Nitrile Assisted, Bronsted Acid Catalyzed Regio and Stereoselective Diarylphosphonylation of Allyl Silyl Ethers

Chun-Yu Ho,^a* Chun-Wa Chan,^a Siu-Kwan Wo,^b Zhong Zuo,^b* Lai-Ying Chan^a

^a Center of Novel Functional Molecules, and ^b School of Pharmacy, Faculty of Medicine,

The Chinese University of Hong Kong, Shatin, New Territories, Hong Kong SAR.

Email : jasonhcy@cuhk.edu.hk

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General Aspect.

p-Toluenesulfonic acid monohydrate and phosphorous trichloride were purchased from Acros Organics and used without further purification. Triphenyl phosphite was purchased from International Laboratory and used as received. Sodium hydride (60% in mineral oil) was purchased from Panreac Sintesis. Triethylamine and phosphorous trichloride were distilled over calcium hydride. All solvents were purchased from LAB-SCAN and used as received or dried according to the following procedures. THF: distilled over sodium/benzophenone. Toluene: distilled over calcium hydride. Analytical thin layer chromatography (TLC) was performed using EM Science silica gel 60 F254 plates. The developed chromatogram was analyzed by UV lamp (254 nm), ethanolic phosphomolybdic acid (PMA) or potassium permanganate (KMnO₄). Liquid chromatography was performed using a forced flow (flash chromatography) of the indicated solvent system on Silicycle Silica Gel (230–400 mesh). ¹H, ¹³C and ³¹P NMR spectra were recorded on Bruker 300 MHz or 400 MHz spectrometers in CDCl₃. Chemical shifts in ¹H NMR spectra are reported in ppm on the δ scale from an internal standard of residual tetramethylsilane (0 ppm). Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t =triplet, q = quartet, m = multiplet, br = broad), coupling constant in hertz (Hz), and integration. Chemical shifts of ¹³C NMR spectra are reported in ppm from the central peak of CDCl₃ (77.16 ppm) on the δ scale. Chemical shifts of ³¹P NMR spectra are reported in ppm referenced to internal P(OPh)₃ as standard on the δ scale (127.8 ppm with respect to 85% aq H₃PO₄ at 0 ppm, R. C. Seiceira, C. M. Higa, A. G. Barreto, J. F. Cajaiba da Silva, Thermochimica Acta 2005, 428, 101. The use of internal standard help us to eliminate minor deviations observed when using external 85% ag H₃PO₄ alone. The δ difference between the two phosphonylation regioisomers can be quite small without the internal standard.). Infrared (IR) spectra were recorded on a Perkin-Elmer Spectrum One FT-IR. High resolution mass spectra (HRMS) were obtained on a Finnigan MAT 95XL GC Mass Spectrometer by Miss. Ng, Hau Yan of the Chinese University of Hong Kong, Department of Chemistry.

2/ Preparation of Allyl Silyl Ethers 1a-k.

The synthesis of allyl silyl ethers **1a-1h**, **1j** and **1k** followed the literature procedure reported by Ho and Jamison.

Ho, C.-Y.; Jamison, T. F. Angew. Chem. Int. Ed. 2007, 46, 782-785.



The allylic alcohol below was synthesized by following the procedure reported by Takai. The allylic alcohol was treated with triethylsilyl trifluoromethanesulfonate in toluene to afford allyl silyl ether **1i**. Isolated yield: 82%.

Takai, K.; Sakamoto, S.; Isshiki, T. Org. Lett. 2003, 5, 653-655.



3/ General Procedure for the Preparation of Triarylphosphites.

The literature procedure reported by Hernández was followed with some modifications.

Hernández, J.; Goycoolea, F. M.; Zepeda-Rivera, D.; Juárez-Onofre, J.; Martínez, K.; Lizardi, J.; Salas-Reyes, M.; Gordillo, B.; Velázquez-Contreras, C.; García-Barradas, O.; Cruz-Sánchezd, S.; Domínguez, Z. *Tetrahedron* **2006**, *62*, 2520-2528.

A 50 mL round bottom flask was equipped with a magnetic stirrer. Under an atmosphere of nitrogen, para-substituted phenols (1.72 mmol, 300 mol%) and sodium hydride (2.16 mmol, 360 mol%) were added to 20 mL dry THF at 0 °C. After 10 min stirring, the solution was warmed to room temperature and continued to stir for 1 h. Then dry triethylamine (0.11 mmol, 20 mol%) and phosphorous trichloride (0.6 mmol, 100 mol%) were added. The mixture was stirred at room temperature for 20 h. After the removal of solvent under reduced pressure, 10 mL dry toluene was added to the solid residue. The milky white mixture was filtered through dry celite under nitrogen. The filtrate was concentrated and purified via flash chromatography on neutral aluminum oxide with dichloromethane (100 mL). It was then dried under reduced pressure at 90 °C for 1 h to afford the desired products as the colourless liquid.

4/ General Procedure for the Allyl Diarylphosphonylation and Arylation.

A mixture of 0.1 mmol substrate^{*} with 500 mol% of P(OPh)₃ and 5 mol% *p*-TsOH•H₂O in 2 mL CH₃CN (for phosphonylation) or 20 mol% *p*-TsOH•H₂O in 2 mL toluene (for arylation) was stirred at 0 °C for 2 h in open air and then stirred at r.t. for 9 h. Solvent was removed under reduced pressure. The yield and the selectivity (average of at least two runs) were determined by ¹H NMR analysis using benzaldehyde as standard. Purification via flash chromatography on silica gel (using chloroform as eluent for phosphonylation or 20 % ethyl acetate in hexane as eluent for arylation, unless otherwise indicated), afforded the desired product as oil. The stereochemistry of the olefin was determined by NOESY using isolated product.

^{*}Vigorous exclusion of other solvents (e.g. chloroform) is necessary, see S5.

Procedure: 1 mL CH₃CN was added to the substrate. Then solvent was removed under reduced pressure.

Procedure for Allyl Diarylphosphonylation of TES protected Baylis-Hillman adduct of methylacrylate and anisaldehyde:

General procedure for the allyl diarylphosphonylation of allyl silyl ethers was followed, except the reaction was conducted at 35 °C.

The TES protected Baylis-Hillman adduct of methyl acrylate and anisaldehyde was prepared according to literature procedure with modifications.



To a solution of 1 mmol Baylis-Hillman adduct in 2 mL toluene, 3 equiv. NEt₃ and 1.2 equiv. TESOTf was added at 0 °C and stir at rt for overnight. Quantative yield.

F. Coelho, W. P. Almeida, C. R. Mateus, L. D. Furtado, J. C. F. Gouveia, ARKIVOC, 443.

5/ Solvent Effect on Allyl Diarylphosphonylation.

	OTES	Cat. TsOH•H ₂ O P(OPh) ₃	P(0)(OF	P(O)(OPh) ₂	<i>p</i> -C ₆ H ₄ OH
p-Aı	nisyl n-Hex -	Solvent r.t.	P-Anisyl	p-Anisyl n-Hex -	<i>p</i> -Anisyl
	1a		2a	2a'	3a
	Entry ^a	Solvent	$\varepsilon_r^{\ b}$	Yield %	2a:2a' ^c
				$(2a+2a':3)^{c}$	
-	1^d	P(OPh) ₃	n.a.	1a recovered	-
	2	P(OPh) ₃	n.a.	90 (38:62)	75:25
	3	Toluene	2.38	85 (5:95)	n.d.
	4	NEt ₃	2.4	1a recovered	-
	5	CHCl ₃	4.8	97 (5:95)	n.d.
	6	THF	7.6	83 (5:95)	n.d.
	7	Acetone	20.7	83 (5:95)	n.d.
	8	CH ₃ NO ₂	35.9	90 (24:76)	75:25
	9	DMF	36.7	89 (14:86)	78:22
-	10	CH ₃ CN	37.5	95 (87:13)	75:25
	11	PhCN	26.0	79 (81:19)	78:22

^{*a*} The standard procedure was followed except that the reactions were carried out at r.t. in parallel, using 20 mol% of *p*-TsOH•H₂O and indicated solvent; ^{*b*} Dielectric constant; ^{*c*} Yield and selectivity were determined by ¹H NMR analysis using benzaldehyde as standard, the olefin stereochemistry was determined as *E*-isomer by NOESY using isolated product; ^{*d*} Control experiment without adding *p*-TsOH·H₂O and run in 500 mol% of P(OPh)₃.

Dielectric constant:

Landolt-Börnstein: Numerical Data and Functional Relationships in Science and Technology -New Series; Wohlfarth, C., Lechner, M. D., Eds; Group 4: Physical Chemistry; Springer Berlin Heidelberg, 2008; Vol.17.

6/ Phosphonylation Experiment using HP(O)(OPh)2 in place of P(OPh)3.



^{*a*} The standard procedure was followed, except that HP(O)(OPh)₂ was used in place of P(OPh)₃; ^{*b*} Yield was determined by ¹H NMR analysis using benzaldehyde as standard; ^{*c*} Olefin stereochemistry of **3a** was determined by NOESY using isolated product.

7/ Competition Experiments using P(OPh)₃, HP(O)(OPh)₂ and PhOH in Catalytic Allyl <u>Arylation.</u>

	p-Anisyl	TES Cat. TsOH•H ₂ O Additive Toluene	H p-Anisyl	DH	
1a			3a		
Entry ^a	Additive	Condition	Conversion	Yield %	$E:Z^c$
			%	$(3)^b$	
1	P(OPh) ₃	0 °C, 2h	0	0	n.a.
2		0 °C, 2h and then r.t.,	92	78	>95:5
		2h			
3	HP(O)(OPh) ₂	0 °C, 2h	81	33	>95:5
4		0 °C, 2h and then r.t.,	87	38	>95:5
		2h			
5	PhOH	0 °C, 2h	100	92	>95:5

^{*a*} The standard procedure was followed, except that HP(O)(OPh)₂ or PhOH was used as indicated above; ^{*b*} Yield was determined by ¹H NMR analysis using benzaldehyde as standard; ^{*c*} Olefin stereochemistry was determined by NOESY using isolated product.

8/ Triarylphosphites.

The spectroscopic data was comparable with the literature:

Nikoletta, S.; Stratakis, M. ARKIVOC 2003, 30-35.

