

## Supporting Information

### Asymmetric synthesis of chiral amine from cyclic imine by bacterial whole-cell catalyst of enantioselective imine reductase

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#### Table of Contents:

Chemicals-----	S1
Screening of Imine-reducing Strains-----	S1
Derivatization of 2-Methylpyrrolidine (2-MP) with 2,3,4,6-Tetra- <i>O</i> -acetyl- $\beta$ -D-glucopyranosyl Isothiocyanate (GITC)-----	S1
Synthesis of Phenyl Isothiocyanate (PITC)-derivatized 2-MP-----	S1
Cultivation of 2-MPN-reducing <i>Streptomyces</i> Strains -----	S1
Production of <i>R</i> - and <i>S</i> -2-MP with whole-cell catalysts -----	S1
Syntheses of <i>R</i> - and <i>S</i> -2-MP hydrochlorides-----	S1
HPLC Chromatograms-----	S2
NMR and MS Spectra of PITC-derived 2-Methylpyrrolidine-----	S3
NMR spectra of <i>R</i> -2-Methylpyrrolidine hydrochloride-----	S4

## Chemicals

2-Methyl-1-pyrroline (2-MPN) and racemic, *R*- and *S*-2-methylpyrrolidine (2-MP) were purchased from Sigma-Aldrich (Tokyo, Japan). 2,3,4,6-Tetra-*O*-acetyl- $\beta$ -D-glucopyranosyl isothiocyanate (GITC) and phenyl isothiocyanate (PITC) were purchased from Wako Pure Chemical Industries (Osaka, Japan).

## Screening of Imine-reducing Strains

After cultivation of each microorganism in a nutrient medium (4 ml) containing 0.1 % (v/v) 2-methyl-1-pyrroline (2-MPN), the cells harvested by centrifugation were washed thrice with 0.85% (w/v) NaCl. Conversion of 10 mM 2-MPN was performed using whole cells from 2 ml of the culture broth in 100 mM phosphate buffer (pH 7.0) at 25°C for 24 h with reciprocal shaking (115 strokes/min). The formation of 2-MP from 2-MPN was initially checked on TLC (silica gel 60 F254 plate, Merck) using developing solvent (*n*-butanol:acetic acid: water=2:1:1) and ninhydrin reagent. Then both concentration and optical purity of 2-MP formed were determined by HPLC after 2-MP was derivatized with GITC.

The following genera and soil isolates (unidentified) were used for screening: **bacterium**; *Achromobacter*, *Acinetobacter*, *Aeromonas*, *Agrobacterium*, *Alcaligenes*, *Arthrobacter*, *Aureobacterium*, *Bacillus*, *Brevibacterium*, *Brevundimonas*, *Cellulomonas*, *Citrobacter*, *Comamonas*, *Corynebacterium*, *Enterobacter*, *Erwinia*, *Escherichia*, *Flavobacterium*, *Gordona*, *Hafnia*, *Hydrogenophaga*, *Iodobacter*, *Micrococcus*, *Moraxella*, *Pantoea*, *Paracoccus*, *Pimelobacter*, *Pseudomonas*, *Ralstonia*, *Rhizobium*, *Rhodobacter*, *Rhodococcus*, *Rhodospirillum*, *Salmonella*, *Serratia*, *Xanthobacter*, **Actinomyces**, *Actinomadura*, *Nocardia* and *Streptomyces*; **fungus**; *Agaricales*, *Alternaria*, *Amylomyces*, *Amylomyces*, *Armillariella*, *Arthrinium*, *Aspergillus*, *Beauveria*, *Byssoschlamys*, *Calonectria*, *Calcarisporium*, *Caldariomyces*, *Chaetomium*, *Chlamydoabsidia*, *Chloridium*, *Cladobotryum*, *Cladosporium*, *Clonostachys*, *Coelomycetes*, *Colletotrichum*, *Coniothyrium*, *Curvularia*, *Dactylaria*, *Dicranophora*, *Dothiora*, *Emericella*, *Eupenicillium*, *Eurotium*, *Eurotium*, *Exophiala*, *Fusarium*, *Ganoderma*, *Gibberella*, *Gliocladium*, *Grifola*, *Halteromyces*, *Hormonema*, *Hypocrea*, *Mariannaea*, *Memnoniella*, *Micronectriella*, *Monocillium*, *Mortierella*, *Mucor*, *Mucorales*, *Mycotypha*, *Myrothecium*, *Nectria*, *Oidiodendron*, *Paecilomyces*, *Penicillium*, *Periconia*, *Phaeococcomyces*, *Phoma*, *Phycomyces*, *Phytophthora*, *Pilaira*, *Pilobolus*, *Plectosphaerella*, *Pringsheimia*, *Protomycocladus*, *Radiomyces*, *Rhizopus*, *Sesquicillium*, *Stachybotrys*, *Sydowia*, *Syncephalastrum*, *Syncephalis*, *Syzygites*, *Thamnostylum*, *Torula*, *Trichoderma*, *Trichophyton*, *Umbelopsis*, *Verticillium*, *Volutella* and *Zygorhynchus*; **yeast**; *American yeast*, *Awamori yeast*, *Baker's yeast*, *Candida*, *Citeromyces*, *Cryptococcus*, *Debaryomyces*, *Hansenula*, *Hanseniaspora*, *Kloeckera*, *Pichia*, *Rhodotorula*, *Saccharomyces*, *Saccamycodes*, *Satumispora*, *Torulopsis* and *Williopsis*.

