SUPPORTING INFORMATION

Omegatides: Constrained Analogs Of Peptide Primary Sequence

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A. General Experimental Methods

All reactions were carried out under an inert atmosphere (nitrogen or argon where stated) with dry solvents under anhydrous conditions. Glassware for anhydrous reactions were dried in an oven at 140 °C for minimum 6 h prior to use. Dry solvents were obtained by passing the previously degassed solvents through activated alumina columns. Yields refer to chromatographically and spectroscopically (¹H-NMR) homogeneous materials, unless otherwise stated. Reagents were purchased at a high commercial quality (typically 97 % or higher) and used without further purification, unless otherwise stated. Analytical thin layer chromatography (TLC) was carried out on Merck silica gel plates with QF-254 indicator and visualized by UV, cerric ammonium molybdate, and/or potassium permanganate stains. Flash column chromatography was performed using silica gel 60 (Silicycle, 230-400 mesh). ¹H and ¹³C spectra were recorded on a Varian Mercury or Inova spectrometer (300 MHz ¹H; 75 MHz ¹³C) and were calibrated using residual undeuterated solvent as an internal reference (CDCl₃: ¹H-NMR = 7.26, ¹³C-NMR = 77.16, DMSO-d6: ¹³C-NMR = 39.52, CD₃OD: ¹H-NMR = 3.31, ¹³C-NMR = 49.00) The following abbreviations or combinations thereof were used to explain the multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, p = pentet, br = broad, app = apparent. Melting points were recorded on an automated melting point apparatus (EZ-Melt, Stanford Research Systems) and are uncorrected. Optical rotations were obtained on a Jasco DIP-360 digital polarimeter at the D-line of sodium.

B. Literature procedures used in synthesis

Compounds **1** {a-c} and **5** {a-c} were synthesized using a literature procedure.^{1, 2} (*S*)-5-methylpyrrolidine-2,4-dione (pyrrolinone **A**) was obtained by a known procedure in 90 % yield, and the spectra match (¹H and ¹³C-NMR) the reported spectra.¹ Compounds **2** {a-c} were prepared accordingly original procedure.²

C. General Procedure for X-Ray Structure Determination

A Leica MZ 75 microscope was used to identify a suitable colorless multi-faceted crystal with very well defined faces with dimensions (max, intermediate, and min) 0.05 mm x 0.03 mm x 0.01 mm from a representative sample of crystals of the same habit. The crystal mounted on a nylon loop was then placed in a cold nitrogen stream maintained at 110 K.

A BRUKER D8-GADDS X-ray (three-circle) diffractometer was employed for crystal screening, unit cell determination, and data collection. The goniometer was controlled using the FRAMBO software suite. The sample was optically centered with the aid of a video camera such that no translations were observed as the crystal was rotated through all positions. The detector was set at 6.0 cm from the crystal sample (MWPC Hi-Star Detector, 512x512 pixel). The X-ray radiation employed was generated from a Cu sealed X-ray tube (K_a = 1.54184 Å with a potential of 40 kV and a current of 40 mA) fitted with a graphite monochromator in the parallel mode (175 mm collimator with 0.5 mm monocapillary optics).

The rotation exposure indicated acceptable crystal quality and the unit cell determination was undertaken. 2100 data frames were taken at widths of 0.5° with an exposure time of 10 seconds. Over 6000 reflections were centered and their positions were determined. These reflections were used in the autoindexing procedure to determine the unit cell. A suitable cell was found and refined by nonlinear least squares and Bravais lattice procedures and reported here in Table 1. No super-cell or erroneous reflections were observed. After careful examination of the unit cell, a standard data collection procedure was initiated. This procedure consists of collection of one hemisphere of data collected using omega scans, involving the collection 0.5° frames at fixed angles for ϕ , 2θ , and χ ($2\theta = -28^{\circ}$, $\chi = 54.73^{\circ}$, $2\theta = -90^{\circ}$, $\chi = 54.73^{\circ}$), while varying

APEX2 "Program for Data Collection on Area Detectors" BRUKER AXS Inc., 5465 East Cheryl Parkway, Madison, WI 53711-5373 USA

omega. Addition data frames were collected to complete the data set. Each frame was exposed for 10 sec. The total data collection was performed for duration of approximately 24 hours at 110K. No significant intensity fluctuations of equivalent reflections were observed.

Data Reduction, Structure Solution, and Refinement

Integrated intensity information for each reflection was obtained by reduction of the data frames with the program SAINT.* The integration method employed a three dimensional profiling algorithm and all data were corrected for Lorentz and polarization factors, as well as for crystal decay effects. Finally the data was merged and scaled to produce a suitable data set. The absorption correction program SADABS** was employed to correct the data for absorption effects. Systematic reflection conditions and statistical tests for the data suggested the space group $P2_1$.

A solution was obtained readily using SHELXTL (SHELXS).* All non-hydrogen atoms were refined with anisotropic thermal parameters. The Hydrogen atoms bound to carbon were placed in idealized positions [C–H = 0.96 Å, $U_{iso}(H) = 1.2 \times U_{iso}(C)$]. The structure was refined (weighted least squares refinement on F^2) to convergence. X-seed was employed for the final data presentation and structure plots.**

^{*} SAINT, "Program for Data Reduction from Area Detectors" "BRUKER AXS Inc., 5465 East Cheryl Parkway, Madison, WI 53711-5373 USA

^{**} SADABS, Sheldrick, G.M. "Program for Absorption Correction of Area Detector Frames", BRUKER AXS Inc., 5465 East Cheryl Parkway, Madison, WI 53711-5373 USA

^{*} SHELXTL, Sheldrick, G.M. (2008). Acta Cryst. A64, 112-122

^{**} Barbour, L.J.,(2001) "X-Seed - A software tool for supramolecular crystallography" *J. Supramol. Chem.* **2001**, *1*, 189-191.

Scheme S1

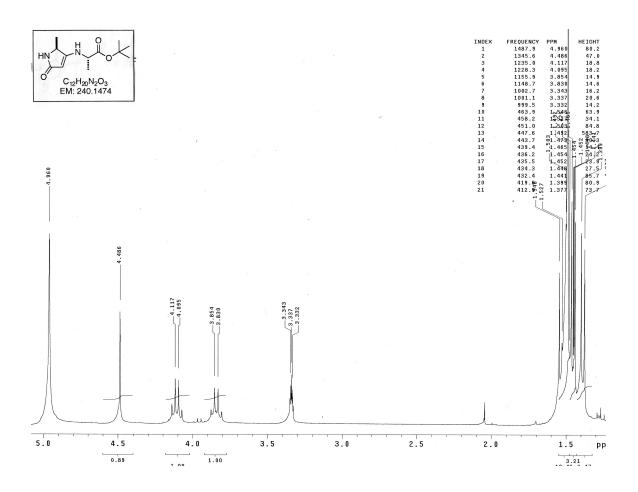
Compound 1a

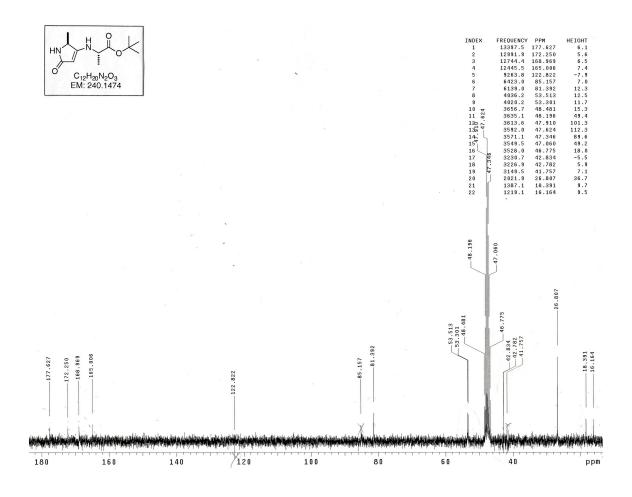
(S)-tert-butyl 2-(((S)-2-methyl-5-oxo-2,5-dihydro-1H-pyrrol-3-yl)amino)propanoate

$$HN$$
 HN
 $CO_2^{\dagger}Bu$

Physical state: solid, 60% mp = 158 - 160 °C, cryst. dichloromethane/hexanes. ¹H-NMR (300 MHz, CD₃OD): 4.49 (s, 1H), 4.11 (q, J = 6.6 Hz, 1H), 3.84 (q, J = 7.2 Hz, 1H), 1.49 (s, 9H), 1.45 (d, J = 7.2 Hz, 3H), 1.39 (d, J = 6.0 Hz, 3H) ¹³C-NMR (75 MHz, CD₃OD): 177.6, 172.3, 169.0, 165.0, 81.4, 53.5, 53.3, 26.8, 18.4, 16.2

