

Electronic Supplementary Information for

Synthesis and Characterization of pH-sensitive, Biotinylated MRI Contrast Agents and Their Conjugates with Avidin

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Table S1. Elution conditions for analytical and semi-preparative HPLC. The flow rates of 1 and 10 mL/min were used for the analytical and semi-preparative HPLC respectively.

Time (minute)	% H ₂ O	% CH ₃ CN
0.0	80	20
5.0	80	20
17.0	20	80
23.0	20	80
26.0	80	20
30.0	80	20

Fluorescence assay

The interaction of biotin (Bio) or **GdL**^{3,4} with avidin (Av) was followed by the displacement of the fluorescence probe 2-anilinonaphthalene-6-sulfonic acid (ANS) using the fluorescence assay as previously described.¹

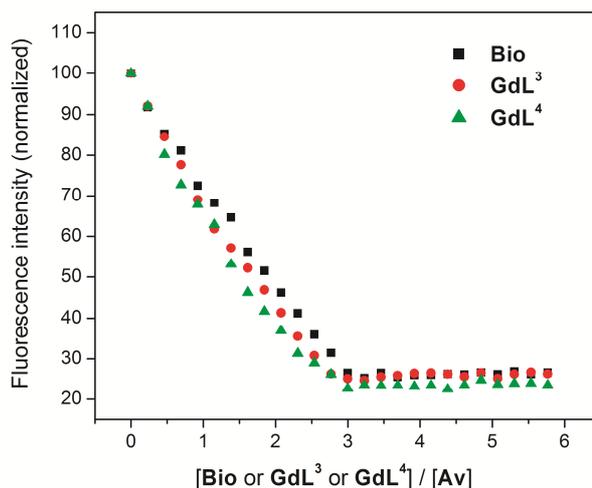
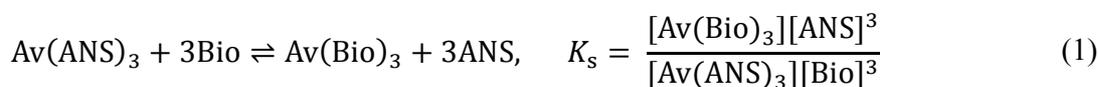
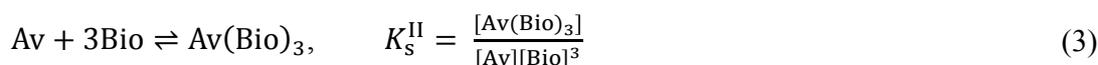
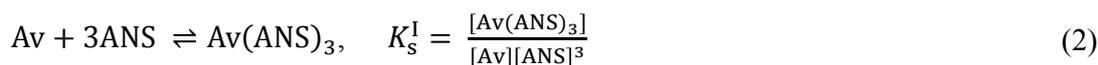


Figure S1. Fluorescence titration of the avidin-ANS complex with biotin and **GdL**^{3,4}. The corrected and normalized fluorescence signal is plotted as a function of the ratio of added biotin/**GdL**^{3,4} to avidin.

Since three, rather than four binding sites are available on the avidin tetramer, this stoichiometry was used to develop a linear relationship used to determine the conditional stability constant value K_s for the reaction of avidin-2,6-ANS complex with biotin (Eq. 1), as well as with **GdL**^{3,4}.



If the basic avidin-ANS and avidin-biotin reactions are considered:



then K_s from Eq. 1 can be rearranged:

$$K_s = \frac{[Av(Bio)_3][ANS]^3}{[Av(ANS)_3][Bio]^3} \times \frac{[Av]}{[Av]} = \frac{K_s^{II}}{K_s^I} \quad (4)$$

to give the linear dependence:

$$\frac{[Av(Bio)_3]}{[Av(ANS)_3]} = K_s \times \frac{[Bio]^3}{[ANS]^3} \quad (5)$$

where $[ANS]$ represents the fixed concentration of ANS (experimental details given in the Experimental section), $[Bio]$ is the concentration of added biotin in each titration point, and $[Av(Bio)_3]$ and $[Av(ANS)_3]$ are the concentrations of avidin-biotin and avidin-ANS complexes, respectively. As these concentrations are proportional to measured fluorescence intensities,² the obtained experimental data (Figure S1) can be used for conditional stability constant determination:

$$\frac{Y_0 - Y}{Y - Y_{lim}} = K_s \times \frac{[Bio]^3}{[ANS]^3} \quad (6)$$

where Y_0 stands for fluorescence intensity at the beginning of titration (no biotin added), Y_{lim} is an average value of all measured fluorescence intensities after the equivalence point was reached, and Y stands for measured fluorescence intensities during the titration (as reaction (1) proceeds). The measured fluorescence intensities were corrected for inner-filter effects³ and normalized. The conditional stability constant values K_s for avidin-biotin, avidin-GdL³ and avidin-GdL⁴ are obtained as slopes from the linear fit based on Eq. 6, with $R^2 > 0.98$ (Figure S2).

