Peptide dendrimer enzyme models for ester hydrolysis and aldolization prepared by convergent thioether ligation

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Materials and Reagents

All reagents were purchased in the highest quality available either from Sigma Aldrich, Bachem, Acors Organics, Carl Roth GmbH or GE Healthcare. PyBOP, amino acids and their derivatives were purchased from Advanced ChemTech (Giessen, Germany) or Novabiochem. For SPPS amino acids were used as the following derivatives: Fmoc-Ala-OH, Fmoc-Amb-OH, Fmoc-Asp(OtBu)-OH, Fmoc-Cys(Trt)-OH, Fmoc-Dap(Fmoc)-OH, Fmoc-Glu(OtBu)-OH, Fmoc-DGlu(OtBu)-OH, Fmoc-Gly-OH, Fmoc-His(Boc)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Lys(Fmoc)-OH, Fmoc-Pro-OH, Fmoc-DPro-OH, Fmoc-Ser(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Val-OH. Tenta Gel S RAM[®] resin (loading: 0.24 or 0.26 mmol·g⁻¹) was purchased from Rapp Polymere (Tübingen, Germany). Peptide dendrimer syntheses were performed manually in polypropylene syringes fitted with a polyethylene frit, a Teflon stopcock and stopper. Ligation reactions were done in solution using standard glass ware. All solvents used in reactions on solid phase and in solution were bought in p.a. quality and distilled prior to use. Analytical RP-HPLC was performed with a Waters Chromatography System (996 Photo diode array detector) using an Atlantis[®] column (dC18, 5 µm, 4.6 × 100 mm, flow rate 1.4 mL·min⁻¹) or a chiral Daicel Chiralpak $AS^{\mathbb{R}}$ column (amylose tris[(S)- α methylbenzylcarbamate] coated on a silica support, $10 \,\mu\text{m}$, $4.6 \times 250 \,\text{mm}$, flow rate 2.0 mL·min⁻¹) for ee determination. Analytical RP-UPLC was performed with an Ultimate 3000 Rapid Separation LC System (DAD-3000RS diode array detector) using an Acclaim[®] RSLC 120 C18 column (2.2 μ m, 120 Å, 3.0 × 50 mm, flow 1.2 ml·min⁻¹) from Dionex. Data recording and processing was either done with Waters Empower2 software (analytical RP-HPLC) or with Dionex Chromeleon Management System Version 6.80 (analytical RP-HPLC). Preparative RP-HPLC was performed with a Waters Prep LC4000 Chromatography System using a Delta-Pak[®] column (C18, 15 µm, pore size 300 Å, flow rate 80 mL·min⁻¹) or an Atlantis OBD column (dC18, 5 μ m, 19 × 100 mm, flow rate 20 mL·min⁻¹). Compounds

were detected by UV absorption at 214 nm using a Waters 486 Tunable Absorbance Detector. The following elution solutions were used for all RP-HPLC: A mQ-deionized H₂O with 0.1% TFA; D mQ-deionized H₂O/HPLC-grade CH₃CN (40:60) with 0.1% TFA. MS spectra, recorded on either a Thermo Scientific LTQ OrbitrapXL, an AB Sciex QTrap or an AB Sciex API 2000 (LC-MS) and amino acid analysis were provided by the MS and protein analytical services of the Department of Chemistry and Biochemistry at the University of Berne. ¹H-NMR were recorded on a Bruker DRX500. Kinetic measurements were carried out using a CytoFluor Series 4000 multi-well plate reader from PerSeptive Biosystems. SDS-PAGE was done with equipment purchased from Bio-Rad.

Dendrimer Spectra Peptide Dendrimer G2C1

(ClAc-His-Ser)₄(Dap-His-Ser)₂Dap-His-Ser-NH₂

From Tenta Gel S RAM[®] resin (500 mg, 0.24 mmol·g⁻¹), G2C1 was obtained as foamy

colourless solid after preparative RP-HPLC (156.7 mg, 53.1 µmol, 44%). Analytical RP-

HPLC: $t_R = 9.071 \text{ min} (A/D \ 100/0 \text{ to } 50/50 \text{ in } 10 \text{ min}, \lambda = 214 \text{ nm})$. MS (ESI+):

C₈₀H₁₀₉Cl₄N₃₅O₂₈ found/calc. 2150.0/2150.8 [M]⁺; 2189.0/2189.9 [M+K]⁺.

Mass spectrum, MS (ESI+):





(ClAc-Pro-Ser)₄(Dap-Pro-Ser)₂Dap-Pro-Ser-NH₂

From Tenta Gel S RAM[®] resin (500 mg, 0.24 mmol·g⁻¹), **G2C2** was obtained as foamy colourless solid after preparative RP-HPLC (104.0 mg, 55.6 µmol, 46%). Analytical RP-UPLC: $t_R = 1.405 \text{ min}$ (A/D 100/0 to 0/100 in 5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{73}H_{109}Cl_4N_{21}O_{28}$ found/calc. 1869.6/1870.6 [M]⁺.

Mass spectrum, MS (ESI+):





(ClAc-His-Ser)₈(Dap-His-Ser)₄(Dap-His-Ser)₂Dap-His-Ser-NH₂

From Tenta Gel S RAM[®] resin (500 mg, 0.24 mmol·g⁻¹), G3C1 was obtained as foamy

yellowish solid after preparative RP-HPLC (116.2 mg, 18.4 µmol, 15%). Analytical RP-

HPLC: $t_R = 9.766 \text{ min} (A/D \ 100/0 \text{ to } 50/50 \text{ in } 10 \text{ min}, \lambda = 214 \text{ nm})$. MS (ESI+):

 $C_{172}H_{233}Cl_8N_{75}O_{60}$ found/calc. 4594.0/4594.8 [M]⁺.

Mass spectrum, MS (ESI+):





(ClAc-Pro-Ser)₈(Dap-Pro-Ser)₄(Dap-Pro-Ser)₂Dap-Pro-Ser-NH₂

From Tenta Gel S RAM[®] resin (500 mg, 0.24 mmol·g⁻¹), G3C2 was obtained as foamy

yellowish solid after preparative RP-HPLC (119.9 mg, 30.0 µmol, 25%). Analytical RP-

UPLC: $t_R = 1.515 \text{ min} (A/D \ 100/0 \text{ to } 0/100 \text{ in } 5 \text{ min}, \lambda = 214 \text{ nm})$. MS (ESI+):

C₁₅₇H₂₃₃Cl₈N₄₅O₆₀ found/calc. 3993.9/3994.4 [M]⁺; 4015.4/4017.4 [M+Na]⁺; 4034.3/4033.5 [M+K]⁺.

