## **Electronic Supplementary Information**

Mutanobactin A from the human oral pathogen *Streptococcus mutans* is a cross-kingdom regulator of the veast-mycelium transition

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General Experimental Procedures. NMR data were obtained on a Varian VNMR spectrometer (500 MHz for <sup>1</sup>H, 125 MHz for <sup>13</sup>C) with a triple resonance probe at 22  $\pm$  0.5 °C. Electrospray-ionization mass spectrometry data were collected on an IonSpec (Varian, Inc.) 9.4 T FT-ICR instrument. ESI MS/MS analyses were performed on a LCT premier (Waters Corp.) time-of-flight instrument. Flash chromatography was performed on a Biotage Isolera One using a 100 g C<sub>18</sub> column with a flow rate of 50 mL/min. HPLC separations were carried out on a Shimadzu system using a SCL-10A VP controller and Gemini 5µm C<sub>18</sub> column (110Å, 250 x 21.2 mm) with a flow rate of 10 mL/min. All solvents were of ACS grade or better. Optical rotation measurement were performed on a Rudolph Research Autopol III automatic polarimeter;  $[\alpha]_D$  values are given in deg·cm<sup>2</sup>·g<sup>-1</sup>.

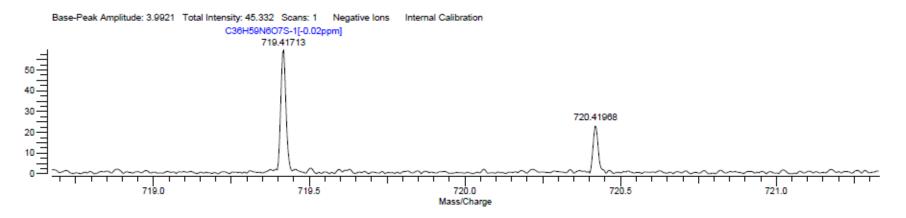
**Preparation and extraction of bacterial culture.** A culture of *Streptococcus mutans* UA159 was prepared by inoculating 15 L of brain-heart infusion (BHI) broth with 100 mL of a stationary phase *S. mutans* UA159 culture. The culture was incubated under microaerobic conditions at 37 °C for 36 h. The culture was extracted three times with equal volumes of ethyl acetate, which was then evaporated *in vacuo* to generate the *S. mutans* UA159 extract.

**Mutanobactin A** (1): white solid;  $[\alpha]_D^{25} = -8.0$  (*c* 0.001 in MeOH);  $\lambda_{max}$  (MeOH) 221 nm ( $\varepsilon$ /dm<sup>3</sup> mol<sup>-1</sup> cm<sup>-1</sup> 38 900); <sup>1</sup>H, <sup>13</sup>C, and <sup>15</sup>N NMR data, see Table S1; HRESIMS (FT-ICR) *m*/*z* [M-H]<sup>-</sup> 719.41713 (calcd. for C<sub>36</sub>H<sub>59</sub>N<sub>6</sub>O<sub>7</sub>S, 719.41714).

