

Supplementary Information:

Organic base effects in NHC promoted *O*- to *C*- carboxyl transfer; chemoselectivity profiles, mechanistic studies and domino catalysis

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I EXPERIMENTAL PROCEDURES

I.2 General information

¹H NMR Spectra were recorded using a Bruker Avance 400 spectrometer and Bruker Avance 300 spectrometer at 400 MHz and 300 MHz respectively, using residual protonated solvent as a reference for internal lock. The chemical shift information (δ_{H}) for each resonance signal are given in units of parts per million (ppm) relative to tetramethylsilane (TMS) where δ_{H} TMS = 0.00 ppm, or to residual (protonated) solvent. The number of protons (n) for a reported resonance signal are indicated by $n\text{H}$ from their integral value and their multiplicity is reported with their coupling constants (J) quoted in Hz. Coupling constants are determined by analysis using iNMR[®] and Topspin[®].

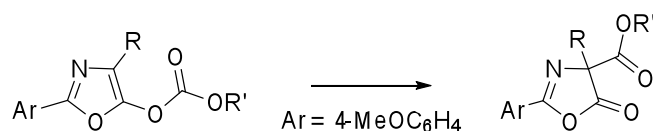
¹³C NMR Spectra were recorded using a Bruker Avance 300 and Bruker Avance 400 spectrometer using the PENDANT sequence at 75.5 MHz and 100 MHz respectively with internal deuterated solvent lock. The chemical shift information (δ_{C}) for each resonance signal is given in units of parts per million (ppm) relative to tetramethylsilane (TMS) where δ_{C} TMS = 0.00 ppm, or to the relevant solvent.

¹⁹F NMR Spectra were recorded using a Bruker Avance 400 spectrometer at 282 MHz. The chemical shift information (δ_{F}) for each resonance signal are given in units of parts per million (ppm) relative to trichlorofluoromethane (CFCl₃) where δ_{F} = 0.00.

HPLC was performed on either a Varian ProStar or Gilson apparatus, using a CHIRALPAK OD-H, AD-H or AS-H silica column, 0.46 cm ϕ $\times$ 25 cm, using hexane and isopropanol as eluents.

I.2 Known Substrates and Rearrangement Products

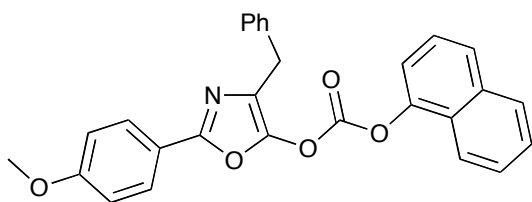
Known azlactone precursors, carbonate substrates and their *C*-carboxyazlactone isomers were prepared following literature procedures with physical and spectroscopic data in agreement with the literature,¹⁻⁵ which can be found at the following references. Spectroscopic data of purified materials were used to identify product distributions in crossover experiments.



R	OR'	Carbonate	<i>C</i> -Carboxy azlactone	Reference
Bn	OPh	4	5	1
Bn	OMe	11	17	1
Bn	OBn	12	18	2
Bn	OCMe ₂ CCl ₃	13	19	2
Me	OPh	23	30	3
<i>n</i> -Bu	OPh	24	31	2,4
<i>i</i> -Bu	OPh	25	32	2
Ph	OPh	26	33	2
4-PhOCO ₂ C ₆ H ₄ CH ₂	OPh	27	34	3
CH ₂ CH ₂ SMe	OPh	28	35	2
<i>i</i> -Pr	OPh	29	36	2
Me	OMe	37	38	2
Me	OCMe ₂ CCl ₃	39	40	2

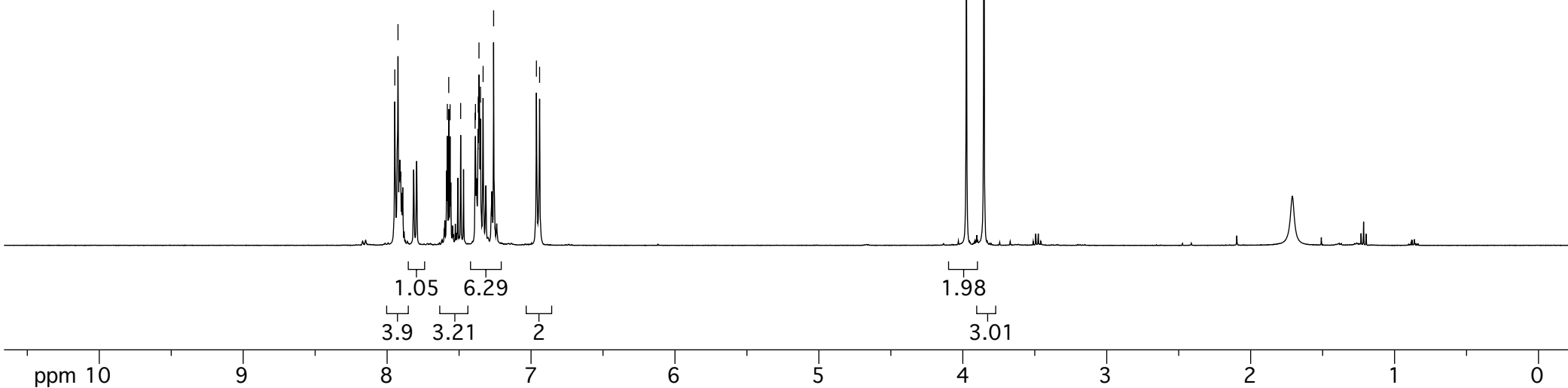
Literature procedures were used for the preparation of compounds **2**,¹ **6**,¹ **7**,¹ **9**,¹ **10**,¹ **47**⁵ and **48**,⁶ giving analytical and spectroscopic data in accordance with the literature.