## Derivatization of 2-MP with GITC

The derivatization of 2-methylpyrrolidine (2-MP) was carried out at 40°C for 1h using each sample (50  $\mu$ L), 0.8% (w/v) GITC (100  $\mu$ L) and 0.2% (w/v) triethylamine (50  $\mu$ L) in acetonitrile. The derivative was analyzed at 254 nm at a flow rate of 1.0 ml/min using HPLC with a Wakosil-II 5C18 RS (4.6 mm $\times$ 150 mm; Wako) and 10 mM KH<sub>2</sub>PO<sub>4</sub>/H<sub>3</sub>PO<sub>4</sub> (pH 2.5)/methanol (55:45 v/v). *S*- and *R*-2-MP derivatives were detected at 24.9 min and 26.3 min, respectively. 2-MP concentration was determined using the calibration curves of authentic *S*- and *R*-2-MP derivatives.

### Synthesis of PITC-derivatized 2-MP

PITC-derivatized 2-MP was synthesized from 2-MP at 40°C using PITC and triethylamine in acetonitrile for NMR and MS analyses. After extraction of reaction product with ethyl acetate, the crude product was purified by silica gel chromatography with the following solvent (*n*-hexane : ethyl acetate=8:1, 4:1, 2:1 and 1:1). The PITC-derivative (54 mg, 0.245 mmol) was obtained from 0.39 mmol *R*-2-MP (>99%e.e.) formed by whole cells of GF3587 with 63% isolated yield.

PITC-derivatized 2-MP, 1-(2-methylpyrrolidine)thiocarboxanilide :  $\delta_{\text{H}}$  (500 MHz;  $\text{CDCl}_3$ ;  $\text{Me}_4\text{Si}$ ) 1.31 (3 H, d,  $J$  6.3  $\text{CHCH}_3$ ), 1.65-2.13 (4 H, m,  $\text{CHCH}_2\text{CH}_2\text{N}$ ), 3.53 (1 H, br s,  $\text{CH}_a\text{H}_b\text{N}$ ), 3.63 (1 H, br s,  $\text{CH}_a\text{H}_b\text{N}$ ), 4.51 (1 H, br s,  $\text{NCHCH}_3$ ), 7.06 (1 H, s,  $\text{NHPh}$ ), 7.15-7.35 (5 H, m, Ph);  $\delta_{\text{C}}$  (125 MHz;  $\text{CDCl}_3$ ;  $\text{Me}_4\text{Si}$ ) 18.8, 23.2, 32.2, 49.6, 56.8, 125.4, 125.6, 128.3, 139.3 and 177.8;  $m/z$  (EI) 220 ( $\text{M}^+$ ; 100%), 187 (65) and 128 (89). All spectral data were coincident with those of PITC- derived authentic racemic 2-methylpyrrolidine.

