

Enantioselective organocatalytic Michael-hemiketalization catalyzed by a
trans-bifunctional indane thiourea catalyst

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Supporting Information

Contents:

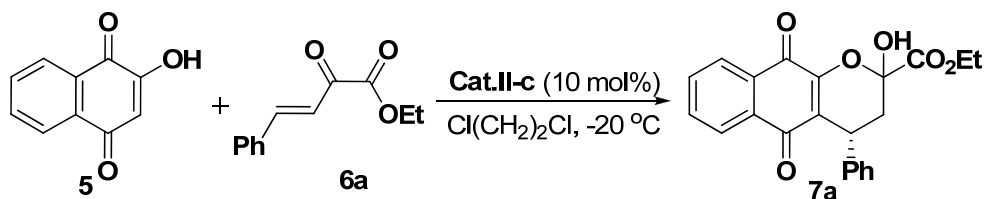
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1. General information

Chemicals and solvents were purchased from commercial suppliers and used as received. ^1H and ^{13}C NMR spectra were recorded on a Bruker ACF300 (300 MHz) or AMX500 (500 MHz) spectrometer. Chemical shifts were reported in parts per million (ppm), and the residual solvent peak was used as an internal reference: proton (chloroform δ 7.26), carbon (chloroform δ 77.0) or tetramethylsilane (TMS δ 0.00) was used as a reference. Multiplicity was indicated as follows: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), dd (doublet of doublet), bs (broad singlet). Coupling constants were reported in Hertz (Hz). Low resolution mass spectra were obtained on a Finnigan/MAT LCQ spectrometer in ESI mode, and a Finnigan/MAT 95XL-T mass spectrometer in EI mode. All high resolution mass spectra were obtained on a Finnigan/MAT 95XL-T mass spectrometer. For thin layer chromatography (TLC), Merck pre-coated TLC plates (Merck 60 F254) were used, and compounds were visualized with a UV light at 254 nm. Further visualization was achieved by staining with KMnO_4 solution, or ninhydrin followed by heating using a heat gun. Flash chromatography separations were performed on Merck 60 (0.040-0.063 mm) mesh silica gel. The enantiomeric excesses of products were determined by chiral phase HPLC analysis. Optical rotations were recorded on Jasco DIP-1000 polarimeter.

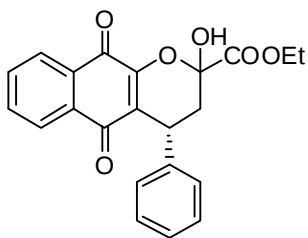
2. Catalyst synthesis and representative reaction procedure for Michael addition reaction

a) Synthesis of Catalyst II-c: Please check the Supporting Information of our published paper: Y. J. Gao, Q. Ren, H. Wu, M. G. Li, J. Wang, *Chem. Commun.* **2010**, DOI: 10.1039/c0cc03489d.



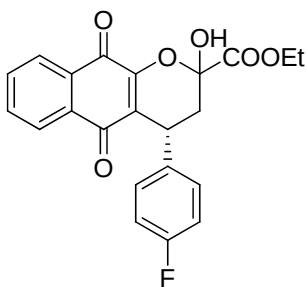
b) General procedure: To a solution of 2-hydroxynaphthalene-1,4-dione **5** (17.5 mg, 0.1 mmol) in 0.45 mL of DCE was added (E)-Ethyl 2-oxo-4-phenylbut-3-enoate **6a** (23 mg, 0.11 mmol) at $-20\text{ }^\circ\text{C}$, followed by adding of 50 μL of pre-cooled catalyst **II-c** solution (4.2 mg in 50 μL of DCE, 0.01 mmol). The mixture was stirred at $-20\text{ }^\circ\text{C}$ for 24 h. The crude product was purified by column chromatography on silica gel, eluted by hexane/EtOAc= 2:1 to afford 36.7 mg (97% yield) of the desired product **7a** as yellow solid.

3. Analytical data of Michael addition reaction products

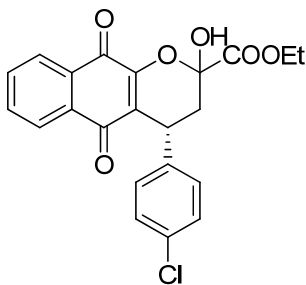


(4R)-Ethyl 2-hydroxy-5,10-dioxo-4-phenyl-3,4,5,10-tetrahydro-2H-benzo[g]chromene-2-carboxylate (7a) (Table 3, entry 1). ^1H NMR (500 MHz, CDCl_3) δ 8.15-8.08 (m, 1H), 8.03-8.0 (m, 0.44 H), 7.90 (dt, $J = 6.6, 3.0$ Hz, 0.48 H), 7.75-7.63 (m, 2.3H), 7.47 (d, $J = 7.3$ Hz, 0.6 H), 7.33-7.18 (m, 4.4 H), 5.01-4.95 (m, 0.74 H), 4.72 (s, 0.16 H), 4.45 (dd, $J = 7.6, 3.5$ Hz, 0.19 H), 4.38-4.25 (m, 2.5 H), 4.15 (dd, $J = 19.2, 9.8$ Hz, 0.32 H), 3.67 (dd, $J = 19.2, 5.7$ Hz, 0.30 H), 2.74 (dd, $J = 14.3, 7.7$ Hz, 0.19 H), 2.46-2.36 (m, 1.27 H), 1.32 (dd, $J = 9.0, 5.5$ Hz, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 192.66, 183.97, 183.24, 183.12, 181.50, 179.30, 178.98, 168.19, 160.68, 153.56, 153.06, 152.57, 142.84, 141.94, 140.99, 135.11, 134.22,

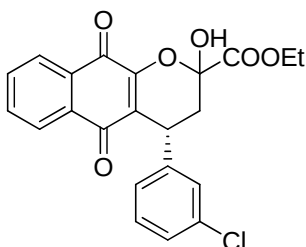
134.15, 133.34, 133.17, 132.96, 132.76, 132.07, 131.89, 130.94, 130.83, 129.07, 128.76, 128.57, 128.33, 128.16, 127.57, 127.22, 127.07, 126.93, 126.68, 126.58, 126.49, 126.43, 126.39, 126.21, 126.05, 125.10, 123.73, 122.98, 95.96, 95.23, 63.54, 63.44, 62.48, 42.24, 37.83, 35.74, 35.28, 34.74, 33.54, 13.93, 13.89; HRMS (ESI) calcd for C₂₂H₁₈NaO₆ (M + Na⁺) 401.1001, found 401.0999; HPLC (Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, λ = 254 nm): t_{major} = 15.37 min, t_{minor} = 20.34 min, *ee* = 96%; $[\alpha]_{\text{D}}^{25}$ = +26.7 (*c* = 1.13 in CHCl₃).



(4R)-Ethyl 4-(4-fluorophenyl)-2-hydroxy-5,10-dioxo-3,4,5,10-tetrahydro-2H-benzo[g]chromene-13-carboxylate (7b) (Table 3, entry 2). ¹H NMR (500 MHz, CDCl₃) δ 8.00 (d, *J* = 94.9 Hz, 2H), 7.69 (d, *J* = 22.1 Hz, 2H), 7.44 (bs, 0.67 H), 7.24 (d, *J* = 23.6 Hz, 1.21 H), 6.98 (bs, 2H), 4.92 (d, *J* = 49.2 Hz, 0.73 H), 4.69 (bs, 0.19 H), 4.32 (t, *J* = 25.7 Hz, 2.63 H), 4.11-4.07 (m, 0.36 H), 3.67 (d, *J* = 14.8 Hz, 0.33 H), 2.72 (bs, 0.19 H), 2.38 (bs, 1 H), 1.34 (t, *J* = 7.1 Hz, 3 H); ¹³C NMR (125 MHz, CDCl₃) δ 183.15, 183.13, 181.41, 179.21, 168.11, 162.52, 160.75, 160.70, 160.57, 153.06, 138.47, 135.18, 134.24, 133.25, 132.07, 130.85, 129.77, 129.18, 128.74, 126.39, 126.25, 115.74, 115.57, 115.27, 95.21, 63.62, 62.53, 42.31, 37.82, 35.07, 34.09, 29.67, 13.93; HRMS (ESI) calcd for C₂₂H₁₇FNao₆ (M + Na⁺) 419.0907, found 419.0895; HPLC (Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, λ = 254 nm): t_{major} = 10.88 min, t_{minor} = 15.99 min, *ee* = 96%; $[\alpha]_{\text{D}}^{25}$ = +27.7 (*c* = 1.37 in CHCl₃).

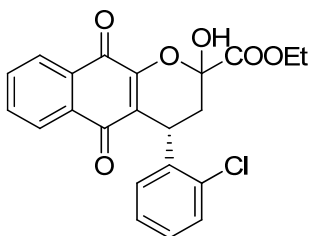


(4R)-Ethyl 4-(4-chlorophenyl)-2-hydroxy-5,10-dioxo-3,4,5,10-tetrahydro-2H-benzo[g]chromene-2-carboxylate (7c) (Table 3, entry 3). ^1H NMR (500 MHz, CDCl_3) δ 8.15-8.01 (m, 1.5 H), 7.92-7.90 (m, 0.48 H), 7.75-7.67 (m, 2 H), 7.40 (t, $J = 8.2$ Hz, 0.69 H), 7.24 (ddd, $J = 29.3, 18.3, 8.0$ Hz, 3.58 H), 4.86 (d, $J = 109.7$ Hz, 1 H), 4.40-4.24 (m, 2.74 H), 4.07 (dd, $J = 19.5, 9.5$ Hz, 0.32 H), 3.68 (dd, $J = 18.9, 5.7$ Hz, 0.31 H), 2.72 (dd, $J = 14.0, 7.7$ Hz, 0.22 H), 2.42-2.30 (m, 1.22 H), 1.34 (t, $J = 7.1$ Hz, 3 H); ^{13}C NMR (75 MHz, CDCl_3) δ 183.37, 179.74, 153.11, 140.67, 134.58, 133.28, 132.49, 132.19, 130.37, 129.01, 128.77, 126.64, 126.30, 123.78, 63.31, 34.29, 13.92; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{17}\text{ClNaO}_6$ ($\text{M} + \text{Na}^+$) 435.0611, found 435.0598; HPLC (Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, $\lambda = 254$ nm): $t_{\text{major}} = 10.77$ min, $t_{\text{minor}} = 15.66$ min, $ee = 97\%$; $[\alpha]_{\text{D}}^{25} = +18.7$ ($c = 1.37$ in CHCl_3).

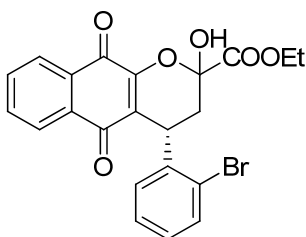


(4R)-Ethyl 4-(3-chlorophenyl)-2-hydroxy-5,10-dioxo-3,4,5,10-tetrahydro-2H-benzo[g]chromene-2-carboxylate (7d) (Table 3, entry 3). ^1H NMR (500 MHz, CDCl_3) δ 8.14-7.91 (m, 2 H), 7.70 (d, $J = 23.3$ Hz, 2 H), 7.40 (d, $J = 51.7$ Hz, 0.68 H), 7.22-7.14 (m, 3.22 H), 4.99 (d, $J = 20.5$ Hz, 0.77 H), 4.79 (s, 0.18 H), 4.40-4.23 (m, 2.69 H), 4.13-4.05 (m, 0.35 H), 3.67 (d, $J = 14.5$ Hz, 0.31 H), 2.71 (d, $J = 6.9$ Hz, 0.19 H), 2.37 (dt, $J = 25.9, 10.2$ Hz, 1.16 H), 1.34 (t, $J = 7.1$ Hz, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 192.28, 182.98, 179.11, 168.02, 153.24, 144.93, 135.19, 134.23, 133.29, 131.99, 130.84, 130.00, 128.28, 127.28, 127.13, 126.95, 126.43, 126.29, 125.61, 124.28, 95.19, 63.62, 62.55, 41.95, 37.60, 35.43, 35.41, 34.90, 34.56, 13.91; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{17}\text{ClNaO}_6$ ($\text{M} + \text{Na}^+$) 435.0611, found 435.0601; HPLC

(Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, λ = 254 nm): t_{major} = 11.07 min, t_{minor} = 13.40 min, ee = 97%; $[\alpha]_{\text{D}}^{25}$ = +7.1 (c = 1.47 in CHCl_3).

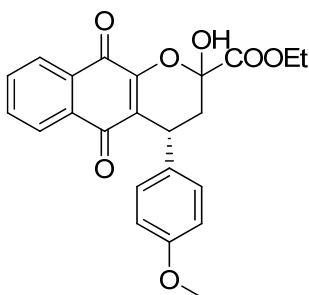


(4S)-Ethyl 4-(2-chlorophenyl)-2-hydroxy-5,10-dioxo-3,4,5,10-tetrahydro-2H-benzo[g]chromene-2-carboxylate (7e) (Table 3, entry 3). ^1H NMR (500 MHz, CDCl_3) δ 8.11-7.92 (m, 2 H), 7.68 (s, 2 H), 7.40 (d, J = 6.0 Hz, 1 H), 7.13 (d, J = 32.8 Hz, 3 H), 4.94-4.74 (m, 2 H), 4.36-4.33 (m, 2 H), 2.69 (s, 0.35 H), 2.42 (d, J = 65.3 Hz, 1.39 H), 1.34 (t, J = 6.6 Hz, 3 H); ^{13}C NMR (75 MHz, CDCl_3) δ 182.89, 179.14, 168.07, 154.00, 153.33, 140.21, 138.83, 134.23, 133.27, 131.89, 130.90, 129.90, 129.60, 127.96, 127.25, 126.44, 126.32, 124.75, 95.91, 95.31, 63.58, 35.11, 32.50, 30.86, 29.66, 13.92; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{17}\text{ClNaO}_6$ ($\text{M} + \text{Na}^+$) 435.0611, found 435.0611; HPLC (Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, λ = 254 nm): t_{major} = 13.36 min, t_{minor} = 16.31 min, ee = 95%; $[\alpha]_{\text{D}}^{25}$ = +29.6 (c = 1.33 in CHCl_3).

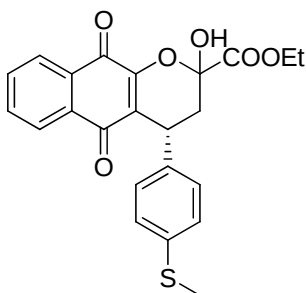


(4S)-Ethyl 4-(2-bromophenyl)-2-hydroxy-5,10-dioxo-3,4,5,10-tetrahydro-2H-benzo[g]chromene-2-carboxylate (7f) (Table 3, entry 6). ^1H NMR (500 MHz, CDCl_3) δ 8.04 (dd, J = 85.6, 26.6 Hz, 2 H), 7.70-7.58 (m, 3 H), 7.19-7.07 (m, 3 H), 4.87-4.72 (m, 2H), 4.34 (dd, J = 15.9, 7.4 Hz, 2 H), 2.55 (dt, J = 212.8, 59.1 Hz, 2 H), 1.34 (t, J = 6.3 Hz, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 182.81, 179.10, 168.03, 154.00, 140.46, 134.19, 133.40, 133.26, 132.91, 131.95, 130.91, 129.72, 128.26, 128.18, 127.92, 126.97, 126.43, 126.30, 124.86, 123.79, 122.63, 95.96, 95.31, 63.61, 60.37, 33.46, 32.76, 21.00, 13.93; HRMS (ESI) calcd

for $C_{22}H_{17}BrNaO_6$ ($M + Na^+$) 479.0106, found 479.0101; HPLC (Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, λ = 254 nm): t_{major} = 14.23 min, t_{minor} = 16.53 min, ee = 94%; $[\alpha]^{25}_D$ = +13.3 (c = 0.87 in $CHCl_3$).

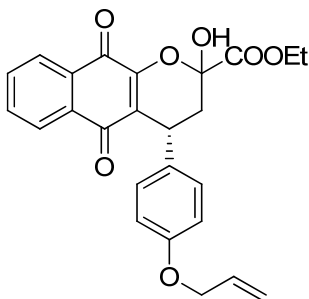


(4R)-Ethyl 2-hydroxy-4-(4-methoxyphenyl)-5,10-dioxo-3,4,5,10-tetrahydro-2H-benzo[g]chromene-2-carboxylate (7g) (Table 3, entry 7). 1H NMR (500 MHz, $CDCl_3$) δ 8.14-7.90 (m, 2 H), 7.68 (d, J = 24.9 Hz, 2 H), 7.39 (d, J = 7.6 Hz, 0.69 H), 7.16 (d, J = 7.9 Hz, 1.26 H), 6.85-6.82 (m, 2 H), 4.86 (d, J = 88.3 Hz, 0.85 H), 4.39-4.23 (m, 2.59 H), 4.09 (dd, J = 18.8, 9.3 Hz, 0.35 H), 3.76 (d, J = 10.7 Hz, 3 H), 3.65 (d, J = 19.2 Hz, 0.41 H), 2.69 (d, J = 7.3 Hz, 0.18 H), 2.39 (d, J = 7.9 Hz, 1 H), 1.33 (t, J = 6.9 Hz, 3 H); ^{13}C NMR (125 MHz, $CDCl_3$) δ 192.78, 184.03, 183.21, 181.56, 179.33, 168.21, 160.77, 158.47, 158.23, 152.89, 152.40, 135.05, 134.72, 134.10, 133.29, 133.11, 132.92, 132.80, 132.14, 130.85, 129.22, 128.61, 128.22, 127.03, 126.36, 126.17, 126.00, 125.32, 124.03, 123.27, 114.19, 113.95, 113.79, 96.02, 95.34, 63.45, 63.37, 62.42, 55.16, 42.44, 37.92, 35.37, 35.00, 33.92, 32.81, 13.91; HRMS (ESI) calcd for $C_{23}H_{20}NaO_7$ ($M + Na^+$) 431.1107, found 431.1101; HPLC (Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, λ = 254 nm): t_{major} = 23.49 min, t_{minor} = 44.30 min, ee = 90%; $[\alpha]^{25}_D$ = +58.7 (c = 1.37 in $CHCl_3$).

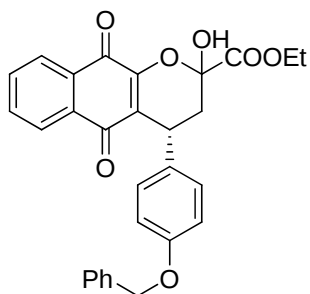


(4R)-Ethyl 2-hydroxy-4-(4-(methylthio)phenyl)-5,10-dioxo-3,4,5,10-tetrahydro-2H-benzo[g]chromene

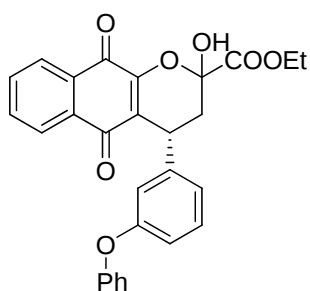
-2-carboxylate (7h) (Table 3, entry 8). ^1H NMR (500 MHz, CDCl_3) δ 8.03 (dd, $J = 74.2, 37.4$ Hz, 2 H), 7.68 (d, $J = 23.0$ Hz, 2 H), 7.39-7.17 (m, 4 H), 5.04 (d, $J = 83.9$ Hz, 1 H), 4.38-4.09 (m, 3 H), 3.67 (d, $J = 17.0$ Hz, 0.27 H), 2.71 (bs, 0.19 H), 2.45-2.36 (m, 4 H), 1.32 (bs, 3 H). ^{13}C NMR (125 MHz, CDCl_3) δ 183.11, 179.26, 168.09, 153.04, 139.68, 136.51, 134.19, 133.15, 130.79, 127.74, 127.05, 126.35, 126.17, 124.89, 95.26, 63.47, 42.07, 37.73, 35.14, 34.21, 15.85, 13.88; HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{20}\text{NaO}_6\text{S}$ ($\text{M} + \text{Na}^+$) 447.0878, found 447.0873; HPLC (Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, $\lambda = 254$ nm): $t_{\text{major}} = 19.14$ min, $t_{\text{minor}} = 31.89$ min, $ee = 93\%$; $[\alpha]_{\text{D}}^{25} = +52.9$ ($c = 1.43$ in CHCl_3).



(4R)-ethyl 4-(4-(allyloxy)phenyl)-2-hydroxy-5,10-dioxo-3,4,5,10-tetrahydro-2H-benzo[g]chromene-2-carboxylate (7i) (Table 3, entry 9). ^1H NMR (500 MHz, CDCl_3) δ 8.16-7.90 (m, 1.66 H), 7.92-7.91 (m, 0.42 H), 7.75-7.58 (m, 2.34 H), 7.38 (d, $J = 8.5$ Hz, 0.73 H), 7.16 (d, $J = 8.2$ Hz, 1 H), 6.87-6.82 (m, 2 H), 6.03 (dt, $J = 24.3, 9.6$ Hz, 1 H), 5.32 (ddd, $J = 27.4, 24.3, 10.7$ Hz, 2 H), 4.94 (dd, $J = 9.6, 5.8$ Hz, 0.29 H), 4.74 (s, 0.35 H), 4.45-4.22 (m, 4.56 H), 4.10 (dd, $J = 19.1, 9.6$ Hz, 0.34 H), 3.64 (dd, $J = 19.2, 6.0$ Hz, 0.35 H), 2.38 (d, $J = 9.5$ Hz, 1 H), 1.34 (t, $J = 7.1$ Hz, 3 H); ^{13}C NMR (75 MHz, CDCl_3) δ 192.79, 184.06, 183.24, 181.60, 179.36, 168.27, 157.55, 157.35, 152.89, 152.35, 135.11, 134.91, 134.15, 133.36, 133.28, 133.17, 132.98, 132.16, 131.72, 130.87, 130.70, 129.25, 128.63, 128.24, 127.09, 126.40, 126.23, 126.06, 125.40, 124.06, 118.04, 117.62, 115.01, 114.79, 114.58, 114.45, 95.91, 95.25, 68.81, 63.60, 63.48, 62.47, 42.47, 37.89, 35.20, 35.03, 33.94, 32.69, 29.68, 13.95; HRMS (ESI) calcd for $\text{C}_{25}\text{H}_{22}\text{NaO}_7$ ($\text{M} + \text{Na}^+$) 457.1263, found 457.1251; HPLC (Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, $\lambda = 254$ nm): $t_{\text{major}} = 20.60$ min, $t_{\text{minor}} = 39.69$ min, $ee = 90\%$; $[\alpha]_{\text{D}}^{25} = +53.9$ ($c = 1.5$ in CHCl_3).

