

Tandem Achmatowicz-Knoevenagel protocol: Diastereoselective synthesis and anticancer evaluation of cyclopenta[b]pyrane derivatives

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Experimental Section

All reagents were used as purchased from commercial suppliers without further purification. The reactions were carried out in oven dried or flamed graduated vessels. Solvents were dried and purified by conventional methods prior use. Flash column chromatography was performed with Silica gel 60, 0.040-0.063 mm (230-400 mesh). Aluminium backed plates pre-coated with silica gel 60 (UV254) were used for thin layer chromatography. ¹H, ¹³C spectra were recorded on 250 MHz / 63 MHz or 400/ 100.7 MHz spectrometers. Splitting patterns are designated as s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; br, broad. Chemical shifts (δ) are given in ppm relative to the resonance of their respective residual solvent peak, CHCl₃ (7.27 ppm, 1H; 77.16 ppm, the middle peak, ¹³C). Elemental analysis was performed on elemental analyzer. The FD mass spectra were recorded using a mass spectrometer connected to a PDO 11/34 (DEC) computer system. All compounds were determined to be >95% pure by high-performance liquid chromatography (HPLC). Purity of compounds

were determined on a Phenomenex Luna C18- (2), 3 mm column, 4.6 mm i.d. × 30 mm length, with 30-75% acetonitrile/water/0.1% trifluoroacetic acid, 1.0 mL/min elution at rt using 210, 254, or 280 nm wavelength.

General procedure of the synthesis of compounds 5a and 5b. A suspension of NaH (174 mg, 4.4 mmol, 65% dispersion in mineral oil) in 10 mL of THF at 0 °C was cautiously treated with *tert*-butyl acetoacetate (0.66 mL, 4 mmol) under argon over a 15 min period. After stirring at this temperature for 30 min, a solution of *n*-BuLi (2.75 mL, 4.4 mmol, 1.6 M in *n*-hexane) was added dropwise over 10 min. The mixture was stirred at 0 °C for 30 min. The resultant milky solution was cooled to -78 °C then 4 mmoles of either furfural (384 mg) or 2-acetylfuran (440 mg) in 10 mL of THF was added. The reaction was then stirred at the same temperature for 1 h, after which the mixture was quenched with a saturated solution of NH₄Cl (15 mL), extracted with EtOAc (3 × 30 mL), dried over Na₂SO₄ and concentrated under vacuum to give a yellowish crude oil. The crude was purified through a short silica gel column using hexane/ EtOAc (9:1) as eluent to afford compounds **5a** (917 mg, 95% yield) and **5b** (977 mg, 91% yield) as a yellowish oils.

***tert*-Butyl 5-(2-furyl)-5-hydroxy-3-oxopentanoate (5a).** ¹H NMR (250 MHz, CDCl₃) δ 7.31 (dd, *J* = 0.8, 1.8 Hz, 1H, H-5'), 6.30 (dd, *J* = 1.8, 3.2 Hz, 1H, H-4'), 6.22 (d, *J* = 3.2 Hz, 1H, H-3'), 5.13 (dd, *J* = 3.3, 8.3 Hz, 1H, H-5), 3.37 (s, 2H, H-2a, H-2b), 3.10 (dd, *J* = 8.3, 17.5 Hz, 1H, H-4a), 2.97 (dd, *J* = 3.3, 17.5 Hz, 1H, H-4b), 1.39 (CO₂C(CH₃)₃); ¹³C NMR (63 MHz, CDCl₃) δ 202.7 (C-2), 166.1 (CO₂C(CH₃)₃), 154.8 (C-2'), 110.3 (C-5'), 110.2 (C-4'), 106.3 (C-3'), 82.3 (CO₂C(CH₃)₃), 63.5 (C-5), 51.1 (C-2), 47.8 (C-4), 27.9 (CO₂C(CH₃)₃); FD-MS *m/z* = 255 (*M* + 1)⁺, Anal. Calcd for C₁₃H₁₈O₅ (254.28); C, 61.40; H, 7.14. Found: C, 61.20; H, 7.31.

***tert*-Butyl 5-(2-furyl)-5-hydroxy-3-oxohexanoate (5b).** ¹H NMR (250 MHz, CDCl₃) δ 7.22 (dd, *J* = 0.9, 1.8 Hz, 1H, H-5'), 6.02 (dd, *J* = 1.8, 3.2 Hz, 1H-4'), 6.15 (dd, *J* = 0.9, 3.3 Hz, 1H, H-3'), 3.25 (bs, 2H, H-2a, H-2b), 3.18 (d, *J* = 16.5 Hz, 1H, H-4a), 2.82 (d, *J* = 16.5 Hz, 1H, H-4b), 1.43 (CH₃), 1.63 (CO₂C(CH₃)₃); ¹³C NMR (63 MHz, CDCl₃) δ 203.0 (C-3), 165.0 (CO₂C(CH₃)₃), 158.5 (C-2'), 110.3 (C-5'),

110.2 (C-4'), 104.7 (C = 3'), 82.0 (CO₂C(CH₃)₃), 70.0 (C-5), 51.7 (C-2), 51.5 (C-4), 28.2 (CO₂C(CH₃)₃), 27.6 (CH₃); FD-MS m/z = 269 (M + 1)⁺, Anal. Calcd for C₁₄H₂₀O₅ (268.31); C, 62.67; H, 7.51. Found: C, 62.51; H, 7.42.

General procedure for the synthesis of compounds 7a and 7b. A solution of **5a** (1270 mg, 5 mmol) or **5b** (1341.5 mg, 5 mmol) in dichloromethane (40 mL) at 0 °C was treated with *m*-chloroperbenzoic acid (1376 mg, 8.0 mmol of 80% aqueous slurry) which was dissolved in 30 ml DCM, dried over Na₂SO₄, filtered, diluted with 50 ml toluene and dried under reduced pressure) and stirred at 0 °C for 6 h. Sodium sulfite solution (30 mL of 10%) was introduced, and the layers were separated after 1 h of rapid mixing. The aqueous phase was washed with saturated sodium bicarbonate solution (30 mL), brine and water (30 mL) prior to drying and solvent evaporation. The crude viscous yellowish oils (**6a** and **6b**) was used in the next step without further purification.

A solution of the crude anomeric mixture of **6a** (852 mg, 3 mmol) or **6b** (810 mg, 3 mmol) in dry toluene (30 ml), freshly prepared piperidinium acetate (3.9 mmol) and 4 °A molecular sieves (250 mg) was stirred at 50 °C for 4 h. The hot mixture was filtered and evaporated under reduced pressure. The brown viscous oil was dissolved in EtOAc and extracted with saturated solution of NaHCO₃, brine, water and dried over Na₂SO₄. The crude was purified on a silica gel column using hexane/ EtOAc (7:3) as eluent to afford compounds **7a** (350 mg, 68% yield) and **7b** (425 mg, 73% yield) as amorphous solids.

tert-Butyl (7aS)-2-hydroxy-6-oxo-2,6,7,7a-tetrahydrocyclopenta[b]pyran-5-carboxylate (7a). ¹H NMR (250 MHz, CDCl₃) δ 7.25 (bt, *J* = 10.1 Hz, 1H, H-3), 6.42 (dd, *J* = 3.3, 10.1 Hz, 1H, H-4), 5.60 (bs, 1H, H-2), 5.16 (dd, *J* = 4.7, 6.8 Hz, 1H, H-7a), 2.84 (dd, *J* = 6.8, 17.6 Hz, 1H, H-7), 2.51 (dd, *J* = 4.6, 17.6 Hz, 1H, H-7'), 15.3 (s, 9H, ^tBuO); ¹³C NMR (63 MHz, CDCl₃) δ 199.1, 199.2 (C-6), 170.5, 170.7 (C-4a), 166.2 (CO₂C(CH₃)₃), 142, 138.2 (C-3), 125.0, 123.0 (C-4), 92.7, 89.1 (C-2), 82.0, 71.5 (CO₂C(CH₃)₃), 65.5 (C-7a), 42.5, 42.3 (C-7), 28.2 (CO₂C(CH₃)₃); FD-MS m/z = 252 (M)⁺, Anal. Calcd for C₁₃H₁₆O₅ (252.10); C, 61.90; H, 6.39. Found: C, 61.85; H, 6.43.

***tert*-Butyl (7a*S*)-2-hydroxy-7a-methyl-6-oxo-2,6,7,7a-tetrahydrocyclopenta[*b*]pyran-5-carboxylate (7b).** ¹H NMR (250 MHz, CDCl₃) δ 7.13 (m, 2H, H-3), 6.32 (m, 2H, H-4), 5.59, 5.51 (bs, 2H, H-2), 2.57 (m, 4H, H-7,7'), 1.42 (bs, 12H, CH₃-7a, CO₂C(CH₃)₃); ¹³C NMR (63 MHz, CDCl₃) δ 198.8, 198.5 (C-6), 170.5, 170.4 (C-4a), 161.1 (CO₂C(CH₃)₃), 141.5, 138.5 (C-3), 125.7, 125.3 (C-5), 122.5, 121.3 (C=4), 89.6, 88.7 (C-2), 82.5 (CO₂C(CH₃)₃), 77.5, 73.9 (C-7a) 52.2, 51.9 (C-7), 28.9, 24.2 (CH₃ = 7a), 28.1, 27.8 (CO₂C(CH₃)₃); FD-MS m/z = 266 (M)⁺, Anal. Calcd for C₁₃H₁₆O₅ (266.12); C, 63.15; H, 6.81. Found: C, 63.09; H, 6.87.

General procedure for the synthesis of compounds 8a, 8b and 9. A solution of the anomeric mixture of **7a** (756 mg, 3 mmol) or **7b** (798 mg, 3 mmol) in MeI (15 mL) at rt was treated with 3.0 mmol of Ag₂O. The progress of reaction was monitored by TLC (12 h). When complete, the reaction mixture was filtered over celite. The filtrate was washed with saturated solutions of sodium sulfite, sodium bicarbonate, brine, dried and evaporated. The residue was purified on silica gel column using hexane/EtOAc (9:1) as an eluent.

***tert*-Butyl (2*S*,7a*S*)-2-methoxy-6-oxo-2,6,7,7a-tetrahydrocyclopenta[*b*]pyran-5-carboxylate (8a).** Yiel: 540 mg, 80%; Significant NMR NOEs are 2-H to 7a-H, 36%; 7a-H to 2-H, 32%. ¹H NMR (250 MHz, CDCl₃) δ 7.20 (dd, *J* = 0.4, 10.1 Hz, 1H, H-3), 6.30 (dd, *J* = 3.3, 10.1 Hz, 1H, H-4), 5.00 (dd, *J* = 3.3 Hz, 1H, H-2), 4.96 (dd, *J* = 4.8, 6.9 Hz, 1H, H-7a), 3.45 (s, 3H, OMe), 2.80 (dd, *J* = 7.0, 17.6 Hz, 1H, H-7), 2.48 (dd, *J* = 4.5, 17.6 Hz, 1H, H-7'), 1.48 (s, 9H, ^tBuO); ¹³C NMR (63 MHz, CDCl₃) δ 197.8 (C-6), 166.1 (C-4a), 160.8 (CO₂C(CH₃)₃), 137.2 (C-3), 123.0 (C-4), 127.0 (C-5), 95.6 (C-2), 82.5 (CO₂C(CH₃)₃), 65.5 (C-7a), 56.4 (OMe), 42.3 (C-7), 28.0 (CO₂C(CH₃)₃); FD-MS m/z = 266 (M)⁺, Anal. Calcd for C₁₄H₁₈O₅ (266.16); C 63.12, H 6.82. Found: C 63.51, H 6.55.

***tert*-Butyl (2*S*,7a*S*)-2-methoxy-7a-methyl-6-oxo-2,6,7,7a-tetrahydrocyclopenta[*b*]pyran-5-carboxylate (8b).** Yiel: 742 mg, 94%; Significant NMR NOEs are 2-H to 7a-CH₃, 31%; 7a-CH₃ to 2-H, 33%. ¹H NMR (250 MHz, CDCl₃) δ 7.26 (dd, *J* = 1.7, 10.0 Hz, 1H, H-3), 6.62 (dd, *J* = 1.6, 10.0 Hz, 1H, H-4), 5.10 (bd, *J* = 3.1 Hz, 1H, H-2), 3.44 (s, 3H, OMe), 2.74 (d, *J* = 17.1 Hz, 1H, H-7), 2.57 (d, *J* = 17.1 Hz, 1H, H-7'), 1.48 (s, 3H, 7a-CH₃), 1.47 (s, 9H, (CO₂C(CH₃)₃)); ¹³C NMR (63 MHz, CDCl₃) δ 197.7 (C-6), 169.4 (C-4a), 160.9

(CO₂C(CH₃)₃), 139.0 (C-2), 123.8 (C-3), 123.7 (C-5), 95.6 (C-2), 82.3 (CO₂C(CH₃)₃), 76.7 (C-7a), 55.1 (OMe), 52.1 (C-7), 28.2 (CO₂C(CH₃)₃), 24.3 (7a-CH₃); FD-MS m/z = 280 (M)⁺, Anal. Calcd for C₁₅H₂₀O₅ (280.32); C 64.27, H 7.19; Found: C 63.95, H 7.32.

tert-Butyl (2R,7aS)-2-methoxy-6-oxo-2,6,7,7a-tetrahydrocyclopenta[b]pyran-5-carboxylate (9). Yield: 138 mg, 20%; Significant NMR NOEs are 2-OMe to 7a-H, 37%; 7a-H to 2-OMe, 29%. ¹H NMR (250 MHz, CDCl₃) δ 7.25 (dd, *J* = 2.1, 10.1 Hz, 1H, H-3), 6.32 (dd, *J* = 1.4, 10.1 Hz, 1H, H-4), 5.34 (dd, *J* = 0.6, 2.0 Hz, 1H, H-2), 4.74 (dd, *J* = 4.7, 6.9 Hz, 1H, H-7a), 3.47 (s, 3H, OMe), 2.83 (dd, *J* = 6.8, 17.7 Hz, 1H, H-7), 2.55 (dd, *J* = 4.6, 17.7 Hz, 1H, H-7'), 1.47 (s, 9H, (CO₂C(CH₃)₃)); ¹³C NMR (63 MHz, CDCl₃) δ 171.5 (CO₂C(CH₃)₃), 169.7 (C-4a), 141.6 (C-3), 124.8 (C-4), 98.7 (C-2), 82.0 (CO₂C(CH₃)₃), 71.4 (C-7a), 56.0 (OMe), 42.6 (C-7), 28.2 (CO₂C(CH₃)₃); FD-MS m/z = 266 (M)⁺, Anal. Calcd for C₁₄H₁₈O₅ (266.12); C 63.15, H 6.81; Found: C 62.98, H 7.11.