position	$\delta_{\rm C}$ , mult. <sup><i>a</i></sup>	$\delta_{ m H}$ (J in Hz)	${\delta_{\mathrm{N}}}^b$
1	50.4, CH	4.43, ddd (3.7, 9.0, 11.0)	
2a	$40.4, CH_2$	1.44, m	
2b	, 2	1.81, ddd (3.9, 10.5, 13.8)	
3	24.2, CH	1.59, m	
4	20.9, CH <sub>3</sub>	0.82, d (6.6)	
5	$23.5, CH_3$	0.92, d (6.7)	
6	170.5, C	0.92, <b>a</b> (0.7)	
7	48.0, CH	4.52, quintet (6.8)	
8	17.7, CH <sub>3</sub>	1.17, d (6.7)	
9	169.7, C	1.17, d (0.7)	
10	61.0, CH	4.12, dd (3.7, 8.9)	
10 11a	29.6, CH <sub>2</sub>	4.12, dd (3.7, 8.9) 1.72, m	
11a 11b	$29.0, CH_2$	2.13, m	
110	24.5 CH		
	24.5, $CH_2$	1.90, m	
13a	$46.8, CH_2$	3.43, m	
13b	171 6 C	3.65, ddd (4.5, 7.5, 9.8)	
14	171.6, C	257 44(92100)	
15	58.8, CH	3.57, dd (8.3, 10.0)	
16	26.2, CH	2.33, m	
17	20.4, CH <sub>3</sub>	0.84, d (6.6)	
18	18.8, CH <sub>3</sub>	0.77, d (6.8)	
19	168.8, C		
20	52.2, CH	4.87 ddd (2.6, 8.0, 9.0)	
21a	$28.5, CH_2$	2.23, dd (2.6, 16.0)	
21b 22	170 4 C	3.19, dd (9.0, 16.0)	
	170.4, C	2.70 m	
23a	43.7, CH <sub>2</sub>	2.79, m	
23b	41.0. CU	3.28, m	
24	41.0, CH	3.25, m	
25 26	61.7, CH	3.87, d (9.8)	
26 27	167.7, C		
27	203.8, C	2.22	
28a	41.4, CH <sub>2</sub>	2.33, m	
28b	22 1 CH	2.44, dd (6.0, 16.6)	
29 20	23.1, CH <sub>2</sub>	1.44, m	
30	28.7, CH <sub>2</sub>	1.20, m	
31	22.1, CH <sub>2</sub>	1.25, m	
32	28.8, $CH_2$	1.23, m	
33	22.1, CH <sub>2</sub>	1.25, m	
34	31.3, CH <sub>2</sub>	1.23, m	
35	28.9, CH <sub>2</sub>	1.27, m	
36	14.0, CH <sub>3</sub>	0.85, t (6.8)	100.0
C1-NH		8.59, d (9.0)	120.0
C7-NH		7.77, d (6.5)	112.0
C10-NH			nd <sup>c</sup>
C15-NH		8.05, d (8.5)	107.5
C20-NH		7.23, d (8.0)	116.0
C23-NH		7.90, dd (5.3, 9.0)	105.5

	NMR Spectroscopic Data (DMSO- <i>d</i> <sub>6</sub> , 500
MHz for	<sup>1</sup> H, 125 MHZ for $^{13}$ C) for mutanobactin A (1)

<sup>*a*</sup>Determined by HSQC experiment at 500 MHz. <sup>*b*</sup>Determined by HMBC experiment at 500 MHz. <sup>*c*</sup> nd: not detected