Tris(4-methylphenyl) phosphite P1

The standard procedure was followed, except that 4-methylphenol was used. Yield: 92%. ¹H NMR (400 MHz, CDCl₃) δ : 7.10 (d, *J* = 8.2 Hz, 6H), 7.02 (d, *J* = 8.2 Hz, 6H), 2.31 (s, 9H). ¹³C NMR (100 MHz, CDCl₃) δ : 149.47, 133.78, 130.26, 120.65 (d, *J* = 7 Hz, ortho), 20.87. ³¹P NMR (121 MHz, CDCl₃) δ : 128.11.

Tris(4-chlorophenyl) phosphite **P2**



The standard procedure was followed, except that 4-chlorophenol was used. Yield: 88%.

¹H NMR (300 MHz, CDCl₃) δ : 7.29 (d, J = 9.0 Hz, 6H), 7.04 (d, J = 9.0 Hz, 6H).

¹³C NMR (100 MHz, CDCl₃) δ: 130.29, 129.96, 129.56, 122.00 (d, J = 5 Hz, ortho), 116.80.

³¹P NMR (121 MHz, CDCl₃) δ: 126.43.

HRMS-EI (m/z): $[M]^+$ calcd for C₁₈H₁₂Cl₃O₃P, 412.9662; found, 412.9692.

9/ Phosphonylation products (Table 1).

Table 1, entry 1:



(major, more polar)

(minor, less polar)

The standard procedure was followed.

HRMS-ESI (m/z): $[M+Na]^+$ calcd for C₂₈H₃₃O₄PNa, 487.2009; found, 487.2013.

2a: ¹H NMR (400 MHz, CDCl₃) δ: 7.33-7.29 (m, 5H), 7.19-7.10 (m, 7H), 6.86 (d, J = 8.7 Hz, 2H), 6.48 (d, J = 6.1 Hz, 1H), 3.82 (s, 3H), 3.05 (d, J = 22.3 Hz, 2H), 2.47-2.42 (m, 2H), 1.54-1.42 (m, 2H), 1.35-1.25 (m, 6H), 0.88-0.83 (m, 3H).

¹³C NMR (100 MHz, CDCl₃) δ: 158.21, 150.54, 150.45, 131.07, 130.95, 130.65, 130.52, 129.68, 124.98, 120.50, 120.46, 113.53, 55.19, 34.46 (d, *J* = 137 Hz), 31.54, 31.27, 29.13, 27.88, 22.54, 14.00.

IR (neat) cm⁻¹: 2920, 2850, 1725, 1592, 1509, 1488, 1273, 1249, 1214, 1189, 1025, 927, 759. ³¹P NMR (121 MHz, CDCl₃) δ: 20.36.

2'a: ¹H NMR (400 MHz, CDCl₃) δ: 7.42 (d, *J* = 8.8 Hz, 2H), 7.31-7.02 (m, 8H), 6.86 (d, *J* = 8.8 Hz, 2H), 6.84 (d, *J* = 8.8 Hz, 2H), 5.69 (d, *J* = 2.8 Hz, 1H), 5.19 (d, *J* = 2.8 Hz, 1H), 4.06 (d, *J* = 24.8 Hz, 1H), 3.80 (s, 3H), 2.07-1.90 (m, 2H), 1.41-1.37 (m, 2H), 1.36-1.20 (m, 6H), 0.89-0.83 (m, 3H).

IR (neat) cm⁻¹: 2926, 2855, 1591, 1509, 1489, 1271, 1214, 1188, 1162, 1025, 927, 759, 688, 617, 589, 500.

³¹P NMR (121 MHz, CDCl₃) δ: 18.42.

Table 1, entry 5:



The standard procedure was followed.

HRMS-ESI (m/z): [M+Na]⁺ calcd for C₂₆H₂₉O₄PNa, 459.1696; found, 459.1691.

2b: ¹H NMR (400 MHz, CDCl₃) δ: 7.33-7.29 (m, 4H), 7.20-7.11 (m, 8H), 6.85 (d, J = 8.7 Hz, 2H), 6.57 (d, J = 6.3 Hz, 1H), 3.81 (s, 3H), 3.06 (d, J = 22.4 Hz, 2H), 2.38 (dd, J = 7.4, 2.4 Hz, 2H), 1.75-1.97 (m, 1H), 0.84 (d, J = 6.6 Hz, 6H).

¹³C NMR (100 MHz, CDCl₃) δ: 158.33, 150.74, 150.65, 132.04, 131.91, 130.43, 130.21, 130.18, 129.89, 125.18, 120.69, 120.64, 113.66, 55.37, 39.76, 34.84 (d, J = 137 Hz), 26.69, 22.48. IR (neat) cm⁻¹: 2921, 2851, 1593, 1510, 1490, 1464, 1270, 1250, 1215, 1190, 1026, 930, 765, 689. ³¹P NMR (121 MHz, CDCl₃) δ: 20.41.

2'b: ¹H NMR (400 MHz, CDCl₃) δ: 7.42 (d, *J* = 8.8 Hz, 2H), 7.31-7.02 (m, 6H), 7.05 (d, *J* = 8.0 Hz, 2H), 6.86 (d, *J* = 8.0 Hz, 2H), 6.72 (d, *J* = 8.8 Hz, 2H), 5.76 (d, *J* = 2.8 Hz, 1H), 5.18 (d, *J* = 2.8 Hz, 1H), 4.04 (d, *J* = 25.2 Hz, 1H), 3.80 (s, 3H), 2.35 (d, *J* = 7.4 Hz, 2H), 1.70-1.81 (m, 1H), 0.84 (d, *J* = 6.6 Hz, 6H).

IR (neat) cm⁻¹: 2954, 2923, 2852, 1727, 1593, 1510, 1490, 1465, 1384, 1274, 1250, 1214, 1190, 1163, 1072, 1026, 930, 760, 689, 617.

³¹P NMR (121 MHz, CDCl₃) δ: 19.02.

Table 1, entry 6:



The standard procedure was followed.

HRMS-ESI (m/z): [M+Na]⁺ calcd for C₂₈H₃₁O₄PNa, 485.1852; found, 485.1845.

2c: ¹H NMR (400 MHz, CDCl₃) δ: 7.32-7.28 (m, 5H), 7.19-7.10 (m, 7H), 6.87 (d, *J* = 8.8 Hz, 2H), 6.72 (d, *J* = 5.2 Hz, 1H), 3.82 (s, 3H), 3.01 (d, *J* = 22.8 Hz, 2H), 2.84-2.78 (m, 1H), 1.76-1.08 (m, 10H).

¹³C NMR (100 MHz, CDCl₃) δ: 158.34, 150.79, 150.69, 135.09, 134.99, 130.14, 130.01, 129.99, 129.87, 129.75, 129.65, 125.14, 120.76, 120.72, 113.73, 55.39, 40.37, 40.31, 31.11, 30.01, 29.23
(d, *J* = 123 Hz), 26.21, 26.08.

IR (neat) cm⁻¹: 2918, 2849, 1724, 1589, 1506, 1488, 1270, 1248, 1214, 1189, 1025, 926, 760, 688. ³¹P NMR (121 MHz, CDCl₃) δ: 21.55.

2'c: ¹H NMR (400 MHz, CDCl₃) δ: 7.43 (d, *J* = 8.8 Hz, 2H), 7.30-7.03 (m, 8H), 6.86 (d, *J* = 8.8 Hz, 2H), 6.79 (d, *J* = 8.6 Hz, 2H), 5.77 (d, *J* = 3.2 Hz, 1H), 5.23 (d, *J* = 3.2 Hz, 1H), 4.12 (d, *J* = 25.3 Hz, 1H), 3.80 (s, 3H), 1.88-1.86 (m, 1H), 1.74-1.05 (m, 10H).

¹³C NMR (100 MHz, CDCl₃) δ: 159.20, 150.93, 150.73, 148.88, 131.21, 131.13, 129.99, 129.87, 129.70, 129.55, 125.00, 124.85, 120.74, 120.64, 114.10, 113.88, 113.81, 55.42, 48.85 (d, *J* = 138 Hz), 45.74, 45.63, 32.80, 32.35, 26.83, 26.68, 26.33.

IR (neat) cm⁻¹: 2919, 2850, 1725, 1591, 1509, 1489, 1454, 1251, 1213, 1187, 1161, 1070, 1025, 927, 759, 678.

³¹P NMR (121 MHz, CDCl₃) δ: 18.67.

Table 1, entry 7:



The standard procedure was followed.

HRMS-ESI (m/z): [M+Na]⁺ calcd for C₂₈H₂₅O₄PNa, 479.1383; found, 479.1412.

2d: ¹H NMR (400 MHz, CDCl₃) δ: 7.32-7.24 (m, 9H), 7.13 (dd, *J* = 8.0, 7.6 Hz, 2H), 7.03 (d, *J* = 8.4 Hz, 4H), 6.86 (d, *J* = 8.8 Hz, 2H), 6.68 (d, *J* = 6.0 Hz, 1H), 6.64 (d, *J* = 8.8 Hz, 2H), 3.73 (s, 3H), 3.38 (d, *J* = 22.0 Hz, 2H).

¹³C NMR (75 MHz, CDCl₃) δ: 159.05, 151.03, 150.91, 140.97, 132.33, 132.16, 130.97, 130.26, 129.64, 129.32, 128.05, 125.59, 121.09, 121.03, 113.95, 55.72, 38.12 (d, J = 137 Hz). IR (neat) cm⁻¹: 2918, 2850, 1722, 1589, 1509, 1488, 1274, 1250, 1213, 1188, 1024, 927, 758, 689. ³¹P NMR (121 MHz, CDCl₃) δ: 19.46.

2'd: ¹H NMR (400 MHz, CDCl₃) δ: 7.51-7.49 (m, 2H), 7.45-7.29 (m, 4H), 7.23-7.14 (m, 4H), 7.09-7.07 (m, 3H), 6.90-6.86 (m, 4H), 6.80 (d, *J* = 8.5 Hz, 2H), 6.15 (d, *J* = 3.1 Hz, 1H) 5.71 (d, *J* = 3.1 Hz, 1H) 4.63 (d, *J* = 25.6 Hz, 1H), 3.79 (s, 3H).

IR (neat) cm⁻¹: 2918, 2850, 1725, 1590, 1509, 1488, 1465, 1384, 1252, 1212, 1182, 1025, 927, 761, 688, 500.

³¹P NMR (121 MHz, CDCl₃) δ: 17.88.

Table 1, entry 8:



The standard procedure was followed.