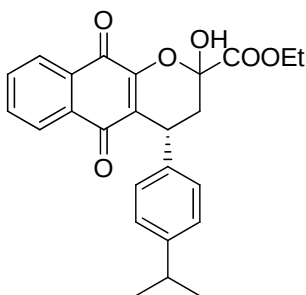


(4R)-Ethyl 4-(4-(benzyloxy)phenyl)-2-hydroxy-5,10-dioxo-3,4,5,10-tetrahydro-2H-benzo[g]chromene-2-carboxylate (7j) (Table 3, entry 10). ^1H NMR (500 MHz, CDCl_3) δ 8.15-7.90 (m, 2 H), 7.74-7.63 (m, 2.59 H), 7.43-7.28 (m, 6.20 H), 7.17-7.15 (m, 1.62 H), 6.93-6.88 (m, 2 H), 5.02-4.93 (m, 3 H), 4.41-4.23 (m, 2.76 H), 4.10 (dd, $J = 19.1, 9.6$ Hz, 0.35 H), 3.64 (dd, $J = 19.2, 6.0$ Hz, 0.32 H), 2.73-2.68 (m, 0.19 H), 2.43-2.37 (m, 1 H), 1.35-1.31 (m, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 192.78, 184.04, 183.22, 181.57, 179.32, 168.23, 160.77, 157.76, 157.56, 152.91, 152.39, 137.05, 135.07, 135.02, 134.19, 134.12, 133.35, 133.31, 133.14, 132.94, 132.83, 132.16, 130.87, 129.28, 129.12, 128.90, 128.66, 128.54, 128.52, 128.38, 128.27, 128.11, 127.92, 127.88, 127.48, 127.40, 127.24, 127.06, 126.49, 126.38, 126.20, 126.03, 125.32, 124.01, 115.09, 114.90, 114.67, 95.98, 95.31, 70.01, 63.51, 63.42, 62.43, 42.46, 37.91, 35.29, 35.04, 33.95, 32.78, 13.93, 13.91; HRMS (ESI) calcd for $\text{C}_{29}\text{H}_{24}\text{NaO}_7$ ($\text{M} + \text{Na}^+$) 507.1420, found 507.1414; HPLC (Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, $\lambda = 254$ nm): $t_{\text{major}} = 24.43$ min, $t_{\text{minor}} = 41.02$ min, $ee = 90\%$; $[\alpha]_D^{25} = +42.2$ ($c = 1.53$ in CHCl_3).

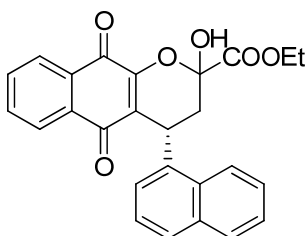


(4R)-Ethyl 2-hydroxy-5,10-dioxo-4-(3-phenoxyphenyl)-3,4,5,10-tetrahydro-2H-benzo[g]chromene-2-carboxylate (7k) (Table 3, entry 11). ^1H NMR (500 MHz, CDCl_3) δ 8.01 (ddd, $J = 47.3, 11.5, 7.4$ Hz, 2 H), 7.73-7.66 (m, 2 H), 7.31-7.18 (m, 3.55 H), 7.08-6.80 (m, 5.3 H), 4.96 (t, $J = 64.9$ Hz, 1 H), 4.41-4.07 (m, 2.7 H), 4.10 (dd, $J = 18.8, 9.3$ Hz, 0.28 H), 3.66 (dd, $J = 19.2, 5.0$ Hz, 0.27 H), 2.72 (dd, $J = 13.9, 7.6$

Hz, 0.19 H), 2.45-2.33 (m, 1.26 H), 1.33 (t, $J = 7.1$ Hz, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 183.19, 179.16, 168.10, 157.01, 153.01, 144.89, 134.16, 133.15, 129.63, 126.22, 123.16, 118.75, 116.90, 95.24, 63.45, 42.08, 37.67, 34.63, 29.64, 13.90; HRMS (ESI) calcd for $\text{C}_{28}\text{H}_{22}\text{NaO}_7$ ($\text{M} + \text{Na}^+$) 493.1263, found 493.1258; HPLC (Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, $\lambda = 254$ nm): $t_{\text{major}} = 14.18$ min, $t_{\text{minor}} = 17.62$ min, $ee = 96\%$; $[\alpha]_{\text{D}}^{25} = +11.7$ ($c = 1.73$ in CHCl_3).

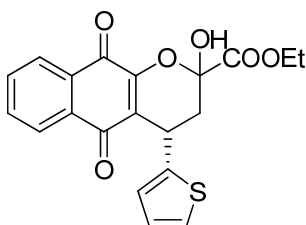


(4R)-Ethyl 2-hydroxy-4-(4-isopropylphenyl)-5,10-dioxo-3,4,5,10-tetrahydro-2H-benzo[g]chromene-2-carboxylate (71) (Table 3, entry 12). ^1H NMR (500 MHz, CDCl_3) δ 8.09-7.91 (m, 2 H), 7.68 (d, $J = 20.8$ Hz, 2 H), 7.38 (s, 1 H), 7.15 (bs, 3 H), 4.81 (d, $J = 154.8$ Hz, 1 H), 4.42-4.14 (m, 2.89 H), 3.64 (d, $J = 15.8$ Hz, 0.31 H), 2.78 (d, $J = 72.2$ Hz, 1 H), 2.40 (d, $J = 8.2$ Hz, 1 H), 1.32 (bs, 3 H), 1.22 (bs, 6 H); ^{13}C NMR (125 MHz, CDCl_3) δ 192.81, 184.01, 183.14, 181.59, 179.35, 168.25, 160.76, 152.98, 147.47, 147.00, 139.96, 138.28, 135.05, 134.09, 133.11, 132.91, 132.17, 130.89, 129.13, 128.08, 127.40, 127.11, 126.79, 126.62, 126.40, 126.19, 126.02, 125.35, 123.95, 104.97, 95.34, 63.46, 62.42, 42.41, 37.94, 35.44, 34.32, 33.61, 23.87, 13.91; HRMS (ESI) calcd for $\text{C}_{25}\text{H}_{24}\text{NaO}_6$ ($\text{M} + \text{Na}^+$) 443.1471, found 443.1465; HPLC (Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, $\lambda = 254$ nm): $t_{\text{major}} = 14.02$ min, $t_{\text{minor}} = 18.78$ min, $ee = 94\%$; $[\alpha]_{\text{D}}^{25} = +39.6$ ($c = 1.0$ in CHCl_3).

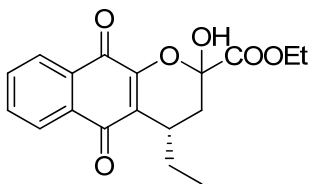


(4R)-Ethyl 2-hydroxy-4-(naphthalen-1-yl)-5,10-dioxo-3,4,5,10-tetrahydro-2H-benzo[g]chromene-2-carboxylate (7m) (Table 3, entry 13). ^1H NMR (500 MHz, CDCl_3) δ 8.28-7.07 (m, 11 H), 5.71 (s, 0.13 H),

5.02 (d, $J = 96.2$ Hz, 1 H), 4.54 (bs, 0.20 H), 4.23-4.13 (m, 2 H), 3.63 (d, $J = 16.7$ Hz, 0.14 H), 2.80 (dd, $J = 13.7, 8.0$ Hz, 0.23 H), 2.53-2.39 (m, 1 H), 1.21 (ddd, $J = 14.5, 10.6, 6.3$ Hz, 3 H). ^{13}C NMR (125 MHz, CDCl_3) δ 192.63, 183.10, 183.00, 179.25, 178.94, 168.15, 154.00, 139.31, 136.93, 135.14, 134.23, 134.15, 133.36, 133.21, 132.97, 132.08, 131.95, 131.00, 130.92, 130.53, 129.41, 129.13, 127.72, 127.53, 127.22, 126.47, 126.42, 126.29, 126.09, 125.73, 125.52, 125.42, 125.28, 125.07, 123.40, 122.58, 95.96, 95.53, 63.44, 62.49, 60.38, 42.95, 36.94, 33.40, 31.95, 29.65, 21.00, 14.15, 13.92, 13.80; HRMS (ESI) calcd for $\text{C}_{26}\text{H}_{20}\text{NaO}_6$ ($\text{M} + \text{Na}^+$) 451.1158, found 451.1152; HPLC (Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, $\lambda = 254$ nm): $t_{\text{major}} = 21.03$ min, $t_{\text{minor}} = 29.86$ min, $ee = 91\%$; $[\alpha]_{\text{D}}^{25} = +45.2$ ($c = 1.5$ in CHCl_3).

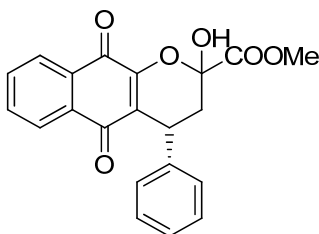


(4S)-Ethyl 2-hydroxy-5,10-dioxo-4-(thiophen-2-yl)-3,4,5,10-tetrahydro-2H-benzo[g]chromene-2-carboxylate (7n) (Table 3, entry 14). ^1H NMR (500 MHz, CDCl_3) δ 8.18-8.04 (m, 1.62 H), 7.97-7.95 (m, 0.27 H), 7.71 (ddd, $J = 29.0, 15.3, 6.9$ Hz, 2.42 H), 7.14 (d, $J = 5.0$ Hz, 0.81 H), 7.06 (d, $J = 3.5$ Hz, 0.53 H), 6.94-6.88 (m, 1.25 H), 5.29 (dd, $J = 9.5, 6.0$ Hz, 0.46 H), 4.86-4.62 (m, 0.77 H), 4.33 (ddd, $J = 21.4, 10.6, 5.2$ Hz, 2 H), 4.12 (dd, $J = 19.1, 9.3$ Hz, 0.55 H), 3.77 (dd, $J = 19.2, 6.0$ Hz, 0.51 H), 2.73 (dd, $J = 14.5, 7.3$ Hz, 0.16 H), 2.59-2.48 (m, 0.73 H), 1.34 (t, $J = 7.1$ Hz, 3 H); ^{13}C NMR (75 MHz, CDCl_3) δ 192.16, 183.61, 183.08, 181.45, 160.53, 152.46, 143.49, 135.22, 134.25, 133.43, 133.23, 133.07, 132.73, 130.79, 129.12, 128.77, 127.15, 126.81, 126.64, 126.56, 126.43, 126.28, 126.16, 125.40, 125.14, 124.89, 124.46, 124.25, 123.78, 123.54, 122.97, 95.64, 95.26, 63.64, 62.56, 43.43, 38.24, 34.94, 30.94, 29.88, 28.36, 13.93; HRMS (ESI) calcd for $\text{C}_{20}\text{H}_{16}\text{NaO}_6\text{S}$ ($\text{M} + \text{Na}^+$) 407.0565, found 407.0560; HPLC (Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, $\lambda = 254$ nm): $t_{\text{major}} = 19.84$ min, $t_{\text{minor}} = 26.87$ min, $ee = 95\%$; $[\alpha]_{\text{D}}^{25} = +52.6$ ($c = 1.17$ in CHCl_3).



(4S)-Ethyl 4-ethyl-2-hydroxy-5,10-dioxo-3,4,5,10-tetrahydro-2H-benzo[g]chromene-2-carboxylate (7o)

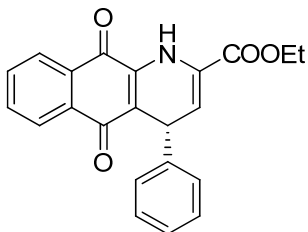
(Table 3, entry 15). ^1H NMR (500 MHz, CDCl_3) δ 8.09 (dd, $J = 20.8, 9.8$ Hz, 2 H), 7.76-7.66 (m, 2 H), 4.68 (d, $J = 84.2$ Hz, 0.55 H), 4.40-4.26 (m, 2H), 3.63 (bs, 0.47 H), 3.50 (dd, $J = 18.0, 8.8$ Hz, 0.49 H), 3.30 (dd, $J = 18.1, 5.5$ Hz, 0.50 H), 3.10-2.98 (m, 0.57 H), 2.33-2.15 (m, 1.39 H), 1.90-1.61 (m, 4.47 H), 1.35 (ddd, $J = 46.0, 25.4, 18.3$ Hz, 3.83 H), 1.05 (t, $J = 7.4$ Hz, 1.17 H), 0.96-0.84 (m, 2.52 H); ^{13}C NMR (75 MHz, CDCl_3) δ 193.49, 184.26, 183.85, 181.27, 179.19, 169.00, 160.91, 153.19, 151.68, 135.03, 134.02, 133.18, 133.09, 132.90, 132.08, 130.91, 129.23, 127.03, 126.57, 126.29, 126.23, 126.07, 125.74, 124.34, 95.96, 63.47, 62.38, 42.30, 32.97, 32.21, 29.21, 28.79, 27.73, 25.79, 25.23, 24.97, 13.94, 12.13, 11.94, 10.65; HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{18}\text{NaO}_6$ ($\text{M} + \text{Na}^+$) 353.1001, found 353.0996; HPLC (Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, $\lambda = 254$ nm): $t_{\text{major}} = 10.50$ min, $t_{\text{minor}} = 12.10$ min, *ee* = 98%; $[\alpha]_{\text{D}}^{25} = +18.5$ ($c = 1.17$ in CHCl_3).



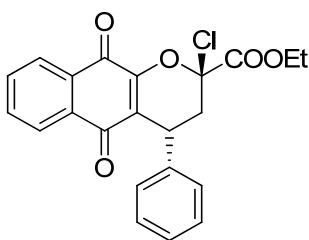
(4R)-Methyl 2-hydroxy-5,10-dioxo-4-phenyl-3,4,5,10-tetrahydro-2H-benzo[g]chromene-2-carboxylate (7p)

(Table 3, entry 16). ^1H NMR (500 MHz, CDCl_3) δ 8.03 (dd, $J = 77.6, 37.5$ Hz, 2 H), 7.68 (d, $J = 23.6$ Hz, 2 H), 7.46 (bs, 0.67 H), 7.30-7.22 (m, 4.4 H), 4.89 (d, $J = 117.6$ Hz, 1 H), 4.43-4.11 (m, 1 H), 3.86 (d, $J = 15.1$ Hz, 3 H), 3.67 (d, $J = 16.4$ Hz, 0.28 H), 2.72 (bs, 0.18 H), 2.46-2.37 (m, 1.22 H); ^{13}C NMR (75 MHz, CDCl_3) δ 192.20, 183.98, 183.09, 181.46, 179.31, 168.60, 152.94, 142.65, 140.95, 135.09, 134.17, 133.33, 133.16, 132.95, 132.73, 132.05, 130.78, 129.05, 128.72, 128.56, 128.34, 128.12, 127.53, 127.20, 127.05, 126.91, 126.67, 126.39, 126.21, 126.04, 125.02, 123.65, 95.39, 53.79, 52.94, 42.32, 37.86, 35.69, 35.46, 34.64, 33.58, 29.62; HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{16}\text{NaO}_6$ ($\text{M} + \text{Na}^+$) 387.0845, found 387.0839; HPLC

(Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, λ = 254 nm): t_{major} = 12.76 min, t_{minor} = 18.37 min, ee = 96%; $[\alpha]_{\text{D}}^{25}$ = +28.7 (c = 1.17 in CHCl_3).



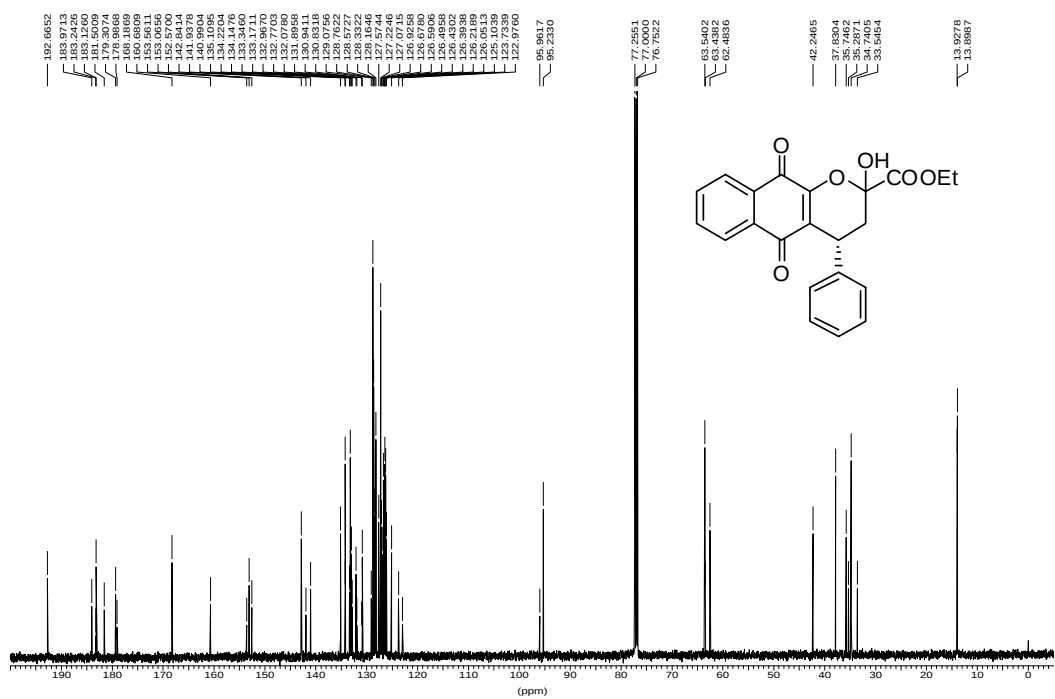
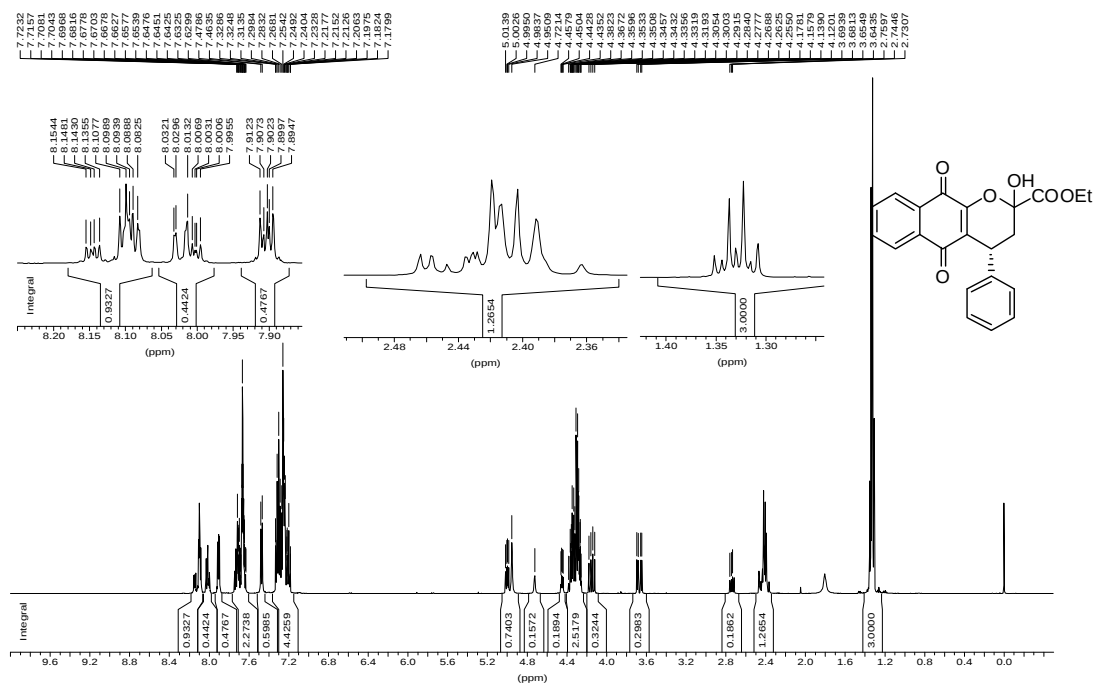
(R)-Ethyl 5,10-dioxo-4-phenyl-1,4,5,10-tetrahydrobenzo[g]quinoline-2-carboxylate (9). A solution of **7a** (38 mg, 0.1 mmol) and ammonium acetate (77 mg, 1.0 mmol) in ethanol (0.5 mL) was heated at 70 °C for 30 min. The solvent was evaporated and the mixture was purified by column chromatography on silica gel, eluted by hexane/EtOAc= 10:1 to provide **9** as a red solid (15 mg, 42% yield). ^1H NMR (500 MHz, CDCl_3) δ 8.07 (dd, J = 7.6, 0.9 Hz, 1 H), 8.00 (dd, J = 7.7, 1.1 Hz, 1 H), 7.70-7.61 (m, 3 H), 7.36 (dd, J = 8.2, 0.9 Hz, 2 H), 7.31 (dd, J = 10.4, 5.0 Hz, 2 H), 7.21 (dd, J = 10.2, 4.3 Hz, 1 H), 6.18 (dd, J = 5.4, 1.9 Hz, 1 H), 5.05 (d, J = 5.4 Hz, 1 H), 4.38-4.27 (m, 2 H), 1.34 (t, J = 7.1 Hz, 3 H); ^{13}C NMR (125 MHz, CDCl_3) δ 182.48, 180.11, 162.13, 144.91, 138.86, 134.69, 132.96, 132.49, 130.36, 128.77, 128.25, 127.22, 126.49, 126.30, 126.10, 115.44, 113.69, 62.04, 38.50, 14.13; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{17}\text{NNaO}_4$ ($\text{M} + \text{Na}^+$) 382.1055, found 382.1050; HPLC (Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, λ = 254 nm): t_{minor} = 20.62 min, t_{major} = 26.70 min, ee = 75%; $[\alpha]_{\text{D}}^{25}$ = +180.7 (c = 0.27 in CHCl_3).