### Cultivation of 2-MPN-reducing *Streptomyces* Strains

*Streptomyces* sp. GF3587 was cultivated at 28°C for 24 h with reciprocal shaking (115 strokes/min) in a 500-ml shaking flask containing 40 ml of the nutrient medium containing 8 g/L glucose, 20 g/L malt extract (Difco, USA), 4 g/L NZ amine (Wako Pure Chemical Industries, Osaka, Japan) and 5 mg/L  $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$  at pH 7.3. Dry cell weight was determined to be 0.51 mg per ml of culture broth per OD (optical density) of 1.0 at 610 nm.

*Streptomyces* sp. GF3546 was cultivated at 28°C for 72 h with reciprocal shaking (115 strokes/min) in a 500-ml shaking flask containing 40 ml of the nutrient medium containing 20 g/L glucose, 30 g/L meat extract (Kyokuto Pharmaceutical Industrial, Takahagi, Ibaraki, Japan), 20 g/L yeast extract (Oriental Yeast, Osaka, Japan), 10 g/L NZ amine and trace elements solution supplemented with 1 mg/L  $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ , 1 mg/L  $\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$  and 1 mg/L  $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$  at pH 7.3. Dry cell weight was determined to be 9.1 mg per ml of culture broth.

### Production of *R*- and *S*-2-MP with whole-cell catalysts

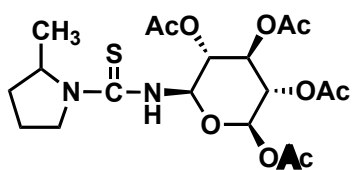
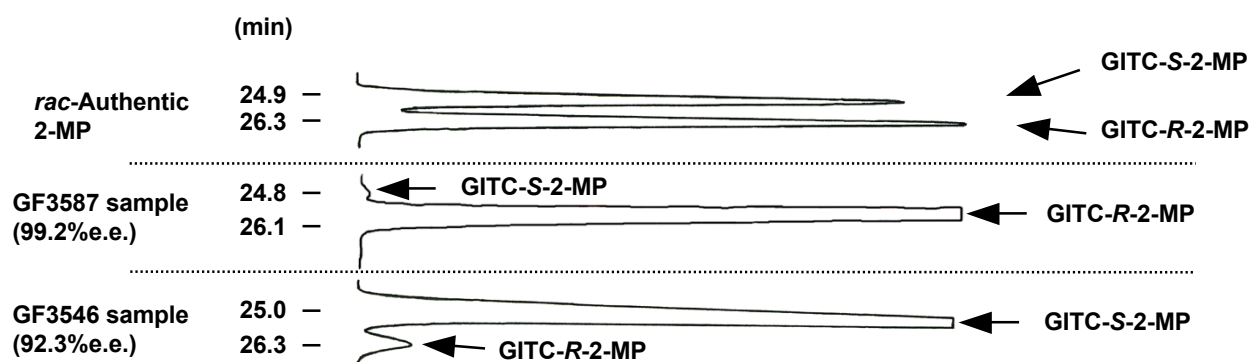
After cultivation, the cells of GF3587 were harvested by centrifugation at 12,000 *g* for 20 min, GF3546 cells were collected by filtration. Each of the cells suspended with an appropriate volume of 0.85% NaCl was used as a whole-cell catalyst (GF3587; 455 mg dry cell weight, GF3546; 83 mg dry cell weight). The reaction was carried out at 25°C in the presence of glucose (2% or 4% w/v) with monitoring 2-MP formation by HPLC.

### Syntheses of *R*- and *S*-2-MP hydrochlorides

*R*- and *S*-2-methylpyrrolidine hydrochloride were synthesized from reaction solution via both protection of amine and deprotection of *N*-Boc group.