HRMS-ESI (m/z): [M+Na]⁺ calcd for C₂₉H₂₇O₄PNa, 493.1539; found, 493.1529.

2e: ¹H NMR (400 MHz, CDCl₃) δ: 7.37-7.14 (m, 17H), 6.86 (d, *J* = 8.7 Hz, 2H), 6.73 (d, *J* = 6.1 Hz, 1H), 3.88 (d, *J* = 2.4 Hz, 2H), 3.79 (s, 3H), 2.92 (d, *J* = 22.1 Hz, 2H).

¹³C NMR (75 MHz, CDCl₃) δ: 158.76, 150.71, 150.59, 138.84, 132.66, 132.49, 130.00, 129.89, 128.93, 129.77, 126.53, 125.74, 125.20, 120.67, 120.61, 113.91, 55.38, 37.13, 34.20 (d, J = 137 Hz).

IR (neat) cm⁻¹: 3061, 3027, 2924, 2836, 1591, 1510, 1490, 1455, 1270, 1251, 1214, 1189, 1162, 1026, 930, 761, 689.

³¹P NMR (121 MHz, CDCl₃) δ: 20.08.

2'e: ¹H NMR (400 MHz, CDCl₃) δ: 7.43 (d, *J* = 8.8 Hz, 2H), 7.37-7.01 (m, 13H), 6.86 (d, *J* = 8.7 Hz, 2H), 6.71 (d, *J* = 7.2 Hz, 2H), 5.81 (d, *J* = 2.8 Hz, 1H), 5.21 (d, *J* = 2.8 Hz, 1H), 4.00 (d, *J* = 25.2 Hz, 1H), 3.80 (s, 3H), 3.49 (d, *J* = 15.2 Hz, 1H), 3.27 (d, *J* = 15.2 Hz, 1H). IR (neat) cm⁻¹: 2954, 2922, 2851, 1723, 1590, 1509, 1489, 1463, 1384, 1250, 1213, 1188, 1073, 1025, 928, 823, 758, 689.

³¹P NMR (121 MHz, CDCl₃) δ: 17.95.

Table 1, entry 9:



(major, more polar)

(minor, less polar)

The standard procedure was followed.

HRMS-ESI (m/z): [M+Na]⁺ calcd for C₃₀H₂₉O₄PNa, 507.1696; found, 507.1688.

2f: ¹H NMR (400 MHz, CDCl₃) δ: 7.32-7.11 (m, 15H), 7.06 (d, *J* = 8.8 Hz, 2H), 6.84 (d, *J* = 8.8 Hz, 2H), 6.52 (d, *J* = 6.2 Hz, 1H), 3.81 (s, 3H), 3.05 (d, *J* = 22.2 Hz, 2H), 2.84-2.77 (m, 4H).

¹³C NMR (75 MHz, CDCl₃) δ: 158.50, 150.67, 150.55, 141.35, 131.90, 131.72, 129.91, 128.52, 126.16, 125.25, 120.70, 120.64, 113.77, 55.40, 35.02 (d, *J* = 137 Hz), 33.25.

IR (neat) cm⁻¹: 2922, 2852, 1592, 1509, 1490, 1273, 1250, 1204, 1190, 1026, 929, 760, 689.

³¹P NMR (121 MHz, CDCl₃) δ: 20.09.

2'f: ¹H NMR (400 MHz, CDCl₃) δ: 7.42 (d, *J* = 8.8 Hz, 2H), 7.20-7.02 (m, 13H), 6.86 (d, *J* = 8.4 Hz, 2H), 6.74 (d, *J* = 8.4 Hz, 2H), 5.74 (d, *J* = 2.4 Hz, 1H), 5.23 (d, *J* = 2.4 Hz, 1H), 4.01 (d, *J* = 25.2 Hz, 1H), 3.82 (s, 3H), 2.52-2.41 (m, 2H), 2.38-2.28 (m, 2H).

IR (neat) cm⁻¹: 2919, 2851, 1724, 1591, 1509, 1489, 1465, 1384, 1274, 1252, 1213, 1189, 1073, 1025, 929, 823, 759, 689.

³¹P NMR (121 MHz, CDCl₃) δ: 17.98.

Table 1, entry 10:



The standard procedure was followed, except that CH_3CN was distilled over calcium hydride before use, 100 mol% *p*-TsOH•H₂O was used, the reaction mixture was stirred at 35 °C for 18 hours and the product was purified with 5% diethyl ether in toluene. Isolated as a mixture of **2g** and **2'g**.

HRMS-ESI (m/z): [M+Na]⁺ calcd for C₂₈H₃₃O₄PNa, 487.2009; found, 487.2039.

2g: ¹H NMR (400 MHz, CDCl₃) δ: 7.37-6.82 (m, 14H), 6.61 (d, *J* = 6.2 Hz, 1H), 3.78 (s, 3H), 3.11 (d, *J* = 22.3 Hz, 2H), 2.43-2.38 (m, 2H), 1.49-1.44 (m, 2H), 1.36-1.21 (m, 6H), 0.86-0.81 (m, 3H).

2'g: ¹H NMR (400 MHz, CDCl₃) δ: 7.75 (d, *J* = 7.8 Hz, 2H), 7.37-6.82 (m, 10H), 6.77 (d, *J* = 8.6 Hz, 2H), 5.67 (d, *J* = 2.9 Hz, 1H), 5.16 (d, *J* = 2.9 Hz, 1H), 4.88 (d, *J* = 25.4 Hz, 1H), 3.77 (s, 3H), 2.17-1.99 (m, 2H), 1.42-1.38 (m, 2H), 1.36-1.21 (m, 6H), 0.86-0.81 (m, 3H).

2g+**2'g**: ¹³C NMR (100 MHz, CDCl₃) δ : 157.26, 157.10, 150.99, 150.90, 150.74, 150.64, 144.29, 132.36, 132.25, 130.85, 130.79, 130.01, 129.82, 129.67, 129.42, 128.85, 128.34, 127.28, 127.15, 126.52, 125.76, 125.14, 124.94, 124.69, 123.12, 120.88, 120.83, 120.78, 120.74, 120.47, 120.43, 120.29, 120.24, 120.19, 115.22, 115.14, 110.69, 110.48, 55.74, 55.41, 41.07 (d, *J* = 140 Hz), 37.16, 37.06, 34.34 (d, *J* = 137 Hz), 31.82, 31.71, 31.48, 29.19, 28.90, 28.03, 27.49, 22.73, 22.69. IR (neat) cm⁻¹: 2927, 2856, 1593, 1490, 1454, 1271, 1247, 1215, 1190, 1026, 929, 755, 689. ³¹P NMR (121 MHz, CDCl₃) δ : 20.53, 19.13.





The standard procedure was followed.

HRMS-EI (m/z): [M]⁺ calcd for C₂₉H₃₅O₅P, 494.22; found, 494.2209.

2h: ¹H NMR (400 MHz, CDCl₃) δ: 7.32-7.28 (m, 3H), 7.21-7.18 (m, 4H), 7.17-7.13 (m, 3H),
7.05 (d, J = 8.4 Hz, 1H), 6.53 (d, J = 6.4 Hz, 1H), 6.48-6.43 (m, 2H), 3.82 (s, 3H), 3.76 (s, 3H),
3.09 (d, J = 22.4 Hz, 2H), 2.44-2.35 (m, 2H), 1.51-1.43 (m, 2H), 1.29-1.17 (m, 6H), 0.87-0.80 (m, 3H).

IR (neat) cm⁻¹: 2955, 2922, 2852, 1732, 1608, 1591, 1491, 1462, 1378, 1262, 1210, 1190, 1160, 1117, 933, 763, 501.

³¹P NMR (121 MHz, CDCl₃) δ: 20.68.

2'h: ¹H NMR (300 MHz, CDCl₃) δ: 7.42 (dd, *J* = 8.4, 2.1 Hz, 1H), 7.32-7.26 (m, 3H), 7.16-7.14 (m, 4H), 7.05 (m, 1H), 6.80 (d, *J* = 7.8 Hz, 2H), 6.50 (d, *J* = 8.4 Hz, 1H), 6.40 (s, 1H), 5.64 (d, *J* = 2.1 Hz, 1H), 5.13 (s, 1H), 4.75 (d, *J* = 25.5 Hz, 1H), 3.80 (s, 3H), 3.74 (s, 3H), 2.11-2.02 (m, 2H), 1.42-1.35 (m, 2H), 1.26-1.21 (m, 6H), 0.90-0.82 (m, 3H).

¹³C NMR (75 MHz, CDCl₃) δ: 160.35, 158.15, 158.03, 151.03, 150.89, 150.82, 144.55, 144.50, 131.39, 131.32, 129.66, 129.42, 124.91, 124.66, 120.79, 120.73, 120.49, 120.43, 115.39, 115.33, 114.89, 114.79, 104.58, 98.61, 55.71, 55.51, 40.54 (d, *J* = 140 Hz), 37.05, 36.92, 31.83, 28.91, 27.49, 22.70, 14.23.

IR (neat) cm⁻¹: 2919, 2851, 1587, 1489, 1455, 1265, 1209, 1189, 1159, 1107, 1026, 926, 761, 688. ³¹P NMR (121 MHz, CDCl₃) δ: 19.39. Table 1, entry 12:



The standard procedure was followed.

HRMS-EI (m/z): [M]⁺ calcd for C₂₈H₃₃O₃P, 448.22; found, 448.2171.

2i: ¹H NMR (400 MHz, CDCl₃) δ: 7.34-7.27 (m, 3H), 7.21-7.12 (m, 5H), 7.12-7.09 (m, 4H),

7.07 (d, J = 8.4 Hz, 2H), 6.51 (d, J = 6.0 Hz, 1H), 3.06 (d, J = 22.0 Hz, 2H), 2.45 (t, J = 8.4, 2H),

2.35 (s, 3H), 2.14-1.98 (m, 2H), 1.52-1.47 (m, 2H), 1.30-1.19 (m, 4H), 0.89-0.82 (m, 3H).

¹³C NMR (100 MHz, CDCl₃) δ: 150.74, 150.64, 136.45, 132.06, 131.94, 131.25, 131.11, 130.00, 129.88, 129.74, 129.02, 128.94, 128.62, 125.18, 120.76, 120.70, 120.66, 120.60, 34.64 (d, *J* = 137 Hz), 31.73, 31.53, 29.31, 22.73, 22.58, 21.31, 14.19.