**Fig. S1** HRESIMS (FT-ICR) data for mutanobactin A (1)

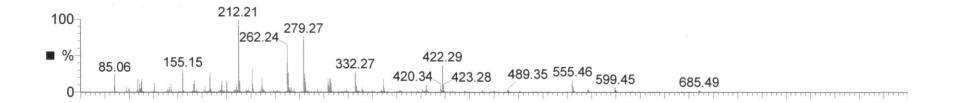
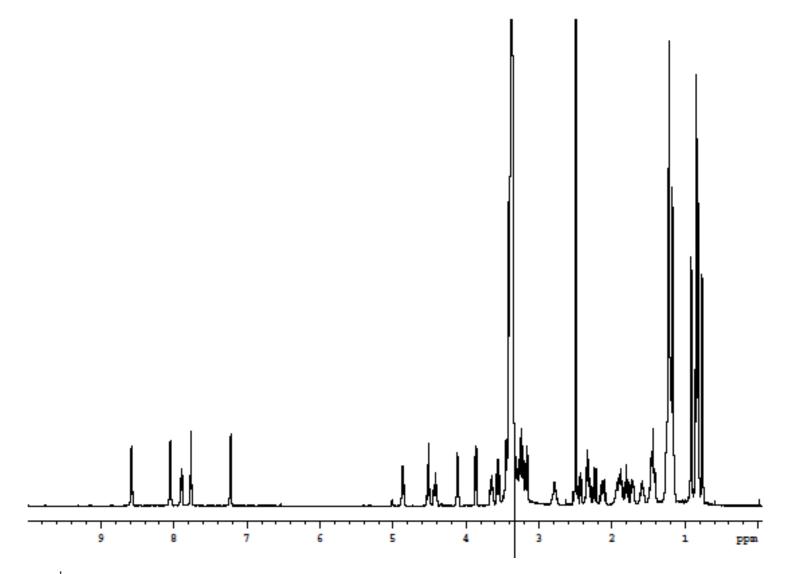
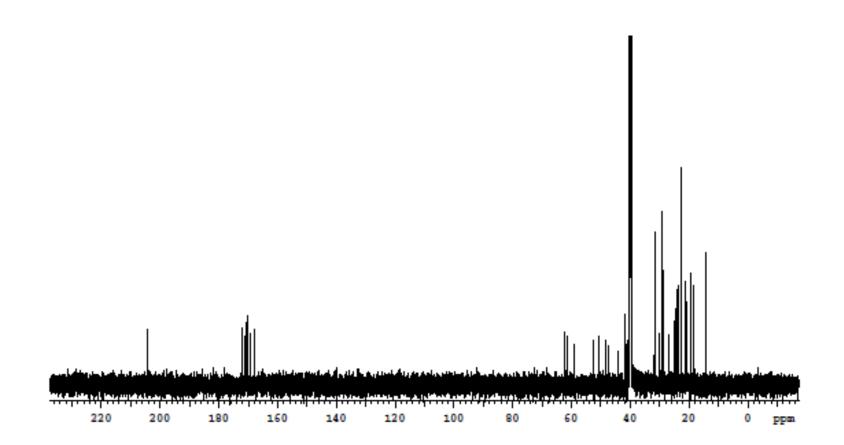


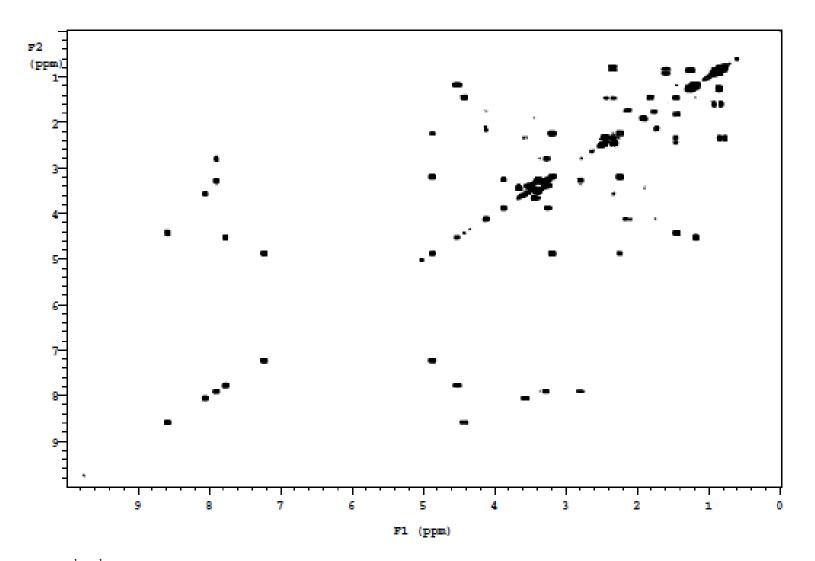
Fig. S2 ESI MS/MS data for mutanobactin A (1)



