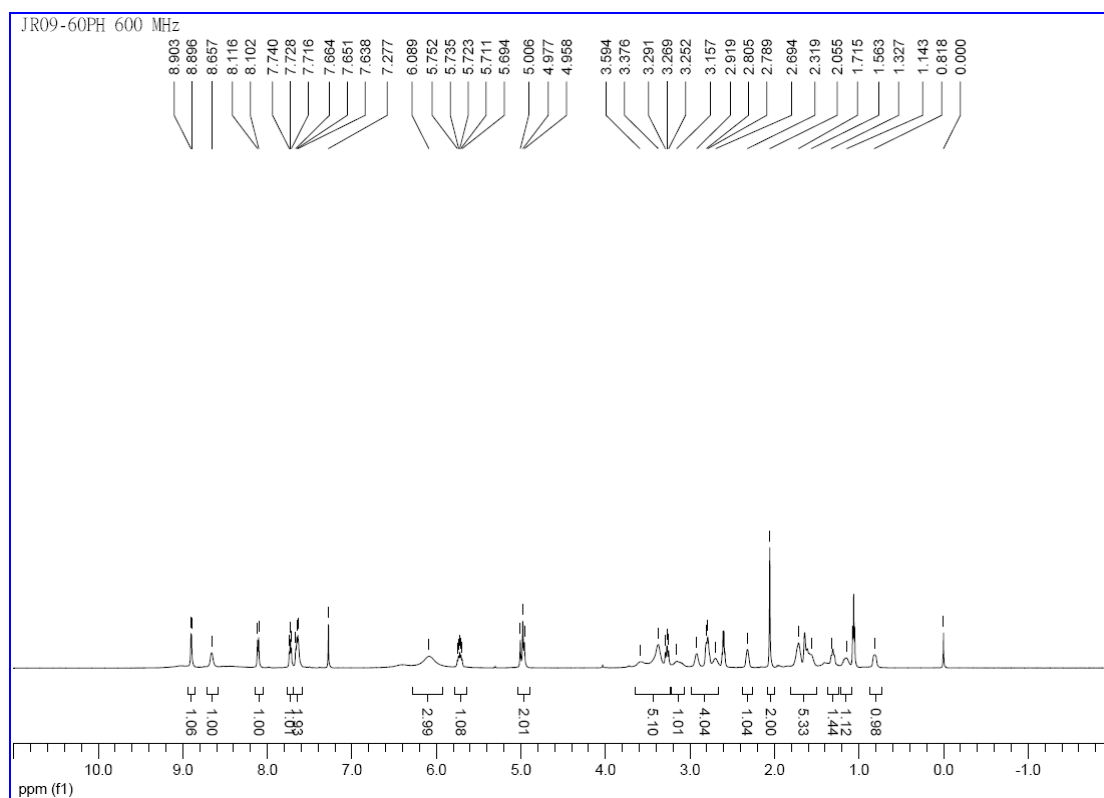
^1H NMR (400 MHz, CDCl_3) spectrum of isothiocyanate **8**



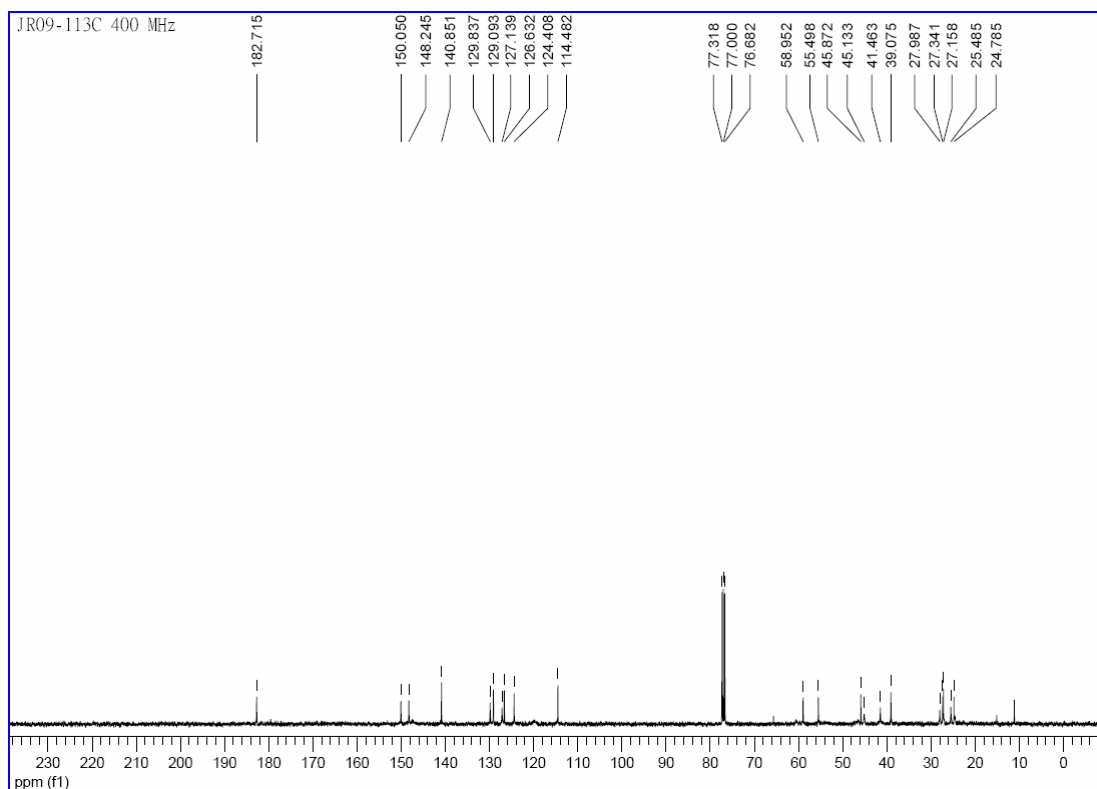
^{13}C NMR spectrum (100 MHz, CDCl_3) of isothiocyanate **8**



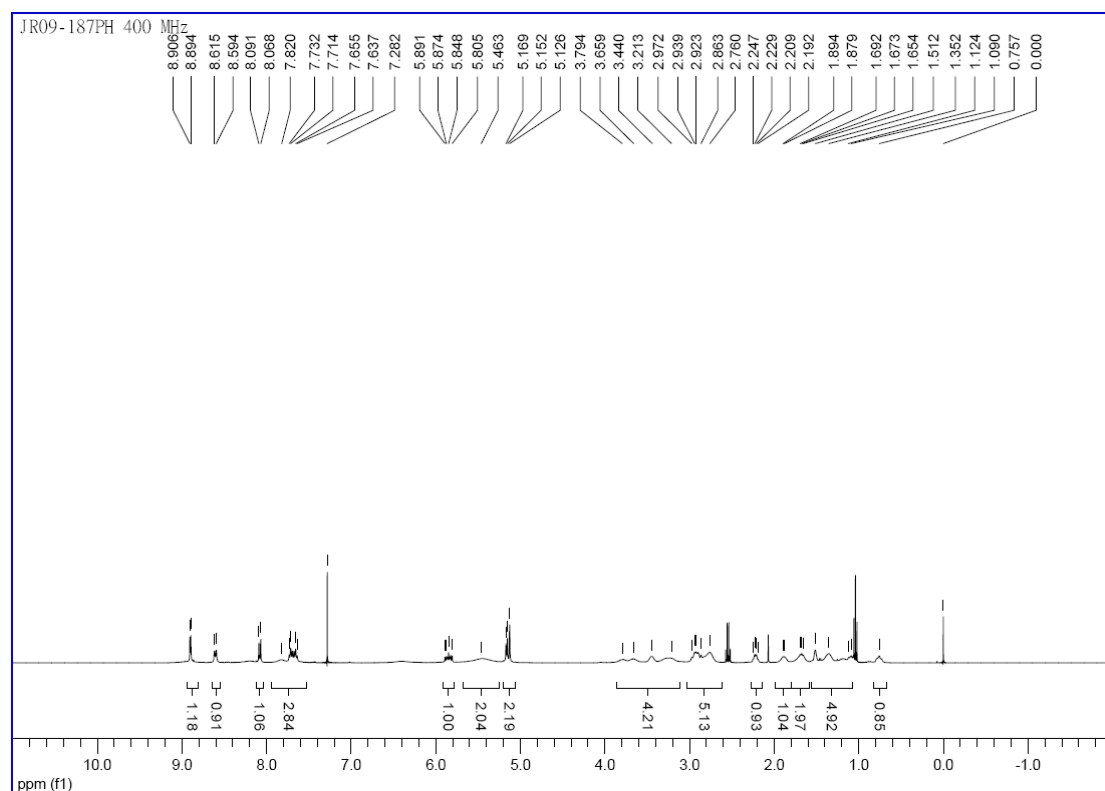
¹H NMR (600 MHz, CDCl₃) spectrum of catalyst 3a



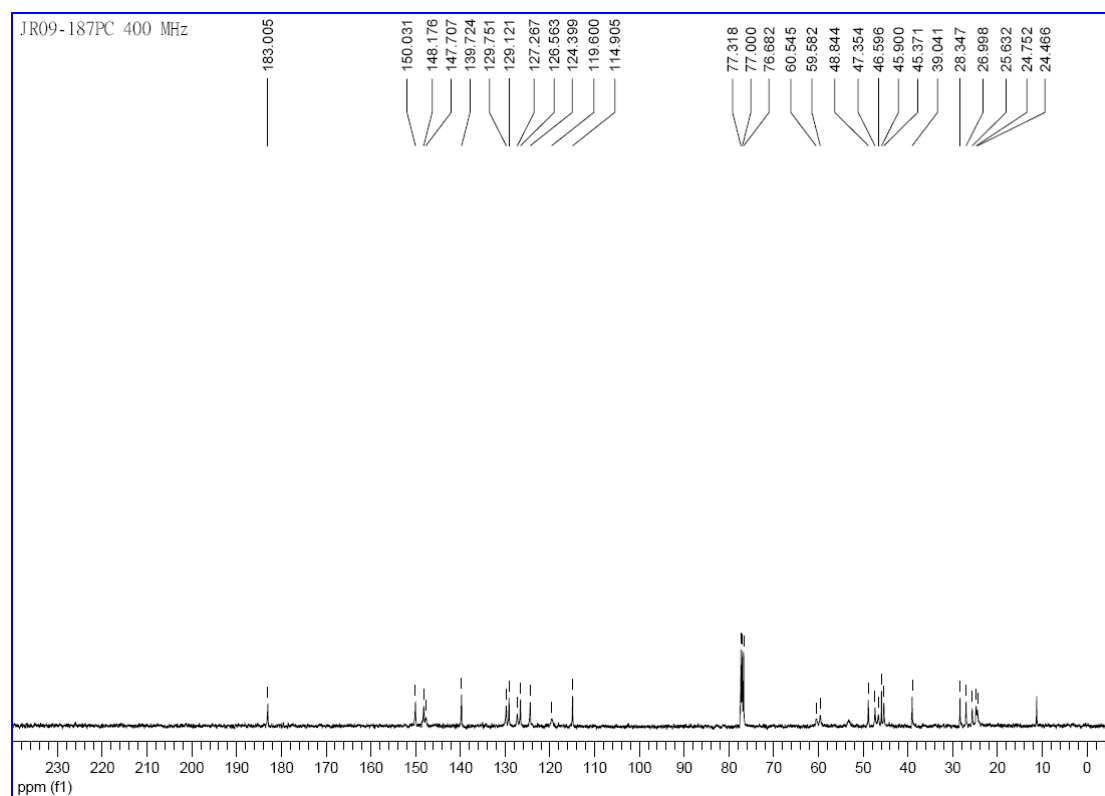
¹³C NMR spectrum (100 MHz, CDCl₃) of catalyst 3a



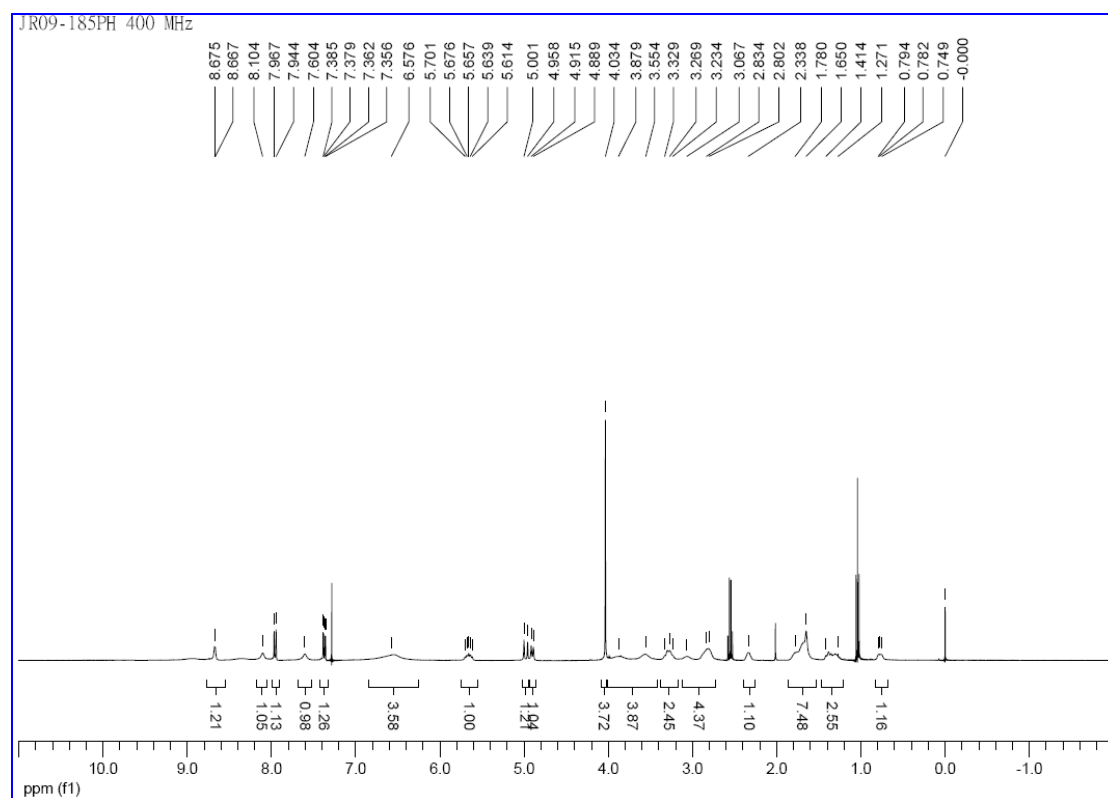
^1H NMR (400 MHz, CDCl_3) spectrum of catalyst 3b



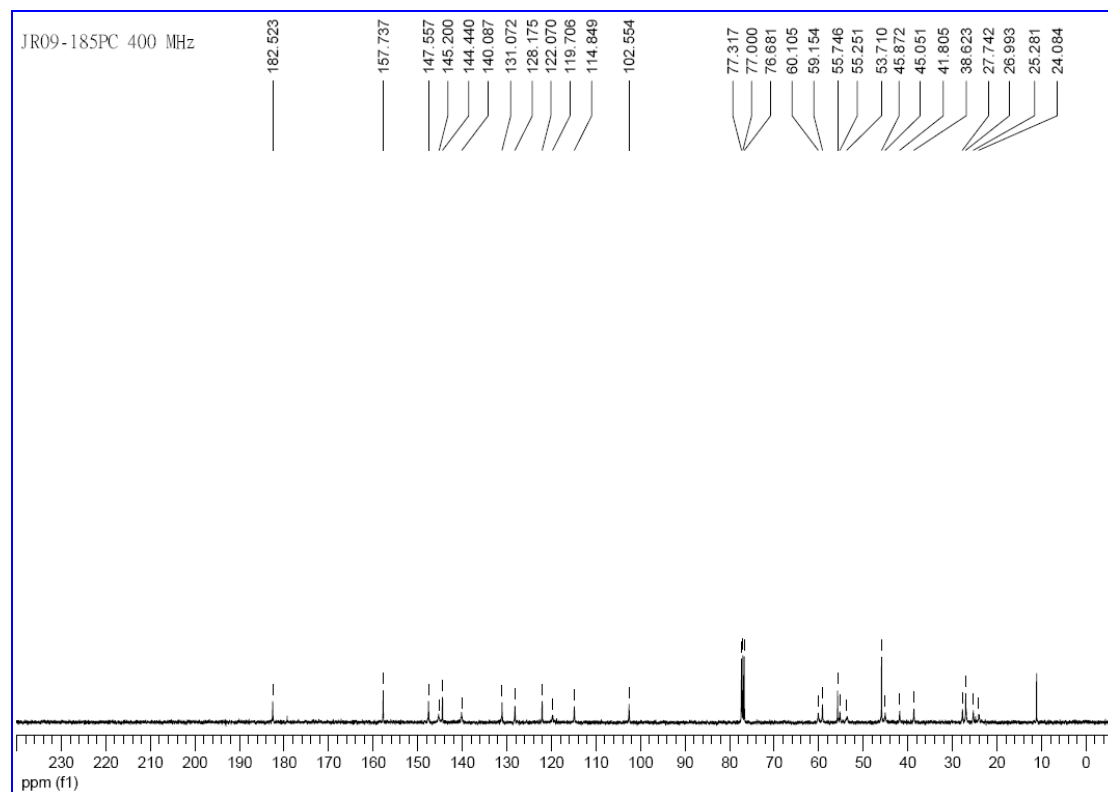
^{13}C NMR spectrum (100 MHz, CDCl_3) of catalyst 3b



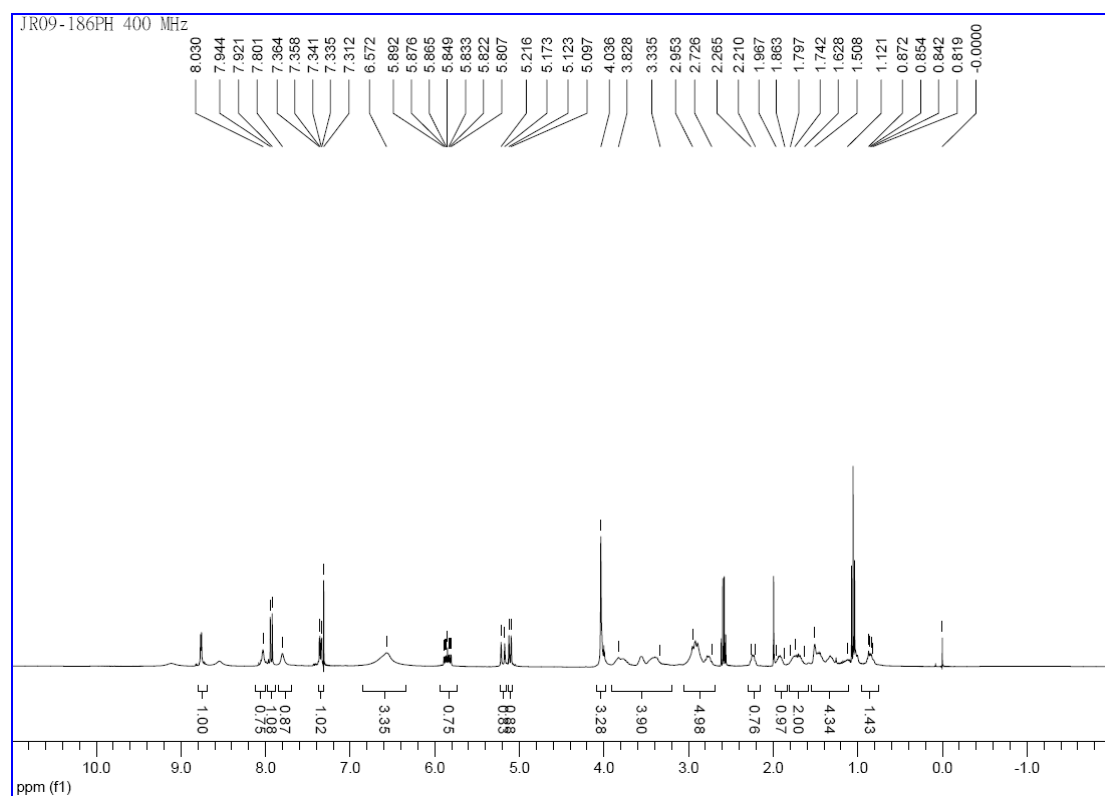
^1H NMR (400 MHz, CDCl_3) spectrum of catalyst 4a



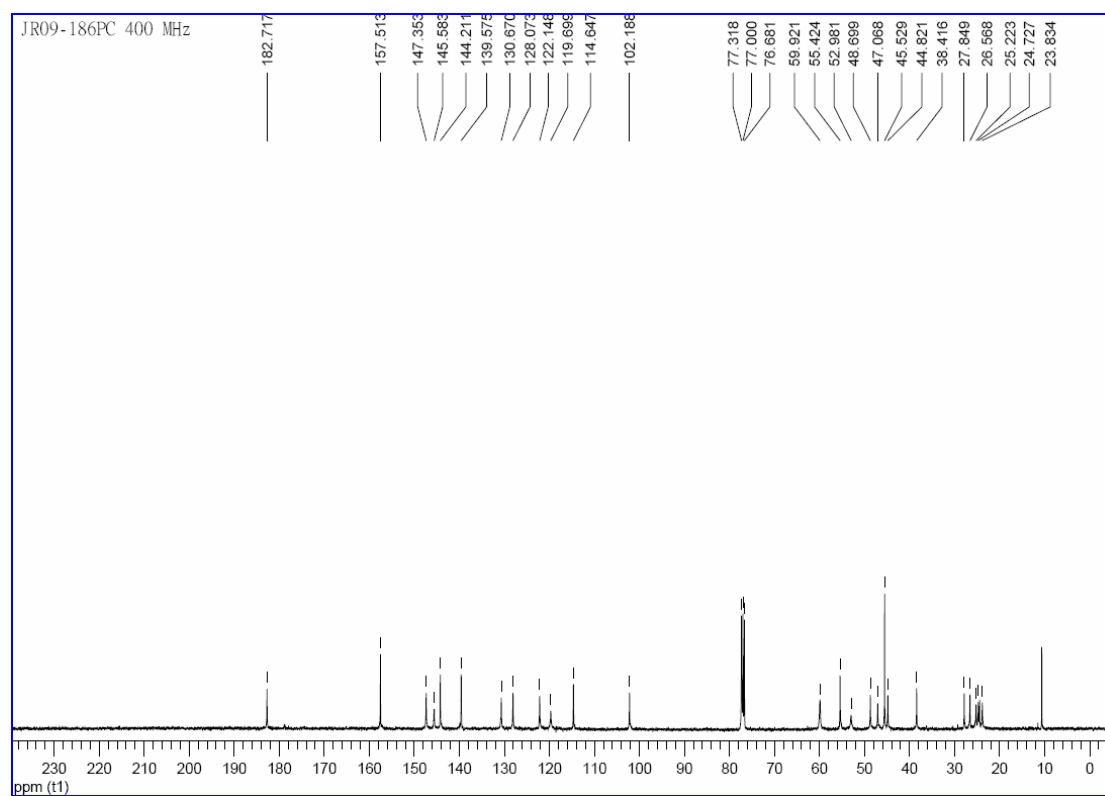
^{13}C NMR spectrum (100 MHz, CDCl_3) of catalyst 4a



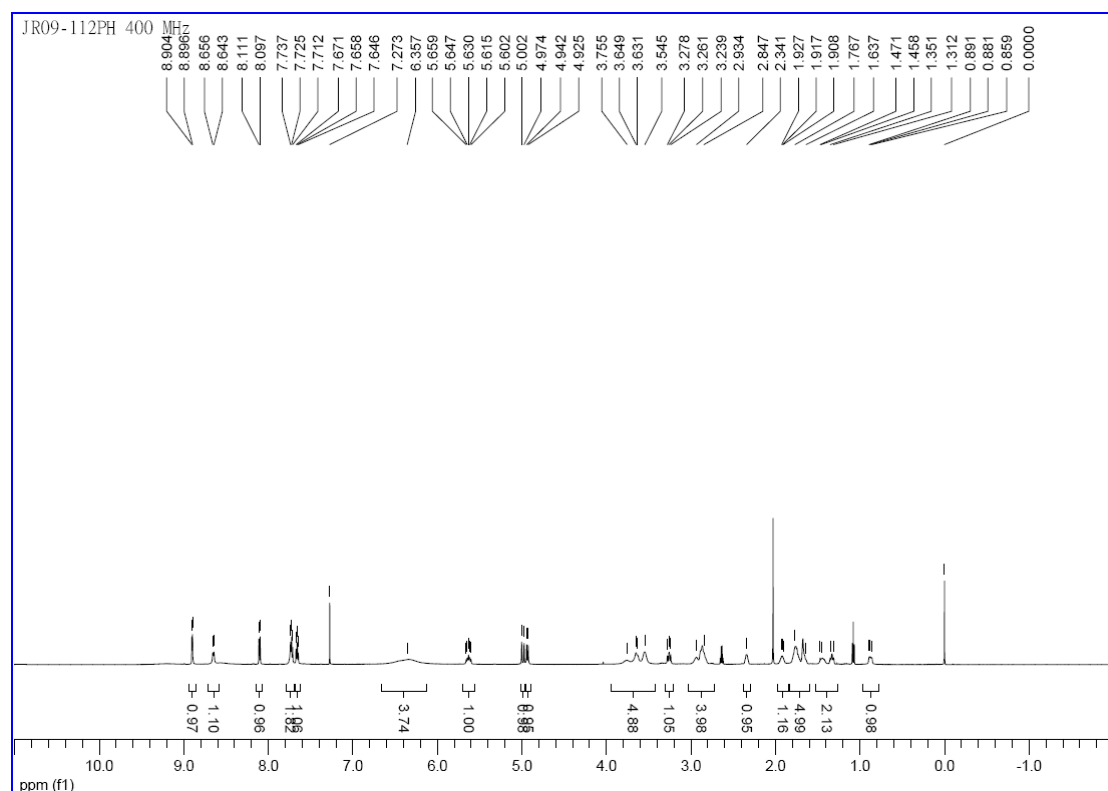
^1H NMR (400 MHz, CDCl_3) spectrum of catalyst 4b