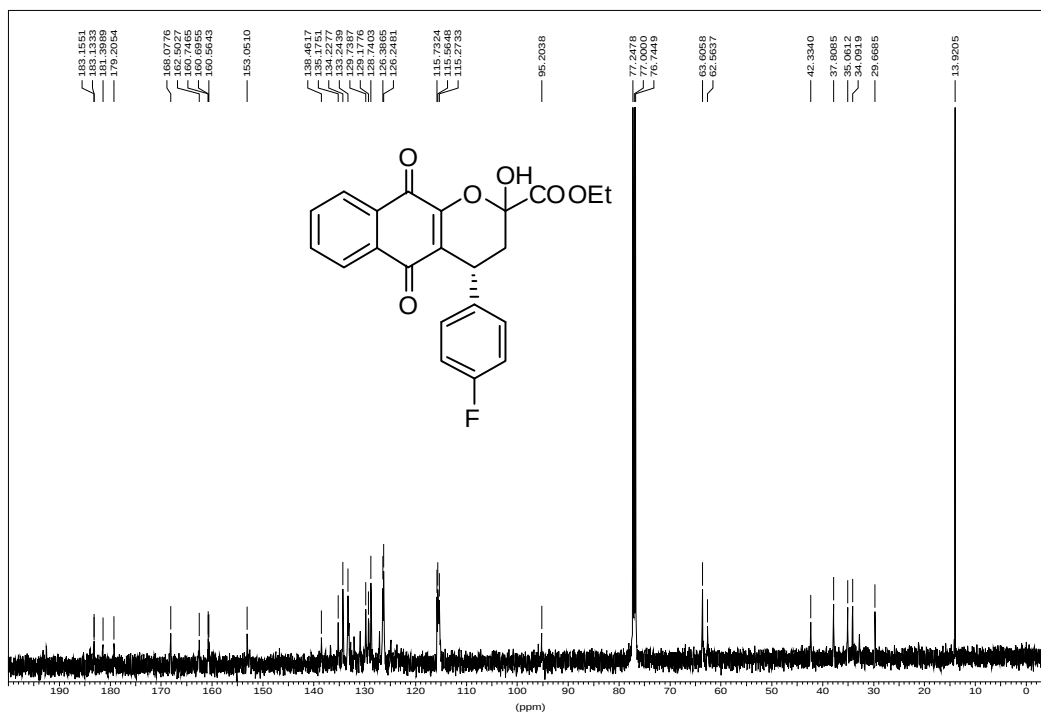
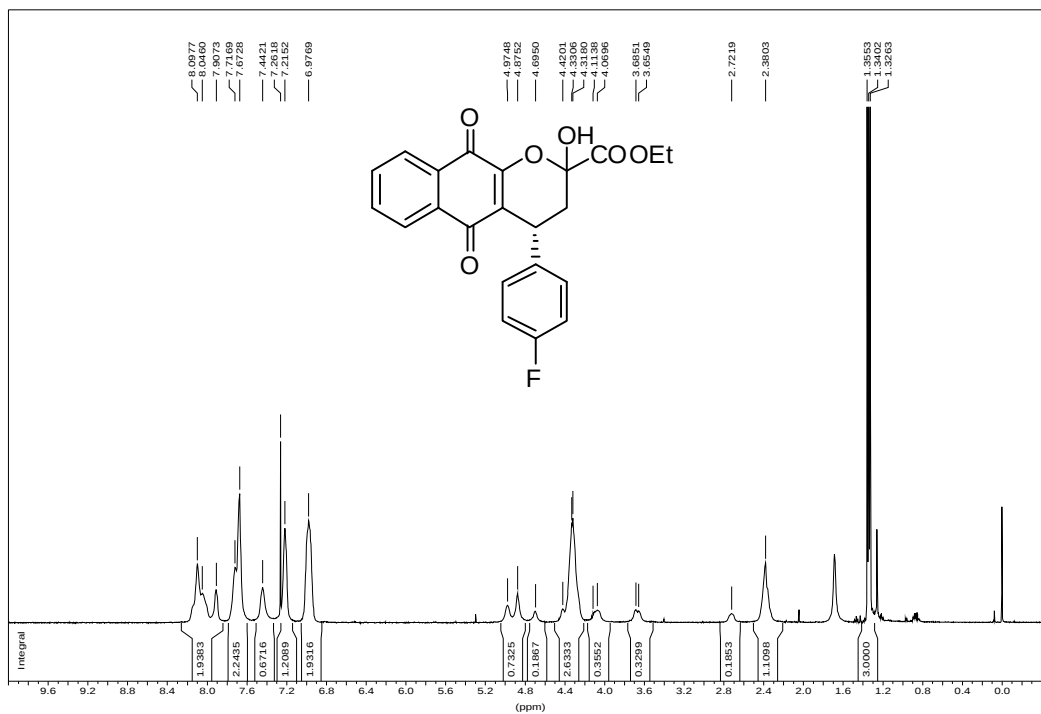
(2R,4R)-Ethyl 2-chloro-5,10-dioxo-4-phenyl-3,4,5,10-tetrahydro-2H-benzo[g]chromene-2-carboxylate (10). To a solution of **7a** (76 mg, 0.2 mmol) in DCM (1 mL) were added triethylamine (0.1 mL) and methanesulfonyl chloride (0.05 mL) at room temperature. After 12 h, the mixture was purified by column chromatography on silica gel, eluted by hexane/EtOAc= 7:1 to provide **10** as yellow oil (32 mg, 40 % yield). ^1H NMR (500 MHz, CDCl_3) δ 8.20-8.18 (m, 1 H), 7.80-7.98 (m, 1 H), 7.76-7.70 (m, 2 H), 7.31-7.28 (m, 2 H), 7.23 (dd, J = 7.1, 5.2 Hz, 3 H), 4.38-4.30 (m, 3 H), 3.00 (dd, J = 14.7, 7.7 Hz, 1 H), 2.83 (dd,

$J = 14.7, 6.1$ Hz, 1 H), 1.34 (t, $J = 7.3$ Hz, 3 H); ^{13}C NMR (126 MHz, CDCl_3) δ 182.91, 177.67, 165.40, 153.12, 141.28, 134.37, 133.69, 131.84, 130.85, 128.65, 127.51, 126.92, 126.64, 126.62, 123.77, 91.46, 63.62, 40.77, 34.96, 13.89; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{17}\text{ClNaO}_5$ ($\text{M} + \text{Na}^+$) 419.0662, found 419.0659; HPLC (Chiralpak IC, *i*-propanol/hexane = 20/80, flow rate 1.0 mL/min, $\lambda = 254$ nm): $t_{\text{minor}} = 32.20$ min, $t_{\text{major}} = 37.06$ min, $ee = 94\%$; $[\alpha]_{\text{D}}^{25} = +149.7$ ($c = 0.33$ in CHCl_3).

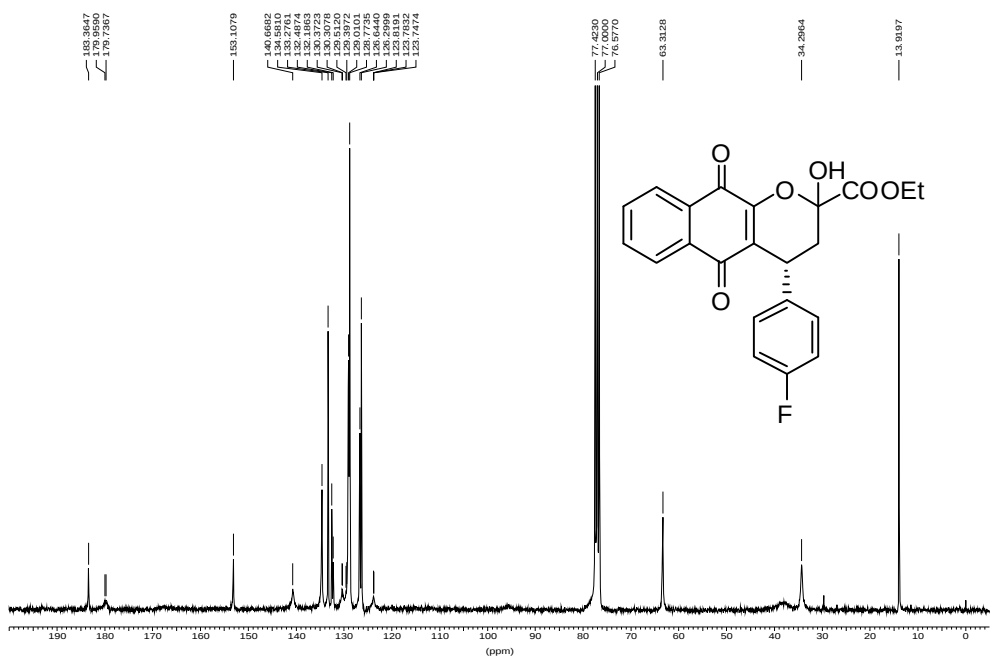
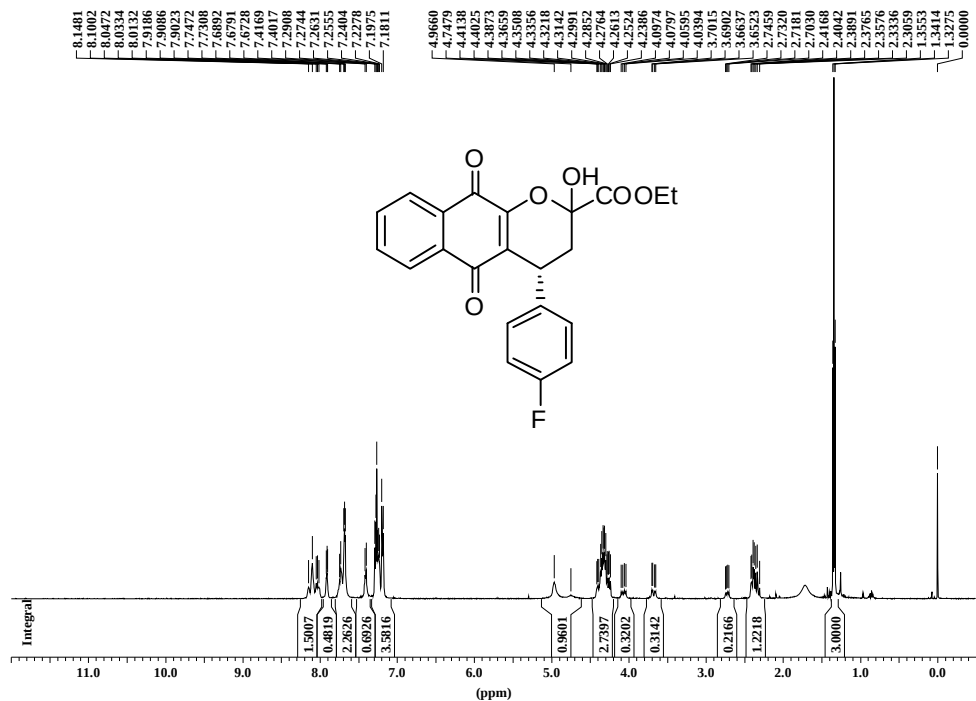
Compound 7a



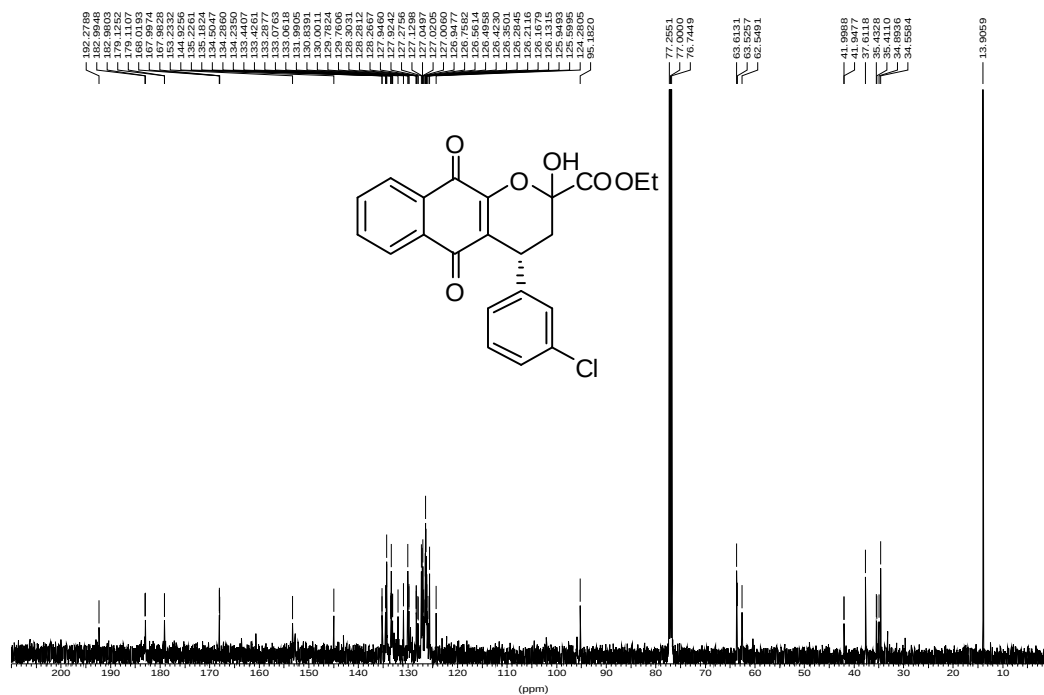
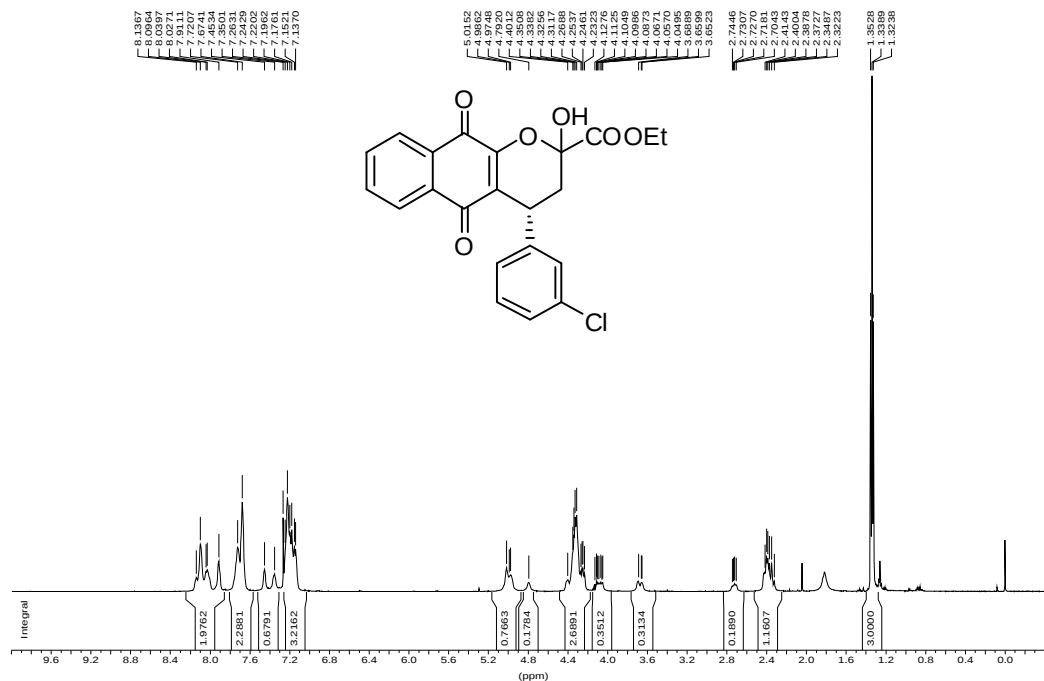
Compound **7b**



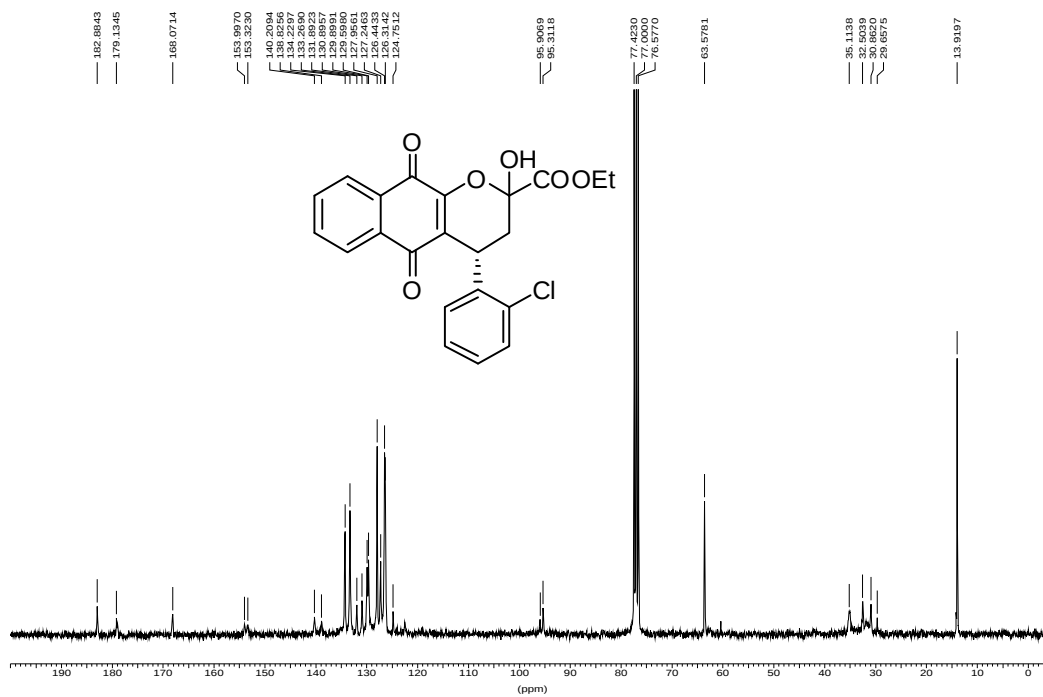
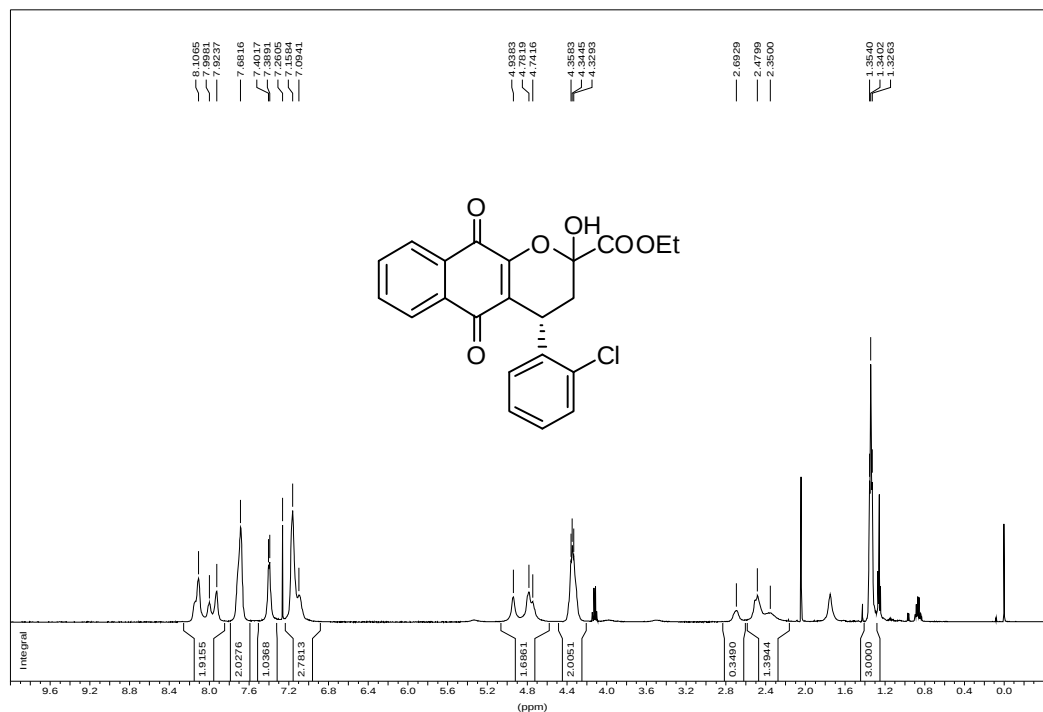
Compound **7c**



