# **Electronic Supplementary Information (ESI)**

# Rapid synthesis of nucleotide pyrophosphate linkages in a ball mill

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#### **General information**

All substrates were purchased from Sigma and used as supplied: AMP-morpholidate (A1127); AMP.H<sub>0.9</sub>.Na<sub>1.1</sub> (A1752); ADP.H<sub>1.6</sub>.Na<sub>1.4</sub> (A2754); ATP.Na<sub>2</sub>.H<sub>2</sub> (A6419); TMP.Na<sub>2</sub> (T7004),  $\beta$ -NMN (N3501) 5-ribose monophosphate (83870). Other reagents or solvents were purchased from commercial suppliers (Sigma, Aldrich or Fluka) and used without further purification. H<sub>2</sub>O was purified to 18.2 M $\Omega$  by reverse osmosis (Barnstead NANOpure Diamond water purification system).

All ball mill reactions were performed using a Retsch Mixer Mill MM  $400^1$  and a 15mm stainless steel ball (13.67g) according to the conditions described below.

Stock solutions of triethylammonium bicarbonate (TEAB) in 18.2 M $\Omega$  H<sub>2</sub>O were prepared by bubbling CO<sub>2</sub> through a sintered frit into a mixture of triethylamine and H<sub>2</sub>O at 0°C overnight to give homogenous solutions and subsequently diluted to 1 M. This stock was stored at 4°C until required (up to 2 days) and then diluted as required to give: 100 mM TEAB (aq.), pH 7.8 (Buffer A); or 100 mM TEAB in 65:35 (v/v) MeCN:H<sub>2</sub>O, pH 8.2 (Buffer B).

To the ball-milled reaction mixtures was added H<sub>2</sub>O (1 mL) giving turbid solutions and treated as below: <u>Analytical HPLC</u>

Analysis of crude reaction mixtures was performed following further dilution (10  $\mu$ l in 1 mL H<sub>2</sub>O) and injection of a 10  $\mu$ l aliquot of this diluted solution onto HPLC (Merck Hitachi D7000 system). Elution from the C-18 reversed phase silica column (Phenomenex Clarity 5 $\mu$ m Oligo-RP - 150 x 4.60 mm) was performed using a gradient of: 0-25 min, 0-20% Buffer B; 25-30 min, isocratic elution at 20% B; flow rate, 1.0 mL min<sup>-1</sup>. Chromatograms were recorded using a Merck Hitachi Diode Array Detector L-7455 monitoring at 260 nm. Compositional analysis was based upon molar extinction coefficients described in the literature<sup>2-4</sup>

# Preparative HPLC

The stock solution was dilution following addition of a further aliquot of  $H_2O$  (0.5 mL) and this diluted solution purified by preparative HPLC using the same system as described above with the following changes: the material was injected in two aliquots (0.75 mL); a larger column was utilised (Phenomenex Clarity 5µm Oligo-RP - 250 x 21.2 mm); the gradient was modified: 0-7.5 min, isocratic elution at 0% B; 7.5-67.5 min, 0-20% B; flow rate, 7.0 mL min<sup>-1</sup>; monitoring was performed at 280 nm

<sup>1</sup>H, <sup>13</sup>C or <sup>31</sup>P NMR NMR spectra were recorded on a Bruker III-400 or DRX-500 at 20°C. All spectra were recorded at ambient temperature in D<sub>2</sub>O. Mass spectra were recorded using a VG Quattro II Triple Quadrupole Mass Spectrometer (Electrospray). Mass spectrometry was performed by Analytical Services and Environmental Projects (ASEP) at Queen's University Belfast.