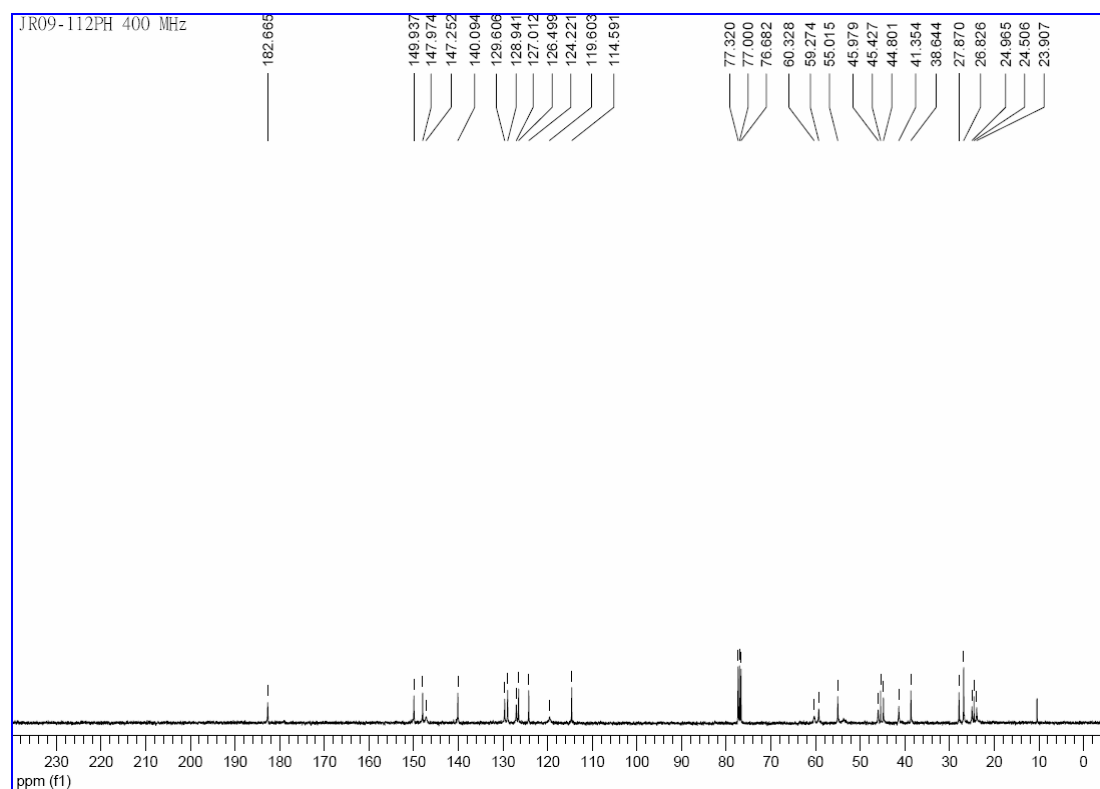
^{13}C NMR spectrum (100 MHz, CDCl_3) of catalyst 4b



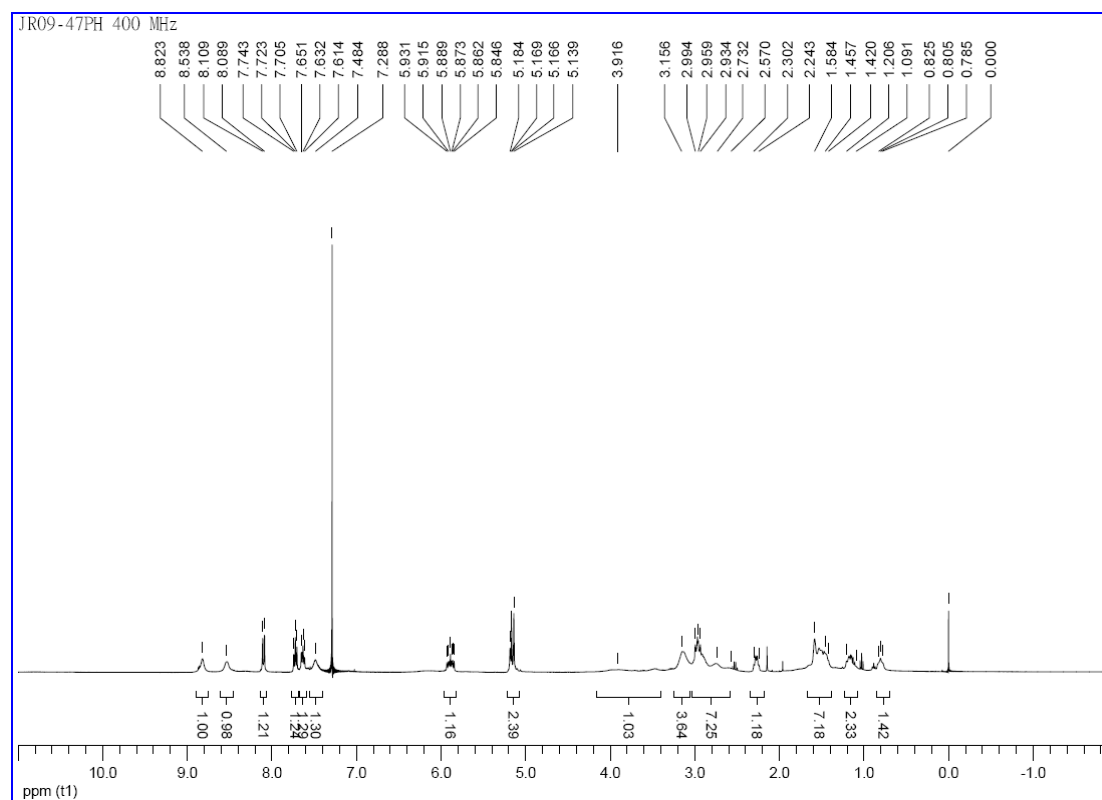
^1H NMR (400 MHz, CDCl_3) spectrum of catalyst 5



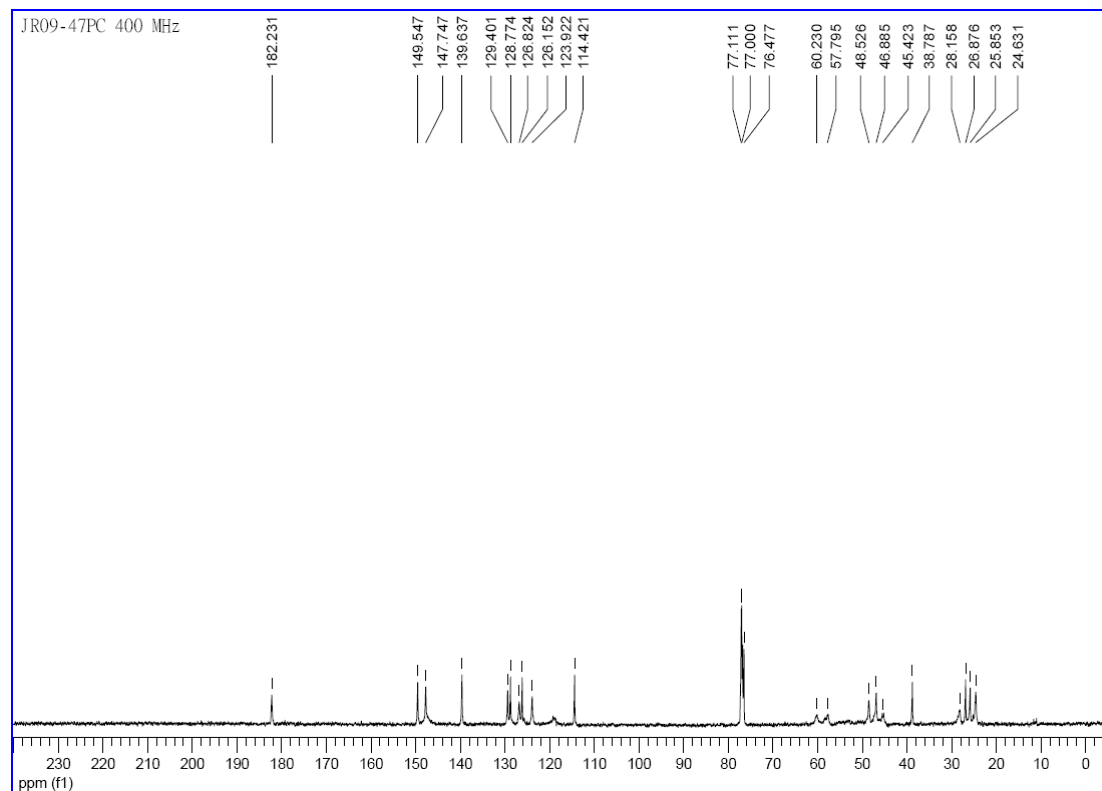
^{13}C NMR spectrum (100 MHz, CDCl_3) of catalyst 5



^1H NMR (400 MHz, CDCl_3) spectrum of catalyst 6

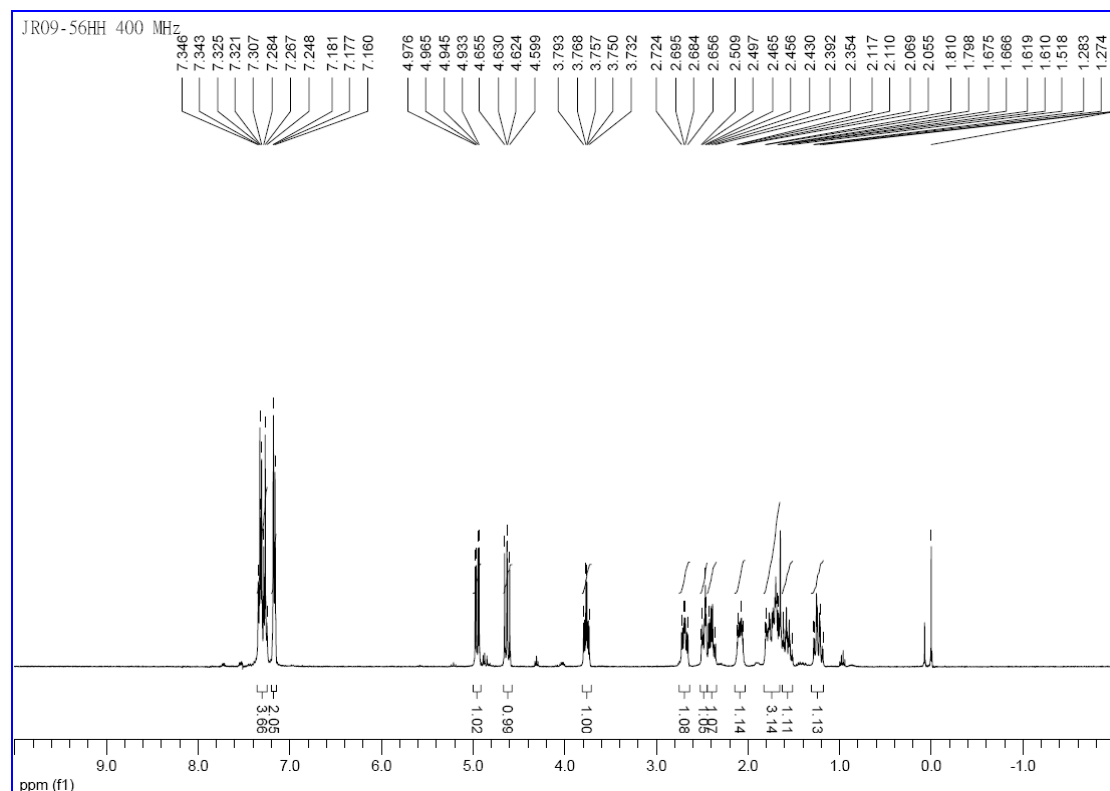


^{13}C NMR spectrum (100 MHz, CDCl_3) of catalyst 6

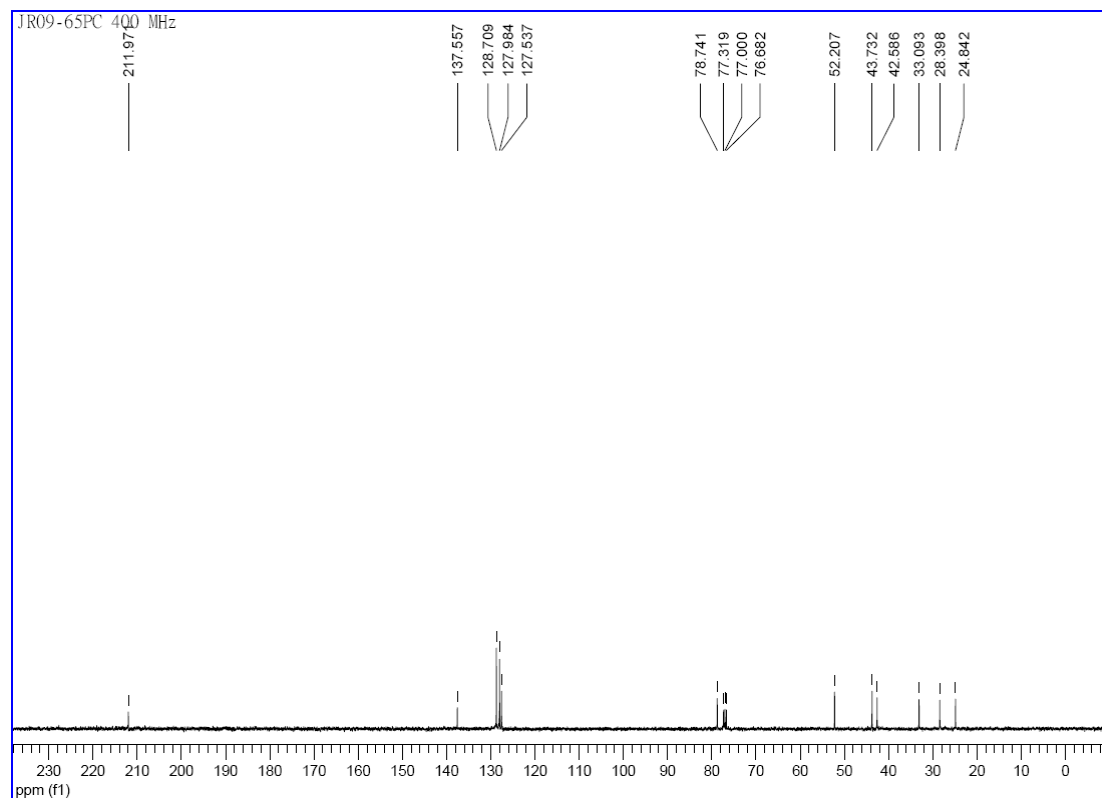


Michael product NMR Spectra

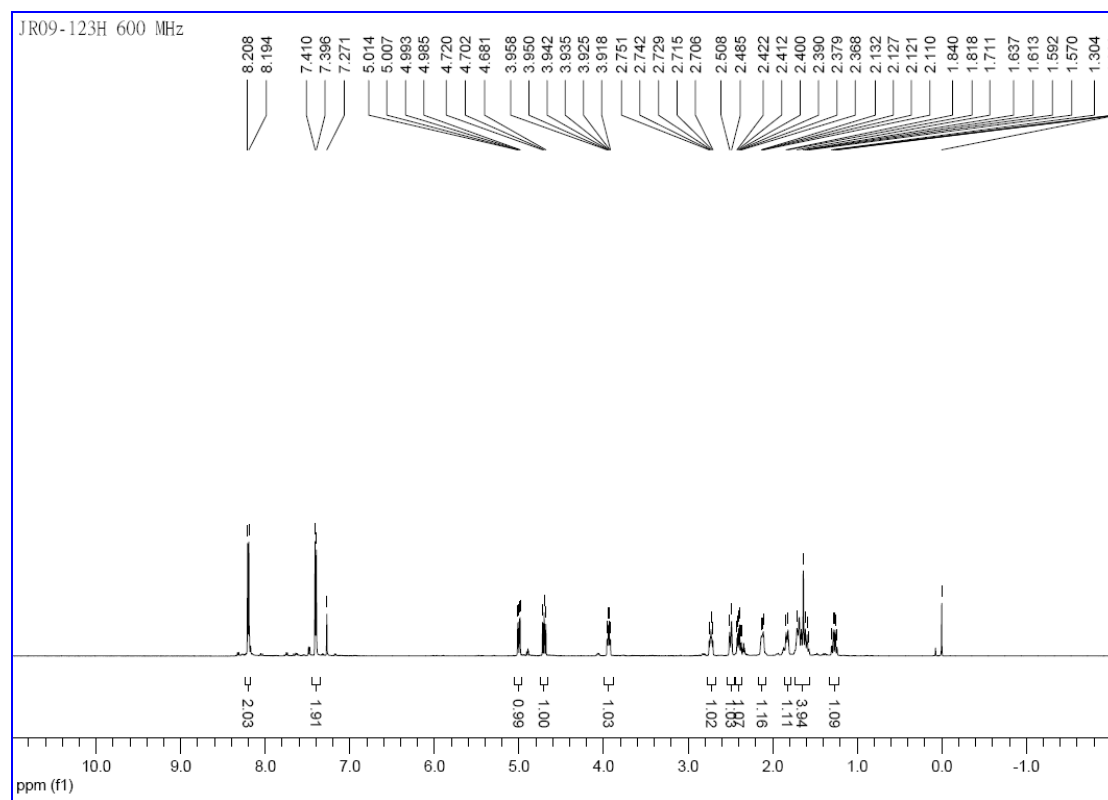
^1H NMR (400 MHz, CDCl_3) spectrum of Michael product 13a



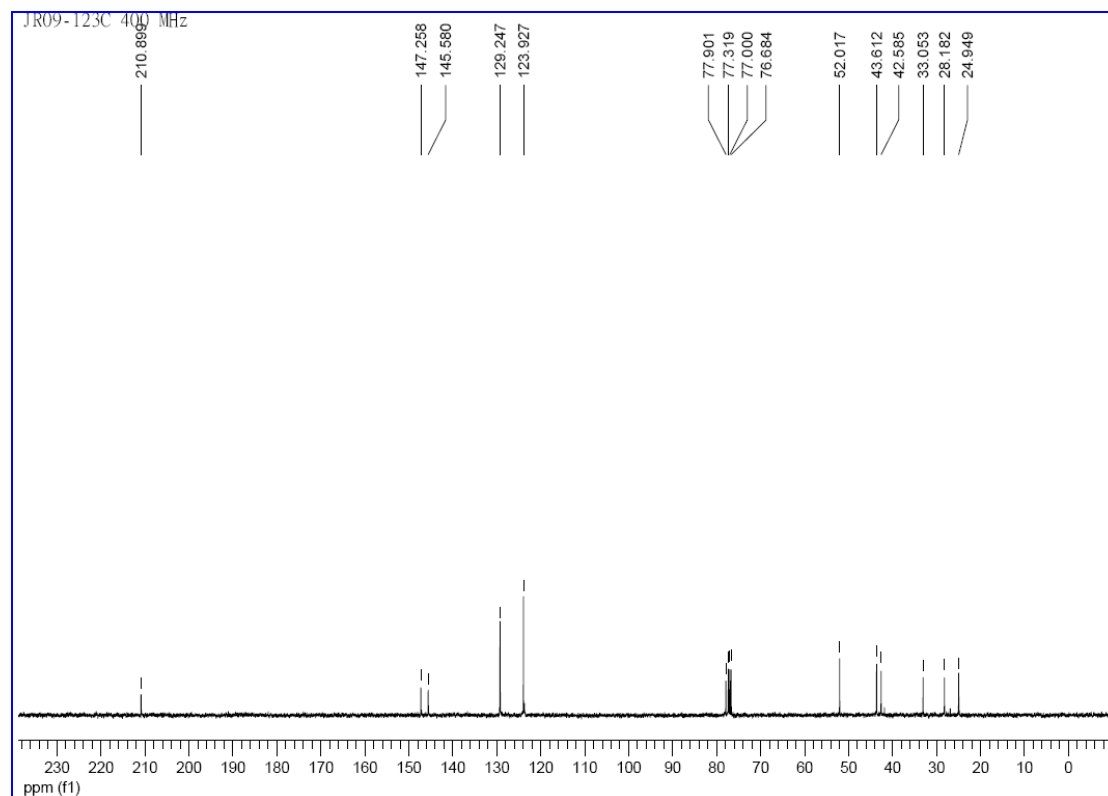
^{13}C NMR spectrum (100 MHz, CDCl_3) of Michael product 13a



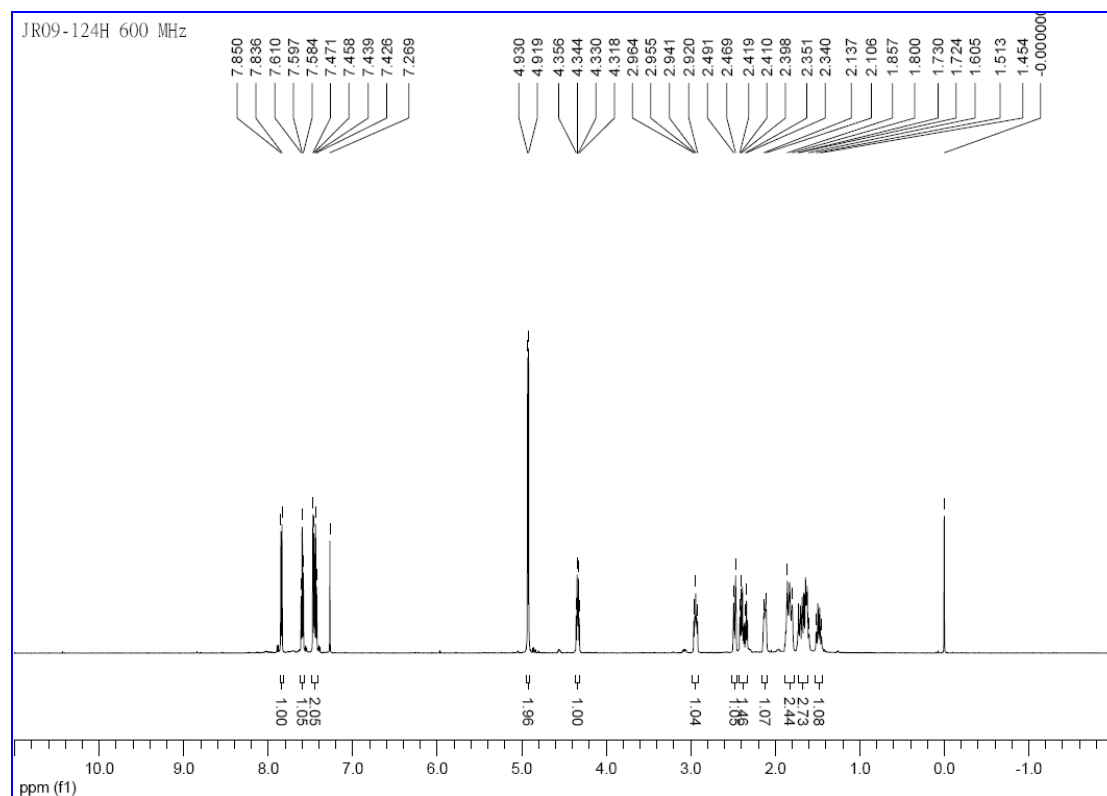
^1H NMR (600 MHz, CDCl_3) spectrum of Michael product 13b



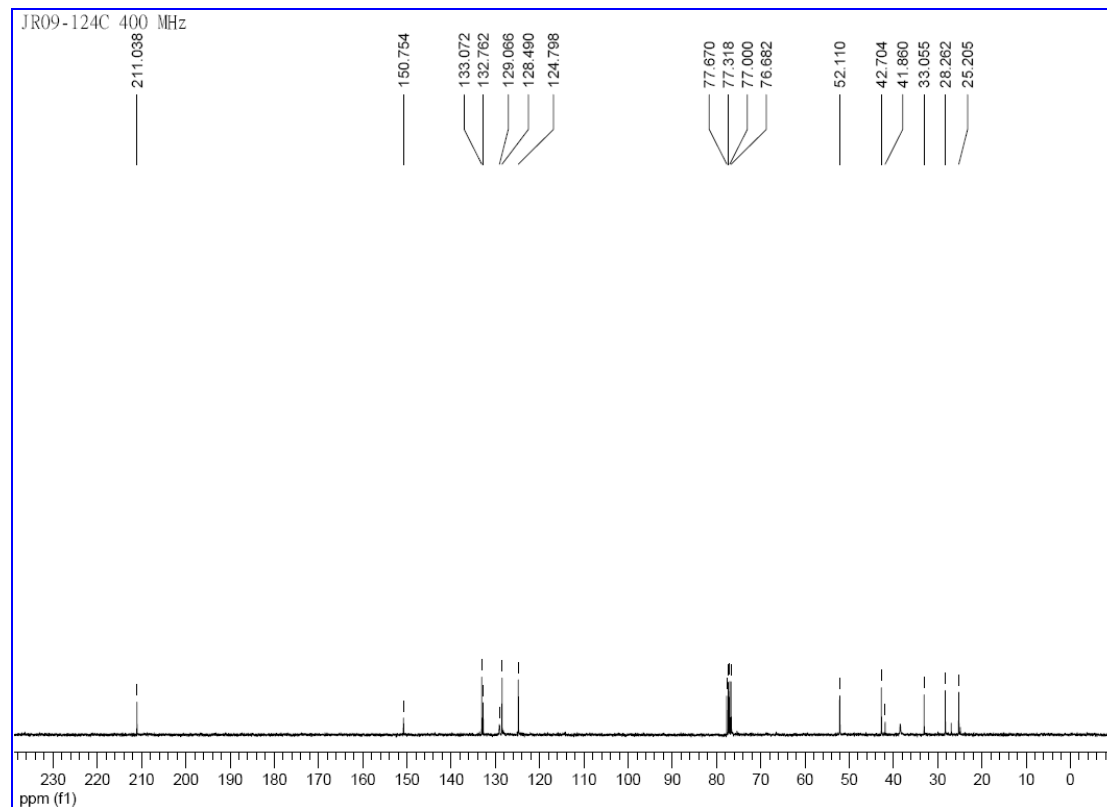
^{13}C NMR spectrum (100 MHz, CDCl_3) of Michael product 13b



