

General experimental information: Anhydrous grade solvents were purchased from Sigma-Aldrich (St Louis, MO) and used without further drying. Reagents purchased from commercial sources were used as supplied. Deblocking, oxidizing and capping mixtures, 2'-OMe phosphoramidites and fluorescein phosphoramidite were purchased from Glen Research (Sterling, VA). Activator 42TM (5-(bis-3,5-trifluoromethylphenyl)-1H-tetrazole) was purchased from SAFC Proligo (St Louis, MO). RNA for thermal melting and biological experiments was purchased from IDT (Coralville, IA). Flash column chromatography was performed using 230 x 400 mesh silica gel (Sorbent Technologies, Atlanta, GA) and thin layer chromatography (tlc) was performed on aluminum-backed silica 60 F254 plates (EMD Biosciences, Gibbstown, NJ). NMR spectra were acquired on a BioSpin 300 MHz spectrometer (Bruker, Billerica, MA). Mass spectra were obtained using a Pulsar QStar Time of Flight spectrometer (AB Sciex, Foster City, CA), equipped with a syringe pump. Phosphinoamidites were analyzed in ESI+ mode in methanol solution at 5.2 kV. Oligoribonucleotides were analyzed in ESI- mode in 1:1 acetonitrile:water at 3.3 kV. Analytical RP-HPLC was performed on an Agilent 1100 series HPLC on a Hypersil ODS column, 4.0 x 250 mm, 5 µm particle size, (Agilent Technologies, Santa Clara, CA) with 0.05M triethylammonium bicarbonate (mobile phase A) and acetonitrile (mobile phase B) at a flow rate of 1.5 mL/min and the following gradient: 0 - 30 minutes, 0-50 % B, 30 – 35 minutes, 50 – 100 % B, 35 – 39 minutes, 100% B. Preparative RP-HPLC was performed on an Agilent 1100 series HPLC on a Zorbax SB-C18 column, 9.4 x 250 mm, 5 µm particle size, (Agilent Technologies, Santa Clara, CA) with the same mobile phases and flow rate and the following gradient: 0 - 50 minutes, 0-30 % B, 50 – 60 minutes, 30 – 50 % B, 60 – 65 minutes, 50 – 100% B. Duplex melting experiments were performed on a Cary 100 Bio UV-visible spectrophotometer (Varian, Santa Clara, CA). The solutions were prepared by dissolving PACE/thioPACE 2'-OMe ORN and complementary RNA in a solution of 0.1 M NaCl/0.01 M Na₂HPO₄ to a concentration of 1.0 µM of duplex. The duplex solution was placed in a 1 cm quartz cuvette and annealed by heating to 95 °C for 3 minutes followed by cooling to room temperature. Absorbance was then measured at 260 nm over a range of 25 – 95 °C at a heating rate of 1 °C per minute. Melting temperature (T_m) was calculated using the derivative method in the Cary UV Thermal software application.

For biological experiments, unless otherwise specified the fluorescence ratios or cell numbers obtained were normalised to equivalent negative control cells treated with transfection media in the absence of oligonucleotides. The results are presented as average values with standard deviations. Data processing was carried out using Microsoft Excel and GraphPad Prism (version 5.0d).