#### Experimental procedures and analytical data

# $P^1$ , $P^2$ -di(adenosin-5'-yl)diphosphate bis(triethylammonium) salt - AppA (**3a**).

A stainless steel jar (25 mL) was charged with a mixture of adenosine-5'phosphoromorpholidate 4-morpholine-N,N'-dicyclohexylcarboxamidine



salt (1) (48 mg, 0.068 mmol), adenosine-5'-monophosphate (**2a**) (25 mg, 0.067 mmol, 1 eq.), MgCl<sub>2</sub>(H<sub>2</sub>O)<sub>6</sub> (21 mg, 0.10 mmol, 1.5 eq.), tetrazole (10 mg, 0.14 mmol, 2.1 eq.), H<sub>2</sub>O (7.5 μL, 0.42 mmol, 6 eq.) and a 15.0 mm stainless steel ball. The vessel was vibrated at 30 Hz for 90 minutes and allowed to cool to room temperature. The reaction mixture was suspended in H<sub>2</sub>O, analysed and then purified by C18 RP-HPLC – see general information. Appropriate fractions were lyophilised to yield pure **3a** (45 mg, 0.051 mmol, 75%). <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O)  $\delta$  = 8.06 (2H, s), 7.88 (2H, s), 5.82 (2H, d, *J* = 5.0 Hz), 4.44 (2H, t, *J* = 5.0 Hz), 4.32 (2H, t, *J* = 4.5 Hz), 4.23 (4H, d, *J* = 11.2 Hz), 4.13 (2H, d, *J* = 11.6 Hz), 2.85 (6H, q, *J* = 7.3 Hz), 1.06 (9H, t, *J* = 7.3 Hz); <sup>13</sup>C NMR (101 MHz, D<sub>2</sub>O)  $\delta$  = 154.66, 152.27, 148.02, 139.09, 117.68, 87.08, 83.30 (d, <sup>2</sup>*J*<sub>*C,P*</sub> = 8.6 Hz), 74.67, 69.86, 65.09, 46.65, 8.20; <sup>31</sup>P (162 MHz, D<sub>2</sub>O)  $\delta$  = -11.30; ES+ MS (C<sub>20</sub>H<sub>27</sub>N<sub>10</sub>O<sub>13</sub>P<sub>2</sub>) (M + H<sup>+</sup>), calc. 677.1234, found 677.1215.



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 $Ap_2A$ 





P<sup>1</sup>,P<sup>3</sup>-di(adenosin-5'-yl)triphosphate tris(triethylammonium) salt –

# <u>ApppA (3b)</u>

A stainless steel jar (25 mL) was charged with a mixture of adenosine-5'-phosphoromorpholidate 4-morpholine-N,N'-dicyclo-hexylcarboxamidine salt (1) (50 mg, 0.070 mmol), adenosine-5'-



diphosphate (**2b**) (30 mg, 0.065 mmol, 0.9 eq.), MgCl<sub>2</sub>(H<sub>2</sub>O)<sub>6</sub> (22 mg, 0.11 mmol, 1.5 eq.), tetrazole (10 mg, 0.14 mmol, 2 eq.), H<sub>2</sub>O (7.5  $\mu$ L, 0.42 mmol, 6 eq.) and a 15.0 mm stainless steel ball. The vessel was vibrated at 30 Hz for 90 minutes and allowed to cool to room temperature. The reaction mixture was suspended in H<sub>2</sub>O, analysed and then purified by C18 RP-HPLC – see general information. Appropriate fractions were lyophilised to yield **3b**·(**TEAB**)<sub>1.5</sub> (39 mg, 0.032 mmol, 49%). <sup>1</sup>H NMR(400 MHz, D<sub>2</sub>O)  $\delta$  = 8.17 (2H, s), 7.91 (2H, s), 5.85 (2H, d, *J* = 3.8 Hz), 4.47 (2H, t, *J* = 4.6 Hz), 4.37 (2H, t, *J* = 4.6 Hz), 4.30 – 4.10 (6H, m), 2.86 (27H, q, *J* = 7.3 Hz), 1.08 (40.3H, t, *J* = 7.3 Hz); <sup>13</sup>C NMR (101 MHz, D<sub>2</sub>O)  $\delta$  = 154.84, 152.55, 148.11, 139.04, 117.75, 87.15, 83.04 (d, <sup>2</sup>*J*<sub>*C,P*</sub> = 9.4), 74.78, 69.57, 64.58 (d, <sup>3</sup>*J*<sub>*C,P*</sub> = 5.4 Hz), 46.11, 8.78; <sup>31</sup>P NMR (162 MHz, D<sub>2</sub>O)  $\delta$  = -11.78 (2 P<sub>a</sub>, d, *J* = 19.7 Hz), -23.25 (1 P<sub>β</sub>, t, *J* = 19.7); ES+ MS, (C<sub>20</sub>H<sub>28</sub>N<sub>10</sub>O<sub>16</sub>P<sub>2</sub>) (M + H<sup>+</sup>), calc. 757.0898, found 757.0906.