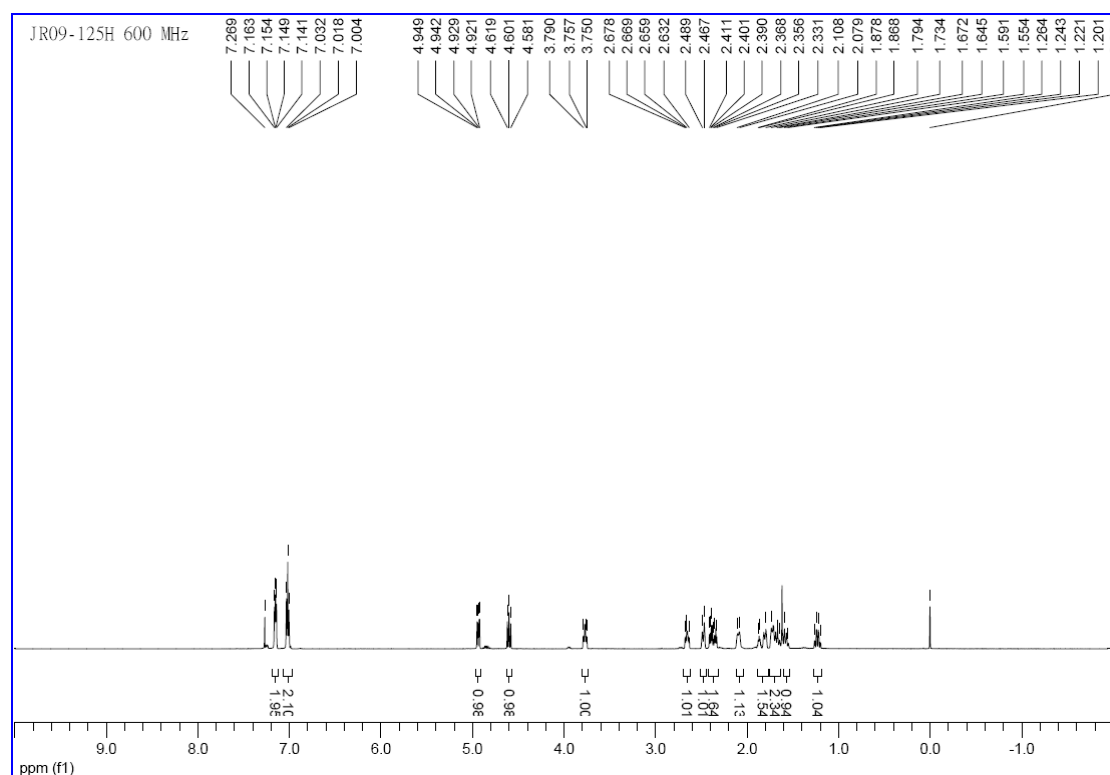
¹H NMR (600 MHz, CDCl₃) spectrum of Michael product 13c



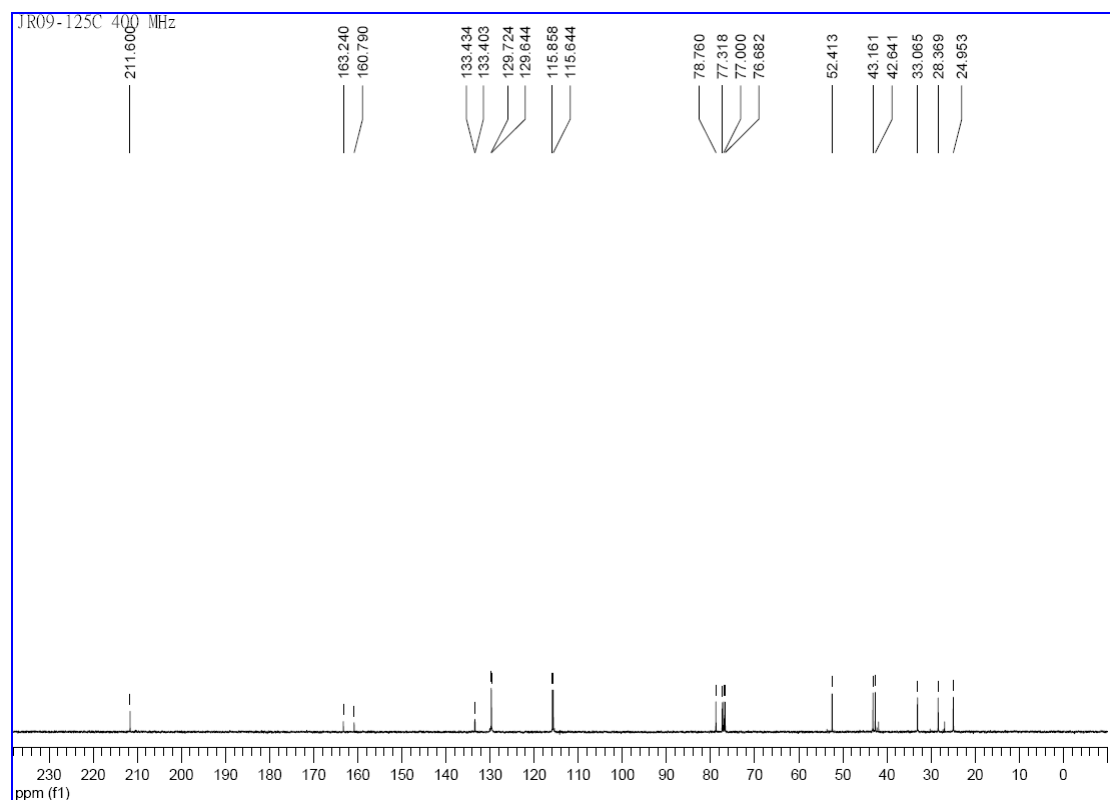
¹³C NMR spectrum (100 MHz, CDCl₃) of Michael product 13c



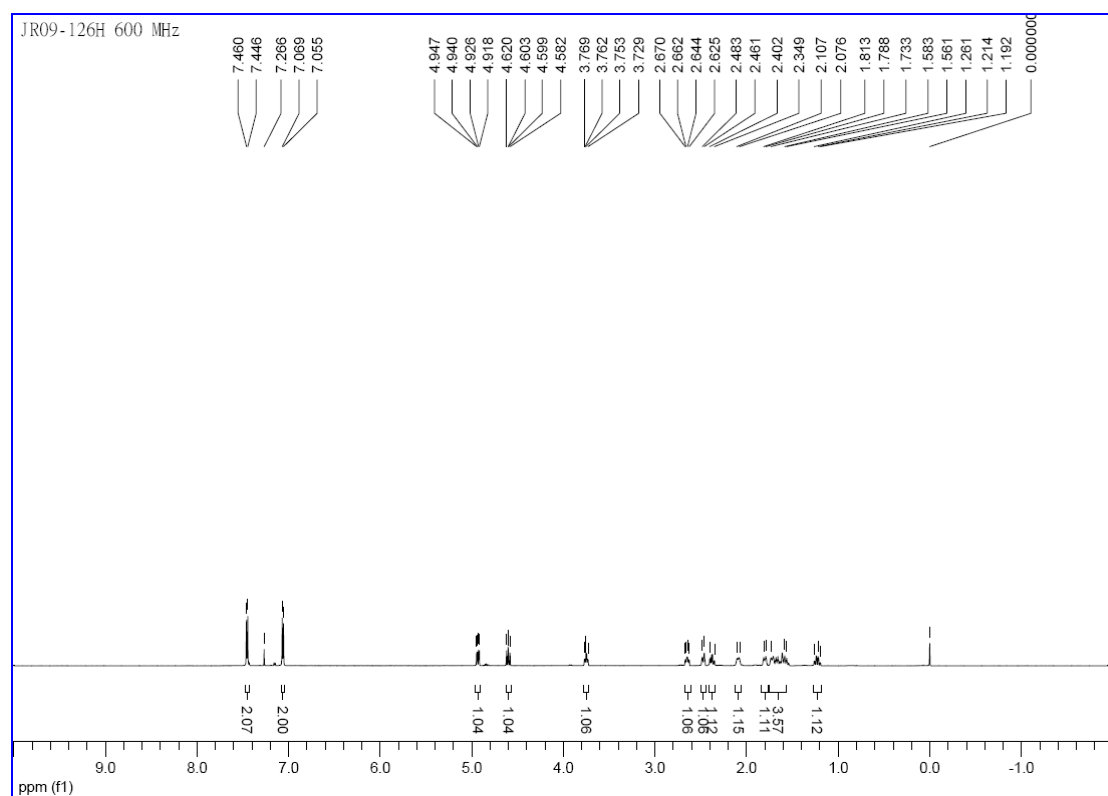
¹H NMR (600 MHz, CDCl₃) spectrum of Michael product 13d



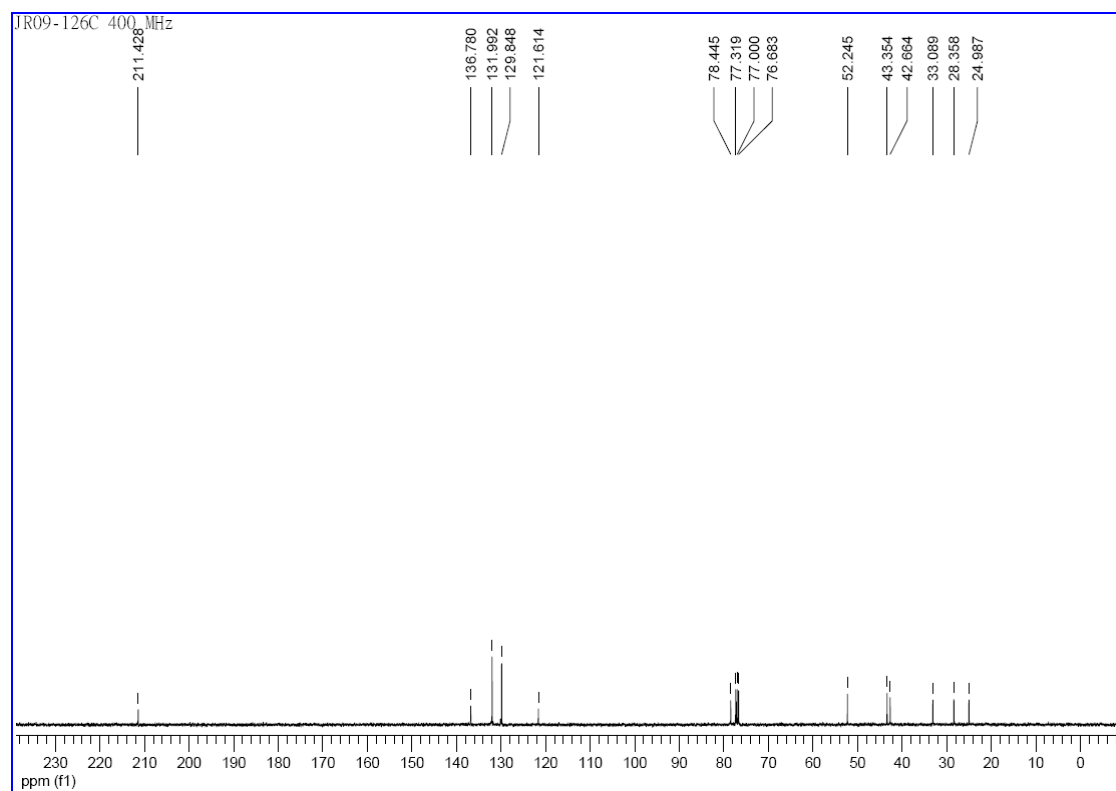
¹³C NMR spectrum (100 MHz, CDCl₃) of Michael product 13d



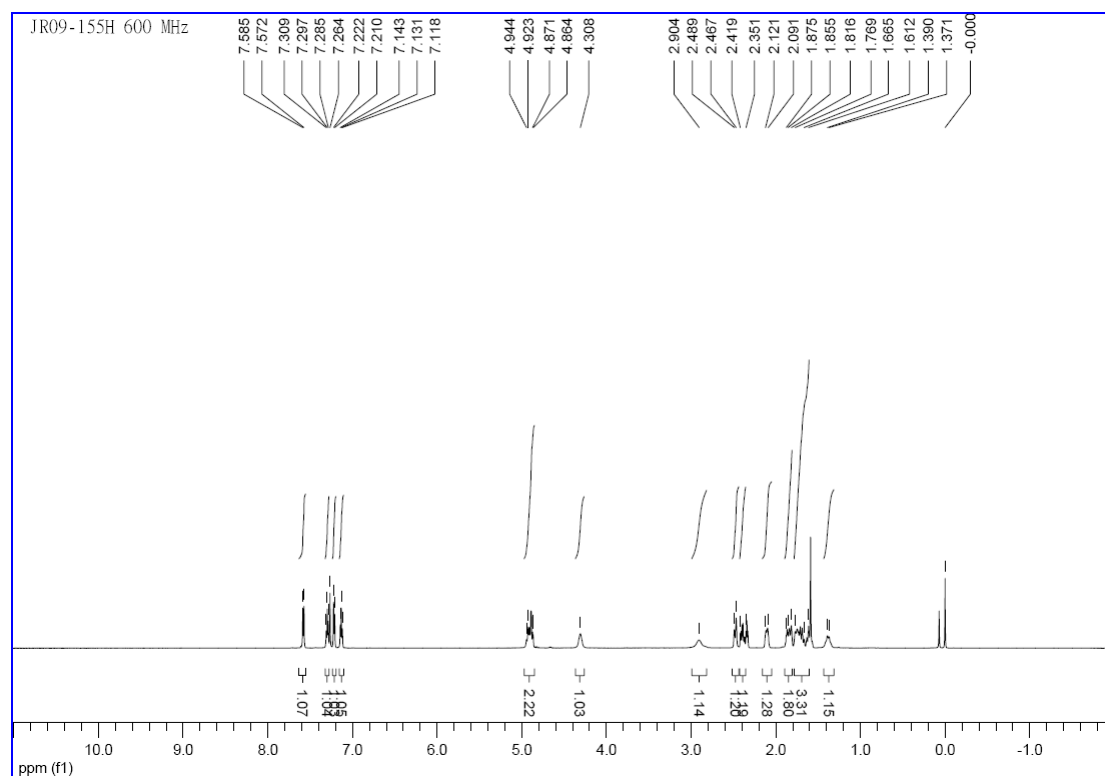
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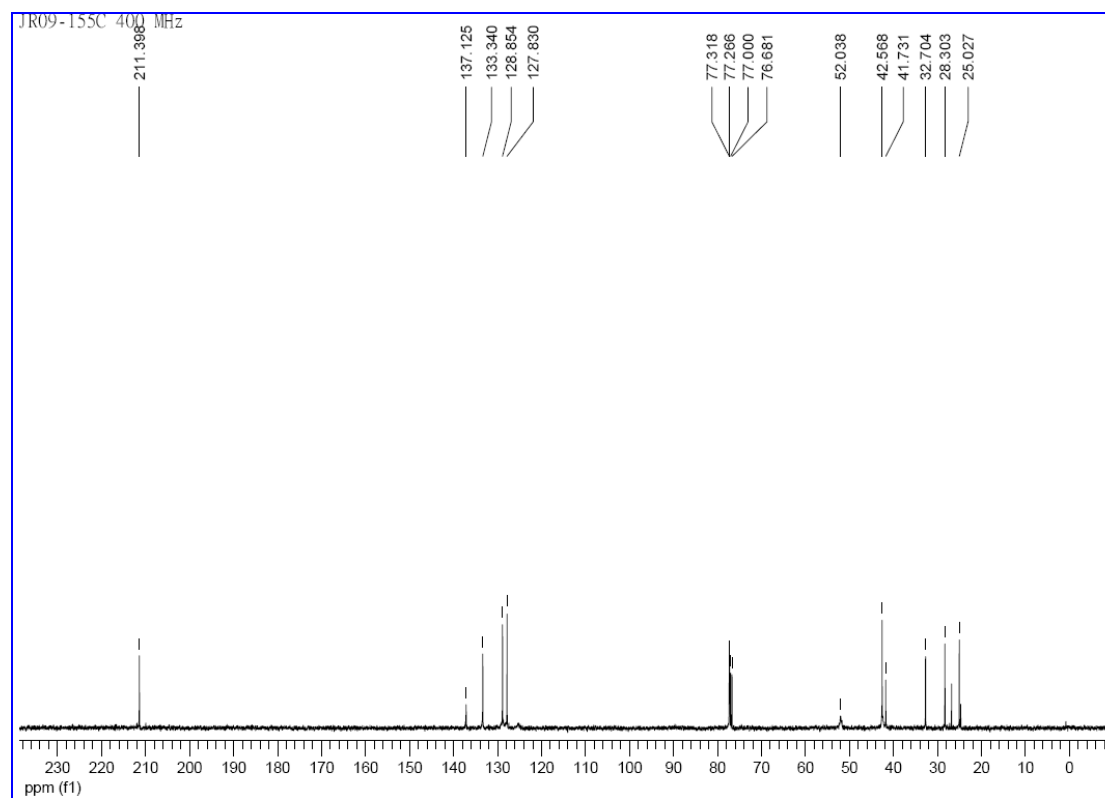
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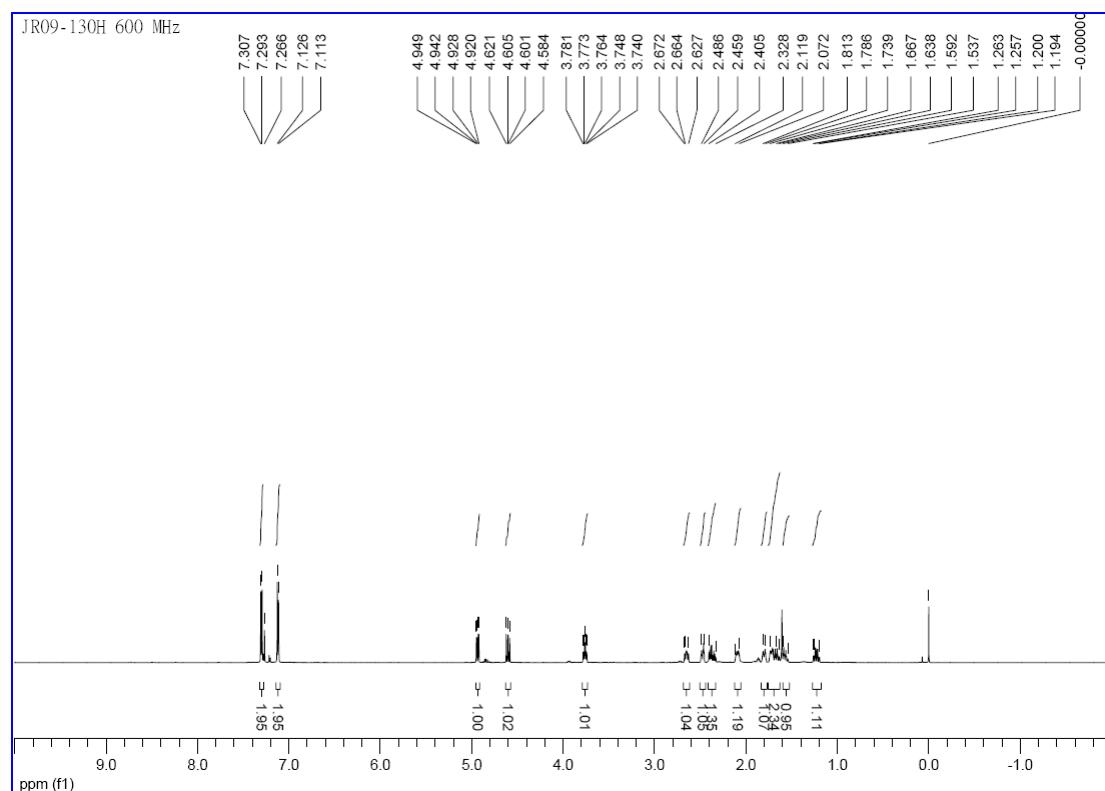
¹H NMR (600 MHz, CDCl₃) spectrum of Michael product 13f



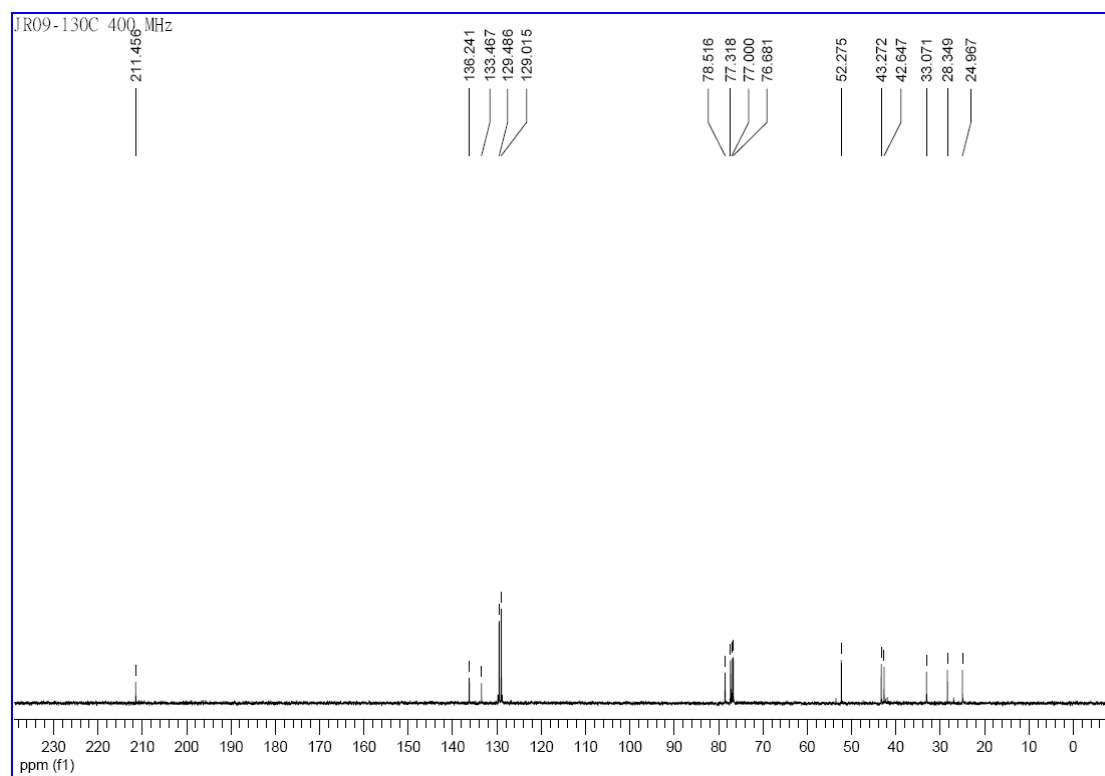
¹³C NMR spectrum (100 MHz, CDCl₃) of Michael product 13f



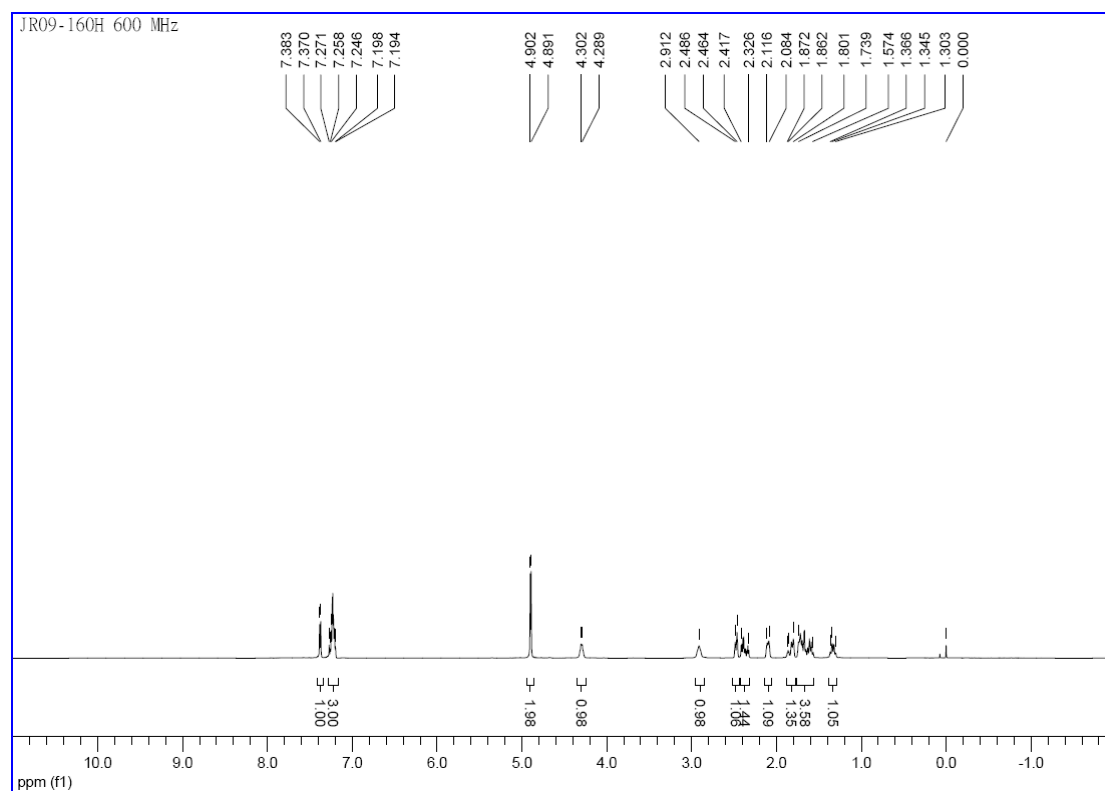
^1H NMR (600 MHz, CDCl_3) spectrum of Michael product 13g