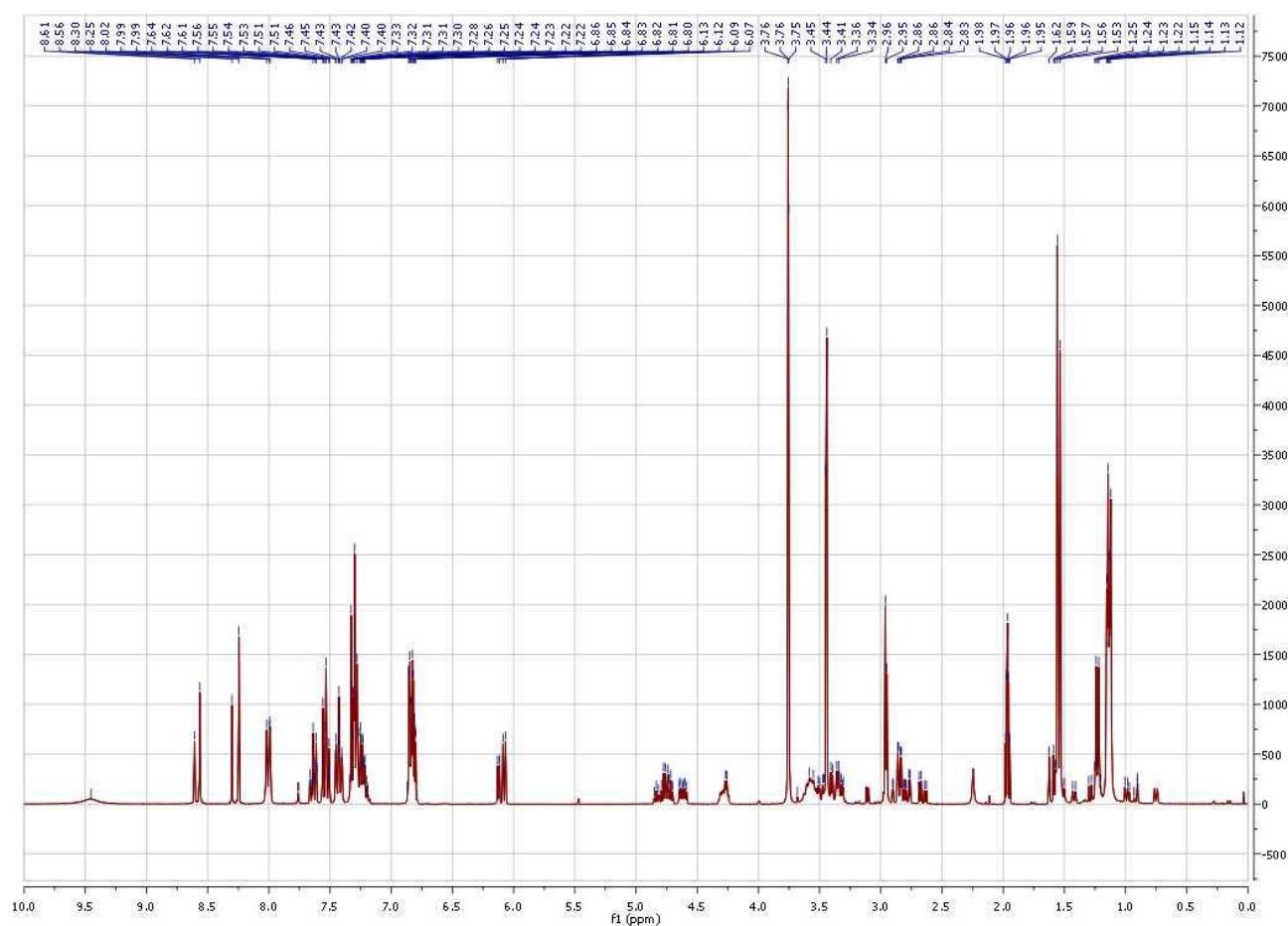


Figure S1 ^1H NMR (300 MHz, CDCl_3) of **2a**

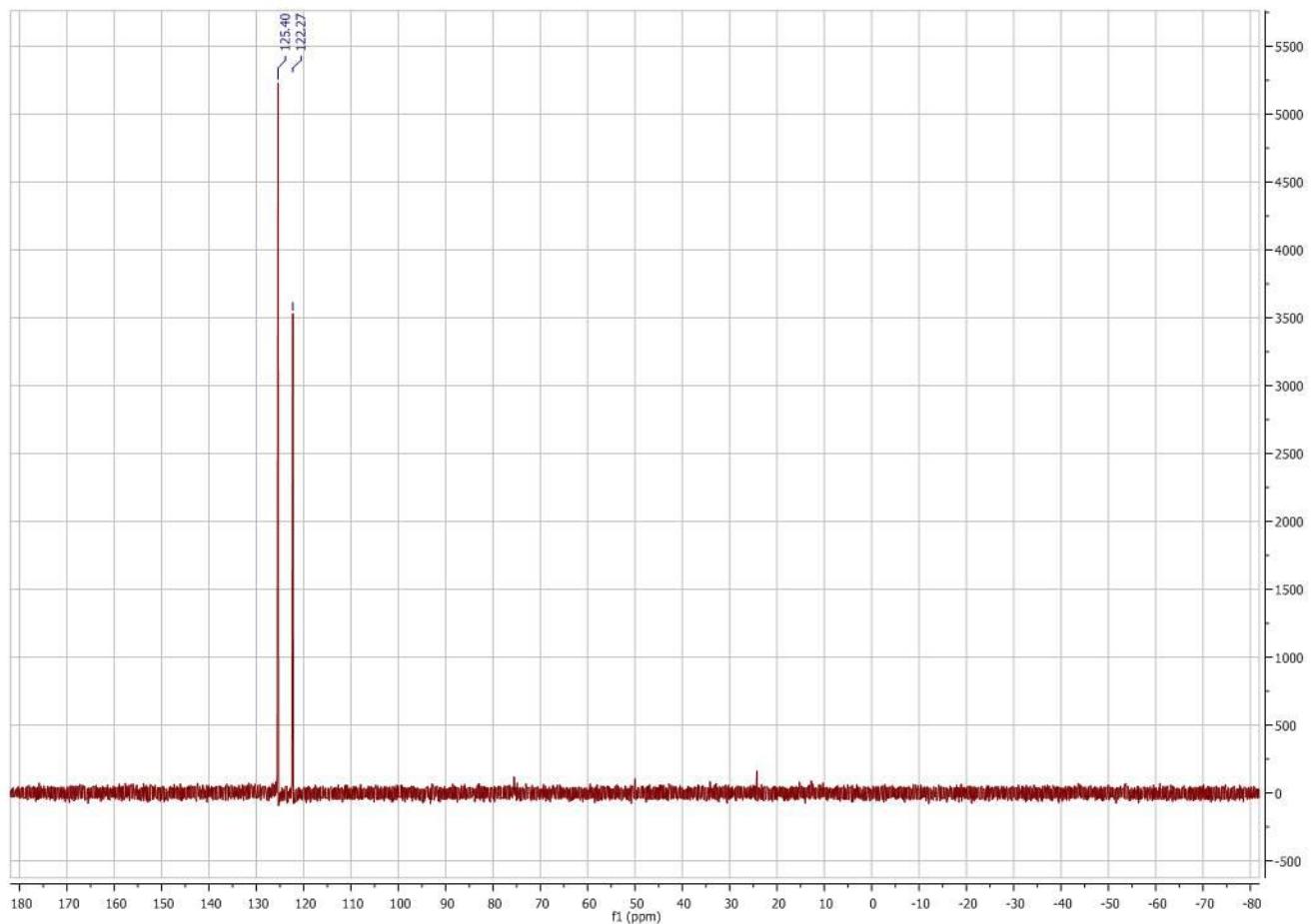


Figure S2 ^{31}P NMR (121 MHz, CDCl_3) of **2a**

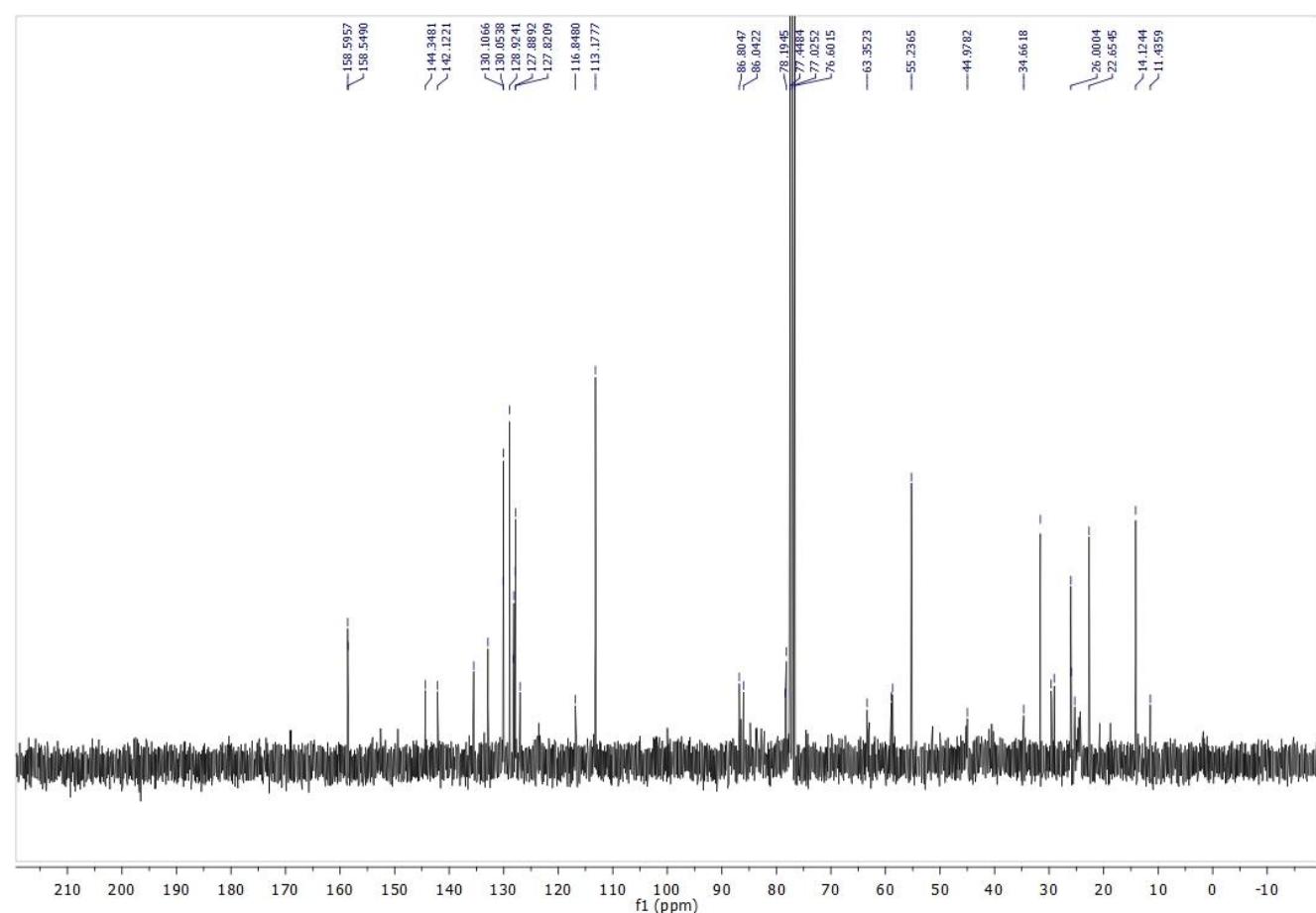


Figure S3 ^{13}C NMR (100 MHz, CDCl_3) of **2a**

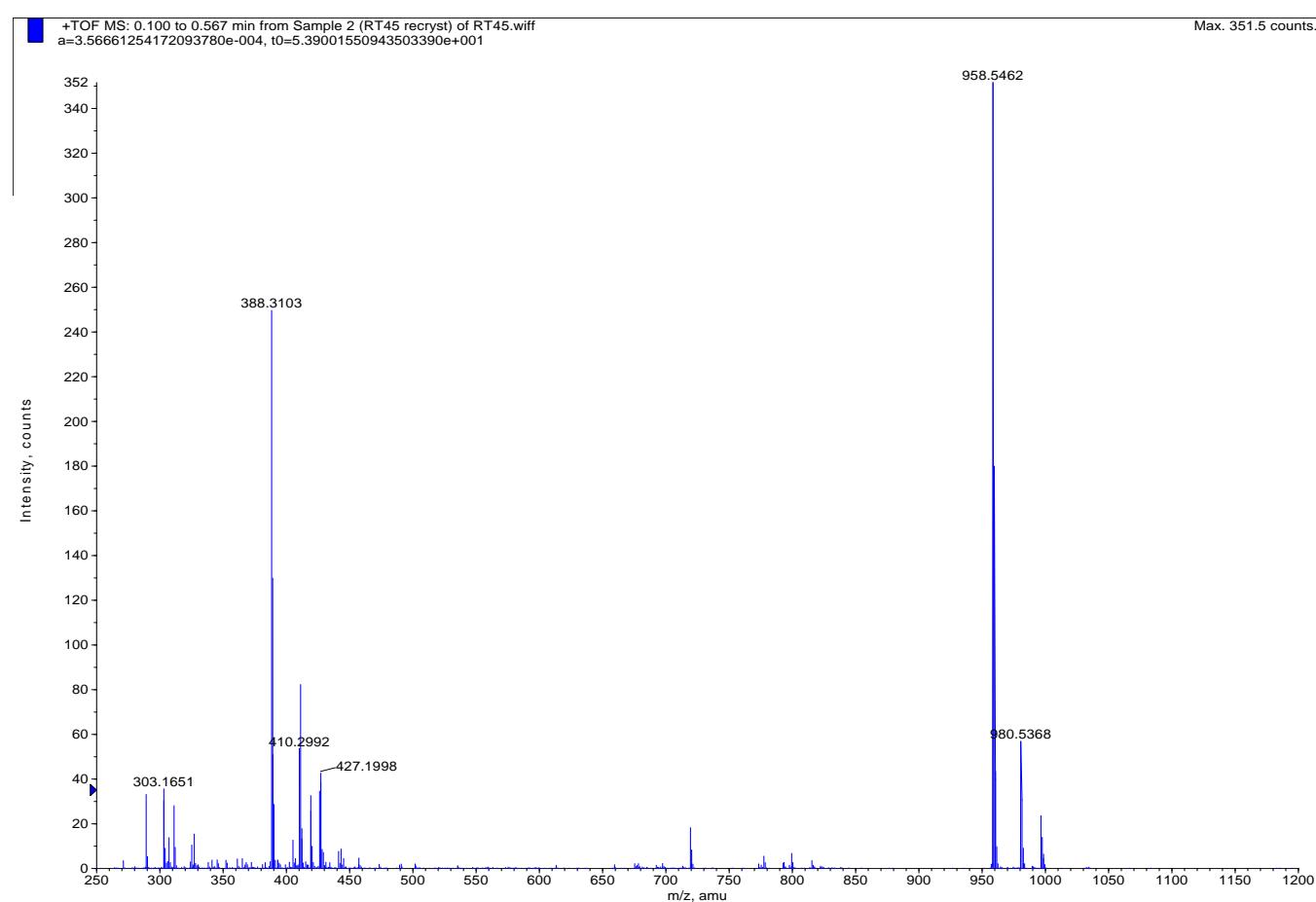


Figure S4 ESI⁺ mass spectrum of **2a**

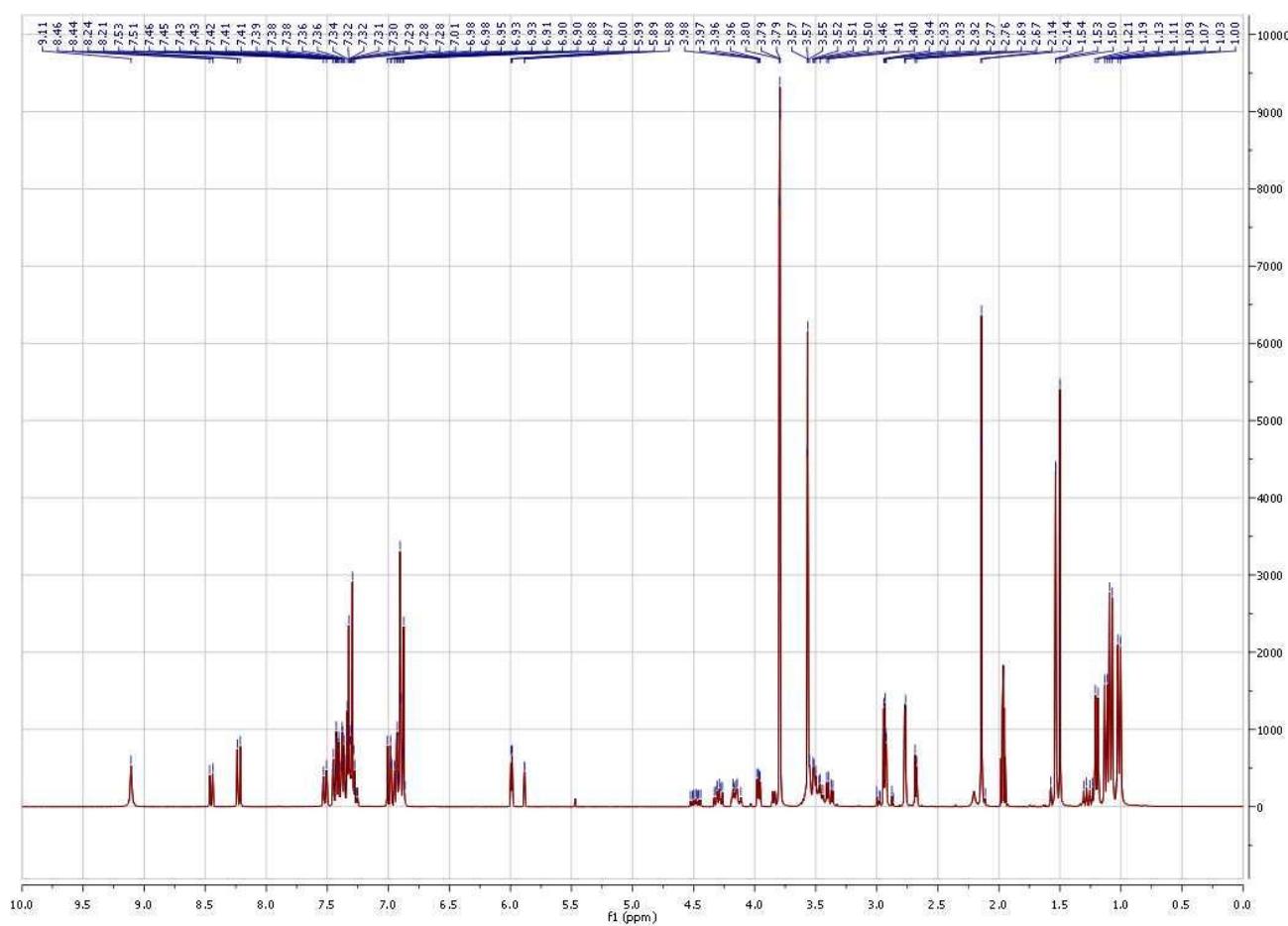


Figure S5 ^1H NMR (300 MHz, CDCl_3) of **2b**

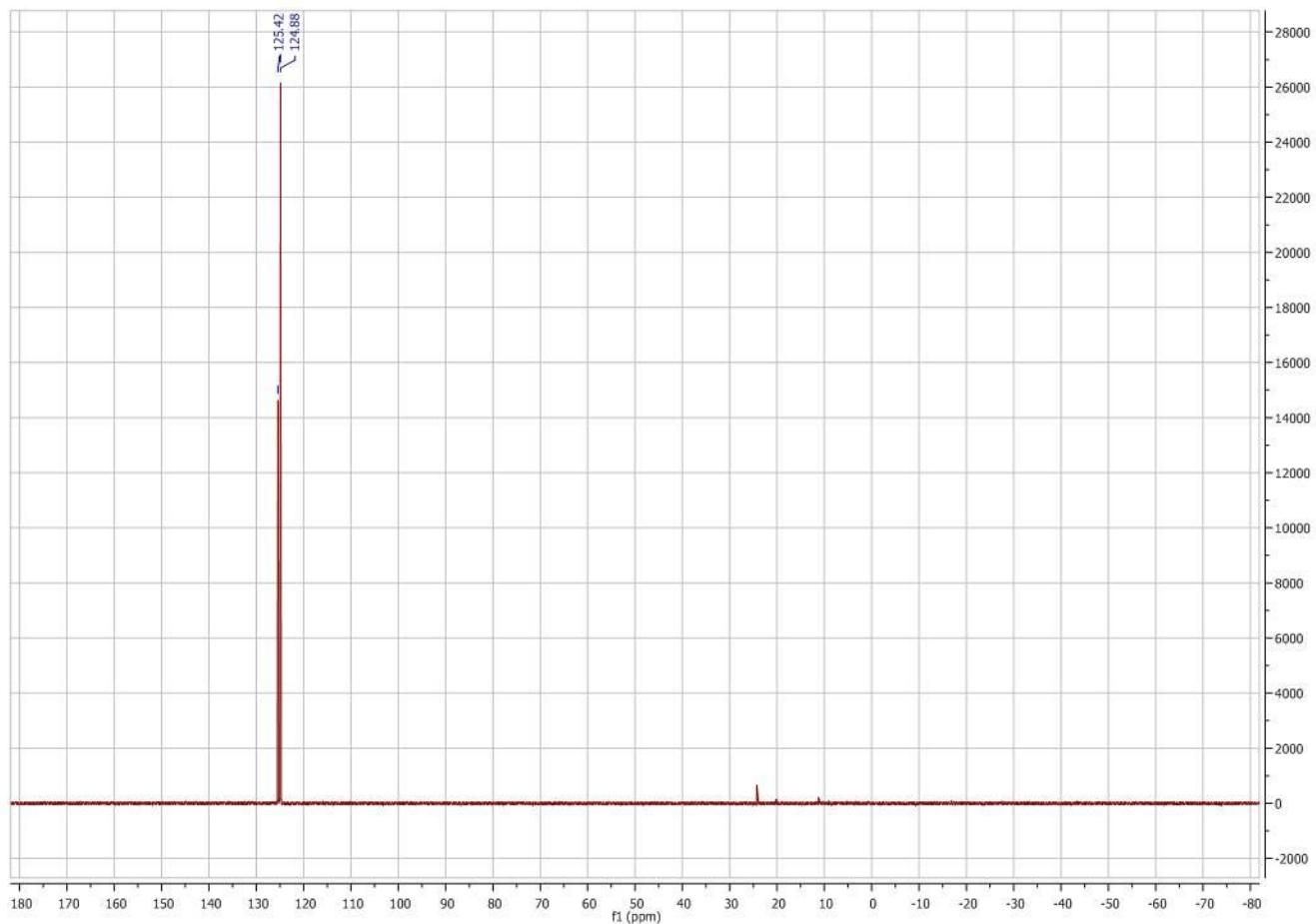


Figure S6 ^{31}P NMR (121 MHz, CDCl_3) of **2b**

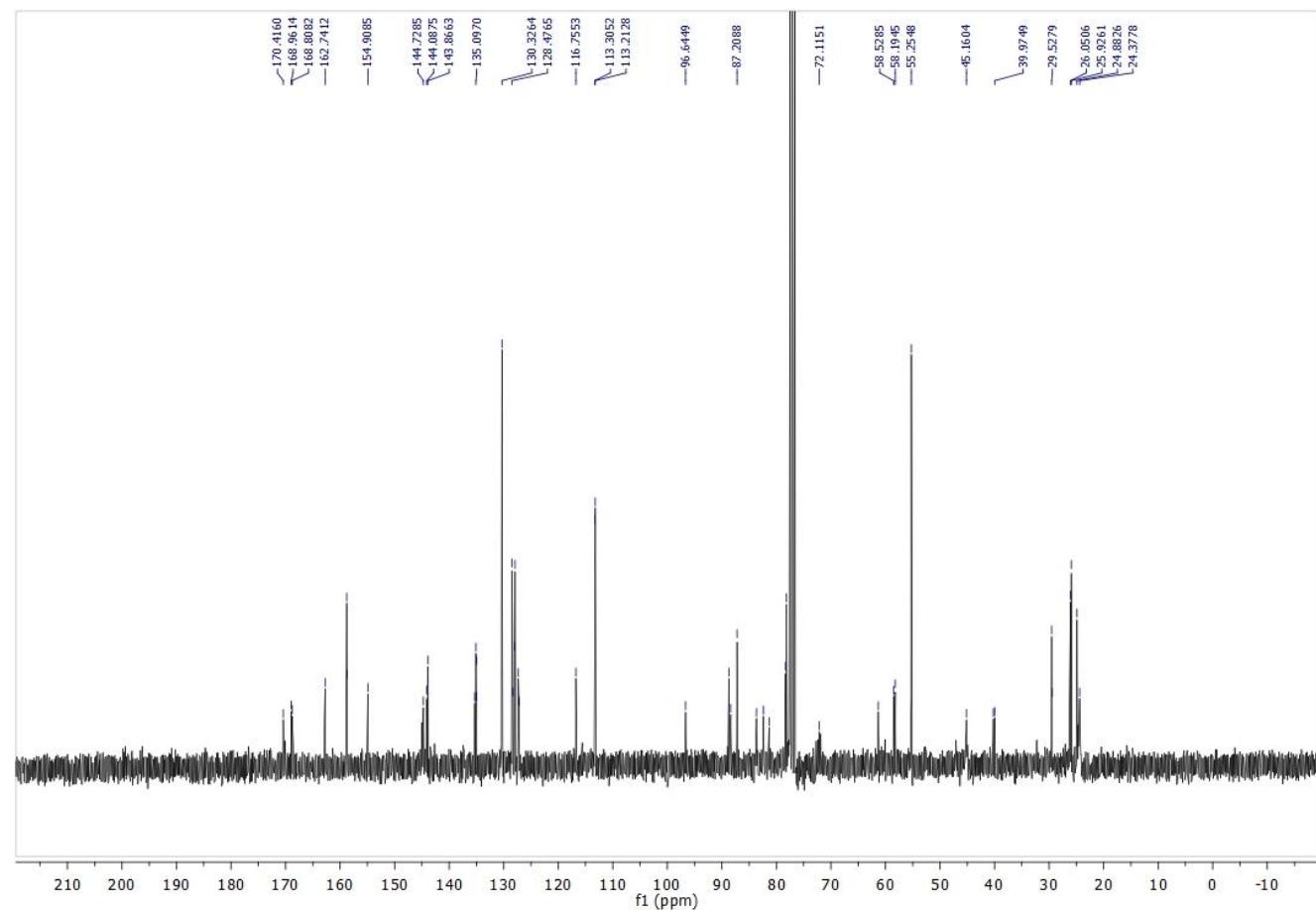


Figure S7 ^{13}C NMR (100 MHz, CDCl_3) of **2b**

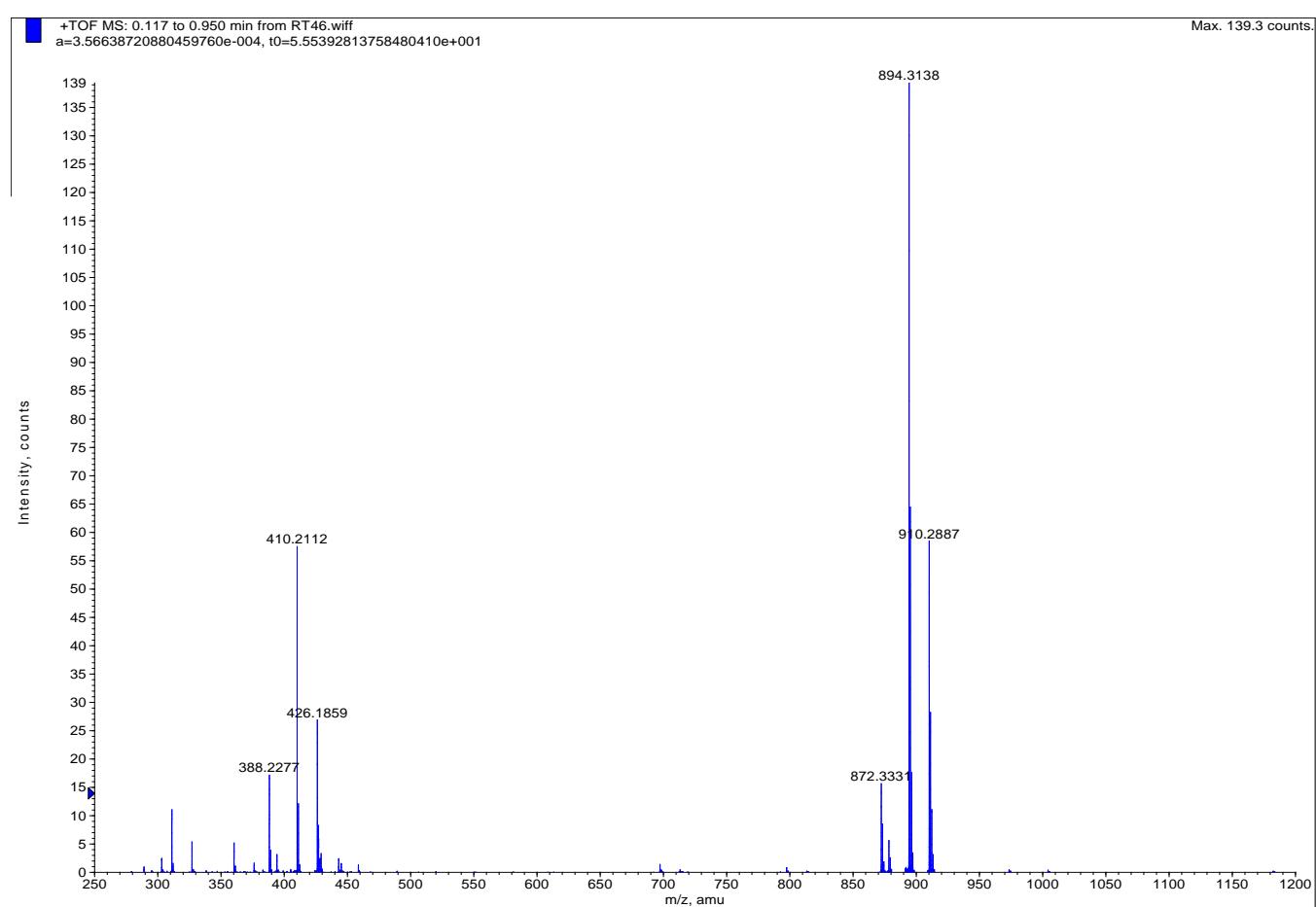


Figure S8 ESI⁺ mass spectrum of **2b**

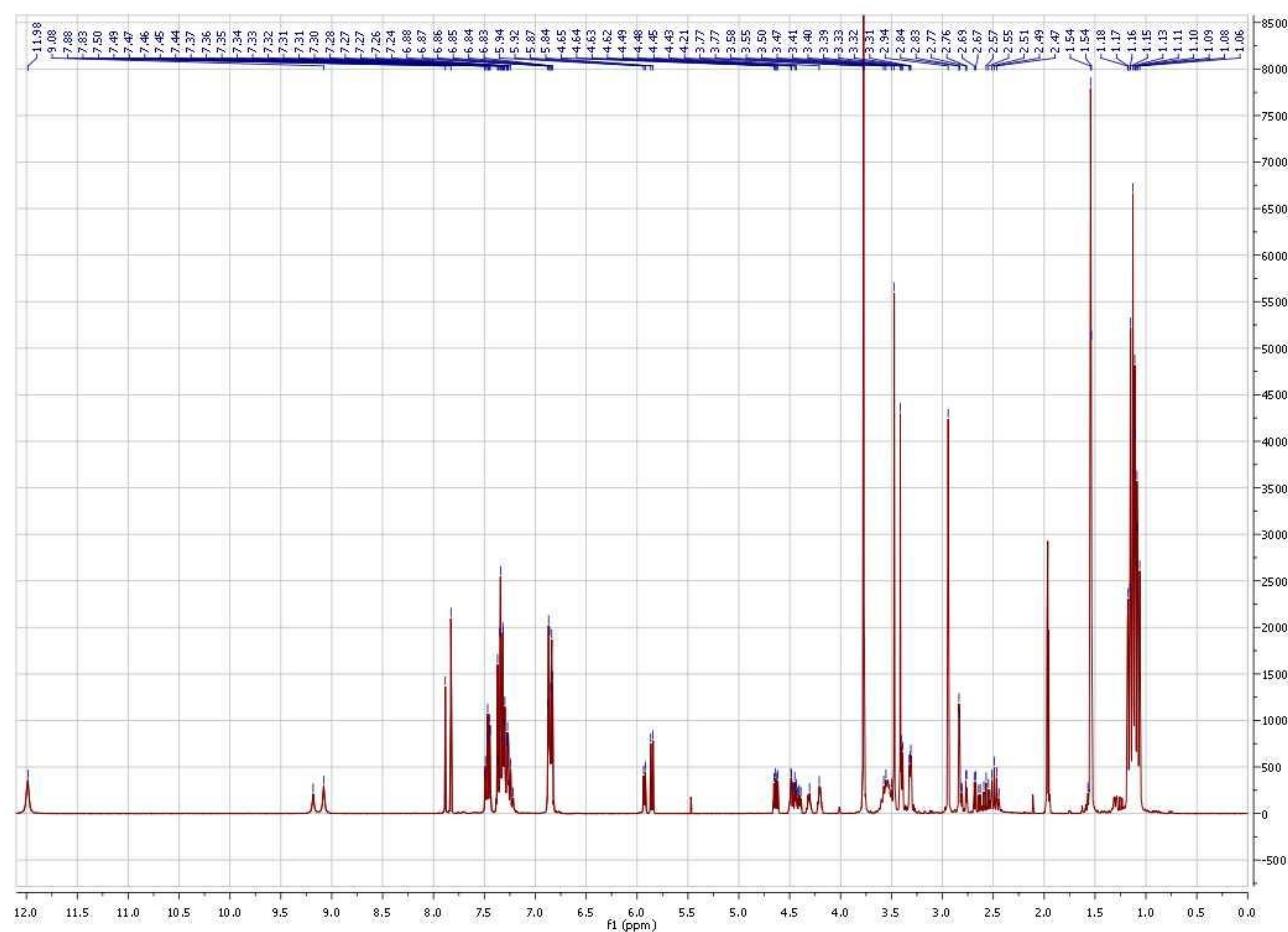


Figure S9 ^1H NMR (300 MHz, CDCl_3) of **2c**

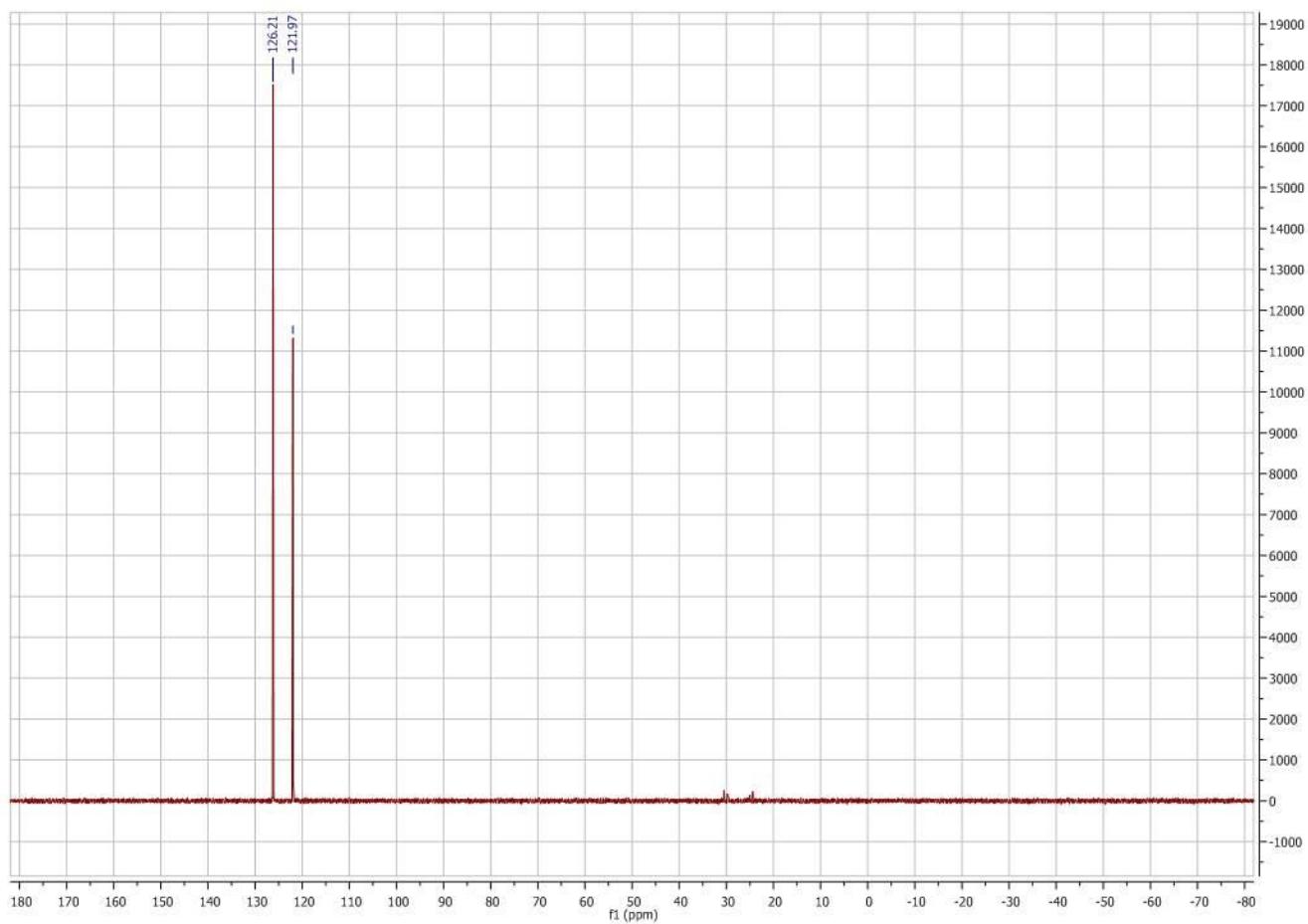


Figure S10 ^{31}P NMR (121 MHz, CDCl_3) of **2c**

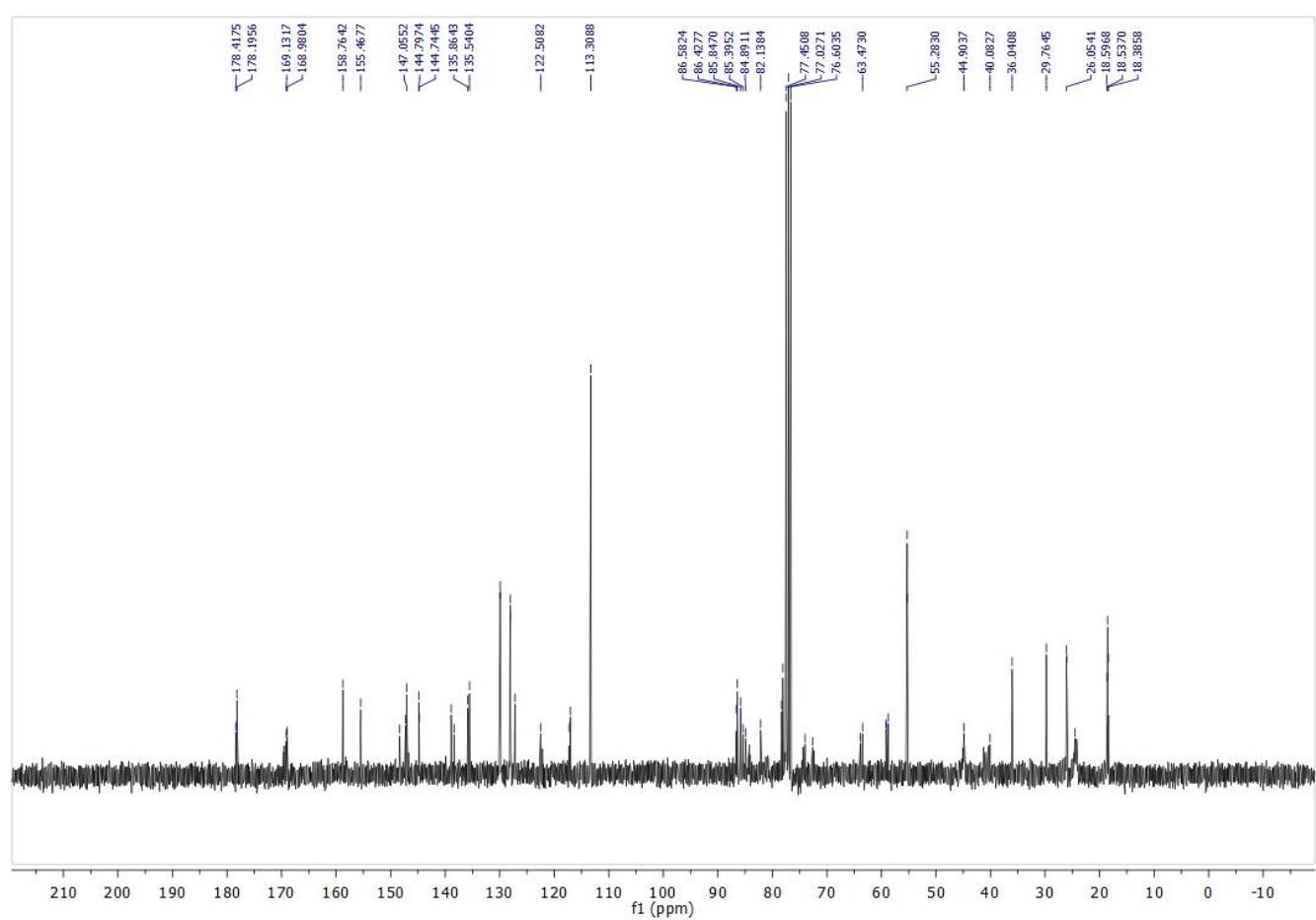


Figure S11 ^{13}C NMR (100 MHz, CDCl_3) of **2c**

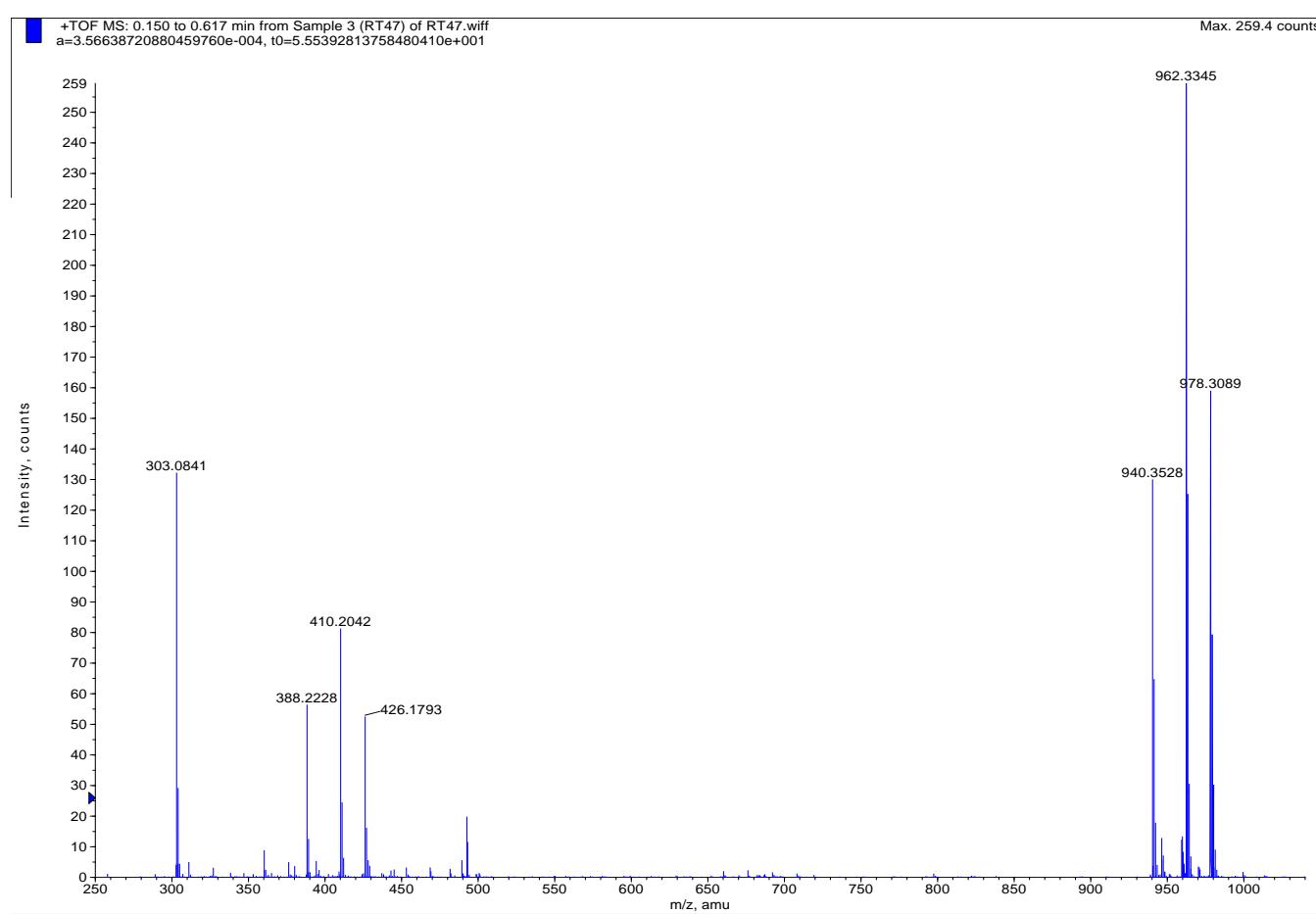


Figure S12 ESI⁺ mass spectrum of **2c**

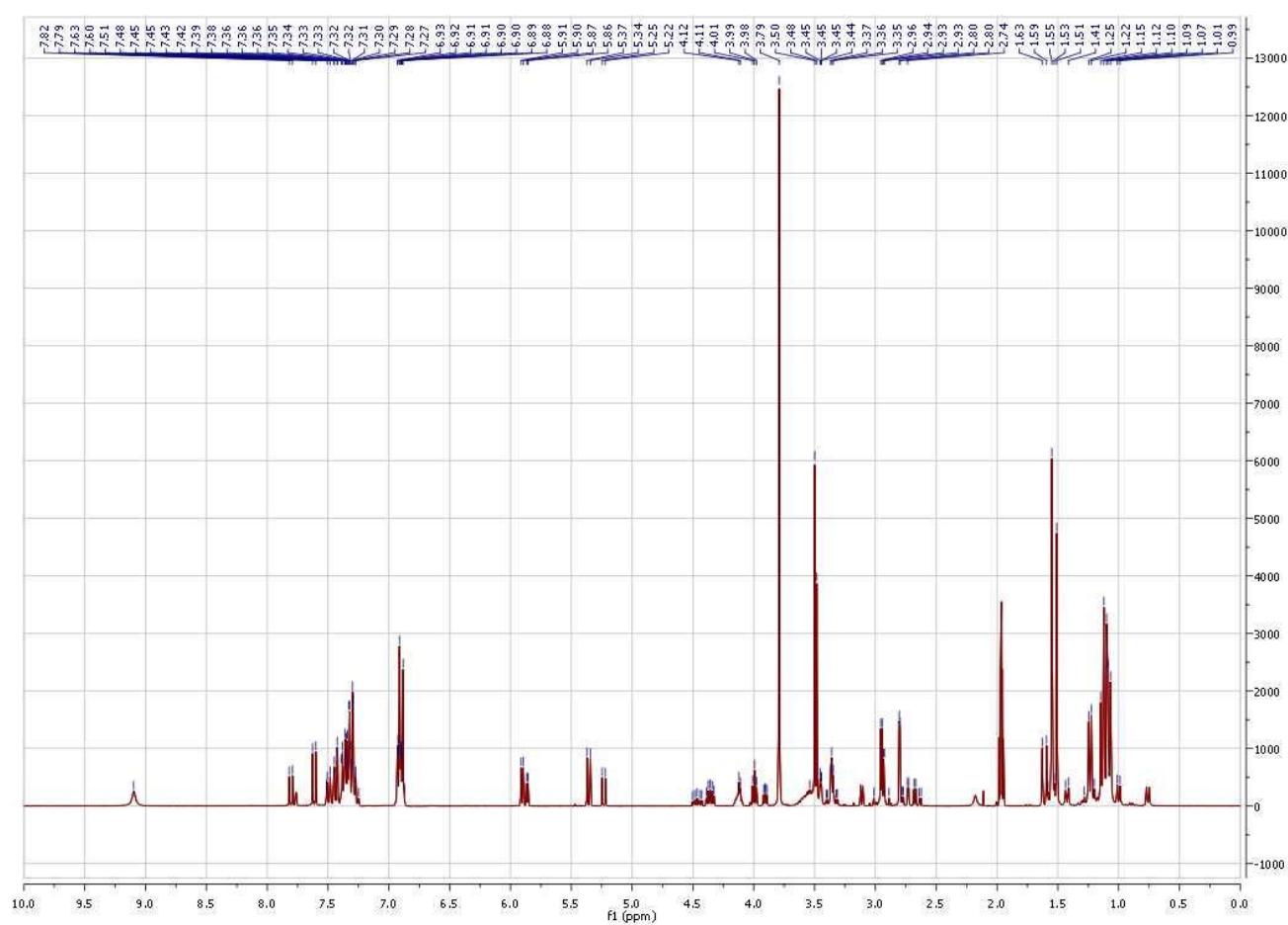


Figure S13 ^1H NMR (300 MHz, CDCl_3) of **2c**

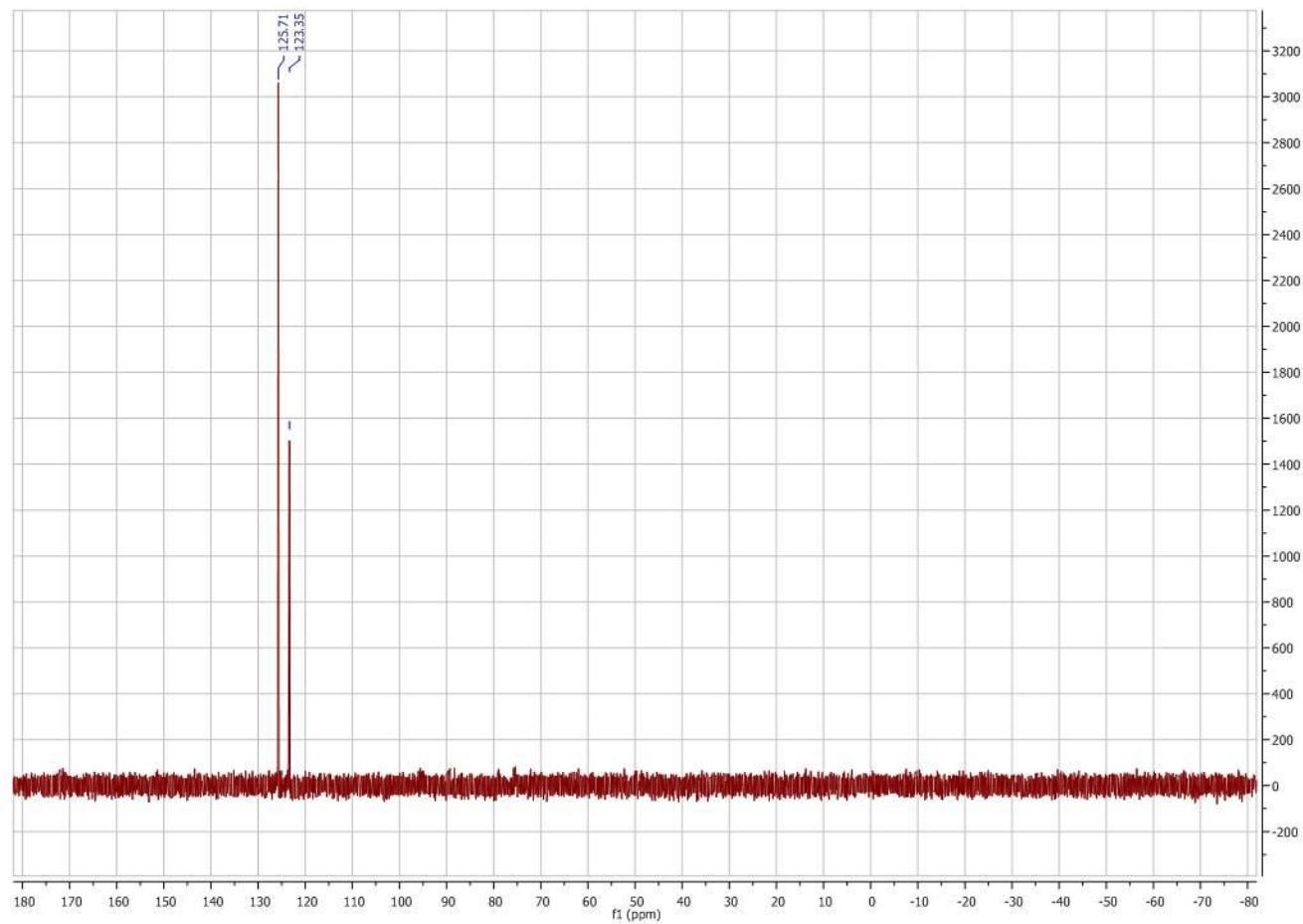


Figure S14 ^{31}P NMR (121 MHz, CDCl_3) of **2d**