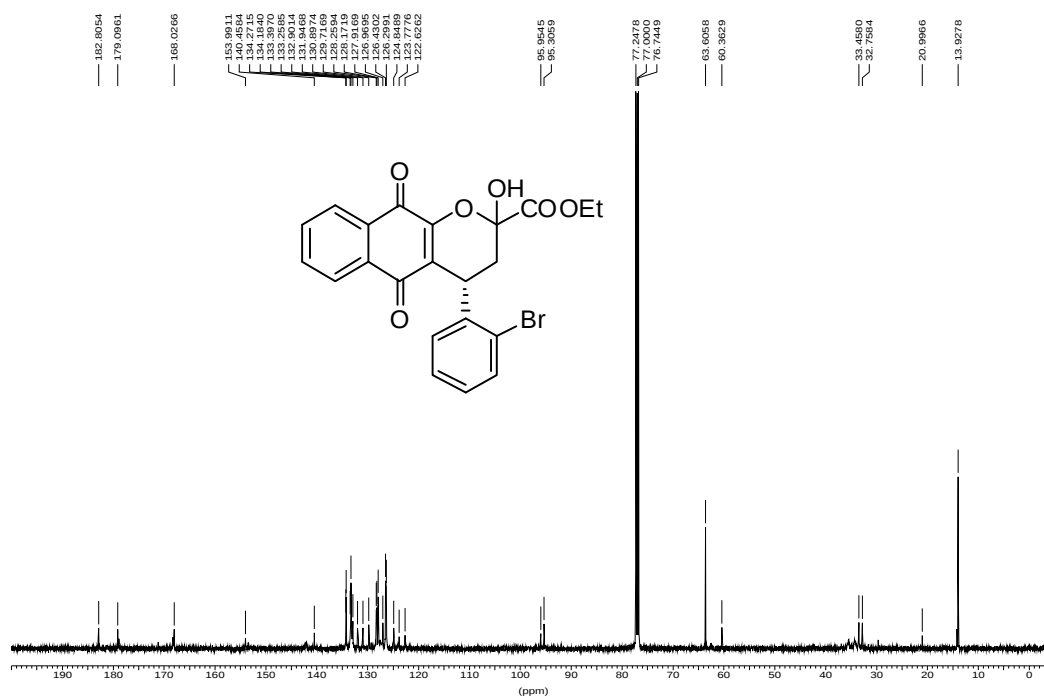
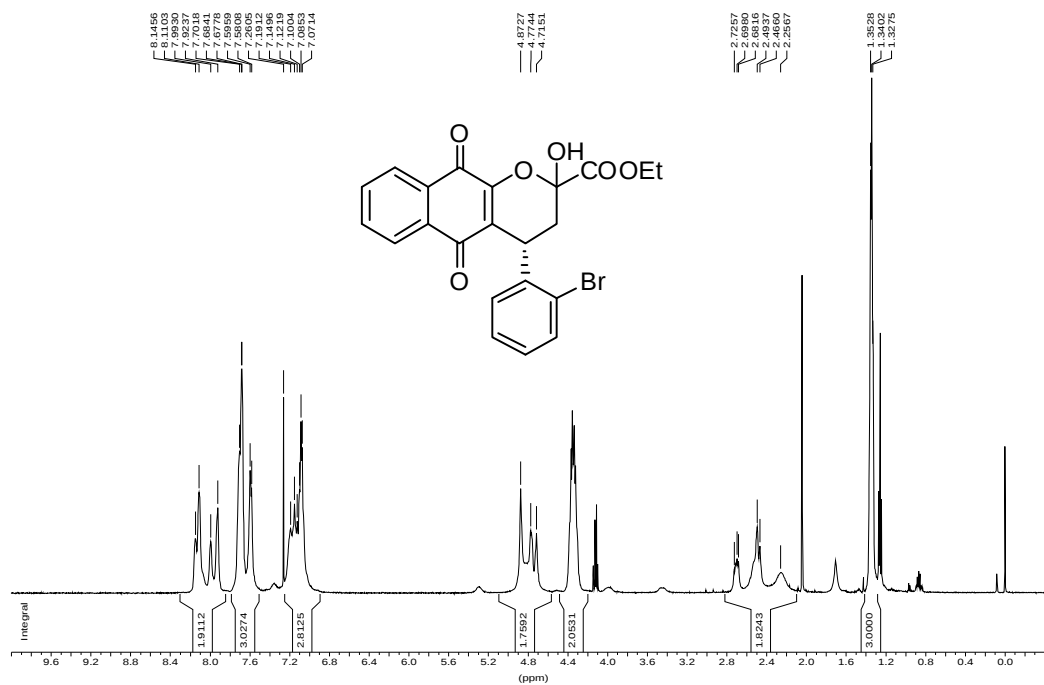
Compound **7d**



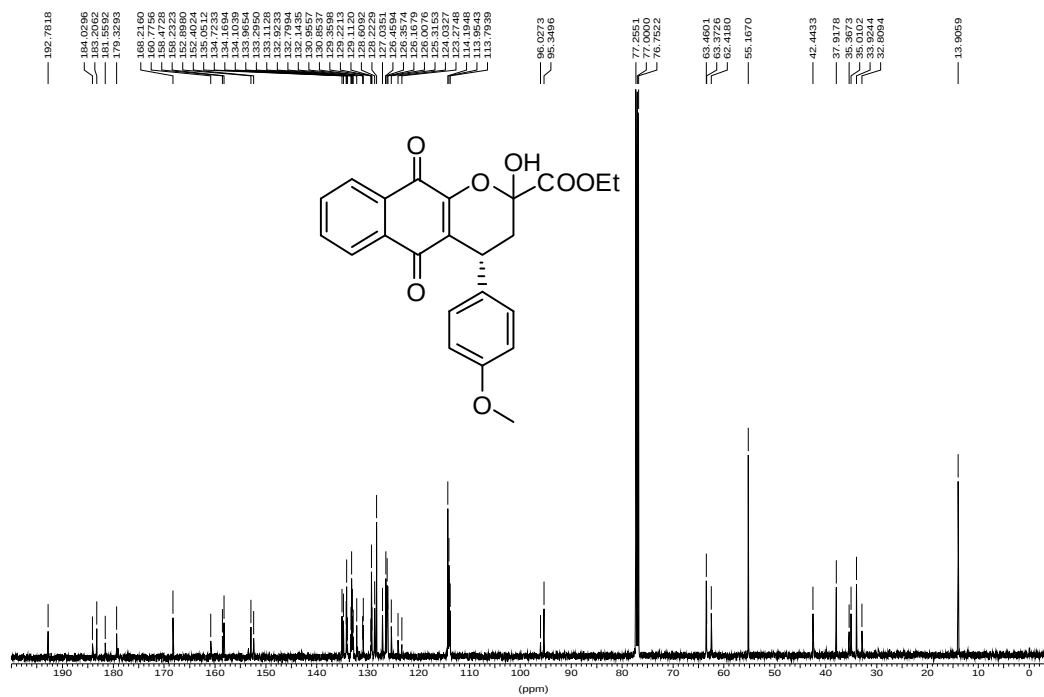
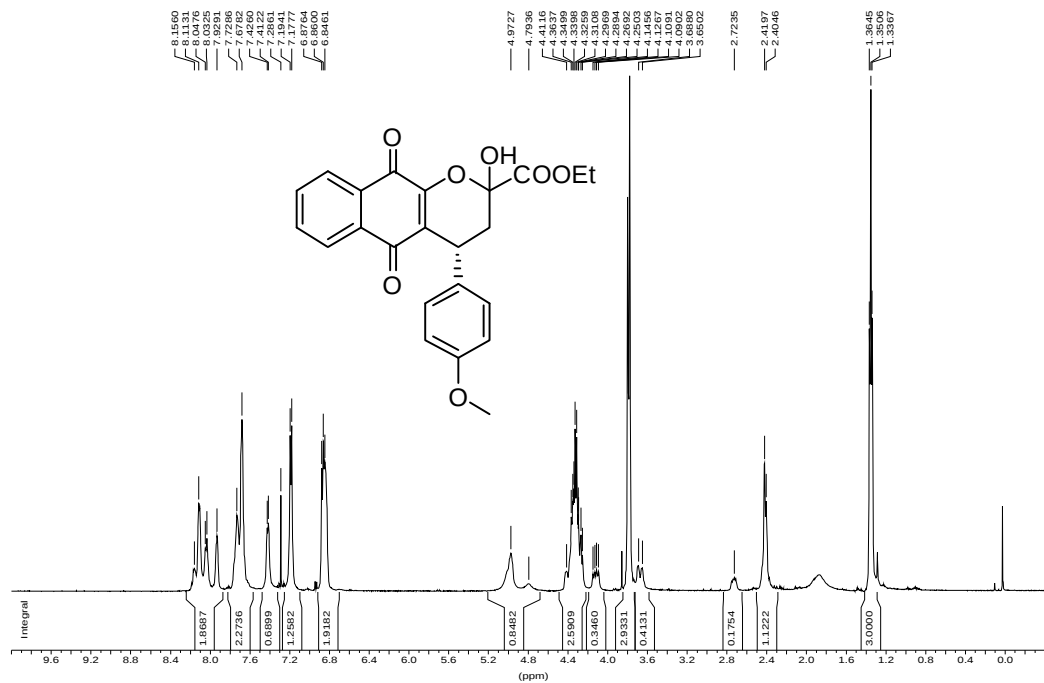
Compound **7e**



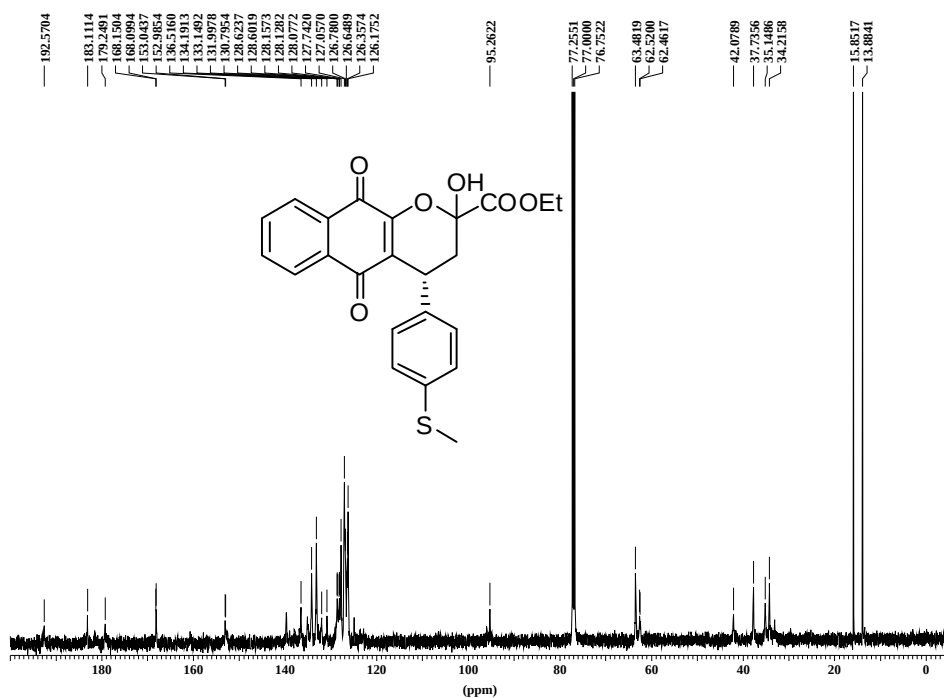
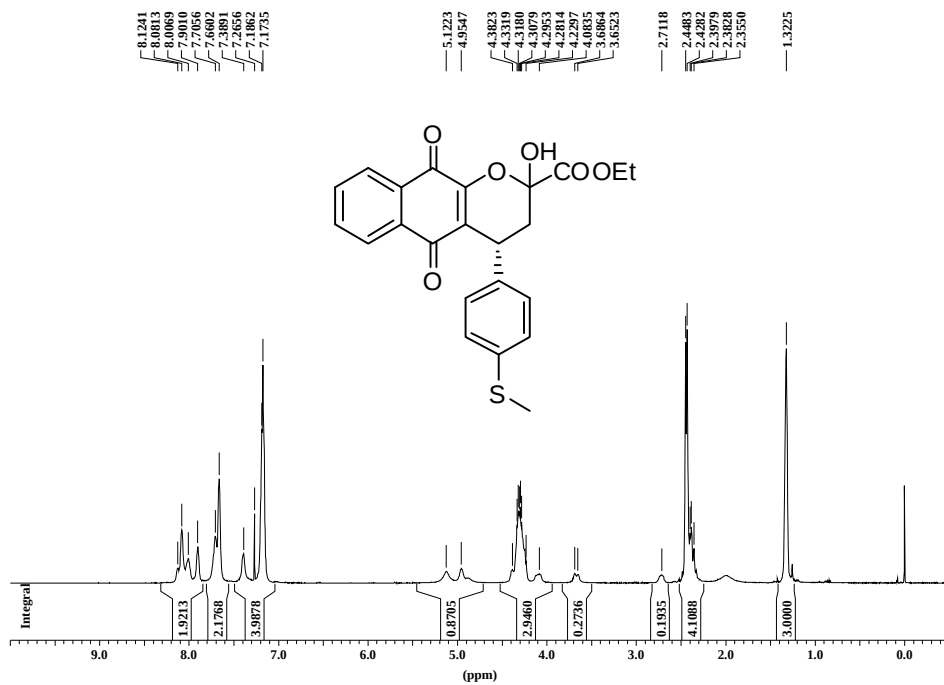
Compound **7f**



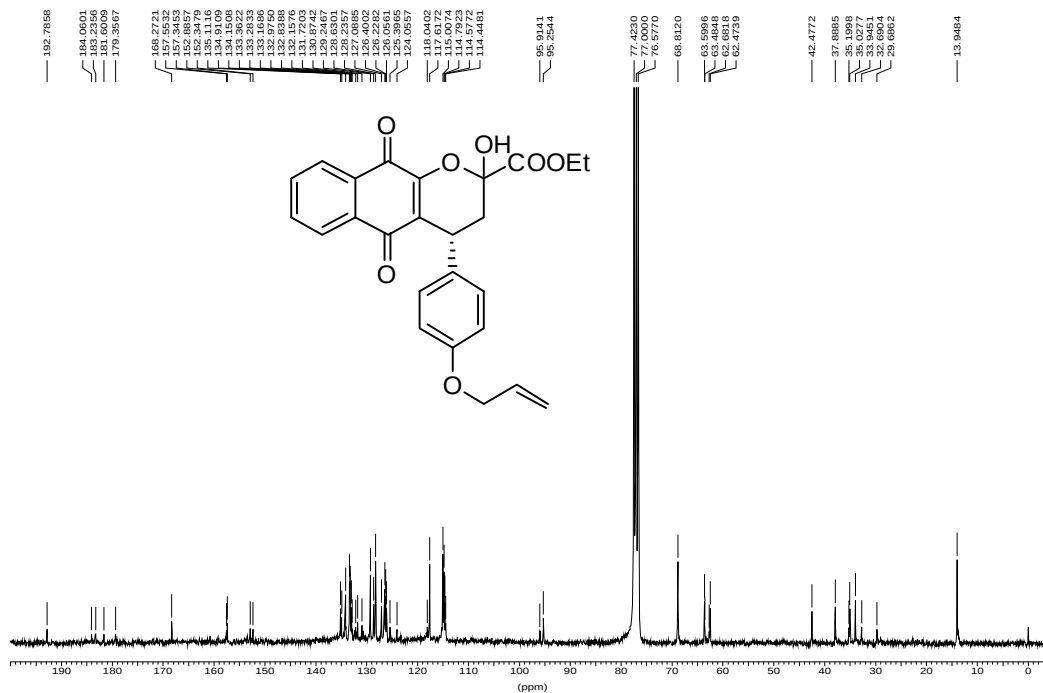
Compound **7g**



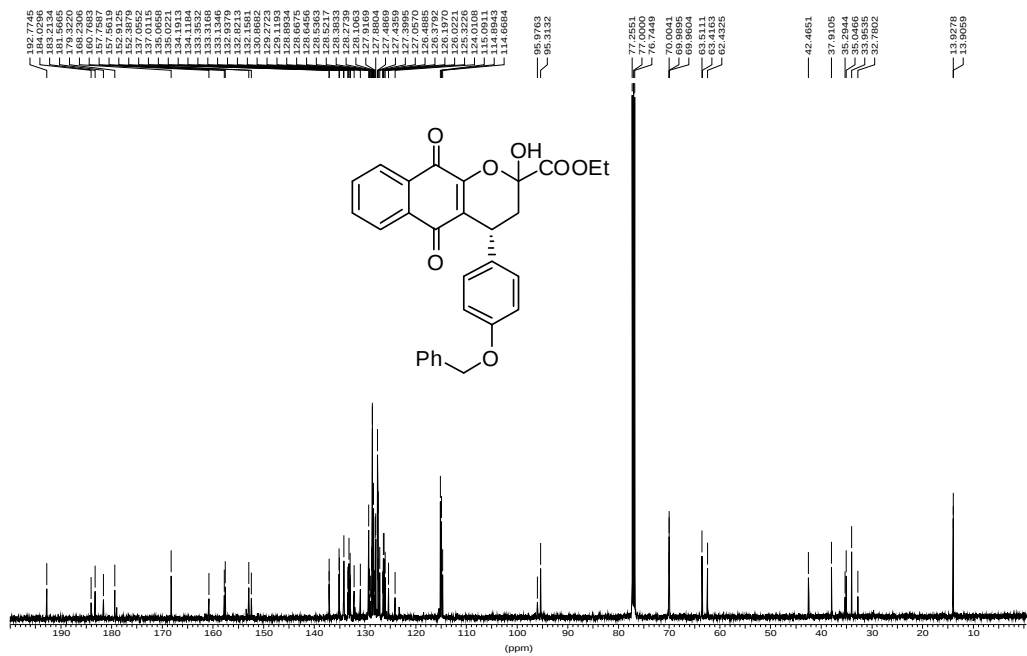
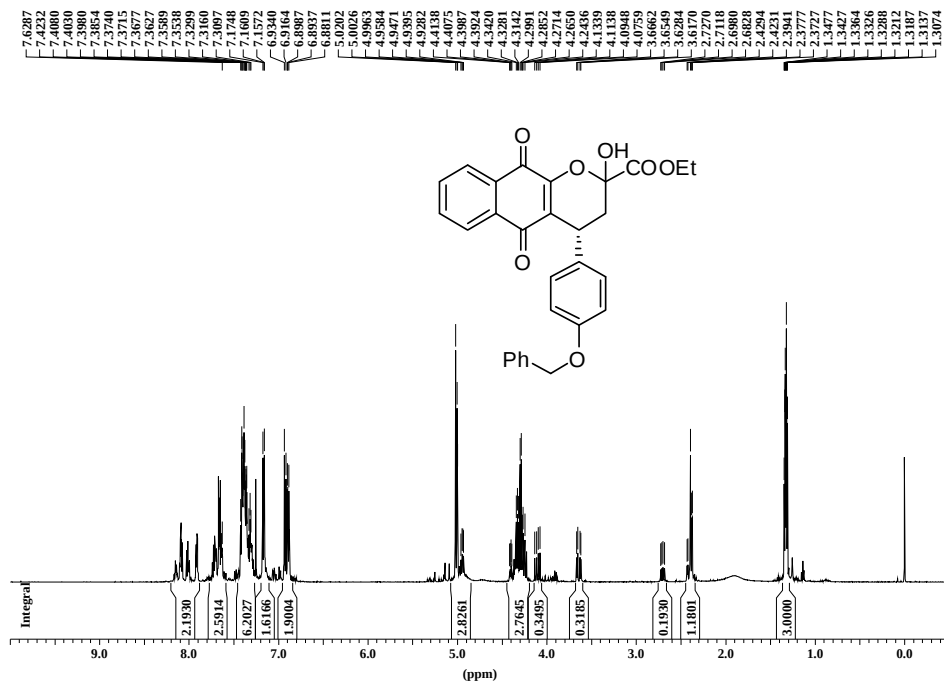
Compound **7h**



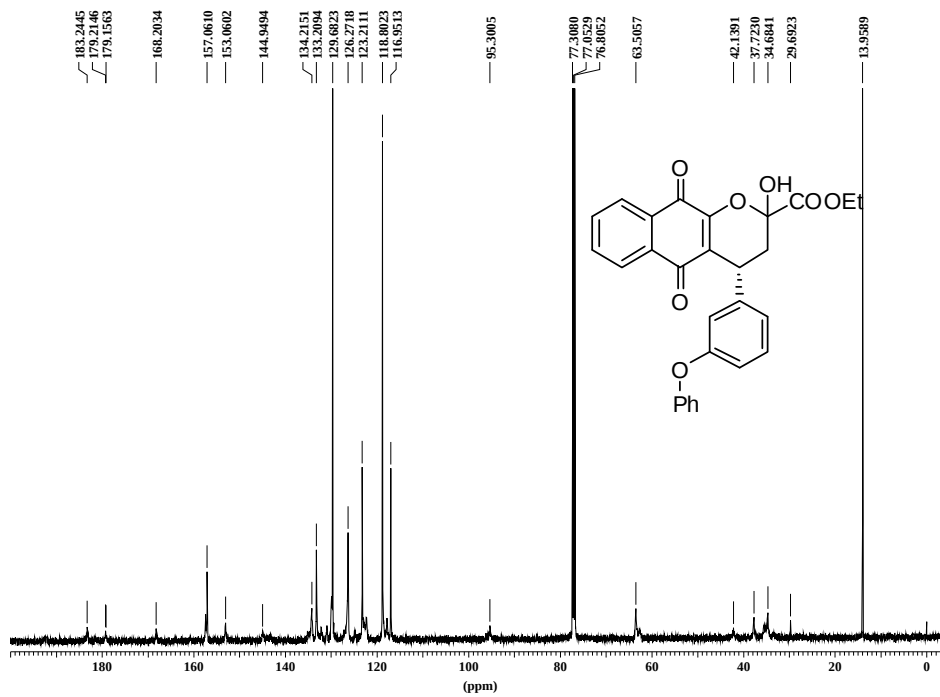
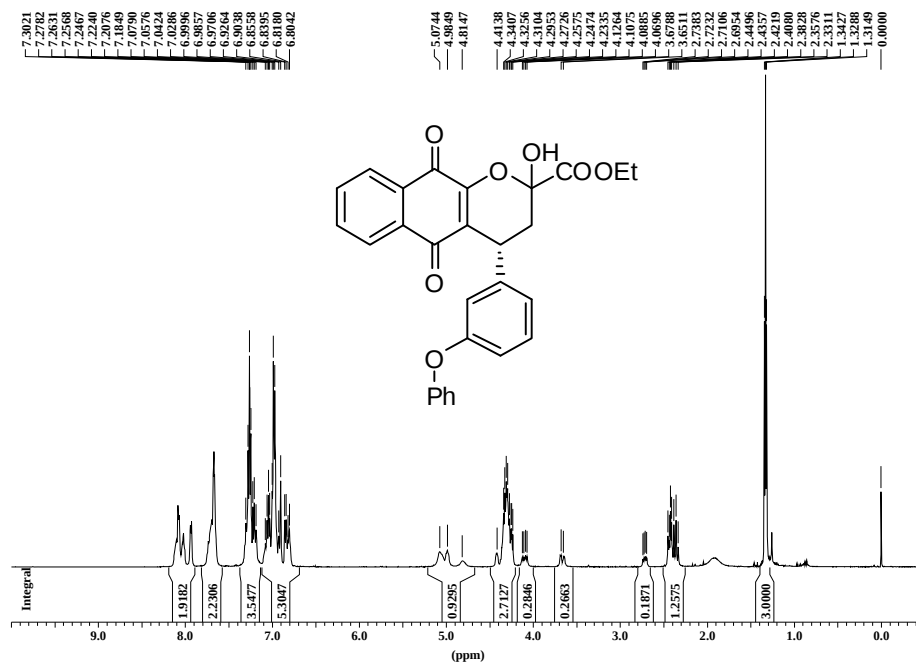
Compound **7i**