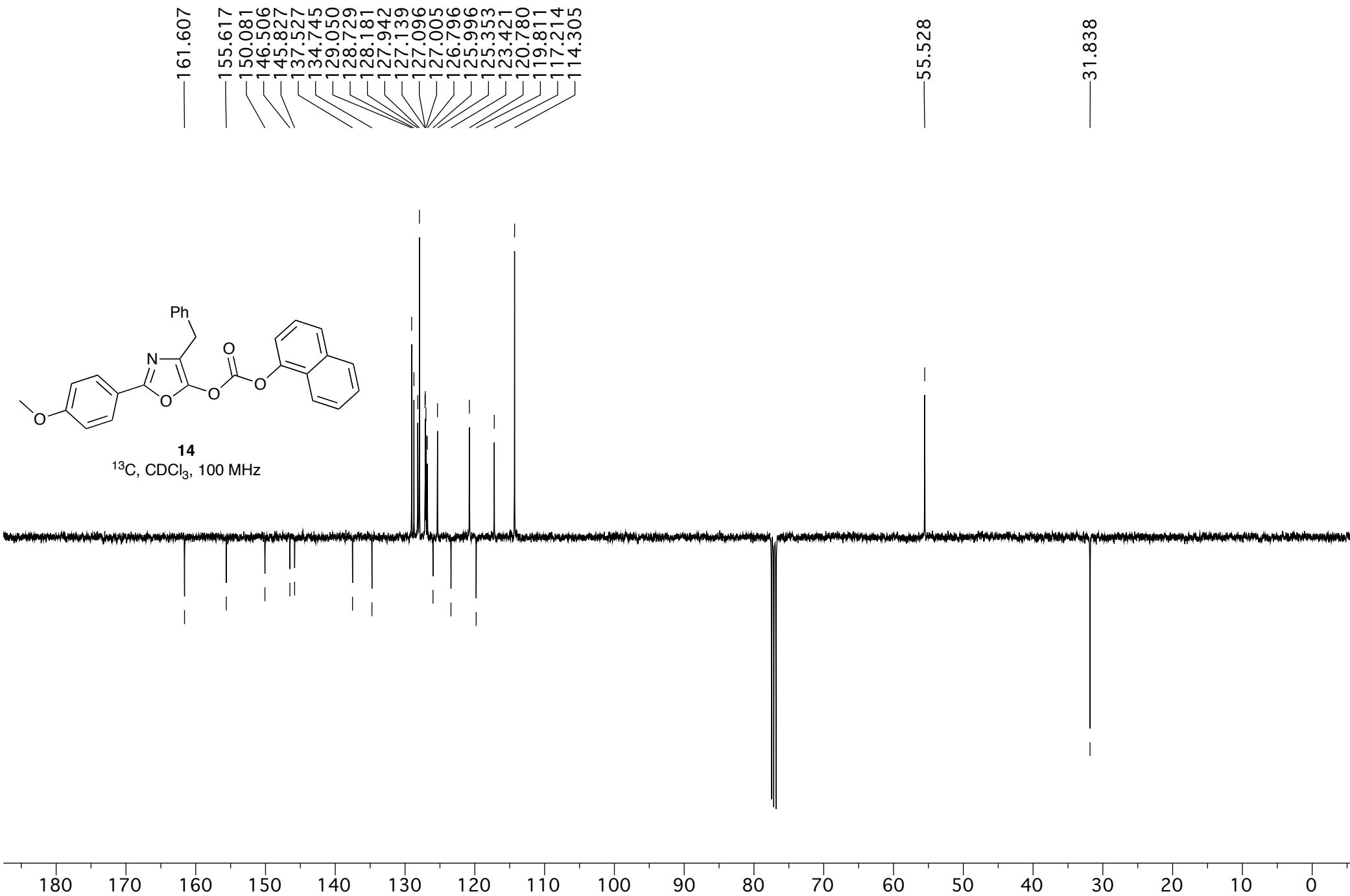
S4

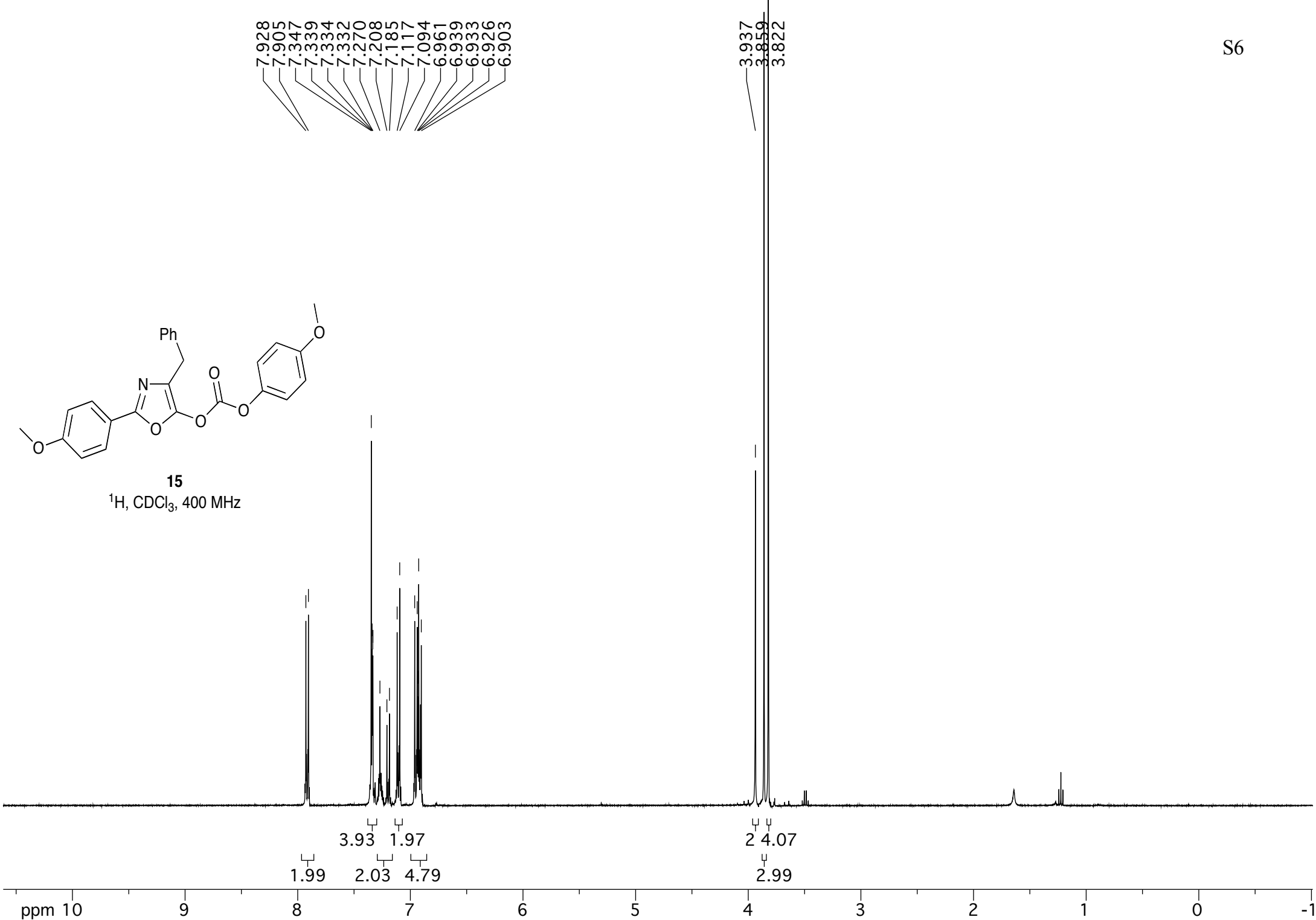
**14** ^1H , CDCl_3 , 400 MHz

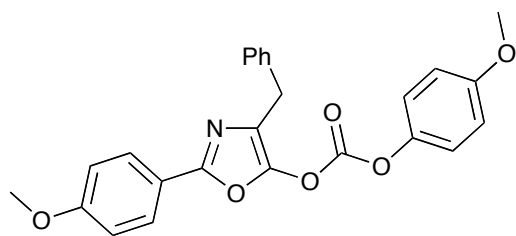
7.947
7.924
7.581
7.572
7.562
7.489
7.388
7.386
7.367
7.362
7.351
7.333
7.260
6.963
6.941

3.975
3.853



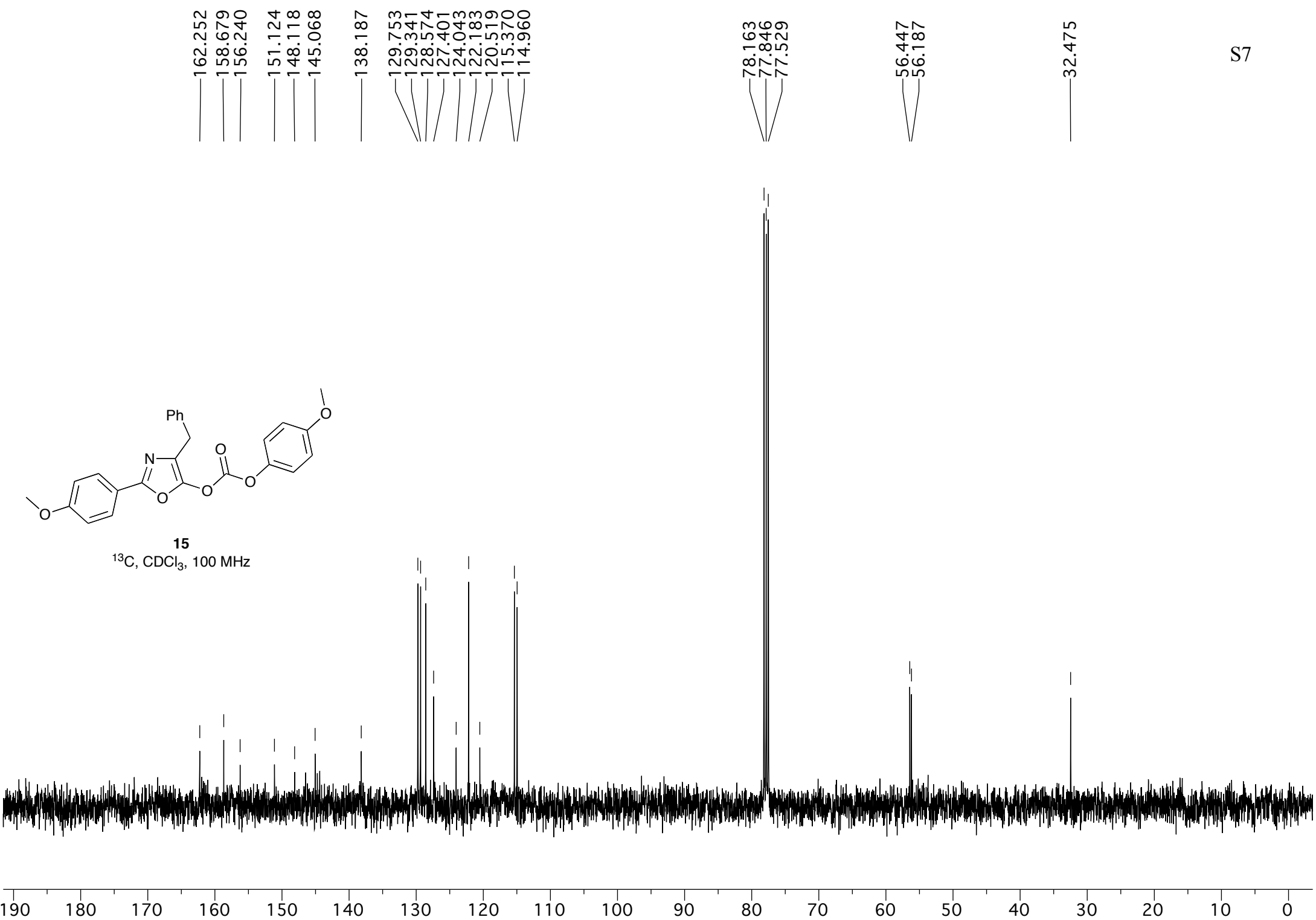


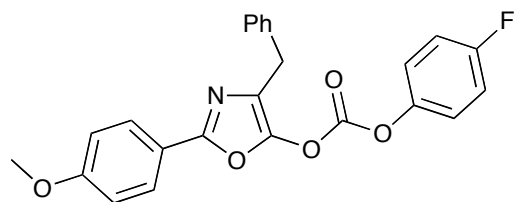
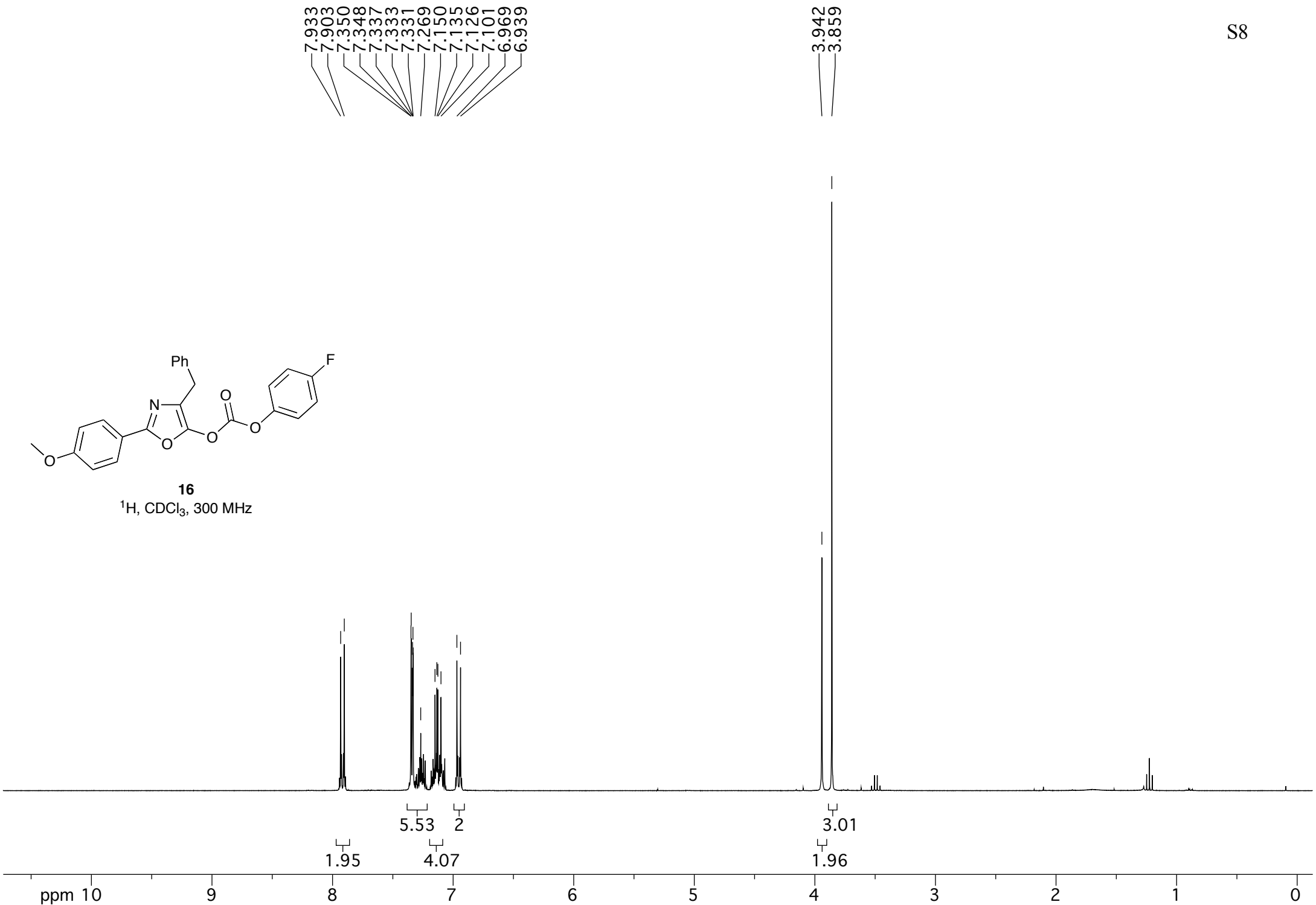