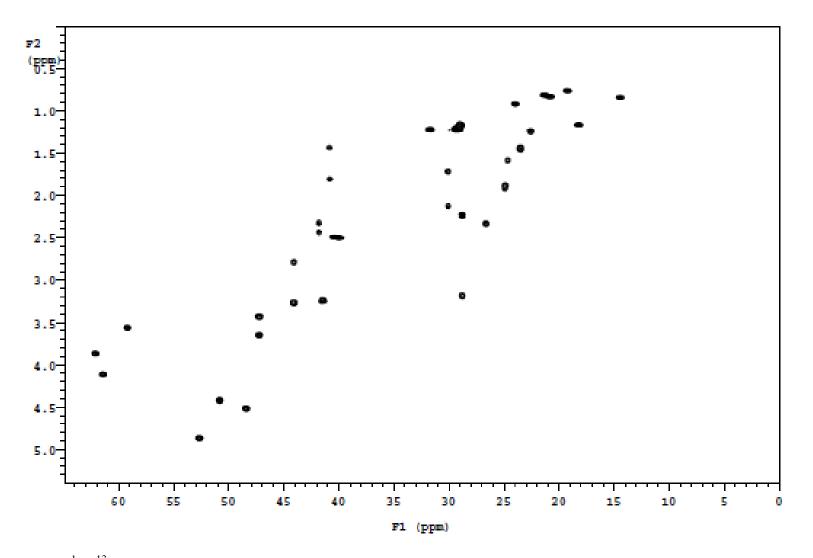
**Fig. S3** <sup>1</sup>H NMR spectrum for mutanobactin A (1)



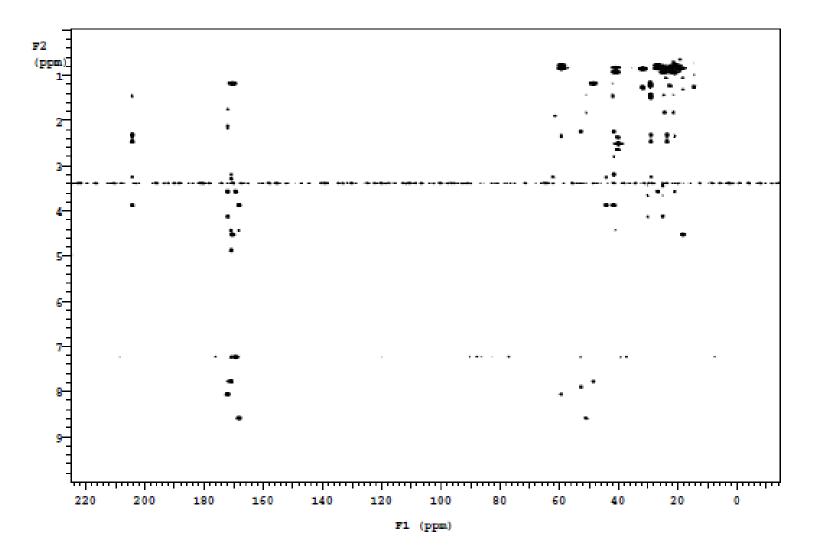
**Fig. S4**  $^{13}$ C NMR spectrum for mutanobactin A (1)



