Exploring *Leishmania major* Inositol Phosphorylceramide Synthase (*Lmj*IPCS): Insights into the ceramide binding domain

ELECTRONIC SUPPLEMENTARY INFORMATION

TABLE OF CONTENTS

Electronic Supplementary Information	1
Table of Contents	1
Chemistry Experimental	2
General Considerations	2
Compound Numbering System	2
Spectroscopic Data	2
Mass Spectrometry	2
Experimental Procedures	3
Synthesis of Intermediate compounds	3
General Procedure for Phase Transfer Catalysis ^{9, 13}	12
Olefin Cross Metathesis	15
General procedure for BOC de-protection - Acylation Reactions	19
Biological Methods	32
Preparation of the Screening Compounds	32
The Assay Protocol of the Inhibition Assays	32
Mass Spectrometry Analyses	32
Cytotoxicity screening	49
References	50

CHEMISTRY EXPERIMENTAL

General Considerations

Compound Numbering System

Numbered compounds in the manuscript have the same number in the ESI. All other compounds are numbered following the pattern S##.

Spectroscopic Data

Infrared spectra were recorded using a golden gate (ATR) on a Perkin-Elmer FT-IR 1600 spectrometer and reported in the following format v_{max} frequency (s, strong; br, broad and w, weak) cm⁻¹. ¹H and ¹³C NMR spectra were acquired in CDCl₃ or CD₃OD unless otherwise stated on a Varian VXR-400 (¹H at 400 MHz, ¹³C at 101 MHz) and reported as follows:

Chemical shift was reported in the following format; δ (ppm) (number of protons, multiplicity, coupling constant J (Hz), assignment).

The residual protic solvent was used as internal reference: CHCl₃ $\delta_{\rm H}$ = 7.26 ppm; CDCl₃ $\delta_{\rm C}$ = 77.16 ppm CHD₂OD $\delta_{\rm H}$ = 3.31 ppm, 1.09 ppm; CD₃OD $\delta_{\rm C}$ = 49.0 ppm

Assignment of stereochemistry was carried out using COSY, HSQC, HMBC and NOESY experiments.

Mass Spectrometry

Low Resolution Mass Spectra were obtained on Waters Micromass LCT Mass spectrometer. Gas-Chromatography Mass Spectra (GC-MS) were taken using a Thermo-Finnigan Trace. High-resolution mass spectra (HRMS) were performed on a Thermo-Finnigan LTQ FT Mass Spectrometer by Durham University Mass Spectroscopy service

Experimental Procedures

Synthesis of Intermediate compounds



S01 (S)-tert-Butyl (1-(methoxy(methyl)amino)-1-oxopropan-2-yl)carbamate¹

Procedure:² To a solution of *N*-(*tert*-butoxycarbonyl)-L-alanine (1.513 g, 8.0 mmol) in anhydrous CH₂Cl₂ (50 ml) was added *N*-methylmorpholine (0.967 ml, 8.8 mmol, 1.1 eq.) and *N*,O-dimethylhydroxylamine hydrochloride (0.859 g, 8.8 mmol, 1.1 eq.) at -15 °C. To the reaction mixture was added 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (1.53 g, 8.8 mmol, 1.1 eq.) over 30 min at the same temperature. The reaction mixture was stirred for 4 hrs then poured into ice and 1N HCI. The resulting mixture was extracted with CH₂Cl₂. The organic layers were combined, and washed with brine, dried over MgSO₄, filtered, and concentrated *in vacuo* to give the crude Weinreb amide as a white solid. Column chromatography on silica gel (from 25 % to 50 % EtOAc in pet. ether) gave 101 (1.69 g, 98 %) as a white solid. v_{max} (ATR) 3293 (s), 2976, 1703, 1658, 1537, 1293, 1173, 1066, 981 cm⁻¹; δ_{H} (700 MHz, CDCl₃) 5.24 (1H, m, NH), 4.68 (1H, m, 2-H), 3.77 (3H, s, OCH₃), 3.20 (3H, s, NCH₃), 1.44 (9H, s, C(CH₃)₃), 1.31 (3H, d, *J* 6.9, 3-H₃); δ_{C} (176 MHz, CDCl₃) 173.8 *C1*, 155.3 NHCOO, 79.7 *C*(CH₃)₃, 61.8 OCH₃, 46.7 *C2*, 32.3 NCH₃, 28.5 C(CH₃)₃, 18.9 C3; *m/z* (ES⁺) 232.9 [M]⁺ C₁₀H₂₀O₄N₂²³ (Expected: 232.1), 255.1 [M+Na]⁺ C₁₀H₂₀O₄N₂²³Na (Expected: 255.1).



13 (S)-tert-Butyl (3-oxopent-4-en-2-yl)carbamate³

Procedure:² Vinyl magnesium bromide (12 ml of 0.6M solution in THF, 7.2 mmol, 4 eq.) was added dropwise at 0 °C to a solution of S01 (400 mg, 1.8 mmol) in anhydrous THF (10 ml). The reaction mixture was allowed to warm up to room temperature. After the reaction mixture was stirred for 1 hr at the same temperature, the reaction mixture was poured into ice cooled 2N HCl. The resulting mixture was extracted with ethyl acetate. The organic layers were combined, washed with brine, dried over MgSO₄, filtered, and concentrated *in vacuo* to give the crude products. Column chromatography on silica gel (from 10 % to 50 % EtOAc in pet. ether) gave 13 (335 mg, 98 %) as a white solid ($R_f \approx 0.37$; EtOAc/pet. ether 25/75). v_{max} (ATR) 3382 (s), 2975, 2931, 1710, 1694, 1612, 1518, 1282, 1246, 1162, 1003 cm⁻¹; δ_{H} (700 MHz, CDCl₃) 6.45 (1H, dd, *J* 17.4, 10.5, 4-*H*), 6.37 (1H, dd, *J* 17.4, 1.3, 5- H_{trans}), 5.88 (1H, d, *J* 10.5, 1.3, 5- H_{cis}), 5.36 (1H, m, NH), 4.61 (1H, p, *J* 7.2, 2-*H*), 1.43 (9H, s, C(CH₃)₃), 1.33 (3H, d, *J* 7.2, , 1- H_3); δ_{C} (176 MHz, CDCl₃) 198.8 C3, 155.3 NHCOO, 132.9 C4, 130.3 C5, 79.8 C(CH₃)₃, 53.2 C2, 28.5 C(CH₃)₃, 18.6 C1; m/z (ES⁺) 222.2 [M+Na]⁺ C₁₀H₁₇O₃N²³Na (Expected: 222.1).



16 *tert*-Butyl ((2S,3R)-3-hydroxypent-4-en-2-yl)carbamate⁴

Procedure:² To a solution of 13 (500 mg, 2.5 mmol) in spectrophotometric grade ethanol (31 ml) was added lithium *tri-tert*-butoxy aluminium hydride (1.40 g, 5.5 mmol, 2.2 eq.) at –78 °C. After the reaction mixture was stirred at the same temperature for 2 hrs, 0.1N HCl (15 ml) was added followed by Celite and EtOAc (15 ml). The resulting slurry was filtered through Celite and the filtering bed was washed with EtOAc (15ml). The two phases were separated and the aqueous phase was re-extracted with EtOAc. The organic extracts were combined, washed with NaHCO₃ and brine, dried over MgSO₄, filtered, and concentrated in vacuo to give the product as a single diastereoisomer 16 as ascertained by crude ¹H NMR. Column chromatography on silica gel (from 25 % to 50 % ethyl acetate in pet. ether) gave 16 (450 mg, 89 %) as a white solid ($R_f \approx 0.3$; EtOAc/pet. ether 25/75). v_{max} (ATR) 3351 (br, s), 2986, 2937, 1679, 1529, 1280, 1161, 1021 cm⁻¹; δ_{H} (700 MHz, CDCl₃) 5.85 (1H, ddd, *J* 17.2, 10.6, 5.5, 4-*H*), 5.33 (1H, dt, *J* 17.2, 1.5, 5-*H*_{trans}), 5.23 (1H, dd, *J* 10.6, 1.5, 5-*H*_{cis}), 4.65 (1H, m, NH), 4.22–4.16 (1H, m, 3-*H*), 3.84 (1H, m, 2-*H*), 1.44 (9H, s, C(CH₃)₃), 1.09 (3H, d, *J* 6.9, 1-*H*₃); δ_{C} (176 MHz, CDCl₃) 156.5 NHCOO, 137.0 C4, 116.7 C5, 79.9 $C(CH_3)_3$, 75.9 C3, 50.9 C2, 28.5 C(CH₃)₃, 15.5 C1; *m*/z (ES⁺) 224.3 [M+Na]⁺ C₁₀H₁₉O₃N²³Na (Expected: 224.1).



19 (S)-*tert*-Butyl (3-oxohex-5-en-2-yl)carbamate^{5, 6}

Procedure:⁶ A solution of *n*-BuLi (1.4 M solution in hexanes, 7.2 ml, 10 mmol, 1.0 eq.) was added dropwise at -10 °C to a solution of *N*-(*tert*-butoxycarbonyl)-L-alanine (1.89 g, 10 mmol) in anhydrous THF (100 ml). The resulting thick gelatinous suspension was stirred at -10 °C for 30 minutes, cooled to -78 °C and treated with a solution of allylmagnesium bromide (1.0 M solution in ether, 23.0 ml, 23 mmol, 2.3 eq.). The resultant light grey slurry was stirred for 1 hour at -78 °C, warmed to room temperature over 1 hour, stirred at this temperature for an additional 30 minutes and then poured into a mixture of sat. aq. NH₄Cl solution, ice and ether. The organic layer was separated and the aqueous layer was extracted with ether. The combined organic layers were washed with brine, dried over MgSO₄, filtered and concentrated under *vacuo* to yield the desired product 19 as a crystalline white solid (2.03 g, 9.5 mmol, 95 %). The product was used without further purification ($R_f \approx 0.80$; EtOAc/pet. ether 67/33). v_{max} (ATR) 3386, 2975, 2932, 1726, 1694, 1642, 1516, 1166 cm⁻¹; $\delta_{\rm H}$ (500 MHz, CDCl₃) 5.91 (1 H, ddt, *J* 17.1, 10.2, 6.9, 5-*H*), 5.15 (3H, m, NH and 6- H_2), 4.41–4.31 (1H, m, 2-H), 3.34–3.21 (2H, m, 4- H_2), 1.43 (9H, s, C(CH₃)₃), 1.33 (3H, d, *J* 7.1, 1- H_3); $\delta_{\rm C}$ (126 MHz, CDCl₃) 207.6 C3, 155.3 NHCOO, 129.9 C5, 119.5 C4, 79.9 C(CH₃)₃, 54.9 C2, 44.2 C3, 28.5 C(CH₃)₃, 17.8 C1; *m/z* (ES⁻) 212.1 [M–H]⁻; (ES⁺) 236.2 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 236.1257, C₁₁H₁₉O₃N²³Na requires M^+ 236.1257.



20 *tert*-Butyl (2*S*,3*R*)-3-hydroxyhex-5-en-2-ylcarbamate⁶

Procedure:⁶ A solution of 19 (2 g, 9.4 mmol) in anhydrous methanol (60 ml) was treated with sodium borohydride (0.72 g, 18.5 mmol, 1.97 eq.) at -78 °C. The resulting mixture was stirred for 90 minutes, carefully quenched by addition of a sat. aq. NH₄Cl solution at -78 °C, warmed to room temperature, and diluted with 1M NaOH solution (24 ml) and ether (60 ml). The organic layer was separated and the aqueous layer was extracted with ether (2 × 20 ml). The combined organic extracts were washed with brine, dried over MgSO₄, filtered and concentrated under vacuum to yield the crude product as a white solid consists of an 82:18 mixture of 20 and the other diastereoisomer respectively as determined by analysis of the ¹H NMR spectra of the crude product. The crude mixture was purified by chromatography (25 % ethyl acetate in pet. ether) to yield 0.73 g of a 70/30 mixture of the diastereoisomers followed by 1.14 g of pure 20 with total yield 92 % (1.87 g, 8.7 mmol) ($R_f 20 \approx 0.30$, $R_f 20' \approx 0.32$; EtOAc/pet. ether 25/75). 20; v_{max} (ATR) 3358 (br, s), 2979, 2940, 1681 (s), 1526 (s), 1174 (s), 1024 (s) cm⁻¹; δ_{H} (500 MHz, CDCl₃) 5.84 (1H, ddt, *J* 7.3, 10.3, 14.4, 5-H), 5.19–5.09 (2H, m, 6-H), 4.75 (1H, br s, NH), 3.71 (2H, m, 2-H and 3-H), 2.38–2.11 (3H, m, OH, 4-H₂), 1.44 (9H, s, C(CH₃)₃), 1.12 (3H, d, *J* 6.8, 1-H₃); δ_{C} (101 MHz, CDCl₃) 156.0 NHCOO, 134.8 C5, 118.2 C6, 79.6 C(CH₃)₃, 73.4 C3, 50.4 C2, 38.5 C4, 28.5 C(CH₃)₃, 14.7 C1; *m*/z (ES⁺) 238.2 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 238.1414, C₁₁H₂₁O₃N²³Na requires *M*⁺ 238.1414.



S02 (S)-2-((tert-Butoxycarbonyl)amino)-3-((tert-butyldimethylsilyl)oxy)propanoic acid^{2, 6}

Procedure:⁶ To a solution of *N*-(*tert*-butoxycarbonyl)-L-serine (512 mg, 2.5 mmol) in anhydrous DMF (12 mL), imidazole (509 mg, 7.5 mmol, 3 eq.) was added and the reaction mixture was cooled to 0 °C followed by addition of TBSCI (490 mg, 3.25 mmol, 1.3 eq.). The resulting mixture was slowly warmed to room temperature and stirred overnight and followed by TLC. Upon total consumption of the starting material (monitored by TLC), the reaction mixture was poured into a mixture of ice cooled 1N HCl (5 mL) and ether (20 mL) to hydrolyze the silyl ester. The organic layer was separated and the aqueous layer was extracted with ether (2 × 20 ml). The combined organic extracts were washed with brine, dried over MgSO4, filtered and concentrated under vacuum to yield the desired product S02 as pale yellow sticky oil (630 mg, 1.97 mmol, 79 %). v_{max} (ATR) 3380-3492 (br), 3451 (s), 2952, 2569, 1724, 1691, 1505 cm⁻¹; δ_{H} (200 MHz, CDCl₃) 8.49 (1H, br s, acidic OH), 5.35 (1H, br d, *J* 8.2, NH), 4.36 (1H, m, 2-H), 4.09 (1H, A of ABX syst., m, 3-HH), 3.83 (1H, B of ABX syst., dd, *J* 10.1, 3.5, 3-HH), 1.44 (9H, s, OC(CH₃)₃), 0.86 (9H, s, SiC(CH₃)₃), 0.03, 0.2 (2 × 3H, s, SiC(H₃)₃); δ_{C} (75 MHz, CDCl₃) 175.3 COOH, 155.5 NHCOO, 80.1 OC(CH₃)₃, 63.7 C3, 55.6 C2, 28.2 OC(CH₃)₃, 25.7 SiC(CH₃)₃, 18.4 SiC(CH₃)₃, -5.5 SiCH₃, -5.6 SiCH₃; *m*/z (ES⁺) 320.3 [M+H]⁺ C₁₄H₃₀O₅N (Expected: 320.2).