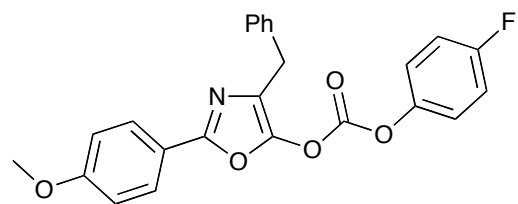


15

^{13}C , CDCl_3 , 100 MHz



**16**¹H, CDCl₃, 300 MHz



16

^{13}C , CDCl_3 , 75 MHz

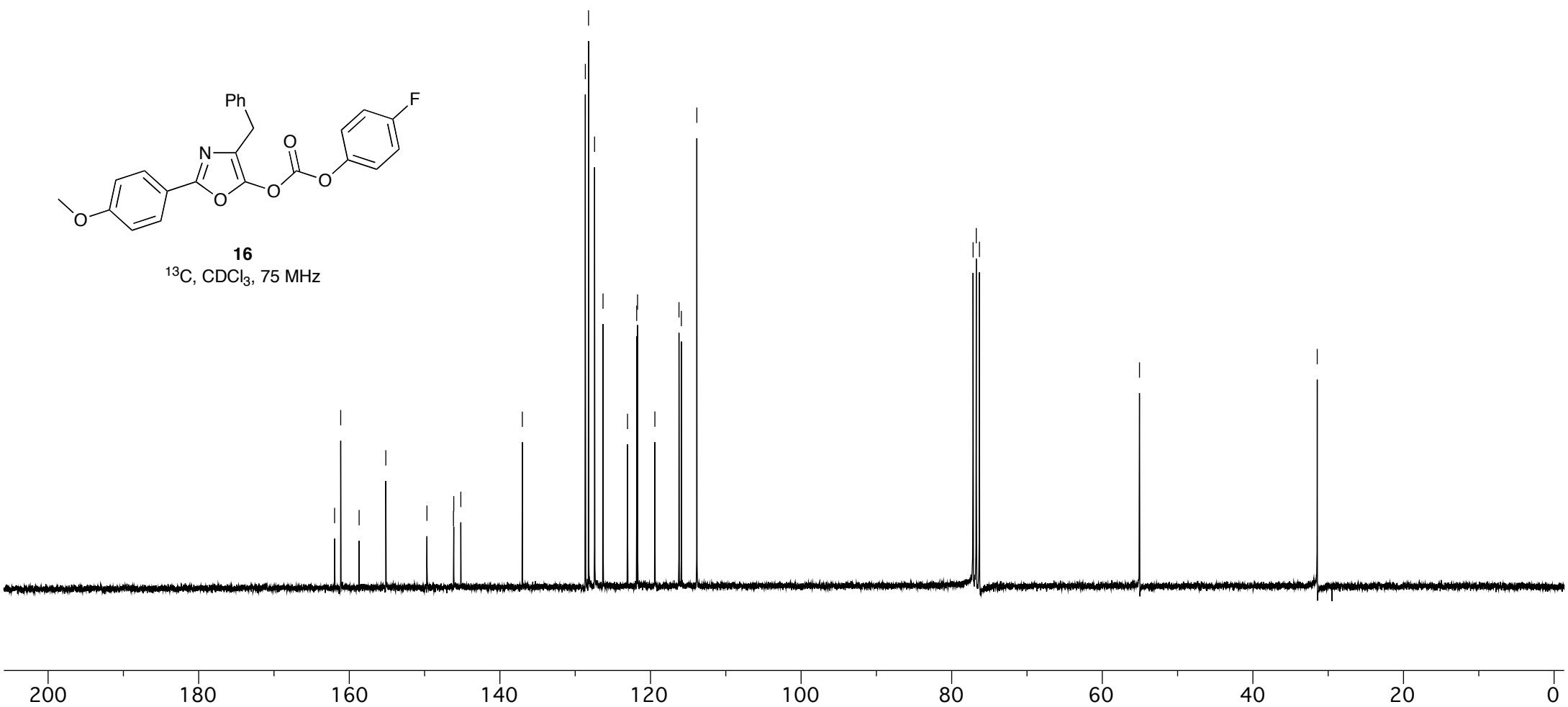
161.954
161.142
158.695
155.150
149.687
146.167
146.126
145.190
137.021
128.654
128.230
127.426
126.304
123.051
121.835
121.719
119.418
116.217
115.902
113.845

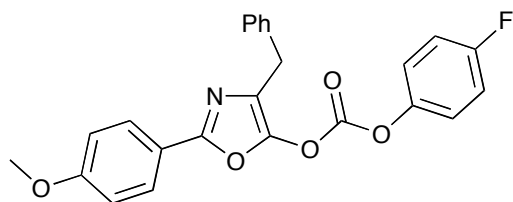
77.160
76.736
76.313

55.059

31.454

S9

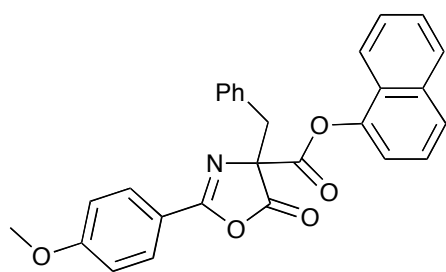




16
 ^{19}F , CDCl_3 , 282 MHz

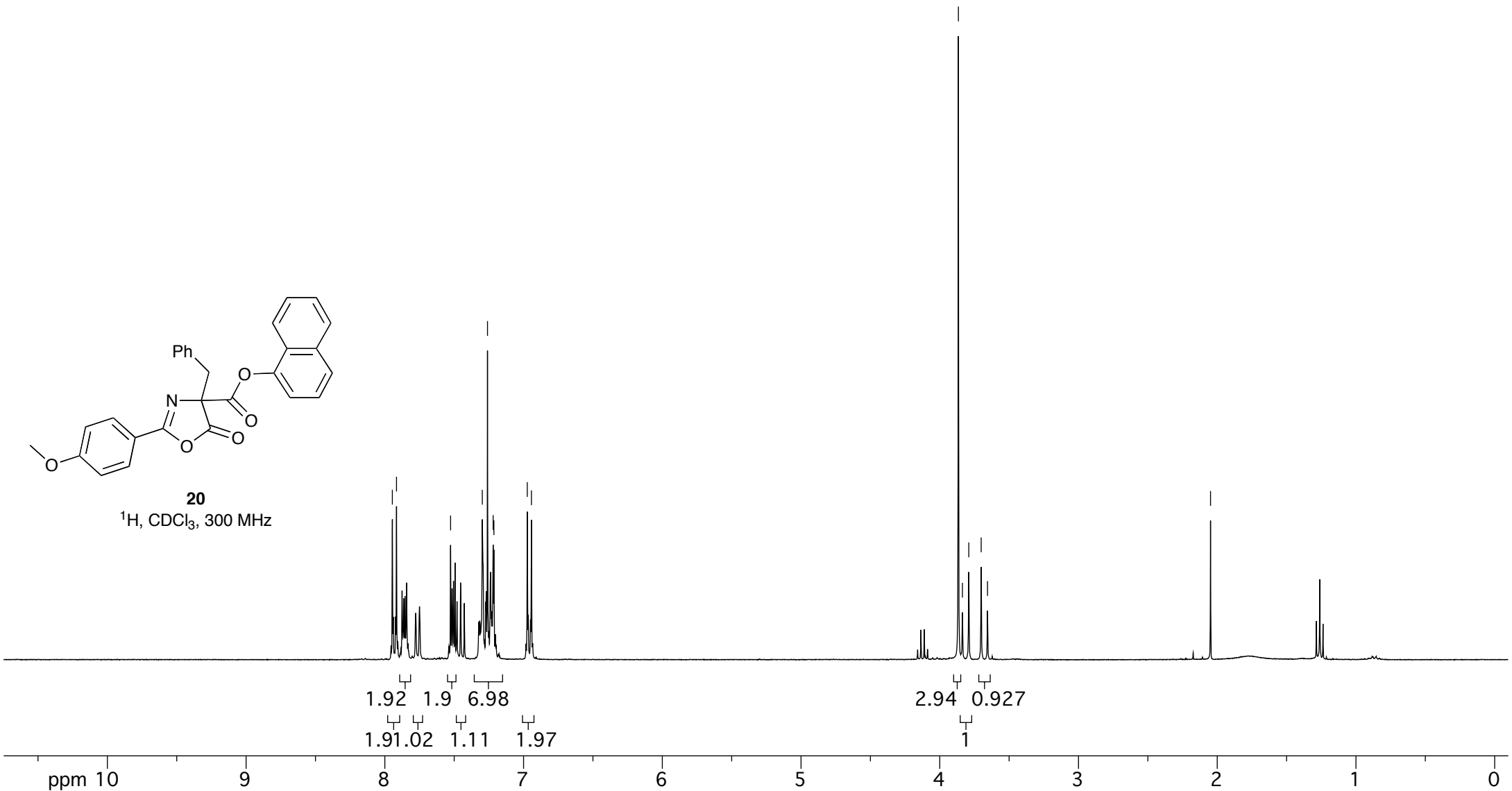
-115.668

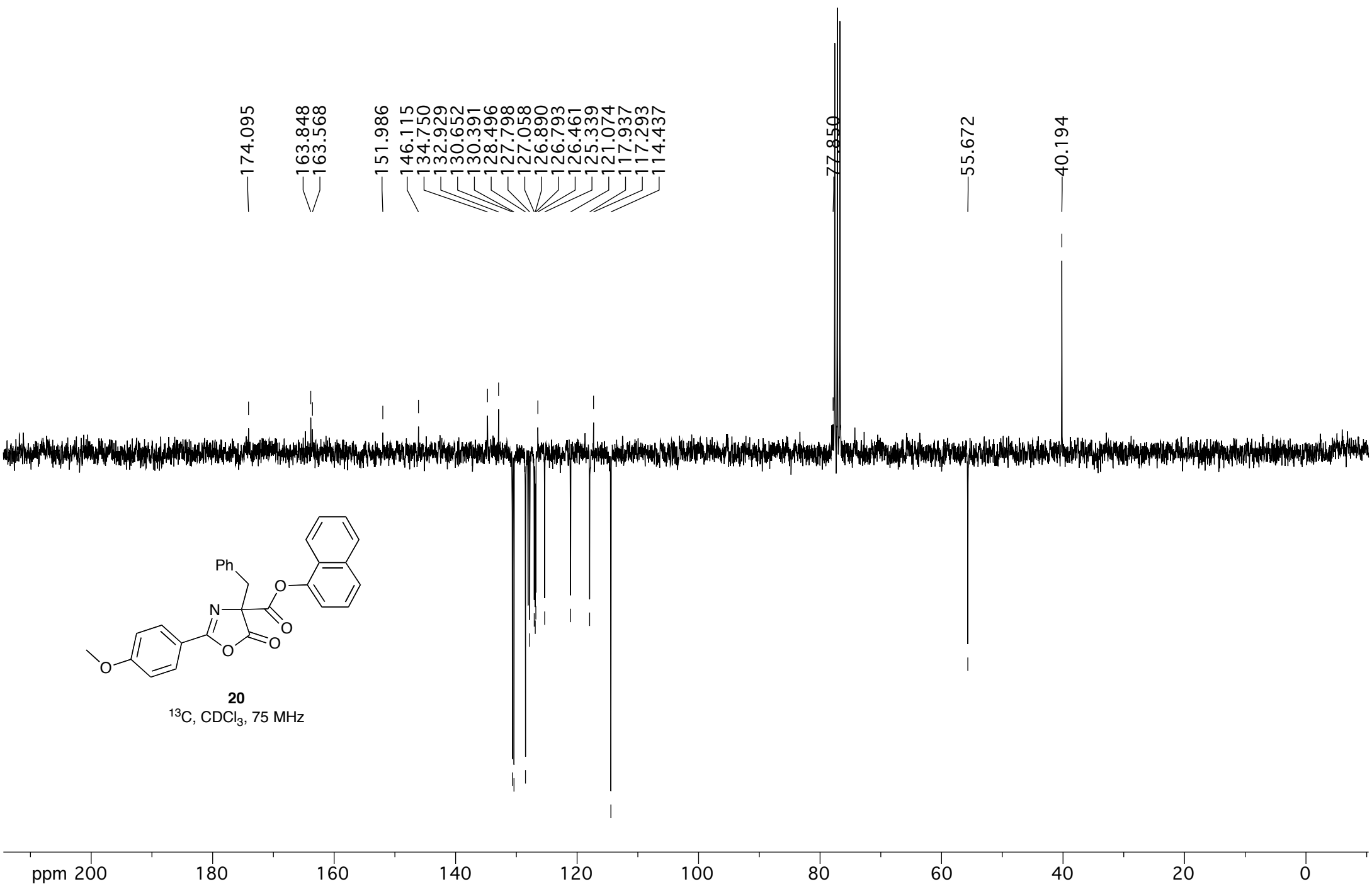
ppm 0 -20 -40 -60 -80 -100 -120 -140 -160 -180 -200 -220 -240