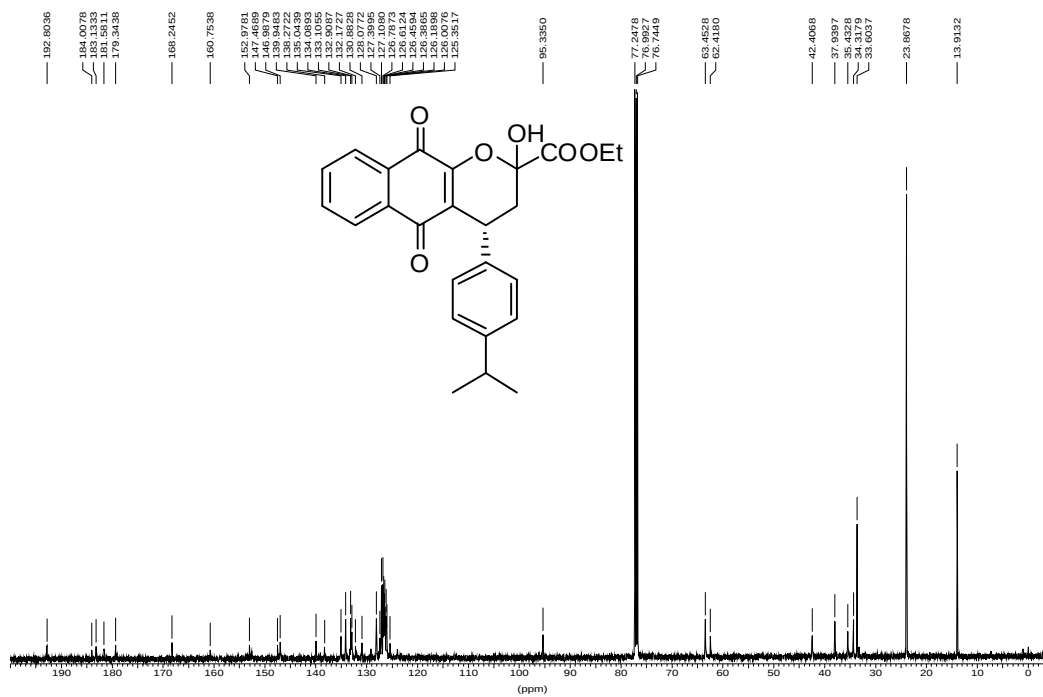
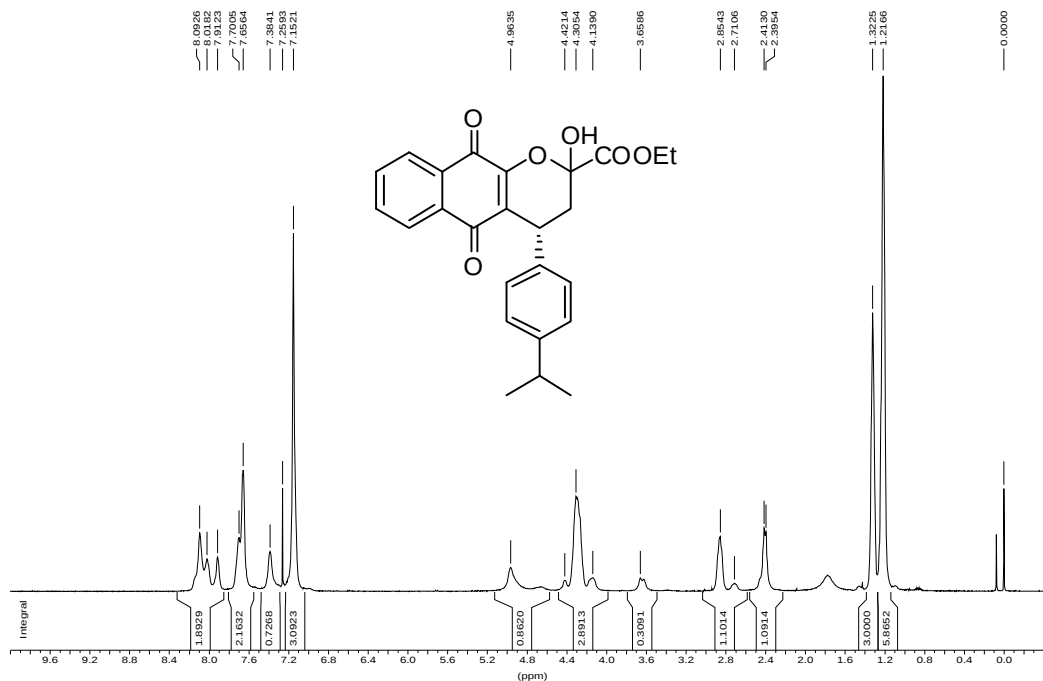
Compound 7j



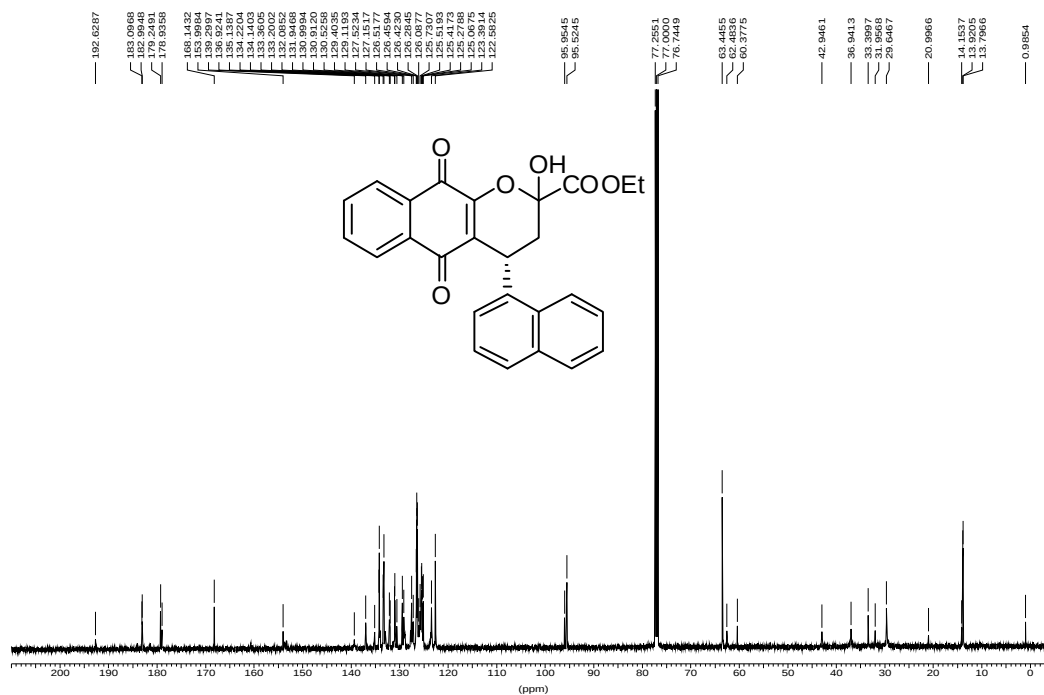
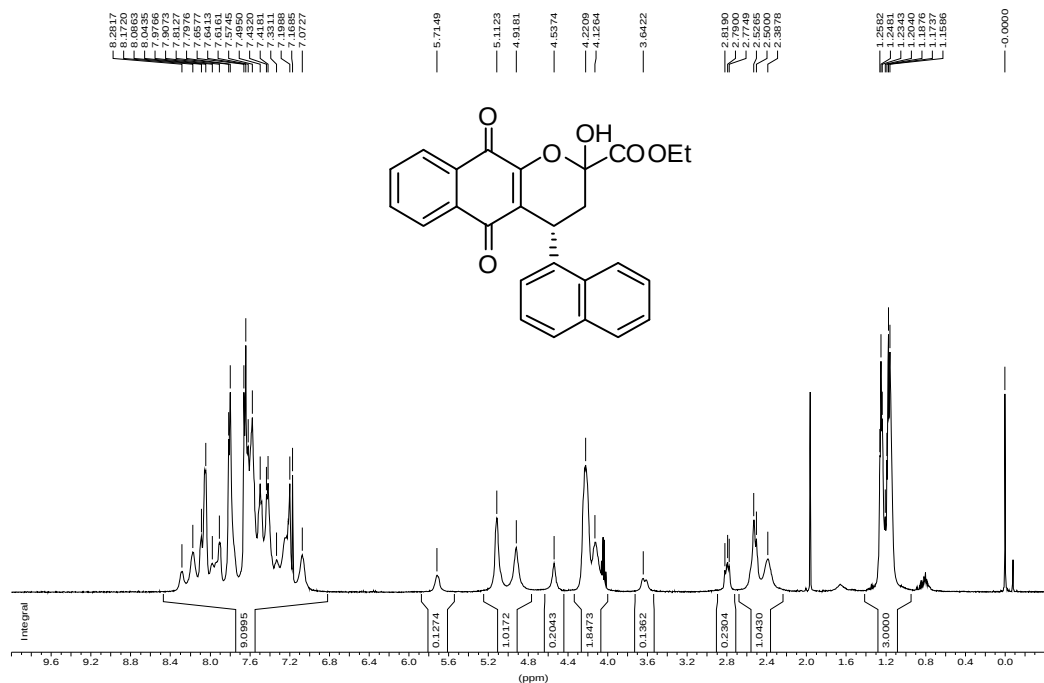
Compound **7k**



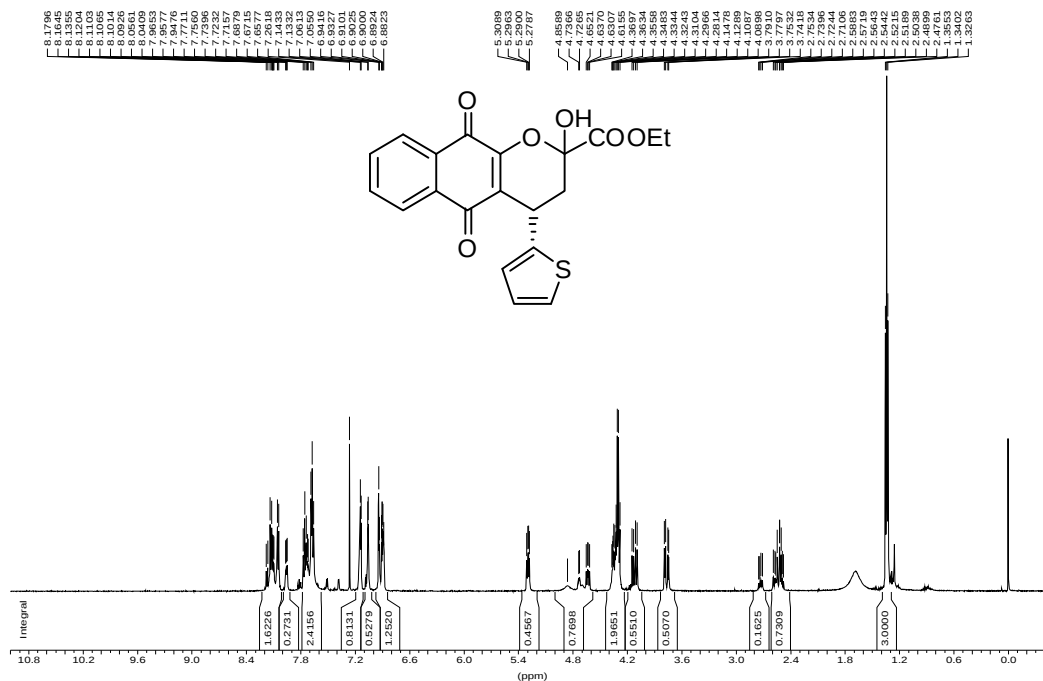
Compound **7l**



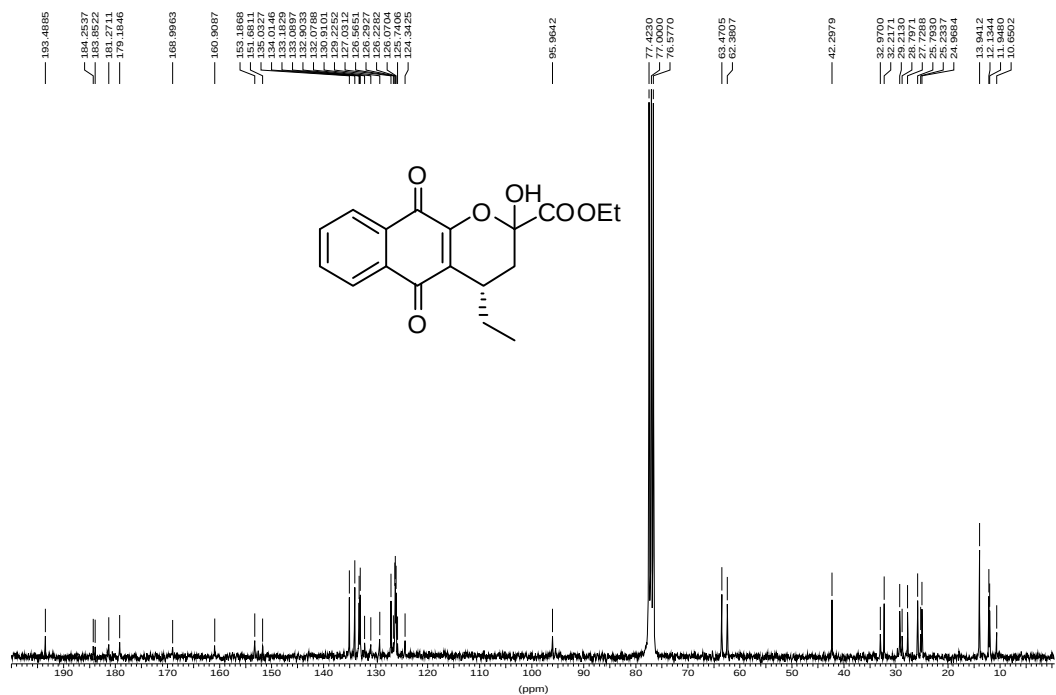
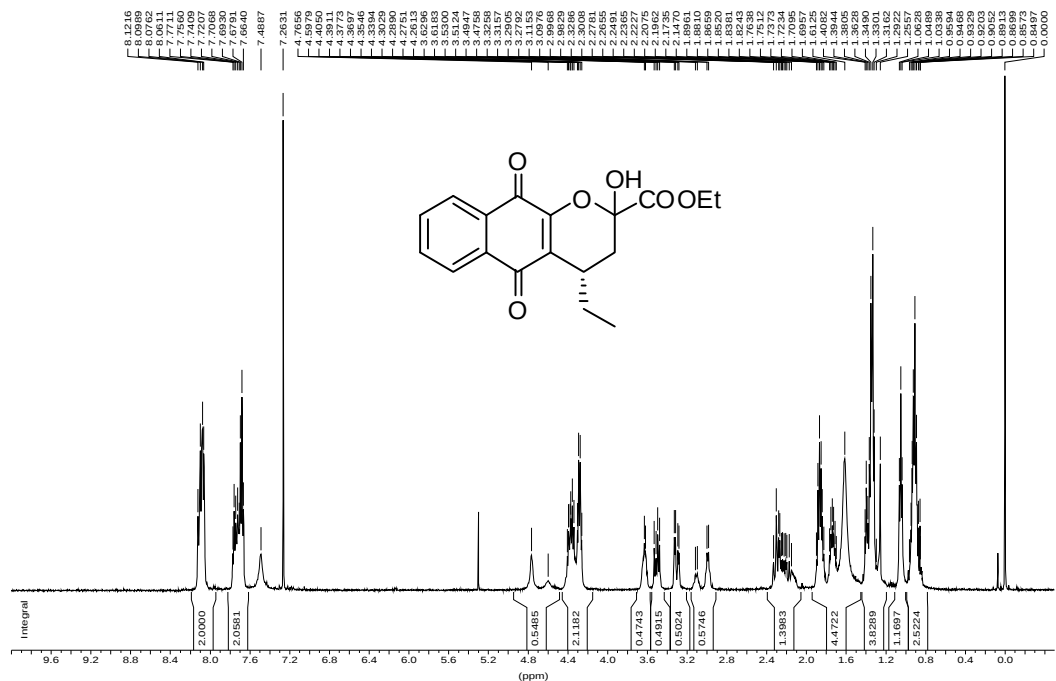
Compound 7m



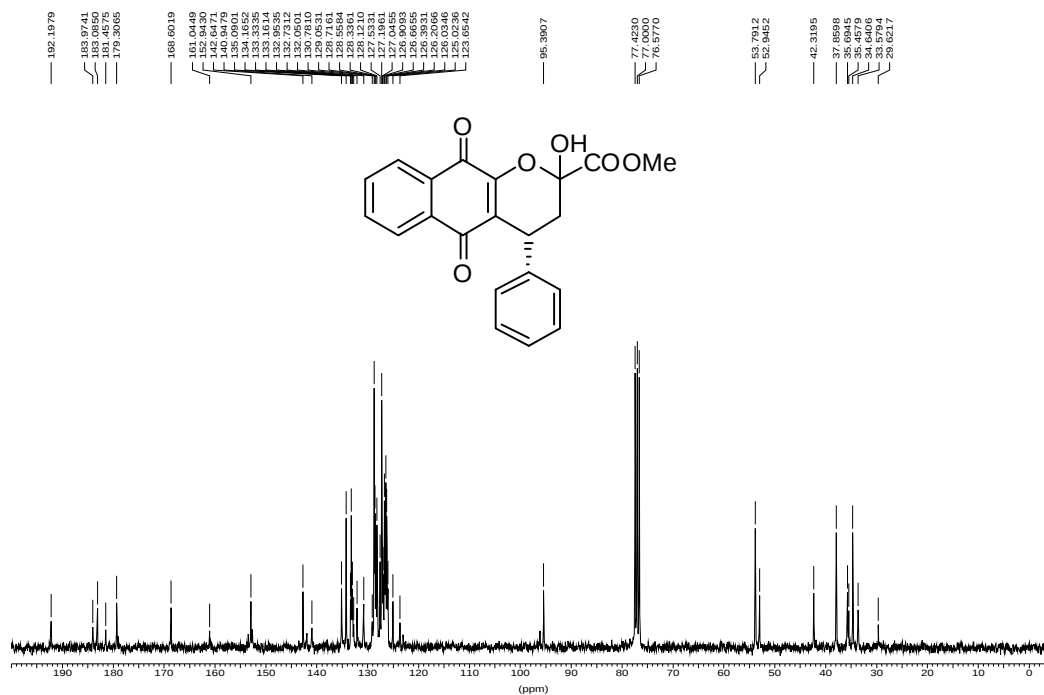
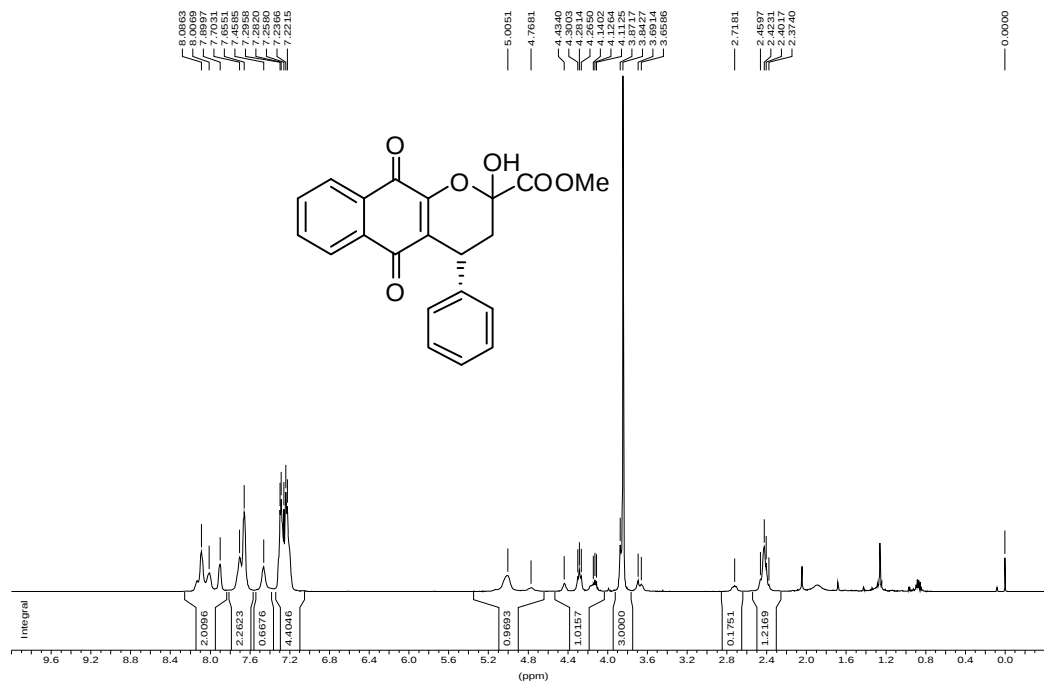
Compound **7n**