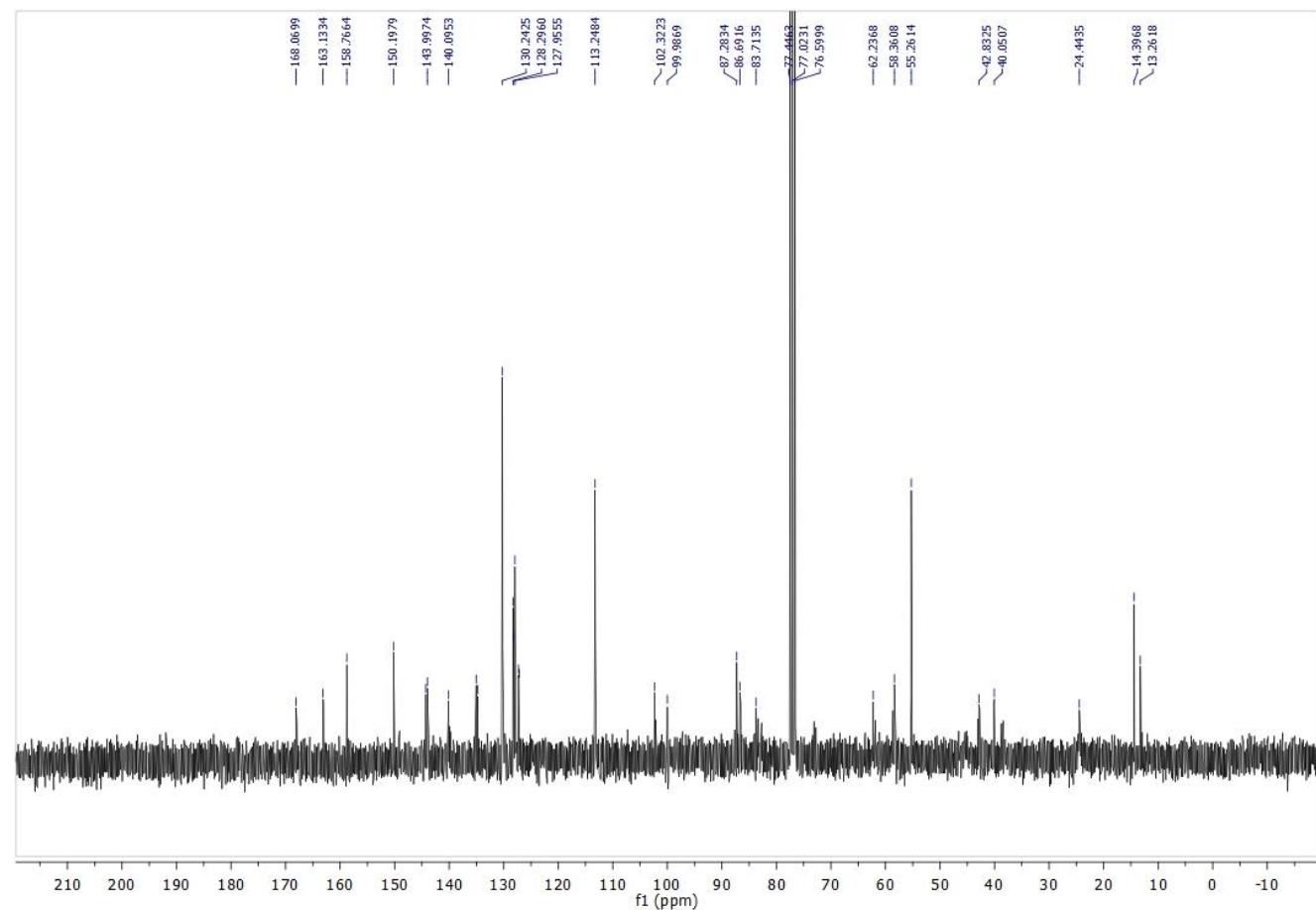


Figure S15 ^{13}C NMR (100 MHz, CDCl_3) of **2d**

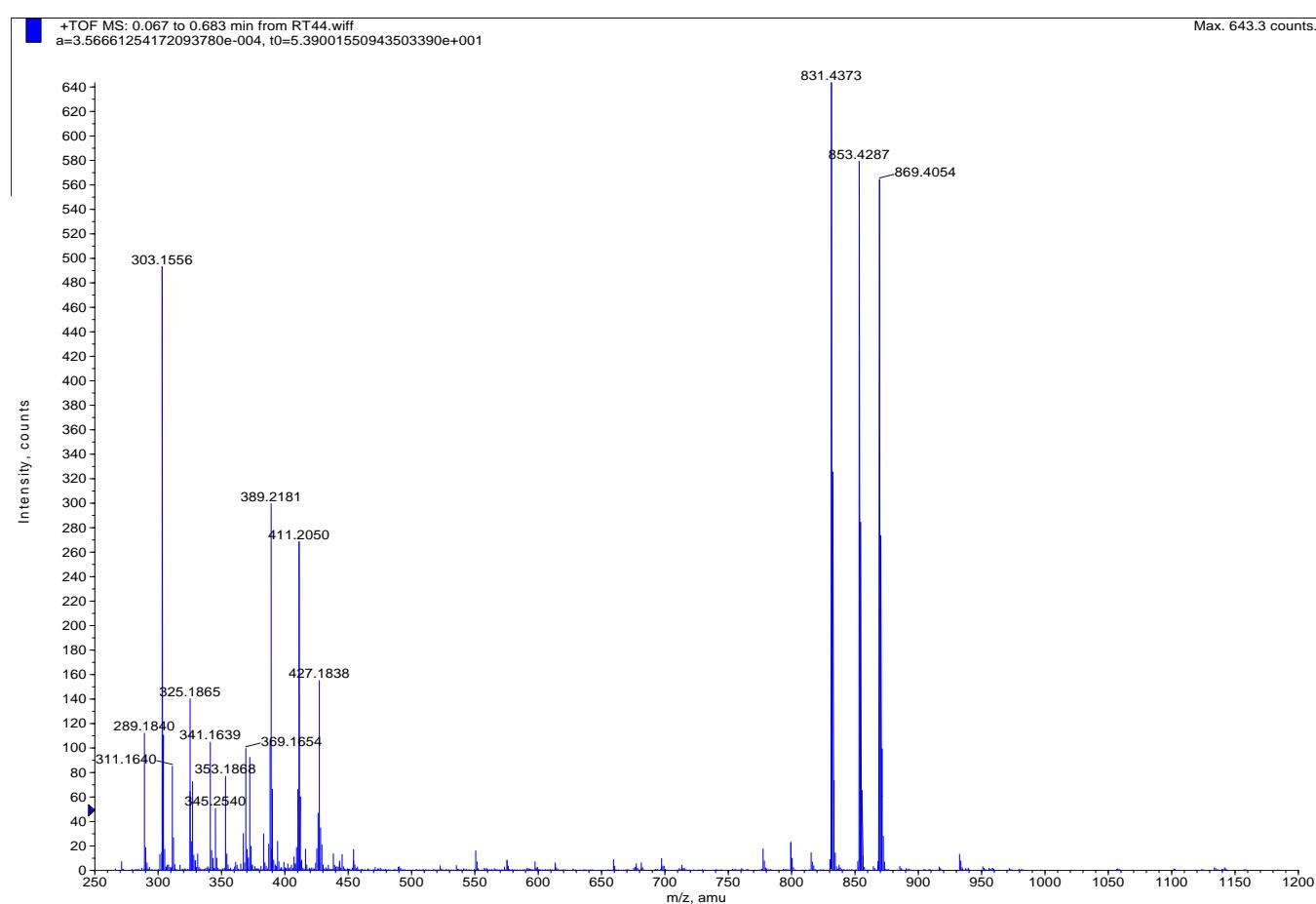


Figure S16 ESI⁺ mass spectrum of **2d**

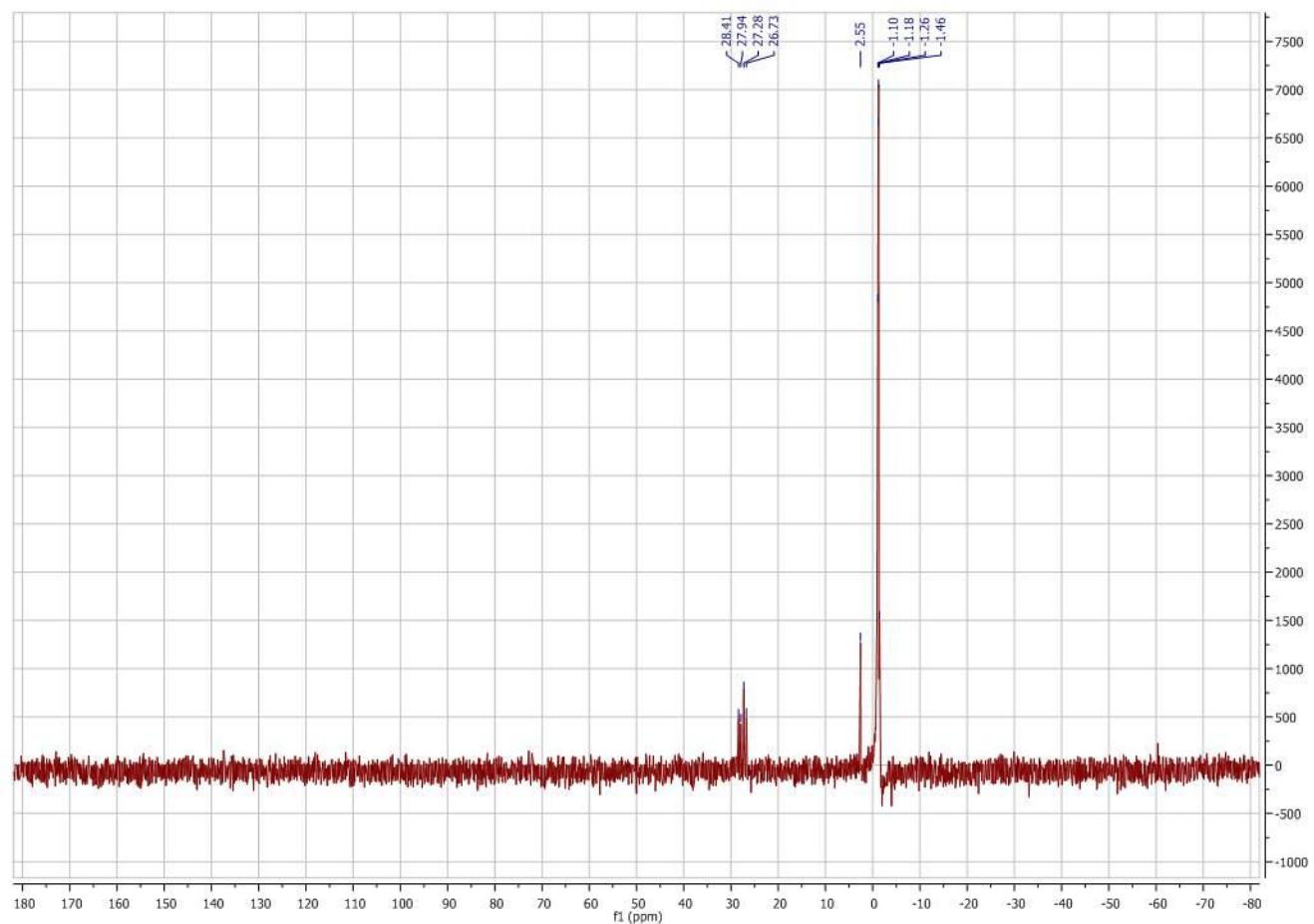


Figure S17 ^{31}P NMR (121 MHz, D_2O) of ORN 4

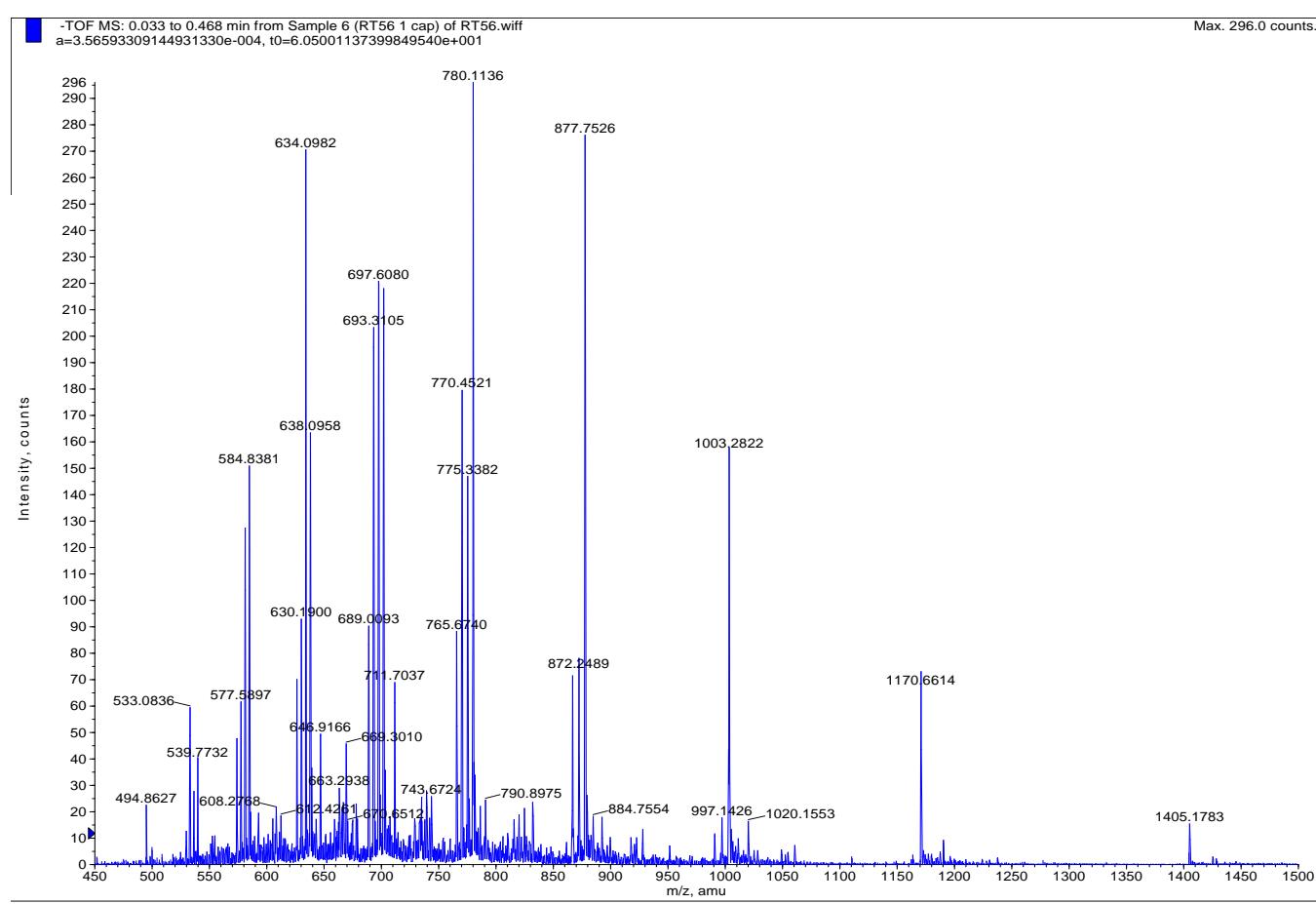
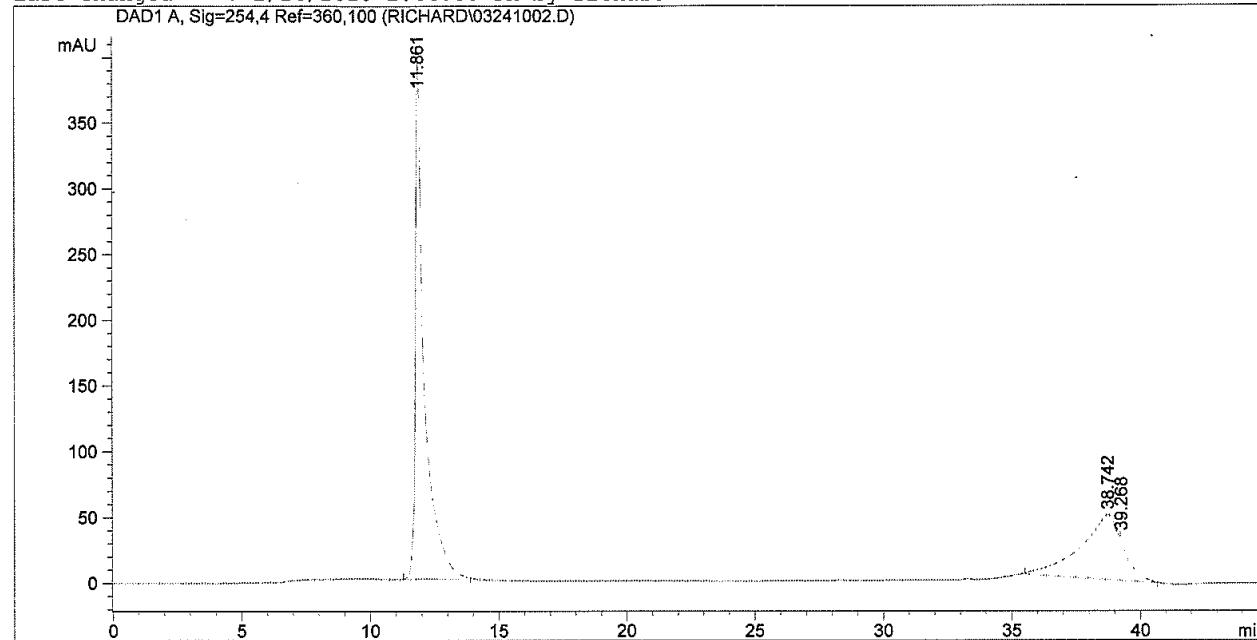


Figure S18 ESI⁻ mass spectrum of ORN 4

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Sample Name   : RT56 1cap DMToff          Location : Vial 1
Acq. Operator  : richard                Inj : 1
                                         Inj Volume : 100 µl
Different Inj Volume from Sequence !    Actual Inj Volume : 10 µl
Sequence File  : C:\HPCHEM\1\SEQUENCE\RICHARD1.S
Method        : C:\HPCHEM\1\METHODS\A-50B.M
Last changed   : 2/24/2010 2:46:50 PM by richard
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Area Percent Report
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Multiplier      : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
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Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.861	BB	0.3442	9737.42090	394.15604	63.5044
2	38.742	BV	1.2629	4761.47510	49.47045	31.0528
3	39.268	VB	0.3092	834.57184	33.90029	5.4428

Totals : 1.53335e4 477.52678