tert-Butyl (2S,4aS,5R,7aS)-2-methoxy-7a-methyl-6-oxooctahydrocyclopenta[b]pyran-5-carboxylate (10). To a stirred solution of **8b** (560 mg, 2.0 mmol) in dry EtOAc/MeOH (1:1, 20 mL) was added Pd/C (56 mg, 15% Pd/C w/w). The mixture was placed under 1.0 atm of H₂ pressure, and the progress of reaction was monitored by TLC. After 2 h, the solid was removed by filtration through a celite pad, which was washed repeatedly with EtOAc. After concentration of the filtrate, the residue was purified on silica gel column (elution with 5% ethyl acetate in hexane) to give **10** as amorphous solid (430 mg, 87%). Significant NMR NOEs are 4a-H to 5-H, 37%; 7a-CH₃ to 2-H, 31%; 7a-CH₃ to 2-H, 35%; 4aH to 2-OMe, 21%. ¹H NMR (250 Mz, CDCl₃) δ 4.73 (dd, *J* = 2.4, 9.8 Hz, 1H, H-2), 3.29 (d, *J* = 6.2 Hz; 1H, H-5), 3.24 (s, 3H, OMe), 2.79 (d, *J* = 18.4 Hz, ; H-7), 2.45 (ddd, *J* = 2.3, 4.6, 6.2 Hz, 1H, H-4a), 2.31 (d, *J* = 18.4 Hz, 1H, H-7'), 1.97 (m, 2H), 1.74 (dm, *J* = 17.0 Hz, 1H) 1.65 (dm, *J* = 13.0 Hz, 1H), 1.49 (s, 3H, CH₃-7a), 1.41 (s, 9H, CO₂C(CH₃)₃); ¹³C NMR (63 MHz, CDCl₃) δ 210.3 (C-6), 171.0 (CO₂C(CH₃)₃), 95.7 (C-2), 82.2 (CO₂C(CH₃)₃), 81.7 (C-7a), 60.2 (C-5), 56.8 (OMe), 55.0 (C-7), 51.5 (C-4a), 34.0 (C-3), 28.4(CO₂C(CH₃)₃), 25.4 (C-4), 24.7 (CH₃-7a); FD-MS m/z = 284 (M)⁺, Anal. Calcd for C₁₅H₂₄O₅ (284.35); C 63.40, H 8.51; Found: C 63.52, H 8.46.

General procedure for the synthesis of compounds 11a-f. The bicyclic β -ketoester **10** derived from the reduction product of **8b** (142 mg, 0.5 mmol) and the desired hydrazine derivative (0.52 mmol) in 20 ml absolute ethanol was refluxed, and the progress of reaction was monitored by TLC. After 6 h, the reaction mixture was concentrated on a rotary evaporator, the residue was purified on silica gel column (elution with 10% ethyl acetate in hexane) to give **11a-f** as amorphous solids.

(3bS,6S,7aS)-6-Methoxy-7a-methyl-2-(4-methylphenyl)-1,2,3b,4,5,6,7a,8-octahydro-3H-pyrano[3',2':3,4]cyclopenta[1,2-c]pyrazol-3-one (11a). Yield: 122 mg, 88%; ^1H NMR (400 Mz, CDCl_3) δ 7.70 (d, $J = 7.3$ Hz, 2H, Ar-H), 7.17 (d, $J = 7.3$ Hz, 2H, Ar-H), 4.64 (dd, $J = 2.7, 7.9$ Hz, 1H, H-2), 3.04 (bd, $J = 6.2$ Hz; 1H), 3.30 (s, 3H, OMe), 2.25 (dd, $J = 4.6, 11.3$ Hz, 1H), 2.20 (m, 3H), 2.17 (s, 3H, Ar-CH₃), 1.90 (m, , 3H), 1.45 (s, 3H, CH₃-7a); ^{13}C NMR (100.7 MHz, CDCl_3) δ 159.6, 143.0, 141.7, 137.4, 133.4, 132.6, 125.9, 115.6, 94.2, 84.1, 60.2, 56.8, 55.0, 34.1, 25.4, 24.7, 20.9; FD-MS $m/z = 314$ (M)⁺, Anal. Calcd for $\text{C}_{18}\text{H}_{22}\text{N}_2\text{O}_3$ (314.38); C, 68.77; H, 7.05; N, 8.91. Found: C, 68.56; H, 7.24; N, 9.07.

4-[(3bS,6S,7aS)-6-Methoxy-7a-methyl-3-oxo-1,3,3b,4,5,6,7a,8-octahydro-2H

pyrano[3',2':3,4]cyclopenta[1,2-c]pyrazol-2-yl]benzotrile (11b). Yield: 91.5 mg, 75% ^1H NMR (400 Mz, CDCl_3) δ 7.81 (dd, $J = 1.2, 7.4$ Hz, 2H, Ar-H), 7.73 (dd, $J = 1.1, 7.4$ Hz, 2H, Ar-H), 4.90 (dd, $J = 3.1, 8.2$ Hz, 1H, H-2), 3.01 (bd, $J = 6.4$ Hz; 1H), 3.41 (s, 3H, OMe), 2.62 (m, 2H), 2.46 (dd, $J = 4.3, 10.1$ Hz, 1H), 2.23 (m, 2H), 1.92 (m, , 2H), 1.49 (s, 3H, CH₃-7a); FD-MS $m/z = 325$ (M)⁺, Anal. Calcd for $\text{C}_{18}\text{H}_{19}\text{N}_3\text{O}_3$ (325.14); C, 66.45; H, 5.89; N, 12.91. Found: C, 66.27; H, 5.95; N, 12.70.

(3bS,6S,7aS)-6-Methoxy-7a-methyl-2-(4-nitrophenyl)-1,2,3b,4,5,6,7a,8-octahydro-3H-pyrano[3',2':3,4]cyclopenta[1,2-c]pyrazol-3-one (11c). Yield: 104 mg, 78%; ^1H NMR (400 Mz, CDCl_3) δ 8.34 (dd, $J = 0.9, 6.9$ Hz, 2H, Ar-H), 8.18 (dd, 1.0, $J = 6.9$ Hz, 2H, Ar-H),

4.85 (dd, $J = 2.9, 7.0$ Hz, 1H, H-2), 3.01 (m, 1H), 3.53 (s, 3H, OMe), 3.01 (dd, $J = 2.1, 4.5$ Hz, 1H), 2.71-2.79 (m, 2H), 2.45(dt, $J = 3.3, 10.1$ Hz, 1H), 2.20 (m, 2H), 1.89 (m, 2H), 1.47 (s, 3H, CH₃-7a); ¹³C NMR (100.7 MHz, CDCl₃) δ 161.6, 144.3, 142.1, 133.0, 122.7, 119.5, 115.0, 94.9, 84.7, 55.7, 49.2, 44.1, 30.8, 22.4, 22.0; FD-MS $m/z = 345$ (M)⁺, Anal. Calcd for C₁₇H₁₉N₃O₅ (345.13); C, 59.12; H, 5.55; N, 12.1. Found: C, 59.22; H, 5.67; N, 12.31.

(3bS,6S,7aS)-6-Methoxy-2-(4-methoxyphenyl)-7a-methyl-1,2,3b,4,5,6,7a,8-octahydro-3H-pyrano[3',2':3,4]cyclopenta[1,2-c]pyrazol-3-one (11d). Yield: 110 mg, 82%; ¹H NMR (400 Mz, CDCl₃) δ 7.61 (dd, $J = 0.9, 6.5$ Hz, 2H, Ar-H), 6.95 (dd, 1.0, $J = 6.5$ Hz, 2H, Ar-H), 4.93 (dd, $J = 2.2, 6.2$ Hz, 1H, H-2), 3.77 (s, 3H, Ar-OMe), 3.00 (m, 1H), 3.40 (s, 3H, OMe), 2.75 (dd, $J = 2.3, 4.4$ Hz, 1H), 1.88 (m, 1H), 2.42(dt, $J = 3.3, 9.5$ Hz, 1H), 2.20 (m, 1H), 1.50 (s, 3H, CH₃-7a); ¹³C NMR (100.7 MHz, CDCl₃) δ 161.75, 132.5, 132.4, 115.1, 112.6, 94.2, 84.6, 55.7, 55.6, 46.1, 39.8, 32.8, 25.7, 25.2; FD-MS $m/z = 330$ (M)⁺, Anal. Calcd for C₁₈H₂₂N₂O₄ (330.16); C, 65.44; H, 6.71; N, 8.48, 12.1. Found: C, 65.53; H, 6.65; N, 8.52.

(3bS,6S,7aS)-2-(4-Chlorophenyl)-6-methoxy-7a-methyl-1,2,3b,4,5,6,7a,8-octahydro-3H-pyrano[3',2':3,4]cyclopenta[1,2-c]pyrazol-3-one (11e). Yield: 120.7 mg, 85%; ¹H-NMR (400 Mz, CDCl₃) δ 7.85 (dd, $J = 0.8, 6.1$ Hz, 2H, Ar-H), 7.30 (dd, 0.9, $J = 6.1$ Hz, 2H, Ar-H), 5.00 (dd, $J = 2.3, 5.7$ Hz, 1H, H-2), 3.03 (m, 1H), 3.99 (s, 3H, OMe), 2.76 (dd, $J = 2.2, 4.1$ Hz, 1H), 2.22-2.46 (m, 3H), 1.89 (m, 3H), 1.47 (s, 3H, CH₃-7a); ¹³C NMR (100.7 MHz, CDCl₃) δ 160.9, 151.8, 151.3, 139.0, 132.4, 116.8, 115.1, 94.7, 84.6, 55.7, 49.3, 46.1, 30.2, 21.8, 21.4; FD-MS $m/z = 334$ (M)⁺, Anal. Calcd for C₁₇H₁₉ClN₂O₃ (334.11); C, 60.99; H, 5.72; Cl, 10.59; N, 8.37. Found: C, 60.85; H, 5.79; Cl, 10.61; N, 8.42.

(3bS,6S,7aS)-2-(1H-Indol-2-yl)-6-methoxy-7a-methyl-1,2,3b,4,5,6,7a,8-octahydro-3H-pyrano[3',2':3,4]cyclopenta[1,2-c]pyrazol-3-one (11f). Yield: 98 mg, 76%; ¹H NMR (400 Mz, CDCl₃) δ 10.62 (bs, 1H), 10.26 (s, 1H), 7.73 (d, $J = 1.8$ Hz, 1H), 7.52 (s, 1H), 7.37 (d,

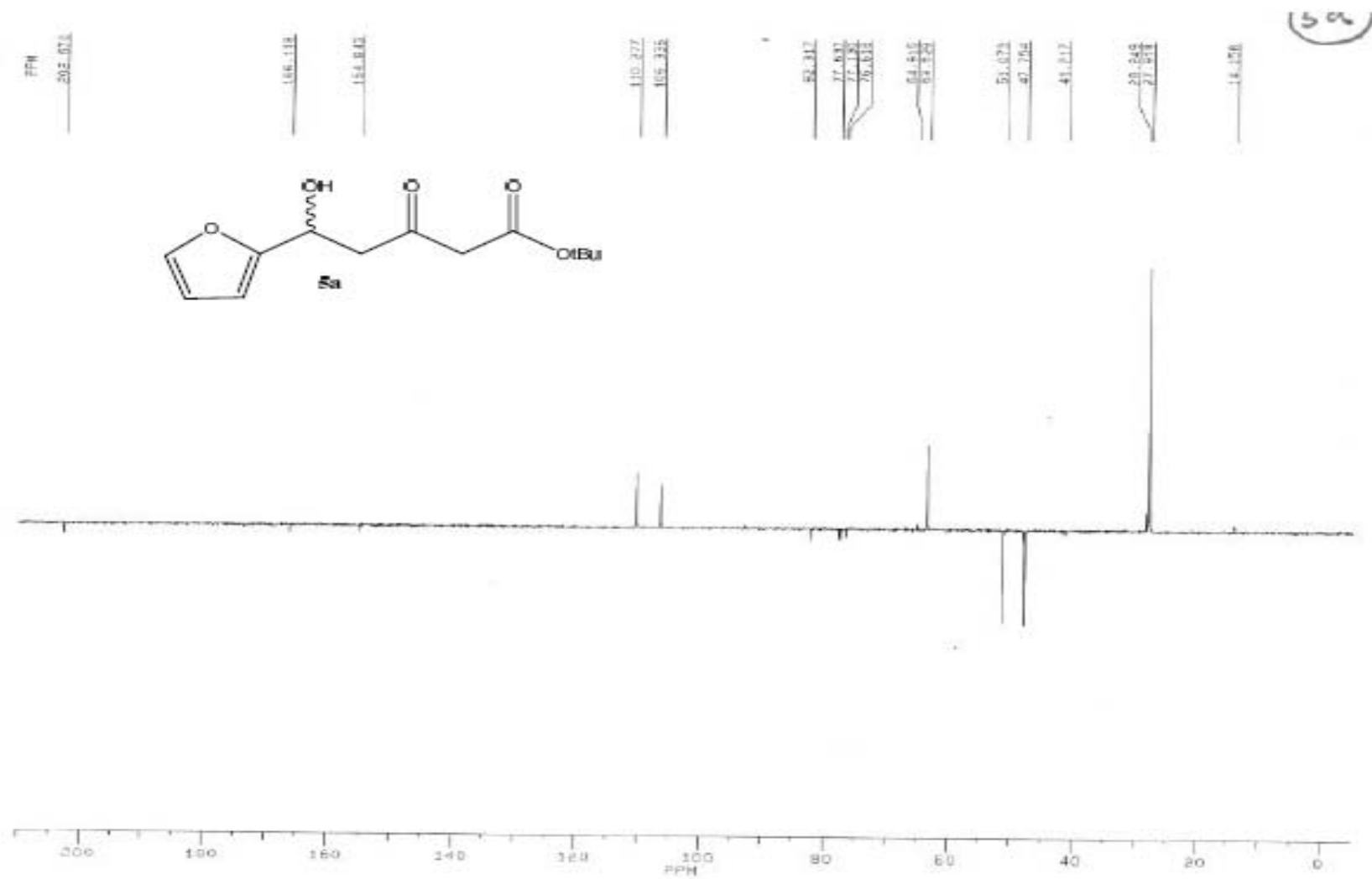
$J = 2.0$ Hz, 1H), 7.16 (t, $J = 1.8, 3.2$ Hz, 1H), 6.50 (t, $J = 1.8, 3.3$ Hz, 1H), 4.84 (dd, $J = 2.1, 5.2$ Hz, 1H, H-2), 3.00 (m, 1H), 3.49 (s, 3H, OMe), 2.77 (m, 1H), 2.21-2.46 (m, 2H), 1.88 (m, 2H), 1.47 (s, 3H, CH₃-7a); FD-MS $m/z = 334$ (M)⁺, Anal. Calcd for C₁₉H₂₁N₃O₃ (339.16); C, 67.24; H, 6.24; N, 12.38. Found: C, 67.33; H, 6.13; N, 12.45.