S03 (S)-tert-Butyl (3-hydroxy-1-(methoxy(methyl)amino)-1-oxopropan-2-yl)carbamate^{2,7}

Procedure:² To a solution of *N*-(*tert*-butoxycarbonyl)-L-serine (3.075 g, 15 mmol) in anhydrous CH₂Cl₂ (100 ml) was added *N*-methylmorpholine (1.812 ml, 16.5 mmol, 1.1 eq.) and *N*,O-dimethylhydroxylamine hydrochloride (1.610 g, 16.5 mmol, 1.1 eq.) at -15 °C. To the reaction mixture was added 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (2.874 g, 16.5 mmol, 1.1 eq.) over 30 min at the same temperature. The reaction mixture was stirred for 4 hrs then poured into ice and 1N HCl. The resulting mixture was extracted with CH₂Cl₂. The organic layers were combined, and washed with brine, dried over MgSO₄, filtered, and concentrated *in vacuo* to give the crude Weinreb amide S03 (3.400 gm, 91%) as a colourless solid. The crude product was at high purity and was used directly without further purification. v_{max} (ATR) 3329, 2935, 1715, 1665, 1498, 1173 cm⁻¹; δ_{H} (500 MHz, CDCl₃) 5.57 (1H, br s, NH), 4.89–4.74 (1H, m, 2-H), 3.86–3.79 (2H, m, 3-H₂), 3.78 (3H, s, OCH₃), 3.24 (3H, s, NCH₃), 2.49 (1H, br s, OH), 1.45 (9H, s, C(CH₃)₃); δ_{C} (126 MHz, CDCl₃) 170.9 C1, 156.0 NHCOO, 80.3 *C*(CH₃)₃, 64.1 C3, 61.8 OCH₃, 52.5 C2, 31.1 NCH₃, 28.5 C(CH₃)₃; *m*/z (ES⁺) 271.2 [M+Na]⁺ C₁₀H₂₀O₅N₂²³Na (Expected: 271.1).



S04 (S)-tert-Butyl (3,8,8,9,9-pentamethyl-4-oxo-2,7-dioxa-3-aza-8-siladecan-5-yl)carbamate²

Procedure:⁷ To a solution of S03 (500 mg, 2.02 mmol) in anhydrous DMF (2.5 mL) were added imidazole (410 mg, 6.12 mmol, 3 eq.) and a catalytic amount of DMAP (25 mg, 0.2 mmol, 0.1 eq.) at 0 °C followed by addition of a solution of TBSCI (349 mg, 2.32 mmol, 1.15 eq.) in anhydrous DMF (2.5 ml). The resulting mixture was allowed to warm up to room temperature, stirred overnight, and poured into sat. aq. NH₄Cl solution (5 ml) and extracted with CH₂Cl₂ (3 × 20 ml). The combined organic extracts were washed with brine, dried over MgSO₄, filtered and concentrated under vacuum to yield the crude product as yellowish viscous oil. Column chromatography on silica gel (from 25% to 50% EtOAc in pet. ether) gave S04 (650 mg, 1.80 mmol, 89%) as a sticky oil ($R_f \approx 0.25$; EtOAc/pet. ether 25/75). v_{max} (ATR) 2931, 2857, 2350, 1711, 1661, 1494, 1468, 1168, 1111, 837 cm⁻¹; $\delta_{\rm H}$ (700 MHz, CDCl₃) 5.35 (1H, br d, *J* 8.5, N*H*), 4.80–4.70 (1H, m, 5-*H*), 3.84 (1H, dd, *J* 9.4, 4.5, 6-*H*H), 3.78 (1H, dd, *J* 9.7, 4.5, 6-HH), 3.74 (3H, s, 1-*H*₃), 3.20 (3H, s, NC*H*₃), 1.43 (9H, s, C(C*H*₃)₃), 0.85 (9H, s, SiC(C*H*₃)₃), 0.02, 0.01 (2 × 3H, s, SiC*H*₃); $\delta_{\rm C}$ (176 MHz, CDCl₃) 170.9, 155.5, 79.7, 63.7, 61.6, 52.6, 32.3, 28.5, 25.9, -5.4, -5.4; *m*/z (ES⁺) 385.2 [M+Na]⁺ C₁₆H₃₄O₅N₂²⁸Si²³Na (Expected: 385.1).



12 (S)-tert-Butyl (1-((tert-butyldimethylsilyl)oxy)-3-oxopent-4-en-2-yl)carbamate²

Procedure:² A (0.6 M) solution of vinyl magnesium bromide (*c.a.* 11 ml, 6.64 mmol, 4 eq.) in THF was added dropwise at 0 °C to a solution of S04 (600 mg, 1.66 mmol) in anhydrous THF (10 ml). The reaction mixture was allowed to warm up to room temperature. After the reaction mixture was stirred for 1 hour at the same temperature, the reaction mixture was poured into ice cooled 2N HCl. The resulting mixture was extracted with ethyl acetate. The organic layers were combined, washed with brine, dried over MgSO₄, filtered, and concentrated in vacuo to give the crude products. Column chromatography on silica gel (from 5% to 20% EtOAc in pet. ether) gave 12 (920 mg, 84%) as a viscous oil ($R_f \approx 0.78$; EtOAc/pet. ether 25/75). v_{max} (ATR) 3460, 2865, 1695, 1489, 1359, 1173 cm⁻¹; δ_{H} (700 MHz, CDCl₃) 6.55 (1H, dd, *J* 17.5, 10.7, 4-CH), 6.34 (1H, dd, *J* 17.5, 1.0, 5-CH_{trans}), 5.83 (1H, d, *J* 10.7, 1.0, 5-CH_{cis}), 5.52 (1H, br d, *J* 6.8, NH), 4.63–4.56 (1H, m, 2-CH), 4.00 (1H, dd, *J* 10.2, 3.1, 1-CHH), 3.85 (1H, dd, *J* 10.2, 4.4, 1-CHH), 2.04 (1H, s, OH), 1.44 (9H, s, C(CH₃)₃), 0.83 (9H, s, SiC(CH₃)₃), 0.02, 0.01 (2 × 3H, s, SiCH₃); δ_{C} (176 MHz, CDCl₃) 196.8, 155.3, 133.1, 129.3, 79.7, 63.4, 59.5, 28.3, 25.7, 18.2, -5.6; *m*/z (ES⁺) 330.5 [M+H]⁺ C₁₆H₃₂O₄N₁²⁸Si (Expected: 330.2).



14 *tert*-Butyl ((2S,3R)-1-((*tert*-butyldimethylsilyl)oxy)-3-hydroxypent-4-en-2-yl)carbamate²

Procedure:² To a solution of the enone 12 (910 mg, 2.77 mmol) in spectrophotometric grade ethanol (35 ml) was added lithium *tri-tert*-butoxy aluminium hydride (1.546 g, 6.1 mmol, 2.2 eq.) at -78 °C. After the reaction mixture was stirred at the same temperature for 2 hrs, 0.1N HCl (16.5 ml) was added followed by Celite and EtOAc (16.5 ml). The resulting slurry was filtered through Celite and the residue was washed with EtOAc (16.5ml). The two phases were separated and the aqueous phase re-extracted with EtOAc. The organic extracts were combined, washed with NaHCO₃ and brine, dried over MgSO₄, filtered, and concentrated *in vacuo* to give the crude product as a single diastereomer 14 as indicated by the ¹H NMR analysis. Column chromatography on silica gel (from 20% to 50% ethyl acetate in pet. ether) gave 14 (870 mg, 95%) as a viscous oil ($R_f \approx 0.40$; EtOAc/pet. ether 25/75). v_{max} (ATR) 3445, 2940, 1699, 1494, 1168, 840 cm⁻¹; δ_{H} (500 MHz, CDCl₃) 5.91 (1H, dd, J 17.1, 10.6, 4.9, 4-H), 5.37 (1H, dd, J 17.1, 1.6, 5- H_{trans}), 5.27 (1H, br d, J 7.9, NH), 5.23 (1H, dd, J 10.6, 1.6, 5- H_{cis}), 4.29–4.22 (1H, m, 3-H), 3.91 (1H, dd, J 10.4, 2.9, 1-HH), 3.74 (1H, dd, J 8.3, 2.9 1-HH), 3.65–3.58 (1H, br s, OH), 3.47 (1H, m, 2-H), 1.44 (9H, s, OC(CH₃)₃), 0.88 (9H, s, SiC(CH₃)₃), 0.05, 0.04 (2 × 3H, s, SiCH₃); δ_{C} (126 MHz, CDCl₃) 155.9, 138.0, 115.9, 79.7, 74.9, 63.6, 54.2, 28.5, 28.5, 25.9, 25.9, 18.2, -5.52, -5.54; m/z (ES⁺) 332.4 [M+H]⁺ C₁₆H₃₃O₄N₁Si₁ (Expected: 332.2), 354.4 [M+Na]^{*} C₁₆H₃₃O₄N₁²⁸Si²³Na (Expected: 354.2).



15 *tert*-Butyl ((2S,3*R*)-1,3-dihydroxypent-4-en-2-yl)carbamate²

Procedure: A solution of 14 (100 mg, 0.3 mmol) in methanol (1.5 ml) was treated with 2N HCl (150 µl) dropwise at -0 °C. The resulting mixture was stirred for 15 minutes at the same temperature and monitored by TLC. Upon complete consumption of the starting material, the reaction was quenched by addition of a brine solution (5 ml) and extracted with EtOAc (2 × 10 ml). The combined organic layers were washed with brine, dried over MgSO₄, filtered and concentrated under vacuum to yield the crude product. Column chromatography on silica gel (from 25% to 50% ethyl acetate in pet. ether and elution with MeOH) gave 15 (60 mg, 2.76 mmol, 92 %) as viscous oil ($R_f \approx 0.13$; EtOAc/pet. ether 50/50). v_{max} (ATR) 3456, 2931, 1705, 1510, 1177, 842 cm⁻¹; δ_H (700 MHz, CDCl₃) 5.94 (1H, ddd, *J* 16.9, 10.6, 5.3, 4-*H*), 5.40 (1H, d, *J* 16.9, 5-*H*_{trans}), 5.35 (1H, br s, N*H*), 5.27 (1H, d, *J* 10.6, 5-*H*_{cis}), 4.39 (1H, br s, 3-*H*), 3.93 (1H, dd, *J* 11.2, 3.5, 1-H*H*), 3.74–3.68 (1H, m, 1-*H*H), 3.68–3.61 (1H, m, 2-*H*), 2.85 (1H, app d, *J* 4.5, 3-O*H*), 2.57 (1 H, br s, 1-O*H*), 1.45 (9H, s, C(C*H*₃)₃); δ_C (176 MHz, CDCl₃) 156.3, 137.6, 116.7, 80.1, 75.1, 62.7, 55.1, 28.5; *m/z* (ES⁺) 240.4 [M+Na]⁺ C₁₀H₁₉O₄N²³Na (Expected: 240.1).



S05 (1S,2S,4S,5R)-1-(Anthracen-9-ylmethyl)-2-((*R*)-hydroxy(quinolin-4-yl)methyl)-5-vinyl-1-azoniabicyclo [2.2.2]octane chloride⁸

Procedure:⁸ To a suspension of cinchonidine (294 mg, 1.0 mmol) in toluene (4 ml) was added 9-(chloromethyl) anthracene (237 mg, 1.05 mmol, 1.05 eg.), and the mixture was stirred at reflux 110 °C for 3 hrs. The mixture was cooled to room temperature, poured into 20 ml of ether. The solid residue was collected by centrifugation and decantation of the liquid phase. The residue was washed by ether (2 × 20 ml) and collected by centrifugation. The mother liquor was concentrated in vacuo and the residue was dissolved in CH₂Cl₂/ether and kept at -20 °C to precipitate another fraction of the product. The combined solid residue was recrystallized from CH₂Cl₂/ether and kept at -20 °C to afford S05 (460 mg, 0.88 mmol, 88%) as light yellow solid ($R_f \approx 0.30$; CH₂Cl₂/MeOH 93/7). δ_H (400 MHz, CDCl₃, 20 mg/ml)⁹ 8.97 (1H, d, J 8.4), 8.86 (1H, d, J 4.3), 8.79 (2H, d, J 8.9), 8.17 (1H, d, J 4.9), 8.06-8.01 (2H, m, J 4.7), 7.71 (1H, d, J 8.2), 7.68–7.64 (1H, m), 7.61 (1H, d, J 7.8), 7.47–7.40 (1H, m), 7.29–7.21 (4H, m), 7.19–7.09 (1H, m), 7.09–7.04 (1H, m), 6.74 (2H, s), 5.42 (1H, ddd, J 16.8, 10.5, 6.1), 5.21 (1H, d, J 16.8), 4.90 (1H, dd, J 10.5, 1.4), 4.85–4.73 (1H, m), 4.72–4.58 (1H, m), 3.97 (1H, d, J 13.0), 2.58 (1H, dd, J 12.9, 10.6), 2.47 (1H, app. t, J 12.6), 2.18–2.06 (1H, m), 1.95–1.77 (2H, m), 1.77–1.56 (1H, m), 1.22–1.11 (1 H, m), 1.11–0.95 (1H, m); δ_C (176 MHz, CDCl₃) 149.01, 136.52, 133.38, 132.76, 131.21, 130.45, 130.39, 128.98, 128.86, 128.38, 127.76, 127.46, 127.23, 125.98, 125.73, 124.93, 124.91, 124.38, 124.06, 120.26, 118.26, 117.86, 67.22, 67.18, 61.58, 54.98, 50.66, 38.64, 25.99, 25.81, 23.50; *m*/z (ES⁺) 485.5 [M]⁺ C₃₄H₃₃ON₂ (Expected: 485.3). Page 8 of 50



29 (1*S*,2*S*,4*S*,5*R*)-2-((*R*)-Allyloxy(quinolin-4-yl)methyl)-1-(anthracen-9-ylmethyl)-5-vinyl-1-azoniabicyclo [2.2.2]octane bromide⁸

Procedure:⁸ To a suspension of S05 (200 mg, 0.39 mmol) in CH₂Cl₂ (5 ml) was added allyl bromide (100 μl, 1.17 mmol, 3.05 eq.) and 50% aq. KOH solution (200 μl, 1.9 mmol, 5 eq.). The resulting mixture was stirred vigorously at room temperature for 4 hrs. The mixture was diluted with water (6.25 ml) and extracted with CH₂Cl₂ (3 × 5 ml). The combined organic extracts were dried over MgSO4, filtered and concentrated *in vacuo*. Recrystallization of the residue from MeOH/ether at -20 °C afforded the desired product 29 (190 mg, 0.32 mmol, 84%) as an orange solid. $\delta_{\rm H}$ (700 MHz, CD₃OD, 25 mg/ml)⁹ 9.07 (1H, d, *J* 4.4), 8.93 (1H, s), 8.77 (1H, d, *J* 9.0), 8.63–8.56 (1H, m), 8.48 (1H, d, *J* 9.1), 8.31–8.26 (2H, m), 8.25 (1H, d, *J* 8.3), 8.01–7.93 (3H, m, *J* 15.0, 7.2), 7.83 (1H, dd), 7.79 (1H, dd, *J* 8.5, 7.2), 7.68 (2H, dd, *J* 14.4, 6.3), 6.98 (1H, s), 6.44 (2H, ddd, *J* 15.9, 11.0, 5.6), 5.92 (1H, d, *J* 13.9), 5.76–5.67 (2H, m), 5.58 (1H, d, *J* 10.4), 5.06–4.98 (2H, m, *J* 13.2), 4.59–4.52 (2H, m, *J* 13.0, 6.0), 4.51–4.42 (2H, m), 3.80 (1H, d, *J* 13.0), 3.30–3.24 (1H, m), 2.94 (1H, td, *J* 11.6, 5.1), 2.54–2.42 (2H, m, *J* 23.5, 13.6), 2.21 (1H, s), 2.00 (1H, s), 1.70–1.58 (2H, m); $\delta_{\rm C}$ (176 MHz, CD₃OD) 151.05, 149.38, 142.94, 138.50, 134.80, 134.67, 134.61, 133.94, 133.10, 133.04, 131.52, 131.31, 131.18, 130.66, 129.41, 129.26, 127.08, 126.66, 126.57, 125.28, 124.88, 121.65, 119.12, 117.80, 71.44, 70.12, 63.55, 57.42, 53.69, 49.53, 39.48, 27.30, 26.26, 23.31; *m*/z (ES⁺) 525.5 [M]⁺ C₃₇H₃₇ON₂ (Expected: 525.3).