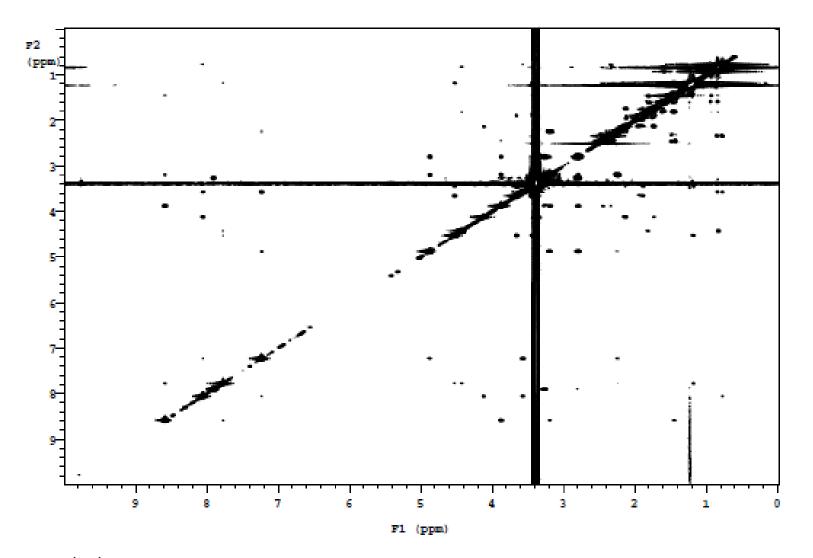
**Fig. S5**  $^{1}$ H $^{-1}$ H COSY NMR spectrum for mutanobactin A (1)



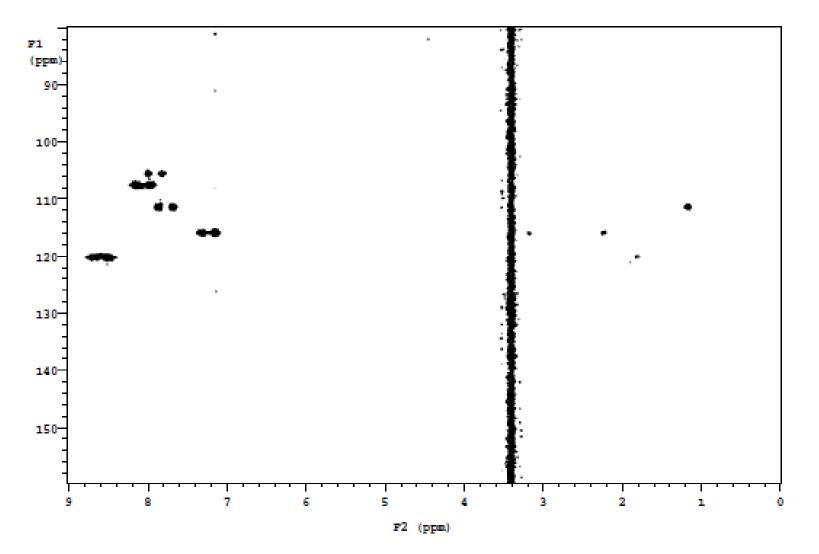
**Fig. S6**  $^{1}$ H $^{13}$ C HSQC NMR spectrum for mutanobactin A (1)



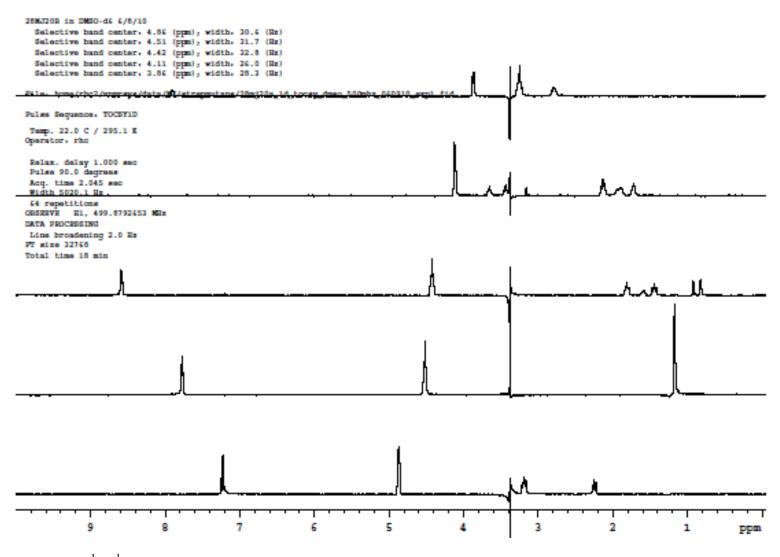
**Fig. S7**  $^{1}$ H $^{-13}$ C HMBC NMR spectrum for mutanobactin A (1)