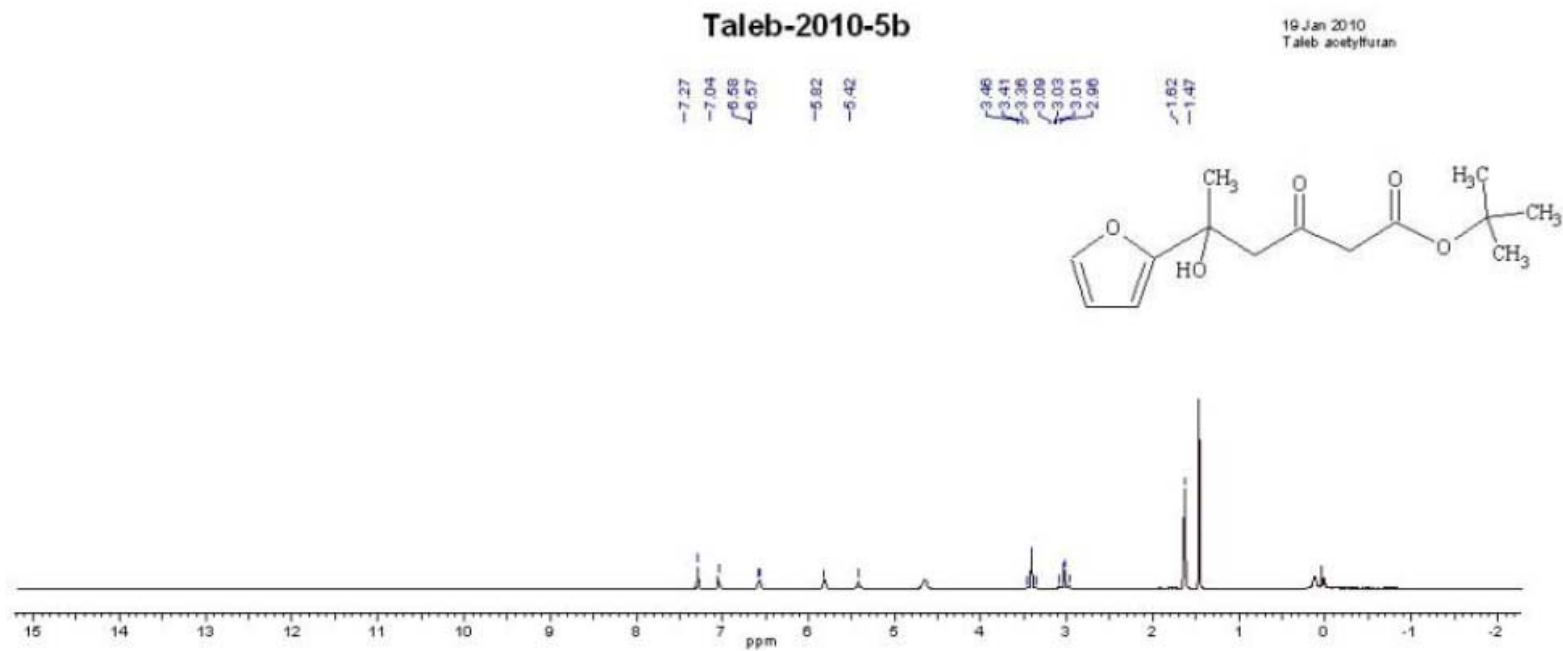
(3bS,6S,7aS)-6-Methoxy-7a-methyl-2-pyridin-2-yl-1,2,3b,4,5,6,7a,8-octahydro-3H-pyrano[3',2':3,4]cyclopenta[1,2-c]pyrazol-3-one (11g). Yield: 119 mg, 79%; ¹H NMR (400 Mz, CDCl₃) δ 8.41 (d, $J = 1.1$ Hz, 1H), 7.92 (d, $J = 0.9$ Hz, 1H), 7.63 (t, $J = 1.3$ Hz, 1H), 6.98 (t, $J = 1.2$ Hz, 1H), 5.19 (t, $J = 4.4$ Hz, 1H, H-2), 3.08 (m, 1H), 3.77 (s, 3H, OMe), 2.65 (m, 2H), 2.21-2.38 (m, 3H), 1.85 (m, 2H), 1.42 (s, 3H, CH₃-7a); FD-MS $m/z = 301$ (M)⁺, Anal. Calcd for C₁₆H₁₉N₃O₃ (301.14); C, 63.77; H, 6.36; N, 13.94. Found: C, 63.65; H, 6.42; N, 13.81.

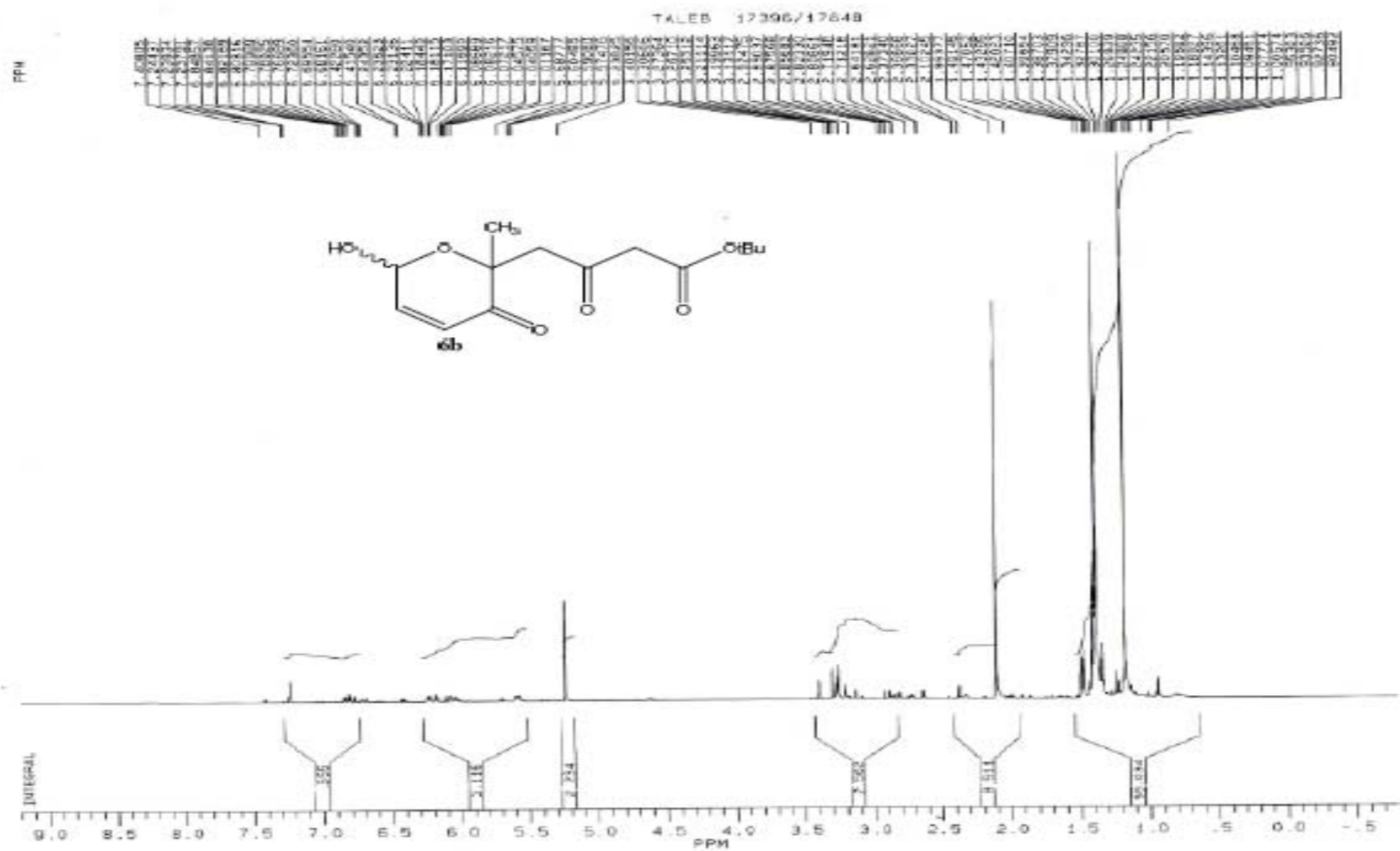
Molecular Modeling. ChemBio3D Ultra 11 was used to calculate the thermodynamically more preferred conformations of compounds **10** and **10a** using force field MM2 method.¹⁸

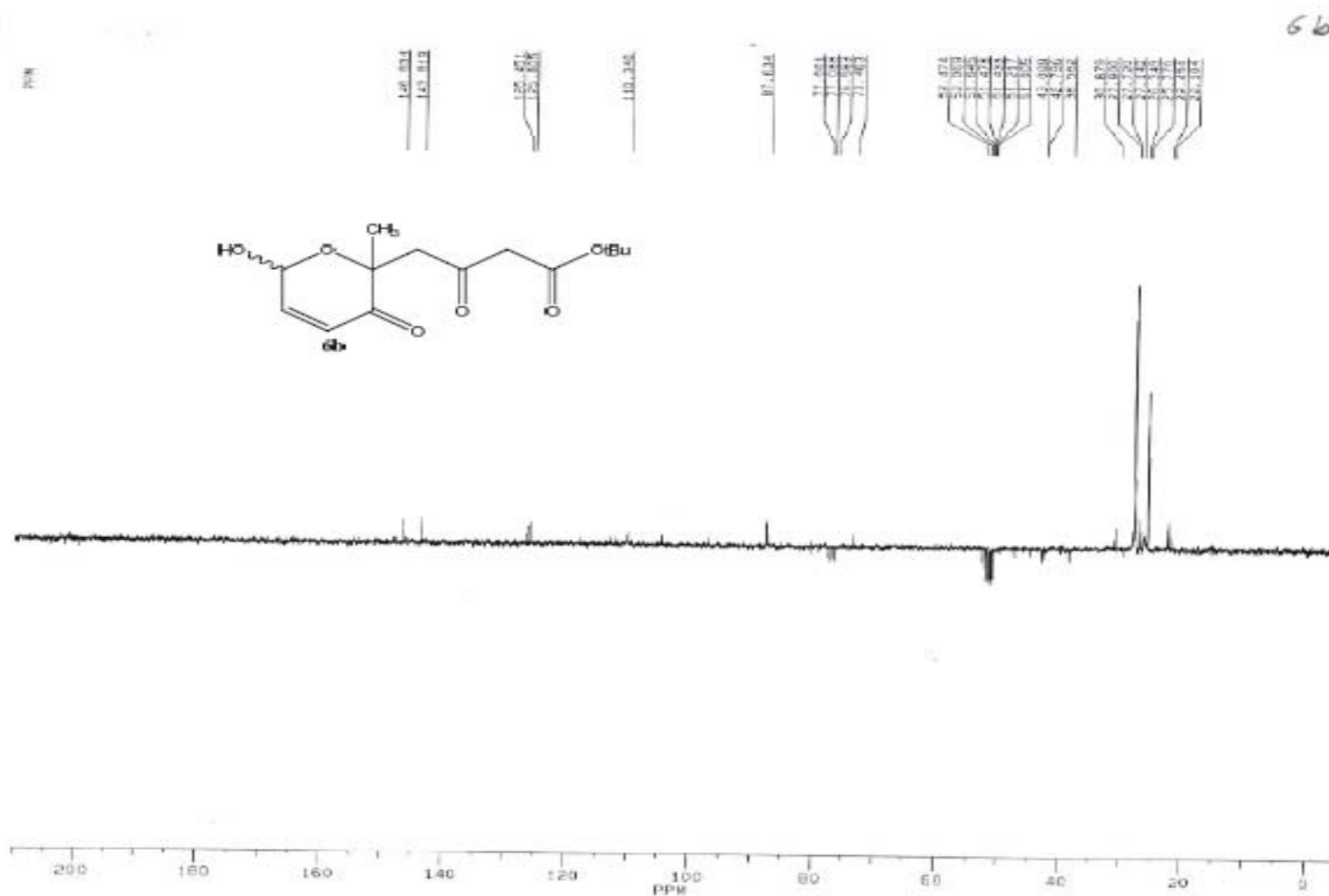
In vitro cytotoxicity assay. The cytotoxic activity of the cyclopenta[b]pyrane derivatives **11a-11g** was determined using a standard (MTT)-based colorimetric assay.¹² This assay quantifies viable cells by observing the reduction of tetrazolium salt, MTT, to formazan crystals by the cells. Based on the absorbance of the cell samples after the test is carried out, cell viability can be measured. Cells were plated with nutritional medium in 96 well plates (2000 and 5000 cells/well for HCT116, SK-N-SH and the non-tumorigenic cell line derived from breast tissue (MCF10A)). After 24 hours, cells were treated with different concentrations (0.1, 0.5, 1, 3 and 10 μ M) of the new compounds, each concentration in 3 repetitions. The plates were incubated with the pyrazolone derivatives for 72 hours. At the end of treatment, cells were washed with PBS solution. Then, 100 μ l of fresh medium and 50 μ l from a stock solution of MTT (3mg/ml PBS)