MS: (ESI, m/z): calculated for $C_{12}H_{20}N_2O_3$ 240.15, found 241.18 (H⁺)





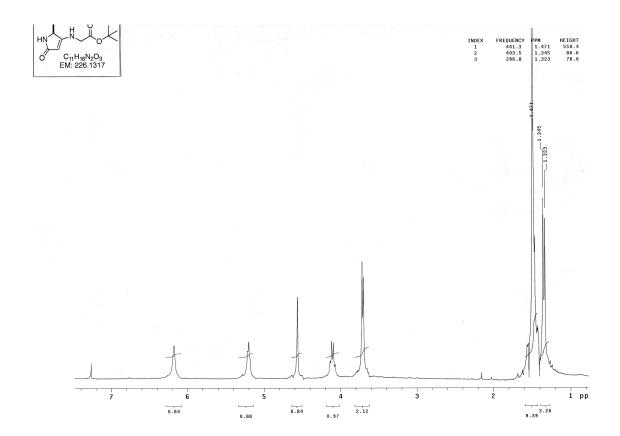
Compound 1b

(S)-tert-butyl 2-((2-methyl-5-oxo-2,5-dihydro-1H-pyrrol-3-yl)amino)acetate

Physical state: oil, 81 %.

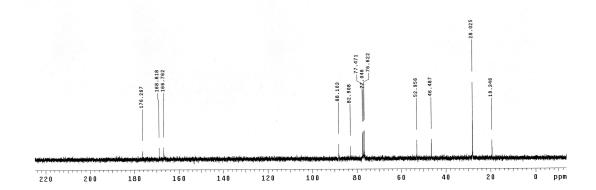
¹H-NMR (300 MHz, CDCl₃): 6.18 (br s, 1H), 5.21 (br s, 1H), 4.50 (s, 1H), 4.16 (q, J = 6.6 Hz, 1H), 3.80 (s, 2H), 1.51 (s, 9H), 1.37 (d, J = 6.9 Hz, 3H) (75 MHz, CDCl₃): 177.3, 168.8, 166.8, 88.1, 82.9, 52.9, 48.5, 28.0, 19.3

MS: (ESI, m/z): calculated for $C_{11}H_{18}N_2O_3$ 226.13, found 227.14 (H⁺)

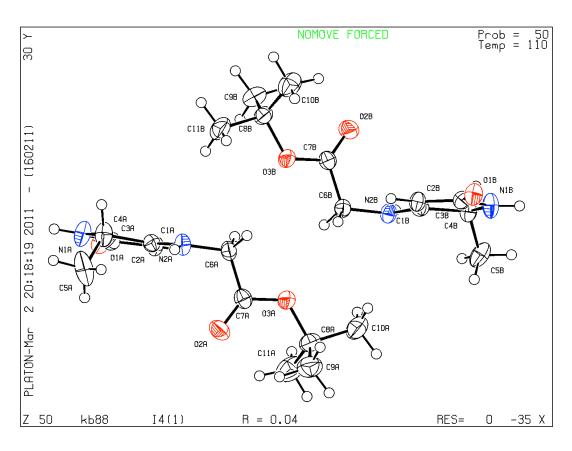








X-Ray 1b



Compound 1c

((S)-tert-butyl 4-methyl-2-(((S)-2-methyl-5-oxo-2,5-dihydro-1 H-pyrrol-3-yl)amino)pentanoate

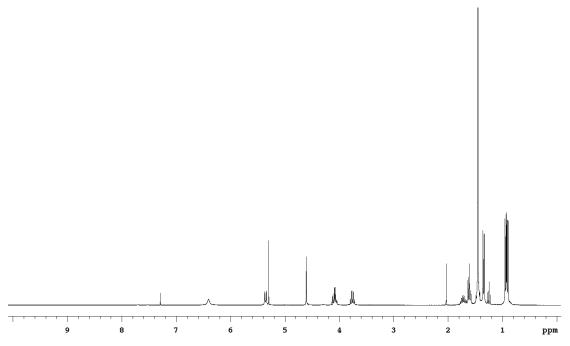
Physical state: White needles (ethyl acetate), mp = 206 °C (decomposes), 60%. $\left[\alpha\right]^{20}_{D} - 93.1$ (*c* 1.0, MeOH)

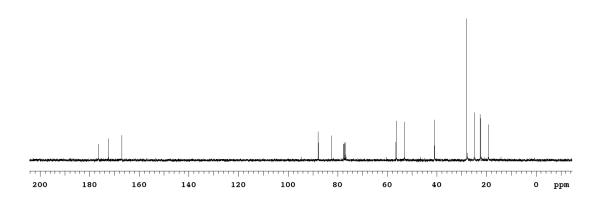
¹H-NMR (300 MHz, CDCl₃) ∂ 6.40 (br s, 1H), 5.36 (d, J =8.1 Hz, 1H), 4.60 (s, 1H), 4.09 (m, 1H), 3.76 (q, J = 7.5 Hz, 1H), 1.72 (m, 1H), 1.61 (t, J = 6.7 Hz, 2H), 1.45 (s, 9H), 1.35 (d, J = 6.9 Hz, 3H), 0.94 (d, J = 6.6 Hz, 3H), 0.91 (d, J = 6.6 Hz, 3H)

¹³C-NMR (75 MHz, CDCl₃) ∂ 176.5, 172.5, 166.9, 87.8, 82.3, 56.4, 53.1, 40.9, 28.0, 24.9, 22.6, 22.3, 19.2

IR (film, cm⁻¹) 3217 (br), 3048, 2959, 2870, 1744, 1645, 1601, 1557, 1368, 1207, 1146, 845, 783, 704