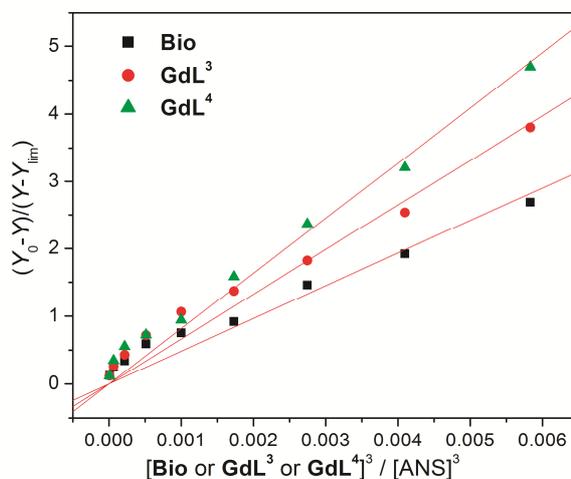


Figure S2. Conditional stability constant (K_s) determination according to Eq. 6.

K_s values are obtained as mean values of five times repeated titrations. Results are shown in Table S2.

Table S2. Conditional stability constant values (K_s) obtained as the slope of the linear fit based on Eq. 6

Sample fluorescence Titration	$K_s \pm SD$		
	Av-Bio	Av-GdL ³	Av-GdL ⁴
1.	302 ± 11	413 ± 24	464 ± 19
2.	417 ± 16	735 ± 30	618 ± 22
3.	527 ± 22	541 ± 21	751 ± 29
4.	379 ± 12	625 ± 23	676 ± 16
5.	487 ± 18	662 ± 33	818 ± 29
$\langle K_s \rangle \pm SD$	$(4.2 \pm 0.9) \times 10^2$	$(6 \pm 1) \times 10^2$	$(7 \pm 1) \times 10^2$

As the dissociation constant of the avidin(monomer)-ANS was already reported ($K_d=203 \mu\text{M}$)^[3], K_s^I value (Eq. 2) is calculated ($K_s^I = \frac{1}{(K_d)^3} = 1.2 \times 10^{11}$), and used for the overall stability

constants K_S^{II} (Eq. 4) calculation, resulting in $K_S^{\text{II}}(\text{Av}(\text{Bio})_3)=5.1 \times 10^{13}$, $K_S^{\text{II}}(\text{Av}(\text{GdL}^3)_3)=7.1 \times 10^{13}$, $K_S^{\text{II}}(\text{Av}(\text{GdL}^4)_3)=8.0 \times 10^{13}$.

MRI phantom experiments

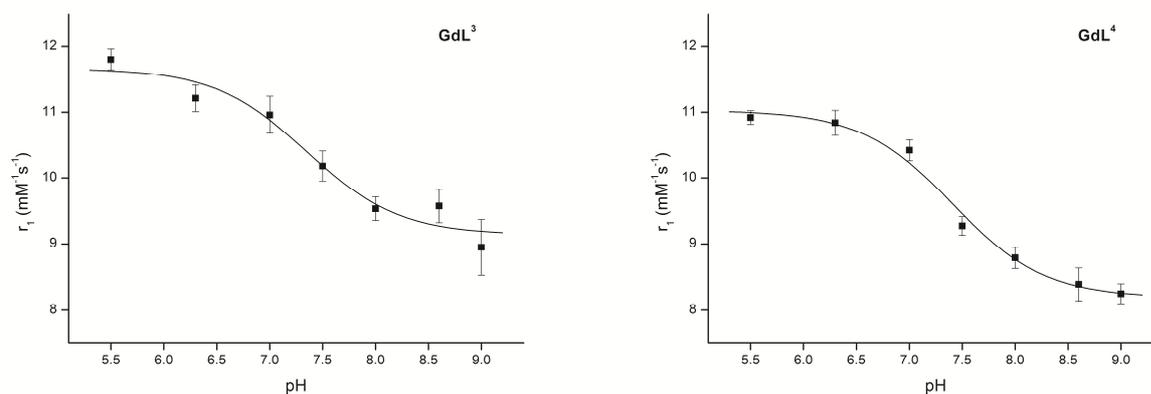


Figure S3. pH dependant r_1 response of GdL^3 (left) and GdL^4 (right) in the absence of avidin (3T MRI scanner, 21 °C). Values are presented as mean \pm SEM of five independent experiments. The lines represent result of the sigmoidal fit and are displayed to aid a better visualization of the pH dependent r_1 decrease.