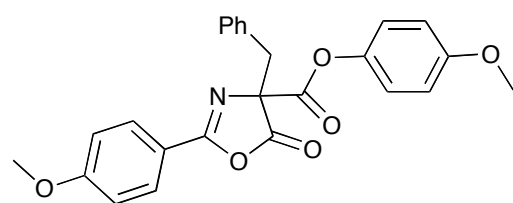
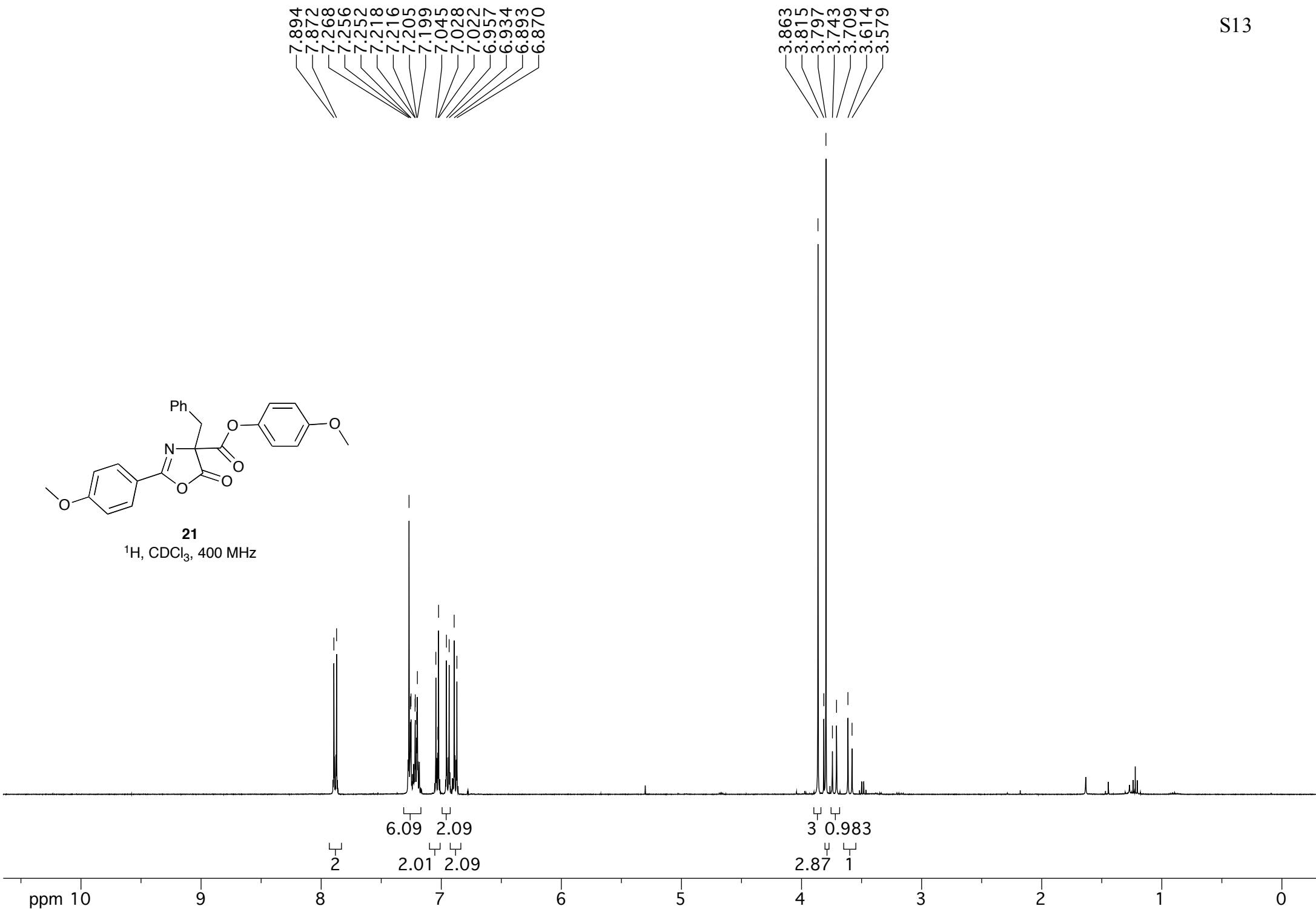


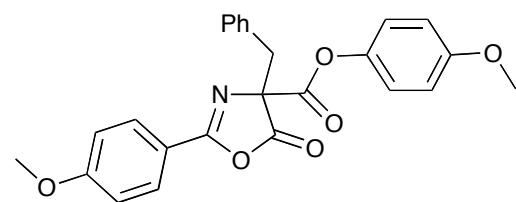
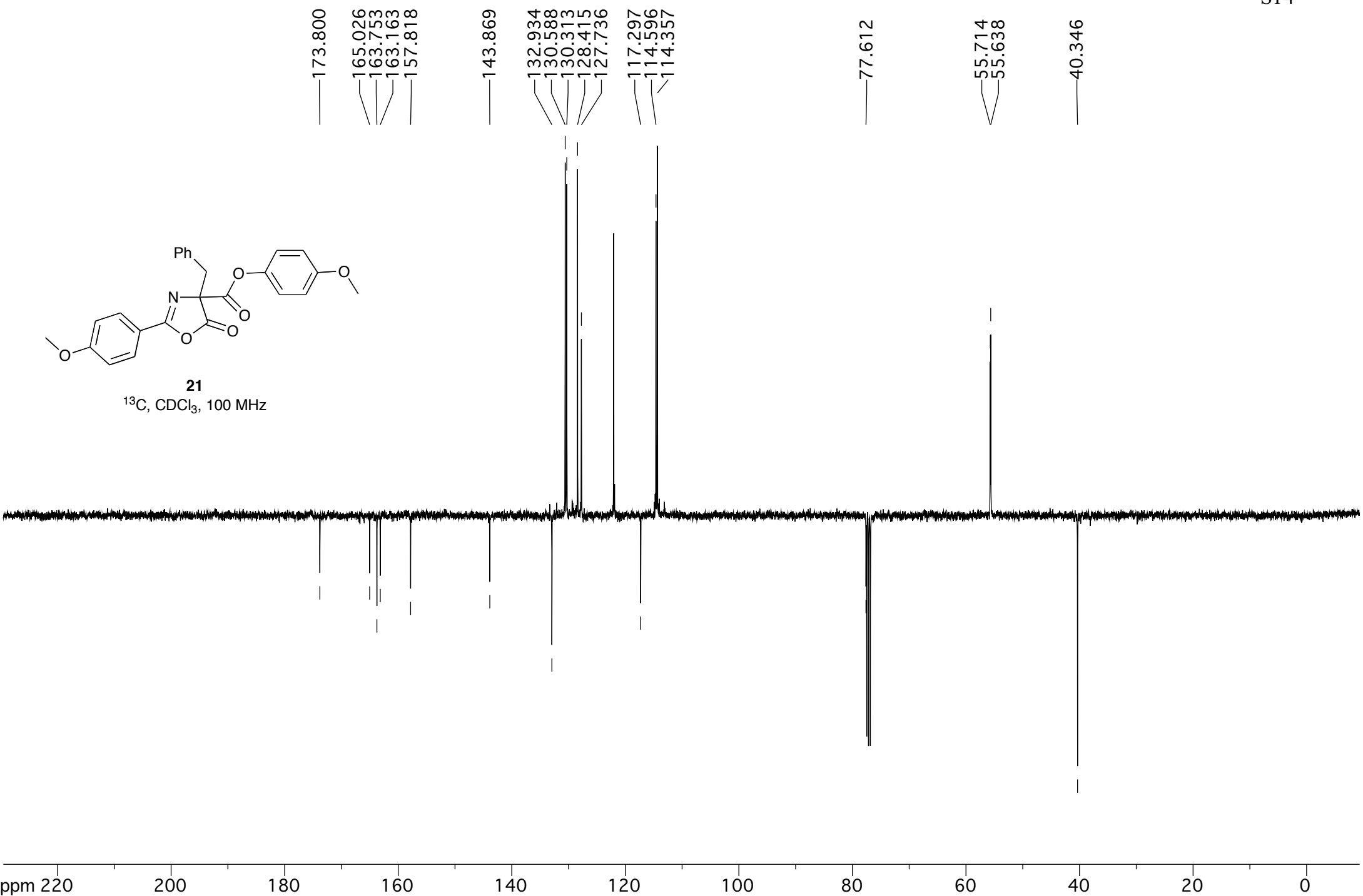
20
 ^1H , CDCl_3 , 300 MHz