S06 (1S,2R,4S,5R)-1-(Anthracen-9-ylmethyl)-2-((S)-hydroxy(quinolin-4-yl)methyl)-5-vinyl-1-azoniabicyclo [2.2.2]octane chloride⁹

Procedure:⁹ To a suspension of cinchonine (294 mg, 1.0 mmol) in toluene (4 ml) was added 9-(chloromethyl)anthracene (237 mg, 1.05 mmol, 1.05 eq.), and the mixture was stirred at reflux 110 °C for 4 days. The mixture was cooled to room temperature, poured into 20 ml of ether. The solid residue was collected by centrifugation and decantation of the liquid phase. The residue was washed by ether (2 × 20 ml) and collected by centrifugation. The mother liquor was concentrated *in vacuo* and the residue was dissolved in CH₂Cl₂/ether and kept at -20 °C to precipitate another fraction of the product. The combined solid residue was recrystallized from CH₂Cl₂/ether and kept at -20 °C to afford S06 (250 mg, 0.48 mmol, 48%) as light yellow solid ($R_f \approx 0.30$; Page 9 of 50 CH₂Cl₂/MeOH 93/7). $\delta_{\rm H}$ (700 MHz, CDCl₃, 25 mg/ml)⁹ 9.24 (1H, d, *J* 8.6), 8.92 (1H, d, *J* 6.7), 8.81 (1H, s), 8.41 (1H, d, *J* 8.7), 8.20 (1H, s), 8.02 (1H, d, *J* 3.7), 7.85 (1H, s), 7.55 (1H, d, *J* 8.1), 7.53 (1H, d, *J* 8.3), 7.45 (1H, d, *J* 8.2), 7.27–7.23 (1H, m), 7.21–7.16 (2H, m), 7.11–7.06 (1H, m, *J* 6.7), 7.06–7.02 (1H, m, *J* 7.2), 6.98–6.89 (3H, m), 6.46 (1H, d, *J* 13.7), 5.56 (1H, ddd, *J* 17.2, 10.5, 6.6), 5.00 (1H, d, *J* 10.5), 4.84 (1H, d, *J* 17.3), 4.75–4.68 (1H, m), 4.44–4.36 (1H, m), 4.24 (1H, t, *J* 11.5), 2.45 (2H, t, *J* 12.0), 2.30 (1H, dd, *J* 20.5, 10.7), 1.92 (1H, t, *J* 12.6), 1.73–1.64 (2H, m, *J* 23.5, 10.1), 1.49 (1H, s), 1.34 (1H, t, *J* 11.1), 0.65 – 0.57 (1H, m); $\delta_{\rm H}$ (176 MHz, CDCl₃) 149.4, 147.0, 145.6, 135.6, 133.1, 132.8, 130.9, 130.4, 130.1, 129.0, 128.5, 128.2, 127.6, 127.3, 126.9, 124.9,124.9, 124.61, 120.1, 118.1, 117.5, 67.8, 66.8, 57.6, 54.3, 54.0, 38.1, 26.4, 24.1, 22.7; *m*/z (ES⁺) 485.8 [M]⁺ C₃₄H₃₃ON₂ (Expected: 485.3).



28 (1*S*,2*R*,4*S*,5*R*)-2-((*S*)-Allyloxy(quinolin-4-yl)methyl)-1-(anthracen-9-ylmethyl)-5-vinyl-1-azoniabicyclo [2.2.2]octane bromide¹⁰

Procedure:⁸ To a suspension of S06 (272 mg, 0.52 mmol) in CH_2Cl_2 (7 ml) was added allyl bromide (141 µl, 1.63 mmol, 3.05 eq.) and 50% aq. KOH solution (286 µl, 2.6 mmol, 5 eq.). The resulting mixture was stirred vigorously at room temperature for 4 hrs. The mixture was diluted with water (9 ml) and extracted with CH_2Cl_2 (3 × 7 ml). The combined organic extracts were dried over MgSO4, filtered and concentrated *in vacuo*. Recrystallization of the residue from MeOH/ether at -20 °C afforded the desired product 28 (275 mg, 0.45 mmol, 87%) as orange solid. δ_{H} (700 MHz, CD₃OD, 18 mg/ml)⁹ 9.04 (1H, d, *J* 4.0), 9.01 (1H, d, *J* 9.0), 8.77 (2H, m), 8.27 (1H, d, *J* 8.7), 8.20–8.14 (3H, m), 8.00 (1H, s), 7.94–7.90 (2H, m), 7.84–7.79 (1H, m), 7.70–7.66 (1H, m), 7.62 – 7.57 (2H, m, *J* 10.4, 5.4), 6.90 (1H, s), 6.46–6.38 (1H, m), 6.13–5.99 (2H, m), 5.93 (1H, ddd, *J* 17.2, 10.6, 6.9), 5.72 (1H, d, *J* 17.2), 5.63 (1H, d, *J* 10.6), 5.19 (1H, d, *J* 10.5), 5.04 (1H, d, *J* 17.3), 4.58 (1H, m), 4.55 (1H, dd, *J* 12.8, 6.0), 4.41–4.34 (3H, m), 3.11 (1H, app. t, *J* 11.5), 2.77 (1H, m), 2.55–2.50 (1H, m), 2.25–2.14 (1H, m), 1.85–1.78 (2H, m), 1.64–1.55 (1H, m), 1.20–1.12 (1H, m), 0.70 (0.45H, t, *J* 7.4); *m/z* (ES⁺) 525.7 [M]⁺ C₃₇H₃₇ON₂ (Expected: 525.3).



22 Methyl 2-((diphenylmethylene)amino)acetate¹¹

Procedure:¹¹ To a suspension of methyl glycine hydrochloride (753 mg, 6 mmol) in anhydrous CH₂Cl₂ (20 ml), an equimolar amount of benzophenone imine (1 ml, 6 mmol) was added and the reaction mixture was stirred at room temperature for 24 hr. The reaction mixture was filtered to remove NH₄Cl and evaporated to dryness on a rotary evaporator. The residue was dissolved in 20 mL of ether, filtered, washed with 20 mL of water, and dried over MgSO4 to yield the crude product. Column chromatography on silica gel (from 10% to 50% ethyl acetate in pet. ether) gave 22 (1.221 g, 4.8 mmol, 80 %) as white waxy solid ($R_f \approx 0.33$; EtOAc/pet. ether 33/67). v_{max} (ATR) 2948, 2890, 2361, 1970, 1752, 1622, 1573, 1384, 1195, 682 cm⁻¹; δ_H (700 MHz, CDCl₃) 7.66 (2H, m, Ar*H*), 7.49–7.42 (3H, m, Ar*H*), 7.40 (1H, m, Ar'*H*), 7.33 (2H, m, Ar'*H*), 7.18 (2H, m, Ar'*H*), 4.22 (2H, s 2-*H*₂), 3.74 (3H, s, OC*H*₃); δ_C (176 MHz, CDCl₃) 172.0, 171.2, 139.3, 136.0, 130.6, 128.9, 128.9, 128.8, 128.2, 127.8, 55.7, 52.1; m/z (ES⁺) 254.4 [M+H]⁺ C₁₆H₁₆O₂N (Expected: 254.1).

General Procedure for Phase Transfer Catalysis^{8, 12}

To an ice cooled solution of the Schiff base 22 in Toluene/CH₂Cl₂ (7:3 v/v), the cinchona catalyst 29 or 28 was added (5 mol%) followed by dropwise addition of allyl bromide (5.0 eq.) at 0 °C. The reaction mixture was stirred for 5 min before the dropwise addition of 50% aq. KOH (5 eq.). The reaction was stirred vigorously overnight from 0°C to room temperature. The reaction was monitored by TLC and upon completion; the reaction mixture was diluted with water (10 ml) and extracted with EtOAc (3 × 5 ml). The organics were dried over MgSO₄, filtered, and evaporated in vacuo, affording the crude products.



 v_{max} (ATR) 2954, 2385, 1704, 1491, 1168, 840 cm⁻¹; δ_{H} (400 MHz, CDCl₃) 7.65–7.61 (2H, m, Ar*H*), 7.47–7.42 (3H, m, Ar*H*), 7.40–7.36 (1H, m, Ar*H*), 7.35–7.30 (2H, m, Ar*H*), 7.19–7.14 (2H, m, Ar*H*), 5.67 (1H, ddt, *J* 17.1, 10.1, 7.2, 4-*H*), 5.07 (1H, dd, *J* 17.1, 2.0, 5-H*H*), 5.02 (1H, dd, *J* 10.1, 2.0, 5-*H*H), 4.16 (1H, dd, *J* 7.8, 5.3, 2-*H*), 3.72 (3H, s, OC*H*₃), 2.75–2.56 (2H, m, 3-*H*₂); δ_{C} (101 MHz, CDCl₃) 172.4 (COOCH₃), 170.8 (Ph₂CN), 139.6 (Ar), 136.4 (Ar), 134.4 (Ar), 130.5 (C4), 128.94 (Ar), 128.78 (Ar), 128.61 (Ar), 128.14 (Ar), 128.03 (Ar), 117.8 (C5), 65.4 (C2), 52.1 (OCH₃), 38.3 (C3).



S09 (S)-Methyl 2-((*tert*-butoxycarbonyl)amino)pent-4-enoate^{13, 14}

Procedure:¹⁵ A mixture of the starting material S07 (735 mg, 2.51 mmol) and 4N HCl (3.1 ml, excess) in MeOH (10 ml) was refluxed for 1 hr. The reaction mixture was allowed to cool down to room temperature. The reaction mixture was extracted by ether (2 × 5 ml). The ether layer was discarded and the aqueous layer was basified by NaHCO₃ to pH 8 followed by addition of Boc2O (Boc anhydride) (824 mg, 3.77 mmol, 1.5 eq.) and the reaction was stirred for 2 hrs at room temperature and monitored by TLC. The reaction volume was reduced under vacuum and extracted with EtOAc (3 × 10 ml). The combined organic extracts were washed with brine, dried over MgSO4, filtered and evaporated in vacuo to afford the crude product. Column chromatography on silica gel (from 20% to

50% ethyl acetate in pet. ether) gave S09 (442 g, 1.9 mmol, 76%) as viscous liquid (Rf ≈ 0.42; EtOAc/pet. ether 33/67). v_{max} (ATR) 3375, 2971, 2879, 1716, 1491, 1173, 840 cm⁻¹; δ_{H} (400 MHz, CDCl₃) 5.68 (1H, ddt, *J* 17.0, 9.7, 7.2, 4-*H*), 5.17–5.08 (2H, m, 5-*H*₂), 5.03 (1H, br d, *J* 6.2, N*H*), 4.37 (1H, dd, *J* 13.3, 6.2, 2-*H*), 3.72 (3H, s, OC*H*₃), 2.61 – 2.39 (2H, m, 3-*H*₂), 1.42 (9H, s, C(C*H*₃)₃); δ_{C} (126 MHz, CDCl₃) 171.2, 155.3, 132.4, 119.1, 79.9, 52.3, 36.9, 28.4; *m*/z (ES⁺) 253.0 [M+Na]⁺ C₁₁H₁₉O₂N₁²³Na₁ (Expected: 253.1).



S10 (*R*)-Methyl 2-((*tert*-butoxycarbonyl)amino)pent-4-enoate^{13, 14}

Procedure:¹⁵ A mixture of the starting material S08 (690 mg, 2.35 mmol) and 4N HCl (3.0 ml, excess) in MeOH (10 ml) was refluxed for 1 hr. The reaction mixture was allowed to cool down to room temperature. The reaction mixture was extracted by ether (2 × 5 ml). The ether layer was discarded and the aqueous layer was basified by NaHCO₃ to pH 8 followed by addition of Boc₂O (Boc anhydride) (775 mg, 3.53 mmol, 1.5 eq.) and the reaction was stirred for 2 hrs at room temperature and monitored by TLC. The reaction volume was reduced under vacuum and extracted with EtOAc (3 × 10 ml). The combined organic extracts were washed with brine, dried over MgSO4, filtered and evaporated in vacuo to afford the crude product. Column chromatography on silica gel (from 20% to 50% ethyl acetate in pet. ether) gave S10 (408 g, 1.78 mmol, 75%) as viscous liquid (Rf ≈ 0.42; EtOAc/pet. ether 33/67). v_{max} (ATR) 3375, 2971, 2879, 1716, 1491, 1173, 840 cm⁻¹; δ_{H} (400 MHz, CDCl₃) 5.68 (1H, ddt, *J* 17.0, 9.7, 7.2, 4-*H*), 5.17–5.08 (2H, m, 5-*H*₂), 5.03 (1H, br d, *J* 6.2, N*H*), 4.37 (1H, dd, *J* 13.3, 6.2, 2-*H*), 3.72 (3H, s, OC*H*₃), 2.61–2.39 (2H, m, 3-*H*₂), 1.42 (9H, s, C(C*H*₃)₃); δ_{C} (126 MHz, CDCl₃) 171.2, 155.3, 132.4, 119.1, 79.9, 52.3, 36.9, 28.4; m/z (ES⁺) 253.1 [M+Na]⁺ C₁₁H₁₉O₂N₁²³Na₁ (Expected: 253.1).

25 (S)-*tert*-Butyl (1-hydroxypent-4-en-2-yl)carbamate¹⁶

Procedure: A solution of S09 (332 mg, 1.45 mmol) in anhydrous ether (5 ml) was added dropwise to a stirred suspension of LiAlH₄ (100 mg, 3.19 mmol, 2.2 eq.) in anhydrous ether (5 ml) at -0 °C. The reaction mixture was allowed to warm up to room temperature, was stirred for additional 1 hr and followed by TLC. Upon completion, the reaction was carefully quenched with dropwise addition of water (1 ml) followed by addition of 15% aq. NaOH (1 ml), water (3 ml), Celite and EtOAc (10 ml). The resulting slurry was filtered through Celite and the filtering bed was washed with EtOAc (2 × 5 ml). The two phases were separated and the organic layer was washed with brine, dried over MgSO₄, filtered and evaporated *in vacuo* to afford the crude product 25 (265 mg, 1.32 mmol, 91%) as a pale yellow oil ($R_f \approx 0.25$; EtOAc/pet. ether 33/67). The product was pure enough and was used without further purification. v_{max} (ATR) 3375, 2971, 2879, 1716, 1491, 1173, 840 cm⁻¹; δ_{H} (700 MHz, CDCl₃) 5.78 (1H, ddt, *J* 17.2, 10.2, 7.1, 4-*H*), 5.11 (2H, m, 5-*H*₂), 4.72 (1H, br s, N*H*), 3.74–3.58 (3H, m, 1-C*H*₂ and 2-*H*), 2.67 (1H, br s, O*H*),

2.31 (1H, m, 3-*H*H), 2.23 (1H, m, 3-H*H*), 1.43 (9H, s, C(C*H*₃)₃); δ_{C} (101 MHz, CDCl₃) 156.56, 134.28, 118.23, 79.89, 65.54, 52.30, 36.10, 28.49; *m*/z (ES⁺) 224.0 [M+Na]⁺ C₁₀H₁₉O₃N₁²³Na₁ (Expected: 224.1).