Mass spectrum, MS (ESI+):





(ClAc-Pro-Ser)₈(Lys-Pro-Ser)₄(Lys-Pro-Ser)₂Lys-Pro-Ser-NH₂

From Tenta Gel S RAM[®] resin (800 mg, 0.24 mmol·g⁻¹), G3C3 was obtained as foamy

colourless solid after preparative RP-HPLC (149.7 mg, 34.9 µmol, 18%). Analytical RP-

UPLC: $t_R = 1.510 \text{ min} (A/D \ 100/0 \text{ to } 0/100 \text{ in } 5 \text{ min}, \lambda = 214 \text{ nm})$. MS (ESI+):

C₁₇₈H₂₇₅Cl₈N₄₅O₆₀ found/calc. 4287.9/4289.0 [M]⁺; 4310.0/4312.0 [M+Na]⁺; 4326.7/4328.1 [M+K]⁺; 4348.8/4351.1 [M+Na+K]⁺; 4364.1/4367.2 [M+2K]⁺.



Analytical RP-UPLC chromatogram:



(Ac-His-Ser)₄(Dap-His-Ser)₂Dap-Cys-NH₂

From Tenta Gel S RAM[®] resin (500 mg, 0.24 mmol·g⁻¹), **G2M1** was obtained as foamy colourless solid after preparative RP-HPLC (123.0 mg, 47.8 µmol, 40%). Analytical RP-HPLC: $t_R = 7.682 \text{ min}$ (A/D 100/0 to 50/50 in 10 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{74}H_{106}N_{32}O_{26}S$ found/calc. 1891.0/1891.9 [M]⁺.

Mass spectrum, MS (ESI+):





(Ac-His-Ser)₄(*Dap*-His-Ser)₂*Dap*-His-Ser-Cys-NH₂

From Tenta Gel S RAM[®] resin (500 mg, 0.24 mmol·g⁻¹), **G2M2** was obtained as foamy colourless solid after preparative RP-HPLC (147.9 mg, 50.7 μ mol, 42%). Analytical RP-HPLC: t_R = 7.569 min (A/D 100/0 to 50/50 in 10 min, λ = 214 nm). MS (ESI+): C₈₃H₁₁₈N₃₆O₂₉S found/calc. 2116.0/2116.1 [M]⁺.

Mass spectrum, MS (ESI+):





(Pro-Ser)₄(Dap-Pro-Ser)₂Dap-Pro-Ser-Cys-NH₂

From Tenta Gel S RAM[®] resin (500 mg, 0.24 mmol·g⁻¹), **G2M3** was obtained as foamy colourless solid after preparative RP-HPLC (118.8 mg, 56.0 μ mol, 47%). Analytical RP-HPLC: t_R = 9.142 min (A/D 100/0 to 50/50 in 10 min, λ = 214 nm). MS (ESI+): C₆₈H₁₁₀N₂₂O₂₅S found/calc. 1668.0/1667.8 [M]⁺; 1709.0/1708.8 [M+2H+K]⁺.

Mass spectrum, MS (ESI+):





(Pro-Asp)₄(Dap-Pro-Asp)₂Dap-Pro-Asp-Cys-NH₂

From Tenta Gel S RAM[®] resin (500 mg, 0.24 mmol·g⁻¹), **G2M4** was obtained as foamy colourless solid after preparative RP-HPLC (110.5 mg, 59.3 µmol, 49%). Analytical RP-HPLC: $t_R = 9.258 \text{ min}$ (A/D 100/0 to 50/50 in 10 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{75}H_{110}N_{22}O_{32}S$ found/calc. 1863.0/1863.9 [M]⁺; 1902.0/1903.0 [M+K]⁺.

Mass spectrum, MS (ESI+):





(Pro-Asp)₄(Dap-Pro-Asp)₂Dap-Cys-NH₂

From Tenta Gel S RAM[®] resin (500 mg, 0.24 mmol·g⁻¹), **G2M5** was obtained as foamy colourless solid after preparative RP-HPLC (109.0 mg, 66.0 µmol, 55%). Analytical RP-HPLC: $t_R = 8.330 \text{ min}$ (A/D 100/0 to 50/50 in 10 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{66}H_{98}N_{20}O_{28}S$ found/calc. 1651.0/1651.7 [M]⁺; 1690.0/1690.8 [M+K]⁺.

Mass spectrum, MS (ESI+):





(Pro-Lys)₄(*Dap*-Pro-Lys)₂*Dap*-Pro-Lys-Cys-NH₂

From Tenta Gel S RAM[®] resin (500 mg, 0.24 mmol·g⁻¹), **G2M6** was obtained as foamy colourless solid after preparative RP-HPLC (195.9 mg, 71.2 µmol, 59%). Analytical RP-HPLC: $t_R = 9.523 \text{ min}$ (A/D 100/0 to 50/50 in 10 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{89}H_{159}N_{29}O_{18}S$ found/calc. 1955.0/1955.5 [M]⁺; 1996.0/1994.6 [M+K]⁺; 2069.0/2069.5 [M+TFA]⁺; 2183.0/2183.5 [M+2TFA]⁺; 2297.0/2297.5 [M+3TFA]⁺; 2411.0/2411.5 [M+4TFA]⁺; 2525.0/2525.5 [M+5TFA]⁺.

Mass spectrum, MS (ESI+):





(DPro-Pro-Glu)₄(Dap-Pro-Asp)₂Dap-Pro-Asp-Cys-NH₂

From Tenta Gel S RAM[®] resin (500 mg, 0.24 mmol·g⁻¹), **G2M7** was obtained as foamy colourless solid after preparative RP-HPLC (149.9 mg, 54.2 µmol, 45%). Analytical RP-UPLC: $t_R = 1.267 \text{ min}$ (A/D 100/0 to 0/100 in 5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{99}H_{146}N_{26}O_{36}S$ found/calc. 2307.5/2308.4 [M]⁺.

Mass spectrum, MS (ESI+):





(Pro-Ser)₈(Dap-Pro-Ser)₄(Dap-Pro-Ser)₂Dap-Pro-Ser-Cys-NH₂

From Tenta Gel S RAM[®] resin (500 mg, 0.24 mmol·g⁻¹), **G3M1** was obtained as foamy

colourless solid after preparative RP-HPLC (149.0 mg, 42.8 µmol, 36%). Analytical RP-

HPLC: $t_R = 8.995 \text{ min} (A/D \ 100/0 \text{ to } 50/50 \text{ in } 10 \text{ min}, \lambda = 214 \text{ nm})$. MS (ESI+):

C₁₄₄H₂₃₀N₄₆O₅₃S found/calc. 3485.0/3485.7 [M]⁺; 3527.0/3526.8 [M+2H+K]⁺; 3568.0/3567.9

[M+4H+2K]⁺; 3610.0/3610.0 [M+7H+3K]⁺; 3650.0/3650.2 [M+8H+4K]⁺.