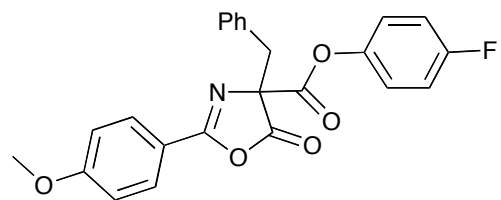
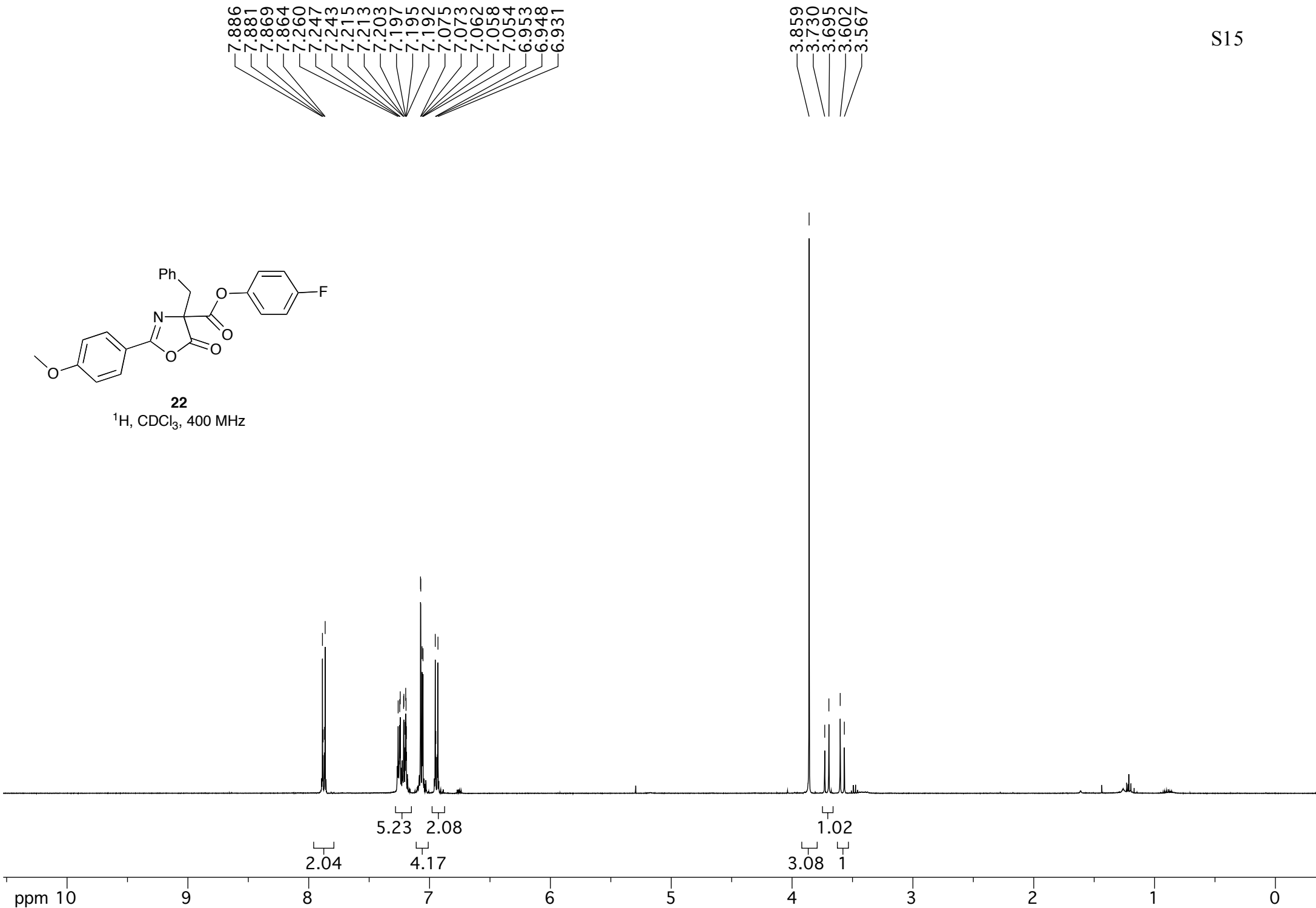
7.946
7.916
7.526
7.297
7.260
7.219
7.214
6.973
6.943
3.866
3.837
3.791
3.702
3.656
2.048

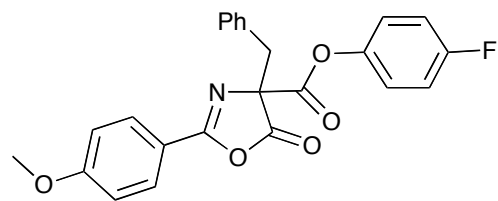
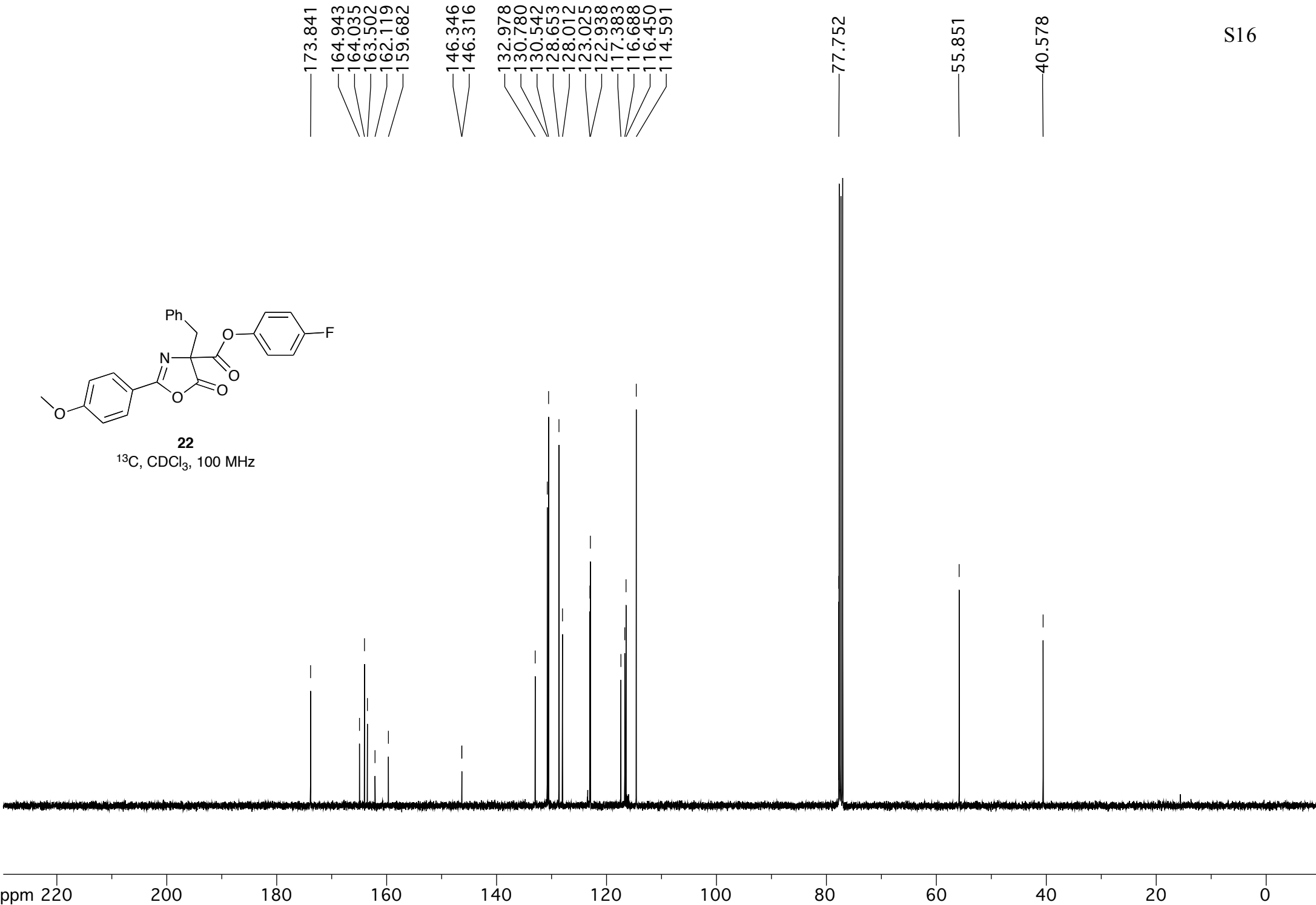