MS (ESI) m/z calcd for $(M+H)^{+}$ $C_{15}H_{27}N_{2}O_{3}$ 283.19; found 283.19





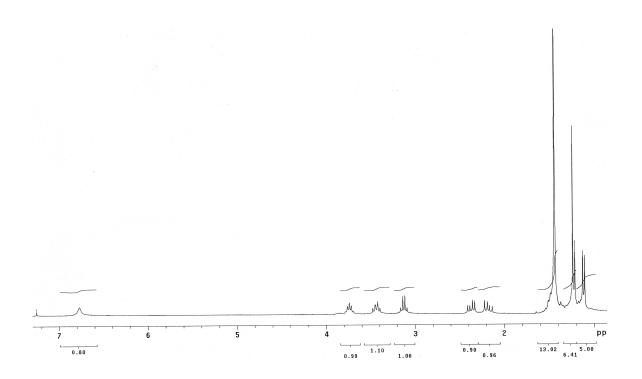
Compound 2a

(S)-tert-butyl 2-(((2S, 3S)-2-methyl-5-oxopyrrolidin-3-yl) amino) propanoate

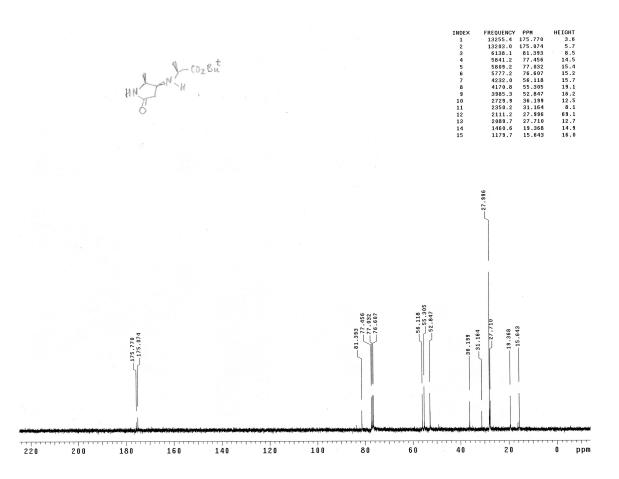
 1 H-NMR (300 MHz, CDCl₃): 6.77 (bs, 1H), 3.80 – 3.65 (m, 1H), 3.55 – 3.35 (m, 1H), 3.13 (q, J = 7.2 Hz, 1H), 2.37 (dd, J = 16.2, J = 7.8 Hz, 1H), 2.18 (dd, J = 16.4, J = 9.5 Hz, 1H), 1.44 (s, 9H), 1.22 (d, J = 6.9 Hz, 3H), 1.11 (d, J = 6.3 Hz, 3H)

¹³C-NMR (75 MHz, CDCl₃): 175.8, 175.1, 81.4, 56.1, 52.8, 36.2, 28.0, 27.7, 19.4, 15.6

MS: (ESI, m/z): calculated for $C_{12}H_{22}N_2O_3$ 242.16, found 243.17 (H+)







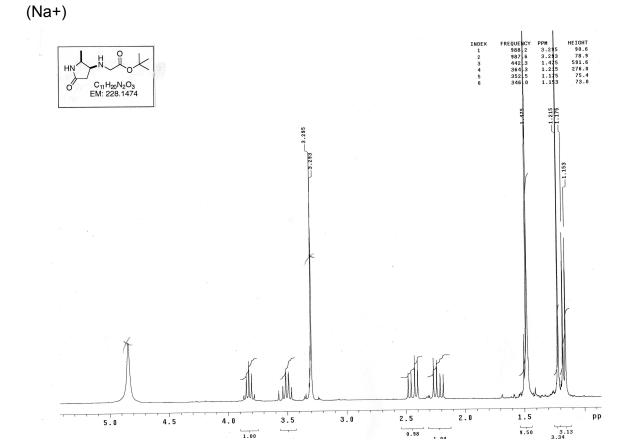
Compound 2b

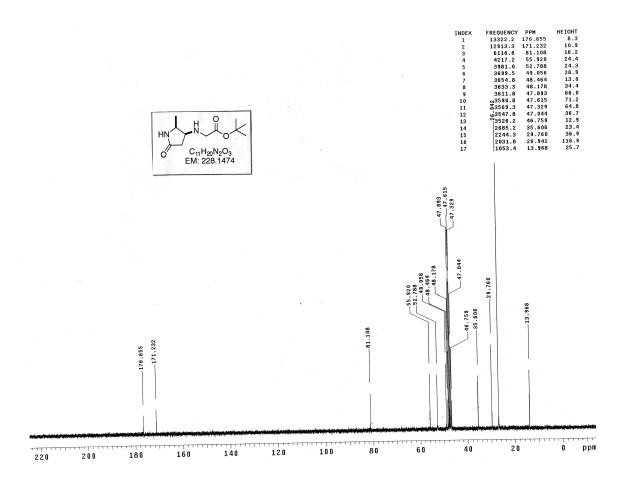
tert-butyl 2-(((2S,3S)-2-methyl-5-oxopyrrolidin-3-yl)amino)acetate

$$HN$$
 CO_2 tBU

¹H-NMR (300 MHz, CDCl₃): 4.84 (s, 1H), 3.88-3.75 (m, 1H), 3.46 – 3.40 (m, 1H), 3.29 (s, 2H), 2.32 (dd, J = 16.5, J = 7.5 Hz, 1H), 2.09 (dd, J = 16.5, J = 7.5 Hz, 1H), 1.35 (s, 9H), 1.05 (d, J = 6.3 Hz, 3H)

¹³C-NMR (75 MHz, CD₃OD): 176.7, 171.2, 81.1, 56.0, 52.8, 35.6, 29.8, 27.0, 14.0 MS: (ESI, m/z): calculated for C₁₁H₂₀N₂O₃ 228.15, found 229.15 (H⁺), 251.11





Compound 2c

(S)-tert-butyl 4-methyl-2-(((2S,3S)-2-methyl-5-oxopyrrolidin-3-yl)amino)pentanoate

 $[\alpha]^{20}$ -10.0 (c 1.0, MeOH)

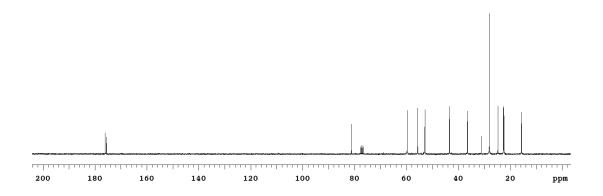
¹H-NMR (300 MHz, CDCl₃) ∂ 7.20 (br s, 1H), 3.70 (m, 1H), 3.35 (m, 1H), 3.01 (app t, J = 7.3 Hz, 1H), 2.32 (dd, J = 16.4, 7.7 Hz, 1H), 2.14 (dd, J = 16.4, 9.6 Hz, 1H), 1.72 (m, 1H), 1.41 (s, 9H), 1.34 (m, 2H), 1.08 (d, J = 6.3 Hz, 3H), 0.85 (d, J = 6.6 Hz, 3H)

¹³C-NMR (75 MHz, CDCl₃) ∂ 176.1, 175.6, 81.2, 59.7, 55.7, 52.9, 43.4, 36.5, 28.0, 24.7, 22.6, 22.5, 15.7

IR (film, cm⁻¹) 3273 (br), 2960, 2932, 1732, 1697, 1682, 1472, 1456, 1435, 1368, 1152, 941, 849

MS (ESI) m/z calculated for $(M+H)^{+}$ $C_{15}H_{29}N_{2}O_{3}$ 285.21; found 285.19

10



2

ppm

General procedure to prepare compound 3

A mixture of enamine **2a** (4 mmol), (triphenylphosphoranyllidene)ketene (8 mmol) and TFA (2.6 mmol) in 5 mL dioxane was heated at 105 – 110°C for 1 h. Upon completion (analysis by NMR), the solvent was removed at reduced pressure, Purification was carried out by flash column chromatography (eluent ethylacetate/methanol).

Note: 3 a-c products are hygroscopic solids and soluble in the water. Isolation could be done by simple extraction of the crude mixture with warm water (50 $^{\circ}$ C), followed by extraction of the impurity triphenylphosphoxide with ether. The water layer contains pure product and triphenylmethylphosphonium ylide as a salt with TFA. Further separation could be done on a SiO₂ column eluting with ethylacetate – methanol mixture 20:1 – 10:1.

Compound 3a

(S)-4-(tert-butoxy)-5-methyl-1-((2S,3S)-2-methyl-5-oxopyrrolidin-3-yl)-1H-pyrrol-2(5H)-one

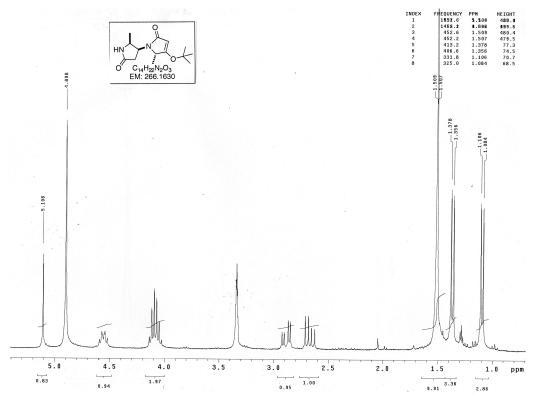
Hygroscopic white solid, mp = 174 - 178 °C, 63%.