Analytical RP-HPLC chromatogram:



(Ac-Glu-Ala)₁₆(*Lys*-Amb-Tyr)₈(*Dap*-Cys(Asp)-x-His-Ser)₄(*Dap*-His-Ser)₂*Dap*-His-Ser-NH₂ From starting materials G2C1 and B1G2 using the general procedure described above (solvent: 0.2 M phosphate buffer pH 8.5), G4E was obtained as colourless solid after preparative RP-HPLC (8.23 mg). Amino Acid Analysis: protein content 52.5% (4.32 mg, 0.40 µmol, yield 28%); found/calc. (%) Glu 17.6/18.8, Ala 17.3/18.8, Dap 11.8/8.2, Tyr 10.0/9.4, Amb 9.5/9.4, Lys 9.2/9.4, His 7.8/8.2, Ser 7.1/8.2, Asp 4.8/4.7, CM-Cys 4.8/4.7 (AMBA t_R = 7.03 min, 1552.4 pmol, Dap coeluted with Phe, 1941.7 pmol). Analytical RP-UPLC: $t_R = 1.467 min (A/D 100/0 to 0/100 in 5 min, \lambda = 214 nm)$. MS (ESI-): C₄₆₄H₆₂₉N₁₁₉O₁₆₀S₄ (4fold ligation product) found/calc. 10562.0/10561.9 [M]⁻; 10624.0/10624.0 [M+Na+K]⁻. No incomplete ligation products observed (3-fold ligation product calc. 8459.1 [M]⁻, 2-fold ligation product calc. 6356.3 [M]⁻). D = 10.25 \cdot 10^{-11} m² \cdot s⁻¹; R_h = 1.98 ± 0.08 nm.



Analytical RP-UPLC chromatogram:



Amino Acid Analysis:



F	T	RT (STD)	PW(50%)	Area	Height	n.a.	Amount	Peak Name
m	in	min	min	mAU*min	mAU	n.a.	pmol	
	1,35	1,35	0,105	8,08	66,57		345,6	EDTA
	1,96	1,96	0,083	12,47	138,31		792,4	Asp
	2,14	2,15	0,083	50,36	479,81		2894,3	Glu
	2,50	2,51	0,083	13,14	124,93		790,2	CM-Cys
	3,30	3,30	0,090	19,23	172,02		1169,1	Ser
	3,72	3,72	0,096	23,23	185,68		1274,4	His
	4,66	4,67	0,100	54,36	423,60		2846,0	Ala
	6,85	6,85	0,129	28,69	220,61		1639,0	Tyr
	9,04	9,02	0,075	0,06	0,85		3,5	C-
	9,36	9,32	n.a.	0,14	1,79		7,3	C-C
	9,71	9,73	0,071	0,12	1,70		12,0	lle
	10,20	10,21	0,125	3,93	27,92		618,3	NH3
	10,73	10,80	0,115	38,97	258,36		1941,7	Phe
	11,86	11,88	0,112	49,75	354,49		1513,2	Lys
Tota	l:			302,528	2456,635		15846,96	



(Ac-Glu-Ala)₃₂(*Lys*-Amb-Tyr)₁₆(*Dap*-Cys(Asp)-x-His-Ser)₈(*Dap*-His-Ser)₄(*Dap*-His-Ser)₂*Dap*-His-Ser-NH₂

From starting materials **G3C1** and **B1G2** using the general procedure described above (solvent: 0.2 M phosphate buffer pH 8.5), **G5E** was obtained as foamy colourless solid after preparative RP-HPLC (13.3 mg). Amino Acid Analysis: protein content 32.3% (4.29 mg, 0.19 µmol, yield 30%); found/calc. (%) Ala 19.2/18.5, Glu 15.2/18.5, Dap 13.1/8.7, Tyr 9.8/9.2, His 9.7/8.7, Lys 9.2/9.2, Amb 9.0/9.2, Ser 8.7/8.7, CM-Cys 4.2/4.6, Asp 1.9/4.6 (Dap coeluted with Phe, 1324.4 pmol). Analytical RP-UPLC: $t_R = 1.480 \text{ min}$ (A/D 100/0 to 0/100 in 5 min, $\lambda = 214 \text{ nm}$). MS (ESI-): $C_{940}H_{1273}N_{243}O_{324}S_8$ (8-fold ligation product, main peak) found/calc. 21417.0/21416.1 [M-H]⁻. $C_{844}H_{1143}ClN_{222}O_{291}S_7$ (7-fold ligation product with one unreacted chloroacetyl group, peak height 33% of main peak) found/calc. 19311.0/19313.3 [M-H]⁻, no 6-fold ligation product observed (calc. 17210.5 [M-H]⁻). D = 8.63 \cdot 10^{-11} m² \cdot s⁻¹; $R_h = 2.35 \pm 0.02 \text{ nm}.$



Analytical RP-UPLC chromatogram:



Amino Acid Analysis:



RT	RT (STD)	PW(50%)	Area	Height	n.a.	Amount	Peak Name
min	min	min	mAU*min	mAU	n.a.	pmol	
1,19	1,20	0,068	0,93	12,44		405,8	EDTA
1,96	1,96	0,066	2,60	34,73		195,7	Asp
2,16	2,16	0,072	23,34	259,76		1541,2	Glu
2,55	2,55	0,073	6,03	65,63		422,2	CM-Cys
3,43	3,43	0,077	13,41	141,19		879,5	Ser
3,83	3,83	0,077	15,06	156,13		975,8	His
4,88	4,88	0,080	29,88	314,07		1940,5	Ala
7,20	7,20	0,083	16,41	168,98		989,3	Tyr
7,42	7,42	0,090	25,38	241,01		906,8	AMBA
11,15	11,15	0,136	7,81	52,47		1112,5	NH3
11,38	11,38	0,088	21,68	207,72		1324,4	Phe
12,54	12,54	0,086	27,26	270,46		933,8	Lys
Total:			189,773	1924,580		11627,32	



(Ac-His-Ser)₁₆(Dap-His-Ser)₈(Dap-Cys-x-His-Ser)₄(Dap-His-Ser)₂Dap-His-Ser-NH₂

From starting materials **G2C1** and **G2M1** using the general procedure described above (solvent: 0.2 M phosphate buffer pH 8.5), **G4H1** was obtained as foamy colourless solid after preparative RP-HPLC (4.02 mg). Amino Acid Analysis: protein content 46.6% (1.87 mg, 0.17 µmol, yield 12%); found/calc. (%) His 47.8/38.3, Ser 31.5/38.3, Dap 16.3/18.5, CM-Cys 4.4/4.9. Analytical RP-UPLC: $t_R = 1.228 \text{ min}$ (A/D 100/0 to 0/100 in 5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{376}H_{529}N_{163}O_{132}S_4$ (4-fold ligation product) found/calc. 9572.0/9572.5 [M]⁺. No incomplete ligation products observed (3-fold ligation product calc. 7717.1 [M]⁺, 2-fold ligation product calc. 5861.6 [M]⁺). D = 7.70 \cdot 10^{-11} \text{ m}^2 \cdot \text{s}^{-1}; $R_h = 2.63 \pm 0.06 \text{ nm}$.