Ap<sub>3</sub>A



 $P^{1}$ ,  $P^{4}$ -di(adenosin-5'-yl)tetraphosphate tetrakis(triethyl-

ammonium) salt - AppppA (3c)

A stainless steel jar (25 mL) was charged with a mixture of adenosine-5'-phosphoromorpholidate 4-morpholine-N,N'-



dicyclohexylcarboxamidine salt (1) (51 mg, 0.072 mmol),

adenosine-5'-triphosphate (2c) (40 mg, 0.073 mmol, 1 eq.), MgCl<sub>2</sub>(H<sub>2</sub>O)<sub>6</sub> (22 mg, 0.11 mmol, 1.5 eq.), tetrazole (10 mg, 0.14 mmol, 2 eq.), H<sub>2</sub>O (7.5 µL, 0.42 mmol, 6 eq.) and a 15.0 mm stainless steel ball. The vessel was vibrated at 30 Hz for 90 minutes and allowed to cool to room temperature. The reaction mixture was suspended in H<sub>2</sub>O, analysed and then purified by C18 RP-HPLC – see general information. Appropriate fractions were lyophilised to yield pure **3c**·(**TEAB**)<sub>0.6</sub> (50 mg, 0.037 mmol, 52 %). <sup>1</sup>H NMR(400 MHz, D<sub>2</sub>O)  $\delta = 8.34$  (2H, s), 8.06 (2 H, s), 5.96 (2H, d, J = 5.9), 4.68 (2H, t, J = 4.5), 4.51 (2H, t, J = 4.5), 4.34 – 4.27 (2H, m), 4.27 - 4.20 (2H, m), 4.20 - 4.11 (2H, m), 2.96 (27.6H, q, J = 7.3 Hz), 1.13 (41.3H, t, J = 7.3 Hz);<sup>13</sup>C NMR (101 MHz, D<sub>2</sub>O)  $\delta$  = 155.10, 152.63, 148.69, 139.58, 118.00, 86.55, 83.89 (d, <sup>2</sup>J<sub>CP</sub> = 9.3), 74.36, 70.31, 65.14 (d,  ${}^{3}J_{C,P} = 5.5$ ), 46.29, 8.51;  ${}^{31}$ P NMR (162 MHz, D<sub>2</sub>O)  $\delta = -11.51$  (2 P<sub>a</sub>, AA'XX'), -23.44 (2 P<sub>b</sub>, AA'XX'); ES+ MS,  $(C_{20}H_{29}N_{10}O_{19}P_4)$  (M + H<sup>+</sup>)calc. 837.0561, found 837.0557.







 $P^{1}$ -(adenosin-5'-yl)- $P^{2}$ -(thymidin-5'''-yl) bis(triethylammonium) salt - AppdT

# <u>(3d)</u>

A stainless steel jar (25 mL) was charged with a mixture of adenosine-5'phosphoromorpholidate 4-morpholine-N,N'- dicyclohexylcarboxamidine salt (1) (100 mg, 0.14 mmol), thymidine-5'-monophosphate (2d) (52 mg, 0.14