IR (neat) cm⁻¹: 2920, 2851, 1591, 1489, 1272, 1214, 1189, 1162, 1025, 928, 761, 688.

³¹P NMR (121 MHz, CDCl₃) δ: 20.28.

2'i: ¹H NMR (400 MHz, CDCl₃) δ: 7.38 (d, *J* = 8.8 Hz, 2H), 7.32-7.10 (m, 8H), 6.86 (d, *J* = 8.8 Hz, 2H), 6.79 (d, *J* = 8.4 Hz, 2H), 5.70 (d, *J* = 2.8 Hz, 1H), 5.18 (d, *J* = 2.8 Hz, 1H), 4.06 (d, *J* = 25.2 Hz, 1H), 2.35 (s, 3H), 2.06-1.97 (m, 2H), 1.41-1.37 (m, 2H), 1.36-1.20 (m, 4H), 0.89-0.83 (m, 5H).

³¹P NMR (121 MHz, CDCl₃) δ: 18.38.

Table 1, entry 13:



The standard procedure was followed.

HRMS-EI (m/z): $[M]^+$ calcd for C₂₇H₂₇O₃P, 406.1692; found, 406.1700.

2j: ¹H NMR (400 MHz, CDCl₃) δ: 7.34-7.29 (m, 6H), 7.22-7.11 (m, 9H), 6.56 (d, *J* = 6.4 Hz, 1H), 3.08 (d, *J* = 22.4 Hz, 2H), 2.50-2.42 (m, 2H), 2.14-1.98 (m, 2H), 1.54-1.45 (m, 2H), 1.44-1.24 (m, 4H), 0.87 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ: 150.73, 150.63, 132.05, 131.93, 131.25, 131.12, 130.01, 129.88, 129.82, 129.74, 129.02, 128.61, 125.19, 120.77, 120.66, 34.63 (d, *J* = 137 Hz), 31.73, 31.52, 29.31, 22.73, 22.58, 21.31, 14.19.

IR (neat) cm⁻¹: 2956, 2925, 2856, 1591, 1490, 1271, 1214, 1189, 1162, 1071, 1025, 1007, 928, 760, 689.

³¹P NMR (121 MHz, CDCl₃) δ: 20.16.

2'j: ¹H NMR (400 MHz, CDCl₃) δ: 7.52 (d, *J* = 8.4 Hz, 2H), 7.32-7.10 (m, 10H), 7.05 (t, *J* = 7.2 Hz, 1H), 6.76 (d, *J* = 8.4 Hz, 2H), 5.72 (d, *J* = 3.0 Hz, 1H), 5.21 (d, *J* = 3.0 Hz, 1H), 4.11 (d, *J* = 24.8 Hz, 1H), 2.06-1.97 (m, 2H), 1.44-1.35 (m, 2H), 1.36-1.20 (m, 4H), 0.90-0.83 (m, 5H). ³¹P NMR (121 MHz, CDCl₃) δ: 18.19. Table 1, entry 14:



The standard procedure was followed.

HRMS-ES (m/z): $[M]^+$ calcd for C₂₇H₃₀ClO₃P, 468.1616; found, 468.1639.

2k: ¹H NMR (400 MHz, CDCl₃) δ: 7.35-7.21 (m, 4H), 7.22-7.09 (m, 10H), 6.57 (d, *J* = 6.4 Hz, 1H), 3.09 (d, *J* = 22.4 Hz, 2H), 2.50-2.43 (m, 2H), 2.11-1.99 (m, 2H), 1.55-1.45 (m, 2H), 1.44-1.22 (m, 4H), 0.87 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ: 150.55, 150.46, 137.77, 131.28, 131.16, 130.86, 130.72, 130.08, 129.89, 124.98, 120.69, 120.60, 113.73, 34.42 (d, *J* = 137 Hz), 31.70, 31.28, 29.14, 27.87, 22.55, 14.01.

IR (neat) cm⁻¹: 2919, 2851, 1726, 1592, 1489, 1466, 1270, 1212, 1187, 1161, 1091, 1025, 929, 760, 688.

³¹P NMR (121 MHz, CDCl₃) δ: 19.86.

2'k: ¹H NMR (400 MHz, CDCl₃) δ: 7.45 (d, *J* = 8.8 Hz, 2H), 7.33-7.29 (m, 4H), 7.22-7.06 (m, 6H), 6.82 (d, *J* = 8.4 Hz, 2H), 5.69 (d, *J* = 3.2 Hz, 1H), 5.21 (d, *J* = 3.2 Hz, 1H), 4.07 (d, *J* = 25.2 Hz, 1H), 2.16-1.94 (m, 2H), 1.42-1.33 (m, 2H), 1.28-1.16 (m, 4H), 0.90-0.81 (m, 5H). ³¹P NMR (121 MHz, CDCl₃) δ: 17.92. Table 1, entry 15:



The standard procedure was followed, except that CH_3CN was distilled over calcium hydride before use, 100 mol% *p*-TsOH•H₂O was used, the reaction mixture was stirred at 35 °C for 18 hours and the product was purified with 5% diethyl ether in toluene.

HRMS-ESI (m/z): [M+Na]⁺ calcd for C₂₈H₃₃O₄PNa, 487.2009; found, 487.2017.

21: ¹H NMR (400 MHz, CDCl₃) δ: 7.33-7.29 (m, 5H), 7.22-7.14 (m, 6H), 6.80-6.77 (m, 2H), 6.71 (s, 1H), 6.53 (d, *J* = 6.1 Hz, 1H), 3.79 (s, 3H), 3.07 (d, *J* = 22.4 Hz, 2H), 2.48-2.41 (m, 2H), 1.48-1.52 (m, 2H), 1.35-1.20 (m, 6H), 0.88-0.82 (m, 3H).

¹³C NMR (100 MHz, CDCl₃) δ: 158.40, 150.74, 131.27, 131.15, 130.85, 130.72, 130.03, 129.88, 125.18, 124.59, 120.99, 120.70, 120.66, 113.73, 55.39, 34.66 (d, *J* = 137 Hz), 31.75, 31.49, 29.85, 29.34, 28.09, 22.74, 14.21.

IR (neat) cm⁻¹: 2920, 2851, 1725, 1591, 1509, 1489, 1249, 1214, 1189, 1162, 1026, 927, 759, 689. ³¹P NMR (121 MHz, CDCl₃) δ: 20.11.

2'I: ¹H NMR (400 MHz, CDCl₃) δ: 7.22-7.05 (m, 11H), 6.85-6.79 (m, 3H), 5.70 (d, *J* = 3.1 Hz, 1H), 5.20 (d, *J* = 3.1 Hz, 1H), 4.08 (d, *J* = 25.0 Hz, 1H), 3.77 (s, 3H), 2.18-2.00 (m, 2H), 1.42-1.38 (m, 2H), 1.31-1.20 (m, 6H), 0.88-0.81 (m, 3H).

¹³C NMR (100 MHz, CDCl₃) δ: 159.78, 150.76, 143.65, 143.60, 135.96, 135.90, 129.74, 129.64, 129.55, 125.13, 124.93, 122.58, 122.50, 120.80, 120.76, 120.62, 120.57, 115.57, 115.50, 115.36, 113.48, 55.38, 50.84 (d, *J* = 138 Hz), 37.04, 36.94, 31.81, 28.92, 27.57, 22.71, 14.20.

IR (neat) cm⁻¹: 2920, 2851, 1726, 1592, 1489, 1456, 1270, 1213, 1188, 1161, 1046, 928, 760, 688. ³¹P NMR (121 MHz, CDCl₃) δ: 18.00. Table 1, entry 16:



The standard procedure was followed.

HRMS-EI (m/z): [M]⁺ calcd for C₂₉H₃₆NO₃P, 477.24; found, 477.2438.

2m: ¹H NMR (400 MHz, CDCl₃) δ: 7.30-7.26 (m, 4H), 7.19-7.13 (m, 6H), 7.11 (d, *J* = 8.8 Hz, 2H), 6.70 (d, *J* = 8.8 Hz, 2H), 6.43 (d, *J* = 6.4 Hz, 1H), 3.05 (d, *J* = 22.4 Hz, 2H), 2.96 (s, 6H), 2.51-2.46 (m, 2H), 1.58-1.50 (m, 2H), 1.35-1.25 (m, 6H), 0.89-0.82 (m, 3H).

¹³C NMR (100 MHz, CDCl₃) δ : 150.76, 150.66, 131.14, 129.85, 129.71, 129.68, 129.49, 125.13, 120.76, 120.72, 112.26, 40.67, 34.90 (d, *J* = 137 Hz), 31.82, 31.68, 29.45, 28.18, 22.77, 14.23. IR (neat) cm⁻¹: 2925, 2854, 1732, 1610, 1593, 1520, 1490, 1353, 1272, 1215, 1190, 1163, 1071, 1025, 928, 762, 689.

³¹P NMR (121 MHz, CDCl₃) δ: 20.41.

2'm: ¹H NMR (400 MHz, CDCl₃) δ: 7.32 (d, *J* = 8.8 Hz, 2H), 7.30-7.28 (m, 1H), 7.19-7.11 (m, 6H), 7.05 (t, *J* = 7.4 Hz, 1H), 6.80 (d, *J* = 8.4 Hz, 2H), 6.69 (d, *J* = 8.8 Hz, 2H), 5.66 (d, *J* = 3.2 Hz, 1H), 5.15 (d, *J* = 3.2 Hz, 1H), 4.00 (d, *J* = 24.8 Hz, 1H), 2.94 (s, 6H), 2.16-1.99 (m, 2H), 1.40-1.36 (m, 2H), 1.25-1.20 (m, 6H), 0.89-0.83 (m, 3H).

¹³C NMR (100 MHz, CDCl₃) δ: 151.02, 150.92, 150.87, 150.77, 150.16, 144.28, 144.24, 130.73, 130.65, 129.84, 129.68, 129.49, 124.96, 124.78, 120.86, 120.82, 120.76, 120.72, 114.64, 114.56, 112.82, 49.84 (d, *J* = 139 Hz), 40.77, 36.93, 36.83, 31.84, 28.96, 27.58, 22.72, 14.21.

IR (neat) cm⁻¹: 2925, 2854, 1611, 1593, 1520, 1490, 1353, 1270, 1214, 1190, 1162, 1070, 1025, 928, 764, 689.

³¹P NMR (121 MHz, CDCl₃) δ: 19.00.



The standard procedure was followed, except that the product was purified with 20% ethyl acetate in hexane. Isolated as a mixture of 2n and 2'n.