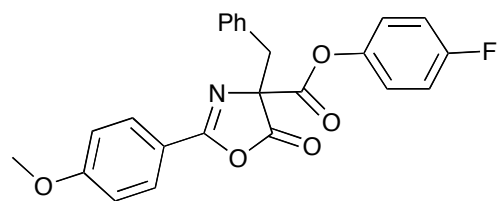


**21**¹H, CDCl₃, 400 MHz

**21** ^{13}C , CDCl_3 , 100 MHz

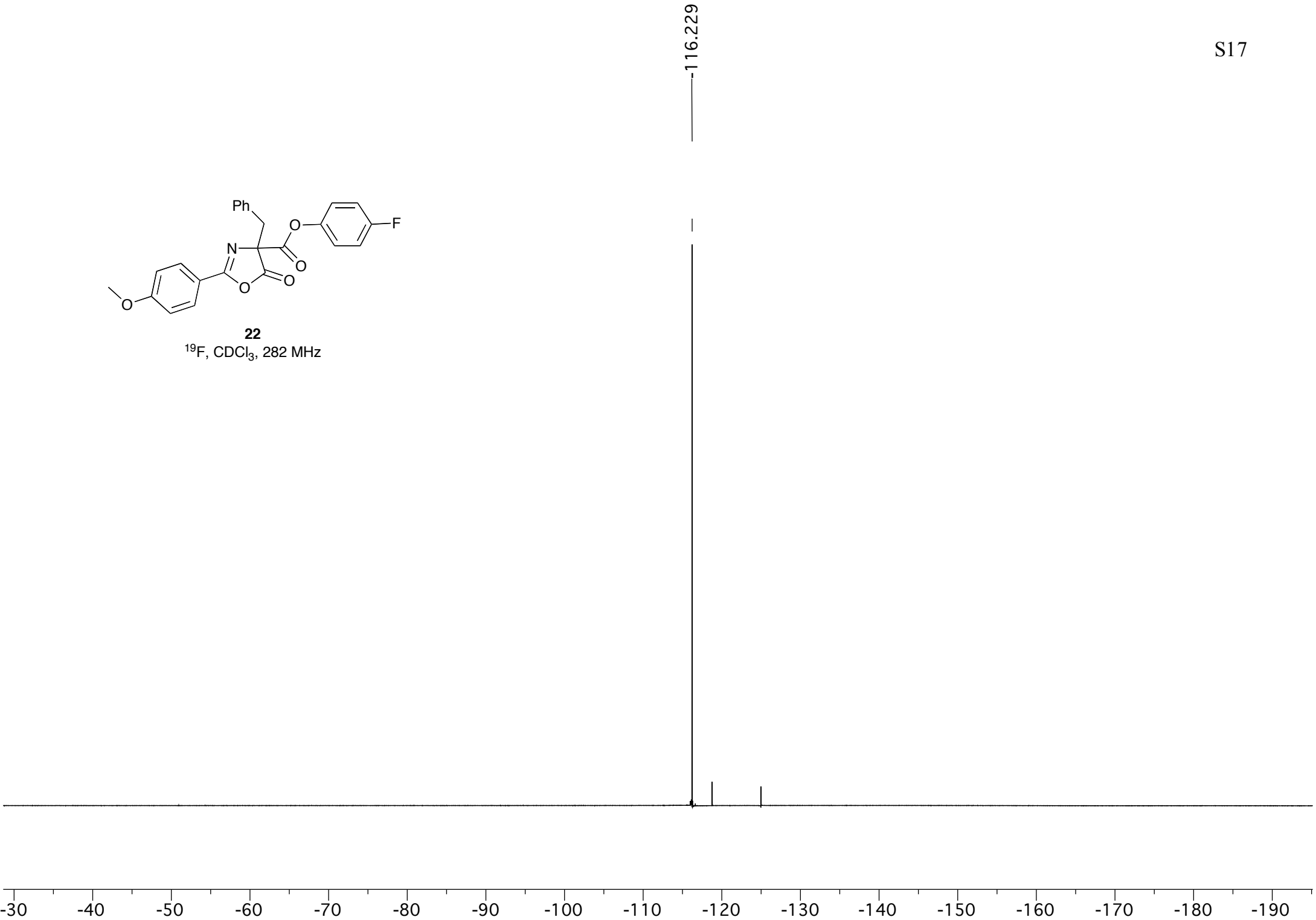
**22** ^1H , CDCl_3 , 400 MHz

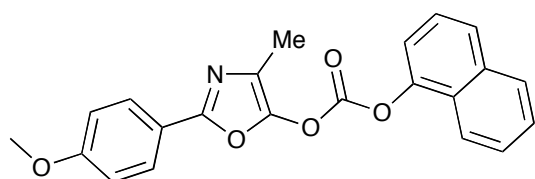
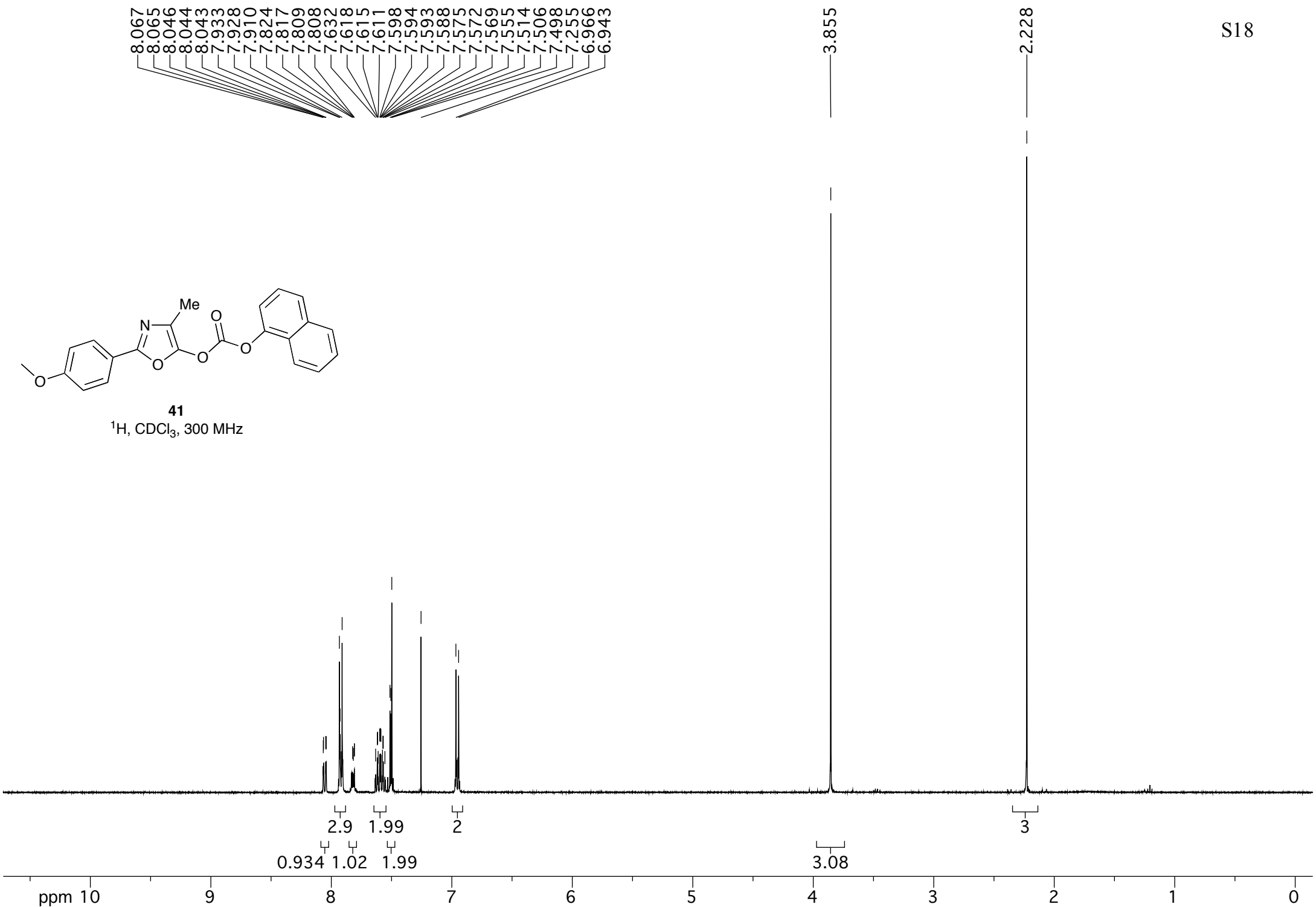
**22** ^{13}C , CDCl_3 , 100 MHz

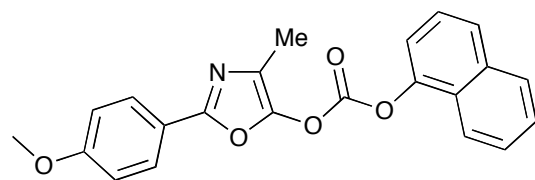


22
 ^{19}F , CDCl_3 , 282 MHz

-116.229

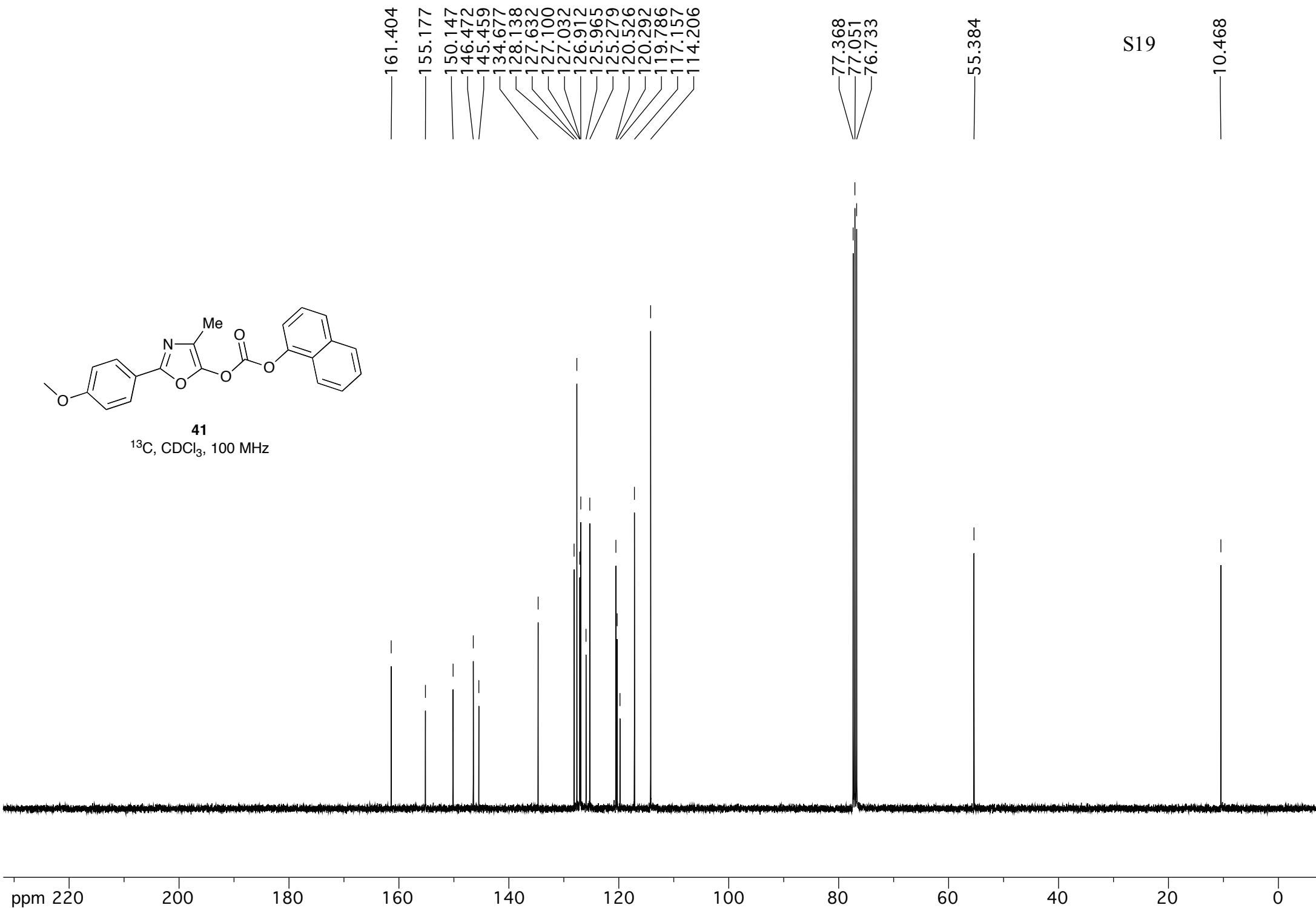


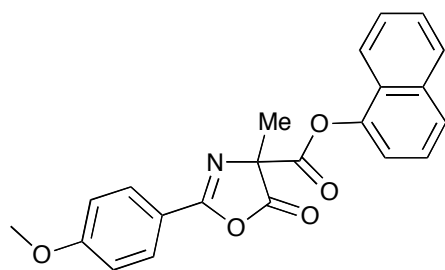
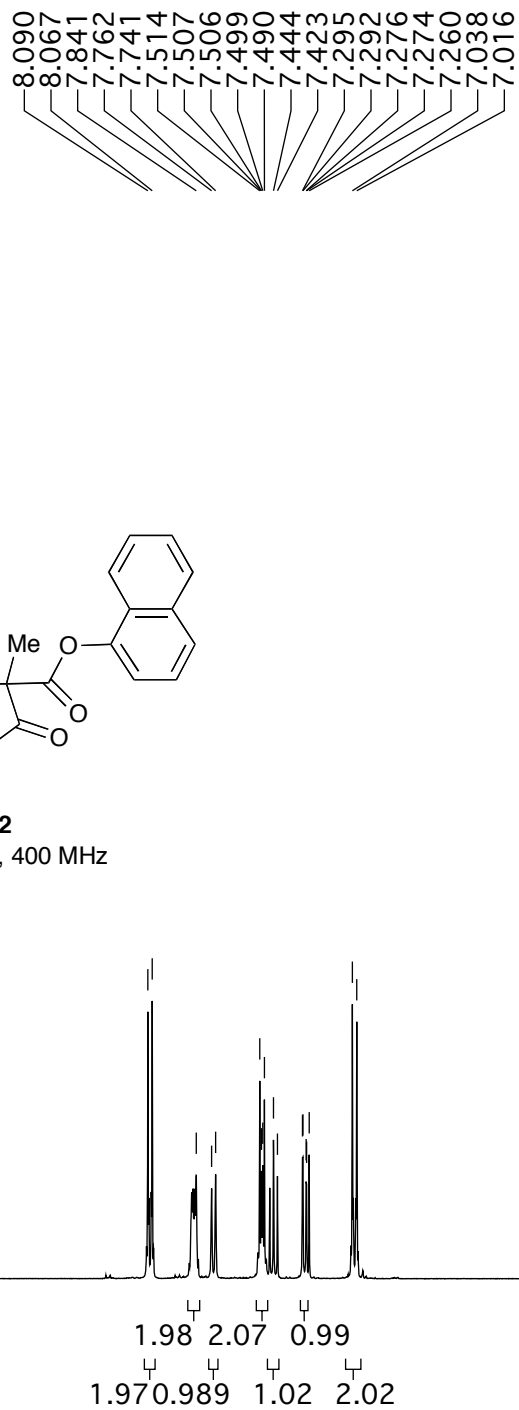
**41** ^1H , CDCl_3 , 300 MHz