24 (R)-tert-Butyl (1-hydroxypent-4-en-2-yl)carbamate¹⁶

Procedure: A solution of S10 (388 mg, 1.69 mmol) in anhydrous ether (6 ml) was added dropwise to a stirred suspension of LiAlH₄ (102 mg, 3.72 mmol, 2.2 eq.) in anhydrous ether (6 ml) at -0 °C. The reaction mixture was allowed to warm up to room temperature, was stirred for additional 1 hr and followed by TLC. Upon completion, the reaction was carefully quenched with dropwise addition of water (1 ml) followed by addition of 15% aq. NaOH (1 ml), water (3 ml), Celite and EtOAc (12 ml). The resulting slurry was filtered through Celite and the filtering bed was washed with EtOAc (2 × 5 ml). The two phases were separated and the organic layer was washed with brine, dried over MgSO₄, filtered and evaporated *in vacuo* to afford the crude product 24 (270 mg, 1.34 mmol, 79%) as a pale yellow oil ($R_f \approx 0.25$; EtOAc/pet. ether 33/67). The product was pure enough and was used without further purification. v_{max} (ATR) 3375, 2971, 2879, 1716, 1491, 1173, 840 cm⁻¹; $\delta_{\rm H}$ (700 MHz, CDCl₃) 5.78 (1H, ddt, *J* 17.2, 10.2, 7.1, 4-*H*), 5.11 (2H, m, 5-*H*₂), 4.72 (1H, br s, N*H*), 3.74–3.58 (3H, m, 1-*H*₂ and 2-*H*), 2.67 (1H, br s, O*H*), 2.31 (1H, m, 3-*H*H), 2.23 (1H, m, 3-HH), 1.43 (9H, s, C(CH₃)₃ $\delta_{\rm C}$ (101 MHz, CDCl₃) 156.56 (NHCOO), 134.28, 118.23, 79.89, 65.54, 52.30, 36.10, 28.49; *m*/z (ES⁺) 224.0 [M+Na]⁺ C₁₀H₁₉O₃N₁²³Na₁ (Expected: 224.1).

Olefin Cross Metathesis

To a solution of Grubbs II (0.3% mol) in CH_2Cl_2 , the starting material was added as a solution in CH_2Cl_2 followed by the addition of the coupling olefin in excess (5 eq.). The reaction mixture was refluxed, typically for 4~8 hrs. The reaction was followed by TLC and quenched by addition of potassium 2-isocyanoacetate (1.32% mol, 4.4 eq. to Grubbs catalyst).¹⁷ The reaction was stirred for 15 min at room temperature then evaporated *in vacuo* to yield the crude mixture which was purified by column chromatography on silica.²

18a tert-Butyl ((2S,3R,E)-3-hydroxyoctadec-4-en-2-yl)carbamate

Yield (52%); v_{max} (ATR) 3375, 2971, 2879, 1716, 1491, 1173, 840 cm⁻¹; δ_{H} (700 MHz, CDCl₃) 5.75–5.66 (1H, m, 5-H), 5.43 (1H, dd, J 15.5, 6.6, 4-H), 4.68–4.60 (1H, m, NH), 4.14–4.06 (1H, m, 3-H), 3.83–3.75 (1H, m, 2-H), 2.07–2.01 (2H, m, 6-H₂), 1.44 (9H, s, C(CH₃)₃), 1.39–1.22 (22H, m, 7-H₂ to 17-H₂), 1.07 (3H, d, J 6.4, 1-H₃), 0.88 (3H, t, J 6.9, 18-H₃); δ_{C} (101 MHz, CDCl₃) 156.3 (NHCOO), 134.2, 128.5, 79.8, 75.9, 32.5, 32.1, 29.9, 29.8, 29.8, 29.7, 29.6, 29.5, 29.3, 28.5, 22.8, 15.6, 14.3; *m*/z (ES⁺) 406.5 [M+Na]⁺; HRMS (ES⁺) found [M+H]⁺ 384.3474, C₂₃H₄₆O₃N₁ requires *M*⁺ 384.3472.



18b tert-Butyl ((2S,3R,E)-3-hydroxynon-4-en-2-yl)carbamate

Yield (67%); v_{max} (ATR) 3375, 2971, 2879, 1716, 1491, 1173, 840 cm⁻¹; δ_{H} (400 MHz, CDCl₃) 5.76–5.65 (1H, m, 5-H), 5.42 (1H, ddt, J 15.4, 6.6, 1.4, 4-H), 4.74–4.61 (1H, m, NH), 4.13–4.05 (1H, m, 3-H), 3.84–3.72 (1H, m, 2-H), 2.01-2.07 (2H, m, 6-H₂), 1.43 (9H, s, C(CH₃)₃), 1.40–1.25 (4H, m, 7-H₂ and 8-H₂), 1.07 (3H, d, J 6.9, 1-H₃), 0.88 (3H, t, J 7.1, 9-H₃); δ_{C} (101 MHz, CDCl₃) 156.4, 134.1, 128.6, 79.7, 75.9, 51.2, 32.1, 31.5, 28.5, 22.3, 15.7, 14.0; m/z (ES⁺) 280.0 [M+Na]⁺; C₁₄H₂₇O₃N₁Na₁; HRMS (ES⁺) found [M+H]⁺ 258.2064, C₁₄H₂₈O₃N₁ requires M^+ 258.2064, [M+Na]⁺ 280.1884, C₁₄H₂₇O₃N₁²³Na₁ requires M^+ 280.1883.



18c tert-Butyl ((2S,3R,E)-3-hydroxy-6-phenylhex-4-en-2-yl)carbamate

Yield (64%) as viscous oil; v_{max} (ATR) 3375, 2971, 2879, 1716, 1491, 1173, 840 cm⁻¹; δ_{H} (400 MHz, CDCl₃) 7.32–7.24 (2H, m, Ar*H*), 7.23–7.14 (3H, m, Ar*H*), 5.93–5.83 (1H, m, 5-*H*), 5.53 (1H, ddt, *J* 15.3, 6.3, 1.4, 4-*H*), 4.74–4.66 (1H, m, N*H*), 4.19–4.13 (1H, m, 3-*H*), 3.86–3.73 (1H, m, 2-*H*), 3.40 (2H, d, *J* 6.8, 6-*H*₂), 1.44 (9H, s, C(CH₃)₃), 1.08 (3H, d, *J* 6.9, 1-*H*₃); δ_{C} (101 MHz, CDCl₃) 156.4, 140.1, 132.2, 130.2, 128.6, 128.9, 126.3, 79.8, 75.6, 51.2, 38.9, 28.5, 15.6; *m*/z (ES⁺) 314.3 [M+Na]⁺; C₁₇H₂₅O₃N₁Na₁; HRMS (ES⁺) found [M+H]⁺ 292.1908, C₁₇H₂₆O₃N₁ requires M^+ 292.1907.



21a tert-Butyl ((2S,3R,E)-3-hydroxynonadec-5-en-2-yl)carbamate

Yield (53%); v_{max} (ATR) 3375, 2971, 2879, 1716, 1491, 1173, 840 cm⁻¹; δ_{H} (700 MHz, CDCl₃) 5.55 (1H, dt, *J* 12.8, 6.7, 6-*H*), 5.43–5.36 (1H, m, 5-*H*), 4.83–4.68 (1H, m, N*H*), 3.73–3.58 (2H, m, 3-*H* and 2-*H*), 3.48 (1H, d, *J* 7.8, O*H*), 2.27–2.15 (4H, m, 4-*H*₂ and 7-*H*₂), 2.14–1.98 (2H, m, 7-*H*₂), 1.44 (9H, s, C(C*H*₃)₃), 1.38–1.22 (94 H, m, 8-*H*₂ to 18-*H*₂ and 1-*H*₃), 0.88 (3H, t, *J* 7.1, 19-*H*₃); δ_{C} (126 MHz, CDCl₃) 156.24, 135.03, 125.50, 79.56, 73.69, 50.35, 37.46, 32.88, 32.15, 29.90, 29.88, 29.85, 29.73, 29.66, 29.58, 29.45, 29.44, 28.63, 22.92, 14.35; *m/z* (ES⁺) 398.5 [M+H]⁺; HRMS (ES⁺) found [M+H]⁺ 398.3627, C₂₄H₄₈O₃N₁ requires *M*⁺ 398.3629; [M+Na]⁺ 420.3446, C₂₄H₄₇O₃N₁²³Na₁ requires *M*⁺ 420.3448.



21b tert-Butyl ((2S,3R,E)-3-hydroxydec-5-en-2-yl)carbamate

Yield (56%); waxy solid; v_{max} (ATR) 3449, 2928, 1715, 1497, 1173, 839 cm⁻¹; δ_{H} (400 MHz, CDCl₃) 5.55 (1H, dt, J 13.2, 6.6, 6-CH), 5.43–5.34 (1H, m, 5-H), 4.83–4.70 (1H, m, NH), 3.74–3.58 (2H, m, 2-H and 3-H), 2.23–1.96 (4H, m, 4-H₂ and 7-H₂), 1.44 (9H, s, C(CH₃)₃), 1.39–1.23 (4H, m, 8-H₂ and 9-H₂), 1.15–1.02 (3H, d, J 6.8, 1-H₃), 0.89 (3H, t, J 7.1, 10-H₃); δ_{C} (101 MHz, CDCl₃) 156.0, 134.8, 125.6, 79.6, 73.5, 50.4, 37.2, 32.3, 31.6, 28.4, 22.2, 13.9; m/z (ES⁺) 294.3 [M+Na]⁺; HRMS (ES⁺) found [M+H]⁺ 272.2220, C₁₅H₃₀O₃N₁ requires M^{+} 272.2220; [M+Na]⁺ 294.2040.



21c tert-Butyl ((2S,3R,E)-3-hydroxy-7-phenylhept-5-en-2-yl)carbamate

Yield (64%) as viscous oil; v_{max} (ATR) 3375, 2971, 2879, 1716, 1491, 1173, 840 cm⁻¹; δ_{H} (700 MHz, CDCl₃) 7.30– 7.27 (2H, m, Ar*H*), 7.21–7.15 (3H, m, Ar*H*), 5.71 (1H, dtd, *J* 15.2, 11.8, 6.6, 6-*H*), 5.53 (1H, dd, *J* 15.2, 6.3, 5-*H*), 4.80 (1H, br s, N*H*), 3.75–3.62 (2H, m, 3-*H* and 2-*H*), 3.36 (2H, d, *J* 6.6, 7-*H*₂), 2.24–2.05 (2H, m, 4-*H*₂), 1.44 (9H, s, C(C*H*₃)₃), 1.10 (3H, d, *J* 6.8, 1-*H*₃); δ_{C} (176 MHz, CDCl₃) 155.88, 140.56, 135.89, 133.24, 128.57, 127.42, 126.16, 79.58, 73.75, 50.37, 39.23, 39.22, 37.17, 28.55, 14.72; *m*/*z* (ES⁺) 328.3 [M+Na]⁺; HRMS (ES⁺) found [M+H]⁺ 306.2063, C₁₈H₂₈O₃N₁ requires *M*⁺ 306.2064; [M+Na]⁺ 328.1883, C₁₈H₂₇O₃N₁²³Na₁ requires *M*⁺ 328.1883.



17a *tert*-Butyl ((2S,3*R*,*E*)-1,3-dihydroxyoctadec-4-en-2-yl)carbamate²

Yield (44%, 50%); v_{max} (ATR) 3410, 2886, 2874, 1687, 1512, 1173 cm⁻¹; δ_{H} (400 MHz, CDCl₃) 5.83–5.73 (1H, m, 5-*H*), 5.53 (1H, ddt, *J* 15.6, 6.5, 1.2, 4-H), 5.30 (1H, br d, *J* 6.4, N*H*), 4.38-4.28 (1H, m, 3-*H*), 3.94 (1H, app. d, *J* 10.0, 1-H*H*), 3.76–3.67 (1H, m, 1-*H*H), 3.63–3.54 (1H, m, 2-*H*), 2.50 (2H, br s, 2 × O*H*), 2.05(2H, q, *J* 7.3, 6-*H*), 1.45 (9H, s, C(C*H*₃)₃), 1.42–1.18 (22H, m, 7-*H*₂ to 17-*H*₂), 0.88 (3H, t, *J* 6.9, 18-*H*₃); δ_{C} (101 MHz, CDCl₃) 158.1, 134.2, 128.9, 79.7, 74.9, 62.9, 56.0, 32.3, 31.9, 29.8, 29.6, 29.5, 29.3, 29.1, 29.1, 28.4, 22.6, 14.1; *m*/z (ES⁺) 400.9 [M+H]⁺; HRMS (ES⁺) found [M+H]⁺ 400.3419, C₂₃H₄₆O₄N₁ requires *M*⁺ 400.3421, [M+Na]⁺ 422.3238, C₂₃H₄₅O₄N₁²³Na₁ requires *M*⁺ 422.3241.



17b *tert*-Butyl ((2S,3*R*,*E*)-1,3-dihydroxynon-4-en-2-yl)carbamate

Yield (59%, 57%); v_{max} (ATR) 3375, 2971, 2879, 1716, 1491, 1173, 840 cm⁻¹; δ_{H} (400 MHz, CDCl₃) 5.76 (1H, dtd, J 15.4, 6.7, 1.1, 5-*H*), 5.51 (1H, ddt, J 15.4, 6.5, 1.3, 4-*H*), 5.32 (1H, br d, N*H*), 4.34–4.21 (1H, m, 3-*H*), 3.90 (1H, d, J 11.4, 1-*H*H), 3.68 (1H, d, J 11.0, 1-H*H*), 3.63–3.51 (1H, m, 2-*H*), 2.97 (2H, br s, 2 × O*H*), 2.05 (2H, dd, J 13.8, 6.7, 6-*H*₂), 1.44 (9H, s, C(C*H*₃)₃), 1.39–1.23 (4H, m, 7-*H*₂ and 8-*H*₂), 0.88 (3H, t, J 7.1, 9-*H*₃); δ_{C} (101 MHz, CDCl₃) 156.4, 134.2, 129.1, 79.9, 74.8, 62.8, 55.6, 32.1, 31.4, 28.5, 22.3, 14.0; *m*/z (ES⁺) 274.6 [M+H]⁺; HRMS (ES⁺) found [M+H]⁺ 274.2012, C₁₄H₂₈O₄N₁ requires *M*⁺ 274.2013, [M+Na]⁺ 296.1831, C₁₄H₂₇O₄N₁²³Na₁ requires *M*⁺ 296.1832.