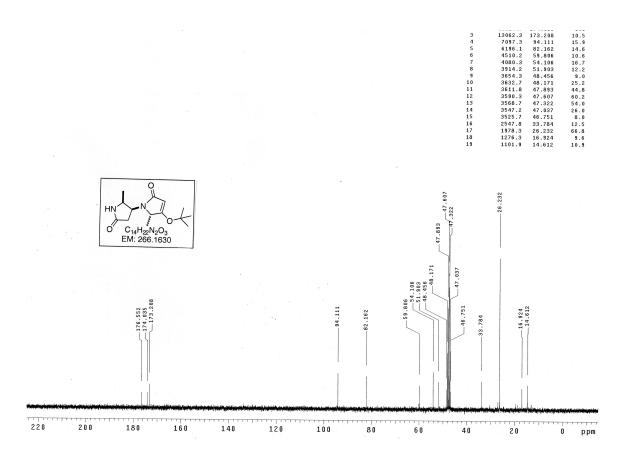
 $[\alpha]^{20}$ _D - 79.7 (*c* 1.0, MeOH)

 1 H-NMR (CD₃OD): 5.11 (s, 1H), 4.60 – 4.45 (m, 1H), 4.16 – 3.96 (m, 3H), 2.89 (dd, J = 16.8, J = 6.0 Hz, 1H), 2.67 (dd, J = 17.1, J = 9.0 Hz, 1H), 1.51 (s, 9H), 1.37 (d, J = 6.6 Hz, 3H), 1.10 (d, J = 6.6 Hz, 3H)

¹³C-NMR (75 MHz, CD₃OD): 176.6, 174.0, 173.2, 94.1, 82.2, 59.8, 54.1, 51.9, 33.8, 26.2, 16.9, 14.6

MS: (ESI, m/z): calculated for $C_{14}H_{22}N_2O_3$ 266.16, found 267.15 (H+), 289.13 (Na+)





Compound 3b

4-(tert-butoxy)-1-((2S,3S)-2-methyl-5-oxopyrrolidin-3-yl)-1H-pyrrol-2(5H)-one

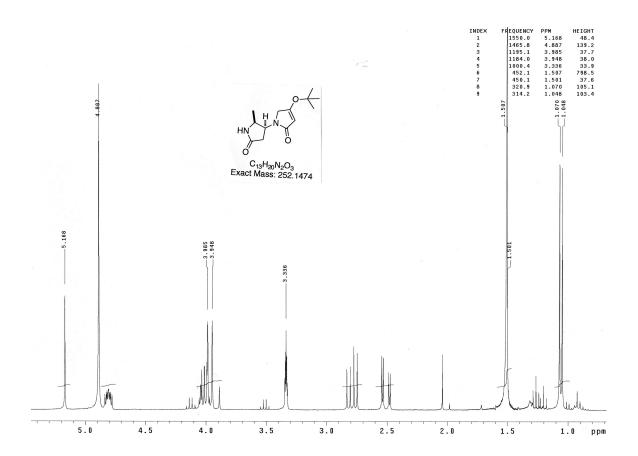
Physical state: Hygroscopic solid, mp = 75 - 80 °C, 80%.

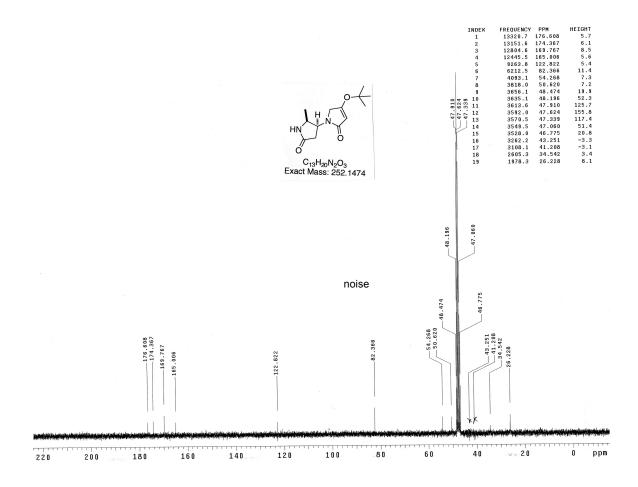
 $[\alpha]^{20}$ _D – 55.6 (*c* 1.0, MeOH)

 1 H-NMR (CD₃OD): 4.85 – 4.76 (m, 1H), 4.08 – 3.88 (m, 3H), 2.79 (dd, J = 17.2, J = 8.2Hz, 1H), 2.51 (dd, J = 17.2, J = 3.8 Hz, 1H), 1.51 (s, 9H), 1.06 (d, J = 6.6 Hz, 3H)

¹³C-NMR (75 MHz, CD₃OD): 176.6, 174.4, 169.8, 165.0, 122.8, 82.4, 54.3, 50.6, 43.3, 41.2

MS: (ESI, m/z): calculated for $C_{13}H_{20}N_2O_3$ 252.15, found 253.18 (H⁺)





Compound 3c

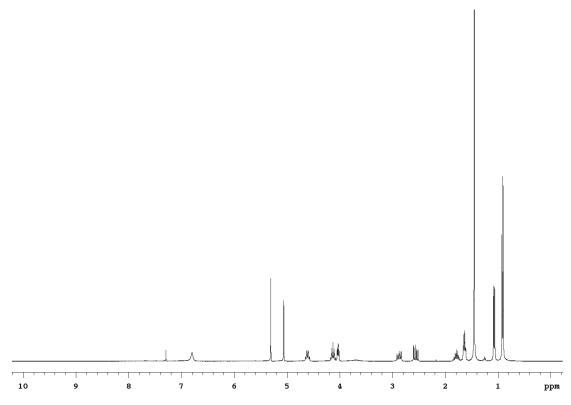
(S)-4-(tert-butoxy)-5-isobutyl-1-((2S,3S)-2-methyl-5-oxopyrrolidin-3-yl)-1H-pyrrol-2(5H)-one

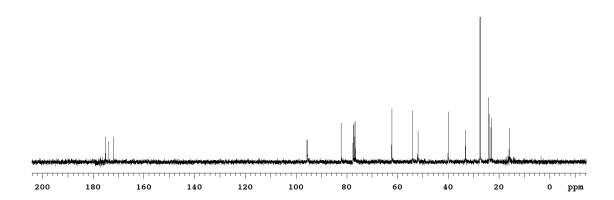
White crystals, 60 %

 $[\alpha]^{20}_{D}$ – 61.0 (*c* 1.0, MeOH)

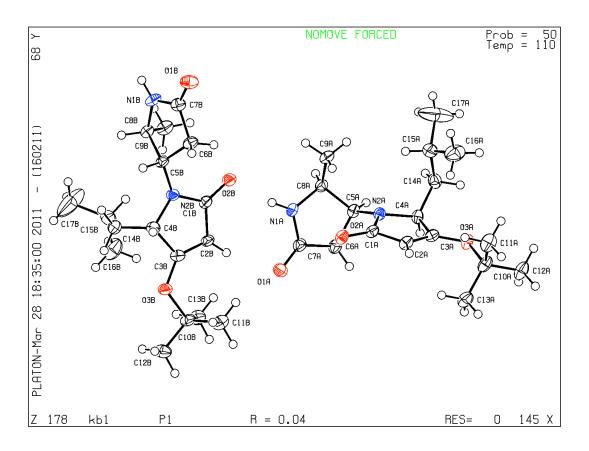
¹H-NMR (CDCl₃): 6.80 (br s, 1H), 5.06 (s, 1H), 4.61 (q, J = 7.6 Hz, 1H), 4.13 (p, J = 6.7 Hz, 1H), 4.03 (t, J = 5.1 Hz, 1H), 2.88 (dd, J = 16.7, 8.1 Hz, 1H), 2.56 (dd, J = 16.5, 8.7 Hz, 1H), 1.87-1.70 (m, 1H), 1.68-1.59 (m, 2H), 1.45 (s, 9H), 1.07 (d, J = 6.6 Hz, 3H), 0.91 (app d, J = 6.6 Hz, 6H)