Analytical RP-UPLC chromatogram:



Amino Acid Analysis:



RT	RT (STD)	PW(50%)	Area	Height	n.a.	Amount	Peak Name
min	min	min	mAU*min	mAU	n.a.	pmol	
1,17	1,17	0,092	6,80	67,03		527,9	EDTA
2,03	2,07	0,074	0,76	10,08		53,6	Asp
2,30	2,30	0,056	0,18	3,08		17,5	Glu
2,73	2,73	0,073	6,95	85,87		483,0	CM-Cys
3,58	3,59	0,080	47,82	532,12		3414,5	Ser
3,81	3,84	n.a.	0,58	7,58		48,9	Gly
4,05	4,06	0,078	69,23	776,90		5188,9	His
5,13	5,15	0,081	0,40	4,71		30,0	Ala
11,81	11,77	0,094	49,62	480,82		1768,3	Dap
Total:						11532,76	



Representative Michaelis-Menten plots for determination of k_{cat} and K_{M} :



(Ac-His-Ser)₁₆(*Dap*-His-Ser)₈(*Dap*-His-Ser-Cys-x-His-Ser)₄(*Dap*-His-Ser)₂*Dap*-His-Ser-NH₂ From starting materials **G2C1** and **G2M2** using the general procedure described above (solvent: 0.2 M phosphate buffer pH 8.5), **G4H2** was obtained as foamy colourless solid after preparative RP-HPLC (2.24 mg). Amino Acid Analysis: protein content 59.7% (1.34 mg, 0.11 µmol, yield 8%); found/calc. (%) His 48.8/39.3, Ser 32.1/39.3, Dap 14.7/16.9, CM-Cys 4.3/4.5. Analytical RP-UPLC: $t_R = 1.230 \text{ min}$ (A/D 100/0 to 0/100 in 5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{412}H_{577}N_{179}O_{144}S_4$ (4-fold ligation product, main peak) found/calc. 10471.0/10470.4 [M+H]⁺; 10511.0/10510.5 [M+2H+K]⁺. $C_{329}H_{460}CIN_{143}O_{115}S_3$ (3-fold ligation product with one unreacted chloroacetyl group, peak height 9% of main peak) found/calc. 8392.0/8390.7 [M+H]⁺, no 2-fold ligation product observed (calc. 6310.1 [M]⁺). D = 6.94 \cdot 10^{-11} m² \cdot s⁻¹; $R_h = 2.92 \pm 0.09 \text{ nm}.$



Analytical RP-UPLC chromatogram:



Amino Acid Analysis:



RT	RT (STD)	PW(50%)	Area	Height	n.a.	Amount	Peak Name
min	min	min	mAU*min	mAU	n.a.	pmol	
1,17	1,17	0,097	8,41	77,42		609,7	EDTA
2,04	2,07	0,073	0,48	6,93		36,9	Asp
2,73	2,73	0,072	9,21	117,09		658,6	CM-Cys
3,58	3,59	0,080	67,61	767,23		4923,1	Ser
4,05	4,06	0,077	99,37	1120,17		7481,6	His
5,13	5,15	0,079	0,63	7,69		49,0	Ala
11,81	11,77	0,093	62,49	613,08		2254,8	Dap
Total:						16013,72	





Representative Michaelis-Menten plots for determination of k_{cat} and K_{M} :



(Ac-His-Ser)₃₂(*Dap*-His-Ser)₁₆(*Dap*-His-Ser-Cys-x-Pro-Ser)₈(*Dap*-Pro-Ser)₄(*Dap*-Pro-Ser)₂(*Dap*-Pro-Ser-NH₂)

From starting materials **G3C2** and **G2M2** using the general procedure described above (solvent: DMF/H₂O (1/1 v/v), 20 eq KI), **G5H** was obtained as foamy colourless solid after preparative RP-HPLC (13.7 mg). Amino Acid Analysis: protein content 64.1% (8.75 mg, 0.38 µmol, yield 50%); found/calc. (%) His 37.2/30.9, Ser 31.8/39.2, Dap 15.6/17.1, Pro 10.9/8.3, CM-Cys 4.5/4.4. Analytical RP-UPLC: $t_R = 1.243 \text{ min}$ (A/D 100/0 to 0/100 in 5 min, $\lambda = 214$ nm). MS (LC-MS+): $C_{821}H_{1169}N_{333}O_{292}S_8$ (8-fold ligation product, main peak) found/calc. 20634.0/20633.6 [M+2H]⁺. $C_{738}H_{1053}N_{297}O_{264}S_7$ (7-fold ligation product with one hydroxyacetyl group, peak height 23% of main peak) found/calc. 18535.0/18534.5 [M+H]⁺, no 6-fold ligation product observed (calc. 16655.3 [M]⁺). D = 8.07 \cdot 10⁻¹¹ m² · s⁻¹; R_h = 2.51 ± 0.07 nm.



Analytical RP-UPLC chromatogram:



Amino Acid Analysis:



RT	RT (STD)	PW(50%)	Area	Height	n.a.	Amount	Peak Name
min	min	min	mAU*min	mAU	n.a.	pmol	
1,17	1,17	0,095	7,30	67,68		533,1	EDTA
2,03	2,07	0,070	0,82	11,25		59,9	Asp
2,73	2,73	0,073	12,46	154,82		870,9	CM-Cys
3,58	3,59	0,081	86,15	961,32		6168,4	Ser
4,05	4,06	0,078	95,58	1079,17		7207,8	His
5,13	5,15	0,073	0,65	8,58		54,7	Ala
5,60	5,64	0,082	34,41	385,45		2109,4	Pro
11,81	11,77	0,092	83,53	824,23		3031,3	Dap
Total:						20035,41	



Representative Michaelis-Menten plots for determination of k_{cat} and K_{M} :



(Pro-Ser)₁₆(*Dap*-Pro-Ser)₈(*Dap*-Pro-Ser-Cys-x-Pro-Ser)₄(*Dap*-Pro-Ser)₂*Dap*-Pro-Ser-NH₂ From starting materials **G2C2** and **G2M3** using the general procedure described above (solvent: DMF/H₂O (1/1 v/v), 20 eq KI), **G4P** was obtained as foamy colourless solid after preparative RP-HPLC (2.05 mg). Amino Acid Analysis: protein content 65.8% (1.35 mg, 0.15 µmol, yield 55%); found/calc. (%) Pro 44.0/39.3, Ser 33.8/39.3, Dap 16.9/16.9, CM-Cys 5.4/4.5. Analytical RP-HPLC: $t_R = 9.156 \text{ min}$ (A/D 100/0 to 50/50 in 10 min, $\lambda = 214 \text{ nm}$). MS (LC-MS+): $C_{345}H_{545}N_{109}O_{128}S_4$ (4-fold ligation product) found/calc. 8393.0/8395.9 [M]⁺. No incomplete ligation products observed (3-fold ligation product calc. 6856.0 [M]⁺, 2-fold ligation product calc. 5316.2 [M]⁺). D = 9.72 \cdot 10⁻¹¹ m² · s⁻¹; R_h = 2.09 ± 0.07 nm.