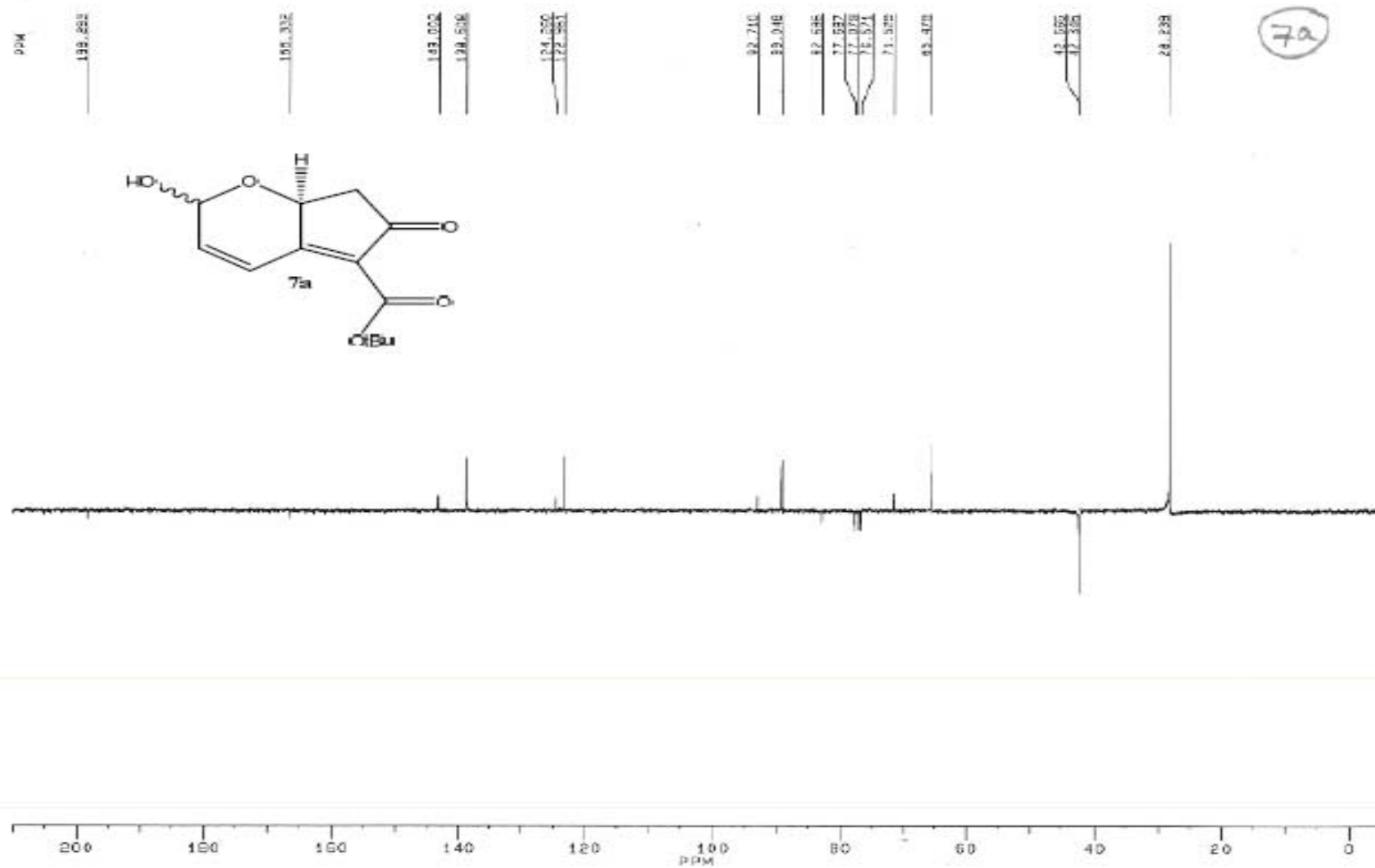
were added to each well. After 4 hours of incubation at 37 °C, the medium was discarded and 100 µl of DMSO solution were added to each well, in order to dissolve the crystals that were formed. After a 30 minute period, the absorbance of the samples was measured by an Elisa reader. The absorbance data were converted to % cell viability. The IC₅₀ were calculated using Graphpad Prism software.

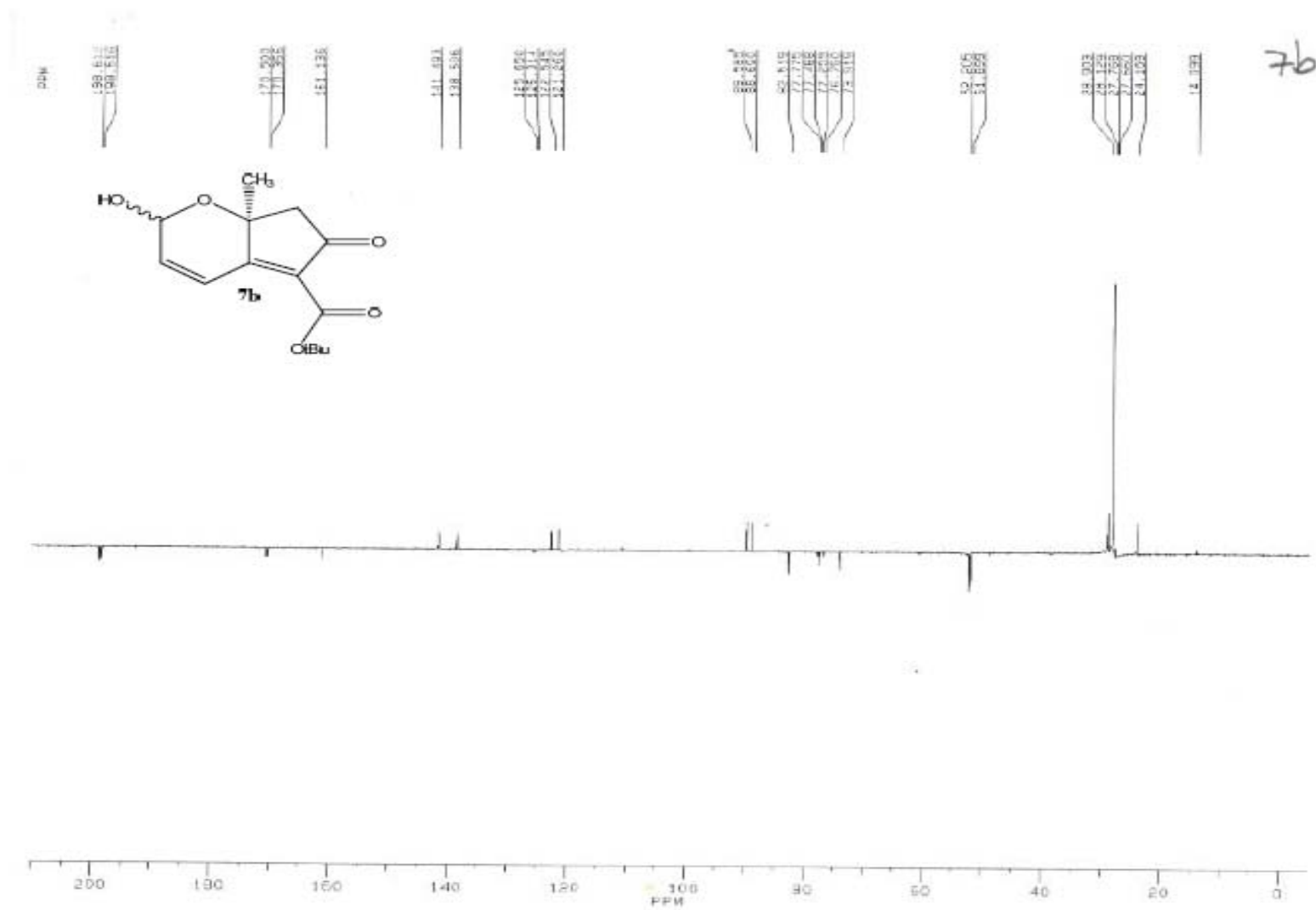


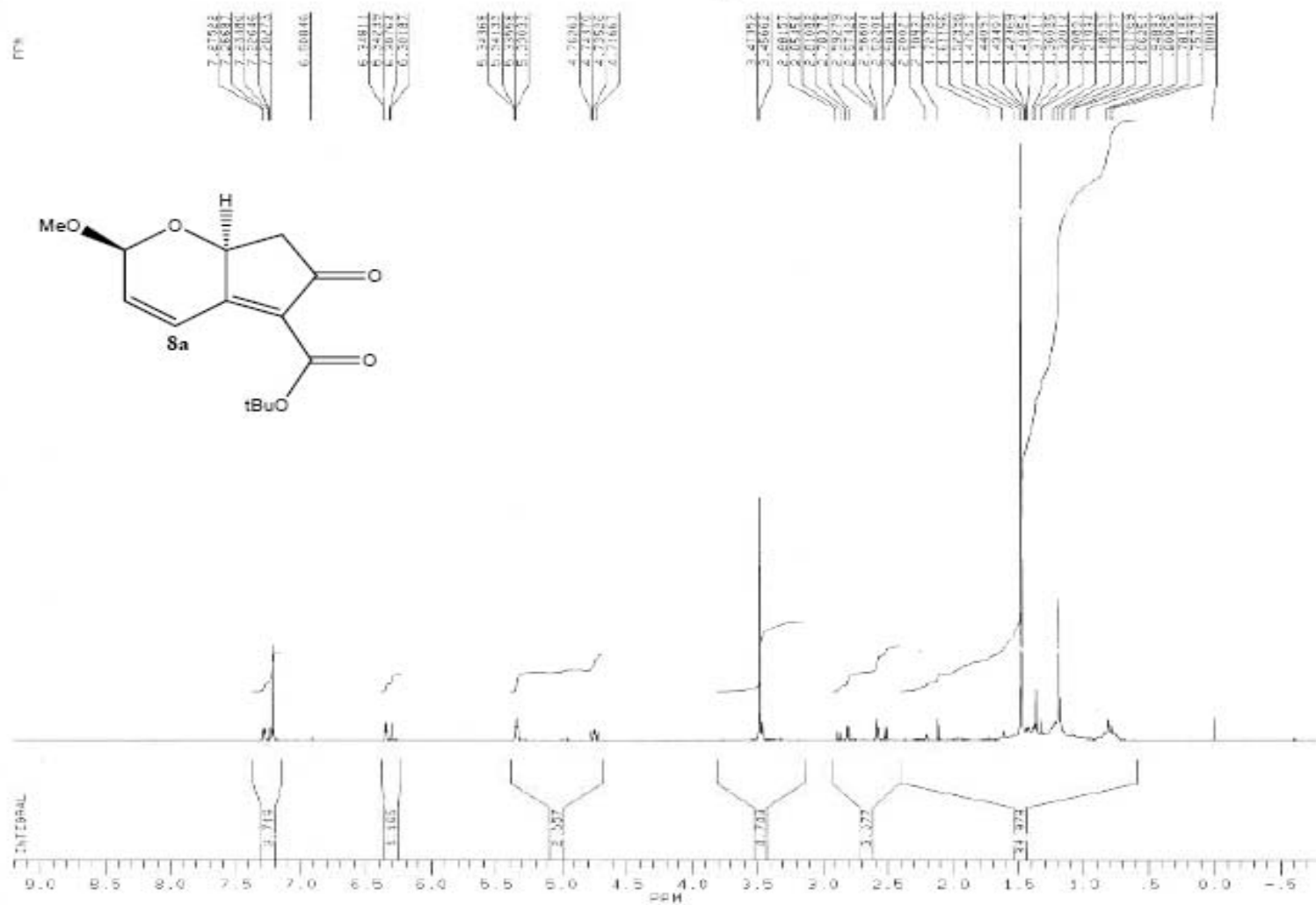












8a

DC151
AU PH
XDC
DATE
TIME

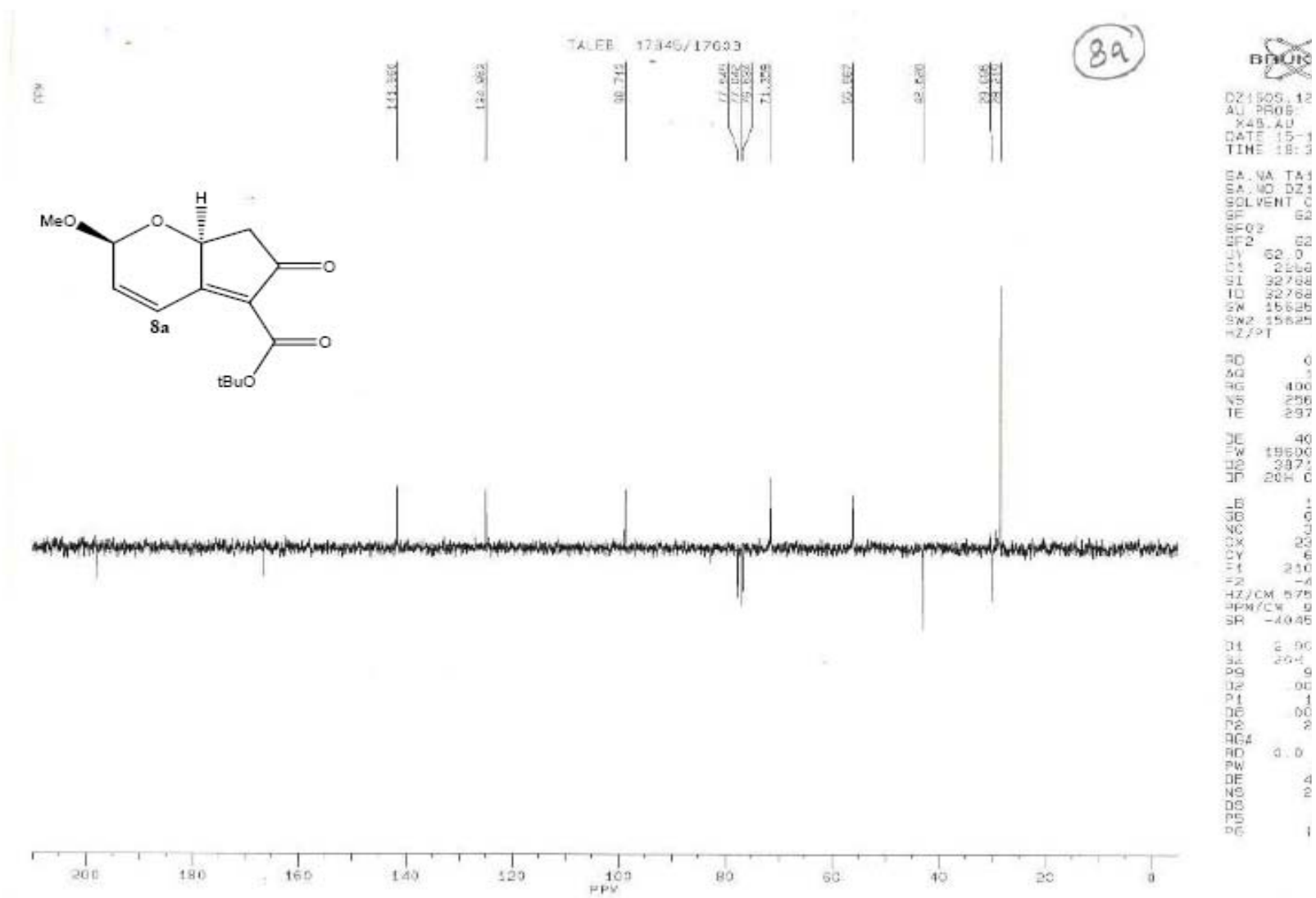
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SA NO
SOLVE
SF
SF02
SFB
SY 10
Q1
Q1 3
TD 3
SM
SMG
HZ/PT