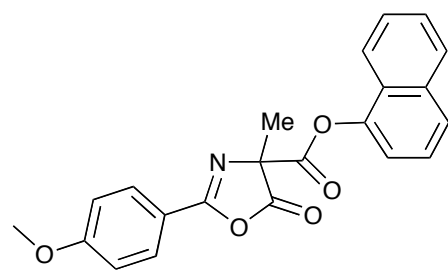


41

^{13}C , CDCl_3 , 100 MHz

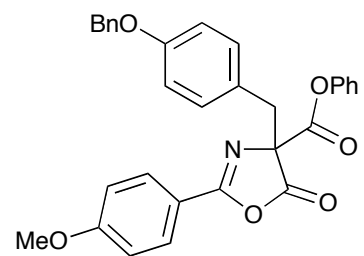
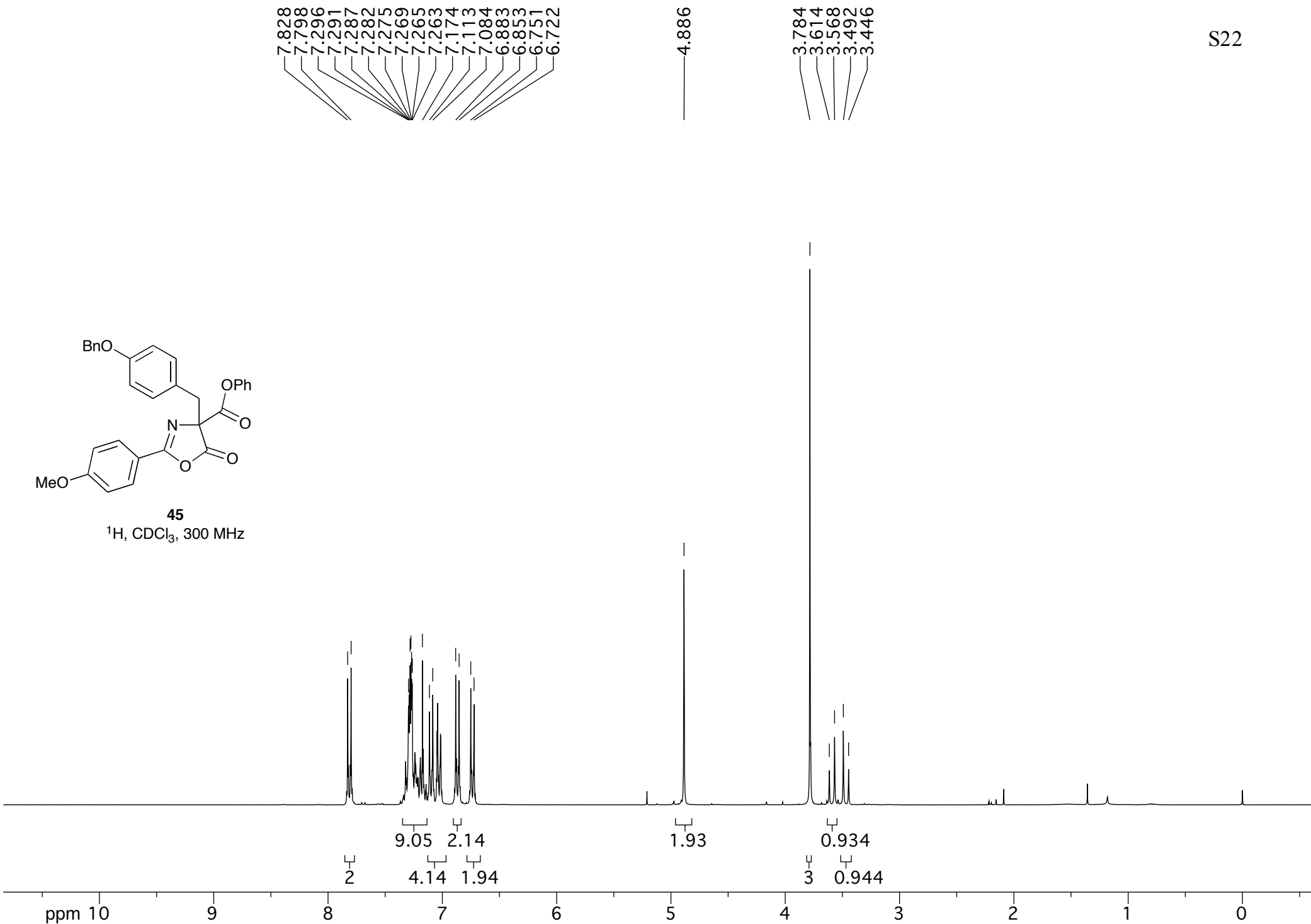


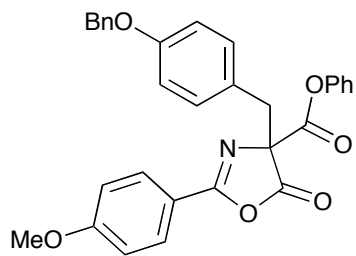
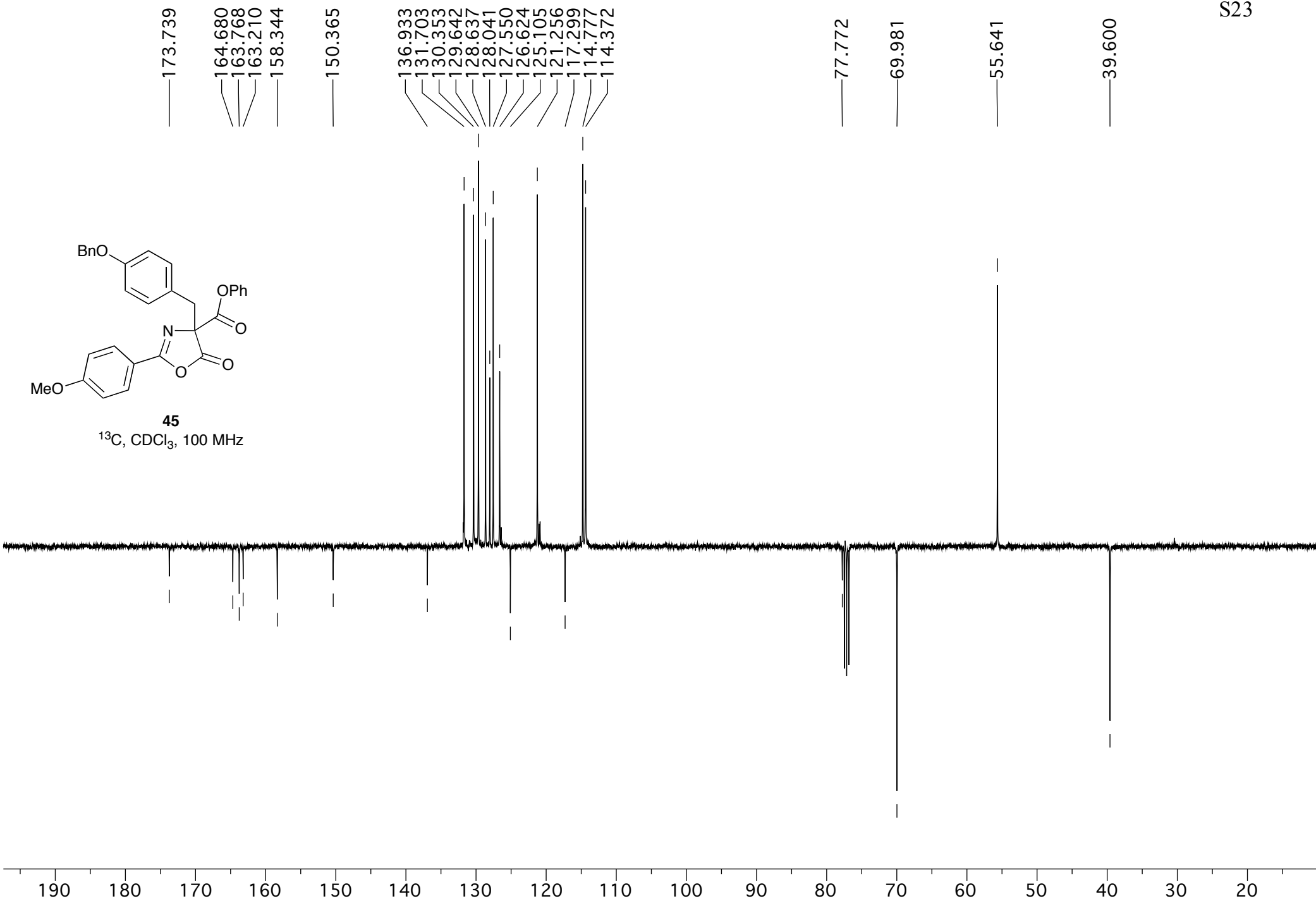
**42** ^1H , CDCl_3 , 400 MHz