Amino Acid Analysis:





(Pro-Asp)₃₂(*Dap*-Pro-Asp)₁₆(*Dap*-Cys-x-Pro-Ser)₈(*Dap*-Pro-Ser)₄(*Dap*-Pro-Ser)₂*Dap*-Pro-Ser-NH₂

From starting materials **G3C2** and **G2M5** using the general procedure described above (solvent: 0.1 M phosphate buffer pH 8.5, 20 eq KI), **G5P1** was obtained as foamy colourless solid after preparative RP-HPLC (10.3 mg). Amino Acid Analysis: protein content 76.5% (7.86 mg, 0.43 µmol, yield 57%); found/calc. (%) Pro 39.4/38.2, Asp 30.7/29.1, Dap 19.0/18.8, Ser 6.3/9.1, CM-Cys 4.7/4.8. Analytical RP-UPLC: $t_R = 1.245 \text{ min}$ (A/D 100/0 to 0/100 in 5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{685}H_{1009}N_{205}O_{284}S_8$ (8-fold ligation product) found/calc. 16916.0/16916.1 [M]⁺. No incomplete ligation products observed (7-fold ligation product calc. 15392.3 [M]⁺, 6-fold ligation product calc. 13868.6 [M]⁺). D = 7.36 \cdot 10^{-11} \text{ m}^2 \cdot \text{s}^{-1}; $R_h = 2.76 \pm 0.05 \text{ nm}$.



Analytical RP-UPLC chromatogram:



Amino Acid Analysis:



RT	RT (STD)	PW(50%)	Area	Height	n.a.	Amount	Peak Name
				IIIAO	n.a.	pinor	
1,29	1,25	0,105	6,73	58,72		216,5	EDTA
1,95	1,95	0,090	125,82	1191,46		8419,4	Asp
2,48	2,48	0,108	17,67	150,15		1287,3	CM-Cys
3,31	3,31	0,124	25,58	187,14		1732,4	Ser
5,12	5,15	0,128	207,01	1438,38		10804,3	Pro
11,07	11,07	0,143	129,03	811,67		5208,7	Dap
Total:			511,837	3837,522		27668,43	



(Pro-Ser)₃₂(*Dap*-Pro-Ser)₁₆(*Dap*-Pro-Ser-Cys-x-Pro-Ser)₈(*Dap*-Pro-Ser)₄(*Dap*-Pro-Ser)₄(*Dap*-Pro-Ser)₂*Dap*-Pro-Ser-NH₂

From starting materials **G3C2** and **G2M3** using the general procedure described above (solvent: DMF/H₂O (1/1 v/v), 20 eq KI), **G5P2** was obtained as foamy colourless solid after preparative RP-HPLC (1.48 mg). Amino Acid Analysis: protein content 62.4% (0.92 mg, 0.05 µmol, yield 40%); found/calc. (%) Pro 44.1/39.2, Ser 33.3/39.2, Dap 17.2/17.1, CM-Cys 5.4/4.4. Analytical RP-HPLC: $t_R = 9.132 \text{ min}$ (A/D 100/0 to 50/50 in 10 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{701}H_{1105}N_{221}O_{260}S_8$ (8-fold ligation product) found/calc. 17043.0/17045.1 [M]⁺. No incomplete ligation products observed (7-fold ligation product calc. 15505.2 [M]⁺, 6-fold ligation product calc. 13965.4 [M]⁺). D = 5.93 \cdot 10⁻¹¹ m² \cdot s⁻¹; R_h = 3.42 ± 0.04 nm.



Electronic Supplementary Material (ESI) for Organic & Biomolecular Chemistry This journal is C The Royal Society of Chemistry 2011



Amino Acid Analysis:



RT	RT (STD)	PW(50%)	Area	Height	n.a.	Amount	Peak Name
min	min	min	mAU*min	mAU	n.a.	pmol	
1,23	1,21	0,090	4,41	41,71		301,5	EDTA
2,62	2,63	0,073	17,97	192,86		1320,3	CM-Cys
3,48	3,50	0,082	129,94	1165,48		8120,9	Ser
5,38	5,41	0,087	210,46	1789,89		10758,5	Pro
11,47	11,49	0,094	138,43	1091,98		4206,2	DAP
Total:			501,211	4281,917		24707,35	



(Pro-Asp)₃₂(*Dap*-Pro-Asp)₁₆(*Dap*-Pro-Asp-Cys-x-Pro-Ser)₈(*Dap*-Pro-Ser)₄(*Dap*-Pro-Ser)₂*Dap*-Pro-Ser-NH₂

From starting materials **G3C2** and **G2M4** using the general procedure described above (solvent: 0.5 M NaHCO₃ buffer pH 8.0, 20 eq KI), **G5P3** was obtained as foamy colourless solid after preparative RP-HPLC (14.7 mg). Amino Acid Analysis: protein content 63.6% (9.33 mg, 0.46 µmol, yield 62%); found/calc. (%) Pro 40.4/39.2, Asp 31.9/30.9, Dap 17.1/17.1, Ser 6.6/8.3, CM-Cys 4.1/4.4. Analytical RP-UPLC: $t_R = 1.245 \text{ min}$ (A/D 100/0 to 0/100 in 5 min, $\lambda = 214 \text{ nm}$). MS (LC-MS+): $C_{757}H_{1105}N_{221}O_{316}S_8$ (8-fold ligation product, main peak) found/calc. 18620.0/18619.7 [M+6H]⁺. $C_{682}H_{996}IN_{199}O_{284}S_7$ (7-fold ligation product with one unreacted iodoactyl group, peak height 26% of main peak) found/calc. 16876.0/16877.7 [M]⁺, no 6-fold ligation product observed (calc. 15141.9). D = 6.50 \cdot 10^{-11} m^2 \cdot s^{-1}; $R_h = 3.12 \pm 0.04 \text{ nm}$.