¹³C-NMR (CDCl₃): 175.2, 173.8, 172.0, 95.6, 82.2, 62.2, 54.1, 51.9, 40.0, 33.2, 27.5, 24.0, 23.9, 23.1, 15.9





X-Ray 3c



General procedure for compounds 4 a-c

Compounds **4** were prepared by acid elimination of *tert*-butyl group from **3 a-c** in a mixture 2:2:0.1 of CH₂Cl₂/TFA/Et₃SiH at room temperature for 3 h in quantitative yields. These are hygroscopic white solids unstable during storage at room temperature. NMR revealed two or more conformers. The products were immediately carried to the next step.

Compound 4a

(S)-4-hydroxy-5-methyl-1-((2S,3S)-2-methyl-5-oxopyrrolidin-3-yl)-1H-pyrrol-2(5H)-one

¹H-NMR (CD₃OD) two forms: 4.60 - 4.50 (m, 0.57H), 4.50 - 4.35 (m, 0.43H), 4.20 - 3.90 (m, 2H), 3.48 (q, J = 6.9 Hz, 1H), 3.02 - 2.75 (m, 1H), 2.74 - 2.55 (m, 1H), 1.37 (d, J = 6.6 Hz, 3H), 1.23 - 1.10 (d, J = 6.6 Hz, 3H), 1.06 (m, 6H) ¹³C-NMR (75 MHz, CD₃OD) two forms: 177.6, 177.1, 176.7, 174.8, 65.6, 65.5, 58.6, 54.2, 53.5, 53.2, 51.9, 34.0, 33.7, 26.2, 16.6, 15.8, 15.0, 14.5 MS: (ESI, m/z): calculated for C₁₀H₁₄N₂O₃ 210.10, found 211.11 (H+)

Compound 4b

4-hydroxy-1-((2S,3S)-2-methyl-5-oxopyrrolidin-3-yl)-1H-pyrrol-2(5H)-one

 1 H-NMR (CD₃OD) two forms: 4.85 – 4.70 (m, 1H), 4.20 – 3.80 (m, 2H), 3.65 – 3.40 (m, 1H), 2.80 – 2.65 (m, 1H), 2.60 – 2.30 (m, 1H), 1.30 – 0.95 (m, 3H)

¹³C-NMR (75 MHz, CD₃OD) two forms: 203.1, 176.6, 173.4, 98.8, 56.3, 56.0, 55.4, 54.3, 54.2, 53.9, 53.7, 53.5, 50.5, 50.3, 50.1, 49.2, 34.6, 34.1, 33.9, 33.7, 13.7, 13.6, 13.4, 13.4

MS: (ESI, m/z): calculated for $C_9H_{12}N_2O_3$ 196.08, found 197.09 (H+)

Compound 4c

(S)-4-hydroxy-5-isobutyl-1-((2S,3S)-2-methyl-5-oxopyrrolidin-3-yl)-1H-pyrrol-2(5H)-one

¹H-NMR (CD₃OD) two forms: 4.50 (q, J = 7.8 Hz, 1H), 4.36 (q, J = 7.3 Hz, 1H), 4.19 (t, J = 10.3 Hz, 1H), 4.15 – 4.00 (m, 3H), 3.00 – 2.75 (m, 2H), 2.75 – 2.55 (m, 2H), 2.00 – 1.75 (m, 2H), 1.75 – 1.60 (m, 3H), 1.60 -1.50 (m, 1H), 1.18 (d, J = 6.6 Hz, 3H), 1.33 (d, J = 6.6 Hz, 3H), 1.07 (d, J = 6.3 Hz, 3H), 1.02 – 0.90 (m, 12 H)

¹³C-NMR (75 MHz, CD₃OD) two forms: 177.2, 176.4, 176.1, 175.4, 171.2, 67.7, 61.5, 54.2, 53.5, 53.4, 52.2, 40.4, 39.3, 33.6, 33.5, 26.2, 23.9, 23.6, 22.7, 22.4, 21.8, 20.5, 15.2, 14.6

MS: (ESI, m/z): calculated for $C_{13}H_{20}N_2O_3$ 252.1474, found 253.1559 (H+), 275,1333 (Na+)

Compound 5aa

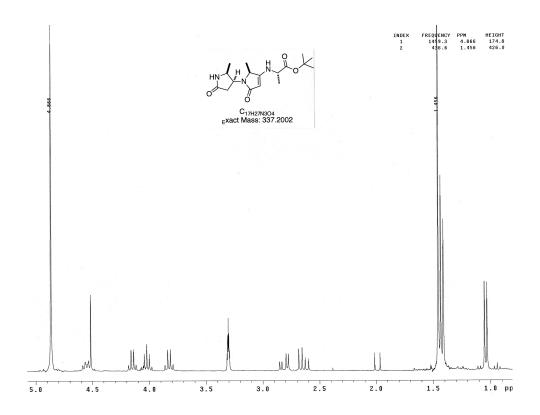
(S)-tert-butyl 2-(((S)-2-methyl-1-((2S,3S)-2-methyl-5-oxopyrrolidin-3-yl)-5-oxo-2,5-dihydro-1H-pyrrol-3-yl)amino)propanoate

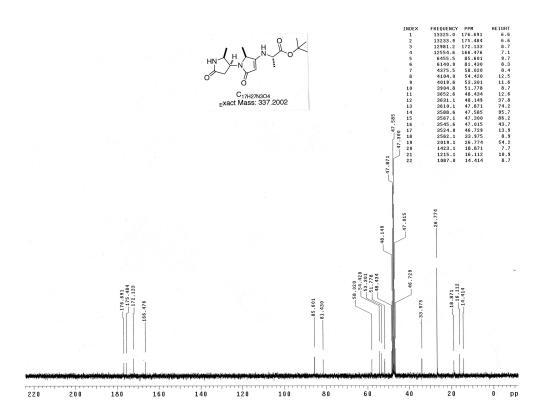
Procedure: As described before for **1 a-c**. White solid, mp = 199 - 200 °C (dichloromethane/hexanes), 50%.