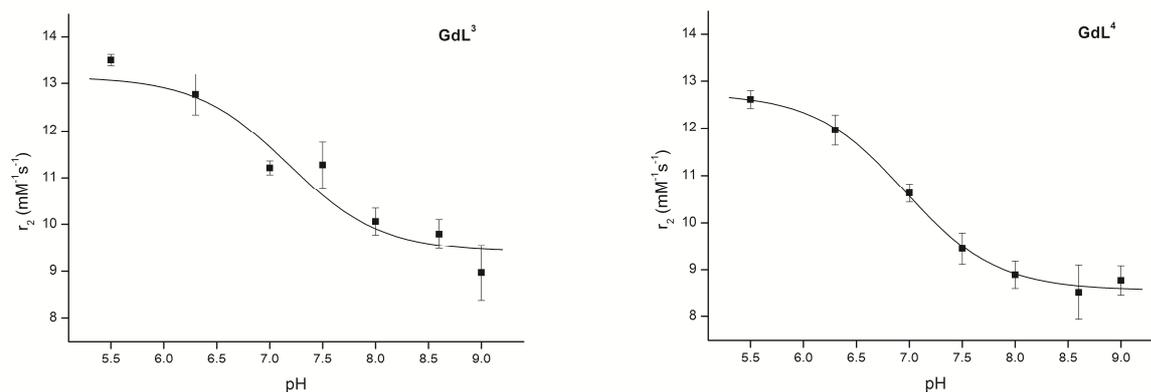


Figure S4. pH dependant r_2 response of GdL^3 (left) and GdL^4 (right) in the absence of avidin (3T MRI scanner, 21 °C). Values are presented as mean \pm SEM of five independent experiments. The lines represent result of the sigmoidal fit and are displayed to aid a better visualization of the pH dependent r_2 decrease.

MRI phantom experiments/E-titrations

The curves obtained from the MRI E-titrations were fitted based on the previously published formula,⁴ however they were slightly modified to use relaxivity values instead of relaxation rates (Eq. 7).

$$r_{1,2}^{obs} = 1000 \times \left\{ (L_0 \times r_{1,2}^f) + 0.5 \times (r_{1,2}^b - r_{1,2}^f) \times \left((n \times c_{Av}) + L_0 + K_a^{-1} - \sqrt{\left((n \times c_{Av}) + L_0 + K_a^{-1} \right)^2 - 4 \times n \times L_0 \times c_{Av}} \right) \right\} \quad (7)$$

where:

$r_{1,2}^{obs}$: observed longitudinal or transversal relaxivity

L_0 : concentration of SCA in M (set to 0.001 M since relaxivities are used)

$r_{1,2}^f$: longitudinal or transversal relaxivity of the free SCA

$r_{1,2}^b$: longitudinal or transversal relaxivity of the avidin bound SCA

n : number of binding sites on the avidin tetramer (set to $n=3$ as obtained from fluorescence displacement assay)

c_{Av} : normalized concentration of avidin in M

K_a : binding constant of respective SCA (set to 7.1×10^{13} and 8.0×10^{13} for **GdL**³ and **GdL**⁴, respectively, as obtained from the fluorescence displacement assay).

References:

1. D. M. Mock, G. Langford, D. Dubois, N. Criscimagna and P. Horowitz, *Anal. Biochem.* 1985, **151**, 178-181.
2. B. Valeur, *Molecular Fluorescence: principles and applications*, Wiley-VCH, Weinheim; Chichester, 2002.
3. D. M. Mock, G. Lankford and P. Horowitz, *Biochim. Biophys. Acta* 1988, **956**, 23-29.
4. T. N. Parac-Vogt, K. Kimpe, S. Laurent, L. Vander Elst, C. Burtea, F. Chen, R. N. Muller, Y. C. Ni, A. Verbruggen and K. Binnemans, *Chem. Eur. J.* 2005, **11**, 3077-3086.