Supplementary Information

- Biological screening information
- ¹H and ¹³ C NMR of most active LasR-antagonists

Bacterial strains and growth conditions

The *P. aeruginosa* strains; PA01 wild type (B.Iglewski) and PAO-JP1 ($\Delta lasI$ mutant)¹ were used in this study. Colonies of the desired strain were grown on a Luria-Bertani (LB) agar plate at 37 °C overnight. The colonies were used to inoculate 10 mL fresh LB media and grown overnight at 37 °C. The overnight cultures were used to inoculate fresh LB media to an initial OD₆₀₀ of 0.05 and cultures were grown with good aeration (300 rpm in an orbital shaker) at 37 °C. Cultures were then used as a source of bacteria for various phenotypic assays. A Jenway 6705 spectrometer 1cm path-length cuvettes were used for all spectrophotometric assays. AHL analogues were stored as 10 mM stock solutions in DMSO at – 20 °C. The AHL analogues in DMSO were added at the desired concentration to the culture at the start of growth. The final DMSO volume remained below 1% and an equivalent volume of DMSO with no AHL analogue was added to control cultures.

Pyocyanin assay

Pyocyanin in the culture supernatant was quantified as previously described.² After growth the OD₆₀₀ was recorded, and the samples were clarified by centrifugation (3,150 xg, 10 min, 20 °C) to remove cell debris. The supernatant (5 mL) was then extracted with chloroform (3 mL) by vortexing. The phases were separated by centrifugation (3,150 xg, 10 min, 20 °C) and the chloroform phase was transferred to a fresh tube and extracted with HCl aq (0.2 N, 1 mL). The phases were separated by centrifugation (3,150 xg, 10 min, 20 °C) and the absorption of the aqueous phase was measured at 520 nm and corrected for the culture OD₆₀₀.

Elastase assay

Elastase in culture supernatant was quantified as previously described,³ with minor modifications. Elastin congo red (5 mg) purchased from Sigma Aldrich was weighed out into 2 mL eppendorf tubes. Elastase congo red buffer (1 mM CaCl₂, 100 mM tris base, pH 7.5) and supernatant (0.1 mL) from the bacterial culture was then added to the eppendorf tube. The resulting suspension was then left to incubate for 13 hours with shaking at 37 °C. The suspension was clarified by centrifugation (3,150 xg, 10 min, 20 °C) and the aqueous phase was separated from the solid residue by pipetting. The absorbance at 492nm was measured and corrected for the culture OD₆₀₀.

References:

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