 1 H-NMR (CD₃OD): 4.60 – 4.50 (m, 1H), 4.52 (s, 1H), 4.15 (q, J = 6.6 Hz, 1H), 4.10 – 3.96 (m, 1H), 3.83 (q, J = 7.2 Hz, 1H), 2.81 (dd, J = 16.8, J = 6.0 Hz, 1H), 2.64 (dd, J = 17.0, J = 8.8 Hz, 1H), 1.46 (s, 9H), 1.42 (d, J = 6.6 Hz, 6H), 1.04 (d, J = 6.6 Hz, 3H)

¹³C-NMR (75 MHz, CD₃OD): 176.7, 175.5, 172.1, 166.5, 85.6, 81.4, 58.0, 54.4, 53.3, 51.8, 34.0, 26.8, 18.9, 16.1, 14.4

MS: (ESI, m/z): calculated for $C_{17}H_{27}N_3O_4$ 337.20, found 338.23 (H+)





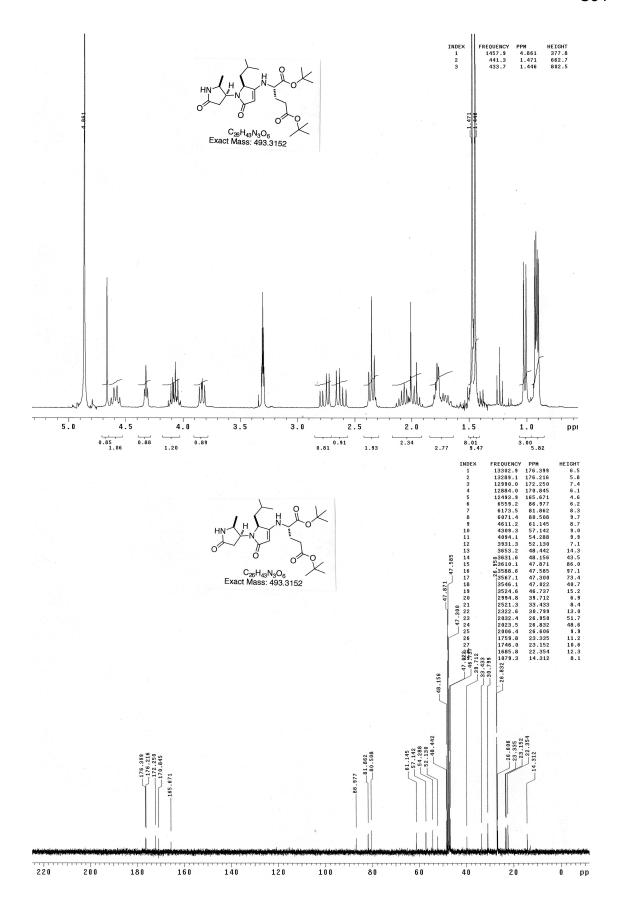
Compound 5cd

(S)-di-tert-butyl 2-(((S)-2-isobutyl-1-((2S,3S)-2-methyl-5-oxopyrrolidin-3-yl)-5-oxo-2,5-dihydro-1H-pyrrol-3-yl)amino)pentanedioate

Procedure: As described before for **1 a-c**. White solid, mp = 51 - 55 °C, eluent ethylacetate/methanol 10:1 - 5:1, 40%.

¹H-NMR (CD₃OD): 4.66 (s, 1H), 4.64 – 4.54 (m, 1H), 4.32 (t, J = 4.0 Hz, 2H), 4.12 – 4.01 (m, 1H), 3.83 (dd, J = 8.1, J = 6.0 Hz, 1H), 2.76 (dd, J = 16.5, J = 7.2 Hz, 1H), 2.62 (dd, J = 16.8, J = 8.7 Hz, 1H), 2.40 – 2.30 (m, 2H), 2.15 – 1.90 (m, 2H), 1.83 – 1.65 (m, 3H), 1.47 (s, 9H), 1.45 (s, 9H), 1.01 (d, J = 6.6 Hz, 3H), 0.92 (d, J = 3.3 Hz, 3H), 0.90 (d, J = 3.3 Hz, 3H) 13 C-NMR (75 MHz, CD₃OD): 176.4, 176.2, 172.3, 170.8, 165.7, 87.0, 81.9, 80.5,

¹³C-NMR (75 MHz, CD₃OD): 176.4, 176.2, 172.3, 170.8, 165.7, 87.0, 81.9, 80.5 61.1, 57.1, 54.3, 52.1, 39.7, 33.4, 30.8, 27.0, 26.8, 26.6, 23.3, 23.2, 22.4, 14.3 MS: (ESI, m/z): calculated for C₂₆H₄₃N₃O₆, 493.31, found 494.32 (H+)



Compound 6aa

(2'S,3'S,4S,5S)-4-((S)-3-(tert-butoxy)-2-methyl-5-oxo-2,5-dihydro-1H-pyrrol-1-yl)-2',5-dimethyl-[1,3'-bipyrrolidine]-2,5'-dione

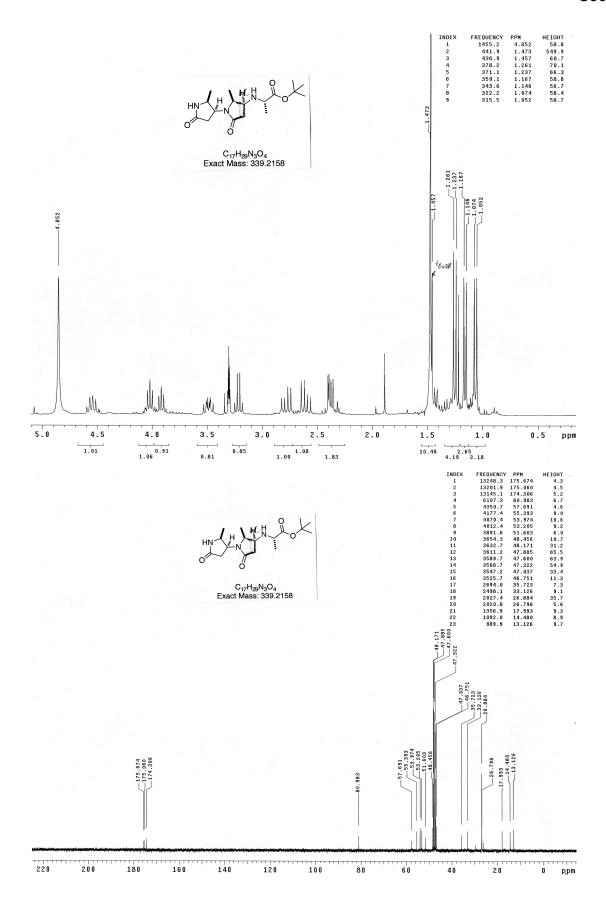
Procedure: As described before for 2 a-c.

Physical state: sticky oil, yield quantitative.

 1 H-NMR (CD₃OD): 4.60 – 4.48 (m, 1H), 4.08 – 3.97 (m, 1H), 3.96 – 3.87 (m, 1H), 3.54 – 3.42 (m, 1H), 3.22 (q, J = 7.0 Hz, 1H), 2.79 (dd, J = 16.5, J = 8.1 Hz, 1H), 2.61 (dd, J = 16.8, J = 8.7 Hz, 1H), 2.48 – 2.30 (m, 2H), 1.47 (s, 9H), 1.25 (d, J = 7.2 Hz, 3H), 1.04 – 0.95 (m, 6H)

¹³C-NMR (75 MHz, CD₃OD): 175.7, 175.1, 174.3, 81.0, 57.7, 55.4, 54.0, 53.2, 51.6, 35.7, 33.1, 26.9, 26.8, 18.0, 14.5, 13.1

MS: (ESI, m/z): calculated for $C_{17}H_{27}N_3O_4$ 339.22, found 340.22 (H+)



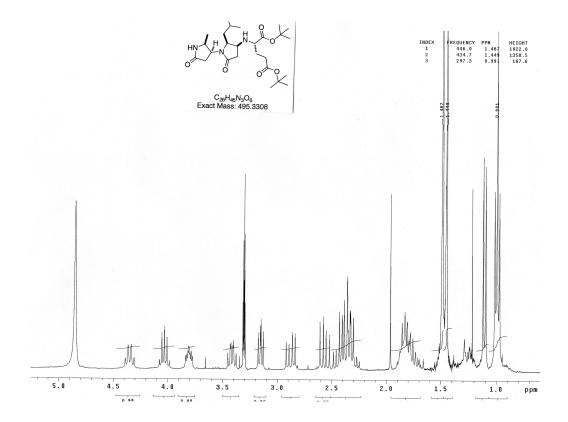
Compound 6cd

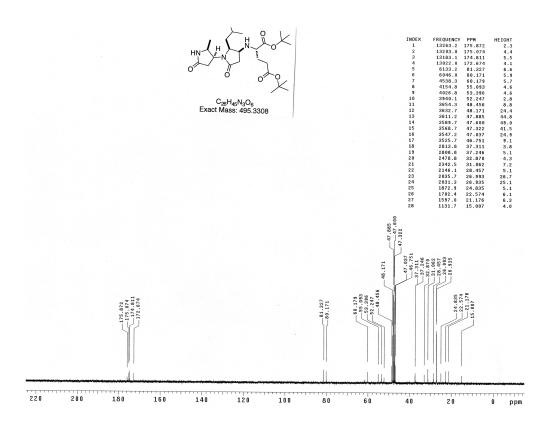
(*S*)-di-*tert*-butyl 2-(((2*S*,2'*S*,3*S*,3'*S*)-2-isobutyl-2'-methyl-5,5'-dioxo-[1,3'-bipyrrolidin]-3-yl)amino)pentanedioate, oil, yield quantitative.