17c tert-Butyl ((2S,3R,E)-1,3-dihydroxy-6-phenylhex-4-en-2-yl)carbamate

Yield (62%, 43%); v_{max} (ATR) 3375, 2971, 2879, 1716, 1491, 1173, 840 cm⁻¹; δ_{H} (400 MHz, CDCl₃) 7.32–7.26 (2H, m, Ar*H*), 7.23–7.19 (1H, m, Ar*H*), 7.18–7.14 (2H, m, Ar*H*), 6.00–5.90 (1H, m, 5-*H*), 5.61 (1H, dd, *J* 15.4, 6.2, 4-*H*), 5.30 (1H, br s, N*H*), 4.40–4.32 (1H, m, 3-*H*), 3.93 (1H, dd, *J* 11.2, 3.5, 1-H*H*), 3.71 (1H, dd, *J* 11.4, 3.5, 1-*H*H), 3.67–3.58 (1H, m, 2-*H*), 3.40 (2H, d, *J* 6.8, 6-*H*), 2.71–2.44 (2H, br m, 2 × O*H*), 1.45 (9H, s, C(C*H*₃)₃); δ_{C} (125 MHz, CDCl₃) 156.7, 139.3, 132.8, 130.7, 128.66, 128.62, 126.4, 80.7, 74.8, 62.8, 38.8, 28.5; *m*/z (ES⁺) 329.9 [M+Na]⁺; HRMS (ES⁺) found [M+H]⁺ 308.1855, C₁₇H₂₆O₄N₁ requires *M*⁺ 308.1856, [M+Na]⁺ 330.1673, C₁₇H₂₅O₄N₁²³Na₁ requires *M*⁺ 330.1676.

27a (S,E)-tert-Butyl (1-hydroxyoctadec-4-en-2-yl)carbamate¹⁸

Yield (53%); v_{max} (ATR) 3375, 2971, 2879, 1716, 1491, 1173, 840 cm⁻¹; δ_{H} (400 MHz, CDCl₃) 5.52 (1H, dt, *J* 13.4, 6.7, 5-*H*), 5.41–5.27 (1H, m, 4-*H*), 4.66 (1H, br s, N*H*), 3.73–3.48 (3H, m, 1-*H*₂ and 2-*H*), 2.57 (1H, br s, O*H*), 2.33–2.10 (2H, m, 3-*H*₂), 2.00 (2H, dt, *J* 14.0, 6.7, 6-*H*₂), 1.44 (9H, s, C(C*H*₃)₃), 1.39–1.14 (22H, m), 0.88 (3H, t, *J* 6.8, 18-*H*₃); δ_{C} (101 MHz, CDCl₃) 155.98, 134.82, 125.20, 79.85, 65.99, 52.74, 34.89, 32.73, 32.07, 29.84, 29.81, 29.78, 29.66, 29.54, 29.51, 29.32, 28.51, 22.84, 14.28; *m*/z (ES⁺) 384.9 [M+H]⁺; HRMS (ES⁺) found [M+H]⁺ 384.3470, C₂₃H₄₆O₃N₁ requires *M*⁺ 384.2472, [M+Na]⁺ 406.3288, C₂₃H₄₅O₃N₁²³Na₁ requires *M*⁺ 406.3292.



26a (*R*,*E*)-*tert*-Butyl (1-hydroxyoctadec-4-en-2-yl)carbamate¹⁸

Yield (62%); v_{max} (ATR) 3375, 2971, 2879, 1716, 1491, 1173, 840 cm⁻¹; δ_{H} (400 MHz, CDCl₃) 5.52 (1H, dtt, *J* 15.4, 6.6, 1.0, 5-*H*), 5.35 (1H, dtt, *J* 15.4, 7.1, 1.3, 4-*H*), 4.66 (1H, br s, N*H*), 3.70–3.53 (3H, m, 1-*H*₂ and 2-*H*), 2.57 (1H, br s, O*H*), 2.30–2.11 (2H, m, 3-*H*₂), 1.99 (2H, dd, *J* 13.7, 6.7, 6-*H*₂), 1.44 (9H, s, OC(C*H*₃)₃), 1.36–1.22 (22H, m, 7-*H*₂ to 17-*H*₂), 0.88 (3H, t, *J* 6.9, 18-*H*₃); δ_{C} (101 MHz, CDCl₃) 155.98, 134.82, 125.20, 79.85, 65.99, 52.74, 34.89, 32.73, 32.07, 29.84, 29.81, 29.78, 29.66, 29.54, 29.51, 29.32, 28.51, 22.84, 14.28; *m*/z (ES⁺) 384.9 [M+H]⁺; HRMS (ES⁺) found [M+H]⁺ 384.3470, C₂₃H₄₆O₃N₁ requires *M*⁺ 384.2472, [M+Na]⁺ 406.3288, C₂₃H₄₅O₃N₁²³Na₁ requires *M*⁺ 406.3292.

General procedure for BOC de-protection - Acylation Reactions

To a solution of the *N*-Boc protected starting material in CH_2Cl_2 , TFA (excess) was added dropwise at room temperature and the reaction was monitored by TLC. Typically the reaction was complete in less than 1 hr. The reaction mixture was dried on the rotary evaporator to remove the excess TFA. The resulting residue (unprotected amine) was dissolved in CH_2Cl_2 (2 ml) and basified to pH 8 with aq. NaHCO₃ followed by the addition of the corresponding acid chloride (1.2 eq.). The reaction was monitored by TLC. Upon completion, the reaction was diluted with sat. aq. NH₄CI. The phases were separated and the aqueous layer was extracted with twice CH_2Cl_2 . The combined organic extracts were washed with brine, dried on anhydrous MgSO₄ and evaporated to yield the crude product. Assessment of the crude product purity was based on crude ¹H NMR. Compounds that were ≥80% pure were used as they soon as produced. Others were purified by standard column chromatography. All compounds used in the screening were characterised as an array (¹H NMR and Mass Spectrometry data).

18an (2S,3R,E)-2-aminooctadec-4-en-3-ol

Yield (62%); δ_{H} (400 MHz, CDCl₃) 7.80 (2H, br s, NH₂), 5.77 (1H, dt, J 15.2, 6.4, 5-H), 5.37 (1H, dd, J 15.2, 6.1, 4-H), 4.41–4.24 (1H, m, 3-H), 3.40–3.28 (2H, m, 2-H and OH), 2.02 (2H, dd, J 13.7, 6.7, 6-H₂), 1.29–1.21 (25H, m, 7-H₂ to 17-H₂ and 1-H₃), 0.87 (3H, t, J 6.8, 18-H₃); *m/z* (ES⁺) 284.3 [M+H]⁺; HRMS (ES⁺) found [M+H]⁺ 284.2947, C₁₈H₃₈O₃N₁ requires *M*⁺ 284.2948.

18az N-((2S,3R,E)-3-hydroxyoctadec-4-en-2-yl)acetamide

Yield (80%); δ_{H} (400 MHz, CDCl₃) 5.75–5.66 (1H, m, 5-*H*), 5.60 (1H, d, *J* 2.4, N*H*), 5.48–5.35 (1H, m, 4-*H*), 4.12 (1H, dd, *J* 6.7, 3.2, 3-*H*), 4.10–4.04 (1H, m, 2-*H*), 2.11–1.95 (5H, m, 6-*H*₂ and COC*H*₃), 1.40–1.11 (22H, m, 7-*H*₂ to 18-*H*₂), 1.08 (3H, d, *J* 6.9, 1-*H*₃), 0.87 (3H, t, *J* 6.8, 18-*H*₃); *m*/z (ES⁺) 348.4 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 348.2871, C₂₀H₃₉O₂N₁²³Na₁ requires *M*⁺ 348.2873.



18ay N-((2S,3R,E)-3-hydroxyoctadec-4-en-2-yl)-2-phenylacetamide

Yield (22%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 7.39–7.23 (5H, m, Ar*H*), 5.64 (1H, dtd, *J* 15.0, 7.0, 0.9, 5-*H*), 5.51 (1H, br d, N*H*), 5.31 (1H, dd, *J* 15.0, 6.3, 4-*H*), 4.13–4.03 (2H, m, 3-*H* and 2-*H*), 3.58 (2H, s, PhC*H*₂), 1.98 (2H, dd, *J* 13.8, 7.0, 6-*H*₂), 1.39-1.18 (22H, m, 7-*H*₂ to 17-*H*₂), 1.01 (3H, d, *J* 6.8, 1-*H*₃), 0.88 (3H, t, *J* 6.9, 18-*H*₃); *m*/z (ES⁺) 424.5 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 424.3182, C₂₆H₄₃O₂N₁²³Na₁ requires *M*⁺ 424.3186.

Page 19 of 50



18ax N-((2S,3R,E)-3-hydroxyoctadec-4-en-2-yl)octanamide

Yield (41%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 5.71 (1H, dtd, *J* 14.9, 6.7, 1.1, 5-*H*), 5.43 (1H, ddt, *J* 15.4, 6.6, 1.4, 4-*H*), 4.64 (1H, br s, N*H*), 4.14–4.06 (1H, m, 3-*H*), 3.85–3.72 (1H, m, 2-*H*), 2.56 (1H, br s, O*H*), 2.04 (2H, dd, *J* 14.9, 7.0, 6-*H*₂), 1.40–1.20 (22 H, m, 7-*H*₂ to 17-*H*₂), 1.07 (3H, d, *J* 6.9, 1-*H*₃), 0.88 (3H, t, *J* 6.9, 18-*H*₃); *m*/z (ES⁺) 432.6 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 432.3806, C₂₆H₅₁O₂N₁²³Na₁ requires *M*⁺ 432.3812.



18bn (2*S*,3*R*,*E*)-2-aminonon-4-en-3-ol

Yield (73%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 5.79–5.69 (1H, m, 5-*H*), 5.39 (1H, dd, *J* 14.9, 5.3, 4-*H*), 5.30 (2H, br s, N*H*₂), 4.32–4.24 (1H, m, 3-*H*), 3.33–3.21 (1H, m, 2-*H*), 2.03 (2H, dd, *J* 12.8, 5.2, 6-*H*₂), 1.41–1.26 (4H, m, 7-*H*₂ and 8-*H*₂), 1.15 (1H, d, *J* 6.5, 1-*H*₃), 0.88 (1H, t, *J* 6.9, 9-*H*₃); *m*/z (ES⁺) 158.1 [M+H]⁺; HRMS (ES⁺) found [M+Na]⁺ 158.1539, C₉H₁₉O₁N₁²³Na₁ requires *M*⁺ 158.1539.



18bz N-((2S,3R,E)-3-hydroxynon-4-en-2-yl)acetamide

Yield (79%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 6.03 (1H, br s, N*H*), 5.74–5.62 (1H, m, 5-*H*), 5.41 (1H, dd, *J* 15.3, 6.4, 4-*H*), 4.16–4.00 (1H, m, 3-*H* and 2-*H*), 3.73 (1H, s, O*H*), 2.14–1.87 (5H, m, 6-*H*₂ and COC*H*₃), 1.44–1.25 (4H, m, 7-*H*₂ and 8-*H*₂), 1.06 (1H, d, *J* 6.8, 1-*H*₃), 0.88 (3H, t, *J* 6.8, 9-*H*₃); *m*/z (ES⁺) 222.2 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 222.1463, C₁₁H₂₁O₂N₁²³Na₁ requires *M*⁺ 222.1465.



18by N-((2S,3R,E)-3-hydroxynon-4-en-2-yl)-2-phenylacetamide

Yield (50%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 7.35–7.22 (5H, m, Ar*H*), 5.69–5.59 (1H, m, 5-*H*), 5.32 (1H, dd, *J* 15.9, 6.3, 4-*H*), 4.19–3.98 (2H, m, 3-*H* and 2-*H*), 3.56 (2H, s, PhC*H*₂), 1.99 (2H, dd, *J* 13.3, 6.6, 6-*H*₂), 1.34–1.21 (2H, m, 7-*H*₂ and 8-*H*₂), 1.01 (1H, d, *J* 6.6, 1-*H*₃), 0.89 (1H, t, *J* 6.5, 9-*H*₃); *m*/z (ES⁺) 298.3 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 298.1775, C₁₇H₂₅O₂N₁²³Na₁ requires *M*⁺ 298.1778.



HCO C-F

18bx N-((2S,3R,E)-3-hydroxynon-4-en-2-yl)octanamide

Yield (23%); δ_H (400 MHz, CDCl₃) 5.76–5.67 (1H, m, 5-H), 5.63 (1H, br d, J 7.7, NH), 5.41 (1H, dd, J 15.5, 6.3, 4-H), 4.11 (2H, m, 3-H and 2-H), 2.20-2.15 (2H, m, COCH₂), 2.10-2.02 (2H, m, 6-H₂), 1.66-1.58 (2H, m, COCH₂CH₂), 1.39–1.22 (12H, m, 7-H₂, 8-H₂ and 4 × octanoyl aliphatic CH₂), 1.09 (3H, d, J 6.8, 1-H₃), 0.92–0.85 (6H, m, 9- H_3 and octanoyl terminal C H_3); m/z (ES⁺) 306.4 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 306.2402, $C_{17}H_{33}O_2N_1^{23}Na_1$ requires M^+ 306.2404.



18cn (2S,3R,E)-2-amino-6-phenylhex-4-en-3-ol

Yield (77%); δ_H (400 MHz, CDCl₃) 7.85 (2H, br s, NH₂), 7.29–7.23 (2H, m, ArH), 7.20–7.16 (1H, m, ArH), 7.14–7.11 (2H, m, ArH), 5.97–5.88 (1H, m, 5-H), 5.45 (1H, dd, J 15.5, 6.2, 4-H), 4.48–4.39 (1H, m, 3-H), 3.43–3.37 (1H, m, 2-*H*), 3.35 (2H, d, *J* 7.7, 6-*H*₂), 2.89 (1H, br s, O*H*), 1.19 (3H, d, *J* 6.5, 1-*H*₃); *m*/z (ES⁺) 192.2 [M+H]⁺; HRMS (ES⁺) found $[M+H]^+$ 192.1383, $C_{12}H_{18}O_1N_1$ requires M^+ 192.1383.



N-((2S,3R,E)-3-hydroxy-6-phenylhex-4-en-2-yl)acetamide 18cz

Yield (67%); δ_H (400 MHz, CDCl₃) 7.31–7.27 (2H, m, ArH), 7.22–7.14 (3H, m, ArH), 5.98 (1H, br s, NH),5.93–5.82 (1H, m, 5-H), 5.50 (1H, dd, J 15.3, 6.2, 4-H), 4.21-4.13 (1H, m, 3-H), 4.06 (1H, dt, J 8.0, 4.0, 2-H), 3.38 (1H, d, J 6.7, 6-H₂), 2.85 (2H, br s, OH), 1.94 (3H, s, COCH₃), 1.08 (3H, d, J 6.9, 1-H₃); m/z (ES⁺) 256.3 [M+Na]⁺; HRMS (ES^{+}) found $[\text{M}+\text{Na}]^{+}$ 256.1307, $C_{14}H_{19}O_2N_1^{-23}\text{Na}_1$ requires M^{+} 256.1308.



N-((2S,3R,E)-3-hydroxy-6-phenylhex-4-en-2-yl)-2-phenylacetamide 18cv

Yield (87%); δ_H (400 MHz, CDCl₃) 7.35–7.10 (10H, m, ArH), 5.91 (1H, br s, NH), 5.86–5.77 (1H, m, 5-H), 5.40 (1H, dd, J 15.4, 6.0, 4-H), 4.18–3.96 (1H, m, 3-H), 3.60–3.49 (1H, m, 2-H), 3.42 (2H, s, COCH₂), 3.32 (2H, d, J 6.7, 6- H_2), 1.01 (3H, d, J 6.8, 1- H_3); m/z (ES⁺) 332.3 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 332.1618, C₂₀H₂₃O₂N₁²³Na₁ requires *M*⁺ 332.1621.

