Supplementary Material

Synthesis and Biological Evaluation of 1,4-Naphthoquinones and Quinoline-5,8-diones as Antimalarial and Schistosomicidal Agent

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15 Enzymatic studies

Enzyme preparation. Recombinant *h*GR was purified as previously reported.¹ One unit of GR activity is defined as the consumption of 1 µmol NADPH per min under conditions of substrate saturation. The enzyme stock solution (266 µm) used for kinetic determinations was > 98% pure as judged from silver stained SDS-PAGE and had specific activities of 200 U/mg (*h*GR). A *h*GR stock II solution was prepared by dilution (1/10) of the enzyme stock solution with *h*GR buffer (pH 6.9).² The *h*GR buffer (pH 6.9) was prepared by dissolving

20 KH₂PO₄ (2.79 g), K₂HPO₄, 3 H₂O (6.04 g), EDTA (0.372 g), and KCl (14.91 g) in 1 L water. The pH was adjusted by dropwise addition of 5 M KOH. Recombinant *Sm*TGR was produced in *Escherichia coli* as previously described.³

Pharmacokinetics

Solubility express. Shake-flask method allowed determination of the apparent thermodynamic solubilities from DMSO stock solutions of compounds. Compound dilutions were prepared to 200 µM in an aqueous 7.4 HEPES buffer. The mix was equilibrated at 20 °C during

- 25 24 hours. After centrifugation, the supernatant was injected in an HPLC column coupled to a LC-MS apparatus (with UV-Vis detection) in order to determine the compound concentration. A reference solution at 200 μM was made in CH₃CN/water and a dilution at 100 μM allowed analysis of the linearity of the UV response. These solutions were also injected in order to calibrate the concentration determination. Comparison of the responses of the injections of three dilutions obtained from a DMSO reference solution allowed determination of the solubility if lower than 200 μM. Measurements were done in duplicate.
- 30 Determination of logD values. The partition coefficient between octanol and water is the most widely used physicochemical determination of lipophilicity. The "shake flask" method was used to determine the partition coefficients. A solution of 100 μM tested compound was prepared from a stock DMSO solution in a "mix" aqueous buffer / octanol. Three octanol / buffer volume ratios were used to cover partition coefficients ranging from -1.5 to +3.2. The partition experience with the best ratio was repeated twice. The mix was equilibrated at 20°C for one hour. Then, the aqueous phase and a reference solution at 100 μM were injected in an HPLC column
- 35 coupled to a LC-MS apparatus (with UV-Vis detection) to determine compound concentration. The partition coefficient was calculated from the peak areas obtained in the chromatograms. Measurements were done in duplicate. For optimal oral absorption, logD values should be between 0.5 and 2.

Determination of CHI values. Compound lipophilicity influences permeability and propensity to nonspecific binding to plasma proteins. Although precise, the classical « shake flask » method is not always appropriate for determination of high hydrophobicity, which can be

- 40 alternatively measured by the Chromatographic Hydrophobicity Index method. This HPLC-based technique leads to rapid and cheap estimates of the partition coefficients. The lipophilicity is determined by partition of the compound between a hydro-organic mobile phase and a reverse stationary phase, typically a C18 column, using a fast gradient. Compound solution samples (200 μM) were prepared in a 50/50 v/v mixture of water and acetonitrile. Five μL were injected onto the HPLC (with UV-Vis detection). CHI values were derived from the retention time and its comparison with 10 reference compounds of known CHI. Measurements were done in duplicate. In
- 45 general, CHI values correlate satisfactorily with logD_{7.4} values. The CHI is approximately equal to the proportion of acetonitrile in the mobile phase when the compound is eluted from the column.

¹ A. Nordhoff, U. S. Bucheler, D. Werner, R. H. Schirmer, *Biochemistry*, 1993, **32**, 4060.

² a) T. Müller, L. Johann, B. Jannack, M. Brückner, D. A. Lanfranchi, H. Bauer, C. Sanchez, V. Yardley, C. Deregnaucourt, J. Schrevel, M. Lanzer, R. H. Schirmer, E. Davioud-Charvet, J. Am. Chem. Soc., 2011, **133**, 11557. b) P. M. Färber, L. D. Arscott, C. H. Williams Jr, K. Becker, R. H. Schirmer, *FEBS Lett.*, 1998, **422**, 311

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