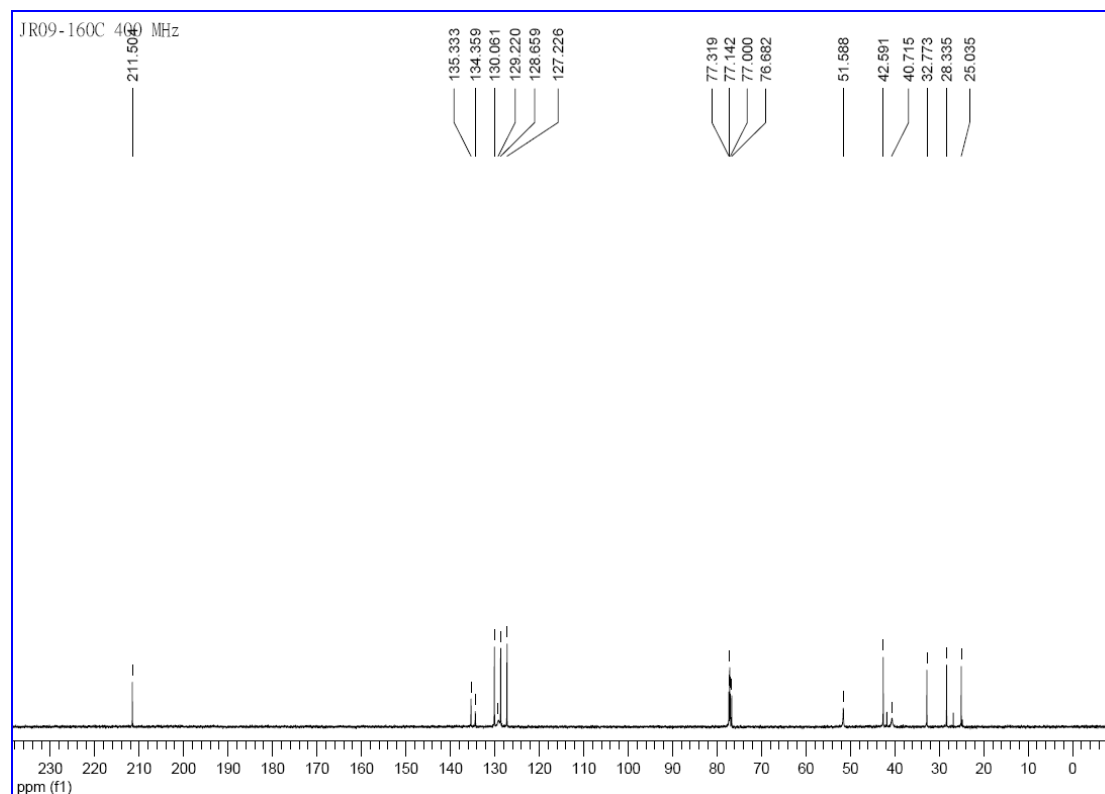
^{13}C NMR spectrum (100 MHz, CDCl_3) of Michael product 13g



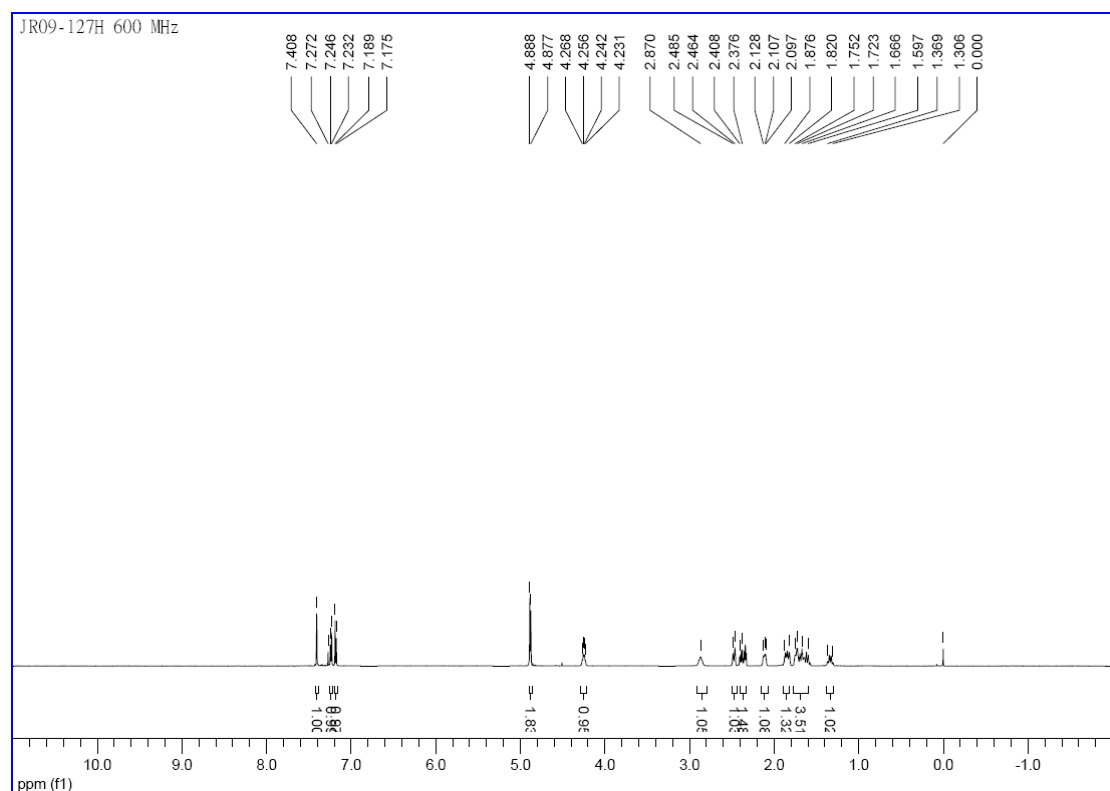
¹H NMR (600 MHz, CDCl₃) spectrum of Michael product 13h



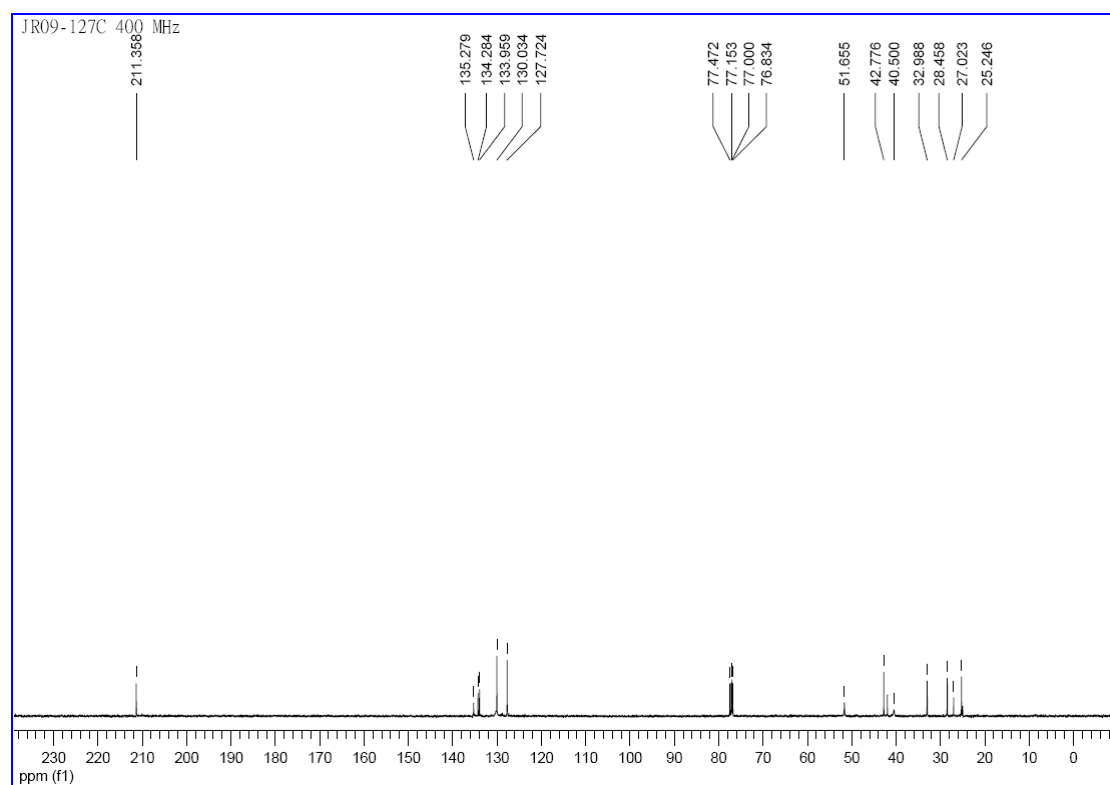
¹³C NMR spectrum (100 MHz, CDCl₃) of Michael product 13h



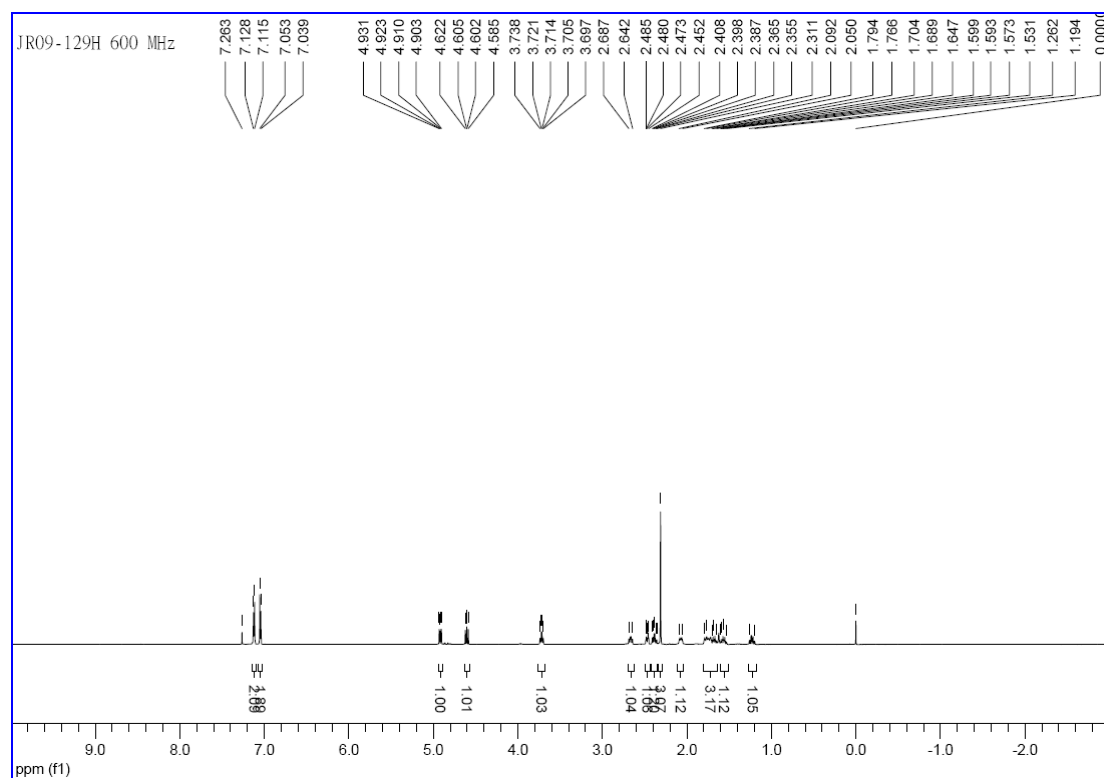
¹H NMR (600 MHz, CDCl₃) spectrum of Michael product 13i



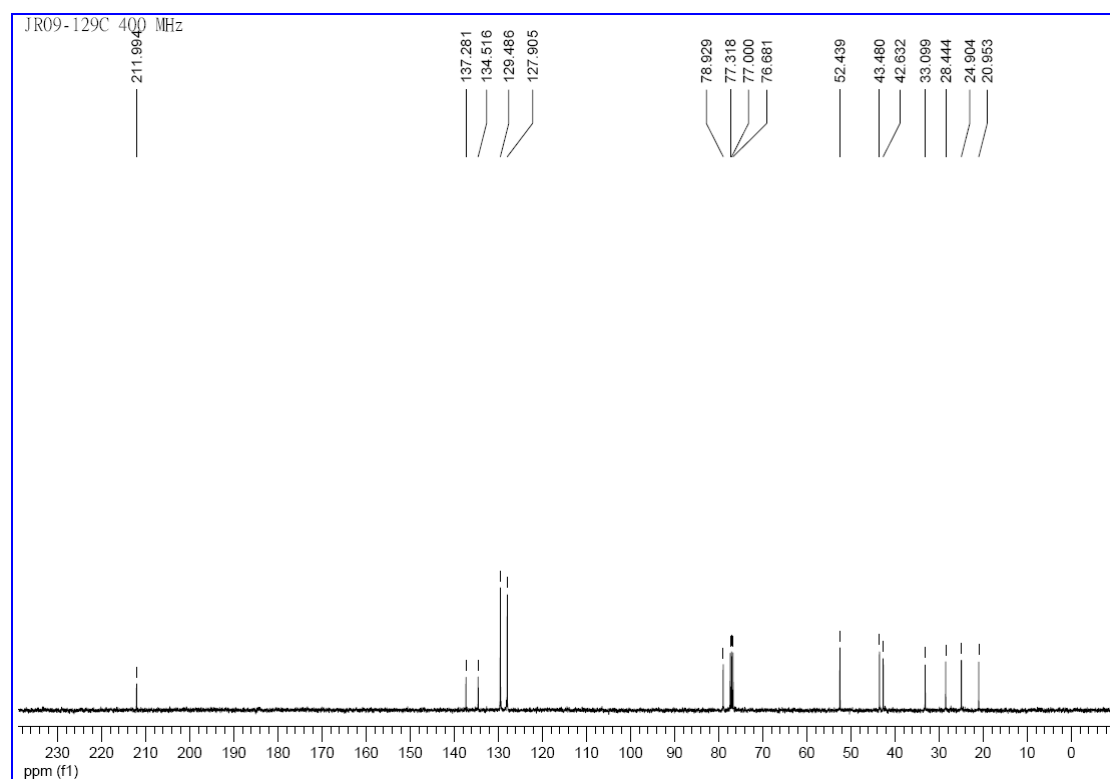
¹³C NMR spectrum (100 MHz, CDCl₃) of Michael product 13i



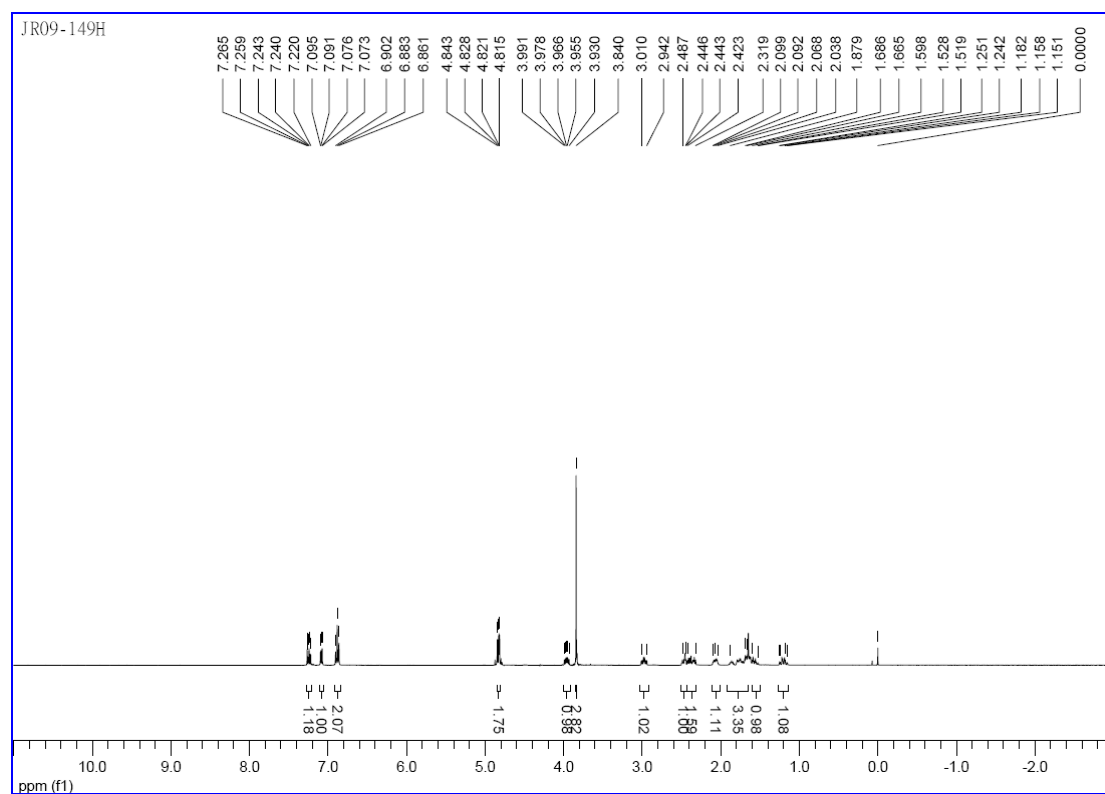
^1H NMR (600 MHz, CDCl_3) spectrum of Michael product 13j



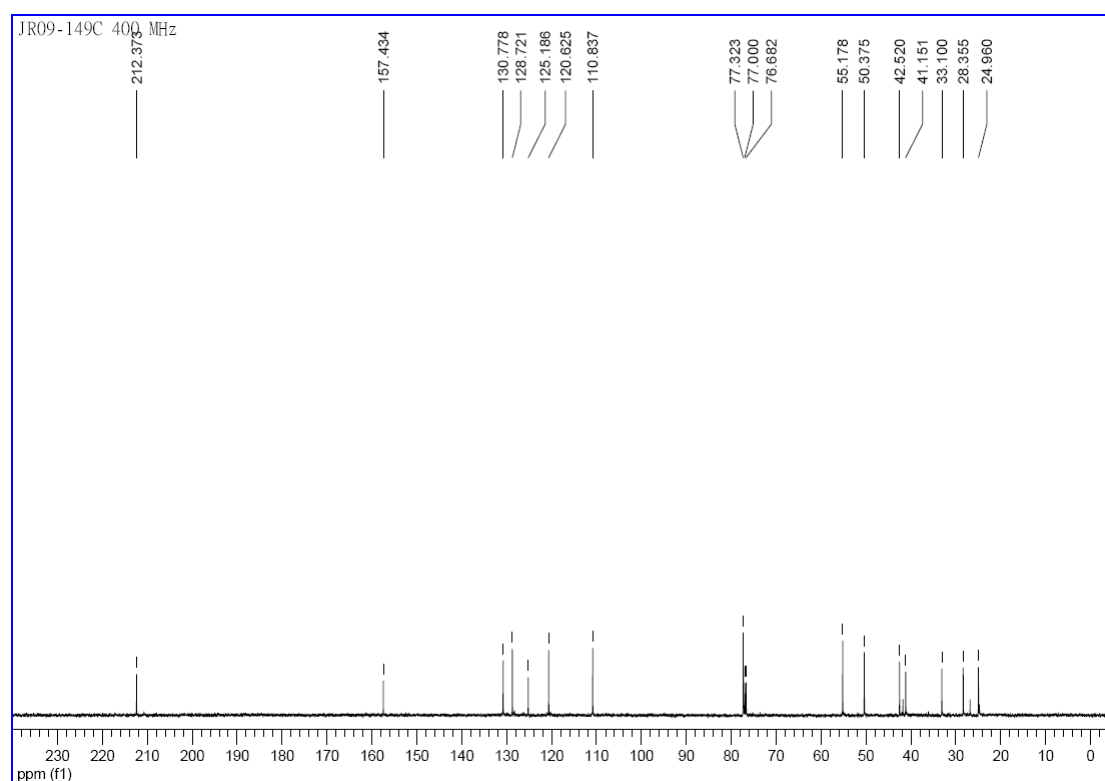
^{13}C NMR spectrum (100 MHz, CDCl_3) of Michael product 13j



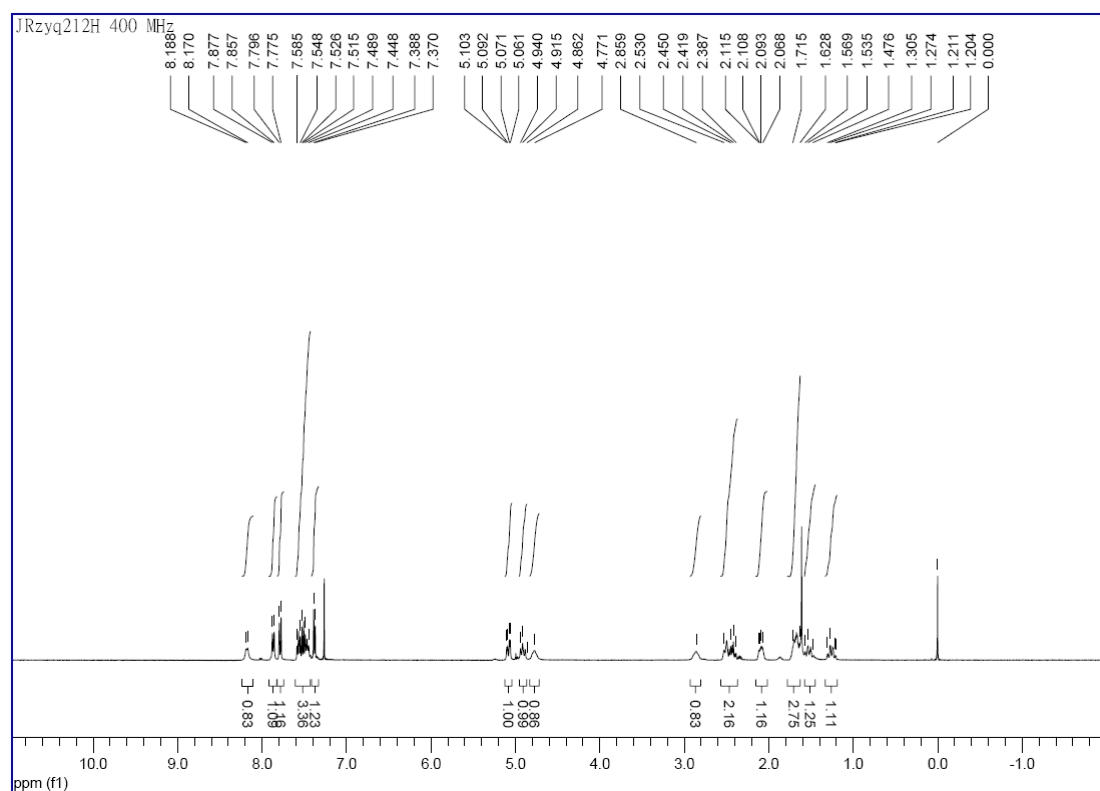
¹H NMR (400 MHz, CDCl₃) spectrum of Michael product 13k



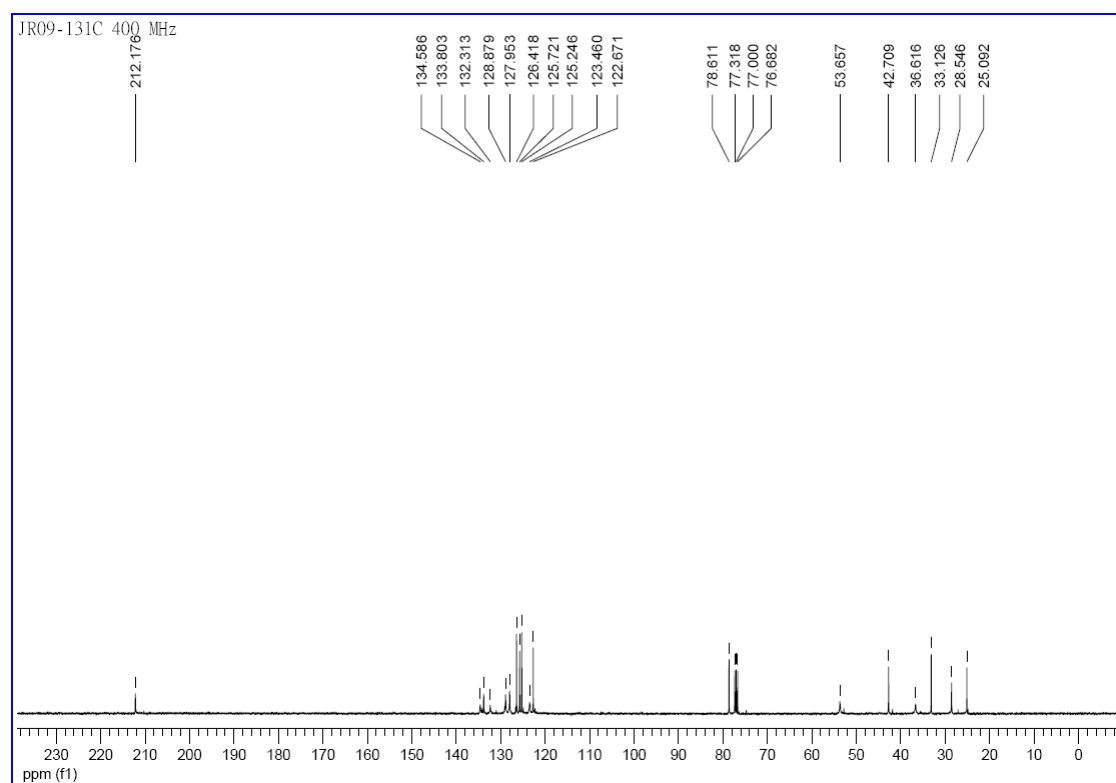
¹³C NMR spectrum (100 MHz, CDCl₃) of Michael product 13k