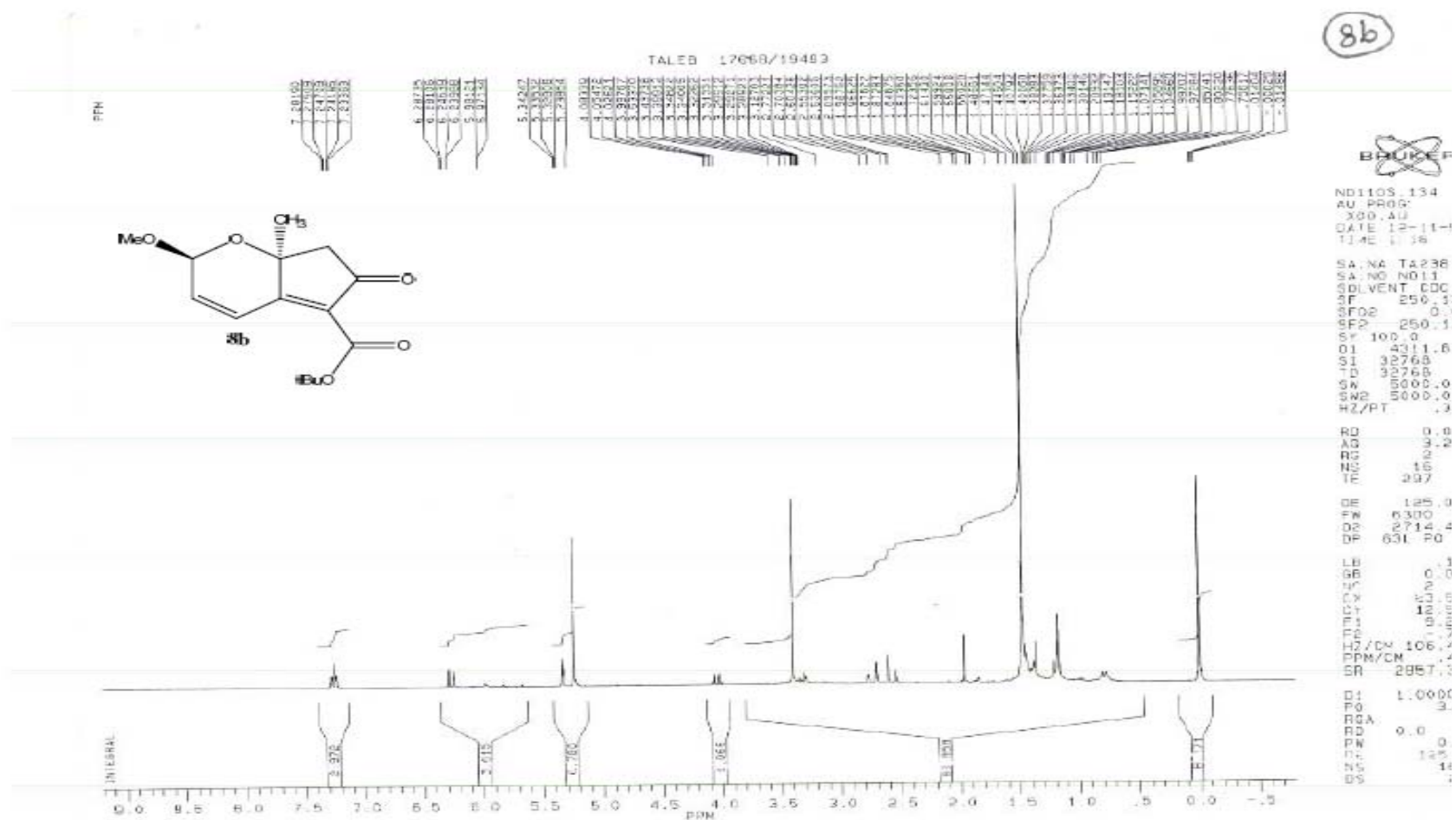
R0
AQ
RG
NS
TE

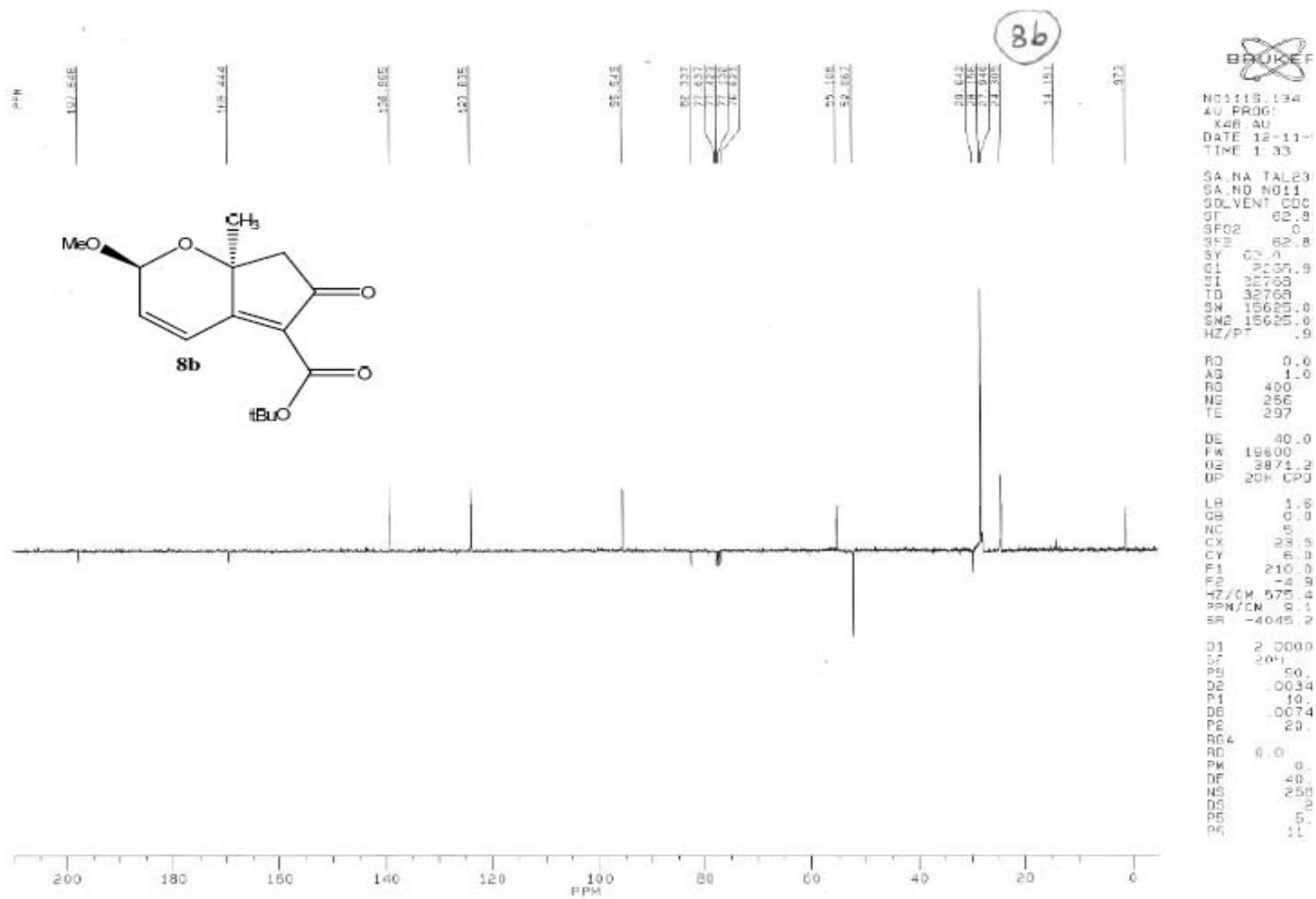
QE
FW
QP
DP 6

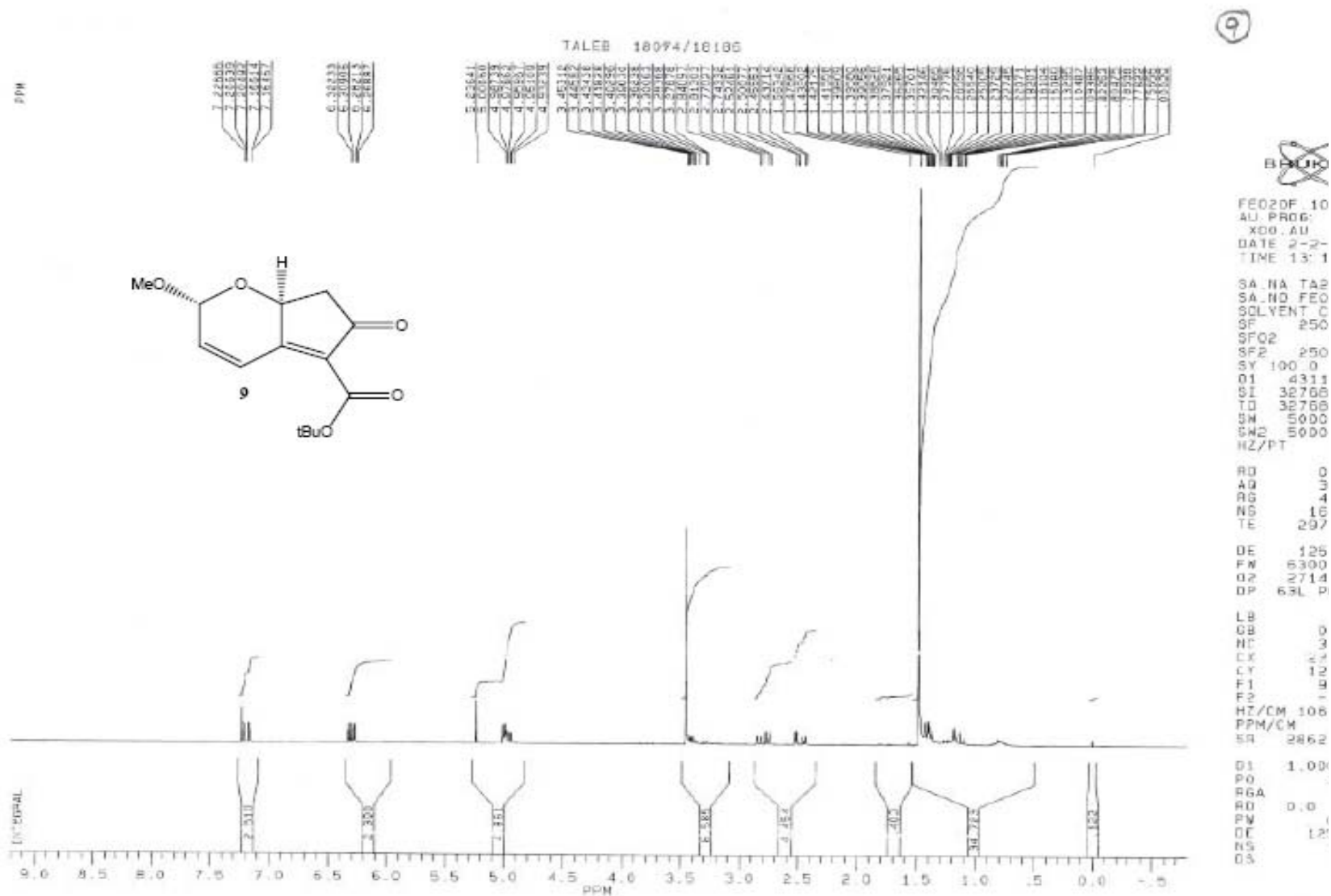
LD
GD
NC
YX
CY
F1
F2
HZ/CH
PDM/C
SR

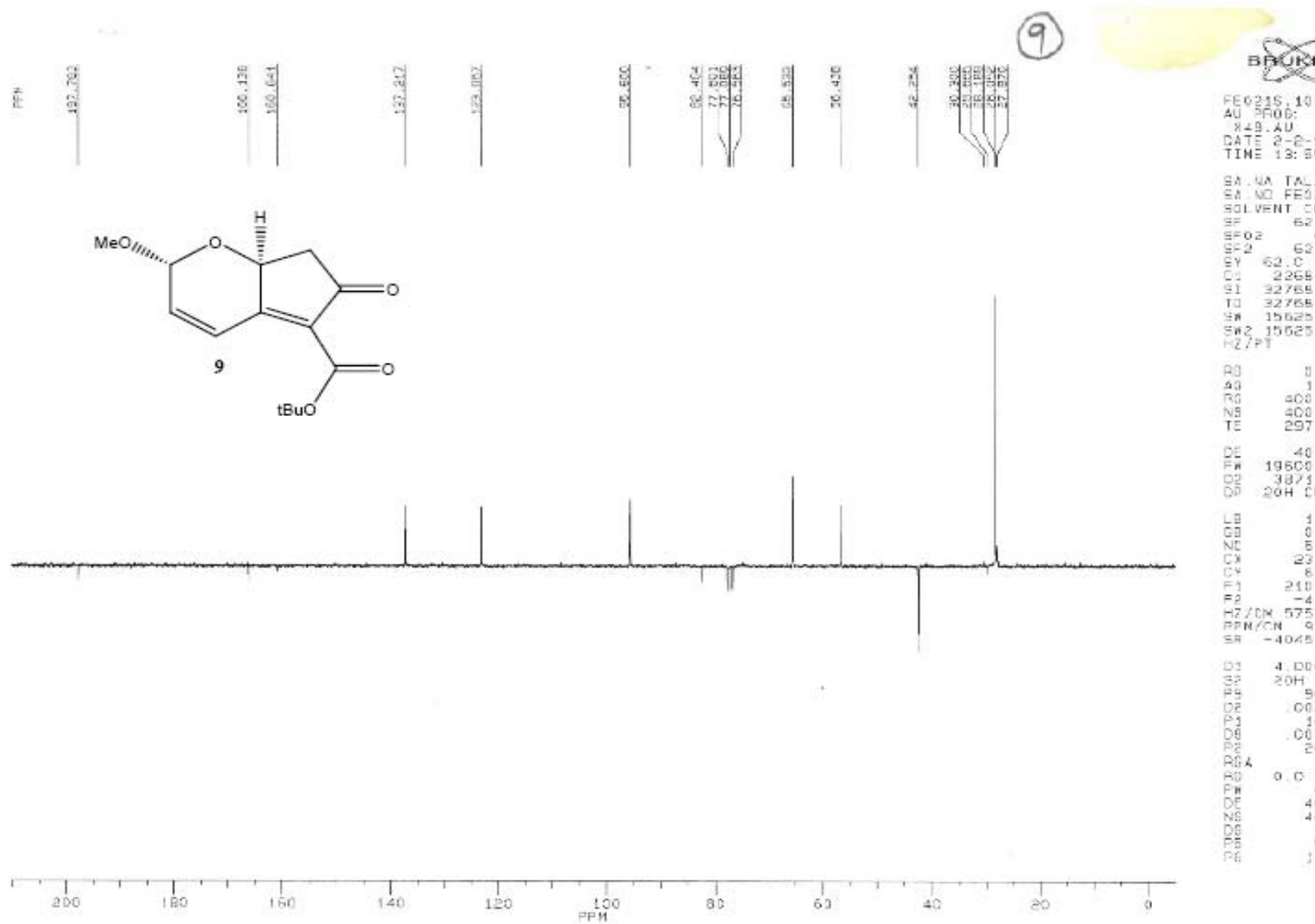
D1
P0
RGA
R0
PW
DC
NS
D3





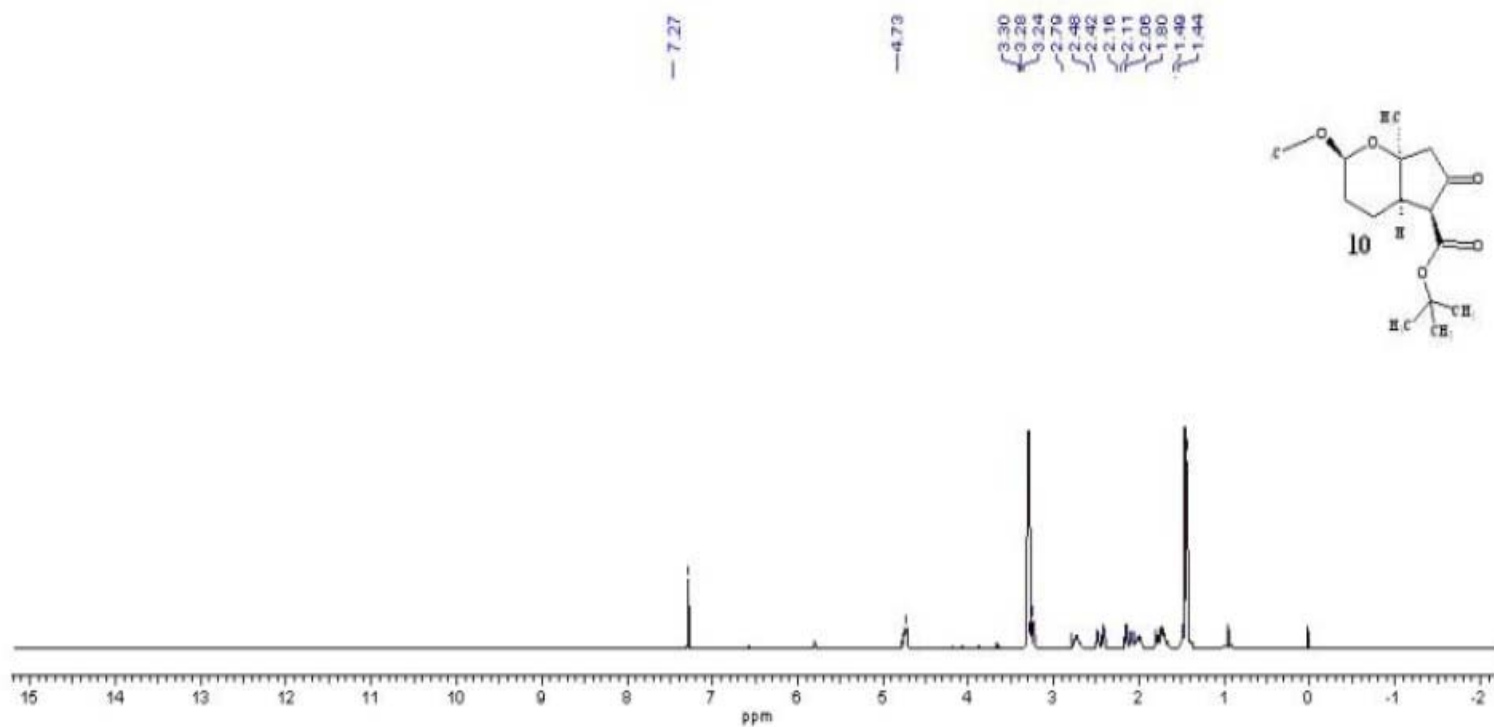