Results obtained with enhanced integrator!

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*** End of Report ***
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Instrument 1 3/24/2010 2:55:58 PM richard

Page 1 of 1

Figure S19 HPLC chromatogram of ORN 4

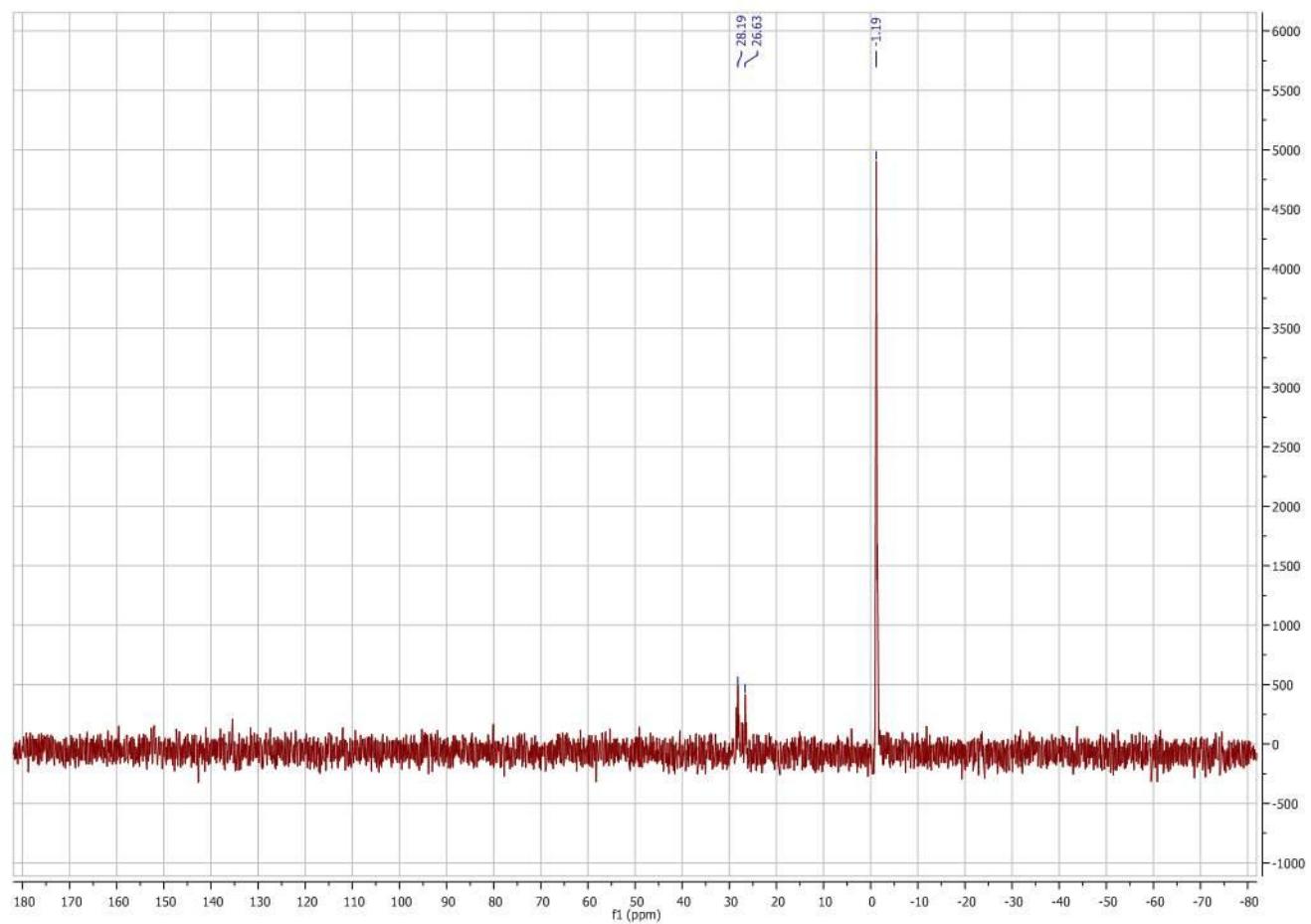


Figure S20 ^{31}P NMR (121 MHz, D_2O) of ORN 5

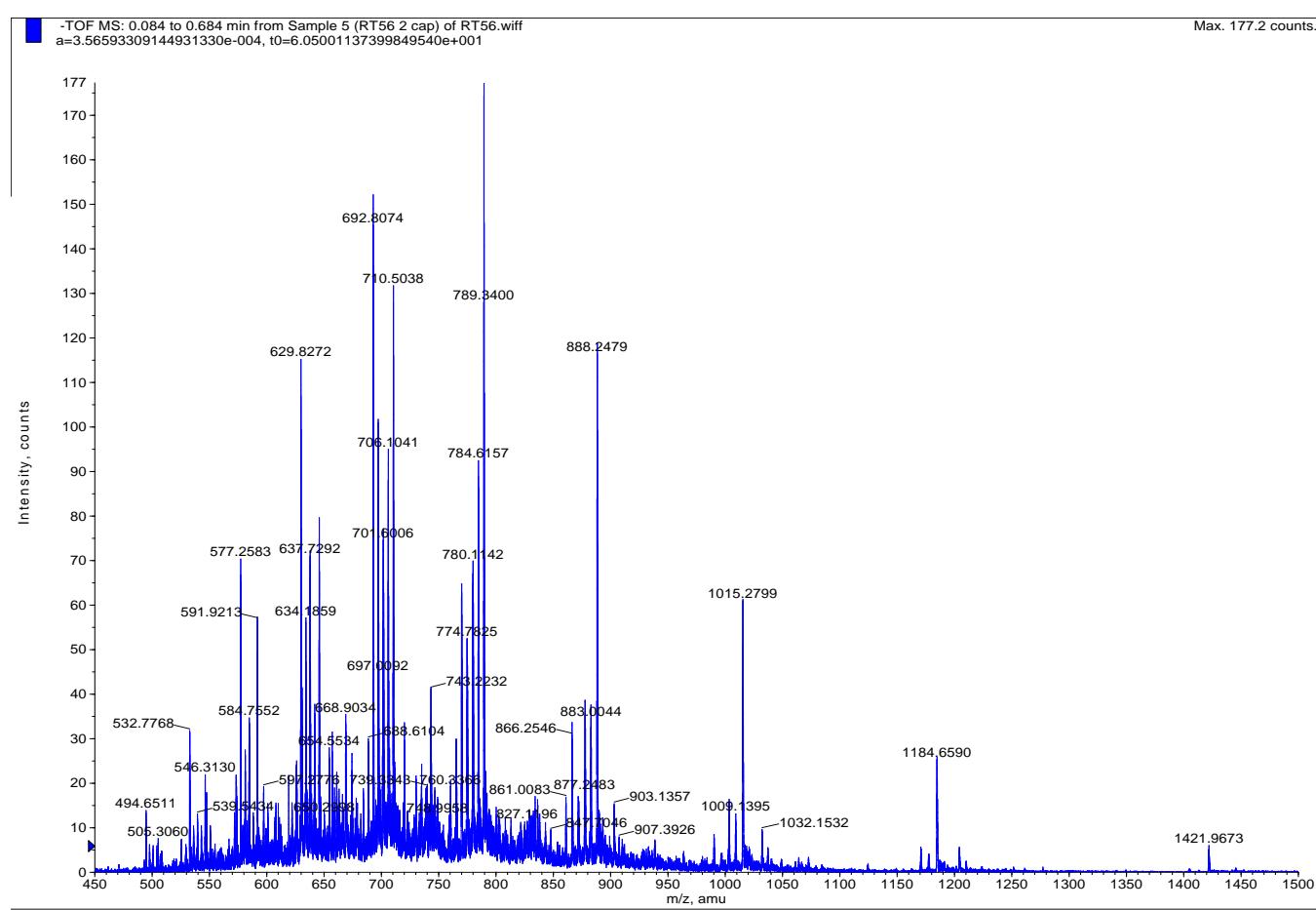
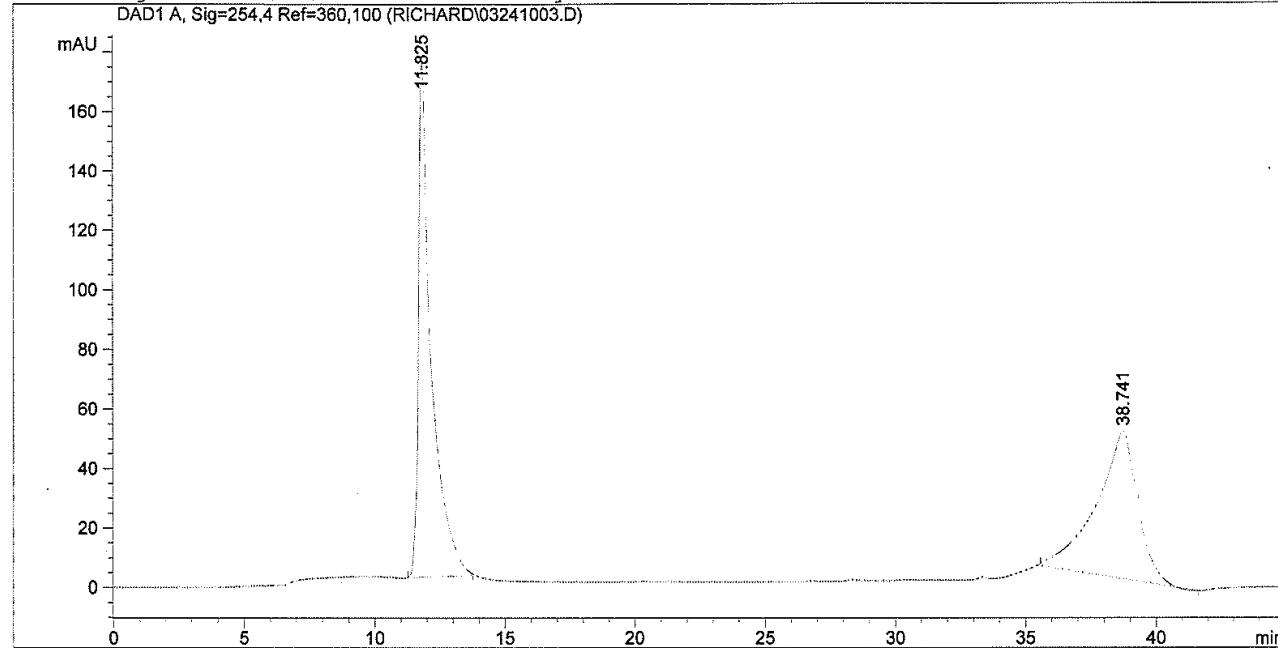


Figure S21 ESI⁻ mass spectrum of ORN 5

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Acq. Operator  : richard                Inj : 1
                                                Inj Volume : 100 µl
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Method        : C:\HPCHEM\1\METHODS\A-50B.M
Last changed   : 2/24/2010 2:46:50 PM by richard
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Area Percent Report
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Multiplier         : 1.0000
Dilution          : 1.0000
Use Multiplier & Dilution Factor with ISTDs
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Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.825	BB	0.4866	6021.92773	173.64264	52.2251
2	38.741	BP	1.4233	5508.79004	49.61862	47.7749

Totals : 1.15307e4 223.26126

Results obtained with enhanced integrator!