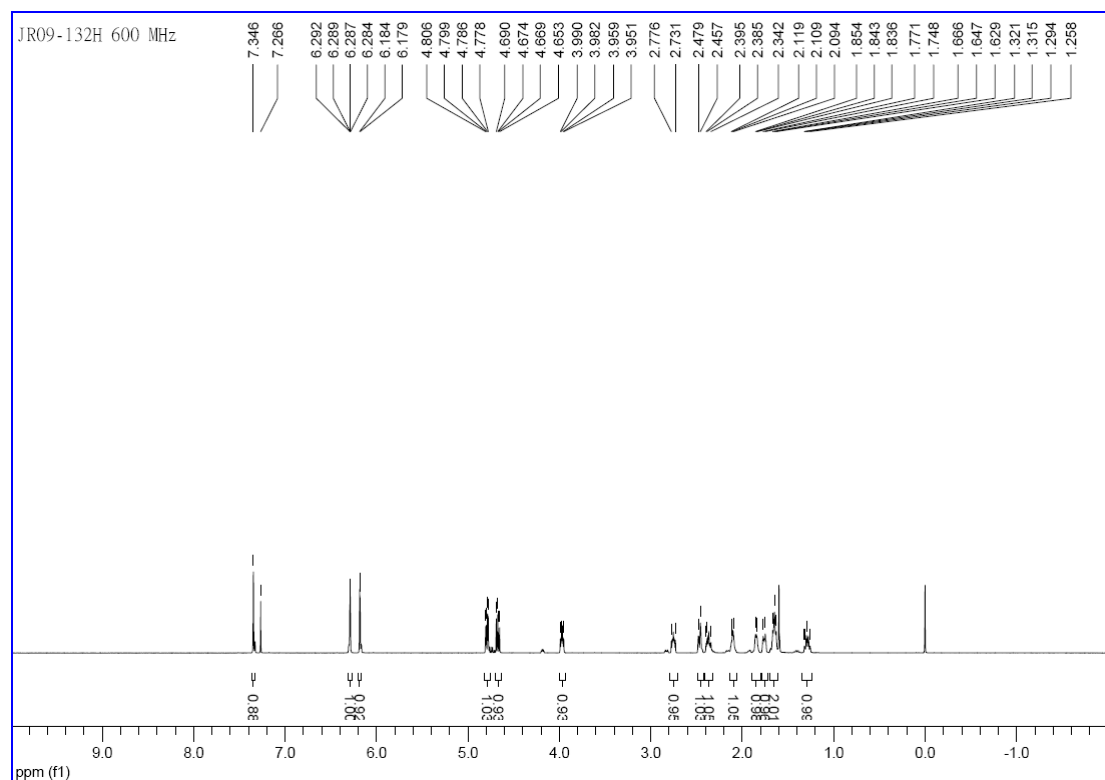
^1H NMR (400 MHz, CDCl_3) spectrum of Michael product 13l



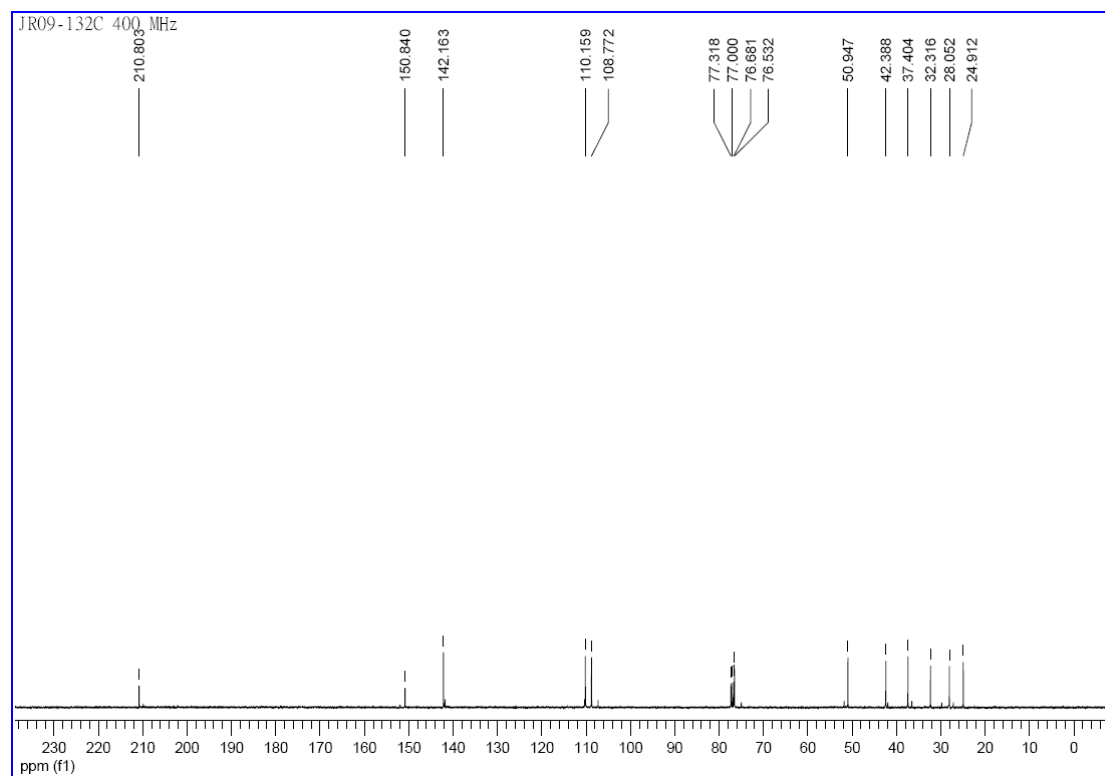
^{13}C NMR spectrum (100 MHz, CDCl_3) of Michael product 13l



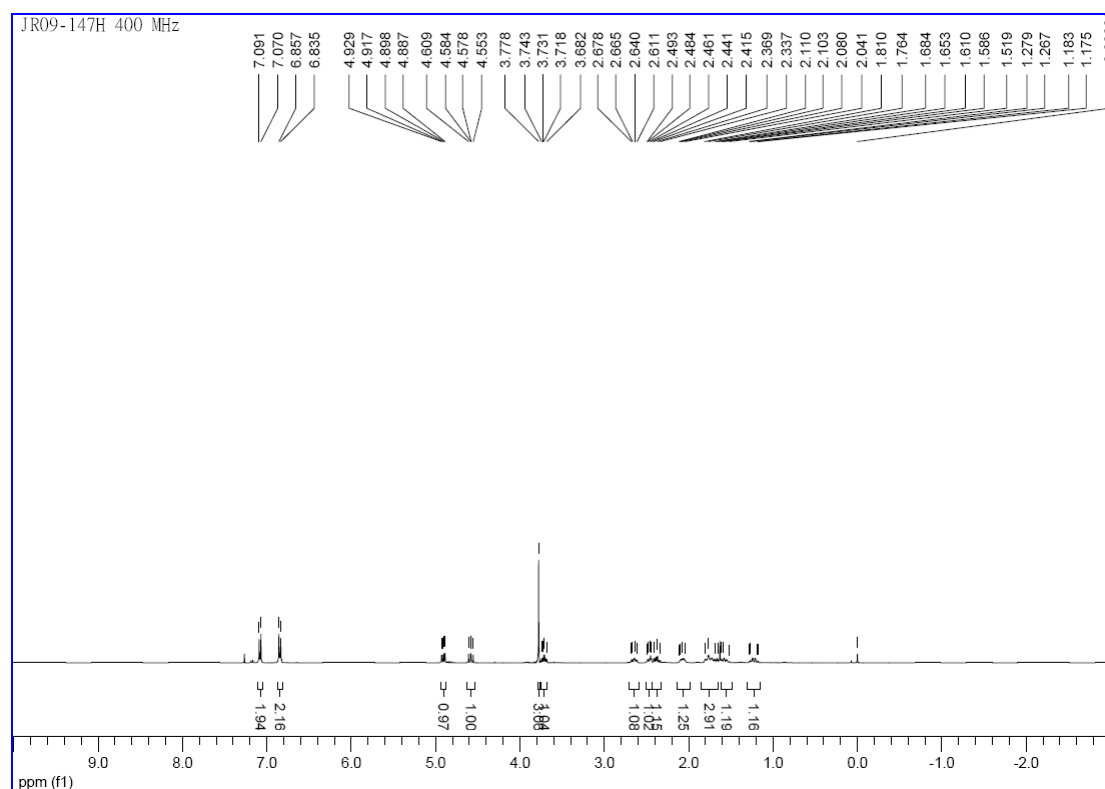
^1H NMR (600 MHz, CDCl_3) spectrum of Michael product 13m



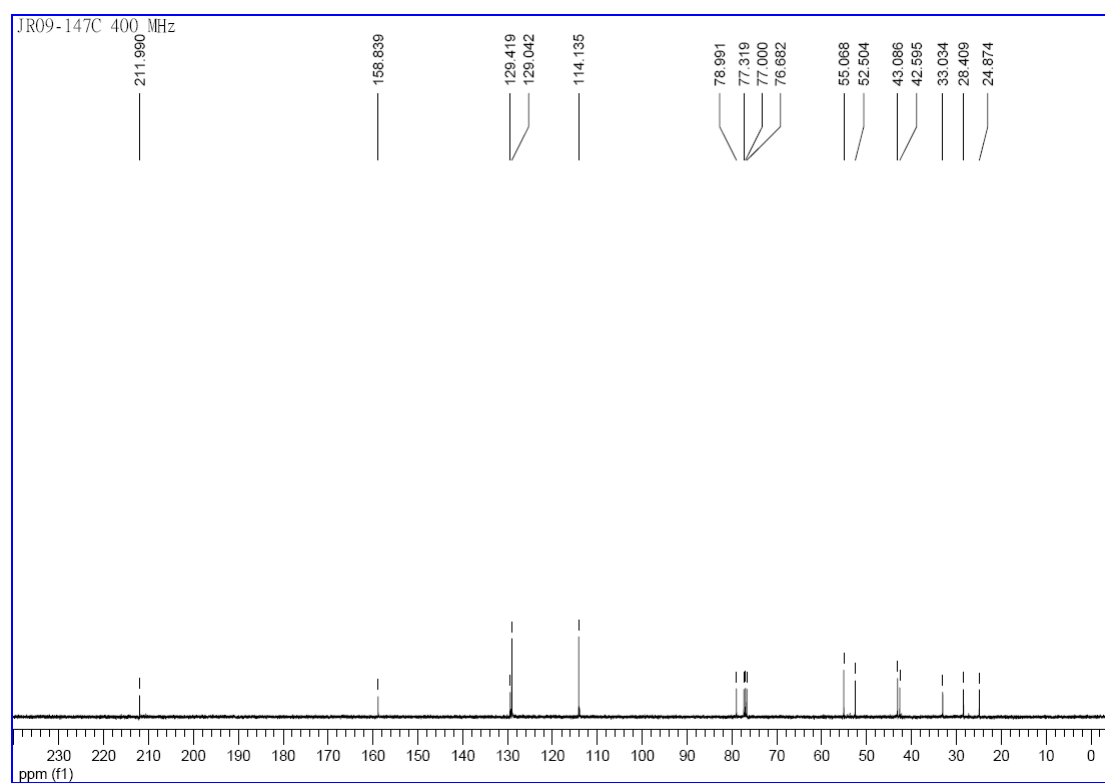
^{13}C NMR spectrum (100 MHz, CDCl_3) of Michael product 13m



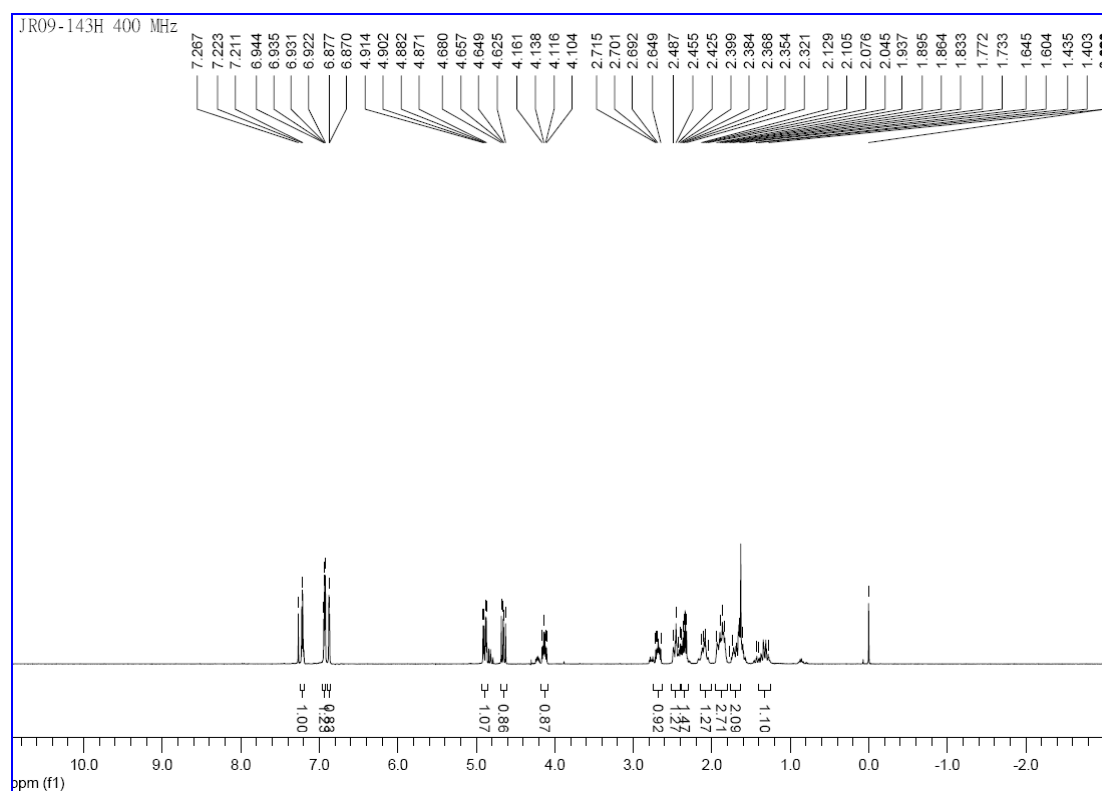
^1H NMR (400 MHz, CDCl_3) spectrum of Michael product 13n



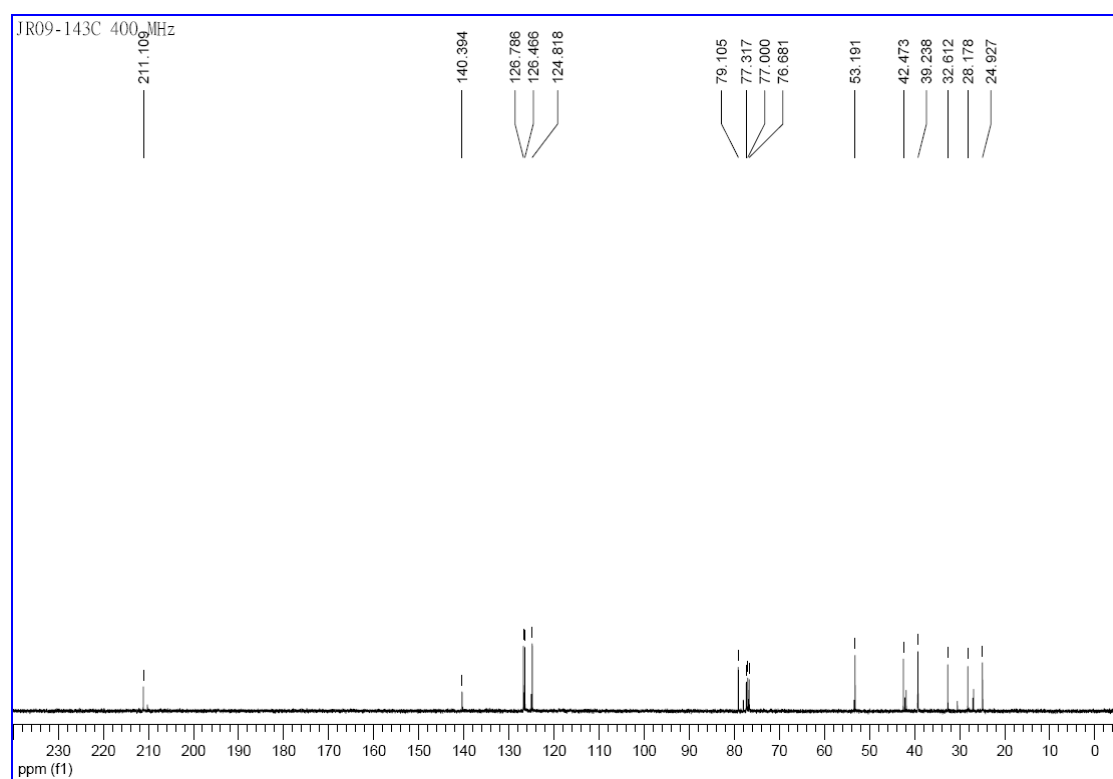
^{13}C NMR spectrum (100 MHz, CDCl_3) of Michael product 13n



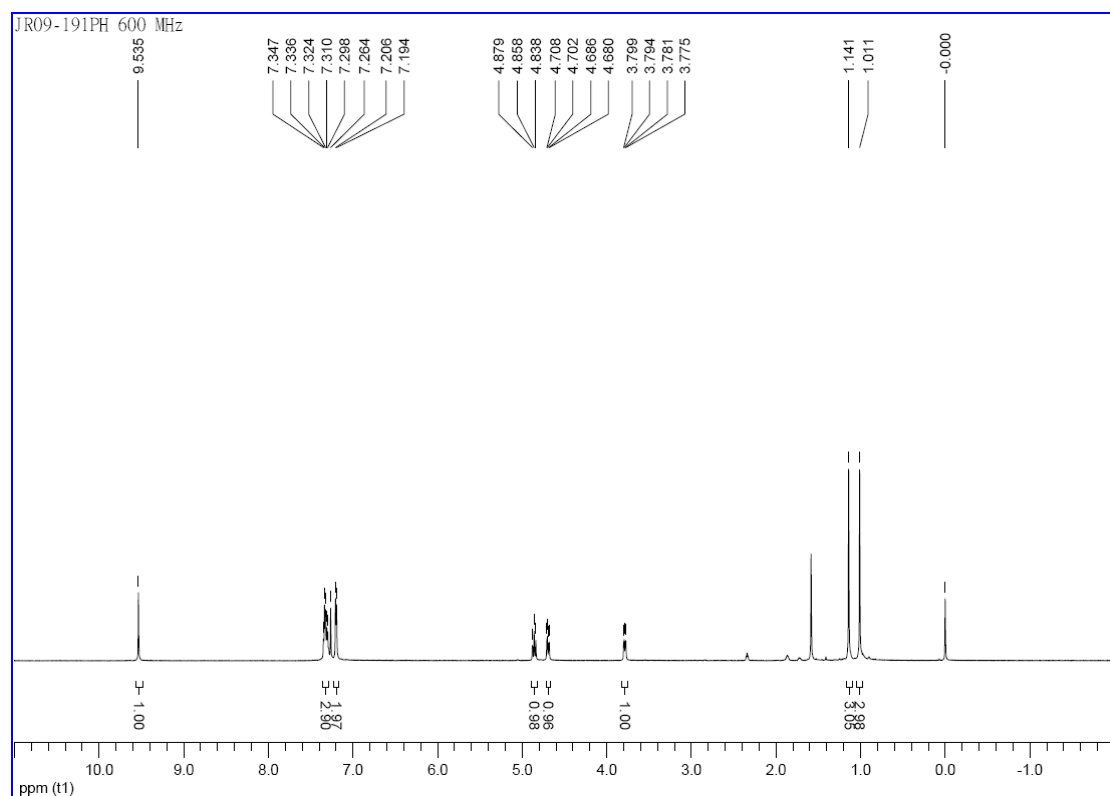
^1H NMR (400 MHz, CDCl_3) spectrum of Michael product 13o



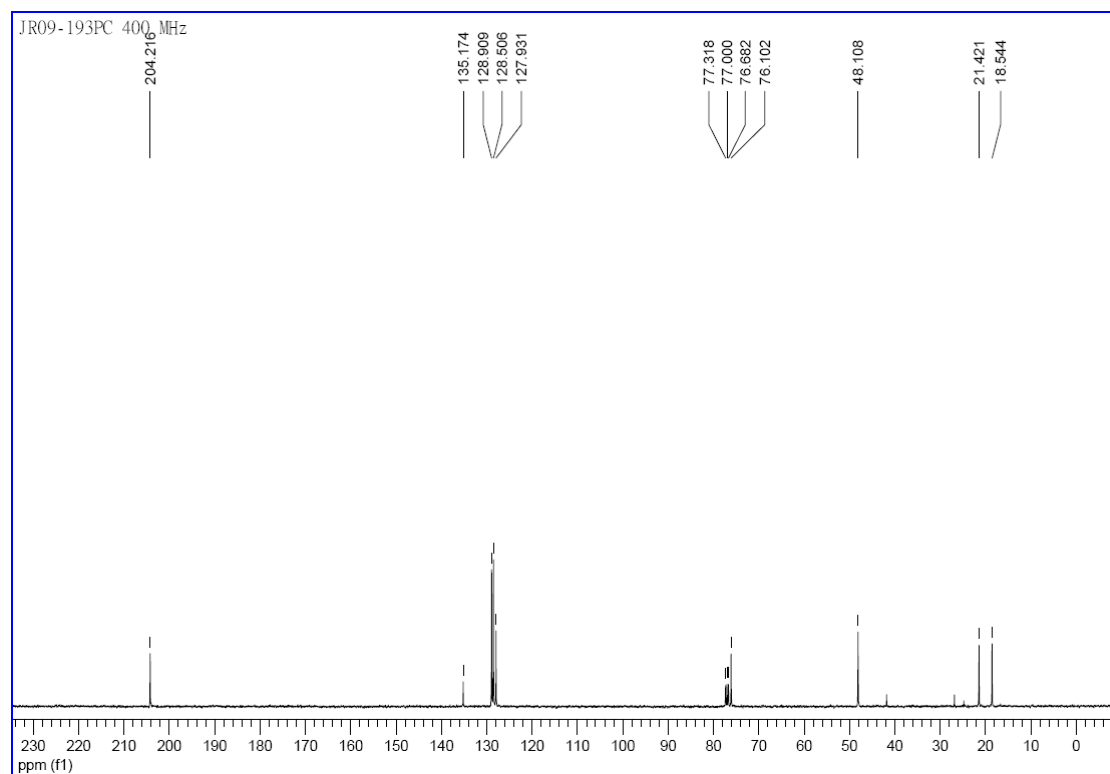
^{13}C NMR spectrum (100 MHz, CDCl_3) of Michael product 13o



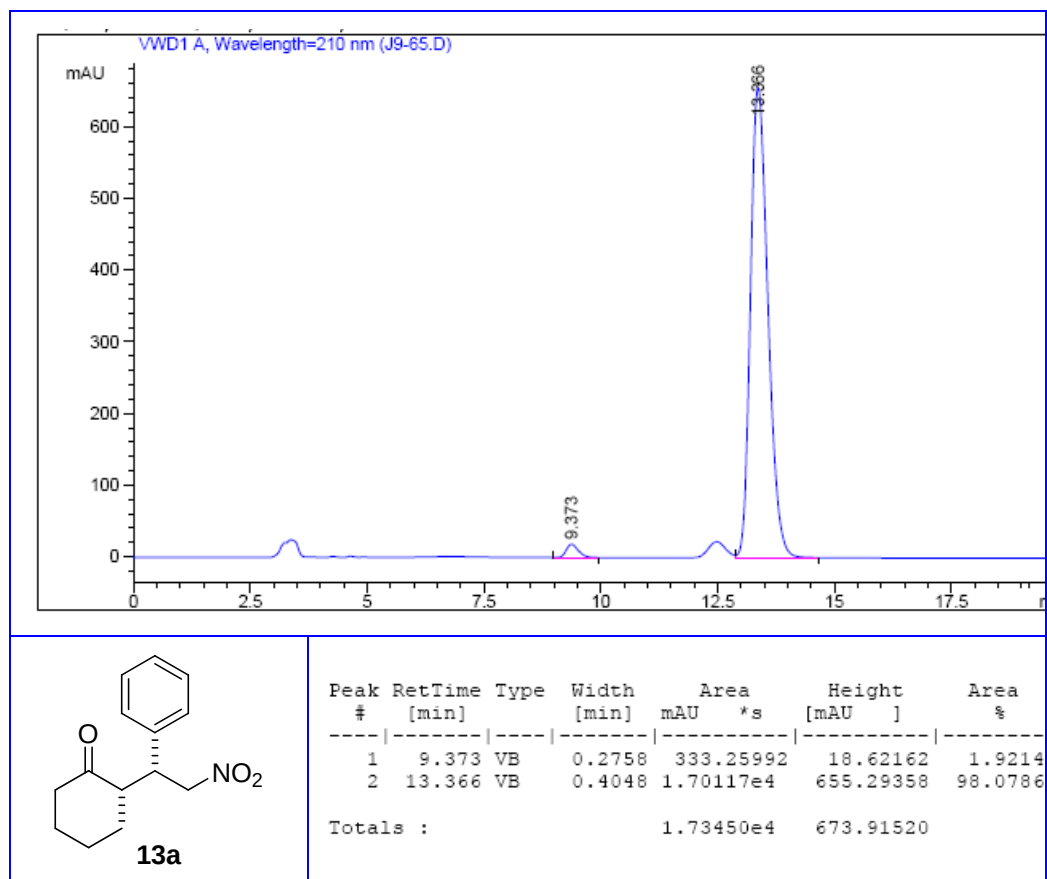
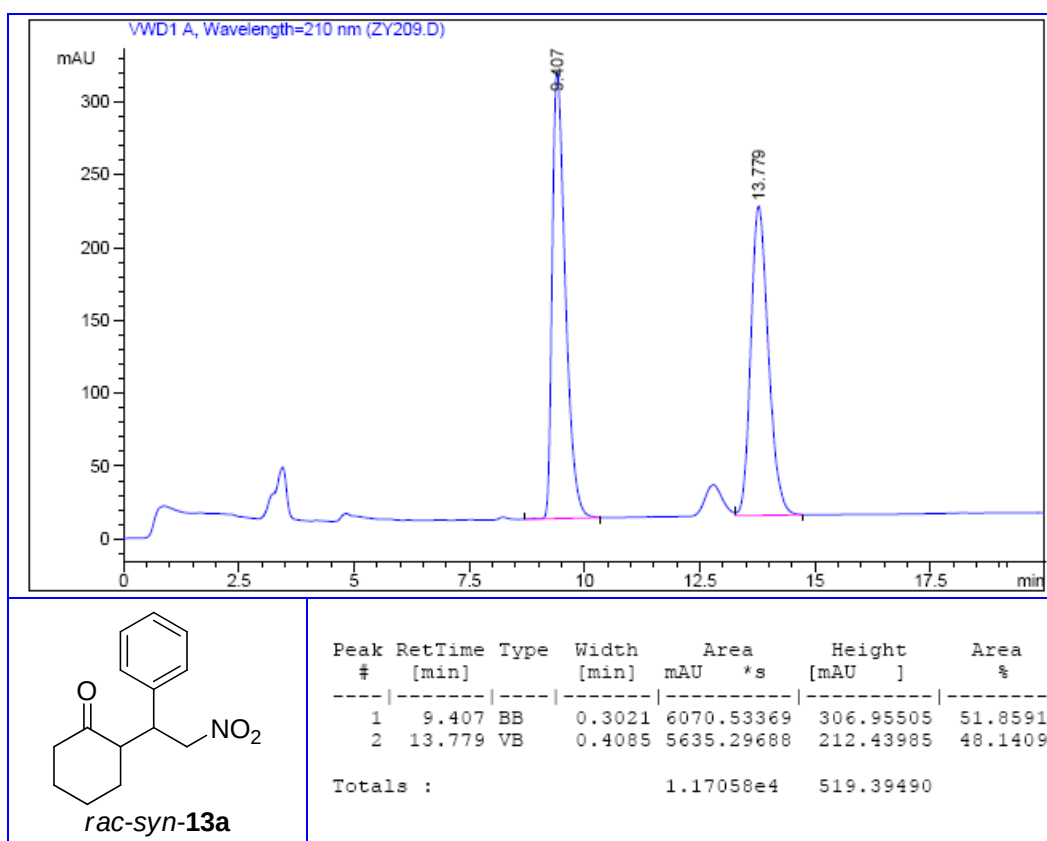
¹H NMR (400 MHz, CDCl₃) spectrum of Michael product 15