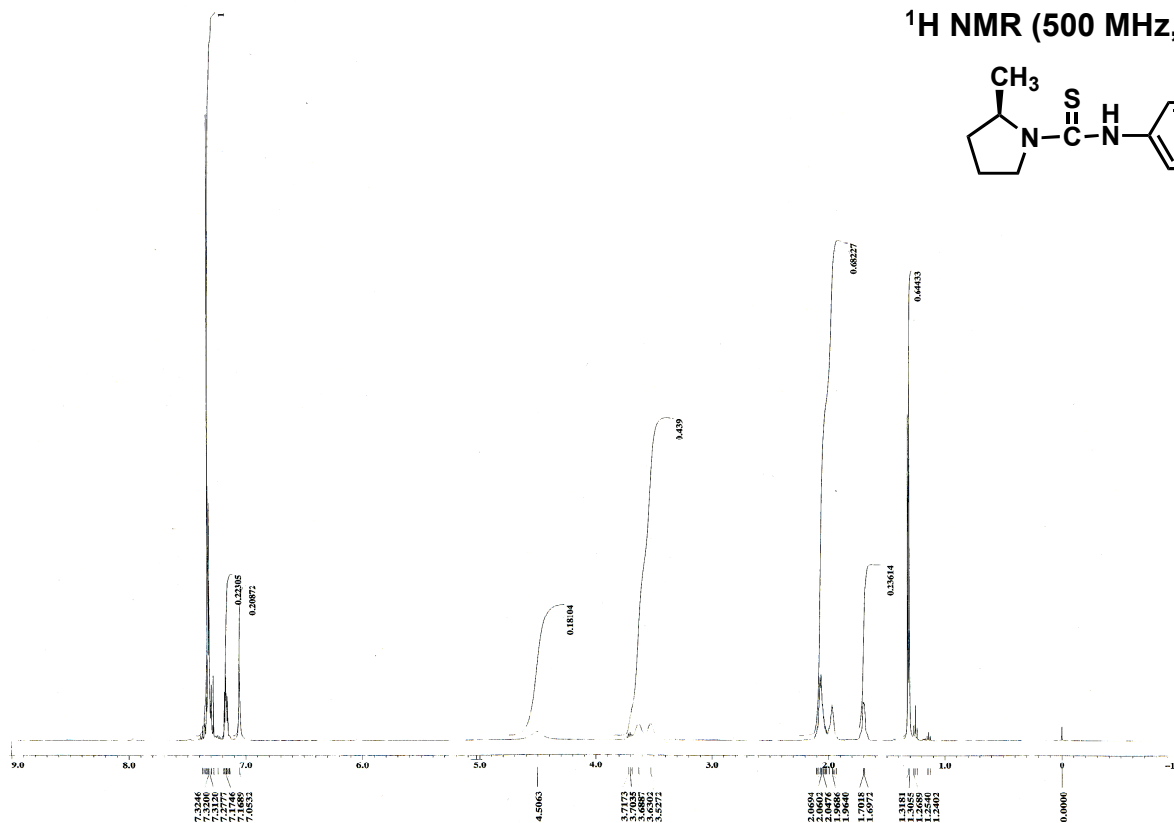
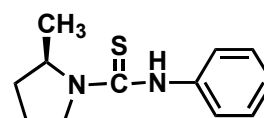
*R*-2-methylpyrrolidine hydrochloride:  $\delta_{\text{H}}$  (600 MHz;  $\text{D}_2\text{O}$ ; DDS) 1.39 (3 H, d,  $J$  6.2), 1.62-1.69 (1 H, m), 1.96-2.13 (2H, m), 2.17-2.26 (1H, m), 3.26-3.38 (2H, m), 3.66-3.74 (1H, m);  $\delta_{\text{C}}$  (150 MHz;  $\text{D}_2\text{O}$ ; DDS) 16.8, 23.2, 31.3, 45.1, 56.6. The spectra of *S*-2-methylpyrrolidine hydrochloride were coincident with those of *R*-2-MP hydrochloride: All spectral data were coincident with those of authentic racemic 2-methylpyrrolidine hydrochloride.