HRMS-ESI (m/z): [M+Na]⁺ calcd for C₂₉H₂₇O₄PNa, 493.1539; found, 493.1523.

2n: ¹H NMR (400 MHz, CDCl₃) δ : 7.60 (d, J = 6.4 Hz, 2H), 7.33-7.07 (m, 12H), 6.90-6.74 (m,

6H), 4.29 (d, *J* = 25.4 Hz, 1H), 3.80 (s, 3H), 1.94 (s, 3H).

2'n: ¹H NMR (400 MHz, CDCl₃) δ: 7.52 (d, *J* = 6.9 Hz, 2H), 7.33-7.07 (m, 11H), 6.97 (d, *J* = 3.1 Hz, 1H), 6.90-6.74 (m, 6H), 4.25 (d, *J* = 25.5 Hz, 1H), 3.80 (s, 3H), 1.94 (s, 3H).

2n+2'n: ¹³C NMR (75 MHz, CDCl₃) δ: 159.29, 156.50, 150.87, 150.74, 150.57, 137.54, 133.24, 131.58, 131.04, 130.80, 130.66, 130.40, 129.83, 129.68, 129.51, 129.16, 128.84, 128.21, 126.80, 125.24, 125.17, 120.80, 120.75, 119.97, 115.23, 114.26, 55.43, 53.51 (d, *J* = 138 Hz), 17.99, 17.90.

IR (neat) cm⁻¹: 2918, 2851, 1726, 1591, 1510, 1489, 1252, 1213, 1185, 1158, 1025, 929, 757, 689. ³¹P NMR (121 MHz, CDCl₃) δ: 18.00, 18.00.

Comparison with correlated compounds (silyl ethers) Mahandru, G. M.; Liu, G.; Montgomery, J. J. Am. Chem. Soc. **2004**, *126*, 3698-3699.

Silyl ether with a structure similar to **2n** instead of **2'n** gives a slightly more deshielded carbinol proton.



Table 1, entry 18:



(major)

(minor)

The standard procedure was followed, except that the tris(4-methylphenyl) phosphite was used instead of triphenyl phosphite and the product was purified with 20% ethyl acetate in hexane. Isolated as a mixture of two isomers.

HRMS-EI (m/z): [M]⁺ calcd for C₂₈H₃₃O₄P, 464.2111; found, 464.2117.

major: ¹H NMR (400 MHz, CDCl₃) δ: 7.13-6.95 (m, 10H), 6.86 (d, *J* = 8.7 Hz, 2H), 6.45 (d, *J* = 6.1 Hz, 1H), 3.81 (s, 3H), 3.02 (d, *J* = 22.2 Hz, 2H), 2.48-2.42 (m, 2H), 2.29 (s, 6H), 1.52-1.44 (m, 2H), 1.44-1.21 (m, 2H), 0.89-0.82 (m, 3H).

minor: ¹H NMR (400 MHz, CDCl₃) δ : 7.41 (d, J = 8.5 Hz, 2H), 7.13-6.95 (m, 6H), 6.86 (d, J = 8.7 Hz, 2H), 6.67 (d, J = 8.5 Hz, 2H), 5.67 (d, J = 2.5 Hz, 1H), 5.17 (d, J = 2.5 Hz, 1H), 4.01 (d, J = 24.9 Hz, 1H), 3.80 (s, 3H), 2.30 (s, 6H), 2.14-1.97 (m, 2H), 1.44-1.21 (m, 4H), 0.89-0.82 (m, 3H).

major+minor:¹³C NMR (100 MHz, CDCl₃) δ : 158.34, 148.50, 148.41, 134.68, 131.37, 131.25, 131.13, 131.06, 130.68, 130.55, 130.35, 130.27, 130.14, 130.10, 129.97, 129.91, 129.88, 120.47, 120.43, 120.39, 120.35, 120.31, 120.27, 114.07, 113.68, 55.39, 55.35, 49.75 (d, *J* = 138 Hz), 36.62, 34.49 (d, *J* = 137 Hz), 31.22, 31.18, 30.29, 30.27, 29.74, 22.77, 22.33, 20.84, 20.79, 14.05. IR (neat) cm⁻¹: 2924, 2855, 1608, 1506, 1465, 1272, 1250, 1221, 1192, 1164, 1105, 1035, 940, 925, 821.

³¹P NMR (121 MHz, CDCl₃) δ: 20.50, 18.59.

Table 1, entry 19:



(major)

(minor)

The standard procedure was followed, except that the tris(4-chlorophenyl) phosphite was used instead of triphenyl phosphite and the product was purified with 20% ethyl acetate in hexane. Isolated as a mixture of two isomers.

HRMS-EI (m/z): $[M]^+$ calcd for C₂₆H₂₇Cl₂O₄P, 504.1019; found, 504.1000.

major: ¹H NMR (400 MHz, CDCl₃) δ: 7.28-7.08 (m, 8H), 6.86 (d, *J* = 8.8 Hz, 2H), 6.68-6.65 (m, 2H), 6.48 (d, *J* = 6.3 Hz, 1H), 3.82 (s, 3H), 3.06 (d, *J* = 22.3 Hz, 2H), 2.45-2.41 (m, 2H), 1.53-1.45 (m, 2H), 1.45-1.22 (m, 2H), 0.89-0.82 (m, 3H).

minor: ¹H NMR (400 MHz, CDCl₃) δ: 7.38 (d, J = 8.8 Hz, 2H), 7.28-7.08 (m, 8H), 6.86 (d, J = 8.8 Hz, 2H), 5.65 (d, J = 3.0 Hz, 1H), 5.19 (d, J = 2.9 Hz, 1H), 4.04 (d, J = 25.1 Hz, 1H), 3.80 (s, 3H), 2.15-1.97 (m, 2H), 1.45-1.22 (m, 4H), 0.89-0.83 (m, 3H).

major+**minor:** ¹³C NMR (100 MHz, CDCl₃) δ : 158.52, 148.97, 148.87, 131.35, 131.22, 131.10, 131.02, 130.81, 130.37, 130.25, 129.95, 129.86, 129.83, 129.67, 129.60, 129.28, 122.07, 122.03, 121.97, 121.93, 121.84, 121.80, 116.77, 114.26, 113.80, 55.42, 55.37, 49.81 (d, *J* = 139 Hz), 36.67, 34.57 (d, *J* = 136 Hz), 31.24, 31.21, 30.24, 31.21, 29.72, 22.73, 22.28, 14.02. IR (neat) cm⁻¹: 2924, 2850, 1725, 1485, 1460, 1270, 1250, 1218, 1193, 1164, 1090, 924, 830. ³¹P NMR (121 MHz, CDCl₃) δ : 21.10, 19.19. Scheme 5, Allyl diarylphosphonylation of TES protected Baylis-Hillman adduct of methylacrylate and anisaldehyde:

General procedure for the allyl diarylphosphonylation of allyl silyl ethers was followed, except the reaction was conducted at 35 °C and was purified with 25% ethyl acetate in hexane.

81% yield (2+2'); 2:2' = 3.2:1; E:Z = 18:82. Olefin stereochemistry were determined by NOESY.



MeO (Minor, **2** *E*-isomer): ¹H NMR (400 MHz, CDCl₃) δ : 7.62-7.54 (m, 2H), 7.42-7.34 (m, 2H), 7.31-7.11 (m, 6H), 7.06 (d, *J* = 7.6 Hz, 1H), 7.01 (d, *J* = 8.0 Hz, 1H), 6.95 (d, *J* = 5.6 Hz, 1H), 6.86 (d, *J* = 8.4 Hz, 2H), 3.82 (s, 3H), 3.65 (s, 3H), 3.37 (d, *J* = 21.6 Hz, 2H). ³¹P NMR (121 MHz, CDCl₃) δ : 18.53.

³¹P NMR (121 MHz, CDCl₃) δ: 17.78.

11/ Arylation products (Table 2).

The products relative substitution patterns were determined by ¹H NMR coupling patterns and by ¹³C NMR chemical shift comparison of related compounds if the aromatic region of ¹H NMR was not clear. The olefin stereochemistry was determined by NOESY.

For ¹³C NMR of 2-methylphenol, 3-methylphenol and 4-methylphenol, see Ilczszyn, M.; Latałka, Z.; Ratajczak, H. *Org. Magn. Reson.* **1980**, *13*, 132-136.

¹³C NMR (25.2 MHz, CDCl₃) δ:

Ortho-	Meta-	Para-
¹³ C δ: 124.0 OH Me	¹³ C δ: 139.3 Me	¹³ C δ: 130.5 Me
153.5, 131.1, 127.7, 124.0,	154.9, 139.3, 130.3, 122.2,	152.6, 130.5, 130.2, 115.3.
121.4, 115.9.	116.1, 112.7.	

Table 2, entry 1:



The standard procedure was followed.

HRMS-EI (m/z): $[M]^+$ calcd for C₂₂H₂₈O₂, 324.2084; found, 324.2077.

¹H NMR (400 MHz, CDCl₃) δ: 7.15 (d, *J* = 8.6 Hz , 2H), 7.10 (d, *J* = 8.4 Hz, 2H), 6.85 (d, *J* = 8.6 Hz, 2H), 6.76 (d, *J* = 8.4 Hz, 2H), 6.21 (s, 1H), 5.05 (brs, 1H), 3.80 (s, 3H), 3.39 (s, 2H), 2.13 (t, *J* = 8.2 Hz, 2H), 1.47-1.41 (m, 2H), 1.28-1.20 (m, 6H), 0.86 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ: 157.97, 154.06, 142.09, 132.48, 131.25, 130.39, 129.97, 126.36, 115.36, 113.75, 55.49, 43.30, 31.85, 30.45, 29.62, 28.35, 22.83, 14.31.

IR (neat) cm⁻¹: 3396, 2925, 1607, 1509, 1464, 1247, 1174, 1035, 822.

Table 2, entry 2:

The standard procedure was followed.

HRMS-EI (m/z): $[M]^+$ calcd for C₂₀H₂₄O₂, 296.1771; found, 296.1774.

¹H NMR (400 MHz, CDCl₃) δ: 7.15 (d, *J* = 8.6 Hz, 2H), 7.11 (d, *J* = 8.4 Hz, 2H), 6.84 (d, *J* = 8.6 Hz, 2H), 6.77 (d, *J* = 8.4 Hz, 2H), 6.29 (s, 1H), 4.92 (brs, 1H), 3.80 (s, 3H), 3.38 (s, 2H), 2.05 (d, *J* = 7.4 Hz, 2H), 1.88-1.98 (m, 1H), 0.82 (d, *J* = 6.6 Hz, 6H).