Analytical RP-UPLC chromatogram:



Amino Acid Analysis:



RT min	RT (STD) min	PW(50%) min	Area mAU*min	Height mAU	n.a. n.a.	Amount pmol	Peak Name
1,27	1,25	0,113	12,40	99,21		365,7	EDTA
1,95	1,95	0,092	111,47	1033,47		7302,9	Asp
2,48	2,48	0,110	13,20	110,19		944,7	CM-Cys
3,31	3,31	0,127	23,89	163,31		1511,8	Ser
5,13	5,15	0,129	177,38	1231,46		9250,0	Pro
11,08	11,07	0,144	97,82	608,95		3907,8	Dap
Total:			436,168	3246,587		23282,87	



(Pro-Lys)₃₂(*Dap*-Pro-Lys)₁₆(*Dap*-Pro-Lys-Cys-x-Pro-Ser)₈(*Dap*-Pro-Ser)₄(

From starting materials G3C2 and G2M6 using the general procedure described above (solvent: 0.5 M NaHCO₃ buffer pH 8.0, 20 eq KI), G5P4 was obtained as crystalline colourless solid after preparative RP-HPLC (16.4 mg). Amino Acid Analysis: protein content 82.3% (13.5 mg, 0.58 µmol, yield 77%); found/calc. (%) Pro 40.8/39.2, Lys 31.5/30.9, Dap 17.8/17.1, Ser 6.4/8.3, CM-Cys 3.5/4.4. Analytical RP-UPLC: $t_R = 1.255 \text{ min}$ (A/D 100/0 to 0/100 in 5 min, $\lambda = 214 \text{ nm}$). MS (LC-MS+): $C_{869}H_{1497}N_{277}O_{204}S_8$ (8-fold ligation product, main peak) found/calc. 19353.0/19352.4 [M+6H]⁺. C₇₈₀H₁₃₄₀N₂₄₈O₁₈₇S₇ (7-fold ligation product with one hydroxyacetyl group, peak height 10% of main peak) found/calc. 17411.0/17411.0 [M+2H]⁺, no 6-fold ligation product observed (calc. 15471.5 [M]⁺). D = 4.27 \cdot 10⁻¹¹ m² \cdot s⁻¹; R_h = 4.74 ± 0.15 nm.



Analytical RP-UPLC chromatogram:



Amino Acid Analysis:



RT	RT (STD)	PW(50%)	Area	Height	n.a.	Amount	Peak Name
min	min	min	mAU [*] min	mau	n.a.	pmol	
1,30	1,25	0,105	2,65	23,48		86,5	EDTA
2,48	2,48	0,111	12,09	102,85		881,8	CM-Cys
3,31	3,31	0,126	24,30	176,90		1637,5	Ser
5,12	5,15	0,131	201,70	1384,30		10398,0	Pro
11,08	11,07	0,142	111,38	707,13		4537,9	Dap
12,26	12,29	0,140	255,27	1634,17		8016,5	Lys
Total:			607,403	4028,835		25558,22	



$(Pro-Ser)_{32}(Dap-Pro-Ser)_{16}(Dap-Pro-Ser-Cys-\textbf{x}-His-Ser)_8(Dap-His-Ser)_4(Dap-His-Ser)_4(Dap-His-Ser)_{16}(Dap-H$

Ser)₂Dap-His-Ser-NH₂

From starting materials **G3C1** and **G2M3** using the general procedure described above (solvent: 0.5 M NaHCO₃ buffer pH 8.0/CH₃CN (2/1 v/v), 20 eq KI), **G5P5** was obtained as foamy colourless solid after preparative RP-HPLC (2.77 mg,). Amino Acid Analysis: protein content 52.2% (1.45 mg, 0.07 µmol, yield 11%); found/calc. (%) Pro 37.5/30.9, Ser 31.2/39.2, His 10.7/8.3, Dap 16.0/17.1, CM-Cys 4.6/4.4. Analytical RP-UPLC: $t_R = 1.144 \text{ min}$ (A/D 100/0 to 0/100 in 5 min, $\lambda = 214 \text{ nm}$). MS (ESI+): $C_{716}H_{1105}N_{251}O_{260}S_8$ (8-fold ligation product, main peak) found/calc. 17645.0/17645.5 [M]⁺. $C_{648}H_{997}N_{229}O_{236}S_7$ (7-fold ligation product with one hydroxyacetyl group, peak height 19% of main peak) found/calc. 15995.0/15995.7 [M]⁺, no 6-fold ligation product observed (calc. 14565.8 [M]⁺). D = 5.57 \cdot 10⁻ ¹¹ m² · s⁻¹; R_h = 3.64 ± 0.10 nm.

Uhlich NAU-197 III F16_100212134638_XT_00001_M_#1_RT: 1.0_AV: 1_NL: 1.09E4 T: FTMS + p NSI Full ms [600.00-2000.00] 17644 98 10500 10000 9500 9000 8500 8000 7500 7000 6500 6000 5500 17588.9 5000 4500 17302.80 4000 17730.01 3500 15837.12 3000 2500 2000 3667 14 1500 1000-17983.08 500-8038.15 16000 m/z 19000 20000 12000 13000 17000 18000

Analytical RP-UPLC chromatogram:



Amino Acid Analysis:



RT	RT (STD)	PW(50%)	Area	Height	n.a.	Amount	Peak Name
min	min	min	mAU*min	mAU	n.a.	pmol	
1,17	1,17	0,093	6,71	65,70		517,5	EDTA
2,03	2,07	0,067	0,54	8,18		43,5	Asp
2,73	2,73	0,073	14,00	170,18		957,3	CM-Cys
3,57	3,59	0,080	90,36	1019,94		6544,6	Ser
4,05	4,06	0,079	30,92	337,59		2254,7	His
5,58	5,64	0,084	133,14	1439,55		7878,0	Pro
11,80	11,77	0,092	92,12	910,10		3347,1	Dap
Total:						21542,77	



Representative Michaelis-Menten plots for determination of k_{cat} and K_{M} :



$(Pro-Asp)_{32}(Dap-Pro-Asp)_{16}(Dap-Pro-Asp-Cys-x-Pro-Ser)_8(Lys-Pro-Ser)_4(Lys-Pro-Ser)_4(Lys-Pro-Ser)_8(Ly$

Ser)₂Lys-Pro-Ser-NH₂

From starting materials **G3C3** and **G2M4** using the general procedure described above (solvent: DMF/H₂O (1/1 v/v), 20 eq KI), **G5P6** was obtained as foamy colourless solid after preparative RP-HPLC (11.8 mg). Amino Acid Analysis: protein content 80.0% (9.41 mg, 0.46 µmol, yield 66%); found/calc. (%) Pro 39.6/39.2, Asp 31.7/30.9, Dap 12.3/13.3, Ser 7.5/8.3, CM-Cys 4.6/4.4, Lys 4.3/3.9 (Dap coeluted with Phe, 3501.1 pmol). Analytical RP-UPLC: $t_R = 1.214 \text{ min}$ (A/D 100/0 to 0/100 in 5 min, $\lambda = 214 \text{ nm}$). MS (LC-MS+): C₇₇₈H₁₁₄₇N₂₂₁O₃₁₆S₈ (8-fold ligation product, main peak) found/calc. 18907.0/18908.2 [M]⁺. C₇₀₃H₁₀₃₈IN₁₉₉O₂₈₄S₇ (7-fold ligation product with one unreacted iodoacetyl group, peak height 28% of main peak) found/calc. 17165.0/17172.3 [M]⁺. C₆₂₈H₉₂₉I₂N₁₇₇O₂₅₂S₆ (6-fold ligation product with two unreacted iodoacetyl groups, peak height 11% of main peak) found/calc. 15467.0/15436.3 [M]⁺. D = 6.61 \cdot 10⁻¹¹ m² \cdot s⁻¹; R_h = 3.07 ± 0.05 nm.