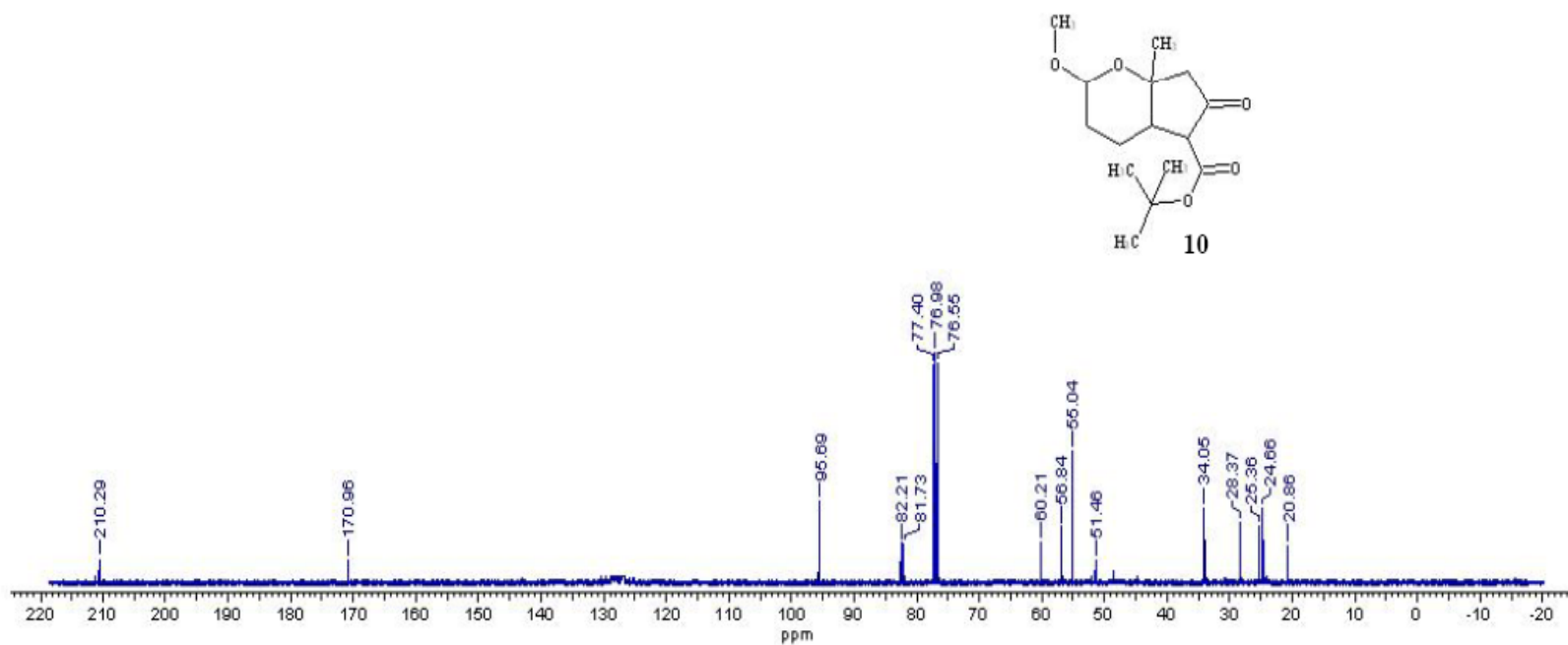


Taleb-2010-10

19 Jan 2010
TA reduction product

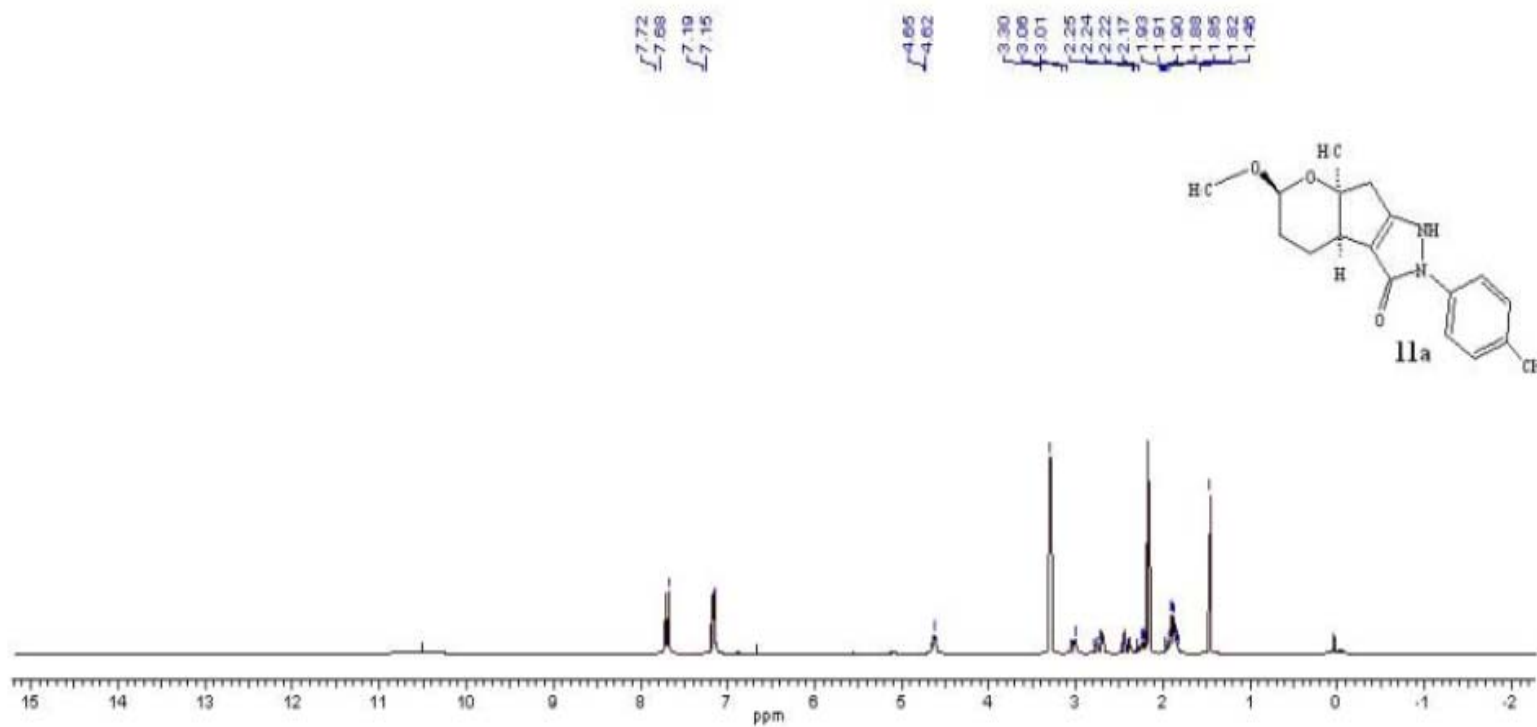


NUCLEUS	13C	NUMBER OF TRANSIENTS	2400	ORIGINAL POINTS COUNT	32708	POINTS COUNT	202144
Pulse Sequence	zgdc30	Solvent	CHLOROFORM-D			Sweep Width (Hz)	18028.85
Temperature (degree C)	27.000						