¹³C NMR (100 MHz, CDCl₃) δ: 157.86, 153.98, 140.98, 132.45, 131.24, 130.29, 130.13, 127.54, 115.27, 113.56, 55.36, 43.23, 38.76, 26.53, 22.65.

IR (neat) cm⁻¹: 3400, 2954, 1608, 1509, 1463, 1246, 1173, 1035, 822.

Table 2, entry 3:



3c ortho-product

The standard procedure was followed, except that the product was purified with chloroform.

HRMS-EI (m/z): $[M]^+$ calcd for C₂₂H₂₆O₂, 322.1927; found, 322.1905.

¹H NMR (400 MHz, CDCl₃) δ : 7.18-7.15 (m, 2H), 7.08 (d, J = 8.5 Hz, 2H), 6.91 (t, J = 7.4 Hz,

1H), 6.86-6.84 (m, 3H), 6.05 (s, 1H), 5.10 (brs, 1H), 3.80 (s, 3H), 3.49 (s, 2H), 2.86-2.79 (m, 1H),

1.80-1.48 (m, 5H), 1.45-1.10 (m, 5H).

¹³C NMR (100 MHz, CDCl₃) δ: 158.17, 154.64, 144.64, 131.55, 130.33, 129.94, 128.10, 125.85,

125.72, 120.98, 116.23, 113.69, 55.38, 40.60, 34.79, 31.64, 26.51, 26.21.

IR (neat) cm⁻¹: 3436, 2926, 1602, 1512, 1458, 1384, 1256, 1176, 1032, 836, 754.

Table 2, entry 4:



The standard procedure was followed, except that the product was purified with chloroform.

HRMS-EI (m/z): $[M]^+$ calcd for C₂₂H₂₀O₂, 316.1458; found, 316.1459.

¹H NMR (400 MHz, CDCl₃) δ: 7.24-7.20 (m, 3H), 7.06-7.04 (m, 4H), 6.84 (d, *J* = 8.6 Hz, 2H), 6.72 (d, *J* = 8.6 Hz, 2H), 6.62 (d, *J* = 8.8 Hz, 2H), 6.35 (s, 1H), 4.70 (brs, 1H), 3.72 (s, 3H), 3.68 (s, 2H).

¹³C NMR (100 MHz, CDCl₃) δ: 158.64, 154.36, 140.07, 139.62, 132.33, 130.89, 130.76, 130.05, 129.14, 128.94, 128.62, 126.49, 115.59, 115.49, 114.13, 55.71, 42.96.

IR (neat) cm⁻¹: 3397, 2918, 1724, 1606, 1510, 1465, 1368, 1286, 1251, 1216, 1176, 1124, 1074, 1040, 825, 759, 701.

Table 2, entry 5:

¹³C δ: 132.01 3e MeO ОН

The standard procedure was followed.

HRMS-EI (m/z): $[M]^+$ calcd for C₂₃H₂₂O₂, 330.1614; found, 330.1617.

¹H NMR (400 MHz, CDCl₃) δ: 7.32-7.29 (m, 2H), 7.26-7.16 (m, 5H), 7.03 (d, *J* = 8.5 Hz, 2H), 6.83 (d, *J* = 8.8 Hz, 2H), 6.76 (d, *J* = 8.5 Hz, 2H), 6.49 (s, 1H), 4.81 (brs, 1H), 3.79 (s, 3H), 3.53 (s, 2H), 3.29 (s, 2H).

¹³C NMR (100 MHz, CDCl₃) δ: 158.32, 154.05, 139.76, 139.30, 132.01, 130.57, 130.45, 129.74, 128.83, 128.63, 138.30, 126.17, 115.28, 113.81, 55.39, 42.65, 36.00.

IR (neat) cm⁻¹: 3401, 3024, 2922, 1607, 1509, 1451, 1248, 1177, 1031, 822, 733.

Table 2, entry 6:



The standard procedure was followed.

HRMS-ESI (m/z): $[M+Na]^+$ calcd for $C_{24}H_{24}O_2Na$, 367.1669; found, 367.1658.

¹H NMR (400 MHz, CDCl₃) δ: 7.25-7.23 (m, 2H), 7.18-7.09 (m, 7H), 6.84 (d, *J* = 8.4 Hz, 2H), 6.77 (d, *J* = 8.2 Hz, 2H), 6.28 (s, 1H), 4.88 (brs, 1H), 3.80 (s, 3H), 3.43 (s, 2H), 2.75-2.71 (m, 2H), 2.50-2.43 (m, 2H).

¹³C NMR (75 MHz, CDCl₃) δ: 158.09, 154.11, 142.07, 140.70, 132.12, 130.78, 130.33, 129.81, 128.45, 128.42, 127.22, 126.00, 115.34, 113.70, 55.40, 43.52, 34.48, 32.33.

IR (neat) cm⁻¹: 3401, 3025, 2925, 2852, 1606, 1509, 1453, 1247, 1177, 1033, 939, 826, 699.

Table 2, entry 7:





(major, more polar)

(minor, less polar)

The standard procedure was followed, except that 40 mol% of *p*-TsOH•H₂O was used.

HRMS-EI (m/z): $[M]^+$ calcd for C₂₂H₂₈O₂, 324.2084; found, 324.2059.

3g: ¹H NMR (400 MHz, CDCl₃) δ: 7.23-7.17 (m, 2H), 7.15 (d, *J* = 8.4 Hz, 2H), 6.90 (dd, *J* = 8.2, 7.4 Hz, 1H), 6.86 (d, *J* = 8.2 Hz, 1H), 6.77 (d, *J* = 8.4 Hz, 2H), 6.35 (s, 1H), 4.73 (brs, 1H), 3.82 (s, 3H), 3.44 (s, 2H), 2.06 (t, *J* = 8.1 Hz, 2H), 1.49-1.39 (m, 2H), 1.30-1.17 (m, 6H), 0.85 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ: 157.30, 153.92, 142.75, 132.61, 130.25, 130.19, 127.73, 127.50, 122.72, 120.14, 115.21, 110.53, 55.59, 42.86, 31.74, 30.30, 29.42, 28.20, 22.73, 14.20.
IR (neat) cm⁻¹: 3398, 2924, 1706, 1612, 1597, 1512, 1487, 1463, 1376, 1288, 1243, 1171, 1109, 1029, 823, 752.

Purified with 10% ethyl acetate in hexane.

3*g ortho*-**product:** ¹H NMR (400 MHz, CDCl₃) δ: 7.23-7.21 (m, 1H), 7.18-7.12 (m, 3H), 6.94-6.85 (m, 4H), 6.54 (s, 1H), 5.86 (brs, 1H), 3.83 (s, 3H), 3.60 (s, 2H), 2.02 (t, *J* = 8.2 Hz, 2H), 1.50-1.39 (m, 2H), 1.34-1.15 (m, 6H), 0.83 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ: 156.99, 142.41, 131.12, 129.65, 128.13, 128.07, 126.46, 123.53, 120.72, 120.11, 116.85, 110.16, 55.37, 40.16, 31.53, 29.93, 29.12, 27.86, 22.54, 14.05. IR (neat) cm⁻¹: 3385, 2953, 2850, 1720, 1594, 1487, 1454, 1369, 1286, 1243, 750.





The standard procedure was followed, except that 40 mol% of *p*-TsOH•H₂O was used.

HRMS-EI (m/z): [M]⁺ calcd for C₂₂H₂₈O, 308.2135; found, 308.2112.

3i: ¹H NMR (400 MHz, CDCl₃) δ: 7.12-7.09 (m, 6H), 6.77 (d, *J* = 8.4 Hz, 2H), 6.24 (s, 1H), 4.74 (brs, 1H), 3.40 (s, 2H), 2.33 (s, 3H), 2.13 (t, *J* = 8.0 Hz, 2H), 1.50-1.40 (m, 2H), 1.31-1.20 (m, 6H), 0.86 (t, *J* = 6.8 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ: 153.96, 142.64, 135.76, 135.56, 132.43, 130.31, 128.91, 128.65, 126.76, 115.25, 43.21, 31.76, 30.41, 29.52, 28.28, 22.75, 21.27, 14.22.

IR (neat) cm⁻¹: 3374, 3020, 2854, 1703, 1612, 1510, 1444, 1376, 1222, 1170, 1099, 824.

Purified with 10% ethyl acetate in hexane.

3i *ortho*-product: ¹H NMR (400 MHz, CDCl₃) δ : 7.15 (d, J = 7.6 Hz, 2H),7.13-7.09 (m, 4H), 6.93-6.87 (m, 1H), 6.84 (d, J = 7.6 Hz, 1H), 6.39 (s, 1H), 5.27 (brs, 1H), 3.54 (s, 2H), 2.33 (s, 3H), 2.20 (t, J = 8.0 Hz, 2H), 1.53-1.44 (m, 2H), 1.30-1.19 (m, 6H), 0.85 (t, J = 7.0 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ : 155.09, 141.60, 136.26, 131.23, 129.01, 128.64, 128.18, 127.19, 125.05, 120.92, 116.17, 39.62, 31.68, 30.62, 29.44, 28.22, 22.70, 21.28, 14.20. IR (neat) cm⁻¹: 3398, 3021, 2926, 1706, 1604, 1488, 1456, 1377, 1243, 1182, 1096, 1020, 807, 752. Table 2, entry 9:





31 ortho-product

(major, more polar)

(minor, less polar)

The standard procedure was followed, except that 40 mol% of *p*-TsOH•H₂O was used.

HRMS-EI (m/z): $[M]^+$ calcd for C₂₂H₂₈O₂, 324.2084; found, 324.2080.

31: ¹H NMR (400 MHz, CDCl₃) δ: 7.24-7.20 (m, 1H), 7.15 (d, *J* = 8.5 Hz, 2H), 6.82-6.74 (m, 5H), 6.25 (s, 1H), 4.78 (brs, 1H), 3.80 (s, 3H), 3.40 (s, 2H), 2.17 (t, *J* = 8.2 Hz, 2H), 1.55-1.40 (m, 2H), 1.30-1.23 (m, 6H), 0.86 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ: 159.49, 154.01, 143.61, 139.92, 132.25, 130.34, 129.15, 126.79, 121.35, 115.37, 115.29, 114.18, 114.03, 112.03, 111.89, 55.30, 43.14, 31.79, 30.52, 29.54, 28.34, 22.74, 14.21.

