Supplementary Data

A novel strategy of chemical modification for rate enhancement of 10-23 DNAzyme: a combination of A9 position and 8-aza-7-deaza-2'-deoxyadenosine analogs

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No.	Modified 10-23 deoxyribozymes	MW (calculated)	MW (found)
DZ-00	5'-d(tgc tct cca GGC TAG CTA CAA CGA cct gca cct)-3'	9994.4	9990.5
DZ-1-5	5'-d(tgc tct cca GGC T1G CTA CAA CGA cct gca cct)-3'	9993.4	9994.8
DZ-1-9	5'-d(tgc tct cca GGC TAG CT1 CAA CGA cct gca cct)-3'	9993.4	9991.1
DZ-1-11	5'-d(tgc tct cca GGC TAG CTA C1A CGA cct gca cct)-3'	9993.4	9993.3
DZ-1-12	5'-d(tgc tct cca GGC TAG CTA CA1 CGA cct gca cct)-3'	9993.4	9991.7
DZ-1-15	5'-d(tgc tct cca GGC TAG CTA CAA CG1 cct gca cct)-3'	9993.4	9989.4
DZ-2-5	5'-d(tgc tct cca GGC T2G CTA CAA CGA cct gca cct)-3'	9994.4	9995.5
DZ-2-9	5'-d(tgc tct cca GGC TAG CT2 CAA CGA cct gca cct)-3'	9994.4	9995.9
DZ-2-11	5'-d(tgc tct cca GGC TAG CTA C2A CGA cct gca cct)-3'	9994.4	9990.1
DZ-2-12	5'-d(tgc tct cca GGC TAG CTA CA2 CGA cct gca cct)-3'	9994.4	9991.2
DZ-2-15	5'-d(tgc tct cca GGC TAG CTA CAA CG2 cct gca cct)-3'	9994.4	9995.5
DZ-3-5	5'-d(tgc tct cca GGC T3G CTA CAA CGA cct gca cct)-3'	10051.5	10051.8
DZ-3-9	5'-d(tgc tct cca GGC TAG CT3 CAA CGA cct gca cct)-3'	10051.5	10051.1
DZ-3-11	5'-d(tgc tct cca GGC TAG CTA C3A CGA cct gca cct)-3'	10051.5	10052.2
DZ-3-12	5'-d(tgc tct cca GGC TAG CTA CA3 CGA cct gca cct)-3'	10051.5	10053.2
DZ-3-15	5'-d(tgc tct cca GGC TAG CTA CAA CG3 cct gca cct)-3'	10051.5	10053.4
DZ-4-5	5'-d(tgc tct cca GGC T4G CTA CAA CGA cct gca cct)-3'	10052.5	10053.5
DZ-4-9	5'-d(tgc tct cca GGC TAG CT4 CAA CGA cct gca cct)-3'	10052.5	10053.4
DZ-4-11	5'-d(tgc tct cca GGC TAG CTA C4A CGA cct gca cct)-3'	10052.5	10053.7
DZ-4-12	5'-d(tgc tct cca GGC TAG CTA CA4 CGA cct gca cct)-3'	10052.5	10050.1
DZ-4-15	5'-d(tgc tct cca GGC TAG CTA CAA CG4 cct gca cct)-3'	10052.5	10053.1
DZ-5-9	5'-d(tgc tct cca GGC TAG CT5 CAA CGA cct gca cct)-3'	10098.6	10099.1

Table S1. MALDI-TOF measurements of modified 10-23 deoxyribozymes modified with 1-5.

substrate.^a

Complex	T _m (°C)
10-23 DNAzyme +D19	51.7
DZ-2-9+D19	50.1
DZ-3-9+D19	50.7
DZ-4-9+D19	51.0
DZ-5-9+D19	51.0
DZ-3-5+D19	49.8

Table S2. T_m measurements of the complexes formed between deoxyribozymes and the full-DNA

^{*a*} D19: the full-DNA substrate, 5'-d (AGG TGC AGG ATG GAG AGC A)-3'. Tm measurement was conducted in the buffer consisting of 50 mM Tris-HCl (pH 7.5) and 2 mM MgCl₂. The solution was heated from 85 °C, after halting for 10 min, it was cooled to 10 °C at a rate of 0.5 °C/min. The UV absorbance was recorded at 260 nm during the cooling process. The T_m values were obtained from the melting curves, and each melting curve was fit to a non-self-complementary two-state model.



Fig. S1 The melting curves of the complex of the DNA substrate and 10-23 DNAzyme (black), DZ-1-9 (purple), DZ-2-9 (green), DZ-3-5 (blue), DZ-3-9 (yellow), DZ-4-9 (red), and DZ-5-9 (pink), respectively. The conditions for the measurement see the footnote under Table S2.



Fig. S2 The cleavage pattern of 10-23 DNAzyme and **DZ-3-9** in the presence of different Mg^{2+} concentrations. The reaction was conducted under single-turnover conditions in a buffer of 50 mM Tris-HCl, pH 7.5, containing 0.2 mM (A), 2 mM (B), or 50 mM (C) Mg^{2+} , as well as the control without Mg^{2+} (D). Time points for aliquots were 0, 15, 30, 45, 60, 120, 240, and 360 min from left to right. In each panel, the upper band was the substrate, and the lower band was the product.



Fig. S3 The effect of divalent metal ions on the cleavage reactions of 10-23 DNAzyme (A) and DZ-3-9 (B) under single turnover conditions, in a buffer of 50 mM Tris-HCl, pH 7.5, containing 0.2 mM divalent metal ion. Time points for aliquots were 0, 10, 20, 30, 40, 50, 60, 90, 120, 150, and 180 min from left to right. In each panel, the upper band is the substrate, and the lower band corresponds to the product.

Compound 3 ¹H NMR



Compound 3 ¹³C NMR



Compound 6¹H NMR



Compound 6 ¹³C NMR



Compound 10¹H NMR



Compound **10**¹³C NMR







Compound 14 ¹H NMR









MALDI-TOF MS













