mmol, 1 eq.), MgCl<sub>2</sub>(H<sub>2</sub>O)<sub>6</sub> (43 mg, 0.21 mmol, 1.5 eq.), tetrazole (20 mg, 0.28 mmol, 2 eq.), H<sub>2</sub>O (15 μL, 0.83 mmol, 6 eq.) and a 15.0 mm stainless steel ball. The vessel was vibrated at 30 Hz for 90 minutes and allowed to cool to room temperature. The reaction mixture was suspended in H<sub>2</sub>O, analysed and then purified by C18 RP-HPLC – see general information. Appropriate fractions were lyophilised to yield pure **3d** (86 mg, 0.10 mmol, 71 %). <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O)  $\delta$  = 8.32 (1H, s), 8.03 (1H, s), 7.35 (1H, s), 6.04 (1H, t, *J* = 6.9), 5.93 (1H, d, *J* = 5.8), 4.63 (1H, t, *J* = 5.5), 4.45 – 4.32 (2H, m), 4.30 – 3.88 (6H, m), 2.85 (12H, q, *J* = 7.3Hz), 2.21 – 2.00 (2H, m), 1.64 (3H, s), 1.06 (18H, t, *J* = 7.3 Hz); <sup>13</sup>C NMR (101 MHz, D<sub>2</sub>O)  $\delta$  = 166.02, 155.30, 152.65, 151.25,148.84, 139.66, 136.95, 118.31, 111.22, 86.69, 85.22 (d, <sup>2</sup>*J*<sub>C,P</sub> = 8.9), 84.65, 83.67 (d, <sup>2</sup>*J*<sub>C,P</sub> = 9.1), 74.14, 70.84, 70.24, 65.38 (d, <sup>3</sup>*J*<sub>C,P</sub> = 4.5), 65.26 (d, <sup>3</sup>*J*<sub>C,P</sub> = 4.6), 38.65, 11.59; <sup>31</sup>P NMR (162 MHz, D<sub>2</sub>O)  $\delta$  = -11.45; ES+ MS (C<sub>32</sub>H<sub>57</sub>N<sub>9</sub>O<sub>14</sub>P<sub>2</sub>) (M + 2Et<sub>3</sub>NH<sup>+</sup>), calc. 855.3657, found 855.3649, (C<sub>26</sub>H<sub>43</sub>N<sub>8</sub>O<sub>14</sub>P<sub>2</sub>) (M + Et<sub>3</sub>NH<sup>+</sup>), calc. 753.2374, found 753.2375.







<u> $\beta$ -Nicotinamide adenine dinucleotide triethylammonium salt -  $\beta$ -NAD<sup>+</sup></u>

<u>(3e)</u>

A stainless steel jar (25 mL) was charged with a mixture of adenosine-5'-phosphoromorpholidate 4-morpholine-*N*,*N*'-



dicyclohexylcarboxamidine salt (1) (99 mg, 0.14 mmol), β-nicotinamide-5'-monophosphate (**2e**) (47 mg, 0.14 mmol, 1 eq.), MgCl<sub>2</sub>(H<sub>2</sub>O)<sub>6</sub> (43 mg, 0.21 mmol, 1.5 eq.), tetrazole (20 mg, 0.29 mmol, 2 eq.), H<sub>2</sub>O (15  $\mu$ L, 0.83 mmol, 6 eq.) and a 15.0 mm stainless steel ball. The vessel was vibrated at 30 Hz for 90 minutes and allowed to cool to room temperature. The reaction mixture was suspended in H<sub>2</sub>O, analysed and then purified by C18 RP-HPLC – see general information. Appropriate fractions were lyophilised to yield **3e**·(**TEAB**)<sub>0.2</sub> (65 mg, 0.081 mmol, 58 %). <sup>1</sup>H NMR (400 MHz, D<sub>2</sub>O)  $\delta$  = 9.22 (1H, s), 9.05 (1H, d, *J* = 6.3), 8.71 (1 H, d, *J* = 8.2), 8.28 (1 H, s), 8.17 – 8.00 (1H, m), 7.96 (1H, d, *J* = 5.7), 5.97 (1H, d, *J* = 5.4), 5.89 (1H, d, *J* = 6.0), 4.67 – 4.62 (2H, m), 4.46 – 4.42 (1H, m), 4.42 – 4.36 (2H, m), 4.35 – 4.31 (1H, m), 4.30 – 4.22 (2H, m), 4.19 – 4.05 (3H, m), 3.04 (7.3H, q, *J* = 7.3Hz), 1.14 (11H, t, *J* = 7.3Hz); <sup>13</sup>C NMR (101 MHz, D<sub>2</sub>O)  $\delta$  = 165.10, 155.16, 152.66, 148.77, 145.69, 142.33, 139.83, 139.62, 133.57, 128.56, 118.17, 99.98, 86.99 (d, <sup>2</sup>*J*<sub>*C,P*</sub> = 8.7), 86.60, 83.79 (d, <sup>2</sup>*J*<sub>*C,P*</sub> = 8.9), 77.57, 73.92, 70.59, 70.34, 65.35 (d, <sup>2</sup>*J*<sub>*C,P*</sub> = 5.2), 64.86 (d, <sup>3</sup>*J*<sub>*C,P*</sub> = 5.1); <sup>31</sup>P NMR (162 MHz, D<sub>2</sub>O)  $\delta$  = -11.56, -11.69 (2 P, ABq, *J* = 20.8). ES+ MS (C<sub>21</sub>H<sub>28</sub>N<sub>7</sub>O<sub>14</sub>P<sub>2</sub>), (M<sup>+</sup>), calc. 664.1170, found 664.1141.