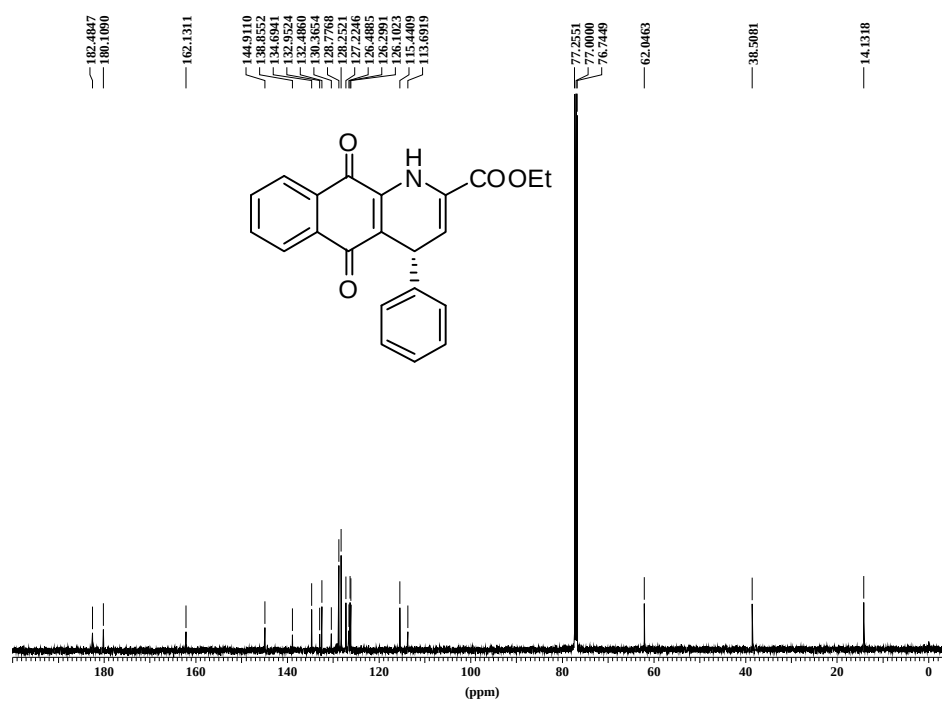
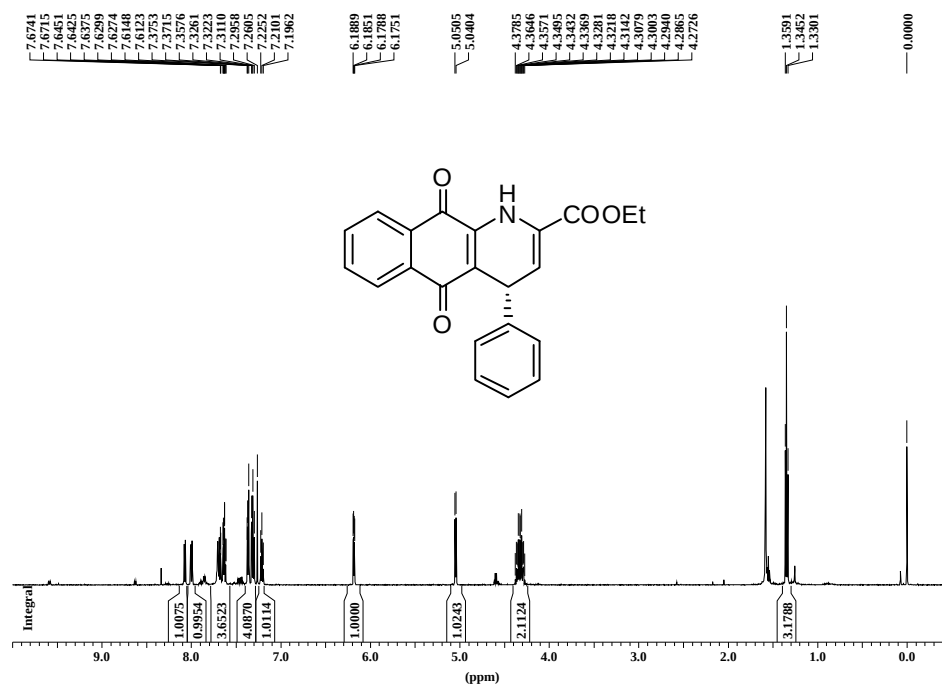
Compound **7o**



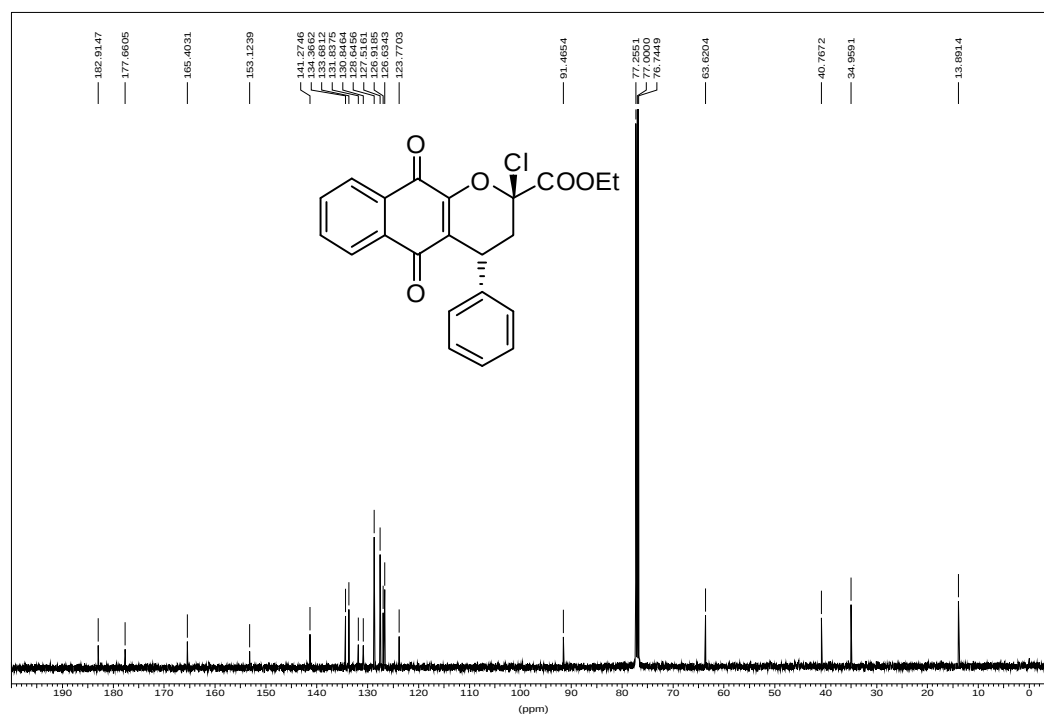
Compound **7p**



Compound 9



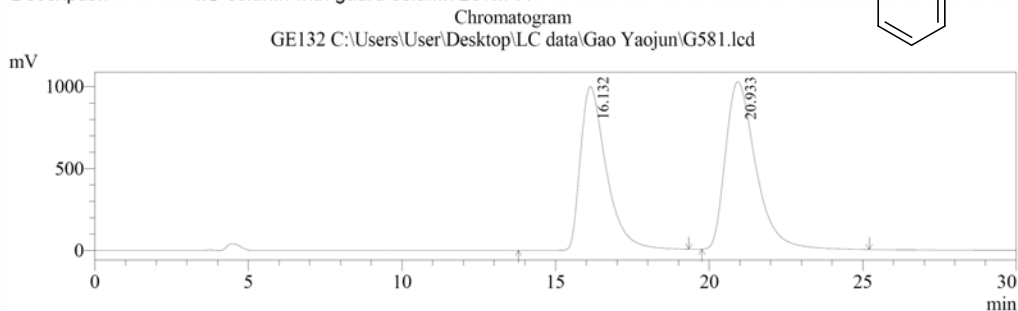
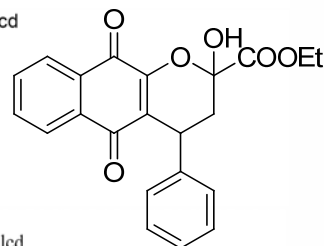
Compound 10



Racemic 7a

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : GE132
 Sample ID : GYJ
 Data File Name : G581.lcd
 Method File Name : 20%IPA, 1ml-min, 40min.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Description : IC column with guard column 20%IPA



PeakTable

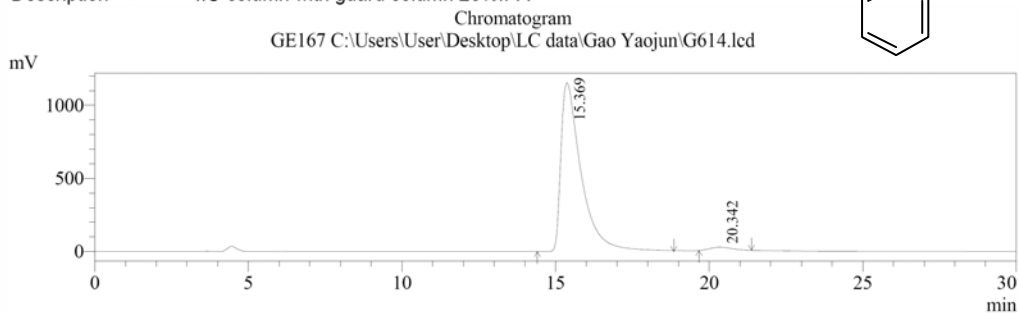
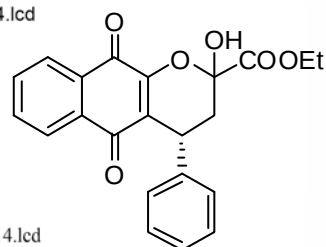
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	16.132	58027528	996826	45.200	49.370
2	20.933	70353214	1022275	54.800	50.630
Total		128380742	2019101	100.000	100.000

Enantiomeric enriched 7a

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : GE167
 Sample ID : GYJ
 Data File Name : G614.lcd
 Method File Name : 20%IPA, 1ml-min, 40min.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Description : IC column with guard column 20%IPA



PeakTable

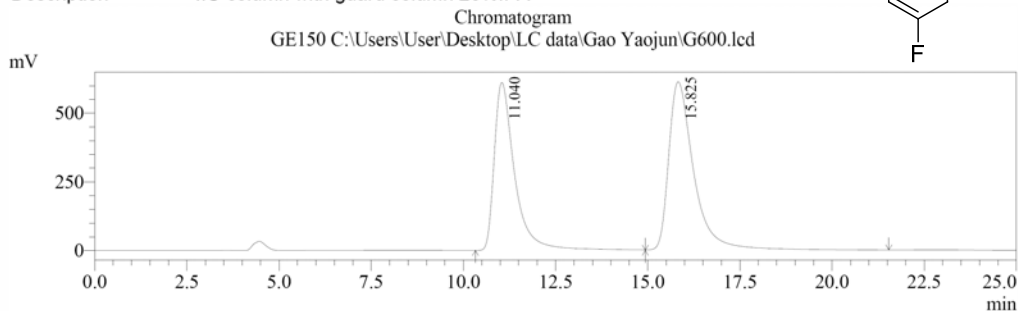
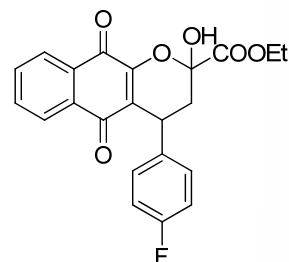
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	15.369	53352132	1154616	98.013	98.153
2	20.342	1081728	21728	1.987	1.847
Total		54433860	1176344	100.000	100.000

Racemic **7b**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE150
Sample ID : GYJ
Data File Name : G600.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column 20%IPA



PeakTable

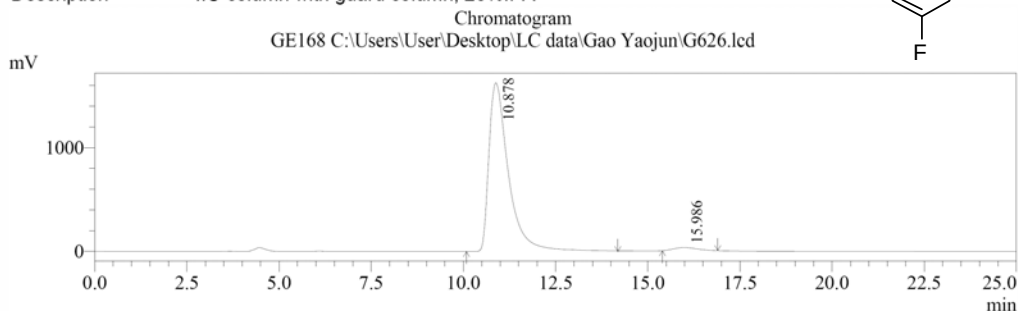
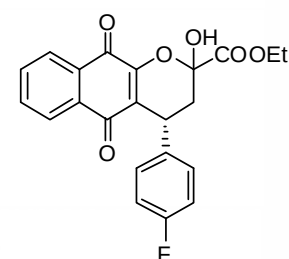
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	11.040	23605344	611346	44.770	49.921
2	15.825	29120359	613280	55.230	50.079
Total		52725703	1224626	100.000	100.000

Enantiomeric enriched **7b**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE168
Sample ID : GYJ
Data File Name : G626.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column, 20%IPA



PeakTable

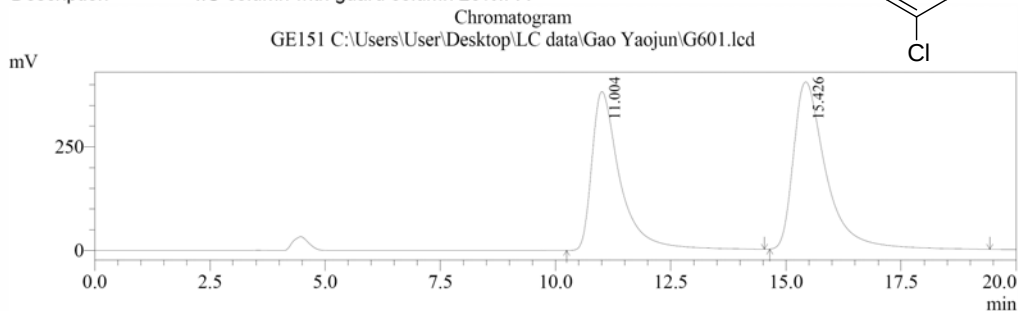
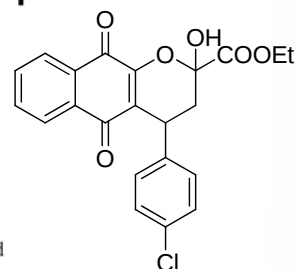
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	10.878	60476187	1627729	97.860	98.167
2	15.986	1322655	30392	2.140	1.833
Total		61798842	1658121	100.000	100.000

Racemic **7c**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : GE151
 Sample ID : GYJ
 Data File Name : G601.lcd
 Method File Name : 20%IPA, 1ml-min, 40min.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Description : IC column with guard column 20%IPA



PeakTable

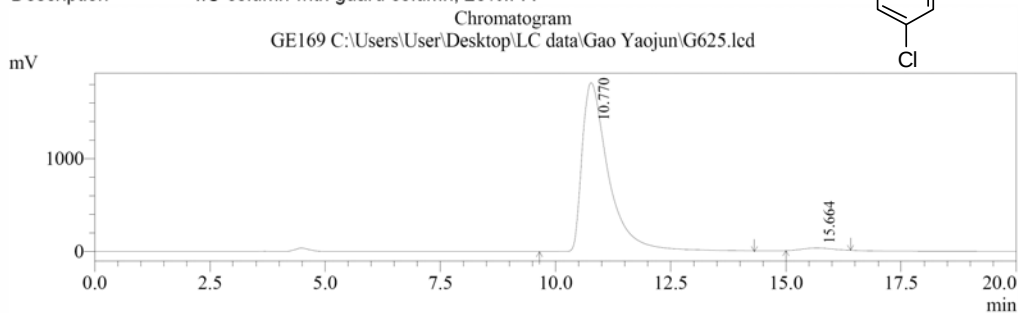
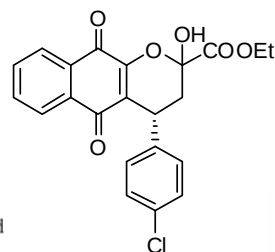
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	11.004	16305463	383048	45.113	48.678
2	15.426	19837917	403852	54.887	51.322
Total		36143380	786900	100.000	100.000

Enantiomeric enriched **7c**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : GE169
 Sample ID : GYJ
 Data File Name : G625.lcd
 Method File Name : 20%IPA, 1ml-min, 40min.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Description : IC column with guard column, 20%IPA



PeakTable

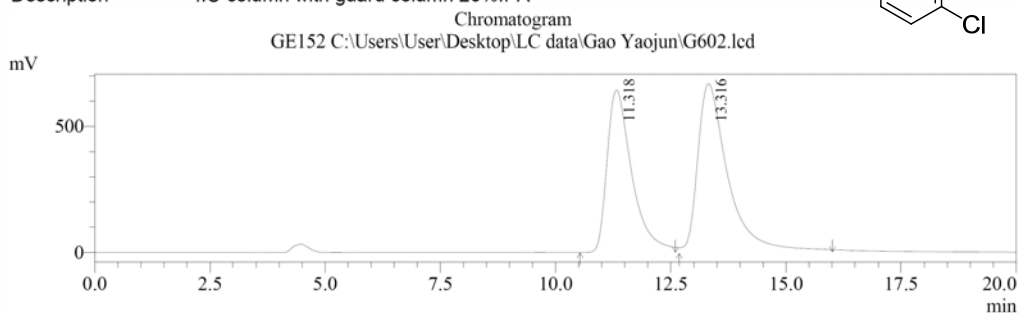
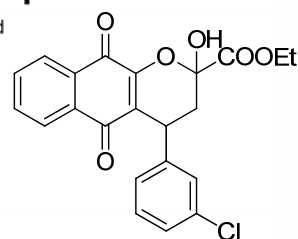
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	10.770	73726804	1818805	98.433	98.536
2	15.664	1173414	27019	1.567	1.464
Total		74900218	1845824	100.000	100.000

Racemic 7d

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : GE152
 Sample ID : GYJ
 Data File Name : G602.lcd
 Method File Name : 20%IPA, 1ml-min, 40min.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Description : IC column with guard column 20%IPA



PeakTable

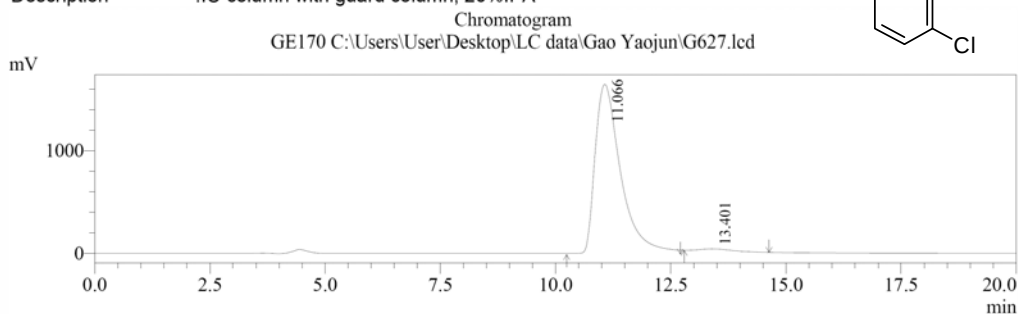
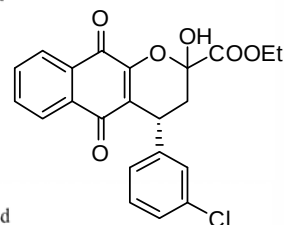
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	11.318	24195943	644441	43.754	48.990
2	13.316	31104255	671013	56.246	51.010
Total		55300198	1315455	100.000	100.000

Enantiomeric enriched 7d

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : GE170
 Sample ID : GYJ
 Data File Name : G627.lcd
 Method File Name : 20%IPA, 1ml-min, 40min.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Description : IC column with guard column, 20%IPA



PeakTable

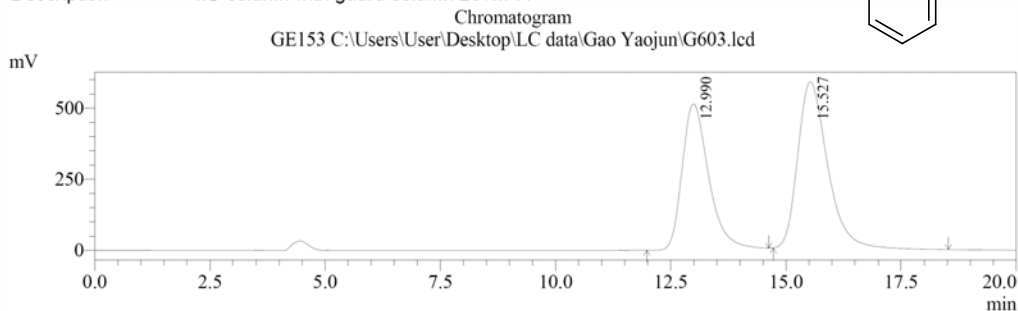
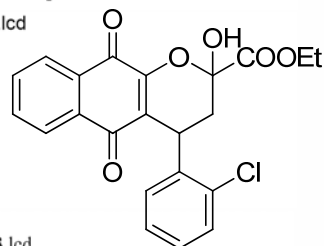
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	11.066	65115945	1655410	98.631	98.753
2	13.401	903685	20911	1.369	1.247
Total		66019629	1676321	100.000	100.000

Racemic **7e**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE153
Sample ID : GYJ
Data File Name : G603.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column 20%IPA



PeakTable

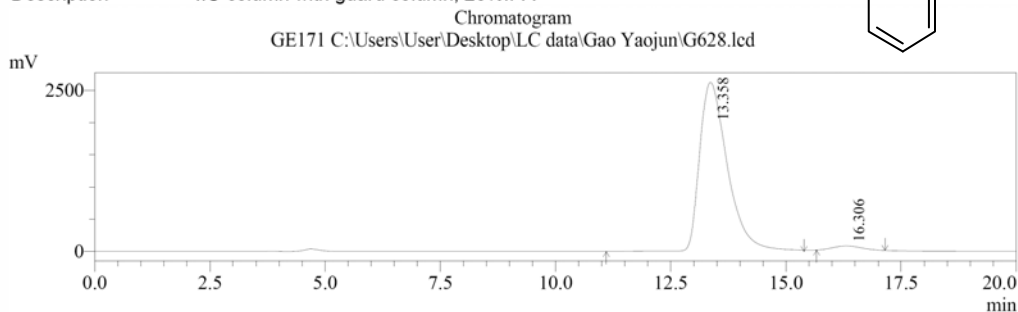
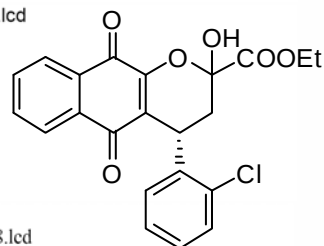
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	12.990	20325869	512309	43.568	46.651
2	15.527	26326955	585859	56.432	53.349
Total		46652824	1098168	100.000	100.000