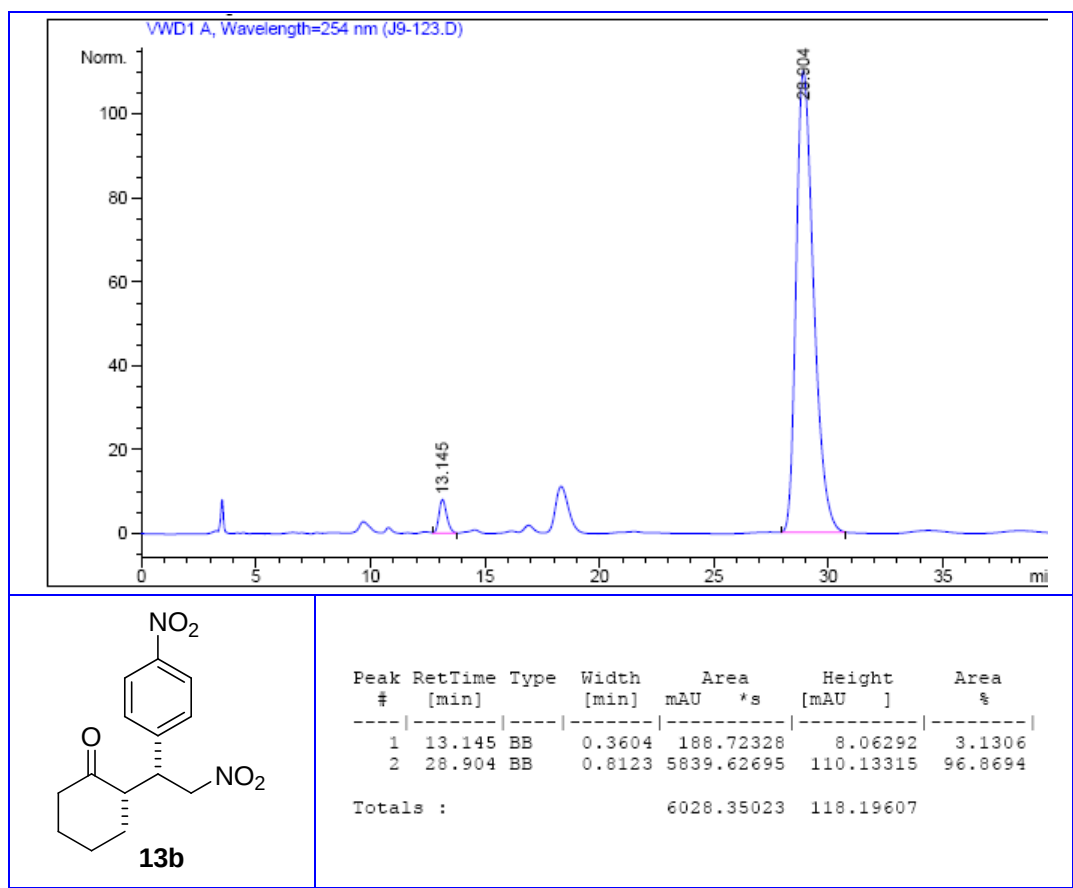
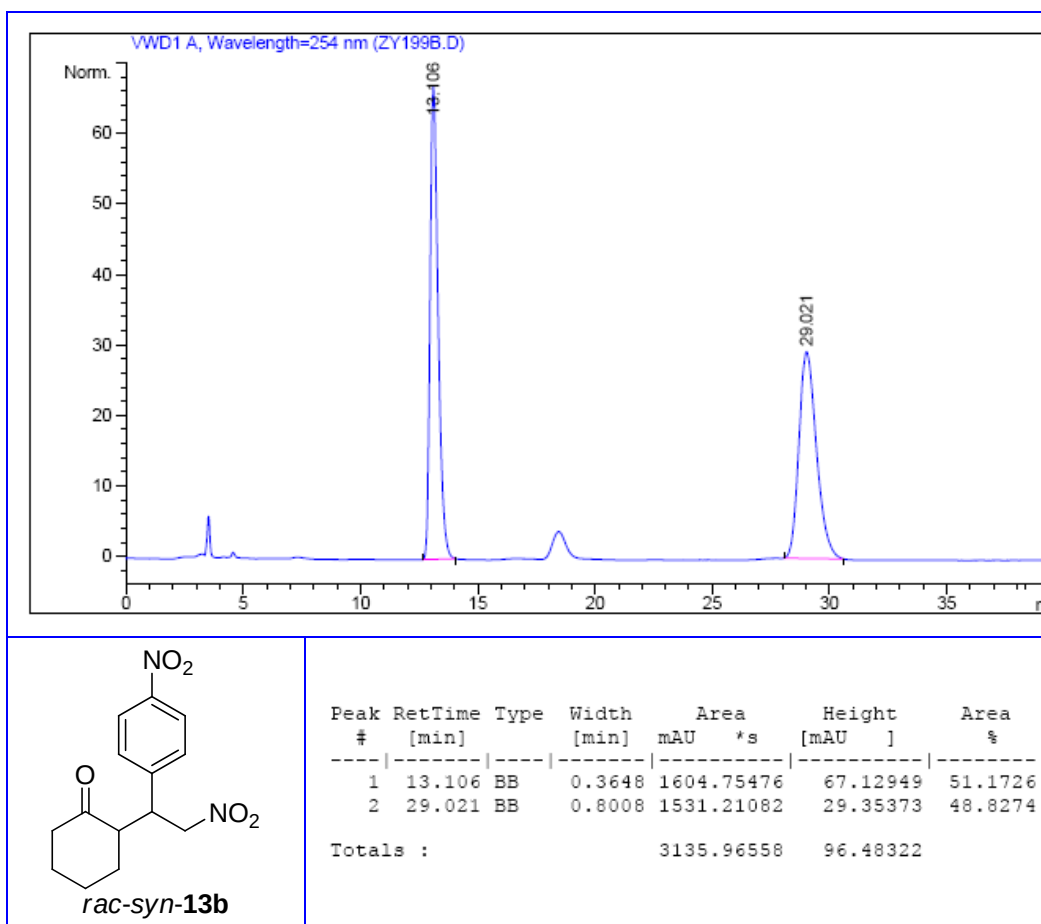


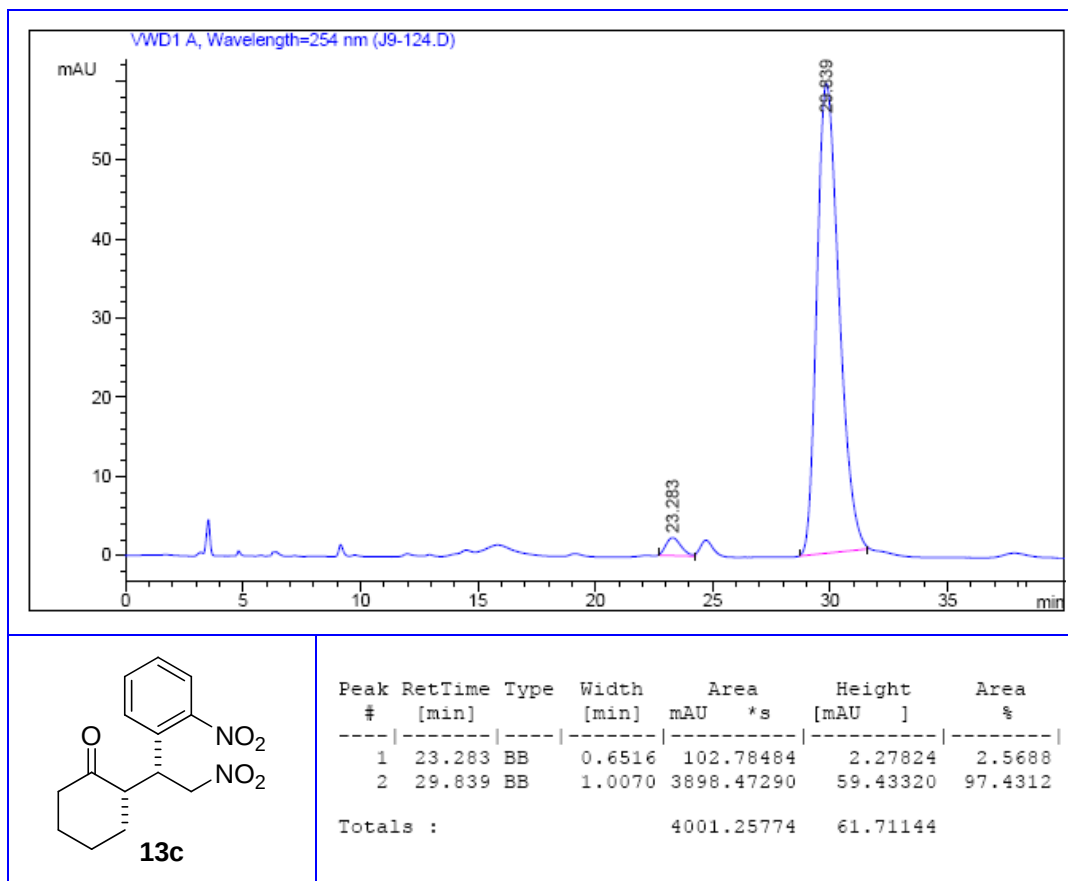
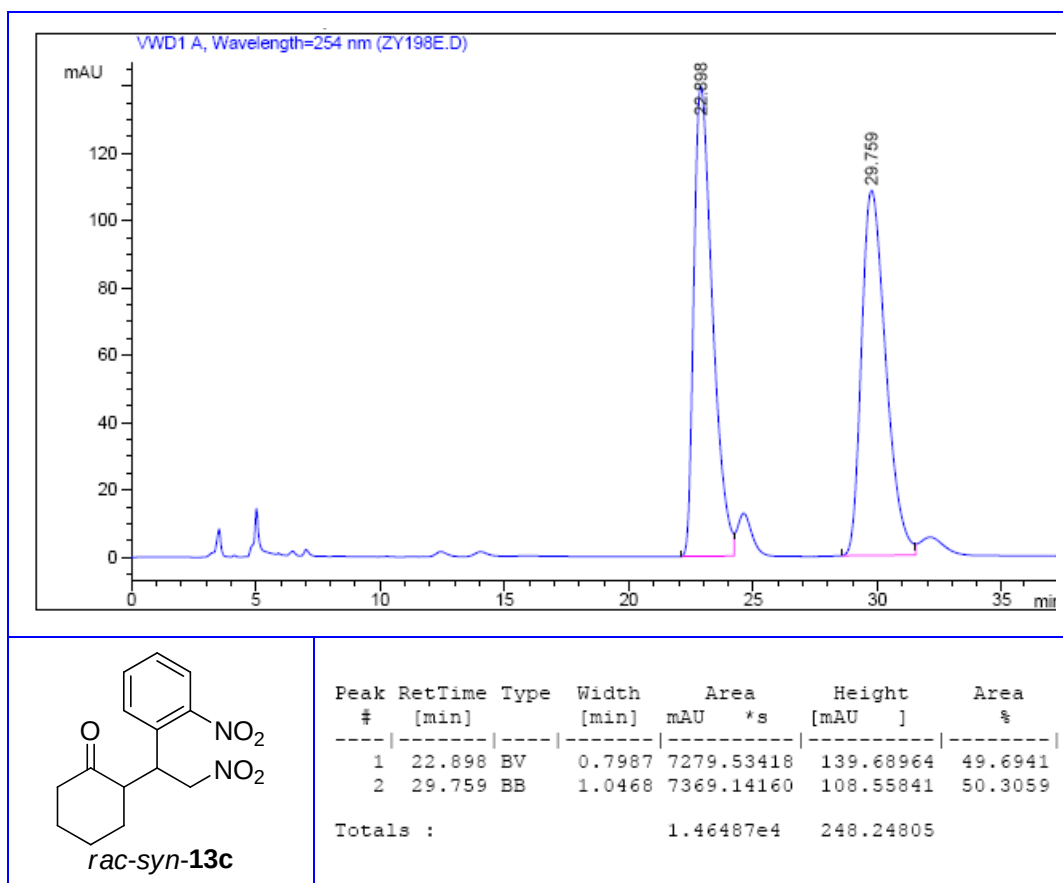
¹³C NMR spectrum (100 MHz, CDCl₃) of Michael product 15

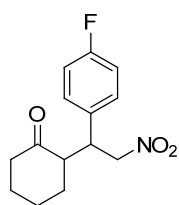
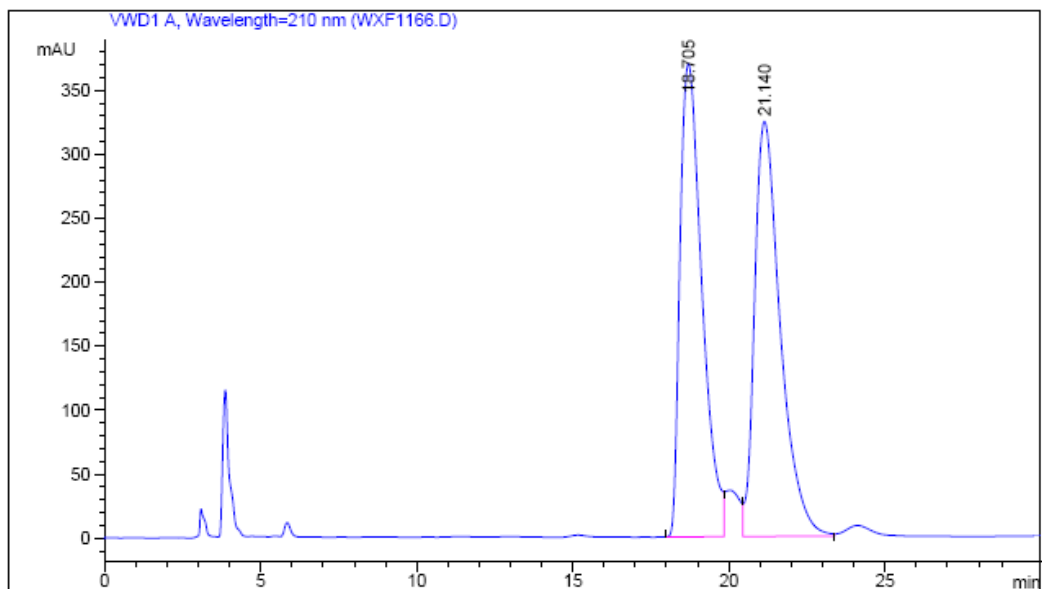


6.0 HPLC Chromatograms

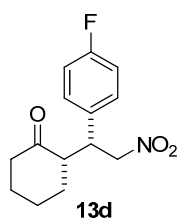
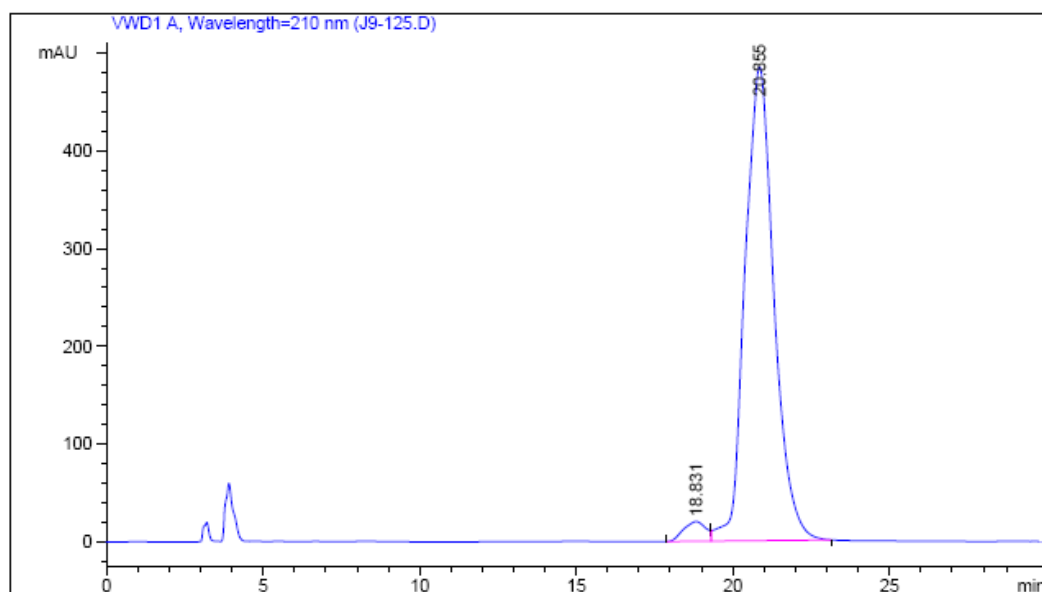




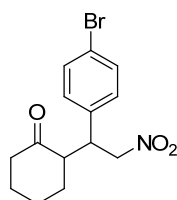
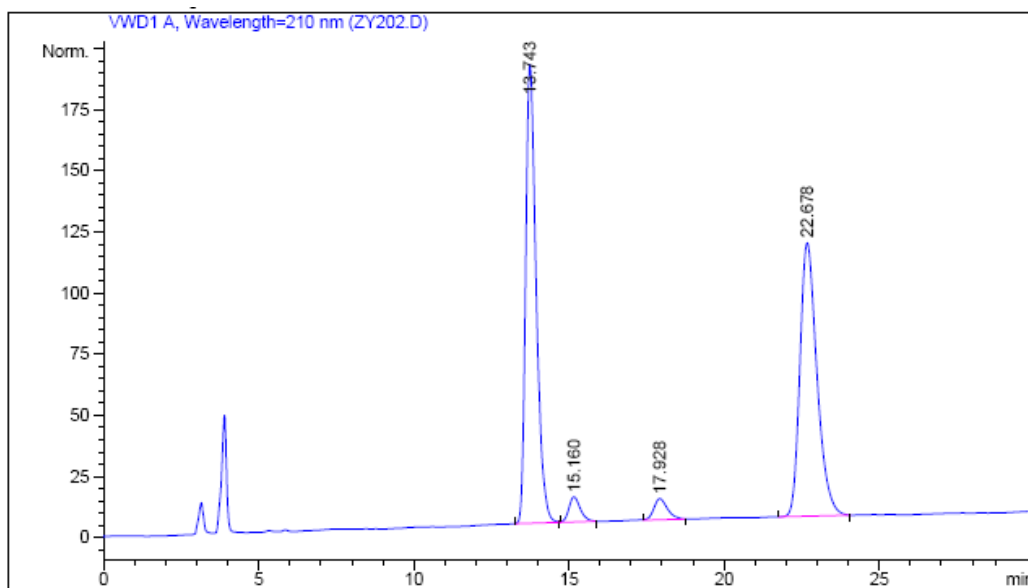




Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	18.705	BV	0.7531	1.80739e4	370.13577	49.1837
2	21.140	VB	0.8739	1.86739e4	324.43109	50.8163
Totals :				3.67478e4	694.56686	

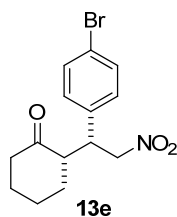
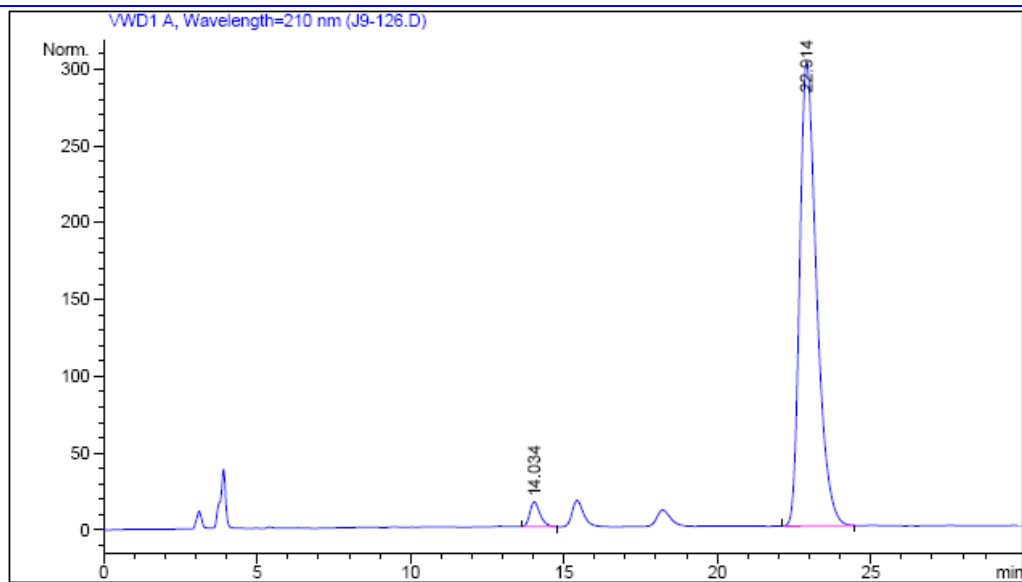


Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	18.831	BV	0.7069	997.02838	20.19934	3.1038
2	20.855	VB	0.9159	3.11256e4	484.21994	96.8962
Totals :				3.21226e4	504.41928	



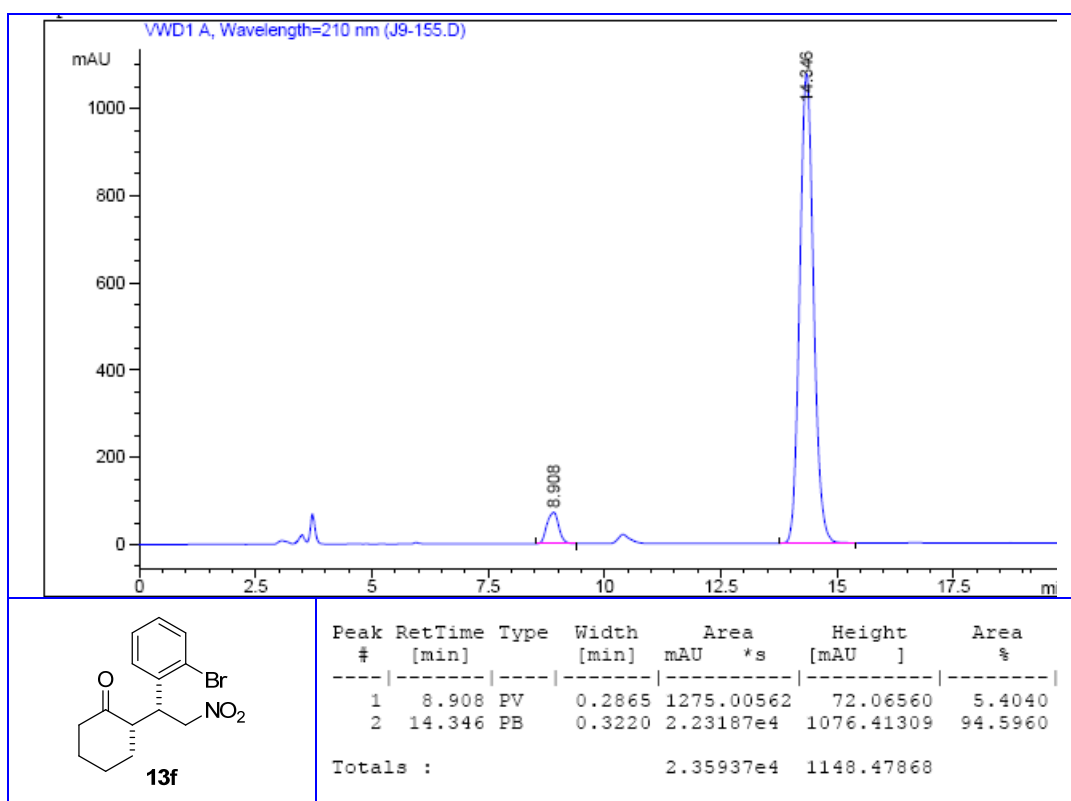
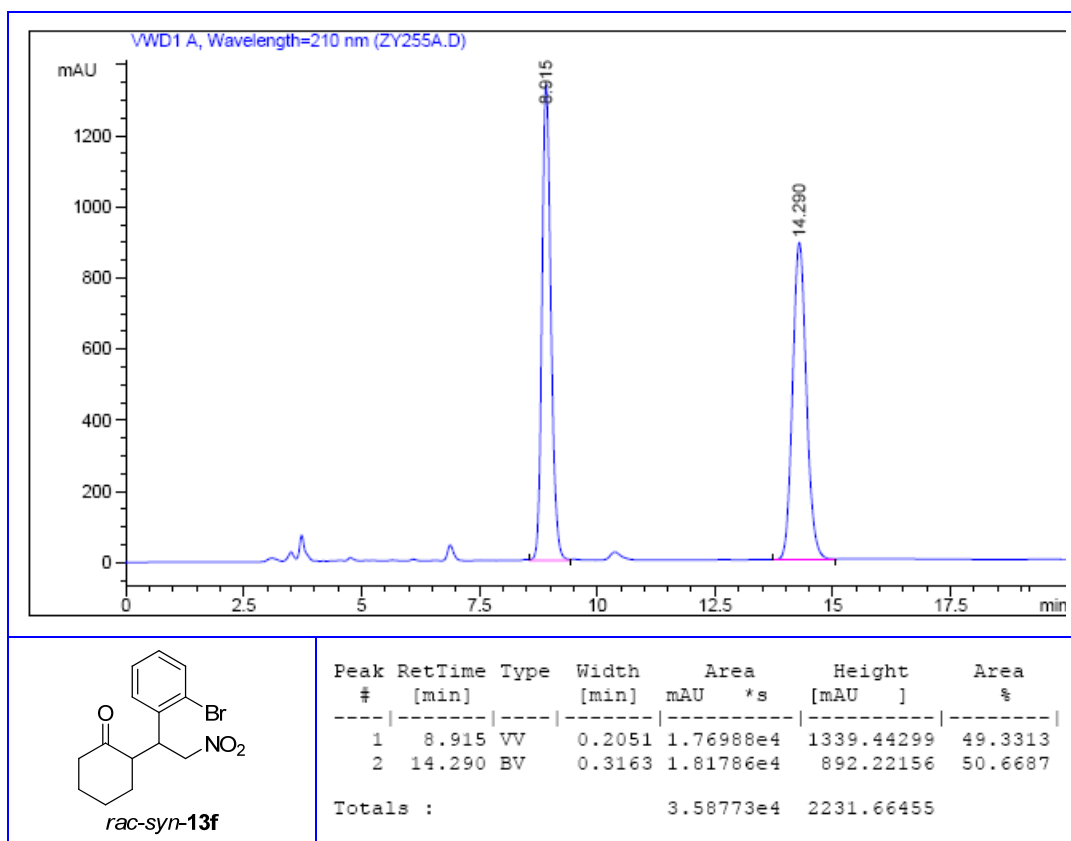
Peak #	RetTime [min]	Type	Width [min]	Area mAU	Area *s	Height [mAU]	Area %
1	13.743	BB	0.3566	4399.88379		187.63319	47.8409
2	15.160	BB	0.3894	265.05792		10.28758	2.8820
3	17.928	BB	0.4558	264.36295		8.64444	2.8745
4	22.678	BB	0.5820	4267.60156		111.84785	46.4026