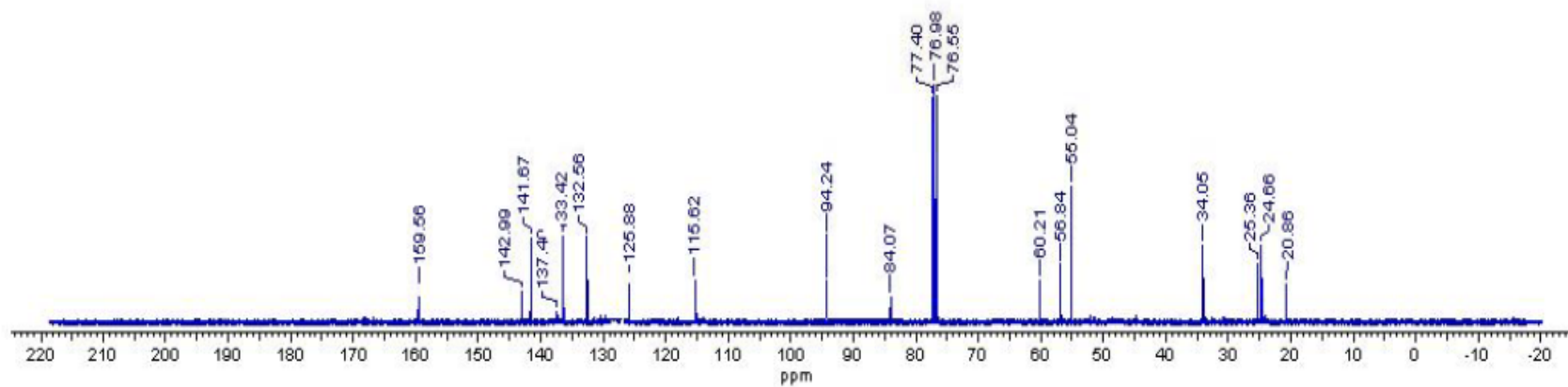
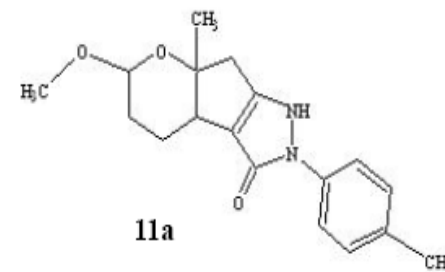


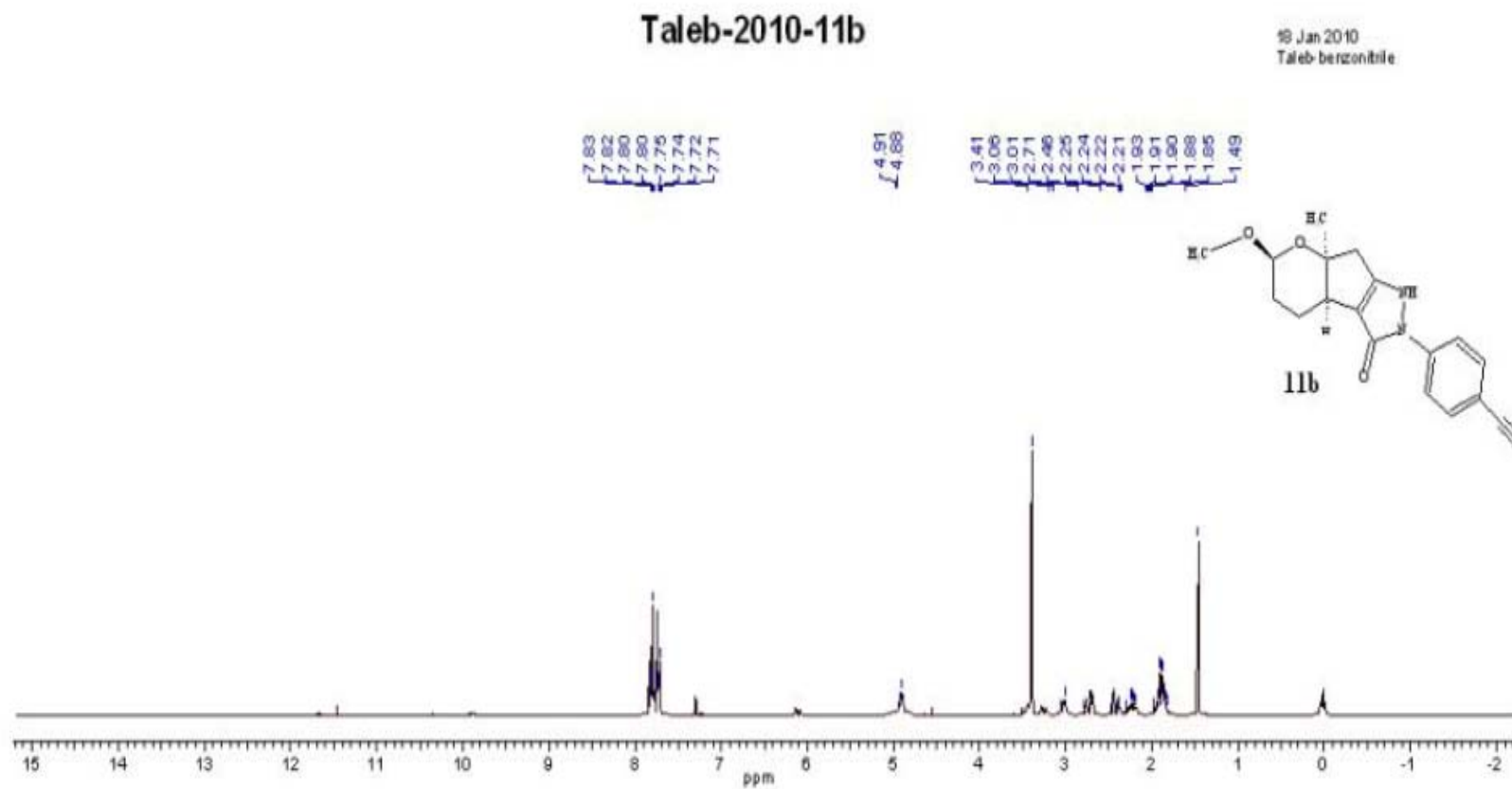
Taleb-2010-11a

18 Jan 2010
TA methylhydrazine 11a



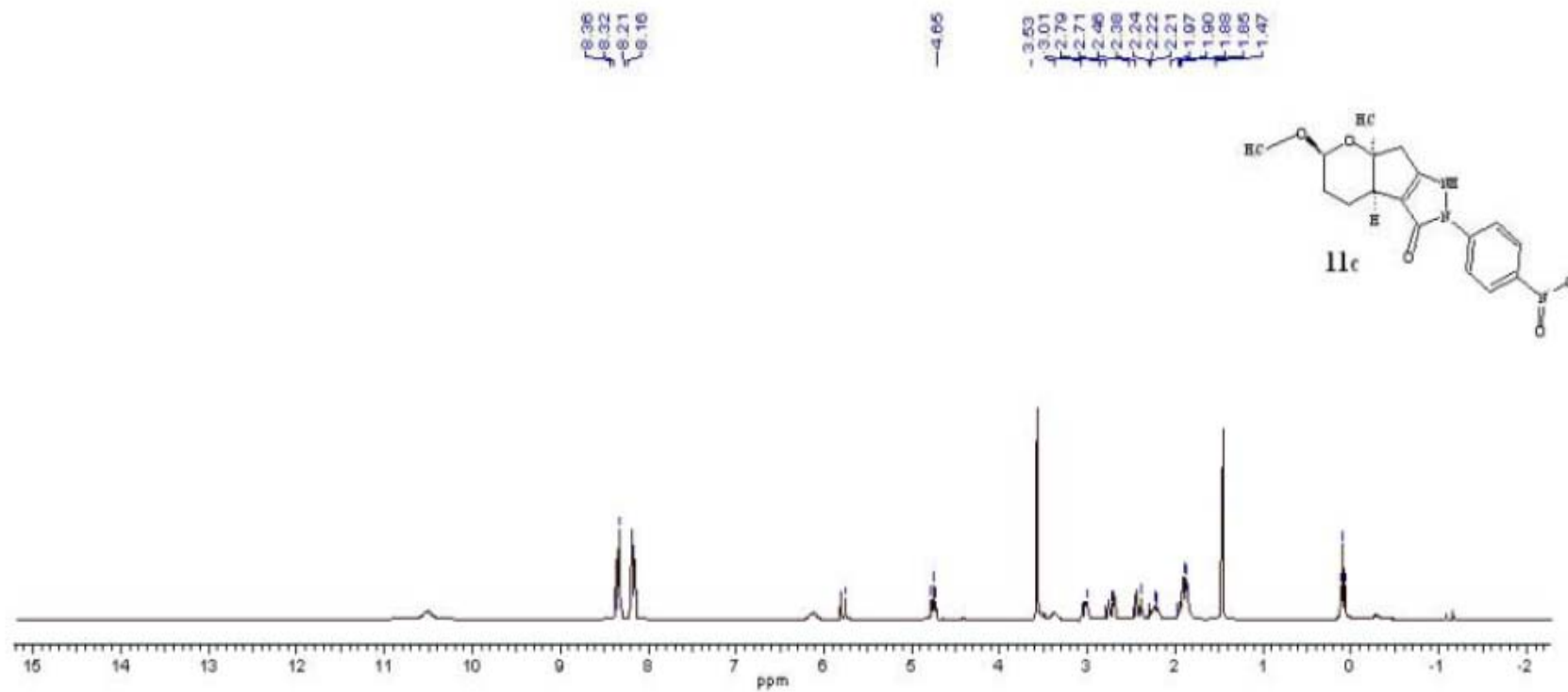
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Nucleus	13C	Number of Transients 2400	Original Points Count 32768	Points Count 262144
Pulse Sequence	zgpg30	Solvent CHLOROFORM-D	Sweep Width (Hz)	18028.85
Temperature (degree C)	27.000			



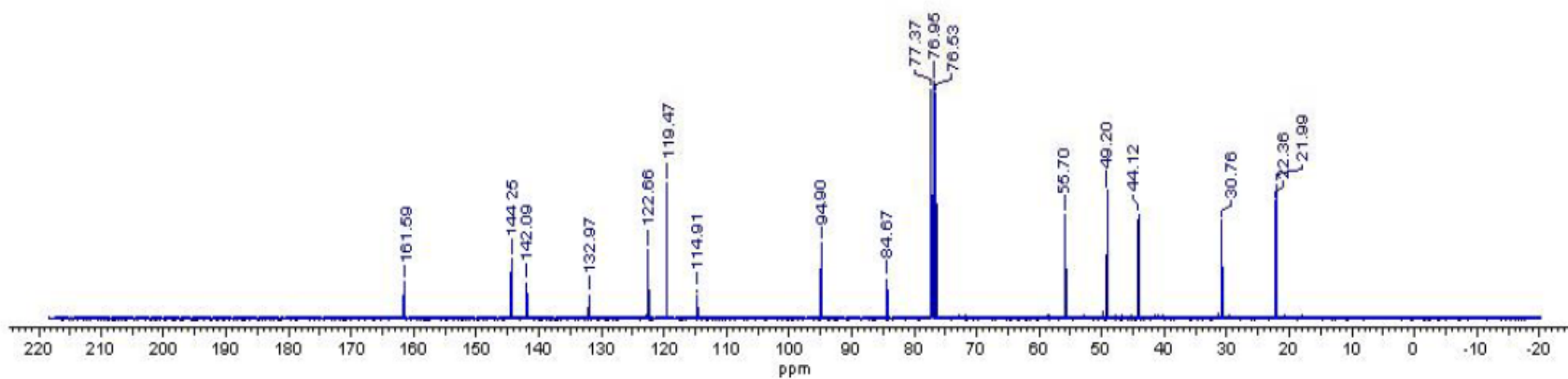
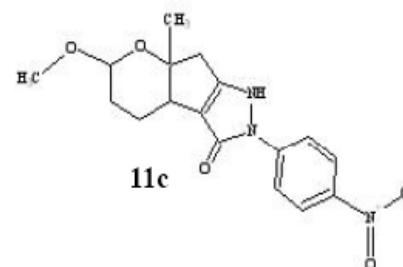


Taleb-2010-11c

19 Jan 2010
Taleb-nitorphenyl



<i>Nucleus</i>	¹³ C	<i>Number of Transients</i>	2400	<i>Original Points Count</i>	32768	<i>Points Count</i>	32768
<i>Pulse Sequence</i>	zgdc30	<i>Solvent</i>	CHLOROFORM-D	<i>Sweep Width (Hz)</i>	18028.85	<i>Temperature (degree C)</i>	27.000



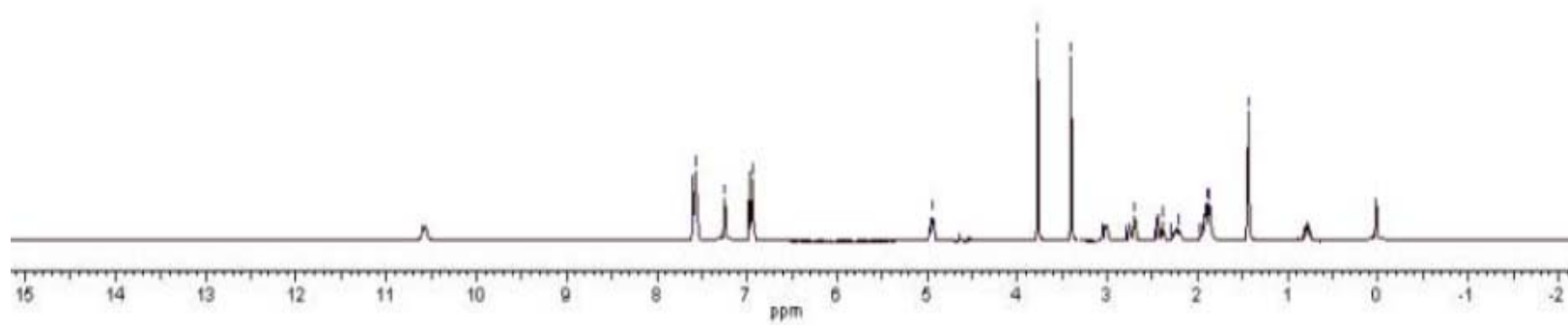
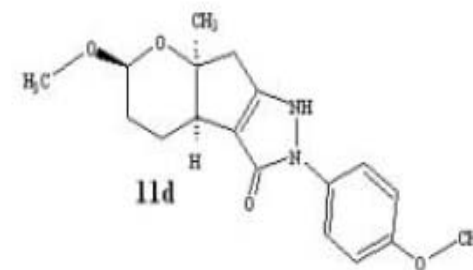
Taleb-2010-11d