 $P^{1}$ -(adenosin-5'-yl)- $P^{2}$ -(ribos-5"-yl) bis(triethylammonium) salt - ADPR

### <u>(3f)</u>

A stainless steel jar (25 mL) was charged with a mixture of adenosine-5'phosphoromorpholidate 4-morpholine-*N*,*N*'-dicyclohexylcarboxamidine salt



(1) (52 mg, 0.073 mmol), ribose-5-monophosphate (**2f**) (35 mg, 0.074 mmol, 1 eq.), MgCl<sub>2</sub>(H<sub>2</sub>O)<sub>6</sub> (23 mg, 1.1 mmol, 1.5 eq.), tetrazole (10 mg, 0.14 mmol, 2 eq.), H<sub>2</sub>O (7.5 µL, 0.42 mmol, 6 eq.) and a 15.0 mm stainless steel ball. The vessel was vibrated at 30 Hz for 90 minutes and allowed to cool to room temperature. The reaction mixture was suspended in H<sub>2</sub>O, analysed and then purified by C18 RP-HPLC – see general information. Appropriate fractions were lyophilised to yield pure **6c** (24 mg, 0.030 mmol, 43 %). <sup>1</sup>H NMR (500 MHz, D<sub>2</sub>O)  $\delta = 8.40$  (1 H, s), 8.12 (1 H, s), 6.02 (1H, dd, J = 5.9, 1.7), 5.20 (0.37H, d, J = 4.1), 5.09 (0.63H, d, J = 2.2), 4.67 – 4.63 (1H, m), 4.49 – 4.36 (1 H, m), 4.27 (1 H, brs), 4.21 – 4.15 (0.37 H, m), 4.11 (2 H, s), 4.07 – 4.03 (0.63 H, m), 4.03 – 3.81 (4 H, m); <sup>13</sup>C NMR (101 MHz, D<sub>2</sub>O)  $\delta = 155.51$ , 152.27, 139.82, 101.12, 96.33, 86.76, 83.93 (d, <sup>2</sup> $J_{C,P} = 9.0$ ), 81.07 (d, <sup>2</sup> $J_{C,P} = 8.9$ ) 75.12, 74.24, 70.66, 70.40, 69.98, 66.29 (d, <sup>3</sup> $J_{C,P} = 4.7$ ), 65.20; <sup>31</sup>P NMR (162 MHz, D<sub>2</sub>O)  $\delta = -11.34$ ; ES+ MS (C<sub>15</sub>H<sub>24</sub>N<sub>5</sub>O<sub>14</sub>P<sub>2</sub>) (M + H<sup>+</sup>), calc. 560.0795, found 560.0796, (C<sub>21</sub>H<sub>39</sub>N<sub>6</sub>O<sub>14</sub>P<sub>2</sub>) (M + TEA<sup>+</sup>), calc. 661.2000, found 661.1995.





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