Procedure: Similar to that described before for **2 a-c**, except that hydrogenation was performed at elevated hydrogen pressure (70 bar) and temperature (70 °C) for 30 h with catalytic acetic acid.

¹H-NMR (CD₃OD): 4.35 (q, J = 9.0 Hz, 1H), 4.10 – 3.96 (m, 1H), 3.86 -3.75 (m, 1H), 3.47 – 3.35 (m, 1H), 3.15 (dd, J = 7.5, J = 6.0 Hz, 1H), 2.88 (dd, J = 16.8, J = 7.5 Hz, 1H), 2.56 (dd, J = 16.8, J = 9.3 Hz, 1H), 2.50 – 2.23 (m, 4H), 1.90 – 1.70 (m, 4H), 1.49 (s, 9H), 1.45 (s, 9H), 1.11 (d, J = 6.6 Hz, 3H), 0.92 (d, J = 3.3 Hz, 3H), 0.90 (d, J = 3.3 Hz, 3H)

¹³C-NMR (75 MHz, CD₃OD): 175.9, 175.1, 174.8, 172.7, 81.3, 80.2, 60.2, 55.1, 53.4, 52.2, 37.3, 37.2, 32.9, 31.1, 28.5, 27.0, 26.9, 24.8, 22.6, 21.2, 15.0 MS: (ESI, m/z): calculated for C₂₆H₄₅N₃O₆, 495.33, found 496.34 (H+)





Compound 7aa

(2'S,3'S,4S,5S)-4-((S)-3-(tert-butoxy)-2-methyl-5-oxo-2,5-dihydro-1H-pyrrol-1-yl)-2',5-dimethyl-[1,3'-bipyrrolidine]-2,5'-dione

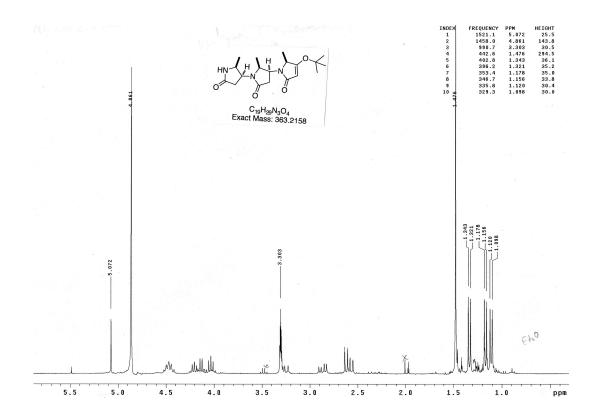
Procedure: As described for **3 a-c** preparation. White crystals, mp = 229 – 230 °C (chloroform/hexanes), yield 80%.

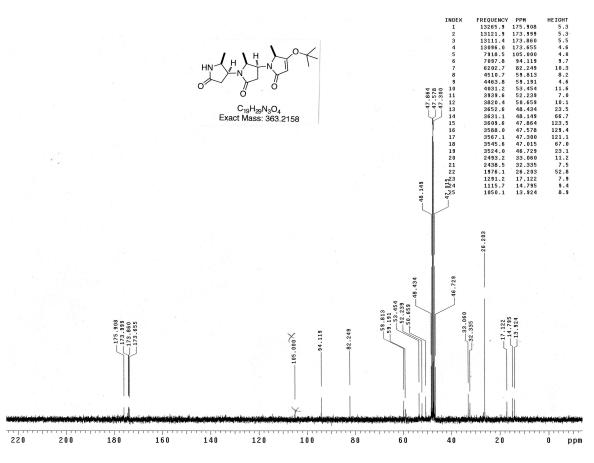
 $[\alpha]^{20}$ _D - 115.1 (*c* 1.0, MeOH)

¹H-NMR (CD₃OD): 5.07 (s, 1H), 4.53 - 4.40 (m, 2H), 4.26 - 3.98 (m, 3H), 3.27 (dd, J = 16.5, J = 11.1 Hz, 1 H), 2.87 (dd, J = 16.8, J = 7.2 Hz, 1H), 2.62 (d, J = 8.7 Hz, 1H), 2.57 (dd, J = 9.0, J = 0.9 Hz, 1H), 1.48 (s, 9H), 1.33 (d, J = 6.6 Hz, 3H), 1.17 (d, J = 6.6 Hz, 3H), 1.11 (d, J = 6.6 Hz, 3H)

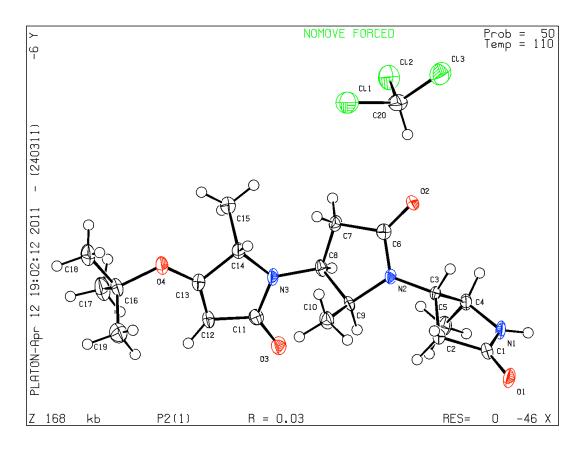
¹³C-NMR (75 MHz, CD₃OD): 176.0, 174.0, 173.9, 173.0, 94.1, 82.2, 59.8, 59.2, 53.5, 52.2, 50.7, 33.1, 32.3, 26.2, 17.1, 14.8, 13.9

MS: (ESI, m/z): calculated for $C_{19}H_{29}N_3O_4$ (+Na) exact mass 386.2061 (Na+), found 386.2056





X-Ray 7aa



Scheme S3

Compound 9

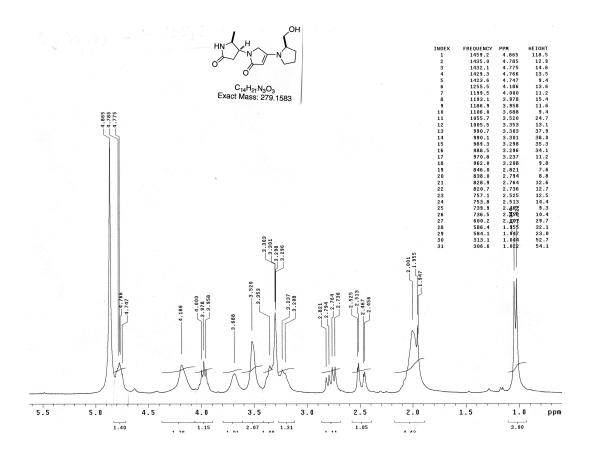
4-((R)-2-(Hydroxymethyl)pyrrolidin-1-yl)-1-((2S,3S)-2-methyl-5-oxopyrrolidin-3-yl)-1H-pyrrol-2(5H)-one

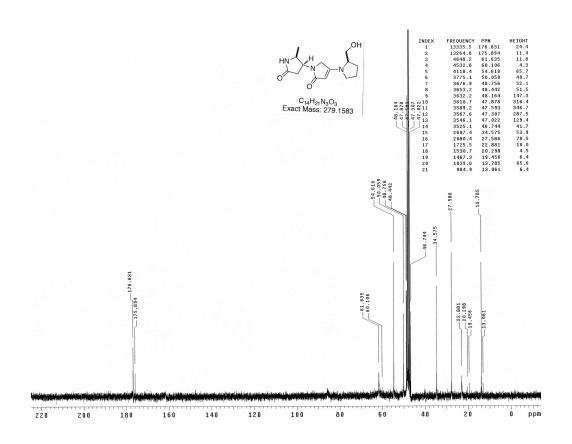
Procedure: As described for **1 a-c**, except that no sodium acetate was added. White sticky oil, 40%, was eluted with ethylacetate/methanol 5:1 - 5:4 mixtures on SiO_2 .