19 Jan 2010
Taleb-paramethoxy

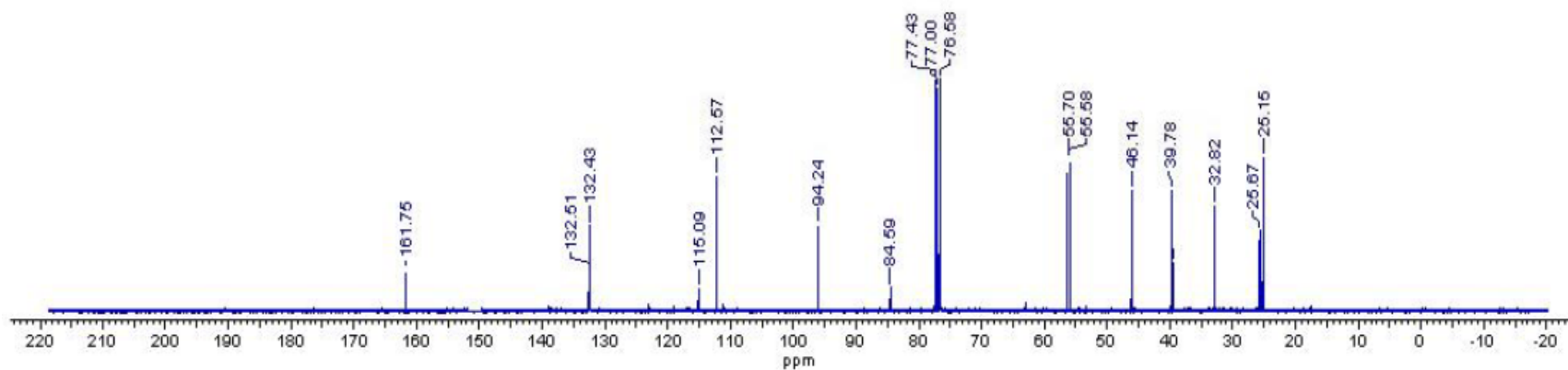
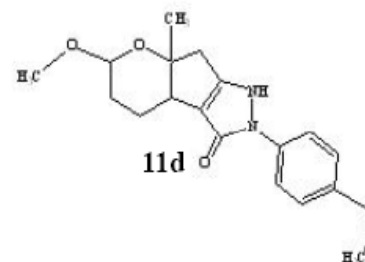
7.61
7.57
7.26
6.97
6.93

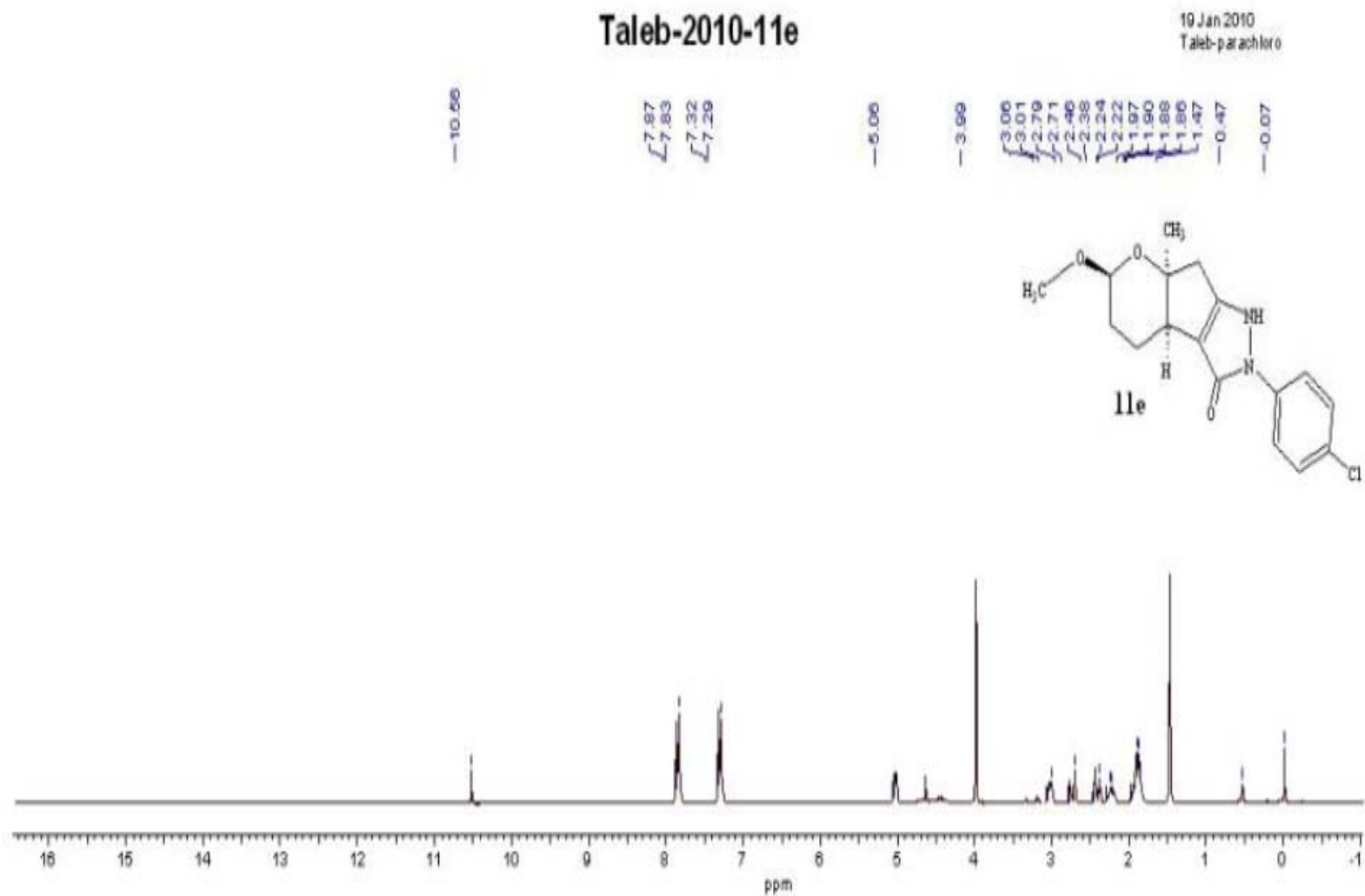
4.93

3.77
3.40
3.06
2.79
2.71
2.46
2.38
2.30
2.22
1.97
1.90
1.88
1.86
1.50

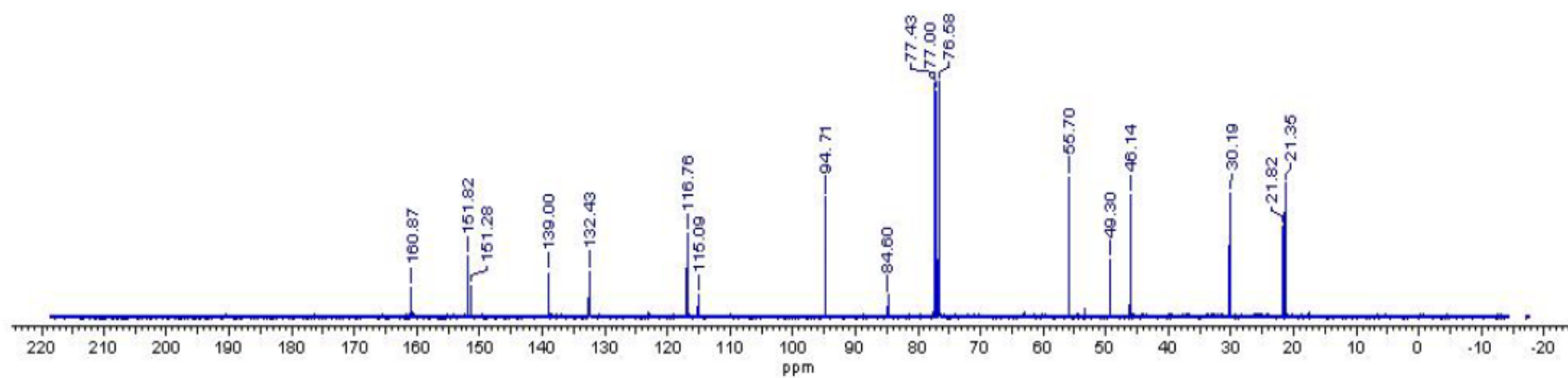
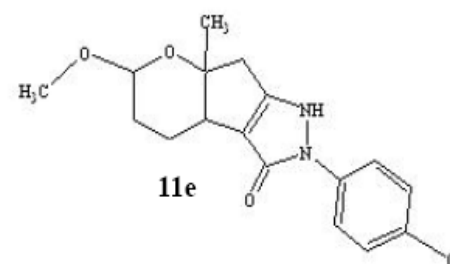


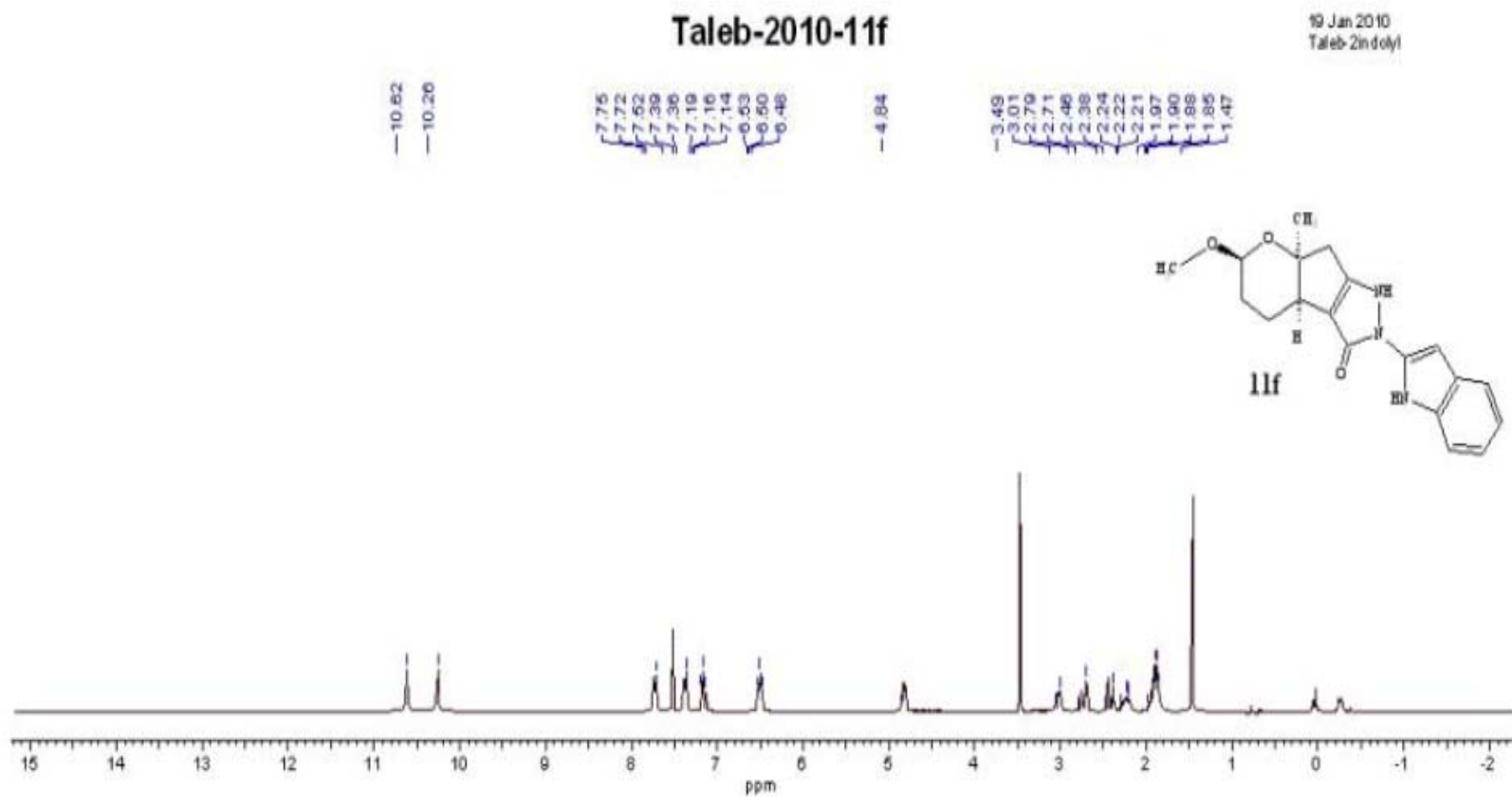
<i>Nucleus</i>	¹³ C	<i>Number of Transients</i>	2400	<i>Original Points Count</i>	32768	<i>Points Count</i>	32768
<i>Pulse Sequence</i>	zgdc30	<i>Solvent</i>	CHLOROFORM-D	<i>Sweep Width (Hz)</i>	18028.85	<i>Temperature (degree C)</i>	27.000





File Name	C:\Documents and Settings\user\desktop\11A-RA-USA-Ampyl-Litranon-5010\11A-Ampyl-Litranon-5010_U1100010			Frequency (MHz)	DS
Nucleus	13C	Number of Transients	2400	Original Points Count	32768
Pulse Sequence	zgdc30	Solvent	CHLOROFORM-D	Sweep Width (Hz)	18028.85
				Temperature (degree C)	27.000





Taleb-2010-11g

19 Jan 2010
Taleb-2-pyridnyl

8.42
8.40
7.84
7.81
7.66
7.63
7.61
7.01
6.88
6.07

5.21
5.19
5.16

3.77
3.09
2.79
2.71
2.49
2.38
2.24
2.22
2.21
1.97
1.80
1.88
1.85
1.42