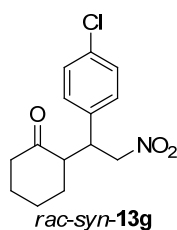
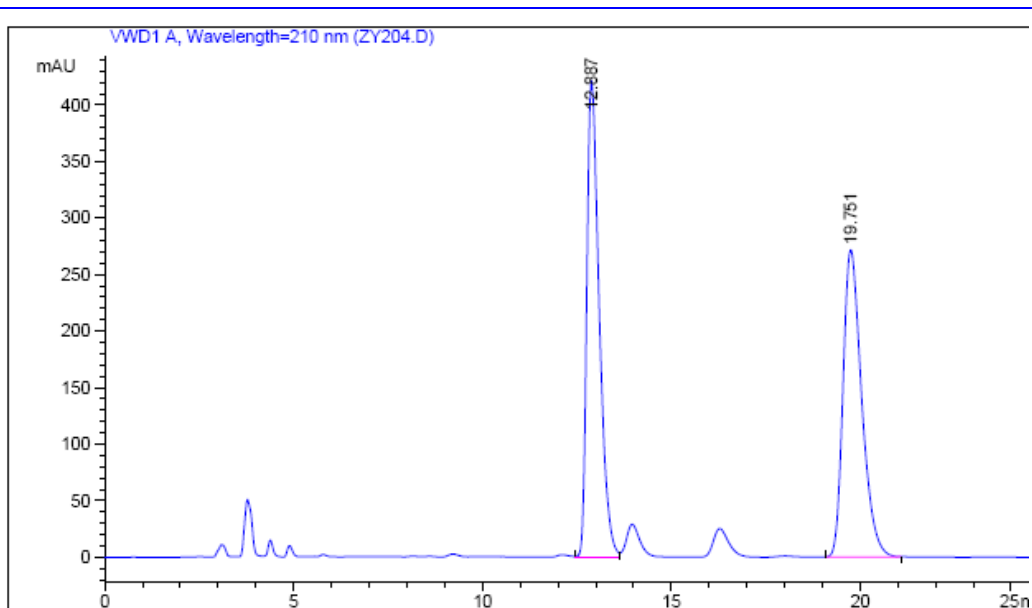
Totals : 9196.90622 318.41306



Peak #	RetTime [min]	Type	Width [min]	Area mAU	Area *s	Height [mAU]	Area %
1	14.034	PP	0.3552	371.19769		16.07867	3.1187
2	22.914	BB	0.5850	1.15313e4		301.17044	96.8813

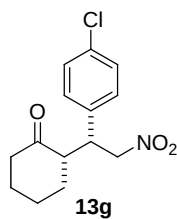
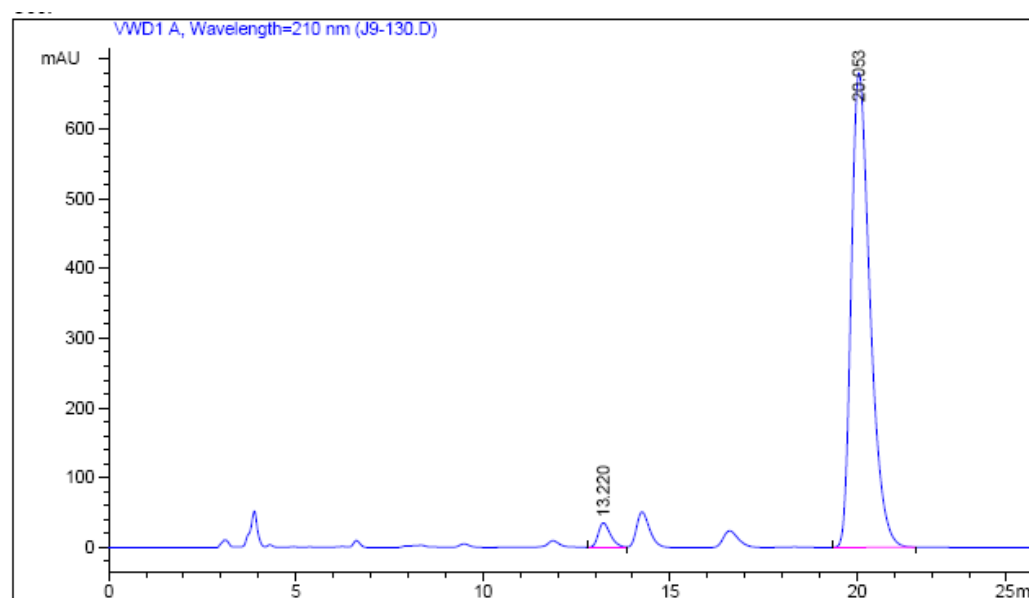
Totals : 1.19025e4 317.24911





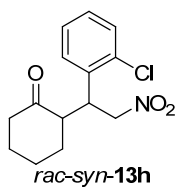
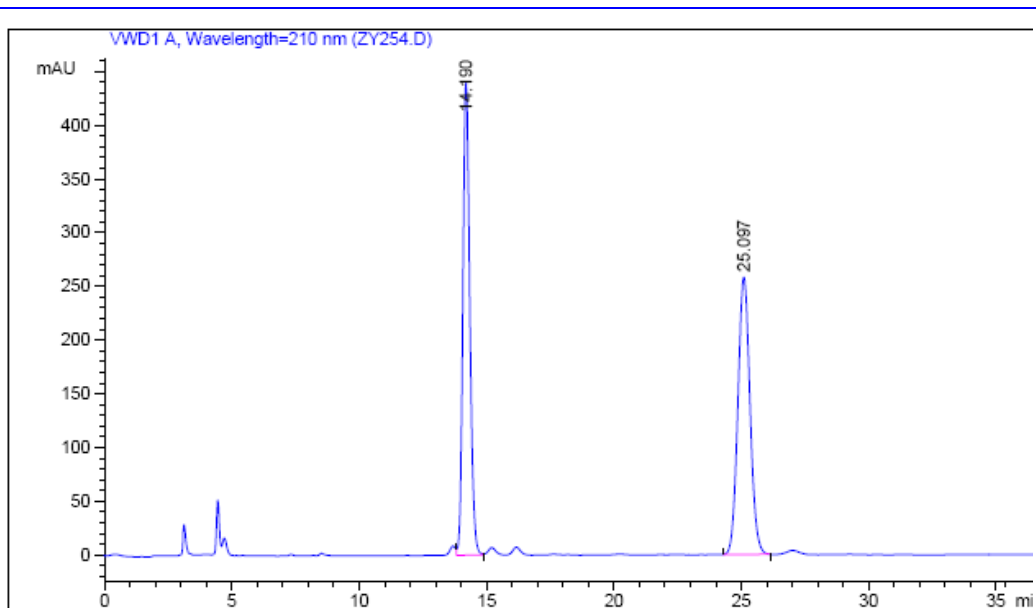
Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	12.887	VV	0.3474	9601.69727	421.28802	50.8179
2	19.751	BB	0.5215	9292.63574	271.55841	49.1821

Totals : 1.88943e4 692.84644

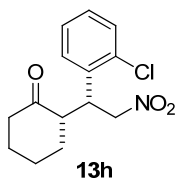
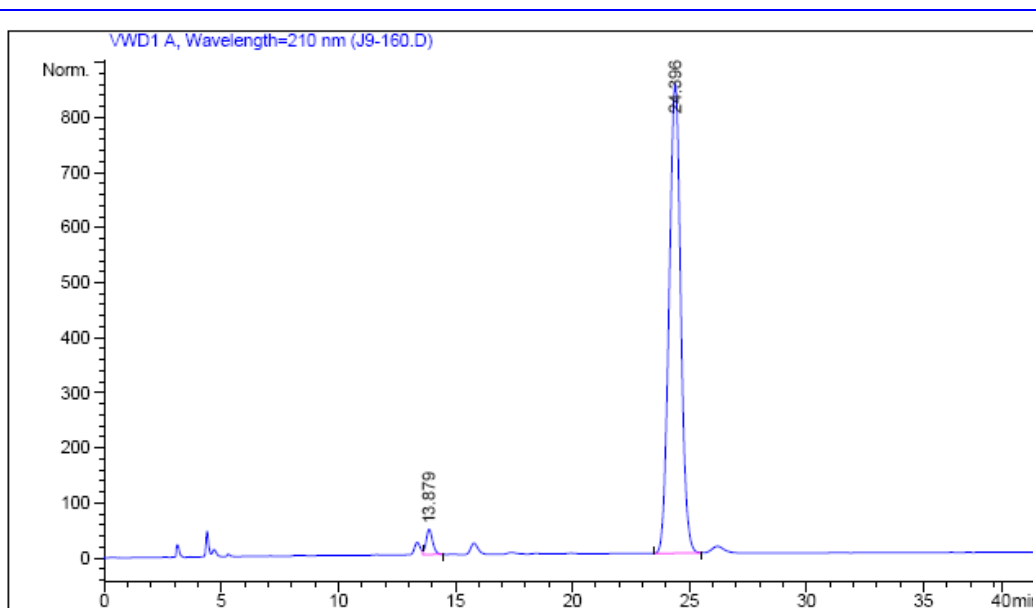


Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	13.220	BV	0.3488	805.84784	35.17484	3.2808
2	20.053	BB	0.5355	2.37569e4	680.41827	96.7192

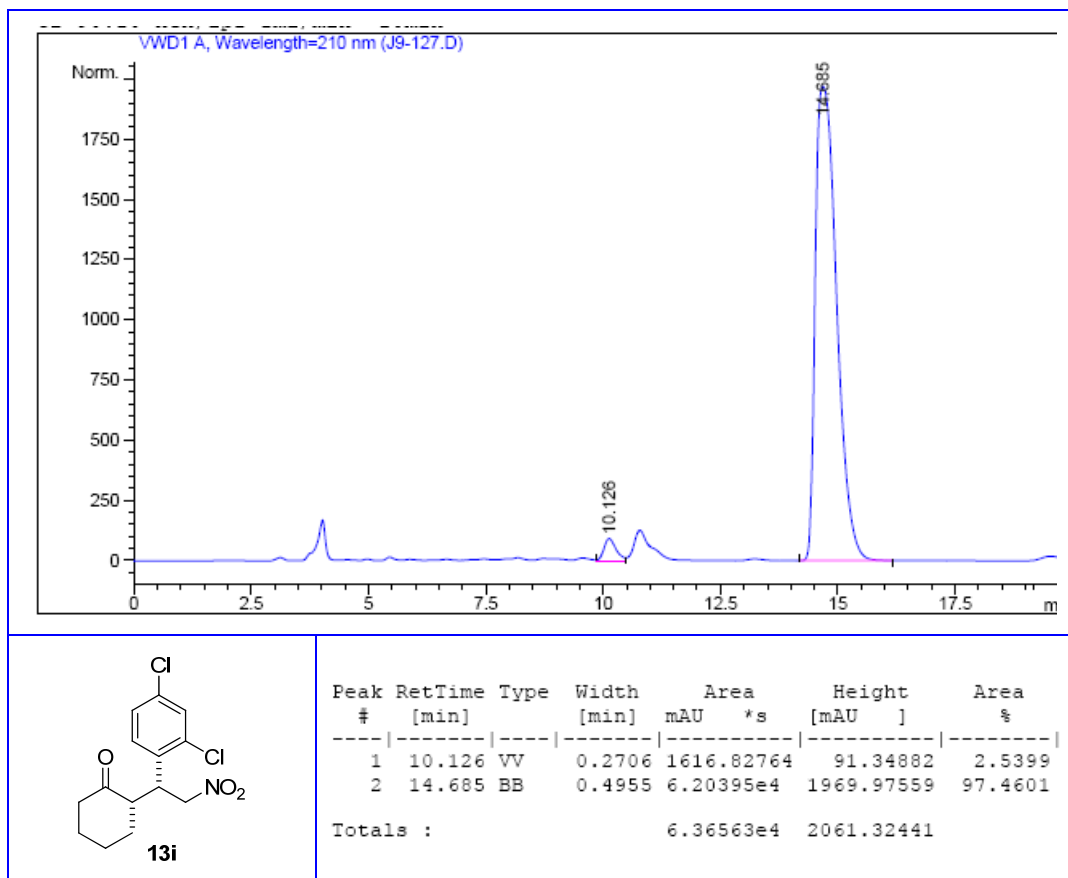
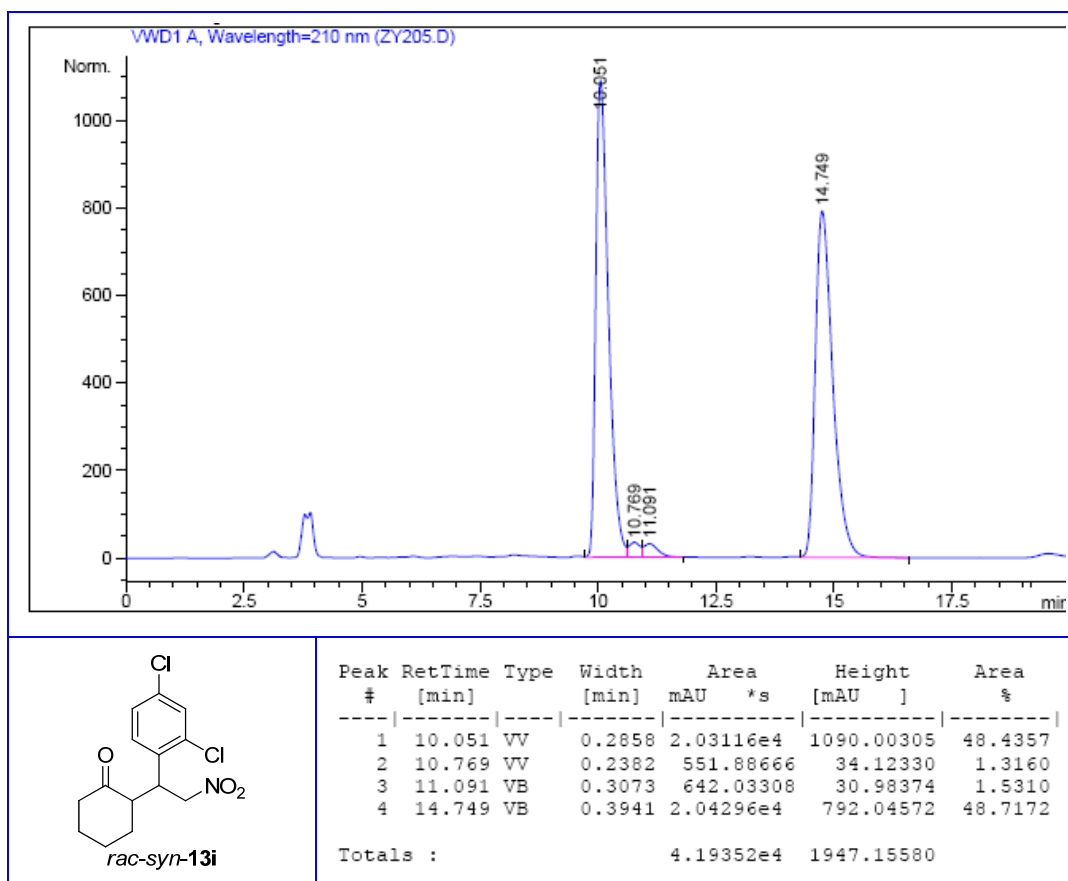
Totals : 2.45627e4 715.59311

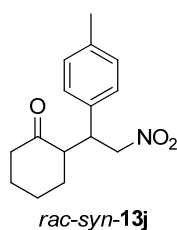
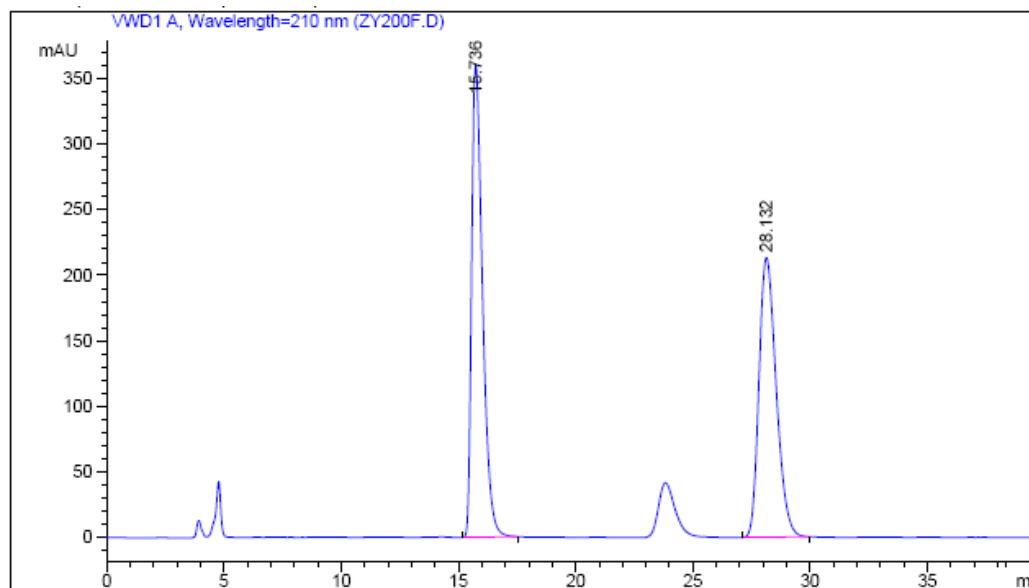


Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	14.190	VB	0.2916	8301.11816	439.61340	49.3708
2	25.097	BB	0.5116	8512.69629	257.97824	50.6292
Totals :				1.68138e4	697.59164	

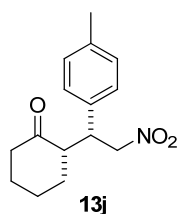
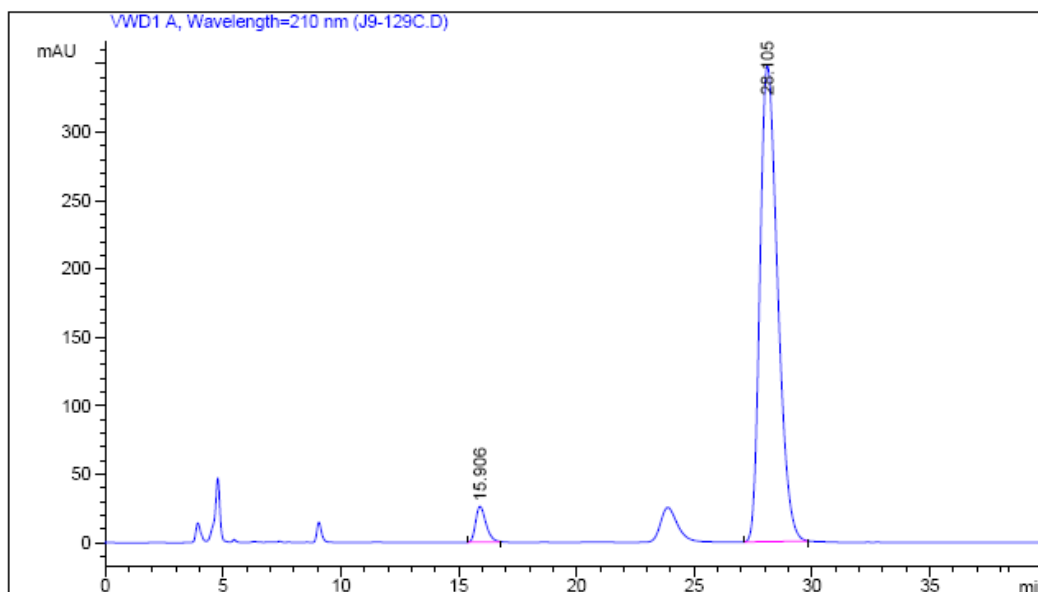


Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	13.879	VB	0.2945	891.68451	46.01762	2.9340
2	24.396	BB	0.5343	2.94997e4	853.37006	97.0660
Totals :				3.03914e4	899.38767	

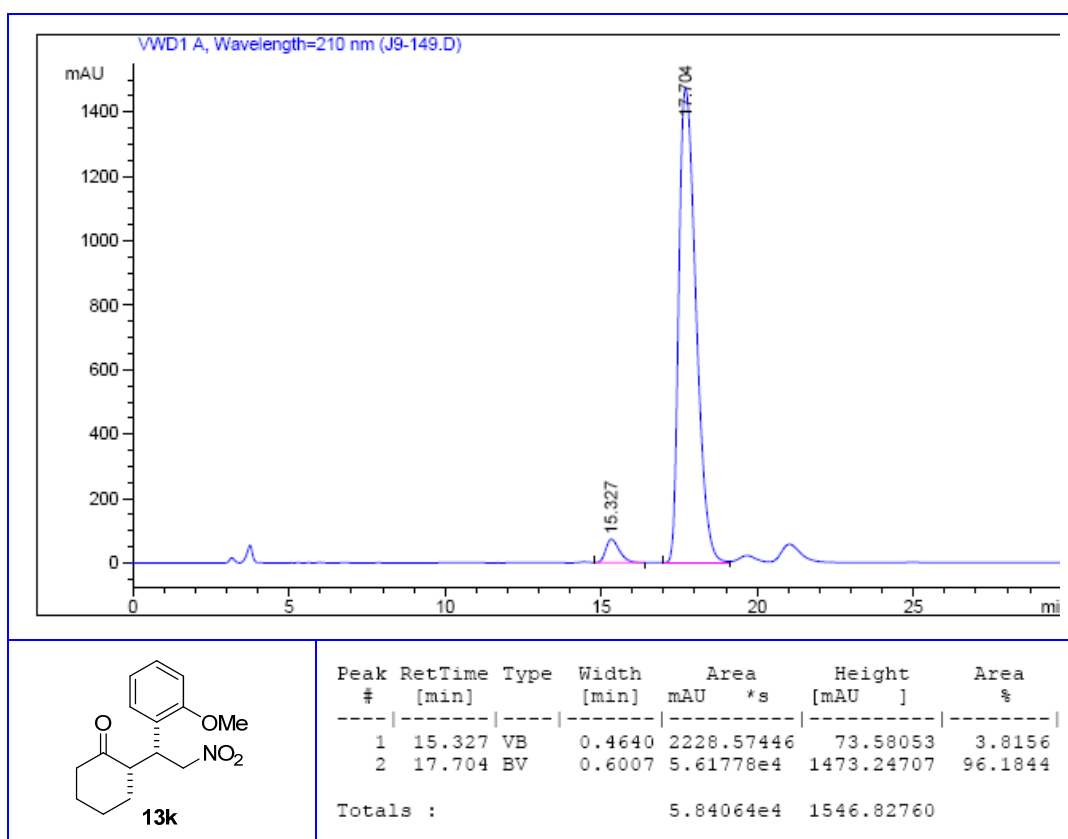
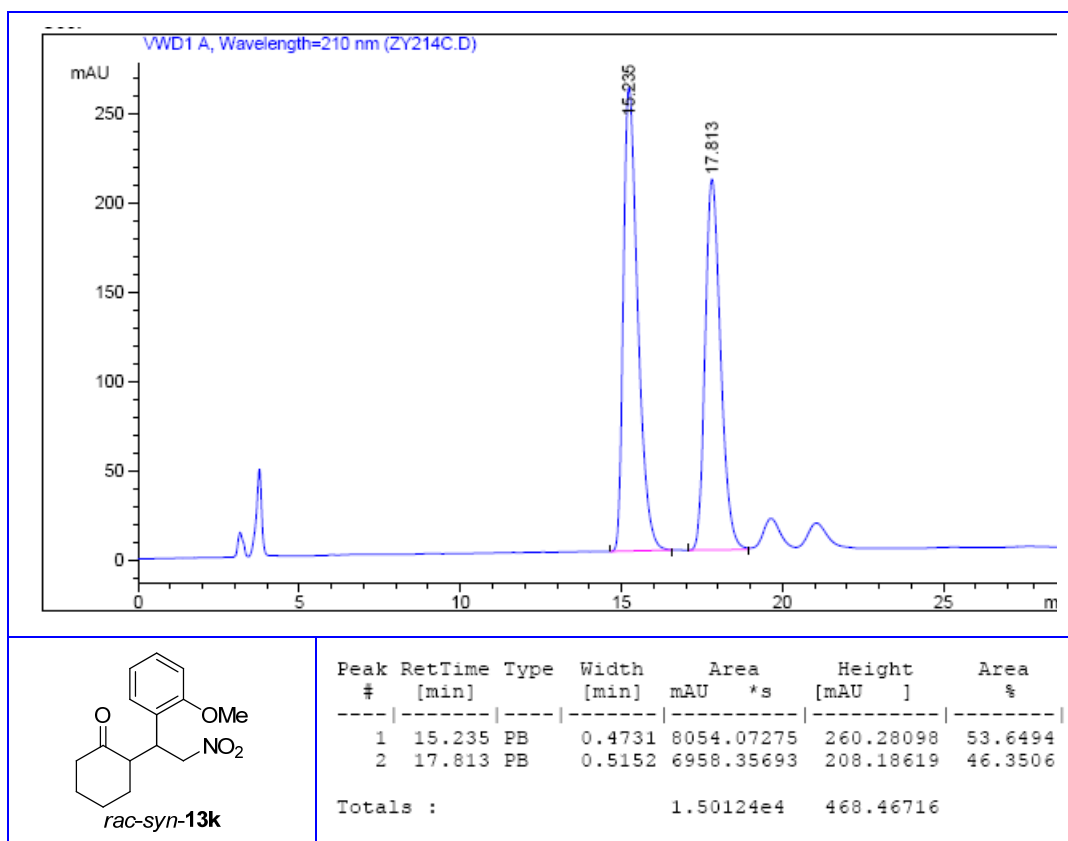