### HPLC Chromatograms of GITC-derivatized 2-Methylpyrrolidine

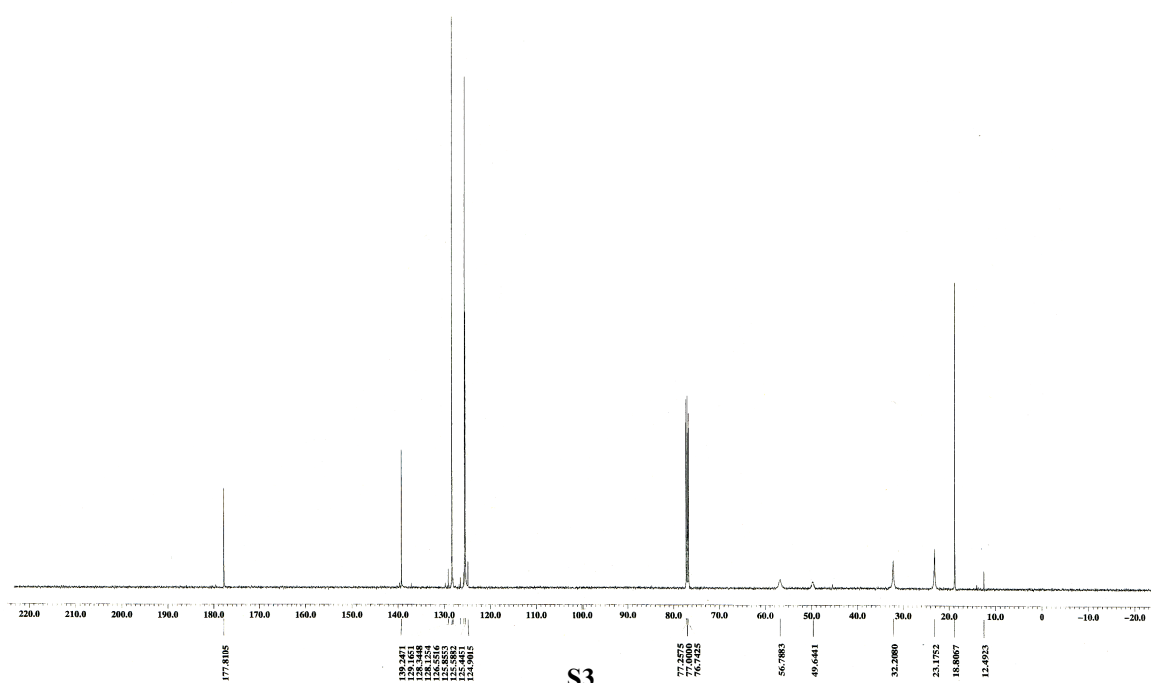
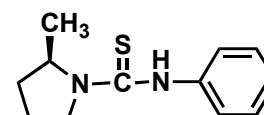


2,3,4,6-tetra-O-acetyl- $\beta$ -D-glucopyranosyl isothiocyanate-2-methylpyrrolidine  
(GITC-2-MP)

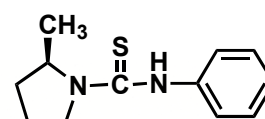
**<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)**



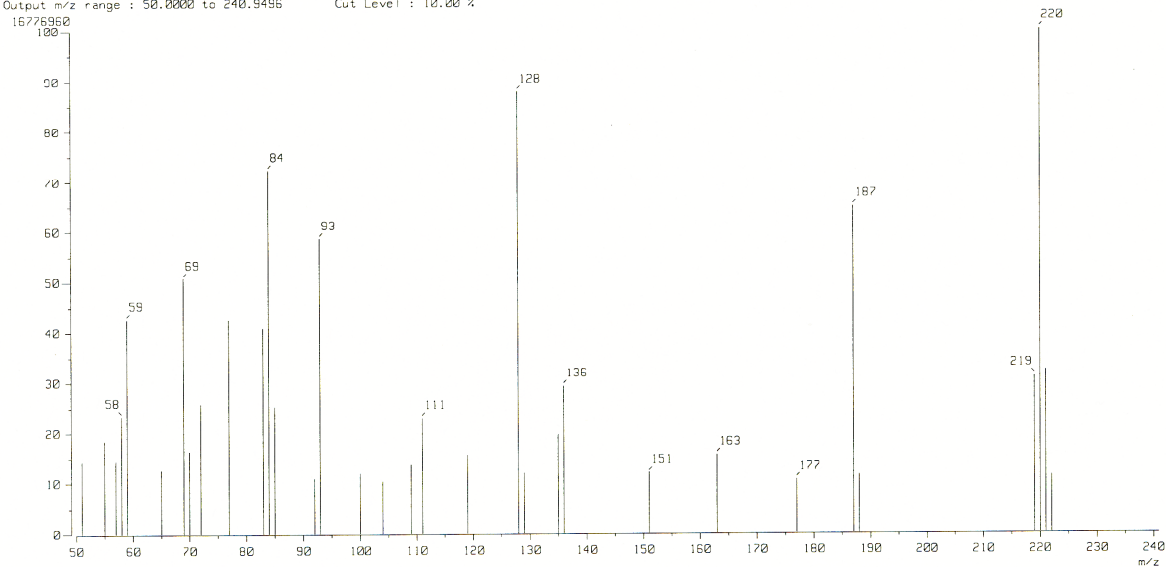
**<sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)**