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Page 1 of 1

Figure S22 HPLC chromatogram of ORN 5

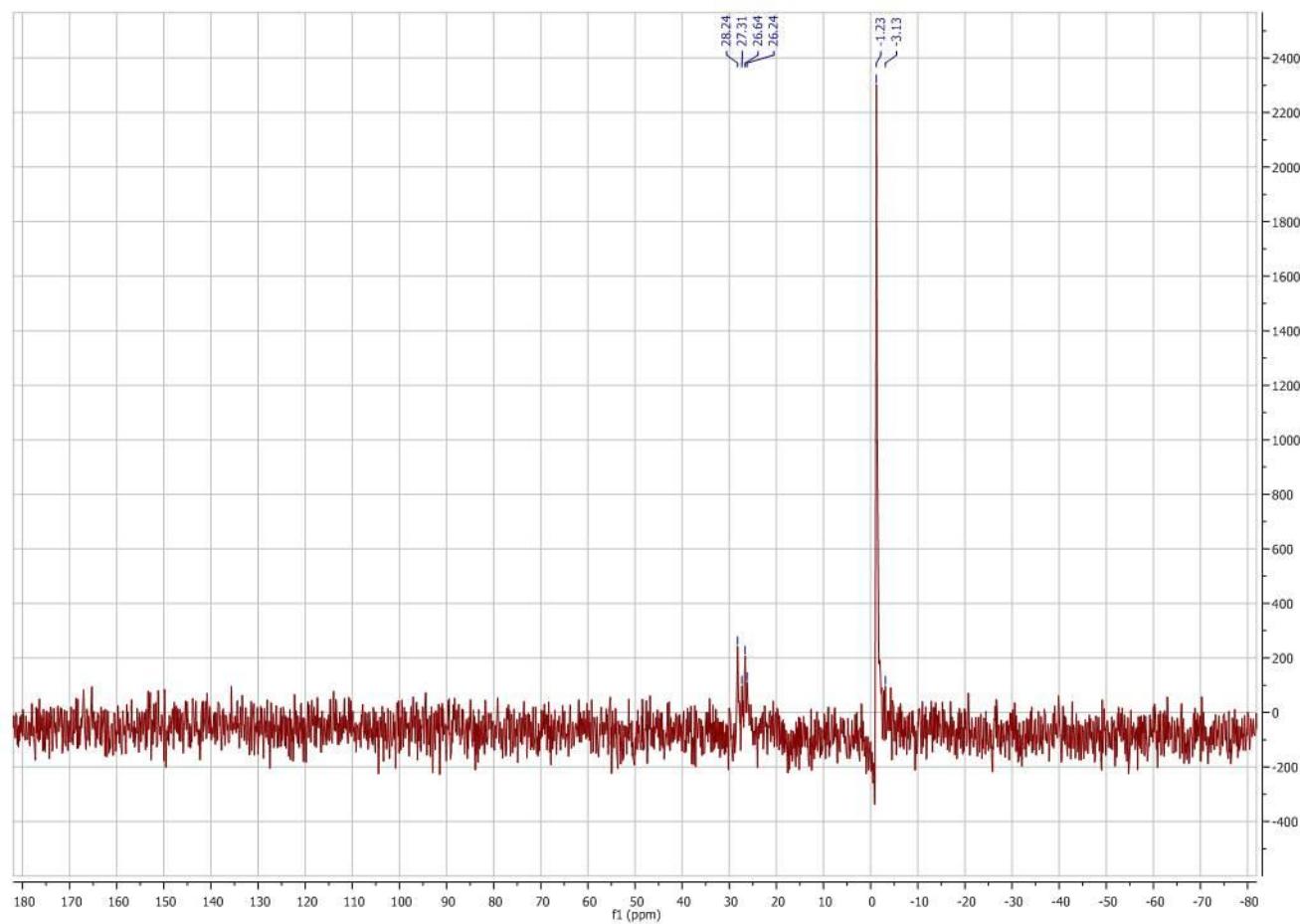


Figure S23 ^{31}P NMR (121 MHz, D_2O) of ORN 6

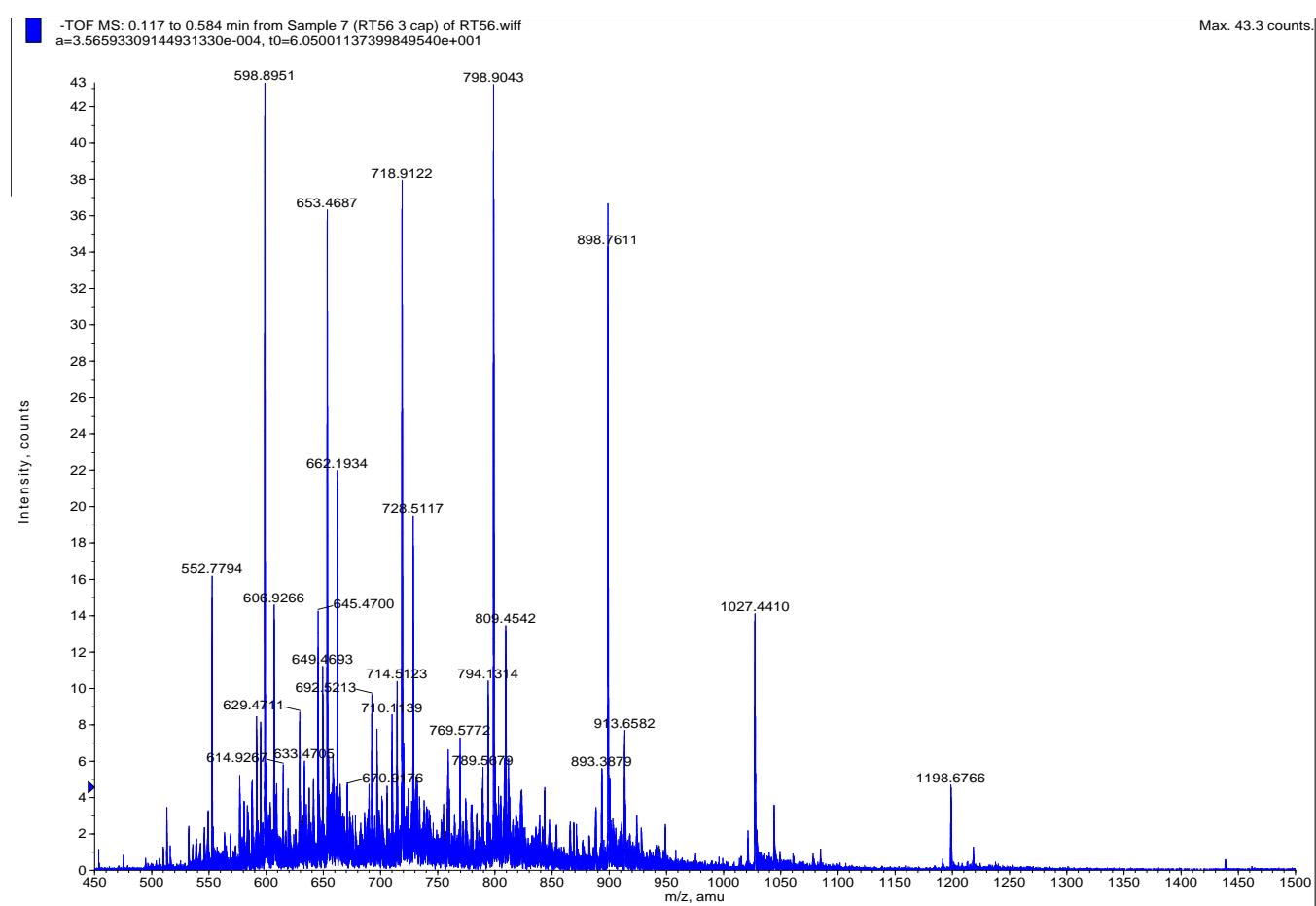
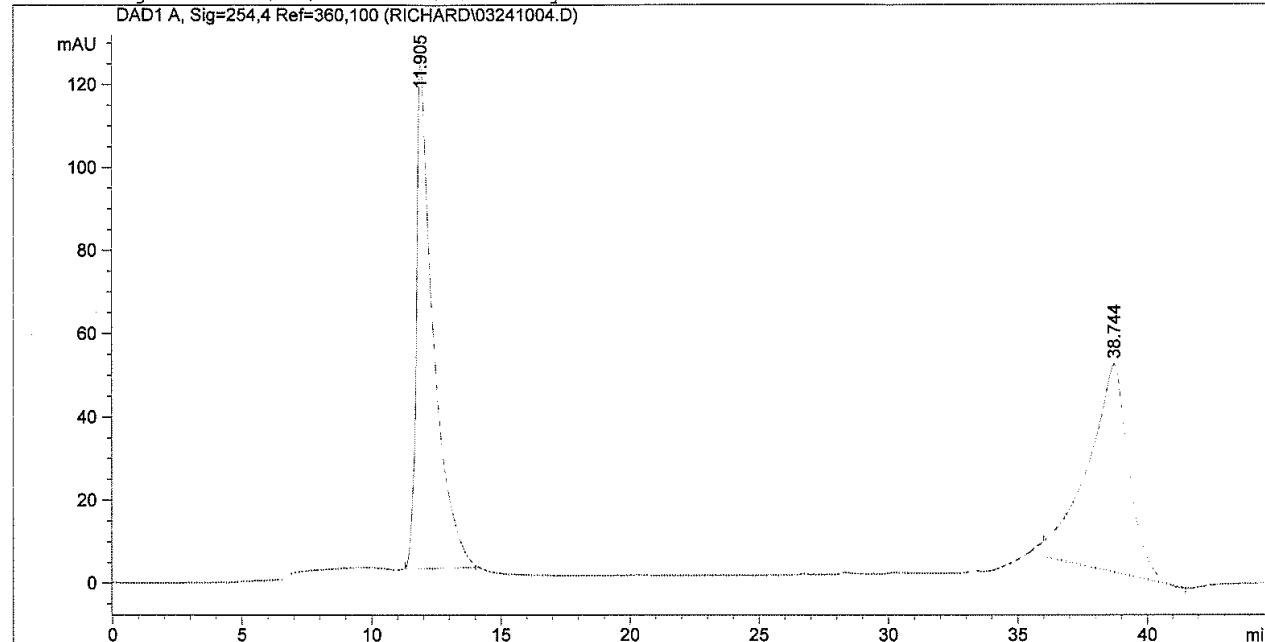


Figure S24 ESI⁻ mass spectrum of ORN 6

Data File C:\HPCHEM\1\DATA\RICHARD\03241004.D

Sample Name: RT56 3cap DMToff

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Acq. Operator   : richard            Inj : 1
                                         Inj Volume : 100 µl
Different Inj Volume from Sequence !  Actual Inj Volume : 10 µl
Sequence File   : C:\HPCHEM\1\SEQUENCE\RICHARD1.S
Method         : C:\HPCHEM\1\METHODS\A-50B.M
Last changed   : 2/24/2010 2:46:50 PM by richard
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Area Percent Report
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Dilution      : 1.0000
Use Multiplier & Dilution Factor with ISTDs
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Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.905	BB	0.6175	5567.13525	122.19295	50.0254
2	38.744	BP	1.4283	5561.47266	50.05218	49.9746

Totals : 1.11286e4 172.24512

Results obtained with enhanced integrator!

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*** End of Report ***

Instrument 1 3/24/2010 4:29:55 PM richard

Page 1 of 1

Figure S25 HPLC chromatogram of ORN 6

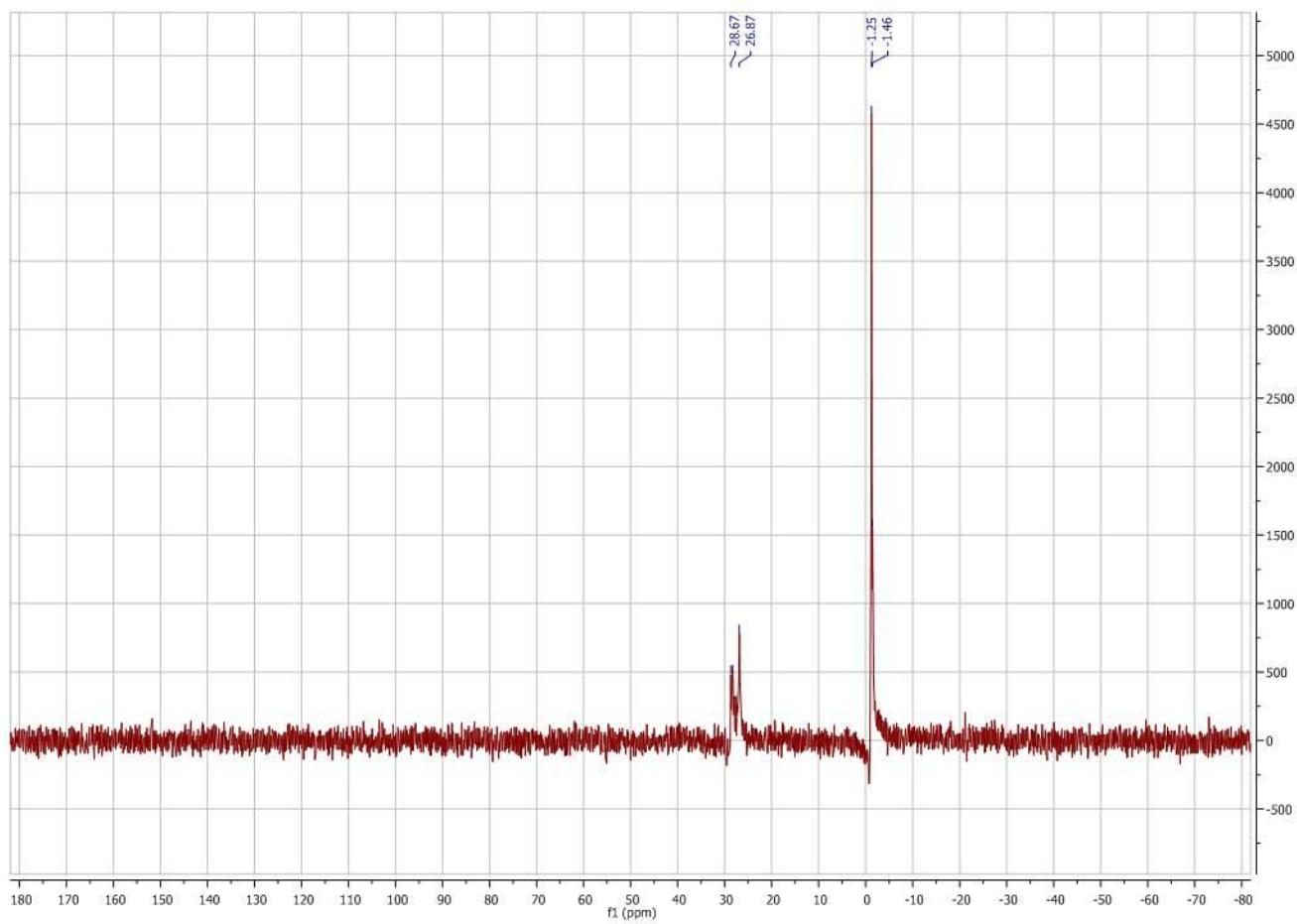


Figure S26 ^{31}P NMR (121 MHz, D_2O) of ORN 7

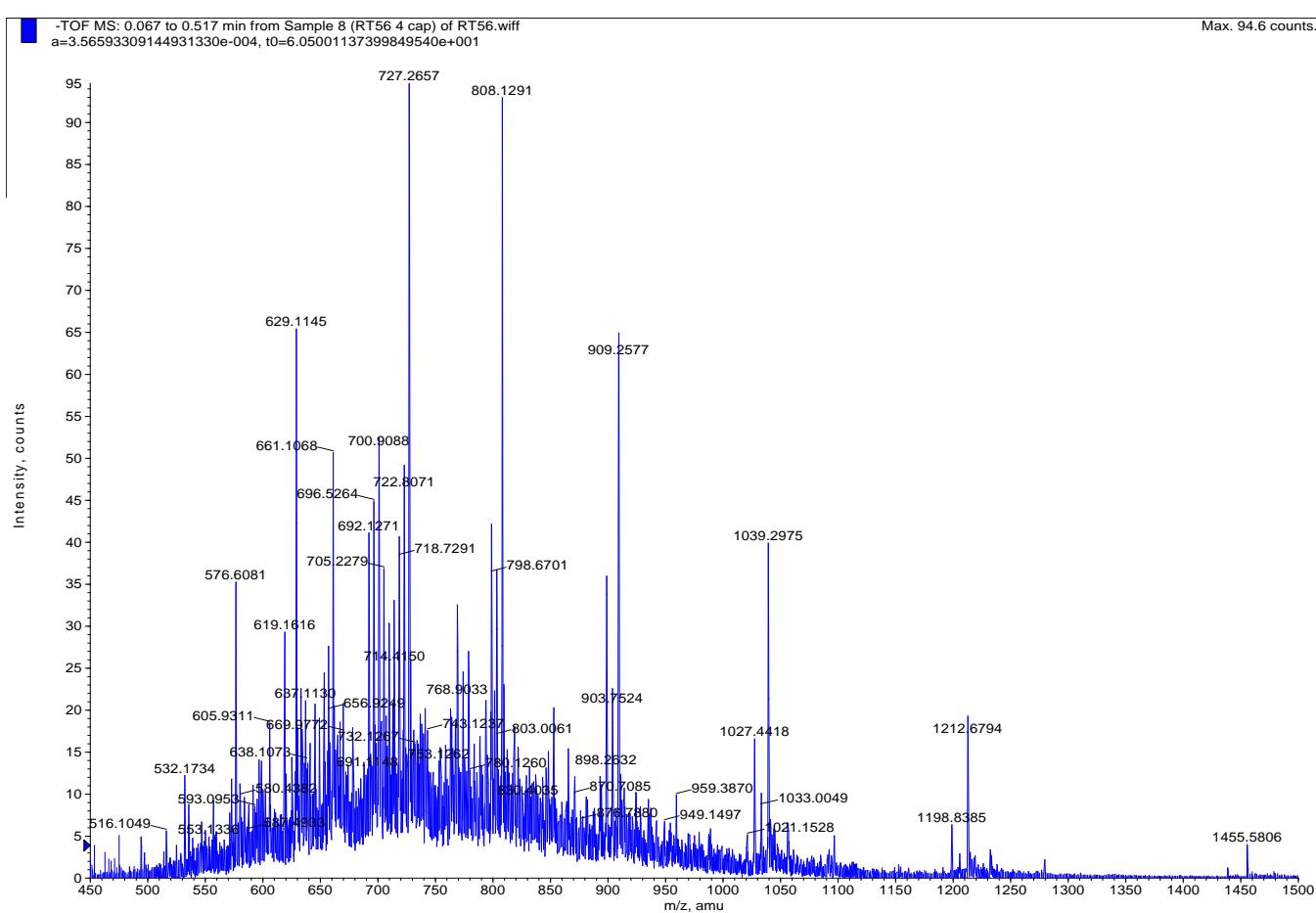
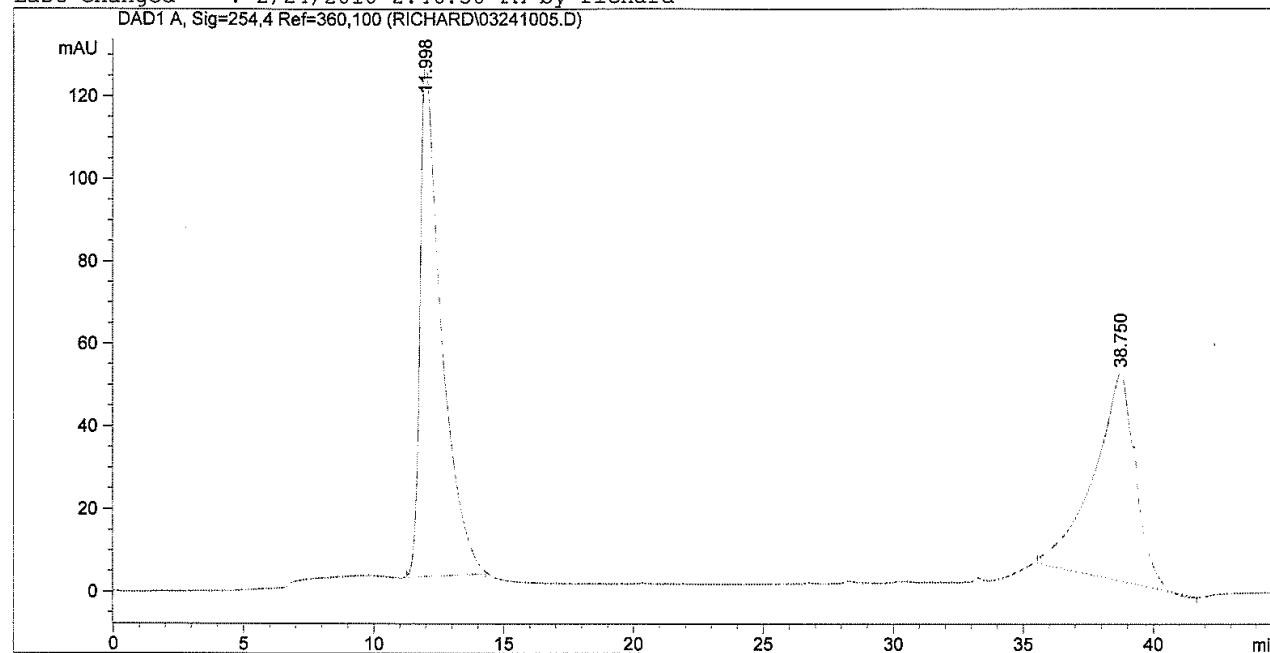


Figure S27 ESI⁻ mass spectrum of ORN 7

Data File C:\HPCHEM\1\DATA\RICHARD\03241005.D

Sample Name: RT56 4cap DMToff

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Injection Date : 3/24/2010 4:31:37 PM      Seq. Line : 5
Sample Name   : RT56 4cap DMToff        Location : Vial 4
Acq. Operator  : richard             Inj : 1
                                                Inj Volume : 100 µl
Different Inj Volume from Sequence !    Actual Inj Volume : 10 µl
Sequence File  : C:\HPCHEM\1\SEQUENCE\RICHARD1.S
Method        : C:\HPCHEM\1\METHODS\A-50B.M
Last changed   : 2/24/2010 2:46:50 PM by richard
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Area Percent Report
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Multiplier        :     1.0000
Dilution         :     1.0000
Use Multiplier & Dilution Factor with ISTDs
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Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.998	BB	0.7957	7148.49951	123.78619	55.8880
2	38.750	BP	1.4340	5642.26270	50.32434	44.1120

Totals : 1.27908e4 174.11053

Results obtained with enhanced integrator!