 $[\alpha]^{20}$ _D - 189.40 ± 1.14 (*c* 1.0, MeOH)

¹H-NMR (CD₃OD): 4.85 - 4.70 (m, 2H), 4.19 (bs, 2H), 4.05 - 3.90 (m, 1H), 3.69 (bs, 1H), 3.52 (bs, 2H), 3.40 - 3.20 (m, 3H), 2.78 (dd, J = 17.3, J = 8.2 Hz, 1H), 2.49 (dd, J = 17.3, J = 3.4 Hz, 1H), 2.00 (bs, 3H), 1.03 (d, J = 6.6 Hz, 3H) ¹³C-NMR (75 MHz, CD₃OD): 176.9, 175.9, 61.6, 54.6, 50.1, 48.8, 34.6, 27.6, 22.9, 20.0, 19.5, 13.8, 13.1

MS: (ESI, m/z): calculated for $C_{14}H_{21}N_3O_3$ exact mass: 279.1583, found 280.1753 (H+)

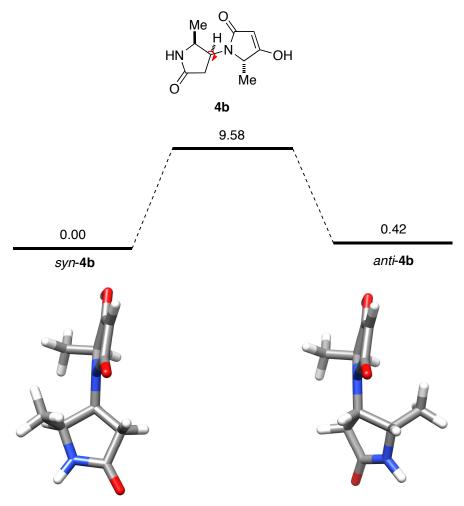




D. Modeling Procedures

DFT Calculation

Reaction path calculations were performed at the B3LYP level of theory with the 6-31G(d') basis set, and a polarized continuum solvation model with a dielectric of H₂O (ϵ =78.3553). All B3LYP calculations were performed using Gaussian 03³. The energy barriers for the compound **4b** were calculated as rotation of the bond (red arrows). ΔG^{o} values were shown in kcal/mol. The population ratio of two conformers is *syn-4b*: *anti-4b* = 1.00: 0.48.



Quenched Molecular Dynamics (QMD)

NAMD⁴ was used for the molecular simulations performed in this work. Explicit atom representations were used throughout the study. The protein structure files (PSF) for all the peptidomimetics were built using Discovery Studio 2.5 (Accelrys Inc) using the CHARMm force field. ⁵

Quenched molecular dynamics simulations were performed using the CHARMm force field as implemented in Discovery Studio 2.5. All four molecules were modeled as neutral compounds in a dielectric continuum of 80 (simulating H_2O). Thus, the starting conformers were minimized using 3000 steps of conjugate gradient. The minimized structures were then subjected to heating, equilibration, and dynamics simulation. Throughout, the equations of motions were integrated using the Verlet algorithm with a time step 1 fs. Each peptidomimetic was heated to 1000 K over 10 ps and equilibrated for another 10 ps at 1000 K, then molecular dynamics runs were performed for a total time of 600 ps with trajectories saved every 1 ps. The resulting 600 structures were thoroughly minimized using 1000 steps of SD followed by 3000 steps of conjugate gradient. Structures with energies less than 3.0 kcal mol^{-1} relative to the global minimum were selected for further analysis.

The VMD⁶ package was used to display, overlay, and classify the selected structures into conformational groups. The best clustering was obtained using a grouping method based on calculation of RMS deviation of a subset of atoms, in this study these were the C^{α} - and C^{β} - atoms. Thus, threshold cutoff values 0.3 Å were selected to obtain families with reasonable homogeneity. The lowest energy conformation from each family was considered to be a typical representative of the family as a whole.

Procedure For Overlays

After minimization in the QMD process, the conformers were grouped into families base on their $C\alpha$ - $C\beta$ coordinates. The process of systematically matching preferred conformers with secondary structures was performed in the

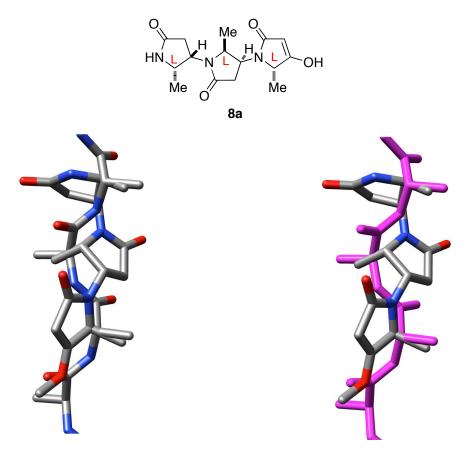
S45

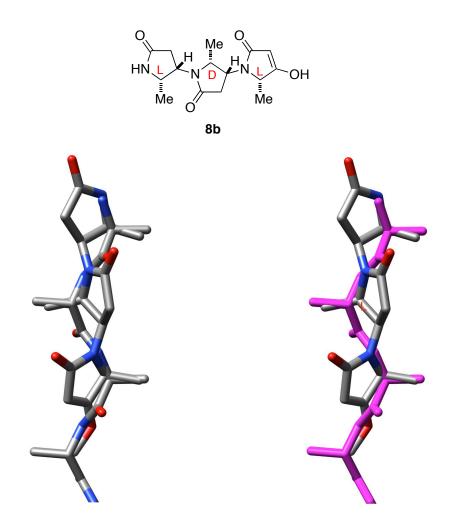
following way. All the conformers within 3.0 kcal/mol were considered to be "preferred". Each of these was overlaid on ideal secondary structures using an in house generated algorithm that compared $C\alpha$ - $C\beta$ coordinates of the side chains which generates a list of structures ranked in terms of the RMSD for the overlay process. The conformers within 3.0 kcal/mol were each overlaid on the ideal secondary structures.

Templates for Secondary Structures

Standard template for overlays with β -strand was obtained from Discovery Studio 2.5.⁵

Overlays of compound 8 conformers with different stereochemistry on the $\beta\text{-strand}$





Procedure for Ramanchandran Plot

Definition of torsional angles: ϕ (H¹-N²-C³-C⁴), ψ (N²-C³-C⁴-N⁵), ϕ ' (C⁴-N⁵-C⁶-C⁷) and ψ ' (N⁵-C⁶-C⁷-O⁸)

For each compound, 600 conformers within 3.0 kcal/mol were obtained by QMD. Ramanchandran plot for the conformers was obtained from UCSF Chimera 1.5.2. www.cgl.ucsf.edu/chimera/

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