Electronic Supplementary Information

Mutated variant of *Candida antarctica* lipase B in (S)-selective dynamic kinetic resolution of secondary alcohols

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Experimental section

Unless otherwise mentioned, all chemicals, were purchased from commercial sources. All reactions were performed in dry glassware under an argon atmosphere. ¹H and ¹³C NMR spectra were recorded at 400 MHz and 100 MHz respectively using a Bruker Avance II spectrometer. Enantiomeric excess was determined by chiral gas chromatography using a CP-Chirasil-DEX CB column (25 m \emptyset * 0.32 mm).

Isopropenyl acetate was dried over $CaCl_2$, distilled and stored over molecular sieves. 1-Phenylheptanol was prepared by NaBH₄-reduction of the corresponding ketone. Ruthenium catalyst **4** was prepared according to a literature procedure.¹ CalB W104A was prepared as previously described, and the enzyme load on the resin was 0.26 %.² The enzyme load of the commercially available CalB (Novozym 435) is approximately 3 %.

Kinetic resolution of 1-phenylpropanol

1a (30 mg, 0.25 mmol) and immobilized CalB W104A (5 mg) were dissolved in toluene (0.5 mL). Once heated to the desired temperature, isopropenyl acetate (83 μ L, 0.75 mmol) was added.

Kinetic resolution of 1-phenylheptanol

1a (96 mg, 0.5 mmol) and immobilized CalB W104A (20 mg) were dissolved in toluene (1 mL). Once heated to the desired temperature, isopropenyl acetate (83 μ L, 0.75 mmol) was added.

General procedure for dynamic kinetic resolution of 1a and 1b

Ru-catalyst 4 (16 mg, 0.025 mmol), CalB W104A (30 mg) and Na₂CO₃ (53 mg, 0.5 mmol) were dissolved in dry toluene (0.5 mL) in a Schlenk-tube. ¹BuOK (0.025 mmol, 100 µL of a 0.5 M solution in dry THF) was added. After 6 minutes of stirring, alcohol 1 (0.5 mmol in 0.5 mL toluene) was added, and after additional 4 minutes isopropenyl acetate (83 µL, 0.75 mmol) was added. When the reaction was finished, the reaction mixture was filtered through a plug of silica and concentrated. Purification by flash column chromatography (Pentane/EtOAc 4:1) yielded the pure acetate 2.

References

¹ B. Martín-Matute, M. Edin, K. Bogár, F. B. Kaynak and J.-E. Bäckvall, J. Am. Chem. Soc., 2005, **127**, 8817-8825.
² M. Vallin, P.-O. Syrén and K. Hult, *ChemBioChem*, 2010, **11**, 411-416.