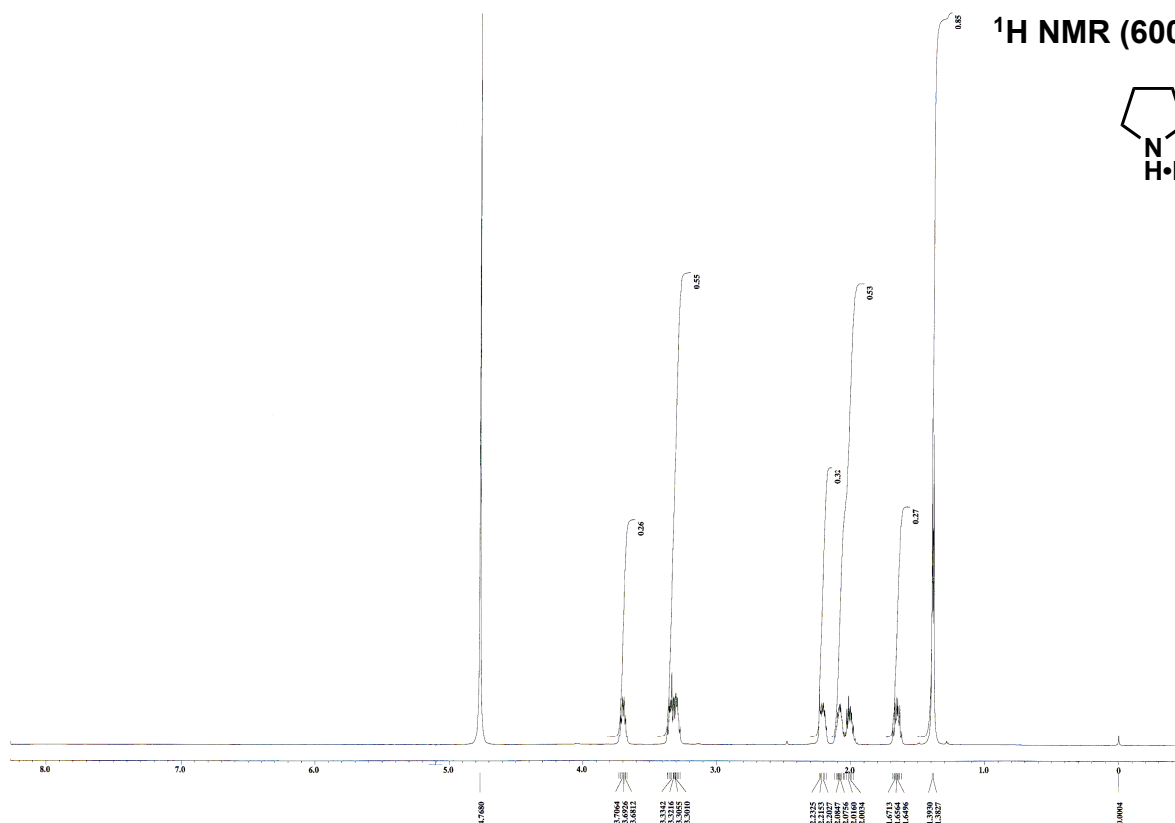
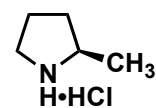
# EI-MS (70 eV)



[ Mass Spectrum ]  
Data : ms01009 Date : 15-Jan-2007 15:20  
Sample : -  
Note : -  
Inlet : Direct Ion Mode : EI+  
Spectrum Type : Normal Ion [MF-Linear]  
RT : 0.00 min Scan# : 1  
BP : m/z 18.0000 Int. : 1559.58  
Output m/z range : 50.0000 to 240.9496 Cut Level : 10.00 %



**<sup>1</sup>H NMR (600 MHz, D<sub>2</sub>O)**



**<sup>13</sup>C NMR (150 MHz, D<sub>2</sub>O)**