Analytical RP-UPLC chromatogram:



Amino Acid Analysis:



RT	RT (STD)	PW(50%)	Area	Height	n.a.	Amount	Peak Name
min	min	min	mAU*min	mAU	n.a.	pmol	
1,20	1,19	0,091	7,17	72,64		448,4	EDTA
1,98	1,97	0,069	144,79	1852,22		9006,1	Asp
2,54	2,54	0,078	20,00	234,09		1297,4	CM-Cys
3,40	3,39	0,087	32,88	345,30		2139,7	Ser
5,29	5,32	0,099	230,18	2078,98		11247,0	Pro
11,54	11,55	0,106	98,99	864,92		5802,3	Phe
12,77	12,75	0,102	36,83	338,42		1222,0	Lys
Total:						31162,91	



(DPro-Pro-Glu)₃₂(Dap-Pro-Asp)₁₆(Dap-Pro-Asp-Cys-x-Pro-Ser)₈(Lys-Pro-Ser)₄(Lys-Pro-Ser)₂Lys-Pro-Ser-NH₂

From starting materials **G3C3** and **G2M7** using the general procedure described above (solvent: DMF/H₂O (1/1 v/v), 20 eq KI), **G5P7** was obtained as foamy colourless solid after preparative RP-HPLC (15.6 mg). Amino Acid Analysis: protein content 60.7% (9.46 mg, 0.40 µmol, yield 56%); found/calc. (%) Pro 51.7/48.4, Glu 15.3/15.0, Asp 11.6/11.3, Dap 10.6/11.3, Ser 6.6/7.0, CM-Cys 4.2/3.8, Lys 3.7/3.3 (Dap coeluted with Phe, 2208.5 pmol). Analytical RP-UPLC: $t_R = 1.304$ min (A/D 100/0 to 0/100 in 5 min, $\lambda = 214$ nm). MS (LC-MS+): C₉₇₀H₁₄₃₅N₂₅₃O₃₄₈S₈ (8-fold ligation product, main peak) found/calc. 22459.0/22464.8 [M]⁺. C₈₇₁H₁₂₉₀IN₂₂₇O₃₁₂S₇ (7-fold ligation product with one unreacted iodoacetyl group, peak height 12% of main peak) found/calc. 20270.0/20271.9 [M]⁺, no 6-fold ligation product observed (calc. 18117.8 [M]⁺). D = 6.31 \cdot 10⁻¹¹ m²·s⁻¹; R_h = 3.21 ± 0.08 nm.



Analytical RP-UPLC chromatogram:



Amino Acid Analysis:



RT	RT (STD)	PW(50%)	Area	Height	n.a.	Amount	Peak Name
min	min	min	mAU*min	mAU	n.a.	pmol	
1,21	1,20	0,077	10,15	113,93		403,9	EDTA
1,89	9 1,88	0,065	34,55	487,42		2421,5	Asp
2,05	5 2,05	0,072	46,85	595,66		3187,4	Glu
2,37	2,37	0,076	11,02	134,10		875,7	CM-Cys
3,21	3,21	0,088	19,44	206,61		1371,5	Ser
4,98	3 5,01	0,098	196,05	1839,30		10773,1	Pro
11,10) 11,11	0,112	65,24	490,87		3745,3	Phe
12,31	12,31	0,106	21,06	188,65		779,3	Lys
Total:						23557,69	



(Pro-Ser)₆₄(Dap-Pro-Ser)₃₂(Dap-Pro-Ser)₁₆(Dap-Pro-Ser-Cys-x-Pro-Ser)₈(Dap-Pro-

Ser)₄(*Dap*-Pro-Ser)₂*Dap*-Pro-Ser-NH₂

From starting materials **G3C2** and **G3M1** using the general procedure described above (solvent: DMF/H₂O (1/1 v/v), 20 eq KI), **G6P1** was obtained as foamy colourless solid after preparative RP-HPLC (17.7 mg). Amino Acid Analysis: protein content 65.8% (11.6 mg, 0.34 µmol, yield 45%); found/calc. (%) Pro 44.2/39.6, Ser 30.3/39.6, Dap 22.9/18.5, CM-Cys 2.6/2.3. Analytical RP-UPLC: $t_R = 1.189 \text{ min}$ (A/D 100/0 to 0/100 in 5 min, $\lambda = 214 \text{ nm}$). MS: $C_{1309}H_{2065}N_{413}O_{484}S_8$ (8-fold ligation product) found/calc. N.d./31588.4 [M]⁺. D = 3.68 \cdot 10^{-11} m^2 \cdot s^{-1}; $R_h = 5.51 \pm 0.07 \text{ nm}$.



Amino Acid Analysis:



RT	RT (STD)	PW(50%)	Area	Height	n.a.	Amount	Peak Name
min	min	min	mAU*min	mAU	n.a.	pmol	
1,20	1,19	0,089	4,82	52,17		472,8	EDTA
2,55	2,55	0,077	10,19	125,26		675,6	CM-Cys
3,40	3,41	0,092	141,75	1357,27		7869,0	Ser
5,29	5,32	0,101	247,89	2223,93		11480,3	Pro
11,46	11,50	0,111	162,16	1275,08		5952,4	DAP
Total:						26450,22	



(Pro-Ser)₆₄(*Dap*-Pro-Ser)₃₂(*Dap*-Pro-Ser)₁₆(*Dap*-Pro-Ser-Cys-**x**-Pro-Ser)₈(*Lys*-Pro-Ser)₄(*Lys*-Pro-Ser)₂*Lys*-Pro-Ser-NH₂

From starting materials **G3C3** and **G3M1** using the general procedure described above (solvent: DMF/H₂O (1/1 v/v), 20 eq KI), **G6P2** was obtained as foamy colourless solid after preparative RP-HPLC (18.2 mg). Amino Acid Analysis: protein content 68.6% (12.5 mg, 0.36 µmol, yield 51%); found/calc. (%) Pro 44.2/39.6, Ser 32.9/39.6, Dap 17.4/16.4, CM-Cys 3.0/2.3, Lys 2.6/2.1 (Dap coeluted with Phe, 4658.9 pmol). Analytical RP-UPLC: $t_R = 1.202$ min (A/D 100/0 to 0/100 in 5 min, $\lambda = 214$ nm). MS: $C_{1330}H_{2107}N_{413}O_{484}S_8$ (8-fold ligation product) found/calc. N.d./31883.0 [M]⁺. D = 4.15 \cdot 10^{-11} m^2 \cdot s^{-1}; $R_h = 4.88 \pm 0.18$ nm.