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*** End of Report ***
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Instrument 1 3/24/2010 5:16:54 PM richard

Page 1 of 1

Figure S25 HPLC chromatogram of ORN 7

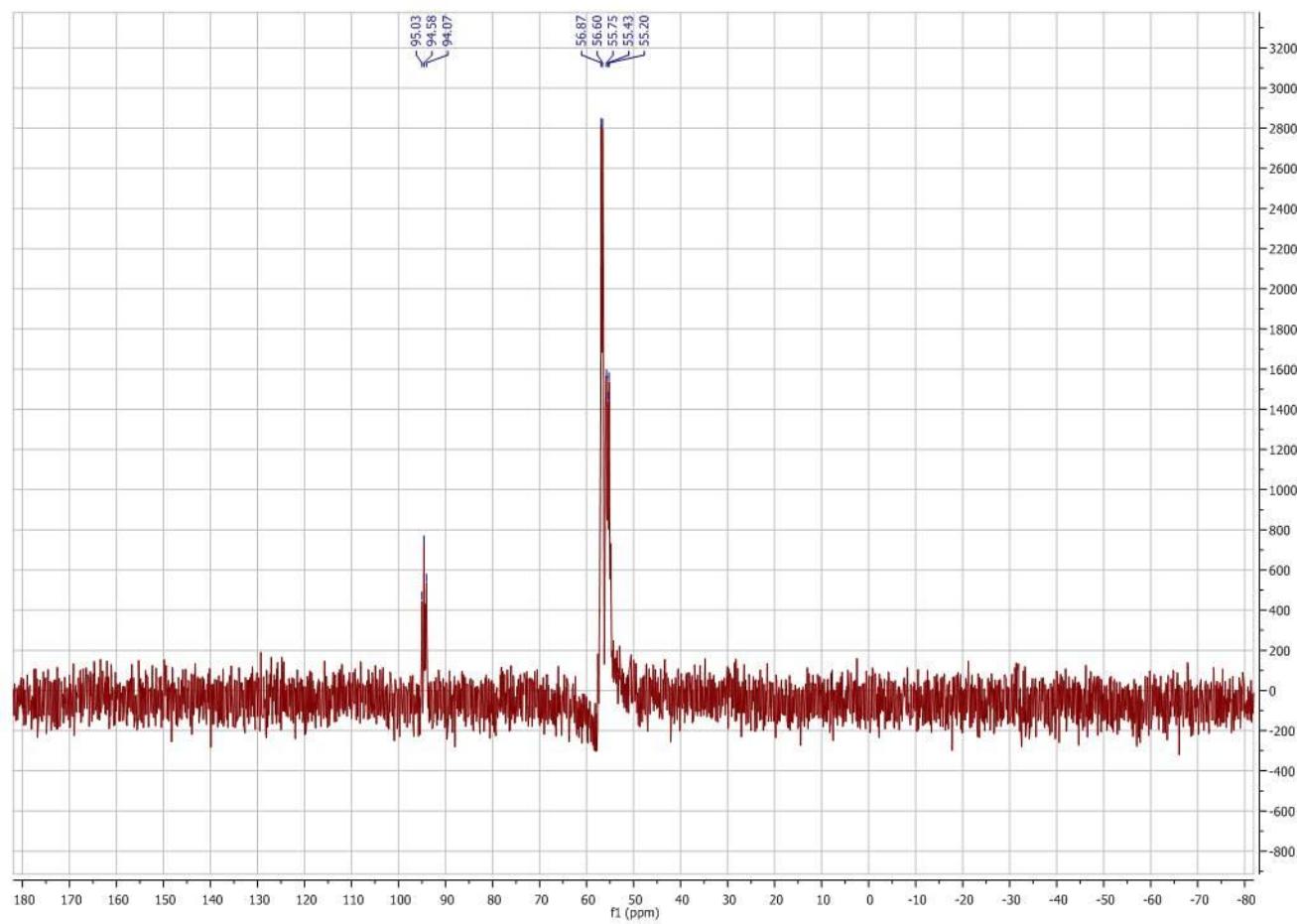


Figure S28 ^{31}P NMR (121 MHz, D_2O) of ORN 9

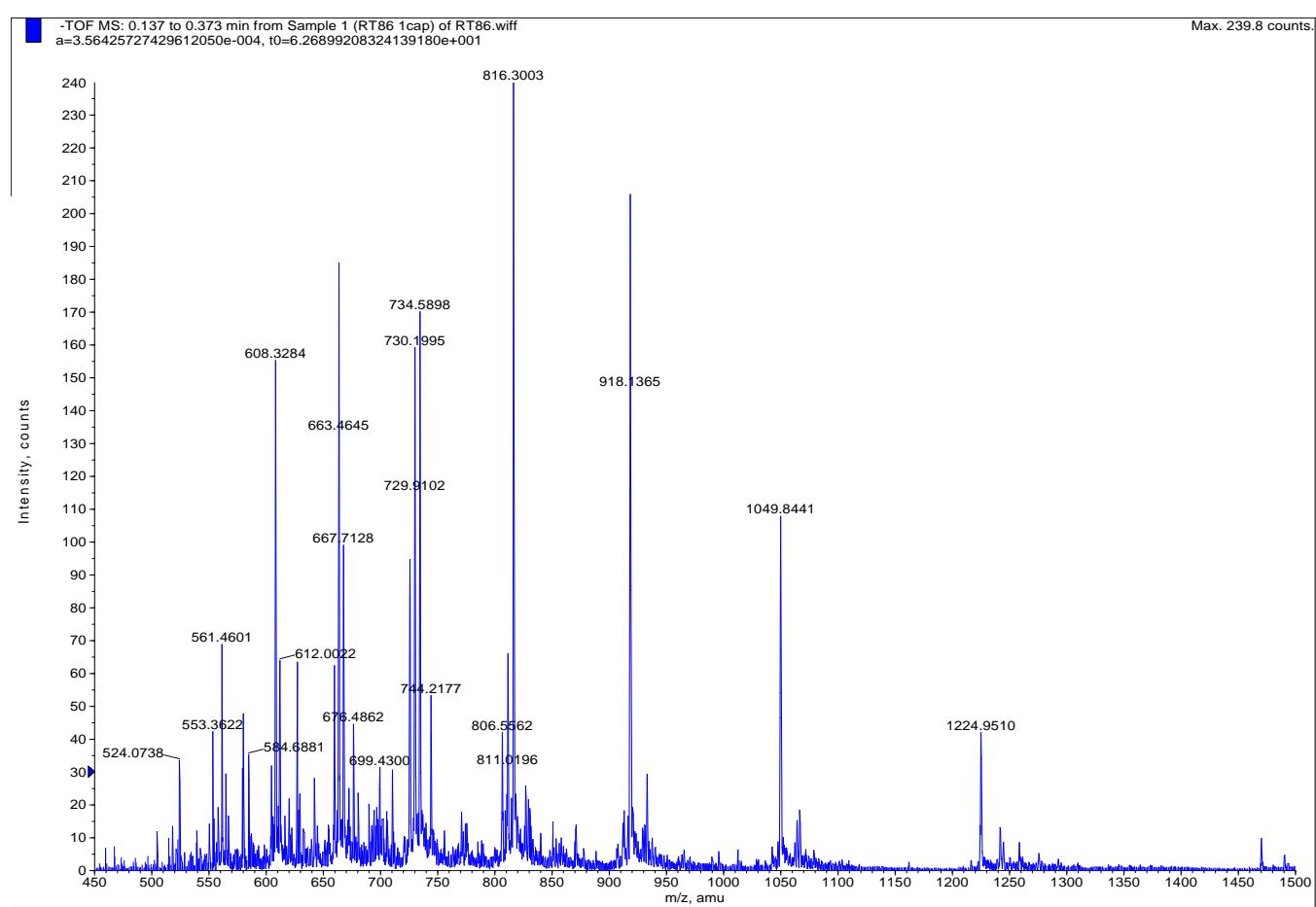
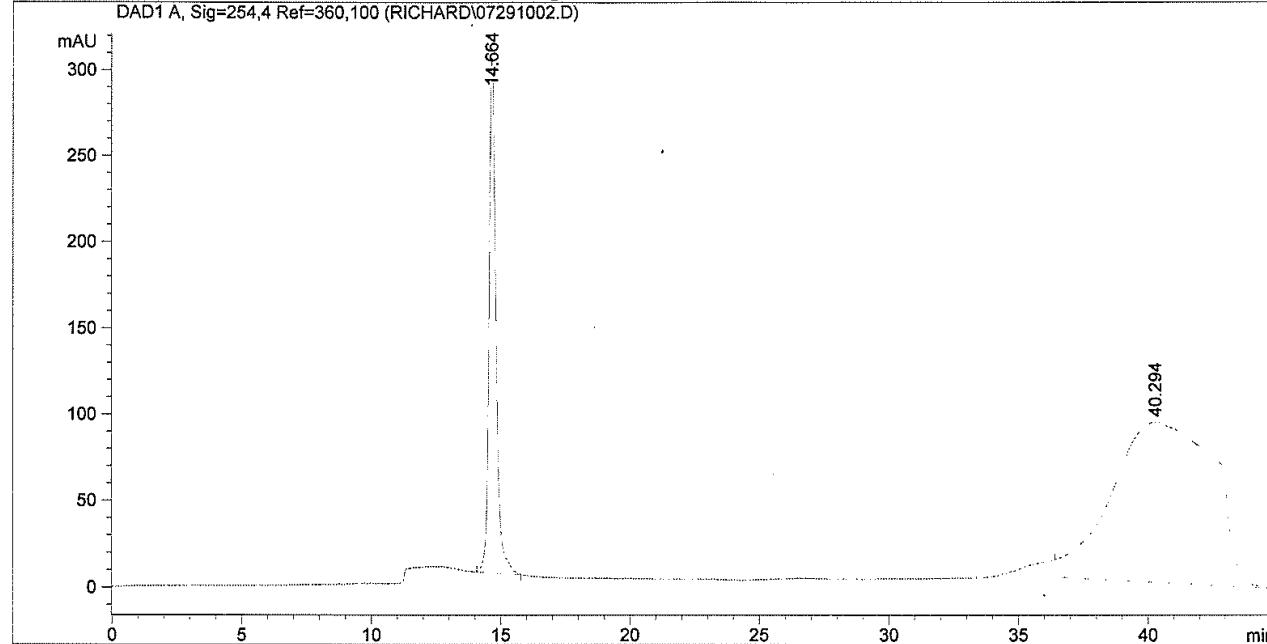


Figure S29 ESI⁻ mass spectrum of ORN 9

Data File C:\HPCHEM\1\DATA\RICHARD\07291002.D

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Injection Date : 7/29/2010 11:27:22 AM Seq. Line : 2
Sample Name : RT86 1cap DMToff Location : Vial 1
Acq. Operator : richard Inj : 1
Inj Volume : 100 μ l
Différent Inj Volume from Sequence ! Actual Inj Volume : 10 μ l
Sequence File : C:\HPCHEM\1\SEQUENCE\RICHARD1.S
Method : C:\HPCHEM\1\METHODS\A-50B.M
Last changed : 6/10/2010 9:15:59 AM by richard



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Area Percent Report
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Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.664	BB	0.2794	5564.75488	298.58466	17.8568
2	40.294	BBA	3.2326	2.55985e4	92.78777	82.1432

Totals : 3.11632e4 391.37243

Results obtained with enhanced integrator!

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*** End of Report ***

Instrument 1 7/29/2010 12:12:34 PM richard

Page 1 of 1

Figure S30 HPLC chromatogram of ORN 9

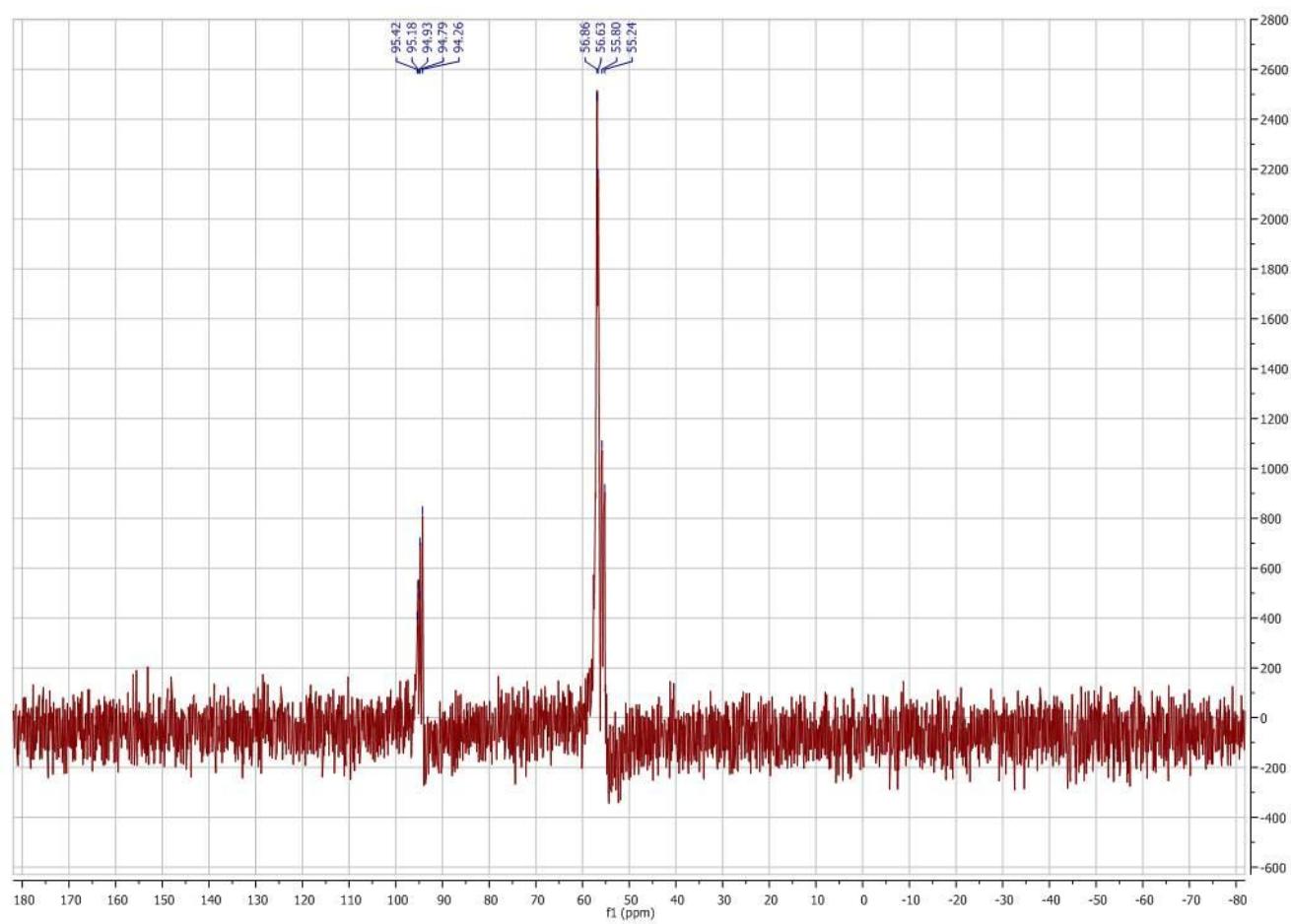


Figure S31 ^{31}P NMR (121 MHz, D_2O) of ORN **10**

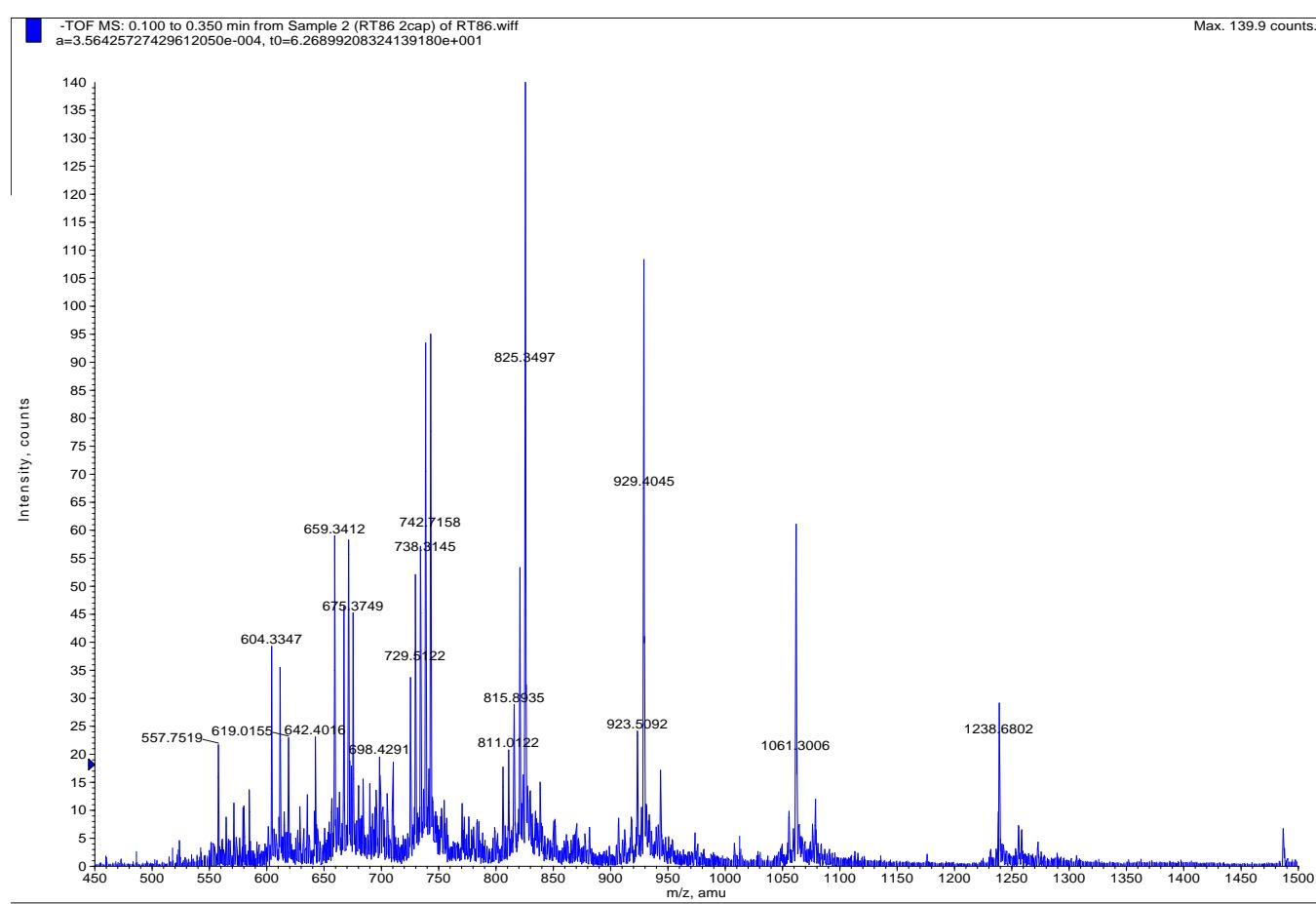
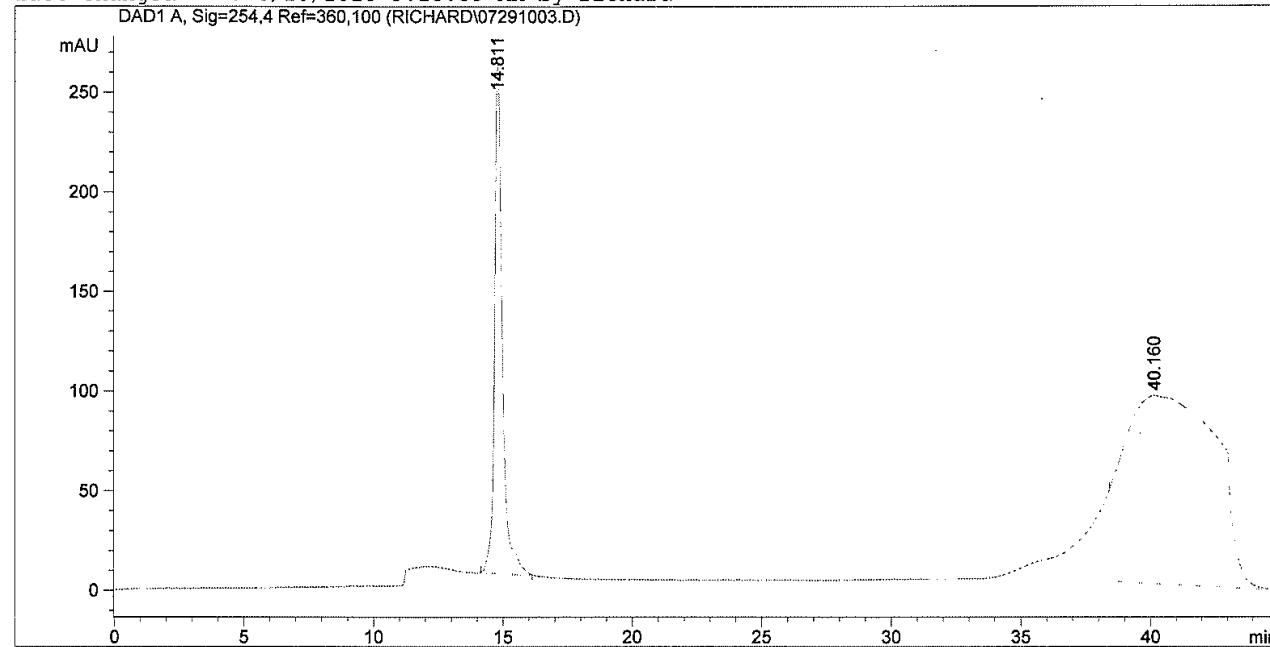


Figure S32 ESI⁻ mass spectrum of ORN 10

Data File C:\HPCHEM\1\DATA\RICHARD\07291003.D

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Sample Name   : RT86 2cap DMToff          Location  : Vial 2
Acq. Operator  : richard                Inj       : 1
                                                Inj Volume : 100 µl
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Sequence File  : C:\HPCHEM\1\SEQUENCE\RICHARD1.S
Method        : C:\HPCHEM\1\METHODS\A-50B.M
Last changed   : 6/10/2010 9:15:59 AM by richard
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Area Percent Report
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Sorted By      : Signal
Multiplier     : 1.0000
Dilution      : 1.0000
Use Multiplier & Dilution Factor with ISTDs
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Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.811	BB	0.3115	5394.86377	256.40179	18.5915
2	40.160	BBA	2.9337	2.36230e4	94.39524	81.4085

Totals : 2.90178e4 350.79704

Results obtained with enhanced integrator!