. NHC0 C₇H₁₅

18cx N-((2S,3R,E)-3-hydroxy-6-phenylhex-4-en-2-yl)octanamide

Yield (90%); δ_{H} (400 MHz, CDCl₃) 7.31–7.24 (2H, m, Ar*H*), 7.22–7.12 (3H, m, Ar*H*), 5.95–5.82 (2H, m, 5-*H* and N*H*), 5.50 (1H, dd, *J* 15.4, 6.2, 4-*H*), 4.20–4.03 (2H, m, 3-*H* and 2-*H*), 3.38 (1H, d, *J* 6.7, 6-*H*₂), 2.17–2.10 (2H, m, COC*H*₂), 1.66–1.51 (2H, m, COCH₂C*H*₂), 1.32–1.20 (8H, m, 4 × C*H*₂), 1.08 (3H, d, *J* 6.8, 1-*H*₃), 0.87 (3H, t, *J* 6.8, octanoyl terminal C*H*₃); *m*/z (ES⁺) 318.2 [M+H]⁺, 340.4 [M+Na]⁺; HRMS (ES⁺) found [M+H]⁺ 318.2431, C₂₀H₃₂O₂N₁ requires *M*⁺ 318.2428.



21bn (2S,3R,E)-2-aminodec-5-en-3-ol

Yield (79%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 5.62–5.45 (1H, m, 6-*H*), 5.41–5.26 (1H, m, 5-*H*), 5.14 (1H, br s, N*H*₂), 3.83–3.71 (1H, m, 2-*H*), 3.31–3.13 (1H, m, 3-*H*), 2.27–1.93 (4H, m, 4-*H*₂ and 7-*H*₂), 1.33–1.29 (4H, m, 8-*H*₂ and 9-*H*₂), 1.16 (3H, d, *J* 6.1, 1-*H*₃), 0.87 (3H, t, *J* 7.0, 10-*H*₃); *m*/z (ES⁺) 172.3 [M+H]⁺, 194.3 [M+Na]⁺; HRMS (ES⁺) found [M+H]⁺ 172.1696, C₁₀H₂₂O₁N₁ requires *M*⁺ 172.1696.



21bz N-((2S,3R,E)-3-hydroxydec-5-en-2-yl)acetamide

Yield (70%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 5.88 (1H, br s, N*H*), 5.55 (1H, dt, *J* 13.4, 6.6, 6-*H*), 5.44–5.34 (1H, m, 5-*H*), 4.01 (1H, ddd, *J* 8.5, 6.8, 3.0, 2-*H*), 3.64 (1H, ddt, *J* 8.5, 4.2, 3.1, 3-*H*), 2.11–1.99 (4H, m, 4-*H*₂ and 7-*H*₂), 1.98 (3H, s, COC*H*₃), 1.38–1.25 (4H, m, 8-*H*₂ and 9-*H*₂), 1.10 (3H, d, *J* 6.8, 1-*H*₃), 0.88 (3H, t, *J* 7.1, 10-*H*₃); *m*/z (ES⁺) 236.3 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 236.1620, C₁₂H₂₃O₂N₁²³Na₁ requires *M*⁺ 236.1621.



21by N-((2S,3R,E)-3-hydroxydec-5-en-2-yl)-2-phenylacetamide

Yield (85%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 7.40–7.22 (5H, m, Ar*H*), 5.73 (1H, br s, N*H*), 5.50 (1H, dt, *J* 15.2, 6.6, 6-*H*), 5.35 (1H, ddd, *J* 15.2, 7.7, 6.3, 5-*H*), 3.99 (1H, ddd, *J* 8.5, 6.9, 3.1, 2-*H*), 3.56 (1H, m, 3-*H* and PhC*H*₂), 2.22–2.08 (2H, m, 4-*H*₂), 2.06–1.92 (2H, m, 7-*H*₂), 1.34–1.28 (4H, m, 8-*H*₂ and 9-*H*₂), 1.04 (3H, d, *J* 6.9, 1-*H*₃), 0.88 (3H, t, *J* 7.1, 10-*H*₃); *m*/z (ES⁺) 312.4 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 312.1935, C₁₈H₂₇O₂N₁²³Na₁ requires *M*⁺ 312.1934.



[≣]NHC0 C₇H₁₅

21bx *N*-((2*S*,3*R*,*E*)-3-hydroxydec-5-en-2-yl)octanamide

Yield (83%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 6.03-5.98 (1H, br s, N*H*), 5.54–5.43 (1H, m, 6-*H*), 5.35–5.25 (1H, m, 5-*H*), 4.23–4.11 (1H, m, 2-*H*), 4.05–3.98 (1H, m, 3-*H*), 2.35–2.27 (2H, m, COC*H*₂), 2.20–2.11 (2H, m, 4-*H*₂), 2.06–1.94 (2H, m, 7-*H*₂), 1.68–1.50 (2H, m, COCH₂C*H*₂), 1.39–1.19 (12H, m, 8-*H*₂, 9-*H*₂ and 4 × aliphatic C*H*₂), 1.14–1.05 (3H, d, *J* 6.9, 1-*H*₃), 0.86 (6H, 10-*H*₃ and octanoyl terminal C*H*₃); *m*/z (ES⁺) 298.5 [M+H]⁺, 320.5 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 320.2563, C₁₈H₃₅O₂N₁²³Na₁ requires *M*⁺ 320.2560.



21cn (2S,3R,E)-2-amino-7-phenylhept-5-en-3-ol

Yield (46%); $\delta_{\rm H}$ (700 MHz, CDCl₃) 7.88 (2H, br s, NH₂), 7.20–7.11 (5H, m, ArH), 5.71–5.65 (1H, m, 6-H), 5.45–5.40 (1H, m, 5-H), 3.93–3.89 (1H, m, 3-H), 3.36 (1H, d, J 7.7, 2-H), 3.32 (2H, d, J 6.7, 7-H₂), 2.28–2.09 (2H, m, 4-H₂), 1.20 (3H, d, J 6.3, 1-H₃). *m*/z (ES⁺) 206.3 [M+H]⁺, 228.3 [M+Na]⁺; HRMS (ES⁺) found [M+H]⁺ 206.1540, C₁₃H₂₀O₁N₁ requires *M*⁺ 206.1539.



21cz N-((2S,3R,E)-3-hydroxy-7-phenylhept-5-en-2-yl)acetamide

Yield (86%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 7.34–7.24 (2H, m, Ar*H*), 7.24-7.15 (3H, m, Ar*H*), 5.79 (1H, br s, N*H*), 5.73 (1H, ddd, *J* 15.2, 8.4, 4.3, 6-*H*), 5.52 (1H, ddd, *J* 15.2, 7.7, 6.4, 5-*H*), 4.03 (1H, dqd, *J* 13.8, 6.9, 3.0, 2-*H*), 3.72–3.65 (1H, m, 3-*H*), 3.37 (1H, d, *J* 6.7, 7-*H*₂), 2.30–2.04 (2H, m, 4-*H*₂), 1.98 (3H, s. COC*H*₃), 1.11 (3H, d, *J* 6.9, 1-*H*₃); *m*/z (ES⁺) 248.4 [M+H]⁺, 270.3 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 270.1464, C₁₅H₂₁O₂N₁²³Na₁ requires *M*⁺ 270.1465.



21cy N-((2S,3R,E)-3-hydroxy-7-phenylhept-5-en-2-yl)-2-phenylacetamide

Yield (27%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 7.43–7.08 (10H, m, Ar*H*), 5.78–5.59 (1H, m, 6-*H*), 5.57–5.37 (1H, m, 5-*H*), 4.07–3.93 (1H, m, 2-*H*), 3.64–3.50 (3H, m, 2-*H* and PhC*H*₂), 3.34 (1H, d, *J* 6.7, 7-*H*₂), 2.25–1.93 (2H, m, 4-*H*₂), 1.03 (3H, d, *J* 6.8, 1-*H*₃); *m*/z (ES⁺) 346.4 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 346.1779, C₂₁H₂₅O₂N₁²³Na₁ requires *M*⁺ 346.1778.

Ρh

[™]_NHC0 C₇H₁₅

21cx N-((2S,3R,E)-3-hydroxy-7-phenylhept-5-en-2-yl)octanamide

Yield (70%); δ_{H} (400 MHz, CDCl₃) 7.31–7.27 (2H, m, Ar*H*), 7.23–7.15 (3H, m, Ar*H*), 5.87–5.65 (1H, m, N*H* and 6-*H*), 5.52 (1H, dt, *J* 14.2, 6.6, 5-*H*), 4.13-3.94 (1H, m, 2-*H*), 3.78–3.59 (1H, m, 3-*H*), 3.37(1H, d, *J* 6.6, 7-*H*₂), 2.32–2.04 (2H, m, COC*H*₂), 1.70–1.54 (2H, m, COC*H*₂C*H*₂), 1.37–1.21 (8H, m, 4 × aliphatic C*H*₂), 1.13 (3H, d, *J* 6.9, 1-*H*₃), 0.87 (3H, t, *J* 6.6, octanoyl terminal C*H*₃); *m*/z (ES⁺) 332.5 [M+H]⁺, 354.4 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 354.2403, C₂₁H₃₃O₂N₁²³Na₁ requires *M*⁺ 354.2404.



21an (2S,3R,E)-2-aminononadec-5-en-3-ol

Yield (70%); δ_{H} (700 MHz, CDCl₃) 8.12 (2H, br s, NH₂), 5.56–5.48 (1H, m, 6-*H*), 5.38 (1H, dd, *J* 14.0, 6.7, 5-*H*), 3.73–3.67 (1H, m, 3-*H*), 3.65–3.60 (1H, m, 2-*H*), 2.15–2.07 (1H, m, 4-*H*H), 2.04–1.95 (3H, m, 4-H*H* and 7-*H*₂), 1.38–1.09 (26H, m, 8-*H*₂ to 18-*H*₂ and 1-*H*₃), 0.86 (3H, t, *J* 7.0, 19-*H*₃). *m/z* (ES⁺) 298.4 [M+H]⁺; HRMS (ES⁺) found [M+H]⁺ 298.3099, C₁₉H₄₀O₁N₁ requires *M*⁺ 298.3110.



21az N-((2S,3R,E)-3-hydroxynonadec-5-en-2-yl)acetamide

Yield (82%); δ_{H} (400 MHz, CDCl₃) 5.96 (1H, br d, *J* 6.0, N*H*), 5.54 (1H, dt, *J* 15.0, 6.6, 6-*H*), 5.45–5.34 (1H, m, 5-*H*), 4.00 (1 H, ddd, *J* 8.5, 6.9, 3.0, 3-*H*), 3.64 (1H, ddd, *J* 8.8, 4.4, 3.0, 2-*H*), 2.23–1.99 (4 H, m, 7-*H*₂ and 4-*H*₂), 1.97 (3H, s, COC*H*₃), 1.38–1.21 (22H, m, 8-*H*₂ to 18-*H*₂), 1.10 (3H, d, *J* 6.8, 1-*H*₃), 0.87 (3H, t, *J* 6.9, 19-*H*₃). *m/z* (ES⁻) 338.4 [M–H]⁻; (ES⁺) 362.4.3 [M+Na]⁺; HRMS (ES⁺) found [M+H]⁺ 340.3219, C₂₁H₄₂O₂N₁ requires *M*⁺ 340.3216.



21ay N-((2S,3R,E)-3-hydroxynonadec-5-en-2-yl)-2-phenylacetamide

Yield (86%); δ_{H} (400 MHz, CDCl₃) 7.37–7.32 (2H, m, Ar*H*), 7.31–7.24 (3H, m, Ar*H*), 5.74 (1H, br d, *J* 8.3, N*H*), 5.50 (1H, dt, *J* 14.7, 6.6, 6-*H*), 5.34 (1H, ddd, *J* 15.2, 7.6, 6.4, 5-*H*), 3.99 (1H, ddd, *J* 8.4, 6.9, 3.1, 3-*H*), 3.73–3.64 (1H, m, 2-*H*), 3.55 (2H, s, PhC*H*₂), 2.15–2.08 (2H, m, 4-*H*₂), 2.06–1.94 (2H, m, 7-*H*₂), 1.36–1.22 (22H, m, 8-*H*₂ to 18-*H*₂), 1.04 (3H, d, *J* 6.8, 1-*H*₂), 0.88 (3H, t, *J* 6.9, 19-*H*₃). *m*/*z* (ES⁻) 414.4 [M-H]⁻; (ES⁺) 438.4 [M+Na]⁺; HRMS (ES⁺) found [M+H]⁺ 416.3533, C₂₇H₄₆O₂N₁ requires *M*⁺ 416.3529.

C13H27

. NHCOC7H15

21ax N-((2S,3R,E)-3-hydroxynonadec-5-en-2-yl)octanamide

Yield (85%); δ_{H} (400 MHz, CDCl₃) 5.85 (1H, br d, *J* 8.4, N*H*), 5.55 (1H, dt, *J* 14.9, 6.6, 6-*H*), 5.39 (1H, ddd, *J* 15.0, 7.5, 6.4, 5-*H*), 4.03 (1H, ddd, *J* 8.7, 7.0, 3.0, 3-*H*), 3.64 (1H, ddd, *J* 8.8, 4.1, 3.0, 2-*H*), 2.33 (2H, t, *J* 7.5, COC*H*₂), 2.19–2.14 (2H, m, 4-*H*₂), 2.00 (2H, dd, *J* 13.7, 6.6, 7-*H*₂), 1.62 (2H, dd, *J* 13.3, 6.0, COCH2C*H*₂), 1.34–1.22 (26H, m, 8-*H*₂ to 18-*H*₂ and 4 × aliphatic C*H*₂), 1.10 (3H, d, *J* 6.8, 1-*H*₃), 0.87 (3H, t, *J* 6.7, 19-*H*₃). *m/z* (ES⁻) 422. [M-H]⁻; (ES⁺) 446.4 [M+Na]⁺; HRMS (ES⁺) found [M+H]⁺ 424.4174, C₂₇H₅₄O₂N₁ requires *M*⁺ 424.4155.



17bn (2S,3R,E)-2-aminonon-4-ene-1,3-diol

Yield (54%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 5.80–5.71 (1H, m, 5-*H*), 5.42 (1H, dd, *J* 15.2, 6.7, 4-*H*), 4.31 (4H, br s, N*H*₂ and 2 × O*H*), 4.25–4.19 (1H, m, 3-*H*), 3.78–3.63 (2H, m, 2-*H* and 1-*H*H), 3.06 (1H, dd, *J* 8.7, 4.2, 1-H*H*), 2.04 (2H, dd, *J* 13.1, 6.5, 6-*H*₂), 1.39–1.26 (4H, m, 7-*H*₂ and 8-*H*₂), 0.88 (3H, t, *J* 7.0, 9-*H*₃); *m*/z (ES⁺) 174.3 [M+H]⁺, 196.4 [M+Na]⁺; HRMS (ES⁺) found [M+H]⁺ 174.1490, C₉H₂₀O₂N₁ requires *M*⁺ 174.1489.



17bz N-((2S,3R,E)-1,3-dihydroxynon-4-en-2-yl)acetamide

Yield (73%); δ_{H} (700 MHz, CDCl₃) 6.44 (1H, br d, *J* 13.7, N*H*), 5.82–5.76 (1H, m, 5-*H*), 5.41 (1H, dd, *J* 15.1, 5.7, 4-*H*), 4.47–4.39 (1H, m, 3-*H*), 3.87–3.77 (2H, m, 2-*H* and 1-*H*H), 3.35–3.29 (1H, m, 1-H*H*), 2.09–2.00 (5H, m, 6-*H*₂ and COC*H*₃), 1.41 – 1.23 (4H, m, 7-*H*₂ and 8-*H*₂), 0.88 (3H, t, *J* 7.3, 9-*H*₃). *m*/z (ES⁺) 238.2 [M+Na]⁺ C₁₁H₂₁O₃N₁²³Na₁; HRMS (ES⁺) found [M+Na]⁺ 238.1419, C₁₁H₂₁O₃N₁²³Na₁ requires *M*⁺ 238.1419.