Enantiomeric enriched **7e**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE171
Sample ID : GYJ
Data File Name : G628.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column, 20%IPA



PeakTable

SPD-20A Ch1 254nm

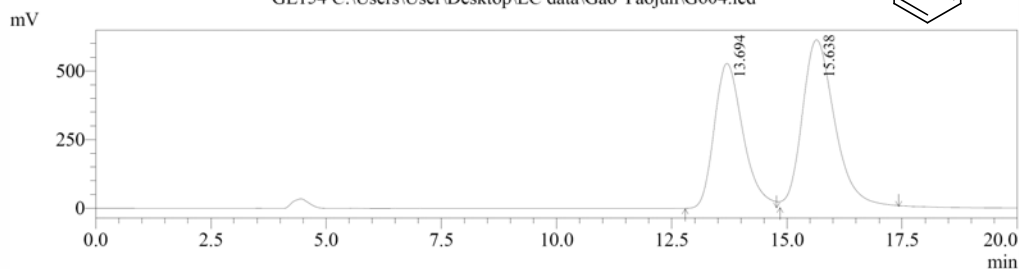
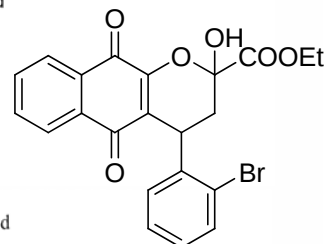
Peak#	Ret. Time	Area	Height	Area %	Height %
1	13.358	111654365	2628505	97.498	97.511
2	16.306	2865352	67094	2.502	2.489
Total		114519717	2695599	100.000	100.000

Racemic **7f**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE154
Sample ID : GYJ
Data File Name : G604.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column 20%IPA

Chromatogram
GE154 C:\Users\User\Desktop\LC data\Gao Yaojun\G604.lcd



PeakTable

SPD-20A Ch1 254nm

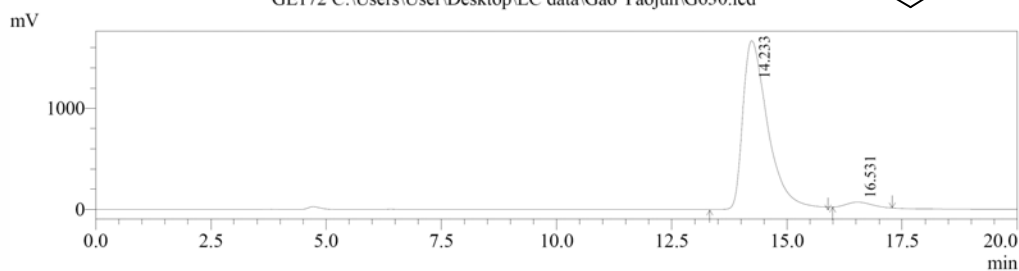
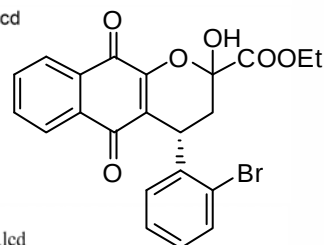
Peak#	Ret. Time	Area	Height	Area %	Height %
1	13.694	23045871	528132	43.485	46.416
2	15.638	29951760	609698	56.515	53.584
Total		52997632	1137830	100.000	100.000

Enantiomeric enriched **7f**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE172
Sample ID : GYJ
Data File Name : G630.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column, 20%IPA

Chromatogram
GE172 C:\Users\User\Desktop\LC data\Gao Yaojun\G630.lcd



PeakTable

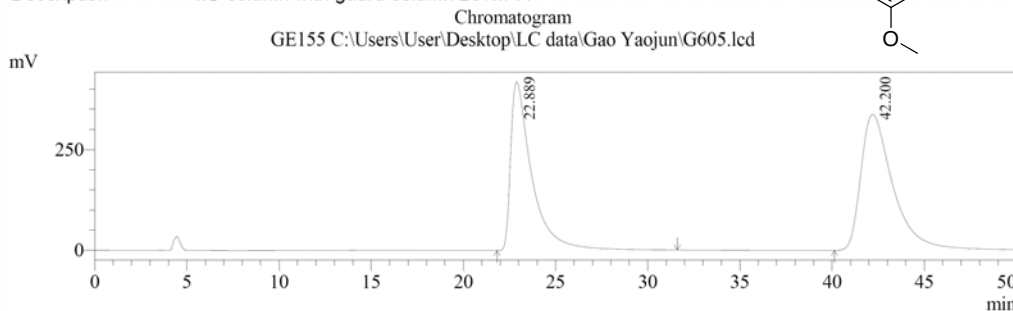
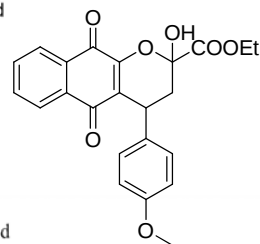
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	14.233	64073132	1674220	97.002	96.918
2	16.531	1980596	53243	2.998	3.082
Total		66053728	1727463	100.000	100.000

Racemic **7g**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : GE155
 Sample ID : GYJ
 Data File Name : G605.lcd
 Method File Name : 20%IPA, 1ml-min, 40min.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Description : IC column with guard column 20%IPA



PeakTable

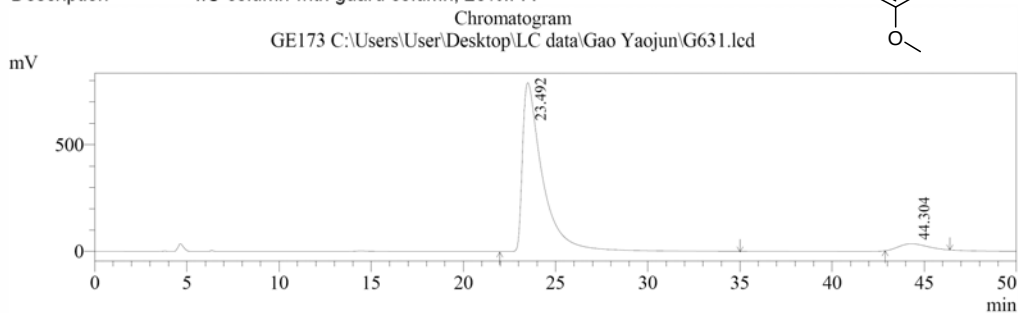
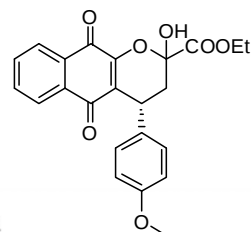
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	22.889	33491839	419009	44.797	55.345
2	42.200	41271794	338080	55.203	44.655
Total		74763632	757090	100.000	100.000

Enantiomeric enriched **7g**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : GE173
 Sample ID : GYJ
 Data File Name : G631.lcd
 Method File Name : 20%IPA, 1ml-min, 40min.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Description : IC column with guard column, 20%IPA



PeakTable

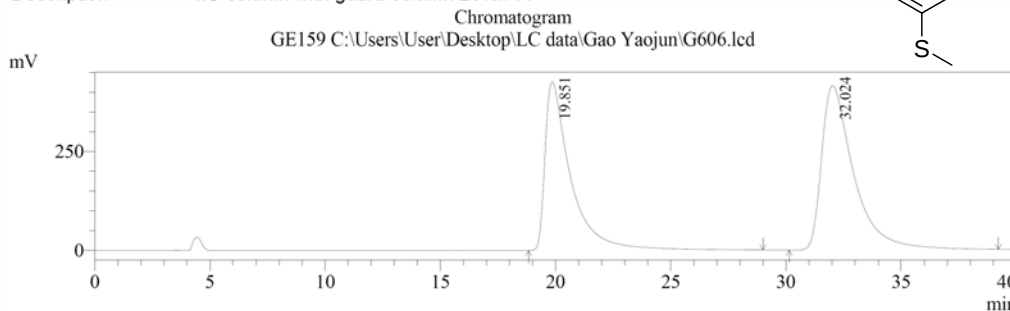
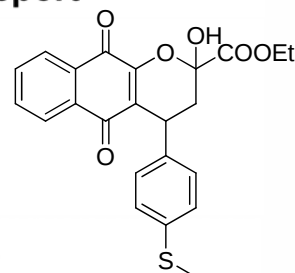
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	23.492	62371081	793640	94.973	96.205
2	44.304	3301384	31308	5.027	3.795
Total		65672465	824948	100.000	100.000

Racemic **7h**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE159
Sample ID : GYJ
Data File Name : G606.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column 20%IPA



PeakTable

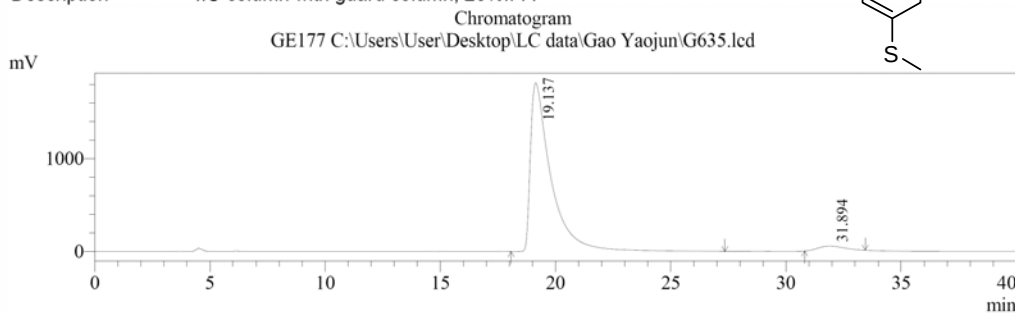
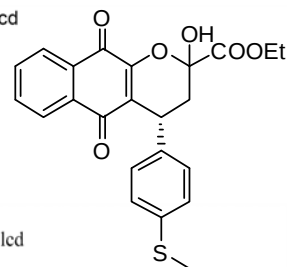
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	19.851	33128744	426704	45.096	50.659
2	32.024	40333228	415604	54.904	49.341
Total		73461972	842308	100.000	100.000

Enantiomeric enriched **7h**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE177
Sample ID : GYJ
Data File Name : G635.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column, 20%IPA



PeakTable

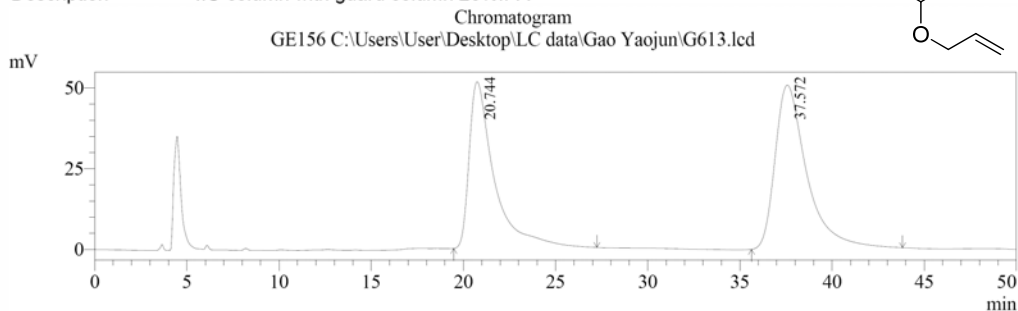
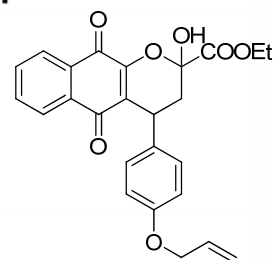
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	19.137	113901747	1819722	96.673	97.316
2	31.894	3920122	50190	3.327	2.684
Total		117821869	1869912	100.000	100.000

Racemic **7i**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE156
Sample ID : GYJ
Data File Name : G613.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column 20%IPA



PeakTable

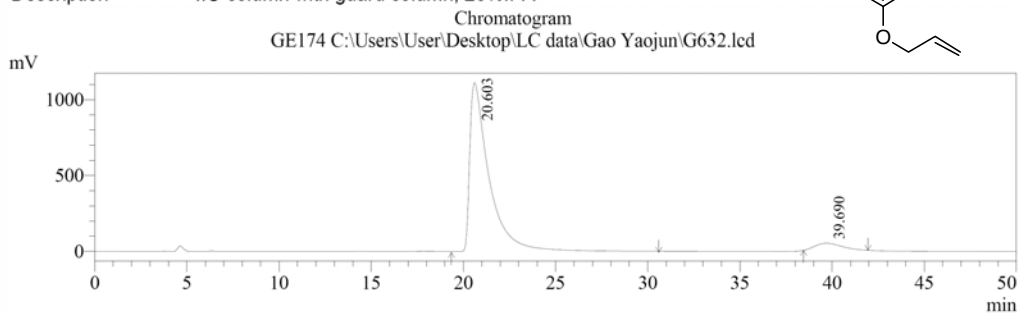
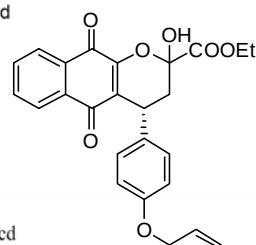
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	20.744	4873659	51459	44.616	50.392
2	37.572	6049880	50658	55.384	49.608
Total		10923540	102117	100.000	100.000

Enantiomeric enriched **7i**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE174
Sample ID : GYJ
Data File Name : G632.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column, 20%IPA



PeakTable

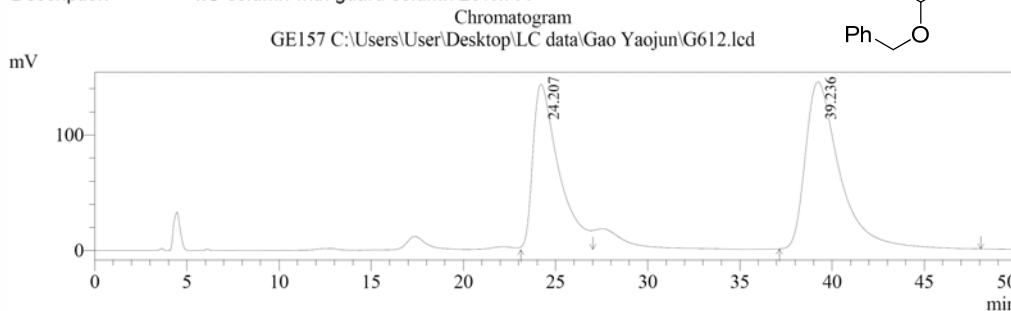
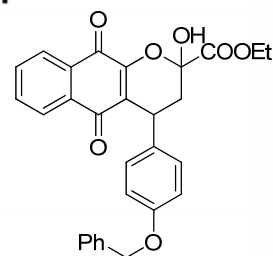
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	20.603	85444348	1117877	94.801	96.011
2	39.690	4686167	46439	5.199	3.989
Total		90130515	1164316	100.000	100.000

Racemic **7j**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : GE157
 Sample ID : GYJ
 Data File Name : G612.lcd
 Method File Name : 20%IPA, 1ml-min, 40min.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Description : IC column with guard column 20%IPA



PeakTable

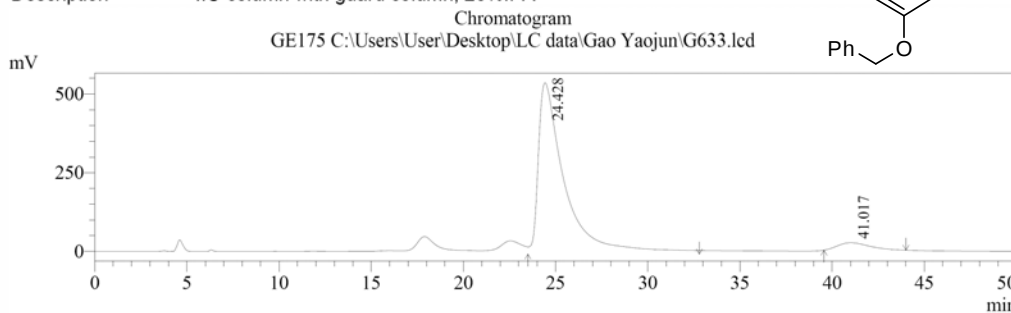
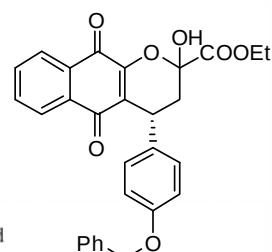
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	24.207	14166045	143069	43.031	49.707
2	39.236	18754644	144758	56.969	50.293
Total		32920689	287827	100.000	100.000

Enantiomeric enriched **7j**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : GE175
 Sample ID : GYJ
 Data File Name : G633.lcd
 Method File Name : 20%IPA, 1ml-min, 40min.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Description : IC column with guard column, 20%IPA



PeakTable

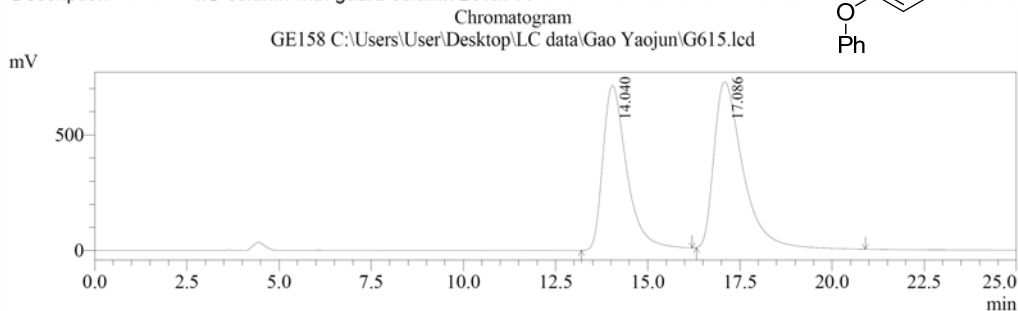
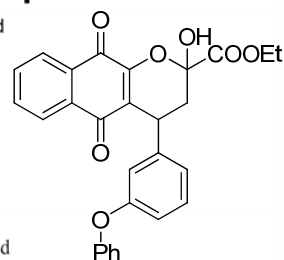
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	24.428	55797401	544116	95.147	95.833
2	41.017	2845886	23661	4.853	4.167
Total		58643287	567777	100.000	100.000

Racemic **7k**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE158
Sample ID : GYJ
Data File Name : G615.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column 20%IPA



PeakTable

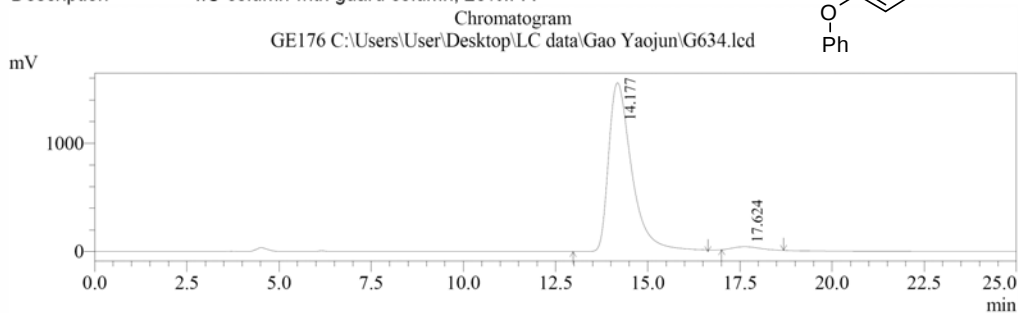
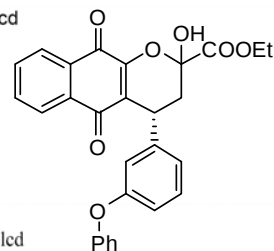
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	14.040	31841727	710657	45.040	49.720
2	17.086	38854642	718654	54.960	50.280
Total		70696369	1429311	100.000	100.000

Enantiomeric enriched **7k**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE176
Sample ID : GYJ
Data File Name : G634.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column, 20%IPA



PeakTable

SPD-20A Ch1 254nm

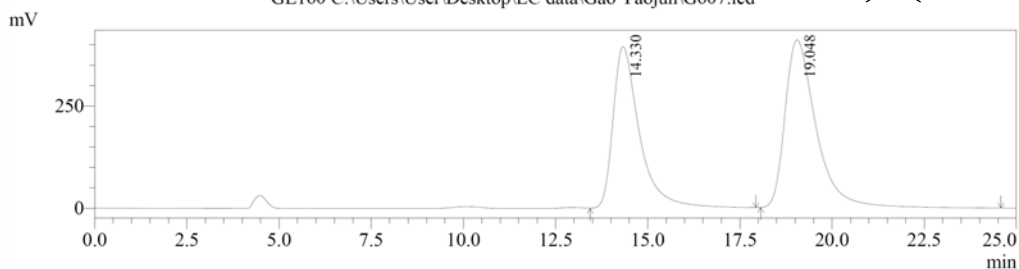
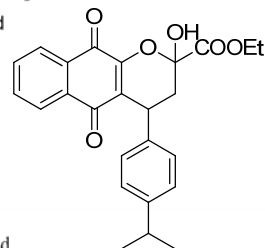
Peak#	Ret. Time	Area	Height	Area %	Height %
1	14.177	68977621	1560348	97.880	98.075
2	17.624	1493834	30631	2.120	1.925
Total		70471455	1590979	100.000	100.000

Racemic **7l**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE160
Sample ID : GYJ
Data File Name : G607.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column 20%IPA