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Instrument 1 7/29/2010 12:59:30 PM richard

Page 1 of 1

Figure S33 HPLC chromatogram of ORN 10

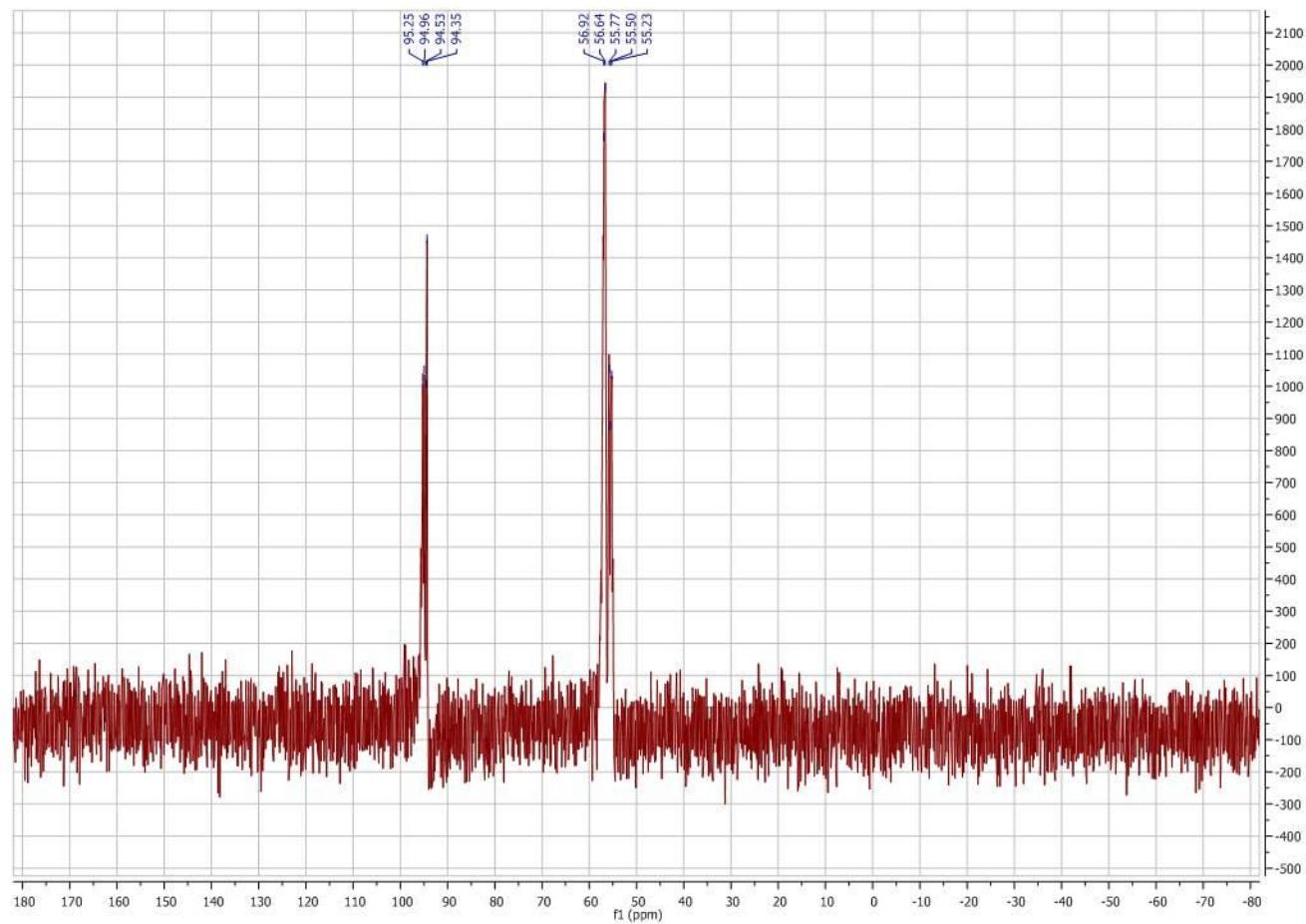


Figure S34 ^{31}P NMR (121 MHz, D_2O) of ORN **11**

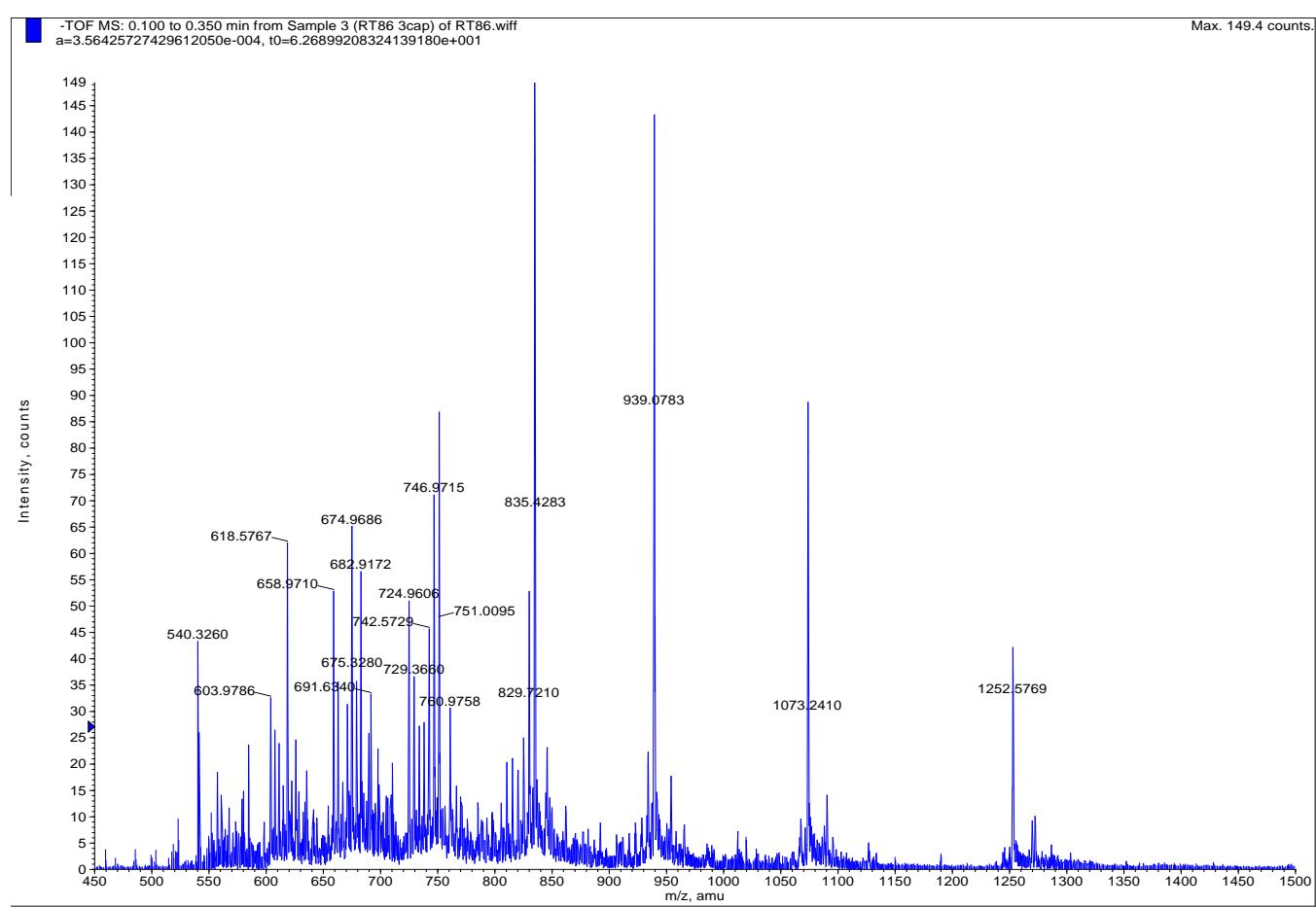
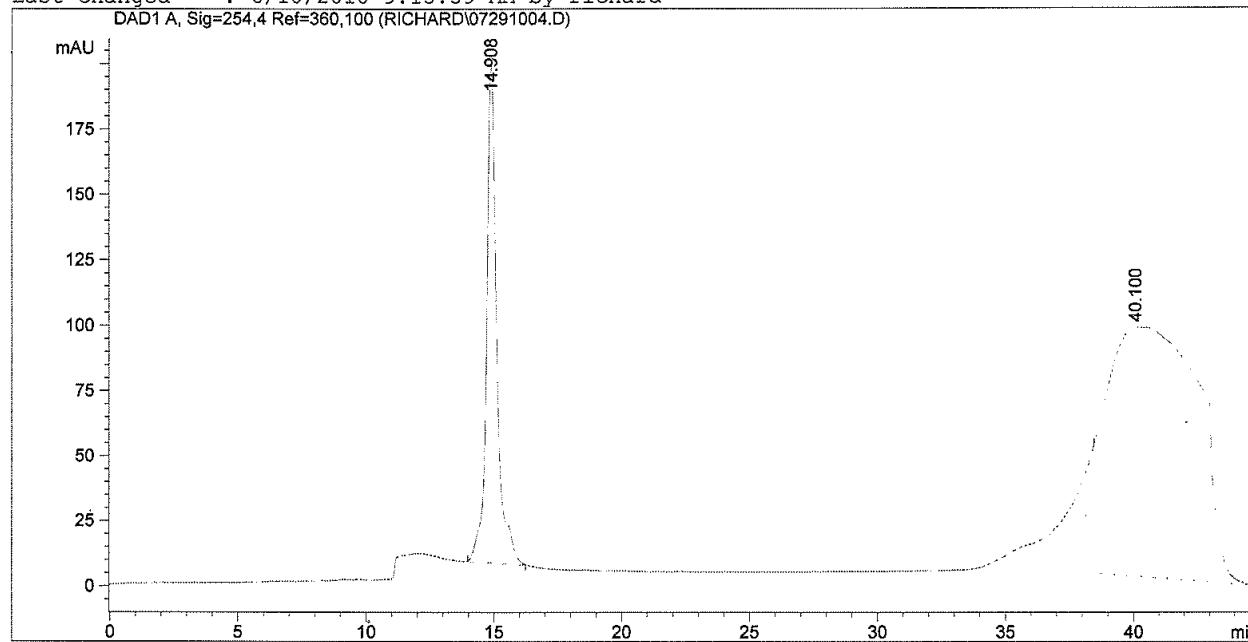


Figure S35 ESI⁻ mass spectrum of ORN 11

Data File C:\HPCHEM\1\DATA\RICHARD\07291004.D

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Sample Name : RT86 3cap DMToff Location : Vial 3
Acq. Operator : richard Inj : 1
Inj Volume : 100 μ l
Different Inj Volume from Sequence ! Actual Inj Volume : 10 μ l
Sequence File : C:\HPCHEM\1\SEQUENCE\RICHARD1.S
Method : C:\HPCHEM\1\METHODS\A-50B.M
Last changed : 6/10/2010 9:15:59 AM by richard



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Area Percent Report
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Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.908	BB	0.4101	5306.27295	190.98944	18.1493
2	40.100	BBA	2.9346	2.39306e4	95.33340	81.8507

Totals : 2.92368e4 286.32284

Results obtained with enhanced integrator!

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*** End of Report ***

Instrument 1 7/29/2010 1:46:31 PM richard

Page 1 of 1

Figure S36 HPLC chromatogram of ORN 11

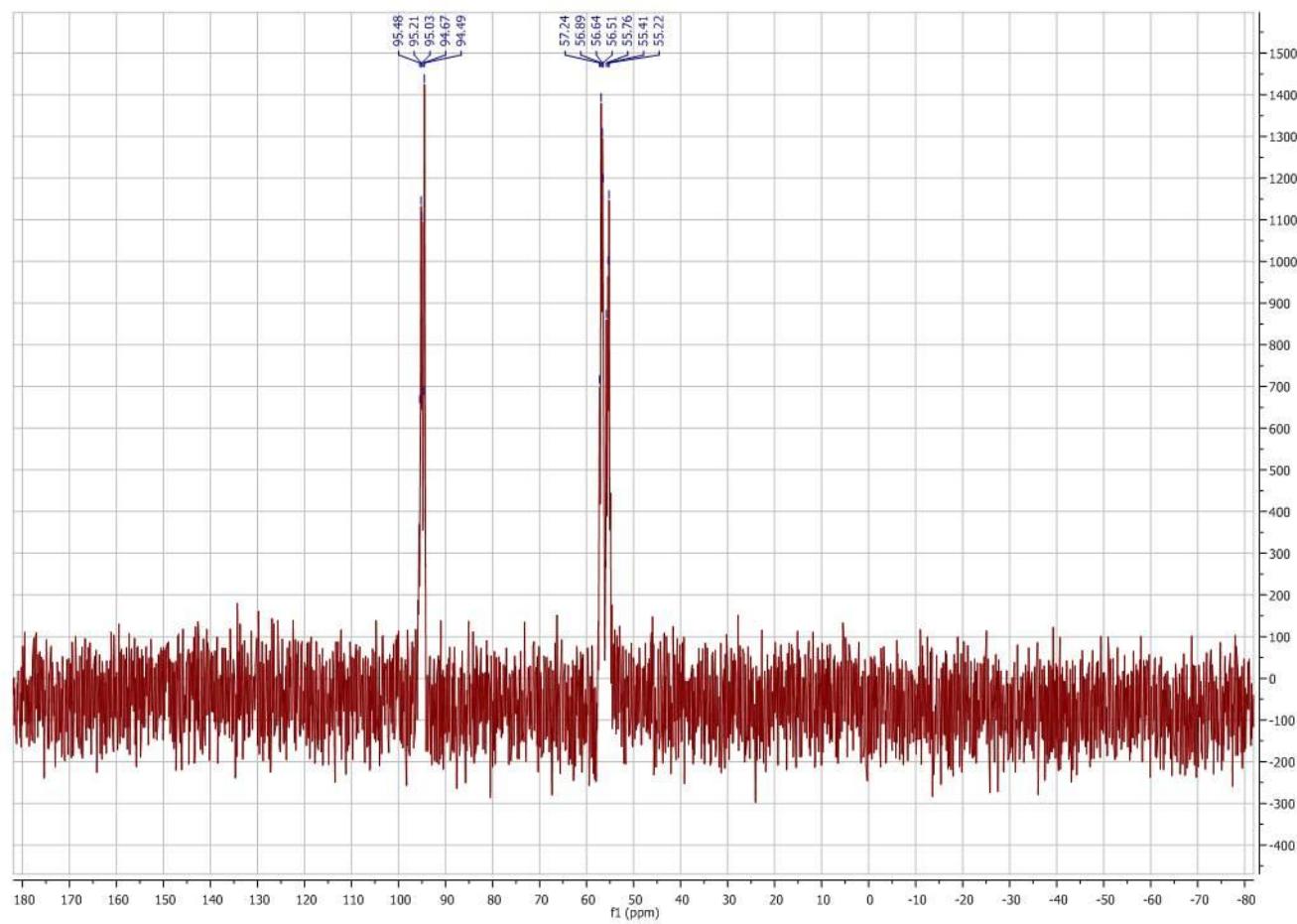


Figure S37 ^{31}P NMR (121 MHz, D_2O) of ORN **12**

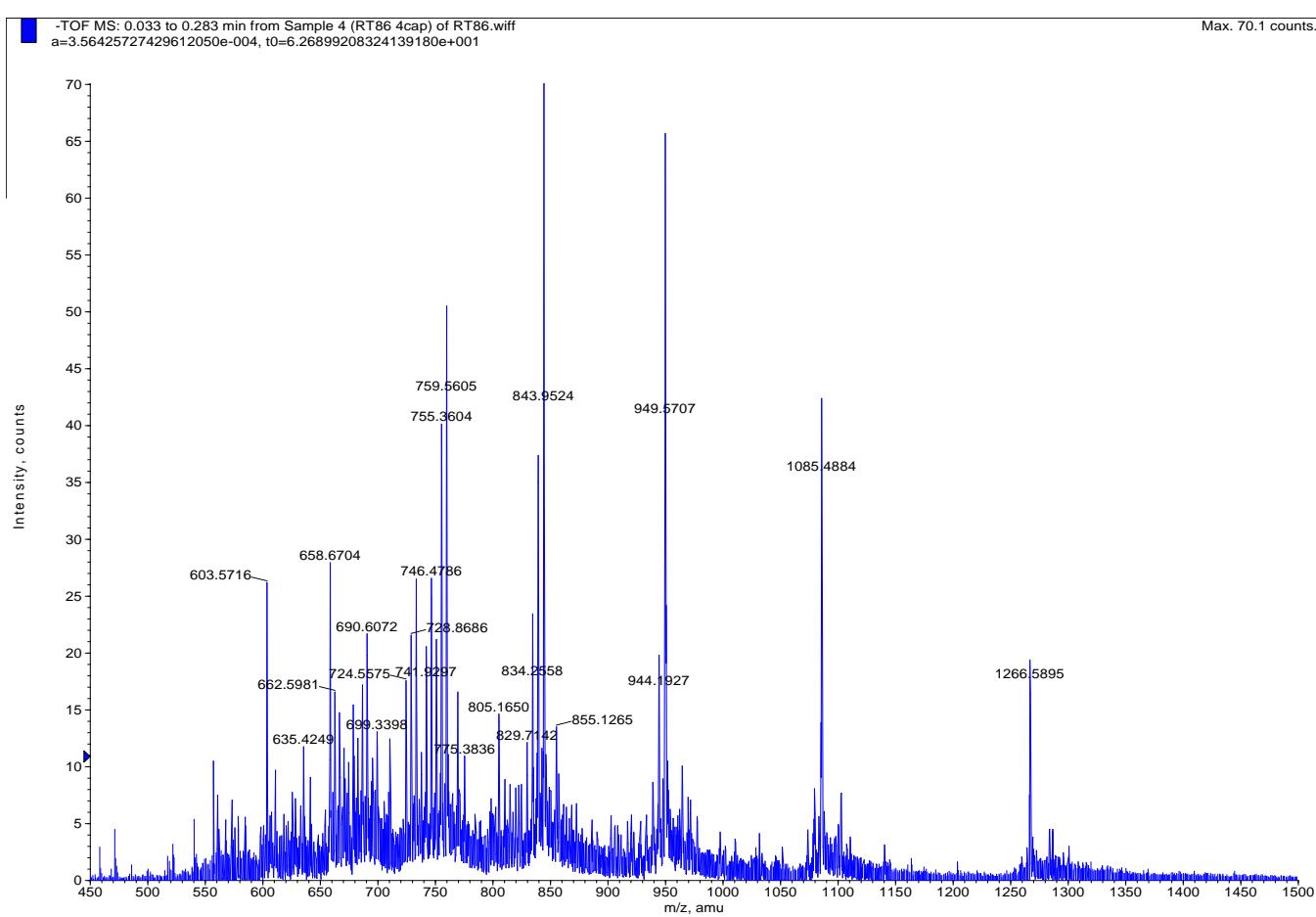
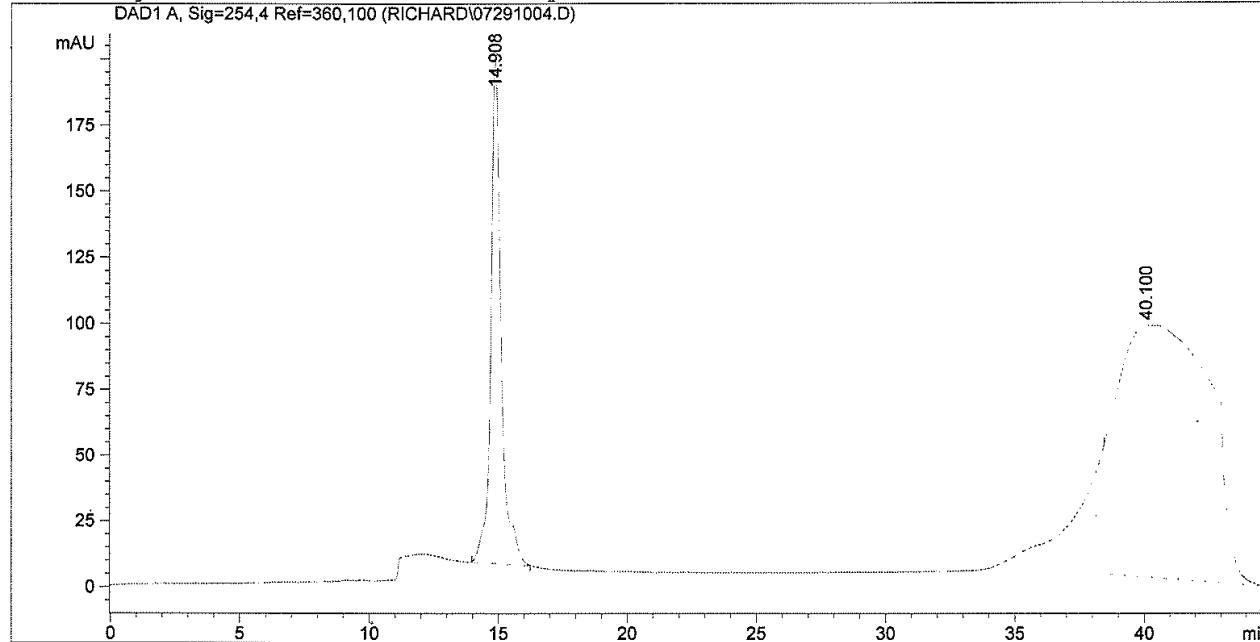