17by N-((2S,3R,E)-1,3-dihydroxynon-4-en-2-yl)-2-phenylacetamide

Yield (65%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 7.35–7.23 (5H, m, Ar*H*), 6.34 (1H, br d, *J* 6.6, N*H*), 5.66 (1H, dt, *J* 14.9, 6.7, 5-*H*), 5.40 (1H, dd, *J* 14.9, 6.5, 4-*H*), 4.23–4.16 (1H, m, 3-*H*), 3.88–3.79 (2H, m, 2-*H* and 1-*H*H), 3.61 (1H, dd, *J* 10.9, 2.7, 1-H*H*), 3.56 (2H, s, PhC*H*₂), 1.98 (1H, dd, *J* 13.5, 6.8, 6-*H*₂), 1.32–1.25 (4H, m, 7-*H*₂ and 8-*H*₂), 0.88 (3H, t, *J* 7.0, 9-*H*₃); *m*/z (ES⁺) 292.2 [M+H]⁺, 314.2 [M+Na]⁺; HRMS (ES⁺) found [M+H]⁺ 292.1907, C₁₇H₂₆O₃N₁ requires *M*⁺ 292.1913.

$\bar{N}HCOC_7H_{15}$

17bx *N*-((2*S*,3*R*,*E*)-1,3-dihydroxynon-4-en-2-yl)octanamide

Yield (68%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 6.44 (1H, br s, N*H*), 5.74 (1H, dt, *J* 15.9, 6.7, 5-*H*), 5.49 (1H, dd, *J* 15.9, 6.0, 4-*H*), 4.30–4.21 (1H, m, 3-*H*), 3.94–3.84 (2H, m, 2-*H* and 1-*H*H), 3.71–3.63 (1H, m, 1-H*H*), 2.24–2.16 (2H, m, COC*H*₂), 2.04 (2H, dd, *J* 13.0, 6.7, 6-*H*₂), 1.64–1.55 (2H, m, COCH₂C*H*₂), 1.34–1.21 (4H, m, 7-*H*₂ and 8-*H*₂), 0.87, 0.86 (2 × 3H, t, *J* 6.5, 9-*H*₃ and octanoyl terminal C*H*₃); *m*/z (ES⁺) 300.3 [M+H]⁺; HRMS (ES⁺) found [M+H]⁺ 300.2536, C₁₇H₃₄O₃N₁ requires *M*⁺ 300.2539.



17an (2S,3R,E)-2-aminooctadec-4-ene-1,3-diol²

Yield (64%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 5.87–5.79 (1H, m, 5-*H*), 5.53 (1H, dd, *J* 15.4, 6.5, 4-*H*), 4.78 (2H, br s, NH₂), 4.41–4.36 (1H, m, 3-*H*), 4.10 (1H, dd, *J* 11.5, 2.7, 1-*H*H), 3.93 (1H, m, 2-*H*), 3.73 (1H, dd, *J* 11.5, 3.5, 1-H*H*), 2.77 (2H, br s, 2 × O*H*), 2.06 (2H, dd, *J* 14.0, 7.0, 6-*H*₂), 1.67 (1 H, dq, *J* 12.2, 6.0, 17-*H*₂), 1.48–1.22 (20H, m, 7-*H*₂ to 16-*H*₂), 0.89 (1H, t, *J* 7.5, 18-*H*₃); *m*/z (ES⁺) 300.5 [M+H]⁺, 322.5 [M+Na]⁺; HRMS (ES⁺) found [M+H]⁺ 300.2902, C₁₈H₃₈O₂N₁ requires *M*⁺ 300.2897.



17az N-((2S,3R,E)-1,3-dihydroxyoctadec-4-en-2-yl)acetamide

Yield (71%); δ_{H} (400 MHz, CDCl₃) 5.81 (1H, dt, *J* 14.6, 7.1, 5-*H*), 5.51 (1H, dd, *J* 15.4, 6.4, 4-*H*), 5.39 (1H, br s, N*H*), 4.38–4.32 (1H, m, 3-*H*), 4.07(1 H, dd, *J* 11.6, 2.7, 2-*H*), 3.83–3.78 (1H, m, 1-H*H*), 3.73 (1H, dd, *J* 11.6, 2.9, 1-*H*H), 2.09–1.98 (5H, m, 6-*H*₂ and COC*H*₃), 1.39–1.20 (22H, m, 7-*H*₂ to 17-*H*₂), 0.87 (1H, t, *J* 6.7, 18-*H*₃); *m*/z (ES⁺) 364.4 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 364.2829, C₂₀H₃₉O₃N₁²³N₁ requires *M*⁺ 364.2822.



17ay N-((2S,3R,E)-1,3-dihydroxyoctadec-4-en-2-yl)-2-phenylacetamide

Yield (44%); δ_{H} (400 MHz, CDCl₃) 7.38–7.25 (5H, m, Ar*H*), 6.21 (1H, br d, *J* 7.0, N*H*), 5.73–5.63 (1H, m, 5-*H*), 5.42 (1H, dd, *J* 15.4, 6.5, 4-*H*), 4.27–4.20 (1H, m, 3-*H*), 3.90–3.83 (2H, m, 2-*H* and 1-*H*H), 3.65 (1H, dd, *J* 11.8, 3.9, 1-H*H*), 3.60 (2H, s, PhC*H*₂), 2.97–2.64 (2H, m, 2 × O*H*), 1.99 (2H, dd, *J* 13.8, 6.9, 6-*H*₂), 1.34–1.16 (22H, m, 7-*H*₂ to 17-*H*₂), 0.88 (1H, t, *J* 6.8, 18-*H*₃); *m*/z (ES⁺) 440.5 [M+Na]⁺; HRMS (ES⁺) found [M+H]⁺ 418.3324, C₂₆H₄₄O₃N₁ requires *M*⁺ 418.3316.

17ax N-((2S,3R,E)-1,3-dihydroxyoctadec-4-en-2-yl)octanamide

Yield (38%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 6.25 (1H, br d, *J* 7.8, N*H*), 5.84–5.74 (1H, m, 5-*H*), 5.53 (1H, dd, *J* 15.5, 6.5, 4-*H*), 4.36–4.30 (1H, m, 3-*H*), 3.96 (1H, dd, *J* 11.1, 3.7, 1-H*H*), 3.93–3.88 (1 H, m, 2-*H*), 3.70 (1H, dd, *J* 11.1, 2.9, 1-*H*H), 2.71 (2H, br s, 2 × O*H*), 2.27–2.20 (2H, m, COC*H*₂), 2.06 (2H, dd, *J* 14.6, 7.4, 6-*H*₂), 1.71–1.57 (2H, m, COCH₂C*H*₂), 1.43–1.18 (28H, m, 7-*H*₂ to 17-*H*₂ and 4'-*H*₂ to 7'-*H*₂), 0.92–0.83 (6H, m, 18-*H*₃ and octanoyl terminal C*H*₃); *m*/z (ES⁺) 426.4 [M+H]⁺; HRMS (ES⁺) found [M+Na]⁺ 448.3761, C₂₆H₄₄O₃N₁ requires *M*⁺ 448.3767.



17cn (2S,3R,E)-2-amino-6-phenylhex-4-ene-1,3-diol

Yield (67%); δ_{H} (400 MHz, CDCl₃) 7.25–7.20 (2H, m, Ar*H*), 7.16–7.07 (3H, m, Ar*H*), 5.88 (1H, dt, *J* 15.0, 6.6, 5-*H*), 5.42 (1H, dd, *J* 15.0, 5.0, 4-*H*), 4.87 (2H, br s, N*H*₂), 4.40–4.29 (1H, m, 3-*H*), 3.76–3.67 (2H, m, 3-*H* and 1-*H*H), 3.41–3.33 (1H, m, 1-H*H*), 3.30 (2H, d, *J* 6.6, 6-*H*₂), 3.21–3.14 (2H, br s, 2 × O*H*); *m*/z (ES⁺) 208.2 [M+H]⁺; HRMS (ES⁺) found [M+H]⁺ 208.1331, C₁₂H₁₈O₂N₁ requires *M*⁺ 208.1338.



17cz N-((2S,3R,E)-1,3-dihydroxy-6-phenylhex-4-en-2-yl)acetamide

Yield (31%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 7.32–7.27 (2H, m, Ar*H*), 7.23–7.14 (3H, m, Ar*H*), 6.32 (1H, br d, N*H*), 5.96 (1H, dt, *J* 15.4, 6.7, 5-*H*), 5.61 (1H, dd, *J* 15.4, 6.0, 4-*H*), 4.38–4.32 (1H, m, 3-*H*), 3.99–3.88 (2H, m, 2-*H* and 1-*H*H), 3.70 (1H, dd, *J* 11.0, 2.8, 1-H*H*), 3.40 (1H, d, *J* 6.7, 6-*H*₂), 2.01 (3H, s, COC*H*₃); *m*/z (ES⁺) 250.1 [M+H]⁺, 272.1 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 272.1248, C₁₄H₁₉O₃N₁²³Na₁ requires *M*⁺ 272.1263.



17cy N-((2S,3R,E)-1,3-dihydroxy-6-phenylhex-4-en-2-yl)-2-phenylacetamide

Yield (85%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 7.34–7.25 (6H, m, Ar*H*), 7.22–7.11 (4H, m, Ar*H*), 6.32 (1H, br d, *J* 7.5, N*H*), 5.82 (1H, dt, *J* 15.0, 6.8, 5-*H*), 5.47 (1H, dd, *J* 15.0, 5.8, 4-*H*), 4.24–4.18 (1H, m, 3-*H*), 3.89–3.84 (1H, m, 2-*H*), 3.81 (1H, dd, *J* 11.1, 4.0, 1-*H*H), 3.59 (1H, dd, *J* 11.1, 3.2, 1-H*H*), 3.51 (2H, s, COC*H*₂), 3.31 (2H, d, *J* 6.8, 6-*H*₂), 2.25 (2H, br s, 2 × O*H*); *m*/z (ES⁺) 326.2 [M+H]⁺; HRMS (ES⁺) found [M+H]⁺ 326.1767, C₂₀H₂₄O₃N₁ requires *M*⁺ 326.1756.

. ŇHC0 C₇H₁₅

17cx N-((2S,3R,E)-1,3-dihydroxy-6-phenylhex-4-en-2-yl)octanamide

Yield (82%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 7.31–7.23 (3H, m, Ar*H*), 7.22–7.18 (1H, m, Ar*H*), 7.17–7.11 (2H, m, Ar*H*), 6.41 (1H, br d, *J* 6.6, N*H*), 5.92 (1H, dt, *J* 15.3, 6.8, 5-*H*), 5.58 (1H, dd, *J* 15.3, 6.1, 4-*H*), 4.33–4.27 (1H, m, 3-*H*), 3.89 (2H, m, 2-*H* and 1-*H*H), 3.66 (1H, dd, *J* 12.6, 4.7, 1-H*H*), 3.38 (2H, d, *J* 6.8, 6-*H*₂), 2.23–2.10 (2H, m, COC*H*₂), 1.63–1.53 (2H, m, COCH₂C*H*₂), 1.32–1.22 (8H, m, 4 × aliphatic C*H*₂), 0.87 (3H, t, *J* 6.8, octanoyl terminal C*H*₃); *m*/z (ES⁺) 334.2 [M+H]⁺; HRMS (ES⁺) found [M+Na]⁺ 334.2393, C₂₀H₃₂O₃N₁ requires *M*⁺ 334.2382.



27an (S,E)-2-aminooctadec-4-en-1-ol

Yield (61%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 5.51 (1H, dt, *J* 13.9, 6.5, 5-*H*), 5.38–5.30 (1H, m, 4-*H*), 3.60 (1H, d, *J* 8.2, 1-H*H*), 3.33 (1H, dd, *J* 10.2, 8.2, 1-*H*H), 2.97-2.83 (1H, m, 2-*H*), 2.34 (3H, br s, N*H*₂ and O*H*), 2.20–2.10 (1H, m, 3-*H*₂), 1.99 (2H, dd, *J* 13.5, 6.6, 6-*H*₂), 1.37–1.20 (22H, m, 7-*H*₂ to 17-*H*₂), 0.87 (3H, t, *J* 6.5, 18-*H*₃); *m*/z (ES⁺) 284.2 [M+H]⁺; HRMS (ES⁺) found [M+H]⁺ 284.2947, C₁₈H₃₈O₁N₁ requires *M*⁺ 284.2953.



27az (S,E)-N-(1-hydroxyoctadec-4-en-2-yl)acetamide

Yield (68%); $\delta_{\rm H}$ (700 MHz, CDCl₃) 5.82 (1H, br d, J 14.8, NH), 5.51 (1H, dt, J 15.0, 6.7, 5-H), 5.34 (1H, dt, J 15.1, 7.1, 4-H), 3.91 (1H, tdd, J 11.1, 7.2, 3.6, 2-H), 3.65 (1H, dd, J 11.0, 3.3, 1-HH), 3.58 (1H, dd, J 11.1, 5.7, 1-HH), 3.25 (1H, br s, OH), 2.27–2.22 (1H, m, 3-HH), 2.17 (1H, dt, J 14.1, 7.1, 3-HH), 1.98 (5H, m, 6-H₂ and COCH₃), 1.35–1.30 (2H, m, 7-H₂), 1.30 – 1.22 (2OH, m, 8-H₂ to 17-H₂), 0.87 (3H, t, J 7.1, 18-H₃); *m/z* (ES⁻) 324.4 [M-H]⁻; (ES⁺) 326.4 [M+H]⁺; HRMS (ES⁺) found [M+H]⁺ 326.3046, C₂₀H₄₀O₂N₁ requires *M*⁺ 326.3035.

C₁₃H₂₇OH NHCOCH₂Ph

27ay (S,E)-N-(1-hydroxyoctadec-4-en-2-yl)-2-phenylacetamide

Yield (82%); $\delta_{\rm H}$ (700 MHz, CDCl₃) 7.36–7.33 (1 H, m, Ar*H*), 7.30–7.28 (1 H, m, Ar*H*), 7.25–7.22 (1 H, m, Ar*H*), 5.58 (1H, br d, *J* 5.6, N*H*), 5.32 (1H, dt, *J* 14.7, 6.6, 5-*H*), 5.23–5.18 (1 H, m, 4-*H*), 3.88 (1H, dtd, *J* 13.2, 6.5, 3.8, 2-*H*), 3.61 (1H, dd, *J* 11.1, 3.5, 1-*H*H), 3.58 (2H, s, PhC*H*₂), 3.54 (1H, dd, *J* 11.1, 6.2, 1-H*H*), 2.19–2.14 (1H, m, 3-*H*H), 2.09–2.04 (1H, m, 3-H*H*), 1.88 (2H, dd, *J* 13.2, 6.5, 6-*H*₂), 1.31–1.23 (22H, m, 7-*H*₂ to 17-*H*₂), 0.88 (3H, t, *J* 7.0, 18-*H*₃); *m*/*z* (ES⁻) 400.5 [M–H]⁻; (ES⁺) 402.4 [M+H]⁺; HRMS (ES⁺) found [M+H]⁺ 402.3356, C₂₆H₄₄O₂N₁ requires *M*⁺ 402.3348.