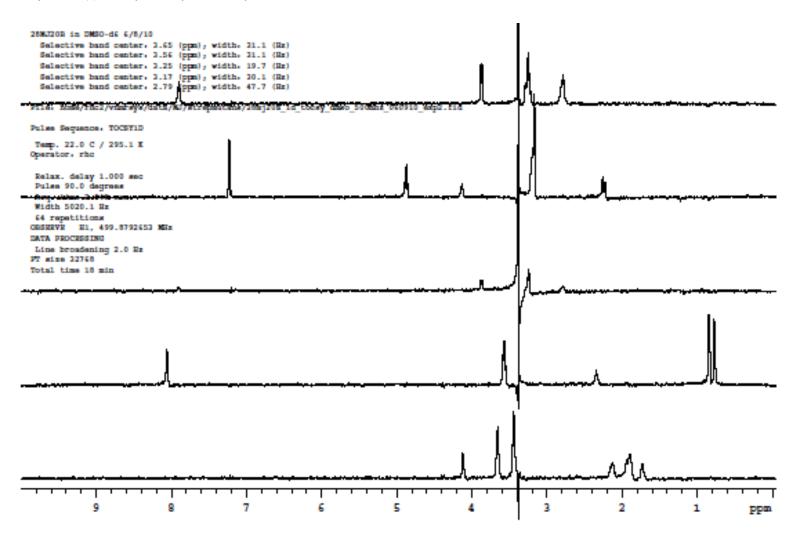
**Fig. S8** <sup>1</sup>H–<sup>1</sup>H NOESY NMR spectrum for mutanobactin A (1)



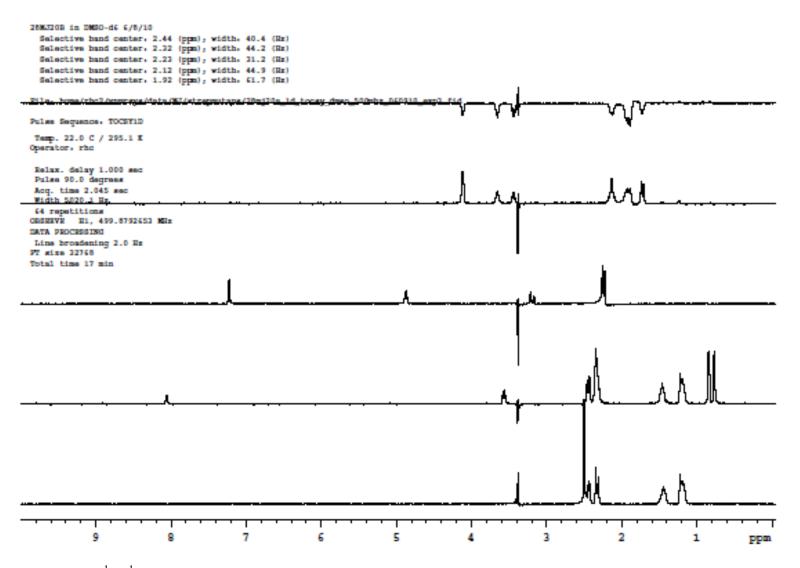
**Fig. S9**  $^{1}$ H $^{15}$ N HMBC NMR spectrum for mutanobactin A (1)



**Fig. S10** 1D  ${}^{1}H{}^{-1}H$  TOCSY NMR spectra for mutanobactin A (1) (panel 1 of 3)



**Fig. S10** 1D  $^{1}$ H $^{-1}$ H TOCSY NMR spectra for mutanobactin A (1) (panel 2 of 3)



**Fig. S10** 1D  ${}^{1}H-{}^{1}H$  TOCSY NMR spectra for mutanobactin A (1) (panel 3 of 3)