Amino Acid Analysis:



RT	RT (STD)	PW(50%)	Area	Height	n.a.	Amount	Peak Name
min	min	min	mAU*min	mAU	n.a.	pmol	
1,21	1,20	0,067	3,49	44,85		159,0	EDTA
2,36	2,37	0,078	10,51	122,43		799,5	CM-Cys
3,21	3,21	0,088	130,82	1329,57		8825,6	Ser
5,00	5,01	0,096	210,28	2026,81		11871,4	Pro
11,09	11,11	0,110	121,18	1035,49		7900,6	Phe
12,31	12,31	0,106	18,72	168,58		696,4	Lys
Total:						30252,51	



Amino Acid Analysis

Samples were hydrolyzed in the gas phase with 6M HCl containing 0.1% (v/v) phenol for 22 h at 115 °C under N₂ vacuum according to Chang and Knecht.¹ The liberated amino acids were coupled with phenylisothiocyanate (PITC), and the resulting phenylthiocarbamoyl (PTC) amino acids were analyzed by RP-HPLC on a Nova Pack C18 column (4 μ m, 3.9 mm × 150 mm, Waters) with a Dionex Summit[®] HPLC system with an automatic injection system according to Bidlingmeyer et al.² The corresponding ammonium acetate buffer replaced the 0.14 M sodium acetate buffer, pH 6.3. Cysteine was detected as carboxymethyl cysteine (CM-Cys); AMBA = Amb = 4-aminomethylbezoic acid. Dap coeluted with Phe. For some compounds the amount of Dap (pmol) was calculated based on a separate standard and is indicated within the characterization data.

Sodium dodecylsulfate polyacrylamide gel electrophoresis (SDS-PAGE)

To cast the gels ($84 \times 73 \times 0.75$ mm), Bio-Rad short plates/spacer plates were filled with resolving gel until 2 cm under the glass rim and covered with mQ-deionized H₂O. When polymerization was finished (~ 20 min) the water was removed and stacking gel was poured on top of the resolving gel. The pocket forming comb was inserted. Polymerization of the stacking gel was generally finished after 5 min. The gels were used in electrophoresis boxes purchased from Bio-Rad. After the addition of electrophoresis buffer, the gel pockets were washed with the latter and compounds (6-36 µg dissolved in a mixture of 12.5 µL mQdeionized H₂O and 2.5 µL sample buffer) or molecular marker (2.5 µL of the prepared solution) were added using a 50 µL glass syringe. Power was supplied by a Consort E452

⁽¹⁾ Chang, J.-Y.; Knecht, R. Anal. Biochem. 1991, 197, 52-58.

⁽²⁾ Bidlingmeyer, B. A.; Cohen, S. A.; Tarvin, T. L. J. Chromatogr. 1984, 336, 93-104.

(200 V) during 0.7 to 1.3 h. To develop the gels, they were removed from the glass plates and stained in a staining/fixation bath for 3 min in a microwave oven at 600 W followed by a background destaining bath (4×15 min). The gels were washed in a mixture of mQ-deionized H₂O/glycerol and slowly dried at rt between two permeable jam foils. Pictures of the gels were obtained by the use of a flat-bed scanner.

Preparation. All stock solutions and buffers were filtered prior to use.

Resolving gel 20% (for two gels). 1.5 M Tris·Base pH 8.7 (2.5 mL), aq. solution of SDS (10%, 0.1 mL), aq. solution of acrylamide (Rotiphorese Gel A[®], 30%, 6.65 mL), aq. solution of bisacrylamide (Rotiphorese Gel B[®], 2%, 0.325 mL), aq. solution of APS (10%, 33 μ L), TMED (5 μ L).

Stacking gel 4.3% (for two gels). mQ-deionized H₂O (2 mL), 0.25 M Tris·Base pH 6.8 with 0.2% of SDS and Coomassie[®] Brilliant Blue R-250 (2.5 mL), aq. solution of acrylamide (30%) and bisacrylamide (0.8%) (0.75 mL), aq. solution of APS (10%, 50 μ L), TMED (10 μ L).

Sample buffer. SDS (5 g), 1M Tris-Base pH 6.8 (15 mL), glycerol (22.5 mL), β -mercatoethanol (12.5 mL), bromphenol blue.

Molecular Marker. Calibration Kit 17-0446-01 purchased from GE Healthcare was used as molecular marker. The protein mixture containing phosphorylase b (97 kDa), albumin (66 kDa), ovalbumin (45 kDa), carbonic anhydrase (30 kDa), trypsin inhibitor (20.1 kDa) and α -lactalbumin (14.4 kDa) was dissolved in mQ-deionized H₂O (160 µL) and sample buffer (40 µL). The solution was heated at 100 °C for 5 min and cooled back down to rt prior to use on the gel.

Electrophoresis buffer. To a solution of glycine (72 g) and Tris-Base (15 g) in mQ-deionized H_2O (975 mL) a solution of SDS (10%, 25 mL) was added.

Staining/fixation bath. MeOH (0.5 L), acetic acid (99%, 0.125 L), mQ-deionized H₂O (0.625

L), Coomassie[®] Brilliant Blue R-250 (2.5 g).

Background destaining bath. MeOH (2.5 L), acetic acid (99%, 0.5 L), mQ-deionized H_2O

(2.5 L).

Catalysis



Figure S1. pH dependent activity profile of G6 dendrimer **G6P1** resulting from a bell-shaped fit assuming bifunctional catalysis (v = c [HA][A]), plotted for pKa(HA) = 7.4.

Table S1. Enantioselectivities (%) observed for aldol 6.

ID	G4P	G5P1	G5P2	G5P3	G5P4	G5P5	G5P6	G5P7	G6P1	G6P2
ee ^a	24	16	18	14	27	15	12	<5	23	19
ee^b	35	24	27	25	n.r. ^c	29	20	9	31	41

Determined after dendrimer-catalyzed aldol reaction in ^{*a*} 0.1 M HEPES buffer pH 7.0/acetone (1/1, v/v) or ^{*b*} DMSO/acetone (1/1, v/v), buffered with *x* eq. of NMM (where *x* = number of formic acid-amine salts for the dendrimer tested) on a chiral Daicel Chiralpak AS[®] column (amylose tris[(S)- α -methylbenzylcarbamate] coated on a silica support, 10 µm, 4.6 × 250 mm, flow rate 2.0 mL·min⁻¹); ^{*c*} no reaction.