IR (neat) cm⁻¹: 3357, 2916, 2850, 1703, 1575, 1510, 1453, 1367, 1257, 1152, 1072, 1040, 820, 781, 693.

Purified with 10% ethyl acetate in hexane.

3*I ortho*-product: ¹H NMR (400 MHz, CDCl₃) δ: 7.21-7.15 (m, 2H), 6.92-6.75 (m, 6H), 6.37 (s, 1H), 5.22 (brs, 1H), 3.80 (s, 3H), 3.54 (s, 2H), 2.22 (t, *J* = 8.1 Hz, 2H), 1.54-1.45 (m, 2H), 1.38-1.15 (m, 6H), 0.89-0.79 (m, 3H).

¹³C NMR (100 MHz, CDCl₃) δ: 159.54, 154.94, 142.54, 139.18, 131.25, 129.26, 128.20, 127.09, 125.03, 121.29, 120.96, 116.13, 114.14, 112.28, 55.32, 39.31, 31.72, 30.79, 29.48, 28.31, 22.71, 14.24, 14.20.

IR (neat) cm⁻¹: 3422, 2926, 2854, 1706, 1597, 1487, 1455, 1261, 1155, 1092, 1042, 873, 753, 696.



(major, more polar)

(minor, less polar)

The standard procedure was followed.

HRMS-EI (m/z): $[M]^+$ calcd for $C_{21}H_{26}O_3$, 326.1876; found, 326.1864.

major: ¹H NMR (400 MHz, CDCl₃) δ : 7.15 (d, J = 8.4 Hz, 2H), 6.86 (d, J = 8.4 Hz, 2H), 6.78 (d, J = 8.8 Hz, 1H), 6.72-6.69 (m, 2H), 6.34 (s, 1H), 4.92 (brs, 1H), 3.81 (s, 3H), 3.77 (s, 3H), 3.49 (s, 2H), 2.22 (t, J = 8.0 Hz, 2H), 1.53-1.46 (m, 2H), 1.33-1.25 (m, 2H), 0.87 (t, J = 7.2 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ : 158.28, 153.78, 148.94, 140.59, 130.22, 129.91, 126.78, 126.30, 116.80, 116.76, 114.01, 113.72, 112.81, 55.86, 55.39, 39.80, 30.52, 30.39, 22.94, 14.07. IR (neat) cm⁻¹: 3401, 2956, 2926, 2871, 1606, 1509, 1464, 1383, 1248, 1203, 1176, 1105, 1039, 1464, 1383, 1248, 1203, 1176, 1105, 1039, 1464, 1383, 1248, 1203, 1176, 1105, 1039, 1464, 1383, 1248, 1203, 1176, 1105, 1039, 1464, 1383, 1248, 1203, 1176, 1105, 1039, 1464, 1405, 140

808.

Purified with 10% ethyl acetate in hexane.

minor: ¹H NMR (400 MHz, CDCl₃) δ: 7.20 (d, *J* = 8.4 Hz, 2H), 6.86 (d, *J* = 8.4 Hz, 2H), 6.74-6.62 (m, 3H), 6.53 (s, 1H), 4.65 (brs, 1H), 3.80 (s, 3H), 3.74 (s, 3H), 3.58 (s, 2H), 2.09 (t, *J* = 8.0 Hz, 2H), 1.53-1.46 (m, 2H), 1.30-1.23 (m, 2H), 0.90 (t, *J* = 7.2 Hz, 3H).

Comparison with correlated compounds (¹³C NMR, CDCl₃)

¹³ C δ: 126.3 OH Me Me OMe ¹³ C δ: 149.2 OH ¹³ C δ: 149.2	¹³ C δ: 147.89 Me C I ¹³ C δ: 125.00 OH I ³ C δ: 147.89 OH I ³ C δ: 147.89	¹³ C δ: 128.0 ¹³ C δ: 128.0
(75 MHz) δ: 153.7, 149.2, 134.9, 126.3, 121.6, 116.7, 112.7, 55.9, 42.2, 15.8, 13.7.	(50 MHz) δ: 153.8, 147. 9, 125.0, 116.7, 115.7, 112.0, 55.9, 16.2.	(75 MHz) δ: 151.8, 148.9, 128.0, 118.0, 112.6, 111.6, 56.1, 16.1.
Trend, R. M.; Ramtohul, Y. K.; Stoltz, B. M. J. Am. Chem. Soc. 2005, 127, 17778.	Mori T.; Grimme S.; Inoue Y. J. Org. Chem. 2007, 72, 6998.	Vyvyan, J. R.; Loitz, C.; Looper, R. E.; Mattingly, C. S.; Peterson, E. A.; Staben, S. T. J. Org. Chem. 2004 , <i>69</i> , 2461.

Table 2, entry 11:



The standard procedure was followed

HRMS-EI (m/z): $[M]^+$ calcd for C₂₁H₂₆O₂, 310.1927; found, 310.1934.

¹H NMR (400 MHz, CDCl₃) δ : 7.15 (d, J = 8.4 Hz, 2H), 6.95-6.93 (m, 2H), 6.86 (d, J = 8.4 Hz,

2H), 6.74 (d, J = 8.4 Hz, 1H), 6.34 (s, 1H), 5.11 (brs, 1H), 3.81 (s, 3H), 3.49 (s, 2H), 2.28 (s, 3H),

2.22 (t, J = 8.0 Hz, 2H), 1.54-1.47 (m, 2H), 1.36-1.26 (m, 2H), 0.87 (t, J = 7.3 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ: 158.25, 152.79, 140.92, 131.76, 130.30, 130.03, 129.91, 128.56, 126.57, 124.82, 115.97, 113.71, 55.38, 39.62, 30.49, 30.37, 22.93, 20.67, 14.06.

IR (neat) cm⁻¹: 3418, 2956, 2930, 1673, 1606, 1464, 1300, 1250, 1177, 1107, 1035, 937, 814.

¹³ C δ: 123.54 Me 1 ³ C δ: 129.91	¹³ C δ: 137.95 Me ^{-C} C ^{-C} Me ⁻¹³ C δ: 128.64
δ: 151.36, 131.61, 129.91, 127.34, 123.54,	δ: 153.42, 137.95, 130.47, 128.64, 116.58,
114.79, 20.36, 15.64.	112.34, 19.82, 18.72.
Wigal, C. T.; McKinley, J. D.; Coyle, J.;	Livant, P.; Xu, W. J. Org. Chem. 1998, 63,
Porter, D. J.; Lehman, D. E. J. Org.	636-641.
<i>Chem.</i> 1995 , <i>60</i> , 8421-8423.	

Comparison with correlated compounds (¹³C NMR, 63 MHz, CDCl₃)

Table 2, entry 12:



The standard procedure was followed.

HRMS-EI (m/z): $[M]^+$ calcd for C₂₀H₂₃ClO₂, 330.1381; found, 330.1389.

¹H NMR (400 MHz, CDCl₃) δ: 7.15 (d, *J* = 8.4 Hz, 2H), 7.13-7.10 (m, 2H), 6.87 (d, *J* = 8.4 Hz, 2H), 6.77 (d, *J* = 8.0 Hz, 1H), 6.35 (s, 1H), 5.31 (brs, 1H), 3.81 (s, 3H), 3.48 (s, 2H), 2.21 (t, *J* = 8.0 Hz, 2H), 1.52-1.44 (m, 2H), 1.35-1.25 (m, 2H), 0.87 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ: 153.59, 148.84, 141.50, 130.76, 130.18, 128.73, 128.36, 127.94,

127.75, 126.72, 121.57, 121.52, 117.40, 55.38, 39.11, 29.38, 28.22, 22.69, 14.19.

IR (neat) cm⁻¹: 3345, 2919, 2850, 1484, 1267, 1189, 1092, 971, 832, 698.

¹³ C δ: 125.22 CI ¹³ C δ: 125.73	¹³ C δ: 137.48 Me ^{-C} Cl ¹³ C δ: 126.00
125 MHz δ: 152.43, 130.65, 126.76, 125.73,	100 MHz δ: 153.91, 137.48, 129.94, 126.00,
125.22, 116.04, 15.69.	118.05, 114.35, 20.17.
Wang, JR.; Manabe, K. Org. Lett. 2009, 11,	Menini, L.; Gusevskaya, E. V. Appl. Catal., A
741-744.	2006 , <i>309</i> , 122-128.

Comparison with correlated compounds ¹³C NMR (CDCl₃):



The standard procedure was followed. HRMS-EI (m/z): [M]⁺ calcd for C₂₀H₂₃ClO₂, 330.1381; found, 330.1367. **major:** ¹H NMR (400 MHz, CDCl₃) δ: 7.14-7.12 (m, 3H), 6.89 (d, *J* = 2.8 Hz, 1H), 6.85 (d, *J* = 8.4 Hz, 2H), 6.70 (dd, *J* = 8.4, 2.8 Hz, 1H), 6.06 (s, 1H), 4.99 (brs, 1H), 3.80 (s, 3H), 3.49 (s, 2H), 2.18 (t, *J* = 8.0 Hz, 2H), 1.54-1.46 (m, 2H), 1.37-1.25 (m, 2H), 0.88 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ: 157.96, 154.62, 140.40, 135.06, 131.88, 131.04, 129.96, 129.82, 126.40, 116.51, 114.18, 113.62, 55.39, 40.03, 30.79, 30.70, 23.02, 14.11.

IR (neat) cm⁻¹: 3362, 2957, 2926, 2855, 1703, 1601, 1511, 1466, 1384, 1252, 1175, 1033, 836.

Purified with 10% ethyl acetate in hexane.

minor: ¹H NMR (400 MHz, CDCl₃) δ: 7.15 (d, *J* = 8.4 Hz, 2H), 7.06 (d, *J* = 8.0 Hz, 1H), 6.91-6.77 (m, 4H), 6.36 (s, 1H), 5.40 (brs, 1H), 3.81 (s, 3H), 3.49 (s, 2H), 2.17 (t, *J* = 8.0 Hz, 2H), 1.53-1.44 (m, 2H), 1.37-1.24 (m, 2H), 0.88 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ: 158.39, 155.75, 140.27, 133.15, 131.96, 130.58, 129.98, 129.90, 127.12, 123.73, 121.12, 121.02, 116.55, 116.04, 113.89. 113.78, 55.40, 39.19, 33.35, 31.74, 30.42, 30.28, 22.91, 22.81, 14.28, 14.04. IR (neat) cm⁻¹: 3400, 2956, 2928, 2858, 1702, 1607, 1577, 1509, 1497, 1465, 1441, 1294, 1249, 1177, 1039, 906, 856, 818.