Chromatogram
GE160 C:\Users\User\Desktop\LC data\Gao Yaojun\G607.lcd



PeakTable

SPD-20A Ch1 254nm

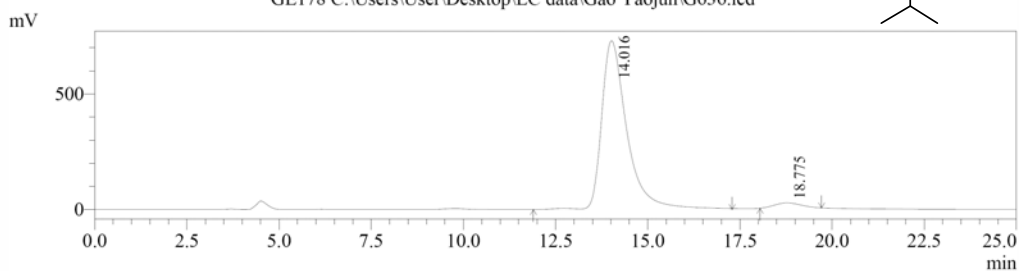
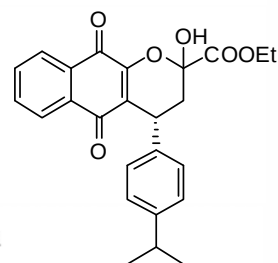
Peak#	Ret. Time	Area	Height	Area %	Height %
1	14.330	19742861	395480	45.017	49.057
2	19.048	24113366	410685	54.983	50.943
Total		43856226	806164	100.000	100.000

Enantiomeric enriched **7l**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE178
Sample ID : GYJ
Data File Name : G636.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column, 20%IPA

Chromatogram
GE178 C:\Users\User\Desktop\LC data\Gao Yaojun\G636.lcd



PeakTable

SPD-20A Ch1 254nm

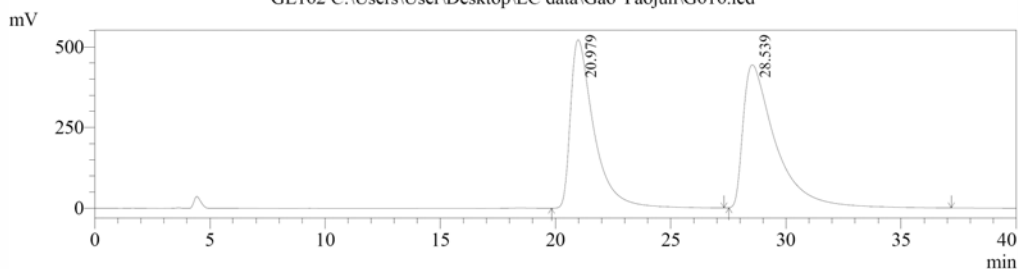
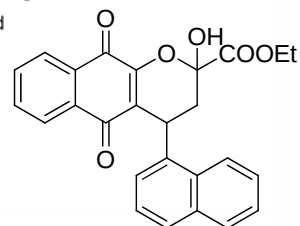
Peak#	Ret. Time	Area	Height	Area %	Height %
1	14.016	35091230	730620	96.937	97.039
2	18.775	1108732	22292	3.063	2.961
Total		36199962	752912	100.000	100.000

Racemic **7m**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE162
Sample ID : GYJ
Data File Name : G616.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column 20%IPA

Chromatogram
GE162 C:\Users\User\Desktop\LC data\Gao Yaojun\G616.lcd



PeakTable

SPD-20A Ch1 254nm

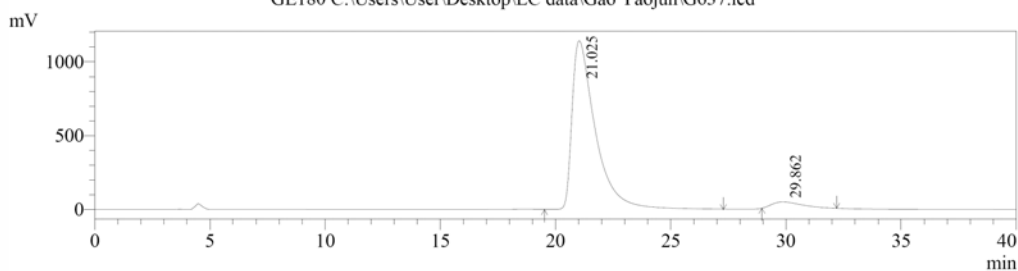
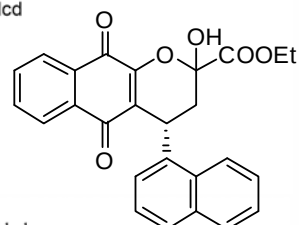
Peak#	Ret. Time	Area	Height	Area %	Height %
1	20.979	37047214	521720	46.046	54.095
2	28.539	43410397	442737	53.954	45.905
Total		80457611	964457	100.000	100.000

Enantiomeric enriched **7m**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE180
Sample ID : GYJ
Data File Name : G637.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column, 20%IPA

Chromatogram
GE180 C:\Users\User\Desktop\LC data\Gao Yaojun\G637.lcd



PeakTable

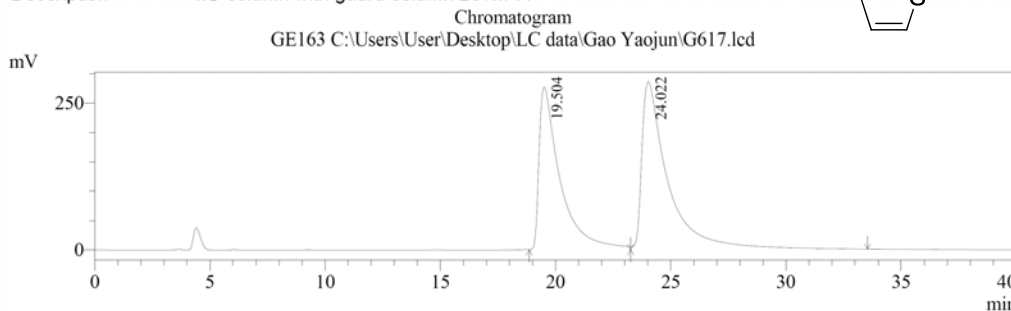
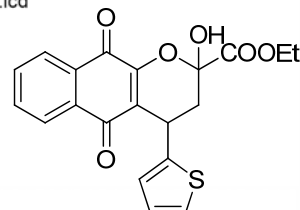
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	21.025	81375113	1143288	95.454	96.387
2	29.862	3875268	42850	4.546	3.613
Total		85250381	1186138	100.000	100.000

Racemic **7n**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE163
Sample ID : GYJ
Data File Name : G617.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column 20%IPA



PeakTable

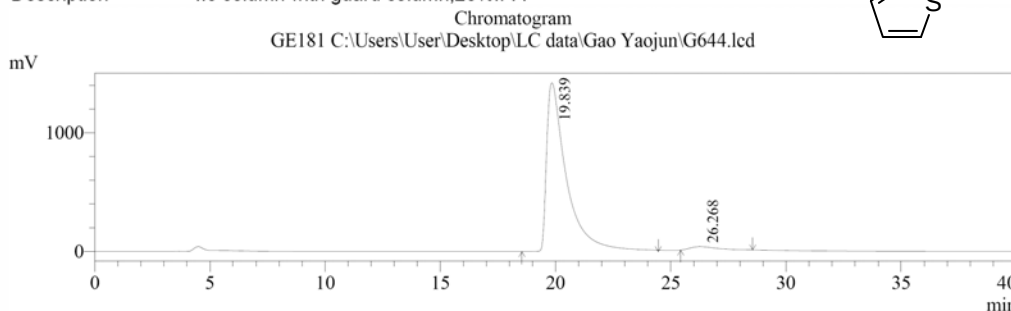
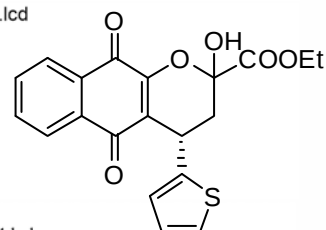
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	19.504	17648165	277097	43.803	49.259
2	24.022	22641644	285438	56.197	50.741
Total		40289810	562534	100.000	100.000

Enantiomeric enriched **7n**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE181
Sample ID : GYJ
Data File Name : G644.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column,20%IPA



PeakTable

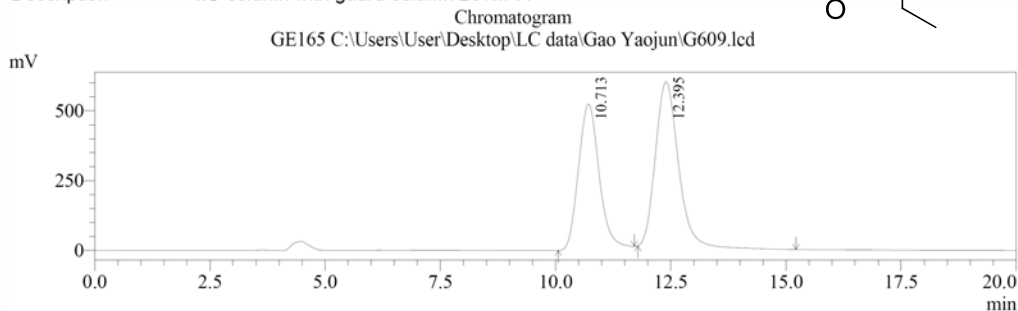
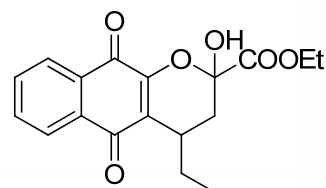
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	19.839	86108325	1423460	97.458	98.048
2	26.268	2246240	28341	2.542	1.952
Total		88354565	1451801	100.000	100.000

Racemic **7o**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE165
Sample ID : GYJ
Data File Name : G609.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column 20%IPA



PeakTable

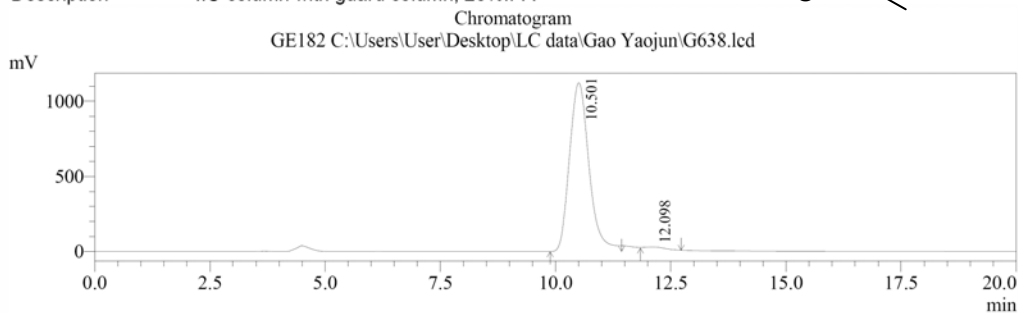
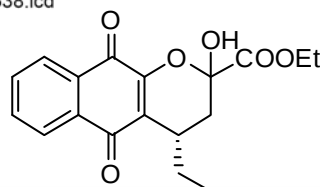
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	10.713	15929469	518919	44.102	46.800
2	12.395	20190049	589875	55.898	53.200
Total		36119518	1108795	100.000	100.000

Enantiomeric enriched **7o**

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE182
Sample ID : GYJ
Data File Name : G638.lcd
Method File Name : 20%IPA, 1ml-min, 40min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column, 20%IPA



PeakTable

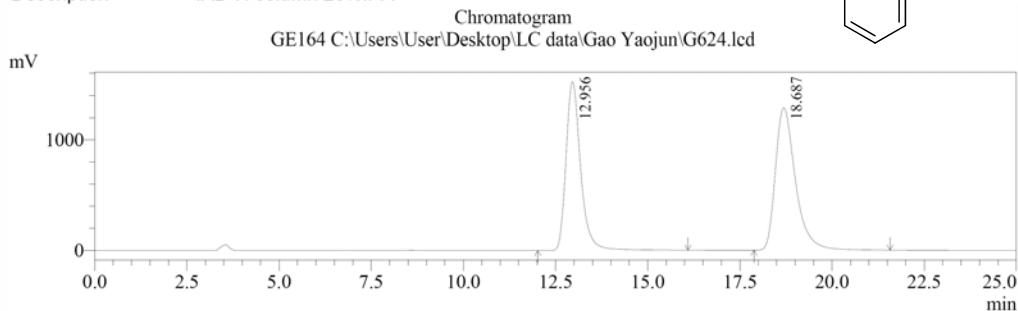
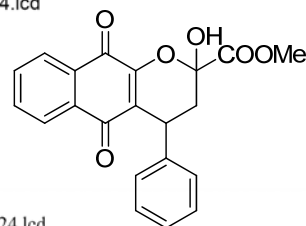
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	10.501	33293008	1124834	99.041	98.955
2	12.098	322322	11879	0.959	1.045
Total		33615330	1136713	100.000	100.000

Racemic 7p

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : GE164
 Sample ID : GYJ
 Data File Name : G624.lcd
 Method File Name : 20%IPA, 1ml-min, 40min.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Description : AD-H column 20%IPA



PeakTable

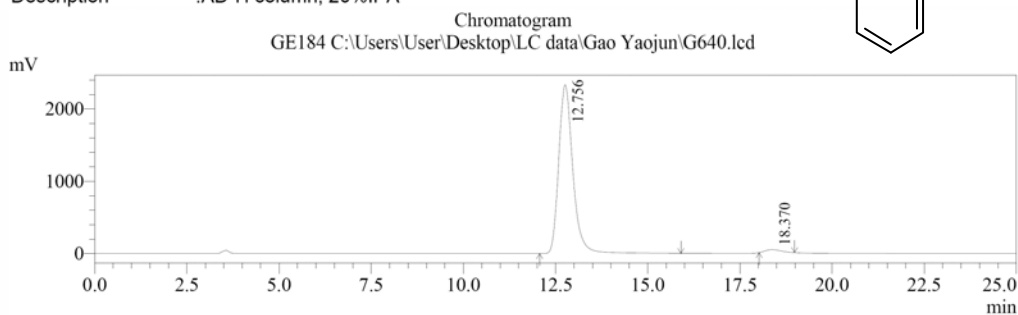
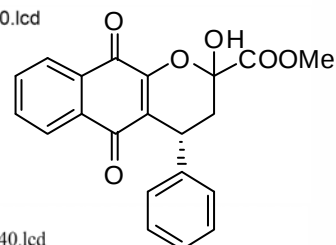
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	12.956	42040733	1524027	46.324	54.205
2	18.687	48713087	1287564	53.676	45.795
Total		90753820	2811591	100.000	100.000

Enantiomeric enriched 7p

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
 Sample Name : GE184
 Sample ID : GYJ
 Data File Name : G640.lcd
 Method File Name : 20%IPA, 1ml-min, 40min.lcm
 Batch File Name :
 Report File Name : Default.lcr
 Description : AD-H column, 20%IPA



PeakTable

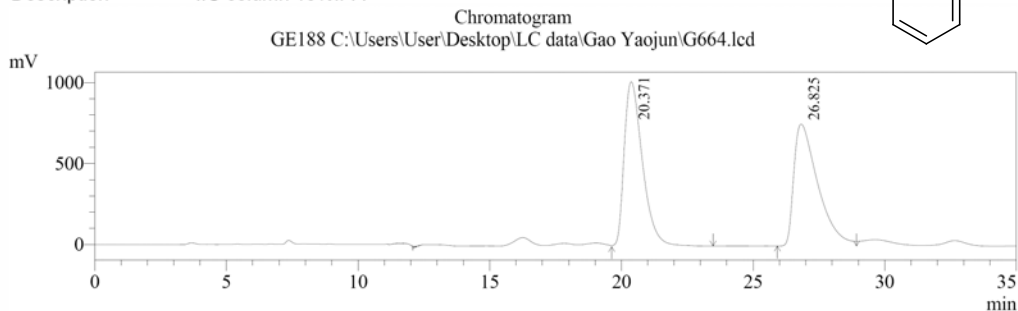
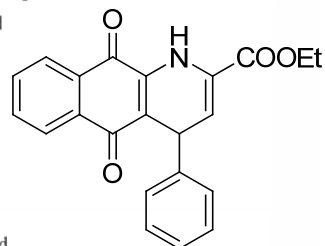
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	12.756	62632992	2334627	98.091	98.251
2	18.370	1218921	41571	1.909	1.749
Total		63851912	2376199	100.000	100.000

Racemic 9

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE188
Sample ID : GYJ
Data File Name : G664.lcd
Method File Name : 10%IPA, 1ml-min, 60min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column 10%IPA



PeakTable

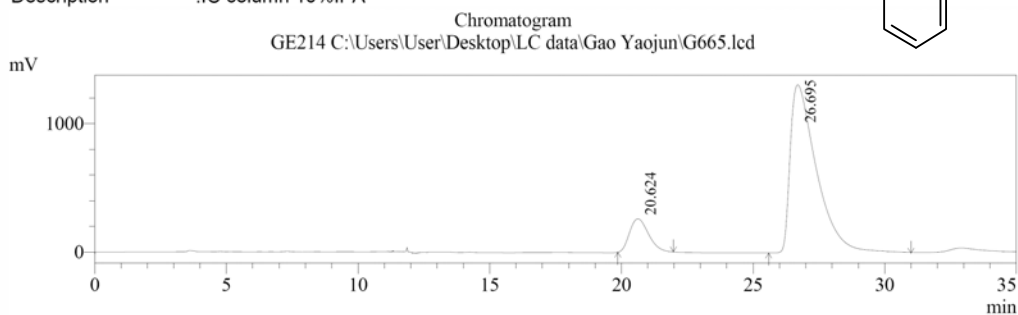
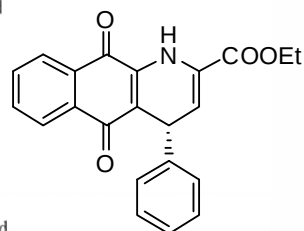
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	20.371	49153785	1016344	50.575	57.453
2	26.825	48035545	752669	49.425	42.547
Total		97189331	1769013	100.000	100.000

Enantiomeric enriched 9

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE214
Sample ID : GYJ
Data File Name : G665.lcd
Method File Name : 10%IPA, 1ml-min, 60min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column 10%IPA



PeakTable

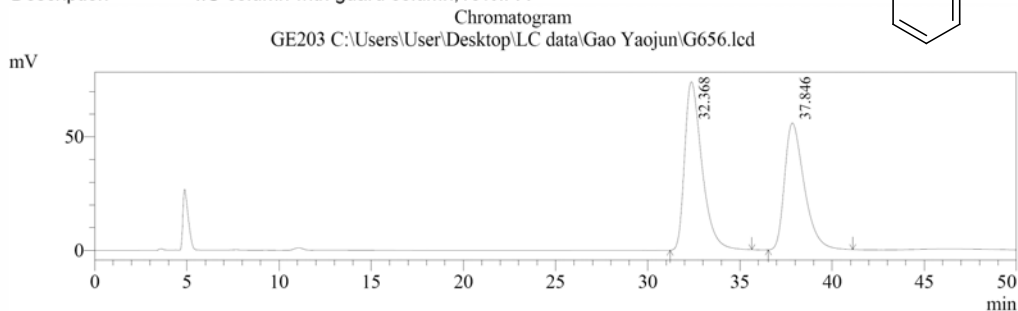
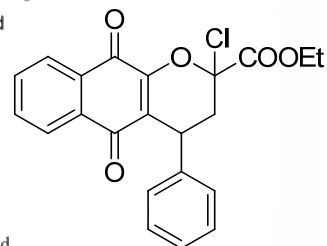
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	20.624	13001897	259790	12.404	16.539
2	26.695	91819719	1310939	87.596	83.461
Total		104821616	1570729	100.000	100.000

Racemic 10

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE203
Sample ID : GYJ
Data File Name : G656.lcd
Method File Name : 10%IPA, 1ml-min, 60min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column with guard column, 10%IPA



PeakTable

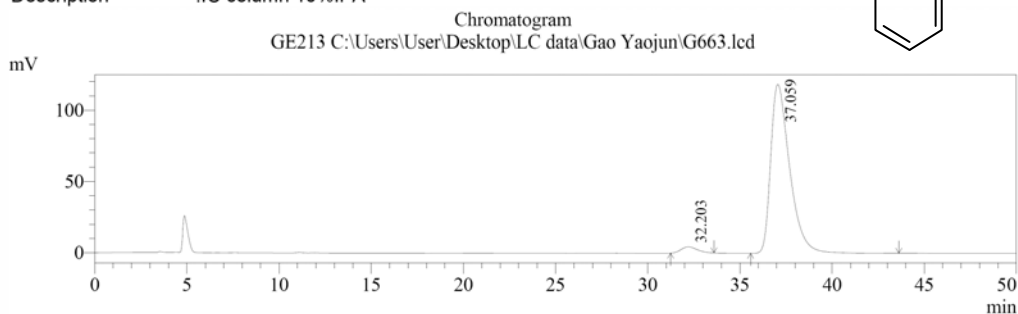
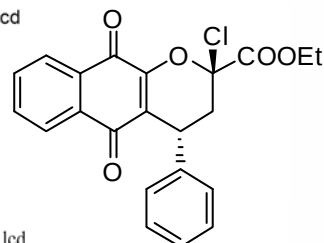
SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	32.368	4840218	74058	53.989	57.067
2	37.846	4124937	55716	46.011	42.933
Total		8965155	129774	100.000	100.000

Enantiomeric enriched 10

==== Shimadzu LCsolution Analysis Report ====

Acquired by : Admin
Sample Name : GE213
Sample ID : GYJ
Data File Name : G663.lcd
Method File Name : 10%IPA, 1ml-min, 60min.lcm
Batch File Name :
Report File Name : Default.lcr
Description : IC column 10%IPA



PeakTable

SPD-20A Ch1 254nm

Peak#	Ret. Time	Area	Height	Area %	Height %
1	32.203	265472	4359	3.025	3.544
2	37.059	8510954	118616	96.975	96.456
Total		8776427	122975	100.000	100.000