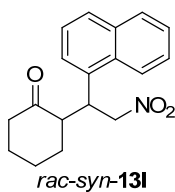
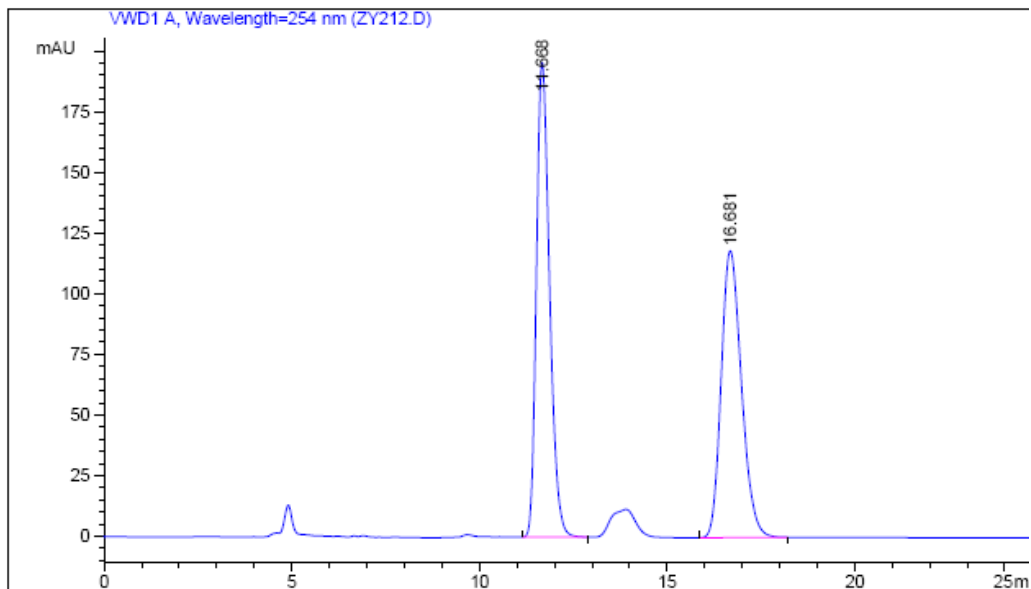


Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	15.736	BB	0.4924	1.16181e4	360.54147	51.7356
2	28.132	BB	0.7887	1.08386e4	213.04189	48.2644
Totals :				2.24567e4	573.58336	

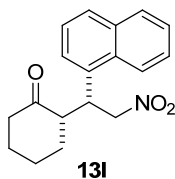
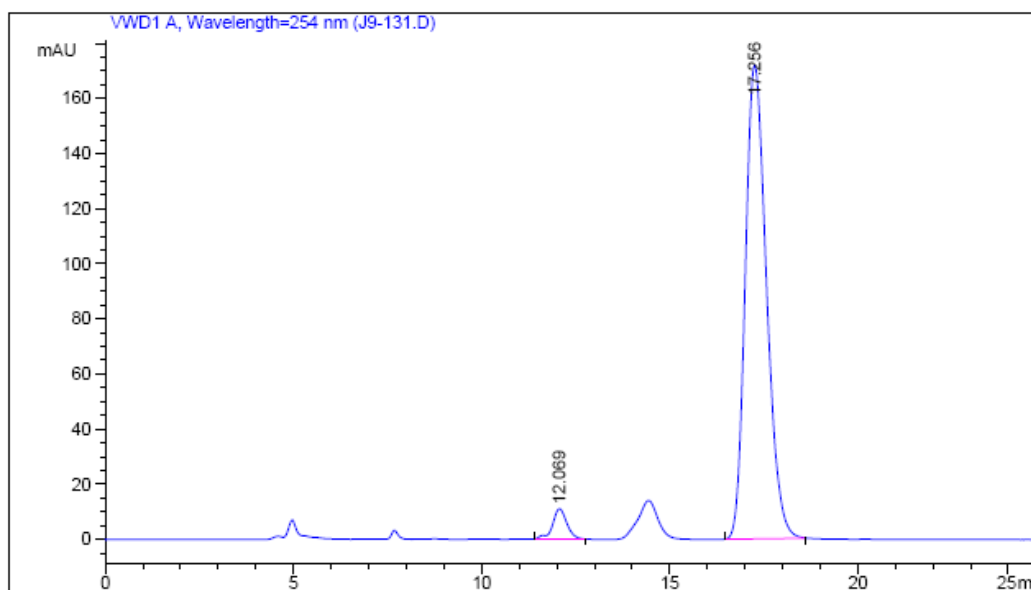


Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	15.906	BB	0.4634	797.02655	26.14407	4.2182
2	28.105	BB	0.8133	1.80981e4	348.09836	95.7818
Totals :				1.88951e4	374.24243	

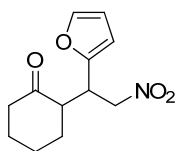
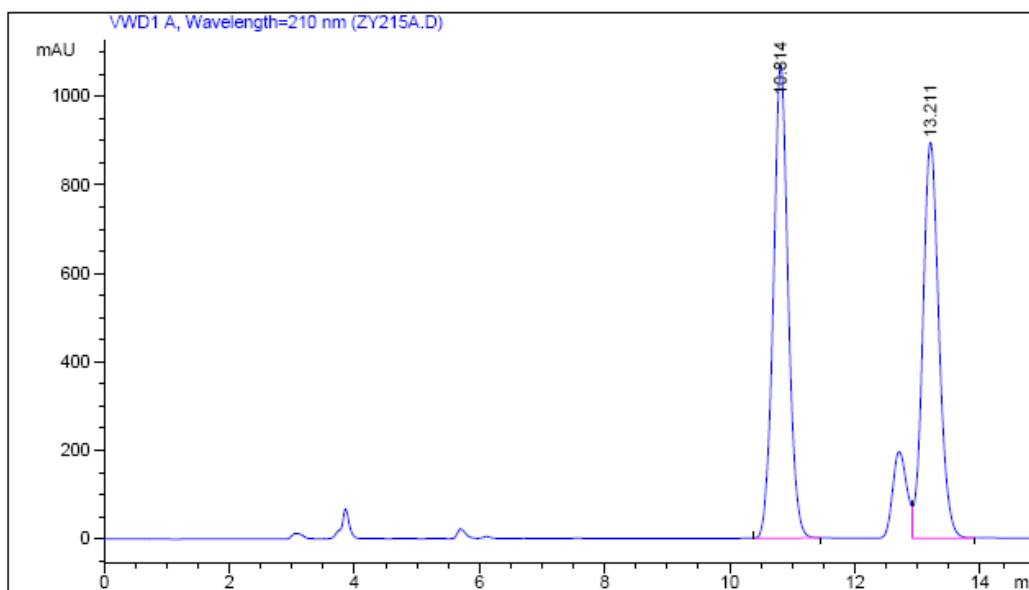




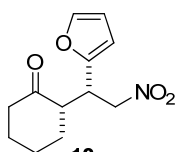
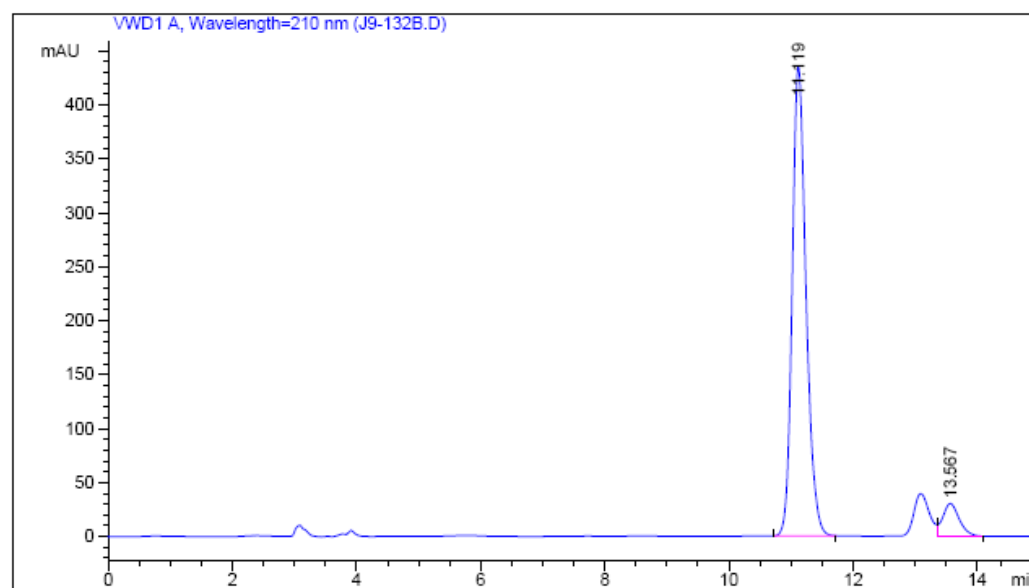
Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	11.668	BB	0.3742	4756.83350	195.47298	51.7687
2	16.681	BB	0.5817	4431.78662	118.15968	48.2313
Totals :				9188.62012	313.63265	



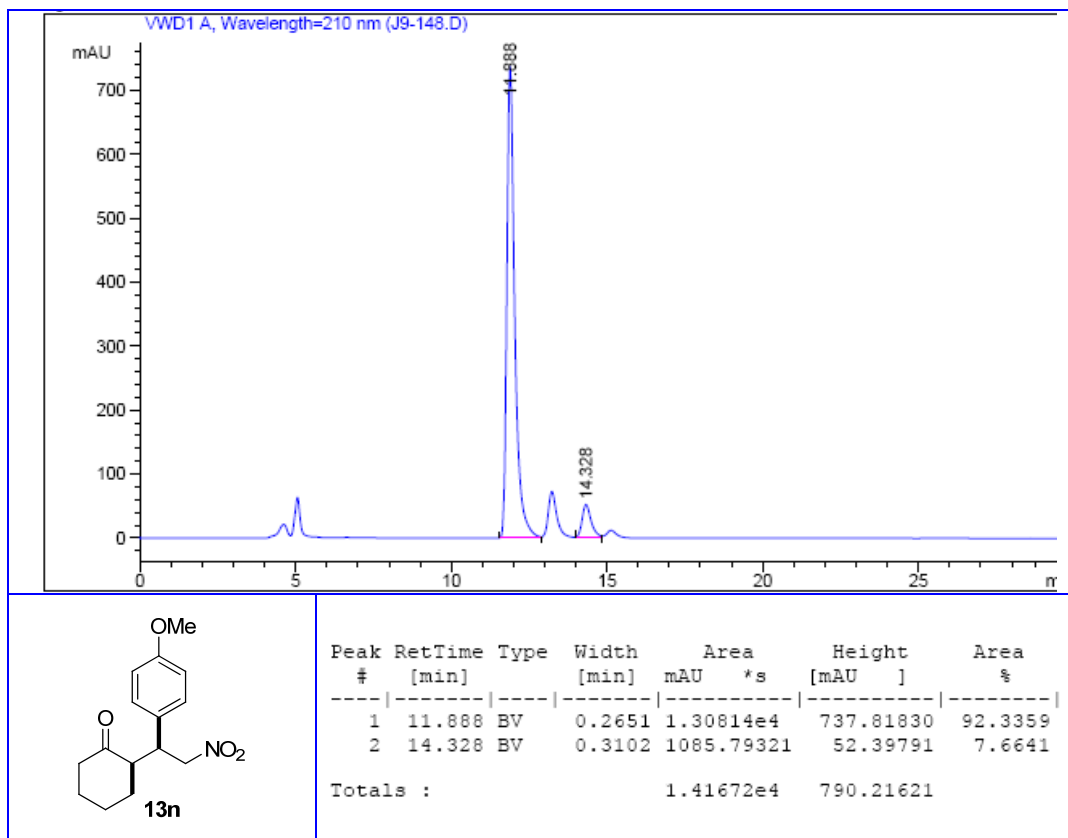
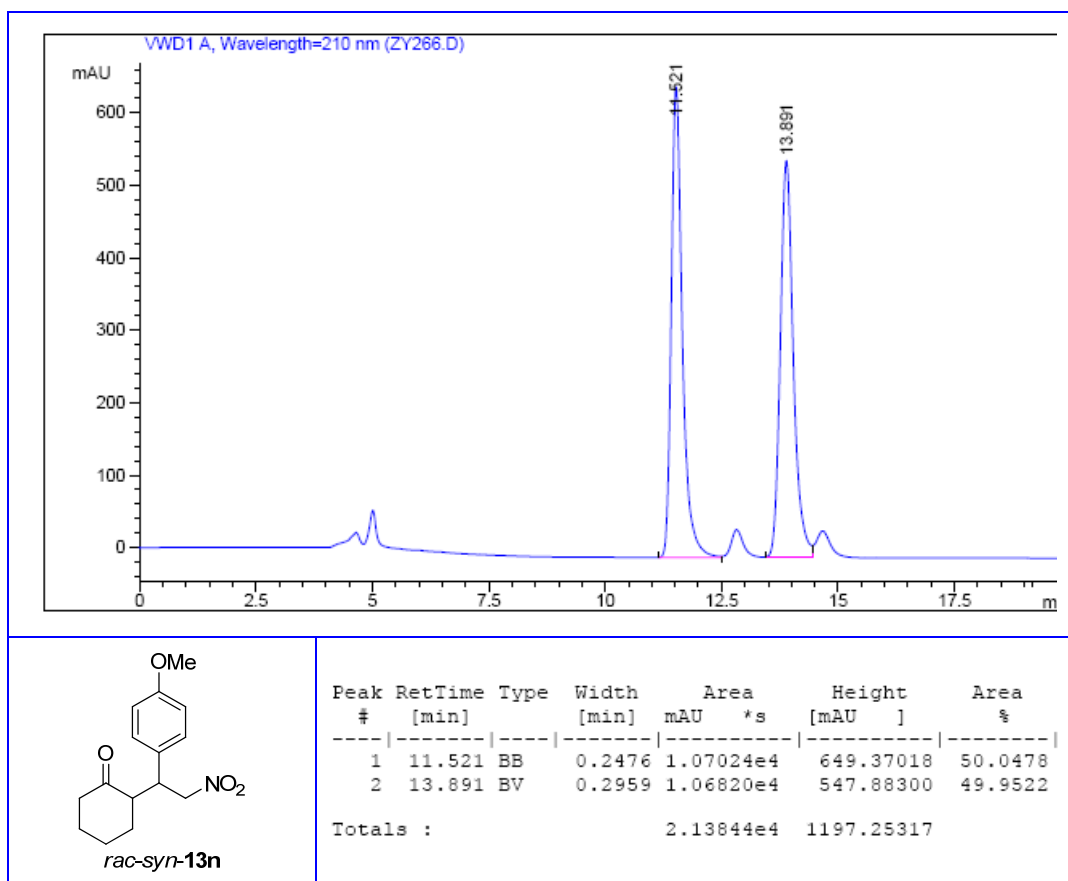
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1	12.069	BB	0.4053	294.62332	11.06538	4.1998
2	17.256	BB	0.6076	6720.60400	171.87505	95.8002
Totals :				7015.22733	182.94042	

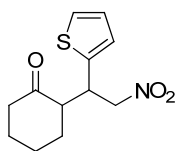
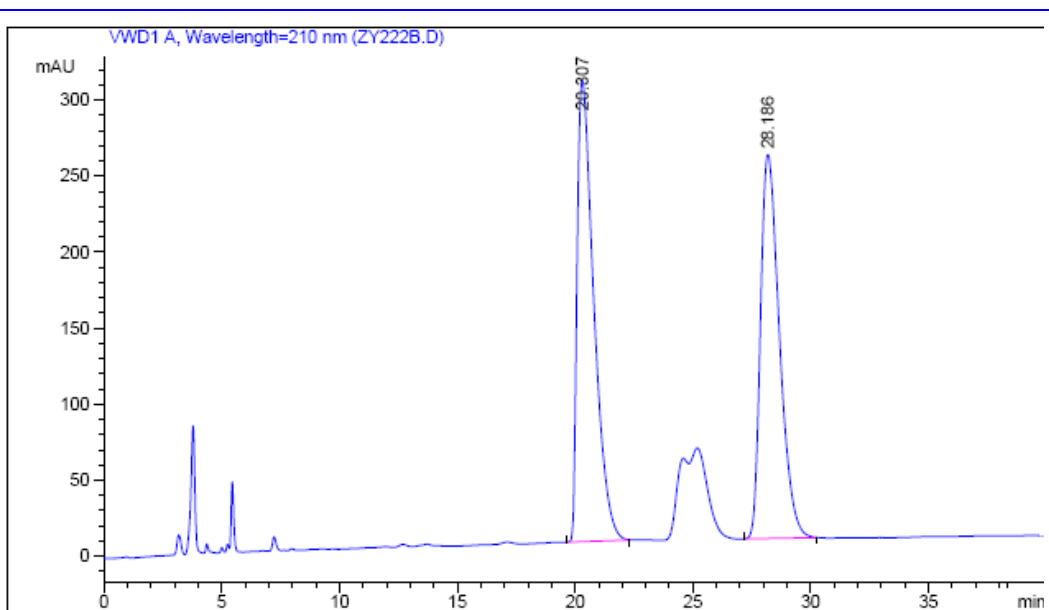


rac-syn-13m



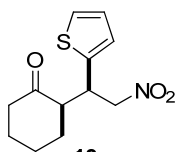
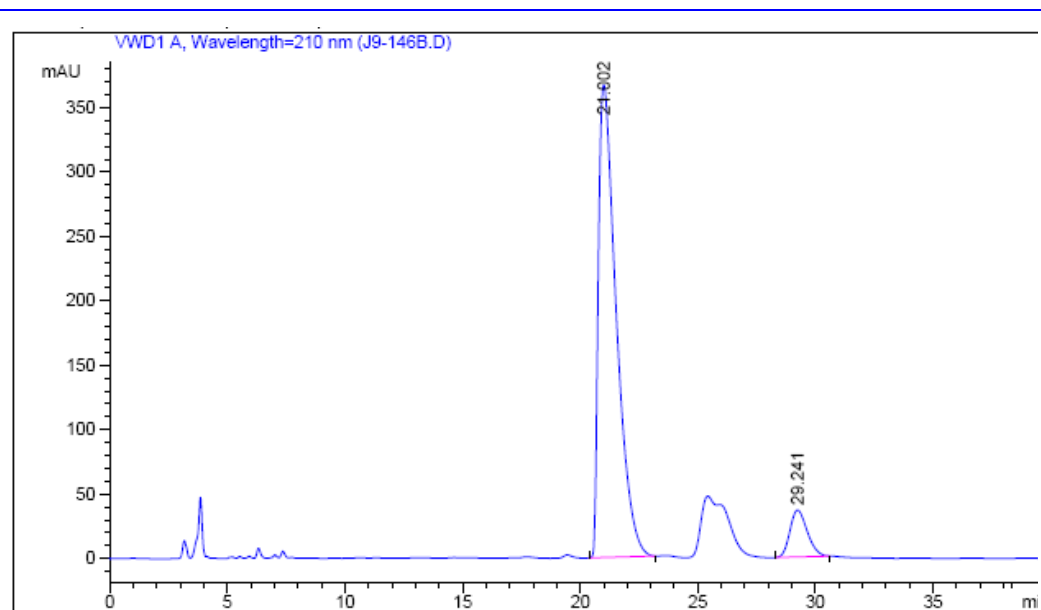
13m





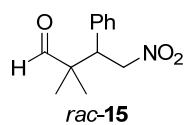
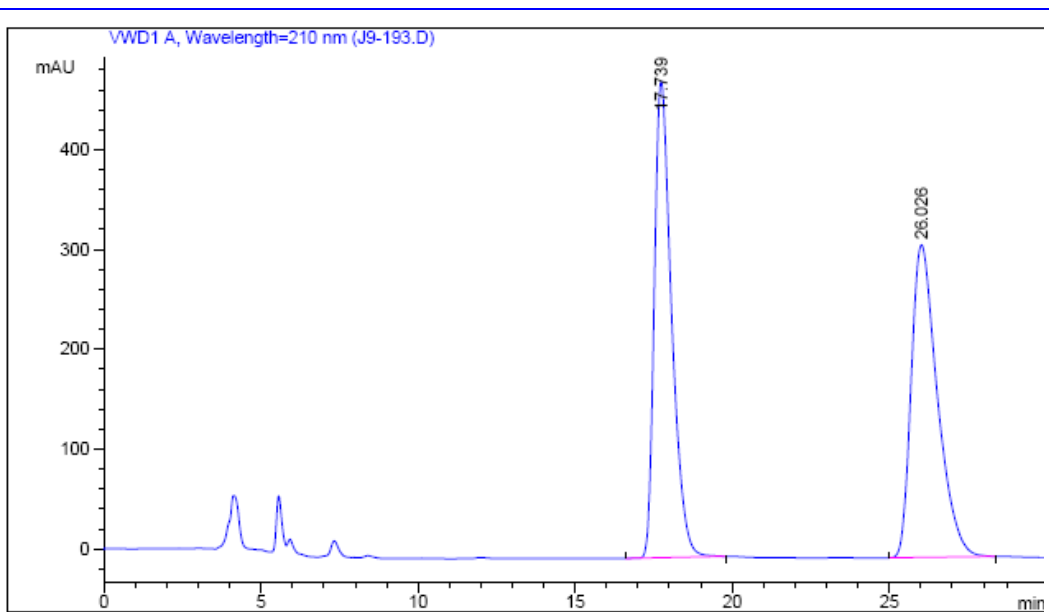
rac-syn-13o

Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	20.307	PB	0.7513	1.48770e4	303.33551	51.4211
2	28.186	BB	0.8602	1.40547e4	252.61896	48.5789
Totals :				2.89317e4	555.95447	

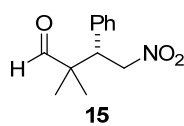
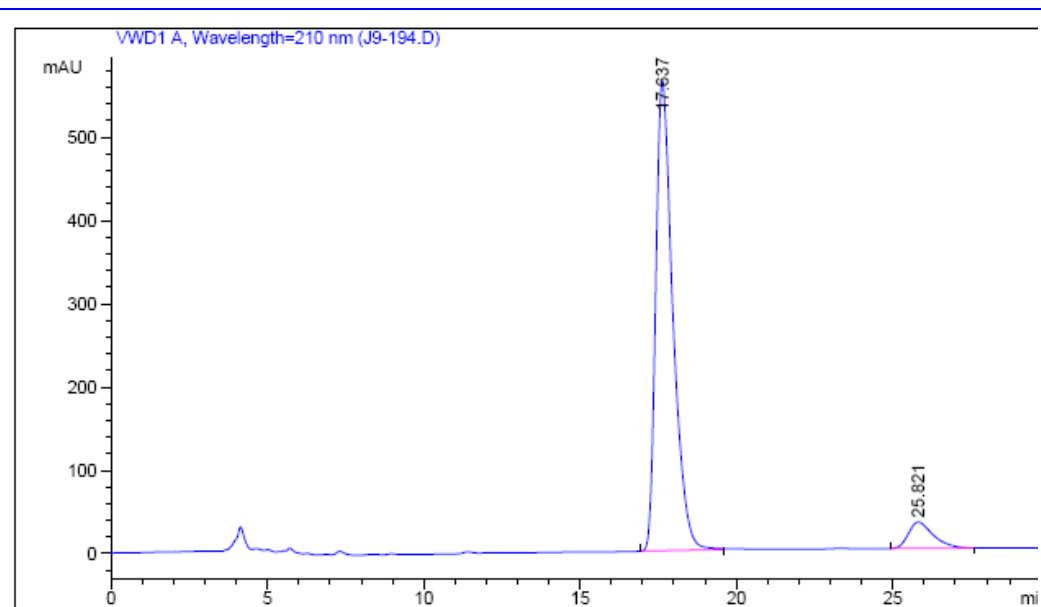


13o

Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	21.002	BP	0.7973	1.93005e4	366.81381	91.1598
2	29.241	PB	0.7905	1871.64771	36.40496	8.8402
Totals :				2.11721e4	403.21877	



Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	17.739	PB	0.5758	1.81896e4	477.16873	50.1249
2	26.026	PB	0.8756	1.80989e4	312.95905	49.8751
Totals :				3.62885e4	790.12778	



Peak #	RetTime [min]	Type	Width [min]	Area mAU *s	Height [mAU]	Area %
1	17.637	BB	0.5721	2.14428e4	563.48169	92.4643
2	25.821	PB	0.8327	1747.54810	31.54666	7.5357
Totals :				2.31903e4	595.02835	