27ax (S,E)-N-(1-hydroxyoctadec-4-en-2-yl)octanamide

Yield (85%); $\delta_{\rm H}$ (700 MHz, CDCl₃) 5.71 (1 H, br s, N*H*), 5.52 (1H, dt, *J* 14.8, 6.8, 5-*H*), 5.37–5.31 (1H, m, 4-*H*), 3.91 (1H, tdd, *J* 10.8, 6.9, 3.6, 2-*H*), 3.65 (1H, dd, *J* 10.9, 3.1, 1-*H*H), 3.58 (1H, dd, *J* 11.0, 6.0, 1-H*H*), 3.28 (1H, br s, O*H*), 2.26 (1H, dt, *J* 13.5, 6.6, 3-*H*H), 2.20–2.14 (3H, m, 3-H*H* and COC*H*₂), 1.99 (2H, dd, *J* 14.3, 7.1, 6-*H*₂), 1.60 (2H, dt, *J* 14.6, 7.4, COCH₂C*H*₂), 1.35–1.22 (30H, m, 7-*H*₂ to 17-*H*₂ and 4 × aliphatic C*H*₂), 0.87 (6H, t, *J* 7.1, 18-*H*₃ and terminal octanoyl C*H*₃); *m*/*z* (ES⁻) 408.5 [M–H]⁻; (ES⁺) 410.5 [M+Na]⁺; HRMS (ES⁺) found [M+H]⁺ 410.3999, C₂₆H₅₂O₂N₁ requires *M*⁺ 410.3998.

26an (*R*,*E*)-2-aminooctadec-4-en-1-ol

Yield (62%); $\delta_{\rm H}$ (400 MHz, CDCl₃) 5.51 (1H, dt, *J* 13.9, 6.5, 5-*H*), 5.38–5.30 (1H, m, 4-*H*), 3.60 (1H, d, *J* 8.2, 1-H*H*), 3.33 (1H, dd, *J* 10.2, 8.2, 1-*H*H), 2.97-2.83 (1H, m, 2-*H*), 2.34 (3H, br s, N*H*₂ and O*H*), 2.20–2.10 (1H, m, 3-*H*₂), 1.99 (2H, dd, *J* 13.5, 6.6, 6-*H*₂), 1.37–1.20 (22H, m, 7-*H*₂ to 17-*H*₂), 0.87 (3H, t, *J* 6.5, 18-*H*₃); *m*/*z* (ES⁺) 284.2 [M+H]⁺; HRMS (ES⁺) found [M+H]⁺ 284.2947, C₁₈H₃₈O₁N₁ requires *M*⁺ 284.2953.



26af (R,E)-2,2,2-trifluoro-N-(1-hydroxyoctadec-4-en-2-yl)acetamide

Yield (38%); δ_{H} (400 MHz, CDCl₃) 6.56 (1H, br d, *J* 7.0, N*H*), 5.56 (1H, dt, *J* 15.0, 6.8, 5-*H*), 5.35 (1H, dt, *J* 15.0, 7.2, 4-*H*), 4.05–3.97 (1H, m, 2-*H*), 3.73 (2H, app. d, *J* 3.8, 1-*H*₂), 2.39–2.25 (2H, m, 3-*H*₂), 2.00 (2H, dd, *J* 13.8, 6.9, 6-*H*₂), 1.38–1.20 (22H, m, 7-*H*₂ to 17-*H*₂), 0.88 (3H, t, *J* 6.9, 18-*H*₃); δ_{F} (376 MHz, CDCl₃) –75.87 CF₃; *m/z* (ES⁻) 378.4 [M-H]⁻; (ES⁺) 402.4 [M+Na]⁺; HRMS (ES⁺) found [M+Na]⁺ 402.2603, C₂₀H₃₆O₂N₁F₃²³Na₁ requires *M*⁺ 402.2608.

C₁₃H₂₇OH NHCOMe

26az (R,E)-N-(1-hydroxyoctadec-4-en-2-yl)acetamide

Yield (88%); $\delta_{\rm H}$ (700 MHz, CDCl₃) 5.82 (1H, br d, *J* 14.8, N*H*), 5.51 (1H, dt, *J* 15.0, 6.7, 5-*H*), 5.34 (1H, dt, *J* 15.1, 7.1, 4-*H*), 3.91 (1H, tdd, *J* 11.1, 7.2, 3.6, 2-*H*), 3.65 (1H, dd, *J* 11.0, 3.3, 1-H*H*), 3.58 (1H, dd, *J* 11.1, 5.7, 1-*H*H), 3.25 (1H, br s, O*H*), 2.27–2.22 (1H, m, 3-H*H*), 2.17 (1H, dt, *J* 14.1, 7.1, 3-*H*H), 1.98 (5H, m, 6-*H*₂ and COC*H*₃), 1.35–1.30 (2H, m, 7-*H*₂), 1.30–1.22 (2OH, m, 8-*H*₂ to 17-*H*₂), 0.87 (3H, t, *J* 7.1, 18-*H*₃); *m*/z (ES⁻) 324.4 [M-H]⁻; (ES⁺) 326.4 [M+H]⁺; HRMS (ES⁺) found [M+H]⁺ 326.3046, C₂₀H₄₀O₂N₁ requires *M*⁺ 326.3035.

26ay (R,E)-N-(1-hydroxyoctadec-4-en-2-yl)-2-phenylacetamide

Yield (47%); $\delta_{\rm H}$ (700 MHz, CDCl₃) 7.36–7.33 (2H, m, Ar*H*), 7.30–7.28 (1H, m, Ar*H*), 7.25–7.22 (2H, m, Ar*H*), 5.58 (1H, br d, J 5.6, N*H*), 5.32 (1H, dt, J 14.7, 6.6, 5-*H*), 5.23–5.18 (1H, m, 4-*H*), 3.88 (1H, dtd, J 13.2, 6.5, 3.8, 2-*H*), 3.61 (1H, dd, J 11.1, 3.5, 1-*H*H), 3.58 (2H, s, PhC*H*₂), 3.54 (1H, dd, J 11.1, 6.2, 1-H*H*), 2.19–2.14 (1H, m, 3-*H*H), 2.09 – 2.04 (1H, m, 3-H*H*), 1.88 (2H, dd, J 13.2, 6.5, 6-*H*₂), 1.31–1.23 (22H, m, 7-*H*₂ to 17-*H*₂), 0.88 (3H, t, J 7.0, 18-*H*₃); *m*/*z* (ES⁻) 400.5 [M–H]⁻; (ES⁺) 402.4 [M+H]⁺; HRMS (ES⁺) found [M+H]⁺ 402.3356, C₂₆H₄₄O₂N₁ requires *M*⁺ 402.3348.

26ax (R,E)-N-(1-hydroxyoctadec-4-en-2-yl)octanamide

Yield (76%); $\delta_{\rm H}$ (700 MHz, CDCl₃) 5.71 (1 H, br s, N*H*), 5.52 (1 H, dt, *J* 14.8, 6.8, 5-*H*), 5.37–5.31 (1H, m, 4-*H*), 3.91 (1H, tdd, *J* 10.8, 6.9, 3.6, 2-*H*), 3.65 (1H, dd, *J* 10.9, 3.1, 1-*H*H), 3.58 (1H, dd, *J* 11.0, 6.0, 1-H*H*), 3.28 (1H, br s, O*H*), 2.26 (1H, dt, *J* 13.5, 6.6, 3-*H*H), 2.20–2.14 (3H, m, 3-H*H* and COC*H*₂), 1.99 (2H, dd, *J* 14.3, 7.1, 6-*H*₂), 1.60 (2H, dt, *J* 14.6, 7.4, COCH₂C*H*₂), 1.35–1.22 (30H, m, 7-*H*₂ to 17-*H*₂ and 4 × aliphatic C*H*₂), 0.87 (6H, t, *J* 7.1, 18-*H*₃ and terminal octanoyl C*H*₃); *m*/*z* (ES⁻) 408.5 [M–H]⁻; (ES⁺) 410.5 [M+Na]⁺; HRMS (ES⁺) found [M+H]⁺ 410.3999, C₂₆H₅₂O₂N₁ requires *M*⁺ 410.3998.

Supplementary Material (ESI) for Organic & Biomolecular Chemistry This journal is (c) The Royal Society of Chemistry 2011

BIOLOGICAL METHODS

Preparation of the Screening Compounds

All compounds were first dissolved in MeOH and 1.0 mM stock solutions were prepared in DMSO/MeOH (9:1 v/v) and 0.20 mM working solutions in DMSO/MeOH (98:2). All prepared solutions were stored at -20 °C until use.

The Assay Protocol of the Inhibition Assays

The hydrophobic nature of most of the synthesised substrate analogues required special attention to ensure their solubility/dispersion in the reaction mixture. For each inhibition reaction, the donor substrate (PI) and test inhibitors were first dried into a reaction Eppendorf tube followed by addition of the reaction buffer, CHAPS and NBD-C₆- ceramide followed by sonication for 3 minutes and incubation at 30 °C for 15 minutes. The reaction was started by the addition of *Lmj*IPCS microsomes (0.6 u). Final concentrations of the test compound, PI and NBD-C₆-ceramide were 20 μ M, 100 μ M and 5 μ M respectively. After 15 minutes the reaction was quenched by the addition of MeOH and the product sphingolipids separated from unreacted NBD-C6-ceramide by ion exchange chromatography.¹⁹ The amount of fluorescent product was then quantified using a fluorescence plate reader. All reactions were done in triplicates with the inhibitory effect quantified by the change in the formation of the labelled product, NBD-C₆-IPC.

Mass Spectrometry Analyses

Analyses were performed on an LTQFT (ThermoFinnigan Corp); an FTICR MS instrument equipped with a 7.0 T magnet. A different assay protocol²⁰ was deployed to minimise CHAPS content using PI-depleted CHAPS-washed microsomal membranes.²¹ The organic extracts were dried and re-suspended in chloroform and introduced into the electrospray ion source by direct infusion from a syringe at a flow rate 3 µl/min. Positive ion measurements were made with the source voltage at 4.0 kV and negative ion measurements were made with the source voltage at 4.0 kV and negative ion measurements were made with the source voltage at 3.5 kV. The tube lens was kept at 100 V and the source temperature at 275 °C for all experiments.

The spectra are presented below in the following format,

A: Full spectrum of the organic extract of the reaction mixture.

B: Expansion of the spectra showing the formation NBD-C₆-IPC.

C: Predicted response of the product that will hypothetically arise if the test compound functions as an alternative substrate.

D: Expansion of the spectra showing the region identified in C above.



27a

А





В







27a

С





D

27a-Phosphoryl Inositol: Not Detected









А

Full Spectrum

В









С





D

18ax-Phosphoryl Inositol: Not Detected







Negative Ion Spectra



А

Full Spectrum

В









С











Page 38 of 50







А

Full Spectrum

В

NBD-C₆-IPC: Detected







С





D







26af







В







26af

С





D







26an







В





Page 43 of 50



26an

С





D







26ay







В























26ax















26ax

С











Page 48 of 50

Cytotoxicity screening

L. major (MHOM/IL/81/Friedlin) promatigotes parasites were maintained at 26 °C in Schneider's Drosophila media (Sigma–Aldrich) supplemented with 15% heat inactivated foetal bovine sera (Biosera).

In 96-well plates (Nunc) parasites at 4×10^5 ml⁻¹ were incubated with compounds in triplicate (including

miltefosine (Cayman Chemical) as a positive control, and untreated parasites and media as negative controls) for

24 h before incubation with Alamar Blue (Invitrogen) for 4 h prior to assessing cell viability

using a fluorescent plate reader (Biotek; 560EX nm/600EMnm).

REFERENCES

- 1. L. Kosynkina, W. Wang and T. C. Liang, *Tetrahedron Lett.*, 1994, **35**, 5173-5176.
- 2. T. Yamamoto, H. Hasegawa, T. Hakogi and S. Katsumura, Org. Lett., 2006, 8, 5569-5572.
- 3. P. K. Chakravarty, W. J. Greenlee, W. H. Parsons, A. A. Patchett, P. Combs, A. Roth, R. D. Busch and T. N. Mellin, *J. Med. Chem.*, 1989, **32**, 1886-1890.
- 4. T. Ibuka, H. Habashita, A. Otaka, N. Fujii, Y. Oguchi, T. Uyehara and Y. Yamamoto, *J. Org. Chem.*, 1991, **56**, 4370-4382.
- 5. M. Toumi, F. Couty and G. Evano, *Tetrahedron Lett.*, 2008, **49**, 1175-1179.
- 6. M. Toumi, F. Couty and G. Evano, *Angewandte Chemie-International Edition*, 2007, **46**, 572-575.
- 7. R. C. So, R. Ndonye, D. P. Izmirian, S. K. Richardson, R. L. Guerrera and A. R. Howell, *J. Org. Chem.*, 2004, **69**, 3233-3235.
- 8. E. J. Corey, F. Xu and M. C. Noe, *Journal of the American Chemical Society*, 1997, **119**, 12414-12415.
- 9. B. Lygo and P. G. Wainwright, *Tetrahedron*, 1999, **55**, 6289-6300.
- 10. J. Aires-de-Sousa, S. Prabhakar, A. M. Lobo, A. M. Rosa, M. J. S. Gomes, M. C. Corvo, D. J. Williams and A. J. P. White, *Tetrahedron-Asymmetry*, 2002, **12**, 3349-3365.
- 11. M. J. Odonnell and R. L. Polt, J. Org. Chem., 1982, 47, 2663-2666.
- 12. R. Chinchilla, C. Nájera and F. J. Ortega, *Tetrahedron-Asymmetry*, 2006, **17**, 3423-3429.
- 13. T. Ohshima, T. Shibuguchi, Y. Fukuta and M. Shibasaki, *Tetrahedron*, 2004, **60**, 7743-7754.
- 14. T. Ohshima, V. Gnanadesikan, T. Shibuguchi, Y. Fukuta, T. Nemoto and M. Shibasaki, *J. Am. Chem. Soc.*, 2003, **125**, 11206-11207.
- 15. S. M. Jones, J. E. Urch, M. Kaiser, R. Brun, J. L. Harwood, C. Berry and I. H. Gilbert, *J. Med. Chem.*, 2005, **48**, 5932-5941.
- 16. H. Matsunaga, T. Ishizuka and T. Kunieda, *Tetrahedron*, 1997, **53**, 1275-1294.
- 17. B. R. Galan, K. P. Kalbarczyk, S. Szczepankiewicz, J. B. Keister and S. T. Diver, *Organic Letters*, 2007, **9**, 1203-1206.
- 18. T. Kawate, N. Fukuta, A. Nishida and M. Nakagawa, *Chem. Pharm. Bull.*, 1997, **45**, 2116-2118.
- 19. J. G. Mina, J. A. Mosely, H. Z. Ali, H. Shams-Eldin, R. T. Schwarz, P. G. Steel and P. W. Denny, *The International Journal of Biochemistry & Cell Biology*, 2010, **42**, 1553-1561.
- 20. J. M. Figueiredo, W. B. Dias, L. Mendonca-Previato, J. O. Previato and N. Heise, *Biochemical Journal*, 2005, **387**, 519-529.
- 21. P. A. Aeed, A. E. Sperry, C. L. Young, M. M. Nagiec and Å. P. Elhammer, *Biochemistry*, 2004, **43**, 8483-8493.