¹³ C δ: 124.0 CI 13C δ: 132.8	¹³ C(1) δ: 121.3 HC HC HC HC HC HC HC HC HC HC HC HC HC
(100 MHz) δ: 154.6, 135.8, 132.8, 131.2, 124.0,	(25.2 MHz) δ: 156.3, 135.1, 130.6, 121.3,
121.0, 116.8, 116.1, 34.5.	116.1, 113.9.
Lin, YL.; Cheng, JY.; Chu, YH.	Ilczszyn, M.; Latałka, Z.; Ratajczak, H.
<i>Tetrahedron</i> 2007 , <i>63</i> , 10949-10957.	Org. Magn. Reson. 1980, 13, 132-136.

Comparison with correlated compounds ¹³C NMR (CDCl₃)

The carbon at *ortho*-position to OH, C(2), was more shielded than that to Cl, C(1). Other examples were shown below:

For the regioselective electrophilic aromatic substitution of 3-chlorophenol, see

 para-chlorination with Freidel-Crafts catalyst: Watson, W. D. J. Org. Chem. 1985, 50, 2145-2148.



2) Bromination: Suresh, P.; Annalakshmi, S.; Pitchumani, K. Tetrahedron 2007, 63, 4959-4967.



Table 2, entry 14:



The standard procedure was followed, except that anisole was used instead of triphenylphosphite.

HRMS-EI (m/z): $[M]^+$ calcd for C₂₃H₃₀O₂, 338.2240; found, 338.2224.

¹H NMR (400 MHz, CDCl₃) δ: 7.16 (d, *J* = 8.4 Hz, 2H), 7.15 (d, *J* = 8.4 Hz, 2H), 6.85 (d, *J* = 8.4 Hz, 2H), 6.84 (d, *J* = 8.4 Hz, 2H), 6.22 (s, 1H), 3.81 (s, 3H), 3.80 (s, 3H), 3.40 (s, 2H), 2.15 (t, *J* = 8.3 Hz, 2H), 1.50-1.42 (m, 2H), 1.29-1.19 (m, 6H), 0.86 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (75 MHz, CDCl₃) δ: 157.88, 141.80, 132.11, 130.90, 129.90, 129.66, 126.06, 113.61, 113.41, 55.18, 43.00, 31.57, 30.17, 29.34, 28.07, 22.55, 14.03.

IR (neat) cm⁻¹: 2953, 2926, 2855, 1712, 1608, 1509, 1464, 1441, 1300, 1247, 1175, 1105, 1037, 824.
Allyl Phenolic Targets	Ar ArOR	Ar ArOR R ¹	
	R ¹ = Ester	R ¹ = Alkyl, Aryl	
The Selected Phenolic Substrates Acceptors	Acetylated Baylis-Hillman Adducts	e.g. Silyl Allyl Ethers	
Phenolic Substrates	ArylKBF₃ Arylboronic Acids	Both protected & <u>unprotected</u> phenols	

Other related informations in the preparation of allyl phenolic compounds, with the concomitant formation of a new C-aryl bond and a new trisubstituted olefin:

It should be noted that treating unprotected phenols with Baylis-Hillman adducts may result mainly O-allylations, for examples, please see: (a) D. Y. Park, S. Gowrisankar, J. N. Kim, *Bull. Korean Chem. Soc.* 2005, *26*, 1440; (b) E. Ramesh, R. Raghunathan, *Tetrahedron Lett.* 2008, 49, 1125.

For detailed potassium organotrifluoroborates or aryl boronic acid coupling with Baylis-Hillman adducts procedures, please see: (a) Kabalka, G. W.; Venkataiah, B.; Dong, G. *Org. Lett.* 2003, *5*, 3803; (b) Kantam, M. L.; Kumar, K. B. S.; Sreedhar, B. *J. Org. Chem.* 2008, *73*, 320; (c) Gendrineau, T.; Demoulin, N.; Navarre, L.; Genet, J.-P.; Darses, S. *Chem. Eur. J.* 2009, *15*, 4710.

12/ Summary on the inhibitory effect of the newly synthesized allyl diarylphosphonates on

the metabolism of oseltamivir (O) to oseltamivir carboxylate (OC) in rat plasma

Experimental procedure

The inhibition experiment was modified from our previous study [Chang et al., Biomedical Chromatography, 2009, 23, 852.] and conducted as follow. To 200 μ L rat plasma 2 μ L of inhibitor working solution in DMSO was added (final 0.18, 0.4, 1, 4 or 10 μ M in plasma) and well mixed. 10 μ L of 100 μ g/mL oseltamivir in H₂O was added, and the mixture was incubated at room temperature for 1 hour. After incubation, 50 μ L of the sample mixture was acidified with 1 mL of 0.1% hydrochloric acid prior to solid phase extraction (Waters Oasis MCX cartridge, 30 mg). The detailed solid phase extraction procedure was reference to the previous study and was not produced here. Analytes were eluted with 1 mL of 1% ammonia in methanol and the solvent was evaporated to dryness by a vacuum concentrator. The residue was reconstituted in 150 μ L mobile phase (0.1% formic acid : methanol, 1:1 v/v) and 20 μ L supernatant were injected for LC-MS/MS analysis (data was acquired by multiple reaction monitoring of O and OC [Chang et al., Biomedical Chromatography, 2009]). Control samples were prepared by adding 2 μ L DMSO (instead of inhibitor) to rat plasma.

Data treatment

The percentage inhibition was calculated as $[(ratio_{control} - ratio_{inhibitor})/ratio_{control}] \times 100\%$, where ratio = Area ratio of OC to O. The % inhibition was then plotted against the logarithm of the concentration of inhibitor in μ M. The logarithm of IC₅₀, the half maximal inhibitory concentration, was evaluated from the Sigmoidal dose-response equation using Prism program (version 3.03, GraphPad Software, Inc.).

Summary

Our newly synthesized allyl diarylphosphonates were first identified as potent carboxylesterase inhibitors with broad range of inhibitory efficiency (reflected by log IC₅₀ from -0.29 to 0.70, or IC₅₀ from 0.52 to 4.99 μ M) and provided room for fine tuning when necessary. Those exhibit much stronger inhibitory effect than those of P(OPh)₃ and H-P(O)(OPh)₂ were quickly identified by using the new phosphonylation methodology developed here. The results obtained here provided us new directions for further optimization, and may lead to the development of a new generation of carboxylesterase inhibitors.

Results and discussion

a. Reference compounds

Reference compound	LOG	IC ₅₀ (µM)	
	Calculated value Standard error		Calculated value
P(OPh) ₃	0.55	0.08	3.53
HP(O)(OPh) ₂	0.97	0.19	9.28

Note: The inhibitory effect of inhibitor on the metabolism of oseltamivir (O) to oseltamivir carboxylate (OC) was presented as Log IC_{50} or IC_{50} . The lower the Log IC_{50} value indicates the lower concentration of inhibitor can be used to exhibit the inhibitory effect and thus the stronger inhibitor.

b. Allyl diarylphosphonates 2 and 2'



\mathbb{R}^1	LOG	IC ₅₀ (µM)	
	Calculated value	Standard error	Calculated value
Н	-0.21	0.09	0.62
<i>n</i> -Hex	-0.26	0.05	0.55
CH ₂ - <i>i</i> -Pr	0.15	0.03	1.40
Ph	0.22	0.07	1.67
CH ₂ Ph	-0.13	0.04	0.73
CH ₂ Bn	0.23	0.09	1.72

2

Remark: $Bn = CH_2Ph$

All the above six allyl diarylphosphonates **2** studied exhibits stronger inhibitory effect (log IC₅₀ ranges from -0.26 to 0.23) than those of reference compounds (log IC₅₀ between 0.55 to 0.97, section a). Among the allyl diarylphosphonates, that with $R^1 = n$ -Hex was found to be most potent inhibitor.



Several allyl diarylphosphonates with general structure of 2' were also examined, however, their inhibitory effects are much lower than those of 2 in general: e.g. for $R^2 = Cy$ and Ph, log IC₅₀ are 0.70 and 0.57, respectively (with standard error: 0.02 and 0.07).

c. Effect of OMe in o-, m-, p-position of allyl diarylphosphonates 2 and amine substituent

P(O)(OPh)₂

n-He>	ć 2		
Substituent	LOG	IC ₅₀	IC ₅₀ (µM)
	Calculated value	Standard error	Calculated value
<i>p</i> -OMe	-0.26	0.05	0.55
<i>m</i> -OMe	0.29	0.07	1.93
o-OMe	-0.29	0.10	0.52
<i>p-,o-di</i> OMe	-0.19	0.09	0.64

Among the monosubstituted OMe compounds, *o*-OMe and *p*-OMe (log IC₅₀ of -0.29 and -0.26, respectively) exhibit stronger inhibition than *m*-OMe (log IC₅₀ of 0.29), but not much enhancement was observed for *o*-, *p*-disubstituted OMe.

In addition, allyl diarylphosphonate with amine substituent (log IC_{50} of 0.46, standard error of 0.14, structure is shown below) lowers the inhibitory effect than those methoxy substituents.



d. α-branch



Inhibitor	LOG IC ₅₀		IC ₅₀ (µM)
	Calculated value Standard error		Calculated value
2'n	-0.13	0.11	0.74

e. Effect of R^4



R ⁴	LOG	IC ₅₀ (µM)	
	Calculated value	Standard error	Calculated value
<i>p</i> -C ₆ H ₄ Me	0.30	0.12	1.98
p-C ₆ H ₄ Cl	0.47	0.09	2.93

In both compounds, their inhibitory effect is weaker than those allyl diarylphosphonates 2 in section b (log IC₅₀ between -0.26 to 0.23).

f. Representative Sigmoidal dose-response curve of allyl diarylphosphonate inhibitor.



Reference:

Chang Q, Chow MSS and Zuo Z. Studies on the influence of esterase inhibitor to the pharmacokinetic profiles of oseltamivir and oseltamivir carboxylate in rats using an improved LC/MS/MS method. *Biomedical Chromatography* **2009**; *23*: 852-857.



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P-(0-()3

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