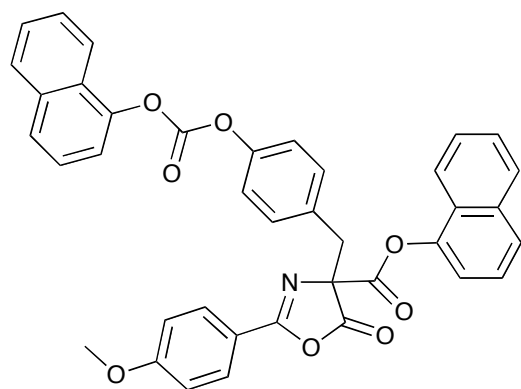


42
 ^{13}C , CDCl_3 , 100 MHz



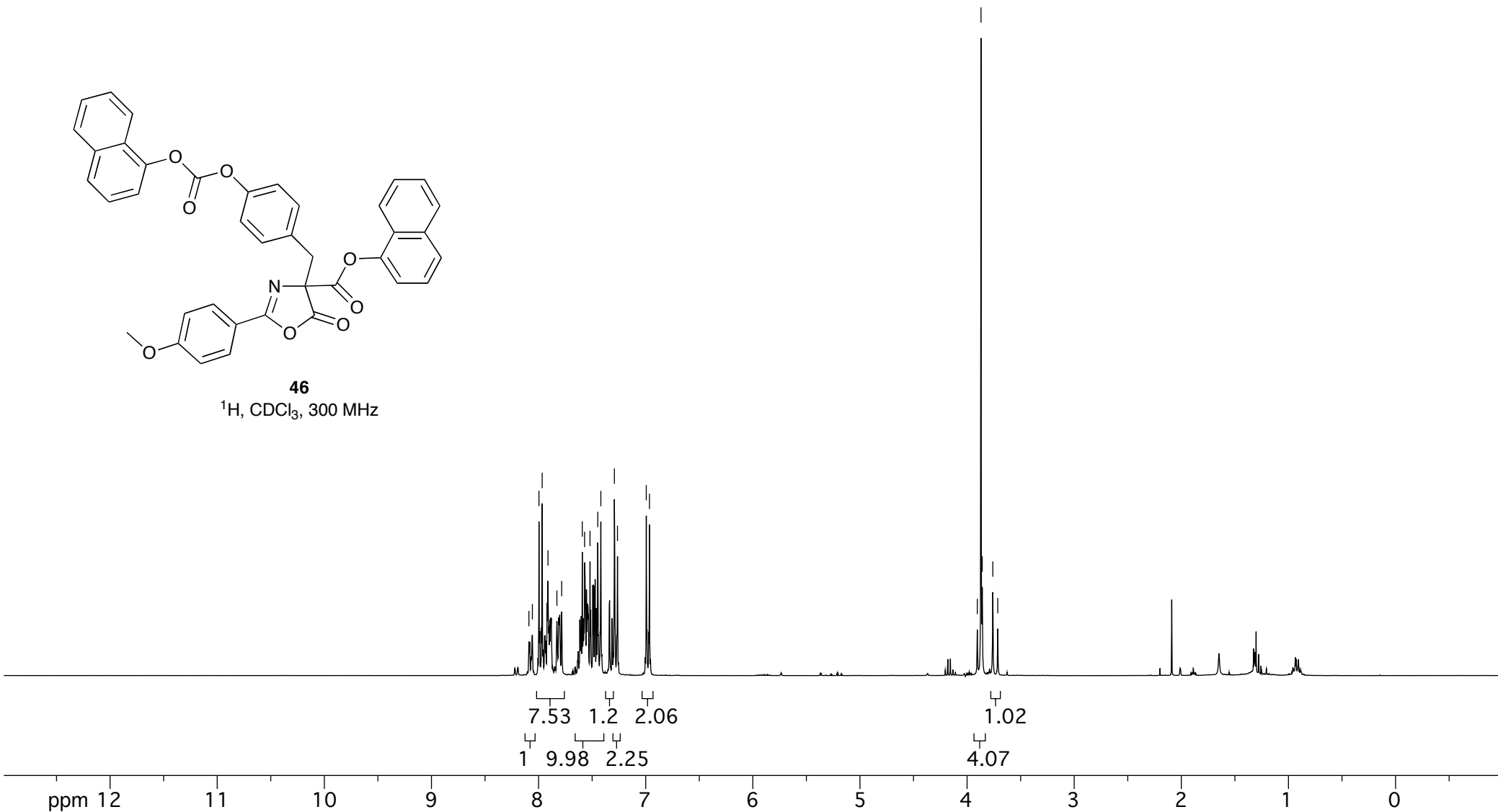
**45** ^1H , CDCl_3 , 300 MHz

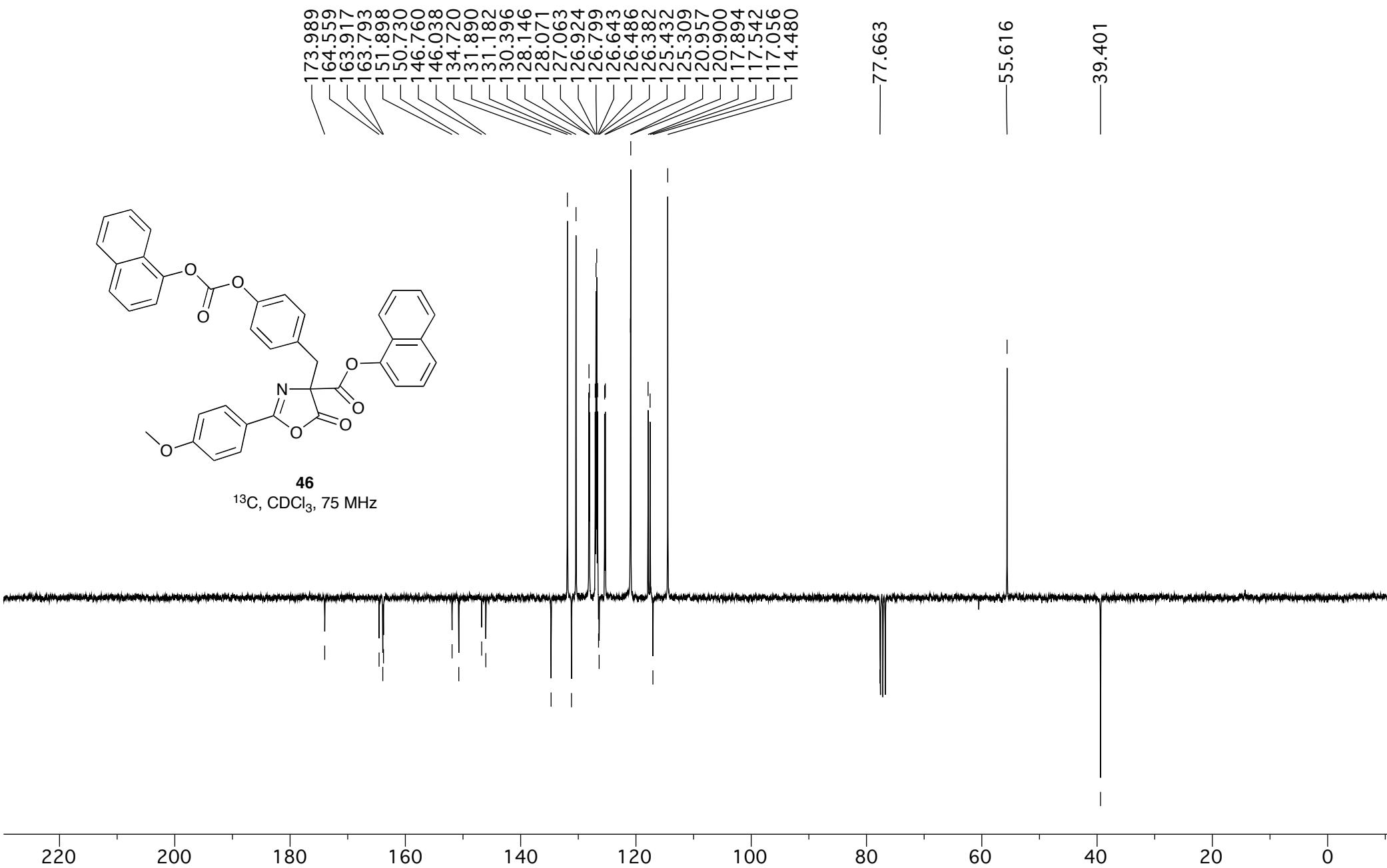
**45** ^{13}C , CDCl_3 , 100 MHz

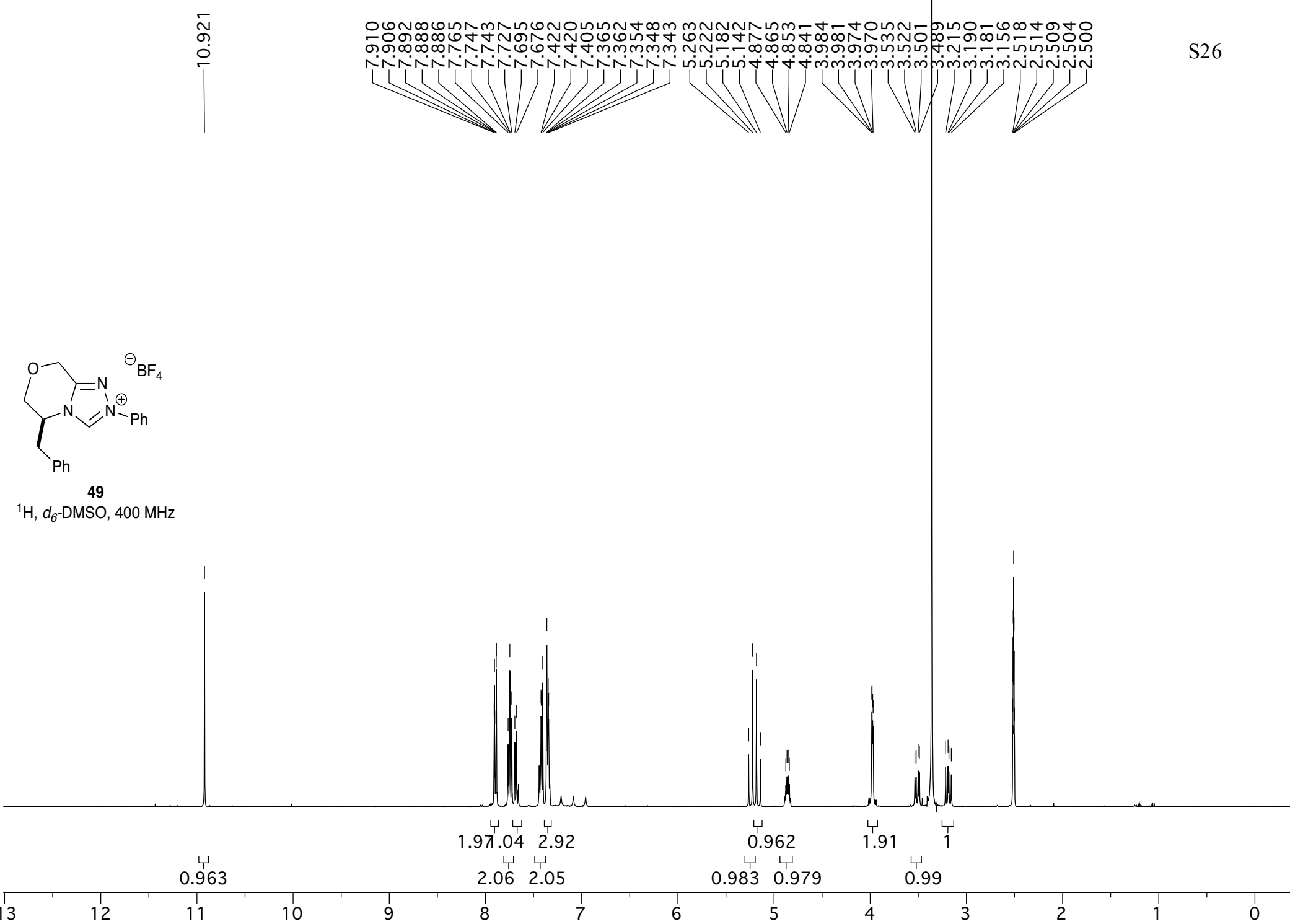
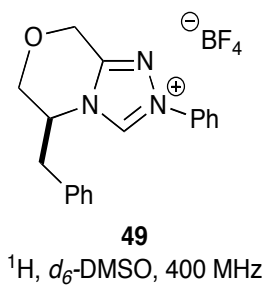
**46**¹H, CDCl₃, 300 MHz

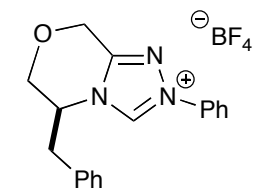
8.090
8.058
7.996
7.966
7.912
7.829
7.784
7.592
7.570
7.520
7.447
7.419
7.292
7.263
6.995
6.965

3.905
3.870
3.859
3.760
3.714







**49** ^{13}C , d_6 -DMSO, 100 MHz

III REFERENCES

- ¹ Thomson, J. E.; Campbell, C. D.; Concellón, C.; Duguet, N.; Rix, K.; Slawin, A. M. Z.; Smith, A. D. *J. Org. Chem.* **2008**, *73*, 2784–2791.
- ² Joannesse, C.; Simal, C.; Concellón, C.; Thomson, J. E.; Campbell, C. D.; Slawin, A. M. Z.; Smith, A. D. *Org. Biomol. Chem.* **2008**, *6*, 2900–2907.
- ³ Shaw, S. A.; Aleman, P.; Vedejs, E. *J. Am. Chem. Soc.* **2003**, *125*, 13368–13369.
- ⁴ Campbell, C. D.; Duguet, N.; Gallagher, K. A.; Thomson, J. E.; Lindsay, A. G.; O'Donoghue, A. C.; Smith, A. D. *Chem. Commun.* **2008**, 3528–3530.
- ⁵ Duguet, N.; Campbell, C. D.; Slawin, A. M. Z.; Smith, A. D. *Org. Biomol. Chem.* **2008**, *6*, 1108–1113.
- ⁶ Kerr, M. S.; de Alaniz, J. R.; Rovis, T. *J. Am. Chem. Soc.* **2002**, *124*, 10298–10299.