Figure S38 ESI⁻ mass spectrum of ORN 12

Data File C:\HPCHEM\1\DATA\RICHARD\07291004.D

Sample Name: RT86 3cap DMToff

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Injection Date : 7/29/2010 1:01:18 PM          Seq. Line : 4
Sample Name   : RT86 3cap DMToff           Location : Vial 3
Acq. Operator  : richard                 Inj : 1
                                         Inj Volume : 100 µl
Different Inj Volume from Sequence !      Actual Inj Volume : 10 µl
Sequence File  : C:\HPCHEM\1\SEQUENCE\RICHARD1.S
Method        : C:\HPCHEM\1\METHODS\A-50B.M
Last changed   : 6/10/2010 9:15:59 AM by richard
```



=====
Area Percent Report
=====

```
Sorted By       : Signal
Multiplier     : 1.0000
Dilution      : 1.0000
Use Multiplier & Dilution Factor with ISTDs
```

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.908	BB	0.4101	5306.27295	190.98944	18.1493
2	40.100	BBA	2.9346	2.39306e4	95.33340	81.8507

Totals : 2.92368e4 286.32284

Results obtained with enhanced integrator!

=====
*** End of Report ***

Instrument 1 7/29/2010 1:46:31 PM richard

Page 1 of 1

Figure S39 HPLC chromatogram of ORN 12

Figure S40 Cell Viability (No Lipids) 1 μ M

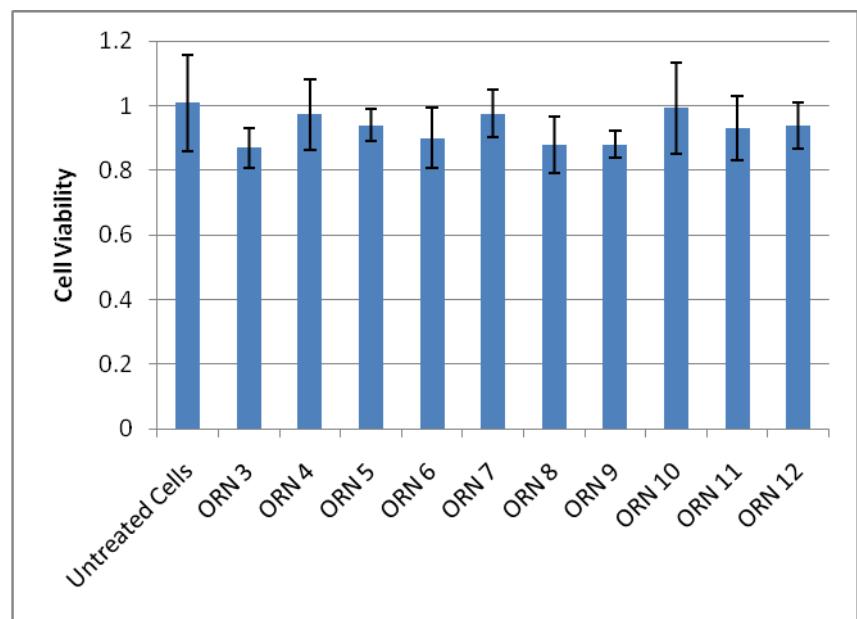
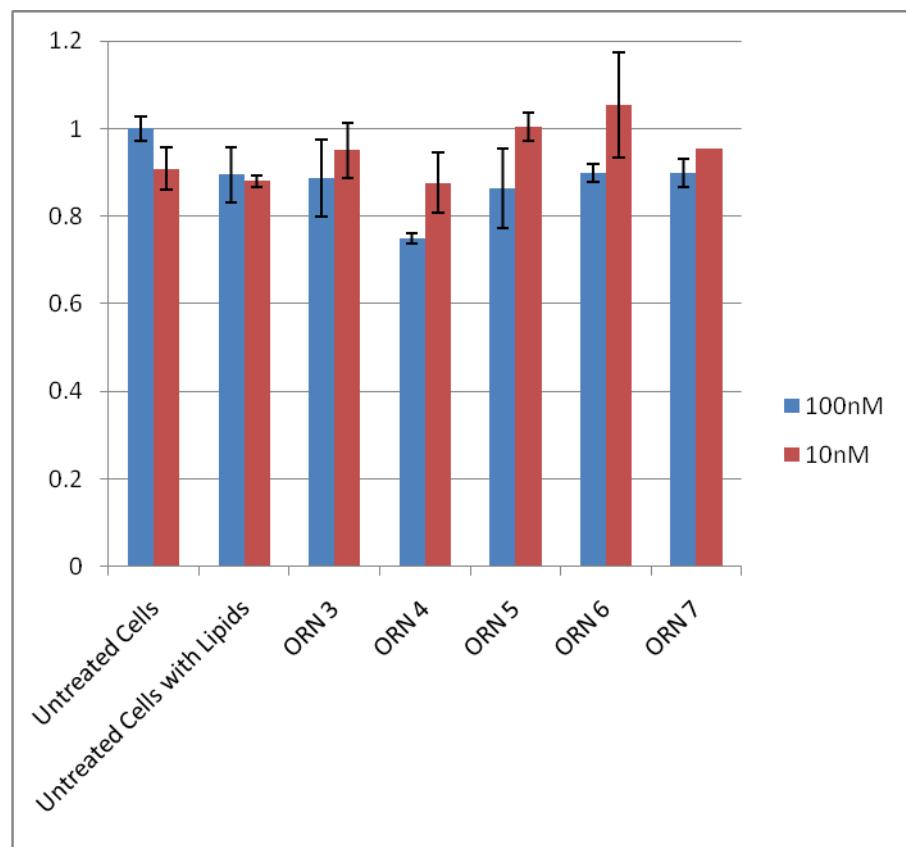
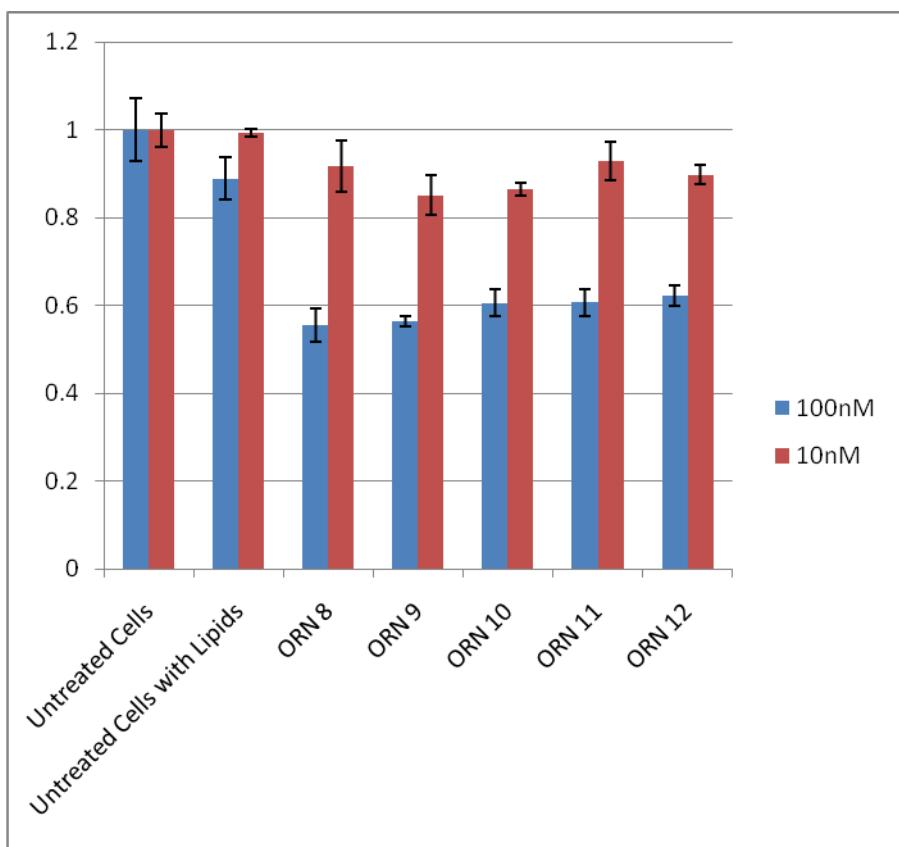


Figure S41 Cell Viability (with Lipids)





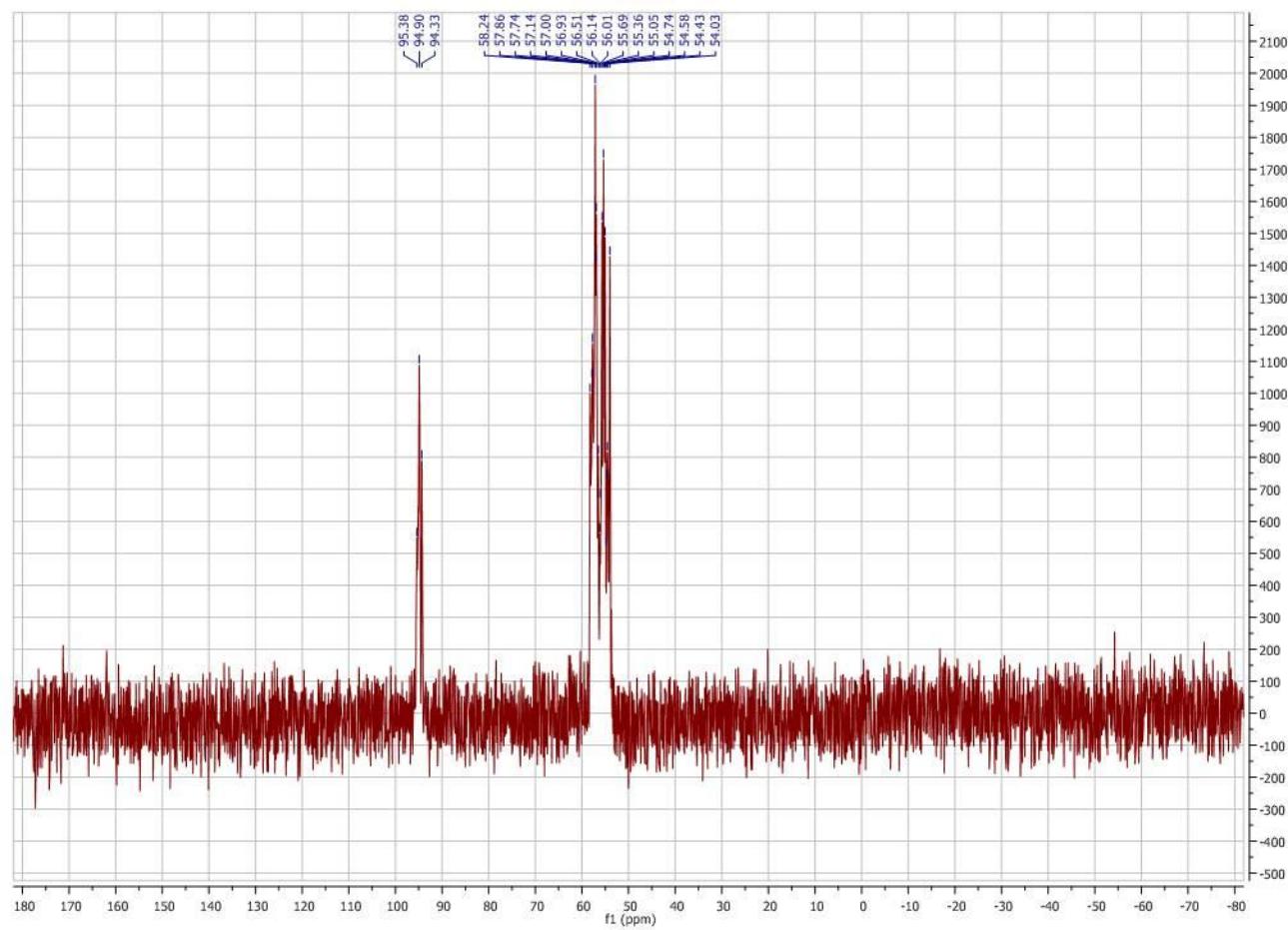


Figure S42 ^{31}P NMR (121 MHz, D_2O) of ORN 13

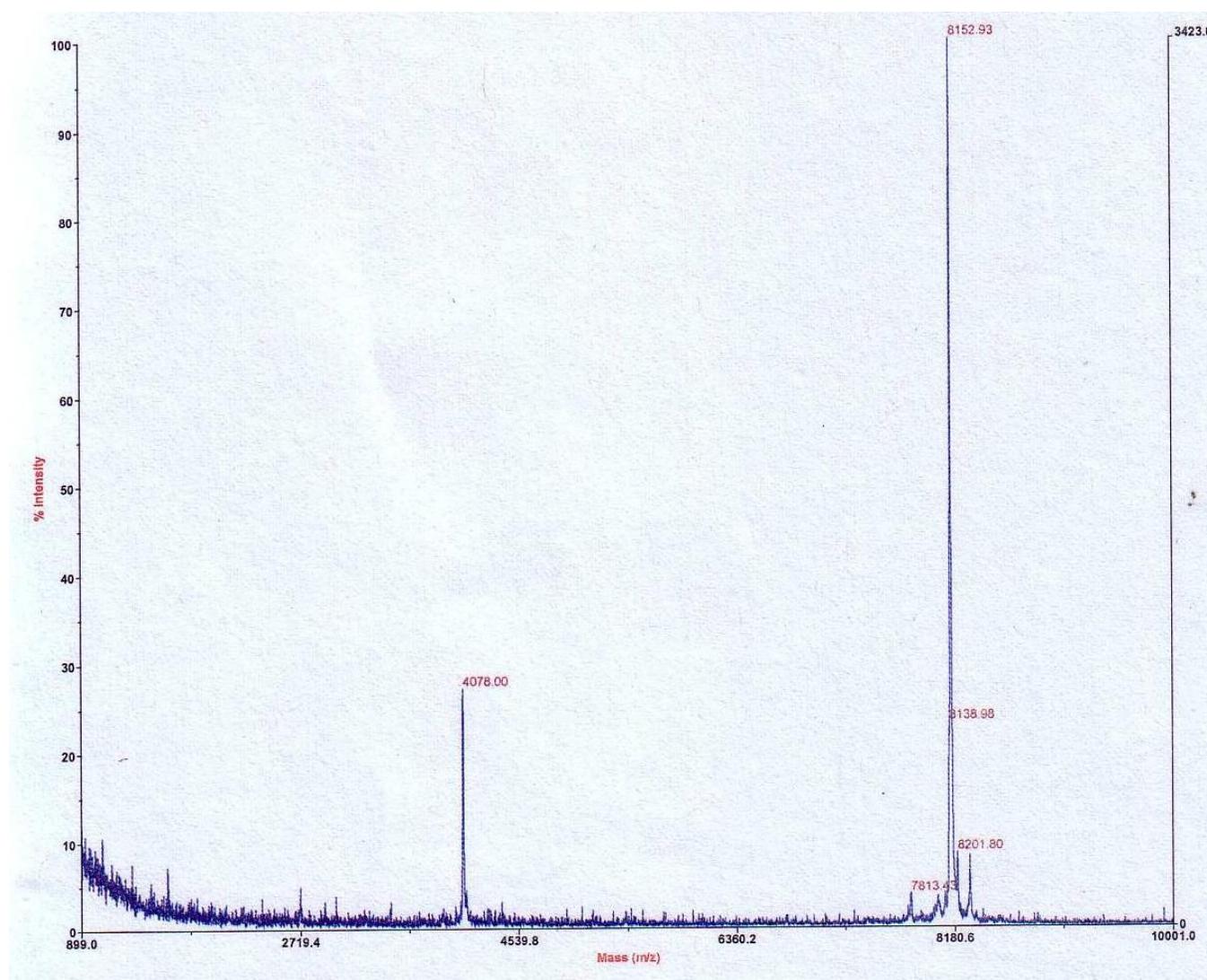
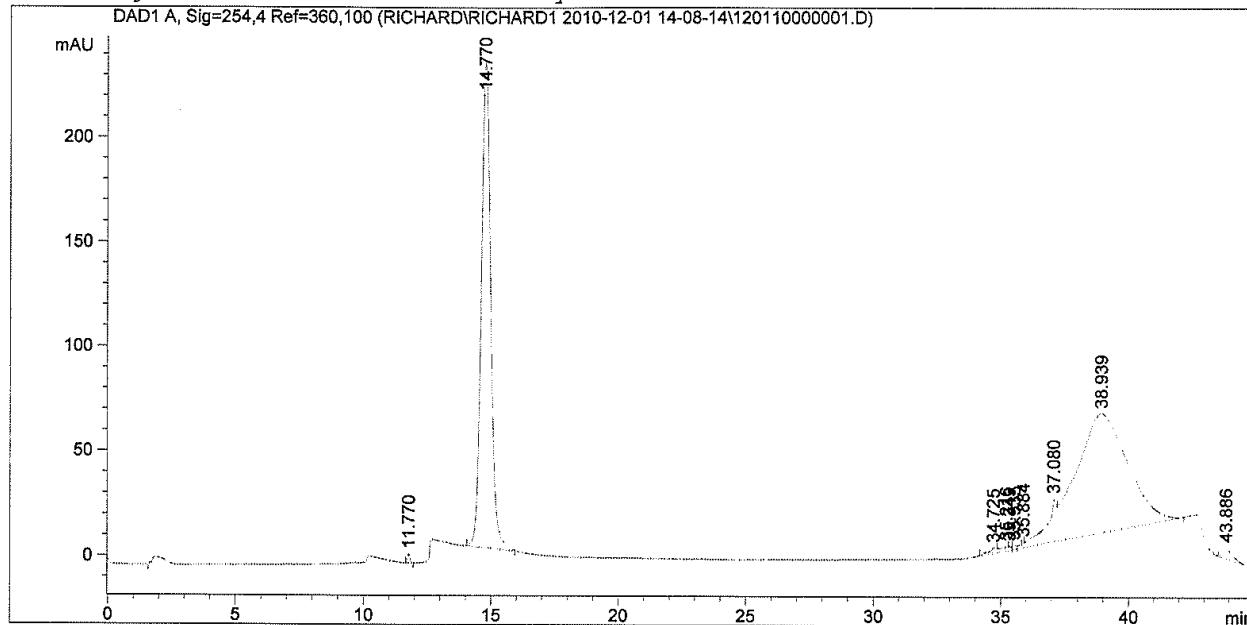


Figure S43 MALDI-TOF mass spectrum of ORN 13

Data File C:\HPCHEM\1\DATA\RICHARD\RICHARD1 2010-12-01 14-08-14\120110000001.D
Sample Name: RT111 miR122 2'OMe PS 4PACE DMToff

=====
Acq. Operator : richard Seq. Line : 1
Acq. Instrument : Instrument 1 Location : Vial 1
Injection Date : 12/1/2010 2:09:56 PM Inj : 1
Inj Volume : 100 μ l
Different Inj Volume from Sequence ! Actual Inj Volume : 10 μ l
Acq. Method : C:\HPCHEM\1\DATA\RICHARD\RICHARD1 2010-12-01 14-08-14\A-50B.M
Last changed : 6/10/2010 10:15:59 AM by richard
Analysis Method : C:\HPCHEM\1\DATA\RICHARD\RICHARD1 2010-12-01 14-08-14\120110000001.D\DA.M
(A-50B.M)
Last changed : 6/10/2010 10:15:59 AM by richard



=====
Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.770	BB	0.0963	27.23801	4.39394	0.1844
2	14.770	BB	0.4025	6070.83984	232.57483	41.1084
3	34.725	BV	0.2649	55.46663	2.62088	0.3756
4	35.216	VV	0.2456	46.60170	2.36790	0.3156
5	35.349	VB	0.0882	12.35856	1.89311	0.0837
6	35.535	BB	0.1236	18.25552	1.97900	0.1236
7	35.884	BV	0.0813	14.92471	2.58642	0.1011
8	37.080	VV	0.3628	606.84460	20.41373	4.1092

Instrument 1 12/1/2010 4:48:19 PM richard

Page 1 of 2

Figure S44 HPLC chromatogram of ORN 13